



UNIVERSITI PUTRA MALAYSIA

***ISOLATION AND CHARACTERIZATION OF CONDENSED TANNIN
FROM
Acacia mangium WILLD. AND ITS ANTIBACTERIAL ACTIVITY***

NASYATUL EKMA BINTI MOHD HUSSIN

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By

NASYATUL EKMA BINTI MOHD HUSSIN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of Master of
Science**

January 2016

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Master of Science

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January 2016

Chairman : Associate Professor Loong Yik Yee, PhD
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Three experiments have been conducted to extract and evaluate the ability of tannins to bind with proteins and also their inhibition towards the growth of several types of pathogenic skin bacteria. The types of tannins and their molecular weight are believed to be the main factors influencing their activity towards proteins and microbes. In the first experiment, condensed tannin was extracted from several parts of *Acacia mangium* including leaves, stems, and pods. Sephadex LH-20 was used to separate condensed tannin (CT) from other phenolic compounds contains in the crude extracts. The amount of purified condensed tannin was measured by modified Vanillin-HCl assay method using catechin as the standard. The concentration of extracted condensed tannin was found to be different in each part. The amount of tannins was calculated based on dry matter (DM) basis. Leaves were found to contain the highest amount which is 3.28% DM, whereas in stems and pods contains 0.98% DM and 1.54% DM, respectively.

In second experiment the protein-binding affinity (PBA) of extracted condensed tannins was determined using a protein precipitation assay with bovine serum albumin (BSA) used as the standard proteins. The affinity was measured based on b-value (the b-value represent the quantity of condensed tannin needed to bind half of the maximum precipitable BSA). The average b-value of isolated CT was 0.53 with standard deviation of 0.048.

In the third experiment, the antibacterial activity of extracted CT was evaluated. There were three sub-experiment were conducted to evaluate the activity. The first experiment was to measure the zone of inhibition of tested bacteria against 100 mg/mL CT extracts by agar diffusion test. Then, the minimum inhibitory were measured by using microdilution test. Lastly in the third experiment, the log reduction of numbers of bacteria based on time incubation and also by concentration was quantified by time-kill assay.

From the study, it showed that all tested bacterium were susceptible to the CT at the concentration of 100 mg/mL, with diameter zone of inhibition of ranged between 12.0 to 13.0 mm. However, the minimum inhibitory concentrations (MIC) were different between the isolates. Compared to resistant strains, *S. aureus* ATCC 29213 was found to be inhibited by CT extract at low concentration (1 mg/mL) and it also was killed after 6 hours exposure with 10 mg/mL CT extracts. On the other hand, all the MRSA isolates had varied inhibitory effects. The MIC values were in the range of 2 mg/mL to 10 mg/mL and most of the isolates were inhibited at 5 mg/mL. Based on the collected data it shows that condensed tannins had the higher content in *A. mangium* leaves. Eventhough, the amount were lower in stems and pods, their affinity to bind protein were almost similar. These compounds also shows a good antibacterial effects against *S. aureus* (including resistant strains).



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

**ISOLATION AND CHARACTERIZATION OF CONDENSED TANNIN FROM
Acacia mangium WILLD. ND ITS ANTIBACTERIAL ACTIVITY**

Oleh

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Januari 2016

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Tiga eksperimen telah dijalankan untuk mengekstrak dan menilai keupayaan tanin untuk mengikat protein dan merencat pertumbuhan beberapa jenis bakteria berbahaya yang biasa dijumpai pada kulit manusia. Jenis tanin serta berat molekul dipercayai merupakan faktor utama mempengaruhi aktiviti tanin ini terhadap protein dan mikrob. Dalam eksperimen pertama, tanin tersejat (CT) telah diekstrak daripada beberapa bahagian pokok *Acacia mangium*, termasuk dedaun, batang, dan buah. Sephadex LH-20 telah digunakan bagi mengasingkan CT daripada beberapa jenis sebatian fenolic lain yang terdapat di dalam ekstrak mentah. Jumlah CT telah dikira menggunakan kaedah Vanillin-HCl assay yang telah diubahsuai, menggunakan cathechin sebagai standard. Kepekatan tanin dilihat berbeza bagi setiap bahagian tersebut. Dedaun mempunyai kandungan tanin tersejat (CT) yang paling tinggi iaitu 3.28% berat kering, manakala dalam batang dan buah mengandungi 0.98% berat kering dan 1.54% berat kering, setiap satunya.

Dalam eksperimen kedua, keupayaan mengikat protein (PBA) oleh ekstrak tanin tersejat (CT) telah dikenalpasti melalui ujian pemendakan protein dengan menggunakan serum albumin lembu (BSA) sebagai protein standard. Keupayaan tersebut telah dikira melalui jumlah nilai-b (nilai b ini mewakili jumlah CT yang diperlukan untuk mengikat separuh dari maksimum BSA yang termendak). Jumlah purata nilai-b untuk tanin yang telah diasingkan daripada setiap bahagian ialah 0.53 dengan sisihan piawainya adalah 0.048.

Dalam eksperimen ketiga, aktiviti anti-bakteria oleh ekstrak CT telah dinilai. Terdapat tiga sub-eksperimen telah dijalankan untuk menilai aktiviti tersebut. Eksperimen pertama adalah untuk mengira zon perencatan bagi bakteria yang diuji ke atas 100 mg/mL ekstrak CT menggunakan ujian difusi/penyebaran agar. Kemudian, efek perencatan yang minimum telah dikira menggunakan kaedah ujian pencairan mikro. Dalam eksperimen terakhir, pengukuran jumlah log bakteria telah dikira menerusi masa inkubasi dan juga menerusi kepekatan, menggunakan kaedah *time-kill*.

Daripada kajian ini, ia telah menunjukkan bahawa semua bakteria yang diuji adalah terdedah kepada CT pada kepekatan 100 mg/mL, dengan zon diameter perencatan dalam lingkungan 12 ke 13 mm. Walau bagaimanapun, kepekatan minima untuk perencatan adalah berbeza diantara jenis-jenis strain. *S. aureus* ATCC 29213 telah dikenalpasti mampu direncatkan oleh ekstrak CT pada kepekatan yang rendah (1 mg/mL) berbanding strain yang mampu tahan/resistan. Ia juga telah mati selepas terdedah selama 6 jam dengan 10 mg/mL ekstrak CT. Pada masa yang lain, kesemua jenis MRSA mempunyai aktiviti perencatan yang berbeza. Nilai MIC adalah dalam lingkungan 2 mg/mL hingga 10 mg/mL dan kebanyakan jenis tersebut telah direncatkan pada 5 mg/mL. Daripada data yang telah disimpan, didapati bahawa kandungan CT dalam daun *A. mangium* adalah tinggi. Walaupun kandungannya adalah rendah dalam batang dan buah, kemampuan ia untuk mengikat protin adalah lebih kurang sama. Ekstrak CT juga menunjukkan ia mempunyai efek anti-bakteria yang baik ke atas *S. aureus* (termasuk strain yang mampu tahan/resistan).

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I certify that a Thesis Examination Committee has met on 18 January 2016 to conduct the final examination of Nasyatul Ekma binti Mohd Hussin on her thesis entitled "Isolation and Characterization of Condensed Tannin from *Acacia mangium* Willd. and its Antibacterial Activity" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

| | |
|---------|------------------------------------|
| BSA | Bovine serum albumin |
| CT | Condensed tannin |
| CTs | Condensed tannins |
| Da | Dalton |
| DM | Dry matter |
| g | gram |
| HT | Hydrolysable tannin |
| M | Molar |
| mg | milligram |
| µg | microgram |
| µL | microliter |
| µm | micrometer |
| nm | nanometer |
| pH | Puissance Hydrogen |
| S | Susceptible |
| I | Intermediate |
| R | Resistance |
| MIC | Minimum inhibitory concentration |
| MBC | Minimum bactericidal concentration |
| HCl | Hydrochloric acid |
| m | meter |
| mm | millimeter |
| mL | millilitre |
| % | percent |
| v/v | volume over volume |
| w/v | weight over volume |
| min | minute |
| °C | Degree Cecius |
| std dev | standard deviation |
| C-C | Carbon-carbon link |
| PBA | Protein binding affinity |
| ATCC | American Type Culture Collection |
| MH | Muller-Hinton |
| TPC | Total plate count |
| ND | Not defined |
| sp. | species |
| PEG | Polyethylene Glycol |
| NaCl | Sodium chloride |
| NaOH | Sodium hydroxide |
| N | Normality |

CHAPTER 1

INTRODUCTION

The wide diversity of structurally unique compounds, produced by tropical plant was documented and well addressed by most researchers in the world as the sources of new mechanisms, materials, therapeutic and prophylactic agents to cure and prevent human diseases (Mazni, 2016). To date, there are more than 500,000 species of higher plant are present in the Earth, but only small percentage were investigated for phytochemically and pharmacological screening (Sharma and Patel, 2013). It is essential to utilize this resource by exploiting its biomolecules that posse's biological activity to bring beneficial use in healthcare and the hope of new cures to important existing diseases.

Tannins are high molecular weight phenolic compounds present in many plants. They are water soluble polyphenolic polymers and have a high capacity to form complexes mainly with proteins. Tannins are molecules that have a potency to bind with proteins, cellulose, starches and minerals. This results in the formation of insoluble compounds which is resistant to the decomposition processes. Tannins can be found in various parts of plants, including barks, wood, leaves, buds, stems, fruit, seed, roots, and plant galls. In all of these structures, tannin help to protect the plant form being infected by bacteria or fungi. One of the mechanisms is by precipitating the enzyme and other proteins produced by the bacteria or fungi (Hassanpour, 2011). There are two groups of tannins; hydrolysable tannins and condensed tannins.

Condensed tannins (CT) were known to be able to interact with biological systems through the induction of some physiological effects, such as antioxidant, anti-allergy, anti-hypertensive, and antimicrobial activities (Romani, 2006). Early as in 1980's, condensed tannins were showed to have antiviral activities on Herpes Simplex Virus (HSV-1 and HSV-2). The effect was due to the inhibition of virus adsorption (Fukuchi, 1986). CT compound also was reported to have antiviral activities against respiratory syncytial virus (RSV), influenza A virus (FLU-A), parainfluenza virus (PIV) comparable to ribavirin, and inhibition towards hepatitis-A was observed (Ubillas, 1994). Those inhibitory mechanisms were believed to be either by the binding of the tannin molecules to the protein coat of the virus or to the host cell membrane (Toss, 1999).

Condensed tannins from the other plants such as *Leucaena leucocephala*, *Phaseolus vulgaris*, *Fagopyrum esculentum*, *Corylus avellana*, and *Juglans nigra*, were shown to have antibacterial activities against some pathogenic bacteria such as *Staphylococcus aureus*, *Escherichia coli*, *Pseudomonas aeruginosa*, and *Salmonella enterica* (Mazni, 2016; Ruzard, 2008).

Acacia is a widespread genus of tropical-subtropical trees and shrubs found in Central/South America through Africa to Southeast Asia and Australia. There

were about 1352 species of *Acacia* species were found worldwide (Maslin, 2003). The predominant *Acacia* species in Malaysia is *Acacia mangium*. It possesses many advantageous growth characteristics such as a high leaf yield, drought tolerance, adaptation to acidic infertile soils and large temperature variations (Marini, 2011). This plant also was shown to contain tannins. However, the reactivity of these tannins was dependent on the structure and their chemical nature to associate with the substrate (Hoong, 2010).

The emergence of antibiotic-resistant bacteria has become a problem to successful treatment for bacterial infections (Silvia, 2013). Recently, many studies have been done to use biologically active compounds extracted from plant to be used as medicine, especially for bacterial infections (Jigna, 2005). The findings on the antimicrobial activity of plant extracts have also been used as the basic knowledge for many applications. This include as preservation in raw or processed food, in pharmaceuticals, also as alternative medicine (Leela and Satirapipathkul, 2011). There are several types of secondary compounds such as tannins, phenols, steroids and alkaloids. These compounds are deposited in various parts of plants (Balandrin, 1985). They play a major role in plant colour, flavour, texture, also as antioxidant and antibacterial (Hernandez, 1999; Negi, 2005).

A variety of methods have been used in a study of antimicrobial activity of plant bioactive compounds. However, the disc diffusion method of antimicrobial susceptibility test was the most practical method and is a method of choice for the average laboratory (Cleudson, 2007). A growth medium, usually Mueller-Hinton agar, was commonly used in agar diffusion test. This medium was considered the best for the susceptibility testing since it has batch-to-batch reproducibility, low concentration of inhibitors and produces satisfactory results for most of the non-fastidious microorganisms.

In the test, antimicrobial agent was dropped on the paper disc. Once it has on the media, the compounds will start to diffuse outwards from the disc and creates a gradient of antimicrobial concentration in the MH agar such that the highest concentration was found close to the disc with decreasing concentrations further away from the disc. The zone around an antibiotic disc that has no growth was referred to as the zone of inhibition since this approximated the minimum antibiotic concentration sufficient to prevent growth of the test isolate (Lalitha, 2004).

This thesis consisted of three experiments. The first was to extract condensed tannin from several parts of *Acacia mangium* including the leaves, stems, and pods, and then Sephadex LH-20 was used to separate condensed tannin (CT) from other phenolic compounds contained in the crude extracts.

In the second experiment, the affinity of extracted condensed tannin to bind with proteins was examined using protein precipitation assay. This step was important to determine the activity of CT in each sample, and lastly, in the third experiment, the antimicrobial property of condensed tannin extract was tested against several pathogenic bacteria including the resistant strains.

Thus, this study was conducted to accomplish the objectives below:

1. To quantify the amount of condensed tannins isolated from different parts of *Acacia mangium*
2. To examine the activity of extractable condensed tannins to precipitate proteins based on the b-value
3. To determine the antimicrobial activity (inhibition zone, minimum inhibitory concentration, and time-killing assay) of condensed tannins towards the growth of pathogenic bacteria including the drug-resistant strains.

REFERENCES

- Adam N, Irene MH, Stig MT. Condensed tannins act against cattle nematodes. *Veterinary Parasitology* 182: 213-220 (2011)
- Afrillia NG, Elin YS, Irda F. Activity of Several Plant Extracts against Drug-Sensitive and Drug-Resistant Microbes. *Procedia Chemistry* 13: 164-169 (2014)
- Anuraga J, Gunjan G, Harinder PSM, Klause B. Divergence between purified hydrolysable and condensed tannin effects on methane emission, rumen fermentation, and microbial population *in vitro*. *Animal Feed Science and Technology* 209: 60-68 (2015)
- Aerts RJ, Barry TN, McNabb WC. Polyphenols and agriculture: beneficial effects of proanthocyanidins in forages. *Agriculture, Ecosystem and Environment* 75: 1-2 (1999)
- Ahn JH, Robertson BM, Elliott RR, Gutteridge RC, Ford CW. Quality assessment of tropical browse legumes; tannin content and protein degradation. *Animal Feed Science and Technology* 27: 147-156 (1989)
- Ali K, Meredith AW, Thosmas HR. Techniques for analysis of plant phenolic compounds. *Molecules* 18: 2328-2375 (2013)
- Alo MN, Anyim C, Igwe JC, Elom M, Uchenna DS. Antibacterial activity of water, ethanol and methanol extracts of *Ocimum gratissimum*, *Vernonia amygdalina*, and *Aframomum melegueta*. *Advance in Applied Science Research* 3 (2): 844-848 (2012)
- Ann EH. The Tannin Handbook. <http://chemistry.muohio.edu/hagerman/> Access 2011
- Arun KW, Arti S, Ajit KS. Determination of minimum inhibitory concentration (MIC) of some novel triazole derivatives. *International Journal of Research in Pharmacy and Chemistry* 1 (4): 1108-1114 (2011)
- Badar N, Arshad M, Farooq U. Characteristics of *Anethum graveolens* (umbelliferae) seed oil: extraction, composition and antimicrobial activity. *International Journal of Agriculture and Biology* 10: 329-332 (2008)
- Balandrin MFJ, Kjoke A, Wurtele E. Natural plant chemicals: source of industrial and medicinal materials. *Science* 228: 1154-1160 (1985)
- Bauer AW, Kirby WMM, Sheriss JC, Turck M. Antibiotic susceptibility testing by a standardized single disk method. *American Journal of Clinical Pathology* 45: 493-496 (1966)
- Blondeau JM, Hansen G, Metzler KL, Borsos S, Chau J. Optimal killing of *Streptococcus pneumoniae* by gemifloxacin, levofloxacin and

- moxifloxacin. In: Gillespie SH, Tillotson GS, editors. Novel perspectives in antibacterial action. London, UK: Royal Society of Medicine Press Ltd (2002). p. 15-26
- Chris B, Karen B, Eko H, Ragil I, Junarto, Caroline M, Anto R. Effect of pruning *Acacia mangium* on growth, form and heart rot. *Forest Ecology and Management* 238: 261-267 (2007)
- Botzenhardt K, G. Ecology and epidemiology of *Pseudomonas aeruginosa* Doring. *Pseudomonas aeruginosa* as an Opportunistic Pathogen: 1-7 (1993)
- Chung KT, Stevens SE, Lin WF, Wei CI. Growth inhibition of selected food-borne bacteria by tannic acid, propyl gallate and related compounds. *Letters in Applied Microbiology* 17: 29–32 (1993)
- Cienciala E, Kucera J, Malmer A. Tree sap flow and stand transpiration of two *Acacia mangium* plantations in Sabah, Borneo. *Journal of Hydrology* 236: 109-120 (2000)
- Cleidson V, Simone MS, Elza FAS, Artur SS. Screening methods to determine antibacterial activity of natural products. *Brazilian Journal of Microbiology* 38: 369-380 (2007)
- Cogen AL, Nizet V, Gallo RL. Skin microbiota: a source of disease or defends? *British Association of Dermatologist*: 1-26 (2008)
- Craven D, Dent D, Ashton MS, Berlyn GP, Hall JS. Seasonal variability of photosynthetic characteristics influences growth of eight tropical tree species at two sites with contrasting precipitation in Panama. *Forest Ecology and Management* 261: 1643-1653 (2011)
- Dalzel SA, and Kerven GL, 1998. A rapid method for the measurement of *Leucaena spp* proanthocyanidins by the proanthocyanidins (Butanol/HCl) assay, *Journal of Science, Food and Agriculture* 78; 405-416
- Daniel B, Cristiane AS, Bouillet JP, Jose LMG, Elke JBNC. *Aucalyptus grandis* and *Acacia mangium* and litter microbial and chemical attributes during early stages of plant development. *Applied Soil Ecology* 63: 57-66 (2013)
- Danish M, Hashim R, Mohamad Ibrahim MN, Sulaiman O. Effect of acidic activating agents on surface area and surface functional groups of activated carbons produced from *Acacia mangium* wood. *Journal of Analytical and Applied Pyrolysis* 104: 418-425 (2013)
- Dawra RK, Makkar HP, Singh B. Protein binding capacity of microquantities of tannins. *Analytical Biochemistry* 170: 50-53 (1988)
- Deshpande LM, Fritsche TR, Jones RN. Molecular epidemiology of selected multi-drug resistant bacteria: a global report from the SENTRY

- antimicrobial Surveillance Program. *Journal of Diagnosis Microbiology and Infection Diseases* 49: 231-236 (2004)
- Doig AJ, William DH, Oelrichs PB, Bacynskyj L. Isolation and structure elucidation of punicalagin, a toxic hydrolysable tannin, from *Terminalia oblongata*. *Journal of the Chemical Society Perkin Transaction 1 I* (8): 2317-2321 (1990)
- Engelkirk P; Duben E, Janet. *Laboratory Diagnosis of Infectious Diseases*. Lippincott Williams & Wilkins: 168. (2008).
- Faiza A, Khalil-ur-Rehman, Asghar M, Sarwar M. Antibacterial activity of various phytoconstituents of neem. *Pakistan Journal of Agriculture Sciences* 46 (3): 209-213 (2009)
- Fickel J, Pitra C, Joest BA, Hofmann RR. A novel method to evaluate the relative tannin-binding capacities of salivary proteins. *Comparative Biochemistry and Physiology Part C*: 225-229 (1999)
- Fremont M, Prin Y, Chauviere M, Diem HG, Pwee KH, Tan TK. A comparison of *Bradyrhizobium* strains using molecular, cultural and field studies. *Plant Science* 141: 81-91 (1999)
- Hagerman AE, Butler LG. The specificity of proanthocyanidin-protein interactions. *The Journal of Biological Chemistry* 256 (9): 4494-4497 (1981)
- Hagerman AE, Butler LG. Choosing appropriate methods and standards for assaying tannin. *Journal of Chemical ecology* 15 (6): 1795-1810 (1989)
- Haruni K, Maarit K, Markku K. *Acacia mangium* Wild: ecology, silviculture and productivity. Center for International Forestry Research; 2011.p.1-3.
- Haslam E. *Plant polyphenols*. Cambridge University Press, Cambridge, UK (1989)
- Hassanpour S, Sadaghian M, Maherisis N, Eshratkhah B, Chaichisemsari M. Effect of condensed tannin on controlling faecal protein excretion in nematode-infected sheep: in vivo study. *Journal or American Sciences* 7 (5): 896-900 (2001)
- Heatley NG. A method for the assay of penicillin. *Journal of Biochemistry* 38: 61-65 (1944)
- Hernandez F, P Melgarejo, FA Tomas-Barberan, Aartes. Evaluation of juice antocyanins during ripening of new selected pomegranate (*Punica granatum*) clones. *European Food Research and Technology* 210: 39-42 (1999)
- Hisanori A, Kazuyasu F, Osamu Y, Takashi O, Keiji I. Antibacterial action of several tannins against *Staphylococcus aureus*. *Journal or Antimicrobial Chemotrapy* 48: 487-491 (2001)

- Hoong YB, Antonio P, Paridah MT, Harald P. Characterization of *Acacia mangium* polyflavonoid tannins by MALDI-TOF mass spectrometry and CP-MAS ¹³C NMR. *European Polymer Journal* 46: 1268-1277 (2010)
- Hoong YB, Paridah MT, Loh YF, Jalaluddin H, Chuah LA. A new source of natural adhesive: *Acacia mangium* bark extracts co-polymerized with phenol-formaldehyde (PF) for bonding Mempisang (*Annonaceae* spp.) veneers. *International Journal of Adhesion and Adhesives* 31: 164-167 (2011)
- James FH, Rachel KL, Mark AK, Mark PD. Impact of condensed tannin size as individual and mixed polymers on bovine serum albumin precipitation. *Food Chemistry* 160: 16-21 (2014)
- Jennifer MA. Determination of minimum inhibitory concentrations. *Journal of Antimicrobial Chemotherapy* 48: 5-16 (2001)
- Jigna P, Darshana J, Sumitra C. Efficacy of aqueous and methanol extracts of some medicinal plants for potential antibacterial activity. *Turkish Journal of Biology* 29: 203-210 (2005)
- Jiro T, John S. Soil macro-fauna in an *Acacia mangium* plantation in comparison to that in the lowlands of Sarawak, Malaysia. *Pedobiologia* 49: 69-80 (2005)
- Jorgensen JH, Ferraro MJ. Antimicrobial Susceptibility Testing: A Review of General Principles and Contemporary Practices. *International Journal of Medical Microbiology* 49: 1749-1755 (2009)
- Joyce LF, Downes J, Stockman K, Andrew JH. Comparison of five methods, including the PDM Epsilometer test (E test), for antimicrobial susceptibility testing of *Pseudomonas aeruginosa*. *Journal of Clinical Microbiology* 30 (10): 2709-2713 (1992)
- Katarina C, Bryan AS, George JM. Skin microflora and bacterial infections of the skin. *The society for investigative Dermatology* 6: 170-174 (2001)
- Karamali K, Teunis van R. Tannins: Classification and Definition. *Natural Product Reports* 18: 641-649 (2001)
- Khidir TOM, Suminar SA, Nisa RM. Optimizing Tannin extraction from antibacterial activity of *Acacia nilotica*. *Journal of international Environmental application & science* 5: 781-800 (2010)
- Kim J, Marshall MR, Wei C. Antibacterial activity of some essential oil components against five foodborne pathogens. *Journal of Agriculture and Food Chemistry* 43: 2839-2845 (1995)
- Lalitha MK. Manual on antimicrobial susceptibility testing. Christian Medical College, Vellore, Tamil Nadu. Pg 1-47. (2004)

- Lederberg, Joshual. *Pseudomonas*. Encyclopedia of Microbiology. Second Edition (3), San Diego; 876-891 (2000)
- Leela T, Satirapipathkul C. Studies on the antibacterial activity of *Quercus Infectoria* Galls. 2011 International Conference on Bioscience, Biochemistry, and Bioinformatics :410-415 (2011)
- Lim SH, Darah I, Jain K. Antimicrobial activities of tannins extract from *Rhizophora apiculata* Barks. Journal of Tropical forest Science 18 (1): 59-65 (2006)
- Liu C, Bayer A, Cosgrove SE, Daum RS, Fridkin SK, Gorwitz RJ, Kaplan SL, Karchmer AW, Levine DP, Murray BE, Rybak M, Talan DA, Chambers HF. Clinical practice guidelines by the infectious diseases society of America for the treatment of methicillin-resistant *Staphylococcus aureus* infections in adults and children: executive summary. Infectious Diseases Society of America 52 (3): 285–292 (2011)
- Long Y, Nan L, Hai R, Jun W. Facilitation by two exotic Acacia: *Acacia auriculiformis* and *Acacia mangium* as nurse plants in South China. Forest Ecology and Management 257: 1786-1793 (2009)
- Magdalena K, Agnieszka K, Anna R, Ryszard A. Extraction and chromatographic separation of tannin fractions from tannin-rich plant arterial. Journal of food and nutrition sciences 57: 471-474 (2007)
- Macauley BJ, Fox LR. Variation in total phenols and condensed tannins in Eucalyptus: leaf phenology and insect grazing. Austral Ecology 5: 31-35 (2006)
- Maisak H, Tipmongkolsilp N, Wongtavatchai J. Minimum Inhibitory Concentration of Antimicrobials against clinical *Vibrio* and *Streptococcus* isolated form aquaculture. Disease in Asian Aquaculture VII: 309-316 (2011)
- Makkar HPS, Dawra RK, Singh B. Protein precipitation assay for quantification of tannins: Determination of Protein in tannin-protein complex. Analytical Biochemistry 166:435-439 (1987)
- Marini AM, Shanmugavelu S, Fadzliarahimi, Abdul Razak M, Mazwan, Siti Masidayu MS, Awis P, Mohd Firdaus B. Proximate and Minerals Composition of *Acacia mangium* Cultivated Vs Wild. The Malaysian Society of Animal Production Proceeding. 31st MSAP Conference (2011)
- Martin L P, Steve van S, Larry GB. A critical evaluation of the Vanillin reaction as an assay for tannin in Sorghum grain. Journal of Agriculture and Food Chemistry 26 (5): 1214-1218 (1978)
- Maslin BR, Miller JT, Siegler DS. Overview of the generic status of Acacia (Leguminosae: Mimosideae). *Australian Systematic Botany* 16: 1-18 (2003).

- McSweeney CS, Palmer B, McNeill DM, Krause DO. Microbial interaction with tannins: nutritional consequences for ruminants. *Animal feed science and technology* 91: 83-93 (2001)
- Meigy NM, Mahendradatta M, Djide N. Test of antimicrobial activity of tannins extract from guava leaves to pathogens microbial. *The International Asian Reserch Journal* 2 (1): 43-50 (2014)
- Meigy NM, Mahendradatta M, Amran L, Djide N. Antimicrobial activity of tannins extract from guava leaves (*Psidium gaujava* L) to pathogens microbial. *International Journal of Scientific & Technology Research* 3(1): 236-241 (2014)
- Min BR, Hart SP. Tannin for suppression of internal parasite. *Journal of animal science*: 102-109 (2003)
- Mookiah S, Tan HY, Sieo CC, Norhani A, Clemente MVLW, Emilia A, Ho YW. Polymerization Degrees, Molecular Weights and Protein-Binding Affinities of Condensed Tannin Fractions from a *Leucaena leucocephala* Hybrid. *Molecules* 19(6): 7990-8010 (2014)
- Mounyr B, Moulay S, Saad KI. Methods for *in vitro* evaluating antimicrobial activity: A review. *Journal of Pharmaceutical Analysis* 6: 71-79 (2016)
- Mrunalini ND, Chavan NS. Antibacterial activity and phytochemical screening of *Cynometra iripa* kostel. *International Journall of Pharma and Bio Sciences* 1 (30): 1-4 (2010)
- Nasyatul E, Marini AM, Shanmugavelu S, Idayusni Y, Masidayu MS, 2014. Crude Tannin Concentration in Local Forages and Feed Ingredients. *Malaysian Journal of Veterinary Research, Volume 5 Supplement 1, 26th Veterinary Association Malaysia Congress*: 354-355
- Neela V, Sasikumar M, Ghazali GR, Zamberi S, Mariana S. In vitro activities of 28 antimicrobial agents against Methicillin-resistant *Staphylococcus aureus* (MRSA) from clinical setting in Malaysia.
- Norazah VK, Lim E, Koh YT, et al. Molecular fingerprinting of fusidic acid- and rifampicin-resistant strains of methicillin-resistant *Staphylococcus aureus* (MRSA) from Malaysian hospitals. *Journal of Medical Microbiology* 51: 1113-1116 (2002)
- Olufunmiso OO, Anthony JA. *In vitro* antibacterial and time-kill assessment of crude methanolic stem bark extract of *Acacia mearsii* de wild against bacteria in Shigellosis. *Molecules* 17: 2103-2118 (2012)
- Osborne NJT, McNeill DM. Characterisation of *Leucaena* condensed tannins by size and protein precipitation capacity. *Journal of the Science of Food and Agriculture* 81 (11) : 1113-1119 (2011)

- Patra AK, Saxena J. A new perspective on the use of plant secondary metabolites to inhibit methanogenesis in the rumen. *Phytochemistry* 71: 11-12 (2010)
- Pollard TD. A guide to simple and Informative binding assays. *Molecular Biology of Cell* 21: 4061-4067 (2010)
- Praveen D, Sharmishtha P. Phytochemical Screening and Antimicrobial Activity of Some Medicinal Plants against Multi-drug Resistant Bacteria from Clinical Isolates. *Indian Journal of Pharmaceutical Sciences* 03: 443-450 (2012)
- Piyarak T, Barbara MR. Determination of minimal bactericidal and effective antibiotic treatment concentration for bacterial contaminants from micropropagated strawberries. *In vitro Cellular & Developmental Biology-Plant* 33: 227-230 (1997)
- Pritchard DA, Tocks DC, O'Sullivan BM, Martin PR, Hurwood IS, O'Rourke PK. The effect of polyethylene glycol on wool growth and live weight of sheep consuming mulga (*Acacia aneura*) diet. *Proceedings of Australian Society of Animal Production* 17: 290-293 (1988)
- Rolando B, Solange S, Carlos EL, Emyr O, Phillip M, Michael KT. Effect of condensed tannin from tropical legume on the activity of fibrolytic enzymes from the rumen fungus *Neocallimatyx hurleyensis*. *Enzyme and Microbial Technology* 39: 281-288 (2006)
- Rama SD, Prasad MNV. Short Communication: Tannins and Related Polyphenols from Ten Common Acacia Species of India. *Bioresource Technology* 36: 189-192 (1991)
- Rameshkumar KB, George V, Shiburaj S. Chemical constituents and antibacterial activity of the leaf oil of *Cinnamomun chemungianum* Mohan et Henry. *Journal of Essential Oil Research* 19: 98-100 (2007)
- Resmi M, Linar Z, Muchtaridi, Supriyatna. Identification and antibacteria activity of methanol extract of *Luffa acutangula* Roxb. *Mediacal and Health Science Journal* 12: 70-77 (2012)
- Romani A, Ieri F, Turchetti B, Mulinacci N, Vincieri FF, Buzzini P. Analysis of condensed tannin and Hydrolysable tannin from commercial plant extract. *Journal of Pharmaceutical and Biomedical Analysis* 41: 415-420 (2006)
- Salmonellosis. The center for food security and public health, Iowa State University, Iowa.p. 1-8 (2005)
- Sambatakou H, Giamerellos-Bourboulis EJ, Grecka P, Chryssouli Z, Giamarellou H. In-vitro activity and killing effect of quinupristin/dalfopristin on nosocomial *Staphylococcus aureus* and interactions with rifampicin and ciprofloxacin against Methicillin-resistant isolates. *Journal of Antimicrobial Chemotherapy* 41: 349-355 (1998)

- Santos-Buelga, Celestino, Victor de Freitas. Influence of Phenolics on Wine Organoleptic Properties. In Wine Chemistry and Biochemistry: 529–70. Springer New York (2009).
- Schoenknecht FD, Washington JA, Gavan TL, Thornsberry C. Rapid Determination of Minimum Inhibitory Concentrations of Antimicrobial Agents by the Autobac Method: A Collaborative Study. Antimicrobial Agents and Chemotherapy: 824-833 (1980)
- Schofield P, Mbugua DM, Pell AN. Analysis of condensed tannins: a review. Animal feed science and technology 91:21-40 (2001)
- Seigler DS. Phytochemistry of Acacia-sensu lato. Biochemical Systematics and Ecology 3: 845-873 (2003)
- Shaan LG, Robert EWH. *Pseudomonas aeruginosa*: new insights into pathogenesis and host defenses. Pathogens and disease 67: 159-173 (2013)
- Sharma VJ, Patel PM. Evaluation of antibacterial activity of methanolic extract of plant *Rivea ornata*. International Research Journal of Pharmacy 4 (1): 233-234 (2013)
- Silanikove N, Perevolotsky A, Provenza FD. Use of tannin-binding chemicals to assay for tannins and their negative post-ingestive effects in ruminants. Animal Feed Science and Technology 91: 69-81 (2001)
- Silvia M, Elba LCA, Tadeu JS, Peixoto S, Antonio MS, Maria NCP, Cristobal NA, Jose AT, Solange IM. Antibacterial activity of crude methanolic extract and fractions obtained from *Larrea tridentata* leaves. Industrial Crops and Products 41: 306-311 (2013)
- Strategic Science Team. Characterization of Salmonella. Ministry of Agriculture and Forestry, Wellington: MAF 2010
- Stefanello MEA, Cervi AC, Ito I, Salvador MJ, Wisniewski A, Simionatto EL. Chemical composition and antimicrobial activity and essential oil of *Eugenia chlorophylla* (Myrtaceae). Journal of Essential Oil Research 20 (1): 75-78 (2008)
- Strumeyer DH, Malin MJ. Condensed Tannins in Grain Sorghum: Isolation, Fractionation, and Characterization. Journal of Agriculture and Food Chemistry 23 (5):909-914 (1975)
- Suraya S, Darah I, Jain K, Lim SH. Antimicrobial and antioxidant activities of condensed tannin from *Rhizophora apiculata* barks. Journal of Chemistry and Pharmacology Research 3 (4): 436-444 (2011)
- Takahashi O, Cai Z, Toda M, Hara Y, Shimamura T. Appearance of antibacterial activity of oxacillin against methicillin resistant *Staphylococcus aureus* (MRSA) in the presence of catechin. Journal of the Japan Association for Infectious Disease 69: 1126–34 (1995).

- Terrill TH, Rowan AM, Douglas GB, Barry TN. Determination of extractable and bound condensed tannin concentrations in forage plants, protein concentrate meals and cereal grains. *Journal of the Science of Food and Agriculture* 58:321-329 (1992)
- Tohru M, Takao D, Yuuki K, Isao A. Color development of proantocyanidins in vanillin-hydrochloric reaction. *Journal of Wood Sciences* 44: 125-130 (1998)
- Tong SYC, McDonald MI, Holt DC, Currie BJ. Global implications of the emergence of community-associated Methicillin-resistant *Staphylococcus aureus* in indigenous populations. *Journal of Clinical Infection Diseases* 46: 1871-1878 (2008)
- Tong SYC, Davis JS, Eichenberger E, Holland TL, Fowler VG. *Staphylococcus aureus* infections: epidemiology, pathophysiology, clinical manifestations, and management. *Clinical Microbiology Reviews* 28 (3): 603–661 (2015)
- Do TTV, Nguyen TM, Inger L. Effect of method of processing foliage of *Acacia mangium* and inclusion of bamboo charcoal in the diet on performance of growing goats. *Animal Feed Science and Technology* 130: 242-256 (2006)
- Waghorn G. Beneficial and detrimental effects of dietary condensed tannins for sustainable sheep and goat production-Progress and challenges. *Animal Feed Science and Technology* 147:-139 (2008)
- Wan Noor Aidawati WN, Rokiah H, Salim H, Othman S, Jia GB, Kushairi MS, Fahmi A, Masatoshi S, Tomoko S. Measurement of some properties of binderless composites manufactured from oil palm trunks and *Acacia mangium*. *Measurement* 50: 250-254 (2014)
- William TJ, James LM. Complexes of the condensed tannins of sainfoin (*Onobrychis viciifoliascop.*) with fraction 1 leaf protein and with submaxillary mucoprotein, and their reversal by polyethylene glycol and pH. *Journal of the Science of Food and Agriculture* 28 (2): 126-136 (1977)
- Yusoff S.; Sharif S.; Noormah MA. Nutrient Composition of Malaysian Feed Materials and Guides to Feeding of Cattle and Goats. Feeding Guide Series 2. Department of Veterinary Services. Ministry of Agriculture and Agro-based Industry Malaysia. Gempita Maju Sdn Bhd; 2005.