



**UNIVERSITI PUTRA MALAYSIA**

***COMPARATIVE IMMUNOPATHOPHYSIOLOGICAL RESPONSES IN  
MICE FOLLOWING DIFFERENT ROUTE OF INOCULATION OF  
*Brucella melitensis* AND ITS LIPOPOLYSACCHARIDE***

**ABDINASIR YUSUF OSMAN**

**FPV 2016 25**



**COMPARATIVE IMMUNOPATHOPHYSIOLOGICAL RESPONSES IN  
MICE FOLLOWING DIFFERENT ROUTE OF INOCULATION OF  
*Brucella melitensis* AND ITS LIPOPOLYSACCHARIDE**

By

**ABDINASIR YUSUF OSMAN**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,  
in Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

**December 2016**

All materials contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



## DEDICATION

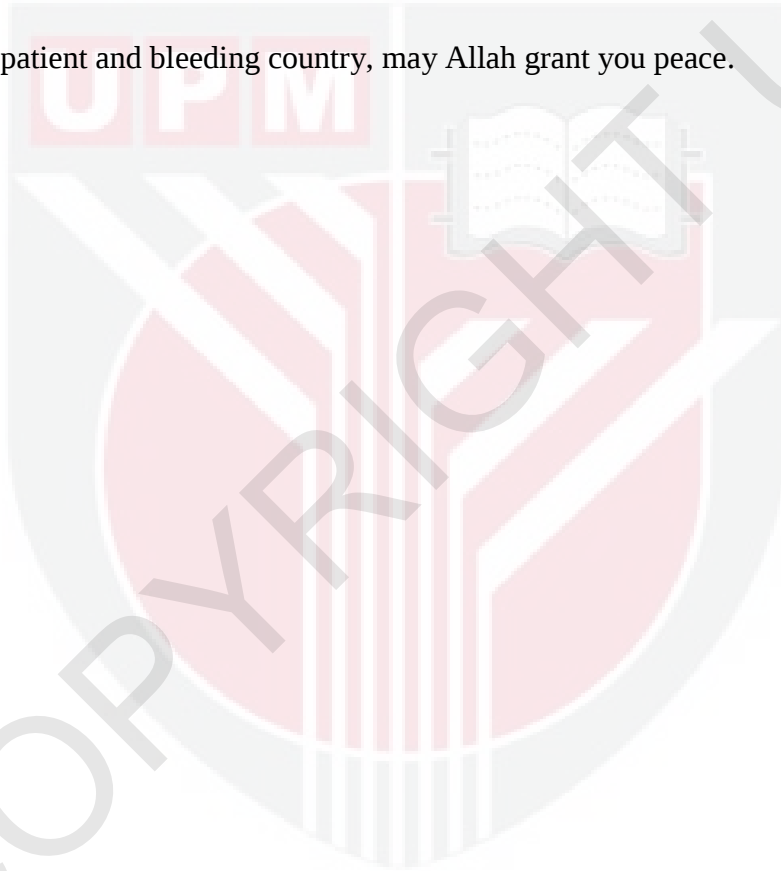
To the Almighty Allah who has been my help, sustainer, provider, guide, encouragement, and my all in all throughout the course of my studies.

To my late father "May Allah blesses him with His supreme benevolence".

To my caring mother and lovely wife who have shown me the unprecedented sacrifice to make sure we reach together the goal of the journey.

To my sister, brothers and all those who passed away in struggle for sovereignty of my fatherland.

To my patient and bleeding country, may Allah grant you peace.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for degree of Doctor of Philosophy

**COMPARATIVE IMMUNOPATHOPHYSIOLOGICAL RESPONSES IN  
MICE FOLLOWING DIFFERENT ROUTE OF INOCULATION OF  
*Brucella melitensis* AND ITS LIPOPOLYSACCHARIDE**

By

**ABDINASIR YUSUF OSMAN**

**December 2016**

**Chairman : Professor Abdul Aziz Saharee, PhD**  
**Faculty : Veterinary Medicine**

*Brucella melitensis*, which causes a small ruminant brucellosis in sheep and goats and Malta fever in humans, is believed to enter the host via ingestion, inhalation or direct contact of the organism with broken skin or mucous membranes. Among the consequences of the different routes of infection are septicaemia, increased permeability of blood vessels and presence of the organism in several organs. However, the oral and the respiratory tract may not be the only portal of entry and route of spread of *B. melitensis*. Circumstantial evidence had suggested the involvement of gastrointestinal, respiratory and reproductive tract in the pathogenesis of *B. melitensis* and its lipopolysaccharide in ruminants. Nevertheless, the pathogenesis and the immunopathophysiology of the disease following different route of infection have not been well documented since previous reports on the disease were limited to incidental observations. The response of gastrointestinal, respiratory, and reproductive tract following oral, intranasal, subcutaneous and intraperitoneal exposure to *B. melitensis* was studied and compared its severity with lipopolysaccharide (LPS) exposure. The cytokine, antibody pattern and sex related hormonal responses following the different route of inoculations to *B. melitensis* and its lipopolysaccharide in mice were also investigated.

The clinical signs observed in these studies include; inappetence, ocular discharge, and ruffled following the different route of exposure to *B. melitensis* and its lipopolysaccharide. Although the severity of the clinical sign varied over time, type of inoculum and route of inoculation, however, mean clinical score were significantly higher in oral and intraperitoneal exposed groups to *B. melitensis* followed by intranasal and subcutaneous groups, respectively. Clinical observations for intranasal and subcutaneous groups were limited mostly to mild and moderate involvement. In contrast to *B. melitensis* infected group, animals challenged with LPS showed mild clinical signs which seemed to be limited in the first 48 h post-

infection. Thereafter, normalization was observed in this group as they were not significantly different from those served as a control group. No significant differences were detected among the different sub groups of LPS infected indicating that the clinical presentation did not differ by route of exposure. Animals in control group did not develop any clinical signs throughout the experimental period.

The pathological alterations varied depending on the route of infection, days post-infection and the type of the organs recorded. Spleen, liver, kidney, lung and the reproductive organs that include uterus, ovary, testes, epididymis and seminal vesicle were the most commonly and severely affected organs with predominance in oral and intraperitoneally infected animals of *B. melitensis* group. These organs presented marked infiltration of inflammatory cells, degeneration, necrosis, haemorrhage and oedema. In intranasal and oral group of *B. melitensis*, lungs were the most affected organ than the other route of infection, with an abundance of fibrin admixed with cellular debris. Emphysema, oedema and marked infiltration of inflammatory cells were also recorded in lungs from 24 hours post-infection until the end point of the experiment. In contrast, histopathological changes of the various organs infected with LPS were almost similar presenting mild degrees of lesion involvement in all routes of infection with special reference in lungs and reticuloendothelial organs. Thus, indicating that LPS have preventive properties toward establishment of pathological lesions. Following the different routes of exposure, *B. melitensis* was isolated from the vital and reproductive organs along with intestinal segments of the mice that developed severe lesions scoring. Higher isolation and detection by PCR was noted predominantly in both reproductive tract and reticuloendothelial-rich organs of oral and intraperitoneal expose groups followed by intranasal and subcutaneous groups to *B. melitensis*, respectively.

Concurrently the cytokine and antibody immune response of mice following different routes of inoculation to *B. melitensis* and its lipopolysaccharide was also evaluated. Both *B. melitensis* and LPS elicited sustained and significantly higher serum IL-1 $\beta$  and IL-6 that has of minor relevance to the route of infection. However, the highest responses were noted in LPS group than *B. melitensis* infected group within the respective route of inoculation. Similarly, the LPS elicited sustained and significantly higher IgM and IgG levels than *B. melitensis* in all different routes of infection. Among the routes of infection, the subcutaneous group yielded highest titers of antibody response followed by intranasal and intraperitoneal groups, respectively. With the presence of severe histopathological evidence along with higher isolation of *B. melitensis* infected group in the reproductive tract, the experiment was conducted to evaluate the serum hormonal changes following different route of exposure to *B. melitensis* and its lipopolysaccharide. Both *B. melitensis* and LPS resulted in significant decrease in the circulating concentrations of serum progesterone, estradiol, and testosterone levels that has significant ( $p<0.05$ ) difference when the effect is compared to those served as a control group.

This study showed that *B. melitensis* organisms were present in various segments and tissues of the gastrointestinal, respiratory, and reproductive tract following the different route of exposure. Therefore, it can be concluded that *B. melitensis*

infection can be transmitted via the gastrointestinal, respiratory and reproductive tract. Oral, intranasal and subcutaneous routes of administration of LPS elicited high serum cytokine and antibody immune response than *B. melitensis* infected group, although the responses of cytokines were variable. Thus, oral, intranasal and subcutaneous infections with  $10^9$  of live *B. melitensis* and its lipopolysaccharide were safer than the intraperitoneal route of inoculation. Both of these routes, in particular subcutaneous route, can be considered as potential alternative route for vaccine administration against *B. melitensis* infection in small ruminants. Similarly, it was concluded that the LPS stimulated significantly the innate and acquired immune system without significant systemic dysfunction, suggesting potentiality of the protective properties of this component as alternative vaccine for brucellosis infection.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan Ijazah Doktor Falsafah

**PERBANDINGAN TINDAK BALAS IMMUNOPATOFISIOLOGIKAL  
DALAM TIKUS BERDASARKAN LALUAN INOKULASI *Brucella melitensis*  
DAN LIPOPOLYSACCHARIDE YANG BERBEZA**

Oleh

**ABDINASIR YUSUF OSMAN**

**Disember 2016**

**Pengerusi : Profesor Abdul Aziz Saharee, PhD**  
**Fakulti : Perubatan Veterinar**

*Brucella melitensis*, yang menyebabkan brucellosis ruminan kecil dalam biri-biri dan kambing serta demam Malta pada manusia, dipercayai memasuki perumah melalui penghadaman, menghidu atau hubungan secara langsung organisma dengan luka pada kulit atau membran mukus. Di antara kesan jangkitan dari laluan yang berbeza adalah septisemia, kebolehtelapan saluran darah dan kehadiran organisma dalam beberapa organ-organ. Walau bagaimanapun, mulut dan saluran pernafasan adalah bukan satu-satunya portal kemasukan dan laluan penyebaran *B. melitensis*. Bukti dari keadaan semasa telah mencadangkan penglibatan saluran pencernaan, pernafasan dan saluran pembiakan dalam patogenesis *B. melitensis* dan lipopolysaccharide dalam ruminan. Walau bagaimanapun, patogenesis dan penyakit immunopatofisiologi penyakit ini berdasarkan laluan jangkitan yang berbeza masih belum didokumenkan atas sebab laporan-laporan penyakit yang terdahulu adalah terhad kepada pemerhatian yang berlaku secara tidak tiba-tiba. Tindak balas pendedahan saluran pencernaan, pernafasan, dan saluran pembiakan diikuti oral, intranasal, subkutaneus dan intraperitoneal kepada *B. melitensis* telah dikaji dan dibandingkan darjah keterukan dengan pendedahan lipopolysaccharide (LPS). Cytokine, corak antibodi dan tindakbalas hormon berkaitan seks berdasarkan laluan inokulasi *B. melitensis* dan lipopolysaccharide yang berbeza dalam mencit juga telah dikaji.

Tanda-tanda klinikal yang diperhatikan dalam kajian ini termasuk; kurang selera makan, penghasilan lelehan dari mata serta bulu haiwan yang tidak terurus berdasarkan pendedahan kepada *B. melitensis* dan lipopolysaccharide melalui saluran yang berbeza. Walaupun darjah keterukan tanda klinikal berubah dari masa ke masa selain jenis serta laluan inokulum, walau bagaimanapun, skor min klinikal adalah lebih tinggi dalam kumpulan oral dan intraperitoneal yang terdedah kepada *B. melitensis* diikuti dengan kumpulan intranasal dan subkutaneus. Pemerhatian klinikal

untuk intranasal dan kumpulan subkutaneus adalah terhad bagi kebanyakan penglibatan yang ringan dan sederhana. Sebaliknya, haiwan yang dijangkiti dengan LPS menunjukkan tanda-tanda klinikal yang sederhana yang mana ianya terhad dalam tempoh 48 jam selepas jangkitan. Sejurus itu, normalisasi diperhatikan berlaku dalam kumpulan ini kerana mereka tidak ketara berbeza daripada kumpulan kawalan. Tiada perbezaan yang signifikan telah dikesan di kalangan kumpulan sub berbeza dijangkiti LPS membuktikan kesan klinikal adalah tidak berbeza berdasarkan laluan pendedahan. Haiwan dalam kumpulan kawalan tidak menunjukkan apa-apa tanda-tanda klinikal sepanjang tempoh eksperimen.

Perubahan-perubahan patologi adalah berbeza-beza bergantung pada laluan jangkitan, bilangan hari selepas jangkitan dan jenis organ yang telah direkodkan. Limpa, hati, buah pinggang, paru-paru serta organ-organ pembiakan termasuklah rahim, ovari, testis, epididimis dan vesikel seminal adalah organ yang paling kerap terjejas teruk yang didominasi oleh haiwan dalam kumpulan yang telah dijangkit melalui laluan oral dan intraperitoneal. Organ-organ ini menunjukkan tingginya kehadiran sel radang, kemerosotan, nekrosis, pendarahan dan edema. Dalam kumpulan *B. melitensis* melalui intranasal dan oral, paru-paru adalah organ yang paling terjejas berbanding jangkitan melalui laluan yang lain, dengan kehadiran fibrin bercampur dengan serpihan selular. Emfisema, edema dan penyusupan sel-sel radang juga telah direkodkan dalam paru-paru dalam tempoh 24 jam selepas jangkitan sehingga titik akhir eksperimen. Sebaliknya, perubahan histopatologi pelbagai organ-organ dijangkiti LPS adalah hampir sama dengan penglibatan lesi secara sederhana dalam semua laluan jangkitan dengan rujukan khas dalam paru-paru dan organ-organ reticulo-endotelial. Oleh itu, ini menunjukkan bahawa LPS mempunyai ciri-ciri pencegahan kearah penghasilan lesi patologi. Berikutan pendedahan laluan yang berbeza, *B. melitensis* telah diasingkan daripada organ-organ penting dan organ pembiakan bersama-sama dengan segmen usus mencit yang menunjukkan markah lesi yang teruk. Pengasingan yang lebih tinggi dan pengesanan oleh PCR telah dikenalpasti di peringkat awal dominasi bagi kedua-dua saluran reproduksi dan organ-organ yang kaya dengan reticuloendotelial dari kumpulan yang terdedah secara oral dan intraperitoneal diikuti oleh masing-masing kumpulan intranasal dan subkutaneus untuk *B. melitensis*.

Serentak dengan itu cytokine dan tindak balas imun antibodi dalam mencit berikutan laluan inokulasi *B. melitensis* dan lipopolysaccharide yang berbeza juga turut dinilai. Kedua-dua *B. melitensis* dan LPS turut mengalami penghasilan serum IL-1 $\beta$  dan IL-6 yang lebih tinggi yang mempunyai kesan yang sedikit kepada laluan jangkitan. Walau bagaimanapun, tindak balas tertinggi diperhatikan dalam kumpulan LPS berbanding kumpulan yang dijangkiti *B. melitensis* bagi aspek laluan inokulasi. Begitu juga, LPS didapati mengalami tahap IgM dan IgG jauh lebih tinggi berbanding *B. melitensis* dalam kesemua laluan jangkitan yang berbeza. Di antara laluan jangkitan tersebut, kumpulan subkutaneus menghasilkan tindak balas antibodi titer tertinggi diikuti masing-masing oleh kumpulan intranasal dan intraperitoneal. Dengan kehadiran bukti histopatologi yang teruk bersama-sama dengan pengasingan *B. melitensis* yang lebih tinggi bagi kumpulan dijangkiti dalam saluran pembiakan, eksperimen tersebut telah dijalankan untuk menilai perubahan hormon serum berdasarkan laluan pendedahan kepada *B. melitensis* dan lipopolysaccharide yang

berbeza. Kedua-dua *B. melitensis* dan LPS menyebabkan penurunan ketara dalam tahap kepekatan serum progesteron, estradiol, dan testosteron yang hanya mempunyai perbezaan yang ketara apabila kesan itu dibandingkan dengan kumpulan kawalan.

Kajian ini telah menunjukkan bahawa organisma *B. melitensis* hadir dalam pelbagai segmen dan tisu saluran pencernaan, pernafasan dan saluran pembiakan berikutan laluan pendedahan yang berbeza. Oleh itu, dapat disimpulkan bahawa jangkitan *B. melitensis* boleh berlaku melalui saluran pencernaan, pernafasan dan pembiakan. Laluan kemasukan LPS secara oral, intranasal dan laluan subkutaneus menghasilkan tinggi cytokine dalam serum dan tindak balas imun antibodi berbanding kumpulan yang dijangkiti dengan *B. melitensis*, walaupun tindak balas cytokines tersebut adalah berbeza-beza. Oleh itu, jangkitan *B. melitensis* dan lipopolysaccharide dengan dos yang besar secara oral, intranasal dan jangkitan subkutaneus adalah lebih selamat daripada laluan inokulasi intraperitoneal. Kedua-dua laluan ini boleh dianggap sebagai laluan alternatif yang berpotensi bagi peberian vaksin terhadap jangkitan *B. melitensis* ruminan kecil. Begitu juga, dapat disimpulkan bahawa LPS dapat merangsang sistem imun secara semula jadi dan imun yang diperlukan tanpa kegagalan fungsi sistemik yang ketara, menunjukkan potensi sebagai pelindung oleh komponen ini sebagai alternatif kepada pengahslan vaksin bagi jangkitan brusellosis.

## ACKNOWLEDGEMENTS

All praises are due to ALLAH, lord of the world for the abundant privileges too numerous to mention and the strength to undergo a training of the mind (Ph.D program).

I wish to sincerely acknowledge the advisory and supervisory guidance of the chair of my committee, Prof. Dr. Abdul Aziz Saharee for his unique style and good research direction, and to Associate Prof. Dr. Faez Firdaus Jesse Abdullah and Assoc. Prof. Dr. Arifah Abdul Kadir for their understanding and constructive criticism right from the conception, through execution to completion of the research. You all remain accessible at all times in the course of my studies; I will forever remain indebted to you.

I would use this opportunity to acknowledge the technical assistance of the following people; who assisted in animal handling and post-mortem, Eng. Liban Mohamed Dado, Dr. Yusuf Abba, Dr. Baba Jalo, Eng. Abdikani Abdullah, Mr. Abdirashid Africa ; In histopathology, Puan (Mrs) Jamilah Jahari, Puan (Mrs) Latifah Mohd Hanan; In PCR analysis, Dr. Kontoh Mohammad, Dr. Bodhrus, Mr. Azalan; in serum analysis; Mr. Yap; Dr. Eric Lim, and Mrs. Amirah; Assoc. Prof.Dr.Goh Yong Meng and Prof. DR. Mohamed Ariff Omar for their guidance in statistical analysis; Prof. Dr. Saleha Abdul Aziz and Haryanti Azura Mohd Wali for their technical assistance with abstract translation to Bahasa Melayu.

To the gratitude of humanity and support of the staff and management of Hospital Pantai Kuala Lumpur, Malaysia who saved the life of my wife. You changed my life through learning that I must not lose faith in humanity. I will forever remain indebted to you.

Special thanks goes to Dr. Panarama and Puan (Mrs) Victoria from Hospital Pantai Kuala Lumpur who are the great examples of humanity. With you, the humanity survives and serves in all its dimensions. As humans, we must love and serve one another to promote the welfare and the stability of society.

To my parents from whom, I learnt hard work and being independent. They have continued to support my course with untiring love. With you around, I feel stable emotionally, psychologically and financially throughout the journey. To my siblings for the consistent calls and concern all through, you all continue to inspired me and keep my spirit high all along my Ph.D Journey.

To my wife for unparalleled sacrifice shown for abandoning her medical studies in Somalia to make sure we raise together our most cherish divine gifts (Shazreena

Abdinasir Yusuf). This concern and many more commitments showed, re-kindled and boost my spirit and determination to succeed.

Lastly, I will especially once more express my profound gratitude to School of Graduate Studies, UPM for the offer of International Graduate Research assistance (IGRF) and all my supervisory committee for the funding the entire Ph.D project.



I certify that a Thesis Examination Committee has met on 2016 to conduct the final examination of Abdinasir Yusuf Osman on his PhD thesis entitled “Comparative immunopathophysiological responses in mice following different routes of inoculation to *Brucella melitensis* and its lipopolysaccharide” in Accordance with the Universities and University College Act 1971 and the constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Examinations Committee were as follows:

**Abdul Wahid Haron, PhD**

Professor  
Faculty of veterinary medicine  
Universiti Putra Malaysia  
(Chairman)

Hazilawati Hamzah, PhD  
Associate Professor  
Faculty of veterinary medicine  
Universiti Putra Malaysia  
(Internal Examiner)

Sabri Mohd Yusoff, PhD  
Associate Professor  
Faculty of veterinary medicine  
Universiti Putra Malaysia  
(Internal Examiner)

Hussni Omar Mohammed  
Professor  
College of Veterinary Medicine  
Cornell University  
Ny 14853-6401 Ithaca, USA

---

**NORITAH OMAR, PhD**

Associate Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: December, 2016

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No: Abdinasir Yusuf Osman, GS36694

## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: \_\_\_\_\_  
Name of  
Chairman of  
Supervisory  
Committee: \_\_\_\_\_

Signature: \_\_\_\_\_  
Name of  
Member of  
Supervisory  
Committee: \_\_\_\_\_

Signature: \_\_\_\_\_  
Name of  
Member of  
Supervisory  
Committee: \_\_\_\_\_

## TABLE OF CONTENTS

|                                                                      | <b>Page</b> |
|----------------------------------------------------------------------|-------------|
| <b>ABSTRACT</b>                                                      | i           |
| <b>ABSTRAK</b>                                                       | iv          |
| <b>ACKNOWLEDGEMENTS</b>                                              | vii         |
| <b>APPROVAL</b>                                                      | ix          |
| <b>DECLARATION</b>                                                   | xi          |
| <b>LIST OF TABLES</b>                                                | xvii        |
| <b>LIST OF FIGURES</b>                                               | xviii       |
| <b>LIST OF ABBREVIATIONS</b>                                         | xxv         |
| <br><b>CHAPTER</b>                                                   |             |
| <b>1 INTRODUCTION</b>                                                | <b>1</b>    |
| 1.1 Research hypotheses                                              | 3           |
| 1.2 Objectives of the study                                          | 3           |
| <br><b>2 LITERATURE REVIEW</b>                                       | <b>4</b>    |
| 2.1 The organism                                                     | 4           |
| 2.1.1 A historical overview                                          | 4           |
| 2.1.2 Bacteriological characteristics and taxonomy                   | 4           |
| 2.1.3 Major virulence factors                                        | 5           |
| 2.2 Epidemiology                                                     | 6           |
| 2.2.1 <i>B. melitensis</i> infection in livestock population         | 6           |
| 2.2.2 Species susceptibility to <i>Brucella melitensis</i> infection | 6           |
| 2.2.3 Global distribution and economic impact                        | 7           |
| 2.2.4 Zoonotic aspects of <i>Brucella melitensis</i> infection       | 7           |
| 2.2.5 Mode of transmission and clinical presentation                 | 9           |
| 2.3 Pathogenesis of <i>Brucella melitensis</i>                       | 10          |
| 2.3.1 Reproductive tract localization                                | 11          |
| 2.3.2 Placentitis and abortions                                      | 12          |
| 2.4 Pathological changes                                             | 12          |
| 2.5 Routes of infection in experimental studies                      | 15          |
| 2.6 Immune response to <i>Brucella melitensis</i> infection          | 16          |
| 2.6.1 Humoral response                                               | 16          |
| 2.6.2 Cytokine response                                              | 18          |
| 2.6.3 Steroid hormones, Biosynthesis and excretion                   | 21          |
| 2.6.4 Steroid Biosynthesis                                           | 22          |
| 2.6.5 Steroid Inactivation and Excretion                             | 23          |
| 2.7 Diagnosis of <i>Brucella melitensis</i>                          | 23          |
| 2.7.1 Serological tests                                              | 23          |
| 2.7.2 PCR Identification                                             | 24          |
| 2.7.3 Determination of cytokines                                     | 25          |
| 2.7.4 Determination of steroid hormones                              | 25          |

|          |                                                                                                                                                                              |           |
|----------|------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|-----------|
| <b>3</b> | <b>IMMUNOPATHOPHYSIOLOGICAL RESPONSES<br/>INDUCED BY <i>Brucella melitensis</i> AND ITS<br/>LIPOPOLYSACCHARIDE VIA INTRAPERITONEAL<br/>ROUTE OF INFECTION IN MOUSE MODEL</b> | <b>27</b> |
| 3.1      | Introduction                                                                                                                                                                 | 27        |
| 3.2      | Materials and Methods                                                                                                                                                        | 28        |
| 3.2.1    | Ethics statement                                                                                                                                                             | 28        |
| 3.2.2    | Animals                                                                                                                                                                      | 28        |
| 3.2.3    | Experimental procedure                                                                                                                                                       | 28        |
| 3.2.4    | Synchronization                                                                                                                                                              | 29        |
| 3.2.5    | Bacterial strain and media                                                                                                                                                   | 29        |
| 3.2.6    | Inoculum preparation of <i>Brucella melitensis</i>                                                                                                                           | 29        |
| 3.2.7    | LPS extraction from <i>B. melitensis</i>                                                                                                                                     | 29        |
| 3.2.8    | Route of exposure                                                                                                                                                            | 30        |
| 3.2.9    | Clinical observation                                                                                                                                                         | 30        |
| 3.2.10   | Histopathological examination                                                                                                                                                | 30        |
| 3.2.11   | Bacteriology                                                                                                                                                                 | 31        |
| 3.2.12   | Polymerase Chain reaction (PCR)                                                                                                                                              | 31        |
| 3.2.13   | Cytokine quantification                                                                                                                                                      | 31        |
| 3.2.14   | Antibody quantification                                                                                                                                                      | 32        |
| 3.2.15   | Hormone analysis                                                                                                                                                             | 33        |
| 3.2.16   | Statistical analysis                                                                                                                                                         | 34        |
| 3.3      | Results                                                                                                                                                                      | 34        |
| 3.3.1    | Clinical observation                                                                                                                                                         | 34        |
| 3.3.2    | Mortality rate                                                                                                                                                               | 35        |
| 3.3.3    | Histopathology                                                                                                                                                               | 36        |
| 3.3.4    | Pro-inflammatory cytokine analysis                                                                                                                                           | 45        |
| 3.3.5    | Antibody Analysis                                                                                                                                                            | 47        |
| 3.3.6    | Hormone analysis                                                                                                                                                             | 49        |
| 3.3.7    | Bacteriological result                                                                                                                                                       | 52        |
| 3.4      | Discussion                                                                                                                                                                   | 53        |
| <b>4</b> | <b>IMMUNOPATHOPHYSIOLOGICAL RESPONSES IN<br/>MICE FOLLOWING ORAL INFECTION TO <i>B. melitensis</i><br/>AND ITS LIPOPOLYSACCHARIDE</b>                                        | <b>57</b> |
| 4.1      | Introduction                                                                                                                                                                 | 57        |
| 4.2      | Materials and Methods                                                                                                                                                        | 58        |
| 4.2.1    | Ethics statement                                                                                                                                                             | 58        |
| 4.2.2    | Animals                                                                                                                                                                      | 58        |
| 4.2.3    | Experimental Design                                                                                                                                                          | 58        |
| 4.2.4    | Synchronization                                                                                                                                                              | 58        |
| 4.2.5    | Inoculum preparation of <i>Brucella melitensis</i>                                                                                                                           | 58        |
| 4.2.6    | LPS extraction from <i>B. melitensis</i>                                                                                                                                     | 59        |
| 4.2.7    | Route of exposure                                                                                                                                                            | 59        |
| 4.2.8    | Clinical observation                                                                                                                                                         | 59        |
| 4.2.9    | Histopathology and lesion scoring                                                                                                                                            | 59        |
| 4.2.10   | Bacteriology                                                                                                                                                                 | 60        |
| 4.2.11   | Cytokine quantification                                                                                                                                                      | 60        |
| 4.2.12   | Antibody quantification                                                                                                                                                      | 61        |
| 4.2.13   | Hormone analysis                                                                                                                                                             | 61        |

|        |                                                                                                                                              |     |
|--------|----------------------------------------------------------------------------------------------------------------------------------------------|-----|
| 4.2.14 | Statistical analysis                                                                                                                         | 62  |
| 4.3    | Results                                                                                                                                      | 62  |
| 4.3.1  | Clinical observation                                                                                                                         | 62  |
| 4.3.2  | Mortality rate                                                                                                                               | 63  |
| 4.3.3  | Histopathology                                                                                                                               | 63  |
| 4.3.4  | Pro-inflammatory cytokine analysis                                                                                                           | 71  |
| 4.3.5  | Antibody analysis                                                                                                                            | 73  |
| 4.3.6  | Hormone analysis                                                                                                                             | 75  |
| 4.3.7  | Bacteriological result                                                                                                                       | 78  |
| 4.4    | Discussion                                                                                                                                   | 80  |
| 5      | <b>IMMUNOPATHOPHYSIOLOGICAL RESPONSES IN MOUSE MODEL FOLLOWING INTRANASAL INFECTION BY <i>B. melitensis</i> AND ITS LIPOPOLYSACCHARIDE</b>   | 84  |
| 5.1    | Introduction                                                                                                                                 | 84  |
| 5.2    | Materials and Methods                                                                                                                        | 85  |
| 5.2.1  | Ethics statement                                                                                                                             | 85  |
| 5.2.2  | Animals                                                                                                                                      | 85  |
| 5.2.3  | Experimental Design                                                                                                                          | 85  |
| 5.2.4  | Synchronization                                                                                                                              | 86  |
| 5.2.5  | Bacterial strain and media                                                                                                                   | 86  |
| 5.2.6  | Inoculum preparation of <i>Brucella melitensis</i>                                                                                           | 86  |
| 5.2.7  | Extraction of lipopolysaccharide (LPS)                                                                                                       | 86  |
| 5.2.8  | Route of exposure                                                                                                                            | 86  |
| 5.2.9  | Clinical observation                                                                                                                         | 87  |
| 5.2.10 | Histopathological examination                                                                                                                | 87  |
| 5.2.11 | Bacteriology                                                                                                                                 | 87  |
| 5.2.12 | Cytokine quantification                                                                                                                      | 87  |
| 5.2.13 | Antibody quantification                                                                                                                      | 88  |
| 5.2.14 | Hormone analysis                                                                                                                             | 88  |
| 5.2.15 | Data analysis                                                                                                                                | 88  |
| 5.3    | Results                                                                                                                                      | 88  |
| 5.3.1  | Clinical observation                                                                                                                         | 88  |
| 5.3.2  | Mortality rate                                                                                                                               | 89  |
| 5.3.3  | Histopathology                                                                                                                               | 90  |
| 5.3.4  | Pro-inflammatory cytokine analysis                                                                                                           | 96  |
| 5.3.5  | Antibody analysis                                                                                                                            | 98  |
| 5.3.6  | Hormone analysis                                                                                                                             | 100 |
| 5.3.7  | Bacteriological result                                                                                                                       | 103 |
| 5.4    | Discussion                                                                                                                                   | 104 |
| 6      | <b>IMMUNOPATHOPHYSIOLOGICAL RESPONSES IN MOUSE MODEL FOLLOWING SUBCUTANEOUS INFECTION BY <i>B. melitensis</i> AND ITS LIPOPOLYSACCHARIDE</b> | 108 |
| 6.1    | Introduction                                                                                                                                 | 108 |
| 6.2    | Materials and Methods                                                                                                                        | 109 |
| 6.2.1  | Ethics statement                                                                                                                             | 109 |
| 6.2.2  | Animals                                                                                                                                      | 109 |

|          |                                                       |            |
|----------|-------------------------------------------------------|------------|
| 6.2.3    | Experimental Design                                   | 109        |
| 6.2.4    | Synchronization                                       | 110        |
| 6.2.5    | Bacterial strain and media                            | 110        |
| 6.2.6    | Inoculum preparation of <i>Brucella melitensis</i>    | 110        |
| 6.2.7    | LPS extraction                                        | 110        |
| 6.2.8    | Route of exposure                                     | 110        |
| 6.2.9    | Clinical observation                                  | 110        |
| 6.2.10   | Histopathological examination                         | 111        |
| 6.2.11   | Bacteriology                                          | 111        |
| 6.2.12   | Cytokine quantification                               | 111        |
| 6.2.13   | Antibody quantification                               | 112        |
| 6.2.14   | Hormone analysis                                      | 112        |
| 6.2.15   | Statistical analysis                                  | 113        |
| 6.3      | Results                                               | 113        |
| 6.3.1    | Clinical observation                                  | 113        |
| 6.3.2    | Histopathology                                        | 114        |
| 6.3.3    | Pro-inflammatory cytokine analysis                    | 122        |
| 6.3.4    | Antibody analysis                                     | 124        |
| 6.3.5    | Hormone analysis                                      | 126        |
| 6.3.6    | Bacteriological result                                | 129        |
| 6.4      | Discussion                                            | 130        |
| <b>7</b> | <b>GENERAL DISCUSSION</b>                             | <b>134</b> |
| <b>8</b> | <b>SUMMARY, COMCLUSSION AND FUTURE RECOMMENDATION</b> | <b>142</b> |
|          | <b>REFERENCES</b>                                     | <b>144</b> |
|          | <b>APPENDICES</b>                                     | <b>168</b> |
|          | <b>BIODATA OF STUDENT</b>                             | <b>237</b> |
|          | <b>LIST OF PUBLICATIONS</b>                           | <b>238</b> |

## LIST OF TABLES

| Table |                                                                                                                                                            | Page |
|-------|------------------------------------------------------------------------------------------------------------------------------------------------------------|------|
| 3.1   | Mean ( $\pm$ SD) Clinical score following intraperitoneal infection with <i>B. melitensis</i> and its lipopolysaccharide (LPS) in mice                     | 35   |
| 3.2   | Percentage of the animals died among the challenged groups following intraperitoneal infection with $10^9$ <i>B. melitensis</i> and its lipopolysaccharide | 35   |
| 3.3   | Mean ( $\pm$ SEM) histopathological score following intraperitoneal infection with <i>B. melitensis</i> and its lipopolysaccharide (LPS) in mice           | 43   |
| 3.4   | Bacterial distribution in mice following intraperitoneal exposure to <i>B. melitensis</i>                                                                  | 53   |
| 4.1   | Mean ( $\pm$ SD) Clinical score following oral infection with <i>B. melitensis</i> and its lipopolysaccharide (LPS) in mice                                | 62   |
| 4.2   | Percentage of the animals died among the treatment groups following oral infection with $10^9$ <i>B. melitensis</i> and its lipopolysaccharide             | 63   |
| 4.3   | Mean ( $\pm$ SEM) histopathological score following oral infection with <i>B. melitensis</i> and its lipopolysaccharide (LPS) in mice                      | 69   |
| 4.4   | Bacterial distribution in mice following oral exposure to <i>B. melitensis</i>                                                                             | 79   |
| 5.1   | Mean ( $\pm$ SD) Clinical score following intranasal infection with <i>B. melitensis</i> and its lipopolysaccharide (LPS) in mice                          | 89   |
| 5.2   | Percentage of the animals died among the groups following intranasal infection with $10^9$ <i>B. melitensis</i> and its lipopolysaccharide                 | 89   |
| 5.3   | Mean ( $\pm$ SEM) histopathological score following intranasal infection with <i>B. melitensis</i> and its lipopolysaccharide (LPS) in mice                | 90   |
| 5.4   | Bacterial distribution in mice following intranasal exposure to <i>B. melitensis</i>                                                                       | 104  |
| 6.1   | Mean ( $\pm$ SD) Clinical score following subcutaneous infection with <i>B. melitensis</i> and its lipopolysaccharide (LPS) in mice                        | 113  |
| 6.2   | Percentage of the animals died following subcutaneous infection with $10^9$ <i>B. melitensis</i> and its lipopolysaccharide                                | 114  |
| 6.3   | Mean ( $\pm$ SEM) histopathological score following subcutaneous infection with <i>B. melitensis</i> and its lipopolysaccharide (LPS) in mice              | 120  |
| 6.4   | Bacterial distribution in mice following subcutaneous exposure to <i>B. melitensis</i> .                                                                   | 130  |

## LIST OF FIGURES

| Figure |                                                                                                                                                                                                                                                                          | Page |
|--------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|------|
| 3.1    | Photomicrograph of liver section showing cellular infiltration (long arrow) and haemorrhage (arrows) in intraperitoneally infected mouse with <i>B. melitensis</i> at day 10 post-infection. (Bar=50µm, H&E X100)                                                        | 37   |
| 3.2    | Spleen of mouse infected intraperitoneally with 10 <sup>9</sup> of <i>B. melitensis</i> showing infiltration of inflammatory cells in the white pulp (long arrow) and haemorrhage (arrow) surrounding the splenic follicle at day 3 post-infection. (Bar=100µm, H&Ex200) | 37   |
| 3.3    | Lung of mouse infected intraperitoneally with 10 <sup>9</sup> of <i>B. melitensis</i> showing interstitial infiltration of inflammatory cells (arrow) and congestion (arrow) at day 17 post-infection. Original magnification (H&E X200)                                 | 38   |
| 3.4    | Kidney of mouse infected intraperitoneally with 10 <sup>9</sup> <i>B. melitensis</i> presenting congestion (long arrow), degeneration (arrow) at day 3 post-infection. Original magnification (H&E X200)                                                                 | 38   |
| 3.5    | Intestine of mouse infected intraperitoneally with 10 <sup>9</sup> of <i>B. melitensis</i> presenting infiltration of inflammatory with disruption of epithelial lining (arrow) at day 17 post-infection. Original magnification (H&E X200).                             | 39   |
| 3.6    | Photomicrograph of the brain showing congestion (arrow) and degeneration of pyramidal neurons (head of arrow) in intraperitoneally infected mouse with <i>B. melitensis</i> at day 24 post-infection. (Bar=100µm, H&E X400)                                              | 39   |
| 3.7    | Photomicrograph of the uterus with congestion (long arrow), degeneration (arrow), and infiltration of inflammatory cells (head of arrow) in animals infected intraperitoneally with <i>B. melitensis</i> at day 17 post-infection. (Bar=100µm, H&E X400)                 | 40   |
| 3.8    | Photomicrograph of necrotic orchitis with diffuse lympho-neutrophilic infiltrations (arrows) in animals infected intraperitoneally with <i>B. melitensis</i> at day 24 post-infection. Original magnification (H&E X100)                                                 | 40   |
| 3.9    | Liver of mouse infected intraperitoneally with lipopolysaccharide extracted from <i>B. melitensis</i> presenting congestion (long arrow), haemorrhage (arrows), degeneration and necrosis (head of arrow) at day 17 post-infection. (Bar=100µm, H&E X400)                | 41   |

|      |                                                                                                                                                                                                                             |    |
|------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|----|
| 3.10 | Photomicrograph of the heart with haemorrhage (arrow), as seen in animals infected intraperitoneally with lipopolysaccharide extracted from <i>B. melitensis</i> at day 10 post-infection. (Bar=100µm, H&E X400)            | 41 |
| 3.11 | Photomicrograph of the pituitary gland showing normal histologic features in animals infected intraperitoneally with lipopolysaccharide extracted from <i>B. melitensis</i> at day 17 post-infection. (Bar=100µm, H&E X200) | 42 |
| 3.12 | Photomicrograph of convoluted seminiferous tubules showing mild congestion in animals infected intraperitoneally with LPS at day 24 post-infection. Original magnification (H&E X100)                                       | 42 |
| 3.13 | Comparison of Interleukin 1-β serum profiles (Mean±SEM) in mice following intraperitoneal infection with <i>B. melitensis</i> and its lipopolysaccharide                                                                    | 45 |
| 3.14 | Comparison of Interleukin-6 serum profiles (Mean±SEM) in mice following intraperitoneal infection with <i>B. melitensis</i> and its lipopolysaccharide                                                                      | 46 |
| 3.15 | Comparison of IgM serum profiles (Mean±SEM) in mice following intraperitoneal infection with 10 <sup>9</sup> of <i>B. melitensis</i> and its lipopolysaccharide                                                             | 47 |
| 3.16 | Comparison of IgG serum profiles (Mean±SEM) in mice following intraperitoneal infection with 10 <sup>9</sup> of <i>B. melitensis</i> and its lipopolysaccharide                                                             | 48 |
| 3.17 | Comparison of progesterone serum profiles (Mean±SEM) in mice following intraperitoneal infection with <i>B. melitensis</i> and its lipopolysaccharide                                                                       | 49 |
| 3.18 | Comparison of estradiol serum profiles (Mean±SEM) in mice following intraperitoneal infection with 10 <sup>9</sup> of <i>B. melitensis</i> and its lipopolysaccharide                                                       | 50 |
| 3.19 | Comparison of Testosterone serum profiles (Mean±SEM) in mice following intraperitoneal infection with 10 <sup>9</sup> of <i>B. melitensis</i> and its lipopolysaccharide                                                    | 51 |
| 3.20 | Gel photograph of PCR products showing bands characteristic of <i>B. melitensis</i> . The presence of bands~ 252 bp are indicative of positive results                                                                      | 52 |
| 4.1  | Photomicrograph of lung section showing cellular infiltration (long arrow) and congestion (arrows) in orally infected mouse with <i>B. melitensis</i> at day 17 post-infection. (Bar=100µm, H&E X200)                       | 64 |

|      |                                                                                                                                                                                                                                                   |    |
|------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|----|
| 4.2  | Photomicrograph of spleen section showing cellular infiltration (long arrow) and congestion (arrows) in orally infected mouse with <i>B. melitensis</i> at day 17 post-infection. (Bar=50µm, H&E X400)                                            | 65 |
| 4.3  | Photomicrograph of liver section showing cellular infiltration (yellow long arrow), congestion (arrows) and degeneration (arrows) in orally infected mouse with <i>B. melitensis</i> at day 17 post-infection. (Bar=50µm, H&E X400)               | 65 |
| 4.4  | Photomicrograph of kidney showing congestion (long arrow) and degeneration (arrow) in orally infected mouse with <i>B. melitensis</i> at day 10 post-infection. (Bar=500µm, H&E X400)                                                             | 66 |
| 4.5  | Photomicrograph of large intestine section showing cellular infiltration (arrow) in orally infected mouse with <i>B. melitensis</i> at day 10 post-infection. (Bar=100µm, H&E X200)                                                               | 66 |
| 4.6  | Photomicrograph of uterus section showing cellular infiltration (long arrow), degeneration and necrosis (arrow) in orally infected mouse with <i>B. melitensis</i> at day 10 post-infection. (Bar=60µm, H&E X400)                                 | 67 |
| 4.7  | Photomicrograph of liver section presenting congestion (long arrow), haemorrhage (arrows) and generalized degeneration and necrosis (head arrows) in orally infected mouse with lipopolysaccharide at day 3 post-infection. (Bar=100µm, H&E X200) | 67 |
| 4.8  | Photomicrograph of brain section presenting congestion (l arrow), in orally infected mouse with lipopolysaccharide at day 10 post-infection. (Bar=50µm, H&E X400)                                                                                 | 68 |
| 4.9  | Photomicrograph of testes section presenting normal histologic features in orally infected mouse with lipopolysaccharide at day 10 post-infection. (Bar=100µm, H&E X200).                                                                         | 68 |
| 4.10 | Photomicrograph of epididymis section presenting normal histologic features in orally infected mouse with LPS at day 10 post-infection. (Bar=100µm, H&E X200)                                                                                     | 69 |
| 4.11 | Comparison of Interleukin 1-β serum profiles in mice following oral infection with 10 <sup>9</sup> of <i>B. melitensis</i> and its lipopolysaccharide                                                                                             | 72 |
| 4.12 | Comparison of Interleukin -6 serum profiles in mice following oral infection with 10 <sup>9</sup> of <i>B. melitensis</i> and its lipopolysaccharide                                                                                              | 73 |
| 4.13 | Comparison of IgM serum profiles (Mean±SEM) in mice following oral infection with 10 <sup>9</sup> of <i>B. melitensis</i> and its lipopolysaccharide                                                                                              | 74 |

|      |                                                                                                                                                                                                                                                |    |
|------|------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|----|
| 4.14 | Comparison of IgG serum profiles (Mean±SEM) in mice following oral infection with $10^9$ of <i>B. melitensis</i> and its lipopolysaccharide                                                                                                    | 75 |
| 4.15 | Comparison of progesterone serum profiles (Mean±SEM) in mice following oral infection with <i>B. melitensis</i> and its lipopolysaccharide                                                                                                     | 76 |
| 4.16 | Comparison of estradiol serum profiles (Mean±SEM) in mice following oral infection with $10^9$ of <i>B. melitensis</i> and its lipopolysaccharide                                                                                              | 77 |
| 4.17 | Comparison of Testosterone serum profiles (Mean±SEM) in mice following oral infection with $10^9$ of <i>B. melitensis</i> and its lipopolysaccharide                                                                                           | 78 |
| 4.18 | Gel photograph of PCR products showing bands characteristic of <i>B. melitensis</i> following oral inoculation. The presence of bands~ 252 bp are indicative of positive results                                                               | 79 |
| 5.1  | Photomicrograph of lung section showing interstitial infiltration (long arrow) and congestion (arrow) in intranasally infected mouse with <i>B. melitensis</i> at day 10 post-infection. (Bar=100µm, H&E X200)                                 | 93 |
| 5.2  | Photomicrograph of liver section showing cellular infiltration (yellow long arrow), and degeneration (head of arrows) in intranasally infected mouse with <i>B. melitensis</i> at day 17 post-infection. (Bar=50µm, H&E X400)                  | 93 |
| 5.3  | Photomicrograph of kidney section showing congestion (arrow), and degeneration (head of arrows) in intranasally infected mouse with <i>B. melitensis</i> at day 3 post-infection. (Bar=50µm, H&E X400)                                         | 94 |
| 5.4  | Photomicrograph of uterus section showing congestion (arrow), and cellular infiltration (head of arrows) in intranasally infected mouse with <i>B. melitensis</i> at day 24 post-infection. (Bar=500µm, H&E X200)                              | 94 |
| 5.5  | Photomicrograph of liver section showing extensive haemorrhage (arrow), and generalized degeneration and necrosis (head of arrows) in intranasally infected mouse with lipopolysaccharide (LPS) at day 10 post-infection. (Bar=50µm, H&E X400) | 95 |
| 5.6  | Photomicrograph of lung section presenting congestion (arrow) and interstitial infiltration (head of arrow) in intranasally infected mouse with lipopolysaccharide (LPS) at day 10 post-infection. (Bar=500µm, H&E X200)                       | 95 |

|      |                                                                                                                                                                                                                                                            |     |
|------|------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|-----|
| 5.7  | Photomicrograph of heart section presenting haemorrhage (long arrow) and degeneration (arrow) in intranasally infected mouse with lipopolysaccharide (LPS) at day 3 post-infection. (Bar=100µm, H&E X200)                                                  | 96  |
| 5.8  | Comparison of Interleukin 1-β serum profiles (Mean±SEM) in mice following intranasal infection with <i>B. melitensis</i> and its lipopolysaccharide                                                                                                        | 97  |
| 5.9  | Comparison of Interleukin 6 serum profiles (Mean±SEM) in mice following intranasal infection with <i>B. melitensis</i> and its lipopolysaccharide                                                                                                          | 98  |
| 5.10 | Comparison of serum IgM profiles (Mean±SEM) in mice following intranasal infection with <i>B. melitensis</i> and its lipopolysaccharide                                                                                                                    | 99  |
| 5.11 | Comparison of serum IgG profiles (Mean±SEM) in mice following intranasal infection with <i>B. melitensis</i> and its lipopolysaccharide                                                                                                                    | 100 |
| 5.12 | Comparison of serum progesterone profiles (Mean±SEM) in mice following intranasal infection with <i>B. melitensis</i> and its lipopolysaccharide                                                                                                           | 101 |
| 5.13 | Comparison of serum estradiol profiles (Mean±SEM) in mice following intranasal infection with <i>B. melitensis</i> and its lipopolysaccharide                                                                                                              | 102 |
| 5.14 | Comparison of serum testosterone profiles (Mean±SEM) in mice following intranasal infection with <i>B. melitensis</i> and its lipopolysaccharide                                                                                                           | 103 |
| 6.1  | Photomicrograph of liver section showing congestion (long arrow), degeneration and necrosis (head of arrows) in subcutaneously infected mouse with <i>B. melitensis</i> at day 3 post-infection. (Bar=100µm, H&E X200)                                     | 115 |
| 6.2  | Photomicrograph of spleen showing hyperplasia (long arrow) and depletion of red pulp (long arrow), haemorrhage (arrow) in subcutaneously infected mouse with <i>B. melitensis</i> at day 3 post-infection. (Bar=100µm, H&E X200)                           | 116 |
| 6.3  | Photomicrograph of lung showing congestion (long arrow), interstitial infiltration of inflammatory cells (yellow arrow), and emphysema (arrow) in subcutaneously infected mouse with <i>B. melitensis</i> at day 10 post-infection. (Bar=100µm, H&E X200)  | 116 |
| 6.4  | Photomicrograph of kidney showing haemorrhage (yellow arrow), cellular infiltration (arrow), and degeneration (head of arrow) as seen in animals following subcutaneous infection with <i>B. melitensis</i> at day 10 post-infection. Bar=100µm, H&E X200) | 117 |

|      |                                                                                                                                                                                                                                                            |     |
|------|------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|-----|
| 6.5  | Small intestine of mice infected subcutaneously with <i>B. melitensis</i> and euthanized 10 day pi, note severe infiltration of inflammatory cells (yellow arrow), disruption of epithelial lining and foci of haemorrhages ( arrow) (Bar=100µm, H&E X200) | 117 |
| 6.6  | Mesenteric lymph node of mice infected subcutaneously with <i>B. melitensis</i> and euthanized 17 day pi, note severe infiltration of inflammatory cells (arrow), and foci of congestion ( head of arrow) (Bar=100µm, H&E X200)                            | 118 |
| 6.7  | Uterus of mice infected subcutaneously with <i>B. melitensis</i> and euthanized 10 day pi, note severe infiltration of inflammatory cells (arrow), and foci of haemorrhages ( head of arrow) (Bar=50µm, H&E X200)                                          | 118 |
| 6.8  | Photomicrograph of convoluted seminiferous tubules showing mild congestion and cellular infiltration in animals infected subcutaneously with <i>B. melitensis</i> at day 24 pi. (Bar=50µm, H&E X20)                                                        | 119 |
| 6.9  | Photomicrograph of liver of mice infected subcutaneously with lipopolysaccharide and euthanized 3 day pi, note generalized degeneration (arrow), mild cellular infiltration (long arrow), and foci of congestion (head of arrow).) (Bar=100µm, H&E X200)   | 119 |
| 6.10 | Photomicrograph of epididymis with cellular infiltration (arrow) as seen in animals following subcutaneous infection with LPS at day 17 post-infection. Original magnification (H&E X20)                                                                   | 120 |
| 6.11 | Comparison of Interleukin 1- $\beta$ serum profiles (Mean $\pm$ SEM) in mice following subcutaneous infection with <i>B. melitensis</i> and its lipopolysaccharide                                                                                         | 123 |
| 6.12 | Comparison of Interleukin 6 serum profiles (Mean $\pm$ SEM) in mice following subcutaneous infection with <i>B. melitensis</i> and its lipopolysaccharide                                                                                                  | 124 |
| 6.13 | Comparison of IgM serum profiles (Mean $\pm$ SEM) in mice following subcutaneous infection with <i>B. melitensis</i> and its lipopolysaccharide                                                                                                            | 125 |
| 6.14 | Comparison of IgG serum profiles (Mean $\pm$ SEM) in mice following subcutaneous infection with <i>B. melitensis</i> and its lipopolysaccharide                                                                                                            | 126 |
| 6.15 | Comparison of progesterone serum profiles (Mean $\pm$ SEM) in mice following subcutaneous infection with <i>B. melitensis</i> and its lipopolysaccharide                                                                                                   | 127 |
| 6.16 | Comparison of estradiol serum profiles (Mean $\pm$ SEM) in mice following subcutaneous infection with <i>B. melitensis</i> and its lipopolysaccharide                                                                                                      | 128 |

- 6.17 Comparison of testosterone serum profiles (Mean $\pm$ SEM) in mice 129  
following subcutaneous infection with *B. melitensis* and its  
lipopolysaccharide



## LIST OF ABBREVIATIONS

|                     |                                        |
|---------------------|----------------------------------------|
| %                   | Percentage                             |
| °C                  | Degree celcius                         |
| µg                  | Microgram                              |
| µL                  | Microlitre                             |
| µm                  | Micrometre                             |
| µM                  | Micrometre                             |
| 1gM                 | Immunoglobulin M                       |
| 1HC                 | Immunohistochemistry                   |
| 1L-β                | Interleukin -1 β                       |
| ANOVA               | Analysis of variances                  |
| APC                 | Antigen presenting cells               |
| ASW                 | Predictive Analysis Software           |
| <i>B.melitensis</i> | <i>Brucella melitensis</i>             |
| BALB/c              | Inbred strain of mouse                 |
| BHIB                | Brain heart infusion broth             |
| CFU                 | Colony forming unit                    |
| CPM                 | Count per minute                       |
| dH <sub>2</sub> O   | distilled water                        |
| DNA                 | Deoxyribonucleic acid                  |
| DPI                 | Days post-infection                    |
| EDTA                | Ethylenediaminetetraacetic Acid        |
| ELISA               | Enzyme Linked Immunosorbent Assay      |
| FAO                 | Food and Agriculture Organization      |
| G                   | Group                                  |
| G                   | Gram                                   |
| GALT                | Gut-associated lymphoid tissues        |
| gDNA                | Genomic deoxyribonucleic acid          |
| GIT                 | Gastrointestinal tract                 |
| H                   | Hours                                  |
| HE                  | Haematoxylin and Eosin                 |
| HPLC                | High-performance liquid chromatography |
| HRP                 | Horseradish peroxidase                 |

|                   |                                                            |
|-------------------|------------------------------------------------------------|
| IACUC             | Animal Care and Use Committee                              |
| IFN- $\gamma$     | Interferon gamma                                           |
| IgA               | Immunoglobulin A                                           |
| IgG               | Immunoglobulin G                                           |
| IL-1 $\beta$      | Interleukin-1 beta                                         |
| IL-6              | Interleukin-6                                              |
| IN                | Intranasal                                                 |
| IP                | Intraperitoneal                                            |
| L                 | Litre                                                      |
| LPS               | Lipopolysaccharide                                         |
| M                 | Moribund                                                   |
| MgCl <sub>2</sub> | Magnesium Chloride                                         |
| Min               | Minutes                                                    |
| ml                | Millilitre                                                 |
| mm                | Millimetre                                                 |
| N                 | Number                                                     |
| NA                | Not applicable                                             |
| NaOH              | Sodium hydroxide                                           |
| NC                | Negative Control                                           |
| Ng                | Nanogram                                                   |
| OD                | Optical density                                            |
| OIE               | World Organization for Animal Health                       |
| OMP               | Outer membrane protein                                     |
| P1                | Primer 1                                                   |
| P2                | Primer 2                                                   |
| PASW              | Predictive Analytics Software                              |
| PBS               | Phosphate buffered saline                                  |
| PBST              | Tween20 phosphate buffered saline                          |
| PC                | Positive Control                                           |
| PCR               | Polymerase chain reaction                                  |
| Pg                | Picogram                                                   |
| pH                | Potential hydrogen/power of hydrogen (acidity or basicity) |
| PI                | Post-infection                                             |
| RIA               | Radioimmunoassay                                           |

|              |                                         |
|--------------|-----------------------------------------|
| RNA          | Ribonucleic Acid                        |
| Rpm          | Revolution per minute                   |
| RT           | Room temperature                        |
| S/c          | Subcutaneous                            |
| Sec          | Seconds                                 |
| sIgA         | Secretory immunoglobulin A              |
| SPSS         | Statistical package for Social Sciences |
| T            | Terminal                                |
| TAE          | Tris-acetate-EDTA                       |
| TBE          | Tris-boric acid-EDTA                    |
| TNF $\alpha$ | Tumour necrosis factor-alpha            |
| TSB          | Trypticase Soy Broth                    |
| UK           | United Kingdom                          |
| UPM          | University Putra Malaysia               |
| USA          | United States of America                |
| USD          | United States Dollar                    |
| UV           | Ultraviolet                             |

## CHAPTER 1

### INTRODUCTION

*Brucella melitensis* is one of the major zoonotic pathogens with significant economic implications as well as considerable human morbidity in many countries including Malaysia (Bamaiyi *et al.*, 2010; Seleem *et al.*, 2010). It is the main causative agent of small ruminant brucellosis (SRB) as it is also infectious to other species including cattle, buffalo and elk (Corbel, 2006; D áz, 2013). The disease remains endemic and neglected in many regions of the world, with predominance in the Mediterranean Basin, Middle East, Africa, Latin America and central Asia (Blasco & Molina-Flores, 2011; Lucero *et al.*, 2008; Thimm, 2013). The global burden of its incidence in human populations remains significantly at alarming rate (Pappas *et al.*, 2006).

The organism is facultative intracellular pathogen cocco-bacilli, non-spore-forming and non-capsulated with up to 3 biovars have been reported. These biovars differ biochemically by their pattern of metabolic activities (Halling *et al.*, 2005). The risk of brucellosis is presumed to be high in nomadic pastoral societies, laboratory workers or veterinarians where close and frequent contact between man and animals is part of the ecology.

The disease affects wild and domestic mammals with special predominance in small ruminants and cattle causing abortion and reduced fertility (Godfroid *et al.*, 2002; Gwida *et al.*, 2010; Megersa *et al.*, 2011). It is notifiable and neglected disease with serious economic repercussion on both humans and animals (Abernethy *et al.*, 2011; Ko & Splitter, 2003; Radostits *et al.*, 2007; Seleem *et al.*, 2010). It is mainly contracted through contact with placenta, foetus, foetal fluids and vaginal discharge from infected animals. In human, it is considered a food borne disease or a disease related to occupational exposures. The routes of infection for both humans and animals are similar of nature which include ingestion, inhalation, or through direct contact of the organism with a break in the skin (Corbel, 2006). Higher incidence of *B. melitensis* is associated with environmental and management factors which include moist, humid conditions, high animal population density, extensive free grazing system and poor husbandry practice (D áz, 2013). The initial symptoms of infected humans are fever, lethargy and night sweats. However, complication may set in as a result of chronic infection, which allows involvement of many organs and systems such as liver, spleen, kidney, and skeleton among others (Young *et al.*, 2014). In domestic animals, the disease is manifested as fertility-related issues. However, the most common symptom is usually abortion during the trimester often followed by retained placenta, weak offspring and metritis which may result in temporary infertility. Others include drop in milk production due to the infection of the udder. Rams experience orchitis and epididymitis. In addition, animals with polyarthritis have been observed in endemic flocks (Corbel, 2006; Radostits *et al.*, 2007).

Despite substantial attempts in the realm of the organism's characteristics, diagnosis and development of vaccines, the disease remains a major issue in animal industry (Bardenstein *et al.*, 2002; Corbel, 2006; Doganay & Aygen, 2003). The pathogenicity is complex and not always well understood. Understanding the pathogenicity and mechanism which *B. melitensis* interact with their hosts to produce clinical manifestation becomes a fundamental issue. Knowledge on immunopathophysiology of *Brucella* and its virulent factor is important to understanding the replication and survival of the bacteria.

The ability of the microbe to cause disease in a susceptible host, however, is determined by multiple virulence factors acting individually or together at different stages of infection (Neta *et al.*, 2010). In this regard, the presence of lipopolysaccharides (LPS) in the outer membrane protein of *B. melitensis* is believed to play a major role in diseases pathogenesis (Lapaque *et al.*, 2005). This unconventional non-endotoxic lipopolysaccharide confers resistance to anti-microbial attacks and modulates the host immune response (Lapaque *et al.*, 2005). The virulence factors are often involved in concealing the bacterial surface from the host's defense mechanisms. Their roles whether or not to directly mediate clinical manifestations of the disease is yet questionable. In the context of protection against *B. melitensis* infection, antibodies specific for the O-antigen of the lipopolysaccharide and production of proinflammatory cytokines are considered to be important for controlling *Brucella* infections (Macedo *et al.*, 2008; Neta *et al.*, 2010).

On the other hand, sex related hormones are essential for regulation of sex differentiation, reproduction, growth, metabolism and immune function (Mellon & Griffin, 2002; Murad *et al.*, 2010). A decrease in serum progesterone and estrogen levels is commonly associated with events leading to abortions in field conditions (Aisemberg *et al.*, 2013).

The clarifications, however, of the exact routes of transmission, sites of infection of *B. melitensis* and its LPS along with the impact of immunopathophysiological aspects in hosts can facilitate understanding of its biological features and control of brucellosis.

This study was, therefore, conducted to compare the establishment of clinical manifestation, the severity of pathological lesions, the role of innate and cellular immune response and the sex related hormonal alterations in mouse model following different route of inoculation of *B. melitensis* and its lipopolysaccharide (LPS).

## 1.1 Research hypotheses

1. Oral route of infection by *B. melitensis* produces comparable degree of injury and clinical manifestation of *B. melitensis* as intranasal, subcutaneous and/or intraperitoneal route of infection.
2. Pattern of bacterial distribution following experimental infection with *B. melitensis* in various organs and/or tissue is similar either orally, intranasally, subcutaneously and/or intraperitoneally infected mice.
3. All routes of administration of lipopolysaccharide (LPS) produce minimal tissue injuries and clinical manifestation compared to *B. melitensis*.
4. All routes of infection elicit comparable cytokine, hormonal changes and antibody immune responses following exposure to *B. melitensis* or its lipopolysaccharide (LPS).

## 1.2 Objectives of the study

1. To determine the clinical signs following experimental infection via different route inoculations of *B. melitensis* and its immunogens (LPS) in mouse model.
2. To determine the antibody levels (IgG and IgM) following assessment of different route inoculations of *B. melitensis* and its immunogens (LPS) in mouse model.
3. To measure the concentration of cytokines following experimental infection of mice with *B. melitensis* and its immunogens (LPS) via different route of inoculation.
4. To determine the concentration of reproductive hormones of both sexes following inoculation of animals with *B. melitensis* and its immunogens (LPS) via different route of exposure.
5. To evaluate the histopathological changes of the infected organs and tissues in mice following different route of exposure of *B. melitensis* and its immunogens (LPS).
6. To isolate and detect the *B. melitensis* by PCR from infected organs and tissues of mice challenged via different route of inoculation

## REFERENCES

- Abdel-Khalik, J., Björklund, E., & Hansen, M. (2013). Development of a solid phase extraction method for the simultaneous determination of steroid hormones in H295R cell line using liquid chromatography–tandem mass spectrometry. *Journal of Chromatography B*, 935, 61-69.
- Abdullah, F. F. J., Nik, N. B., Saad, M. Z., Haron, A. W., Omar, A. R., Sabri, J., Adamu, L., Osman, A. Y., & Saharee, A. A. (2013). Clinico-pathological Changes Associated with " *Brucella melitensis*" Infection and its Bacterial Lipopolysaccharides (LPS) in Male Mice. *International Journal of Animal and Veterinary Advances*, 5(5), 165-170.
- Abernethy, D., Moscard-Costello, J., Dickson, E., Harwood, R., Burns, K., McKillop, E., McDowell, S., & Pfeiffer, D. (2011). Epidemiology and management of a bovine brucellosis cluster in Northern Ireland. *Preventive veterinary medicine*, 98(4), 223-229.
- Acha, P. N., & Szyfres, B. (2003). *Zoonoses and communicable diseases common to man and animals* (Vol. 3): Pan American Health Org.
- Acocella, G., Bertrand, A., Beytout, J., Durrande, J. B., Rodriguez, J.-A. G., Kosmidis, J., Micoud, M., Rey, M., Zapata, M. R., & Roux, J. (1989). Comparison of three different regimens in the treatment of acute brucellosis: a multicenter multinational study. *Journal of Antimicrobial Chemotherapy*, 23(3), 433-439.
- Aisemberg, J., Vercelli, C. A., Bariani, M. V., Billi, S. C., Wolfson, M. L., & Franchi, A. M. (2013). Progesterone is essential for protecting against LPS-induced pregnancy loss. LIF as a potential mediator of the anti-inflammatory effect of progesterone. *PLoS One*, 8(2), e56161.
- Ajuwon, K. M., & Spurlock, M. E. (2005). Palmitate activates the NF- $\kappa$ B transcription factor and induces IL-6 and TNF $\alpha$  expression in 3T3-L1 adipocytes. *The Journal of nutrition*, 135(8), 1841-1846.
- Al-Garadi, M. A., Khairani-Bejo, S., Zunita, Z., & Omar, A. (2011). Isolation and identification of *Brucella melitensis* in goats. *J Anim Vet Adv*, 10(8), 972-979.
- Al-Majali, A. M., Talafha, A. Q., Ababneh, M. M., & Ababneh, M. M. (2009). Seroprevalence and risk factors for bovine brucellosis in Jordan. *Journal of veterinary science*, 10(1), 61-65.
- Al-Mariri, A. (2010). Protection of BALB/c mice against *Brucella melitensis* 16M infection induced by vaccination with live *Escherchia coli* expression *Brucella* P39 protein. *Vaccine*, 28(7), 1766-1770.
- Alton, G. G., Nielsen, K., & Duncan, J. (1990). *Brucella suis*. *Animal brucellosis*., 411-422.

- Álvarez, J., Sáez, J. L., García, N., Serrat, C., Pérez-Sancho, M., González, S., Ortega, M. J., Gou, J., Carbajo, L., & Garrido, F. (2011). Management of an outbreak of brucellosis due to *B. melitensis* in dairy cattle in Spain. *Research in veterinary science*, 90(2), 208-211.
- Anka, M. S., Hassan, L., Adzhar, A., Khairani-Bejo, S., Mohamad, R. B., & Zainal, M. A. (2013). Bovine brucellosis trends in Malaysia between 2000 and 2008. *BMC veterinary research*, 9(1), 1.
- Baldi, P. C., Miguel, S. E., Fossati, C. A., & Wallach, J. C. (1996). Serological follow-up of human brucellosis by measuring IgG antibodies to lipopolysaccharide and cytoplasmic proteins of *Brucella* species. *Clinical infectious diseases*, 22(3), 446-455.
- Baldwin, C., Goenka, R., López-Goñi, I., & Moriyón, I. (2004). Host cellular immune responses against *Brucella* spp. evaluated using the mouse model. *Brucella: molecular and cellular biology*, 341-367.
- Baldwin, C. L., & Goenka, R. (2006). Host immune responses to the intracellular bacteria *Brucella*: does the bacteria instruct the host to facilitate chronic infection? *Critical Reviews™ in Immunology*, 26(5).
- Bamaiyi, P., Hassan, L., Siti-Khirani, B., Adzhar, A., & Rachmat, R. (2010). *The Seroprevalence of Brucella melitensis in goats of Malaysia from year 2000 to 2008*. Paper presented at the 22nd Veterinary Association of Malaysia Congress and 4th Wildlife Society of Zoo and Wildlife Medicine International Meeting. Veterinary Association of Malaysia, City.
- Barbier, T., Nicolas, C., & Letesson, J. (2011). *Brucella* adaptation and survival at the crossroad of metabolism and virulence. *FEBS letters*, 585(19), 2929-2934.
- Bardenstein, S., Mandelboim, M., Ficht, T. A., Baum, M., & Banai, M. (2002). Identification of the *Brucella melitensis* vaccine strain Rev. 1 in animals and humans in Israel by PCR analysis of the PstI site polymorphism of its omp2 gene. *Journal of clinical microbiology*, 40(4), 1475-1480.
- Barquero-Calvo, E., Chaves-Olarte, E., Weiss, D. S., Guzmán-Verri, C., Chacón-Díaz, C., Rucavado, A., Moriyón, I., & Moreno, E. (2007). *Brucella abortus* uses a stealthy strategy to avoid activation of the innate immune system during the onset of infection. *PLoS One*, 2(7), e631.
- Bartels, E. M., Falbe Wätjen, I., Littrup Andersen, E., Danneskiold-Samsøe, B., Bliddal, H., & Ribel-Madsen, S. (2011). Rheumatoid factor and its interference with cytokine measurements: problems and solutions. *Arthritis*, 2011.
- Bassil, N., Alkaade, S., & Morley, J. E. (2009). The benefits and risks of testosterone replacement therapy: a review. *Ther Clin Risk Manag*, 5(3), 427-448.

- Beh, K. (1974). Quantitative distribution of *Brucella* antibody amongst immunoglobulin classes in vaccinated and infected cattle. *Research in veterinary science*, 17(1), 1-4.
- Bekele, M., Mohammed, H., Tefera, M., & Tolosa, T. (2011). Small ruminant brucellosis and community perception in Jijiga district, Somali Regional State, eastern Ethiopia. *Tropical Animal Health and Production*, 43(4), 893-898.
- Bhattacharjee, A. K., Izadjoo, M. J., Zollinger, W. D., Nikolich, M. P., & Hoover, D. L. (2006). Comparison of protective efficacy of subcutaneous versus intranasal immunization of mice with a *Brucella melitensis* lipopolysaccharide subunit vaccine. *Infection and immunity*, 74(10), 5820-5825.
- Blasco, J. (2010). Control and eradication strategies for *Brucella melitensis* infection in sheep and goats. *Prilozi*, 31(1), 145-165.
- Blasco, J., Gamazo, C., Winter, A., de Bagüés, M. J., Marín, C., Barberán, M., Moriyón, I., Alonso-Urmeneta, B., & Díaz, R. (1993). Evaluation of whole cell and subcellular vaccines against *Brucella ovis* in rams. *Veterinary immunology and immunopathology*, 37(3-4), 257-270.
- Blasco, J. M., & Molina-Flores, B. (2011). Control and eradication of *Brucella melitensis* infection in sheep and goats. *Veterinary Clinics of North America: Food Animal Practice*, 27(1), 95-104.
- Bolstad, N., Warren, D. J., & Nustad, K. (2013). Heterophilic antibody interference in immunometric assays. *Best Practice & Research Clinical Endocrinology & Metabolism*, 27(5), 647-661.
- Bricker, B. J. (2002). Diagnostic strategies used for the identification of *Brucella*. *Veterinary microbiology*, 90(1), 433-434.
- Bruce, S. D. (1888). *The micrococcus of Malta fever*: publisher not identified.
- Cassataro, J., Estein, S. M., Pasquevich, K. A., Velikovsky, C. A., de la Barrera, S., Bowden, R., Fossati, C. A., & Giambartolomei, G. H. (2005). Vaccination with the recombinant *Brucella* outer membrane protein 31 or a derived 27-amino-acid synthetic peptide elicits a CD4<sup>+</sup> T helper 1 response that protects against *Brucella melitensis* infection. *Infection and immunity*, 73(12), 8079-8088.
- Cassataro, J., Pasquevich, K. A., Estein, S. M., Laplagne, D. A., Velikovsky, C. A., de la Barrera, S., Bowden, R., Fossati, C. A., Giambartolomei, G. H., & Goldbaum, F. A. (2007). A recombinant subunit vaccine based on the insertion of 27 amino acids from Omp31 to the N-terminus of BLS induced a similar degree of protection against *B. ovis* than Rev. 1 vaccination. *Vaccine*, 25(22), 4437-4446.
- Catalfamo, M., Le Saout, C., & Lane, H. C. (2012). The role of cytokines in the pathogenesis and treatment of HIV infection. *Cytokine & growth factor reviews*, 23(4), 207-214.

- Çelebi, G., Külah, C., Kiliç, S., & Üstündağ, G. (2007). Asymptomatic *Brucella* bacteraemia and isolation of *Brucella melitensis* biovar 3 from human breast milk. *Scandinavian journal of infectious diseases*, 39(3), 205-208.
- Cha, S.-B., Rayamajhi, N., Kang, M.-L., Lee, W.-J., Shin, M.-K., & Yoo, H.-S. (2010). Comparative study of gamma interferon production in mice immunized with outer membrane proteins and whole bacteria of *Brucella abortus* *Jpn J Infect Dis*, 63(1), 49-51.
- Chan, M. S., Carey, A. L., Watt, M. J., & Febbraio, M. A. (2004). Cytokine gene expression in human skeletal muscle during concentric contraction: evidence that IL-8, like IL-6, is influenced by glycogen availability. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, 287(2), R322-R327.
- Chen, M., Chen, J., Liao, W., Zhu, S., Yu, J., Leung, W. K., Hu, P., & Sung, J. J. (2003). Immunization with attenuated *Salmonella typhimurium* producing catalase in protection against gastric *Helicobacter pylori* infection in mice. *Helicobacter*, 8(6), 613-625.
- Cherrier, M., Matsumoto, A., Amory, J., Ahmed, S., Bremner, W., Peskind, E., Raskind, M., Johnson, M., & Craft, S. (2005). The role of aromatization in testosterone supplementation effects on cognition in older men. *Neurology*, 64(2), 290-296.
- Chiswick, E. L., Duffy, E., Japp, B., & Remick, D. (2012). Detection and quantification of cytokines and other biomarkers. *Leucocytes: Methods and Protocols*, 15-30.
- Chomarat, P., Banchereau, J., Davoust, J., & Palucka, A. K. (2000). IL-6 switches the differentiation of monocytes from dendritic cells to macrophages. *Nature immunology*, 1(6), 510-514.
- Clark, D. A., Chaouat, G., & Gorczynski, R. M. (2002). Thinking Outside the Box: Mechanisms of Environmental Selective Pressures on the Outcome of the Materno-fetal Relationship. *American Journal of Reproductive Immunology*, 47(5), 275-282.
- Clark, D. A., Manuel, J., Lee, L., Chaouat, G., Gorczynski, R. M., & Levy, G. A. (2004). Ecology of Danger-dependent Cytokine-boosted Spontaneous Abortion in the CBA× DBA/2 Mouse Model. I. Synergistic Effect of LPS and (TNF- $\alpha$ + IFN- $\gamma$ ) on Pregnancy Loss. *American Journal of Reproductive Immunology*, 52(6), 370-378.
- Colmenero, J. D., Munoz-Roca, N. L., Bermudez, P., Plata, A., Villalobos, A., & Reguera, J. M. (2007). Clinical findings, diagnostic approach, and outcome of *Brucella melitensis* epididymo-orchitis. *Diagnostic microbiology and infectious disease*, 57(4), 367-372.

- Commander, N. J., Spencer, S. A., Wren, B. W., & MacMillan, A. P. (2007). The identification of two protective DNA vaccines from a panel of five plasmid constructs encoding *Brucella melitensis* 16M genes. *Vaccine*, 25(1), 43-54.
- Corbel, M. J. (2006). *Brucellosis in humans and animals*: World Health Organization.
- Cox, J. H., Ferrari, G., & Janetzki, S. (2006). Measurement of cytokine release at the single cell level using the ELISPOT assay. *Methods*, 38(4), 274-282.
- Cram, L. F., Zapata, M.-I., Toy, E. C., & Baker 3rd, B. (2002). Genitourinary infections and their association with preterm labor. *American family physician*, 65(2), 241-248.
- Davis, D. S., Templeton, J. W., Ficht, T. A., Williams, J. D., Kopec, J. D., & Adams, L. G. (1990). *Brucella abortus* in captive bison. I. Serology, bacteriology, pathogenesis, and transmission to cattle. *Journal of Wildlife Diseases*, 26(3), 360-371.
- De Bagues Maria-Pilar, J., Dudal, S., Dornand, J., & Gross, A. (2005). Cellular bioterrorism: how *Brucella* corrupts macrophage physiology to promote invasion and proliferation. *Clinical Immunology*, 114(3), 227-238.
- De Massis, F., Di Girolamo, A., Petrini, A., Pizzigallo, E., & Giovannini, A. (2005). Correlation between animal and human brucellosis in Italy during the period 1997–2002. *Clinical microbiology and infection*, 11(8), 632-636.
- Delpino, M. V., Estein, S. M., Fossati, C. A., Baldi, P. C., & Cassataro, J. (2007). Vaccination with *Brucella* recombinant DnaK and SurA proteins induces protection against *Brucella abortus* infection in BALB/c mice. *Vaccine*, 25(37), 6721-6729.
- Díaz, A. E. (2013). Epidemiology of brucellosis in domestic animals caused by *Brucella melitensis*, *Brucella suis* and *Brucella abortus* *Revue scientifique et technique (International Office of Epizootics)*, 32(1), 43-51, 53-60.
- Dienz, O., Eaton, S. M., Bond, J. P., Neveu, W., Moquin, D., Noubade, R., Briso, E. M., Charland, C., Leonard, W. J., & Ciliberto, G. (2009). The induction of antibody production by IL-6 is indirectly mediated by IL-21 produced by CD4<sup>+</sup> T cells. *The Journal of experimental medicine*, 206(1), 69-78.
- Dinareello, C. A. (2009). Immunological and inflammatory functions of the interleukin-1 family. *Annual review of immunology*, 27, 519-550.
- Ding, E. L., Song, Y., Malik, V. S., & Liu, S. (2006). Sex differences of endogenous sex hormones and risk of type 2 diabetes: a systematic review and meta-analysis. *Jama*, 295(11), 1288-1299.
- Doganay, M., & Aygen, B. (2003). Human brucellosis: an overview. *International Journal of Infectious Diseases*, 7(3), 173-182.

- Ducrotoy, M. J., Conde-Álvarez, R., Blasco, J. M., & Moriyón, I. (2016). A review of the basis of the immunological diagnosis of ruminant brucellosis. *Veterinary immunology and immunopathology*, 171, 81-102.
- Dunne, A., & O'Neill, L. A. (2003). The interleukin-1 receptor/Toll-like receptor superfamily: signal transduction during inflammation and host defense. *Sci. STKE*, 2003(171), re3-re3.
- Duran-Ferrer, M., Leon, L., Nielsen, K., Caporale, V., Mendoza, J., Osuna, A., Perales, A., Smith, P., De-Frutos, C., & Gomez-Martin, B. (2004). Antibody response and antigen-specific gamma-interferon profiles of vaccinated and unvaccinated pregnant sheep experimentally infected with *Brucella melitensis*. *Veterinary microbiology*, 100(3), 219-231.
- Edalatmanesh, M. A. (2014). Evaluation of sex hormones (FSH, Estrogen and Testosterone) changes during follicular and luteal phases and sexual dysfunction in women with Multiple Sclerosis. *Journal of Jahrom University of Medical Sciences*, 12(4), 49-56.
- Eddahri, F., Denanglaire, S., Bureau, F., Spolski, R., Leonard, W. J., Leo, O., & Andris, F. (2009). Interleukin-6/STAT3 signaling regulates the ability of naive T cells to acquire B-cell help capacities. *Blood*, 113(11), 2426-2433.
- Edmonds, M. D., Cloeckaert, A., & Elzer, P. H. (2002). *Brucella* species lacking the major outer membrane protein Omp25 are attenuated in mice and protect against *Brucella melitensis* and *Brucella ovis*. *Veterinary microbiology*, 88(3), 205-221.
- Elzer, P. H., Phillips, R. W., Kovach, M. E., Peterson, K. M., & Roop, R. (1994). Characterization and genetic complementation of a *Brucella abortus* high-temperature-requirement A (htrA) deletion mutant. *Infection and immunity*, 62(10), 4135-4139.
- Enright, F., Araya, L., Elzer, P., Rowe, G., & Winter, A. (1990). Comparative histopathology in BALB/c mice infected with virulent and attenuated strains of *Brucella abortus*. *Veterinary immunology and immunopathology*, 26(2), 171-182.
- Eze, M. O., Yuan, L., Crawford, R. M., Parnavitana, C. M., Hadfield, T. L., Bhattacharjee, A. K., Warren, R. L., & Hoover, D. L. (2000). Effects of opsonization and gamma interferon on growth of *Brucella melitensis* 16M in mouse peritoneal macrophages in vitro. *Infection and immunity*, 68(1), 257-263.
- Falkenstein, E., Tillmann, H.-C., Christ, M., Feuring, M., & Wehling, M. (2000). Multiple actions of steroid hormones—a focus on rapid, nongenomic effects. *Pharmacological reviews*, 52(4), 513-556.

- Fasshauer, M., Kralisch, S., Klier, M., Lossner, U., Bluher, M., Klein, J., & Paschke, R. (2004). Insulin resistance-inducing cytokines differentially regulate SOCS mRNA expression via growth factor-and Jak/Stat-signaling pathways in 3T3-L1 adipocytes. *Journal of Endocrinology*, 181(1), 129-138.
- Faupel-Badger, J. M., Fuhrman, B. J., Xu, X., Falk, R. T., Keefer, L. K., Veenstra, T. D., Hoover, R. N., & Ziegler, R. G. (2010). Comparison of liquid chromatography-tandem mass spectrometry, RIA, and ELISA methods for measurement of urinary estrogens. *Cancer Epidemiology Biomarkers & Prevention*, 19(1), 292-300.
- Febbraio, M. A., & Pedersen, B. K. (2002). Muscle-derived interleukin-6: mechanisms for activation and possible biological roles. *The FASEB Journal*, 16(11), 1335-1347.
- Fernández-Lago, L., Orduña, A., & Vizcaíno, N. (2005). Reduced interleukin-18 secretion in *Brucella abortus* 2308-infected murine peritoneal macrophages and in spleen cells obtained from B. *abortus* 2308-infected mice. *Journal of medical microbiology*, 54(6), 527-531.
- Fettelschoss, A., Kistowska, M., LeibundGut-Landmann, S., Beer, H.-D., Johansen, P., Senti, G., Contassot, E., Bachmann, M. F., French, L. E., & Oxenius, A. (2011). Inflammasome activation and IL-1 $\beta$  target IL-1 $\alpha$  for secretion as opposed to surface expression. *Proceedings of the National Academy of Sciences*, 108(44), 18055-18060.
- Ficht, T. (2010). *Brucella* taxonomy and evolution. *Future microbiology*, 5(6), 859-866.
- Foster, G., Osterman, B. S., Godfroid, J., Jacques, I., & Cloeckeaert, A. (2007). *Brucella ceti* sp. nov. and *Brucella pinnipedialis* sp. nov. for *Brucella* strains with cetaceans and seals as their preferred hosts. *International journal of systematic and evolutionary microbiology*, 57(11), 2688-2693.
- Franco, M. P., Mulder, M., Gilman, R. H., & Smits, H. L. (2007). Human brucellosis. *The Lancet infectious diseases*, 7(12), 775-786.
- Frost, R. A., Nystrom, G. J., & Lang, C. H. (2003). Lipopolysaccharide and proinflammatory cytokines stimulate interleukin-6 expression in C2C12 myoblasts: role of the Jun NH2-terminal kinase. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, 285(5), R1153-R1164.
- Fugier, E., Pappas, G., & Gorvel, J.-P. (2007). Virulence factors in brucellosis: implications for aetiopathogenesis and treatment. *Expert Reviews in Molecular Medicine*, 9(35), 1-10.
- Gall, D., & Nielsen, K. (2004). Serological diagnosis of bovine brucellosis: a review of test performance and cost comparison. *Revue scientifique et technique (International Office of Epizootics)*, 23(3), 989-1002.

- Garner, M. M., Lambourn, D. M., Jeffries, S. J., Hall, P. B., Rhyan, J. C., Ewalt, D. R., Polzin, L. M., & Cheville, N. F. (1997). Evidence of *Brucella* infection in Parafilaroides lungworms in a Pacific harbor seal (*Phoca vitulina richardsi*). *Journal of Veterinary Diagnostic Investigation*, 9(3), 298-302.
- Giambartolomei, G. H., Delpino, M. V., Cahanovich, M. E., Wallach, J. C., Baldi, P. C., Velikovskiy, C. A., & Fossati, C. A. (2002). Diminished production of T helper 1 cytokines correlates with T cell unresponsiveness to *Brucella* cytoplasmic proteins in chronic human brucellosis. *Journal of Infectious Diseases*, 186(2), 252-259.
- Godfroid, J., Nielsen, K., & Saegerman, C. (2010). Diagnosis of brucellosis in livestock and wildlife. *Croatian medical journal*, 51(4), 296-305.
- Godfroid, J., Saegerman, C., Wellemans, V., Walravens, K., Letesson, J.-J., Tibor, A., Mc Millan, A., Spencer, S., Sanna, M., & Bakker, D. (2002). How to substantiate eradication of bovine brucellosis when aspecific serological reactions occur in the course of brucellosis testing. *Veterinary microbiology*, 90(1), 461-477.
- Golding, B., Scott, D. E., Scharf, O., Huang, L.-Y., Zaitseva, M., Lapham, C., Eller, N., & Golding, H. (2001). Immunity and protection against *Brucella abortus*. *Microbes and infection*, 3(1), 43-48.
- Golding, B., Scott, D. E., Scharf, O., Huang, L.-Y., Zaitseva, M., Lapham, C., Eller, N., & Golding, H. (2001). Immunity and protection against *Brucella abortus*. *Microbes and infection*, 3(1), 43-48.
- Goletiani, N. V., Keith, D. R., & Gorsky, S. J. (2007). Progesterone: review of safety for clinical studies. *Experimental and clinical psychopharmacology*, 15(5), 427.
- Gonzalez, D., Grilló, M.-J., De Miguel, M.-J., Ali, T., Arce-Gorvel, V., Delrue, R.-M., Conde-Álvarez, R., Muñoz, P., López-Goñi, I., & Iriarte, M. (2008). Brucellosis vaccines: assessment of *Brucella melitensis* lipopolysaccharide rough mutants defective in core and O-polysaccharide synthesis and export. *PLoS One*, 3(7), e2760.
- Gonzalez-Fernandez, D., Grillo, M. J., Miguel, M., Ali, T., Arce-Gorvel, V., Delrue, R. M., Conde-Alvarez, R., Muñoz, P., Lopez-Goñi, I., & Iriarte, M. (2008). Brucellosis vaccines: assessment of *Brucella melitensis* lipopolysaccharide rough mutants defective in core and O-polysaccharide synthesis and export.
- Gopaul, K. K., Koylass, M. S., Smith, C. J., & Whatmore, A. M. (2008). Rapid identification of *Brucella* isolates to the species level by real time PCR based single nucleotide polymorphism (SNP) analysis. *BMC microbiology*, 8(1), 1.
- Gorvel, J. P., & Moreno, E. (2002). *Brucella* intracellular life: from invasion to intracellular replication. *Veterinary microbiology*, 90(1), 281-297.

- Gorvel, J. P., Moreno, E., & Moriyón, I. (2009). Is *Brucella* an enteric pathogen? *Nature Reviews Microbiology*, 7(3), 250-250.
- Greene, C. E., Carmichael, L. E., & Greene, C. (2006). Canine brucellosis. *Infectious diseases of the dog and cat*, 369-381.
- Grillo, M., Barberán, M., & Blasco, J. (1997). Transmission of *Brucella melitensis* from sheep to lambs. *The Veterinary Record*, 140(23), 602-605.
- Grilló, M. J., Manterola, L., De Miguel, M. J., Muñoz, P. M., Blasco, J. M., Moriyón, I., & López-Goñi, I. (2006). Increases of efficacy as vaccine against *Brucella abortus* infection in mice by simultaneous inoculation with avirulent smooth bvrS/bvrR and rough wbkA mutants. *Vaccine*, 24(15), 2910-2916.
- Grilló, M. J., Manterola, L., De Miguel, M. J., Muñoz, P. M., Blasco, J. M., Moriyón, I., & López-Goñi, I. (2006). Increases of efficacy as vaccine against *Brucella abortus* infection in mice by simultaneous inoculation with avirulent smooth bvrS/bvrR and rough wbkA mutants. *Vaccine*, 24(15), 2910-2916.
- Grilló, M.-J., Blasco, J. M., Gorvel, J. P., Moriyón, I., & Moreno, E. (2012). What have we learned from brucellosis in the mouse model? *Veterinary research*, 43(1), 1.
- Guest, P. C., Schwarz, E., Krishnamurthy, D., Harris, L. W., Leweke, F. M., Rothermundt, M., van Beveren, N. J., Spain, M., Barnes, A., & Steiner, J. (2011). Altered levels of circulating insulin and other neuroendocrine hormones associated with the onset of schizophrenia. *Psychoneuroendocrinology*, 36(7), 1092-1096.
- Guilloteau, L. A., Dornand, J., Gross, A., Olivier, M., Cortade, F., Le Vern, Y., & Kerboeuf, D. (2003). Nramp1 is not a major determinant in the control of *Brucella melitensis* infection in mice. *Infection and immunity*, 71(2), 621-628.
- Guilloteau, L. A., Dornand, J., Gross, A., Olivier, M., Cortade, F., Le Vern, Y., & Kerboeuf, D. (2003). Nramp1 is not a major determinant in the control of *Brucella melitensis* infection in mice. *Infection and immunity*, 71(2), 621-628.
- Gupta, V., Rout, P., & Vihan, V. (2007). Induction of immune response in mice with a DNA vaccine encoding outer membrane protein (omp31) of *Brucella melitensis* 16M. *Research in veterinary science*, 82(3), 305-313.
- Gupta, V., Shivasharanappa, N., Kumar, V., & Kumar, A. (2014). Diagnostic evaluation of serological assays and different gene based PCR for detection of *Brucella melitensis* in goat. *Small Ruminant Research*, 117(1), 94-102.
- Gwida, M., Al Dahouk, S., Melzer, F., Rösler, U., Neubauer, H., & Tomaso, H. (2010). Brucellosis—regionally emerging zoonotic disease? *Croatian medical journal*, 51(4), 289-295.

- Gwida, M., Al Dahouk, S., Melzer, F., Rösler, U., Neubauer, H., & Tomaso, H. (2010). Brucellosis—regionally emerging zoonotic disease? *Croatian medical journal*, 51(4), 289-295.
- Haag, A. F., Myka, K. K., Arnold, M. F., Caro-Hernández, P., & Ferguson, G. P. (2010). Importance of lipopolysaccharide and cyclic  $\beta$ -1, 2-glucans in *Brucella*-mammalian infections. *International journal of microbiology*, 2010.
- Hafez, E., Jainudeen, M., & Rosnina, Y. (2000). Hormones, growth factors, and reproduction. *Reproduction in Farm Animals*, 7th Edition, 31-54.
- Haisenleder, D. J., Schoenfelder, A. H., Marcinko, E. S., Geddis, L. M., & Marshall, J. C. (2011). Estimation of estradiol in mouse serum samples: evaluation of commercial estradiol immunoassays. *Endocrinology*, 152(11), 4443-4447.
- Halling, S. M., Peterson-Burch, B. D., Bricker, B. J., Zuerner, R. L., Qing, Z., Li, L.-L., Kapur, V., Alt, D. P., & Olsen, S. C. (2005). Completion of the genome sequence of *Brucella abortus* and comparison to the highly similar genomes of *Brucella melitensis* and *Brucella suis*. *Journal of Bacteriology*, 187(8), 2715-2726.
- Handa, R. J., & Weiser, M. J. (2014). Gonadal steroid hormones and the hypothalamo–pituitary–adrenal axis. *Frontiers in neuroendocrinology*, 35(2), 197-220.
- Handelsman, D. (2001). Estrogens and falling sperm counts. *Reproduction, Fertility and Development*, 13(4), 317-324.
- Hartady, T., Saad, M. Z., Bejo, S. K., & Salisi, M. S. (2014). Clinical human brucellosis in Malaysia: a case report. *Asian Pacific Journal of Tropical Disease*, 4(2), 150-153.
- He, Y. (2012). Analyses of *Brucella* pathogenesis, host immunity, and vaccine targets using systems biology and bioinformatics. *Frontiers in cellular and infection microbiology*, 2, 2.
- Heumann, D., & Roger, T. (2002). Initial responses to endotoxins and Gram-negative bacteria. *Clinica Chimica Acta*, 323(1), 59-72.
- High, K. P., Prasad, R., Marion, C. R., Schurig, G. G., Boyle, S. M., & Sriranganathan, N. (2007). Outcome and immune responses after *Brucella abortus* infection in young adult and aged mice. *Biogerontology*, 8(5), 583-593.
- Hinić, V., Brodard, I., Thomann, A., Holub, M., Miserez, R., & Abril, C. (2009). IS 711-based real-time PCR assay as a tool for detection of *Brucella* spp. in wild boars and comparison with bacterial isolation and serology. *BMC veterinary research*, 5(1), 1.
- Holmes, A. G., Watt, M., Carey, A. L., & Febbraio, M. A. (2004). Ionomycin, but not physiologic doses of epinephrine, stimulates skeletal muscle interleukin-6 mRNA expression and protein release. *Metabolism*, 53(11), 1492-1495.

- Hort, G., Weisenburger, J., Borsdorf, B., Peters, C., Banai, M., Hahn, H., Jacob, J., & Mielke, M. (2003). Delayed type hypersensitivity-associated disruption of splenic periarteriolar lymphatic sheaths coincides with temporary loss of IFN- $\gamma$  production and impaired eradication of bacteria in *Brucella abortus*-infected mice. *Microbes and infection*, 5(2), 95-106.
- Hort, G., Weisenburger, J., Borsdorf, B., Peters, C., Banai, M., Hahn, H., Jacob, J., & Mielke, M. (2003). Delayed type hypersensitivity-associated disruption of splenic periarteriolar lymphatic sheaths coincides with temporary loss of IFN- $\gamma$  production and impaired eradication of bacteria in *Brucella abortus*-infected mice. *Microbes and infection*, 5(2), 95-106.
- Hort, G., Weisenburger, J., Borsdorf, B., Peters, C., Banai, M., Hahn, H., Jacob, J., & Mielke, M. (2003). Delayed type hypersensitivity-associated disruption of splenic periarteriolar lymphatic sheaths coincides with temporary loss of IFN- $\gamma$  production and impaired eradication of bacteria in *Brucella abortus*-infected mice. *Microbes and infection*, 5(2), 95-106.
- Hsing, A. W., Stanczyk, F. Z., Bélanger, A., Schroeder, P., Chang, L., Falk, R. T., & Fears, T. R. (2007). Reproducibility of serum sex steroid assays in men by RIA and mass spectrometry. *Cancer Epidemiology Biomarkers & Prevention*, 16(5), 1004-1008.
- Hsu, H.-C., Yang, P., Wang, J., Wu, Q., Myers, R., Chen, J., Yi, J., Guentert, T., Tousson, A., & Stanus, A. L. (2008). Interleukin 17-producing T helper cells and interleukin 17 orchestrate autoreactive germinal center development in autoimmune BXD2 mice. *Nature immunology*, 9(2), 166-175.
- Hu, J., Zhang, Z., Shen, W.-J., & Azhar, S. (2010). Cellular cholesterol delivery, intracellular processing and utilization for biosynthesis of steroid hormones. *Nutrition & metabolism*, 7(1), 1.
- Huber, J., & Nicoletti, P. (1986). Comparison of the results of card, rivanol, complement-fixation, and milk ring tests with the isolation rate of *Brucella abortus* from cattle. *American journal of veterinary research*, 47(7), 1529-1531.
- Ibis, C., Sezer, A., Batman, A. K., Baydar, S., Eker, A., Unlu, E., Kuloglu, F., Cakir, B., & Coskun, I. (2007). Acute abdomen caused by brucellar hepatic abscess. *Asian Journal of Surgery*, 30(4), 283-285.
- Isobe, N., Akita, M., Nakao, T., Yamashiro, H., & Kubota, H. (2005). Pregnancy diagnosis based on the fecal progesterone concentration in beef and dairy heifers and beef cows. *Animal reproduction science*, 90(3), 211-218.
- Jesse, F. F. A., Abba, Y., Tijjani, A., Sadiq, M. A., Konto, M., Adamu, L., Wahid, A. H., MohdAzmi, M. L., Eric, L. T. C., & Ab Rahman, M. F. (2016). Gonadophyseal lesions and reproductive hormonal changes in *Brucella melitensis*-infected mice and its lipopolysaccharides (LPSs). *Comparative Clinical Pathology*, 25(1), 31-36.

- Jove, M., Iñaniga, A., Laguna, J. C., & Vázquez-Carrera, M. (2005). Palmitate-induced interleukin 6 production is mediated by protein kinase C and nuclear-factor  $\kappa$ B activation and leads to glucose transporter 4 down-regulation in skeletal muscle cells. *Endocrinology*, 146(7), 3087-3095.
- Júnior, C. C., Moustakas, V., Xavier, M., Costa, E., Costa, L., Silva, T., Paixão, T., Borges, A., Gouveia, A., & Santos, R. (2012). Andrological, pathologic, morphometric, and ultrasonographic findings in rams experimentally infected with *Brucella ovis*. *Small Ruminant Research*, 102(2), 213-222.
- Kahl-McDonagh, M., Arenas-Gamboa, A., & Ficht, T. (2007). Aerosol infection of BALB/c mice with *Brucella melitensis* and *Brucella abortus* and protective efficacy against aerosol challenge. *Infection and immunity*, 75(10), 4923-4932.
- Kakar, S. (2015). Cytokines evolution: Role in various diseases. *Current Medicine Research and Practice*, 5(4), 176-182.
- Kallen, K.-J., Grötzinger, J., Lelièvre, E., Vollmer, P., Aasland, D., Renné, C., Müllberg, J., zum Büschenfelde, K.-H. M., Gascan, H., & Rose-John, S. (1999). Receptor recognition sites of cytokines are organized as exchangeable modules Transfer of the leukemia inhibitory factor receptor-binding site from ciliary neurotrophic factor to interleukin-6. *Journal of Biological Chemistry*, 274(17), 11859-11867.
- Kamimura, D., Ishihara, K., & Hirano, T. (2003). IL-6 signal transduction and its physiological roles: the signal orchestration model *Reviews of physiology, biochemistry and pharmacology* (pp. 1-38): Springer.
- Kaufmann, A. F., Meltzer, M. I., & Schmid, G. P. (1997). The economic impact of a bioterrorist attack: are prevention and postattack intervention programs justifiable? *Emerging infectious diseases*, 3(2), 83.
- Khani, Y., Mollajan, A., & Rahimi, F. (2015). Inappropriate Dietary and Occupational Patterns: Major Risk Factors Associated With Brucellosis in the Area Covered by Karaj Health Center No. 2. *International Journal of Enteric Pathogens*, 3(4).
- Khosla, S., Amin, S., Singh, R. J., Atkinson, E. J., Melton III, L. J., & Riggs, B. L. (2008). Comparison of sex steroid measurements in men by immunoassay versus mass spectroscopy and relationships with cortical and trabecular volumetric bone mineral density. *Osteoporosis International*, 19(10), 1465-1471.
- Kim, H. O., Kim, H.-S., Youn, J.-C., Shin, E.-C., & Park, S. (2011). Serum cytokine profiles in healthy young and elderly population assessed using multiplexed bead-based immunoassays. *Journal of translational medicine*, 9(1), 1.
- Kim, S., Lee, D. S., Watanabe, K., Furuoka, H., Suzuki, H., & Watarai, M. (2005). Interferon- $\gamma$  promotes abortion due to *Brucella* infection in pregnant mice. *BMC microbiology*, 5(1), 1.

- Kleemann, R., Zadelaar, S., & Kooistra, T. (2008). Cytokines and atherosclerosis: a comprehensive review of studies in mice. *Cardiovascular research*.
- Kleemann, R., Zadelaar, S., & Kooistra, T. (2008). Cytokines and atherosclerosis: a comprehensive review of studies in mice. *Cardiovascular research*.
- Ko, J., & Splitter, G. A. (2003). Molecular host-pathogen interaction in brucellosis: current understanding and future approaches to vaccine development for mice and humans. *Clinical microbiology reviews*, 16(1), 65-78.
- Kopf, M., Baumann, H., Freer, G., Freudenberg, M., Lamers, M., Kishimoto, T., Zinkernagel, R., Bluethmann, H., & Köhler, G. (1994). Impaired immune and acute-phase responses in interleukin-6-deficient mice. *Nature*, 368(6469), 339-342.
- Korhonen, H., Syväoja, E. L., Ahola-Luttila, H., Sivelä, S., Kopola, S., Husu, J., & Kosunen, T. (1995). Bactericidal effect of bovine normal and immune serum, colostrum and milk against *Helicobacter pylori*. *Journal of Applied Bacteriology*, 78(6), 655-662.
- Kortepeter, M. G., & Parker, G. W. (1999). Potential biological weapons threats. *Emerging infectious diseases*, 5(4), 523.
- Krone, N., Hughes, B. A., Lavery, G. G., Stewart, P. M., Arlt, W., & Shackleton, C. H. (2010). Gas chromatography/mass spectrometry (GC/MS) remains a pre-eminent discovery tool in clinical steroid investigations even in the era of fast liquid chromatography tandem mass spectrometry (LC/MS/MS). *The Journal of steroid biochemistry and molecular biology*, 121(3), 496-504.
- Kushnir, M. M., Rockwood, A. L., & Bergquist, J. (2010). Liquid chromatography–tandem mass spectrometry applications in endocrinology. *Mass spectrometry reviews*, 29(3), 480-502.
- Lapaque, N., Forquet, F., De Chastellier, C., Mishal, Z., Jolly, G., Moreno, E., Moriyon, I., Heuser, J. E., He, H. T., & Gorvel, J. P. (2006). Characterization of *Brucella abortus* lipopolysaccharide macrodomains as mega rafts. *Cellular microbiology*, 8(2), 197-206.
- Lapaque, N., Moriyon, I., Moreno, E., & Gorvel, J.-P. (2005). *Brucella* lipopolysaccharide acts as a virulence factor. *Current opinion in microbiology*, 8(1), 60-66.
- Lim, J. J., Kim, D. H., Lee, J. J., Kim, D. G., Min, W., Lee, H. J., Rhee, M. H., & Kim, S. (2012). Protective effects of recombinant *Brucella abortus* Omp28 against infection with a virulent strain of *Brucella abortus* 544 in mice. *Journal of veterinary science*, 13(3), 287-292.
- Liu, G., Qi, M., Hutchinson, M. R., Yang, G., & Goldys, E. M. (2016). Recent advances in cytokine detection by immunosensing. *Biosensors and Bioelectronics*, 79, 810-821.

- Lopez, A., Hitos, F., Perez, A., & Navarro-Fierro, R. (1984). Lung lesions in bovine fetuses aborted by *Brucella abortus* *Canadian journal of comparative medicine*, 48(3), 275.
- Lucero, N., Ayala, S., Escobar, G., & Jacob, N. (2008). *Brucella* isolated in humans and animals in Latin America from 1968 to 2006. *Epidemiology and infection*, 136(04), 496-503.
- Luo, D., Ni, B., Li, P., Shi, W., Zhang, S., Han, Y., Mao, L., He, Y., Wu, Y., & Wang, X. (2006). Protective immunity elicited by a divalent DNA vaccine encoding both the L7/L12 and Omp16 genes of *Brucella abortus* in BALB/c mice. *Infection and immunity*, 74(5), 2734-2741.
- Macedo, G. C., Magnani, D. M., Carvalho, N. B., Bruna-Romero, O., Gazzinelli, R. T., & Oliveira, S. C. (2008). Central role of MyD88-dependent dendritic cell maturation and proinflammatory cytokine production to control *Brucella abortus* infection. *The Journal of Immunology*, 180(2), 1080-1087.
- Magnani, D., Harms, J., Durward, M., & Splitter, G. (2009). Nondividing but metabolically active gamma-irradiated *Brucella melitensis* is protective against virulent *B. melitensis* challenge in mice. *Infection and immunity*, 77(11), 5181-5189.
- Manterola, L., Tejero-Garcés, A., Ficapal, A., Shopayeva, G., Blasco, J., Marin, C., & Lopez-Goni, I. (2003). Evaluation of a PCR test for the diagnosis of *Brucella ovis* infection in semen samples from rams. *Veterinary microbiology*, 92(1), 65-72.
- Mantur, B. G., Biradar, M. S., Bidri, R. C., Mulimani, M. S., Veerappa, K., Kariholu, P., Patil, S. B., & Mangalgi, S. S. (2006). Protean clinical manifestations and diagnostic challenges of human brucellosis in adults: 16 years' experience in an endemic area. *Journal of medical microbiology*, 55(7), 897-903.
- Martins, R. D. C., Irache, J. M., Blasco, J. M., Muñoz, M. P., Marín, C. M., Grilló, M. J., De Miguel, M. J., Barberán, M., & Gamazo, C. (2010). Evaluation of particulate acellular vaccines against *Brucella ovis* infection in rams. *Vaccine*, 28(17), 3038-3046.
- Martirosyan, A., Moreno, E., & Gorvel, J. P. (2011). An evolutionary strategy for a stealthy intracellular *Brucella* pathogen. *Immunological reviews*, 240(1), 211-234.
- Martirosyan, A., Moreno, E., & Gorvel, J. P. (2011). An evolutionary strategy for a stealthy intracellular *Brucella* pathogen. *Immunological reviews*, 240(1), 211-234.
- Mazokopakis, E., Christias, E., & Kofteridis, D. (2003). Acute brucellosis presenting with erythema nodosum. *European journal of epidemiology*, 18(9), 913-915.

- Megersa, B., Biffa, D., Abunna, F., Regassa, A., Godfroid, J., & Skjerve, E. (2011). Seroprevalence of brucellosis and its contribution to abortion in cattle, camel, and goat kept under pastoral management in Borana, Ethiopia. *Tropical Animal Health and Production*, 43(3), 651-656.
- Megid, J., Antonio Mathias, L., & A Robles, C. (2010). Clinical manifestations of brucellosis in domestic animals and humans. *The Open Veterinary Science Journal*, 4(1).
- Mellon, S. H., & Griffin, L. D. (2002). Neurosteroids: biochemistry and clinical significance. *Trends in Endocrinology & Metabolism*, 13(1), 35-43.
- Mellon, S. H., & Griffin, L. D. (2002). Synthesis, regulation, and function of neurosteroids. *Endocrine research*, 28(4), 463-463.
- Memish, Z., Mah, M. W., Al Mahmoud, S., Al Shaalan, M., & Khan, M. Y. (2000). *Brucella* bacteraemia: clinical and laboratory observations in 160 patients. *Journal of Infection*, 40(1), 59-63.
- Memish, Z., & Venkatesh, S. (2001). *Brucellar* epididymo-orchitis in Saudi Arabia: a retrospective study of 26 cases and review of the literature. *BJU international*, 88(1), 72-76.
- Mense, M. G., Borschel, R. H., Wilhelmsen, C. L., Pitt, M. L., & Hoover, D. L. (2004). Pathologic changes associated with brucellosis experimentally induced by aerosol exposure in rhesus macaques (*Macaca mulatta*). *American journal of veterinary research*, 65(5), 644-652.
- Mense, M. G., Verg, L. L. V. D., Bhattacharjee, A. K., Garrett, J. L., Hart, J. A., Lindler, L. E., Hadfield, T. L., & Hoover, D. L. (2001). Bacteriologic and histologic features in mice after intranasal inoculation of *Brucella melitensis*. *American journal of veterinary research*, 62(3), 398-405.
- Menshaw, A., Perez-Sancho, M., Garcia-Seco, T., Hosein, H. I., García, N., Martinez, I., Sayour, A. E., Goyache, J., Azzam, R. A., & Dominguez, L. (2014). Assessment of genetic diversity of zoonotic *Brucella* spp. recovered from livestock in Egypt using multiple locus VNTR analysis. *BioMed Research International*, 2014.
- Mert, A., Ozaras, R., Tabak, F., Bilir, M., Yilmaz, M., Kurt, C., Ongoren, S., Tanriverdi, M., & Ozturk, R. (2003). The sensitivity and specificity of *Brucella* agglutination tests. *Diagnostic microbiology and infectious disease*, 46(4), 241-243.
- Mestas, J., & Hughes, C. C. (2004). Of mice and not men: differences between mouse and human immunology. *The Journal of Immunology*, 172(5), 2731-2738.
- Miranda, R. T., Gimeno, A. E., Rodriguez, T. F., de Arriba, J. J., Olmo, D. G. a., & Solera, J. (2001). Acute cholecystitis caused by *Brucella melitensis*: case report and review. *Journal of Infection*, 42(1), 77-78.

- Moreno, E., Cloeckeaert, A., & Moriyón, I. (2002). *Brucella* evolution and taxonomy. *Veterinary microbiology*, 90(1), 209-227.
- Moriyón, I., Grilló, M. J., Monreal, D., González, D., Marín, C., López-Goñi, I., Mainar-Jaime, R. C., Moreno, E., & Blasco, J. M. (2004). Rough vaccines in animal brucellosis: structural and genetic basis and present status. *Veterinary research*, 35(1), 1-38.
- Morris, M. C., Gilliam, E. A., & Li, L. (2015). Innate immune programming by endotoxin and its pathological consequences. *Frontiers in immunology*, 5, 680.
- Morrison, D. C. (1983). Bacterial endotoxins and pathogenesis. *Review of Infectious Diseases*, 5(Supplement 4), S733-S747.
- Muma, J., Toft, N., Oloya, J., Lund, A., Nielsen, K., Samui, K., & Skjerve, E. (2007). Evaluation of three serological tests for brucellosis in naturally infected cattle using latent class analysis. *Veterinary microbiology*, 125(1), 187-192.
- Munoz, P., Marin, C., Monreal, D., Gonzalez, D., Garin-Bastuji, B., Diaz, R., Mainar-Jaime, R., Moriyon, I., & Blasco, J. (2005). Efficacy of several serological tests and antigens for diagnosis of bovine brucellosis in the presence of false-positive serological results due to *Yersinia enterocolitica* O: 9. *Clinical and Diagnostic Laboratory Immunology*, 12(1), 141-151.
- Murad, M. H., Elamin, M. B., Garcia, M. Z., Mullan, R. J., Murad, A., Erwin, P. J., & Montori, V. M. (2010). Hormonal therapy and sex reassignment: A systematic review and meta-analysis of quality of life and psychosocial outcomes. *Clinical endocrinology*, 72(2), 214-231.
- Murphy, E. A., Parent, M., Sathiyaseelan, J., Jiang, X., & Baldwin, C. L. (2001). Immune control of *Brucella abortus* 2308 infections in BALB/c mice. *FEMS Immunology & Medical Microbiology*, 32(1), 85-88.
- Nasruddin, N. S., Mazlan, M., Saad, M. Z., Hamzah, H., & Sabri, J. (2014). Histopathology and immunohistochemistry assessments of acute experimental infection by *Brucella melitensis* in bucks. *Open Journal of Pathology*, 4(02), 54.
- Neta, A. V. C., Mol, J. P., Xavier, M. N., Paixão, T. A., Lage, A. P., & Santos, R. L. (2010). Pathogenesis of bovine brucellosis. *The Veterinary Journal*, 184(2), 146-155.
- Nielsen. (2002). Diagnosis of brucellosis by serology. *Veterinary microbiology*, 90(1), 447-459.
- Nielsen, Gall, D., Smith, P., Kelly, W., Yeo, J., Kenny, K., Heneghan, T., McNamara, S., Maher, ., & O'Connor, . (2001). Fluorescence polarization assay for the diagnosis of bovine brucellosis: adaptation to field use. *Veterinary microbiology*, 80(2), 163-170.

- Nielsen, Smith, P., Yu, W., Elmgren, C., Halbert, G., Nicoletti, P., Perez, B., Conde, S., Samartino, L., & Nicola, A. (2008). Validation of a second generation competitive enzyme immunoassay (CELISA) for the diagnosis of brucellosis in various species of domestic animals. *Veterinary immunology and immunopathology*, 125(3), 246-250.
- Nielsen, K., Gall, D., Smith, P., Kelly, W., Yeo, J., Kenny, K., Heneghan, T., McNamara, S., Maher, ., & O'Connor, . (2001). Fluorescence polarization assay for the diagnosis of bovine brucellosis: adaptation to field use. *Veterinary microbiology*, 80(2), 163-170.
- O'Callaghan, D., & Whatmore, A. M. (2011). *Brucella* genomics as we enter the multi-genome era. *Briefings in functional genomics*, 10(6), 334-341.
- O'Callaghan, D., & Whatmore, A. M. (2011). *Brucella* genomics as we enter the multi-genome era. *Briefings in functional genomics*, 10(6), 334-341.
- Oliveira, S. C., de Oliveira, F. S., Macedo, G. C., de Almeida, L. A., & Carvalho, N. B. (2008). The role of innate immune receptors in the control of *Brucella abortus* infection: toll-like receptors and beyond. *Microbes and infection*, 10(9), 1005-1009.
- Oliveira, S. C., Giambartolomei, G. H., & Cassataro, J. (2011). Confronting the barriers to develop novel vaccines against brucellosis. *Expert review of vaccines*, 10(9), 1291-1305.
- Oliveira, S. C., & Splitter, G. A. (1996). Immunization of mice with recombinant L7L12 ribosomal protein confers protection against *Brucella abortus* infection. *Vaccine*, 14(10), 959-962.
- Olsen, & Tatum, F. (2010). Bovine brucellosis. *Veterinary Clinics of North America: Food Animal Practice*, 26(1), 15-27.
- Olsen, S., Boyle, S., Schurig, G., & Sriranganathan, N. (2009). Immune responses and protection against experimental challenge after vaccination of bison with *Brucella abortus* strain RB51 or RB51 overexpressing superoxide dismutase and glycosyltransferase genes. *Clinical and Vaccine Immunology*, 16(4), 535-540.
- Osman, A. Y. (2016). Immuno-pathophysiological responses of mouse model to experimental infection with *Brucella melitensis* and its lipopolysaccharides via intraperitoneal route. *Microbial Pathogenesis*,
- Padilla Poester, F., Nielsen, K., Ernesto Samartino, L., & Ling Yu, W. (2010). Diagnosis of brucellosis. *The Open Veterinary Science Journal*, 4(1).
- Paixão, T. A., Roux, C. M., Den Hartigh, A. B., Sankaran-Walters, S., Dandekar, S., Santos, R. L., & Tsolis, R. M. (2009). Establishment of systemic *Brucella melitensis* infection through the digestive tract requires urease, the type IV

secretion system, and lipopolysaccharide O antigen. *Infection and immunity*, 77(10), 4197-4208.

Pallares, P., & Gonzalez-Bulnes, A. (2009). A new method for induction and synchronization of oestrus and fertile ovulations in mice by using exogenous hormones. *Laboratory animals*, 43(3), 295-299.

Palme, R., Rettenbacher, S., Touma, C., El-Bahr, S., & Möstl, E. (2005). Stress hormones in mammals and birds: comparative aspects regarding metabolism, excretion, and noninvasive measurement in fecal samples. *Annals of the New York Academy of Sciences*, 1040(1), 162-171.

Papatsoris, A. G., Mpadra, F. A., Karamouzis, M. V., & Frangides, C. Y. (2002). Endemic brucellar epididymo-orchitis: a 10-year experience. *International Journal of Infectious Diseases*, 6(4), 309-313.

Pappas, G., & Papadimitriou, P. (2007). Challenges in *Brucella* bacteraemia. *International journal of antimicrobial agents*, 30, 29-31.

Pappas, G., Papadimitriou, P., Akritidis, N., Christou, L., & Tsianos, E. V. (2006). The new global map of human brucellosis. *The Lancet infectious diseases*, 6(2), 91-99.

Pasquevich, K. A., Estein, S. M., Samartino, C. G., Zwerdling, A., Coria, L. M., Barrionuevo, P., Fossati, C. A., Giambartolomei, G. H., & Cassataro, J. (2009). Immunization with recombinant *Brucella* species outer membrane protein Omp16 or Omp19 in adjuvant induces specific CD4<sup>+</sup> and CD8<sup>+</sup> T cells as well as systemic and oral protection against *Brucella abortus* infection. *Infection and immunity*, 77(1), 436-445.

Pasquevich, K. A., Estein, S. M., Samartino, C. G., Zwerdling, A., Coria, L. M., Barrionuevo, P., Fossati, C. A., Giambartolomei, G. H., & Cassataro, J. (2009). Immunization with recombinant *Brucella* species outer membrane protein Omp16 or Omp19 in adjuvant induces specific CD4<sup>+</sup> and CD8<sup>+</sup> T cells as well as systemic and oral protection against *Brucella abortus* infection. *Infection and immunity*, 77(1), 436-445.

Patterson, J., Deyoe, B., & Stone, S. (1976). Identification of immunoglobulins associated with complement fixation, agglutination, and low pH buffered antigen tests for brucellosis. *American journal of veterinary research*, 37(3), 319-324.

Pavone, I. R. (2015). Animal Experimentation and Animal Welfare in the Context of the European Union: Reflections on the Directive 2010/63/EU and Its Transposition in Italy. *BioLaw Journal-Rivista di BioDiritto*, 5(3), 75-97.

Payne, A. H., & Hales, D. B. (2004). Overview of steroidogenic enzymes in the pathway from cholesterol to active steroid hormones. *Endocrine reviews*, 25(6), 947-970.

- Phillips, R., Elzer, P., Robertson, G., Hagius, S., Walker, J., Fatemi, M., Enright, F., & Roop, R. (1997). A *Brucella melitensis* high-temperature-requirement A (htrA) deletion mutant is attenuated in goats and protects against abortion. *Research in veterinary science*, 63(2), 165-167.
- Poester, F., Samartino, L., & Santos, R. (2013). Pathogenesis and pathobiology of brucellosis in livestock. *Rev Sci Tech*, 32(32), 105-115.
- Pouliot, K., Pan, N., Wang, S., Lu, S., Lien, E., & Goguen, J. D. (2007). Evaluation of the role of LcrV-Toll-like receptor 2-mediated immunomodulation in the virulence of *Yersinia pestis*. *Infection and immunity*, 75(7), 3571-3580.
- Radostits, O., Gay, C., Hinchcliff, K., & Constable, P. (2007). Brucellosis caused by *Brucella abortus*. *Veterinary Medicine*. 10th edn., Elsevier Saunders, London, UK.
- Ragan, V. E. (2002). The animal and plant health inspection service (APHIS) brucellosis eradication program in the United States. *Veterinary microbiology*, 90(1), 11-18.
- Rajashekara, G., Glover, D. A., Krepps, M., & Splitter, G. A. (2005). Temporal analysis of pathogenic events in virulent and avirulent *Brucella melitensis* infections. *Cellular microbiology*, 7(10), 1459-1473.
- Ramin, B., Malhotra, J., Schreiber, Y., & MacPherson, P. (2013). Infective endocarditis in a new immigrant. *Canadian Family Physician*, 59(6), 644-646.
- Rauh, M. (2010). Steroid measurement with LC-MS/MS. Application examples in pediatrics. *The Journal of steroid biochemistry and molecular biology*, 121(3), 520-527.
- Rhyan, J. C., Quinn, W. J., Stackhouse, L. S., Henderson, J. J., Ewalt, D. R., Payeur, J. B., Johnson, M., & Meagher, M. (1994). Abortion caused by *Brucella abortus* biovar 1 in a free-ranging bison (*Bison bison*) from Yellowstone National Park. *Journal of Wildlife Diseases*, 30(3), 445-446.
- Riffle, B. W., Henderson, W. M., & Laws, S. C. (2013). Measurement of steroids in rats after exposure to an endocrine disruptor: Mass spectrometry and radioimmunoassay demonstrate similar results. *Journal of pharmacological and toxicological methods*, 68(3), 314-322.
- Robinson, A., & Production, A. (2003). *Guidelines for coordinated human and animal brucellosis surveillance*: FAO Rome, Italy.
- Sam, I.-C., Karunakaran, R., Kamarulzaman, A., Ponnampalavanar, S., Omar, S. S., Ng, K., Yusof, M. M., Hooi, P., Jafar, F., & Abubakar, S. (2012). A large exposure to *Brucella melitensis* in a diagnostic laboratory. *Journal of Hospital Infection*, 80(4), 321-325.

- Samadi, A., Ababneh, M., Giadinis, N., & Lafi, S. (2010). Ovine and caprine brucellosis (*Brucella melitensis*) in aborted animals in Jordanian Sheep and goat flocks. *Veterinary medicine international*, 2010.
- Samartino, L., Truax, R., & Enright, F. (1994). Invasion and replication of *Brucella abortus* in three different trophoblastic cell lines. *Journal of Veterinary Medicine, Series B*, 41(1-10), 229-236.
- Sathiyaseelan, J., Goenka, R., Parent, M., Benson, R. M., Murphy, E. A., Fernandes, D. M., Foulkes, A. S., & Baldwin, C. L. (2006). Treatment of *Brucella*-susceptible mice with IL-12 increases primary and secondary immunity. *Cellular immunology*, 243(1), 1-9.
- Scheller, J., Chalaris, A., Schmidt-Arras, D., & Rose-John, S. (2011). The pro-and anti-inflammatory properties of the cytokine interleukin-6. *Biochimica et Biophysica Acta (BBA)-Molecular Cell Research*, 1813(5), 878-888.
- Scholz, H. C., Hubalek, Z., Sedl ček, I., Vergnaud, G., Tomaso, H., Al Dahouk, S., Melzer, F., Kämpfer, P., Neubauer, H., & Cloeckeaert, A. (2008). *Brucella microti* sp. nov., isolated from the common vole *Microtus arvalis*. *International journal of systematic and evolutionary microbiology*, 58(2), 375-382.
- Schröder, C., Jacob, A., Tonack, S., Radon, T. P., Sill, M., Zucknick, M., Rüffer, S., Costello, E., Neoptolemos, J. P., & Crnogorac-Jurcevic, T. (2010). Dual-color proteomic profiling of complex samples with a microarray of 810 cancer-related antibodies. *Molecular & cellular proteomics*, 9(6), 1271-1280.
- Seleem, M. N., Boyle, S. M., & Sriranganathan, N. (2010). Brucellosis: a re-emerging zoonosis. *Veterinary microbiology*, 140(3), 392-398.
- Shackleton, C. (2010). Clinical steroid mass spectrometry: a 45-year history culminating in HPLC–MS/MS becoming an essential tool for patient diagnosis. *The Journal of steroid biochemistry and molecular biology*, 121(3), 481-490.
- Shahdaezadeh, S., Edalatmanesh, M., & Moghadasi, M. (2015). evaluation of sex hormones (fsh, estrogen and testosterone) changes during follicular and luteal phases and sexual dysfunction in women with multiple sclerosis.
- Silva, T., Costa, E. A., Paixão, T. A., Tsolis, R. M., & Santos, R. L. (2011). Laboratory animal models for brucellosis research. *BioMed Research International*, 2011.
- Silva, T., Costa, E. A., Paixão, T. A., Tsolis, R. M., & Santos, R. L. (2011). Laboratory animal models for brucellosis research. *BioMed Research International*, 2011.
- Sintayehu, G., Melesse, B., Abayneh, D., Sintayehu, A., Melaku, S., Alehegne, W., Mesfin, S., De Blas, I., Casal, J., & Allepuz, A. (2015). Epidemiological survey of brucellosis in sheep and goats in selected pastoral and agro-pastoral

lowlands of Ethiopia. *Revue scientifique et technique (International Office of Epizootics)*, 34(3), 881-893.

Smits, H. L., & Kadri, S. M. (2005). Brucellosis in India: a deceptive infectious disease. *Indian Journal of Medical Research*, 122(5), 375.

Stanczyk, F. Z., & Clarke, N. J. (2010). Advantages and challenges of mass spectrometry assays for steroid hormones. *The Journal of steroid biochemistry and molecular biology*, 121(3), 491-495.

Steensberg, A., Keller, C., Starkie, R. L., Osada, T., Febbraio, M. A., & Pedersen, B. K. (2002). IL-6 and TNF- $\alpha$  expression in, and release from, contracting human skeletal muscle. *American Journal of Physiology-Endocrinology and Metabolism*, 283(6), E1272-E1278.

Steimer, T., Python, A., Schulz, P. E., & Aubry, J.-M. (2007). Plasma corticosterone, dexamethasone (DEX) suppression and DEX/CRH tests in a rat model of genetic vulnerability to depression. *Psychoneuroendocrinology*, 32(5), 575-579.

Stelwagen, K., Carpenter, E., Haigh, B., Hodgkinson, A., & Wheeler, T. (2009). Immune components of bovine colostrum and milk. *Journal of animal science*, 87(13\_suppl), 3-9.

Stenken, J. A., & Poschenrieder, A. J. (2015). Bioanalytical chemistry of cytokines—A review. *Analytica chimica acta*, 853, 95-115.

Sutra, L., Caffin, J., & Dubray, G. (1986). Role of milk immunoglobulins in the *Brucella* milk ring test. *Veterinary microbiology*, 12(4), 359-366.

Tabynov, K., Kydyrbayev, Z., Ryskeldinova, S., Yespembetov, B., Zinina, N., Assanzhanova, N., Kozhamkulov, Y., Inkarebekov, D., Gotskina, T., & Sansyzbay, A. (2014). Novel influenza virus vectors expressing *Brucella* L7/L12 or Omp16 proteins in cattle induced a strong T-cell immune response, as well as high protectiveness against *B. abortus* infection. *Vaccine*, 32(18), 2034-2041.

Tachibana, M., Watanabe, K., Yamasaki, Y., Suzuki, H., & Watarai, M. (2008). Expression of heme oxygenase-1 is associated with abortion caused by *Brucella abortus* infection in pregnant mice. *Microbial Pathogenesis*, 45(2), 105-109.

Taleski, V., Zerva, L., Kantardjiev, T., Cvetnic, Z., Erski-Biljic, M., Nikolovski, B., Bosnjakovski, J., Katalinic-Jankovic, V., Panteliadou, A., & Stojkoski, S. (2002). An overview of the epidemiology and epizootology of brucellosis in selected countries of Central and Southeast Europe. *Veterinary microbiology*, 90(1), 147-155.

Tan, Y., & Kagan, J. C. (2014). A cross-disciplinary perspective on the innate immune responses to bacterial lipopolysaccharide. *Molecular cell*, 54(2), 212-223.

- Tarantola, A., Abiteboul, D., & Rachline, A. (2006). Infection risks following accidental exposure to blood or body fluids in health care workers: a review of pathogens transmitted in published cases. *American journal of infection control*, 34(6), 367-375.
- Thimm, B. M. (2013). *Brucellosis: Distribution in man, domestic and wild animals*: Springer Science & Business Media.
- Tobias, L., Cordes, D., & Schurig, G. (1993). Placental pathology of the pregnant mouse inoculated with *Brucella abortus* strain 2308. *Veterinary Pathology Online*, 30(2), 119-129.
- Tuck, S., & Francis, R. (2008). Testosterone, bone and osteoporosis *Advances in the Management of Testosterone Deficiency* (Vol. 37, pp. 123-132): Karger Publishers.
- Velikovsky, C. A., Goldbaum, F. A., Cassataro, J., Estein, S., Bowden, R. A., Bruno, L., Fossati, C. A., & Giambartolomei, G. H. (2003). *Brucella* lumazine synthase elicits a mixed Th1-Th2 immune response and reduces infection in mice challenged with *Brucella abortus* 544 independently of the adjuvant formulation used. *Infection and immunity*, 71(10), 5750-5755.
- Verma, S. K., Jain, S., & Kumar, S. (2012). Immunogenicity and protective potential of a bacterially expressed recombinant dihydrolipoamide succinyltransferase (rE2o) of *Brucella abortus* in BALB/c mice. *World Journal of Microbiology and Biotechnology*, 28(7), 2487-2495.
- Villarino, A. V., Tato, C. M., Stumhofer, J. S., Yao, Z., Cui, Y. K., Hennighausen, L., O'Shea, J. J., & Hunter, C. A. (2007). Helper T cell IL-2 production is limited by negative feedback and STAT-dependent cytokine signals. *The Journal of experimental medicine*, 204(1), 65-71.
- Vishvanath, N., & Kole, C. (2008). *Genome mapping and genomics in animal-associated microbes*: Springer Science & Business Media.
- Watanabe, K., Iwai, N., Tachibana, M., Furuoka, H., Suzuki, H., & Watarai, M. (2008). Regulated upon activation normal T-cell expressed and secreted (RANTES) contributes to abortion caused by *Brucella abortus* infection in pregnant mice. *Journal of Veterinary Medical Science*, 70(7), 681-686.
- Weigert, C., Brodbeck, K., Staiger, H., Kausch, C., Machicao, F., Häring, H. U., & Schleicher, E. D. (2004). Palmitate, but not unsaturated fatty acids, induces the expression of interleukin-6 in human myotubes through proteasome-dependent activation of nuclear factor- $\kappa$ B. *Journal of Biological Chemistry*, 279(23), 23942-23952.
- Wetendorf, M., & DeMayo, F. J. (2014). Progesterone receptor signaling in the initiation of pregnancy and preservation of a healthy uterus. *The International journal of developmental biology*, 58, 95.

- Whatmore, A. M., Davison, N., Cloeckaert, A., Al Dahouk, S., Zygmunt, M. S., Brew, S. D., Perrett, L. L., Koylass, M. S., Vergnaud, G., & Quance, C. (2014). *Brucella papionis* sp. nov., isolated from baboons (*Papio* spp.). *International journal of systematic and evolutionary microbiology*, 64(12), 4120-4128.
- Whicher, J., & Evans, S. (1990). Cytokines in disease. *Clinical chemistry*, 36(7), 1269-1281.
- Wolfson, M. L., Aisemberg, J., Salazar, A. I., Rubio, A. P. D., Vercelli, C. A., & Franchi, A. M. (2013). Progesterone reverts LPS-reduced FAAH activity in murine peripheral blood mononuclear cells by a receptor-mediated fashion. *Molecular and cellular endocrinology*, 381(1), 97-105.
- Wyatt, H. (2005). How Themistocles Zammit found Malta Fever (brucellosis) to be transmitted by the milk of goats. *Journal of the Royal Society of Medicine*, 98(10), 451-454.
- Xavier, M., Paixão, T., Poester, F., Lage, A., & Santos, R. (2009). Pathological, immunohistochemical and bacteriological study of tissues and milk of cows and fetuses experimentally infected with *Brucella abortus*. *Journal of comparative pathology*, 140(2), 149-157.
- Yanagi, M., & Yamasato, K. (1993). Phylogenetic analysis of the family Rhizobiaceae and related bacteria by sequencing of 16S rRNA gene using PCR and DNA sequencer. *FEMS Microbiology Letters*, 107(1), 115-120.
- Yang, X., Hudson, M., Walters, N., Bargatze, R. F., & Pascual, D. W. (2005). Selection of protective epitopes for *Brucella melitensis* by DNA vaccination. *Infection and immunity*, 73(11), 7297-7303.
- Yoneyama, H., & Ichida, T. (2005). Recruitment of dendritic cells to pathological niches in inflamed liver. *Medical molecular morphology*, 38(3), 136-141.
- You, L. (2004). Steroid hormone biotransformation and xenobiotic induction of hepatic steroid metabolizing enzymes. *Chemico-biological interactions*, 147(3), 233-246.
- Young, E. J. (1983). Human brucellosis. *Review of Infectious Diseases*, 5(5), 821-842.
- Young, E. J., Roushan, M. R. H., Shafae, S., Genta, R. M., & Taylor, S. L. (2014). Liver histology of acute brucellosis caused by *Brucella melitensis*. *Human pathology*, 45(10), 2023-2028.
- Young, E. J., Roushan, M. R. H., Shafae, S., Genta, R. M., & Taylor, S. L. (2014). Liver histology of acute brucellosis caused by *Brucella melitensis*. *Human pathology*, 45(10), 2023-2028.
- Zampetaki, A., Zhang, Z., Hu, Y., & Xu, Q. (2005). Biomechanical stress induces IL-6 expression in smooth muscle cells via Ras/Rac1-p38 MAPK-NF-κB

signaling pathways. *American Journal of Physiology-Heart and Circulatory Physiology*, 288(6), H2946-H2954.

Zhao, Z., Li, M., Luo, D., Xing, L., Wu, S., Duan, Y., Yang, P., & Wang, X. (2009). Protection of mice from *Brucella* infection by immunization with attenuated *Salmonella* enterica serovar typhimurium expressing A L7/L12 and BLS fusion antigen of *Brucella*. *Vaccine*, 27(38), 5214-5219.

