



UNIVERSITI PUTRA MALAYSIA

***ANTI-ALLODYNIC AND ANTIHYPERALGESIC ACTIVITIES OF
ZERUMBONE IN CHRONIC CONSTRICTION INJURY-INDUCED
NEUROPATHIC PAIN AND ITS POSSIBLE MECHANISM OF ACTION***

NURUL ATIQA BINTI ZULAZMI

FPSK(M) 2016 38



**ANTI-ALLODYNIC AND ANTIHYPERALGESIC ACTIVITIES OF
ZERUMBONE IN CHRONIC CONSTRICTION INJURY-INDUCED
NEUROPATHIC PAIN AND ITS POSSIBLE MECHANISM OF ACTION**

By

NURUL ATIQA BINTI ZULAZMI

**Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirement for the degree of Master of Science**

April 2016

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



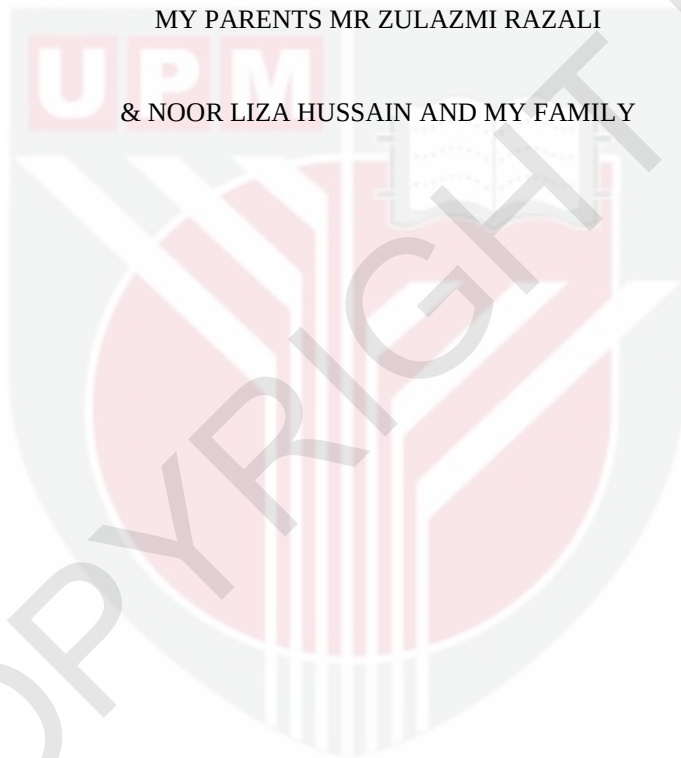
DEDICATION

WITH APPRECIATION AND RESPECT,

THIS THESIS IS DEDICATED

TO

MY PARENTS MR ZULAZMI RAZALI
& NOOR LIZA HUSSAIN AND MY FAMILY



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the Degree of Master of Science

**ANTI-ALLODYNIC AND ANTIHYPERALGESIC ACTIVITIES OF
ZERUMBONE IN CHRONIC CONSTRICTION INJURY-INDUCED
NEUROPATHIC PAIN AND ITS POSSIBLE MECHANISM OF ACTION**

By

NURUL ATIQA BINTI ZULAZMI

April 2016

Chairman : Enoch Kumar Perimal, PhD
Faculty : Medicine and Health Science

The present study was conducted to investigate the potential of zerumbone, a bioactive sesquiterpene of *Zingiber zerumbet* (L) Smith (*Z. zerumbet*) in anti-allodynic and antihyperalgesic properties in neuropathic pain induced by chronic constriction injury (CCI) in mice. We used this model and demonstrated the comparison of different number of sciatic nerve ligation (1, 2, 3 and 4). The outcome revealed that a single ligation CCI animal model on the sciatic nerve is enough to mimic the symptoms of neuropathic pain such as hyperalgesia and allodynia with almost similar effect with the usual 4 ligations sciatic nerve in CCI animal model. Indeed, acute and repeated intraperitoneal (i.p) administration (on 14th day and continued administration once daily for 7 days) of zerumbone (10, 50, 100 mg/kg) significantly exhibited dose-dependent inhibition of chronic constriction injury induced-neuropathic pain in mice, when evaluated using von Frey filament test, cold plate, Randall-Selitto and Hargreaves plantar test ($p < 0.05$). The compound was found to exert anti-allodynic and antihyperalgesic properties in chronic constriction injury (CCI) model and the optimum dose for zerumbone was found out to be 10 mg/kg. The anti-allodynic and antihyperalgesic effect of zerumbone (10 mg/kg i.p) were also significantly reversed by pre-treatment of L-arginine (10 mg/kg), 1H [1,2,4] Oxadiazole [4,3a] quinoxalin-1-one (ODQ), soluble guanosyl cyclase blocker (2 mg/kg i.p) and glibenclamide (ATP-sensitive potassium channel blocker) (10 mg/kg i.p). In addition, the histology of nerve morphology also showed a profound attenuation of mast cells around nerve fibres after treated 7 days daily with 10 mg/kg i.p and 50 mg/kg i.p of zerumbone ($p < 0.05$). Even though, zerumbone failed to produce significant result with vehicle-treated group on the swelling of nerve fibre but the anti-inflammatory effect of zerumbone was able to attenuate the inflammatory cells surrounding the nerve fibre. Together, these findings proved that zerumbone produce pronounced anti-allodynic and antihyperalgesic effects in modified CCI model of neuropathic pain in mice that involves inhibition of L-arginine-NO-Cyclic GMP (cGMP) pathway followed by the opening of ATP-sensitive K^+ channel at the peripheral level. Hence, considering that few effective drugs that are available in the market for the treatment of neuropathic pain, this study indicates that zerumbone a potentially interesting in the development of new clinically relevant drugs for the management of chronic pain.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

**AKTIVITI ANTI-ALODINIK DAN ANTIHIPERALGESIK ZERUMBONE
TERHADAP KECEDEeraan PENYEMPITAN KRONIK YANG
MENYEBABKAN KESAKITAN SARAF DAN MEKANISMA
DISEBALIKNYA.**

Oleh

NURUL ATIQA BINTI ZULAZMI

April 2016

Pengerusi : Enoch Kumar Perimal, PhD
Fakulti : Perubatan dan Sains Kesihatan

Kajian ini dijalankan bagi menunjukkan potensi zerumbone sebagai bahan bioaktif *Zingiber zerumbet* (L.) Smith (*Z. zerumbet*) dalam menghasilkan kesan anti-alodinik dan antihiperalgik terhadap sakit neuropatik yang dipengaruhi oleh model haiwan dengan konstriksi saraf yang kronik (CCI). Beberapa bilangan ikatan (1,2,3 dan 4) pada bahagian saraf siatik telah dihasilkan bagi memilih model CCI yang terbaik dan satu ikatan saraf siatik telah berjaya menghasilkan kesakitan neuropatik yang sama seperti dihasilkan oleh empat bilangan ikatan saraf siatik iaitu ikatan sebenar model haiwan CCI sebelum ini. Penemuan ini diteruskan dengan memberi rawatan zerumbone kepada model CCI yang terpilih. Suntikan akut intra-peritoneal dan berulang kali (pada hari ke-14 dan suntikan berturutan selama 7 hari) zerumbone (10, 50, 100 mg/kg) menghasilkan kesan perencatan bersandar dos terhadap CCI apabila dinilai menggunakan von Frey Ujian Filamen, Plantar Sejuk, Randall-Selitto, dan Ujian Plantar Hargreaves ($p < 0.05$). Zerumbone ternyata memberi kesan anti-alodinik dan antihiperalgik terhadap model CCI. Manakala, dos terbaik zerumbone yang memberikan kesan optimum ialah 10 mg/kg. Kesan anti-alodinik dan antihiperalgik zerumbone (10 mg/kg ip) telah dibalikkan dengan suntikan pra-rawatan L-arginina (10 mg/kg), 1*H* [1, 2, 4] Oxadiazole [4,3*a*] quinoxalin-1-one (ODQ) (penyekat Guanosil Siklase) (2 mg/kg ip) dan Glibenclamide (penyekat saluran kalium) (10 mg/kg ip). Manakala, data terhadap histologi morfologi saraf juga menunjukkan pengurangan bilangan sel mast di gentian saraf akibat CCI apabila dirawat selama 7 hari dengan 10 mg/kg ip dan 50 mg/kg ip zerumbone ($p < 0.05$). Walaubagaimanapun, zerumbone gagal membaiki struktur saraf tetapi kesan anti-radang zerumbone telah berjaya menghapuskan sel-sel radang sekitar gentian saraf. Kesimpulannya, penemuan ini telah membuktikan bahawa zerumbone menghasilkan kesan anti-alodinik dan antihiperalgik yang ketara dalam model CCI yang menyebabkan kesakitan neuropatik melalui penglibatan laluan L-arginina-NO-Cyclic GMP (cGMP) yang diikuti oleh pembukaan saluran ATP sensitif K^+ pada peringkat periferi. Oleh itu, memandangkan hanya beberapa ubat berkesan yang terdapat di pasaran bagi merawat kesakitan neuropatik, kajian aktiviti zerumbone telah membuktikan bahawa zerumbone mungkin berpotensi dalam pembangunan ubat-ubatan klinikal yang baru untuk pengurusan sakit kronik ini.

ACKNOWLEDGEMENTS

Assalamualaikum w.b.t

Alhamdulillah, all praise to Allah, for all His graciousness and blessings that have made it possible for me to finish up my dissertation. All my success is due to HIS Mercy.

I wish to express my profound gratitude to my supervisor, Dr. Enoch Kumar Perimal for being a good mentor to me. I have been extremely fortunate to have a supervisor who gave me the freedom to explore on my own and at the same time gave guidance to recover when my steps faltered. Without his exemplary guidance, monitoring and constant encouragement, throughout my years of study here, this dissertation would not have been possible.

I would like to thank my supervisory committee members, Prof Mohd. Roslan Sulaiman, and Dr Ahmad Akira Omar Farouk for their insightful comments and constructive criticisms at different stages of my research that helped me a lot to focus on my ideas for the research project. Besides, I would also take this opportunity to express my sincere gratitude to Banulata Gopalsamy and Voon Fui Ling whom have helped me to stay sane through these difficult years. Their support and care helped me overcome setbacks and stay focused throughout my graduate study. I greatly value their friendship.

I would like to express my deepest sense of gratitude for the help and guidance that I received from the staff of Physiology lab, Pharmacology lab, Histopathology lab and Anatomy lab. They helped me a lot to handle the machines there and provided me with an excellent atmosphere for doing research. A bunch of thanks is also dedicated to my fellow labmates for the stimulating discussions, sharing ideas, working together to clean up the lab and all the fun we have had during these years. In particular, I am also grateful to my supervisor during my final year project who enlightened me on the first glance of research.

Most importantly, none of this would have been possible without the love and patience from my family. My parents Zulazmi Razali and Noor Liza Hussain have been a constant source of love, concern, support and strength for all these years.

My sincere thanks is also dedicated to the authority of Medical and Health Sciences Faculty UPM for providing me with a good environment and facilities to complete this project. Finally, thanks to my friends and families for their help and wishes for the successful completion of this project.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Enoch Kumar Perimal, PhD

Senior Lecturer,
Faculty of Medical and Health Sciences
Universiti Putra Malaysia
(Chairman)

Mohd Roslan Sulaiman, PhD

Professor,
Faculty of Medical and Health Sciences
Universiti Putra Malaysia
(Member)

Ahmad Akira Omar Ahmad Farouk, PhD

Senior Lecturer,
Faculty of Medical and Health Sciences
Universiti Putra Malaysia
(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean
School of Graduates Studies,
Universiti Putra Malaysia.

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- There is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Nurul Atiqah Binti Zulazmi, GS37408

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____

Name of Chairman
of Supervisory
Committee:

Dr. Enoch Kumar Perimal

Signature: _____

Name of Member
of Supervisory
Committee:

Professor Dr. Mohd Roslan Sulaiman

Signature: _____

Name of Member
of Supervisory
Committee:

Dr. Ahmad Akira Omar Ahmad Farouk

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	ii
ACKNOWLEDGEMENTS	iii
APPROVAL	iv
DECLARATION	vi
LIST OF TABLES	xi
LIST OF FIGURES	xii
LIST OF ABBREVIATIONS	xiv
 CHAPTER	
 1 INTRODUCTION	 1
1.1 Neuropathic pain	1
1.2 Current Research Interest	2
1.3 Objectives	2
1.4 Hypothesis	3
 2 LITERATURE REVIEW	 4
2.1 <i>Zingiber zerumbet</i> sp.	4
2.1.1 Background of <i>Z.zerumbet</i>	4
2.1.2 Application of <i>Z.zerumbet</i>	4
2.2 Zerumbone	5
2.2.1 Potential of Zerumbone	5
2.3 Pain	6
2.3.1 Phases of pain	6
2.3.2 Types of pain	7
2.3.3 Chronic pain	7
2.4 Neuropathic pain	7
2.4.1 Definition of Neuropathic pain	8
2.4.2 Classification of Neuropathic pain	9
2.4.3 Peripheral Mechanism of Neuropathic pain	11
2.4.4 Central Mechanism of Neuropathic pain	11
2.4.5 Pharmacologic treatment	14
2.4.6 Nitric Oxide in Neuropathic pain	15
 3 EFFECTS OF DIFFERENT NUMBER OF SCIATIC NERVE LIGATION ON CCI-INDUCED NEUROPATHIC PAIN IN MICE	 19
3.1 Introduction	19
3.2 Methodology	19
3.2.1 Experimental animals	19
3.2.2 Motor coordinations	20
3.2.3 Surgical procedure	20
3.2.4 Groups and Timeline	20

	3.2.5 Assesment on allodynia	24
	3.2.6 Assesment on hyperalgesia	24
	3.2.7 Statistical Analysis	25
	3.3 Results	25
	3.3.1 Response towards allodynia	25
	3.3.2 Response towards hyperalgesia	29
	3.4 Discussion	32
	3.5 Conclusion	33
4	THE ANTI-ALLODYNIC AND ANTIHYPERALGESIC EFFECTS IN DIFFERENT DOSAGES OF ZERUMBONE TREATMENT ON THE CCI-INDUCED NEUROPATHIC PAIN IN MICE	34
	4.1 Introduction	34
	4.2 Methodology	34
	4.2.1 Experimental animals	34
	4.2.2 Motor coordinations	34
	4.2.3 Preparation of zerumbone from plant material	35
	4.2.3.1 Extraction of plant material	35
	4.2.3.2 Preparation of zerumbone for experiment	35
	4.2.4 Preparation of drugs and chemicals	36
	4.2.5 Surgical procedure	36
	4.2.6 Groups and Timeline	36
	4.2.7 Assesment on allodynia effect	38
	4.2.8 Assesment on hyperalgesia effect	38
	4.2.9 Statistical Analysis	38
	4.3 Results	38
	4.3.1 Anti-allodynic effect	38
	4.3.2 Anti-hyperalgesic effect	42
	4.3.4 Effective dose (ED50)	45
	4.4 Discussion	48
	4.5 Conclusion	51
5	THE INVOLVEMENT OF L-ARGININE-NITRIC OXIDE-cGMP-K⁺ ATP PATHWAY IN ZERUMBONE ANTI-ALLODYNIC AND ANTIHYPERALGESIC ACTIVITIES	52
	5.1 Introduction	52
	5.2 Methodology	52
	5.2.1 Experimental animals	52
	5.2.2 Motor coordinations	52
	5.2.3 Preparation of zerumbone,drugs and chemicals	53
	5.2.4 Surgical procedure	53
	5.2.5 Groups and Timeline	53

5.2.6	Allodynia effect	56
5.2.7	Hyperalgesia effect	56
5.2.8	Statistical Analysis	56
5.3	Results	56
5.3.1	L-arginine-Nitric oxide pathway	56
5.3.2	Cyclic Guanosine Monophosphate pathway	58
5.3.3	Potassium channel pathway	60
5.4	Discussion	62
5.5	Conclusion	64
6	THE EFFECTS OF ZERUMBONE TREATMENT ON THE HISTOLOGY OF SCIATIC NERVE	65
6.1	Introduction	65
6.2	Methodology	65
6.2.1	Experimental animals	65
6.2.2	Motor coordinations	65
6.2.3	Preparation of zerumbone, drugs and chemicals	66
6.2.4	Surgical procedure	66
6.2.5	Groups and Timeline	66
6.2.6	Nerve isolation	68
6.2.7	Tissue processing and H&E staining	68
6.2.8	Inflammatory cell counts	68
6.2.9	Sciatic nerve fibre derangement	68
6.2.10	Statistical analysis	69
6.3	Results	69
6.3.1	Nerve morphology	69
6.3.2	Neutrophils and Mast cells counts	73
6.3.3	Scoring for Nerve derangement	75
6.4	Discussion	76
6.5	Conclusion	76
7	GENERAL DISCUSSIONS AND CONCLUSIONS AND RECOMMENDATION FOR FUTURE RESEARCH	79
	REFERENCES	82
	APPENDICES	104
	BIODATA OF STUDENT	111
	LIST OF PUBLICATIONS	112

LIST OF TABLES

Table		Page
3-1	Experimental Groups for effects on different number of sciatic nerve ligation.	21
4-1	Experimental Groups For Anti-Allodynic And Antihyperalgesic Effect Of Zerumbone Study.	36
5-1	Experimental Groups for the involvement of L-arginine-Nitric Oxide-cGMP-K ⁺ ATP pathway in zerumbone anti-allodynic and antihyperalgesic activities	54
6-1	Experimental Groups for the zerumbone effects on the sciatic nerve histology study.	66
6-2	Scoring For Sciatic Nerve fibre derangement.	69
A1	Pain threshold for tactile allodynia (mean $\pm$ se) in different groups at respective days of observation	105
A2	Pain threshold for cold allodynia (mean $\pm$ se) in different groups at respective days of observation.	105
A3	Pain threshold for mechanical hyperalgesia (mean $\pm$ se) in different groups at respective days of observation	106
A4	Pain threshold for thermal hyperalgesia (mean $\pm$ se) in different groups at respective days of observation.	106
A5	The response of zerumbone administered intra-peritoneally on tactile allodynia inhibition (mean $\pm$ se) of CCI-induced mice model.	107
A6	The response of zerumbone administered intra-peritoneally on cold allodynia inhibition (mean $\pm$ se) of CCI-induced mice model	107
A7	The response of zerumbone administered intra-peritoneally on mechanical hyperalgesia inhibition (mean $\pm$ se) of CCI-induced mice model	108
A8	The response of zerumbone administered intra-peritoneally on thermal hyperalgesia inhibition (mean $\pm$ se) of CCI-induced mice model	108

LIST OF FIGURES

Figure		Page
2-1	Factors that contribute to the patterns of activity generated by the body self neuromatrix.	10
2-2	Peripheral sensitization after nerve injury showed degenerating sensory axon because of the ligation.	13
2-3	NO and cGMP Pathway (Schmidtke <i>et al.</i> , 2009).	18
3-1	Ligation on the sciatic nerve procedure.	22
3-2	Timeline experiment for effects on different number of sciatic nerve ligation.	23
3-3	Time-course of tactile allodynia for four different ligation CCI group with sham on ipsilateral paw.	27
3-4	Time-course of tactile allodynia for four different ligation CCI group with sham on the contralateral paw.	27
3-5	Time-course of cold allodynia for four different ligation CCI group with sham.	28
3-6	Time-course of mechanical hyperalgesia for four different ligation CCI group with sham on the ipsilateral paw.	30
3-7	Time-course of mechanical hyperalgesia for four different ligation CCI group with sham on the ipsilateral paw.	30
3-8	Time-course of thermal hyperalgesia for four different ligation cci group with sham on the contralateral paw.	31
3-9	Time-course of thermal hyperalgesia for four different ligation CCI group with sham on the contralateral paw.	31
4-1	Timeline for the response of zerumbone towards allodynia and hyperalgesia exhibited by CCI-induced mice model.	37
4-2	Response of Zerumbone on CCI-induced neuropathic pain in mice on day 14 and day 21 von Frey assessment on the ipsilateral paw.	40
4-3	Response of Zerumbone on CCI-induced neuropathic pain in mice on day 14 and day 21 von Frey assessment on the contralateral paw.	40
4-4	Response of Zerumbone on CCI-induced neuropathic pain in mice on day 14 and day 21 Cold plate assessment by scoring method.	41
4-5	Response of Zerumbone on CCI-induced neuropathic pain in mice on day 14 and day 21 Randall Selitto assessment on the ipsilateral paw.	43
4-6	Response of Zerumbone on CCI-induced neuropathic pain in mice on day 14 and day 21 Hargreaves Plantar Test assessment on the contralateral paw.	43
4-7	Response of Zerumbone on CCI-induced neuropathic pain in mice on day 14 and day 21 Randall Selitto assessment on the ipsilateral paw.	44
4-8	Response of Zerumbone on CCI-induced neuropathic pain in mice on day 14 and day 21 Hargreaves Plantar test assessment on the contralateral paw.	44
4-9	The dose-response curve of Zerumbone on tactile allodynia	46

	test in CCI-induced mice.	
4-10	The dose-response curve of Zerumbone on cold allodynia test in CCI-induced mice.	46
4-11	The dose-response curve of Zerumbone on mechanical hyperalgesia test in CCI-induced mice.	47
4-12	The dose-response curve of Zerumbone on thermal hyperalgesia test in CCI-induced mice.	47
5-1	Timeline for the Involvement Of L-Arginine-Nitric Oxide-cGMP-K ⁺ ATP Pathway in Zerumbone anti-allodynic and antihyperalgesic activities.	55
5-2	Effect of pre-treatment of mice with L-Arginine (10 mg /kg, i.p.) and L-NOARG (10 mg /kg, i.p.) on the anti-allodynic effect of zerumbone (10 mg / kg, i.p.) in CCI-induced neuropathic pain.	57
5-3	Effect of pre-treatment of mice with Glibenclamide (10 mg/kg i.p) on the antihyperalgesic effect of zerumbone (10 mg /kg, i.p) In CCI-induced neuropathic pain.	57
5-4	Effect of pre-treatment of mice with ODQ (2 mg/kg i.p) on the anti-allodynic effect of zerumbone (10 mg / kg, i.p.) in CCI-induced neuropathic pain.	59
5-5	Effect of pre-treatment of mice with ODQ (2 mg/kg i.p) on the antihyperalgesic effect of zerumbone (10 mg / kg, i.p.) in CCI-induced neuropathic pain.	59
5-6	Effect of pre-treatment of mice with Glibenclamide (10 mg/kg i.p) on the anti-allodynic effect of zerumbone (10 mg / kg, i.p.) in CCI-induced neuropathic pain.	61
5-7	Effect of pre-treatment of mice with Glibenclamide (10 mg/kg i.p) on the antihyperalgesic effect of zerumbone (10 mg / kg, i.p.) in CCI-induced neuropathic pain.	61
6-1	Timeline for the effects of zerumbone on the sciatic nerve histology study.	67
6-2	Longitudinal sections with H&E staining; vehicle group.	70
6-3	Longitudinal sections with H&E staining; sham group.	70
6-4	Longitudinal sections with H&E Staining; morphine group.	71
6-5	Longitudinal sections with H&E staining; gabapentin group.	71
6-6	Longitudinal sections with H&E staining; zerumbone 50 mg/Kg Group.	72
6-7	Longitudinal sections with H&E staining; zerumbone 10 mg/Kg group.	72
6-8	The number of neutrophils measured between groups.	74
6-9	The number of mast cells measured between groups.	74
6-10	Score of nerve morphology based on the score given in each sample.	75
7-1	Graphical Summary of the research project.	81
A1	The rhizomes of <i>Zingiber zerumbet</i> smith where the zerumbone is isolated (Yob <i>et al.</i> , 2011)	104
A2	Animal Ethics approved by IACUC UPM.	109
A3	Published article from Fitoterapia for the research project.	110

LIST OF ABBREVIATIONS

IASP	International Association Study of Pain
nNOS	Neuronal nitric-oxide synthases
iNOS	Inducible nitric-oxide synthases
cGMP	central guanosine 3,5 -cyclic monophosphate
PKG	protein kinase G
sGC	soluble guanylate cyclase
K ⁺ ATP	ATP-sensitive potassium
CCI	Chronic Constriction Injury
RZZ	<i>Zingiber zerumbet</i>
HTC	Hepatoma Tissue Culture
PNS	peripheral nervous system
CNS	central nervous system
NGF	nerve growth factor
NMDA	N-methyl-D-aspartate
AMPA	α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid
IUPAC	International Union of Pure and Applied Chemistry
GABA	Gamma-aminobutyric acid
NO	Nitric Oxide
NOS	Nitric Oxide Synthase
PKC	Protein Kinase C
DMSO	dimethylsulfoxide
ANOVA	one way analysis variance
L-NOARG	L-NG-nitro arginine
L-arginine	L-arginine hydrochloride
ODQ	Oxadiazole [4, 3-a] quinoxaline-1-one
NaCl	Sodium Chloride
H&E	Hematoxylin and Eosin
MCP-1	monocyte chemoattractant protein-1

CHAPTER 1

INTRODUCTION

1.1 Neuropathic pain

Neuropathic pain has become a common issue that affected millions of people around the world (Tsuda *et al.*, 2005). Apart from that, The Ministry of Health Malaysia in the Quick Reference Guide on Management of Cancer Pain, 2010 classified neuropathic pain as a difficult type of pain syndrome that is critically in need of adjuvant analgesics and additional interventions. International Association Study of Pain (IASP) defined neuropathic pain as a pain that is caused by a lesion or disease on the somatosensory system. Nevertheless, this type of pain is often associated with disease or injury to the peripheral and central nervous system that usually causes abnormal processing of sensory input (Jensen *et al.*, 2001; Dworkin *et al.*, 2003).

Neuropathic pain is categorized as a pathophysiologic pain which is dissimilar from physiologic pain in many ways and this includes the mechanism that is underlying it (Iyengar *et al.*, 2004; Pasero, 2004). Additionally, neuropathic pain is described as constant pain arising from the changes that originate in pain sensory system after peripheral nerve injury (Iyengar *et al.*, 2004). Moreover, this pain can be indicated by the sensory impairment such as unpleasant abnormal sensation (dyesthesia), increase response to a normal painful stimulus (hyperalgesia) and pain that is caused by normal non-painful stimulus (allodynia) (Jaggi *et al.*, 2011). While other related symptoms include the altered pain sensation, numbness, burning and continuous or intermittent evoked or spontaneous pain (Ueda and Rashid, 2003).

However, existing treatment such as non-steroidal anti-inflammatory drugs (NSAIDs) and opiates showed little success with a limited response in neuropathic pain (Dowdall *et al.*, 2005). Plus, the commonly used medications for neuropathic pain also cause numerous side effects, unpredictable effectiveness, complex doses, delayed analgesic onset and somehow can reduce the patient's quality of life (Meyer-Rosberg *et al.*, 2001; Dworkin *et al.*, 2007). Although, the mechanisms of neuropathic pain are not yet fully understood, substances that have potential inhibitory effect on this type of pain with minimal or no adverse effect are of analgesic research interest.

1.2 Current Research Interest

Complications and problems that arise perpetuate a clear of unmet therapeutic needs and a high demand for the field of drug discovery (Gutierrez *et al.*, 2012). In conjunction with that, herbal medicines and alternative medicine has been used by the general population today because of dissatisfaction on ineffective conventional treatment and their adverse effects (Astin, 1998). *Zingiber zerumbet* is considered as a large genus of herbs that belongs to the family of Zingiberaceae. This species consist

about 141 species that is scattered mainly in Asia and confined to the tropics in Asia, Malaysia and Pacific Islands (Burkill, 1966; Jantan *et al.*, 2003). The name 'lempoyang' were given by Malaysians and Indonesians and this plant has also been traditionally used as folkloric medicine (Habsah *et al.*, 2000).

Zerumbone is a bioactive sesquiterpene that is found to be one of the major components in the rhizome of *Zingiber zerumbet* Smith (Sulaiman *et al.*, 2010b) that has been studied extensively in in-vivo and in-vitro studies (Yob *et al.*, 2011). Surprisingly, there are some medicinal properties that has been previously reported for zerumbone such as antinociceptive (Perimal *et al.*, 2011), anti-inflammatory (Sulaiman *et al.*, 2010a) and antitumor activities (Murakami *et al.*, 2004; Huang *et al.*, 2005). Even though, several researchers portrayed some information about the capabilities of zerumbone with pathways underlying physiological pain, but still no research has discussed its effects on neuropathic pain and the mechanism behind it.

One of the important pathways associated with neuropathic pain mechanism is the L-arginine-nitric oxide-cGMP-K⁺ATP pathway. Nitric oxide is synthesized by neuronal (nNOS) or inducible (iNOS) nitric-oxide synthases via central guanosine 3,5 -cyclic monophosphate (cGMP)-protein kinase G (PKG) pathway activation that mediates numerous neuropathic pain symptoms (Meller *et al.*, 1992). Ionic channel modulation by NO can be produced either indirectly through the classical pathway of soluble guanylate cyclase (sGC) activation and generation of cGMP, or directly through a pathway involving S-nitrosylation of target proteins (Ahern *et al.*, 2002). Nonetheless, pharmacological studies in vivo imply that K⁺ATP channels in peripheral sensory neurons may be activated indirectly via the NO/cGMP/PKG pathway (Soares and Duarte, 2001; Alves *et al.*, 2004a; Sachs *et al.*, 2004). Some drugs produce peripheral analgesia via NO-dependent activation of ATP-sensitive potassium (K⁺ATP) channels (Rodrigues and Duarte, 2000; Lázaro-Ibáñez *et al.*, 2001; Granados-Soto *et al.*, 2002; Alves *et al.*, 2004b). Therefore, this study attempted to illustrate the effects of zerumbone towards neuropathic pain on the neuropathic animal model and provide better understanding of some mechanism of action underlying it.

1.3 Objectives

General objective:

This research was conducted to investigate the anti-allodynic and antihyperalgesic effects of zerumbone in an animal model of neuropathic pain.

Specific objectives:

- i) To investigate and compare the pain developed by number of different sciatic nerve ligations in Chronic Constriction Injury (CCI) animal model of neuropathic pain as a pilot study.

- ii) To evaluate the anti-allodynic and antihyperalgesic effects of different doses of zerumbone in tactile, thermal and mechanical test in animal model of neuropathic pain.
- iii) To elucidate the role of L-Arginine-NO-cGMP-K⁺ATP channel pathway in the zerumbone analgesic activities in vivo.
- iv) To assess the effect of zerumbone treatment on the sciatic nerve morphology.

1.4 Hypothesis

Zerumbone significantly inhibits the allodynia and hyperalgesia caused by CCI animal model of neuropathic pain.

REFERENCES

- Abdul, A. B. H., Al-Zubairi, A. S., Tailan, N. D., Wahab, S. I. A., Zain, Z. N. M., Ruslay, S. & Syam, M. M. (2008). Anticancer Activity of Natural Compound (Zerumbone) Extracted from *Zingiber zerumbet* in Human HeLa Cervical Cancer Cells. *International Journal of Pharmacology*, 4(3), 160-168.
- Ahern, G. P., Klyachko, V. A. & Jackson, M. B. (2002). cGMP and S-nitrosylation: two routes for modulation of neuronal excitability by NO. *Trends in Neurosciences*, 25(10), 510-517.
- Altman, R. D. & Marcussen, K. C. (2001). Effects of a ginger extract on knee pain in patients with osteoarthritis. *Arthritis and Rheumatism*, 44(11), 2531-2538.
- Alves, D. P., Soares, A. C., Francischi, J. N., Castro, M. S. A., Perez, A. C. & Duarte, I. D. G. (2004a). Additive antinociceptive effect of the combination of diazoxide, an activator of ATP-sensitive K⁺ channels, and sodium nitroprusside and dibutyl-yl-cGMP. *European Journal of Pharmacology*, 489(1-2), 59-65.
- Alves, D. P., Tatsuo, M. A. F., Leite, R. & Duarte, I. D. G. (2004b). Diclofenac-induced peripheral antinociception is associated with ATP-sensitive K⁺ channels activation. *Life Sciences*, 74(20), 2577-2591.
- Andersson, D. A., Gentry, C., Moss, S. & Bevan, S. (2008). Transient Receptor Potential A1 Is a Sensory Receptor for Multiple Products of Oxidative Stress. *The Journal of Neuroscience: the official journal of the Society for Neuroscience*, 28(10), 2485-2494.
- Appendino, G., Minassi, A., Collado, J. A., Pollastro, F., Chianese, G., Tagliatela-Scafati, O., Ayyari, M., Garcia, V. & Muñoz, E. (2015). The Thia-Michael Reactivity of Zerumbone and Related Cross-Conjugated Dienones: Disentangling Stoichiometry, Regiochemistry, and Addition Mode with an NMR-Spectroscopy-Based Cysteamine Assay. *European Journal of Organic Chemistry*, 2015(17), 3721-3726.
- Astin, J. A. (1998). Why patients use alternative medicine: Results of a national study. *The journal of the American Medical Association*, 279(19), 1548-1553.
- Attal, N. (2000). Chronic Neuropathic Pain: Mechanisms and Treatment. *The Clinical Journal of Pain*, 16(3), 118-130.
- Attal, N., Cruccu, G., Baron, R., Haanpää, M., Hansson, P., Jensen, T. & Nurmikko, T. (2010). EFNS guidelines on the pharmacological treatment of neuropathic pain: 2010 revision. *European Journal of Neurology*, 17(9), 1113-1188
- Attal, N., Jazat, F., Kayser, V. & Guilbaud, G. (1990). Further evidence for 'pain-related' behaviours in a model of unilateral peripheral mononeuropathy. *Pain*, 41(2), 235-251.

- Baby, S., Dan, M., Thaha, A. R. M., Johnson, A. J., Kurup, R., Balakrishnapillai, P. & Lim, C. K. (2009). High content of zerumbone in volatile oils of *Zingiber zerumbet* from southern India and Malaysia. *Flavour and Fragrance Journal*, 24(6), 301-308.
- Backonja, M.M., Irving, G. & Argoff, C. (2006). Rational multidrug therapy in the treatment of neuropathic pain. *Current Pain and Headache Reports*, 10(1), 34-38.
- Bandell, M., Story, G. M., Hwang, S. W., Viswanath, V., Eid, S. R., Petrus, M. J., Earley, T. J. & Patapoutian, A. (2004). Noxious cold ion channel TRPA1 is activated by pungent compounds and bradykinin. *Neuron*, 41(6), 849-857.
- Baraldi, P. G., Preti, D., Materazzi, S. & Geppetti, P. (2010). Transient Receptor Potential Ankyrin 1 (TRPA1) Channel as Emerging Target for Novel Analgesics and Anti-Inflammatory Agents. *Journal of Medicinal Chemistry*, 53(14), 5085-5107.
- Baron, R. (2000). Peripheral Neuropathic Pain: From Mechanisms to Symptoms. *The Clinical Journal of Pain*, 16(2), 12-20.
- Baron, R. (2006). Mechanisms of Disease: neuropathic pain-a clinical perspective. *Nature Clinical Practice Neurology*, 2(2), 95-106.
- Baron, R., Binder, A. & Wasner, G. (2010). Neuropathic pain: diagnosis, pathophysiological mechanisms, and treatment. *The Lancet Neurology*, 9(8), 807-819.
- Beckman, J. S., Beckman, T. W., Chen, J., Marshall, P. A. & Freeman, B. A. (1990). Apparent hydroxyl radical production by peroxynitrite: implications for endothelial injury from nitric oxide and superoxide. *Proceedings of the National Academy of Sciences*, 87(4), 1620-1624.
- Bennett, G. J. & Xie, Y. K. (1988). A peripheral mononeuropathy in rat that produces disorders of pain sensation like those seen in man. *Pain*, 33(1), 87-107.
- Bennett, M. I., Smith, B. H., Torrance, N. & Lee, A. J. (2006). Can pain can be more or less neuropathic? Comparison of symptom assessment tools with ratings of certainty by clinicians. *Pain*, 122(3), 289-294.
- Besson, J. M. (1999). The neurobiology of pain. *The Lancet*, 353(9164), 1610-1615.
- Beydoun, A. & Backonja, M.M. (2003). Mechanistic Stratification of Antineuralgic Agents. *Journal of Pain and Symptom Management*, 25(5), 18-30.
- Bhuiyan, M. N. I., Chowdhury, J. U. & Begum, J. (2008). Chemical investigation of the leaf and rhizome essential oils of *Zingiber zerumbet* (L.) Smith from Bangladesh. *Bangladesh Journal of Pharmacology*, 4(1), 9-12.
- Binshtok, A. M., Wang, H., Zimmermann, K., Amaya, F., Vardeh, D., Shi, L., Brenner, G. J., Ji, R.R., Bean, B. P., Woolf, C. J. & Samad, T. A. (2008). Nociceptors Are Interleukin-1 β Sensors. *The Journal of Neuroscience*, 28(52), 14062-14073.

- Bolotina, V. M., Najibi, S., Palacino, J. J., Pagano, P. J. & Cohen, R. A. (1994). Nitric oxide directly activates calcium-dependent potassium channels in vascular smooth muscle. *Nature*, 368(6474), 850-853.
- Bordia, A., Verma, S. K. & Srivastava, K. C. (1997). Effect of ginger (*Zingiber officinale* Rosc.) and fenugreek (*Trigonella foenumgraecum* L.) on blood lipids, blood sugar and platelet aggregation in patients with coronary artery disease. *Prostaglandins, Leukotrienes and Essential Fatty Acids*, 56(5), 379-384.
- Böger, R. H., & Bode-Böger, S. M. (2001). The clinical pharmacology of L-arginine. *Annual review of pharmacology and toxicology*, 41(1), 79-99.
- Bridges, D., Thompson, S. W. N. & Rice, A. S. C. (2001). Mechanisms of neuropathic pain. *British Journal of Anaesthesia*, 87(1), 12-26.
- Brück, W. (1997). The Role of Macrophages in Wallerian Degeneration. *Brain Pathology*, 7(2), 741-752.
- Burkill, I. H. (1966). A dictionary of the economic products of the Malay Peninsula. A Dictionary of the Economic Products of the Malay Peninsula., 2(2nd edition), 2444.
- Bujalska, M. & Gumulka, S. (2008). Effect of cyclooxygenase and nitric oxide synthase inhibitors on vincristine induced hyperalgesia in rats. *Pharmacology Reports*, 60(5), 735-741.
- Callsen-Cencic, P., Hoheisel, U., Kaske, A., Mense, S. & Tenschert, S. (1999). The controversy about spinal neuronal nitric oxide synthase: under which conditions is it up- or downregulated? *Cell and Tissue Research*, 295(2), 183-194.
- Campbell, J. N. & Meyer, R. A. (2006). Mechanisms of Neuropathic Pain. *Neuron*, 52(1), 77-92.
- Cao, H. & Zhang, Y.Q. (2008). Spinal glial activation contributes to pathological pain states. *Neuroscience and Biobehavioral Reviews*, 32(5), 972-983.
- Carrier, G. O., Fuchs, L. C., Winecoff, A. P., Giulumian, A. D. & White, R. E. (1997). Nitrovasodilators relax mesenteric microvessels by cGMP-induced stimulation of Ca-activated K channels. *American Journal of Physiology Heart and Circulatory Physiology*, 273(1), 76-84.
- Casals-Díaz, L., Vivó, M. & Navarro, X. (2009). Nociceptive responses and spinal plastic changes of afferent C-fibers in three neuropathic pain models induced by sciatic nerve injury in the rat. *Experimental Neurology*, 217(1), 84-95.
- Chabrier, P. E., Demerlé-Pallardy, C. & Auguet, M. (1999). Nitric oxide synthases: targets for therapeutic strategies in neurological diseases. *Cellular and Molecular Life Sciences*, 55(8-9), 1029-1035.

- Chacur, M., Matos, R. J. B., Alves, A. S., Rodrigues, A. C., Gutierrez, V., Cury, Y. & Britto, L. R. G. (2010). Participation of neuronal nitric oxide synthase in experimental neuropathic pain induced by sciatic nerve transection. *Brazilian Journal of Medical and Biological Research*, 43, 367-376.
- Chen, B.Y., Lin, D. P.C., Wu, C.Y., Teng, M.C., Sun, C.Y., Tsai, Y.T., Su, K.C., Wang, S.R. & Chang, H.H. (2011). Dietary zerumbone prevents mouse cornea from UVB-induced photokeratitis through inhibition of NF- κ B, iNOS, and TNF- α expression and reduction of MDA accumulation. *Molecular Vision*, 17, 854-863.
- Chen, I. N., Chang, C.C., Ng, C.C., Wang, C.Y., Shyu, Y.T. & Chang, T.L. (2008). Antioxidant and Antimicrobial Activity of Zingiberaceae Plants in Taiwan. *Plant Foods for Human Nutrition*, 63(1), 15-20.
- Choi, S., Choi, H. J., Cheong, Y., Lim, Y. J., Park, H. K. (2013). Internal-Specific Morphological Analysis of Sciatic Nerve Fibres in a Radiofrequency-Induced Animal Neuropathic Pain Model. *PLoS ONE* 8(9) 1-11.
- Chu, Y.C., Guan, Y., Skinner, J., Raja, S. N., Johns, R. A. & Tao, Y.X. (2005). Effect of genetic knockout or pharmacologic inhibition of neuronal nitric oxide synthase on complete Freund's adjuvant-induced persistent pain. *Pain*, 119(1-3), 113-123.
- Cirino, G., Fiorucci, S. & Sessa, W. C. (2003). Endothelial nitric oxide synthase: the Cinderella of inflammation? *Trends in Pharmacological Sciences*, 24(2), 91-95.
- Costigan, M., Scholz, J. & Woolf, C. J. (2009). Neuropathic Pain: A Maladaptive Response of the Nervous System to Damage. *Annual Review of Neuroscience*, 32, 1-32.
- Čížková, D., Lukáčová, N., Maršala, M. & Maršala, J. (2002). Neuropathic pain is associated with alterations of nitric oxide synthase immunoreactivity and catalytic activity in dorsal root ganglia and spinal dorsal horn. *Brain Research Bulletin*, 58(2), 161-171.
- Cunha, T. M., Verri Jr., W. A., Vivancos, G. G., Moreira, I. F., Reis, S., Parada, C. A., Cunha, F. Q. & Ferreira, S. H. (2004). An electronic pressure-meter nociception paw test for mice. *Brazilian Journal of Medical and Biological Research*, 37, 401-407.
- Cury, Y., Picolo, G., Gutierrez, V. P. & Ferreira, S. H. (2011). Pain and analgesia: The dual effect of nitric oxide in the nociceptive system. *Nitric Oxide*, 25(3), 243-254.
- De Alba, J., Clayton, N. M., Collins, S. D., Colthup, P., Chessell, I. & Knowles, R. G. (2006). GW274150, a novel and highly selective inhibitor of the inducible isoform of nitric oxide synthase (iNOS), shows analgesic effects in rat models of inflammatory and neuropathic pain. *Pain*, 120(1-2), 170-181.
- De Vry, J., Kuhl, E., Franken-Kunkel, P. & Eckel, G. (2004). Pharmacological characterization of the chronic constriction injury model of neuropathic pain. *European Journal of Pharmacology*, 491(2-3), 137-148.

- Déciga-Campos, M. & López-Muñoz, F. J. (2004). Participation of the L-arginine–nitric oxide–cyclic GMP–ATP-sensitive K⁺ channel cascade in the antinociceptive effect of rofecoxib. *European Journal of Pharmacology*, 484(2–3), 193-199.
- Derbyshire, E. & Martin, D. (2004). Neutropenia occurring after starting gabapentin for neuropathic pain. *Clinical Oncology*, 16(8), 575-576.
- Dev, S. (1960). Studies in sesquiterpenes—XVI: Zerumbone, a monocyclic sesquiterpene ketone. *Tetrahedron*, 8(3–4), 171-180.
- Dev, S., Anderson, J. E., Cormier, V., Damodaran, N. P. & Roberts, J. D. (1968). Nuclear magnetic resonance spectroscopy. The conformational mobility of humulene and zerumbone. *Journal of the American Chemical Society*, 90(5), 1246-1248.
- Devor, M. (2006). Sodium Channels and Mechanisms of Neuropathic Pain. *The Journal of Pain*, 7(1), 3-12.
- Dias, Q. M., Rossaneis, A. C., Fais, R. S. & Prado, W. A. (2013). An improved experimental model for peripheral neuropathy in rats. *Brazilian Journal of Medical and Biological Research*, 46, 253-256.
- Dowdall, T., Robinson, I. & Meert, T. F. (2005). Comparison of five different rat models of peripheral nerve injury. *Pharmacology Biochemistry and Behavior*, 80(1), 93-108.
- Dubový, P. (2011). Wallerian degeneration and peripheral nerve conditions for both axonal regeneration and neuropathic pain induction. *Annals of Anatomy - Anatomischer Anzeiger*, 193(4), 267-275.
- Ducreux, D., Attal, N., Parker, F. & Bouhassira, D. (2006). Mechanisms of central neuropathic pain: a combined psychophysical and fMRI study in syringomyelia. *Brain*, 129(4), 963-976.
- Duncan, G. H., Catherine Bushnell, M. & Lavigne, G. J. (1989). Comparison of verbal and visual analogue scales for measuring the intensity and unpleasantness of experimental pain. *Pain*, 37(3), 295-303.
- Duñg, N. X., Chính, T. D., Rañg, D. D. & Leclercq, P. A. (1993). The Constituents of the Rhizome Oil of *Zingiber zerumbet* (L.) Sm. from Vietnam. *Journal of Essential Oil Research*, 5(5), 553-555.
- Dunham, N. W. & Miya, T. S. (1957). A note on a simple apparatus for detecting neurological deficit in rats and mice. *Journal of the American Pharmaceutical Association*, 46(3), 208-209.
- Durrenberger, P. F., Facer, P., Casula, M. A., Yiangou, Y., Gray, R. A., Chessell, I. P., Day, N. C., Collins, S. D., Bingham, S. & Wilson, A. W. (2006). Prostanoid receptor EP1 and Cox-2 in injured human nerves and a rat model of nerve injury: a time-course study. *BioMed Central Neurology*, 6(1), 1.

- Dworkin, R. H., Backonja, M., Rowbotham, M. C., Allen, R. R., Argoff, C. R., Bennett, G. J. & Hewitt, D. J. (2003). Advances in neuropathic pain: diagnosis, mechanisms, and treatment recommendations. *Archives of Neurology*, 60(11), 1524-1534.
- Dworkin, R. H., O'Connor, A. B., Backonja, M., Farrar, J. T., Finnerup, N. B., Jensen, T. S., Kalso, E. A., Loeser, J. D., Miaskowski, C., Nurmikko, T. J., Portenoy, R. K., Rice, A. S. C., Stacey, B. R., Treede, R.D., Turk, D. C. & Wallace, M. S. (2007). Pharmacologic management of neuropathic pain: Evidence-based recommendations. *Pain*, 132(3), 237-251.
- Farghaly, H. S., Abd-Ellatief, R. B., Moftah, M. Z., Mostafa, M. G., Khedr, E. M. & Kotb, H. I. (2014). The Effects of Dexmedetomidine Alone and in Combination with Tramadol or Amitriptyline in a Neuropathic Pain Model. *Pain Physician*, 17(2), 187-195.
- Field, M. J., Oles, R. J., Lewis, A. S., McCleary, S., Hughes, J. & Singh, L. (1997). Gabapentin (neurontin) and S-(+)-3-isobutylgaba represent a novel class of selective antihyperalgesic agents. *British Journal of Pharmacology*, 121(8), 1513-1522.
- Finnerup, N. B. & Jensen, T. S. (2006). Mechanisms of disease: mechanism-based classification of neuropathic pain—a critical analysis. *Nature Clinical Practice Neurology*, 2(2), 107-115.
- Fukuoka, K., Sawabe, A., Sugimoto, T., Koga, M., Okuda, H., Kitayama, T., Shirai, M., Komai, K., Komemushi, S. & Matsuda, K. (2004). Inhibitory Actions of Several Natural Products on Proliferation of Rat Vascular Smooth Muscle Cells Induced by Hsp60 from *Chlamydia pneumoniae* J138. *Journal of Agricultural and Food Chemistry*, 52(20), 6326-6329.
- Fu, S. Y. & Gordon, T. (1997). The cellular and molecular basis of peripheral nerve regeneration. *Molecular Neurobiology*, 14(1-2), 67-116.
- Gatchel, R. J. (2004a). Award for Distinguished Professional Contributions to Applied Research. *American Psychologist*, 59(8), 791-805.
- Gatchel, R. J. (2004b). Comorbidity of Chronic Pain and Mental Health Disorders: The Biopsychosocial Perspective. *American Psychologist*, 59(8), 795-805.
- Gatchel, R. J., Peng, Y. B., Peters, M. L., Fuchs, P. N. & Turk, D. C. (2007). The biopsychosocial approach to chronic pain: Scientific advances and future directions. *Psychological Bulletin*, 133(4), 581-624.
- Gee, N. S., Brown, J. P., Dissanayake, V. U. K., Offord, J., Thurlow, R. & Woodruff, G. N. (1996). The Novel Anticonvulsant Drug, Gabapentin (Neurontin), Binds to the Subunit of a Calcium Channel. *Journal of Biological Chemistry*, 271(10), 5768-5776.

- Ghanouni, P., Behera, D., Xie, J., Chen, X., Moseley, M. & Biswal, S. (2012). In vivo USPIO magnetic resonance imaging shows that minocycline mitigates macrophage recruitment to a peripheral nerve injury. *Molecular Pain*, 8(49), 1-7.
- Gilron, I. & Max, M. B. (2005). Combination pharmacotherapy for neuropathic pain: current evidence and future directions. *Expert Review of Neurotherapeutics*, 5(6), 823-830.
- Gonzalez, S., Fernando, R. N., Perrin-Tricaud, C. & Tricaud, N. (2014). In vivo introduction of transgenes into mouse sciatic nerve cells in situ using viral vectors. *Nature Protocols*, 9(5), 1160-1169.
- Granados-Soto, V., Argüelles, C. F. & Ortiz, M. I. (2002). The peripheral antinociceptive effect of resveratrol is associated with activation of potassium channels. *Neuropharmacology*, 43(5), 917-923.
- Guan, Y., Yaster, M., Raja, S. N. & Tao, Y.-X. (2007). Genetic knockout and pharmacologic inhibition of neuronal nitric oxide synthase attenuate nerve injury-induced mechanical hypersensitivity in mice. *Molecular Pain*, 3, 29.
- Gunn, C. C. (1989). Neuropathic pain: a new theory for chronic pain of intrinsic origin. *Acupuncture in Medicine*, 6(2), 50-53.
- Gutierrez, V. P., Zambelli, V. O., Picolo, G., Chacur, M., Sampaio, S. C., Brigatte, P., Konno, K. & Cury, Y. (2012). The peripheral L-arginine–nitric oxide–cyclic GMP pathway and ATP-sensitive K⁺ channels are involved in the antinociceptive effect of crotalphine on neuropathic pain in rats. *Behavioural Pharmacology*, 23(1), 14-24.
- Habsah, M., Amran, M., Mackeen, M. M., Lajis, N. H., Kikuzaki, H., Nakatani, N., Rahman, A. A. & Ali, A. M. (2000). Screening of Zingiberaceae extracts for antimicrobial and antioxidant activities. *Journal of Ethnopharmacology*, 72(3), 403-410.
- Hasnah, M.S. (1991). Chemical constituents of some medicinal plants of Zingiberaceae. *Proceedings of the conference on the medicinal products from tropical rain forest; Forest Research Institute Malaysia, Kuala Lumpur*, 2: 299-304.
- Hains, B. C., Saab, C. Y., Klein, J. P., Craner, M. J. & Waxman, S. G. (2004). Altered sodium channel expression in second-order spinal sensory neurons contributes to pain after peripheral nerve injury. *The Journal of Neuroscience*, 24(20), 4832-4839.
- Harden, R. N. (1999). Gabapentin: A new tool in the treatment of neuropathic pain. *Acta Neurologica Scandinavica*, 100, 43-47.
- Hargreaves, K., Dubner, R., Brown, F., Flores, C. & Joris, J. (1988). A new and sensitive method for measuring thermal nociception in cutaneous hyperalgesia. *Pain*, 32(1), 77-88.

- Hervera, A., Negrete, R., Leáñez, S., Martín-Campos, J. & Pol, O. (2010). The role of nitric oxide in the local antiallodynic and antihyperalgesic effects and expression of δ -opioid and cannabinoid-2 receptors during neuropathic pain in mice. *Journal of Pharmacology and Experimental Therapeutics*, 334(3), 887-896.
- Hervera, A., Negrete, R., Leáñez, S., Martín-Campos, J. M. & Pol, O. (2011). Peripheral effects of morphine and expression of mu-opioid receptors in the dorsal root ganglia during neuropathic pain: nitric oxide signaling. *Molecular Pain*, 7, 25.
- Hess, D. T., Matsumoto, A., Kim, S. O., Marshall, H. E. & Stamler, J. S. (2005). Protein S-nitrosylation: purview and parameters. *Nature Reviews Molecular Cell Biology*, 6(2), 150-166.
- Hökfelt, T., Zhang, X. & Wiesenfeld-Hallin, Z. (1994). Messenger plasticity in primary sensory neurons following axotomy and its functional implications. *Trends in Neurosciences*, 17(1), 22-30.
- Hollingshead, J., Dühmke, R. M. & Cornblath, D. R. (2005). Tramadol for neuropathic pain. *The Cochrane Database of Systematic Reviews*, (3), 3726-3726.
- Holthusen, H. & Arndt, J. (1995). Nitric oxide evokes pain at nociceptors of the paravascular tissue and veins in humans. *The Journal of Physiology*, 487(1), 253-258.
- Holtum, R. E. (1950). The zingiberaceae of the Malay Peninsula. *Gard. Bull., Singapore*, 13(1), 1-249.
- Honoré, P. H., Basnet, A., Kristensen, P., Andersen, L. M., Neustrup, S., Møllgaard, P., Eljaja, L. & Bjerrum, O. J. (2011). Predictive validity of pharmacologic interventions in animal models of neuropathic pain. *Scandinavian Journal of Pain*, 2(4), 178-184.
- Hossain, M. E., Bhattacharjee, S. C. & Islam, M. D. E. (2005). Chemical Investigation on *Zingiber zerumbet* Sm. *Frontiers in Natural Product Chemistry*, 1(1), 185-187.
- Huang, G.C., Chien, T.Y., Chen, L.G. & Wang, C.C. (2005). Antitumor effects of zerumbone from *Zingiber zerumbet* in P-388D1 cells in vitro and in vivo. *Planta Medica*, 71(3), 219-224.
- Hucho, T. & Levine, J. D. (2007). Signaling Pathways in Sensitization: Toward a Nociceptor Cell Biology. *Neuron*, 55(3), 365-376.
- Hudson, L. J., Bevan, S., Wotherspoon, G., Gentry, C., Fox, A. & Winter, J. (2001). VR1 protein expression increases in undamaged DRG neurons after partial nerve injury. *European Journal of Neuroscience*, 13(11), 2105-2114.
- Hughes, S. R., Williams, T. J. & Brain, S. D. (1990). Evidence that endogenous nitric oxide modulates oedema formation induced by substance P. *European Journal of Pharmacology*, 191(3), 481-484.

- Ialenti, A., Ianaro, A., Moncada, S. & Di Rosa, M. (1992). Modulation of acute inflammation by endogenous nitric oxide. *European Journal of Pharmacology*, 211(2), 177-182.
- Indu, B. J., Ng, L. T. & Malaysia, I. P. d. K. P. (2000). *Herbs: The Green Pharmacy of Malaysia*. Vinpress.
- Iyengar, S., Webster, A. A., Hemrick-Luecke, S. K., Xu, J. Y. & Simmons, R. M. A. (2004). Efficacy of Duloxetine, a Potent and Balanced Serotonin-Norepinephrine Reuptake Inhibitor in Persistent Pain Models in Rats. *Journal of Pharmacology and Experimental Therapeutics*, 311(2), 576-584.
- Jaffrey, S. R. & Snyder, S. H. (1995). Nitric Oxide: A Neural Messenger. *Annual Review of Cell and Developmental Biology*, 11(1), 417-440.
- Jaggi, A. S., Jain, V. & Singh, N. (2011). Animal models of neuropathic pain. *Fundamental and Clinical Pharmacology*, 25(1), 1-28.
- Jang, D. S., Min, H.Y., Kim, M.S., Han, A.R., Windono, T., Jeohn, G.H., Kang, S. S., Lee, S. K. & Seo, E.K. (2005). Humulene Derivatives from *Zingiber zerumbet* with the Inhibitory Effects on Lipopolysaccharide-Induced Nitric Oxide Production. *Chemical and Pharmaceutical Bulletin*, 53(7), 829-831.
- Jantan, I. B., Yassin, M. S. M., Chin, C. B., Chen, L. L. & Sim, N. L. (2003). Antifungal Activity of the Essential Oils of Nine Zingiberaceae Species. *Pharmaceutical Biology*, 41(5), 392-397.
- Jensen, T. S. & Baron, R. (2003). Translation of symptoms and signs into mechanisms in neuropathic pain. *Pain*, 102(1-2), 1-8.
- Jensen, T. S., Gottrup, H., Sindrup, S. H. & Bach, F. W. (2001). The clinical picture of neuropathic pain. *European Journal of Pharmacology*, 429(1-3), 1-11.
- Johnson, R. W., Wasner, G., Saddier, P. & Baron, R. (2007). Postherpetic neuralgia: epidemiology, pathophysiology and management. *Expert Review of Neurotherapeutics*, 7(11), 1581-1595.
- Julius, D. & Basbaum, A. I. (2001). Molecular mechanisms of nociception. *Nature*, 413(6852), 203-210.
- Kaeidi, A., Esmaeili-Mahani, S., Sheibani, V., Abbasnejad, M., Rasouljan, B., Hajializadeh, Z. & Afrazi, S. (2011). Olive (*Olea europaea* L.) leaf extract attenuates early diabetic neuropathic pain through prevention of high glucose-induced apoptosis: In vitro and in vivo studies. *Journal of Ethnopharmacology*, 136(1), 188-196.
- Kamaldin, M., Akhtar, M., Mohamad, A., Lajis, N., Perimal, E., Akira, A., Ming-Tatt, L., Israf, D. & Sulaiman, M. (2013). Peripheral Antinociception of a Chalcone, Flavokawin B and Possible Involvement of the Nitric Oxide/Cyclic Guanosine Monophosphate/Potassium Channels Pathway. *Molecules*, 18(4), 4209.

- Kawabata, A., Manabe, S., Manabe, Y. & Takagi, H. (1994). Effect of topical administration of l-arginine on formalin-induced nociception in the mouse: a dual role of peripherally formed NO in pain modulation. *British Journal of Pharmacology*, **112**(2), 547-550.
- Kawano, T., Zoga, V., Kimura, M., Liang, M.Y., Wu, H.E., Gemes, G., McCallum, J. B., Kwok, W.M., Hogan, Q. H. & Sarantopoulos, C. D. (2009). Nitric oxide activates ATP-sensitive potassium channels in mammalian sensory neurons: action by direct S-nitrosylation. *Molecular Pain*, 5(12), 12.
- Keong, Y. S., Alitheen, N. B., Mustafa, S., Aziz, S. A., Rahman, M. A. & Ali, A. M. (2010). Immunomodulatory effects of zerumbone isolated from roots of *Zingiber zerumbet*. *Pakistan Journal of Pharmaceutical Sciences*, 23(1), 75-82.
- Khalid, M. H., Akhtar, M. N., Mohamad, A. S., Perimal, E. K., Akira, A., Israf, D. A., Lajis, N. & Sulaiman, M. R. (2011). Antinociceptive effect of the essential oil of *Zingiber zerumbet* in mice: possible mechanismss. *Journal of Ethnopharmacology*, 137(1), 345-351.
- Kim, B.S., Yoo, J. J. & Atala, A. (2004). Peripheral nerve regeneration using acellular nerve grafts. *Journal of Biomedical Materials Research Part A*, 68A(2), 201-209.
- Kim, C. F. & Moalem-Taylor, G. (2011). Interleukin-17 Contributes to Neuroinflammation and Neuropathic Pain Following Peripheral Nerve Injury in Mice. *The Journal of Pain*, 12(3), 370-383.
- Kinghorn, A., Farnsworth, N., Soejarto, D., Cordell, G., Swanson, S., Pezzuto, J., Wani, M., Wall, M., Oberlies, N., Kroll, D., Kramer, R., Rose, W., Vite, G., Fairchild, C., Peterson, R. & Wild, R. (2003). Novel Strategies for the Discovery of Plant-Derived Anticancer Agents. *Pharmaceutical Biology*, 41(s1), 53-67.
- Kirana, C., McIntosh, G. H., Record, I. R. & Jones, G. P. (2003). Antitumor Activity of Extract of *Zingiber aromaticum* and Its Bioactive Sesquiterpenoid Zerumbone. *Nutrition and Cancer*, 45(2), 218-225.
- Kishi, T., Hirooka, Y., Mukai, Y., Shimokawa, H. & Takeshita, A. (2003). Atorvastatin causes depressor and sympatho-inhibitory effects with upregulation of nitric oxide synthases in stroke-prone spontaneously hypertensive rats. *Journal of Hypertension*, 21(2), 379-386.
- Kitto, K. F., Haley, J. E. & Wilcox, G. L. (1992). Involvement of nitric oxide in spinally mediated hyperalgesia in the mouse. *Neuroscience Letters*, 148(1-2), 1-5.
- Koesling, D., Russwurm, M., Mergia, E., Mullershausen, F. & Friebe, A. (2004). Nitric oxide-sensitive guanylyl cyclase: structure and regulation. *Neurochemistry International*, 45(6), 813-819.
- Koltzenburg, M. & Scadding, J. (2001). Neuropathic pain. *Current opinion in neurology*, 14(5), 641-647.

- Koshimizu, H., Araki, T., Takai, S., Yokomaku, D., Ishikawa, Y., Kubota, M., Sano, S.i., Hatanaka, H. & Yamada, M. (2002). Expression of CD47/integrin-associated protein induces death of cultured cerebral cortical neurons. *Journal of Neurochemistry*, 82(2), 249-257.
- Kumar, A., Meena, S. & Pottabathini, R. (2014). Effect of Ashwagandha (*Withania somnifera*) against chronic constriction injury induced behavioral and biochemical alterations: Possible involvement of nitric oxide mechanism. *International Journal of Nutrition, Pharmacology, Neurological Diseases*, 4(3), 131.
- Kumar, A., Meena, S., Kalonia, H., Gupta, A. & Kumar, P. (2011). Effect of nitric oxide in protective effect of melatonin against chronic constriction sciatic nerve injury induced neuropathic pain in rats. *Indian Journal of Experimental Biology*, 49(9), 664.
- Langner, E., Greifenberg, S. & Gruenwald, J. (1998). Ginger: history and use. *Advances in Therapy*, 15(1), 25-44.
- Larsen, K., Ibrahim, H., Khaw, S. & Saw, L. (1999). *Gingers of Peninsular Malaysia and Singapore*. Natural History Publications (Borneo). 137.
- Latremoliere, A. & Woolf, C. J. (2009). Central Sensitization: A Generator of Pain Hypersensitivity by Central Neural Plasticity. *The Journal of Pain : official journal of the American Pain Society*, 10(9), 895-926.
- Lau, F. H. & Chung, K. C. (2004). Silas Weir Mitchell, MD: the physician who discovered causalgia. *Journal of Hand Surgery*, 29(2), 181-187.
- Lázaro-Ibáñez, G. G., Torres-López, J. E. & Granados-Soto, V. (2001). Participation of the nitric oxide–cyclic GMP–ATP-sensitive K^+ channel pathway in the antinociceptive action of ketorolac. *European Journal of Pharmacology*, 426(1–2), 39-44.
- Levy, D. & Strassman, A. M. (2004). Modulation of Dural Nociceptor Mechanosensitivity by the Nitric Oxide-Cyclic GMP Signaling Cascade. Vol. 92.
- Levy, D. & Zochodne, D. W. (1998). Local nitric oxide synthase activity in a model of neuropathic pain. *European Journal of Neuroscience*, 10(5), 1846-1855.
- Levy, D. & Zochodne, D. W. (2004). NO Pain: Potential Roles of Nitric Oxide in Neuropathic Pain. *Pain Practice*, 4(1), 11-18.
- Levy, D., Höke, A. & Zochodne, D. W. (1999). Local expression of inducible nitric oxide synthase in an animal model of neuropathic pain. *Neuroscience Letters*, 260(3), 207-209.
- Levy, D., Kubes, P. & Zochodne, D. W. (2001). Delayed peripheral nerve degeneration, regeneration, and pain in mice lacking inducible nitric oxide synthase. *Journal of Neuropathology and Experimental Neurology*, 60(5), 411-421.

- Liu, Y.L., Zhou, L.J., Hu, N.W., Xu, J.T., Wu, C.Y., Zhang, T., Li, Y.Y. & Liu, X.G. (2007). Tumor necrosis factor- α induces long-term potentiation of C-fiber evoked field potentials in spinal dorsal horn in rats with nerve injury: The role of NF-kappa B, JNK and p38 MAPK. *Neuropharmacology*, 52(3), 708-715.
- Lorenzetti, B. B. & Ferreira, S. H. (1996). Activation of the arginine-nitric oxide pathway in primary sensory neurons contributes to dipyrone-induced spinal and peripheral analgesia. *Inflammation Research*, 45(6), 308-311.
- Lucas, K. A., Pitari, G. M., Kazerounian, S., Ruiz-Stewart, I., Park, J., Schulz, S., Chepenik, K. P. & Waldman, S. A. (2000). Guanylyl Cyclases and Signaling by Cyclic GMP. *Pharmacological Reviews*, 52(3), 375-414.
- Luo, Z. D., Calcutt, N. A., Higuera, E. S., Valder, C. R., Song, Y.H., Svensson, C. I. & Myers, R. R. (2002). Injury Type-Specific Calcium Channel $\alpha 2\delta$ -1 Subunit Up-Regulation in Rat Neuropathic Pain Models Correlates with Antiallodynic Effects of Gabapentin. *Journal of Pharmacology and Experimental Therapeutics*, 303(3), 1199-1205.
- Luo, Z. D., Chaplan, S. R., Higuera, E. S., Sorkin, L. S., Stauderman, K. A., Williams, M. E. & Yaksh, T. L. (2001). Upregulation of dorsal root ganglion $\alpha 2\delta$ calcium channel subunit and its correlation with allodynia in spinal nerve-injured rats. *The Journal of Neuroscience*, 21(6), 1868-1875.
- Mao, J., Price, D. D. & Mayer, D. J. (1995). Experimental mononeuropathy reduces the antinociceptive effects of morphine: implications for common intracellular mechanisms involved in morphine tolerance and neuropathic pain. *Pain*, 61(3), 353-364.
- Marchand, F., Perretti, M. & McMahon, S. B. (2005). Role of the immune system in chronic pain. *Nature Reviews Neuroscience*, 6(7), 521-532.
- Martínez-Salio, A., Porta-Etessam, J., Berbel-Garcia, A., Garcia-Morales, I., de la Peña-Mayor, P. & Vicente-Fatela, L. (2001). Antiepileptic drugs and neuropathic pain. *Revista de neurologia*, 32(4), 345-350.
- Martini, R., Fischer, S., López-Vales, R. & David, S. (2008). Interactions between Schwann cells and macrophages in injury and inherited demyelinating disease. *Glia*, 56(14), 1566-1577.
- Martinov, T., Mack, M., Sykes, A. & Chatterjea, D. (2013). Measuring Changes in Tactile Sensitivity in the Hind Paw of Mice Using an Electronic von Frey Apparatus. *Journal of Visualized Experiments: JoVE* (82), 51212.
- Martucci, C., Trovato, A. E., Costa, B., Borsani, E., Franchi, S., Magnaghi, V., Panerai, A. E., Rodella, L. F., Valsecchi, A. E., Sacerdote, P. & Colleoni, M. (2008). The purinergic antagonist PPADS reduces pain related behaviours and interleukin-1 β , interleukin-6, iNOS and nNOS overproduction in central and peripheral nervous system after peripheral neuropathy in mice. *Pain*, 137(1), 81-95.

- Matthews, E. A. & Dickenson, A. H. (2001). Effects of spinally delivered N- and P-type voltage-dependent calcium channel antagonists on dorsal horn neuronal responses in a rat model of neuropathy. *Pain*, 92(1–2), 235-246.
- Ma, W. & Eisenach, J. C. (2003). Cyclooxygenase 2 in infiltrating inflammatory cells in injured nerve is universally up-regulated following various types of peripheral nerve injury. *Neuroscience*, 121(3), 691-704.
- Meller, S. T. & Gebhart, G. F. (1993). Nitric oxide (NO) and nociceptive processing in the spinal cord. *Pain*, 52(2), 127-136.
- Meller, S. T., Pechman, P. S., Gebhart, G. F. & Maves, T. J. (1992). Nitric oxide mediates the thermal hyperalgesia produced in a model of neuropathic pain in the rat. *Neuroscience*, 50(1), 7-10.
- Melzack, R. (2001). Pain and the neuromatrix in the brain. *Journal of Dental Education*, 65(12), 1378-1382.
- Melzack, R. (2005). Evolution of the Neuromatrix Theory of Pain. The Prithvi Raj Lecture: Presented at the Third World Congress of World Institute of Pain, Barcelona 2004. *Pain Practice*, 5(2), 85-94.
- Melzack, R., & Casey, K. L. (1968). Sensory, motivational, and central control determinants of pain. In *The Skin Senses*, 423-439.
- Meotti, F. C., Missau, F. C., Ferreira, J., Pizzolatti, M. G., Mizuzaki, C., Nogueira, C. W. & Santos, A. R. S. (2006). Anti-allodynic property of flavonoid myricitrin in models of persistent inflammatory and neuropathic pain in mice. *Biochemical Pharmacology*, 72(12), 1707-1713.
- Merskey, H. & Bogduk, N. (1994). Classification of chronic pain, IASP Task Force on Taxonomy. Seattle, WA: International Association for the Study of Pain Press. (Also available online at www.iasp-pain.org).
- Merskey, H. E. (1986). Classification of chronic pain: Descriptions of chronic pain syndromes and definitions of pain terms. *Pain*, 3, 226.
- Meyer-Rosberg, K., Kvarnström, A., Kinnman, E., Gordh, T., Nordfors, L.O. & Kristofferson, A. (2001). Peripheral neuropathic pain—a multidimensional burden for patients. *European Journal of Pain*, 5(4), 379-389.
- Ming-Tatt, L., Khalivulla, S. I., Akhtar, M. N., Lajis, N., Perimal, E. K., Akira, A., Ali, D. I. & Sulaiman, M. R. (2013). Anti-hyperalgesic effect of a benzilidene-cyclohexanone analogue on a mouse model of chronic constriction injury-induced neuropathic pain: Participation of the κ -opioid receptor and KATP. *Pharmacology Biochemistry and Behavior*, 114–115(0), 58-63.
- Mitchell, S. W. (1872). *Injuries of nerves and their consequences*. JB Lippincott.

- Mitchell, S. W., Morehouse, G. R. & Keen, W. W. (2007). Gunshot wounds and other injuries of nerves. 1864. *Clinical Orthopaedics and Related Research*, 458, 35-39.
- Moalem, G. & Tracey, D. J. (2006). Immune and inflammatory mechanisms in neuropathic pain. *Brain Research Reviews*, 51(2), 240-264.
- Moncada, S. (1997). Nitric Oxide in the Vasculature: Physiology and Pathophysiology. *Annals of the New York Academy of Sciences*, 811(1), 60-69.
- Moore, K. A., Kohno, T., Karchewski, L. A., Scholz, J., Baba, H. & Woolf, C. J. (2002). Partial peripheral nerve injury promotes a selective loss of GABAergic inhibition in the superficial dorsal horn of the spinal cord. *The Journal of Neuroscience*, 22(15), 6724-6731.
- Murakami, A., Hayashi, R., Takana, T., Kwon, K. H., Ohigashi, H. & Safitri, R. (2003). Suppression of dextran sodium sulfate-induced colitis in mice by zerumbone, a subtropical ginger sesquiterpene, and nimesulide: separately and in combination. *Biochemical Pharmacology*, 66(7), 1253-1261.
- Murakami, A., Takahashi, D., Kinoshita, T., Koshimizu, K., Kim, H. W., Yoshihiro, A., Nakamura, Y., Jiwajinda, S., Terao, J. & Ohigashi, H. (2002). Zerumbone, a Southeast Asian ginger sesquiterpene, markedly suppresses free radical generation, proinflammatory protein production, and cancer cell proliferation accompanied by apoptosis: the α,β -unsaturated carbonyl group is a prerequisite. *Carcinogenesis*, 23(5), 795-802.
- Murakami, A., Takahashi, M., Jiwajinda, S., Koshimizu, K. & Ohigashi, H. (1999). Identification of Zerumbone in *Zingiber zerumbet* Smith as a Potent Inhibitor of 12-O-Tetradecanoylphorbol-13-acetate-induced Epstein - Barr virus Activation. *Bioscience, Biotechnology, and Biochemistry*, 63(10), 1811-1812.
- Murakami, A., Tanaka, T., Lee, J.Y., Surh, Y.J., Kim, H. W., Kawabata, K., Nakamura, Y., Jiwajinda, S. & Ohigashi, H. (2004). Zerumbone, a sesquiterpene in subtropical ginger, suppresses skin tumor initiation and promotion stages in ICR mice. *International Journal of Cancer*, 110(4), 481-490.
- Muthuraman, A. & Singh, N. (2011). Attenuating effect of Acorus calamus extract in chronic constriction injury induced neuropathic pain in rats: an evidence of anti-oxidative, anti-inflammatory, neuroprotective and calcium inhibitory effects. *BMC Complementary and Alternative Medicine*, 11(1), 1-14.
- Nadal, X., Baños, J. E., Kieffer, B. L. & Maldonado, R. (2006). Neuropathic pain is enhanced in δ -opioid receptor knockout mice. *European Journal of Neuroscience*, 23(3), 830-834.
- Naik, A. K., Tandan, S. K., Kumar, D. & Dudhgaonkar, S. P. (2006). Nitric oxide and its modulators in chronic constriction injury-induced neuropathic pain in rats. *European Journal of Pharmacology*, 530(1-2), 59-69.

- Nhareet, S. & Nur, S. (2003). Anti-inflammatory property of ethanol and water extracts of *Zingiber Zerumbet*. Indian Journal of Pharmacology, 35(3), 181-182.
- Nicholson, B. (2000). Gabapentin use in Neuropathic Pain syndromes. Journal of the Peripheral Nervous System, 5(4), 245-245.
- Nickel, F. T., Seifert, F., Lanz, S. & Maihöfner, C. (2012). Mechanisms of neuropathic pain. European Neuropsychopharmacology, 22(2), 81-91.
- Obata, K., Katsura, H., Sakurai, J., Kobayashi, K., Yamanaka, H., Dai, Y., Fukuoka, T. & Noguchi, K. (2006). Suppression of the p75 Neurotrophin Receptor in Uninjured Sensory Neurons Reduces Neuropathic Pain after Nerve Injury. The Journal of Neuroscience, 26(46), 11974-11986.
- Ocaña, M., Cendán, C. M., Cobos, E. J., Entrena, J. M. & Baeyens, J. M. (2004). Potassium channels and pain: present realities and future opportunities. European Journal of Pharmacology, 500(1-3), 203-219.
- Okuducu, H. & Onal, S. (2005). Is nitric oxide involved in the antinociceptive activity of tramadol? Findings in a rat model of neuropathic pain. AGRI-ISTANBUL, 17(4), 31.
- Ossipov, M. H. & Porreca, F. (2005). Challenges in the Development of Novel Treatment Strategies for Neuropathic Pain. NeuroRx, 2(4), 650-661.
- Ozaki, Y., Kawahara, N. & Harada, M. (1991). Anti-inflammatory effect of *Zingiber cassumunar* Roxb. and its active principles. Chemical and Pharmaceutical Bulletin, 39(9), 2353-2356.
- PAIN, C. M. (1957). Randall LO, Selitto JJ. Arch Int Pharmacodyn, 3, 409-19.
- Pan, H.C., Chen, C.J., Cheng, F.C., Ho, S.P., Liu, M.J., Hwang, S.M., Chang, M.H. & Wang, Y.C. (2009a). Combination of G-CSF Administration and Human Amniotic Fluid Mesenchymal Stem Cell Transplantation Promotes Peripheral Nerve Regeneration. Neurochemical Research, 34(3), 518-527.
- Pan, H.C., Cheng, F.C., Chen, C.J., Lai, S.Z., Lee, C.W., Yang, D.Y., Chang, M.H. & Ho, S.P. (2007). Post-injury regeneration in rat sciatic nerve facilitated by neurotrophic factors secreted by amniotic fluid mesenchymal stem cells. Journal of Clinical Neuroscience, 14(11), 1089-1098.
- Pan, H.C., Cheng, F.C., Chen, C.J., Lai, S.Z., Liu, M.J., Chang, M.H., Wang, Y.C., Yang, D.Y. & Ho, S.P. (2009b). Dietary supplement with fermented soybeans, natto, improved the neurobehavioral deficits after sciatic nerve injury in rats. Neurological Research, 31(5), 441-452.
- Pan, H.C., Chin, C.S., Yang, D.Y., Ho, S.P., Chen, C.J., Hwang, S.M., Chang, M.H. & Cheng, F.C. (2009c). Human Amniotic Fluid Mesenchymal Stem Cells in Combination with Hyperbaric Oxygen Augment Peripheral Nerve Regeneration. Neurochemical Research, 34(7), 1304-1316.

- Pan, H. C., Yang, D. Y., Ho, S. P., Sheu, M. L., Chen, C. J., Hwang, S. M. & Cheng, F. C. (2009d). Escalated regeneration in sciatic nerve crush injury by the combined therapy of human amniotic fluid mesenchymal stem cells and fermented soybean extracts, *Natto Journal of Biomedical Science*, 16(75), 10-1186.
- Pasero, C. & McCaffery, M. (1999). *Pain: Clinical Manual*. Vol. 9: Mosby Incorporated.
- Pasero, C. (2004). Pathophysiology of neuropathic pain. *Pain Management Nursing*, 5, Supplement (0), 3-8.
- Perimal, E. K., Akhtar, M. N., Mohamad, A. S., Khalid, M. H., Ming, O. H., Khalid, S., Tatt, L. M., Kamaldin, M. N., Zakaria, Z. A., Israf, D. A., Lajis, N. & Sulaiman, M. R. (2011). Zerumbone-Induced Antinociception: Involvement of the l-Arginine-Nitric Oxide-cGMP -PKC-K⁺ATP Channel Pathways. *Basic and Clinical Pharmacology and Toxicology*, 108(3), 155-162.
- Perry, L. M. & Metzger, J. (1980). *Medicinal plants of east and Southeast Asia: attributed properties and uses*. MIT press.
- Prast, H. & Philippu, A. (2001). Nitric oxide as modulator of neuronal function. *Progress in Neurobiology*, 64(1), 51-68.
- Raivich, G. & Makwana, M. (2007). The making of successful axonal regeneration: genes, molecules and signal transduction pathways. *Brain Research Reviews*, 53(2), 287-311.
- Robinson, I. & Meert, T. F. (2005). Stability of neuropathic pain symptoms in partial sciatic nerve ligation in rats is affected by suture material. *Neuroscience Letters*, 373(2), 125-129.
- Rodrigues, A. R. A. & Duarte, I. D. G. (2000). The peripheral antinociceptive effect induced by morphine is associated with ATP-sensitive K⁺ channels. *British Journal of Pharmacology*, 129(1), 110-114.
- Rotshenker, S. (2011). Wallerian degeneration: the innate-immune response to traumatic nerve injury. *Journal of Neuroinflammation*, 8(1), 109.
- Ruslay, S., Abas, F., Shaari, K., Zainal, Z., Maulidiani, Sirat, H., Israf, D. A. & Lajis, N. H. (2007). Characterization of the components present in the active fractions of health gingers (*Curcuma xanthorrhiza* and *Zingiber zerumbet*) by HPLC-DAD-ESIMS. *Food Chemistry*, 104(3), 1183-1191.
- Sabu, M. (2003). Revision of the genus *Zingiber* in South India. *Folia Malaysiana*, 4(1), 25-52.
- Sacerdote, P., Franchi, S., Trovato, A. E., Valsecchi, A. E., Panerai, A. E. & Colleoni, M. (2008). Transient early expression of TNF- α in sciatic nerve and dorsal root ganglia in a mouse model of painful peripheral neuropathy. *Neuroscience Letters*, 436(2), 210-213.

- Sachs, D., Cunha, F. Q. & Ferreira, S. H. (2004). Peripheral analgesic blockade of hypernociception: Activation of arginine/NO/cGMP/protein kinase G/ATP-sensitive K⁺ channel pathway. *Proceedings of the National Academy of Sciences of the United States of America*, 101(10), 3680-3685.
- Sakinah, S. A., Handayani, S. T. & Hawariah, L. P. (2007). Zerumbone induced apoptosis in liver cancer cells via modulation of Bax/Bcl-2 ratio. *Cancer Cell International*, 7(4), 1-11.
- Siddall, P. J. & Cousins, M. J. (1997). Spinal Pain Mechanisms. *Spine*, 22(1), 98-104.
- Santos-Nogueira, E., Redondo Castro, E., Mancuso, R. & Navarro, X. (2012). Randall-Selitto Test: A New Approach for the Detection of Neuropathic Pain after Spinal Cord Injury. *Journal of Neurotrauma*, 29(5), 898-904.
- Schisler, R. E. & Groninger, H. (2012). Topical Capsaicin for Neuropathic Pain. *Journal of Palliative Medicine*, 15(8), 946-947.
- Schmidtke, A., Tegeder, I. & Geisslinger, G. (2009). No NO, no pain? The role of nitric oxide and cGMP in spinal pain processing. *Trends in Neurosciences*, 32(6), 339-346.
- Scholz, J. & Woolf, C. J. (2002). Can we conquer pain? *Nature Neuroscience*, 5, 1062-1067.
- Scholz, J. & Woolf, C. J. (2007). The neuropathic pain triad: neurons, immune cells and glia. *Nature Neuroscience*, 10(11), 1361-1368.
- Scholz, J., Mannion, R. J., Hord, D. E., Griffin, R. S., Rawal, B., Zheng, H. & Abdi, S. (2009). A novel tool for the assessment of pain: validation in low back pain. *PLoS Medicine*, 6(4), 446.
- Sekiguchi, M., Sekiguchi, Y., Konno, S.i., Kobayashi, H., Homma, Y. & Kikuchi, S.I. (2009). Comparison of neuropathic pain and neuronal apoptosis following nerve root or spinal nerve compression. *European Spine Journal*, 18(12), 1978-1985.
- Shaik-Dasthagirisahab, Y. B., Varvara, G., Murmura, G., Saggini, A., Potalivo, G., Caraffa, A., Antinolfi, P., Tete, S., Tripodi, D., Conti, F., Cianchetti, E., Toniato, E., Rosati, M., Conti, P., Speranza, L., Pantalone, A., Saggini, R., Theoharides, T. C. & Pandolfi, F. (2013). Vascular endothelial growth factor (VEGF), mast cells and inflammation. *Int J Immunopathol Pharmacol*, 26(2), 327-35.
- Sirirugsa, P. (1999). Thai Zingiberaceae: species diversity and their uses. *World (total)*, 52, 1-500.
- Sivilotti, L. & Woolf, C. J. (1994). The contribution of GABAA and glycine receptors to central sensitization: disinhibition and touch-evoked allodynia in the spinal cord. *Journal of Neurophysiology*, 72(1), 169-179.

- Smith, B. H., Elliott, A. M., Chambers, W. A., Smith, W. C., Hannaford, P. C. & Penny, K. (2001). The impact of chronic pain in the community. *Family Practice*, 18(3), 292-299.
- Snyder, S. (1992). Nitric oxide: first in a new class of neurotransmitters. *Science*, 257(5069), 494-496.
- Soares, A. C. & Duarte, I. D. G. (2001). Dibutyl-cyclic GMP induces peripheral antinociception via activation of ATP-sensitive K⁺ channels in the rat PGE₂-induced hyperalgesic paw. *British Journal of Pharmacology*, 134(1), 127-131.
- Somchit, M. N., Mak, J. H., A., Bustamam, A., Zuraini, A., Arifah, A. K., Adam, Y. & Zakaria, Z. A. (2012). Zerumbone isolated from *Zingiber zerumbet* inhibits inflammation and pain in rats. *Journal of Medicinal Plants Research*, 6, 177-180.
- Sommer, C. & Kress, M. (2004). Recent findings on how proinflammatory cytokines cause pain: peripheral mechanisms in inflammatory and neuropathic hyperalgesia. *Neuroscience Letters*, 361(1-3), 184-187.
- Sommer, C. (2003). Painful neuropathies. *Current Opinion in Neurology*, 16(5), 623-628.
- Sousa, A. M. & Prado, W. A. (2001). The dual effect of a nitric oxide donor in nociception. *Brain Research*, 897(1-2), 9-19.
- Srivastava, A. K., Srivastava, S. K. & Shah, N. C. (2000). Essential Oil Composition of *Zingiber zerumbet* (L.) Sm. from India. *Journal of Essential Oil Research*, 12(5), 595-597.
- Stillman, M. (2006). Clinical approach to patients with neuropathic pain. *Cleveland Clinic Journal of Medicine*, 73(8), 726.
- Stoll, G. & Müller, H. W. (1999). Nerve Injury, Axonal Degeneration and Neural Regeneration: Basic Insights. *Brain Pathology*, 9(2), 313-325.
- Stoll, G., Griffin, J. W., Li, C. Y. & Trapp, B. D. (1989). Wallerian degeneration in the peripheral nervous system: participation of both Schwann cells and macrophages in myelin degradation. *Journal of Neurocytology*, 18(5), 671-683.
- Sulaiman, M. R., Perimal, E. K., Akhtar, M. N., Mohamad, A. S., Khalid, M. H., Tasrip, N. A., Mokhtar, F., Zakaria, Z. A., Lajis, N. H. & Israf, D. A. (2010a). Anti-inflammatory effect of zerumbone on acute and chronic inflammation models in mice. *Fitoterapia*, 81(7), 855-858.
- Sulaiman, M. R., Perimal, E. K., Zakaria, Z. A., Mokhtar, F., Akhtar, M. N., Lajis, N. & Israf, D. (2009). Preliminary analysis of the antinociceptive activity of zerumbone. *Fitoterapia*, 80(4), 230-232.
- Sulaiman, M. R., Tengku Mohamad, T. A. S., Shaik Mossadeq, W. M., Moin, S., Yusof, M., Mokhtar, A. F., Zakaria, Z. A., Israf, D. A. & Lajis, N. (2010b).

- Antinociceptive activity of the essential oil of *Zingiber zerumbet*. *Planta medica*, 76(2), 107-112.
- Sutton, K. G., Martin, D. J., Pinnock, R. D., Lee, K. & Scott, R. H. (2002). Gabapentin inhibits high-threshold calcium channel currents in cultured rat dorsal root ganglion neurones. *British Journal of Pharmacology*, 135(1), 257-265.
- Szczudlik, A., Dobrogowski, J., Wordliczek, J., Stępień, A., Krajnik, M., Leppert, W., Woroń, J., Przekłasa-Muszyńska, A., Kocot-Kępska, M., Zajączkowska, R., Janecki, M., Adamczyk, A. & Malec-Milewska, M. (2014). Diagnosis and management of neuropathic pain: Review of literature and recommendations of the Polish Association for the Study of Pain and the Polish Neurological Society – Part Two. *Neurologia i Neurochirurgia Polska*, 48(6), 423-435.
- Tandrup, T., Woolf, C. J. & Coggeshall, R. E. (2000). Delayed loss of small dorsal root ganglion cells after transection of the rat sciatic nerve. *The Journal of Comparative Neurology*, 422(2), 172-180.
- Tang, Q., Svensson, C. I., Fitzsimmons, B., Webb, M., Yaksh, T. L. & Hua, X.Y. (2007). Inhibition of spinal constitutive NOS-2 by 1400W attenuates tissue injury and inflammation-induced hyperalgesia and spinal p38 activation. *European Journal of Neuroscience*, 25(10), 2964-2972.
- Taskinen, H. S., Olsson, T., Bucht, A., Khademi, M., Svelander, L. & Røytta, M. (2000). Peripheral nerve injury induces endoneurial expression of IFN- γ , IL-10 and TNF- α mRNA. *Journal of Neuroimmunology*, 102(1), 17-25.
- Taskinen, H.S. & Røytta, M. (2000). Increased expression of chemokines (MCP-1, MIP-1 α , RANTES) after peripheral nerve transection. *Journal of the Peripheral Nervous System*, 5(2), 75-81.
- Terenghi, G., Riveros-Moreno, V., Hudson, L. D., Ibrahim, N. B. N. & Polak, J. M. (1993). Immunohistochemistry of nitric oxide synthase demonstrates immunoreactive neurons in spinal cord and dorsal root ganglia of man and rat. *Journal of the Neurological Sciences*, 118(1), 34-37.
- Thiagarajan, V. R., Shanmugam, P., Krishnan, U. M. & Muthuraman, A. (2014). Ameliorative potential of *Vernonia cinerea* on chronic constriction injury of sciatic nerve induced neuropathic pain in rats. *An Acad Bras Cienc*, 86(3), 1435-1449.
- Thomas, D. A., Ren, K., Besse, D., Ruda, M. A. & Dubner, R. (1996). Application of nitric oxide synthase inhibitor, N ω -nitro-L-arginine methyl ester, on injured nerve attenuates neuropathy-induced thermal hyperalgesia in rats. *Neuroscience Letters*, 210(2), 124-126.
- Tölle, T. R. (2010). Challenges with current treatment of neuropathic pain. *European Journal of Pain Supplements*, 4(S2), 161-165.
- Tos, P., Ronchi, G., Papalia, I., Sallen, V., Legagneux, J., Geuna, S. & Giacobini-Robecchi, M. (2009). Chapter 4 Methods and Protocols in Peripheral Nerve

Regeneration Experimental Research: Part I—Experimental Models. In, International Review of Neurobiology Vol. Volume 87: Academic Press, pp 47-79.

Tsuda, M., Inoue, K. & Salter, M. W. (2005). Neuropathic pain and spinal microglia: a big problem from molecules in 'small' glia. *Trends in Neurosciences*, 28(2), 101-107.

Üçeyler, N. & Sommer, C. (2008). Cytokine regulation in animal models of neuropathic pain and in human diseases. *Neuroscience Letters*, 437(3), 194-198.

Ueda, H. & Rashid, M. H. (2003). Molecular mechanisms of neuropathic pain. *Drug News Perspect*, 16(9), 605.

Uhlenius, C., Linderöth, B., Meyerson, B. A. & Wallin, J. (2006). Spinal NMDA receptor phosphorylation correlates with the presence of neuropathic signs following peripheral nerve injury in the rat. *Neuroscience Letters*, 399(1-2), 85-90.

Uuml, Ltekin, H., I. & Ahmedov, V. (2006). The Roles of the Opioidergic System and Nitric Oxide in the Analgesic Effect of Venlafaxine. *The Pharmaceutical Society of Japan*, 126(2), 117-121.

Vale, M. L., Rolim, D. E., Cavalcante, I. F., Ribeiro, R. A. & Souza, M. H. L. P. (2007). Role of NO/cGMP/KATP pathway in antinociceptive effect of sildenafil in zymosan writhing response in mice. *Inflammation Research*, 56(2), 83-88.

Verge, V. M., Xu, Z., Xu, X. J., Wiesenfeld-Hallin, Z. & Hökfelt, T. (1992). Marked increase in nitric oxide synthase mRNA in rat dorsal root ganglia after peripheral axotomy: in situ hybridization and functional studies. *Proceedings of the National Academy of Sciences*, 89(23), 11617-11621.

Vivancos, G. G., Parada, C. A. & Ferreira, S. H. (2003). Opposite nociceptive effects of the arginine/NO/cGMP pathway stimulation in dermal and subcutaneous tissues. *British Journal of Pharmacology*, 138(7), 1351-1357.

Wagner, W. L., Herbst, D. R. & Sohmer, S. H. (1999). *Manual of the Flowering Plants of Hawai'i*, Vols. 1 and 2. University of Hawai'i and Bishop Museum Press.

Wall, P. D. (1979). On the relation of injury to pain the John J. Bonica Lecture. *Pain*, 6(3), 253-264.

Walter, R. D. (1970). S. Weir Mitchell, MD, neurologist: a medical biography. Thomas.

Wang, J., Zhang, L.C., Lv, Y.W., Ji, Y., Yan, X.J. & Xue, J.P. (2008). Involvement of the nitric oxide-cyclic GMP-protein kinase G-K⁺ channel pathway in the antihyperalgesic effects of bovine lactoferrin in a model of neuropathic pain. *Brain Research*, 1209(0), 1-7.

- Webber, C. & Zochodne, D. (2010). The nerve regenerative microenvironment: early behavior and partnership of axons and Schwann cells. *Experimental Neurology*, 223(1), 51-59.
- Wiesenfeld-Hallin, Z., Hao, J. X., Xu, X. J. & Hokfelt, T. (1993). Nitric oxide mediates ongoing discharges in dorsal root ganglion cells after peripheral nerve injury. *Journal of Neurophysiology*, 70(6), 2350-2353.
- Woolf, C. J. & Decosterd, I. (1999). Implications of recent advances in the understanding of pain pathophysiology for the assessment of pain in patients. *Pain*, 82, Supplement 1(0), 141-147.
- Woolf, C. J. & Mannion, R. J. (1999). Neuropathic pain: aetiology, symptoms, mechanisms, and management. *The Lancet*, 353(9168), 1959-1964.
- Woolf, C. J. & Salter, M. W. (2000). Neuronal plasticity: increasing the gain in pain. *Science*, 288(5472), 1765-1769.
- Woolf, C. J., Bennett, G. J., Doherty, M., Dubner, R., Kidd, B., Koltzenburg, M., Lipton, R., Loeser, J. D., Payne, R. & Torebjork, E. (1998). Towards a mechanism-based classification of pain? *Pain*, 77(3), 227-229.
- Wu, G., Ringkamp, M., Hartke, T. V., Murinson, B. B., Campbell, J. N., Griffin, J. W. & Meyer, R. A. (2001). Early onset of spontaneous activity in uninjured C-fiber nociceptors after injury to neighboring nerve fibers. *The Journal of neuroscience: the official Journal of the Society for Neuroscience*, 21(8), 140.
- Xian, M., Ito, K., Nakazato, T., Shimizu, T., Chen, C.K., Yamato, K., Murakami, A., Ohigashi, H., Ikeda, Y. & Kizaki, M. (2007). Zerumbone, a bioactive sesquiterpene, induces G2/M cell cycle arrest and apoptosis in leukemia cells via a Fas- and mitochondria-mediated pathway. *Cancer Science*, 98(1), 118-126.
- Yob, N. J., Jofrry, S. M., Affandi, M. M. R. M. M., Teh, L. K., Salleh, M. Z. & Zakaria, Z. A. (2011). *Zingiber zerumbet* (L.) Smith: A Review of Its Ethnomedicinal, Chemical, and Pharmacological Uses. *Evidence-Based Complementary and Alternative Medicine*, 2011, 12.
- Yoon, M. H. & Yaksh, T. L. (1999). Evaluation of interaction between gabapentin and ibuprofen on the formalin test in rats. *Anesthesiology*, 91(4), 1006-1013.
- Zaeoung, S., Plubrukarn, A. & Keawpradub, N. (2005). Cytotoxic and free radical scavenging activities of Zingiberaceous rhizomes. *Songklanakarin Journal of Science and Technology* 27: 799-812
- Zakaria, Z. A., Mohamad, A. S., Chear, C. T., Wong, Y. Y., Israf, D. A., & Sulaiman, M. R. (2010). Antiinflammatory and antinociceptive activities of Zingiber zerumbet methanol extract in experimental model systems. *Medical Principles and Practice*, 19(4), 287-294. Zimmermann, M. (1983). Ethical guidelines for investigations of experimental pain in conscious animals. *Pain*, 16(2), 109-110.

Zimmermann, M. (2001). Pathobiology of neuropathic pain. *European Journal of Pharmacology*, 429(1–3), 23-37.

Zuo, Y., Perkins, N. M., Tracey, D. J. & Geczy, C. L. (2003). Inflammation and hyperalgesia induced by nerve injury in the rat: a key role of mast cells. *Pain*, 105(3), 467-479.

