



**UNIVERSITI PUTRA MALAYSIA**

***HEPATOPROTECTIVE EFFECT OF *Muntingia calabura* L. LEAF  
EXTRACTS***

**NUR DIYANA BINTI MAHMOOD**

**FPSK(M) 2016 25**



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By

**NUR DIYANA BINTI MAHMOOD**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfilment of the Requirements for the Degree of Master of Science**

**July 2016**

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

## HEPATOPROTECTIVE EFFECT OF *Muntingia calabura* L. LEAF EXTRACTS

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July 2016

**Chairman:** Associate Professor Zainul Amiruddin Zakaria, PhD  
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*Muntingia calabura* is a plant of the family Elaeocarpaceae, a tropical species and known to the Malays as 'Kerukup Siam'. Traditionally, the Peruvian believed that *M. calabura*'s leaves could reduce gastric ulcers and swelling of prostate gland as well as alleviate headache and cold. Therefore, the objective of this study was to determine the hepatoprotective activity of methanolic extract of *M. calabura* leaves (MEMC) and its partitions using rat models. The hepatoprotective potential of MEMC was investigated using paracetamol (PCM)- and carbon tetrachloride (CCl<sub>4</sub>)-induced hepatotoxicity in rats. Briefly, male Sprague Dawley rats ( $n = 6$  per group) were divided into groups and administered orally once daily with 10% dimethyl sulfoxide (DMSO) (negative control), 50 mg/kg N-acetylcysteine (NAC) (positive control), or MEMC (50, 250, and 500 mg/kg) for 7 consecutive days, followed by hepatotoxicity induction using PCM or CCl<sub>4</sub>. MEMC was later partitioned into 3 fractions: petroleum ether extract (PEMC), ethyl acetate extract (EAMC) and aqueous extract (AQMC). The protective effect of PEMC, EAMC and AQMC were tested on PCM-induced hepatotoxicity rat model. Blood samples were subjected to biochemical analysis to evaluate alanine transferase (ALT), aspartate amino transferase (AST), and alkaline phosphatase (ALP) levels; some parts of the liver were subjected to microscopic analysis. Fresh samples of liver tissues were used to determine the superoxide dismutase (SOD) and catalase (CAT) activities. All extracts (MEMC, PEMC, EAMC and AQMC) were tested for antioxidant activity study using the 2, 2-diphenyl-1-picrylhydrazyl radical scavenging assay (DPPH), superoxide dismutase scavenging activity assay (SOD), and oxygen radical absorbance capacity assay (ORAC), and anti-inflammatory study using xanthine oxidase (XO) and lipoxigenase (LOX) assays. Total phenolic content (TPC), phytochemical screening, and high-performance liquid chromatography (HPLC) analysis were also performed. From the histological observation, lymphocyte infiltration and marked necrosis were observed in the DMSO-treated group (negative control). MEMC at 500 mg/kg significantly reduce the toxic effect of PCM and CCl<sub>4</sub> on the liver by reducing the weight of the liver; histological observation demonstrated normalization of the histopathological changes, preserving hepatocytes structure, causing a significant decline in ALT and AST levels ( $P < 0.05$ ) and increase in the SOD and CAT activities. Among the partitions, AQMC at 250 mg/kg showed significant reduction in the toxic effect of PCM by causing significant decline in the ALT and AST levels. AQMC were then tested at doses of 50 mg/kg and 500 mg/kg and the result

demonstrated reduction in the liver enzymes in a dose-dependent manner with augmentation of SOD and CAT activities. MEMC had the highest TPC value, followed by EAMC, PEMC, and AQMC. MEMC and AQMC demonstrated strong free radical scavenging activity in the DPPH and SOD assays. AQMC showed the highest ORAC value while MEMC was the lowest among the extracts. All extracts in the present study demonstrated strong anti-inflammatory activity via inhibition of LOX. Phytochemical screening of the extracts showed that MEMC, PEMC and EAMC contained flavonoids, tannins, and steroids. Only PEMC showed the presence of triterpenes. However, the phytochemical screening showed that AQMC contained fewer compounds. HPLC analysis suggests that MEMC and AQMC contained flavonoid-based compounds. In conclusion, MEMC exerted potential hepatoprotective activity that can be attributed to its antioxidant activity, and AQMC was considered to have the best activity among other partitions, which warrants further investigation.



Abstrak thesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

## **KESAN HEPATOPROTEKTIF OLEH EKSTRAK DAUN *Muntingia calabura* L.**

Oleh

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**Julai 2016**

**Pengerusi:           Profesor Madya Zainul Amiruddin Zakaria, PhD**  
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*Muntingia calabura* ialah sejenis tumbuhan tropikal dari keluarga Elaeocarpaceae dan dikenali sebagai Kerukup Siam oleh masyarakat Melayu. Secara tradisi, masyarakat Peru percaya bahawa daun *M. calabura* mampu mengurangkan ulser gastrik dan bengkak pada kelenjar prostat termasuk juga meredakan sakit kepala dan selsema. Oleh itu, objektif kajian ini adalah untuk menentukan aktiviti hepatoprotektif ekstrak methanol daripada daun *M. calabura* dan pecahannya dengan menggunakan model tikus. Potensi hepatoprotektif dari ekstrak metanol daun *M. calabura* (MEMC) telah diuji menggunakan rangsangan hepatotoksiti paracetamol (PCM)- dan karbon tetraklorida (CCl<sub>4</sub>) pada tikus. Secara ringkas, tikus 'Sprague Dawley' jantan ( $n = 6$  bagi setiap kumpulan) dibahagikan kepada beberapa kumpulan dan diberi makan secara oral sekali sehari dengan 10% dimetil sulfoxide (DMSO) (kawalan negatif), 50 mg/kg N-acetylcysteine (NAC) (kawalan positif), atau MEMC (50, 250, dan 500 mg / kg) selama 7 hari, diikuti dengan aruhan hepatotoksiti menggunakan PCM atau CCl<sub>4</sub>. MEMC seterusnya dibahagikan kepada 3 pecahan: ekstrak petroleum eter (PEMC), ekstrak etil asetat (EAMC) dan ekstrak air (AQMC). Kesan perlindungan oleh PEMC, EAMC dan AQMC telah diuji dengan model aruhan PCM ke atas tikus. Sampel darah yang telah diambil dibuat kajian biokimia untuk menganalisis paras enzim seperti alanine transferase (ALT), aspartate aminotransferase (AST), and alkaline phosphatase (ALP). Manakala sebahagian daripada sampel hati pula diuji secara mikroskopik. Sampel hati yang segar pula diuji untuk aktiviti superoxide dismutase (SOD) dan catalase (CAT). Semua ekstrak (MEMC, EAMC dan AQMC) diuji untuk kajian antioksidan menggunakan cerakin 2, 2-difenil-1-picrylhydrazyl radikal (DPPH), pengujian perangkap aktiviti superoxide dismutase (SOD), dan cerakin penyerapan oksigen radikal kapasiti (ORAC), dan kajian anti-radang menggunakan analisis aktiviti xanthine oxidase (XO) dan lipoxigenase (LOX). Kandungan jumlah fenol (TPC), pemeriksaan fitokimia, dan analisa kromatografi cecair berprestasi tinggi (HPLC) juga telah dilaksanakan. Dari segi pemerhatian histologi penyusupan limfosit dan nekrosis diperhatikan dalam kumpulan rawatan DMSO (kawalan negatif). MEMC pada dos 500 mg/kg menunjukkan pengurangan yang signifikan kepada kesan toksik oleh PCM dan CCl<sub>4</sub> ke atas hati, dengan menyebabkan penurunan berat hati, pemantauan histology menunjukkan pemulihan struktur sel-sel hati, dan menyebabkan penurunan paras ALT dan AST secara signifikan ( $P < 0.05$ ) dan peningkatan aktiviti SOD dan CAT. Antara semua pecahan, AQMC pada dos 250 mg/kg menunjukkan pengurangan yang

signifikan ke atas kesan toksik oleh PCM yang dengan menurunkan paras ALT dan AST secara signifikan. AQMC diuji pada dos 50 mg/kg dan 500 mg/kg dan keputusan menunjukkan penurunan paras enzim hati dengan kebergantungan pada peningkatan dos dan juga peningkatan aktiviti enzim SOD dan CAT. MEMC mempunyai nilai TPC paling tinggi diikuti oleh EAMC, PEMC, dan AQMC. MEMC dan AQMC menunjukkan aktiviti yang paling tinggi dalam pengujian perangkap DPPH dan SOD. AQMC menunjukkan nilai ORAC yang tinggi manakala MEMC menunjukkan nilai yang paling rendah berbanding ekstrak lain. Semua ekstrak menunjukkan anti radang yang kuat dalam menghalang LOX. Pemeriksaan fitokimia ekstrak menunjukkan MEMC, PEMC dan EAMC mempunyai flavonoid, tannin, dan steroid. Hanya PEMC yang mempunyai triterpene. Walau bagaimana, AQMC hanya mempunyai sedikit sebatan. Analisa HPLC mencadangkan bahawa MEMC dan AQMC mempunyai sebatan daripada flavonoid. Kesimpulannya, MEMC mempunyai potensi sebagai agen hepatoprotektif yang juga bergantung kepada aktiviti antioksidan, dan AQMC dianggap mempunyai aktiviti yang terbaik di antara pecahan ekstrak, yang memerlukan siasatan lanjut.

## ACKNOWLEDGEMENT

Alhamdulillah, All Praises are for Allah subhanahu wa ta`ala for helping me in coordinating my life.

I wish to extend my deepest appreciation to my supervisor, Associate Professor Dr. Zainul Amiruddin Zakaria, for his guidance, valuable advice, patience, support and continuous supervision throughout the course of this project. I truly thank him for giving me the opportunity to be his postgraduate student.

I would like to express my gratitude and appreciation to my co-supervisor, Associate Professor Dr Norhafizah Mohtarrudin, for her kindness to teach and guide me in histological study that is definitely important in order to complete this project.

My sincere appreciation goes to the Faculty of Medicine and Health Sciences, Universiti Putra Malaysia, for giving me the opportunity to carry out this project.

My special dedication to Encik Ramli bin Suhaimi, staff of Pharmacology Laboratory and of Histology Laboratory for their kind cooperation, excellent facilities and support, which helped me to perform the analyses smoothly. I would like to thank all my fellow friends, Farah Hidayah bt Kamisan, Mohammad Fauzi Fahmi bin Kamarolzaman, Siti Syariah bt Mamat, Roihana bt Rodzi, Tavamani Balan, Farhana bte Yahya, Nur Liana bt Md Nasir, NoorSyaza Eddrina bt Kamsani, Noor Wahida bt Suhaimy, Mohd Khusairi bin Azmi, Mohd Hijaz bin Sani, and Salahuddin bin Mumtaz Ahmad for their kindness, patience and cooperation in completing this project successfully.

Words are not enough to express my gratitude to my beloved family, especially my dearest parents, Mahmood bin Ibrahim and Fatimah binti Mamat for their love, comfort, encouragement, support and advice that truly motivated me to accomplish my Master's degree successfully. I owe them a depth gratitude which can never be repaid. May Allah shower His countless blessings upon them.

Thank you so much.



I certify that a Thesis Examination Committee has met on 18 July 2016 to conduct the final examination of Nur Diyana binti Mahmood on her thesis entitled "Hepatoprotective Effect of *Muntingia calabura* L. Leaf Extracts" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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## LIST OF ABBREVIATIONS

AAPH	2,2'-azobis-2-methyl-propanimidamide, dihydrochloride
AQMC	Aqueous extract of <i>Muntingia calabura</i>
CYP450	Cytochrome P450
CYP2E1	Cytochrome P450 2E1
EAMC	Ethyl acetate extract of <i>Muntingia calabura</i>
IC <sub>50</sub>	Median inhibitory concentration
IL-1	interleukin-1
MEMC	Methanol extract of <i>Muntingia calabura</i>
NSAIDs	Non-steroidal anti-inflammatory drugs
ORAC	Oxygen radical absorbance capacity
PCM	Paracetamol
PEMC	Petroleum ether extract of <i>Muntingia calabura</i>
PPAR- $\alpha$	Peroxisome proliferator-activated receptor alpha
TNF	Tumor necrosis factor
WHO	World Health Organization

# CHAPTER 1

## INTRODUCTION

### 1.1 Research background

Liver is a vital organ in the body that plays a role in regulation of diverse processes including metabolism, secretion, storage, and detoxification of waste metabolites (endogenous) and toxic compounds (exogenous) (Madrigal-Santillán *et al.*, 2014). Continual damage to the liver by acute liver insult will eventually result in the development of hepatic fibrogenesis, which promotes the abnormal structural changes to the tissue known as cirrhosis (Qua and Goh, 2011). The increasing incidence of mortality due to the liver diseases has been reported to be the tenth leading cause of death in the United States of America (Jiaquan *et al.*, 2016). Meanwhile, the prevalence of liver cirrhosis in Malaysia is 15 in every 10,000 population, and the distributions of underlying etiology vary regionally with viral hepatitis being much higher compared to the European countries (Ng *et al.*, 2011).

For millennia, natural products have been used traditionally to treat and/or prevent various liver diseases in many parts of the world. Regardless of the abundance of the number of modern drugs in the pharmaceutical market, natural products have begun to gain importance and popularity in the world for promoting health care as well as disease prevention, and been used as conventional or complementary medicines for treatable and incurable diseases (Zhang *et al.*, 2013). The World Health Organization (WHO) estimated that up to four billion people representing 80% of developing societies of the world's population consider on natural products as their most preferred healthcare option (Amiri *et al.*, 2012; Ekor, 2014).

Natural products become an interest in drug discovery as it contributes unique structural diversity, which provides the opportunities for discovering mainly novel low molecular weight lead compounds that have desirable properties (Dias *et al.*, 2012). An analysis from 1981 to 2010 by Newman and Cragg (2012) showed that more than two-thirds of the drug active compounds recently introduced are likely derived from natural sources, and only about 30% are completely synthetic origin (Newman and Cragg, 2012).

Since less than 10% of the 250,000 species of the world's biodiversity has been evaluated for medicinal purposes (Ramasamy *et al.*, 2011; McChesny *et al.*, 2007), many more species await to be discovered for therapeutic benefits. Malaysia, a well known tropical country, has been acknowledged worldwide as a land of floral and faunal prosperity, and is believed to be a reservoir of a large collection of potential medicinal plants.

*Muntingia calabura* from the family of Elaeocarpaceae, is one of the native plants in Malaysia that has been widely tested and documented for its promising

pharmacological properties. Numerous investigations showed that the leaves possess various pharmacological activities, including antiulcer, antipyretic, antinociceptive and anti-inflammatory activities along with antioxidant and antiproliferative properties (Mahmood *et al.*, 2014). However, hepatoprotective properties of this plant have not yet been explored. As such, further research on this activity is significant to nominate another plant to the list of potential medicinal hepatoprotective plant-based products.

## **1.2 Problem statement**

Liver diseases remain one of the major threats to community health and a worldwide burden. Particularly, drug-induced hepatotoxicity is a major cause of hepatic damage or dysfunction. Paracetamol (PCM), a mild analgesic and antipyretic drug developed in the past few decades, causes severe liver injuries in human and experimental animals when taken overdose. Liver cirrhosis develops from excessive alcohol consumption is the common alcoholic liver disease in western countries (Byass, 2014). Zain *et al* (2006) reported that in Singapore, PCM represented 55% of overdosing cases in year 1999 among patients admitted to the hospital for intentional self-poisoning. In spite of remarkable advances in modern medicine, there are hardly reliable drugs that protect the liver from damage and/or help in regeneration of hepatic cells. Due to adverse effects of drug, a great deal of research has been conducted to explore natural products from plants as promising hepatoprotective agents. It is known that agents possessing anti-inflammatory activity have the potential to act as an antioxidant (Yanpallewar *et al.*, 2002). Previous studies on *M. calabura* reported the presence of antioxidant and anti-inflammatory activity that is relevant to hepatoprotective activity. Considering these reports, the antioxidant and anti-inflammatory activity indicate different pathways assisting the hepatoprotective effect. ROS generated from drug or chemical metabolism appear to be the fundamental mechanisms underlying most human ailments. The antioxidant and anti-inflammatory properties of plants facilitate the free radical scavenging process and regulate the inflammatory response, respectively, which are believed to initiate their damaging effects on the liver. Therefore, this study is expected to discover the capacity of *M. calabura* for hepatoprotective activity.

## **1.3 Objective(s)**

### **1.3.1 General objective**

To determine the hepatoprotective activity of methanol extract of *Muntingia calabura* (MEMC) leaves and its partitions using rat models.



### 1.3.2 Specific objectives

- To determine the hepatoprotective effect of MEMC against paracetamol (PCM) and carbon tetrachloride (CCl<sub>4</sub>)-induced liver toxicity in rats,
- To determine the most effective partition of MEMC (petroleum ether, ethyl acetate and aqueous extracts) on liver toxicity study,
- To elucidate the possible mechanisms of hepatoprotection of MEMC and the most effective partition

### 1.4 Research hypotheses

1. It is hypothesize that methanol extracts of *M. calabura* leaves (MEMC) will exhibit a hepatoprotective activity in paracetamol (PCM)- and carbon tetrachloride (CCl<sub>4</sub>)- induced liver toxicity assays.
2. AQMC is expected to have good hepatoprotective activity in PCM-induced liver toxicity.

## REFERENCES

- Adeneye, A. A. 2009. Protective activity of the stem bark aqueous extract of *Musanga cercropioides* in carbon tetrachloride- and acetaminophen- induced acute hepatotoxicity in rats. *African Journal Traditional, Complementary, and Alternative Medicines*. 6: 131-138.
- Aluko, B. T., Oloyede, O. I. and Afolayan, A. J. (2013). Hepatoprotective activity of *Ocimum americanum* L leaves against paracetamol-induced liver damage in rats. *American Journal of Life Sciences*.1: 37-42.
- Amarowicz, R., Pegg, R. B., Rahimi-Moghaddam, P., Barl, B., and Weil, J. A. (2004). Free radical scavenging capacity and antioxidant activity of selected plant species from the Canadian prairies. *Food Chemistry*. 84: 551-562.
- Amin, Z. A., Bolgen, M., Alshawsh, M. A., Ali, H. M., Hadi, A. H. A., and Abdulla, M. A. (2012). Protective role of *Phyllanthus niruri* extract against thioacetamide induced liver cirrhosis in rat model. *Evidence-Based Complementary and Alternative Medicine*. 2012:1-9.
- Amiri, M. S., Jabbarzadeh, P. and Akhondi, M. (2012). An ethnobotanical survey of medicinal plants used by indigenous people in Zangelanlo district, Northeast Iran. *Journal of Medicinal Plants Research*. 6: 749-753.
- Ao, Z. H., Xu, Z. H., Lu, M. Z., Xu, H. Y., Zhang, X. M. and Dou, W. F. (2009). *Niuchangchih (Antrodia camphorate)* and its potential in treating liver diseases. *Journal of Ethnopharmacology*. 121: 194-212.
- Babu, D., Gurumurthy, P., Borra, S. K. and Cherian, K. M. (2013). Antioxidant and free radical scavenging activity of triphala determined by using different *in vitro* models. *Journal of Medicinal Plants Research*. 7: 2898-2905.
- Balan, T., Sani, M. H. M., Suppaiah, V., Mohtarrudin, N., Suhaili, Z., Ahmad, Z., Zakaria, Z. A. (2013). Antiulcer activity of *Muntingia calabura* leaves involves the modulation of endogenous nitric oxide and nonprotein sulfhydryl compounds. *Pharmaceutical Biology*. 52: 410-418.
- Battacharjee, R. and Sil, P. C. (2006). The protein fraction of *Phyllanthus niruri* plays a protective role against acetaminophen induced hepatic disorder via its antioxidant properties. *Phytotherapy Research*. 20: 595-601.
- Bellik, Y., Boukra, L., Alzahrani, H. A., Bakhotmah, B., Abdellah, F., Hammoudi, S. M., and Iguer-Ouada, M. (2013). Molecular mechanism underlying anti-inflammatory and anti-allergic activities of phytochemicals: An update. *Molecules*. 18: 322-353.

- Bessems, J.G. and Vermeulen N.P. (2001) Paracetamol (acetaminophen)-induced toxicity: Molecular and biochemical mechanisms, analogues and protective approaches. *Critical Reviews in Toxicology*. 31:55-138.
- Bhagwat, S., Haytowitz, D. B. and Holden, J. M. (2007). USDA database for the Oxygen Radical Absorbance Capacity (ORAC) of selected foods. In *American Institute for Cancer Research Launch Conference*. Washington, DC.
- Blois, M.S. (1958). Antioxidant determinations by the use of a stable free radical. *Nature*. 181: 1199–1200.
- Brind, A. M. (2006). Drugs that damage the liver. *Medicine*. 35(1): 26-30.
- Bupesh, G., Amutha, C., Vasanth, S., Manoharan, N., Raja, R. S., Krishnamoorthy, R., and Subramanian, P. (2012). Hepatoprotective efficacy of *Hypnea muciformis* ethanolic extract on CCl<sub>4</sub> induced toxicity in rats. *Brazilian Archives of Biology and Technology*. 55(6): 857-863.
- Byass, P.(2014). The global burden of liver disease: A challenge for methods and for public health. *Biomedical Central Medicine Journal*, 12: 159-161.
- Campbell, I. (2006). Liver: functional anatomy and blood supply. *Anaesthesia And Intensive Care Medicine*. 7: 49-51.
- Carocho, M. and Ferreira, I. C. F. R. (2013). A review on antioxidant, prooxidants and related controversy: Natural and synthetic compounds, screening and analysis methodologies and future perspectives. *Food and Chemical Toxicology*. 51: 15-25.
- Chang, W. S., Lin, C. C. and Chiang, H. C. (1996). Superoxide anion scavenging effect of coumarins. *American Journal of Chinese Medicine*. 24: 11–17.
- Chen, J. J., Lin, R. W., Duh, C. Y., Huang, H. Y., and Chen, I. S. (2004). Flavones and cytotoxic constituents from the stem bark of *Muntingia calabura*. *Journal of the Chinese Chemical Society*. 51:665–670.
- Chen, J. J., Lee, H. H., Duh, C. Y., and Chen, I. S. (2005). Cytotoxic chalcones and flavonoids from the leaves of *Muntingia calabura*. *Planta Medica*. 71: 31-34.
- Chen, J. J., Lee, H. H., Shih, C. D., Liao, C. H., Chen, I. S., and Chou, T. H. (2007). New dihydrochalcones and anti-platelet aggregation constituents from the leaves of *Muntingia calabura*. *Planta Medica*. 73:572-577.
- Chen, Y. H., Lin, F. Y., Liu, P. L., Huang, Y. T., Chiu J. H., Chang Y. C., Man, K. M., Hong, C. H., Ho, Y. Y., and Lai, M. T. (2009). Antioxidative and hepatoprotective effects of magnolol on acetaminophen-induced liver damage in rats. *Archives of Pharmacal Research*. 32: 221.-228.

- Choudhari, A. S., Suryavanshi, S. A., Ingle, H. and Kaul-Ghanekar, R. (2011). Evaluating the antioxidant potential of aqueous and alcoholic extracts of *Ficus religiosa* using ORAC assay and assessing their cytotoxic activity in cervical cancer cell lines. *Biotechnology, Bioinformatics and Bioengineering*. 1:443-450.
- Cichoż-Lach, H., and Michalak, A. (2014). Oxidative stress as a crucial factor in liver diseases. *World Journal of Gastroenterology*. 20: 8082-8091.
- Clawson, G. A. (1989). Mechanisms of carbon tetrachloride hepatotoxicity. *Pathology and Immunopathology Research*. 8:104-112.
- Conforti, F., Sosa, S., Marrelli, M., Menichini, F., Statti, G. A., Uzunov, D., Tubaro, A., Menichini, F. and Loggia, R. D. (2008). *In vivo* anti-inflammatory and *in vitro* antioxidant activities of Mediterranean dietary plants. *Journal of Ethnopharmacology*. 116: 144-151.
- Convention on Biological Diversity (CBD) Country Profile. <http://www.cbd.int/countries/profile/default.shtml?country=my#status> accessed on 9<sup>th</sup> January 2014.
- Cover, C., Liu, J., Farhood, A., Malle, E., Waalkes, M. P., Bajt, M. L., and Jaeschke, H. (2006). Pathophysiological role of the acute inflammatory response during acetaminophen hepatotoxicity. *Toxicology and Applied Pharmacology*. 216: 98-107.
- Dai, J. and Mumper, R. J. (2010). Plant phenolics: extraction, analysis and their antioxidant and anticancer properties. *Molecules*. 15: 7313-7352.
- Dancygier, H. (2009). Clinical hepatology: Principles and practice of hepatobiliary diseases. *Springer Science and Business Media*. Springer-Verlag Berlin Heidelberg Publisher, Berlin, pp 103-125.
- Dash, D. K., Yeligar, V. C., Nayak, S. S., Ghosh, T., Rajalingam, D., Sengupta, P., Maiti, B. C., Maity, T. K. (2007). Evaluation of hepatoprotective and antioxidant activity of *Ichnocarpus frutescens* (Linn.) R.Br. on paracetamol-induced hepatotoxicity in rats. *Tropical Journal of Pharmaceutical Research*. 6:755-765.
- DeAngelis, R. A, Markiewski, M. M, and Lambris, J. D. (2006). Liver regeneration: A link to inflammation through complement. *Advances in Experimental Medicine and Biology*. 586: 17-34.
- Dias, D. A., Urban, S., Roessner, U. (2012). A historical overview of natural products in drug discovery. *Metabolites*. 2: 303-336.
- Domitrović, R., Jakovac, H., Marchesi, V. V., Vladimir-Knežević, S., Cvijanović, O., Tadić, Ž., Romić, Ž and Rahelić, D. (2012). Differential hepatoprotective mechanisms of rutin and quercetin in CCl<sub>4</sub>-intoxicated BALB/cN mice. *Acta Pharmacologica Sinica*. 33: 1260-1270.

- Dong, M. W. (2006). *Modern HPLC for practicing scientists*. John Wiley & Sons Inc Publisher, New Jersey.
- Duh, P. D., Lin, S. L., and Wu, S. C. (2011). Hepatoprotective of *Graptopelatum paraguayense* E. Walther on CCl<sub>4</sub>-induced liver damage and inflammation. *Journal of Ethnopharmacology*. 134: 379-385.
- Ekor, M. (2014). The growing use of herbal medicines: issues relating to adverse reactions and challenges in monitoring safety. *Frontiers in Pharmacology*. 4: 177. <http://doi.org/10.3389/fphar.2013.00177>.
- El-Beshbishy, H. A., Mohamadin, A. M., Nagy, A. A. and Abdel-Naim, A. B. (2010). Amelioration of tamoxifen-induced liver injury in rats by grape seed extract, blackseed extract and curcumin. *Indian Journal of Experimental Biology*. 48: 280– 288.
- Fakurazi, S., Sharifudin S. A., and Arulselvan, P. (2012). *Moringa oleifera* hydroethanolic extracts effectively alleviate acetaminophen-induced hepatotoxicity in experimental rats through their antioxidant nature. *Molecules*. 17: 8334-8350.
- Fakurazi, S., Hairuszah, I., and Nanthini, U. (2008). *Moringa oleifera* Lam prevents acetaminophen induced liver injury through restoration of glutathione level. *Food and Chemical Toxicology*. 46: 2611–2615.
- Gawlik-Dziki, U., Swieca, M., Sugier, D., and Cichocka, J. (2011). Comparison of *in vitro* lipoxygenase, xanthine oxidase inhibitory and antioxidant activity of *Arnica Montana* and *Arnica chamissonis* tinctures. *Acta Scientiarum Polonorum Hortorum Cultus*. 10: 15-27.
- Giada, M. I. R. (2013). Food phenolic compounds: Main classes, sources and their antioxidant power. InTech Publisher, Croatia, pp 88-112.
- Graefe, E. U., Wittig, J., Mueller, S., Riethling, A. K., Uehleke, B., Dremelow, B., Pforte, H., Jacobasch, G., Derendorf, H., and Veit, M. (2001). Pharmacokinetics and bioavailability of quercetin glycosides in humans. *Journal of Clinical Pharmacology*. 41: 492-499.
- Gupta, R. C., Sharma, V., Sharman, N., Kumar, N., and Singh, B. (2008). *In vitro* antioxidant activity from leaves of *Oroxylum indicum* (L.) Vent.- a north Indian highly threatened and vulnerable medicinal plant. *Journal of Pharmacy Research*. 1: 65-72.
- Halliwel, B. (2008). Are polyphenols antioxidants or pro-oxidants? What do we learn from cell culture and *in vivo* studies?. *Archives of Biochemistry and Biophysics*. 476: 107-112.
- Handa, S. S., Sharma, A., Chakraborti, K. K. (1986). Natural products and plants as liver protecting drugs. *Fitoterapia*. 57: 307–345.

- Harborne, J. B. (1998). *Phytochemical methods: A guide to modern techniques of plant analysis*. Chapman and Hall, London, pp 54–84.
- Haschek, W. M., Rousseaux, C. G., and Wallig, M. A. (2009). *Fundamentals of toxicologic pathology*. Academic Press Publisher, USA, pp 196-237.
- Hashem, H. E. (2012). Light and electron microscopic study of the possible protective effect of *Nigella sativa* on metalaxyl induced hepatotoxicity in adult albino rats. *Journal of Cell Science and Therapy*. 3: 1-6.
- Hemat, R. A. S. (2004). *Principle of orthomolecularism. Urology and Health*. Urotex Publisher, USA.
- Huan, S. K. H., Wang, K. T., Lee, C. J., Sung, C. H., Chien, T. Y., and Wang, C. C. (2012). Wu-Chia-Pi solution attenuates carbon tetrachloride-induced hepatic injury through the antioxidative abilities of its components acteoside and quercetin. *Molecules*. 17: 14673-14684.
- Huang, D., Ou, B., Hampch-Woodill, M., Flanagan, J. A. and Prior, R. L. (2002). Highthroughput assay of oxygen radical absorbance capacity (ORAC) using a multichannel liquid handling system coupled with a microplate fluorescence reader in 96-well format. *Journal of Agricultural and Food Chemistry*. 5: 4437–4444.
- Huang, X. J., Choi, Y. Kyu., Im, H. S., Yarimaga, O., Yoon, Euisik., and Kim, H. S. (2006). Aspartate aminotransferase (AST/GOT) and alanine aminotransferase (ALT/GPT) detection techniques. *Sensor*. 6: 756-782.
- Huang, J., Wang, S., Zhu, M., Chen, J., and Zhu, X. (2011). Effects of genistein, apigenin, quercetin, rutin and astilbin on serum uric acid levels and xanthine oxidase activities in normal and hyperuricemic mice. *Food and Chemical Toxicology*. 49: 1943-1947.
- Hudaib, M. M., Tawaha, K. A., Mohammad, M. K., Assaf, A. M., Issa, A. Y., Alali, Q. F., Aburjai, T. A., and Bustanii, Y. K. (2011). Xanthine oxidase inhibitory activity of the methanolic extracts of selected Jordanian medicinal plants *Pharmacognosy Magazine*. 7: 320-324.
- Hussain, Md. S., Fareed, S., Ansar, S., Rahman, Md. A., Ahmad, I. Z., and Saeed, M. (2012). Current approaches toward production of secondary plant metabolites. *Journal of Pharmacy and Bioallied Sciences*. 4: 10-20.

<http://flipper.diff.org/app/items/3869> accessed on 10th July 2015.

<http://library.med.utah.edu/WebPath/LIVEHTML/LIVER006.html> accessed on 20th July 2016.

<http://vrachfree.ru/en/diseases-en/item/8702-liver-necrosis-en> accessed on 20th July 2016.

[http://www.nature.com/labinvest/journal/v92/n3/fig\\_tab/labinvest2011193f9.jpg](http://www.nature.com/labinvest/journal/v92/n3/fig_tab/labinvest2011193f9.jpg)

accessed on 10th July 2015.

- Ibrahim, I. A. A., Abdulla, M. A., Abdelwahab, S. I., Al-Bayat, F., Majid, N. A. (2012). Leaves extract of *Muntingia calabura* protects against gastric ulcer induced by ethanol in Sprague-dawley rats. *Clinical and Experimental Pharmacology*. doi:10.4172/2161.
- Ikram, E. H. K., Eng, K. H., Jalil, M. A. M., Ismail, A., Idris, Salma., Azlan, A., Mohd Nazri, H. S., Diton, M. N. A., and Mohd Mokhtar, R. A. (2009). Antioxidant capacity and total phenolic content of Malaysian underutilized fruits. *Journal of Food Composition and Analysis*. 22: 388-393.
- Jiaquan, X., Murphy, S. L., Kochanek, K. D. and Bastian, B. A. (2016). Deaths: Final Data for 2013. *National Vital Statistics Reports*. 64: 1-118.
- Kalegari, M., Bruel-Gemin, C. A., Araújo-Silva, G., Neves de Brito, N. J., Lopez, J. A., de Oliveira Tozetto, S., Almeida Md. G., Miguel M. D., Stien, D., and Miguel O. G. (2013). Chemical composition antioxidant activity and hepatoprotective potential of *Rourea induta* Planch. (Connaraceae) against CCl<sub>4</sub> induced liver injury in female rats. *Nutrition* 30: 713-718.
- Kamisan, F.H., Yahya, F., Ismail, N.A., Din, S.S., Mamat, S.S., Zabidi, Z., Zainulddin, W.N., Mohtarrudin, N., Husain, H., Ahmad, Z. and Zakaria, Z.A. (2013). Hepatoprotective activity of methanol extract of *Melastoma malabathricum* leaf in rats. *Journal of Acupuncture & Meridian Studies* 6: 52-55.
- Kaneda, N., Pezzuto, J. M., Soejarto, D. D., Kinghorn, A. D., Farnsworth, N. R. (1991). Plant anticancer agents, XLVIII. New cytotoxic flavanoids from *Muntingia calabura* roots. *Journal of Natural Products*. 54: 196-206.
- Karelina, T. A., Zhudenkova, K. V., Demin, O. O., Svetlichny, D. V., Agoram, B., Fairman, D., and Demin, O. O. (2012). Regulation of leukotriene and 5-oxoETE synthesis and the effect of 5-lipoxygenase inhibitors: a mathematical modeling approach. *Biomed Central System Biology*. 6: 1-41.
- Karthayini and Suresh, K. (2012). Pharmacognostic evaluation, *in vitro* antioxidant and *in vivo* anti-inflammatory studies of *Muntingia calabura* Linn. *Journal of Global Trends in Pharmaceutical Science*. 3: 805-811.
- Kazakevich, Y. V. and Lobrutto, R. (2007). HPLC for pharmaceutical scientists. John Wiley & Sons, New Jersey.
- Kennedy, D. O., and Wightman, E. L. (2011). Herbal extracts and phytochemicals: Plant secondary metabolites and the enhancement of human brain function. *American Society for Nutrition*. 2: 32-50.

- Khasawneh, M., Elwy, M. H., Fawzi, N. M., Hamza, A. A., Chevidenkandy, A. R. and Hassan, H. A. (2014). Antioxidant Activity and Lipoxygenase Inhibitory Effect of *Caralluma arabica* and Related Polyphenolic Constituents. *American Journal of Plant Sciences*. 5: 1623-1631.
- Koeppen, B. M. and Stanton, B. A. (2008). *Berne and Levy Physiology*. Mosby Inc Publisher, St. Louis.
- Kokate, K. C. (1997). *Practical pharmacognosy*. Vallabh Prakashan, Delhi. p. 218.
- Krishna, M. (2013). Microscopic anatomy of the liver. *Clinical Liver Disease*. 2:S1-S5.
- Krishnakumar, M. N., Latha, P. G., Suja, S. R., Shine, V. J., Shyamal, S., Anuja, G. I., Sini S., Pradeep, S., Shikha, P., Somasekharan Unni, P. K. and Rajasekharan, S. (2008). Hepatoprotective effect of *Hibiscus hispidissimus Griffith*, ethanolic extract in paracetamol and CCl<sub>4</sub> induced hepatotoxicity in Wistar rats. *Indian Journal of Experimental Biology*. 46: 653-659.
- Kumar, S., and Pandey, A. K. (2013). Chemistry and biological activities of flavonoids: An overview. *The Scientific World Journal*. <http://dx.doi.org/10.1155/2013/162750>.
- Kurt, C. K. and Kathleen, A. (2011). Chapter 12: Biochemical and metabolic principles. *Annals of Internal Medicine*. Goldfrank's Toxicologic Emergency,
- Landim, L. P., Feitoza, G. S. and da Costa, J. G. M. (2013). Development and validation of a HPLC method for the quantification of three flavonoids in a crude extract of *Dimorphandra gardneriana*. *Brazilian Journal of Pharmacognosy*. 23: 58-64.
- Lavhale, M. S. and Mishra, S. H. (2007). Evaluation of free radical scavenging activity of *Butea monosperma Lam*. *Indian Journal of Experimental Biology*. 45: 376-384.
- Lee, W. M. (2004). Acetaminophen and the U.S. acute liver failure study group: Lowering the risks of hepatic failure. *Hepatology* 40: 6-9.
- Li, R., Guo, W., Fu, Z., Ding, G., Zou, Y. and Wang, Z. (2011). Hepatoprotective action of *Radix Paeoniae Rubra* aqueous extract against CCl<sub>4</sub>-induced hepatic damage. *Molecules*. 16: 8684-8693.
- Li, S., Tan, H. Y., Wang, N., Zhang, Z. J., Lao, L., Wong, C. W. and Feng, Y. (2015). The Role of Oxidative Stress and Antioxidants in Liver Diseases. *International Journal of Molecular Sciences*. 16: 26087-26124.
- Lin, Y. C., Cheng, K. M., Huang, H. Y., Chao, P. Y., Hwang, J. M., Lee, H. H., Lu, C. Y., Chiu, Y. W. and Liu, J. Y. (2014). Hepatoprotective activity of Chhit-Chan-Thau extract powder against carbon tetrachloride-induced liver injury in rats. *Journal of Food and Drug Analysis*. 22: 220-229.



- Loganayaki, N., Suganya, N. and Manian, S. (2012). Evaluation of edible flowers of agathi (*Sesbania grandiflora* L. Fabaceae) for in vivo anti-inflammatory and analgesic, and in vitro antioxidant potential. *Food Science and Biotechnology*. 21: 509-517.
- Lupea, A. X., Pop, M. and Cacig, S. (2008). Structure-radical scavenging activity relationships of flavonoids from *Ziziphus* and *Hydrangea* extracts. *Revista de Chimie*. 59: 309-313.
- Madkour, F. F. and Abdel-Daim, M. M. (2013). Hepatoprotective and antioxidant activity of *Dunaliella salina* in paracetamol-induced acute toxicity in rats. *Indian Journal of Pharmaceutical Science*. 75: 642-648.
- Madrigal-Santillán, E., Madrigal-Bujaidar, E., Álvarez-González, I., Sumaya-Martínez, M. T., Gutiérrez-Salinas, J., Bautista, M., Morales- González, García-Luna, M., Aguilar-Faisal, J. L., and Morales- González, J. (2014). Review of natural products with hepatoprotective effects. *World Journal of Gastroenterology*. 20: 14787-14804.
- Mahadevan, V. (2014). Anatomy of the liver. *Surgery*. <http://dx.doi.org/10.1016/j.mpsur.2014.10.004>
- Mahmood, N. D., Nasir, N. L. M., Rofiee, M. S., Tohid, S. F. M., Ching, S. M., The, L. K., Salleh, M. Z. and Zakaria, Z. A. (2014). *Muntingia calabura*: A review on its traditional uses, chemical properties, and pharmacological observations. *Pharmaceutical Biology*. 52: 1598-1623.
- Malhi, H. and Gores, G. J. (2008). Cellular and molecular mechanisms of liver injury. *Gastroenterology*. 134: 1641-1654.
- Malik, A., Anis, I., Khan, S. B., Ahmed, E., Ahmed, Z., Nawaz, S. A. and Choudhary, M. I. (2004). Enzymes inhibiting lignans from *Vitex negundo*. *Chemical and Pharmaceutical Bulletin*. 52:1269–1272.
- Mamat, S. S., Kamarolzaman, M. F., Yahya, F., Mahmood, N. D., Shahril, M. S., Jakius, K. F., Mohtarrudin, N. and Zakaria, Z. A. (2013). Methanol extract of *Melastoma malabathricum* leaves exerted antioxidant and liver protective activity in rats. *BMC complementary and alternative medicine*. 13: 326.
- Markiewski, M. M, DeAngelis, R. A, and Lambris, J. D. (2006). Liver inflammation and regeneration: Two distinct biological phenomena or parallel pathophysiological processes. *Molecular Immunology*. 43: 45-56.
- Masih, N, G. and Sigh, B. S. (2012). Phytochemical screening of some plants used in herbal based cosmetic preparations. *Section A Health Perspectives*. Springer-Verlag Berlin Heidelberg, Berlin, pp111-112.
- McChesney, J. D., Venkataraman, S. K. and Henri, J. T. (2007). Plant natural products: Back to the future or into extinction?. *Phytochemistry* 68: 2015–2022.

- McGavock, H. (2011). How drugs work: Basic pharmacology for healthcare professionals. Third Edition. Radcliffe Publisher.
- McGill, M. R., Williams, C. D., Xie, Y., Ramachandran, A. and Jaeschke, H. 2012. Acetaminophen-induced liver injury in rats and mice: Comparison of protein adducts, mitochondrial dysfunction, and oxidative stress in the mechanism of toxicity. *Toxicology and Applied Pharmacology*, 264: 378-394.
- Megha, R. N., Sridevi, K. and Rao, S. N. (2006). Preliminary phytochemical analysis of fresh juice and aqueous extract of *Coleus amboinicus* Linn leaves. *International Journal of Applied Biology and Pharmaceutical Technology*. 7: 216-220.
- Mensor, L. L., Menezes, F. S., Leltão, G. G., Reis, A. S., dos Santos, T. C., Coube, C. S. and Leltão, S. G. (2001). Screening of Brazilian plant extracts for antioxidant activity by the use of DPPH free radical method. *Phytotherapy Research*. 15: 127-130.
- Meyer, B. N., Ferrigni, N. R. and Putnam, J. E. (1982). Brine shrimp: A convenient general bioassay for active plant constituents. *Planta Medica*. 45: 31-34.
- Middleton, E., Kandaswami, C. and Theoharides, C. T. (2000). The effects of plant flavonoids on mammalian cells: Implication for inflammation, heart disease, and cancer. *The American Society for Pharmacology and Experimental Therapeutics*. 52: 674-735.
- Mitra, V. and Metcalf, J. (2009). Functional anatomy and blood supply of the liver. *Physiology*. 10: 332-333.
- Mohan, C. G., Deepak, M., Viswanatha, G. L., Savinay, G., Hanumantharaju, V., Rajendra, C. E. and Praveen, D. H. (2013). Anti-oxidant and anti-inflammatory activity of leaf extracts and fractions of *Mangifera indica*. *Asian Pacific Journal of Tropical Medicine*. 6: 311-314.
- Negi, J. S., Singh, P., Pant, G. J. N. and Rawat, M. S. M. (2011). High-performance liquid chromatography analysis of plant saponins: An update 2005-2010. *Pharmacognosy Review*. 5: 155-158.
- Nemudzivhadi, V. and Masoko, P. (2008). *In vitro* assessment of cytotoxicity, antioxidant, and anti-inflammatory activities of *Ricinus communis* (Euphorbiaceae) leaf extracts. *Evidence-Based Complementary and Alternative Medicine*. 2014: 1-8.
- Newman, D.J. and Cragg, G.M. (2012). Natural products as sources of new drugs over the 30 years from 1981 to 2010. *Journal of Natural Products*. 75: 311-335.
- Ng, T. H., Khor, B. P., Ruben, Lau, K. and Wong, H. C. (2011). Clinico-epidemiology of liver cirrhosis patients treated at Hospital Tengku Ampuan Rahimah Klang. *Medical Journal of Malaysia*. 66: 35.

- Nidhi, S., Robin, S. and Sunil, K. (2012). Different model of hepatotoxicity and related liver diseases: A review. *International Journal of Pharmacy*, 3: 86-95.
- Nijveldt, R. J., Van Nood, E., Van Hoorn, D. E., Boelens, P. G., Van Norren, K. and Van Leeuwen, P. A. (2001). Flavonoids: a review of probable mechanisms of action and potential applications. *American Journal of Clinical Nutrition*. 74: 418-425.
- Nithianantham, K., Shyamala, M., Chen, Y., Latha, L. Y., Jothy, S. L. and Sasidharan, S. (2011). Hepatoprotective potential of *Clitoria ternatea* Leaf extract against paracetamol induced damage in mice. *Molecules*. 16: 10134-10145.
- Nithianantham, K., Ping, K. Y., Latha, L. Y., Jothy, S. L., Darah, I., Chen, Y., Chew, A.L. and Sasidharan, S. (2013). Evaluation of hepatoprotective effect of methanolic extract of *Clitoria ternatea* a (Linn.) flower against acetaminophen-induced liver damage. *Asian Pacific Journal of Tropical Medicine*. 3: 314-319.
- Nivethetha, M., Jayasari, J., Brindha, P. (2009). Effects of *Muntingia calabura* L. on isoproterenol-induced myocardial infarction. *Singapore Medical Journal*. 50: 300-302.
- Noori, S., Nasir, K. and Mahboob, T. (2009). Effects of cocoa powder on oxidant/antioxidant status in liver, heart and kidney tissues of rats. *The Journal of Animal and Plant Sciences*. 19: 174-178.
- Noro, T., Miyase, T. and Kuroyanagi, M. (1983). Monoamine oxidase inhibitor from the rhizomes of *Kaempferia galanga* L. *Chemical and Pharmaceutical Bulletin*. 31: 2708-2711.
- OECD. (2001). *OECD Guidelines for Testing of Chemicals*; Organisation for Economic Co-operation and Development: Paris, France, 2001; No. 423.
- Orhan, I. E., Sener, B. and Musharral, S. G. (2012). Antioxidant and hepatoprotective activity appraisal of four selected *Fumaria* species and their total phenol and flavonoid quantities. *Experimental and Toxicologic Pathology*. 64: 205-209.
- Osadebe, P. O., Okoye, F. B. C., Uzor, P. F., Nnamani, N. R., Adiele, I. E. and Obiano, N. C. (2012). Phytochemical analysis, hepatoprotective and antioxidant activity of *Alchornea cordifolia* methanol leaf extract on carbon tetrachloride-induced liver damage in rats. *Asian Specific of Tropical Medicine*. 2012: 289-293.
- Otsuka, H. (2005). Purification by solvent extraction using partition coefficient. *Natural Products Isolation*, Humana Press, pp269-273.
- Pithayanukul, P., Nithitanakool, S. and Bavovada, R. (2009). Hepatoprotective potential of extracts from seeds of *Areca catechu* and nutgalls of *Quercus infectoria*. *Molecules*. 14: 4987-5000.
- Porchezian, E. and Ansari, S. H. (2005). Hepatoprotective activity of *Abutilon indicum* on experimental liver damage in rats. *Phytomedicine* 12: 62-64.

- Preethi, K., Vijayalakshmi, N., Shamna, R. and Sasikumar, J. M. (2010). *In vitro* antioxidant activity of extracts from fruits of *Muntingia calabura* Linn. from India. *Pharmacognosy Journal*. 2:11–18.
- Preethi, K., Premasudha, P. and Keerthana, K. (2012). Anti-inflammatory activity of *Muntingia calabura* fruits. *Journal of Pharmacognosy*. 4:51–56.
- Prior, R. L., Wu, X. and Schaich, K. (2005). Standardized methods for the determination of antioxidant and phenolics in foods and dietary supplements. *Journal of Agricultural and Food Chemistry*. 53: 4290- 4302.
- Qua, C. S. and Goh, K. L. (2011). Liver cirrhosis in Malaysia: Peculiar epidemiology in a multiracial Asian country. *Journal of Gastroenterology and Hepatology*. 26: 1333-1337.
- Rajasekaran, A. and Periyasamy, M. (2012). Hepatoprotective effect of ethanolic extract of *Trichosanthes lobata* on paracetamol-induced liver toxicity in rats. *Chinese Medicine* 7: 1-6.
- Raj Kapoor, B., Venugopal, Y., Anbu, J., Harikrishnan, N., Gobinath, M. and Ravichandran, V. (2008). Protective effect of *Phyllanthus polyphyllus* on acetaminophen induced hepatotoxicity in rats. *Pakistan Journal of Pharmaceutical Sciences*. 21: 57-62.
- Ramasamy, S., Wahab, N. A., Abidin, N. Z. and Manickam, S. (2011). Cytotoxicity evaluation of five selected Malaysian Phyllanthaceae species on various human cancer cell lines. *Journal of Medicinal Plants Research* 5: 2267-2273.
- Ranawat, L., Bhatt, J. and Patel, J. (2010). Hepatoprotective activity of ethanolic extracts of bark of *Zanthoxylum armatum* DC in CCl<sub>4</sub> induced hepatic damage in rats. *Journal of Ethnopharmacology*. 127: 777-780.
- Rogers, A. B. and Dintzis, R. Z. (2012). Chapter 13; Liver and gallbladder. *Comparative anatomy and histology*. Elsevier Inc. p. 193-01.
- Rongey, C. and Kaplowitz, N. (2006). Current concepts and controversies on the treatment of alcoholic hepatitis. *World Journal of Gastroenterology*. 12: 6909-6921.
- Rubinstein, D. (1962). Epinephrine release and liver glycogen levels after carbontetrachloride administration. *American Journal of Physiology*. 203: 1033-1037.
- Saad, R. A., EL-Bab, M. F. and Shalaby, A. A. (2013). Attenuation of acute and chronic liver injury by melatonin in rats. *Journal of Taibah University for Science*. 7: 88–96.
- Sabyasachi Sircar. (2008), *Principles of Medical Physiology*. First Edition. Germany Thieme Medical Publishers, New York.

- Salama, S. M., Abdulla, M. A., Alrashdi, A. S. and Hadi, A. H. A. (2013). Mechanism of hepatoprotective effect of *Boesenbergia rotunda* in thioacetamide-induced liver damage in rats. *Evidence-Based Compelementary and Alternative Medicine*. 2013: 1-13.
- Sandhar, H. K., Kumar, B., Prasher, S., Tiwari, P., Salhan, M. and Sharma, P. (2011). A review of phytochemistry and pharmacology of flavonoids. *Internationale Pharmaceutica Scientia*. 1: 25-41.
- Sangvikar, S., Malgaonkar, M., Sharma, C., Kumar, S. and Murthy, S. N. (2015). Comparative phytochemical screening of qualitative and quantitative parameters of *Bixa orellana* L. *World Journal of Pharmacy and Pharmaceutical Sciences*. 4: 1001-1017.
- Sani, M. H., Zakaria, Z. A., Balan, T., Teh, L. K., Salleh, M. Z. (2012). Antinociceptive activity of methanol extract of *Muntingia calabura* leaves and the mechanisms of action involved. *Evidence-Based Complementary Alternative Medicine*. 2012: 1-10.
- Santillan, E. M., Bujaidar, E. M., Gonzalez, I. A., Martinez M. T. S. M., Salinas, J. G., Bautista, M., Gonzalez, A. M., Rubio, M. G. L. G., Faizal, J. L. A. and Gonzalez, J. A. M. (2014). Review of natural products with hepatoprotective effects. *World Journal of Gastroenterology*. 20: 14787-14804.
- Sellamuthu, P. S., Arulselvan, P., Kamalraj, S., Fakurazi, S., and Kandasamy, M. (2013). Protective nature of Mangiferin on oxidative stress and antioxidant status in tissues of streptozotocin-induced diabetic rats. *ISRAN Pharmacology*. 2013: 1-10.
- Sharifudin, S. A., Fakurazi, S., Hidayat, M. T., Hairuszah, I., Mohd Moklas, M. A., and Arulselvan, P. (2013). Therapeutic potential of *Moringa oleifera* extracts against acetaminophen-induced hepatotoxicity in rats. *Pharmaceutical Biology*. 51: 279-288.
- Sibi, G., Naveen, R., Dhananjaya, K., Ravikumar, K. R., and Mallesha, H. (2012). Potential use of *Muntingia calabura* L. extracts against human and plant pathogens. *Journal of Pharmacognosy*. 4: 44-47.
- Siddiqua, A., Premakumari, K. B., Sultana, R., Vithya, and Savitha. (2010). Antioxidant activity and estimation of total phenolic content of *Muntingia calabura* by colorimetry. *International Journal of Chemical Technology Research*. 2:205-208.
- Singh, G., Goyal, R., and Sharma, P. L. 2012. Pharmacological potential of silymarin in combination with hepatoprotective plants against experimental hepatotoxicity in rats. *Asian Journal of Pharmaceutical and Clinical Research*, 5: 128-133.
- Singleton, V. L. and Rossi J. A. Jr. (1965). Colorimetry of total phenolics with phosphomolybdcphosphotungstic acid reagents. *American Journal of Enology and Viticulture* 16: 144-158.

- Sowndhararajan, K. and Kang, S. C. (2013). Protective effect of ethyl acetate fraction of *Acacia ferruginea* DC. Against ethanol-induced gastric ulcer in rats. *Journal of Ethnopharmacology*.148: 175-181.
- Su B. N., Park, E. J., Vigo, J. S., Graham, J. G., Cabieses, F., Fong, H. H. S., Pezzuto, J. M. and Kinghorn, A. D. (2003). Activity-guided isolation of the chemical constituents of *Muntingia calabura* using a quinine reductase induction assay. *Phytochemistry*. 63:335–341.
- Sufian, A. S., Ramasamy, K., Ahmat, N, Zakaria, Z. A. and Mohd Yusof, M. I. (2013). Isolation and identification of antibacterial and cytotoxic compounds from the leaves of *Muntingia calabura* L. *Journal of Ethnopharmacology*.146: 198-204.
- Swierkosz, T. A., Jordan, L., McBride, M., McGough, K., Devlin, J. and Botting, R. M. (2002) Actions of paracetamol on cyclooxygenases in tissue and cell homogenates of mouse and rabbit. *Medical Science Monitor*. 8: 496–503.
- Syed, S. H. and Namdeo, A. G. (2014). Current status of natural products for the treatment of liver disease-A review. *International Journal of Phytopharmacy*. 4:37-43.
- Teoh, N. C., and Farrel, G. C. (2003). Hepatic ischemia reperfusion injury: Pathogenic mechanisms basis for hepatoprotection. *Journal of Gastroenterology and Hepatology*. 18: 891-902.
- Thapa, B. R. and Walia, A. (2007). Liver function tests and their interpretation. *Indian Journal of Pediatrics*. 74: 663-671.
- Tiwari, P., Kumar, B., Kaur, M., Kaur, G. and Kaur, H. (2011). Phytochemical screening and extraction: A review. *Internationale Pharmaceutica Scientia*. 1: 98-106.
- Tsimogiannis, D., Samiotaki, M., Panayotou, G. and Oreopoulou, V. (2007). Characterization of flavonoid subgroups and hydroxy substitution by HPLC MS/MS. *Molecules* 12: 593–606.
- Umukoro, S. and Ashorobi, R. B. (2006). Evaluation of anti-inflammatory and membrane stabilizing property of aqueous leaf extract of *Momordica charantia* in rats. *African Journal of Biomedical Research*. 9: 11-124.
- Vimala, S., Rohana, S., Rashih, A. A. and Juliza, M. (2012). Antioxidant Evaluation in Malaysian Medicinal Plant: *Persicaria minor* (Huds.) Leaf. *Science Journal of Medicine and Clinical Trials*, 2012.
- Wang, J., Yuand, X., Jin, Z., Tian, Y., and Song, H. (2007). Free radical and reactive oxygen species scavenging activities of peanut skins extract. *Food chemistry*. 104: 242-250.

- Webb, C and Twedt, D. (2008). Oxidative stress and liver disease. *Veterinary Clinics Small Animal Practice*. 38: 125-135.
- Wisastra, R. and Dekker, F. J. (2014). Inflammation, cancer and oxidative lipoxigenase activity are intimately linked. *Cancers*. 6: 1500-1521.
- World Health Organization. (2002). Traditional medicine strategy launched. *Bulletin of the World Health Organization*. 80: 7.
- World Health Organization. (2015) Climate change and human health. <http://www.who.int/globalchange/ecosystems/biodiversity/en/>
- Xu, B. J. and Chang, S. K. (2007). A comparative study on phenolic profiles and antioxidant activities of legumes as affected by extraction solvents. *Journal of Food Science*. 72: S159-166.
- Yanpallewar, S. U., Sen, S., Tapas, S., Kumar, M., Raju, S. S. and Acharya, S. B. (2002). Effect of *Azadirachta indica* on paracetamol-induced hepatic damage in albino rats. *Phytomedicine*. 10: 391-396.
- Yasunaka, K., Abe, F., Nagayama, A., Okabe, H., Lozada-Perez, L., Lopez-Villafranco, E., Muniz, E. E., Aquilar, A., Reyes-Chilpa, R. (2005). Antibacterial activity of crude extracts from Mexican medicinal plants and purified coumarins and xanthenes. *Journal of Ethnopharmacology*. 97: 293-299.
- Yumita, A., Suganda, A. G. and Sukandar, E. Y. (2013). Xanthine oxidase inhibitory activity of some Indonesian medicinal plants and active fraction of selected plants. *International Journal of Pharmacy and Pharmaceutical Sciences*. 4: 293-296.
- Yusof, M. I. M., Salleh, M. Z., Kek, T. L., Ahmat, N., Nik Azmin, N. F., and Zakaria, Z. A. (2013). Activity-guided isolation of bioactive constituents with antinociceptive activity from *Muntingia calabura* L. leaves using the formalin test. *Evidence-Based Complementart and Alternative Medicine*. 2013: 1-9.
- Zabidi, Z., Wan Zainulddin, W. N., Mamat, S. S., Shamsahal Din, S., Kamisan, F. H., Yahya, F., Ismail, N.A. and Zakaria, Z. A. (2012). Antiulcer activity of methanol extract of *Melastoma malabathricum* leaves in rats. *Medical Principles and Practice* 21: 501-503.
- Zain, M. Z., Fathelrahman, A. I. and Ab Rahman, A. F. (2006). Characteristics and outcomes of paracetamol poisoning cases at a general hospital in Northern Malaysia. *Singapore Medical Journal*. 47: 134-137.
- Zakaria, Z. A., Sulaiman, M. R., Jais, A. M. M., Somchit, M. Z. and Jayaraman, K. V. (2006). The antinociceptive activity of *Muntingia calabura* aqueous extract and the involvement of L-arginine/nitric oxide/cyclic guanosine monophosphate pathway in its observed activity in mice. *Fundamental and Clinical Pharmacology*. 20: 365-372.

- Zakaria, Z. A., Fatimah, C.A., Jais, A. M. M., Zaiton, H., Henie, E. F. P., Sulaiman, M. R., Somchit, M. N., Thenamutha, M. and Kasthuri, D. (2006b). The *in vitro* antibacterial activity of *Muntingia calabura* extract. *Journal of Pharmacology*. 2: 290-293.
- Zakaria, Z. A., Jais, A. M. M., Mastura, M. Jusoh, M. S. H., Mohamed, A. M., Jamil, M. N. S., Rofiee, M. S. and Sulaiman, M. R. (2007). *In vitro* antistaphylococcal activity of the extracts of several neglected plants in Malaysia. *Journal of Pharmacology*. 3: 428-431.
- Zakaria, Z. A., Kumar, G. H., Zaid, S. N. H., Ghani, M. A., Hassan, M. H., Hazalin, N. A. M. N., Khamis, M. M., and Sulaiman, M. R. (2007a). Analgesic and antipyretic actions of *Muntingia calabura* leaves chloroform extract in animal models. *Oriental Pharmacy and Experimental Medicine*. 7:34–40.
- Zakaria, Z. A., Hassan, M. H., Aqmar, N. M. N. H, Ghani, M. A., Zaid, M. S. N. H., Sulaiman, M. R., Kumar, G. H. and Fatimah, C. A. (2007d). Effects of various nonopioid receptor antagonist on the antinociceptive activity of *Muntingia calabura* extracts in mice. *Methods and Findings in Experimental Clinical Pharmacology*.29: 515-520.
- Zakaria, Z. A., Mustapha, S., Sulaiman, M. R., Jais, A. M. M., Somchit, M. N. and Fatimah, C. A. (2007e). The antinociceptive action of aqueous extract from *Muntingia calabura* leaves: The role of opioid receptors. *Medical Principles and Practice*.16: 130–136.
- Zakaria, Z. A., Hazalin, N. A. M. N., Zaid, S. N. H. M., Abdul Ghani, M., Hassan, M. H., Gopalan, H. K. and Sulaiman, M. R. (2007f). Antinociceptive, anti-inflammatory and antipyretic effects of *Muntingia calabura* aqueous extract in animal models. *Journal of Natural Medicines*. 61:443–448.
- Zakaria, Z. A., Mohamed, A. M., Jamil, N. S. M., Rofiee, M. S., Hussain, M. K., Sulaiman, M. R., Teh, L. K. and Salleh, M. Z. (2011). *In vitro* antiproliferative and antioxidant activities of the extracts of *Muntingia calabura* leaves. *American Journal of Chinese Medicine*. 39:1–18.
- Zakaria, Z. A., Rofiee, M. S., Somchit, M. N., Zuraini, A., Sulaiman, M. R., Teh, L. K., Salleh, M.Z. and Long, K. (2011c). Hepatoprotective activity of dried-and fermented-processed virgin coconut oil. *Evidence-Based Complementary and Alternative Medicine*. 5: 2526-2536.
- Zhang A., Sun H. and Wang X. (2013). Recent advances in natural products from plants for treatment of liver diseases. *European Journal of Medicinal Chemistry*. 63: 570-577.
- Zhou, G., Chen, Y., Liu, S., Yao, X., and Wang, Y. 2013. In vitro and in vivo hepatoprotective and antioxidant activity of ethanolic extract from *Meconopsis integrifolia* (Maxim.) Franch. *Journal of Ethnopharmacology*, 148: 664- 670.
- Zhu, R, Wang, Y, Liangqing, Z, and Guo, Q. (2012). Oxidative stress and liver disease. *Hepatology Research*. 42: 741-749.