



**UNIVERSITI PUTRA MALAYSIA**

***FUNCTIONAL CHARACTERIZATION OF SUPEROXIDE DISMUTASE IN  
Candida dubliniensis AND EFFECT OF DECYL METHYL CARBINOL***

**GOH MENG CHUAN**

**FPSK(M) 2012 56**

**FUNCTIONAL CHARACTERIZATION OF SUPEROXIDE DISMUTASE IN  
*Candida dubliniensis* AND EFFECT OF DECYL METHYL CARBINOL**

By  
**GOH MENG CHUAN**

**Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfilment of the Requirements for the Degree of Master of Science**

**October 2012**

## DEDICATION

**May my reverends, my parents, my teachers, my sisters, my relatives, my friends  
and my love one, may you be free from harm and danger, may you be well and  
happy always.**



Abstract of thesis presented to senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

**FUNCTIONAL CHARACTERIZATION OF SUPEROXIDE DISMUTASE IN  
*Candida dubliniensis* AND EFFECT OF DECYL METHYL CARBINOL**

By

**GOH MENG CHUAN**

**October 2012**

**Chairman: Associate Professor Chong Pei Pei, PhD**

**Faculty: Medicine and Health Sciences**

*Candida dubliniensis* is a *Candida* species that is closely related to *Candida albicans*. Although they share high similarity in both genotypic and phenotypic characteristics including the capability to undergo mycelia transformation from yeast to hypha/pseudohyphae, *C. dubliniensis* is found to be less virulent, more susceptible to antifungal drugs and more sensitive to oxidative stress environment when compared to *C. albicans*. Superoxide dismutase (Sod) is an important enzyme that disproportionates superoxide free radical anions that are generated from oxidative stress. Although *C. dubliniensis* consists of all the *SOD* ortholog sequences which have been found in *C. albicans*, *C. dubliniensis* is unable to sustain significant growth at temperature higher than 40°C when compared to *C. albicans*. Apart from that, quorum sensing molecule (QSM) has been found to be proficient in suppressing morphological switching from yeast to hyphal transition in both *C. albicans* and *C. dubliniensis*. Decyl methyl carbinol (2-dodecanol), a recently identified QSM molecule that has been tested in *C. albicans* is found to have suppressed the yeast to hyphal transformation in hyphal-induced condition. 2-dodecanol also found to suppress certain hyphal-specific genes (HSGs). The current study is focused on the morphological changes of *C. dubliniensis* at different

temperatures. The *SOD* genes in *C. dubliniensis* also have been sequenced and characterized and the expression profile at 37°C and 42°C are studied in comparison to *C. albicans*. The genes expression profiles were quantified using relative quantification real time PCR. Two (2)-dodecanol was used to treat both *C. dubliniensis* and *C. albicans*. The morphological transition, toxicity effect and growth rate in both *C. dubliniensis* & *albicans* were recorded under the effect of 2-dodecanol. *C. dubliniensis* HSGs expression profiles were recorded after exposing 2-dodecanol to *C. dubliniensis* under the hyphal-induced condition. A clinical isolate that is isolated from University Malaya Medical Centre was also molecularly characterized in this study. In the results, *C. dubliniensis* has formed pseudohyphal at 42°C but unable to grow at temperatures higher than 42°C. The *SOD* sequences from *C. dubliniensis* were posted with 85-95% similarity when aligned with the orthologs sequences from *C. albicans*. Despite sharing a certain level of similarities in the *SOD* sequences, a total of 5 *SOD* genes in *C. dubliniensis* were up-regulated in 42°C when compared to *C. albicans* that with only 2 *SOD* genes being up-regulated. This study discovers that, the over-expression of Sod in *C. dubliniensis* could lead to the accumulations of high concentration of hydrogen peroxide and cause *C. dubliniensis* unable to survive in high temperatures. 2-dodecanol, however, showed with the effect to preserve both *C. albicans* and *C. dubliniensis* in round shaped yeast form. Nevertheless, *C. dubliniensis* is significantly more sensitive to high concentration of 2-dodecanol in terms of fungicidal and fungistatic effect. HSGs expression in *C. dubliniensis* under the exposure to 2-dodecanol also vary from what has been seen in *C. albicans* and constant expression of *cdHSP90* can be the main factor that kept *C. dubliniensis* in the yeast form. Interestingly, in this study, we also discovered that the clinical isolate that resembled both *C. dubliniensis* & *C. albicans* are the potential inter-species or intra-species between these two closely related species.

Abstract of thesis presented to senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

**FUNCTIONAL CHARACTERIZATION OF SUPEROXIDE DISMUTASE IN  
*Candida dubliniensis* AND EFFECT OF DECYL METHYL CARBINOL**

By

**GOH MENG CHUAN**

**October 2012**

**Chairman: Associate Professor Chong Pei Pei, PhD**

**Faculty: Medicine and Health Sciences**

*Candida dubliniensis* is a *Candida* species that is closely related to *Candida albicans*. Although they share high similarity in both genotypic and phenotypic characteristics including the capability to undergo mycelia transformation from yeast to hypha/pseudohyphae, *C. dubliniensis* is found to be less virulent, more susceptible to antifungal drugs and more sensitive to oxidative stress environment when compared to *C. albicans*. Superoxide dismutase (Sod) is an important enzyme that disproportionates superoxide free radical anions that are generated from oxidative stress. Although *C. dubliniensis* consists of all the *SOD* ortholog sequences which have been found in *C. albicans*, *C. dubliniensis* is unable to sustain significant growth at temperature higher than 40°C when compared to *C. albicans*. Apart from that, quorum sensing molecule (QSM) has been found to be proficient in suppressing morphological switching from yeast to hyphal transition in both *C. albicans* and *C. dubliniensis*. Decyl methyl carbinol (2-dodecanol), a recently identified QSM molecule that has been tested in *C. albicans* is found to have suppressed the yeast to hyphal transformation in hyphal-induced condition. 2-dodecanol also found to suppress certain hyphal-specific genes (HSGs). The current study is focused on the morphological changes of *C. dubliniensis* at different

temperatures. The *SOD* genes in *C. dubliniensis* also have been sequenced and characterized and the expression profile at 37°C and 42°C are studied in comparison to *C. albicans*. The genes expression profiles were quantified using relative quantification real time PCR. Two (2)-dodecanol was used to treat both *C. dubliniensis* and *C. albicans*. The morphological transition, toxicity effect and growth rate in both *C. dubliniensis* & *albicans* were recorded under the effect of 2-dodecanol. *C. dubliniensis* HSGs expression profiles were recorded after exposing 2-dodecanol to *C. dubliniensis* under the hyphal-induced condition. A clinical isolate that is isolated from University Malaya Medical Centre was also molecularly characterized in this study. In the results, *C. dubliniensis* has formed pseudohyphal at 42°C but unable to grow at temperatures higher than 42°C. The *SOD* sequences from *C. dubliniensis* were posted with 85-95% similarity when aligned with the orthologs sequences from *C. albicans*. Despite sharing a certain level of similarities in the *SOD* sequences, a total of 5 *SOD* genes in *C. dubliniensis* were up-regulated in 42°C when compared to *C. albicans* that with only 2 *SOD* genes being up-regulated. This study discovers that, the over-expression of Sod in *C. dubliniensis* could lead to the accumulations of high concentration of hydrogen peroxide and cause *C. dubliniensis* unable to survive in high temperatures. 2-dodecanol, however, showed with the effect to preserve both *C. albicans* and *C. dubliniensis* in round shaped yeast form. Nevertheless, *C. dubliniensis* is significantly more sensitive to high concentration of 2-dodecanol in terms of fungicidal and fungistatic effect. HSGs expression in *C. dubliniensis* under the exposure to 2-dodecanol also vary from what has been seen in *C. albicans* and constant expression of *cdHSP90* can be the main factor that kept *C. dubliniensis* in the yeast form. Interestingly, in this study, we also discovered that the clinical isolate that resembled both *C. dubliniensis* & *C. albicans* are the potential inter-species or intra-species between these two closely related species.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**PENCIRIAN FUNGSI SUPEROXIDE DISMUTASE DALAM *Candida dubliniensis*  
DAN KESAN DECYL METHYL CARBINOL**

By

**GOH MENG CHUAN**

**Oktober 2012**

**Pengerusi: Professor Madya Chong Pei Pei, PhD**

**Fakulti: Perubatan dan Sains Kesihatan**

*Candida dubliniensis* merupakan species *Candida* yang paling berkait rapat dengan *Candida albicans*. Walaupun mereka berkongsi banyak persamaan dari segi genotip and fenotip yang termasuk keupayaan untuk menjalani transformasi daripada bentuk yis kepada hyphal ataupun pseudohyphal, *C. dubliniensis* didapati kurang berkesan dari segi kebisaan terhadap sel, lebih terjejas kepada ubat-ubatan antikulat dan kekurangan keupayaan untuk hidup dalam persekitaran yang mengandungi tekanan oksidatif yang tinggi berbanding dengan *C. albicans*. Superoxide dismutase (Sod) merupakan enzim yang penting untuk menurunkan atau mengoksidakan ion negatif superoxide radikal yang dijana daripada tekanan oksidatif. Walaupun *C. dubliniensis* mempunyai semua urutan ortholog gen *SOD* daripada *C. albicans*, *C. dubliniensis* tidak dapat menyesuaikan dirinya apabila terdedah kepada suhu yang lebih tinggi ataupun sama dengan 40°C dan akan berhenti mereplikasi atau mati. Selain itu, molekul quorum sensing (QSM) juga didapati berkesan untuk melindungi *C. albicans* dan *C. dubliniensis* daripada menjalankan transformasi yis kepada bentuk hyphal ataupun pseudohyphal. Salah satu QSM yang baru temu dunia, decyl methyl carbinol (2-dodecanol) didapati dapat menghalang pembentukan hyphal atau pseudohyphal daripada bentuk yis. 2-dodecanol



juga dapat menindas ungkapan gen hyphal spesifik (HSGs) tertentu. Dalam kajian ini, tumpuan perhatian telah diberi kepada perubahan *C. dubliniensis* dari segi sel morfologi ketika disimpan pada suhu yang berbeza. Kami juga telah mengatur dan membandingkan urutan gen *SOD* dari *C. dubliniensis* dengan urutan gen *SOD* dari *C. albicans*. Ekspresi gen *SOD* daripada *C. dubliniensis* dalam suhu persekitaran 37°C dan 42°C juga dirakamkan dengan menggunakan kaedah kuantifikasi relatif *real time* PCR. Selain itu, 2-dodecanol telah digunakan untuk mengkaji kesannya ke atas peralihan morfologi, ketoksikan dan kadar pertumbuhan dalam *C. dubliniensis* dan *C. albicans*. Ekspresi gen HSGs daripada *C. dubliniensis* juga direkodkan dengan menggunakan kaedah kuantifikasi relatif *real time* PCR. *C. dubliniensis* didapati telah bertumbuh kepada pseudohyphal pada suhu 42°C dan tidak dapat bertumbuh lagi dalam suhu yang lebih tinggi daripada 42°C. Gen *SOD* daripada *C. dubliniensis* juga didapati mempunyai persamaan di antara 85 hingga 95 peratus jika berbanding dengan gen *ortholog* daripada *C. albicans*. Walaubagaimanapun, 5 gen *SOD* dari *C. dubliniensis* telah menunjukkan peningkatan yang jelas dalam suhu 42°C berbanding dengan hanya 2 gen *SOD* dari *C. albicans*. Hipotesis daripada kajian ini telah mencadangkan ungkapan gen *SOD* yang berlebihan di dalam *C. dubliniensis* telah meningkatkan kepekatan *hydrogen peroxide* yang sangat tinggi di dalam *C. dubliniensis* dan menyebabkan ia tidak dapat menyesuaikan dirinya dalam suhu yang tinggi. 2-dodecanol, bagaimanapun, telah menunjukkan kesannya yang mengekalkan *C. dubliniensis* dan *C. albicans* dalam bentuk yis. Tetapi, *C. dubliniensis* lebih sensitif kepada kepekatan 2-dodecanol yang tinggi. Kepekatan 2-dodecanol yang tinggi akan menghalang *C. dubliniensis* daripada menghasilkan generasi yang baru atau akan membunuhkannya. HSGs gen daripada *C. dubliniensis* juga mempunyai cara gen ekspresi yang berlainan jika berbanding dengan *C. albicans*. Daripada kesimpulan kajian ini, ekspresi *cdHSP90* yang tinggi mungkin disebabkan oleh perangsangan 2-dodecanol

dan kemungkinan merupakan faktor besar yang mengekalkan *C. dubliniensis* dalam bentuk yis. Kajian ini juga mempunyai pengesanan yang menarik, di mana kemungkinan yang besar species *Candida* daripada hospital Pusat Perubatan Universiti Malaya yang mempunyai ciri-ciri pertengahan diantara *C. dubliniensis* dengan *C. albicans*, adalah species yang baru.



## ACKNOWLEDGEMENT

First and foremost, I would like to express my heartiest thanks to my supervisor, Associate Professor Dr. Chong Pei Pei for her invaluable guidance, encouragement and endless support throughout this challenging study. Her constructive criticisms have been crucial in ensuring the success of this project as well as the writing of this thesis.

Special thanks to my co-supervisor, Professor Dr. Ng Kee Peng for providing the clinical specimens that involved in this study. My sincere thanks to my laboratory mates, Phelim, Crystale, Chee Hong, Alireza, Nabil, Asykin, Pey Yee, Alan, Matun, Shira and Puspa throughout the study. Your assistance, idea sharing and understanding are delightful. To the laboratory staff, Kak Ruhaidah, Fatimah and Intan for their kindness to maintain the lab in a good condition and accompany me during over-time lab work.

I would like to express my gratitude to Research University Grants (RUGS) for supporting my research and the fellowship through out my study.

Last but not least, a big thank you for my parents, my sisters and Chieng Ping for their support and encouragement that helps me to go through my difficult time during my study.

I certify that a Thesis Examination Committee has met on 17<sup>th</sup> October 2012 to conduct the final examination of Goh Meng Chuan on his thesis entitled “Functional Characterization of Superoxide Dismutase and Decyl Methyl Carbinol Effect on *Candida dubliniensis*” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the master degree.

Members of the Thesis Examination Committee were as follows:



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

**Chong Pei Pei, PhD**

Associate Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Chairman)

**Rozita Rosli, PhD**

Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Member)

**Ng Kee Peng, MD, PhD**

Professor  
Faculty of Medicine  
Universiti of Malaya  
(Member)

---

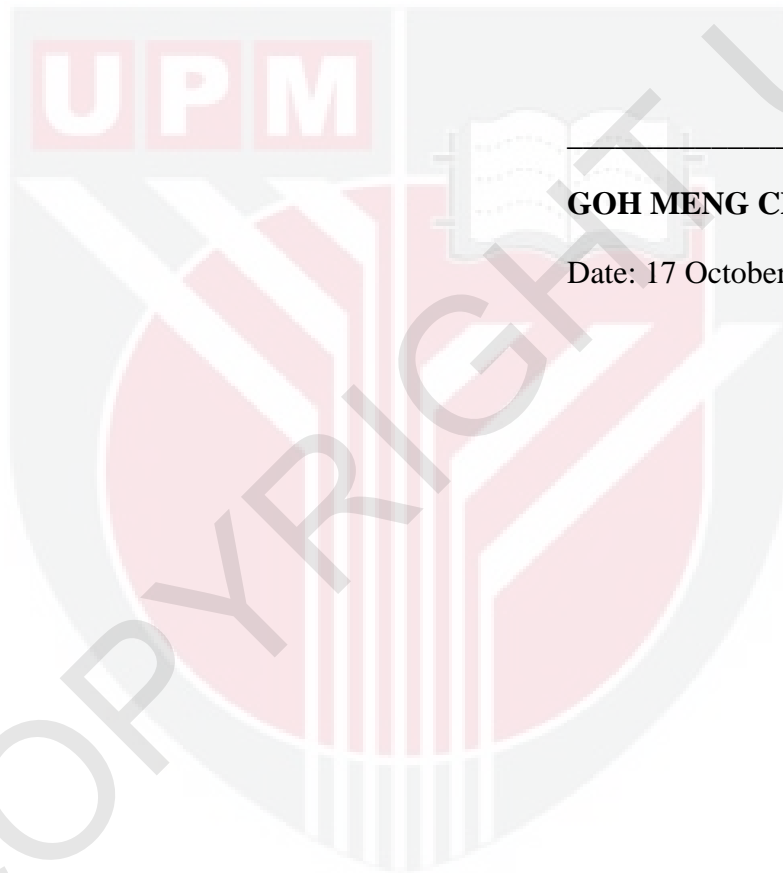
**BUJANG BIN KIM HUAT, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

## DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.



---

**GOH MENG CHUAN**

Date: 17 October 2012

## TABLE OF CONTENTS

	<b>Page</b>
<b>DEDICATION</b>	ii
<b>ABSTRACT</b>	iii
<b>ABSTRAK</b>	v
<b>ACKNOWLEDGEMENTS</b>	viii
<b>APPROVAL</b>	ix
<b>DECLARATION</b>	xi
<b>LIST OF TABLES</b>	xvi
<b>LIST OF FIGURES</b>	xvii
<b>LIST OF ABBREVIATIONS</b>	xxi
<b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	<b>1</b>
<b>2 LITERATURE REVIEW</b>	<b>6</b>
2.1 <i>Candida</i> spp.	6
2.1.1 Candidiasis	7
2.1.1.1 Superficial candidiasis	8
2.1.1.2 Systemic candidiasis	9
2.1.2 Pathogenic <i>Candida</i> spp.	9
2.1.3 <i>Candida albicans</i>	10
2.1.4 <i>Candida dubliniensis</i>	11
2.2. Virulence factors of <i>Candida</i> spp.	12
2.2.1 Morphological switching	13
2.2.2 Adherence	15
2.2.3 Hydrolytic enzymes	16
2.2.4 Biofilm formation	16
2.3 Oxidative stress and superoxide dismutase	17
2.3.1 Reactive oxygen species	17
2.3.2 Superoxide dismutase	17
2.3.3 Importance of superoxide dismutase in pathogenic fungus	19
2.3.4 <i>Candida</i> spp. superoxide dismutase	19
2.3.5 <i>Candida albicans</i> copper- and zinc- containing superoxide dismutase	20
2.3.6 <i>Candida albicans</i> manganese- containing superoxide dismutase	21
2.4 Cells signalling	22
2.4.1 Signalling pathway in <i>Candida</i> spp.	22
2.4.2 Pathway involved in morphological switching	23
2.4.3 Quorum sensing in microorganism	24
2.4.3.1 Quorum sensing molecules	25
2.4.3.2 Decyl methyl carbinol	25

2.5	Hyphal-Specifics Genes	26
2.5.1	Silence Information Regulator 2 Gene	26
2.5.2	Hyphal-Wall Protein 1 Gene	27
2.5.3	Secreted Aspartyl Proteinase 4 Gene	28
2.5.4	Agglutinin-Like Sequence 1 Gene	28
2.5.5	Heat Shock Protein 90 Gene	29
<b>3</b>	<b>METHODOLOGY</b>	<b>30</b>
3.1	Study design	30
3.2	Strains and cultures maintenance	33
3.2.1	Strains	33
3.2.2	Maintenance of cultures	34
3.2.3	High temperature growth environment	35
3.2.4	Hyphal induced conditioned medium preparation	36
3.3	Antifungal susceptibility test	37
3.3.1	E-Test	37
3.3.2	RPMI-1640 agar plate preparation	38
3.3.3	Fluconazole susceptibility testing	38
3.4	DNA extraction and purifications	39
3.4.1	DNA preparation from <i>Candida</i> spp. cultures	40
3.4.2	Conventional DNA extraction	40
3.4.3	Ethanol precipitation	41
3.5	Total RNA extraction	41
3.5.1	Phase determination	42
3.5.2	RNA isolation	42
3.5.3	RNA concentration measurement, purity and integrity check	42
3.5.4	DNase treatment and cDNA preparation	43
3.5.5	cDNA purity check	44
3.5.6	Primer design	44
3.6	Polymerase Chain Reaction	50
3.6.1	Conventional PCR	50
3.6.2	Agarose gel electrophoresis	51
3.6.3	Gradient PCR	51
3.6.4	Molecular identification of <i>Candida</i> spp.	52
3.6.5	Random Amplification Polymorphic DNA PCR	53
3.6.6	Real time PCR	54
3.6.7	Real time PCR standard curves construction	55
3.6.8	Relative quantitative real time RT PCR	56
<b>4</b>	<b>RESULTS</b>	<b>57</b>
4.1	Phenotypic difference of <i>C. dubliniensis</i> , <i>C. albicans</i> and clinical isolate 141	57
4.2	Antifungal susceptibility test	59



4.3	Determination of genetic relationship between clinical isolate 141, <i>C. albicans</i> and <i>C. dubliniensis</i>	62
4.3.1	Molecular characterization via Internal Transcribed Spacer sequence	62
4.3.2	Random Amplification Polymorphic DNA PCR	65
4.4	Effect of temperature shift on different <i>Candida</i> spp. growth rate and genes expression	67
4.4.1	Colonies morphological changes in different temperatures	67
4.4.2	Cellular morphology changes in 37°C and 42°C	69
4.4.3	Comparison between <i>Candida</i> spp. cell density count and growth rate in 37°C and 42°C cultures.	71
4.4.4	<i>C. dubliniensis</i> <i>SOD</i> genes sequence comparison	74
4.4.4.1	Cytosolic copper- and zinc- containing superoxide dismutase ( <i>SOD1</i> )	74
4.4.4.2	Mitochondrial manganese- containing superoxide dismutase ( <i>SOD2</i> )	77
4.4.4.3	Cytosolic manganese- containing superoxide dismutase ( <i>SOD3</i> )	79
4.4.4.4	Cell surface copper- and zinc- containing superoxide dismutase ( <i>SOD4</i> , <i>SOD5</i> and <i>SOD6</i> )	81
4.4.5	<i>SOD</i> genes expression profiling	84
4.4.5.1	RNA isolation	84
4.4.5.2	Relative quantitative real time PCR	87
4.4.5.3	<i>SOD1</i> gene expression in <i>C. dubliniensis</i> and <i>C. albicans</i>	92
4.4.5.4	<i>SOD2</i> gene expression in <i>C. dubliniensis</i> and <i>C. albicans</i>	92
4.4.5.5	<i>SOD3</i> gene expression in <i>C. dubliniensis</i> and <i>C. albicans</i>	93
4.4.5.6	<i>SOD4</i> gene expression in <i>C. dubliniensis</i> and <i>C. albicans</i>	94
4.4.5.7	<i>SOD5</i> gene expression in <i>C. dubliniensis</i> and <i>C. albicans</i>	94
4.4.5.8	<i>SOD6</i> gene expression in <i>C. dubliniensis</i> and <i>C. albicans</i>	95
4.4.5.9	<i>SOD1Chap</i> and <i>SODC5</i> putative gene expression in <i>C. dubliniensis</i>	95
4.5	Effect of quorum sensing molecule on different <i>Candida</i> spp.	102
4.5.1	2-dodecanol sensitivity testing	102
4.5.2	Cellular morphology changes under 2-dodecanol effect	103
4.5.3	Comparison of cell density count and growth rate of different <i>Candida</i> spp. upon exposure to 2-dodecanol	109
4.5.4	Hyphal-specific genes expression profile upon exposure to 2-dodecanol	115
4.5.4.1	RNA isolation	115
4.5.4.2	Gene expression in <i>C. dubliniensis</i> MYA-178 upon exposure to 2-dodecanol	116

4.5.4.3	<i>SIR2</i> gene expression in <i>C. dubliniensis</i> after exposure to 2-dodecanol	120
4.5.4.4	<i>HWP1</i> gene expression in <i>C. dubliniensis</i> after exposure to 2-dodecanol	120
4.5.4.5	<i>SAP4</i> gene expression in <i>C. dubliniensis</i> after exposure to 2-dodecanol	121
4.5.4.6	<i>ALS1</i> gene expression in <i>C. dubliniensis</i> after exposure to 2-dodecanol	121
4.5.4.7	<i>SOD1</i> gene expression in <i>C. dubliniensis</i> after exposure to 2-dodecanol	122
4.5.4.8	<i>HWP90</i> gene expression in <i>C. dubliniensis</i> after exposure to 2-dodecanol	122
<b>5</b>	<b>DISCUSSION</b>	<b>123</b>
5.1	Morphological, biochemical and phylogenetics differences between <i>C. dubliniensis</i> and <i>C. albicans</i>	123
5.2	Phenotypic difference of <i>C. dubliniensis</i> , <i>C. albicans</i> and clinical isolate 141	124
5.3	Antifungal susceptibility test	125
5.4	Determination of genetic relationship between clinical isolate 141, <i>C. albicans</i> and <i>C. dubliniensis</i> by Internal Transcribed Spacer 1	126
5.5	Determination of genetic relationship between clinical isolate 141, <i>C. albicans</i> and <i>C. dubliniensis</i> by RAPD PCR	127
5.6	Differences in <i>Candida</i> spp.	129
5.7	Effect of temperature shift on different <i>Candida</i> spp. growth rate and gene expression	131
5.8	Colony morphology, cellular phenotype and cell density changes in 37°C and 42°C	131
5.9	<i>C. dubliniensis</i> superoxide dismutase sequence comparison	134
5.10	<i>SOD</i> genes expression profiling	135
5.11	Differences in <i>SOD</i> genes expression in <i>C. dubliniensis</i> and <i>C. albicans</i>	139
5.12	Effect of 2-dodecanol towards <i>C. dubliniensis</i>	140
5.13	Colony morphology, cellular phenotype and cell density changes in <i>C. dubliniensis</i> , <i>C. albicans</i> and clinical isolate 141 under the exposure of 2-dodecanol	140
5.14	Hyphal-specific genes expression profiling in <i>C. dubliniensis</i> under the exposure of 2-dodecanol	143
5.15	Differences in <i>HSG</i> genes expression in <i>C. dubliniensis</i>	146
<b>6</b>	<b>CONCLUSIONS AND FUTURE RECOMMENDATIONS</b>	<b>148</b>
6.1	Conclusions	148
6.2	Future recommendations	150
	<b>REFERENCES</b>	<b>151</b>
	<b>APPENDICES</b>	<b>167</b>
	<b>BIODATA OF STUDENT</b>	<b>196</b>



## LIST OF TABLES

Table		Page
2.1	Agent of opportunistic mycoses.	8
3.1	<i>Candida</i> spp. strains that involved in this study.	33
3.2	The total medium mixtures for culture with 2-dodecanol (0.02% and 0.005%) and without 2-dodecanol.	37
3.3	Specific designed primer pairs flanking the homologous sequences of putative SOD genes from <i>C. dubliniensis</i> CD36 database from GenBank.	46
3.4	Specific designed primer pairs based on the homologous sequences of putative SOD genes from <i>C. dubliniensis</i> CD36 database from GenBank.	47
3.5	Specific designed primer pairs based on the SOD genes sequences from <i>C. albicans</i> from GenBank.	48
3.6	Specific designed primer pairs based on the homologous sequences from <i>C. dubliniensis</i> CD36 database from GenBank.	49
4.1	Indication of the fluconazole susceptibility E-Test reference range for <i>Candida</i> spp.	60
4.2	Selected BLAST hit of the most identical <i>Candida</i> spp. strains when compared to the ITS sequence of clinical isolates 141 after sequence blast in GenBank.	63
4.3	Genetic similarities of <i>Candida</i> spp. determined by calculating similarity coefficient (SAB) value.	65
4.4	Comparison between the percentages of growth rate per hour for <i>C. albicans</i> 14053, clinical isolate 141 and <i>C. dubliniensis</i> MYA-178 in SDB medium incubated at 37°C and 42°C.	73
4.5	Cells density count of <i>C. albicans</i> 14053, <i>C. dubliniensis</i> MYA-178 and Clinical Isolates Coded 141 in different concentrations of 2-dodecanol in a six-well plate.	111

## LIST OF FIGURES

Figure		Page
2.1	Three different phenotypic forms of <i>C. albicans</i> .	14
2.2	Demonstrated <i>C. albicans</i> morphological switching related signalling pathways by Berman and Sudbery (2002).	24
3.1	The distribution of different <i>Candida</i> spp. stripped on each agar plate.	35
3.2	Examples of MIC value judgement.	39
4.1	Phenotypic structure of <i>C. albicans</i> 14053, clinical isolate 141 and <i>C. dubliniensis</i> MYA-178 at different conditions.	58
4.2	Fluconazole E-Test result for <i>C. albicans</i> 14053, clinical isolate 141 and <i>C. dubliniensis</i> MYA-178 on RPMI-1640 medium buffered with MOPS.	61
4.3	Multiple ITS sequences alignment for clinical isolates 141 with two most identical <i>Candida</i> spp. after blast in NCBI database.	64
4.4	RAPD profile of <i>C. albicans</i> 14053, clinical isolates 141 and <i>C. dubliniensis</i> MYA-178 generated by using 3 different random primers, namely: (A) PST, (B) OPA09 and (C) OPA02 primer.	66
4.5	Dendogram of <i>C. albicans</i> 14053, <i>C. dubliniensis</i> MYA-178 and Clinical isolates 141 generated from RAPD-PCR profiles.	66
4.6	Different incubation temperatures showed different extents of growth in the different <i>Candida</i> spp.	68
4.7	Phenotypic structure of <i>C. albicans</i> 14053, clinical isolates 141 and <i>C. dubliniensis</i> MYA-178 at different conditions.	70
4.8	Direct comparison between the growth rates of <i>C. albicans</i> 14053, clinical isolates 141 and <i>C. dubliniensis</i> MYA-178 incubated at 37°C and 42°C.	73
4.9	Nucleotide and deduced <i>C. dubliniensis</i> MYA-178 <i>SOD1</i> transcript region generated from this study.	76

4.10	Nucleotide and deduced <i>C. dubliniensis</i> MYA-178 <i>SOD2</i> transcript region generated from this study.	78
4.11	Nucleotide and deduced <i>C. dubliniensis</i> MYA-178 <i>SOD3</i> transcript region generated in this study.	80
4.12	Nucleotide and deduced <i>SOD4</i> , <i>SOD5</i> , and partial <i>SOD6</i> , transcript region in <i>C. dubliniensis</i> MYA-178.	83
4.13	Nucleotide and deduced partial <i>SOD6</i> , partial <i>SOD1Chap</i> and partial <i>SODC5</i> transcript region in <i>C. dubliniensis</i> MYA-178.	84
4.14	Formaldehyde agarose gel showed the integrity of the total RNA extracted from <i>C. albicans</i> 14053 and <i>C. dubliniensis</i> MYA-178.	86
4.15	Representative of the relative quantification real time PCR cycling profile of 5 pair's of primers ( <i>CdAct100</i> , <i>CdSOD1</i> , <i>CdSOD2</i> , <i>CdSOD3</i> and <i>CdSOD4</i> ) that amplified different genes from cDNA of <i>C. dubliniensis</i> MYA-178 at different time points.	88
4.16	Representative of the relative quantification real time PCR cycling profile of another 4 pairs of primers ( <i>CdSOD5</i> , <i>CdSOD6</i> , <i>CdSOD1Chap</i> and <i>CdSODC5</i> ) that amplified different genes from cDNA of <i>C. dubliniensis</i> MYA-178 at different time points.	89
4.17	Representative of the relative quantification real time PCR cycling profile of another 4 pairs of primers ( <i>CaAct100</i> , <i>CaSOD1</i> , <i>CaSOD2</i> and <i>CaSOD3</i> ) that amplified different genes from cDNA of <i>C. albicans</i> 14053 at different time points.	90
4.18	Representative of the relative quantification real time PCR cycling profile of another 3 pairs of primers ( <i>CaSOD4</i> , <i>CaSOD5</i> and <i>CaSOD6</i> ) that amplified different genes from cDNA of <i>C. albicans</i> 14053 at different time points.	91
4.19	Four different <i>SOD</i> genes ( <i>CdSOD1</i> , <i>CdSOD2</i> , <i>CdSOD3</i> and <i>CdSOD4</i> ) expression were compared between parallel yeast forms samples incubated at 24 hours and 48 hours respectively.	97
4.20	Another four different <i>SOD</i> genes ( <i>CdSOD5</i> , <i>CdSOD6</i> , <i>CdSOD1Chap</i> and <i>CdSODC5</i> ) expression was compared between parallel yeast form samples incubated for 24 and 48 hours respectively.	98

4.21	Four different <i>SOD</i> genes ( <i>CdSOD1</i> , <i>CdSOD2</i> , <i>CdSOD3</i> and <i>CdSOD4</i> ) expression was compared between <i>C. dubliniensis</i> MYA-178 cultures incubated at 42°C for 6, 12, 24 and 48 hours respectively.	99
4.22	Four different <i>SOD</i> genes ( <i>CdSOD5</i> , <i>CdSOD6</i> , <i>CdSOD1Chap</i> and <i>CdSODC5</i> ) expression was compared between <i>C. dubliniensis</i> MYA-178 cultures incubated in 42°C at 6, 12, 24 and 48 hours respectively.	100
4.23	Six different <i>SOD</i> genes ( <i>CaSOD1</i> , <i>CaSOD2</i> , <i>CaSOD3</i> , <i>CaSOD4</i> , <i>CaSOD5</i> , and <i>CaSOD6</i> ) expression was compared between <i>C. albicans</i> 14053 cultures incubated at 42°C at 6, 12, 24 and 48 hours respectively.	101
4.24	Comparative growth of <i>C. dubliniensis</i> MYA-178 and <i>C. albicans</i> 14053 under the exposure to different concentrations of 2-dodecanol on SDA agar plate.	104
4.25	Cellular morphologies of <i>C. albicans</i> 14053, Clinical isolate 141 and <i>C. dubliniensis</i> MYA-178 incubated in hyphal-induced condition that consist of 5 mL of RPMI-1640 medium with 10% of FBS, 5% of CO <sub>2</sub> , in 37°C for 24 hours.	107
4.26	Morphological changes of <i>C. albicans</i> 14053, Clinical isolate 141 and <i>C. dubliniensis</i> MYA-178 after incubated in 5 mL of RPMI-1640 medium with 10% of FBS, 5% of CO <sub>2</sub> , 0.02% of 2-dodecanol for <i>C. albicans</i> 14053 and Clinical isolate 141, 0.005% of 2-dodecanol for <i>C. dubliniensis</i> MYA-178 in 37°C for 3, 6, 12 and 24 hours respectively.	108
4.27	Cell density counts of <i>C. albicans</i> 14053 versus time in different concentrations of 2-dodecanol in a six-well plate.	112
4.28	Cell density counts of Clinical isolate 141 versus time in different concentrations of 2-dodecanol out in a six-well plate.	113
4.29	Cell density counts of <i>C. dubliniensis</i> MYA-178 versus time in different concentrations of 2-dodecanol in a six-well plate.	114
4.30	Representative formaldehyde agarose gel showed the integrity of the total RNA extracted from <i>C. dubliniensis</i> MYA-178 under 2-dodecanol treatment.	116
4.31	Representative of the relative quantification real time PCR cycling profile of 4 pairs of primers ( <i>CdAct100</i> , <i>CdSIR2</i> , <i>CdHWP1</i> and <i>CdSOD1</i> ) that amplified respective genes from the cDNA of <i>C. dubliniensis</i> MYA-178 at different time points.	117



- 4.32 Representative of the relative quantification real time PCR cycling profile of another 3 pairs of primers (CdHSP90, CdALS1 and CdSAP4) that amplified respective genes from cDNA of *C. dubliniensis* MYA-178 extracted at different time points. 118
- 4.33 Relative expression of *CdSIR2*, *CdALS1*, *CdSOD1*, *CdHWPI*, *CdSAP4* and *CdHSP90* genes in *C. dubliniensis* MYA-178 after exposure to 0.005% of 2-dodecanol in RPMI-1640 medium containing 10% of FBS, incubated in 37°C oven with 5% of CO<sub>2</sub>. 119





## LIST OF ABBREVIATIONS

ALS	Agglutinin-Like Sequence
ATCC	American Type Culture Collection
BLAST	Basic Local Alignment Search Tool
bp	base-pair
cAMP	cyclic adenosine monophosphate
cDNA	complementary Deoxyribonucleic Acid
<i>C.</i>	<i>Candida</i>
<i>Ca</i>	<i>Candida albicans</i>
<i>Cd</i>	<i>Candida dubliniensis</i>
CDK	Cyclin-dependent kinase
cm	centimeter
CO <sub>2</sub>	carbon dioxide
CPH	<i>Candida</i> PseudoHyphal
C <sub>T</sub>	threshold cycle
DEPC	diethyl pyrocarbonate
DNA	Deoxyribonucleic acid
dNTPs	deoxynucleotide triphosphates
E	Amplification Efficiency
EDTA	ethylenediaminetetracetic acid
EFG	Enhanced Filamentous Growth

FBS	Fetal Bovine Serum
g	gram
H <sub>2</sub> O <sub>2</sub>	Hydrogen Peroxide
HSP	Heat Shock Protein
HWP	Hyphal Wall Protein
ITS	Internal Transcribed Spacer
kb	kilo-base
L	liter
M	Molar
MAP	Mitogen-activated Protein
mb	mega-base
mg	miligram
MgCl <sub>2</sub>	Magnesium Chloride
MIC	Minimum Inhibitory Concentration
mL	mililiter
mM	milimolar
mm	milimeter
MMLV	Moloney Murine Leukaemia Virus
mRNA	messenger Ribonucleic Acid
NCBI	National Center for Biotechnology Information
NJ	Neighbour-Joining

nm	nanometer
OD	Optical Density
O <sub>2</sub>	Oxygen
O <sub>2</sub> <sup>·-</sup>	Superoxide anion radical
PBS	phosphate buffer saline
PCR	polymerase chain reaction
pg	picogram
PL	Phospholipase
QSM	Quorum Sensing Molecule
R <sup>2</sup>	correlation coefficient
RAPD	random amplification of polymorphic DNA
RAS	Rat sarcoma
ROS	Reactive Oxygen Species
RNA	Ribonucleic Acid
rpm	revolution per minute
RPMI	Roswell Park Memorial Institute
RT	reverse transcription
S <sub>AB</sub>	similarity coefficient
SAP	Secreted Aspartyl Proteinase
SDA	Sabouraud Dextrose Agar
SDB	Sabouraud Dextrose Broth

SIR	Silent Information Regulator
SOD	Superoxide Dismutase
spp.	Species
TAE	Tris acetate EDTA
TBE	Tris Boric EDTA
UMMC	University of Malaya Medical Centre
UV	Ultra Violet
V	volt
v/v	volume/volume
w/v	weight/volume
141	clinical isolate 141
2-dodecanol	decyl methyl carbinol
%	percentage
°C	degree Celsius
μg	microgram
μL	microliter
·OH	hydroxyl radical

## CHAPTER 1

### INTRODUCTION

*Candida* species (spp.) are eukaryotic microorganisms that belong to the kingdom of Fungi, phylum of Ascomycota, Subphylum of Saccharomycotina, class of Saccharomycetes, Order of Saccharomycetales and family of Saccharomycetaceae. The capability of some fungal genera to undergo morphological changes had been reported. To date, more than 200 *Candida* spp. have been discovered and described (Kauffman *et al.*, 2011). Among them, only a few had been reported to cause medical complications. These include *C. albicans*, *C. parapsilosis*, *C. tropicalis*, *C. glabrata*, *C. guiliermondii*, *C. kefyr*, *C. krusei*, *C. lusitanae* and *C. dubliniensis*.

*Candida* spp. can cause a wide range of infections in humans. Most often, *Candida* spp. are considered as opportunistic pathogens and not true pathogens. *C. albicans* is considered as an inhabitant in human skin and mucosal surfaces and is part of the normal microflora. The spectrum of infections caused by *Candida* spp. includes superficial candidiasis (such as vaginal candidiasis, oral thrush, etc), candidemia and invasive candidiasis (deep-seated organ infections). The importance of *Candida* spp. can be seen in its prevalence in blood stream infections in hospitals, where *Candida* spp. ranked as the fourth most frequently isolated microorganism after bacteria species (Edmond *et al.*, 1999). The mortality rates associated with systemic candidiasis is relatively high if left untreated (Pacheco-Rios *et al.*, 1997; Friedman *et al.*, 2000; Akpan and Morgan, 2002).

The hidden identity of *C. dubliniensis* has been unearthed in the last decade, differentiating it from the most dominant species among pathogenic *Candida* spp., *C.*

*albicans*. This strange species which has made the oral cavity of immunocompromised patients as their home is often found to be less virulent, more susceptible to antifungal drugs and possess reduced survival rate under stress environment compared to *C. albicans* (Gilfillan *et al.*, 1998). Although *C. dubliniensis* rarely causes systemic infection compared to *C. albicans*, antifungal drug resistant *C. dubliniensis* strains had been recovered from infected patients (Moran *et al.*, 1997; Ruhnke *et al.*, 2000). Since then, many studies focusing on unravelling additional characteristics of this atypical *Candida* spp. and differentiating it from the most closely related *Candida* spp., *C. albicans* have been published.

Besides *C. dubliniensis*, *C. stellatoidea* and *C. africana* have also been reported to be closely related to *C. albicans* previously and were presumed to be separate species from *C. albicans*. Unlike *C. dubliniensis*, after years of studies, both species have now been classified as two of the sub-strains of *C. albicans* (McCullough *et al.*, 1999; Romeo and Criseo, 2009). Scientists believe that more and more subtypes of *Candida* spp. will be revealed in the future and this phenomenon has highlighted the importance to differentiate these species from one another in order to understand this genus better.

From the previous reports, one interesting finding is that *C. dubliniensis* seems to lack the ability to grow at temperatures higher than 40°C when compared to *C. albicans* (Sullivan *et al.*, 1995). On another hand, *C. albicans* have been found to consist of 6 different isoforms of superoxide dismutase (Sod), an anti-oxidant enzyme that is well recognized with its function to disproportionate superoxide free radical anions that are generated from the cells themselves when exposed to high temperature (Frealde *et al.*, 2005). The *SOD* genes have been identified and characterized in many organisms from

human, animals, plants and microbes. The function of *SOD* genes usually is to protect the host from the reactive oxygen species (ROS). Looking into a newly published genome database of *C. dubliniensis* strain CD36, it is found that *C. dubliniensis* genome library consisted of 8 putative *SOD* genes which may potentially mimic the superoxide dismutase enzymatic functions of *C. albicans*. This has triggered the hypothesis that some of the Sod enzymes in *C. dubliniensis* might possess different expression profiles when compared to *C. albicans* that lead to the disability of *C. dubliniensis* to grow at higher temperature.

Comparing the genetic properties of *C. albicans* and *C. dubliniensis*, the genome size of *C. dubliniensis* is 14.6 Mb while *C. albicans* is varies from 14.3 – 15.6 Mb, depending on the strains (Chibana *et al.*, 2005; Butler *et al.*, 2009). Both are diploid *Candida* spp., and share more than 98% similarity of the orthologous genes (Magee *et al.*, 2008).

Another interesting fact of *C. dubliniensis* is it is capable of undergoing morphological changes to form germ tube to hyphal or pseudohyphal forms and this ability to switch between yeast form and hyphal form has been known as one of the critical virulence factors for pathogenic *C. albicans*. This characteristic also serves as one of the diagnostic tools to distinguish *C. albicans* with other *Candida* spp. in laboratories, although it is no longer the most effective diagnostic method. Studies have pointed out that some of the hypha-specific genes in *C. albicans* are also found in *C. dubliniensis* genome. This suggests that *C. dubliniensis* can also be as pathogenic as *C. albicans*. Recently, cell to cell signalling has been studied intensively in all types of organisms and quorum sensing molecules have been reported to control communication between cells or within the cell itself. The potential of Decylmethyl carbinol (2-dodecanol) as a quorum sensing

molecule has caught the attention of researchers and has been tested in *C. albicans* where it had inhibited *C. albicans* from undergoing morphological transition (Davis-Hanna *et al.*, 2008; Lim *et al.*, 2009). As *C. dubliniensis* is closely related to *C. albicans*, 2-dodecanol may be able to block *C. dubliniensis* from undergoing phenotypic changes.

### **Research problems:**

Globally, *C. albicans* is still the most predominant species of *Candida* isolated from clinical specimens, although some other *Candida* spp. seem to have survival advantages over this species, such as inherent drug resistance to azole antifungal drugs. *C. krusei* and *C. glabrata* are two species that have inherited reduced-susceptibility to fluconazole, itraconazole and ketoconazole (Koga-Ito *et al.*, 2010; Pfaller *et al.*, 2004) Unlike *C. dubliniensis*, these 2 species are quite different from *C. albicans* as they are unable to form pseudohypha or true hypha. More importantly, *C. glabrata* is a haploid *Candida* spp. whereas *C. albicans* and *C. dubliniensis* are both diploid *Candida* spp. It is important to conduct more fundamental studies to understand what makes *C. albicans* such a successful opportunistic pathogen, so that potential therapeutic targets or enzymes can be identified as future strategies to eradicate this pathogen. In light of this, *C. dubliniensis* has been chosen as a model organism to study in comparison to *C. albicans*, as *C. dubliniensis* is rarely found to be a causative organism of systemic candidiasis.

### **Hypothesis:**

As *C. dubliniensis* had shown unique differences when compared to *C. albicans*, the hypothesis of this study focused on three parts. The first part of the study described the



growth differences between *C. dubliniensis* and *C. albicans* plus a clinical isolate that possesses close relationship between these two species. The second part of the study mainly aimed to shed light on the effect of 42°C towards the growth pattern, morphological changes and *SOD* gene expression in *C. dubliniensis* and *C. albicans*. The putative *SOD* gene sequences from *C. dubliniensis* were also compared with the sequences of *SOD* genes from *C. albicans*. The third focal point of the study was on the effect of the quorum-sensing molecule, 2-dodecanol towards the growth pattern, morphological changes and the expression of hypha-specific genes in *C. dubliniensis*.

#### **Objectives of the study:**

The general objectives of this study were to study the characteristics and behaviour of *C. dubliniensis* in a high temperature environment and when exposed to 2-dodecanol in comparison with *C. albicans*.

The specific aims of this study had been designed to focus on (1) to molecularly identify a clinical isolate that is similar to both *C. dubliniensis* and *C. albicans*, (2) to compare the different morphological and growth profiles between *C. dubliniensis* and *C. albicans* when grown in 37°C and 42°C, (3) to determine the *C. dubliniensis* *SOD* putative gene expression levels at 37°C and 42°C growth temperatures, (4) to identify the effect of 2-dodecanol in *C. dubliniensis* and (5) to determine the hypha-specific genes (HSGs) expression in *C. dubliniensis* upon exposure to 2-dodecanol.

## REFERENCES

- Abegg, M. A., P. V. Alabarse, A. Casanova, J. Hoscheid, T. B. Salomon, F. S. Hackenhaar, T. M. Medeiros, and M. S. Benfato (2010), Response to oxidative stress in eight pathogenic yeast species of the genus *Candida*, *Mycopathologia*, 170(1), 11-20.
- Akpan, A., and R. Morgan (2002), Oral candidiasis, *Postgrad Med J*, 78(922), 455-459.
- Anderson, J., L. Cundiff, B. Schnars, M. X. Gao, I. Mackenzie, and D. R. Soll (1989), Hypha formation in the white-opaque transition of *Candida albicans*, *Infect Immun*, 57(2), 458-467.
- Arif, M., D. R. Pani, N. W. Zaidi, and U. S. Singh (2011), PCR-Based Identification and Characterization of *Fusarium* sp. Associated with Mango Malformation, *Biotechnol Res Int*, 2011, 141649.
- Asmundsdottir, L. R., H. Erlendsdottir, B. A. Agnarsson, and M. Gottfredsson (2009), The importance of strain variation in virulence of *Candida dubliniensis* and *Candida albicans*: results of a blinded histopathological study of invasive candidiasis, *Clin Microbiol Infect*, 15(6), 576-585.
- Baires-Varguez, L., A. Cruz-Garcia, L. Villa-Tanaka, S. Sanchez-Garcia, L. A. Gaitan-Cepeda, L. O. Sanchez-Vargas, G. Quindos, and C. Hernandez-Rodriguez (2007), Comparison of a randomly amplified polymorphic DNA (RAPD) analysis and ATB ID 32C system for identification of clinical isolates of different *Candida* species, *Rev Iberoam Micol*, 24(2), 148-151.
- Baker, J. G., I. F. Salkin, D. H. Pincus, and R. F. D'Amato (1981), Diagnostic characters of an atypical *Candida*, *J Clin Microbiol*, 13(1), 199-203.
- Bautista-Munoz, C., X. M. Boldo, L. Villa-Tanaca, and C. Hernandez-Rodriguez (2003), Identification of *Candida* spp. by randomly amplified polymorphic DNA analysis and differentiation between *Candida albicans* and *Candida dubliniensis* by direct PCR methods, *J Clin Microbiol*, 41(1), 414-420.
- Beck-Sague, C., and W. R. Jarvis (1993), Secular trends in the epidemiology of nosocomial fungal infections in the United States, 1980-1990. National Nosocomial Infections Surveillance System, *J Infect Dis*, 167(5), 1247-1251.
- Beno, D. W., and H. L. Mathews (1993), Quantitative measurement of lymphocyte mediated growth inhibition of *Candida albicans*, *J Immunol Methods*, 164(2), 155-164.
- Berman, J., and P. E. Sudbery (2002), *Candida albicans*: a molecular revolution built on lessons from budding yeast, *Nat Rev Genet*, 3(12), 918-930.

- Boerlin, P., F. Boerlin-Petzold, C. Durussel, M. Addo, J. L. Pagani, J. P. Chave, and J. Bille (1995), Cluster of oral atypical *Candida albicans* isolates in a group of human immunodeficiency virus-positive drug users, *J Clin Microbiol*, 33(5), 1129-1135.
- Borg-von Zepelin, M., S. Beggah, K. Boggian, D. Sanglard, and M. Monod (1998), The expression of the secreted aspartyl proteinases Sap4 to Sap6 from *Candida albicans* in murine macrophages, *Mol Microbiol*, 28(3), 543-554.
- Boucher, H., S. Mercure, S. Montplaisir, and G. Lemay (1996), A novel group I intron in *Candida dubliniensis* is homologous to a *Candida albicans* intron, *Gene*, 180(1-2), 189-196.
- Bourne, H. R., D. A. Sanders, and F. McCormick (1990), The GTPase superfamily: a conserved switch for diverse cell functions, *Nature*, 348(6297), 125-132.
- Braun, B. R., and A. D. Johnson (2000), TUP1, CPH1 and EFG1 make independent contributions to filamentation in *Candida albicans*, *Genetics*, 155(1), 57-67.
- Brown, A. J., and N. A. Gow (1999), Regulatory networks controlling *Candida albicans* morphogenesis, *Trends Microbiol*, 7(8), 333-338.
- Brown, D. H., Jr., A. D. Giusani, X. Chen, and C. A. Kumamoto (1999), Filamentous growth of *Candida albicans* in response to physical environmental cues and its regulation by the unique CZF1 gene, *Mol Microbiol*, 34(4), 651-662.
- Burchett, S. A., M. L. Volk, M. J. Bannon, and J. G. Granneman (1998), Regulators of G protein signaling: rapid changes in mRNA abundance in response to amphetamine, *J Neurochem*, 70(5), 2216-2219.
- Burt, E. T., R. Daly, D. Hoganson, Y. Tsurulnikov, M. Essmann, and B. Larsen (2003), Isolation and partial characterization of Hsp90 from *Candida albicans*, *Ann Clin Lab Sci*, 33(1), 86-93.
- Butler, G., et al. (2009), Evolution of pathogenicity and sexual reproduction in eight *Candida* genomes, *Nature*, 459(7247), 657-662.
- Calderone, R. A., and P. C. Braun (1991), Adherence and receptor relationships of *Candida albicans*, *Microbiol Rev*, 55(1), 1-20.
- Calderone, R. A., and W. A. Fonzi (2001), Virulence factors of *Candida albicans*, *Trends Microbiol*, 9(7), 327-335.
- Carrillo, D., J. Vicente-Soler, J. Fernandez, T. Soto, J. Cansado, and M. Gacto (1995), Activation of cytoplasmic trehalase by cyclic-AMP-dependent and cyclic-AMP-independent signalling pathways in the yeast *Candida utilis*, *Microbiology*, 141 ( Pt 3), 679-686.

- Chaffin, W. L., J. L. Lopez-Ribot, M. Casanova, D. Gozalbo, and J. P. Martinez (1998), Cell wall and secreted proteins of *Candida albicans*: identification, function, and expression, *Microbiol Mol Biol Rev*, 62(1), 130-180.
- Chandra, J., D. M. Kuhn, P. K. Mukherjee, L. L. Hoyer, T. McCormick, and M. A. Ghannoum (2001), Biofilm formation by the fungal pathogen *Candida albicans*: development, architecture, and drug resistance, *J Bacteriol*, 183(18), 5385-5394.
- Chary, P., D. Dillon, A. L. Schroeder, and D. O. Natvig (1994), Superoxide dismutase (sod-1) null mutants of *Neurospora crassa*: oxidative stress sensitivity, spontaneous mutation rate and response to mutagens, *Genetics*, 137(3), 723-730.
- Chen, H., M. Fujita, Q. Feng, J. Clardy, and G. R. Fink (2004), Tyrosol is a quorum-sensing molecule in *Candida albicans*, *Proc Natl Acad Sci U S A*, 101(14), 5048-5052.
- Chen, Y. C., S. C. Chang, C. C. Sun, L. S. Yang, W. C. Hsieh, and K. T. Luh (1997), Secular trends in the epidemiology of nosocomial fungal infections at a teaching hospital in Taiwan, 1981 to 1993, *Infect Control Hosp Epidemiol*, 18(5), 369-375.
- Chibana, H., N. Oka, H. Nakayama, T. Aoyama, B. B. Magee, P. T. Magee, and Y. Mikami (2005), Sequence finishing and gene mapping for *Candida albicans* chromosome 7 and syntenic analysis against the *Saccharomyces cerevisiae* genome, *Genetics*, 170(4), 1525-1537.
- Cho, T., T. Aoyama, M. Toyoda, H. Nakayama, H. Chibana, H. Kaminishi, and R. A. Calderone (2008), Farnesol as a quorum-sensing molecule in *Candida albicans*, *Nippon Ishinkin Gakkai Zasshi*, 49(4), 281-286.
- Ciardo, D. E., G. Schar, E. C. Bottger, M. Altwegg, and P. P. Bosshard (2006), Internal transcribed spacer sequencing versus biochemical profiling for identification of medically important yeasts, *J Clin Microbiol*, 44(1), 77-84.
- Coleman, D. C., D. J. Sullivan, D. E. Bennett, G. P. Moran, H. J. Barry, and D. B. Shanley (1997), Candidiasis: the emergence of a novel species, *Candida dubliniensis*, *AIDS*, 11(5), 557-567.
- Coleman, D. C., D. E. Bennett, D. J. Sullivan, P. J. Gallagher, M. C. Henman, D. B. Shanley, and R. J. Russell (1993), Oral *Candida* in HIV infection and AIDS: new perspectives/new approaches, *Crit Rev Microbiol*, 19(2), 61-82.
- d'Enfert, C., and B. Hube (2007), *Candida: Comparative and Functional Genomics*, 428 pp., Caister Academic Press.
- Daniels, K. J., T. Srikantha, S. R. Lockhart, C. Pujol, and D. R. Soll (2006), Opaque cells signal white cells to form biofilms in *Candida albicans*, *EMBO J*, 25(10), 2240-2252.

- Davidson, J. F., B. Whyte, P. H. Bissinger, and R. H. Schiestl (1996), Oxidative stress is involved in heat-induced cell death in *Saccharomyces cerevisiae*, *Proc Natl Acad Sci U S A*, 93(10), 5116-5121.
- Davis-Hanna, A., A. E. Piispanen, L. I. Stateva, and D. A. Hogan (2008), Farnesol and dodecanol effects on the *Candida albicans* Ras1-cAMP signalling pathway and the regulation of morphogenesis, *Mol Microbiol*, 67(1), 47-62.
- De Larkin, V. P. (1957), Polyoxyethylene dodecanol vaporization in the treatment of respiratory infections of infants and children, *N Y State J Med*, 57(16), 2667-2672.
- Decanis, N., N. Tazi, A. Correia, M. Vilanova, and M. Rouabhia (2011), Farnesol, a fungal quorum-sensing molecule triggers *Candida albicans* morphological changes by downregulating the expression of different secreted aspartyl proteinase genes, *Open Microbiol J*, 5, 119-126.
- Diaz-Guerra, T. M., E. Mellado, M. Cuenca Estrella, F. Laguna, and J. L. Rodriguez-Tudela (1999), Molecular characterization by PCR-fingerprinting of *Candida dubliniensis* strains isolated from two HIV-positive patients in Spain, *Diagn Microbiol Infect Dis*, 35(2), 113-119.
- Edmond, M. B., S. E. Wallace, D. K. McClish, M. A. Pfaller, R. N. Jones, and R. P. Wenzel (1999), Nosocomial bloodstream infections in United States hospitals: a three-year analysis, *Clin Infect Dis*, 29(2), 239-244.
- El Barkani, A., O. Kurzai, W. A. Fonzi, A. Ramon, A. Porta, M. Frosch, and F. A. Muhlschlegel (2000), Dominant active alleles of RIM101 (PRR2) bypass the pH restriction on filamentation of *Candida albicans*, *Mol Cell Biol*, 20(13), 4635-4647.
- Enjalbert, B., G. P. Moran, C. Vaughan, T. Yeomans, D. M. Maccallum, J. Quinn, D. C. Coleman, A. J. Brown, and D. J. Sullivan (2009), Genome-wide gene expression profiling and a forward genetic screen show that differential expression of the sodium ion transporter *Ena21* contributes to the differential tolerance of *Candida albicans* and *Candida dubliniensis* to osmotic stress, *Mol Microbiol*, 72(1), 216-228.
- Fabrizio, P., S. D. Pletcher, N. Minois, J. W. Vaupel, and V. D. Longo (2004), Chronological aging-independent replicative life span regulation by *Msn2/Msn4* and *Sod2* in *Saccharomyces cerevisiae*, *FEBS Lett*, 557(1-3), 136-142.
- Fabrizio, P., C. Gattazzo, L. Battistella, M. Wei, C. Cheng, K. McGrew, and V. D. Longo (2005), Sir2 blocks extreme life-span extension, *Cell*, 123(4), 655-667.
- Fitzpatrick, D. A., M. E. Logue, and G. Butler (2008), Evidence of recent interkingdom horizontal gene transfer between bacteria and *Candida parapsilosis*, *BMC Evol Biol*, 8, 181.
- Fleischhacker, M., J. Pasligh, G. Moran, and M. Ruhnke (2010), Longitudinal genotyping of *Candida dubliniensis* isolates reveals strain maintenance,



- microevolution, and the emergence of itraconazole resistance, *J Clin Microbiol*, 48(5), 1643-1650.
- Frealle, E., C. Noel, E. Viscogliosi, D. Camus, E. Dei-Cas, and L. Delhaes (2005), Manganese superoxide dismutase in pathogenic fungi: an issue with pathophysiological and phylogenetic involvements, *FEMS Immunol Med Microbiol*, 45(3), 411-422.
- Frealle, E., C. Noel, N. Nolard, F. Symoens, M. S. Felipe, E. Dei-Cas, D. Camus, E. Viscogliosi, and L. Delhaes (2006), Manganese superoxide dismutase based phylogeny of pathogenic fungi, *Mol Phylogenet Evol*, 41(1), 28-39.
- Friedman, S., S. E. Richardson, S. E. Jacobs, and K. O'Brien (2000), Systemic *Candida* infection in extremely low birth weight infants: short term morbidity and long term neurodevelopmental outcome, *Pediatr Infect Dis J*, 19(6), 499-504.
- Frohner, I. E., C. Bourgeois, K. Yatsyk, O. Majer, and K. Kuchler (2009), *Candida albicans* cell surface superoxide dismutases degrade host-derived reactive oxygen species to escape innate immune surveillance, *Mol Microbiol*, 71(1), 240-252.
- Fu, Y., A. S. Ibrahim, D. C. Sheppard, Y. C. Chen, S. W. French, J. E. Cutler, S. G. Filler, and J. E. Edwards, Jr. (2002), *Candida albicans* Als1p: an adhesin that is a downstream effector of the EFG1 filamentation pathway, *Mol Microbiol*, 44(1), 61-72.
- Gale, C. A., C. M. Bendel, M. McClellan, M. Hauser, J. M. Becker, J. Berman, and M. K. Hostetter (1998), Linkage of adhesion, filamentous growth, and virulence in *Candida albicans* to a single gene, INT1, *Science*, 279(5355), 1355-1358.
- Garrison, T. R., Y. Zhang, M. Pausch, D. Apanovitch, R. Aebersold, and H. G. Dohlman (1999), Feedback phosphorylation of an RGS protein by MAP kinase in yeast, *J Biol Chem*, 274(51), 36387-36391.
- Gee, S. F., S. Joly, D. R. Soll, J. F. Meis, P. E. Verweij, I. Polacheck, D. J. Sullivan, and D. C. Coleman (2002), Identification of four distinct genotypes of *Candida dubliniensis* and detection of microevolution in vitro and in vivo, *J Clin Microbiol*, 40(2), 556-574.
- Gil, C., R. Pomes, and C. Nombela (1988), A complementation analysis by parasexual recombination of *Candida albicans* morphological mutants, *J Gen Microbiol*, 134(6), 1587-1595.
- Gilfillan, G. D., D. J. Sullivan, K. Haynes, T. Parkinson, D. C. Coleman, and N. A. Gow (1998), *Candida dubliniensis*: phylogeny and putative virulence factors, *Microbiology*, 144 ( Pt 4), 829-838.
- Gow, N. A. (1997), Germ tube growth of *Candida albicans*, *Curr Top Med Mycol*, 8(1-2), 43-55.

- Hall, R. A., K. J. Turner, J. Chaloupka, F. Cottier, L. De Sordi, D. Sanglard, L. R. Levin, J. Buck, and F. A. Muhlschlegel (2011), The quorum-sensing molecules farnesol/homoserine lactone and dodecanol operate via distinct modes of action in *Candida albicans*, *Eukaryot Cell*, 10(8), 1034-1042.
- Hannula, J., M. Saarela, B. Dogan, J. Paatsama, P. Koukila-Kahkola, S. Pirinen, H. L. Alakomi, J. Perheentupa, and S. Asikainen (2000), Comparison of virulence factors of oral *Candida dubliniensis* and *Candida albicans* isolates in healthy people and patients with chronic candidosis, *Oral Microbiol Immunol*, 15(4), 238-244.
- Henriques, M., M. Martins, J. Azeredo, and R. Oliveira (2007), Effect of farnesol on *Candida dubliniensis* morphogenesis, *Lett Appl Microbiol*, 44(2), 199-205.
- Hogan, D. A., A. Vik, and R. Kolter (2004), A *Pseudomonas aeruginosa* quorum-sensing molecule influences *Candida albicans* morphology, *Mol Microbiol*, 54(5), 1212-1223.
- Holmberg, K., and R. D. Meyer (1986), Fungal infections in patients with AIDS and AIDS-related complex, *Scand J Infect Dis*, 18(3), 179-192.
- Hornby, J. M., E. C. Jensen, A. D. Lisee, J. J. Tasto, B. Jahnke, R. Shoemaker, P. Dussault, and K. W. Nickerson (2001), Quorum sensing in the dimorphic fungus *Candida albicans* is mediated by farnesol, *Appl Environ Microbiol*, 67(7), 2982-2992.
- Hoyer, L. L., S. Scherer, A. R. Shatzman, and G. P. Livi (1995), *Candida albicans* ALS1: domains related to a *Saccharomyces cerevisiae* sexual agglutinin separated by a repeating motif, *Mol Microbiol*, 15(1), 39-54.
- Hoyer, L. L., J. Clevenger, J. E. Hecht, E. J. Ehrhart, and F. M. Poulet (1999), Detection of Als proteins on the cell wall of *Candida albicans* in murine tissues, *Infect Immun*, 67(8), 4251-4255.
- Hoyer, L. L., R. Fundyga, J. E. Hecht, J. C. Kapteyn, F. M. Klis, and J. Arnold (2001), Characterization of agglutinin-like sequence genes from non-*albicans* *Candida* and phylogenetic analysis of the ALS family, *Genetics*, 157(4), 1555-1567.
- Hube, B., M. Monod, D. A. Schofield, A. J. Brown, and N. A. Gow (1994), Expression of seven members of the gene family encoding secretory aspartyl proteinases in *Candida albicans*, *Mol Microbiol*, 14(1), 87-99.
- Hube, B., D. Sanglard, F. C. Odds, D. Hess, M. Monod, W. Schafer, A. J. Brown, and N. A. Gow (1997), Disruption of each of the secreted aspartyl proteinase genes SAP1, SAP2, and SAP3 of *Candida albicans* attenuates virulence, *Infect Immun*, 65(9), 3529-3538.

- Hwang, C. S., Y. U. Baek, H. S. Yim, and S. O. Kang (2003), Protective roles of mitochondrial manganese-containing superoxide dismutase against various stresses in *Candida albicans*, *Yeast*, 20(11), 929-941.
- Hwang, C. S., G. E. Rhie, J. H. Oh, W. K. Huh, H. S. Yim, and S. O. Kang (2002), Copper- and zinc-containing superoxide dismutase (Cu/ZnSOD) is required for the protection of *Candida albicans* against oxidative stresses and the expression of its full virulence, *Microbiology*, 148(Pt 11), 3705-3713.
- Hwang, C. S., G. Rhie, S. T. Kim, Y. R. Kim, W. K. Huh, Y. U. Baek, and S. O. Kang (1999), Copper- and zinc-containing superoxide dismutase and its gene from *Candida albicans*, *Biochim Biophys Acta*, 1427(2), 245-255.
- Ibrahim, A. S., F. Mirbod, S. G. Filler, Y. Banno, G. T. Cole, Y. Kitajima, J. E. Edwards, Jr., Y. Nozawa, and M. A. Ghannoum (1995), Evidence implicating phospholipase as a virulence factor of *Candida albicans*, *Infect Immun*, 63(5), 1993-1998.
- Jackson, A. P., et al. (2009), Comparative genomics of the fungal pathogens *Candida dubliniensis* and *Candida albicans*, *Genome Res*, 19(12), 2231-2244.
- Jamieson, D. J., S. L. Rivers, and D. W. Stephen (1994), Analysis of *Saccharomyces cerevisiae* proteins induced by peroxide and superoxide stress, *Microbiology*, 140 ( Pt 12), 3277-3283.
- Johnson E.M. (2009) Race and Emerging *Candida* Species. In *Current Fungal Infection Reports*. 3, 152-159.
- Jones, S., G. White, and P. R. Hunter (1994), Increased phenotypic switching in strains of *Candida albicans* associated with invasive infections, *J Clin Microbiol*, 32(11), 2869-2870.
- Kang, Y. S., Y. Lee, H. Jung, C. O. Jeon, E. L. Madsen, and W. Park (2007), Overexpressing antioxidant enzymes enhances naphthalene biodegradation in *Pseudomonas* sp. strain As1, *Microbiology*, 153(Pt 10), 3246-3254.
- Kao, A. S., et al. (1999), The epidemiology of candidemia in two United States cities: results of a population-based active surveillance, *Clin Infect Dis*, 29(5), 1164-1170.
- Kauffman, C. A., J. F. Fisher, J. D. Sobel, and C. A. Newman (2011), *Candida* urinary tract infections--diagnosis, *Clin Infect Dis*, 52 Suppl 6, S452-456.
- Kim, K. Y., S. Y. Lee, Y. S. Cho, I. C. Bang, K. H. Kim, D. S. Kim, and Y. K. Nam (2007), Molecular characterization and mRNA expression during metal exposure and thermal stress of copper/zinc- and manganese-superoxide dismutases in disk abalone, *Haliotis discus discus*, *Fish Shellfish Immunol*, 23(5), 1043-1059.
- Kimmel, J. R., H. Markowitz, and D. M. Brown (1959), Some chemical and physical properties of erythrocyte, *J Biol Chem*, 234(1), 46-50.



- Klotz, S. A., D. J. Drutz, J. L. Harrison, and M. Huppert (1983), Adherence and penetration of vascular endothelium by *Candida* yeasts, *Infect Immun*, 42(1), 374-384.
- Koga-Ito, C. Y., E. Y. Komiyama, C. A. De Paiva Martins, T. C. Vasconcellos, A. O. Cardoso Jorge, Y. R. Carvalho, R. F. Do Prado, and I. Balducci (2010), Experimental systemic virulence of oral *Candida dubliniensis* isolates in comparison with *Candida albicans*, *Candida tropicalis* and *Candida krusei*, *Mycoses*.
- Kothavade, R. J., and M. H. Panthaki (1998), Evaluation of phospholipase activity of *Candida albicans* and its correlation with pathogenicity in mice, *J Med Microbiol*, 47(2), 99-102.
- Krcmery, V., Jr. (1996), Emerging fungal infections in cancer patients, *J Hosp Infect*, 33(2), 109-117.
- LaFayette, S. L., C. Collins, A. K. Zaas, W. A. Schell, M. Betancourt-Quiroz, A. A. Gunatilaka, J. R. Perfect, and L. E. Cowen (2010), PKC signaling regulates drug resistance of the fungal pathogen *Candida albicans* via circuitry comprised of Mkc1, calcineurin, and Hsp90, *PLoS Pathog*, 6(8).
- Lamarre, C., J. D. LeMay, N. Deslauriers, and Y. Bourbonnais (2001), *Candida albicans* expresses an unusual cytoplasmic manganese-containing superoxide dismutase (SOD3 gene product) upon the entry and during the stationary phase, *J Biol Chem*, 276(47), 43784-43791.
- Lan, C. Y., G. Newport, L. A. Murillo, T. Jones, S. Scherer, R. W. Davis, and N. Agabian (2002), Metabolic specialization associated with phenotypic switching in *Candida albicans*, *Proc Natl Acad Sci U S A*, 99(23), 14907-14912.
- Lanchares, J. L., and M. L. Hernandez (2000), Recurrent vaginal candidiasis changes in etiopathogenical patterns, *Int J Gynaecol Obstet*, 71 Suppl 1, S29-35.
- Lane, S., C. Birse, S. Zhou, R. Matson, and H. Liu (2001), DNA array studies demonstrate convergent regulation of virulence factors by Cph1, Cph2, and Efg1 in *Candida albicans*, *J Biol Chem*, 276(52), 48988-48996.
- Leberer, E., D. H Marcus, D. Dignard, L. Johnson, S. Ushinsky, D. Y. Thomas, and K. Schroppel (2001), Ras links cellular morphogenesis to virulence by regulation of the MAP kinase and cAMP signalling pathways in the pathogenic fungus *Candida albicans*, *Mol Microbiol*, 42(3), 673-687.
- Lemos-Carolino, M., A. Madeira-Lopes, and N. Van Uden (1982), The temperature profile of the pathogenic yeast *Candida albicans*, *Z Allg Mikrobiol*, 22(10), 705-709.
- Lescuyer, P., S. Picot, V. Bracchi, J. Burnod, J. Austin, A. Perard, and P. Ambroise-Thomas (1997), Detection of RAPD markers correlated with chloroquine resistance in *Plasmodium falciparum*, *Genome Res*, 7(7), 747-753.

- Li, Y., W. Xu, M. W. McBurney, and V. D. Longo (2008), SirT1 inhibition reduces IGF-I/IRS-2/Ras/ERK1/2 signaling and protects neurons, *Cell Metab*, 8(1), 38-48.
- Li, Y., et al. (1995), Dilated cardiomyopathy and neonatal lethality in mutant mice lacking manganese superoxide dismutase, *Nat Genet*, 11(4), 376-381.
- Lian, C. H., and W. D. Liu (2007), Differential expression of *Candida albicans* secreted aspartyl proteinase in human vulvovaginal candidiasis, *Mycoses*, 50(5), 383-390.
- Lim, C. S., W. F. Wong, R. Rosli, K. P. Ng, H. F. Seow, and P. P. Chong (2009), 2-dodecanol (decyl methyl carbinol) inhibits hyphal formation and SIR2 expression in *C. albicans*, *J Basic Microbiol*, 49(6), 579-583.
- Lin, J. C., K. Duell, and J. B. Konopka (2004), A microdomain formed by the extracellular ends of the transmembrane domains promotes activation of the G protein-coupled alpha-factor receptor, *Mol Cell Biol*, 24(5), 2041-2051.
- Lingappa, B. T., M. Prasad, Y. Lingappa, D. F. Hunt, and K. Biemann (1969), Phenethyl alcohol and tryptophol: autoantibiotics produced by the fungus *Candida albicans*, *Science*, 163(863), 192-194.
- Liu, T. T., R. E. Lee, K. S. Barker, L. Wei, R. Homayouni, and P. D. Rogers (2005), Genome-wide expression profiling of the response to azole, polyene, echinocandin, and pyrimidine antifungal agents in *Candida albicans*, *Antimicrob Agents Chemother*, 49(6), 2226-2236.
- Lo, H. J., J. R. Kohler, B. DiDomenico, D. Loebenberg, A. Cacciapuoti, and G. R. Fink (1997), Nonfilamentous *C. albicans* mutants are avirulent, *Cell*, 90(5), 939-949.
- Loaiza-Loeza, S., B. Parra-Ortega, J. C. Cancino-Diaz, B. Illades-Aguilar, C. H. Hernandez-Rodriguez, and L. Villa-Tanaca (2009), Differential expression of *Candida dubliniensis*-secreted aspartyl proteinase genes (CdSAP1-4) under different physiological conditions and during infection of a keratinocyte culture, *FEMS Immunol Med Microbiol*, 56(3), 212-222.
- Lorenz, M. C., and J. Heitman (1997), Yeast pseudohyphal growth is regulated by GPA2, a G protein alpha homolog, *EMBO J*, 16(23), 7008-7018.
- Low, C. F., P. P. Chong, P. V. Yong, C. S. Lim, Z. Ahmad, and F. Othman (2008), Inhibition of hyphae formation and SIR2 expression in *Candida albicans* treated with fresh *Allium sativum* (garlic) extract, *J Appl Microbiol*, 105(6), 2169-2177.
- Magee, B. B., M. D. Sanchez, D. Saunders, D. Harris, M. Berriman, and P. T. Magee (2008), Extensive chromosome rearrangements distinguish the karyotype of the hypovirulent species *Candida dubliniensis* from the virulent *Candida albicans*, *Fungal Genet Biol*, 45(3), 338-350.

- Markowitz, H., G. E. Cartwright, and M. M. Wintrobe (1959), Studies on copper metabolism. XXVII. The isolation and properties of an erythrocyte cuproprotein (erythrocyte cuproprotein), *J Biol Chem*, 234(1), 40-45.
- Martchenko, M., A. M. Alarco, D. H Marcus, and M. Whiteway (2004), Superoxide dismutases in *Candida albicans*: transcriptional regulation and functional characterization of the hyphal-induced SOD5 gene, *Mol Biol Cell*, 15(2), 456-467.
- Maxwell, M. J., S. A. Messer, R. J. Hollis, L. Boyken, S. Tendolkar, D. J. Diekema, and M. A. Pfaller (2003), Evaluation of Etest method for determining fluconazole and voriconazole MICs for 279 clinical isolates of *Candida* species infrequently isolated from blood, *J Clin Microbiol*, 41(3), 1087-1090.
- McCord, J. M., and I. Fridovich (1969), Superoxide dismutase. An enzymic function for erythrocyte cuproprotein (hemocuproprotein), *J Biol Chem*, 244(22), 6049-6055.
- McCullough, M. J., K. V. Clemons, and D. A. Stevens (1999), Molecular and phenotypic characterization of genotypic *Candida albicans* subgroups and comparison with *Candida dubliniensis* and *Candida stellatoidea*, *J Clin Microbiol*, 37(2), 417-421.
- Meyers, J. D. (1990), Fungal infections in bone marrow transplant patients, *Semin Oncol*, 17(3 Suppl 6), 10-13.
- Miller, M. B., and B. L. Bassler (2001), Quorum sensing in bacteria, *Annu Rev Microbiol*, 55, 165-199.
- Moran, G. P., D. J. Sullivan, M. C. Henman, C. E. McCreary, B. J. Harrington, D. B. Shanley, and D. C. Coleman (1997), Antifungal drug susceptibilities of oral *Candida dubliniensis* isolates from human immunodeficiency virus (HIV)-infected and non-HIV-infected subjects and generation of stable fluconazole-resistant derivatives in vitro, *Antimicrob Agents Chemother*, 41(3), 617-623.
- Mulley, A. G. (2006). *Primary Care Medicine: office evaluation and management of the adult patient*, A. H. Goroll. pp. 802-3. Philadelphia: Wolters Kluwer Health.
- Naglik, J. R., F. Fostira, J. Ruprai, J. F. Staab, S. J. Challacombe, and P. Sundstrom (2006), *Candida albicans* HWP1 gene expression and host antibody responses in colonization and disease, *J Med Microbiol*, 55(Pt 10), 1323-1327.
- Nantel, A., et al. (2002), Transcription profiling of *Candida albicans* cells undergoing the yeast-to-hyphal transition, *Mol Biol Cell*, 13(10), 3452-3465.
- Nas, T., A. Kalkanci, I. Fidan, K. Hizel, S. Bolat, S. Yolbakan, E. Yilmaz, S. Ozkan, and S. Kustimur (2008), Expression of ALS1, HWP1 and SAP4 genes in *Candida albicans* strains isolated from women with vaginitis, *Folia Microbiol (Praha)*, 53(2), 179-183.
- Nguyen, M. H., J. E. Peacock, Jr., A. J. Morris, D. C. Tanner, M. L. Nguyen, D. R. Snyderman, M. M. Wagener, M. G. Rinaldi, and V. L. Yu (1996), The changing face of

- candidemia: emergence of non-*Candida albicans* species and antifungal resistance, *Am J Med*, 100(6), 617-623.
- Ollert, M. W., and R. A. Calderone (1990), A monoclonal antibody that defines a surface antigen on *Candida albicans* hyphae cross-reacts with yeast cell protoplasts, *Infect Immun*, 58(3), 625-631.
- Pacheco-Rios, A., C. Avila-Figueroa, D. Nobigrot-Kleinman, and J. I. Santos (1997), Mortality associated with systemic candidiasis in children, *Arch Med Res*, 28(2), 229-232.
- Paganelli, M., P. Romandini, G. Bertoloni, M. Beltramini, L. Tallandini, and B. Salvato (1989), Induction of superoxide dismutase by methanol and structural modifications in *Candida albicans*, *Yeast*, 5 Spec No, S431-435.
- Pappas, P. G. (2006), Invasive candidiasis, *Infect Dis Clin North Am*, 20(3), 485-506.
- Peltroche-Llacsahuanga, H., S. Schmidt, M. Seibold, R. Lutticken, and G. Haase (2000a), Differentiation between *Candida dubliniensis* and *Candida albicans* by fatty acid methyl ester analysis using gas-liquid chromatography, *J Clin Microbiol*, 38(10), 3696-3704.
- Peltroche-Llacsahuanga, H., N. Schnitzler, S. Schmidt, K. Tintelnot, R. Lutticken, and G. Haase (2000b), Phagocytosis, oxidative burst, and killing of *Candida dubliniensis* and *Candida albicans* by human neutrophils, *FEMS Microbiol Lett*, 191(1), 151-155.
- Perez-Martin, J., J. A. Uria, and A. D. Johnson (1999), Phenotypic switching in *Candida albicans* is controlled by a SIR2 gene, *EMBO J*, 18(9), 2580-2592.
- Pfaffl, M. W. (2001), A new mathematical model for relative quantification in real-time RT-PCR, *Nucleic Acids Res*, 29(9), e45.
- Pfaller, M. A., and D. J. Diekema (2007), Epidemiology of invasive candidiasis: a persistent public health problem, *Clin Microbiol Rev*, 20(1), 133-163.
- Pfaller, M. A., R. N. Jones, S. A. Messer, M. B. Edmond, and R. P. Wenzel (1998), National surveillance of nosocomial blood stream infection due to species of *Candida* other than *Candida albicans*: frequency of occurrence and antifungal susceptibility in the SCOPE Program. SCOPE Participant Group. Surveillance and Control of Pathogens of Epidemiologic, *Diagn Microbiol Infect Dis*, 30(2), 121-129.
- Pfaller, M. A., S. A. Messer, L. Boyken, S. Tendolkar, R. J. Hollis, and D. J. Diekema (2004), Geographic variation in the susceptibilities of invasive isolates of *Candida glabrata* to seven systemically active antifungal agents: a global assessment from the ARTEMIS Antifungal Surveillance Program conducted in 2001 and 2002, *J Clin Microbiol*, 42(7), 3142-3146.
- Pfaller, M. A., D. J. Diekema, R. N. Jones, H. S. Sader, A. C. Fluit, R. J. Hollis, and S. A. Messer (2001), International surveillance of bloodstream infections due to *Candida*



- species: frequency of occurrence and in vitro susceptibilities to fluconazole, ravuconazole, and voriconazole of isolates collected from 1997 through 1999 in the SENTRY antimicrobial surveillance program, *J Clin Microbiol*, 39(9), 3254-3259.
- Pinjon, E., D. Sullivan, I. Salkin, D. Shanley, and D. Coleman (1998), Simple, inexpensive, reliable method for differentiation of *Candida dubliniensis* from *Candida albicans*, *J Clin Microbiol*, 36(7), 2093-2095.
- Pinjon, E., C. J. Jackson, S. L. Kelly, D. Sanglard, G. Moran, D. C. Coleman, and D. J. Sullivan (2005), Reduced azole susceptibility in genotype 3 *Candida dubliniensis* isolates associated with increased CdCDR1 and CdCDR2 expression, *Antimicrob Agents Chemother*, 49(4), 1312-1318.
- Pinto, P. M., M. A. Resende, C. Y. Koga-Ito, and M. Tendler (2004), Genetic variability analysis among clinical *Candida* spp. isolates using random amplified polymorphic DNA, *Mem Inst Oswaldo Cruz*, 99(2), 147-152.
- Poulter, R., K. Jeffery, M. J. Hubbard, M. G. Shepherd, and P. A. Sullivan (1981), Parasexual genetic analysis of *Candida albicans* by spheroplast fusion, *J Bacteriol*, 146(3), 833-840.
- Ramage, G., and J. L. Lopez-Ribot (2005), Techniques for antifungal susceptibility testing of *Candida albicans* biofilms, *Methods Mol Med*, 118, 71-79.
- Ramage, G., E. Mowat, B. L. Jones, C. Williams, and J. L. Lopez-Ribot (2009), Our current understanding of fungal biofilms. *C Rev in Microbiol*, 35: 340-355.
- Ramage, G., K. Vande Walle, B. L. Wickes, and J. L. Lopez-Ribot (2001), Biofilm formation by *Candida dubliniensis*, *J Clin Microbiol*, 39(9), 3234-3240.
- Ramage, G., S. P. Saville, B. L. Wickes, and J. L. Lopez-Ribot (2002), Inhibition of *Candida albicans* biofilm formation by farnesol, a quorum-sensing molecule, *Appl Environ Microbiol*, 68(11), 5459-5463.
- Redwood, A. J. 1997. *Cytokine gene expression patterns and immune responses to systemic Candida albicans infection in inbred mice*. PhD Thesis, Curtin University of Technology, Australia.
- Rhie, G. E., C. S. Hwang, M. J. Brady, S. T. Kim, Y. R. Kim, W. K. Huh, Y. U. Baek, B. H. Lee, J. S. Lee, and S. O. Kang (1999), Manganese-containing superoxide dismutase and its gene from *Candida albicans*, *Biochim Biophys Acta*, 1426(3), 409-419.
- Rippon, J. W. 1974. Candidosis. In *The Pathogenic Fungi and The Pathogenic Actinomycetes*. pp. 175-204. Philadelphia: W. B. Saunders Company.
- Romandini, P., C. Bonotto, G. Bertoloni, M. Beltramini, and B. Salvato (1994), Superoxide dismutase, catalase and cell dimorphism in *Candida albicans* cells exposed to methanol and different temperatures, *Comp Biochem Physiol Pharmacol Toxicol Endocrinol*, 108(1), 53-57.

- Romeo, O., and G. Criseo (2009), Morphological, biochemical and molecular characterisation of the first Italian *Candida africana* isolate, *Mycoses*, 52(5), 454-457.
- Ruhnke, M., A. Schmidt-Westhausen, and J. Morschhauser (2000), Development of simultaneous resistance to fluconazole in *Candida albicans* and *Candida dubliniensis* in a patient with AIDS, *J Antimicrob Chemother*, 46(2), 291-295.
- Sadhu, C., M. J. McEachern, E. P. Rustchenko-Bulgac, J. Schmid, D. R. Soll, and J. B. Hicks (1991), Telomeric and dispersed repeat sequences in *Candida* yeasts and their use in strain identification, *J Bacteriol*, 173(2), 842-850.
- Salomé, L. L., P. O. Berenice, B. M. Consuelo, C. R. César, H. R. C. Hugo and V. T. Lourdes (2007), The Proteolytic System of *Candida Dubliniensis*, *American Journal of Infectious Diseases*, 3(2): 76-83.
- Sanglard, D., B. Hube, M. Monod, F. C. Odds, and N. A. Gow (1997), A triple deletion of the secreted aspartyl proteinase genes SAP4, SAP5, and SAP6 of *Candida albicans* causes attenuated virulence, *Infect Immun*, 65(9), 3539-3546.
- Sato, T., T. Watanabe, T. Mikami, and T. Matsumoto (2004), Farnesol, a morphogenetic autoregulatory substance in the dimorphic fungus *Candida albicans*, inhibits hyphae growth through suppression of a mitogen-activated protein kinase cascade, *Biol Pharm Bull*, 27(5), 751-752.
- Schroppel, K., M. Rotman, R. Galask, K. Mac, and D. R. Soll (1994), Evolution and replacement of *Candida albicans* strains during recurrent vaginitis demonstrated by DNA fingerprinting, *J Clin Microbiol*, 32(11), 2646-2654.
- Schweizer, A., S. Rupp, B. N. Taylor, M. Rollinghoff, and K. Schroppel (2000), The TEA/ATTS transcription factor CaTec1p regulates hyphal development and virulence in *Candida albicans*, *Mol Microbiol*, 38(3), 435-445.
- Scott, M. D., S. R. Meshnick, and J. W. Eaton (1987), Superoxide dismutase-rich bacteria. Paradoxical increase in oxidant toxicity, *J Biol Chem*, 262(8), 3640-3645.
- Shapiro, R. S., and L. Cowen (2010), Coupling temperature sensing and development: Hsp90 regulates morphogenetic signalling in *Candida albicans*, *Virulence*, 1(1), 45-48.
- Shapiro, R. S., P. Uppuluri, A. K. Zaas, C. Collins, H. Senn, J. R. Perfect, J. Heitman, and L. E. Cowen (2009), Hsp90 orchestrates temperature-dependent *Candida albicans* morphogenesis via Ras1-PKA signaling, *Curr Biol*, 19(8), 621-629.
- Sharkey, L. L., M. D. McNemar, S. M. Saporito-Irwin, P. S. Sypherd, and W. A. Fonzi (1999), HWP1 functions in the morphological development of *Candida albicans* downstream of EFG1, TUP1, and RBF1, *J Bacteriol*, 181(17), 5273-5279.

- Shchepin, R., J. M. Hornby, E. Burger, T. Niessen, P. Dussault, and K. W. Nickerson (2003), Quorum sensing in *Candida albicans*: probing farnesol's mode of action with 40 natural and synthetic farnesol analogs, *Chem Biol*, 10(8), 743-750.
- Shchepin, R., D. H. Navarathna, R. Dumitru, S. Lippold, K. W. Nickerson, and P. H. Dussault (2008), Influence of heterocyclic and oxime-containing farnesol analogs on quorum sensing and pathogenicity in *Candida albicans*, *Bioorg Med Chem*, 16(4), 1842-1848.
- Shen, J., L. E. Cowen, A. M. Griffin, L. Chan, and J. R. Kohler (2008), The *Candida albicans* pescadillo homolog is required for normal hypha-to-yeast morphogenesis and yeast proliferation, *Proc Natl Acad Sci U S A*, 105(52), 20918-20923.
- Silva, G. M., F. R. Silveira, and F. Pires Mde (2007), Adherence to HeLa cells, typing by killer toxins and susceptibility to antifungal agents of *Candida dubliniensis* strains, *Braz Oral Res*, 21(1), 87-91.
- Simona, E. S., P. Diana, I. Robertina, A. Ionela, S. Ileana, and V. D. Tatiana (2009), Molecular identification of some yeast strains involved in oral candidosis, *Romanian Biotechnological Letters*, 14(1), 4180-4186.
- Slutsky, B., M. Staebell, J. Anderson, L. Risen, M. Pfaller, and D. R. Soll (1987), "White-opaque transition": a second high-frequency switching system in *Candida albicans*, *J Bacteriol*, 169(1), 189-197.
- Smit, M. S., M. M. Mokgoro, E. Setati, and J. M. Nicaud (2004), Preparation of dodecanol-tolerant strains of *Yarrowia lipolytica*, *Biotechnol Lett*, 26(10), 849-854.
- Smith, J. M. B. 1989. Candidosis. In *Opportunistic Mycoses of Man and Other Animal*. C. A. B. Wallingford. pp. 11-35. United Kingdom. International Mycology Institute.
- Soll, D. R., C. J. Langtimm, J. McDowell, J. Hicks, and R. Galask (1987), High-frequency switching in *Candida* strains isolated from vaginitis patients, *J Clin Microbiol*, 25(9), 1611-1622.
- Soll, D. R., R. Galask, J. Schmid, C. Hanna, K. Mac, and B. Morrow (1991), Genetic dissimilarity of commensal strains of *Candida* spp. carried in different anatomical locations of the same healthy women, *J Clin Microbiol*, 29(8), 1702-1710.
- Sonneborn, A., B. Tebarth, and J. F. Ernst (1999), Control of white-opaque phenotypic switching in *Candida albicans* by the Efg1p morphogenetic regulator, *Infect Immun*, 67(9), 4655-4660.
- Staab, J. F., C. A. Ferrer, and P. Sundstrom (1996), Developmental expression of a tandemly repeated, proline-and glutamine-rich amino acid motif on hyphal surfaces on *Candida albicans*, *J Biol Chem*, 271(11), 6298-6305.

- Staab, J. F., S. D. Bradway, P. L. Fidel, and P. Sundstrom (1999), Adhesive and mammalian transglutaminase substrate properties of *Candida albicans* Hwp1, *Science*, 283(5407), 1535-1538.
- Steffan, P., J. A. Vazquez, D. Boikov, C. Xu, J. D. Sobel, and R. A. Akins (1997), Identification of *Candida* species by randomly amplified polymorphic DNA fingerprinting of colony lysates, *J Clin Microbiol*, 35(8), 2031-2039.
- Stokes, C., G. P. Moran, M. J. Spiering, G. T. Cole, D. C. Coleman, and D. J. Sullivan (2007), Lower filamentation rates of *Candida dubliniensis* contribute to its lower virulence in comparison with *Candida albicans*, *Fungal Genet Biol*, 44(9), 920-931.
- Stoldt, V. R., A. Sonneborn, C. E. Leuker, and J. F. Ernst (1997), Efg1p, an essential regulator of morphogenesis of the human pathogen *Candida albicans*, is a member of a conserved class of bHLH proteins regulating morphogenetic processes in fungi, *EMBO J*, 16(8), 1982-1991.
- Sudbery, P., N. Gow, and J. Berman (2004), The distinct morphogenic states of *Candida albicans*, *Trends Microbiol*, 12(7), 317-324.
- Sullivan, D., and D. Coleman (1998), *Candida dubliniensis*: characteristics and identification, *J Clin Microbiol*, 36(2), 329-334.
- Sullivan, D., D. Bennett, M. Henman, P. Harwood, S. Flint, F. Mulcahy, D. Shanley, and D. Coleman (1993), Oligonucleotide fingerprinting of isolates of *Candida* species other than *C. albicans* and of atypical *Candida* species from human immunodeficiency virus-positive and AIDS patients, *J Clin Microbiol*, 31(8), 2124-2133.
- Sullivan, D., K. Haynes, J. Bille, P. Boerlin, L. Rodero, S. Lloyd, M. Henman, and D. Coleman (1997), Widespread geographic distribution of oral *Candida dubliniensis* strains in human immunodeficiency virus-infected individuals, *J Clin Microbiol*, 35(4), 960-964.
- Sullivan, D. J., T. J. Westerneng, K. A. Haynes, D. E. Bennett, and D. C. Coleman (1995), *Candida dubliniensis* sp. nov.: phenotypic and molecular characterization of a novel species associated with oral candidosis in HIV-infected individuals, *Microbiology*, 141 ( Pt 7), 1507-1521.
- Swoboda, R. K., G. Bertram, S. Budge, G. W. Gooday, N. A. Gow, and A. J. Brown (1995), Structure and regulation of the HSP90 gene from the pathogenic fungus *Candida albicans*, *Infect Immun*, 63(11), 4506-4514.
- Tay, S. T., H. C. Chai, S. L. Na, and K. P. Ng (2005), Molecular subtyping of clinical isolates of *Candida albicans* and identification of *Candida dubliniensis* Malaysia, *Mycopathologia*, 159(3), 325-329.



- van Loon, A. P., B. Pesold-Hurt, and G. Schatz (1986), A yeast mutant lacking mitochondrial manganese-superoxide dismutase is hypersensitive to oxygen, *Proc Natl Acad Sci U S A*, 83(11), 3820-3824.
- Vanden Bossche, H. (1997), Mechanisms of antifungal resistance, *Rev Iberoam Micol*, 14(2), 44-49.
- Vilela, M. M., K. Kamei, A. Sano, R. Tanaka, J. Uno, I. Takahashi, J. Ito, K. Yarita, and M. Miyaji (2002), Pathogenicity and virulence of *Candida dubliniensis*: comparison with *C. albicans*, *Med Mycol*, 40(3), 249-257.
- Walsh T. J. and D. M. Dixon, 1996. "Deep Mycoses". In Baron S *et al.* eds. (via NCBI Bookshelf). *Baron's Medical Microbiology* (4th ed.). Univ of Texas Medical Branch. ISBN 0-9631172-1-1.  
<http://www.ncbi.nlm.nih.gov/books/bv.fcgi?rid=mmed.section.4006>.
- Waters, C. M., and B. L. Bassler (2005), Quorum sensing: cell-to-cell communication in bacteria, *Annu Rev Cell Dev Biol*, 21, 319-346.
- Westwater, C., E. Balish, and D. A. Schofield (2005), *Candida albicans*-conditioned medium protects yeast cells from oxidative stress: a possible link between quorum sensing and oxidative stress resistance, *Eukaryot Cell*, 4(10), 1654-1661.
- Zheng, X. D., R. T. Lee, Y. M. Wang, Q. S. Lin, and Y. Wang (2007), Phosphorylation of Rga2, a Cdc42 GAP, by CDK/Hgc1 is crucial for *Candida albicans* hyphal growth, *EMBO J*, 26(16), 3760-3769.