



**UNIVERSITI PUTRA MALAYSIA**

***COMPARISON BETWEEN SEVERE AND SURVIVAL MODELS OF  
MALARIA INFECTION FOR BETTER UNDERSTANDING OF THE  
UNDERLYING DISEASE PATHOGENESIS***

**CHONG WING CHUI**

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**MASTER OF SCIENCE  
UNIVERSITI PUTRA MALAYSIA**

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By

**CHONG WING CHUI**

**This thesis submitted to the School of Graduate Studies,  
Universiti Putra Malaysia, in fulfillment of the  
Requirements for the Degree of Master of Science**

**November 2013**

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

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**CHONG WING CHUI**

**November 2013**

**Chair: Associate Professor Rusliza Basir, PhD**

**Faculty: Medicine and Health Sciences**

Different host-parasite combination will result in varied malaria presentation and outcome. In this preliminary study, ICR mice and *Sprague dawley* rats were inoculated intraperitoneally with  $2 \times 10^7$  parasitized red blood cells (pRBCs) obtained from donor mouse pre-infected with *P. berghei* ANKA (PbA). Control to malaria-infected animals received an equivalent volume and dilution of normal RBC through the same route of administration. The susceptible mice which serve as severe model demonstrated a parasitaemia level of approximately 75% together with a progressive decrease in body weight and temperature before succumb to the infection. Malarial rats that survived the infection developed moderate level of parasitaemia (45%) and insignificant changes in terms of illness symptoms and behavioural responses followed with gradual resolution. Multiple organ dysfunctions is also an important contributor towards malaria-associated mortality, hence histopathological examination

was performed on H&E stained tissue obtained from five vital organs including brain, liver, spleen, kidney and lung. Some of the major histopathological alterations observed in both mice and rats including sequestration of pRBCs, accumulation of hemozoin pigment and macrophage engulfing elements in microvasculature, disorganization of spleen architecture, and hyalinized membrane in alveolar wall. Systemic concentration of pro-inflammatory cytokines (TNF- $\alpha$ , IFN $\gamma$  and IL-1 $\alpha$ ) and anti-inflammatory cytokines (IL-10, IL-4 and IL-13) were also quantified in mice and rats by using commercial ELISA kit. A distinctive difference obtained from analyzed data showed significant elevation of plasma TNF- $\alpha$  in malarial mice which contradict with low production of TNF- $\alpha$  along with significant concentrations of IL-10, IL-4 and IL-13 in serum of malarial rats. This pattern of cytokine release has tipped towards the regulation of inflammatory response and survival during malaria infection. The protective role of the above said anti-inflammatory cytokines is not well elucidated, nor is there a relevance of these cytokines with the disease pattern or extent of vital organ dysfunction during the infection. Hence, the effects of systemic augmentation of IL-10, IL-4 and IL-13 on pathological conditions of malaria were investigated. *P. berghei* ANKA (PbA)-infected mice were treated with either the recombinant mouse (rm) IL-10, rmIL-4 or rmIL-13 and treatment with these cytokines has successfully delayed the mortality rate in all malarial mice with development of parasitaemia was reduced to as much as 20% during peak parasitaemia in relative to malaria mice receiving PBS. Body weight and rectal temperature also showed lesser extent of reduction. Results revealed the amelioration of malaria histopathological conditions in all examined organs upon treatment. Sequestration of pRBCs was

reduced in the brain tissue. Hypertrophied Kupffer cells and sinusoids dilatation that were significant during malaria infection was lessened by treatment. The red and white pulp elements and central germinal structure were regained in spleen. Glomerular and tubular appearance of the treated malarial kidney appeared to be normal whilst the lung tissue of treated malarial mice were revealed to be normal without formation of hyalinized membrane and pigment deposition in alveolar septa. Taken together, it can be concluded that IL-10, IL-4 and IL-13 play significant role(s) during malaria infection and they may well serve as potential immunotherapeutic target strategy in malaria therapy.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

**PERBANDINGAN ANTARA MODEL JANGKITAN MALARIA TENAT DAN BERDAYA HIDUP UNTUK PEMAHAMAN YANG LEBIH BAIK TENTANG PATOGENESIS YANG MENDASARI PENYAKIT**

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Kombinasi hos-parasit yang berbeza akan menghasilkan jenis malaria yang berlainan. Dalam kajian awal ini, mencit ICR dan tikus *Sprague dawley* diinokulasikan secara intraperiteum dengan  $2 \times 10^7$  sel-sel darah merah berparasit (pRBCs) yang diperolehi dari mencit penderma yang dijangkiti terlebih dahulu dengan *P. berghei* ANKA (PbA). Kawalan kepada haiwan terjangkit menerima isipadu dan pencairan sel-sel darah merah normal yang setara melalui cara administrasi yang sama. Mencit rentan yang berfungsi sebagai model tenat mempamerkan tahap parasitemia lebih kurang 75% beserta dengan pengurangan berat dan suhu badan yang progresif sebelum tewas terhadap jangkitan. Tikus malaria yang mampu bertahan terhadap jangkitan menghasilkan tahap parasitemia yang sederhana dan perubahan-perubahan yang tidak ketara dari segi simptom penyakit dan gerakbalas-gerakbalas tingkahlaku diikuti dengan penyembuhan secara beransur-ansur. Kegagalan fungsi organ berganda juga adalah merupakan penyumbang penting kepada mortaliti yang dikaitkan dengan



malaria, jadi pemeriksaan histopatologi dilakukan ke atas tisu terwarna H&E yang diperolehi daripada lima organ utama termasuk otak, hati, limpa, ginjal dan paru-paru. Perubahan-perubahan histopatologi utama yang diperhatikan dalam kedua-dua tikus dan mencit termasuklah sekuestrasi pRBCs, longgokan pigmen hemozoin dan elemen-elemen diliputi makrofaj di dalam mikrovaskulatur, arkitektur limpa yang berselerak dan membran terhalang dalam dinding alveolar. Kepekatan sistemik sitokin-sitokin proinflamasi (TNF $\alpha$ , IFN $\gamma$  dan IL-1 $\alpha$ ) dan sitokin-sitokin antiinflamasi (IL-10, IL-4 dan IL-13) di dalam mencit dan tikus juga ditentukan menggunakan peralatan ELISA komersil. Perbezaan ketara diperolehi dari data yang dianalisis menunjukkan peningkatan signifikan TNF $\alpha$  plasma dalam mencit malaria di mana ini berlawanan dengan pengeluaran TNF $\alpha$  yang rendah beserta dengan kepekatan IL-10, IL-4 dan IL-13 yang signifikan dalam serum tikus malaria. Corak pembebasan sitokin ini memberikan petunjuk terhadap pengawalaturan gerakbalas inflamasi dan kelangsungan hidup semasa jangkitan malaria. Fungsi perlindungan sitokin-sitokin antiinflamasi yang disebutkan di atas tidak diterangkan dengan baik, dan tiada hubungkait sitokin-sitokin ini dengan corak penyakit atau kegagalan fungsi organ utama semasa jangkitan. Oleh yang demikian, kesan penambahan sistemik IL-10, IL-4 dan IL-13 ke atas keadaan patologi malaria diselidiki. Mencit terjangkit *P. berghei* ANKA (PbA) dirawat dengan rekombinan mencit (rm) IL-10, rmIL-4 atau rmIL-13 dan rawatan dengan ketiga-ketiga sitokin ini telah berjaya melewati kadar mortaliti dalam kesemua mencit malaria dengan perkembangan parasitemia berkurangan sebanyak 20% berbanding mencit malaria yang dirawat dengan PBS. Berat dan suhu badan juga menunjukkan kadar pengurangan yang rendah. Keputusan menunjukkan

pembaikan keadaan histopatologi malaria di dalam kesemua organ-organ yang diperiksa selepas rawatan. Sekuestrasi pRBC berkurangan di dalam tisu otak. Sel-sel kupffer yang terhipertrofi dan dilatasi sinusoid yang signifikan semasa jangkitan malaria berkurangan dengan rawatan. Elemen-elemen palpa merah dan putih dan juga struktur pusat germinal di dalam limpa pulih semula. Keadaan glomerulus dan tubul ginjal malaria yang dirawat kelihatan normal, manakala tisu paru-paru menciit malaria yang dirawat kelihatan normal tanpa pembentukan membran terhialin dan longgokan pigment di dalam septa alveolar. Secara keseluruhan, ianya boleh disimpulkan bahawa IL-10, IL-4 dan IL-13 memainkan peranan yang signifikan semasa jangkitan malaria dan mungkin boleh menjadi sasaran imunoterapeutik yang berpotensi dalam terapi malaria.

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I certify that a Thesis Examination Committee has met on 21 November 2013 to conduct the final examination of Chong Wing Chui on her thesis entitled “Comparison between survival and severe models of malaria infection for better understanding of the underlying disease pathogenesis” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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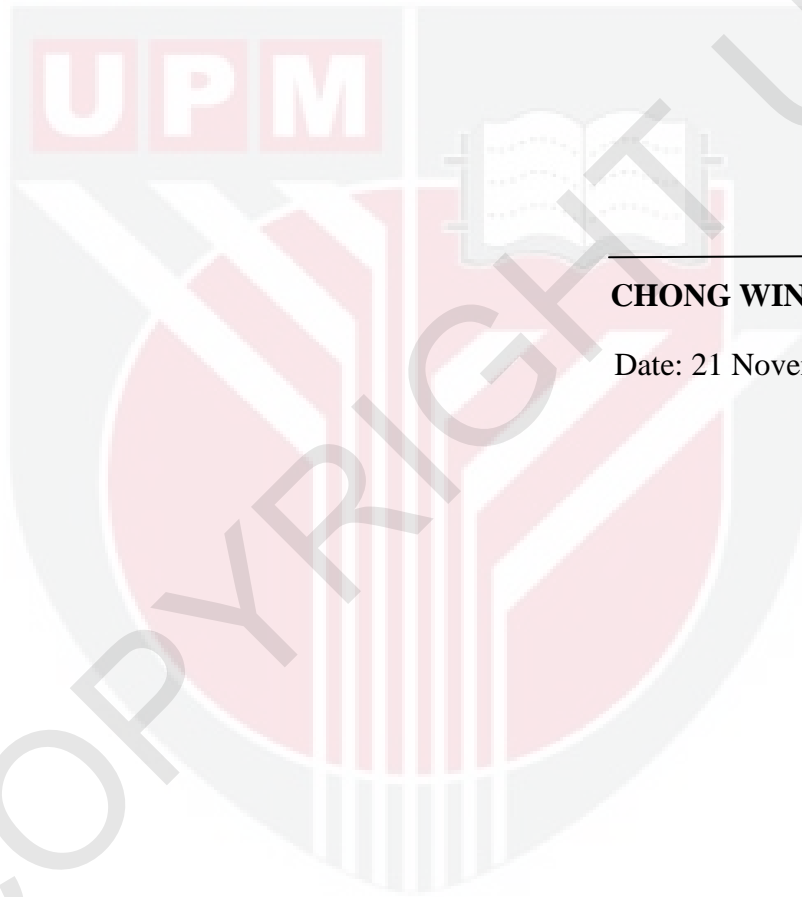
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## DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted to any other degree at Universiti Putra Malaysia or at any other institution.



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**CHONG WING CHUI**

Date: 21 November 2013

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## LIST OF ABBREVIATIONS

ACT	:	artemisinin-based combination therapies
ACUC	:	animal care and use committee
ADCC	:	antibody dependent cellular cytotoxicity
ADCI	:	antibody dependent cellular inhibition
APC	:	antigen presenting cells
ARDS	:	acute respiratory distress
ATN	:	acute tubular necrosis
BBB	:	blood brain barrier
°C	:	degree Celcius
CM	:	cerebral malaria
CSA	:	chondroitin sulphate A
dH <sub>2</sub> O	:	deionized water
DNA	:	double helix nucleic acid
ELISA	:	Enzyme Linked Immunosorbent Assay
<i>et al</i>	:	elsewhere or and others
etc	:	et cetera
g	:	gram
GPI	:	glycosyl phosphotidyl inositol
H&E	:	hematoxylin and eosin
HLA	:	human leukocyte antigen
hr	:	hour

HRP	:	horseradish peroxidase
ICAM-1	:	intercellular adhesion molecule-1
ICR mice	:	imprinting control region mice
IFN $\gamma$	:	interferon gamma
Ig	:	immunoglobulin
IL-	:	interleukin
IL1R1	:	Interleukin1 receptor 1
iNOS	:	inducible nitric oxide synthase
i.p	:	intraperitoneal
i.v	:	intravenous
kDa	:	kilodalton
kg	:	kilogram
L	:	liter
LPS	:	lipopolysaccharide
MHC	:	major histocompatibility complex
min	:	minute
mL	:	milliliter
mm	:	millimeter
mmol	:	millimolar
n	:	number of observation
ng	:	nanogram
NK Cells	:	Natural Killer cells
NKT	:	Natural Killer T lymphocyte

nm	:	nanometer
NO	:	Nitric Oxide
PbA	:	<i>Plasmodium berghei</i> ANKA
PBS	:	phosphate buffered saline
%	:	percent
pg	:	picogram
pRBC	:	parasitized red blood cell
RBC	:	red blood cell
RNI	:	reactive nitrogen intermediates
ROI	:	reactive oxygen intermediates
rpm	:	revolution per minute
rmIL-4	:	recombinant mouse Interleukin-4
rmIL-10	:	recombinant mouse Interleukin-10
rmIL-13	:	recombinant mouse Interleukin-13
S.E.M	:	standard error of the mean
Stat6	:	signal transducer and activator of transcription 6
TGF- $\beta$	:	transforming growth factor-beta
Th <sub>1</sub>	:	T helper type 1
Th <sub>2</sub>	:	T helper type 2
TNF- $\alpha$	:	tumor necrosis factor-alpha
TNFR2	:	tumor necrosis factor receptor 2
Treg cells	:	T regulatory cells
$\mu$ g	:	microgram

$\mu\text{L}$  : microliter  
VCAM-1 : vascular cell adhesion molecule-1  
WHO : World Health Organization



# CHAPTER 1

## INTRODUCTION

### 1.1 Background

The contest between malaria and human has begun since mankind evolved, and it continues unabated till present, with no resolution in sight (Mohr, 2002). In the company of more than 200 million clinical cases and estimated 700000 deaths per annum, malaria is one of the major vectorborne diseases which now rivals HIV/AIDS as the world most deadly infection (Guerin *et al.*, 2002; WHO, 2010). To date, there are five *Plasmodium* protozoa namely *Plasmodium vivax* (*P. vivax*), *Plasmodium falciparum* (*P. falciparum*), *Plasmodium malariae* (*P. malariae*), *Plasmodium ovale* (*P. ovale*), and the recent discovered simian *Plasmodium knowlesi* (*P. knowlesi*) that infect human with malaria through the Anopheles mosquito (Campbell, 2009; Lee *et al.*, 2011; White 2008). Infections caused by *P. vivax* and *P.falciparum* are reported extensively worldwide, along with its high mortality rate and drug resistance. Until recently, *P. knowlesi* infection in human has become a significant malaria infection in Southeast Asia that may lead to deadly complications (Cox-Singh *et al.*, 2008; Snow *et al.*, 2005). With widespread air travel, malaria is now prevailing and threatening developed countries too, with two-thirds of the global population at risk (Vernicket *al.*, 2004).

Once malaria parasite intrudes successfully into the host, liver cells are the bed where they first grow and multiply. Then they multiplies asexually in the red blood cells at

which most of the clinical symptoms such as chills, headaches and fever begin to occur (Davis, 2010). Pathogenesis of malaria can be divided into severe and survival model (Craig *et al.*, 2012). It is noteworthy that the relationship between the host's genetic profile and the susceptibility towards malaria is intricately intertwined. Conferred genetic resistance of malaria not only enhanced by modifications of immune system such as the major histocompatibility complex gene, but also by certain haemoglobin inherited disorders or erythrocyte polymorphisms (Lopez *et al.*, 2010). It is also believed that the involvement of environmental factors, parasite genetic factor, and multi-gene interaction play pivotal roles as well (Kwiatkowski, 2005).

Control of malaria infection is critical due to rapid increment of mosquito insecticide resistance and parasite drug resistance. To date, many attempts have been made to facilitate the development of a more effective treatment and efficacious malaria vaccine. Incomplete understanding of protective immunity and its core induction process has hindered the progress of effective malaria control measures (Kumar *et al.*, 2002; Richie *et al.*, 2002). In this regard, a greater effort on investigating prospective targets and mechanisms of immunity to malaria on both severe and survival models may provide better description on pathological versus protective responses and this provide more efficacious immunological interventions (Langhorne *et al.*, 2008).

Animal models have been principal, in that they provide much valuable information to understanding malaria, particularly the accumulation of infected red blood cells in a number of organs and microvascular obstruction induced by cytoadherence and

inflammation (Craig *et al.*, 2012). To date, most of the current anti-malarial strategies are targeting on *Plasmodium* parasites, however malaria remained to be endemic due to anti-malarial drug resistance (Lee *et al.*, 2013). To solve this problem, it is hypothesized that cytokines may be one of the potential targets for immunotherapeutic strategy in malaria. Albeit the well-documented pathogenesis studies in malaria infection, there is still a large gap to be filled for the understanding of the host's immune responses during malaria infection. In accordance with this, two different models of mice and rats representing the severe and survival mode of pathogenesis, respectively were conducted throughout this research to add evidence to immunological studies of the infection. With that, the development of future malaria treatment and vaccines are plausible.

## **1.2 Objectives**

The main objective of this study is to investigate the effectiveness of anti-inflammatory cytokines in preventing severe malarial infection.

The specific objectives are:

- i. to establish survival and severe models in mice and rats, respectively.
- ii. to determine and differentiate behavioural, clinical, histopathology, pro- and anti-inflammatory cytokine levels and survival in these models.

## REFERENCES

- About malaria-biology*; Center for Disease Control and Prevention: Atlanta, GA, 2010.
- Adam, E., Pierrot, C., Lafitte, S., Godin, C., Saoudi, A., Capron, M. and Khalife, J. (2003). The age-related resistance of rats to *Plasmodium berghei* infection is associated with differential cellular and humoral immune responses. *International Journal of Parasitology*. 33(10): 1067-1078.
- Adams, S., Brown, H. and Turner, G. (2002). Breaking down the blood-brain-barrier: signaling a pathway to cerebral malaria? *Trends in Parasitology*. 18(8): 360-366.
- Afzal, M.S., Ullah, S., Farooqi, Z.U.R., Anjum, S., Shafi, T., Ahmed, T., Ashraf, M. and Qadri, I. (2012). Association of IL-10 polymorphism and malarial susceptibility in Pakistani population. *Asian Biomedicine*. 6(3): 337-342.
- Aikawa, M., Iseki, M., Barnwell, J.W., Taylor, D., Oo, M.M. and Howard, R.J. (1990). The pathology of human cerebral malaria. *American Journal of Tropical Medicine and Hygiene*. 43(2): 30-37.
- Alexander, C. and Rietschel, E.T. (2001). Bacterial lipopolysaccharides and innate immunity. *Journal of Endotoxin Research*. 7(3): 167-202.
- Amani, V., Vigario, A.M., Belnoue, E., Marussig, M., Fonseca, L., Mazier, D. and Renia, L. (2000). Involvement of IFN- $\gamma$  receptor-mediated signaling in pathology and anti-malarial immunity induced by *Plasmodium berghei* infection. *European Journal of Immunology*. 30(6): 1646-1655.
- Amante, F.H., Haque, A., Stanley, A.C., Rivera, F.L., Randall, L.M., Wilson, Y.A., Yeo, G., Pieper, C., Crabb, B.S., de Koning-Ward, T.F., Lundie, R.J., Good, M.F., Pinzon-Charry, A., Pearson, M.S., Duke, M.G., McManus, D.P., Loukas, A., Hill, G.R. and Engwerda, C.R. (2010). Immune-mediated mechanisms of parasite tissue sequestration during experimental cerebral malaria. *The Journal of Immunology*. 18(6): 3632-3642.



- Andrade, B.B., Reis-Filho, A., Souza-Neto, S.M., Clarencio, J., Camargo, L.M., Barral, A. and Barral-Netto, M. (2010). Severe *Plasmodium vivax* malaria exhibits marked inflammatory imbalance. *Malaria Journal*. 9:13.
- Angulo, I., and Fresno M. (2002). Cytokines in the pathogenesis of and protection against malaria. *Clinical and Drug Laboratory Immunology*. 9(6): 1145-1152.
- Anidi, I.U., Servinsky, L.E., Rentsendorj, O., Gao, J., Scott, A.L. and Pearse, D.B. (2011). Lung endothelial barrier dysfunction in murine malaria: role of CD36 and soluble guanylyl cyclase (SGC). *American Journal of Respiratory and Critical Care Medicine*. 183: A3755.
- Anstey, N.M., Jacups, S.P., Cain, T., Pearson, T., Ziesing, P.J., Fisher, D.A., Currie, B.J., Marks, P.J. and Maguire, G.P. (2002). Pulmonary manifestations of uncomplicated falciparum and vivax malaria: Cough small airway obstruction, impaired gas transfer and increased pulmonary phagocytic activity. *The Journal of Infectious Diseases*. 185(9): 1326-1334.
- Arevalo-Herrera, M., Soto, L., Perlaza, B.L., Cespedes, N., Vera, O., Lenis, A.M., Bonelo, A., Corradin, G. and Herrera, S. (2011). Antibody-mediated and cellular immune responses induced in naïve volunteers by vaccination with long synthetic peptides derived from the *Plasmodium vivax* circumsporozoite protein. *American Journal of Tropical Medicine and Hygiene*. 84(Suppl 2): 35-42.
- Armani, V., Boubou, M.I., Pied, S., Marussig, M., Walliker, D., Mazier, D. and Renia, L. (1998). Cloned lines of *Plasmodium berghei* ANKA differ in their abilities to induce experimental cerebral malaria. *Infection and Immunity*. 66(9): 4093-4099.
- Artavanis-Tsakonas, K. and Riley, E.M. (2002). Innate immune response to malaria: rapid induction of IFN-gamma from human NK cells by live *Plasmodium falciparum*-infected erythrocytes. *The Journal of Immunology*. 169(6): 2956-2963.
- Artavanis-Tsakonas, K., Tongren, J.E. and Riley E.M. (2003). The war between malaria parasite and the immune system: immunity, immunoregulation and immunopathology. *Clinical and Experimental Immunology*. 133(2): 145-152.

- Bagot, S., Boubou, M.I., Campino, S., Behrschmidt, C., Gorgetto, O., Guenet, J.L., Penha-Goncalves, C., Mazier, O., Pied, S. and Cazenave, P.A. (2002). Susceptibility to experimental cerebral malaria induced by *Plasmodium berghei* ANKA in inbred mouse strains recently derived from wild stock. *Infection and Immunity*. 70(4): 2049-2056.
- Baheti, R., Laddha, P. and Gehlot, R.S. (2003). Liver involvement in falciparum malaria – A histo-pathological analysis. *Journal of Indian Academy of Clinical Medicine*. 4(1): 34-38.
- Bakir, H.Y., Tomiyama, C. and Abo, T. (2011). Cytokine profile of murine malaria: stage-related production of inflammatory and anti-inflammatory cytokines. *Biomedical Research*. 32(3): 203-208.
- Ball, H.J., MacDougall, H.G., McGregor, I.S. and Hunt, N.H. (2004). Cyclooxygenase-2 in the pathogenesis of murine cerebral malaria. *The Journal of Infectious Disease*. 189(4): 751-758.
- Bashyam, H. (2007). IFN $\gamma$ : issuing macrophages a license to kill. *The Journal of Experimental Medicine*. 204(1): 3.
- Barillas-Mury, C. and Kumar, S. (2005). Plasmodium-mosquito interactions: a tale of dangerous liaisons. *Cellular Microbiology*. 7(11): 1539-1545.
- Barsoum, R.S. (2000). Malarial acute renal failure. *Journal of American Society of Nephrology*. 11(11): 2147-2154.
- Bartoloni, A. and Zammarchi, L. (2012). Clinical aspects of uncomplicated and severe malaria. *Mediterranean Journal of Hematology and Infectious Diseases*. 4(1): e20120.
- Bechmann, I., Kwidzinski, E., Kovac, A.D., Simburger, E., Horvath, T., Gimsa, U., Dirnagi, U., Priller, J. and Nitsch, R. (2001). Turnover of rat brain perivascular cells. *Experimental Neurology*. 168(2): 242-249.
- Belkaid, Y. (2007). Regulatory T cells and infection: a dangerous necessity. *Nature Reviews Immunology*. 7(11): 875-888.
- Bhalla, A., Suri, V. and Singh, V. (2006). Malarial hepatopathy. *Journal of Postgraduate Medicine*. 52(4): 315-320.

- Blanque, R., Meakin, C. and Gardner, C.R. (1996). Hypothermia as an indicator of the acute effects of lipopolysaccharides: comparison with serum levels of IL1 beta, IL6 and TNF alpha. *General Pharmacology*. 27(6): 973-977.
- Blanque, R., Meakin, C., Millet, S. and Gardner, C.R. (1998). Seletive enhancement of LPS-induced serum TNF alpha production by carrageenan pretreatment in mice. *General Pharmacology*. 31(2): 301-306.
- Boelan, A., Platvoet-ter Schiphorst, M.C. and Wiersinga, W.M. (1997). Immunoneutralization of interleukin-1, tumor necrosis factor, interleukin-6 or interferon does not prevent the LPS-induced sick euthyroid syndrome in mice. *Journal of Endocrinology*. 153(1): 115-122.
- Boeuf, P.S., Loizon, S., Awandare, G.A., Tetteh, J.K., Addae, M.M., Adjei, G.O., Goka, B., Kutzhals, J.A., Puijalon, O., Hviid, L., Akanmori, B.D. and Behr, C. (2012). Insights into deregulated TNF and IL-10 production in malaria: implications for understanding severe malarial anaemia. *Malaria Journal*. 11: 253.
- Bosmann, G.J., Willekens, F.L. and Were, J.M. (2005). Erythrocyte aging: a more than superficial resemblance to apoptosis. *Cellular Physiology and Biochemistry*. 16(1-3): 1-8.
- Brown, H., Turner, G., Rogerson, S., Tembo, M., Mwenechanya, J., Molyneux, M. and Taylor, T. (1999). Cytokine expression in the brain in human cerebral malaria. *Journal of Infectious Diseases*. 180(5): 1742-1746.
- Bueno, L.L., Morais, C.G., Araujo, F.F., Gomes, J.A.S., Cornea-Oliveira, R., Soares, I.S., Lacerda, M.V., Fujiwara, R.T. and Braga, E.M. (2010). *Plasmodium vivax*: induction of CD4<sup>+</sup>CD25<sup>+</sup>FoxP3<sup>+</sup> regulatory T cells during infection are directly associated with level of circulating parasites. *PLoS ONE*. 5(3): e9623.
- Buffet, P.A., Safeukui, I., Milon, G., Mercereau-Puijalon, O. and David, P.H. (2009). Retention of erythrocytes in the spleen: a double-edged process in human malaria. *Current Opinion in Hematology*. 16(3): 157-164.
- Buffet, P.A., Safeukui, I., Peplane, G., Brousse, V., Plendki, V., Thellier, M., Turner, G.D. and Mercereau-Puijalon, O. (2011). The pathogenesis of *Plasmodium*

*falciparum* malaria in humans: insight from splenic physiology. *Blood*. 117(2): 381-392.

Bultinck, J., Sips, P., Vakaet, L., Brouckaert, P. and Cauwels, A. (2006). Systemic NO production during (septic) shock depends on parenchymal and not on hematopoietic cells: in vivo iNOS expression pattern in (septic) shock. *FASEB Journal*. 20(13): 2363-2365.

Cabantous, S., Poudiougou, B., Oumar, A.A., Traore, A., Abdoulaye, B., Vitte, J., Bongrand, P., Marguet, S., Doumbo, O. and Dessen, A.J. (2009). Genetic evidence for the aggravation of *Plasmodium falciparum* malaria by interleukin 4. *Journal of Infectious Diseases*. 200(10): 1530-1539.

Campbell, C.C. (2009). Malaria control – addressing challenges to ambitious goals. *New England Journal of Medicine*. 361(5): 522-523.

Carvalho, L.J., Ferreira-da-Cruz, M.F., Daniel-Ribeiro, C.T., Pelajo-Machado, M. and Lenzi, H.L. (2007). Germinal center architecture disturbance during *Plasmodium berghei* ANKA infection in CBA mice. *Malaria Journal*. 6: 59.

Chang, K.H. and Stevenson, M.M. (2004). Effect of anemia and renal cytokine production on erythropoietin production during blood-stage malaria. *Kidney International*. 65(5): 1640-1646.

Charoenpan, P., Indraprasit, S., Kiatboonsri, S., Surachittanont, O. and Tanomsup, S. (1990). Pulmonary edema in severe falciparum malaria: Hemodynamic study and clinicophysiological correlation. *Chest*. 97(5): 1190-1197.

Chomarat, P. and Banchereau, J. (1998). Interleukin-4 and interleukin-13: their similarities and discrepancies. *International Reviews of Immunology*. 17(1-4): 1-52.

Chotivanich, K., Udomsangpetch, R., McGready, R., Proux, S., Newton, P., Pukrittayakamee, S., Looareesuwan, S. and White, N.J. (2002). Central role of the spleen in malaria parasite clearance. *The Journal of Infectious Diseases*. 185(10): 1538-1541.

Chua, C.L.L., Brown, G., Hamilton, J.A., Rogerson, S. and Boeuf, P. (2013). Monocytes and macrophages in malaria: protection or pathology? *Trends in Parasitology*. 29(1): 26-34.

- Clark, I.A. and Chaudhri, G. (1988). Tumour necrosis factor may contribute to the anaemia of malaria by causing dyserythropoiesis and erythrophagocytosis. *British Journal of Haematology*. 70(1): 99-103.
- Clark, I.A., Iischer, S., MacMicking, J.D. and Cowden, W.B. (1990). TNF and *Plasmodium berghei* ANKA-induced cerebral malaria. *Immunology Letters*. 25(1-3): 195-198.
- Cook, G.C. (1994). Malaria in the liver. *Journal of Postgraduate Medicine*. 70: 780-784.
- Cordeiro, R.S., Cunha, F.Q., Filho, J.A., Flores, C.A., Vasconcelos, H.N. and Martins, M.A. (1983). *Plasmodium berghei*: physiopathological changes during infections in mice. *Annals of Tropical Medicine and Parasitology*. 77(5): 455-465.
- Couper, K.N., Phillips, R.S., Brombacher, F. and Alexander, J. (2005). Parasite-specific IgM plays a significant role in the protective immune response to asexual erythrocytic stage *Plasmodium chabaudi* AS infection. *Parasite Immunology*. 27(5): 171-180.
- Couper, B.N., Blount, D.G. and Riley, E.M. (2008). Interleukin 10: The master regulator of immunity to infection. *Journal of Immunology*. 180(9): 5771-5777.
- Cox-Singh, J., Davis, T.M., Lee, K.S., Shamsul, S.S., Matusop, A., Ratnam, S., Rahman, H.A., Conway, D.J. and Singh, B. (2008). *Plasmodium knowlesi*: malaria in humans is widely distributed and potentially life threatening. *Clinical Infectious Diseases*. 46(2): 165-171.
- Craig, A. and Scherf, A. (2001). Molecules on the surface of *Plasmodium falciparum* infected erythrocyte and their role in malaria pathogenesis and immune evasion. *Molecular and Biochemical Parasitology*. 115(2): 129-143.
- Craig, A.G., Grau, G.E., Janse, C., Kazura, J.W., Milner, D., Barnwell, J.W., Turner, G.D.H. and Langhorne, J. (2012). The role of animal models for research on severe malaria. *PLoS Pathogens*. 8(2): e1002401.

- Curfs, J.H., Schetters, T.P., Hermsen, C.C., Jerusalem, C.R., van Zon, A.A. and Eling, W.M. (1989). Immunological aspects of cerebral lesions in murine malaria. *Clinical and Experimental Immunology*. 75(1): 136-140.
- Curfs, J.H., van der Meer, J.W., Sauerwein, R.W. and Eling, W.M. (1990). Low dosage of interleukin-1 protect mice against lethal cerebral malaria. *Journal of Experimental Medicine*. 172(5): 1287-1291.
- Curfs, J.H., van der Meide, P.H., Billiau, A., Meuwissen, J.H. and Eling, W.M. (1993). *Plasmodium berghei*: recombinant interferon and the development of parasitemia and cerebral lesions in malaria-infected mice. *Experimental Parasitology*. 77(2): 212-223.
- Das, B.S. (2008). Renal failure in malaria. *Journal of Vector Borne Diseases*. 45(2): 83-97.
- Dascombe, M.J. and Sidara, J.Y. The absence of fever in rat malaria is associated with increased turnover of 5-hydroxytryptamine in the brain. *Advances in pharmacological Sciences*, Birkhauser Verlag Basel, 1994, pp. 47-48.
- Davis, C. (2010). Malaria. [www.medicinenet.com/malaria/article.html](http://www.medicinenet.com/malaria/article.html). Accessed 24th Nov 2012.
- Day, N.P., Hien, T.T., Schollaardt, T., Loc, P.P., Chuong, L.V., Chau, T.T., Mai, N.T., Phu, N.H., Sinh, D.X., White, N.J. and Ho, D.M. (1999). The prognostic and pathophysiologic role of pro and anti-inflammatory cytokines in severe malaria. *The Journal of Infectious Diseases*. 180(4): 1288-1297.
- Delacollette, C., Taelman, H. and Wery, M. (1995). An etiologic study of hemoglobinuria and blackwater fever in the Kivu Mountains, Zaire. *Annales Societe Belge Medecine Tropicale*. 75(1): 51-63.
- Del Prete, G., De Carli, M., Almerigogna, F., Giudizi, M.G., Biagotti, R. and Romagnani, S. (1993). Human IL-10 is produced by both type 1 helper (Th1) and type 2 helper (Th2) T cell clones and inhibit their antigen-specific proliferation and cytokine production. *Journal of Immunology*. 150(2): 353-360.



- Del Portillo, H.A., Ferrer, M., Brugat, T., Martin-Jaular, L., Langhorne, J. and Lacerda, M.V.G. (2012). The role of the spleen in malaria. *Cellular Microbiology*. 14(3): 343-355.
- Depinay, N., Franetich, J.F., Gruner, A.C., Mauduit, M., Chavette, J., Luty, A.J.F., van Germert, G., Sauerwein, R.W., Siksik, J., Hannoun, L., Mazier, D., Snounou, G. and Renia, L. (2011). Inhibitory effects of TNF- $\alpha$  on malaria pre-erythrocytic stage development: influence of host hepatocyte/parasite combinations. *PLoS ONE*. 6(3): e17464.
- De Souza, J.B. and Riley, E.M. (2002). Cerebral malaria: the contribution of studies in animal models to our understanding of immunopathogenesis. *Microbes and Infection*. 4(3): 291-300.
- Dinarello, C.A. (1997). Induction of interleukin-1 and interleukin-1 receptor antagonist. *Seminars in Oncology*. 24(3 Suppl 9): 81-93.
- Dinarello, C.A. (2000). Proinflammatory cytokines. *CHEST*. 118(2): 503-508.
- Dinarello, C.A. (2009). Immunological and inflammatory functions of the interleukin-1 family. *Annual Review of Immunology*. 27: 519-550.
- Dinarello, C.A. and Wolff, S.M. (1993). The role of interleukin-1 in disease. *The New England Journal of Medicine*. 328(10): 744.
- Dolinay, T., Kim, Y.S., Howrylak, J., Hunninghake, G.M., An, C.H., Fredenburgh, L., Massaro, A.F., Rogers, A., Gazourian, L., Nakahira, K., Haspel, J.A., Landazury, R., Eppanapally, S., Christie, J.D., Meyer, N.J., Ware, L.B., Christiani, D.C., Ryter, S.W., Baron, R.M. and Choi, A.M. (2012). Inflammasome-regulated cytokines are critical mediators of acute lung injury. *American Journal of Respiratory and Critical Care Medicine*. 185(11): 1225-1234.
- Dondorp, A.M. (2005). Pathophysiology, clinical presentation and treatment of cerebral malaria. *Neurology Asia*. 10: 67-77.
- Dondorp, A.M., Kager, P.A., Vreeken, J. and White, N.J. (2000). Abnormal blood flow and red cell deformability in severe falciparum malaria. *Parasitology Today*. 16(6): 228-232.

- Dong, C. and Flavell, R.A. (2001). Th1 and Th2 cells. *Current Opinion in Hematology*. 8(1): 47-51.
- Doolan, D.L., Dobano, C. and Baird, J.K. (2009). Acquired immunity to malaria. *Clinical Microbiology Reviews*. 22(1): 13-36.
- Doolan, D.L. and Martinez-Alier, N. (2006). Immune response to pre-erythrocytic stages of malaria parasites. *Current Molecular Medicine*. 6(2): 169-185.
- Doubal, F.N., MacLulich, M.J., Ferguson, K.J., Dennis, M.S. and Wardlaw, J.M. (2010). Enlarged perivascular spaces on MRI are a feature of cerebral small vessel disease. *Stroke*. 41(3): 450-454.
- Duarte, M.I., Corbett, C.E., Boulos, M. and Amato Neto, V. (1985). Ultrastructure of the lung in falciparum malaria. *American Journal of Tropical Medicine and Hygiene*. 34(1): 31-35.
- Ehrich, J.H. and Eke, F.U. (2007). Malaria-induced renal damage: facts and myths. *Pediatric Nephrology*. 22(5): 626-637.
- Ehrich, J.H. and Voller, A. (1972). Studies on kidneys of mice infected with rodent malaria. I. Deposition of gamma-globulins in glomeruli in the early stage of disease. *Tropical Medicine and Parasitology*. 23(2): 147-152.
- Eiam-Ong, S. (2003). Malaria nephropathy. *Seminars in Nephrology*. 23(1): 21-33.
- Engwerda, C.R., Beattie, L. and Amante, F.H. (2005). The importance of spleen in malaria. *Trends in Parasitology*. 21(2): 75-80.
- Favre, N., Ryffel, B., Bordmann, G. and Rudin, W. (1997). The course of *Plasmodium chabaudi chabaudi* infections in interferon- $\gamma$  receptor deficient mice. *Parasite Immunology*. 19(8): 375-383.
- Ferreira, A., Schofield, L., Enea, V., Schellekens, H., van der Meide, P., Collins, W.E., Nussenzweig, R.S. and Nussenzweig, V. (1986). Inhibition of development of exoerythrocytic forms of malaria parasites by  $\gamma$ -interferon. *Science*. 232(4752): 881-884.



- Ferreira, M.U., Nunes, M.S. and Wunderlich, G. (2004). Antigenic diversity and immune evasion by malaria parasites. *Clinical and Diagnostic Laboratory Immunology*. 11(6): 987-995.
- Finkelman, F.D., Shea-Donohue, T., Morris, S.C., Gildea, L., Strait, R., Madden, K.B., Schopf, L. and Urban, J.F. (2004). Interleukin 4 and interleukin 13 mediated host protection against intestinal nematode parasites. *Immunological Reviews*. 201: 139-155.
- Finney, O.C., Riley, E.M. and Walther, M. (2010). Regulatory T cells in malaria – friend or foe? *Trends in Immunology*. 31(2): 63-70.
- Fiorentino, D.F., Zlotnik, A., Mosmann, T.R., Howard, M. and O’Gara, A. (1991). IL-10 inhibits cytokine production by activated macrophages. *Journal of Immunology*. 147(11): 3815-3822.
- Franke-Fayard, B., Fonager, J., Braks, A., Khan, S.M. and Janse, C.J. (2010). Sequestration and tissue accumulation of human malaria parasites: can we learn anything from rodent models of malaria? *PLoS Pathogens*. 6(9): e1001032. doi:10.1371/journal.ppat.1001032.
- Franke-Fayard, B., Janse, C.J., Cunha-Rodrigues, M., Ramesar, J., Buscher, P., Que, I., Lowik, C., Voshol, P.J., den Boer, M.A., van Duinen, S.G., Febbraio, M., Mota, M.M. and Waters, A.P. (2005). Murine malaria parasite sequestration: CD36 is the major receptor, but cerebral pathology is unlinked to sequestration. *Proceedings of the National Academy of Sciences USA*. 102(32): 11468-11473.
- Frevert, U. (2004). Sneaking in through the back entrance: the biology of malaria liver stages. *Trends in Parasitology*. 20(9): 417-424.
- Fried, M., Muga, R.O., Misore, A.O. and Duffy, P.E. (1998). Malaria elicits type 1 cytokines in the human placenta: IFN- $\gamma$  and TNF- $\alpha$  associated with pregnancy outcomes. *Journal of Immunology*. 160(5): 2523-2530.
- Fritsche, G., Larcher, C., Schennach, H. and Weiss, G. (2001). Regulatory interactions between iron and nitric oxide metabolism for immune defense against *Plasmodium falciparum* infection. *The Journal of Infectious Diseases*. 183(9): 1388-1394.

- Fukao, T., Frucht, D.M., Yap, G., Gadina, M., O'Shea, J.J. and Koyasu, S. (2001). Inducible expression of Stat4 in dendritic cells and macrophages and its critical role in innate and adaptive immune responses. *Journal of Immunology*. 166(7): 4446-4455.
- Gachot, B., Wolff, M., Nissack, G., Veber, B. and Vachon, F. (1995). Acute lung injury complicating imported *Plasmodium falciparum* malaria. *CHEST*. 108(3): 746-749.
- Garroud, O., Perraut, R., Riveau, G., Nutman, T.B. (2003). Class and subclass selection in parasite-specific antibody responses. *Trends in Parasitology*. 19(7): 300-304.
- Gessner, A., Mohrs, K. and Mohrs, M. (2005). Mast cells, basophils, eosinophils acquire constitutive IL-4 and IL-13 transcripts during lineage differentiation that are sufficient for rapid cytokine production. *Journal of Immunology*. 174(2): 1063-1072.
- Gilles, H.M. and Warrell, D.A. (1993). Pathology and pathophysiology of human malaria. In Francis, N. and Warrell, D.A. *Bruce Chwatt's Essential Malariology*(pp. 50-59).London: Edward Arnold Publisher.
- Gioannini, T.L. and Weiss, J.P. (2007). Regulations of interactions of Gram-negative bacterial endotoxins with mammalian cells. *Immunologic Research*. 39(1-3): 49-60.
- Gozal, D. (1992). The incidence of pulmonary manifestations during *Plasmodium falciparum* malaria in non immune subjects. *Annals of Tropical Medicine and Parasitology*. 43: 6-8.
- Grau, G.E., Fajardo, L.F., Allet, B., Lambert, P.H. and Vassalli. P. (1987). TNF (cachetin) as an essential mediator in murine cerebral malaria. *Science*. 237(4819): 1210-1212.
- Grau, G.E., Heremans, H., Piguet, P.F., Pointaire, P., Lambert, P.H., Billiau, A. and Vassalli, P. (1989). Monoclonal antibody against interferon gamma can prevent experimental cerebral malaria and its associated overproduction of tumor necrosis factor. *Proceedings of the National Academy of Sciences USA*. 86(14): 5572-5574.

- Greenberger, M.J., Strieter, R.M., Kunkel, S.L., Danforth, J.M., Goodman, K.E. and Standiford, T.J. (1995). Neutralization of IL-10 increases survival in a murine model of *Klebsiella pneumoniae*. *Journal of Immunology*. 155(2): 722-729.
- Groom, A.C., Schmidt, E.E. and MacDonald, I.C. (1991). Microcirculatory pathways and blood flow in spleen: new insights from washout kinetics, corrosion casts, and quantitative intravital videomicroscopy. *Scanning Microscopy*. 5(1): 159-173.
- Groux, H., Bigler, M., De Vries, J.E. and Roncarolo, M.G. (1996). Interleukin-10 induces a long-term antigen-specific anergic state in human CD4<sup>+</sup> T cells. *Journal of Experimental Medicine*. 184(1): 19-29.
- Grun, J.L., Long, C.A. and Weidanz, W.P. (1985). Effects of splenectomy on antibody-independent immunity to *Plasmodium chabaudiadami* malaria. *Infection and Immunity*. 48(3): 853-858.
- Guerin, P., Olliaro, P., Nosten, F., Druilhe, P., Laxminarayan, R., Binka, F., Kilama, W., Ford, N. and White, N. (2002). Malaria: current status of control, diagnosis, treatment, and a proposed agenda for research and development. *The Lancet Infectious Diseases*. 2(9): 564-73.
- Hafalla, J.C.R., Cockburn, I.A. and Zavala, F. (2006). Protective and pathogenic roles of CD8<sup>+</sup> T cells during malaria infection. *Parasite Immunology*. 28(1-2): 15-24.
- Haldar, K., Murphy, S.C., Milner, D.A. and Taylor, T.E. (2007). Malaria: mechanisms of erythrocytic infection and pathological correlates of severe disease. *Annual Review of Pathology-Mechanisms of Disease*. 2: 217-249.
- Hanada, T. and Yoshimura, A. (2002). Regulation of cytokine signaling and inflammation. *Cytokine Growth Factor Reviews*. 13(4-5): 413-421.
- Haque, A., Best, S.E., Amante, F.H., Ammerdorffer, A., Labastida, F., Pereira, T., Ramm, G.A. and Engwerda, C.R. (2011). High parasite burden cause liver damage in mice following *Plasmodium berghei* ANKA infection independently of CD8<sup>+</sup> T cell-mediated immune pathology. *Infection and Immunity*. 79(5): 1882-1888.
- Harkness, J.E., Turner, P.V., Woude, S.V. and Wheler, C.L (2010). *Harkness and Wagner's biology and medicine of rabbits and rodents*. (5<sup>th</sup> Ed) (pp. 167 & 260).

American College of Laboratory Animal Medicine. New Jersey: Wiley-Blackwell Publisher.

- Hearn, J., Rayment, N., Landon, D.N., Katz, D.R. and de Souza, J.B. (2000). Immunopathology of cerebral malaria: morphological evidence of parasite sequestration in murine brain microvasculature. *Infection and Immunity*. 68(9): 5364-5376.
- Hernandez-Valladares, M., Naessens, J., Nagda, S., Musoke, A.J., Rihet, P., Ole-Moiyoi, O.K. and Iraqi, F.A. (2004). Comparison of pathology in susceptible A/J and resistant C7BL/6J mice after infection with different sub-strains of *Plasmodium chabaudi*. *Experimental Parasitology*. 108(3-4): 134-141.
- Hill, A.V. (2006). Pre-erythrocytic malaria vaccines: towards greater efficacy. *Nature Reviews of Immunology*. 6(1): 21-32.
- Himmerilrich, H., Launois, P., Maillard, I., Biedermann, T., Tacchini-Cottier, F., Locksley, R.M., Rocken, M. and Louis, J.A. (2000). In Balb/c mice, IL-4 production during the initial phase of infection with *Leishmania major* is necessary and sufficient to instruct Th2 cell development resulting in progressive disease. *Journal of Immunology*. 164(9): 4819-4825.
- Hoareau, L., Bencharif, K., Rondeau, P., Murumalla, R., Ramanan, P., Tallet, F., Delarue, P., Cesar, M., Roche, R. and Festy, F. (2010). Signaling pathways involved in LPS induced TNFalpha production in human adipocytes. *Journal of Inflammation*. 7:1.
- Huemann, D. and Roger, T. (2002). Initial responses to endotoxins and Gram-negative bacteria. *Clinica Chimica Acta*. 323(1-2): 59-72.
- Hugosson, E., Montgomery, S.M., Premiji, Z., Troye-Blomberg, M. and Bjorkman, A. (2004). Higher IL-10 levels are associated with less effective clearance of *Plasmodium falciparum* parasites. *Parasite Immunology*. 26(3): 111-117.
- Hunt, N.H. and Grau, G.E. (2003). Cytokines: accelerators and brakes in the pathogenesis of cerebral malaria. *Trends in Immunology*. 24(9): 491-499.
- Iademarco, M.F., Barks, J.L. and Dean, D.C. (1995). Regulation of vascular cell adhesion molecule-1 expression by IL-4 and TNF-alpha in cultured endothelial cells. *The Journal of Clinical Investigation*. 95(1):264-271.

- Idro, R., Jenkins, N.E. and Newton, C.R.J. (2005). Pathogenesis, clinical features, and neurological outcomes of cerebral malaria. *The Lancet Neurology*. 4(12): 827-840.
- Jager, W., Bourcier, K., Rijkers, G.T., Prakken, B.J. and Seyfert-Margolis, V. (2009). Prerequisites for cytokine measurements in clinical trials with multiplex immunoassays. *BMC Immunology*. 10:52.
- Jakobsen, P.H., Bate, C.A., Taverne, J. and Playfair, J.H. (1995). Malaria: toxins, cytokines and disease. *Parasite Immunology*. 17(5): 223-231.
- Jarra, W., Hills, L.A., March, J.C. and Brown, K.N. (1986). Protective immunity to malaria. Studies with cloned lines of *Plasmodium chabaudi chabaudi* and *P. berghei* in CBa/Ca mice II. The effectiveness and inter- or intra-species specificity of the passive transfer of immunity with serum. *Parasite Immunology*. 8(3): 239-254.
- Jeong, J.Y., Kim, S.H., Lee, H.J. and Sim, J.S. (2002). Atypical low-signal-intensity renal parenchyma causes and patterns. *Radiographics*. 22(4): 833-846.
- Jennings, G. and Elia, M. (1987). Effects of *E. coli* endotoxin on temperature, oxygen consumption and brown adipose tissue thermogenesis in rats and mice. *Bioscience Reports*. 7(6): 517-523.
- Jennings, V.M., Actor, J.K., Lal, A.A. and Hunter, R.L. (1997). Cytokine profile suggesting that murine cerebral malaria is an encephalitis. *Infection and Immunity*. 65(11): 4883-4887.
- Jurlander, J, Lai, C.F., Tan, J., Chou, C.C., Geisler, C.H., Schriber, J., Blumenson, L.E., Narula, S.K., Baumann, H. and Caliguri, M.A. (1997). Characterization of interleukin-10 receptor expression on B-cell chronic lymphocytic leukemia cells. *Blood*. 89(11): 4146-4152.
- Kakkilaya, B. S. (2009). Complications : anemia.  
[www.malariasite.com/malaria/Complications6.htm](http://www.malariasite.com/malaria/Complications6.htm) (2010).
- Kambayashi, T., Jacob, C.O. and Strassman, G. (1996). IL-4 and IL-13 modulate IL-10 release in endotoxin-stimulated murine peritoneal mononuclear phagocytes. *Cellular Immunology*. 171(1): 153-158.

- Kaplan, M.H., Schindler, U., Smiley, S.T. and Grusby, M.J. (1996). Stat6 is required for mediating responses to IL-4 and for development of Th2 cells. *Immunity*. 4(3): 313-319.
- Kelso, A. (1995). Th1 and Th2 subsets: paradigms lost? *Immunology Today*. 16(8): 374-379.
- Kossodo, S., Monso, C., Juillard, P., Velu, T., Goldman, M. and Grau, G.E. (1997). IL-10 modulates susceptibility in experimental cerebral malaria. *Immunology*. 91(4): 536-540.
- Kotowicz, K., Callard, R.E., Friedrich, K., Matthews, D. J. and Klein, N. (1996). Biological activity of IL-4 and IL-13 on human endothelial cells: functional evidence on both cytokines act through the same receptor. *International Immunology*. 8(12): 1915-1925.
- Kremsner, P.G., Grundmann, H., Neifer, S., Silwa, K., Sahlmuller, G., Hegenscheid, B. and Bienzel, U. (1991). Pentoxifylline prevents murine cerebral malaria. *The Journal of Infectious Diseases*. 164(3): 605-608
- Kremsner, P.G., Winkler, S., Brandts, C., Wildling, E., Jenne, L., Graninger, J., Prada, J., Bienzele, U., Juillard, P. and Grau, G.E. (1995). Prediction of accelerated cure in *Plasmodium falciparum* malaria by the elevated capacity of tumor necrosis factor production. *The American Journal of Tropical Medicine and Hygiene*. 53(5): 532-538.
- Krishna, S., Waller, D.W., ter Kulle, F., Kwiatkowski, D., Crawley, J., Craddock, C.F., Nosten, F., Chapman, D., Brewster, D. and Holloway, P.A. (1994). Lactic acidosis and hypoglycemia in children with severe malaria: pathophysiological and prognostic significance. *Transactions of the Royal Society of Tropical Medicine and Hygiene*. 88(1): 67-73.
- Krucken, J., Delic, D., Pauen, H., Wojtalla, A., El-Kahragy, M., Dkhil, M.A., Mosmann, H. and Wunderlich, F. (2009). Augmented particle trapping and attenuated inflammation in the liver by protective vaccination against *Plasmodium chabaudi* malaria. *Malaria Journal*. 8: 54.



- Kumar, S., Epstein, J.E., Richie, T.L., Nkrumah, F.K., Soisson, L., Carucci, D.J. and Hoffman, S.L. (2002). A multilateral effort to develop DNA vaccines against falciparum malaria. *Trends in Parasitology*. 18(3): 129-135.
- Kumaratilake, L.M. and Ferrante, A. (1992). IL-4 inhibits macrophage-mediated killing of *Plasmodium falciparum* *in vitro*: a possible parasite-immune evasion mechanism. *The Journal of Immunology*. 149(1): 194-199.
- Kumaratilake, L.M. and Ferrante, A. (2000). Opsonization and phagocytosis of *Plasmodium falciparum* merozoites measured by flow cytometry. *Clinical and Diagnostic Laboratory Immunology*. 7(1): 9-13.
- Kutzhals, J.A., Adabayeri, V., Goka, B.Q., Akanmori, B.D., Oliver-Commey, J.O., Nkrumah, F.K., Behr, C. and Hviid, L (1998). Low plasma concentrations of interleukin 10 in severe malarial anemia compared with cerebral and uncomplicated malaria. *Lancet*. 353(9155): 848.
- Kwiatkowski, D. (1990). Tumour necrosis factor, fever and fatality in falciparum malaria. *Immunology Letters*. 25(1-3): 213-216.
- Kwiatkowski, D., Hill, A.V., Sambou, I., Twumasi, P., Castracane, J., Manogue, K.R., Cerami, A., Brewster, D.R. and Greenwood, B.M. (1990). TNF concentration in fatal cerebral, non-fatal cerebral, and uncomplicated *Plasmodium falciparum* malaria. *Lancet*. 17(8725): 1201-1204.
- Kwiatkowski, D.P. (2005). How malaria has affected the human genome and what human genetics can teach us about malaria. *American Journal of Human Genetics*. 77(2): 171-190.
- Lackner, P., Beer, R., Helbok, R., Broessner, G., Engelhardt, K., Brenneis, C., Schmutzhard, E. and Pfaller, K. (2006). Scanning electron microscopy of neuropathology of murine cerebral malaria. *Malaria Journal*. 5: 116.
- Lai, Y.H., Heslan, J.M., Poppema, S., Elliott, J.F. and Mosmann, T.R. (1996). Continuous administration of IL-13 to mice induced extramedullary hemopoiesis and monocytosis. *The Journal of Immunology*. 156(9): 3166-3173.
- Lai, Y.H. and Mosmann, T.R. (1999). Mouse IL-13 enhances antibody-production *in vivo* and acts directly on B cells *in vitro* to increase survival and hence antibody production. *The Journal of Immunology*. 162(1): 78-87.

- Langermans, J.A., Nibbering, P.H., Van Vuren-Van Der Hulst, M.E. and Van Furth, R. (2001). TGF- $\beta$  suppresses IFN $\gamma$ -induced toxoplasmatatic activity in murine macrophages by inhibition of TNF- $\alpha$  production. *Parasite Immunology*. 23(4): 169-175.
- Langhorne, J., Ndungu, F.M., Sponaas, A. and Marsh, K. (2008). Immunity to malaria: more questions than answers. *Nature Immunology*. 9(7): 725-732.
- Lee, K.S., Divis, P.C., Zakaria, S.K., Matusop, A., Julin, R.A., Conway, D.J., Cox-Singh, J. and Singh, B. (2011). *Plasmodium knowlesi*: reservoir hosts and tracking the emergence in humans and macaques. *PLoS Pathogens*. 7(4): e1002015.
- Lee, S.J., Seo, E. and Cho, Y. (2013). Proposal for a new therapy for drug-resistant malaria using *Plasmodium* synthetic lethality inference. *International Journal for Parasitology: Drugs and Drugs Resistance*. 3: 119-128.
- Lee, Y.W., Eum, S.Y., Chen, K.C., Henning, B. and Toborek, M. (2004). Gene expression profile in interleukin-4 stimulated human vascular endothelial cells. *Molecular Medicine*. 10(1-6): 19-27.
- Letterio, J.J. and Roberts, A.B. (1997). TGF- $\beta$ : a critical modulator of immune cell function. *Clinical Immunology and Immunopathology*. 84(3): 244-250.
- Li, C., Corraliza, I. and Langhorne, J. (1999). A defect in interleukin-10 leads to enhanced malarial disease in *Plasmodium chabaudi chabaudi* infection in mice. *Infection and Immunity*. 67(9): 4435-4442.
- Li, C., Sanni, I.A., Omer, F., Riley, E.M. and Langhorne, J. (2003). Pathology of *Plasmodium chabaudi chabaudi* infection and mortality in interleukin-10 deficient mice are ameliorated by anti-tumor necrosis factor and exacerbated by anti-transforming growth factor  $\beta$  antibodies. *Infection and Immunity*. 71(9): 4850-4856.
- Li, X., Chen, H., Oo, T.H., Daly, T.M., Bergman, L.W., Liu, S.C., Chishti, A.H. and Oh, S.S (2004). A co-ligand complex anchors *Plasmodium falciparum* merozoites to the erythrocyte invasion receptor band 3. *The Journal of Biological Chemistry*. 279(7): 5765-5771.



- Lichtman, A.H., Chin, J., Schmidt, J.A. and Abbas, A.K. (1988). Role of interleukin-1 in the activation of T lymphocytes. *Proceedings of the National Academy of Sciences USA*. 85(24): 9699-9703.
- Linke, A., Kuhn, R., Muller, W., Honarvar, N., Li, C. and Langhorne, J. (1996). *Plasmodium chabaudi chabaudi*: differential susceptibility of gene-targeted mice deficient in IL-10 to an erythrocytic-stage infection. *Experimental Parasitology*. 84(2): 253-263.
- Lloyd, C.M., Wozencraft, A.O. and Williams, D.G. (1993). Cell-mediated pathology during murine malaria-associated nephritis. *Clinical and Experimental Immunology*. 94(3): 398-402.
- Lopez, C., Saravia, C., Gomez, A., Hoebeke, J. and Patarroyo, M.A. (2010). Mechanisms of genetically-based resistance to malaria. *Gene*. 467(1-2): 1-12.
- Lou, J., Lucas, R. and Grau, G.E. (2001). Pathogenesis of cerebral malaria: recent experimental data and possible applications for human. *Clinical Microbiology Reviews*. 14(4): 810-820.
- Lovegrove, F.E., Gharib, S.A., Pena-Castillo, L., Patel, S.N., Ruzinski, J.T., Hughes, T.R., Liles, W.C. and Kain, K.C. (2008). Parasite burden and CD36-mediated sequestration are determinants of acute lung injury in an experimental malaria model. *PLoS Pathogens*. 4(5): e1000068.
- Lucas, R., Juillard, P., Decoster, E., Redard, M., Burger, D., Donati, Y., Giroud, C., Monso-Hinard, C., De-Kesel, T., Buurman, W.A., Moore, M.W., Dayer, J.M., Fiers, W., Bluethmann, H. and Grau, G.E. (1997). Crucial role for tumour necrosis factor (TNF) receptor 2 and membrane bound TNF in experimental cerebral malaria. *European Journal of Immunology*. 27(7): 1719-1725.
- Mackintosh, C.L., Beeson, J.G. and Marsh, K. (2004). Clinical features and pathogenesis of severe malaria. *Trends in Parasitology*. 20(12): 597-603.
- Maguire, G.P., Handojo, T., Pain, M.C.F., Kenangalem, E., Price, R.N., Tjitra, E. and Anstey, N.M. (2005). Lung injury in uncomplicated and severe *falciparum* malaria: A longitudinal study in Papua, Indonesia. *The Journal of Infectious Diseases*. 192(11): 1966-1974.

- Maitland, K. and Newton, C.R.J.C. (2005). Acidosis of severe falciparum malaria: heading for a shock? *Trends in Parasitology*. 21(1): 11-16.
- Malaguarnera, L. and Musumeci, S. (2002). The immune response to *Plasmodium falciparum* malaria. *The Lancet Infectious Diseases*. 2(8): 472-478.
- Malaria Prevention and Treatment*; UNICEF: New York, 2000.
- Martin-Jaular, L., Ferrer, M., Calvo, M., Rosanas-Urgell, A., Kalko, S., Graewe, S., Soria, G., Cortadellas, N., Ordi, J., Planas, A., Burns, J., Heussler, V. and del Portillo, H.A. (2011). Strain-specific spleen remodeling in *Plasmodium yoelii* infections in Balb/c mice facilitates adherence and spleen macrophage clearance escape. *Cellular Microbiology*. 13(1): 109-122.
- Matthay, M.A. and Zimmerman, G.A. (2005). Acute lung injury and the acute respiratory distress syndrome. *American Journal of Respirator Cell and Molecular Biology*. 33(4): 319-327.
- Mattsby-Baltzer, I., Ahlstrom, B., Edebo, L. and De-man, P. (1996). Susceptibility of LPS-responsiveness and -hyperresponsive ItyS mice to infection with rough mutants of *Salmonella typhimurium*. *Infection and Immunity*. 64(4): 1321-1327.
- McCall, M.B. and Sauerwein, R.W. (2010). Interferon- $\gamma$  – central mediator of protective immune responses against the pre-erythrocytic and blood stage of malaria. *Journal of Leukocyte Biology*. 88(6): 1131-1143.
- Medina, T.S., Costa, S.P.T., Oliveira, M.D., Ventura, A.M., de Souza, J.M., Gomes, T.F., Vollinoto, A.C.R., Povoá, M.M., Silva, J.S., Cunha, M.G. (2011). Increased interleukin-10 and interferon- $\gamma$  levels in *Plasmodium vivax* malaria suggest a reciprocal regulation which is not altered by IL-10 gene promoter polymorphism. *Malaria Journal*. 10: 264.
- Meding, S.J., Cheng, S.C., Simon-Haarhaus, B. and Langhorne, J. (1990). Role of  $\gamma$  interferon during infection with *Plasmodium chabaudi chabaudi*. *Infection and Immunity*. 58(11): 3671-3678.
- Mehta, K.S., Halankar, A.R., Makwana, P.D., Torane, P.P., Satijia, P.S. and Shah, V.B. (2001). Severe acute renal failure in malaria. *Journal of Postgraduate Medicine*. 47(1): 24-26.

- Mendis, K., Naotunne, T., Karunaweera, N.D., Del Giudice, G., Grau, G.E. and Carter, R. (1990). Anti-parasite effects of cytokines in malaria. *Immunology Letters*. 25(1-3): 217-220.
- Menendez, C., Fleming, A.F. and Alonso, P.L. (2000). Malaria-related anemia. *Parasitology Today*. 16(11): 469.
- Miller, L.H., Baruch, D.I., March, K. and Doumbo, O.K. (2002). The pathogenic basis of malaria. *Nature*. 415(6872): 673-679.
- Mitchell, A.J., Hansen, A.M., Hee, L., Ball, H.J., Potter, S.M., Walker, J.C. and Hunt, N.H. (2005). Early cytokine production is associated with protection from murine cerebral malaria. *Infection and Immunity*. 73(9): 5645-5653.
- Miyakoda, M., Kimura, D., Yuda, M., Chinzei, Y., Shibata, Y., Honma, K. and Yui, K. (2008). Malaria-specific and nonspecific activation of CD8+ T cells during blood stage of *Plasmodium berghei* infection. *Journal of Immunology*. 181(2): 1420-1428.
- Mohan, K. and Stevenson, M.M. (1998). Acquired immunity to asexual blood stages. In I.W. Sherman. *Malaria: Parasite biology, pathogenesis and protection* (pp. 467-494). Washington DC: ASM Press.
- Mohr, N. (2002). *Malaria: evolution of a killer*. Seattle: Serif & Pixel Press.
- Moreb, J. and Zucali, J.R. (1992). The therapeutic potential of interleukin 1 and tumor necrosis factor on hematopoietic stem cells. *Leukemia and Lymphoma*. 8(4-5): 267-275.
- Moore, K.W., de Waal Malefyt, R., Coffman, R.I. and O'Garra, A. (2001). Interleukin-10 and the interleukin-10 receptor. *Annual Review of Immunology*. 19: 683-765.
- Morrot, A., Hafalla, J.C., Cockburn, I.A., Carvalho, L.H. and Zavala, F. (2005). IL-4 receptor expression on CD8+ T cells is required for the development of protective memory responses against liver stages of malaria parasites. *The Journal of Experimental Medicine*. 202(4): 551-560.
- Mosmann, T.R. and Sad, S. (1996). The expanding universe of T-cell subsets: Th1, Th2 and more. *Immunology Today*. 17(3): 138-146.

- Muchamuel, T., Menon, S., Pisacane, P., Howard, M.C. and Cockayne, D.A. (1997). IL-13 protects mice from lipopolysaccharide-induced lethal endotoxemia: correlation with own-modulation of TNF- $\alpha$ , IFN- $\gamma$  and IL-12 production. *The Journal of Immunology*. 158(6): 2898-2903.
- Munoz, C., Carlet, J., Fitting, C., Misset, B., Bleriot, J.P. and Cavaillon, J.M. (1991). Dysregulation of in vitro cytokine production by monocytes during sepsis. *The Journal of Clinical Investigation*. 88(5): 1747-1754.
- Murata, T., Obiri, N.I. and Puri, R.K. (1998). Structure of and signal transduction through interleukin 4 and interleukin 13 receptors. *International Journal of Molecular Medicine*. 1(3): 551-557.
- Naito, M., Hasegawa, G., Ebe, Y. and Yamamoto, T. (2004). Differentiation and function of Kupffer cells. *Medical Electron Microscopy*. 37(1): 16-28.
- Newton, C.R.J.C., Taylor, T.E. and Whitten, R.O. (1998). Pathophysiology of fatal falciparum malaria in African children. *American Journal of Tropical Medicine and Hygiene*. 58(5): 673-683.
- Newton, C.R.J.C. and Warell, D.A. (1998). A neurological manifestations of falciparum malaria. *Annals of Neurology*. 43(6): 695-702.
- Nguansangiam, S., Day, N.P.J., Hien, T.T., Mai, N.T.H., Chaisri, U., Riganti, M., Dondorp, A.M., Lee, S.J., Phu, N.H., Turner, G., White, N.J., Ferguson, D. and Pongponratn, E. (2007). A quantitative ultrastructural study of renal pathology in fatal *Plasmodium falciparum* malaria. *Tropical Medicine and International Health*. 12(9): 1037-1050.
- Niikura, M., Kamiya, S., Nakane, A., Kita, K. and Kobayashi, F. (2010). IL-10 plays a crucial role for the protection of experimental cerebral malaria by co-infection with non-lethal malaria parasites. *International Journal for Parasitology*. 40(1): 101-108.
- Niikura, M., Inoue, S. and Kobayashi, F. (2011). Role of Interleukin-10 in malaria: Focusing on coinfection with lethal and nonlethal murine malaria parasites. *Journal of Biomedicine and Biotechnology*. doi: 10.1155/2011/383962.

- Nitcheu, J., Bonduelle, O., Combadiere, C., Tefit, M., Seilhean, D., Mazier, D. and Combadiere. (2003). Perforin-dependent brain-infiltrating cytotoxic CD8+ T lymphocytes mediate experimental cerebral malaria pathogenesis. *Journal of Immunology*. 170(4): 2221-2228.
- Nobes, M.S., Ghabrial, H., Simms, K.M., Smallwood, R.B., Morgan, D.J. and Sewell, R.B. (2002). Hepatic Kupffer cell phagocytic function in rats with erythrocytic-stage malaria. *Journal of Gastroenterology and Hepatology*. 17(5): 598-605.
- Ockenhouse, C.F., Tegoshi, T., Maeno, Y., Benjamin, C., Ho, M., Ei Khan, K., Thway, Y., Win, K., Aikawa, M. and Lobb, R.R. (1992). Human vascular endothelial cell adhesion receptors for *Plasmodium falciparum*-infected erythrocytes: roles for endothelial leukocyte adhesion molecule 1 and VCAM-1. *The Journal of Experimental Medicine*. 176(4): 1183-1189.
- Ohashi, J., Naka, I., Doi, A., Patarapotikul, J., Hananantachai, H., Tangpukdee, N., Looareesuwan, S. and Tokunaga, K. (2005). A functional polymorphism in the IL1B gene promoter, IL1B -31 C>T, is not associated with cerebral malaria in Thailand. *Malaria Journal*. 4: 38.
- Omer, F.M., Kurtzhals, J.A.L. and Riley, E.M. (2000). Maintaining the immunological balance in parasitic infections: a role for TGF- $\beta$ ? *Parasitology Today*. 16(1): 18-23.
- Omer, F.M. and Riley, E.M. (1998). Transforming growth factor-beta production is inversely correlated with severity of immune infection. *Journal of Experimental Medicine*. 188(1): 39-48.
- Ong'echa, J.M., Davenport, G.C., Vulule, J.M., Hitner, J.B. and Perkins, D.J. (2011). Identification of inflammatory biomarkers for pediatric malarial anemia severity using novel statistical methods. *Infection and Immunity*. 79(11): 4674-4680.
- Oster, C.N., Koontz, L.C. and Wyler, D.J. (1980). Malaria in asplenic mice: effects of splenectomy, congenital asplenia, and splenic reconstitution in the course of infection. *American Journal of Tropical Medicine and Hygiene*. 29(6): 1138-1142.

- Othoro, C., Lal, A.A., Nahlen, B., Koech, D., Orago, A.S. and Udhayakumar, V. (1999). A low interleukin-10, tumor necrosis factor alpha ratio is associated with anemia in children residing in a holoendemic malaria region in western Kenya. *Journal of Infectious Diseases*. 179(1): 279-282.
- Ouma, C., Davenport, G.C., Awandare, G.A., Keller, C.C., Were, T., Otieno, M.F., Vulule, J.M., Martonson, J., Ong'echa, J.M., Ferrell, R.E. and Perkins, D.J. (2008). Polymorphic variability in the interleukin (IL)-1beta promoter conditions susceptibility to severe malarial anemia and functional changes in IL-1beta production. *Journal of Infectious Diseases*. 198(8): 1219-1226.
- Owusu-Agyei, S., Koram, K.A., Baird, J.K., Utz, G.C., Binka, F.N., Nkrumah, F.K., Fryauff, D.J. and Hoffman, S.L. (2001). Incidence of symptomatic and asymptomatic *Plasmodium falciparum* infection following curative therapy in adult residents of northern Ghana. *American Journal of Tropical Medicine and Hygiene*. 65(3): 197-203.
- Pain, A., Ferguson, D.J., Kai, O., Urban, B.C., Lowe, B., Marsh, K. and Roberts, D.J. (2001). Platelet-mediated clumping of *Plasmodium falciparum*-infected erythrocytes is a common adhesive phenotype and is associated with severe malaria. *Proceedings of the National Academy of Sciences USA*. 98(4): 1805-1810.
- Pais, T.F., Figueiredo, C., Peixoto, R., Braz, M.H. and Chatterjee, S. (2008). Necrotic neurons enhance microglial neurotoxicity through induction of glutaminase by a MyD88-dependent pathway. *Journal of Neuroinflammation*. 5: 43.
- Parant, M., Le-Contel, C., Parant, F. and Chedid, L. (1991). Influence of endogenous glucocorticoid and endotoxin-induced production of circulating TNF $\alpha$ . *Lymphokine and Cytokine Research*. 10(4): 265-271.
- Paul, W.E. (1991). Interleukin-4: prototypic immunoregulatory lymphokine. *Blood*. 77(9): 1859-1870.
- Perkins, D.J., Wene, T., Davenport, G.C., Kempaiah, P., Hittner, J.B. and Ong'echa, J.M. (2011). Severe malarial anemia: innate immunity and pathogenesis. *International Journal of Biological Sciences*. 7(9): 1427-1442.



- Perlmann, H., Kumar, S., Vinets, J.M., Kullberg, M., Miller, L.H. and Perlmann, P. (1995). Cellular mechanisms in the immune response to malaria in *Plasmodium vinckei*-infected mice. *Infection and Immunity*. 63(10): 3987-3993.
- Perlmann, P. and Troye-Blomberg, M. (2002). Malaria and the immune system in humans. *Chemical Immunology*. 80: 229-242.
- Peyron, F., Burdin, N., Ringwald, P., Vuillez, J.P. and Banchereau, J. (1994). High levels of circulating interleukin-10 in human malaria. *Clinical and Experimental Immunology*. 95(2): 300-303.
- Plebanski, M. and Hill, A.V. (2000). The immunology of malaria infection. *Current Opinion in Immunology*. 12(4): 437-441.
- Pongponratn, E., Riganti, M., Punpoowong, B. and Aikawa, M. (1991). Microvascular sequestration of parasitized erythrocytes in human falciparum malaria: a pathological study. *American Journal of Tropical Medicine and Hygiene*. 44(2): 168-175.
- Pouvelle, B., Buffet, P.A., Lepolard, C., Scherf, A. and Gysin, J. (2000) Cytoadhesion of *Plasmodium falciparum* ring-stage-infected erythrocytes. *Nature Medicine*. 6(11): 1264-1268.
- Prada, J., Malinowski, J., Muller, S., Bienzle, U. and Kremsner, P.G. (1996). Effects of *Plasmodium vinckei* haemozoin on the production of oxygen radicals and nitric oxides in murine macrophages. *American Journal of Tropical Medicine and Hygiene*. 54(6): 620-624.
- Prakash, D., Fesel, C., Jain, R., Cazenave, P., Mishra, G.C. and Pied, S. (2006). Clusters of cytokines determine malaria severity in *Plasmodium falciparum*-infected patients from endemic areas of central India. *The Journal of Infectious Diseases*. 194(2): 198-207.
- Prasad, R.N. and Virk, K.J. (1993). Malaria as a cause of diarrhea – a review. *PNG Medical Journal*. 36(4): 337-341.
- Prommano, O., Chaisri, U., Turner, G.D., Wilairatana, P., Ferguson, D.J., Viriyavejakul, P., White, N.J. and Pongponratn, E. (2005). A quantitative ultrastructural study of the liver and the spleen in fatal falciparum malaria.

*Southeast Asian Journal of Tropical Medicine and Public Health.* 36(6): 1359-1370.

Quin, S.J. and Langhorne, J. (2001). Different regions of the malaria merozoite surface protein 1 of *Plasmodium chabaudi* elicit distinct T-cell and antibody isotype responses. *Infection and Immunity.* 69(4): 2245-2251.

Richie, T.L. and Saul, A. (2002). Progress and challenges for malaria vaccines. *Nature.* 415(6872): 694-701.

Riley, E.M., Wahl, S., Perkins, D.J. and Schofield, L. (2006). Regulating immunity to malaria. *Parasite Immunology.* 28(1-2): 35-49.

Ringwald, P., Peyron, F., Vuilleux, J.P., Touze, J.E., Le Bras, J. and Deleron, P. (1991). Levels of cytokines in plasma during *Plasmodium falciparum* malaria attacks. *Journal of Clinical Microbiology.* 29(9): 2076-2078.

Roberts, J.D. (2010). Anemia in malaria, [www.uptodate.com/patients/content/topic.do?topicKey=~DaDDD.hh5Z/r5MK](http://www.uptodate.com/patients/content/topic.do?topicKey=~DaDDD.hh5Z/r5MK) (2010). Accessed 20<sup>th</sup> Oct 2012.

Rockett, K.A., Awburn, M.M., Rockett, E.J. and Clark, I.A. (1994). Tumor necrosis factor and interleukin-1 synergy in the context of malaria pathology. *American Journal of Tropical Medicine and Hygiene.* 50(6): 735-742.

Rui-Mei, L., Kara, A.U., and Sinniah, R. (1998). Dysregulation of cytokine expression in tubulointerstitial nephritis associated with murine malaria. *Kidney International.* 53(4): 845-852.

Rungruang, T., Chaweeborisuit, P. and Klosek, S.K. (2010). Effect of malaria infection and dexamethasone on spleen morphology and histology. *Southeast Asian Journal of Tropical Medicine and Public Health.* 41(6): 1290-1296.

Saeftef, M., Krueger, A., Arriens, S., Heussler, V., Racz, P., Fleischer, B., Brombacher, F. and Hoerauf, A. (2004). Mice deficient in IL-4 or IL-4 receptor alpha have higher resistance to sporozoite infection with *Plasmodium berghei* (ANKA) than do naïve wild-type mice. *Infection and Immunity.* 72(1): 322-331.

Schoenborn, J.R. and Wilson, C.B. (2007). Regulation of interferon-gamma during innate and adaptive immune response. *Advance Immunology.* 96: 41-101.



- Schroder, K., Hertzog, P.J., Ravasi, T. and Hume, D.A. (2004). Interferon-gamma: an overview of signals, mechanisms and functions. *Journal of Leukocyte Biology*. 75(2): 163-189.
- Schwarzer, E., Alessio, M., Ulliers, D. and Arese, P. (1998). Phagocytosis of the malarial pigment, haemozoin, I, pairs expression of major histocompatibility class II antigen, CD54 and CD11c in human monocytes. *Infection and Immunity*. 66(4): 1601-1606.
- Schwenk, R., Asher, L.V., Chalom, I., Lanar, D., Sun, P., White, K., Keil, D., Kester, K.E., Stoute, J., Happner, D.G. and Krzych, U. (2003). Opsonization by antigen-specific antibodies as a mechanism of protective immunity induced by *Plasmodium falciparum* circumsporozoite protein-based vaccine. *Parasite Immunology*. 25(1): 17-25.
- Seixas, E., Oliveira, P., Moura Nunes, J.F. and Coutinho, A. (2008). An experimental model for fatal malaria due to TNF- $\alpha$ -dependent hepatic damage. *The Journal of Parasitology*. 135(6): 683-690.
- Senthikumaar, P. and Sarojini, S. (2013). Hematological studies in malaria affected patients in North Chennai, Tamil Nadu. *European Journal of Experimental Biology*. 3(1): 199-205.
- Shaffer, N., Grau, G.E., Hedberg, K., Davachi, F., Lyamba, B., Hightower, A.W., Breman, J.G. and Phuc, N.D. (1991). Tumor necrosis factor and severe malaria. *Journal of Infectious Diseases*. 163(1): 96-101.
- Shimoda, K., van Deursen, J., Sangster, M.Y., Sarawar, S.R., Carson, R.T., Tripp, R.A., Chu, C., Quelle, F.W., Nosaka, T., Vignali, D.A., Doherty, P.C., Grosveld, G., Paul, W.E. and Ihle, J.N. (1996). Lack of IL-4-induced Th2 response and IgE class switching in mice with disrupted Stat6 gene. *Nature*. 380(6575): 630-633.
- Siewe, L., Bollati-Fogolin, M., Wickenhauser, C., Krieg, T., Muller, W. and Roers, A. (2006). Interleukin-10 derived from macrophages and/or neutrophils regulates the inflammatory responses to LPS but not the response to CpG DNA. *European Journal of Immunology*. 36(12): 3248-3255.

- Smith, E.C. and Taylor-Robinson, A.W. (2003). Parasite-specific immunoglobulin isotypes during lethal and nonlethal murine malaria infections *Parasitology Research*. 89(1): 26-33.
- Snow, R.W., Guerra, C.A., Noor, A.M., Myint, H.Y. and Hay, S.I. (2005). The global distribution of clinical episodes of *Plasmodium falciparum* malaria. *Nature*. 434(7030): 214-217.
- Spits, H. and de Waal Malefyt, R. (1992). Functional characterization of human interleukin-10. *International Archives of Allergy and Applied Immunology*. 99(1): 8-15.
- Sterzel, R.B., Ehrich, J.H., Lucia, H., Thomson, D. and Kashgarian, M. (1982). Mesangial disposal of glomerular immune deposits in acute malarial glomerulonephritis. *Laboratory Investigation*. 46(2): 209-214.
- Tatle, M. and Malik, G.B. (1990). Pulmonary pathology in severe malaria infection in health and protein deprivation. *Journal of Tropical Medicine and Hygiene*. 93(6): 377-382.
- Taylor-Robinson, A.W. (2010). Regulation of immunity to *Plasmodium*: Implications from mouse models for blood stage malaria vaccine design. *Experimental Parasitology*. 126(3): 406-414.
- Taylor-Robinson, A.W. and Phillips, R.S. (1996). Reconstitution of B-cell-depleted mice with B cells restores Th2-type immune responses during *Plasmodium chabaudi chabaudi* infection. *Infection and Immunity*. 64(1): 366-370.
- Taylor-Robinson, A.W. and Smith, E.C. (1999). A role for cytokines in potentiation of malaria vaccines through immunological modulation of blood stage infection. *Immunological Reviews*. 171: 105-123.
- Taylor, W.R.J., Hanson, J., Turner, G.D.H., White, N.J. and Dondorp, A.M. (2012). Respiratory manifestations of malaria. *CHEST*. 142(2): 492-505.
- Te Velde, A.A., Hujibens, R.J., de Vries, J.E. and Figdor C.G. (1990). IL-4 decreases Fc gamma R membrane expression and Fc gamma R-mediated cytotoxic activity of human monocytes. *Journal of Immunology*. 144(8): 3046-3051.

*The Global Malaria Burden*; UNICEF: New York, USA, 2000.

- The University of Liverpool, (n.d). Biology of the malaria parasite. [www.liv.ac.uk/geography/research\\_projects/epidemics/MAL\\_biology.htm](http://www.liv.ac.uk/geography/research_projects/epidemics/MAL_biology.htm)(2010). Accessed 10<sup>th</sup> Nov 2012.
- Torre, D., Speranza, F. and Martegani, R. (2002). Role of proinflammatory and anti-inflammatory cytokines in the immune response to *Plasmodium falciparum* malaria. *The Lancet Infectious Diseases*. 2(12): 719-720.
- Touzani, O., Boutin, H., Chuquet, J. and Rothwell, N. (1999). Potential mechanisms of interleukin-1 involvement in cerebral ischaemia. *Journal of Neuroimmunology*. 100(1-2): 203-215.
- Tracey, K.J. and Cerami, A. (1993). Tumor necrosis factor: an updated review of its biology. *Critical Care Medicine*. 21(10 Suppl): 415-422.
- Trang, T.T., Phu, N.H., Vinh, H., Hien, T.T., Cuong, B.M., Chau, T.T., Mai, N.T., Waller, P.J. and White, N.J. (1992). Acute renal failure in patients with severe falciparum malaria. *Clinical Infectious Diseases*. 15(5): 874-880.
- Udomsangpetch, R., Chivapat, S., Viriyavejakul, P., Riganti, M., Wilairatana, P., Pangpanratin, E. and Looareesuwan, S. (1997). Involvement of cytokines in the histopathology of cerebral malaria. *American Journal of Tropical Medicine and Hygiene*. 57(5): 501-506.
- University of Leeds. (1995). Cytokines mediating inflammatory and effector functions, <http://nic.sav.sk/logos/books/scientific/node32.html> (2010). Accessed 1<sup>st</sup> Dec 2012.
- Urban, B.C., Hien, T.T., Day, N.P., Phu, N.H., Roberts, R., Pongponratn, E., Jones, M., Mai, N.T.H., Bethell, D., Turner, G.D.H., Ferguson, D., White, N.J. and Roberts, D.J. (2005). Fatal *Plasmodium falciparum* malaria causes specific patterns of splenic architectural disorganization. *Infection and Immunity*. 73(4): 1986-1994.
- Urquhart, A.D. (1994). Putative pathophysiological interactions of cytokines and phagocytic cells in severe human falciparum malaria. *Clinical Infectious Diseases*. 19(1): 117-131.

- Van den Steen, P.E., Deroost, K., Deckers, J., Van Herck, E., Struyf, S. and Opdenakker, G. (2013). Pathogenesis of malaria-associated acute respiratory distress syndrome. *Trends in Parasitology*. 29(7): 346-358.
- Van der Heyde, H.C., Batchelder, J.M., Sandor, M. and Weidanz, W.P. (2006). Splenic gammadelta T cells regulated by CD4+ T cells are required to control chronic *Plasmodium chabaudi* malaria in the B-cell deficient mice. *Infection and Immunity*. 74(5): 2717-2725.
- Vannier, E., Miller, L.C. and Dinarello, C.A. (1992). Coordinated anti-inflammatory effects of interleukin 4: interleukin 4 suppresses interleukin 1 production but up-regulates gene expression and synthesis of interleukin 1 receptor antagonist. *Proceedings of the Academy of Sciences USA*. 89(9): 4076-4080.
- Vernick, K.D. and Waters, A.P. (2004). Genomics and malaria control. *The New England Journal of Medicine*. 351(18): 1901-1904.
- Wahl, S.M. (1994). Transforming growth factor- $\beta$ : the good, the bad and the ugly. *The Journal of Experimental Medicine*. 180(5): 1587-1590.
- Walker, P., Salako, L.A., Sowunmi, A., Thomas, J.O., Sodeine, O. and Bondi, F.S. (1992). Prognostic risk factors and post mortem findings in cerebral malaria in children. *Transactions of the Royal Society of Tropical Medicine and Hygiene*. 86(5): 491-493.
- Walley, A.J., Aucan, C., Kwiatkowski, D. and Hill, A.V. (2004). Interleukin-1 gene cluster polymorphisms and susceptibility to clinical malaria in a Gambian case-control study. *European Journal of Human Genetics*. 12(2): 132-138.
- Walther, M., Woodruff, J., Edele, F., Jeffries, D., Tongren, J.E., King, E., Andrews, L., Bejon, P., Gilbert, S., De Souza, J.B., Sinden, R., Hill, A.V. and Riley, E.M. (2006). Innate immune responses to human malaria: heterogeneous cytokine responses to blood-stage *Plasmodium falciparum* correlate with parasitological and clinical outcomes. *The Journal of Immunology*. 177(8): 5736-5745.
- Wassmer, S.C., de Souza, J.B., Frere, C., Candal, F.J., Juhan-Vague, I. and Grau, G.E. (2006). TGF- $\beta_1$  released from activated platelets can induce TNF-stimulated human brain endothelium apoptosis: a new mechanism for microvascular lesion during cerebral malaria. *The Journal of Immunology*. 176(2): 1180-1184.

- Weiss, L. (1990). The spleen in malaria: the role of barrier cells. *Immunology Letters*. 259(1-3): 165-172.
- White, N.J., Turner, G.D.H., Medana, I.M., Dondorp, A.M. and Day, N.P.J. (2009). The murine cerebral malaria phenomenon. *Trends in Parasitology*. 26(1): 11-15.
- White, N.J. (2010). Artemisinin resistance – the clock is ticking. *Lancet*. 376(9758): 2051-2052.
- White, V.A. (2011). Malaria in Malawi – inside a research autopsy study of pediatric cerebral malaria. *Archives of Pathology and Laboratory Medicine*. 135(2): 220-226.
- Wilairatana, P., Tangpuckdee, N., Krudsood, S., Pongponratn, E. and Riganti, M. (2008). Gastrointestinal and liver involvement in falciparum malaria. *Thai Journal of Gastroenterology*. 9(3): 124-127.
- Wills-Karp, M., Luyimbazi, J., Xu, X., Schofield, B., Neben, T.Y., Karp, C.L. and Donaldson, D.D. (1998). Interleukin-13: central mediator of allergic asthma. *Science*. 282(5397): 2258-2261.
- Winkler, S., Willheim, M., Baier, K., Schmid, D., Aichelburg, A., Graninger, W. and Kreamer, P.G. (1999). Frequency of cytokine-producing T cells in patients of different age groups with *Plasmodium falciparum* malaria. *Journal of Infectious Diseases*. 179(1): 209-216.
- Wood, N., Whitters, M.J., Jacobson, B.A., Witek, J., Sypek, J.P., Kasaian, M., Eppihimer, M.J., Unger, M., Tanaka, T., Goldman, S.J., Collins, M., Donaldson, D.D. and Grusby, M.J. (2003). Enhanced interleukin (IL)-13 responses in mice lacking IL-13 receptor alpha 2. *Journal of Experimental Medicine*. 197(6): 703-709.
- Woodward, E.A., Prele, C.M., Nicholson, S.E., Kolesnik, T.B. and Hart, P.H. (2010). The anti-inflammatory effects of IL-4 are not mediated by suppressor of cytokine signaling-1 (SOCS1). *Journal of Immunology*. 131(1): 118-127.
- WHO: *World Malaria Report 2010*; World Health Organization: Geneva, Switzerland 2010.

- Wykes, M.N. and Good, M.F. (2009). What have we learnt from mouse models for the study of malaria? *European Journal of Immunology*. 39(8): 2004-2006.
- Wynn, T.A. (2003). Interleukin-13 effector functions. *Annual Reviews of Immunology*. 21: 425-456.
- Yanez, D.M., Manning, D.D., Cooley, A.J., Weidanz, W.P. and van der Heyde. (1996). Participation of lymphocyte subpopulations in the pathogenesis of experimental murine cerebral malaria. *Journal of Immunology*. 157(4): 1620-1624.
- Young, H.A. and Hardy, K.J. (1995). Role of interferon gamma in immune cell regulation. *Journal of Leukocyte Biology*. 58(4): 373-381.
- Zander, D.S. and Farver, C.F. (2008). Pulmonary pathology. *A volume in foundations in diagnostic pathology series*(pp. 288-290). Pennsylvania: Churchill Livingstone Elsevier Inc.
- Zemse, S.M., Chin, W.C., Hilger, R.H.P. and Webb, R.C. (2010). IL-10 inhibits the in vivo and in vitro adverse effects of TNF- $\alpha$  on the endothelium of murine aorta. *American Journal of Physiology – Heart and Circulatory Physiology*. 299(4): H 1160-1167.
- Zhu, J., Yamane, H., Cote-Sierra, J., Guo, L. and Paul, W.E. (2006). GATA-3 promotes Th2 responses through three different mechanisms: induction of Th2 cytokine production, selective growth of Th2 cells and inhibition of Th1 cell-specific factors. *Cell Research*. 16(1): 3-10.
- Zingman, B.S. and Viner, B.L. (1993). Splenic complications in malaria: case report and review. *Clinical Infectious Diseases*. 16(2): 223-232.
- Zurawski, G. and de Vries, J.E. (1994). IL-13, an IL-4-like cytokine that acts on monocytes and B cells, but not on T cells. *Immunology Today*. 15(1): 19-26.