



UNIVERSITI PUTRA MALAYSIA

***CHARACTERIZATION OF Enterococcus faecium AND Enterococcus faecalis
CLINICAL ISOLATES IN A MALAYSIAN HOSPITAL.***

WENG POH LENG

FPSK(M) 2013 46



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By

WENG POH LENG

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of Master of Science

December 2013

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

CHARACTERIZATION OF *Enterococcus faecium* AND *Enterococcus faecalis* CLINICAL ISOLATES IN A MALAYSIAN HOSPITAL.

By

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December 2013

Chair: Assoc. Prof. Malina Osman, PhD

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Enterococcus faecium and *Enterococcus faecalis* have been well documented as ubiquitous, Gram-positive cocci, opportunistic nosocomial pathogens. In recent decades, they have been recognized as the primary agent for nosocomial infections in many countries worldwide. The aims of the study were to determine the molecular characterization of *E. faecium* and *E. faecalis* isolated from patients in a Malaysian hospital focused on enterococcal surface protein (*esp*) gene, biofilm formation, association of *esp* gene with antibiotic resistance and genetic relatedness amongst the isolates collected. Samples were collected from a local tertiary hospital from May 2009 to March 2010 and subjected to characterization via biochemical identification, antibiotic susceptibility tests and DNA extraction. PCR was performed for genotypic identification and *esp* gene detection and MLST of isolates. Association of biofilm with the presence of *esp* gene was also examined with crystal violet assay. Enzymatic digestion with *Sma*I and PFGE typing were used to determine genetic relatedness of some strains. In this study, *E. faecalis* (n=52) was found to be most

commonly isolated amongst 80 isolates followed by *E. faecium* (n=28). The higher resistance rates were exhibited by *E. faecium* in decreasing order: tazobactam-piperacillin (96.4%), ampicillin (92.9%), high-level gentamicin (89.3%) and penicillin (82.1%). Whereas *E. faecalis* demonstrated slightly lower resistance rates: high-level gentamicin (25.0%), penicillin (7.7%), ampicillin (1.9%) and tazobactam-piperacillin (1.9%) respectively. No vancomycin and teicoplanin resistant enterococci was found. The prevalence of *esp* gene was found higher in *E. faecium* (78.5%) compared to *E. faecalis* (46.2%). However, the prevalence of this gene was more predominantly found in isolates that were resistant to ampicillin (74.1%) and tazobactam-piperacillin (65.8%). Ampicillin-resistant strains and *esp* gene were strongly associated with the genetic clustering in clonal complex-17. A significant strength of relationship between *esp* gene and biofilm formation was strongly observed in *E. faecium* than in *E. faecalis*. Some isolates without the *esp* gene were also found to form biofilms and these findings suggest *esp* might play a significant role although multiple factors might also be involved apart from environmental conditions. PFGE typing revealed high genetic diversity of enterococcal isolates and no indication of outbreaks. Clonally related ST types (ST6, ST16, ST28, *E. faecalis*; ST17, ST 78, ST203, *E. faecium*), which are circulating globally and two new ST types (ST399, *E. faecalis*; ST601, *E. faecium*) were obtained via MLST. ST6, ST16, ST28 and ST179 of *E. faecalis* were documented and of particular concern ST6 is associated with clonal-complex 2, a representative of hospital adapted complexes and the most often reported amongst hospital isolates ST type. ST type of *E. faecium*: ST17, ST78 and ST203, have been widely linked to CC17, which is associated worldwide spread and hospital outbreaks. The conclusion drawn from this study is that *E. faecium* exhibited high resistance rate, which is expected and observed in other countries. Strong

association of *esp* gene with biofilm formation in *E. faecium* would suggest *esp* gene as a potential marker for line-associated infections. In addition, detection of several ST types in both species should prompt proper surveillance system to be carried out in the near future.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PERINCIAN *Enterococcus faecium* DAN *Enterococcus faecalis* ISOLAT KLINIKAL DARIPADA SATU HOSPITAL DI MALAYSIA.

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Enterococcus faecium dan *Enterococcus faecalis* telah didokumentasikan sebagai bakteria Gram positif cocci yang lasak dan sentiasa ada di mana jua. Bakteria ini merupakan patogen nosokomial opportunis yang biasa diperolehi dan dalam beberapa dekad kebelakangan ini telah menjadi penyebab utama jangkitan nosokomial di pelbagai negara seluruh dunia. Objektif kajian ini adalah untuk menyiasat ciri-ciri molekular *E. faecium* dan *E. faecalis* yang diperoleh daripada pesakit di sebuah hospital di Malaysia yang fokuskan kepada gen enterococcal surface protein (*esp*), pembentukan biofilem, perhubungan antara gen *esp* dengan rintangan antibiotic dan perkaitan genetic antara sampel. Sampel dikutip daripada sebuah hospital tertier tempatan di Malaysia dari Mei 2009 sehingga Mac 2010 dan sampel diperiksa melalui pengesahan biokimia, ujian rintangan terhadap antibiotic dan pengeskrakan DNA. PCR juga dilakukan pada isolat untuk pengesahan genotipik dan gen *esp* serta MLST. Kaitan antara biofilem dan kehadiran gen *esp*

dikaji melalui cerakin kristal ungu (crystal violet assay). Pencernaan enzim dengan *Sma*I dan analisa PFGE digunakan untuk mengkaji perkaitan antara strain. Didapati *E. faecalis* (n=52) merupakan isolat yang paling sering ditemui daripada 80 isolat yang dikaji dan diikuti oleh *E. faecium* (n=28). Kadar rintangan antibiotik mengikut susunan menurun yang dipamerkan oleh *E. faecium* adalah: tazobactam-piperacillin (96.4%), ampicillin (92.9%), gentamicin aras tinggi (89.3%) and penicillin (82.1%). *E. faecalis* pula menunjukkan kadar rintangan yang lebih rendah iaitu: gentamicin aras tinggi (25.0%), penicillin (7.7%), ampicillin (1.9%) and tazobactam-piperacillin (1.9%) masing-masing. Tiada rintangan terhadap antibiotik vancomycin dan teicoplanin dijumpai di dalam enterococci. Prevalens gen *esp* didapati lebih tinggi pada *E. faecium* (78.5%) daripada *E. faecalis* (46.2%). Tetapi, prevalens gen ini didapati lebih banyak tertumpu di isolat yang rintang kepada ampicillin (74.1%) dan tazobactam-piperacillin (65.8%). Bakteria yang jenis rintang kepada ampicillin dan gen *esp* dikait rapat dengan penggugusan genetik pada klonal kompleks-17. Didapati bahawa terdapat hubungan yang ketara dan kuat antara gen *esp* dan pembentukan biofilem pada *E. faecium* berbanding dengan *E. faecalis*. Sesetengah isolat tanpa gen *esp* didapati boleh membenruk biofilem dan penemuan ini mencadangkan gen *esp* memainkan peranan yang penting, namun pelbagai faktor terlibat dalam proses pembentukan biofilem selain faktor persekitaran. Jenis ST (ST6, ST16, ST28, *E. faecalis*; ST17, ST 78, ST203, *E. faecium*) yang berhubung-kait secara klonal yang tersebar secara global, serta menemui dua jenis ST yang baru (ST399, *E. faecalis*; ST601, *E. faecium*) melalui MLST. ST6, ST16, ST28 dan ST179 daripada *E. faecalis* telah didokumentasikan di dalam pangkalan data MLST di mana ST6 dihubungkaitkan dengan klonal kompleks-2, iaitu wakil gugusan kompleks yang telah beradaptasi dengan hospital, dan merupakan jenis ST yang

paling kerap dilaporkan di kalangan isolat hospital. Dalam analisis MLST *E. faecium* pula, ST17, ST78 dan ST203 terkenal sebagai kumpulan utama klonal kompleks-17 yang dihubungkaitkan dengan penularan jangkitan dan peletusan di hospital. Kesimpulannya, daripada kajian ini, *E. faecium* menunjukkan kadar rintangan yang tinggi seperti mana dijangka dan diperhatikan di negara-negara lain. Hubungkait yang ketara antara gen *esp* dan pembentukkan biofilem mencadangkan gen *esp* mempunyai potensi sebagai penanda untuk jangkitan berkaitan saluran parenteral intravena. Tambahan pula, pengesanan beberapa jenis ST di dalam kedua-dua spesis sepatutnya mengingatkan supaya sistem pengawasan yang sepatutnya dilaksanakan pada masa akan datang.

I certify that a Thesis Examination Committee has met on **2nd December 2013** to conduct the final examination of **Weng Poh Leng** on her thesis entitled "**Characterization of *Enterococcus faecium* and *Enterococcus faecalis* clinical isolates in a Malaysian Hospital**" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

%	percentage
bp	base pair
DNA	deoxyribonucleic acid
EDTA	ethylene diamine tetraacetic acid
h	hour
kb	kilobase
kg	kilogram
min	minute
MLST	multi locus sequence typing
mm	milimetre
mM	milimolar
mg	miligram
ml	mililitre
ng	nanogram
nm	nanometer
°C	degree celcius
PBS	phosphate buffered saline
PCR	polymerase chain reaction
PFGE	pulsed-field gel electrophoresis
RNA	ribonucleic acid
s	seconds
ST	sequence type
TE	Tris-EDTA
TBE	tris-borate EDTA

Taq	<i>Thermus aquaticus</i>
U	unit
μl	microlitre
μg	microgram
UV	ultra violet
V	volt
v/v	volume per volume
w/v	weight per volume



CHAPTER 1

GENERAL INTRODUCTION

1.1 Introduction

Enterococci are mainly isolated from patients with bacteremia, urinary tract infections or soft skin tissue infections and many more. In Malaysia, resistant strains of enterococci specifically vancomycin-resistant enterococci (VRE) have been isolated from animal and food sources (Radu *et al.*, 2001). The first case of hospital-acquired VRE was reported in 1996 by Riley, Parasakthi and Teh (Riley *et al.*, 1996).

Two main species have been identified related to the enterococcal infections in humans accounting for 75% and 25% prevalence such as *Enterococcus faecalis* (*E. faecalis*) and *Enterococcus faecium* (*E. faecium*) respectively (Shankar *et al.*, 1999). Nosocomial infections caused by enterococci have been reported to demonstrate an ascending trend of high morbidity and mortality, with the incidence of bacteremia has increased as much as 77% in 1980s (Lautenbach, *et al.*, 1999). Enterococcal surface protein (esp) which was firstly discovered by Shankar *et al.* (1999) revealed this novel surface protein was a significant enrichment in infection-derived isolates. In a study conducted by Valdezate *et al.* (2009) the esp gene was likely shown to be present in *E. faecium* and *E. faecalis* related infections. These enriched esp *E. faecium* isolates were likely to be in the Clonal Complex-17 (CC17) group, a clonal complex group of which highly associated with hospital outbreaks and it is part of the worldwide vancomycin-resistant enterococci (VRE) epidemic clone (Khan *et al.*, 2008).

Strain typing of bacterial pathogens with increased virulence and/or transmissibility and antibiotic resistance (example of *E. faecium*, Clonal Complex-17 group) not only raised concerns but also highlighted the necessity to have effective identification methods of these strains and track their spread (Enright and Spratt, 1999). The current investigations encompassing the interface of taxonomy, genetics, evolutionary and epidemiology with the use of molecular typing such as multi-locus sequence typing (MLST) was to illustrate the importance of genetic variability in a single microbial species. And the examination of either gene polymorphism or genome has implications for the identification of microbial genetic types (Belkum et al., 2001).

There are still insufficient data to elucidate the current *E. faecium* and *E. faecalis* infections in Malaysian hospitals in comparison to other countries (Abadía-Patiño, 2010). Therefore the aims of this study were to determine the molecular characterisation of *E. faecium* and *E. faecalis* isolated from patients in a Malaysian hospital with special focus on enterococcal surface protein (esp) gene, biofilm formation, association of esp gene with antibiotic resistance and genetic relatedness amongst the isolates collected in the study.

1.2 General objectives

- i) To determine the molecular characteristics of *Enterococcus faecium* and *Enterococcus faecalis* isolated from patients by Pulsed-Field Gel Electrophoresis and Multi Locus Sequence Typing in a Malaysian hospital.

1.3 Specific objectives

- i) To investigate the demographic of isolates, prevalence of enterococcal related infections and part of the microbiological characteristics (antibiotic profiles) of enterococci isolated from patients with enterococci related infections.
- ii) To determine the prevalence of esp gene (associated with antibiotic profiles) among the isolates.
- iii) To examine the association between esp gene and biofilm formation.
- iv) To study the genetic relatedness among the isolates.

REFERENCES

- Abadía-Patiño, L. (2010). Prevalence of Resistant Enterococci in Developing Countries. In *Antimicrobial Resistance in Developing Countries* (pp. 233-247). New York: Springer.
- Arciola, C.A., Baldassarri, L., Campoccia, D., Creti, R., Pirini, V., Huebner, J. & Montanaro, L. (2008). Strong biofilm production, antibiotic multi-resistance and high *gelE* expression in epidemic clones of *Enterococcus faecalis* from orthopaedic implant infections. *Biomaterials*. 29(5): 580-586.
- Arias, C.A. & Murray B.E. (2012). The rise of the Enterococcus: beyond vancomycin resistance. *Nature Review Microbiology*. 10(4): 266-278.
- Baldassarri, L., Bertuccini, L., Ammendolla, M. G., Gherardi, G. & Creti, R. (2001). Variant *esp* gene in vancomycin-sensitive *Enterococcus faecium*. *Lancet*. 357: 1802.
- Bens, C.C., Voss, A. & Klaassen, C.H. (2006). Presence of a novel DNA methylation enzyme in methicillin-resistant *Staphylococcus aureus* isolates associated with pig farming leads to uninterpretable results in standard pulsed-field gel electrophoresis analysis. *Journal of Clinical Microbiology*. 44: 1875–1876.
- Bidet, P., Lalande, V., Salauze, B., Burghoffer, B., Avesani, V., Delmee, M., Rossier, A., Barbut, F. & Petit, J.C. (2000). Comparison of PCR-ribotyping, arbitrarily primed PCR, and pulse-field gel electrophoresis for typing *Clostridium difficile*. *Journal of Clinical Microbiology*. 38: 2484-2487.
- Billström, H., Lund, B., Sullivan, Á., & Nord, C.E. (2008). Virulence and antimicrobial resistance in clinical *Enterococcus faecium*. *International Journal Antimicrobial Agents*. 32: 374-377.
- Billström, H., Top, J., Edlund, C. & Lund B. (2009). Frequent occurrence of multidrug-resistant CC17 *Enterococcus faecium* among clinical isolates in Sweden. *Journal of Applied Microbiology*. 108: 1810-1816.
- Birren, B. & Lai, E. (1993). Pulse Field Gel Electrophoresis, A Practical Guide, California: Academic Press Inc.
- Borgmann, S., Schulte, B., Wolz, C., Gruber, H., Werner, G., Goerke, C., Klare, I., Heeg, P. & Autenrieth I.B. (2007). Discrimination between epidemic and non-epidemic glycopeptide-resistant *E. faecium* in a post-outbreak situation. *Journal of Hospital Infection*. 67: 49-55.
- Bonten, M.J.M., Willems, R. & Weinstein, R.A. (2001). Vancomycin-resistant enterococci: why are they here, and where do they come from? *The Lancet, Infectious Disease*. 1: 314-325.

- Butaye, P., Devriese, L.A., Goossens, H., Ieven, M. & Haesebrouck F. (1999). Enterococci with Acquired Vancomycin Resistance in Pigs and Chickens of Different Age Groups. *Antimicrobial Agents and Chemotherapy*. 43: 365-366.
- Brown, M.R.W., Allison, D.G. & Gilbert, P. (1988). Resistance of bacterial biofilms to antibiotics: a growth-rate related effect? *Journal of Antimicrobial and Chemotherapy*. 22(6) :777-780.
- Caplin, J.L., Hanlon, G.W. & Taylor, H.D. (2008). Presence of vancomycin and ampicillin-resistant *Enterococcus faecium* of epidemic clonal complex-17 in wastewaters from the south coast of England. *Environmental Microbiology*. 10(4), 885–892.
- Cetinkaya, Y., Falk, P. & Mayhall, G. (2000). Vancomycin-Resistant Enterococci. *Clinical Microbiology Reviews*. 13(4): 686-707.
- Chouchani, C., Salabi, A.E., Marrakchi, R., Ferchichi, L. & Walsh, T.R. (2011). First report of mefA and msrA/msrB multidrug efflux pumps associated with bla_{TEM-1} β-lactamase in *Enterococcus faecalis*. *International Journal of Infectious Disease*. doi:10.1016/j.ijid.2011.09.024
- Chow, J.W. (2000). Aminoglycoside resistance in enterococci. *Clinical Infectious Disease*. 31: 586-589.
- Chu, G., Vollrath, D., & Davis, R.W. (1986). Separation of large DNA molecules by contour-clamped homogenous electric fields. *Science*. 241: 1582-1585.
- Clinical and Laboratory Standards Institute (CLSI) (2012). *Performance Standards for Antimicrobial Susceptibility Testing, Seventeenth Informational Supplement M100-S17*, vol. 27, CLSI, Wayne, Pa, USA.
- Coburn, P.S. & Gilmore, M.S. (2003). The *Enterococcus faecalis* cytolysin: a novel toxin active against eukaryotic and prokaryotic cells. *Cellular Microbiology*. 5(10): 661-669.
- Coque, T.M., Willems, R., Cantón, R., Del Campo, R., & Baquero, F. (2002). High occurrence of esp among ampicillin-resistant and vancomycin-susceptible *Enterococcus faecium* clones from hospitalized patients. *Journal of Antimicrobial Chemotherapy*. 50(6): 1035-1038.
- Coque, T.M., Willems, R.J.L., Fortun, J., Top, J., Diz, S., Loza, E., Canton, R. & Baquero, F. (2005). Population structure of *Enterococcus faecium* Causing Bacteremia in a Spanish University Hospital: Setting the Scene for a Future Increase in Vancomycin Resistance? *Antimicrobial Agents and Chemotherapy*. 49(7): 2693-2700.
- Couvarlin, P. (2006). Vancomycin Resistance in Gram-Positive Cocci. *Clinical Infectious Diseases*. 42: S25-34.

- Costerton, J.W., Stewart, P.S. & Greenberg, E.P., (1999). Bacterial Biofilms: A Common Cause of Persistent Infections. *Science*. 284:1318-1322.
- Damborg, P., Top, J., Hendrickx, A.P.A., Dawson, S., Willems, R.J.L. & Guardabassi, L. (2009). Dogs Are a Reservoir of Ampicillin-Resistant *Enterococcus faecium* Lineages Associated with Human Infections. *Applied and Environmental Microbiology*. 75(8): 2360–2365.
- D'Azevedo, P. A., Dias, C. A. G., & Teixeira, L. M. (2006). Genetic diversity and antimicrobial resistance of enterococcal isolates from southern region of Brazil. *Revista do Instituto de Medicina Tropical de São Paulo* 48(1): 11–16.
- Deshpande, L., Fritsche, T.R., Moet, G.J., Biedenbach, D.J. & Jones, R.N. (2007). Antimicrobial resistance and molecular epidemiology of vancomycin-resistant enterococci from North America and Europe: a report from the SENTRY antimicrobial surveillance program. *Diagnostic Microbiology and Infectious Disease*. 58(2): 163-170.
- Devriese, L., Baele, M. & Butaye, P. (2006). The Genus Enterococcus: Taxonomy. *Prokaryotes*. 4:163-174.
- Donlan, R.M. (2002). Biofilms: Microbial Life on Surfaces. *Emerging Infectious Diseases*. 8(9): 881-890.
- Donlan, R.M. & Costerton, J.W. (2002). Biofilms: Survival Mechanisms of Clinically Relevant Microorganisms. *Clinical Microbiology Reviews*. 15(2): 167-193.
- Dutka-Malen, S., Blaimont, B., Wauters, G & Courvalin, P. (1994). Emergence of high-level resistance to glycopeptides in *Enterococcus gallinarum* and *Enterococcus casseliflavus*. *Antimicrobial Agents and Chemotherapy*. 38: 1675- 1677.
- Dworniczek, E., Wojciech, L., Sobleszczanska, B. & Seniuk, A. (2005). Virulence of *Enterococcus* isolates collected in Lower Silesia (Poland). *Scandinavia Journal of Infectious Disease*. 37: 630-636.
- Eaton, T.J. & Gasson, M.J. (2002). A variant enterococcal surface protein Espfm in *Enterococcus faecium*; distribution among food, commensal, medical, and environmental isolates. *FEMS Microbiology Letters*. 216: 269-275.
- Enright, M.C. & Spratt B.G. (1999) Multilocus sequence typing. *Trends In Microbiology*. 7(12): 482-487.
- Feil, E., Li, C.B., Aanensen, D.M., Hanage, W.P. & Spratt, B.G. (2004). eBURST: Inferring Patterns of Evolutionary Descent among Clusters of Related Bacterial Genotypes from Multilocus Sequence Typing Data. *Journal of Bacteriology*. 186(5): 1518-1530.

Feil, E. & Chan, M. (2001). The BURST algorithm (Based Upon Related Sequence Types) www.pubmlst.org/analysis/burst/burst.shtml
(Accessed 21 March 2012)

Fisher, K. & Phillips, C. (2009). The ecology, epidemiology and virulence of *Enterococcus*. *Microbiology*. 155: 1749-1757.

Freitas, A.R., Novais, C., Ruiz-Garbajosa, P., Coque, T.M. & Peixe, L. (2009). Dispersion of Multidrug-Resistant *Enterococcus faecium* Isolates Belonging to Major Clonal Complex in Different Portuguese Settings. *Applied and Environmental Microbiology*. 75(14): 4904-4908.

Garsin, D.A., Sifri, C.D., Mylonakis, E., Qin, X., Singh, K.V., Murray, B.E., Calderwood ,S.B. & Ausubel, F.M. (2001). A simple model host for identifying Gram-positive virulence factors. *Proceedings of National Academy of Sciences*. 98: 10892-10897.

Ge, Y., MacDonald, D., Hait, H., Lipsky, B., Zasloff, M. & Holroyd, K. (2002). Microbiological profile of infected diabetic foot ulcers. *Diabetic Medicine*. 19:1032-1035

Getachew, Y.M., Hassan, L., Zakaria, Z., Saleha, A.A., Kamaruddin, M.I. & Che Zalina, M.Z. (2009). Characterization of vancomycin-resistant *Enterococcus* isolates from broilers in Selangor, Malaysia. *Tropical Biomedicine* 26(3): 280–288.

Gutschik, E., Moller, S. & Christensen, N. (1979). Experimental endocarditis in rabbits, Significance of the proteolytic capacity of the infecting strains of *Streptococcus faecalis*. *Acta Pathologica Microbiologica Scandinavica*. 87:353-362.

Goering, R.V., Dockrell, H.M., Wakelin, D., Zuckerman, M., Chiodini, P.L., Roitt, I.M. & Mims, C. (2008). Mims' Medical Microbiology (4th edition) Philadelphia: Mosby-Year Book Europe Ltd.

Goering, R.V. (2010). Pulsed field gel electrophoresis: A review of application and interpretation in the molecular epidemiology of infectious disease. *Infection, Genetics and Evolution*. 10: 866–875.

Gram, C. (1884). Über die isolirte Farung der Schizomycetin in Schnitt-und Trockenpräparaten. *Fortschritte der Medizin*. 2:185–189.

Hällgren, A., Abednazari, H., Ekdahl, C., Hanberger, H., Nilsson, M., Samuelsson, A., Svensson, E., Nilsson, L.E. & the Swedish ICU Study Group. (2001) Antimicrobial susceptibility patterns of enterococci in intensive care units in Sweden evaluated by different MIC breakpoints systems. *Journal of Antimicrobial Chemotherapy*. 48: 53-62.

- Hällgren, A., Claesson, C., Saeedi, B., Monstein, H., Hanberger, H. & Nilsson, L.E. (2009). Molecular detection of aggregation substance, enterococcal surface protein, and cytolytic genes and in vitro adhesion to urinary catheters of *Enterococcus faecalis* and *Enterococcus faecium* of clinical origin. *International Journal of Medical Microbiology*. 299: 323-332.
- Hanage, W.P., Feil, E.J., Brueggemann, A.B. & Spratt, B.G. (2004). *Multilocus Sequence Typing: Strain Characterization, Population Biology, and Patterns of Evolutionary Descent In Molecular Microbiology: Diagnostic Principles and Practice*. (Eds. Persing, D.H., Tenover, F.C., Versalovic, J., Tang, Y., Unger, E.R., Relman, D.A. and White, T.J.) Washington, D.C.: ASM Press.
- Heikens, E., Bonten, M.J.M. & Wilems, R.J.L. (2007). Enterococcal surface protein Esp is important for biofilm formation of *Enterococcus faecium* E1162. *Journal of Bacteriology*. 189: 8233-8240.
- Hollingshead, S. & Vapnek, D. (1985). Nucleotide sequence analysis of a gene encoding a streptomycin/spectinomycin adenyltransferase. *Plasmid*. 13: 17-30.
- Homan, W.L., Tribe, D., Poznanski, S., Li, M., Hogg, G., Spalburg, E., van Embden, J.D.A. & Willems, R.J.L. (2002). Multilocus Sequence Typing Scheme for *Enterococcus faecium*. *Journal of Clinical Microbiology*. 40: 1963- 1971.
- Hunter, S.B., Vauterin, P., Lambert-Fair, M.A., Van Duyne, M.S., Kubota, K., Graves, L., Wrigley, D., Barrett, T. & Ribot, E. (2005). Establishment of a Universal Size Standard Strain for Use with the PulseNet Standardized Pulsed-Field Gel Electrophoresis Protocols: Converting the National Databases to the New Size Standard. *Journal of Clinical Microbiology*. 43(3): 1045-1050.
- Huycke, M. M., Sahm, D. F. & Gilmore, M. S. (1998). Multiple-drug resistant enterococci: the nature of the problem and an agenda for the future. *Emerging Infectious Disease*. 4: 239-249.
- Jolley, K.A. & Maiden, M.C.J. (2010). BIGSdb: Scalable analysis of bacterial genome variation at the population level. *BMC Bioinformatics*. 11:595.
- Kariyama, R., Mitsuhashi, R., Chow, J.W., Clewell, D.B. & Kumon, H. (2000). Simple and Reliable Multiplex PCR Assay for Surveillance Isolates of Vancomycin-Resistant Enterococci. *Journal of Clinical Microbiology*. 38(8): 3092-3095.
- Kam, K.M., Luey, C.K. Tsang, Y.M., Law, C.P., Chu, M.Y., Cheung, T.L.. & Chiu, A.W. (2003). Molecular subtyping of *Vibrio cholerae* O1 and O139 by pulsed-field gel electrophoresis in Hong Kong: correlation with epidemiological events from 1994 to 2002. *Journal of Clinical Microbiology*. 41: 4502-4511.
- Karmarkar, M.G., Gershom, E.S., Kaul, S., Mankeshwar, A.A. & Mehta, P.R. (2005). In *Study of Drug Resistance in Clinical Isolates of Enterococci with Special*

Reference to High Level Aminoglycoside Resistance, β Lactamase Production and Lateral Transfer of Drug Resistance – New Insights Into an Old Enemy, Proceedings of the 15th Lancefield International Symposium on Streptococci and Streptococcal Diseases, Palm Cove, Australia, 25-29 September 2005. International Congress Series Streptococci, Volume 289 , pp. 111-114, 2006.

Khan, M.A., van der Wal, M., Farell, D.J., Cossins, L., van Belkum, A., Alaidan, A. & Hays, J.P. (2008). Analysis of VanA Vancomycin-resistant *Enterococcus faecium* isolates from Saudi Arabian hospitals reveals presence of clonal cluster 17 and two new Tn 1546 lineage types. *Journal of Antimicrobial Chemotherapy*. 62: 279-283.

Kirkwood, B.R. & Sternem J.A.C. (2003). *Essential Medical Statistics*, 2nd Edition, USA: Blackwell Science Ltd.

Klma, C.L., Alexander, T.W., Selinger, L.B., Read, R.R., Shewan, P.E., Gow, S.P., Booker, C.W. & McAllister, T.A. (2010). Comparison of repetitive PCR and pulsed-field gel electrophoresis for the genotyping of *Mannheimia haemolytica*. *Journal of Microbiological Methods*. 81: 39–47.

Kreft, B., Marre, R., Schramm, U. & Wirth, R. (1992). Aggregation substance of *Enterococcus faecalis* mediates adhesion to cultured renal tubular cells. *Infection and Immunity*. 60:25-30.

Kühn, I.L., Burman, G., Haeggman, S., Tullus, K. & Murray, B.E. (1995). Biochemical fingerprinting compared with ribotyping and pulse-field gel electrophoresis of DNA for epidemiological typing of enterococci. *Journal of Clinical Microbiology*. 33: 2812-2817.

Lautenbach, E., Bilker, W.B. & Brennan, P.J. (1999). Enterococcal Bacteremia: Risk Factors for Vancomycin Resistance and Predictors of Mortality. *Infection Control and Hospital Epidemiology*. 20(5): 318-323.

Leavis, H.L., Willems, R.J.L., Top, J., Spalburg, E., Mascini, E.M., Fluit, A.C. Hoepelman, A., de Neeling, A.J. & Bonten, M.J.M. (2003). Epidemic and Nonepidemic Multidrug-Resistant *Enterococcus faecium*. *Emerging Infectious Diseases*. 9(9): 108-1115.

Leavis, H., Top, J., Shankar, N., Borgen, K., Bonten, M., van Embden, J. & Willems, R.J.L. (2004). A Novel Putative Enterococcal Pathogenicity Island Linked to the *esp* Virulence Gene of *Enterococcus faecium* and Associated with Epidemicity. *Journal of Bacteriology*. 186(3): 672-682.

Leavis, H.L., Bonten, M.J.M. & Willems, R.J.L. (2006). Identification of high-risk enterococcal clonal complexes: global dispersion and antibiotic resistance. *Current Opinion in Microbiology*. 9(5): 454-460.

Levinson, W. (2004). *Medical Microbiology and Immunology, Examination and Board Review*, 8th edition, San Francisco : The McGraw-Hill Companies.

Lloyd, S., Zervos, M., Mahayni, R., & Lundstrom, T. (1998). Risk factors for enterococcal urinary tract infection and colonization in a rehabilitation facility. *American Journal of Infection Control*. 26(1):35–39.

Magiorakos, A.P., Srinivasan, A., Carey, R.B., Carmelli, Y., Falagas, M.E., Giske, C.G., Harbath, S., Hindler, J.F., Kahlmeter, G., Olsson-Liljequist, B. et al. (2012). Multidrug-resistant, extensively drug-resistant and pandrug-resistant bacteria: an international expert proposal for interim standard definitions for acquired resistance. *Clinical Microbiology and Infection*. 18(3): 268-281.

Mahon, C.R., Lehman D.C. & Manuselis, G. (2007). *Textbook of Diagnostic Microbiology*. St. Louis: Saunders.

Mato, R., Almeida, F., Pires, R., Rodrigues, P., Ferreira, T. & Santos-Sanches, I. (2009). Assessment of high-level gentamicin and glycopeptide-resistant *Enterococcus faecalis* and *E. faecium* clonal structure in a Portuguese hospital over a 3-year period. *European Journal of Clinical Microbiology and Infectious Disease*. 28: 855-859.

Marraffini, L.A. & Schneewind, O. (2006). Targeting proteins to the cell wall of sporulating *Bacillus anthracis*. *Molecular Microbiology*. 62: 1402-1417.

Meredith, K., Bolhuis, A. & O'Neill, M.A.A. (2009). Enterococcal surface protein Esp affects antibiotic sensitivity in *Enterococcus faecium*. *International Journal of Antimicrobial Agents*. 14: 380-393.

Ministry of Health, Malaysia, Annual Report. Information and Documentation Systems Unit: Malaysia, 2009.

Ministry of Health, Malaysia, National Antibiotic Guideline 2008.

Ministry of Health, Malaysia, Annual Report. Medical Development Division: Malaysia, 2006.

Moellering, R.C. Jr & Weinberg, A.N. (1971). Studies on antibiotic synergism against enterococci. *Journal of Clinical Investigation*. 50: 2580-2584.

Mohamed, J.A. & Huang, D.B. (2007). Biofilm formation by enterococci. *Journal of Medical Microbiology*. 56: 1581-1588.

Moreno, M.R.F., Leisner, J.J., Tee, L.K., Ley, C., Radu, S., Rusul, G., Vancanneyt, M. & De Vuyst, L. (2002). Microbial analysis of Malaysian tempeh, and characterization of two bacteriocins produced by isolates of *Enterococcus faecium*. *Journal of Applied Microbiology*. 92: 147-157.

Moses, V., Jerobin, J., Nair, A., Sathyendara, S. & Balaji, V. (2012). Enterococcal Bacteremia is Associated with Prolonged Stay in the Medical Intensive Care Unit. *Journal of Global Infectious Diseases*. 4(1): 26-30.

- Murray, P.R., Baron, E.J., Jorgensen, J.H., Pfaller, M.A. & Yolken, R.H. (2003). *Manual of Clinical Microbiology*, 8th edition, Volume 1, Washington: American Society of Microbiology.
- Oancea, C., Klare, I., Witte, W. & Werner, G. (2004). Conjugative transfer of the virulence gene, *esp*, among isolates of *Enterococcus faecium* and *Enterococcus faecalis*. *Journal of Antimicrobial Chemotherapy*. 54: 232-235.
- Ong, C.H.S., Asaad, M., Lim, K.C. & Ngeow, Y.F. (2002). Infrequent occurrence of vancomycin-resistant enterococci in poultry from Malaysian wet markets *Malaysian Journal of Pathology*. 24(2) : 91 – 94.
- Paulsen, I.T., Banerjee, L., Myers, G.S., Nelson, K.E., Seshadri, R., Read, T.D., Fouts, D.E., et al. (2003). Role of mobile DNA in the evolution of vancomycin- resistant *Enterococcus faecalis*. *Science*. 299(5615) 2071-2074.
- Peterson, L.R. (2005). Squeezing the Antibiotic Balloon: The impact of Antimicrobial Classes on Emerging Resistance. *Clinical Microbiology and Infection*. 11(Suppl. 5): 4-16.
- Pillai, S.K., Sakoulas, G., Eliopoulos, G.M., Moellering, R.C. Jr., Murray, B.E. & Inouye, R.T. (2004) Effects of glucose on *fsr*-mediated biofilm formation in *Enterococcus faecalis*. *Journal of Infectious Disease*. 190: 967-970.
- Poh, C.H., Oh, H.M. & Tan, A.L. (2006). Epidemiology and clinical outcome of enterococcal bacteraemia in an acute care hospital. *Journal of Infection*. 52: 383-386.
- Poole, K. (2004). Resistance to *b*-lactam antibiotics. *Cellular and Molecular Life Sciences*. 61: 2200–2223.
- Raja, N.S., Karunakaran, R., Ngeow, Y. F. & Awang, R. (2005). Community-acquired vancomycin-resistant *Enterococcus faecium*: a case report from Malaysia. *Journal of Medical Microbiology*. 54:901–903.
- Radu, S., Toosa, H., Rahim, R.A., Reezal, A., Ahmad, M., Hamid, A.M., Rusul, G. & Nishibuchi, M. (2001). Occurrence of the *vanA* and *vanC2/C3* genes in *Enterococcus* species isolated from poultry sources in Malaysia. *Diagnostic Microbiology and Infectious Disease*. 39: 145–153.
- Ramadhan, A.A. & Hegedus, E. (2005). Biofilm formation and *esp* gene carriage in enterococci. *Journal of Clinical Pathology*. 58: 685-686.
- Reyes, K., Malik, R., Moore, C., Donabedian, S., Perri, M., Johnson, L. & Zervos, M. (2010). Evaluation of Risk Factors for Coinfection or Cocolonization

with Vancomycin-Resistant Enterococcus and Methicillin-Resistant *Staphylococcus aureus*. *Journal of Clinical Microbiology*. 48(2): 628-630.

Riley, P.A., Parasakthi, N. & The, A. (1996). *Enterococcus faecium* with High Level Vancomycin Resistance Isolated from the Blood Culture of a Bone Marrow Transplant Patient in Malaysia. *Medical Journal of Malaysia*. 51: 383-385.

Romling, U. & Tummler, B. (2000). Achieving 100% typeability of *Pseudomonas aeruginosa* by pulsed-field gel electrophoresis. *Journal of Clinical Microbiology*. 38: 464–465.

Ruiz-Garbazosa, P., Bonten, M. J. M., Robinson, D. A. et al. (2006). Multilocus sequence typing scheme for *Enterococcus faecalis* reveals hospital-adapted genetic complexes in a background of high rates of recombination. *Journal of Clinical Microbiology*. 44(6): 2220–2228.

Son R., Nimita, F., Rusul, G., Nasreldin, E., Samuel, L. & Nishibuchi, M. (1999). Isolation and molecular characterization of vancomycin-resistant *Enterococcus faecium* in Malaysia. *Letters in Applied Microbiology*. 29: 118–122

Salgado, C. D. (2008). The risk of developing a vancomycin-resistant *Enterococcus* bloodstream infection for colonized patients. *American Journal of Infection Control*. 36(10): S175.e5-S175.e8.

Santagati, M., Campanile, F., & Stefani, S. (2012) Genomic diversification of enterococci in hosts: the role of the mobilome. *Frontiers in Microbiology*. Volume 3, Article 95. doi:10.3389/fmicb.2012.00095

Sava, I., Heikens, E. & Huebner, J. (2010). Pathogenesis and immunity in enterococcal infections. *Clinical Microbiology and Infection*. 16 (6): 533–540.

Schwartz, D.C. & Cantor, C.R. (1984). Separation of yeast chromosome-sized DNAs by pulse field gradient gel electrophoresis. *Cell*. 37(1): 67-75.

Shankar, V., Baghdyan, A.S., Huycke, M.M., Lindahl, G. & Gilmore, M.S. (1999). Infection-Derived *Enterococcus faecalis* Strains Are Enriched in esp, a Gene Encoding a Novel Surface Protein. *Infection and Immunity*. 67(1):193-200.

Shankar, N., Baghdyan, A.S. & Gilmore, M.S. (2002). Modulation of virulence within a pathogenicity island in vancomycin- resistant *Enterococcus faecalis*. *Nature*. 417: 746–750.

Solomkin, J.S., Mazuski, J.E., Baron, E.J., Sawyer, R.G., Nathens, A.B., DiPiro, J.T., Buchman, T., Dellinger, E.P., et al. (2003). Guidelines for the selection of Anti-infective Agents for Complicated Intra-abdominal Infections. *Clinical Infectious Diseases*. 37: 997-1005.

- Stephanovic, S., Vukovic, D., Savic, B.D. & Svabic-Vlahovic, M. (2000). A modified microtiter-plate test for quantification of staphylococcal biofilm formation. *Journal of Microbiological Methods*. 40(2): 175-179.
- Suci, P.A., Mittelman, M.W., Yu, F.P. & Geesey, G.G. (1994). Investigation of ciprofloxacin penetration into *Pseudomonas aeruginosa* biofilms. *Antimicrobial Agents and Chemotherapy*. 38: 2125-2133.
- Tacconelli, E. & Cataldo, M.A. (2008). Vancomycin-resistant enterococci (VRE): transmission and control. *International Journal of Antimicrobial Agents*. 31:99-106.
- Talebi, M., Rahimi, F., Katouli, M., Milby, R. & Pourshafie, M.R. (2008). Epidemiological link between wastewater and human vancomycin-resistant *Enterococcus faecium* isolates. *Current Microbiology*. 56(5): 468-473.
- Tenover, F., Arbeit, R.D., Goering, R.V., Mickelsen, P.A., Murray, B.E., Persing, D.H. & Swaminathan, B. (1995). Interpreting chromosomal DNA restriction patterns produced by pulsed-field gel electrophoresis: criteria for bacterial strain typing. *Journal of Clinical Microbiology*. 33(9): 2233-2239.
- Tenover, F. (2006). Mechanisms of Antimicrobial Resistance in Bacteria. *The American Journal of Medicine*. 119 (6A): S3-S10.
- Toledo-Arana, A., Valle, J., Solano, C., Arrizubieta, M.J., Cucarella, C., Lamata, M., Amorena, B., Leiva, J., Penades, J.R. & Lasa, I. (2001). The Enterococcal surface protein Esp, Is Involved In *Enterococcus faecalis* Biofilm Formation. *Applied and Environmental Microbiology*. 67(10): 4538-4545.
- Tomayko, J. F. & Murray, B. E. (1995). Analysis of *Enterococcus faecalis* isolates from intercontinental sources by multilocus enzyme electrophoresis and pulsed-field gel electrophoresis. *Journal of Clinical Microbiology*. 33(11): 2903-2907.
- Top, J., Willems, R. & Bonten, M. (2008). Emergence of CC17 *Enterococcus faecium*: from commensal to hospital-adapted pathogen. *FEMS Immunology and Medical Microbiology*. 52: 297-308.
- Urwin, R. & Maiden, M.C.J. (2003). Multi-locus sequence typing: a tool for global epidemiology. *Trends in Microbiology*. 11(10): 479-487.
- Valdezate, S., Labaryu, C., Navarro, A., Mantecón, M.A., Ortega, M., Coque, T.M., García, M. & Saéz-Nieto, J. (2009). Large Clonal Outbreak Of Multidrug-resistant CC17 ST17 *Enterococcus faecium* containing Tn5382 in a Spanish Hospital. *Journal of Antimicrobial Chemotherapy*. 63: 17-20.
- van Belkum, A., van den Braak, N., Thomassen, R., Verbugh, H. & Endtz, H. (1996). Vancomycin resistant Enterococci in cats and dogs. *Lancet*. 348: 1038-1039.

- van Belkum, A., Struelens M., de Visser, A., Verbugh, H. & Tibayrenc, M. (2001) Role of Genomic Typing in Taxonomy, Evolutionary Genetics, and Microbial Epidemiology. *Clinical Microbiology Review*. 14(3): 547-560.
- van den Braak, N., Power, E., Anthony, R., Endtz, H.P., Verbugh, H.A. & van Belkum, A. (2000). Random Amplification of Polymorphic DNA versus Pulse Field Gel Electrophoresis of *Sma*I DNA Macrorestriction Fragments for Typing Strains of Vancomycin Resistant Enterococci. *FEMS Microbiology Letters*. 192:45-52.
- van Wamel, W.J.B., Hendrickx, A.P.A., Bonten, M.J.M., Top, J., Posthuma, G. & Willems, R.J.L. (2007). Growth Condition-Dependent *Esp* Expression by *Enterococcus faecium* Affects Initial Adherence and Biofilm Formation. *Infection and Immunity*. 75(2): 924-931.
- van Schaik, W. & Willems, R.J.L. (2010). Genomic-based insights into the evolution of enterococci. *Clinical Microbiology and Infection*. 16(6): 527-532.
- Wang, J.T., Chang, S.C., Wang, H.Y., Chen, P.C., Shiau, Y.R. & Lauderdale, T.L. (2013). High Rates of multidrug resistance in *Enterococcus faecalis* and *E. faecium* isolated from inpatients and outpatients in Taiwan. *Diagnostic Microbiology and Infectious Disease*. 75(4): 406-411.
- Weiss, C.A., Statzm C.L., Dahms, R.A., Remucal, M.J., Dunn, D.L. & Beilman, G.J. (1999). Six Years of Surgical Wound Infection Surveillance at a Tertiary Care Center. Review of the Microbiologic and Epidemiological Aspects of 20007 Wounds. *Archives of Surgery*. 134:1041-1048
- Wells, C.L. Moore, E.A., Hoag, J.A., Hirt, H., Dunny, G.M. & Erlandsen, S.L. (2000). Inducible expression of *Enterococcus faecalis* aggregation substance surface protein facilitates bacterial internalization by cultured enterocytes. *Infection and Immunity*. 68: 7190-7194.
- Weng, P.L., Hamat, R.A., Cheah, Y.K., Zainol, N., Aziz, M.N. & Shamsudin, M. N. (2012). Vancomycin-resistant *Enterococcus faecium* of multi locus sequence type 18 in Malaysia. *Medical Journal of Malaysia*. 67: 639-640.
- Werner, G., Coque, T.M., Hammerum, A.M., Hope, R., Hryniwicz, W., Johnson, A., Klare, I., Kristinsson, K.G., Leclercq, R., Lester, C.H., et al. (2008). Emergence and Spread of Vancomycin Resistance among Enterococci in Europe. *Eurosurveillance* 13(47): 1-11.
- Willems, R. J. L., Homan, W., Top, J., van Santen-Verheuvel, M., Tribe, D., Manzioros, X., Gaillard, C., Vandenbroucke-Grauls, C.M.J.E., Mascini, E.M., van Embden, J.D.A. & Bonten, M.J.M (2001). Variant *esp* gene as a marker of a distinct genetic lineage of vancomycin resistant *Enterococcus faecium* spreading in hospitals. *Lancet*. 357: 853–855.
- Willems, R.J.L., Top, J., van Santen, M., Robinson, D.A., Coque, T.M., Baquero, F., Grundmann, H. & Bonten, M.J.M. (2005). Global Spread of Vancomycin-

resistant *Enterococcus faecium* from Distinct Nosocomial Genetic Complex. *Emerging Infections Diseases*. 11(6): 82-828.

Willems, R.J.L. & van Schaik, W. (2009). Transition of *Enterococcus faecium* from commensal organism to nosocomial pathogen. *Future Microbiology*. 4(9): 1125-1135.

Willems, R.J.L., Top J., van Schaik, W., Leavis, H., Bonten, M., Siren, J., Hanage, W.P. & Corander, J. (2012). Restricted Gene Flow among Hospital Subpopulation of *Enterococcus faecium*. *mBio* 3(4): e00151-12.

Woodford, N., Soltani, M. & Hardy, K. J. (2001). Frequency of esp in *Enterococcus faecium* isolates. *Lancet* 358: 584.

Zubaidah, A.W., Ariza, A. & Azmi, S. (2006). Hospital-acquired vancomycin-resistant enterococci: now appearing in Kuala Lumpur Hospital. *Medical Journal of Malaysia*. 61: 487-489.