

UNIVERSITI PUTRA MALAYSIA

IN VITRO EFFECTS OF Phaleria macrocarpa (Boerl.) MAHKOTA DEWA FRUIT AQUEOUS AND METHANOL-CHLOROFORM EXTRACTS ON GLUCOSE UPTAKE AND METABOLISM

NURUL AKMARYANTI ABDULLAH

FPSK(M) 2014 4



IN VITRO EFFECTS OF *Phaleria macrocarpa* (Boerl.) MAHKOTA DEWA FRUIT AQUEOUS AND METHANOL-CHLOROFORM EXTRACTS ON GLUCOSE UPTAKE AND METABOLISM



By

NURUL AKMARYANTI ABDULLAH

Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirements for the Degree of Master of Science

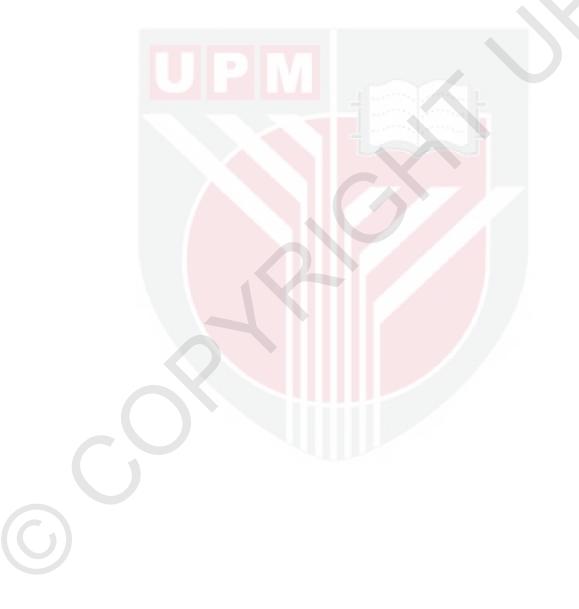
February 2014



COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirements for the degree of Master of Science

IN VITRO EFFECTS OF *Phaleria macrocarpa* (Boerl.) MAHKOTA DEWA FRUIT AQUEOUS AND METHANOL-CHLOROFORM EXTRACTS ON GLUCOSE UPTAKE AND METABOLISM

By

NURUL AKMARYANTI ABDULLAH

February 2014

Chair: Associate Professor Mohamad Aziz Dollah, PhD

Faculty: Faculty of Medicine and Health Sciences

Diabetes mellitus is the most common metabolic disease worldwide. One of the main pathophysiological defects of diabetes is insulin resistance due to impairment in the insulin-signaling pathway leading to a failure of the insulin stimulated glucose uptake in the target cells. Traditionally, plants such as Phaleria macrocarpa have been used to treat diabetes. The mechanism of how this *Phaleria macrocarpa* exert an anti-diabetic effect still obscure and thus a research consisted of five studies was conducted in vitro with the objectives to evaluate the anti-diabetic properties of *Phaleria macrocarpa* aqueous and methanol-chloroform fruit extracts using hepatocytes (HepG2 cells) as a model. In these studies, HepG2 cells were cultured in RPMI media in the presence of Phaleria macrocarpa fruit extracts (0, 0.01, 0.1 and 1 mg/mL) or metformin (1 mg/mL) or insulin (100 nM). Metformin and insulin were used as positive control. There are five studies including glucose uptake assay, inhibitor studies using wortmannin (PI3-kinase inhibitor) and genistein (protein tyrosine kinase inhibitor), glycogen synthesis assay and glycogen synthase activity assay. The present studies showed that both Phaleria macrocarpa extracts (aqueous and methanol-chloroform) had demonstrated the ability to enhance glucose uptake activity by up to 5-folds (p<0.05). That is similarly to the metformin when compared to the control. However, insulin showed the highest in glucose uptake activity (7-folds). The efficacy of aqueous extract was found to be slightly better than the commercial drug, metformin. In inhibitors study, the glucose uptake activity in cell cultures pre-treated with wortmannin or genistein and later with Phaleria macrocarpa extracts were significantly reduced (p<0.05) suggesting a possible involvement of PI3-kinase pathway and protein tyrosine kinase pathway in Phaleria macrocarpa-induced glucose uptake activity. The percentages of



inhibition at all doses of *Phaleria macrocarpa* extracts were comparable with the percentage of inhibition on insulin action. These indicated that the Phaleria *macrocarpa* action was mimicking the action of insulin. Glycogen synthesis was stimulated in HepG2 cells by more than 1-fold after treatment with Phaleria macrocarpa extracts. Metformin showed no effect on glycogen synthesis whereas insulin caused a maximum increased in glycogen synthesis activity in two hours. Both aqueous and methanol-chloroform of *Phaleria macrocarpa* fruit (p<0.05) glycogen synthase activity. The extracts significantly increased significant changes in enzyme activities were observed at as low as 0.01 mg/mL of *Phaleria macrocarpa* extracts while the maximum effects was observed at 1.0 mg/mL aqueous extract. This was similar to insulin effect and thus Phaleria *macrocarpa* was able to increase glycogen synthase activity at the same rate as insulin. Moreover, metformin also demonstrated significant increase in glycogen synthase activity (p<0.05) when compare to control, however its activity was lower than insulin. These studies concluded that *Phaleria macrocarpa* extracts have the ability to increase glucose uptake, glycogen synthesis and glycogen synthase activity similar to insulin action. Thus, Phaleria macrocarpa fruit extracts have insulin mimic activity. Therefore, with further investigation including clinical trial, Phaleria macrocarpa has a therapeutic potential as an anti-diabetic agents.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan ijazah Sarjana Sains

Kesan Estrak Akues dan Metanol-Kloroform Buah *Phaleria macrocarpa* (Boerl.) Mahkota Dewake Atas Pengambilan dan Metabolisme Glukosa In Vitro

Oleh

NURUL AKMARYANTI ABDULLAH

Februari 2014

Pengerusi: Profesor Madya Mohamad Aziz Dollah, PhD

Fakulti: Fakulti Perubatan dan Sains Kesihatan

Penyakit diabetes adalah penyakit metabolik yang tersebar meluas di seluruh dunia.Salah satu punca utama penyakit diabetes adalah disebabkan oleh rintangan hormon insulin yang berpunca dari penurunan dalam penghantaran isyarat insulin yang kemudian membawa kepada kegagalan insulin untuk merangsang pengambilan glukosa oleh sel-sel tubuh. Secara tradisional, Phaleria macrocarpa ialah sejenis tumbuhan yang telah digunakan untuk merawat diabetes. Mekanisme bagaimana Phaleria macrocarpa boleh memberi kesan anti-diabetik masih tidak diketahui. Oleh yang demikian, lima kajian telah dijalankan secara in-vitro menggunakan sel hepatosit (HepG2) sebagai model dengan objektif untuk menilai ciri-ciri anti-diabetik estrak akues dan ekstrak metanol-kloroform buah Phaleria macrocarpa. Dalam kajian ini, sel HepG2 dieram dalam media RPMI dan diberi masing-masing estrak buah Phaleria macrocarpa (0, 0.01, 0.1 dan 1 mg/mL) atau insulin (100 nM) atau metformin (1 mg/mL).Metformin dan insulin digunakan sebagai kawalan positif. Terdapat lima kajian termasuk kajian pengambilan glukosa, kajian perencat meggunakan wortmannin (perencat PI3-kinase) dan genistein (perencat protein tyrosine kinase), sintesis glikogen dan kajian aktiviti enzim glikogen sintase. Kajian ini menunjukkan bahawa kedua-dua Phaleria macrocarpa ekstrak (akues dan metanol-kloroform) mempunyai keupayaan untuk meningkatkan aktiviti pengambilan glukosa sehingga 5 kali ganda (p<0.05) dari kawalan. Kesan ini adalah sama seperti kesan metformin. Walau bagaimanapun, insulin menunjukkan aktiviti pengambilan glukosa paling tinggi (7 kali ganda). Keberkesanan ekstrak air didapati sedikit lebih baik daripada ubat komersial iaitu metformin.Dalam kajian kedua dan ketiga, aktiviti pengambilan glukosa dalam sel didik yang diberi wortmannin atau genistein menyebabkan aktiviti pemgambilan glukosa oleh sel didik berkurang dengan ketara (p<0.05). Ini menunjukkan terdapat penglibatan PI3-kinase dan tyrosine kinase dalam aktiviti



pengambilanglukosa oleh HepG2 sel. Peratusan perencatan kemasukan glukosa oleh semua dos ekstrak Phaleria macrocarpa adalah setara dengan peratusan perencatan terhadap tindakan insulin. Ini menunjukkan bahawa, tindakan Phaleria macrocarpa adalah menyamai dengan tindakan insulin.Sintesis glikogen yang dirangsang di dalam sel HepG2 adalah lebih daripada 1 kali ganda selepas diberi ekstrak Phaleria macrocarpa. Metformin tidak menunjukkan sebarang kesan ke atas sintesis glikogen manakala insulin menunjukkan kesan maksimum terhadap aktiviti sintesis glikogen dalam masa dua jam. Kedua-dua esktrak akues dan metanol-kloroform Phaleria macrocarpa meningkatkan aktiviti enzim glikogen synthase dengan ketara (p<0.05). Perubahan ketara aktiviti enzim adalah berkadar dengan dos Phaleria macrocarpa ekstrak dan menyamai dengan kesan insulin.Selain itu, metformin juga menunjukkan peningkatan yang ketara dalam aktiviti enzim glikogen synthase (p<0.05) apabila dibandingkan dengan kumpulan kawalan, namun aktivitinya adalah lebih rendah daripada insulin.Kajian ini membuktikan bahawa ekstrak Phaleria macrocarpa mempunyai keupayaan untuk meningkatkan pengambilan glukosa, sintesis glikogen dan aktiviti enzim glikogen synthase pada sel HepG2 yang menyamai dengan tindakan insulin.Oleh itu, kajian ini dapat membuktikan peranan dan mekanisme tindakan Phaleria macrocarpa untuk mengurangkan gula dalam darah. Oleh itu, untuk mengetahui lebih lanjut mengenai potensi Phaleria macrocarpa sebagai agen anti-diabetik, adalah dicadangkan membuat siasatan lanjut melibatkan percubaan klinikal.

ACKNOWLEDGMENTS

First, I would like to express my sincere gratitude to my supervisor, Associate Professor Dr. Mohamad Aziz Dollah. His wise opinion, extensive knowledge, devotion and enthusiasm gave significant positive influence on the development of this project. He also is the one who introduced me to the field of Radiation Biology, which I now began to love. I have always appreciated his patience and willingness to make time to discuss the obstacles I encountered throughout the project. I also thank him for never complaining and losing patience during supervision as well as correcting my thesis draft. He always stood behind me in the difficult times and provides me with the kindest and wisest words of advice that made hard phase thousand times easier.

Secondly, I would like to thank both my committee members Associate Professor Dr.Sabrina Sukardi and Dr. Abdah Akim for their encouragement and guidance especially on Physiology and Biochemistry parts of my project. I would also like to thank Radioisotope Laboratory staff, Mr Fadzlee Mohd Noh for helping me especially on machines preparations throughout my graduate years.

My sincere thanks goes to my fellow lab-mates and friends especially Hazirah, Nurhayatie, Thilaga, Azimah, Nor Afiqah, Saadah, Dila and Sarah for their help, motivation, and useful suggestions from time to time. I would be missing you all very much.

Finally and most importantly, I offer a heartfelt thank you to my dearest family for their endless love, faith and support. Thank you for believing in me all these years. I would like to dedicate this dissertation to them.



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Mohamad Aziz Dollah, PhD

Associate Professor Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Chairman)

Sabrina Sukardi, PhD

Associate Professor Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Member)

Abdah Md Akim, PhD

Senior Lecturer Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean, School of Graduate Studies Universiti Putra Malaysia.

Date: 21 April 2014

DECLARATION

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrent for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fullyowned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before this thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:	Date:	

Name and Matric No: Nurul Akmaryanti Abdullah (GS29293)

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- the supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:	Signature:
Name of	Name of
Chairman of	Chairman of
Supervisory	Supervisory
Committee:	Committee:
Signature:	Signature:
Name of	Name of
Chairman of	Chairman of
Supervisory	Supervisory
Committee:	Committee:

TABLE OF CONTENTS

	Page
ABSTRACT	ii
ABSTRAK	iv
ACKNOWLEDGEMENTS	vi
APPROVAL	vii
DECLARATION	ix
LIST OF TABLES	xiii
LIST OF FIGURES	XV
LIST OF ABBREVIATIONS	xvii
CHAPTER	
	1
1INTRODUCTION	1
2 LITERATURE REVIEW	
2.1 Glucose Homeostasis	4
2.2 Metabolic Pathway of Glucose in the Liver	6
2.3 Insulin and Its Signaling Pathway	7
2.3.1 PI3-kinase-Akt Pathway	8
2.3.2 Translocation of GLUT4	9
2.4 Diabetes Mellitus and Classification	10
2.4.1 Type 1 Diabetes Mellitus	11
2.4.2 Type 2 Diabetes Mellitus	11
2.5 Metformin	12
2.6 Phaleria macrocarpa	13
2.7 Previous Scientific Study of Phaleria macrocarpa	14
2.8 Anti-diabetic Properties of <i>Phaleria macrocarpa</i>	15
3 MATERIALS AND METHODS	
3.1 Chemicals and Reagents	17
3.2 Preparation of <i>Phaleria macrocarpa</i> dried fruit slices	17
3.3 Preparation of <i>Phaleria macrocarpa</i> Extracts	
3.3.1 Preparation of <i>Phaleria macrocarpa</i> Aqueous Extract	17
3.3.2 Preparation of <i>Phaleria macrocarpa</i> Methanol-Chloroform	17
Extract	
3.4 Cell Cultures	18
3.4.1 Cells Cultures Protocols and Maintenances	18
3.4.2 Determine Cells number and Cells Viability	19
3.5 Experiment 1: Study on the effect of <i>Phaleria macrocarpa</i> extract on	22
glucose uptake in HepG2 cells.	
3.6 Experiment 2: Study on the effect on glucose uptake activity after incubated with wortmannin (inhibitor) in HepG2	24
cells.	
3.7 Experiment 3: Study on the effect on glucose uptake activity after incubated with genistein (inhibitor) in HepG2 cells.	26
3.8 Experiment 4: Study on the effect of <i>Phaleria macrocarpa</i> extract on	28

	glycogen synthesis in HepG2 cells. 3.9 Experiment 5: Study on the effect of <i>Phaleria macrocarpa</i> extract on	30
	glycogen synthase activity in HepG2 cells.	20
	3.10 Statistical Analysis	33
4	RESULTS	
	4.1 Experiment 1: Effect of <i>Phaleria macrocarpa</i> extracts on glucose uptake in HepG2 cells.	34
	4.2 Experiment 2: Effect of <i>Phaleria macrocarpa</i> extracts on glucose uptake in HepG2 cells after incubated with wortmannin (inhibitor).	37
	4.3 Experiment 3: Effect of <i>Phaleria macrocarpa</i> extract on glucose uptake in HepG2 cells after incubated with genistein (inhibitor).	40
	4.4 Experiment 4: Effect of <i>Phaleria macrocarpa</i> extracts on glycogen synthesis in HepG2 cells.	43
	4.5 Experiment 5: Effect of <i>Phaleria macrocarpa</i> extracts on glycogen synthase activity in HepG2 cells.	46
5	DISCUSSION	50
6	CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	56

REFERENCES	57
APPENDICES	73
BIODATA OF STUDENT	86
LIST OF PUBLICATIONS	

C

LIST OF TABLES

Table		Page
3.1	Experimental design table used for <i>in-vitro</i> study.	20
4.1	Effect of <i>Phaleria macrocarpa</i> aqueous and methanol- chloroform extracts on glucose uptake in HepG2 cells.	35
4.2	Effect of <i>Phaleria macrocarpa</i> extracts on glucose uptake of HepG2 cells pre-treated with PI3-kinase inhibitor (Wortmannin).	38
4.3	Effect of <i>Phaleria macrocarpa</i> extracts on glucose uptake of HepG2 cells pre-treated with protein kinase inhibitor (Wortmannin).	41
4.4	Effect of <i>Phaleria macrocarpa</i> aqueous and methanol- chloroform extracts on glycogen synthesis in HepG2 cells.	44
4.5	Effect of <i>Phaleria macrocarpa</i> aqueous and methanol- chloroform extracts on glycogen synthase activity in HepG2 cells.	47
A1	Analysis of Variance (ANOVA) on the effect of <i>Phaleria</i> macrocarpa extract on glucose uptake in HepG2 cells.	73
A2	Analysis of Variance (ANOVA) on the effect of <i>Phaleria macrocarpa</i> extract on glucose uptake in HepG2 cells after incubated with wortmannin (inhibitor).	73
A3	Analysis of Variance (ANOVA) on the effect of <i>Phaleria macrocarpa</i> extract on glucose uptake in HepG2 cells after incubated with genistein (inhibitor).	74
A4	Analysis of Variance (ANOVA) on the effect of <i>Phaleria macrocarpa</i> extract on glycogen synthesis in HepG2 cells.	74
A5	Analysis of Variance (ANOVA) on the effect of <i>Phaleria macrocarpa</i> extract on glycogen synthase activity in HepG2 cells.	75
A6	Effect of the <i>Phaleria macrocarpa</i> aqueous and methanol- chloroform extracts on glucose uptake in HepG2 cells.	76

 \bigcirc

- A7 Effect of *Phaleria macrocarpa* extracts on glucose uptake of 77 HepG2 cells pre-treated with PI-3 Kinase inhibitor (Wortmannin).
- A8 Effect of *Phaleria macrocarpa* extracts on glucose uptake of 78 HepG2 cells pre-treated with protein kinase inhibitor (Genistein).
- A9 Effect of the *Phaleria macrocarpa* aqueous and methanolchloroform extracts on glycogen synthesisin HepG2 cells.
- A10 Effect of the *Phaleria macrocarpa* aqueous and methanolchloroform extracts on glycogen synthase activity in HepG2 cells.

80

79

LIST OF FIGURES

Figure		Page
2.1	Insulin signaling pathway regulating glucose transport.	10
2.2	Chemical structure of metformin.	12
2.3	Ripe fruits of <i>Phaleria macrocarpa</i> and dried slices of <i>Phaleria macrocarpa</i> fruits with seeds removed.	13
3.1	Flowchart of the overall study.	21
3.2	Flowchart for glucose uptake assay.	23
3.3	Flowchart for Inhibitor Study (Wortmannin)	25
3.4	Flowchart for Inhibitor Study (Genistein)	27
3.5	Flowchart for Glycogen Synthesis Assay	29
3.6	Flowchart for Glycogen Glycogen Synthase Enzyme Assay	32
4.1	Effect of various concentrations of <i>Phaleria macrocarpa</i> aqueous and methanol-chloroform extracts, insulin and metformin on relative glucose uptake in HepG2 cells.	36
4.2	Effect of various concentrations of <i>Phaleria macrocarpa</i> aqueous and methanol-chloroform extracts and insulin pre-treated with wortmannin on percentage of inhibition of glucose uptake in HepG2 cells.	39
4.3	Effect of various concentrations of <i>Phaleria macrocarpa</i> aqueous and methanol-chloroform extracts and insulin pre-treated with genistein on percentage reduction of glucose uptake in HepG2 cells.	42
4.4	Effect of various concentrations of <i>Phaleria macrocarpa</i> aqueous and methanol-chloroform extracts, insulin and metformin on glycogen synthesis in HepG2.	45
4.5	Effect of various concentrations of <i>Phaleria macrocarpa</i> aqueous and methanol-chloroform extracts, insulin and metformin on glycogen synthase activity in HepG2 cells.	48
A6	Phaleria macrocarpa ripe fruit.	81
A7	Phaleria macrocarpa seed.	81

C

A8 *Phaleria macrocarpa* fruit slice with seed removed. 82

82

A9 HepG2 cells (Adherent types cells).



LIST OF ABBREVATIONS

	Akt	Protein kinase
	ARC	Arcuate nucleus
	ATCC	American Type Culture Collection
	cAMPK	Cyclic adenosine monophosphate
	COX-2	Cyclooxygenase 2
	CPLA2	Cytosolic phospholipase A.
	DMSO	Dimethylsulfoxide
	EFGR	Epidermal growth factor receptor
	ERK	Extracellular signal-regulated kinase
	FDA	Food and Drug Administration
	G6P	Glucose-6-phosphate
	GLUT	Glucose transporter
	GSK 3β	Glycogen synthase kinase-3β
	HDL	High density lipoprotein
	HepG2	Hepatocellular carcinoma cells
	HER2	Human epidermal receptor-2
	HGP	Hepatic glucose production
	HL-60	Human promyelocytic leukemia cells
	IDF	International Diabetes Federation
	IGF	Insulin growth factor
	IR	Insulin receptor
	IRS 1/2	Insulin receptor substrate 1/2
	IRTK	Insulin receptor tyrosine kinase
	KIF3	Kinesins superfamily-3
	LC-MS	Liquid Chromatography/Mass Spectrometry
	LDL	Low density lipo protein
	LKB1	Liver kinase B1
	mRNA	Messenger ribonucleic acid
	MTT	Methylthiazol Tetrazolium Assay
	NEFAs	Non-esterified fatty acids
	NKC	Natural killer cells

PBS	Phosphate buffered saline		
PCSK9	Proprotein convertase subtilisin kexin 9		
PDK1	Phosphoinositide-dependent kinase-1		
PEPCK	Phosphoenolpyruvate carboxykinase		
PET	Positron Emission Tomography		
PI3-kinase	Phosphatidylinositol-3-kinase		
PIP3	Phosphatidylinositol (3,4,5)- trisphosphate		
РКВ	Protein-kinase B		
РКС	Protein-kinase C		
PTK	Protein tyrosine kinase		
SDS	Sodium dodecyl sulfate		
STAT3	Signal transducers and activators of transcription 3		
UDP	Uridine di-phosphate		
UTP	Uridine tri-phosphate		
VEGF-C	Vascular endothelial growth factor C		

C

LIST OF ANNOTATIONS

- > More than
- < Less than
- cm² Centimeter square
- dpm Disintegration per minute
 - g gram
- mg/mL Miligram per mililiter
 - mL Mililiter
 - nM Nanomolar
 - °C Degree celcius
- S.E.M Standard error of mean
- μCi/mL Micro curie per mililiter
 - μL Microliter

CHAPTER 1

INTRODUCTION

1.0 Overview

Diabetes is a complex and multi-factorial disease mainly caused by insulin deficit that leads to abnormalities in carbohydrate, fat and protein metabolism on target tissues (American Diabetes Association, 2010). Diabetes is considered the most prevalent disorder in the world. In 2011, there were already 336 million patients with diabetes mellitus worldwide. This value is expected to increase up to 552 million cases by the year of 2030 (International Diabetes Federation, 2011). Malaysia is very likely heading towards epidemic proportions based on the result in Malaysian National HealthMorbidity Survey III 2006 reported by Letchuman et al. (2010), which showed that Malaysia has alreadyreached the projected prevalence for year 2025. The number of diabetic patients increased from 8.3% in 1996 to 11.6 % in 2006.

Diabetes mellitus is a common metabolic disorder of the endocrine system. It is described by high blood glucose (sugar). Glucose is the main source of energy for the cells and tissues especially brain. Both adults and children require an intake of 130 grams of dietary glycemic carbohydrates per day to cover the glucose requirement of the brain (EFSA Panel on Dietetic Products Nutrition and Allergies, 2010). In a normal healthy people, blood glucose is strictly maintained by a close ranged, around 3.9 to 6.1 mmol/L (Philip, 2007). However, in diabetic patients, this range cannot be controlled by the body and always causes extra glucose circulating in the blood.

Insulin is the hormone that responsible in maintaining blood glucose level by stimulating phosphorylation and utilization of glucose. Liver is the primary site of insulin action. After a meal, insulin stimulates glucose uptake and store surplus glucose in the form of glycogen in the liver. Insulin initiates the glucose uptake into the cells by activating the insulin receptor (IR). It is then phosphorylates and recruits different substrate adaptors such as the insulin receptor substrates family (IRS)including IRS-1 and IRS-2. This initiatesthe phosphatidylinositol-3-kinase (PI3-kinase) pathwayandconducts the modulation of other proteins necessary for the metabolic effects of insulin (Klover and Mooney, 2004). PI3-kinase is known to play an important role in glucose transporter translocation (Lizcano and Alessi 2002). The inhibition of catalytic subunit of PI3-kinase, p110 with inhibitors for example wortmannin, can totallyblock most actions of insulin (Okada et al., 1994). Previous studies postulate that defect in glucose transporter translocation and impaired in insulin signaling cascade have been identified in patients with type 2 diabetes (Leng et al., 2004).

By looking at the prevalence mentioned earlier, many researchers interested to investigate new anti-diabetic drugs that can treat hyperglycemia better than the commercially available drugs due to its clinical side effects such as gastrointestinal side effect and increase cardiovascular risk(Hoffmann et al., 2003; Nissen and Wolski, 2007). Therefore, many diabetic patients are also recommended to receive complementary and alternative medicine therapies.

Herbal medicines have been practiced many years ago and research showed their value in treatment and prevention of various kinds of diseases. Recently, much attention has been paid to the discovery of natural products that may be advantageous in reducing the risks of diabetes. Normally, these natural products are usually has less toxicity, with little side effects than over the counter drugs (Jung et al., 2006). Diabetes is known can be controlled using natural products and therefore, the discovery and development of novel drugs for diabetic mellitus based on naturalproducts are very important (Dham et al., 2006). The available literature shows that there are more than 400 plant species possess anti-hyperglycemic activity (Rai, 1995).

Phaleria macrocarpa typically known as Crown God originated from Papua, Indonesia has been used traditionally to treat diabetic mellitus and other diseases such as rheumatic, kidney, gout, heart, hypertension and eczema (Lisdawati, 2002). Phaleria macrocarpa fruit is rich in alkaloids, saponins and flavoids while the leaves contain mangiferin, and saponins compound, which are classified into triterpenoid and steroid saponins, based on the structure of aglycon (Zhang et al., 2006; Saufi et al., 2008). Both of these compounds are reported to have anti-inflammatory and cytotoxic effects (Gotama et al., 1999). The isolated chemical constituents of *Phaleria macrocarpa* fruits extract include Icariside C3, magniferin, and gallic acid (Oshimi et al., 2008). It is found that mangiferin has a wide range of therapeutic effects including anti-diabetic, anti-HIV, anti-cancer, immunomodulatory and anti-oxidant activities (Yoshimi et al., 2001). Phaleria macrocarpa is one of the herbs that has been claimed traditionally to reduce high blood glucose levels. Sri Sugiwati et al., (2006) demonstrated that an oral administration of Phaleria macrocarpaleaves extract exhibited hypoglycemic effect and its ability to act as α -glucosidase inhibitors. Study in our laboratory has successful screened this plant on itsanti-diabetic properties. After 28 days of administration with Phaleria macrocarpafruit aqueous extract, blood glucose level of diabetes-induced rats found to reduce from 25 mmol/L to5 mmol/L (Raudhah and Dollah, 2008).In addition, it was reported that the bioassay-activity guided of anti-diabetic study of the methanol extract of Phaleria macrocarpa fruit showed that the flavonoids sub-fraction, which contains 22% of magniferin, has the highest anti-diabetic activity on diabetic rats (Rabyah et al., 2012). However, the mechanism of action of which Phaleria macrocarpa reduces the blood glucose still remains to be elucidated.

Hence, the present study focuses on finding the possible mechanism of actions of *Phaleria macrocarpa* extracts on glucose metabolism by determining in-vitro the glucose uptake, glycogen synthesis and glycogen synthase enzyme activity. In this study, the possible mechanism of action of *Phaleria macrocarpa* extracts on glucose uptake was also identified using specific inhibitors.

1.1 Objectives

1.1.1 General Objective

To investigate the ability of *Phaleria macrocarpa* aqueous and methanolchloroform fruits extracts to exert glucose metabolism effect in HepG2 cells by measuring the glucose uptake activity and its possible mechanism using inhibitors (wortmannin and genistein), glycogen synthesis and glycogen synthase enzyme activity.

1.1.2 Specific Objectives

- i. To study on the effect of *Phaleria macrocarpa* fruits extracts on glucose uptake by measuring the uptake activity of 2-Deoxy D-[1- $2,^{3}$ H (N)] glucose in HepG2 cells.
- ii. To determine on the effect of *Phaleria macrocarpa* fruits extracts on glucose uptake in HepG2 cells following incubation with wortmannin (PI3-kinase inhibitor).
- iii. To determine on the effect of *Phaleria macrocarpa* fruits extracts on glucose uptake in HepG2 cells following incubation with genistein (protein tyrosine kinase inhibitor).
 - iv. To measure on the effect of *Phaleria macrocarpa* fruits extracts on glycogen synthesis by measuring the activity of D-[¹⁴C]glucose in HepG2 cells.
 - v. To study on the effect of *Phaleria macrocarpa* fruits extract on glycogen synthase by measuring the activity of [³H]-UDP glucose in HepG2 cells.

1.2 Hypothesis

Phaleria macrocarpa extracts stimulate glucose uptake, glycogen synthesis and glycogen synthase activity in HepG2 cells and mimic insulin.

REFERENCES

- Adam, Z., Hamid, M., Ismail, A. and Khamis, S. (2009). Effect of *Ficus deltoidea* extracts on hepatic basal and insulin stimulated glucose uptake. *Journal of Biological Sciences* 9:796-803.
- Agung, B.S., Faried, A., Arifin, M.Z., Wiriadisastra, K. and Ohta, T. (2008). Herbal Medicine Isolation, *Phaleria macrocarpa* for primary glioblastoma multiforme cells. *Annual Epidemiology* 18:708–741.
- Akiyama, T., Ishida, J., Nakagawa, S. and Ogawara, H. (1987). Genistein, a specific inhibitor of tyrosine-specific protein kinases. *Journal of Biology and Chemistry* 262:5592–5595.
- Alessi, D.R., James, S.R., Downes, C.P., Holmes, A.B., Gaffney P.R, Reese, C.B. and Cohen, P. (1997). Characterization of a 3-phosphoinositide-dependent protein kinase which phosphorylates and activates protein kinase Balpha. *Journal of Current Biology*7:261-269.
- Ali, R.B., Atangwho, I.J., Kaur, N., Abraika, O.S., Ahmad, M. and Mahmud, R. (2012).Bioassay-guided antidiabetic study of *Phaleria macrocarpa* fruit extract.*Molecules*17:4986–5002.
- American Diabetes Association. (2010). Diagnosis and Classification of Diabetes Mellitus.*Diabetes Care*33:62-69.
- American Diabetes Association. (2011). Diagnosis and Classification of Diabetes Mellitus.*Diabetes Care* 34:63-69.
- Andersen, D.K., Ruiz, C.L. and Burant, C.F. (1994). Insulin reactions of hepatic glucose transporter is impaired in chronic pancreatis. *Annual Surgery*219:679-686.
- Angiosperm Phylogeny Group. (2003). An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG II. *Botanical Journal of the Linnean Society* 141:399-436.
- Anil Kumar, K.L. and Marita, A.R. (2000). Troglitazone prevents and reverses dexamethasone induced insulin resistance on glycogen synthesis in 3T3 adipocytes. *British Journal of Pharmacology*130: 351-358.
- Antonella, P. and Stefano, C. (2006). Central Control of Glucose Homeostasis.*The Review of Diabetic Studies* 3:54-60.
- Aronoff, S.L., Berkowitz, K., Shreiner, B. and Want, L. (2004). Glucose Metabolism and Regulation: Beyond Insulin and Glucagon. *Diabetic Spectrum* 17:183-190.

- Association American Diabetes. (2010). Diagnosis and Classification of Diabetes Mellitus. *Diabetes Care* 33:62-69.
- Atkinson, M.A. and Eisenbarth, G.S. (2001). Type 1 diabetes: new perspectives on disease pathogenesis and treatment. *Lancet*.358:221–229.
- Bauzine, M., Van, B. and Maassen, A. (2005). Genistein directly inhibits GLUT4mediated glucose uptake in 3T3-L1 adipocytes. *Biohemistry Biophysic Research Community* 326:11–14.
- Bell, G.I.; Pilkis, S.J., Weber, I.T. and Polonsky, K.S. (1996). Glucokinase Mutations, Insulin Secretion, and Diabetes Mellitus. Annual Review of Physiology 58:171-186.
- Berg, J.M, Tymoczko, J.L and Stryer, L. (2002). Biochemistry (5th Edition ed.). New York: W H Freeman.
- Bhatia, E. and Aggarwal, A. (2007). Insulin therapy for patients with type 1 diabetes. *Supplement of Japi* 55:29-34.
- Bode, M.W. (2004). Medical Management of Type 1 Diabetes. American Diabetes Association.
- Bouche, C., Serdy, S., Kahn, C.R. and Goldfine, A.B. (2004). The cellular fate of glucose and its relevance in type 2 diabetes. *Endocrinology Reviews* 25: 807–830.
- Bouskila, M., Hirshman, M.F., Jensen, J., Goodyear, L.J. and Sakamoto, K. (2008).Effects of Insulin and Transgenic Overexpression of UDP-glucose Pyrophosphorylase on UDP-glucose and Glycogen Accumulation in Skeletal Muscle Fiber.American Journal of Physiology-Endocrinology And Metabolism 294: 28-35.
- Brady, M.J. (2003). Measurement of Glycogen Synthesis and Glycogen Synthase Activity in 3T3-L1 Adipocytes. In O. S., Diabetes Mellitus:Methods and Protocols. Totowa: Humana Press Inc.
- Burgess, S.C., He, T., Yan, Z., Sherry, A.D., Malloy, C.R., Browning, J.D. and Magnuson, M.A. (2007). Cytosolic phosphoenolpyruvate carboxykinase does not solely control the rate of hepatic gluconeogenesis in the intact mouse liver. *Cell Metabolism* 5:313–320.

Cahill, G.J. (1971). Physiology of insulin in man. Diabetes 20:785-799.

- Calera, M.R., Martinez, C., Liu, H., Jack, A.K., Birnbaum, M.J. and Pilch, P.F. (1998). Insulin increases the association of Akt-2 with Glut4-containing vesicles. *Journal of Biology and Chemistry*273:7201–7204.
- Cazarolli, L.H., Zanatta, L., Alberton, E.H., Figueiredo, M.S.R.B., Folador P. and Damazio R.G., Pizzolatti M.G.b, Silva F.R.M.B. (2008). Flavonoids: Cellular

and molecular mechanism of action in glucose homeostasis. *Mini-Reviews in Medicinal Chemistry* 8:1032-1038.

- Chambers, J.K., Macdonald, L.E., Sarau, H.M., Ames, R.S., Freeman, K., Foley J.J., Zhu, Y. and McLaughlin, M.M. (2000). A G Protein-coupled Receptor for UDP-glucose.*The Journal of Biological Chemistry*275:10767-10771.
- Chan, J.C., Malik, V. and Jia, W. (2009). Diabetes in Asia: epidemiology, risk factors, and pathophysiology. *Journal of American Medical Association*301:2129–2140.
- Chen, T.W., Pollak, M. and Hung, H. (1998). Inhibition of Insulin-like Growth Factor Signaling Pathways in Mammary Gland by Pure Antiestrogen ICI 182,7801. Journal of Clinical Cancer Research. 7:2545-2554.
- Cheatham, B. and Kahn, C.R. (1995). Insulin action and the insulin signalling network. *Endocrinology Review* 16:117-142.
- Cherrington, A.D., Edgerton, D. and Sindelar, D.K. (1998). The direct and indirect effects of insulin on hepatic glucose production in vivo.*Diabetologia* 41:987-996.
- Chong, S.C., Dollah, M.A., Chong, P.P. and Maha, A. (2012)*Phaleria macrocarpa* (Scheff.) Boerl fruit aqueous extract enhances LDL receptor and PCSK9 expression in vivo and in vitro. Journal of Ethnopharmacology 137:817–827.
- Chynna, S., William, A.H., Stephen, G., Umesh, M., Melissa, C., Kristina, I.R., David, D. and Jeffery, B. (2004). Insulin Secretion in Type 1 Diabetes. *Diabetes*53:426-433.
- Clarke, J.F., Young, P.W., Yonezawa, K., Kasuga, M. and Holman, G.D. (1994). Inhibition of the translocation of GLUT1 and GLUT4 in 3T3-L1 cells by the phosphatidylinositol 3-kinase inhibitor, wortmannin. Journal of Biochemistry 300:631-635.
- Combs, T.P., Berg, A.H., Obici, S., Scherer, P.E. and Rossetti, L. (2001). Endogenous glucose production is inhibited by the adipose-derived protein Acrp30. *Journal of Clinical Investigation*108:1875-1881.
- Csepanyi-Komi, R., Levay, M. and Ligeti, E. (2012).Small G proteins and their regulators in cellular signalling.*Molecular and Cellular Endocrinology* 353:10-20.
- Cushman, J.W. and Wardzala, L.J. (1980). Potential mechanism of insulin action on glucose transport in the isolated rat adipose cell. *Journal of Biochemistry and Biology* 25:4578-4562.
- Cusi, K., Consoli, A. and DeFronzo, R.A. (1996). Metabolic effects of metformin on glucose and lactate metabolism in noninsulin-dependent diabetes mellitus. *Journal of Clinical Endocrinology Metabolism* 81:4059–4067.

- Czech, M.P. and Massaguc, J. (1982). Subunit structure and dynamics of the insulin receptor. *Federation Proceedings* 41:2719-2723.
- DeFronzo, R.A., Gunnarsson, R., Bjorkman, O., Olsson, M. and Wahren, J. (1985).Effects of insulin on peripheral and splanchnic glucose metabolism in noninsulin-dependent (type II) diabetes mellitus.*Journal of Clinical Investigation* 76:149-155.
- Detaille D., Wiernsperger N., Devos P. (1998). Potentiating effect of metformin on insulin-induced glucose uptakeand glycogen metabolism within Xenopus oocytes. *Diabetologia*. 41: 2-8
- Dimitriadis, G., Mitroub, P., Lambadiaria, V., Maratoub, E. and Sotirios, A. R. (2011). Insulin Effects in Muscle and Adipose Tissue. *Diabetes Research and Clinical Practice* 1:52-59.
- Dham, S., Shah, V. S., Hirsch, S. and Banerji, M. A. (2006). The role of complementary and alternative medicine in diabetes. *Current Diabetes Reports* 6:251–258.
- Dineshkumar, B., Mitra, A. and Manjunatha, M. (2010). Studies on the anti-diabetic and hypolipidemic potentials of mangiferin (Xanthone Glucoside) in STZinduced Type I and Type II diabetic model rats. *International Journal of Advanced Pharmacological Sciences*.1:75-85.
- Duncan, R. E., Ahmadian, M., Jaworski, K., Sarkadi-Nagy, E. and Sul, S.H. (2007).Regulation of lipolysis in adipocytes. *Annual Review Nutrition* 27:79-101.
- EFSA Panel on Dietetic Products, Nutrition and Allergies (NDA) (2010). Scientific Opinion on Dietary Reference Values for fats, including saturated fatty acids, polyunsaturated fatty acids, monounsaturated fatty acids, trans fatty acids, and cholesterol. *EFSA Journal* 8:1461
- Eldar-Finkelman, H. and Yarkoni, M. (2002).Insulin resistance and glycogen synthesis, roles in liver muscle and adipose tissue. New York: Harwood Academic Press.
- Faried, A., Kurnia, D., Faried, N., Usman, T., Miyazaki, H. and Kuwano, K.H. (2007) Anticancer effects of gallic acid isolated from Indonesian herbal medicine, *Phaleria macrocarpa* (Scheff.) Boerl, on human cancer cell lines.*International Journal of Oncology* 30:605-613.
- Fernandes, A. P. (2011). "Understanding post-prandial glucose clearance by peripheral organs: the role of the hepatic parasympathetic system." *Journal of Neuroendocrinology* 23:1288-1295.
- Folli, F., Saad, M.J., Backer, J.M. and Kahn, C.R. (1993).Regulation of phosphatidylinositol 3-kinase activity in liver and muscle of animal models of

insulin-resistant and insulin-deficient diabetes mellitus. Journal of Clinical Investigation 92:1787-1794.

- Frank, Q.N. and Mary C.Q. (1993). Allosteric Regulation of Glycogen Synthase in Liver. *A Physiological Dilema* 268:13286-13290
- Frank, T., Masur, K. and Zanker K.S. (2008).Pathophysiology of Diabetes Mellitus Type 2: Roles of Obesity, Insulin Resistance and Cell Dysfunction. *Diabetes* and Cancer.19:1-18
- Froguel, P. and Velho, G. (1999). Molecular Genetics of Maturity-Onset Diabetes of the young. *Trends in Endocrinology and Metabolism* 10:142-146.
- Gerich, J.E. (1993). Control of glycaemia. *Baillieres Clinical Endocrinology Metabolism* 7:51–586.
- Gerish, J.E. (2000). Physiology of Glucose Homeostasis. *Diabetes Obesity Metabolism* 2:345-350.
- Giannarelli, R., Aragona, M., Coppelli, A. and Del Prato, S. (2003). Reducing Insulin Resistance with Metformin :The Evidence Today. *Diabetes and Metabolism* 29:6528-6535.
- Girard, J. (2006). The Inhibitory Effects of Insulin on Hepatic Glucose Production Are Both Direct and Indirect. *Diabetes* 2:65-69.
- Gotama, I.B.I, Sugiarto, S. and Nurhadi, M. (1999). Inventory of Indonesian Medicinal Plants. (Vol. 5). Jakarta: Ministry of Health.
- Graeme, I. B., Toshiaki, K., John, B.B., Charles, F.B., Denis, L., Hirofumi, F. and Susumu, S. (1990). Molecular Biology of Mammalian Glucose Transporters. *Diabetes Care* 13:199-208.
- Gray, A.M. and Flatt, P.R.(1997).Insulin-releasing and insulin-like activity of *Agaricus campestris* (mushroom).*Journal of Endocrinology* 157 259–266.
- Grimes, C.A. and Jope, R.S. (2001). The multifaceted roles of glycogen synthase kinase 3β in cellular signaling. Progress in Neurobiology 65: 391–426.
- Gunton, J. D. (2003). Metformin rapidly increases insulin receptor activation in human liver and signals preferentially through insulinreceptor substrate-2. *Journal Clinical Endocrinology Metabolism* 88:1323-1332.
- Gupta, D., Varma, S. and Khandelwal, R.L. (2007).Long-term effects of tumor necrosis factor- treatment on insulin signaling pathway in HepG2 cells and HepG2 cells overexpressing constitutively active Akt/PKB.Journal of Cellular Biochemistry100:593-607.
- Gupta, R.N., Pareek, A., Suthar, M., Rathore, G.S., Basniwal P.K. and Jain D. (2009). Study of glucose uptake activity of Helicteres isora Linn, fruits in L-6

cell lines. International Journal of Diabetes Development Countries29:170–173.

- Ha, T.H., Trung, T.N., Hien, T.T. and Dao, T.T. (2010). Selected compounds derived from Moutan Cortex stimulated glucose uptake and glycogen synthesis via AMPK activation in human HepG2 cells. *Journal of Ethnopharmacology* 131:417-424.
- Haddad, P. M. (2006). Antidiabetic Plants of North Africa and the Middle East. Boca Raton, Florida: CRC Press.
- Hans, P.M.M. and Jonathan, D. (2010). Measuring GLUT4 translocation in mature muscle fibers. *Endocrinology and Metabolism* 2:169-179.
- Harmanto, N. (2003). Mahkota Dewa: Obat Pusaka Para Dewa. Jakarta: Mahkota Dewa: Obat Pusaka Para Dewa.
- Hendra R. (2011). Flavonoid Analyses and Antimicrobial Activity of Various Parts of *Phaleria macrocarpa* (Scheff.) Boerl Fruit. *International Journal of Molecular Sciences*.12:3422-3431.
- Hendra, R., Ahmad, S., Oskoueian, E., Sukari, A. and Shukor, M.Y. (2011). Antioxidant, Anti-inflammatory and Cytotoxicity of *Phaleria macrocarpa* (Boerl.) Scheff Fruit. *BMC Complementary and Alternative Medicine*11:1-10.
- Heydrick, S.J., Jullien, D., Gautier, N., Tanti, J.F., Giorgetti, S., Van Obberghen E. and Le Marchand-Brustel, Y. (1993). Defect in skeletal muscle phosphatidylinositol-3-kinase in obese insulin-resistant mice. *Journal of Clinical Investigation*, 91, 1358-1366.
- Hirokawa, N. (2000). Stirring up development with the heterotrimeric kinesin KIF3. *Traffic* 1:29-34.
- Hoffmann, I.S., Roa, M., Torrico, F. and CubedduL, X. (2003). Ondansetron and metformin-induced gastrointestinal side effects. *American Journal of Therapeutics*, 10, 447–451.
- Hong, G., Julie, A. L. (2004). Kinetic measurements of phosphoglucomutase by direct analysis of glucose-1-phosphate and glucose-6-phosphate using ion/molecule reactions and Fourier transform ion cyclotron resonance mass spectrometry. *Analytical Biochemistry* 329:269–275.
- Horton, J.D., Goldstein, J.L. and Brown, M.S. (2002). SREBPs: Activators of the complete program of cholesterol and fatty acid synthesis in the liver. *Journal of Clinical Investigations* 10:1125 –1131.

Hou, D. (1960). Thymelaeaceae. Leyden: Noordhoff International Publishing.. Noordhoff. International Publishing, Leyden. Pg: 1–48.

- Hundal, R.S., Krssak, M., Dufour, S., Laurent D. and Lebon, V., (2000). Mechanism by which metformin reduces glucose production in type 2 diabetes. *Diabetes* 49:2063-2069.
- Hubbard, S.R. (2013). The Insulin Receptor: Both a Prototypical and Atypical Receptor Tyrosine Kinase. *Cold Spring Perspectives in Biology* 5:10-15.
- Imamura, T.P., Vollenweider, K. E., Clodi, M., shibashi K., Nakashima, K., Ugi, S., Adams J.H. and Olefsky, J.M. (1999). G alpha-q/11 protein plays a key role in insulin-induced glucose transport in 3T3-L1 adipocytes. *Molecular Cell Biology* 19:6765-6774.
- International Diabetes Federation. (2013). Diabetes Atlas.2013 International Diabetes Federation.
- International Diabetes Federation. (2011). Diabetes Atlas.2011 International Diabetes Federation.
- Iozzo, P., Hallsten, K., Oikonen, V., Virtanen, K.A., Kemppainen J., Solin, O., Ferrannini, E., Knuuti, J. and Nuutila, P. (2003). Insulin-mediated hepatic glucose uptake is impaired in type 2 diabetes: evidence for a relationship with glycemic control. *Journal Clinical Endocrinology Evidence*. 2055 –2060.
- Irianti, T., Fakhrudin, N. and Efendy, K. (2008). The inhibition effect of water extract of mahkota dewa fruit (*Phaleria macrocarpa* (Scheff.) Boerl.) compared to vitamin C on tyrosine photodegradation. *Berkala Ilmiah Biologi*. 7:75-81.
- Johnson, J.L., Rupasinghe, S.G., Stefani, F., Schuler, M.A., Gonzalez de and Mejia E. (2011). Citrus flavonoids luteolin, apigenin, and quercetin inhibit glycogen synthase kinase-3β enzymatic activity by lowering the interaction energy within the binding cavity. *Journal of Medicinal Food* 14:325-333.
- Jung, M., Park, M., Lee, H.C., Kang, Y.H. and Kim, S.K. (2006). Antidiabetic agent from medicinal plants. *Current Medical Chemistry* 13:1203-1218.
- Jung, S.H., Ha, Y.J., Shim, E.K., Choi, S.Y., Jin, J.L. and Yun-Choi, H.S. (2007). Insulin-mimetic and insulin-sensitizing activities of a pentacyclic triterpenoid insulin receptor activator. *Journal of Biochemistry* 403:243-250.
- Kahn, C.R., White, M.F., Shoelson, S.E., Backer, J.M., Araki E., Cheatham, B., Csermely P., Folli F., Goldstein, B.J., Huertas P., Rothenberg, P.L., Saad M.J.A., Siddle, K., Sun X-J, Wilden, P.A., Yamada, K. and Kahn, S.A. (1993). The insulin receptor and its substrate: molecular determinants of early events in insulin action. *Recent Progress in Hormone Research* 48:291-339.
- Kalekar, S.A., Munshi, R.P., Bhalerao, S.S. and Thatte, U.M. (2013). Insulin sensitizing effect of 3 Indian medicinal plants: an in vitro study. Indian Journal of Pharmacology 45:30-33.

- Karttunen, P., Uusitupa, M. and Lamminsivu, U. (1983). The Pharmacokinetics of metformin: A comparison of The Properties of A Rapid-Release and A Sustained-Release Preparation. *International Journal of Clinical Pharmacology, Therapy and Toxicology* 21:31-36.
- Katz, J., Golden, S. and Wals, P.A. (1976). Stimulation of hepatic glycogen synthesis by amino acids. *Proceedings of the National Academy of Sciences* 73: 3433– 3438.
- Kausch, C., Hamann, A., Uphues, I., Niendorf, A. and Müller-Wieland, D. (2005). Association of impaired phosphatidylinositol 3-kinase activity in GLUT1containing vesicles with malinsertion of glucose transporters into the plasma membrane of fibroblasts from a patient with severe insulin resistance and clinical features of Werner syndrome. *Journal of Clinical Endocrinology and Metabolism* 85:905–918.
- Keller, S.R., Scott, H.M., Mastick, C.C., Aebersold, R. and Lienhard, G.E. (1995). Cloning and characterization of a novel insulin-regulated membrane aminopeptidase from Glut4 vesicles. *Journal of Biology and Chemistry* 270:23612-23618.
- Kim, D.I., Lim, S.K. and Park, M.J. (2007). The involvement of phosphatidylinositol 3-kinase/Akt signaling in high glucose-induced downregulation of GLUT-1 expression in ARPE cells. *Journal of Life Science* 80:626–632.
- Kim, W.J., Veriansyah, B., Lee, Y.W. and Kim, J. (2010). Extraction of mangiferin from Mahkota Dewa (*Phaleria macrocarpa*) using subcritical water. *Journal of Industrial and Engineering Chemistry* 16:425–430.
- Kimball, S.R., Farrell, P.A. and Jefferson, L.S. (2002). Role of insulin in translational control of protein synthesis in skeletal muscle by amino acids or exercise. *Journal of Applied Physiology*93:168 –1180.
- Kimmel, B. and Inzucchi, S.E. (2005). Oral agents for type 2 diabetes: an update. *Clinical Diabetes*23:64–76.
- Khan, A.H. and Pessin, J.E. (2002).Insulin regulation of glucose uptake: a complex interplay of intracellular signalling pathways.*Diabetologia* 45: 1475-8143.
- Klover, P.J. and Mooney, R.A. (2004). Hepatocytes: Critical for glucose homeostasis. *International Journal of Biochemistry and Cell Biology* 36:753-758.
- König, M., Bulik, S. and Holzhütter, H.G. (2012). Quantifying the contribution of the liver to glucose homeostasis: a detailed kinetic model of human hepatic glucose metabolism. *PLoS Computer Biology*6:1-43.
- Kotani K., Ogawa W., Matsumoto M., Kitamura T and Sakaue H. (1998).Requirement of atypical protein kinase clambda for insulin stimulation

of glucose uptake but not for Akt activation in 3T3-L1 adipocytes.*Molecular Cell Biology* 18: 6971–6982.

- Krssak, M., Brehm, A., Bernroider, E., Anderwald, C., Nowotny P., Dalla, Man C., Cobelli, C., Cline, G.W, Shulman, GI, Waldhäusl, W and Roden, M. (2004). Alterations in postprandial hepatic glycogen metabolism in type 2 diabetes. *Diabetes* 3048-3056.
- Lauritzen, H.P., Ploug, T., Prats, C., Tavare, J.M. and Galbo, H. (2006). Imaging of insulin signaling in skeletal muscle of living mice shows major role of Ttubules. *Diabetes* 55:1300-1306.
- Lawrence, J.C. and Roach, P.J. (1997). New Insights Into the Role and Mechanism of Glycogen Synthase Activation by Insulin.*Diabetes*46:541-547.
- Lee J., Ju J., Park S., Hong S.J. and Yoon S. (2012). Inhibition of IGF-1 signaling by genistein: modulation of E-cadherin expression and downregulation of β -catenin signaling in hormone refractory PC-3 prostate cancer cells. *Nutrition Cancer*. 64:153-162.
- Leng, Y., Karlsson, H.R. and Zierath, J.R. (2004). Insulin signaling defects in type 2 diabetes. *Revision Endocrinology Metabolism Disorder* 5:111-117.
- Letchuman, G., Nazaimoon, W., Mohamad, W. W. and Chandran, L. (2010). Prevalance of Diabetes in the Malayian National Health Morbidity Survey III 2006. *Medical Journal of Malaysia* 65:173-179.
- Lisdawati, A. (2002). Senyawa lignan dari fraksi etil acetat daging buah mahkota dewa [*Phaleria macrocarpa* (Scheff.) Boerl.]. Magister Thesis,. Jakarta: Universitas Indonesia.
- Liu, F., Kim, J., Li, Y., Liu, X., Li, J. and Chen, X. (2001). An extract of Lagerstroemia speciosa L. has insulin-like glucose uptake-stimulatory and adipocyte differentiation-inhibitory activities in 3T3-L1 cells. *Journal of Nutrition* 131:2242-2247.
- Lizcano, J.M. and Alessi, D.R. (2002). The insulin signaling pathway. *Current Biology* 12:236-238.
- Loizou, C.L., Ozanne, S.E. and Hales, C.N. (1999). The effect of insulin on □5 desaturation in hepG2 human hepatoma cells and L6 rat muscle myoblasts.*Prostaglandins, Leukotrienes and Essential Fatty Acids* 61:89–95.
- Magnusson, I., Rothman, D., Katz, L., Shulman, R. and Shulman, G. (1992). Increased rate of gluconeogenesis in type II diabetes mellitus: a 13C nuclear magnetic resonance study. *Journal of Clinical Investigation* 90:1323-1327.
- Manning, B.D. and Cantley, L.C. (2007). AKT/PKB signaling: navigating downstream. *Cell*. 129:1261-1274.

- Markuns, J.F., Wojtaszewski, J.F.P. and Goodyear, L.J. (1999). nsulin and exercise decrease glycogen synthase kinase-3 activity by different mechanisms in rat skeletal muscle. *The Journal of Biological Chemistry* 274: 24896–24900.
- Masur K., Thevenod F. and Zanker K.S. (2008). Pathophysiology of Diabetes Mellitus Type 2: Roles of Obesity, Insulin Resistance and B-Cell Dysfunction. *Diabetes and Cancer* 19:1-18.
- Mathis, D. and Benoist, C. (2004). Back to central tolerance. *Immunity* 20:509-516.
- Matsumoto, M., Pocai, A, Rossetti, L., Depinho, R.A. and Accili, D. (2007).Impaired regulation of hepatic glucose production in mice lacking the forkhead transcription factor Foxo1 in liver.*Cell Metabolism* 6:208–216.
- Muhammad, G. S. (2008). The effect of mahkota dewa (*Phaleria macrocarpa* (Scheff) Boerl) leaf etanolic extract on splenic NK 1.1 cells activity. *Periodic Medical Science* 40:109-118.
- Muhammad, S. and Indrianti, P. (2012). Nanoencapsulation of The Flavanoids Isolated from Phaleria macrocarpa Leaf by Casein Micelle.*International Journal of Pharma and Bio Sciences*. 472 - 478.
- Muruganandan, S., Srinivasan, K., Gupta, S., Gupta, P.K. and Lal, J. (2005). Effect of mangiferin on hyperglycemia and atherogenicity in streptozotocin diabetic rats.*Journal of Ethnopharmacology*97:497-501.
- Musi, N., Hirshman, M.F. and Nygren, J. (2002). Metformin Increases AMP-Activated Protein Kinase Activity in Skeletal Muscle of Subjects with Type 2 Diabetes. *Diabetes* 51:2074-2081.
- Myers, J.M.G. and White M.F. (1996). Insulin signal transduction and the IRS proteins. *Annual Review Pharmacology Toxicology* 36:615–658.
- Myers, M.G. and White, M.F. (1995). New frontiers in insulin receptor substrate signaling. *Trends in Endocrinology and Metabolism*6:209–215.
- Nissen, S.E. and Wolski, K. (2007). ffect of rosiglitazone on the risk of myocardial infarction and death from cardiovascular causes. *The New England Journal of Medicine* 356:2457–2471.
- Niswender ,K.D., Gallis, B., Blevins, J.E., Corson, M.A., Schwartz, M.W. and Baskin, D.G. (2003). Immunocytochemical detection of phosphatidylinositol 3-kinase activation by insulin and leptin. *Journal of Histochemistry and Cytochemistry* 51:275-283.
- Nor Fariza, J., Fadzureena, A., Zunoliza, A., Luqman C., Pin, K.Y. and Adawiah, I. (2012). Anti-inflammatory Activity of the Major Compound from Methanol Extract of *Phaleria macrocarpa* Leaves. *Journal of Applied Sciences*12:1195-1198.

- Nuttall, F.Q. and Gannon, M.C, (1993). Allosteric Regulation of Glycogen Synthase in the Liver. *The Journal of Biological Biochemistry* 268:13286-13290.
- Nuttall, F.Q., Gilboe, D.P., Gannon, M.C., Niewoehner, C.B. and Tan A.W.H. (1988). Regulation of glycogen synthesis in the liver. *American Journal of Medicine* 28:77-85.
- Nuttall, F.Q., Ngo, A. and Gannon, M.C. (2008) Regulation of hepatic glucose production and the role of gluconeogenesis in humans: is the rate of gluconeogenesis constant? *Diabetes Metabolism Research Revision* 24:438–458.
- Obici, S., Zhang, B.B., Karkanias, G. and Rossetti, L. (2002). Hypothalamic insulin signaling is required for inhibition of glucose production. *Natural Medicine* 8:1376-1382.
- Ojuka, E.O. (2004). Role of Calsium and AMP kinase in the Regulation of Mitochondrial Biogenesis and GLUT4 Levels in Muscle. *Proceedings of Nutrition Society* 63:275-278.
- Okada, T., Sakuma, L., Fukui, Y., Hazeki, O. and Ui, M. (1994).Blockage of chemotactic peptide-induced stimulation of neutrophils by wortmannin as a result of selective inhibition of phosphatidylinositol 3-kinase.*Journal of Biology and Chemistry* 269:3563–3567.
- Oshimi, S., Zaima, K., Matsuno, Y., Hirasawa, Y., Iizuka, T. and Studiawan, H. (2008). Studies on the constituents from the fruits of *Phaleria* macrocarpa. Journal of Natural Medicine 62:207–210.
- Otto, M,.Breinholt, J. and Westergaard, N. (2003). Metformin inhibits glycogen synthesis and gluconeogenesis in cultured rat hepatocytes. *Diabetes, Obesity and Metabolism*5:189-194.
- Pagliassoti, M.J. and Horton, T.J. (1994). Hormonal and neural regulation of hepatic glucose uptake in the role of liver in maintaining glucose homeostasis. Davids and Cherington. Austin, Texas.
- Perriello, G., Misericordia, P., Volpi, E., Santucci, C., Ferrannini E., Ventura M., Santeusiano F., Brunetti P. and Bolli G. (1994). Acute antihyperglycemic mechanisms of metformin in NIDDM: evidence for suppression of lipid oxidation and hepatic glucose production. *Diabetes*. 43:920–928.
- Philip E.C. (2007). Hypoglycemia, functional brain failure and brain death. *Journal* of Clinical Investigations 117:868-870.
- Pinent, M., Bladé, M.C., Salvadó, M.J., Arola, L. and Ardévol, A. (2005). Metabolic fate of glucose on 3T3-L1 adipocytes treated with grape seed-derived procyanidin extract (GSPE). Comparison with the effects of insulin. *Journal of Agricultural and Food Chemistry* 53:5932-5935.

- Pocai, A., Obici, S., Schwartz, G. and Rossetti, L. (2005). A brain-liver circuit regulates glucose homeostasis. *Cell Metabolism* 1:53-61.
- Quesada, I., Tuduri, E., Ripoll, C. and Nadal, A. (2008). Physiology of the pancreatic α-cell and glucagon secretion: role in glucose homeostasis and diabetes. *Journal of Endocrinology*.199:5-19.
- Rabyah, A., Atangwho, J., Navneet, K., Mariam, A., Roziahanim, M. and Asmawi, M.Z. (2013). In vitro and in vivo effects of standardized extract and fractions of *Phaleria macrocarpa* fruits pericarp on lead carbohydrate digesting enzymes. *BMC Complementary and Alternative Medicine*.13:1-11.
- Rabyah, B.A., Item, J.A., Navneet, K., Elsnoussi, A.H., Ali, M.J., Asmawi, M.Z. and Mahmud, R. (2012). Hypoglycemic and anti-hyperglycemic study of *Phaleria macrocarpa* fruits pericarp. Journal Medicinal Plant Research 6:1982-1990.
- Radziuk, J. and Pye, S. (2001). Hepatic glucose uptake, gluconeogenesis and the regulation of glycogen synthesis. *Diabetes Metabolism Research* 17:250–272.
- Rai, M.K. (1995) A review on some antidiabetic plants of India. Ancient Science Life 14:42-54.
- Raudhah, R. and Dollah, M.A. (2008). Hypoglycemic effect of Phaleria macrocarpa fruit aqueous extract in diabetic induced rats. Serdang: Final Year Project, Universiti Putra Malaysia.
- Rinayanti, A., Radji, M., Munim, A. and Suyatna, F.D. (2013). Screening Angiotensin Converting Enzyme (ACE) Inhibitor Activity of Antihypertensive Medicinal Plants from Indonesia. *International Journal of Pharmacy Teaching* & *Practices*4:527-532.
- Ripudaman, S. Hundal, Kitt, F., Adam, B., Mayerson, P. S., Randhawa, S.I., Steven E.S. and Gerald, I.S. (2000). Mechanism by which high-dose aspirin improves glucose metabolism in type 2 diabetes. *Journal of Clinical Investigations* 109:1321–1326.
- Roach, P J., Cheng, C., Huang, D., Lin, A., Mu, J., Skurat, A.V., Wilson W. and Zhai L. (1998).Novel aspects of the regulation of glycogen storage.*Journal of Basic Clinical Physioliogy and Pharmacology* 9:139-151.
- Rodney, A.R. and David, R.B. (2012). Medical Physiology: Principles for Clinical Medicine. Indiana: Lippincott Williams & Wilkins.
- Rondinone, C.M., Wang, L.M., Lonnroth, P., Wesslau, C., Pierce J.H. and Smith U. (1997). Insulin receptor substrate (IRS) 1 is reduced and IRS-2 is the main docking protein for phosphatidylinositol 3-kinase in adipocytes from subjects with non-insulin-dependent diabetes mellitus. *Proceedings of the National Academy of Sciences* 94:4171-4175.

- Rosenstock, J, Sugimoto, D., Strange, P., Stewart, J.A., Soltes-Rak, E. and Dailey, G. (2006). Triple therapy in type 2 diabetes: insulin glargine or rosiglitazone added to combination therapy of sulfonylurea plus metformin in insulin-naive patients. *Diabetes Care* 29:554–559.
- Saadat, P., Zainudin, C.Z. and Dollah, M.A. (2013). Effect of *Phaleria macrocarpa* on sexual function of rats.*Avicenna Journal of Phytomedicine* 3: 371-377.
- Sabyasachi, S. (2008) Principal of Medical Physiology. New York,(USA). Germany: Georg Thieme Verlaq.
- Sadagurski, M. and White, M.F. (2013). Integrating Metabolism and Longevity Through Insulin and IGF1 Signaling. *Endocrinology and Metabolism Clinics* of North America 127-148.
- Sakla, M. S., Shenouda, N. S., Ansell P. J., Macdonald R.S. and Lubahn, D.B. (2007). Genistein affects HER2 protein concentration, activation, and promoter regulation in BT- 474 human breast cancer cells. *Endocrine* 32:69–78.
- Saltiel, A.R. and Pessin, J.E. (2002). Insulin signaling pathways in time and space.*Trends Cell Biology* 12:65 –71.
- Sangeetha, K.N., Sujatha, S., Muthusamy, V.S., Anand, S., Nithya N. and Velmurugan D. (2010). 3beta- taraxerol of Mangifera indica, a PI3K dependent dual activator of glucose transport and glycogen synthesis in 3T3-L1 adipocytes.*Biochimica et Biophysica Acta*1800:359–366.
- Sano H., P.G. (2011). Insulin-stimulated GLUT4 protein translocation in adipocytes requires the Rab10 guanine nucleotide exchange factor Dennd4C. *Journal of Biology and Chemistry* 286:16541–16545.
- Saufi, A., Von Heimendahl, C.B., Alfermann, A.W. and Fuss, E. (2008). Stereochemistry of lignans in *Phaleria macrocarpa* (Scheff.) Boerl. Z *Naturforsch C*. 16:13-16.
- Shepherd, P.R., Withers, D.J. and Siddle, K. (1998). Phosphoinositide 3-kinase: the key switch mechanism in insulin signalling. *Journal of Biochemistry* 333:471-490.
- Shodikin, A. (2009). Antimicrobial activity of ethanol extract of Mahkota Dewa (*Phaleria macrocarpa*) fruits and leaves against pseudomonas aeruginosa by agar dilution and scanning electron microscopy. Surabaya: Faculty of Medicine, University of Airlangga.
- Sindellar, D.K., Chu, C.A., Rohlie, M., Neal, D.W., Swift, L.L. and Cherrington A.D. (1997). The role of fatty acids in mediating the effects of peripheral insulin on hepatic glucose production in the conscious dog. *Diabetes* 46:187-196.

- Singh, A., Ebenso, E.E. and Quraishi, M.A. (2012). Theoretical and Electrochemical Studies of Metformin as Corrosion Inhibitor for Mild Steel in Hydrochloric Acid Solution. *International Journal of Electrochemical Science*7:4766 – 4779.
- Smith, R.M., Tiesinga, J.J., Shah, N., Smith, J.A. and Jarett, L. (1993). Genistein inhibits insulin-stimulated glucose transport and decreases immunocytochemical labeling of GLUT4 carboxyl-terminus without affecting translocation of GLUT4 in isolated rat adipocytes: additional evidence of GLUT4 activation by insulin. Archives of Biochemistry and Biophysics.300: 238–246.
- Srivastava ,A.K. and Pandey, S.K. (1998). Potential mechanism(s) involved in the regulation of glycogen synthesis by insulin. *Molecular and Cellular Biochemistry*182:135-141.
- Sri Sugiwati. S., Siswati, S. and Efy, A. (2006). Antihyperglycemic Activity of the Mahkota Dewa [*Phaleria macrocarpa* (Scheff.) Boerl.] Leaf Extracts as an Alpha-Glucosidase Inhibitor. *Makara Kesehatan* 13:74-78.
- Sujatha S., Anand, S., Sangeetha, K.N., K. Shilpa, J., Lakshmi, A., Balakrishnan,
 1.And Lakshmi, B.S. (2010).Biological evaluation of (3b)-STIGMAST-5-EN3-OL as potent anti-diabetic agent in regulating glucose transport using in vitro model.*International Journal of Diabetes Mellitus* 2:101-109.
- Takanaga, H., Chaudhuri, B. and Frommer, W. B. (2008). GLUT1 and GLUT9 as major contributors to glucose influx in HepG2 cells identified by a high sensitivity intramolecular FRET glucose sensor. *Biochemistry and Biophysic.Acta* 1778: 1091–1099.
- Tappy, L., Jéquier, E. and Schneiter, P. (2000). Autoregulation of Glucose Production. News Physiology Sciences 15:198–202.
- Tjandrawinata, R.R., Arifin, P.F., Tandrasasmita, O.M., Rahmi D. and Aripin A., (2010). DLBS1425, a *Phaleria macrocarpa* (Scheff.) Boerl. extract confers anti proliferative and proapoptosis effects via eicosanoid pathway. *Journal of Experimental therapeutics &Oncology* 3:187-201.
- Triastuti A. (2006). Efek Antiangiogenik Ekstrak Etanol Buah Mahkota Dewa *Phaleria macrocarpa* ada Membran Korio Alantois (CAM) Embrio Ayam Yang Terinduksi bFGF, Lapen.Prodi Farmasi UII. Yogyakarta.
- Vera, J.C., Reyes, A.M., Carcamo, J.G., Velasquez, F.V., Rivas, C.L., Zhang R.H., Strobel P., Iribarren R., Scher H.I., Slebe J.C. and Golde D.W. (1996). Genistein is a natural inhibitor of hexose and dehydroascorbic acid transport through the glucose transporter, GLUT1. *Journal of Biology Chemistry* 271:8719–8724.

- Vijayakumar, M.V., Singh, S., Chhipa, R.R. and Bha, M.K. (2005). The hypoglycaemic activity of fenugreek seed extract is mediated through the stimulation of an insulin signalling pathway. *British Journal of Pharmacology.*, 146:41-48.
- Villar-Palasí, C. and Guinovart, J.J. (1997). The role of glucose 6-phosphate in the control of glycogen synthase. *FASEB* 11:544 -558.
- Wahren, J. and Ekberg, K. (2007). Splanchnic regulation of glucose production. *Annual Review Nutrition*27:329-345.
- Wang, J., Eltoum, I. E. and Lamartiniere, C. A. (2004). Genistein alters growth factor signaling in transgenic prostate model (TRAMP). *Molecular Cell Endocrinology* 171–180.
- Waters, S.B., D'Auria, M., Martin, S.S., Nguyen, C., Kozma, L.M. and Luskey, K.L. (1997). The amino terminus of insulin-responsive aminopeptidase causes Glut4 translocation in 3T3-L1 adipocytes. *Journal of Biology and Chemistry* 272:23323-23327.
- Watson, R. T., and Pessin, J. E. (2006). Bridging the GAP between insulin signaling and GLUT4 translocation. *Trends Biochemistry Science* 31:215–222.
- Watson, R.T., Kanzaki, M. and Pessin, J.E. (2004). Regulated membrane trafficking of the insulin-responsive glucose transporter 4 in adipocytes. *Journal of Endocrinology Revision* 25:177-204.
- Wauwe, J.V. and Haefner, B. (2003). Glycogen synthase kinase-3 as drug target: from wallflower to center of attention. *Drug News and Perspectives* 16:557– 565.
- Wiernsperger, N.F. and Biley, C.J. (1994). The Antihyperglycemic Effect of Metformin. *Drugs* 59:31-39.
- Wilcock, C. and Bailey, C.J. (1999). Accumalation of Metformin by tissues of the Normal and Diabetic Mouse. *Xenobiotica*24:49-57.
- White, M.F. (1988). Mutation of the insulin receptor at tyrosine 960 inhibits signal transmission but does not affect its tyrosine kinase activity. *Cells* 54:641–649.
- Wright, D.C., Geiger, P.C. and Han, D.H. (2006). Are tyrosine kinases involved in mediating contraction-stimulated muscle glucose transport? *American Journal* of Physiology Endocrinology Metabolism 290:123-128.
- Wymann, M.P., Bulgarelli-Leva, G., Velebil, M.J., Pirola, L., Vanhaesebroeck B., Waterfield M.D. and Panayotou, G. (1996). Wortmannin inactivates phosphoinositide 3-kinase by covalent modification of Lys-802, a residue involved in the phosphate transfer reaction. *Molecular and Celullar Biology* 16:1722–1733.

- Yosie, A., Effendy, M.A.W., Sifzizul, T.M. and Habsah, M. (2011). Antibacterial, radical scavanging activities and cytotoxicity properties of *Phaleria* macrocarpa (Scheff.) Boerl leaves in HepG2 cell lines. *International Journal Pharmacology Science Research* 1700–1706.
- Yoshida, T., Okuno, A., Tanaka, J., Takahashi, K., Nakashima, R., Kanda, S., Ogawa, J., Hagisawa, Y. and Fujiwara, T. (2009).Metformin primarily decreases plasma glucose not by gluconeogenesis suppression but by activating glucose utilization in a non-obese type 2 diabetes Goto-Kakizaki rats.*European Journal of Pharmacology* 623:141-147
- Yoshimi, N., Matsunaga, K., Katayama, M., Yamada, Y., Kuno, T., Qiao, Z, Hara, A., Yamahara, J. and Mori, H. (2001). *Cancer Letter* 163:164-170.
- Zhang, W.Y., Lee, J.J., Kim, I.S., Kim, Y. and Myung, C.S. (2011). Stimulation of Glucose Uptake and Improvement of Insulin Resistance by Aromadendrin.*Pharmacology*88:266–274.
- Zhang, Y.B., Xu, X.J. and Liu, H.M. (2006). Chemical constituents from Mahkota dewa. *Journal of Asian Natural Product*, 8, 119-123.
- Zhou, H., Zhang, T., Harmon, J.S., Bryan, J. and Robertson, R.P. (2007). Zinc, not insulin, regulates the rat α-cell response to hypoglycemia in vitro. *Diabetes*.56: 1107-1112.