



UNIVERSITI PUTRA MALAYSIA

***TOXICOLOGY EVALUATIONS OF *Rhaphidophora decursiva* (Roxb.)
Schott EXTRACT IN SPRAGUE DAWLEY***

SITI SURIANI BINTI ARSAD

FPSK(M) 2013 45



**TOXICOLOGY EVALUATIONS OF *Rhaphidophora decursiva* (Roxb.) Schott
EXTRACT IN SPRAGUE DAWLEY**

By

SITI SURIANI BINTI ARSAD

**Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfillment of the Requirements for the Master of Science**

October 2013

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Master of Science

TOXICOLOGY EVALUATIONS OF *Rhaphidophora decursiva* (Roxb.) Schott EXTRACT IN SPRAGUE DAWLEY

By

SITI SURIANI BINTI ARSAD

October 2013

Chairman : Norhaizan Mohd Esa, PhD

Faculty : Medicine and Health Sciences

Rhaphidophora decursiva (Roxb.) Schott has been widely used among Chinese community in Malaysia for treating colon cancer. The study aims to evaluate the toxic effects of the methanol extract of *R. decursiva* after single dose toxicity study (14-day acute toxicity study), repeated 28-day subacute toxicity and 90-day subchronic toxicity study in male Sprague Dawley rats, and also to determine the anti-proliferative activity of *R. decursiva* extract on colon cancer cell lines (HT-29) and normal cell lines (3T3). For toxicity study, rats were divided into 4 groups consisting of 6 rats per group for each acute, subacute and subchronic toxicity evaluations, with a total number of 72 rats. All control groups received distilled water (vehicle). For the acute toxicity, the 3 treatment groups received oral single dose of the plant extract at 700 mg/kg, 2800 mg/kg and 3500 mg/kg, respectively, and sacrificed at day 14 post administration of the plant extract. For subacute toxicity, the 3 treatment groups received daily oral dose of the plant extract at 70 mg/kg, 140 mg/kg and 210 mg/kg for 28 days. As no lethality was observed in subacute toxicity, similar doses were used for the 3 treatment groups in 90-day subchronic toxicity. The toxicity of *R. decursiva* extract was evaluated by the incident of lethality, cage side observations, body weight measurement, absolute and relative weight for kidneys and liver, hematological parameters such as erythrocyte count (RBC), hemoglobin (Hb), hematocrit (Hct), mean corpuscular volume (MCV), mean corpuscular hemoglobin (MCH), mean corpuscular hemoglobin concentration (MCHC), leucocyte count (WBC) and platelet count (Plt), then serum biochemistry parameters such as urea, creatinine (Crea), albumin (Alb), aspartate aminotransferase (AST), alanine aminotransferase (ALT), and alkaline phosphatase (ALP) and histopathological analysis of kidneys and liver. No lethality or adverse toxic signs were seen during the experimental periods in all toxicity studies. Behavior, body weight, absolute and relative organ weight also showed no significant changes in all three

toxicity studies. Hematological analyses revealed no statistically significant differences between groups in the studies. Serum biochemistry analyses showed no significant changes in acute and subchronic toxicity studies. Similar results were observed in subacute toxicity study. For histopathological findings, no significant changes ($p > 0.05$) showed in liver tissue as well as kidneys tissues for all toxicity studies. Anti-proliferative activity of methanol extract of *R. decursiva* (Roxb.) Schott on HT-29 showed the highest sensitivity at 72 hours incubation period with IC_{50} value of 40 $\mu\text{g/mL}$, followed by 80 $\mu\text{g/mL}$ and 260 $\mu\text{g/mL}$, respectively for 48 hours and 24 hours incubation period. Besides, the extract was found not to be cytotoxic on 3T3 (IC_{50} cannot be determined). In conclusion, based on clinical appearance, clinical pathological, organ pathology and histopathological results, the methanol extract of *R. decursiva* did not cause any toxic effects to male Sprague Dawley rats, and the oral lethal dose (LD_{50}) of the extract is more than 3500 mg/kg, while the no-observed-adverse-effect level (NOAEL) for the extract is 210 mg/kg per day for 90 days. Besides, according to the ability of *R. decursiva* extracts to achieve 50% inhibition of HT-29 cell lines, the findings from this study suggest that *R. decursiva* extract is safe to be consumed as no toxic effect was found when tested both in *in vivo* and *in vitro* studies.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan Ijazah Master Sains

PENILAIAN TOKSIKOLOGI EKSTRAK *Rhaphidophora decursiva* (Roxb.) Schott KE ATAS SPRAGUE DAWLEY

Oleh

SITI SURIANI BINTI ARSAD

Oktober 2013

Pengerusi : Norhaizan Mohd Esa, PhD

Fakulti : Perubatan dan Sains Kesihatan

Rhaphidophora decursiva (Roxb.) Schott telah digunakan secara meluas di kalangan masyarakat Cina di Malaysia untuk merawat kanser usus. Kajian ini dijalankan bertujuan untuk menilai kesan ketoksikan oleh ekstrak metanol tumbuhan selepas kajian toksik dos tunggal (kajian ketoksikan akut 14 hari), seterusnya berulang selama 28 hari (ketoksikan subakut) dan kajian ketoksikan subkronik selama 90 hari ke atas tikus jantan Sprague Dawley, serta menentukan anti-proliferatif aktiviti ke atas sel kanser usus (HT-29) dan normal sel (3T3). Untuk kajian tahap ketoksikan, tikus telah dibahagikan kepada 4 kumpulan yang terdiri daripada 6 ekor tikus untuk setiap kumpulan bagi setiap ujikaji ketoksikan akut, subakut dan subkronik, dengan jumlah bilangan 72 ekor tikus. Semua kumpulan kawalan menerima air suling (vehicle). Untuk ketoksikan akut, 3 kumpulan menerima dos tunggal dengan kepekatan ekstrak tumbuhan 700 mg/kg, 2800 mg/kg dan 3500 mg/kg, masing-masing, dan semua tikus tersebut dimatikan pada 14 hari selepas diberikan ekstrak tumbuhan secara oral (suapan). Bagi kajian ketoksikan subakut, 3 kumpulan yang telah diuji dengan menggunakan dos ekstrak tumbuhan pada kepekatan 70 mg/kg, 40 mg/kg dan 210 mg/kg selama 28 hari. Oleh kerana tiada kematian tikus semasa kajian ketoksikan subakut, maka dos yang sama telah digunakan untuk 3 kumpulan dalam ketoksikan subkronik (90 hari). Ketoksikan ekstrak *R. decursiva* telah dikaji melalui bilangan kematian tikus, pemerhatian bagi kelakuan tikus, pengukuran berat badan dan berat organ yang diambil seperti buah pinggang dan hati, keputusan hematologi seperti kiraan eritrosit (RBC), hemoglobin (Hb), hematokrit (Hct), isipadu kopuskular purata (MCV), hemoglobin kopuskular purata (MCH), konsentrasi hemoglobin kopuskular purata (MCHC), kiraan leukosit (WBC) dan kiraan platelet (Plt), seterusnya keputusan serum biokimia seperti urea, kreatinin (Cre), albumin (Alb), aspartate aminotransferase (AST), alanine aminotransferase (ALT), dan alkaline phosphatase (ALP), dan analisis histopatologi ke atas tisu buah pinggang dan hati juga

dilakukan. Keputusan menunjukkan tiada kematian atau tanda-tanda toksik dilihat semasa tempoh eksperimen dalam semua kajian ketoksikan. Pemerhatian dari segi kelakuan, perubahan berat badan dan organ juga menunjukkan tiada sebarang perubahan ketara dalam tiga kajian ketoksikan ini. Hematologi analisis menunjukkan tiada perbezaan yang signifikan di antara kumpulan yang diuji dengan ekstrak *R. decursiva*. Serum analisis biokimia juga tidak menunjukkan sebarang perubahan yang signifikan dalam kajian ketoksikan akut dan subkronik. Keputusan yang sama telah diperhatikan dalam kajian ketoksikan subakut. Untuk keputusan histopatologi, tiada perubahan signifikan ($p>0.05$) diperhatikan dalam tisu buah pinggang serta tisu hati bagi semua kajian ketoksikan. Aktiviti anti-proliferatif *R. decursiva* (Roxb.) Schott ekstrak pada HT-29 menunjukkan kesan paling sensitif dengan nilai IC_{50} sebanyak 40 $\mu\text{g/mL}$ selepas 72 jam tempoh pendedahan terhadap ekstrak. Manakala tempoh pendedahan ekstrak selama 48 jam menunjukkan kesan kurang sensitif ke atas sel HT-29 dengan nilai IC_{50} sebanyak 80 $\mu\text{g/mL}$ diikuti dengan tempoh pendedahan ekstrak selama 24 jam menunjukkan kesan paling kurang sensitif dengan nilai IC_{50} yang tinggi iaitu 260 $\mu\text{g/mL}$. Sebaliknya, kajian yang dijalankan ke atas sel 3T3 menunjukkan ekstrak *R. decursiva* tidak memberi kesan toksik kerana tiada nilai IC_{50} . Kesimpulannya, berdasarkan keputusan klinikal darah, patologi organ dan histopatologi, ekstrak methanol *R. decursiva* tidak menyebabkan sebarang kesan toksik kepada tikus jantan Sprague Dawley, dan nilai 'lethal dose' (LD_{50}) ekstrak *R. decursiva* adalah lebih daripada 3500 mg/kg, manakala tahap 'no-observed-adverse-effect level' (NOAEL) bagi ekstrak *R. decursiva* ialah 210 mg/kg sehari selama 90 hari. Selain itu, mengikut keupayaan ekstrak *R. decursiva* untuk mencapai 50% perencatan terhadap sel HT-29, penemuan daripada kajian ini mencadangkan bahawa ekstrak *R. decursiva* adalah selamat untuk dimakan kerana tiada kesan toksik ditemui apabila diuji secara *in vivo* dan *in vitro*.

ACKNOWLEDGEMENTS

In the name of Allah, the most benevolent and most merciful. I would like to take this opportunity to thank all those who gave great support to me while doing this thesis. First and foremost, I would like to express my deepest gratitude to my main supervisor, Assoc. Prof. Dr. Norhaizan Mohd Esa for her generous guidance, patience, advice and endless support that contributed significantly towards the completion of this project. Her careful reviews and constructive criticism have been crucially important for this thesis.

My sincere gratitude is also accorded to my co-supervisors, Dr. Hazilawati Hamzah and Prof. Fauziah Othman for their constructive advice, priceless comment and invaluable advice throughout the entire course of this research.

Special thanks are expressed to the laboratory staff of Department of Nutrition and Dietetics of Faculty of Medicine and Health Sciences, UPM especially, Mr. Syed Hasbullah Bin Syed Kamaruddin and Mr. Eddy Ghadaffie Bin Jamiauddin for their help, patience and guidance throughout my research study.

Finally, yet importantly, I would like to express my heartiest appreciation to all my family members for their understanding and support throughout my studies. Also, thank you to all my friends and laboratory mates for support, help and guidance.

Above all, I thank God for His mercy and blessing on me.

I certify that a Thesis Examination Committee has met on 29 October 2013 to conduct the final examination of Siti Suriani binti Arsad on her thesis entitled "Toxicology Evaluations of *Rhaphidophora decursiva* (Roxb.) Schott Extract in Sprague Dawley" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Asmah binti Rahmat, PhD

Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Chairman)

Sharida binti Fakurazi, PhD

Associate Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Internal Examiner)

Loh Su Peng, PhD

Associate Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Internal Examiner)

Nor Fadilah Rajab, PhD

Associate Professor
Universiti Kebangsaan Malaysia
Malaysia
(External Examiner)



NORITAH OMAR, PhD

Associate Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 21 January 2013

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree. The members of the Supervisor Committee are as follows:

Norhaizan Mohd Esa, PhD

Associate Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Chairman)

Fauziah Othman, PhD

Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

Hazilawati Hamzah, PhD

Senior Lecturer
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

DECLARATION

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustration and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia (Research); Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: 27th March 2014

Name and Matric No.: **Siti Suriani Binti Arsad (GS29459)**

Declaration by Members of Supervisory Committee

This is to confirm that:

- The research conducted and the writing of this thesis was under our supervision;
- Supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____ Signature: _____

Name of
Chairman of
Supervisory
Committee: Associate Professor
Dr. Norhaizan Mohd
Esa

Name of
Member of
Supervisory
Committee: Professor
Dr. Fauziah Othman

Signature: _____

Name of
Member of
Supervisory
Committee: Dr. Hazilawati Hamzah

TABLE OF CONTENTS

	Page
ABSTRACT	iii
ABSTRAK	v
ACKNOWLEDGEMENTS	vii
APPROVAL	viii
DECLARATION	x
LIST OF TABLES	xv
LIST OF FIGURES	xvi
LIST OF ABBREVIATIONS	xvii
CHAPTER	
1	
INTRODUCTION	1
1.1 Background	1
1.2 Problem statements	3
1.3 Significant of the study	4
1.4 Objectives	5
1.4.1 General objectives	5
1.4.2 Specific objectives	5
1.5 Limitations of the study	5
2	
LITERATURE REVIEW	6
2.1 Herbal medicine	6
2.2 Chinese traditional herbal medicine	8
2.2.1 Raphidophora plants	9
2.3 Cancer	11
2.4 Anti-proliferative study	12
2.5 Toxicity study	13
2.5.1 Acute toxicity study	13
2.5.2 Subacute toxicity study	14
2.5.3 Subchronic toxicity study	14
2.6 Serum biochemistry analysis	15
2.6.1 Liver function test	15
2.6.1.1 Alanine Aminotransferase (ALT)	15
2.6.1.2 Aspartate Aminotransferase (AST)	16
2.6.1.3 Alkaline Phosphatase (ALP)	16
2.6.1.4 Albumin	17
2.6.2 Renal function test	17
2.6.2.1 Creatinine and urea	17
2.7 Hematology parameters	18
2.7.1 Erythron parameters	18
2.7.2 Leucocytes	18
2.7.3 Platelets	19

3	MATERIALS AND METHODS	20
	3.1 Materials	20
	3.1.1 Instruments	20
	3.1.2 Sample, chemicals and reagents	20
	3.2 Methodology	21
	3.2.1 Sample preparation	21
	3.2.1.1 <i>Rhaphidophora decursiva</i> (Roxb.) Schott sample	21
	3.2.1.2 Sample extraction	21
	3.2.2 <i>In vivo</i> study	21
	3.2.2.1 Location of study	21
	3.2.2.2 Experimental animals	22
	3.2.2.3 Experimental designs	22
	3.2.2.4 Toxicity study	24
	3.2.2.4.1 Acute toxicity	24
	3.2.2.4.2 Subacute toxicity	24
	3.2.2.4.3 Subchronic toxicity	24
	3.2.2.5 Cage side observation	24
	3.2.2.6 Body weight measurement	24
	3.2.2.7 Hematology analysis	25
	3.2.2.8 Serum biochemistry analysis	25
	3.2.2.9 Histopathology	25
	3.2.3 <i>In vitro</i> study	26
	3.2.3.1 Cell culture	26
	3.2.3.2 Thawing	26
	3.2.3.3 Subculture	26
	3.2.3.4 Plating	26
	3.2.3.5 Treatment	27
	3.2.3.6 MTT assay	27
	3.2.4 Data Interpretation (Statistical Evaluation)	28
4	RESULTS	29
	4.1 Acute toxicity study (14 days)	29
	4.1.1 Cage side observation and body weight measurement	29
	4.1.2 Hematology analysis	30
	4.1.3 Serum biochemistry analysis	33
	4.1.4 Histopathology analysis	37
	4.2 Subacute toxicity study (28 days)	44
	4.2.1 Cage side observation and body weight measurement	44
	4.2.2 Hematology analysis	45
	4.2.3 Serum biochemistry analysis	50
	4.2.4 Histopathology analysis	54
	4.3 Subchronic toxicity (90 days)	56
	4.3.1 Cage side observation and body weight measurement	56
	4.3.2 Hematology analysis	57
	4.3.3 Serum biochemistry analysis	63
	4.3.4 Histopathology analysis	67

	4.4 Anti-proliferative study (MTT assay)	71
5	DISCUSSIONS	73
	5.1 <i>In vivo</i> study	73
	5.1.1 Toxicity studies	73
	5.1.1.1 Cage side observation, body weight and Organ measurement	74
	5.1.1.2 Hematology analysis	75
	5.1.1.3 Serum biochemistry analysis	76
	5.1.1.4 Histopathology analysis	77
	5.2 <i>In vitro</i> study	79
	5.2.1 Anti-proliferative study	79
6	CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	82
	6.1 Conclusion	82
	6.2 Recommendations	82
	REFERENCES	84
	APPENDICES	100
	BIODATA OF STUDENT	104
	LIST OF PUBLICATIONS	105

LIST OF TABLES

Table		Page
2.1	Herbal products from different cultures	7
3.1	The doses for acute, subacute and subchronic toxicity study	10
4.1	Organ weights for acute toxicity study	30
4.2	WBC, RBC, Hb and Hct values of experimental rats for acute toxicity study	31
4.3	MCV, MCH, MCHC and Plt values of experimental rats for acute toxicity study	33
4.4	ALT, AST and ALP counts of experimental rats for acute toxicity Study (14 days)	34
4.5	Cre, Alb and Urea values of experimental rats for acute toxicity Study (14 days)	36
4.6	Lesion scores of kidney and liver for all groups analyzed using Kruskal-Wallis test (14 days)	38
4.7	Results of Mann-Whitney U test for comparisons between groups for lesions in the organs (14 days)	39
4.8	Organ weights for subacute toxicity study	43
4.9	WBC, RBC, Hb and Hct values of experimental rats for subacute toxicity study	45
4.10	MCV, MCH, MCHC and Plt values of experimental rats for subacute toxicity study	47
4.11	ALT, AST and ALP counts of experimental rats for subacute toxicity study (28 days)	49
4.12	Cre, Alb and Urea values of experimental rats for subacute toxicity study (28 days)	51
4.13	Lesion scores of kidney and liver for all groups analyzed using Kruskal-Wallis test (28 days)	53
4.14	Results of Mann-Whitney U test for comparisons between groups for lesions in the organs (28 days)	54
4.15	Organ weights for subchronic toxicity study	57
4.16	WBC, RBC, Hb and Hct values of experimental rats for subchronic toxicity study (90 days)	59
4.17	MCV, MCH, MCHC and Plt values of experimental rats for subchronic toxicity study (90 days)	62
4.18	ALT, AST and ALP counts of experimental rats for subchronic toxicity study (90 days)	64
4.19	Cre, Alb and Urea values of experimental rats for subchronic toxicity study (90 days)	66
4.20	Lesion scores of kidney and liver for all groups analyzed using Kruskal-Wallis test (90 days)	68
4.21	Results of Mann-Whitney U test for comparisons between groups for lesions in the organs (90 days)	69

LIST OF FIGURES

Figure	Page
2.1 <i>Rhaphidophora decursiva</i> (Roxb.) Schott plant	10
4.1 Body weight changes in experimental rats for acute toxicity study	29
4.2 Histological section of kidney tissue for group C (section stained with H&E, x400)	40
4.3 Histological section of kidney tissue and liver tissue for group C (section stained with H&E, x400).	41
4.4 Body weight changes in experimental rats for subacute toxicity study	42
4.5 Histological section of kidney tissue and liver tissue for group D (section stained with H&E, x400).	55
4.6 Body weight changes in experimental rats for subchronic toxicity study	56
4.7 Histological section of kidney tissue for group D (section stained with H&E, x400) and liver tissue for group D (section stained with H&E, x200).	70
4.8 Percentage of viability of HT-29 cell lines against concentration of <i>R. decursiva</i> extract	71
4.9 Percentage of viability of 3T3 cell lines against concentration of <i>R. decursiva</i> extract	72

LIST OF ABBREVIATIONS

µL	microlitre
µmol/L	micromole per litre
ACF	aberrant crypt foci
ACS	American Cancer Society
ACUC	Animal Care Use Committee
AIDS	Acquired immunodeficiency syndrome
Alb	albumin
ALP	alkaline phosphate
ALT	alanine aminotransferase
ANOVA	analysis of variance
AST	aspartate aminotransferase
ATCC	American Type Culture Collection
ATP	adenosine triphosphate
CC	cellular cast
CM	conservative medicine
CO ₂	carbon dioxide
Cre	creatinine
DHI	5,6- dihydroxyindole
dL	decilitre
DMEM	dulbecco's modified eagle medium
DMSO	dimethyl sulfoxide
EDTA	ethylenediaminetetraacetic Acid
EEC	European Economic Community
FBS	fetal bovine serum
fL	femtoliters
FRIM	Forest Research Institute of Malaysia
GHS	Globally Harmonized System
g	gram
g/L	gram per litre
GC	granular cast
Hb	hemoglobin
Hct	hematocrit
CPV	cytoplasmic vacuolation
IARC	International Agency for Research on Cancer
L	litre
LD ₅₀	50% lethal dose
MCH	mean corpuscular hemoglobin
MCHC	mean corpuscular hemoglobin concentration
MCV	mean corpuscular volume
mg/kg	milligram/kilogram
mL	millilitre
mmol/L	milimole per litre

MTT	(3-[4,5-dimethylthiazol-2-yl]-2-5-diphenyltetrazoliumbrimide)
NCR	National Cancer Registry
NCSM	National Cancer Society of Malaysia
NCCAM	National Center for Complementary and Alternative Medicine
NOAEL	no-observable adverse effect levels
NOEL	no-observable effect
OECD	Organization for Economic Cooperation and Development
PBS	phosphate buffer saline
P	protein cast
Pg	picogram
Plt	platelet count
RBC	red blood cell
RDW	red blood cell distribution width
Rpm	revolutions per minute
RPMI	Roswell Park Memorial Institute
S.E.M	standard error of mean
SARS	Severe Acute Respiratory Syndrome
SPSS	Statistical Package of Social Science
SW	sinusoidal widening
U/L	units/litre
UK	United Kingdom
UKM	Universiti Kebangsaan Malaysia
UMS	Universiti Malaysia Sarawak
UNESCO	United Nations Educational, Scientific and Cultural Organization
UPM	Universiti Putra Malaysia
UTM	Universiti Teknologi Malaysia
USA	United State of America
USM	Universiti Sains Malaysia
WBC	white blood cell
WHO	World Health Organization

CHAPTER 1

INTRODUCTION

1.1 Background

Plants and herbs have been used since the earliest of times to cure various ailments. By the middle of the nineteenth century, about 80% of all medicines were derived from herbs. Generally, herbs refer to yearly, seasonal or biennial seed-producing, non-woody plants that die at the end of each harvest season. The National Center for Complementary and Alternative Medicine (NCCAM) (2011) defined herb as a plant or part of a plant that is used for its smell, taste or healing characteristics. The flowers, stems, seeds or roots of a plant can bring benefits in medical or aromatherapy attributes. McCann (2004) stated that generally, health care experts acknowledge herbs as a raw drug that can be used to treat infections, avoid illnesses or sustain good health. Nonetheless, herbs can be made into extracts, drinks, teas, pills, oils and shampoos.

McCann (2003) also reported that herbs have significance in current day biomedical antidotes. In a quarter of biomedical medications regularly prescribed nowadays, at least one active ingredient comes from plants and the rest of the substances are chemically produced in laboratories. Many medical herbs have long been used as anticancer agents (Tiwari *et al.* 2000; Duong Van Huyen *et al.* 2003). Recent evidence from studies by Prozeski *et al.*, (2001), Kawada *et al.* (2001) and Moalic *et al.* (2001) reported that a number of natural products originated from plants have antitumor properties. Besides this, a study by Wojdylo *et al.* (2007) has also shown that herbs promote antioxidant activity, digestion, anti-inflammatory, anti-microbial, prevent genetic mutation effects and have anti-cancer agents.

Herbal medicine are substance that come from one or more plants and brings healing qualities or health benefits to human in forms of raw or processed ingredients. Herbal medicine as the major remedy in traditional medical systems, have been used in medical practice for thousands of years and have made a great contribution to maintain human health. World Health Organisation (WHO) (1993) informed that a majority of the world's population in developing nations still depend on herbal medicines to fulfill its wellbeing needs. A study by MacManus (2008) has shown that many herbal medicines are effective since they contain pharmacologically active substance derived from plants. Li *et al.* (2009) explained that extracts from the herbs are often used in line with long-established cancer treatments to improve survival rate and quality of life because it is found to be cheaper than conventional treatments. Ali *et al.* (2005) disclosed that in Malaysia and Indonesia, factory-made traditional herbal antidotes such as the popular 'jamu' and 'makjun' are easily available and taken in frequently, up to 3 times daily to promote good health. Furthermore, the Indonesian 'jamu' is also available in Singapore, Australia and Holland where there is a large community of Indonesians (Ali *et al.* 2005).

Joshi & Kaul (2001) further added that traditional herbal medicines are gaining popularity globally and these products purchases have raised significantly. This is due to the fact that research has reported that essences from several herbal medicines or blend have an anticancer potential *in vitro* or *in vivo*. Various extracts either water extract or alcohol extract of herbs that are used by researchers have shown that there are some anti-proliferative effect on cancer cell lines. Instead, earlier studies also have shown that there are some antioxidant mechanisms in herbs that act as an anticancer agent (Prasad *et al.* 1999). However currently, poison related to herbal drugs is becoming more of a concern in terms of their increasing intake in developed nations. WHO (2000) and International Agency for Research on Cancer (IARC) (2002) pointed out that appropriate instructions, guidelines and assessment have been published to maintain the safe practice of these products. Saad *et al.* (2006) revealed that the usage of a number of herbal and herbal derived goods could result in a hepatic toxicity. Thus, herbal scientists or manufacturers must understand the association between pharmacological reactions and the chances of the herbs dynamic compounds interacting with other drugs when both are taken together.

Timbrell (2000) put forward that researches in toxicology play a significant role in pharmaceutical sciences in which it uncovers the needed comprehension on how the human body works and drugs interactions within it. Salsburg (1986) defined the LD₅₀ value as the approximation of the potency of toxicant or poisons to subjects which results in fifty percent of lethality and generally stated in mg/kg body weight. However, other conditions such as growth of cancer and organ damage resulted from substances are not measured. This is because there is doubt in the forecast of other outcomes such as change of enzyme activity and organ impairment based on the LD₅₀ value. A comprehensive evaluation of blood serum tests such as liver and kidney function test, hematology and histopathology, food and water intake need to be performed on subjects to investigate the occurrence of secondary stage of toxicity resulted by chemicals. Information from a toxicity study will be used as reference for the selection of doses for repeated-dose study, presenting initial detection of target organ toxicity or any contrary effects after consumption.

Rhaphidophora decursiva (Roxb.) Schott is used widely in the herbal and traditional medicinal preparations especially among the Chinese community. They believe that this plant can be used to treat colon cancer and they commonly consumed it in the form of herbal tea. However, there are still no scientific studies that have been done to prove the effectiveness of this plant in treating colon cancer. Therefore, many studies need to be done especially in terms of toxicity studies before it can be promoted for utilization or be commercialized. Information from a toxicity study will be a useful tool for the indication of doses for repeated-dose study, offering initial detection of target organ toxicity or any harmful effects after use.

1.2 Problem statements

Herbal medicines have been widely utilized as effective remedies for the prevention and treatment of multiple health conditions for centuries by almost every known culture. The first documented records of herbal medicine use date back 5,000 years (NCCAM, 2005) in China. Similarly, India's Ayurvedic medicine tradition is thought to be more than 5,000 years old and herbal medicine remain an essential component of its practice (Astin *et al.* 2000). Today, the populations of certain countries still depend on herbal medicines to address their healthcare needs.

Worldwide it is estimated that 80% of the population uses herbs; in the developing world rates could be as high as 95% (Nahin *et al.* 2007). WHO estimates that the global market is approximately US \$83 billion annually (Nahin *et al.* 2007). The use of herbal medicines continues to expand globally, in parallel to an increasing acceptance of herbal remedies by consumers. Despite the fact that herbal remedies are not classified as drugs by the United States (US) Food and Drug Administration (FDA) Dietary Supplements Health and Education Act, 1994, the attitude of the general population toward herbal medicine is that this kind of therapy is natural and therefore safe (Zaffani *et al.* 2006). So despite the potential for harmful side effects (De Smet, 1995) and interactions with conventional drugs (Williamson, 2003; Chavez *et al.* 2006), natural products are often taken on a self-medication basis, without the advice of pharmacists or physicians (Eisenberg *et al.* 1998). The safety and efficacy of many herbal medicines used remain essentially unknown. There are limited clinical trials to determine efficacy and safety of traditional herbal medicines. This lack of research does not impede most from using them, given that these remedies are often grounded in long standing cultural traditions.

Similarly, Chinese community in Malaysia believes that *R. decursiva* is effective in curing colon cancer but there is also no scientific researches have been done before. In prior research, it was found that *R. decursiva* actually contains antimalarial compounds by performing antimalarial bioassay-directed fractionation, which has led to the isolation of 'decursivine', a new active indole alkaloid, from the leaves and stems of *R. decursiva* (Zhang *et al.* 2002). There are possibilities of toxicity effect present due to the long term use and unpredictable amounts of the substance that produces the therapeutic effect (Li *et al.* 2009). Saper *et al.* (2008) reported that 20% of Ayurvedic medicines purchased via the Internet contained detectable levels of lead, mercury, and arsenic. Herbs that have caused major adverse events include creosote bush (hepatotoxicity) (Slade & Keating, 2007) and kava (hepatotoxicity) (Kwon *et al.* 2006). Using the proper parts of the plant and the appropriate process for obtaining the ingredients could prevent toxicity, as seen in kava-induced toxicity (White *et al.* 2007).

1.3 Significant of the study

Natural products, especially plants, have been used for the treatment of various diseases for thousands of years (Shoeb, 2006). Although many research-related medicinal plants have been made over the past 30 years, a large number of plants still have not been studied (Kakuko *et al.* 2005). Malaysia have various kind of plants used for medicinal purposes. There are lots of herb plants surviving in the Malaysian rain forest. Some familiar and unfamiliar species allow the researchers to study more in terms of their beneficial aspect to human health.

The substance of the organic components as well as the effects such as anti-inflammatory and anticancer activities in plants may differ. Wu *et al.* (2002) stated that some studies revealed that substances in natural products bring positive outcomes in cancer treatment when weighed against with chemotherapy or existing hormonal therapies. Furthermore, Lee *et al.* (2004) added that specifically, oriental therapeutic plants are believed to be one of the main potential sources due to their variation in types and purposes.

Nowadays, most research has focused on certain plant species that have the potential benefits to the human body, and until now, many other species under the same family group also have not been undiscovered as to bringing beneficial effect on health. For instance, some studies have discovered the potential of *Rhaphidophora* species that belong to the *Araecea* family in the prevention and curative abilities for certain health problems. This species may include *Rhaphidophora korthalsii*, *Rhaphidophora glauca*, *Rhaphidophora elliptica*, *Rhaphidophora cryptantha*, *Rhaphidophora pertusa*, *Rhaphidophora decursiva*, *Rhaphidophora hookeri*, *Raphidophora hongkongensis*, and many more. Each of them has their own biological active components that play an important role as a curing agent for certain health problems. To date, only a few studies have focused on this species since they are rarely found and not yet commercialized including *R. decursiva* which is also known as Pa Shu Long or Shan Shu Long. This plant can be grouped as unfamiliar species that needs to be investigated for their bioactive compound.

In view of the complexity of herbal medicines and their inherent biological variations, it is necessary to evaluate their safety, efficacy, and quality (Castro *et al.* 2009). With the increasing usage of herbal therapies, significant concerns have been raised over the lack of quality control and scientific evidence of the efficacy and safety of these agents (Firenzuoli & Gori, 2007). In particular, safety of herbal prescriptions has become an increasingly important issue (Tang *et al.* 2008) and is generally evaluated via toxicological assessment of all medicinal plant materials. *R. decursiva* which is rarely known among people will increase in value due to the known safety dosage through this study and it can be used for further research especially anticancer study.

1.4 Objectives

1.4.1 General objective

To study the toxicology evaluations of *Rhaphidophora decursiva* (Roxb.) Schott extract on Sprague Dawley rats.

1.4.2 Specific objectives

1. To determine the effect of doses and time on physical haematological, biochemical and histopathological of liver and renal functions.
2. To evaluate the toxicological potential of the plant extract on liver and kidney.
3. To determine the IC₅₀ value of *Rhaphidophora decursiva* (Roxb.) Schott extract on the proliferation of normal cell line (3T3) and colon cancer (HT-29) cell line.

Limitations of the study

In the light of the fact that this plant can be found in different regions of the country as well as in other countries, however the plant samples that were used in this study were only collected in one selected region in Malaysia which may not represent the same species of the plant found in other parts of the world and thus, cannot be generalized by other researchers. Besides this, the plant samples may not represent the whole part of the plant because the study was only carried out on the leaves part. Therefore, the assumptions for the effect of the whole plant cannot be obtained. In addition to this, there are several aspects that may affect the quality of the samples which may include the transportation, processing and storage that had caused the samples not to be really fresh.

REFERENCES

- Adeneye, A.A., Ajagbonna, O.P., Adeleke, T.I. & Bello, S.O. (2006). Preliminary toxicity and phytochemical studies of the stem bark aqueous extract of *Musangacecropioides* in rats. *J. Ethnopharmacol.* 105; 374-379.
- Alade, G.O., Akanmu, M.A., Obuotor, E.M., Osasan, S.A., & Omobuwajo, O.R. (2009). Acute and oral subacute toxicity of methanolic extract of *Bauhinia monandra* leaf in rats. *Afr. J. Pharm. Pharmacol.* 3; 354-358.
- Al-Mamary, M, Al-Habori M, Al-Aghbari A.M, & Baker M.M (2002). Investigation into the toxicological effects of *Catha edulis* leaves: a short term study in animals. *Phytother. Res.* 16:127 – 132.
- Ali, N., Hashim, N. H., Saad, B., Safan, K., Nakajima, M., & Yoshizawa, T. (2005). Evaluation of a method to determine the natural occurrence of aflatoxins in commercial traditional herbal medicines from Malaysia and Indonesia. *Food Chem Toxicol*, Vol. 43, Dec, pp. 1763-1772, 0278-6915
- Akanmu, M.A., Iwalewa, E.O., Elujoba, A.A. & Adelusola, K.A. (2004). Toxicity potentials of *Cassia fistula* fruits as laxative with reference to senna. *Afr. J. Biomed. Res.*7; 23-26.
- Aniagu, S.O., Nwinyi, F.C., Olanubi, B., Akumka, D.D., Ajoku, G.A., Izebe, K.S., Agala, P., Agbani, E.O., Enwerem, N.M., Iheagwara, C., & Gamaniel, K.S. (2004). Is *Berlina grandiflora* (Leguminosae) toxic in rats? *Phytomedicine.* 11; 352–360.
- Asante-Duah, K. Public Health Risk Assessment for Human Exposure to Chemicals (illustrated ed.); Kluwer Academic Publishers: Dordrecht, The Netherlands, 2002; Volume 6.
- Ashafa A.O.T, Yakubu M.T., Grierson D.S. & Afolayan A.J. (2009). Effects of aqueous leaf extract from the leaves of *Chrysocoma ciliate* L. on some biochemical parameters of Wistar rats. *Afr. J. Biotechnol.*, 8: 1425-1430.
- Astin, J.A, Pelletier, K.R, Marie, A. & Haskell, W.L. (2000). Complementary and alternative medicine use among elderly persons: One-year analysis of a Blue Shield Medicare supplement. *J Gerontol Biol Sci Med Sci* 55(1):M4–M9.
- Aziz, Z. & Tey, N.P. (2009). Herbal medicines: Prevalence and predictors of use among Malaysian adults. *Complementary Ther. Med.*, 17: 44-50.

- Balan P, Han K.S, Rutherford S.M, Singh H. & Moughan P.J (2009). Orally administered ovine serum immunoglobulins influence growth performance, organ weights, and gut morphology in growing rats. *J. Nutr* 139:244–249.
- Ballmer P.E, Walshe D., McNurlan M.A, Watson H., Brunt P.W. & Garlick P.J. (1993). Albumin synthesis rates in cirrhosis: correlation with Child–Turcotte classification. *Hepatology*. 18:292–297.
- Bertam, J. S. (1999). Carotenoids and gene regulation. *Nutritional Review* 57(6): 182-191.
- Bird R.P. (1995). Role of aberrant crypt foci in understanding the pathogenesis of colon cancer. *Cancer Lett* 93:55 –71.
- Boullata, J.L., & Nace, A.M. (2000). Safety issues with herbal medicine. *Pharmacotherapy*, 257-69.
- Boyce, P. C. & J. Bogner. 2000. An account of neotenic species of *Rhaphidophora* Hassk. (Araceae-Monsteroideae-Monstereae) in New Guinea and Australia. *Gard Bull Singapore* 52:89–100.
- Boyce, P.C. & Bogner, J. (2000). An account of neotenic species of *Rhaphidophora* Hassk. (Araceae-Monsteroideae-Monstereae) in New Guinea and Australia. *Gard Bull Singapore* 52: 91-93.
- Boyce, P.C. (2000). The genus *Rhaphidophora* Hassk. (Araceae-Monsteroideae-Monstereae) in the southern and western Indonesian archipelago. *Gard Bull Singapore* 52: 101–183.
- Boyce, P.C. (2001). The genus *Rhaphidophora* Hassk. (Araceae-Monsteroideae-Monstereae) in Borneo. *Gard Bull Singapore* 53: 19–74.
- Boyer, T.D., Manns, M.P., & Sanyal, J.A (2011). *Hepatology: A text book of liver disease*, sixth ed. Philadelphia: Elsevier Inc. pp. 206-220.
- Cardellina, J.H., Fuller R.W., Gamble, WR, Westergaard., C., Boswell, J., Munro, M.H.G., Currens, M. & Boyd, M. (1999). Evolving strategies for the selection dereplication and prioritization of antitumor and HIV-inhibitory natural products extracts. In: Bohlin, L., Bruhn, J.G. (Eds.), *Bioassay Methods in Natural Product Research and Development*. Kluwer Academic Publishers Dordrecht, pp. 25–36
- Carol, S.A. (1995). Acute, Subchronic and Chronic Toxicology. In *CRC Handbook of Toxicology*; Michael, J.D., Manfred, A.H., Eds.; CRC Press Inc.: Boca Raton, FL, USA, pp. 51-104.

- Castro, L.S., Perazzo, F.F. & Maistro, E.L. (2009). Genotoxicity testing of *Ambelania occidentalis* (Apocynaceae) leaf extract *in vivo*. *Genet Mol Res.* 8; 440–7.
- Chan, K. (2003). Some aspects of toxic contaminants in herbal medicines. *Chemosphere*, 52: 1361-1371.
- Chan, T.Y.K. (1997). Monitoring the safety of herbal medicine. *Drug Safety* 17: 209–215.
- Chaves, C.B., & Silva, R.A. (1998). Contribuição ao do laboratório de patologia clínica aplicada da antibioticoterapia. In: Silva, P. (Ed.), *Farmacologia. Guanabara Koogan, Rio de Janeiro*, pp. 949–958.
- Cheng, Y. L., Chang, W. L., Lee, S. C., Liu, Y.G., Chen, C.J., Lin, S.Z., Tsai, N. M., Yu, D. S., Yen, C. Y. & Harn, H. J. (2004). Acetone extract of *Angelica sinensis* inhibits proliferation of human cancer cells via inducing cell cycle arrest and apoptosis. *Life Sci* 75(13): 1579-1594.
- Corns, C.M. (2003). Herbal remedies and clinical biochemistry. *Ann Clin Biochem* 40:489-507.
- Crook, M.A. (2006). *Clinical Chemistry and Metabolic Medicine*. 7th ed. London: Hodder Arnold.
- De Smet, P.A.G.M. (1995). Health risks of herbal remedies. *Drug Saf.* 3:81–93.
- Degott C, & Potet F. (1984). Peliosis hepatitis and sinusoidal dilatation. *Arch Anat Cytol Pathol.* 32:296–300.
- Demma, J., Gebre-Mariam, T., Asres, K., Evgetie, W. & Engindawork, E. (2006). Toxicological study on *Glinus lotoides*: a traditionally used technical herb in ethiopia. *J. Ethnopharmacol* 111: 451–457.
- Desai, A.K. & Grossberg, G.T. (2003). Herbals and botanicals in geriatric psychiatry, *Am J Geriatr Psychiatry.* 11: 498-506.
- Diallo, A., Gbeassor, M., Vovor, A., Eklugadegbeku, K. & Aklikokou, K. (2008). Effect of *Tectona grandis* on phenylhydrazine-induced anaemia in rats. *Fitoterapia*, 79: 332-336.
- Duong Van Huyen, J.P., Delignat, S. & Kazatchkine, M.D. (2003). Comparative study of the sensitivity of lymphoblastoid and transformed monocyte cell lines to the cytotoxic effect of *Viscum album* extracts of different origin. *Chemotherapy*, 49:298-302.

- Dybing, E., Doe, J., Groten, J., Kleiner, J. & O'Brien, J. (2002). Hazard characterization of chemicals in food and diet: dose response, mechanism and extrapolation issues. *Food Chem Toxicol* 42: 237-282.
- Eaton, D.L. & Klaassen, C.D. (1996). Principles of toxicology. In: Klaassen, C.D.(Ed.), Casarett and Doull's Toxicology: *The Basic Science of Poisons*, 5th ed. McGrawHill, p. 13.
- Edmondsun, J. M., Armstrong, L. S. & Martinez, A. O. (1988). A rapid and simple MTT-based spectrophotometric assay for determining drug sensitivity in monolayer cultures. *J Tissue Cul Meth* 11(1): 15-17.
- Edward, M.B., Maynard, I.C. & McNair Scott, T.F. (1941). Serum protein concentration as a guide to the treatment of dehydration in diarrheal diseases. *J Pediatr* 18(6): 709-726.
- Ernst, E. (1998). Harmless herb. A review of the recent literature. *Am J Med* 104: 170–178.
- Everds, N. (2004). Chapter 17: Hematology of the mouse. In *The Laboratory Mouse (Handbook of Experimental Animals)*. Hedrich, H. J., & Bullock, G. (Eds.) Elsevier Academic Press: London, UK.
- Farah Dina, P., Hazilawati, H., Rosly, S.M., Shanmugavelu, S. & Noordin, M.M. (2011). Expression of circulating CD146 associated with endovascular dysfunction in adenine-induced chronic renal failure in rats using an EvaGreen real-time RT-PCR assay. *Pertanika J Trop Agric Sci* 34(2), 381-391.
- Firenzuoli, F. & Gori, L. (2007). Herbal medicine today: clinical and research issues. *Evid Based Complement Alternat Med.* 4; 37–40.
- Florian, M.F., Claude, A.J., Markus, A. & Urs, T., (1999). The MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide] assay is a fast and reliable method for colometric determination of fungal cell densities. *Appl Environ Microbiol* 65(8): 3727-3729.
- Gerard, C.C.L. (2002). Overview of cancer in Malaysia. *Jpn J Clin Oncol* 32(1):37-42.
- Gorman, C. (1992). The Power of Potions, *Time*, April 20, p.52 – 53.
- Gregus, Z. & Kiwassen, C. (1995) Mechanism of toxicity. In: Klaassen C.D. (Ed). *The Basic Science of Poisons*, 5th ed. New York, McGraw-Hill 35-74.
- Hagerstrand, I. (1975). Distribution of alkaline phosphatase activity in healthy and diseased human liver tissue. *Acta Path Microbiol Scand A* 83:519-526.

- Halim, S.Z., Abdullah, N. R., Afzan, A., Abdul Rashid, B. A., Jantan, I. & Ismail, Z., (2011). Acute Toxicity Study of Carica papaya leaf extract in Sprague Dawley rats. *J Med Plants Res* 5: 1867- 1872.
- Hall, R.B. (2001): Principles of clinical pathology for toxicology studies. In: principles and methods of toxicology. 4th edition by A. Wallace Hayes, Tyler and Francis Philadelphia. pp.1001-1034.
- Harizal, S.N., Mansor, S.M., Hasnan, J.; Tharakan, J.K.J. & Abdullah, J. (2010). Acute toxicity study of the standardized methanolic extract of *Mitragyna speciosa* Korth in Rodent. *J Ethnopharmacol* 131: 404-409.
- Harris, R.B., Zhou J., Youngblood, B.D., Rybkin, I.I., Smagin, G.N. & Ryan, D.H. (1998). Effect of repeated stress on body weight and body composition of rats fed low- and high-fat diets. *Am J Physiol Reg-I* 275:1928–1938.
- Hazelwood, R.L. & Wilson, W.O. (1962). Comparison of the hematological alterations induced in the pigeon and rat by fasting and heat stress. *Comparative Biochem Physiol* 7: 211-219.
- Hazilawati, H., Abdullah, M., Hutheyfa, A.H., Rosly, S.M., Jasni, S., Noordin, M.M. & Shanmugavelu, S. (2009a). High concentration of *N*-Methyl-*N*-Nitrourea (MNU) induced intravascular haemolytic anaemia rather than leukaemia-lymphoma in Sprague Dawley rats. In *Proceedings of the 1st Malaysian Association of Veterinary Pathology Conference* (pp. 95-99). Harbour View Hotel, Kuching, Sarawak: Malaysian Association of Veterinary Pathology.
- Hazilawati, H., Farah Dina, P., Rosly, S.M., Shanmugavelu, S. & Noordin, M.M. (2010). *Phyllanthus niruri* reduces renal azotaemia in rats induced with chronic renal failure. *Med J Malays* 65:132-134.
- Hazilawati, H., Rosly, S.M., Tarmizi, A. S., Subramanian, K., Johaimi, J., Nurul Syakirah, M.H., Rasedee, A. & Shanmugavelu, S. (2009b). Comparison of blood urea nitrogen and serum creatinine in gentamicin-induced nephrotoxicity in rats and mice. In *Proceedings of the 21st Veterinary Association Malaysia Scientific Congress* (pp. 346-349). The Legend Water Hotel, Port Dickson, Negeri Sembilan: Veterinary Association Malaysia.
- Hilaly, J., El Israili, Z.H. & Lyoussi, B. (2004). Acute and chronic toxicological studies of *Ajuga iva* in experimental animals. *J Ethnopharmacol* 91:43–50.
- Hillaire, S., Bonte E. & Deninger, M.H. (2002). Idiopathic non-cirrhotic intrahepatic portal hypertension in the West: a reevaluation in 28 patients. *Gut* 51:275–280.
- Hoareau, L. & DaSilva, E. J. (1999). Medicinal plants: a re-emerging health aid. *Electron J Biotechn* 2(2): 56-70

- Huang, D.J., Lin, C.D., Chen, H.J. & Lin, Y. H. (2004). Antioxidant and antiproliferative activities of water spinach (*Ipomoea aquatic Forsk*) constituents. *Bot Bull Acad Sinica* 46: 99-106.
- Huang, D.J., Chen, H.J., Lin, C.D. & Lin, Y. H. (2005). Antioxidant and antiproliferative activities of sweet potato (*Ipomoea babatas*) (L.J. Lam 'tainong 57') constituents. *Bot Bull Acad Sinica* 45: 179-186.
- Isnard, B. C., Deray, G., Baumelou, A., Le Quintree, M. & Vanherweghem, J. L. (2004). Herbs and the kidney. *Am J Kidney Dis* 44:1-11.
- International Agency for Research on Cancer (IARC), (2002). *IARC Monographs on the Evaluation of Carcinogenic Risks to Humans*. Vol. 82. *Some Traditional Herbal Medicines, Some Mycotoxins, Naphthalene and Styrene*. Lyon, IARC Press, pp. 69.
- Jain. A., Soni, M., Deb, L., Jain, A., Rout, S.P., Gupta, V.B. & Krishna, K.L. (2008). Antioxidant and hepatoprotective activity of ethanolic and aqueous extract of *Momordica dioica* Roxb leaves. *J Ethnopharmacol* 115, 61-66.
- Jo, E. H., Hong, H. D., Ahn, N. C., Jung, J. W., Yang, S. R., Park, J. S. Kim, S. H., Lee, Y.S., & Kang, K. S., (2004). Modulations of the Bcl-2/Bax family were involved in the chemopreventive effects of licorice root (*Glycyrrhiza uralensis* Fisch) in MCF-7 human breast cancer cell. *J Agric Food Chem* 52: 1715-1719.
- Joshi, B.S., & Kaul.P.N. (2001). Alternative medicine:Herbal drugs and their critical appraisal-Part I. *Prog Drug Res J* 56: 1-76.
- Kakuko, Y., Fumiko, A., Ogayama, A., N.Hikaru, O., Lucio, L.P., Edith, L., Elizabeth, M., Abigail, A. & Ricardo,R.C. (2005). Antibacterial activity of crude extract from Mexican medicinal plants and purified coumarins and xanthenes *J Ethnopharmacol* 97(2): 293-299.
- Kawada, S., Harada, M., Obara, Y., Kobayashi, S., Koyasu, K. & Oda, S. (2001). Karyosystematic analysis of Japanese talpine moles in the genera *Euroscaptor* and *Mogera* (Insectivora, Talpidae). *Zool Sci* 18: 1003-1010.
- Kim, J.Y., Park, K.W., Kwand-Deog Moon, K.D., Lee, M.K., Choi, J., Yee, S.T., Shim, K.H. & Seo, K.I. (2008). Induction of apoptosis in HT-29 colon cancer cells by crude saponin from *Platcodi Radix*. *Food Chem Toxicol* 46(12):3753-3758.
- Klaassen, C.D. (2001). Principles of Toxicology. In Casarett and Doull's Toxicology: The Basic Science of Poisons, 5th ed.; McGraw-Hill: New York, NY, USA, pp. 13.

- Knight, A.P. and Walter R.G. (2001). Plants Affecting Blood: Bracken Fern Poisoning. In A.P. Knight & R.G. Walter (Eds.). *A Guide to Plant Poisoning of Animals in North America*. Burbank, CA, USA: Teton NewMedia. pp. 195-197
- Kritikar, K.R. & Basu, B.D. (2001). Indian Medical Plant, vol.11. 2nd ed. Oriental Enterprises, Dehradun, pp.3578-3622.
- Kumar, R.A., Sridevi, K., Kumar, N.V., Nanduri, S. & Rajapopal, S., (2004). Anticancer and immunostimulatory compounds from *Andrographis paniculata*. *J Ethnopharmacol* 92: 291-295.
- Kumarnsit, E., Keawpradub, N. & Nuankaew, W. (2006). Acute and long term effects of alkaloid extract of *Mitragyna speciosa* on food and water intake and bodyweight in rats. *Fitoterapia* 77: 339-345.
- Kummalue, T., O-Charoenrat, P., Jiratchariyakul, W., Chanchai, K. Pattanapanyasat M. & Sukapirom, K. (2007). Antiproliferative effect of *Erycibe elliptilimba* on human breast cancer cell lines. *J Ethnopharmacol* 110: 439-443.
- Kwon, Y.D., Pittler, M.H. & Ernst, E. (2006). Acupuncture for peripheral joint osteoarthritis: A systematic review and meta-analysis. *Rheumatology (Oxford)*. 45(11):1331-7.
- Laffon, A., Moreno, A. & Gutierrez-Bucero, A. (1989). Hepatic sinusoidal dilatation in rheumatoid arthritis. *J Clin Gastro* 11:653-7.
- Lanfranco, G. (1992). Popular Use of Medicinal Plants in the Maltese Islands. *Insula* 1:34-35.
- Lazo, M., Selvin, E. & Clark, J. M., (2008). Brief Communication: Clinical Implications of Short-Term Variability in Liver Function Test Results. *Ann Inter Medicine*, 148:5.
- Lee, J.H., Choi, Y.H., Kang H.S. & Choi, B.T. (2004). An aqueous extract of *Platycodon radix* inhibits LPS-induced NF-kappaB nuclear translocation in human cultured airway epithelial cells. *Int. J Mol Med* 13: 843-847.
- Lemma, A. (1991). The Potentials and Challenges of Endod, the Ethiopian Soapberry Plant for Control of Schistosomiasis. In: *Science in Africa: Achievements and Prospects*. American Association for the Advancement of Sciences (AAAS), Washington, D.C., USA.
- Li, S., Long, C., Liu, F., Lee, S., Guo, Q., Li, R. & Liu, Y. (2006). Herbs for medicinal baths among the traditional Yao communities of China. *J Ethnopharmacol* 108(1): 59-67.

- Li, H., Hao, Z., Wang, X., Huang, L. & Li, J. (2009). Antioxidant activities of extracts and fractions from *Lysimachia foenum-graecum* Hance. *Bioresour Technol* 100: 970–974.
- Loeb, W.F. & Quimby, F.W. (1999) *Clinical Chemistry of Laboratory Animals*. 2nd ed., Philadelphia: Taylor & Francis, Philadelphia, PA. pp.405.
- Lu, F.C. & Kacew S. (2002). *Lu's basic toxicology*, 4th ed. London: Taylor and Francis.
- MacManus, M.P. (2008). Unproven medical devices and cancer therapy: big claims but no evidence. *Biomed Imaging Interv J* 4(3):25.
- Maton, A., Jean, H., McLaughlin, C.W., Warner, M.Q., Lattart, D. & Wright, J.D. (1993) *Human Biology and Health*. Englewood Cliffs, New Jersey, Prentice Hall.
- Mayne, P.D. (1996). *Clinical Chemistry in Diagnosis and treatment*. 6th ed (International Students Edition). Arnold London/Oxford University Press Inc. New York.
- McCann J. (2003). *Herbal Medicine Handbook* 2nd ed. Philadelphia: Lippincott.
- McGee, H. (1998). *In victu veritas*, *Nature* 392:649 – 650.
- McLellan, E.A., Medline, A. & Bird, R.P. (1991). Sequential analysis of the growth and morphological characteristics of aberrant crypt foci: putative preneoplastic lesions. *Cancer Res* 51:5270 –5274 4.
- Millan, J.L. (1988). Oncodevelopmental expression and structure of alkaline phosphatase genes. *Anticancer Research* 8: 995 –1004.
- Moalic, S., Liagre B, Corbiere, C., Bianchi, A., Danca, M., Bordji, K. & Beneytout, J.L. (2001). A plant steroid diosgenin, induces apoptosis, cell cycle arrest and cox activity in osteosarcoms cells. *FEBS Lett* 506: 205-230.
- Mohammed, M.A.M. (2006). *Effect of Morinda citrifolia (linn.) n phase i and ii drug metabolism and its molecular mechanism elucidation in rat liver*. Master of Science Thesis, Universiti Sains Malaysia. Malaysia.
- Mohd-Fuat, A.R., Kofi, E.A. & Allan, G.G. (2007). Mutagenic and cytotoxic properties of three herbal plants from Southeast Asia. *Trop Biomed* 24(2): 49-59.
- Moore, D. M. (2000): Hematology of rabbits. In: Schalm's Veterinary Hematology. 5th ed. (Feldman, B. F., J. G. Zinkel, N. C. Jain, Eds.). Lippincott Williams & Wilkins. Baltimore, Maryland, USA. pp. 1100-1106.

- Moshi, M.J. (2007). Brine shrimp toxicity evaluation of some Tanzanian plants used traditionally for the treatment of fungal infections. *Afr J Tradit Complem Altern Med* 4:219-225.
- Mukinda, J.T. & Syce, J.A. (2007). Acute and cronic toxicity of the aqueous extract of *Artemisia afra* in Rodent. *J Ethnopharmacol* 111; 138–144.
- Myhre, M.J. (2000). “Herbal Remedies, Nephropathies, and Renal Disease”. *Nephrol Nurs J* 27:5.
- Nahin, R.L., Dahlhamer, J.M., Taylor, B.L., Barnes, P.M., Stussman, B.J. & Simile, C.M. (2007). Health behaviors and risk factors in those who use complementary and alternative medicine. *BMC Public Health* 7(147):217.
- Nancy, E. (2004). In *The Laboratory Mouse*: Edited by Hans JH, Gilian B. Peter P. Elsevier Academic Press. UK, pp. 271-285.
- Natioanl Cancer Registry (NCR) (2002). *Cancer Incidence in Malaysia*. [Online]. Available: <http://www.cancer.org.my/> [accessed on January 2012].
- National Center for Complementary and Alternative Medicine. Expanding horizons of health care: Strategic plan 2005–2009. [Online]. Available from: <http://nccam.nih.gov/about/plans/2005>. [Accessed on January 2012].
- National Center for Complementary and Alternative Medicine, (NCCAM), (2011). *National Center for Complementary and Alternative Medicine: Botanical Dietary Supplement*. [Online]. Available: <http://www.nccam.nih.gov> [accessed on January 2012]
- Negishi, M., Aida, K. & Yoshioka, H. (1993). Sexually dimorphic expression of P-450 genes. In: Omura T, Ishimura Y, Fujii-Kuriyama Y, editors. *Cytochrome P-450*, 2nd ed. Kodansha, Tokyo, Japan, pp.230 – 238.
- Newman, D. J. & Price, C. P., (1999). Renal function and nitrogen metabolites. In: Burtis C.A., Ashwood E.R., (eds) *Tietz Text book of Clinical Chemistry*, 3rd ed. W.B. Saunders Company, Philadelphia, pp. 1204-1270.
- Newman, D.J., Cragg, G.M. & Snader, K.M. (2003). Natural Products as Sources of New Drugs. *J Natl Prod* 66:1022-1103.
- Nicholson, J.P., Wolmarans, M.R. & Park, G.R., (2000). The role of albumin in critical illness. *Br J Anaesth* 85 (4): 599-610.
- Norhaizan, M.E. & Phuah, S.C. (2009). *Rhaphidophora decursiva* leaves: Phenolic content and antioxidant activity. *J Trop Agri Food Sci* 37:61– 66.

- Nor Zuriati, M. (2010). *Effect of Rhabdophora decursiva (Roxb.). Schott extract on the proliferation of colon cancer (HT-29), liver cancer (HepG2), breast cancer (MCF-7) and ovary cancer (Caov-3) cell lines*. Thesis B. Sc (Nutrition and Community Health). Universiti Putra Malaysia, Serdang.
- Obici, S., Otobone, J.F., da Silva Sela, V.R., Ishida, K., da Silva, J.C., Nakamura, C.V., Cortez, D.A.G. & Audi, E.A. (2008). Preliminary toxicity study of dichloromethane extract of Kielmeyera coriacea stems in mice and rats. *J Ethnopharmacology* 115:131–139.
- Organization for Economic Cooperation and Development (OECD), (1995). *Guideline for Testing of Chemicals. Repeated Dose 28-Day Oral Toxicity Study in Rodents*; Organisation for Economic Co-operation and Development: Paris, France.
- Organization for Economic Cooperation and Development (OECD), (1998). *OECD Guideline for Testing of Chemicals (TG 408). Repeated Dose 90- day Oral Toxicity Study in Rodents*. OECD/OECD.
- Organization for Economic Cooperation and Development (OECD), (2001). *OECD Guideline for Testing of Chemicals (TG 423). Acute Oral Toxicity-Fixed Dose Procedure*. OECD/OECD.
- Organization for Economic Cooperation and Development (OECD), (2008). *OECD Guideline for Testing of Chemicals (TG 407). Repeated Dose 28- Day Oral Toxicity Study in Rodents*. OECD/OECD.
- Olson, H., Betton, G., Robinson, D., Thomas, K., Monro, A., & Kolaja, G. (2000). Concordance of toxicity of pharmaceuticals in humans and in animals. *Reg Toxicol Pharmacol* 32:56-67.
- Othman, A., Ismail, A., Ghani, A.N. & Adenan, I. (2007). Antioxidant capacity and phenolic content of cocoa beans. *Food Chem* 100: 1523–1530.
- Ozkan, A. & Fishkin, K. (2003). Cytotoxicity of low dose epirubicin- HCl combined with lymphokine activated killer cells against hepatocellular carcinoma cell line hepatoma G2. *Turk J Med Sci* 34: 11-19.
- Pari, L. & Murugan, P. (2004). Protective role of tetrahydrocurcumin against Erythromycin estolate-induced hepatotoxicity. *Pharmacol Res* 49; 481-6.
- Parkin, D.M., Bray, F., Ferlay, J. & Pisani, P. (2005). *Global Cancer Statistics, 2002. Cancer J Clin* 55:74-108.
- Peters, T.S. (2005). Do preclinical testing strategies help predict human hepatotoxic potentials?. *Toxicol Pathol* 33:146–154.

- Philip A.B., (2002). Acute Systemic Toxicity. *ILAR J* 43: 27-30.
- Prasad, K.N., Cole, W.C., Hovland, A.R., Prasad, K.C., Nahreini, P., Kumar, B., Edwards Prasad, J. & Andreatta, C.P. (1999). Multiple antioxidants in the prevention and treatment of neurodegenerative disease: analysis of biologic rationale. *Curr Opin Neuro* 12:761–770.
- Pratumvinit, B., Srisapoomi, T., Worawattananon, P., Opartkiattikul, N., Jiratchariyakul, W. & Kummalue, T. (2009). In vitro antineoplastic effect of *Ficus hispida* L. plant against breast cancer cell lines. *J Med Plants Res* 3(4): 255 – 261.
- Prozesky, E.A., Meyer, J.J.M. & Louw, A.I. (2001). In vitro antiplasmodial activity and cytotoxicity of ethnobotanically selected South African plants. *J Ethnopharmacol* 76: 239-245.
- Raja, M.M.M., Raja, A., Imran, M.M., Santha, A.M.I. & Devaseba, K. (2011). Enzymes Application in Diagnostic Prospects. *J Biotechnol* 10(1): 51-59.
- Ramaiah, S.K. (2007). A toxicologist guide to the diagnostic interpretation of hepatic biochemical parameters. *Food Chem Toxicol* 45:1551 –155 7.
- Rang, H.P., Dale, M.M. & Ritter, J.M. (1999). Absorption and distribution of drugs. In: Pharmacology, Fourth ed. Churchill Livingstone Edinburg, London, NY, Philadelphia, Sydney, Toronto, pp. 68.
- Raza, M., Al-Shabanah, O.A., El-Hadiyah, T.M. & Al-Majed, A.A. (2002). Effect of prolonged vigabatrin treatment on haematological and biochemical parameters in plasma, liver and kidney of Swiss albino mice. *Scientia Pharmaceutica* 70: 135-145.
- Rebecca, S., Deepa Naishadham, M.A. & Ahmedin Jemal, D.V.M. (2011). Cancer statistics, 2012. CA: *Am J Clin Nutr* 00:000–000.
- Rhiouani, H.R., Nazari, P., Kamli-Nejad, M. & Lyoussi, B. (2008). Acute and subchronic oral toxicity of an aqueous extract of leaves of *Herniaria glabra* in rodents. *J Ethnopharmacol* 118:378-386.
- Rispin, A., Farrar, D., Margosches, E., Gupta, K., Stitzel, K., Carr, G., Greene, M., Meyer, W. & McCall, D. (2002). Alternative methods for the median lethal dose (LD₅₀) test: The up-and-down procedure for acute oral toxicity. *ILAR J* 43(4): 233-243.
- Rothschild, M.A., Oratz, M. & Schreiber, S.S. (1988): Serum albumin. *Hepatology* 8:385-401.

- Saad, B., Azaizeh, H., Abu-Hijleh, G. & S. Said. (2006). Safety of traditional Arab herbal medicine. *Evid. Based Comp. Alter Med* 3: 433-439.
- Saadoun, D., Cazals-Hatem, D., Denninger, M.H., Boudaoud, L., Pham, B.N., Mallet, V., Condat, B., Brière, J. & Valla, J. (2004). Association of idiopathic hepatic sinusoidal dilatation with the immunological features of the antiphospholipid syndrome. *Int J Gastroentol Hepatol* 53(10): 1516–1519.
- Salawu, O.A., Chindo, B.A., Tijani, A.Y., Obidike, I.C., Salawu, T.A. & James Akingbasote, A. (2009). Acute and sub-acute toxicological evaluation of the methanolic stem bark extract of *Crossopteryx febrifuga* in rats. *Afr J Pharm Pharmacol* 3:621-626.
- Salazar, M., Martinez, E., Madrigal, E., Luiz, L.E. & Chamorro, G.A. (1998). Subchronic toxicity study in mice fed *Spirulina maxima*. *J Ethnopharmacol* 62:235-241.
- Salsburg, D.S. (1986) *Statistics for toxicologists*. New York: Marcel Dekker, pp. 3-93.
- Samy, J., Sugumaran, M. & Lee, K. (2005). *Herbs of Malaysia* (ed. K.M. Wong). Pub-Times Editions - Marshall Cavendish. pp.244
- Sánchez-Muniz, F.J., Higón, E., Cava, F. & Viejo, J.M. (1992). Prevention of dietary hypercholesterolemia in rats using sunflower-oil-fried sardines. Effects on cholesterol and serum enzymes. *J Agri Food Chem* 40:2226-223.
- Sasidharan, S., Darah, I. & Jain, K. (2008). In vivo and in vitro toxicity study of *Gracilaria changii*. *Pharm Biol* 46: 413-417.
- Sasikumar, J.M. & Doss, P.A. (2006). In vitro antioxidant and antibacterial activity of *Rhaphidophora pertusa* stem. *Fitoterapia* 77:605–607.
- Schindhelm, R.K., Dekker, J.M., Nijpels, G., Bouter, L.M., Stehouwer, C.D., Heine, R.J. & Diamant, M. (2007). Alanine aminotransferase predicts coronary heart disease events: A 10-year follow-up of the Hoorn Study. *Atherosclerosis* 191: 391-396.
- Schindhelm RK, Diamant M, Dekker JM, Tushuizen ME, Teerlink T. & Heine RJ. (2006). Alanine aminotransferase as a marker of non-alcoholic fatty liver disease in relation to type 2 diabetes mellitus and cardiovascular disease. *Diabetes Metab Res* 22:437-43.
- Shahin, S.A., Naresh, K., Abhinav, L., Angad, S., Hallihosur, S., Abhishek, S. & Utpal, B. (2008). *Food Res Int* 41(1): 1-15.
- Shoeb M. (2006). Anticancer agents from medicinal plants. *Bangalore J Pharm* 1:35-41.

- Slade, S.C. & Keating, J.L. (2007). Unloaded movement facilitation exercise compared to no exercise or alternative therapy on outcomes for people with nonspecific chronic low back pain: A systematic review. *J Manipulative Physiol Ther* (4):301– 11.
- Stephens, J.C., Reich, D.E., Goldstein, D.B., Shin, H.D., Smith, M.W., Carrington, M., Winkler, C., Huttley, G., Allikmets, R., Schriml, L., Gerrard, B., Malasky, M., Ramos, M.D., Morlo, t.S., Tzetzis, M., Oddoux, C., Giovine, F.S., Nasioulas, G., Chandler, D., Aseev, M., Hanson, M., Kalaydjieva, L., Glavac, D., Gasparini, Kanavakis, E., Claustres, M., Kambouris, M., Ostrer, H., Duff, G., Baranov, V., Sibul, H., Metspalu, A., Goldman, D., Martin, N., Duffy, D., Schmidtke, J., Estivill, X., O'Brien, S.J., Dean, M. (1998). Dating the origin of the CCR5-D32 AIDS-resistance allele by the coalescence of haplotypes. *Am J Hum Genet* 62:1507–1515.
- Swamy, S.M.K. & Tan, B.K.H. (2000). Cytotoxic and immunopotentiating effects of ethanolic extract of *Nigella sativa* L. seeds. *J Ethnopharmacol* 70:1-7.
- Tan, A.S. & Berridge, M.V. (2000). Superoxide produced by activated neutrophils efficiently reduces the tetrazolium salt, WST-1 to produce a soluble formazan: a simple colometric assay for measuring respiratory burst activation and for screening anti-inflammatory agents. *J Immunol Methods* 238: 59-68.
- Tan, P.V., Mezui, C., Enow-Orock, G., Njikam, N., Dimo, T. & Bitolog, P. (2008). Teratogenic effects, acute and sub chronic toxicity of the leaf aqueous extract of *Ocimum suave* Wild (Lamiaceae) in rats. *J Ethnopharmacol* 115:232–237.
- Tang, J.L., Liu, B.Y. & Ma, K.W. (2008). Traditional Chinese medicine. *Lancet*. 372: 1938–40.
- Tempesta, M.S. and S. King. (1994). Tropical plants as a source of new pharmaceuticals, in P.S. Barnacal (ed) *Pharmaceutical Manufacturing International: The International Review of Pharmaceutical Technology Research and Development*. Sterling Publications Ltd., London.
- Teo, S.D., Stirling, S., Thomas, A., Kiorpes, A. and Vikram, K. (2002). A 90-day oral gavage toxicity study of D-methylphenidate and D, L methylphenidate in Sprague-dawley rats. *Toxicol* 179: 183-196.
- Timbrell, J. (2000). *Principles of biochemical toxicology*. London: Taylor&Francis Ltd, pp. 25-172.
- Tiwari, K. N., Sharma N.C., Tiwari, V., & Singh, B.D. (2000). Micropropagation of *Centella asiatica* (L.), a valuable medicinal herb. *Plant, Cell, Tiss, Org* 63(1): 179-185

- Tolman, K.G. & Rej, R. (1999). Liver Function. In: Tietz Text Book of Clinical Chemistry, Burtis, C.A. and E.R. Ashwood (Eds.). 3rd Edn., W.B. Saunders Co., Philadelphia, PA., USA., pp. 1125-1177.
- Tortora, G., & Derrickson, B. (2006). Principles of Anatomy and Physiology (11th edition). New Jersey: John Wiley and Sons Inc.
- Toyota, M., & Ihara, M. (1998). Recent progress in the chemistry of nonmonoterpenoid indole alkaloids. *Nat Prod* 15:307-340.
- Trevan, J.W. (1927). The error of determination of toxicity. Proceedings of the Royal Society (London), Series B 101, 483-514.
- United Nations Educational, Scientific and Cultural Organization (UNESCO), (1996). *Culture and Health*, Orientation Texts – World Decade for Cultural Development 1988 – 1997, Document CLT/DEC/PRO – 1996, Paris, France, p. 129.
- Urabe, K., Aroca, P., Tsukamoto, K., Mascagna, D., Palumbo, A., Prota, G., & Hearing, V.J. (1994). The inherent cytotoxicity of melanin precursors: a revision. *Biochimica et Biophysica Acta (BBA) – Molecular Cell Research* 1221: 272 - 278.
- Vaes, L.P.J. and Chyka, P.A. (2000). Interaction of warfarin with garlic, ginger, ginkgo, or ginseng: nature of the evidence. *Ann Pharmacol* 34:1478–1482.
- Vaghasiya, Y.K., Shukla, V.J., Chanda, S.V. (2011). Acute oral toxicity study of *Pluchea arguta* boiss extract in mice. *J Pharmacol Toxicol* 6: 113-123.
- Van de Greef, J., (2003). The role of analytical sciences in medical systems biology. *Curr Opin Cell Biol* 8:944-954.
- Wahlberg, B.J., Burholt, D.R., Kornblith, P., Richards, T.J., Bruffsky, A., Herberman, R.B., & Vujanovic, N.L. (2001). Measurement of NK activity by the microcytotoxicity assay (MCA): a new application for an old assay. *J Immunol Methods* 253: 69-81.
- Wang, G., Zhao, J., Liu, J., Huang, Y., Zhong, J., & Tang, W. (2007). Enhancement of IL-2 and IFN- γ expression and NK cells activity involved in the anti-tumor effect of ganoderic acid Me *in vivo*. *Int Immunopharmacol* 7: 864-870.
- Weingand, K., Brown, G., & Hall, R. (1996): Harmonization of Animal Clinical Pathology Testing in Toxicity and safety studies. *Fund Appl Toxicol* 29:198-201.

- White, A., Foster, N.E., Cummings, M. & Barlas, P. (2007). Acupuncture treatment for chronic knee pain: A systematic review. *Rheumatology (Oxford)*. 46(3):384–90.
- Whiting, P.W., Clouston, A., & Kerlin, P. (2002). Black cohosh and other herbal remedies associated with acute hepatitis. *Med J Aust* 177: 440–443.
- Wieckowska, A., Zein, N.N., Yerian, L.M., Lopez, A.R., McCullough A.J., & Feldstein, A.E. (2006). *In vivo* assessment of liver cell apoptosis as a novel biomarker of disease severity in non-alcoholic fatty liver disease. *Hepatology* 44: 27-33.
- Wilson, J.W. (1979). Inherited elevation of alkaline phosphatase activity in the absence of disease. *N England J Med* 301:983-984.
- Wiseman, N. (2004). Designations of medicines. *Evid based Complement Alternat Med* 1(3):327-329.
- Wojdylo, A., Oszmianski, J., & Czemerys, R. (2007). Antioxidant activity and phenolic compounds in 32 selected herbs. *Food Chem* 105: 940-949.
- Wong, K.T & Tan, B.K.H. (1996). In vitro cytotoxicity and immunomodulating property of *Rhaphidophora korthalsii*. *J Ethnopharmacol* 52:53–57.
- World Health Organization (WHO), (1993). Research Guidelines for Evaluating the Safety and Efficacy of Herbal Medicines. Geneva.
- World Health Organization (WHO), (2000). General Guidelines for Methodologies on Research and Evaluation of Traditional Medicine. Geneva. WHO/EDM/TRM/2000.1.
- World Health Organization (WHO), (2008). <http://www.who.int/dgspeech/index.html> WHO Congress on traditional medicine. [Accessed on January 2012]
- Wu, S.N., Chen, C.C., Li, H.F., Lo, Y.K., & Chen, S.A. (2002): Chiang, H.T. Stimulation of BKCa channel in cultured smooth muscle cells of human trachea by magnolol. *Thorax* 57: 67-74
- Wurochekke, A.U., Anthony, A.E., & Obidah, W. (2008): Biochemical effects on Liver and kidney of rats administered aqueous stem bark extract of *Xemenia americana*. *Afr J Biotechnol* 7(16): 2777-2780.
- Xiao, J.B., Chen, X.Q., Zhang, Y.W., Jiang, X.Y., & Xu, M. (2006). Cytotoxicity of *Marchantia convolute* leaf extract to human liver and lung cancer cells. *Braz J Med Biol Res* 39: 731-738.

- Yakubu, M.T., Akanji, M.A., & Oladiji, A.T. (2007). Haematological evaluation in male albino rats following chronic administration of aqueous extract of *Fadogia agrestis* stem. *Phcog Mag* 3: 34.
- Yeap, S.K., Alitheen, N.B., Ali, A.M., Omar, A.R., Suraini, A.A., & Muhajir, A.H., (2007). Effect of *Rhaphidophora korthalsii* methanol extract on human peripheral blood mononuclear cell (PBMC) proliferation and cytolytic activity toward HepG2. *J Ethnopharmacol* 114(3): 406-411.
- Zaffani, S., Cuzzolin, L. & Benoni, G. (2006). Herbal products: behaviors and beliefs among Italian women. *Pharmacoepidemiol. Drug Saf.* 15:354–359.
- Zhang, H., Qiu, S., Tamez, P., Tan, G.T., Aydogmus, Z., Van Hung, N., Cuong, N.M. & Fong, H.H.S. (2002). Antimalarial agents from plants II. Decursivine, a new antimalarial indole alkaloid from *Rhaphidophora decursiva*. *Pharma Biol* 40(3): 221–224.
- Zhang, H.J., Tamez, P.A., Hoang, V.D., Ghee, T.T, Hung, N.V., Xuan, L.T., Huong, L., Fong, H.S. & Pezzuto, J.M. (2001). “Antimalarial Componds from *Raphidiphora decursiva*”. *J Nat Prod* 64(6):772-777.
- Zhou, H.L., Deng, Y.M., & Xie, Q.M. (2006). Themodulatory effects of the volatile oil of ginger on the cellular immune response in vitro and in vivo in mice. *J Ethnopharmacol* 105:301–305.