



UNIVERSITI PUTRA MALAYSIA

***IDENTIFICATION OF POTENTIAL PLASMA PROTEIN
MARKERS FOR GESTATIONAL DIABETES MELLITUS***

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**IDENTIFICATION OF POTENTIAL PROTEIN MARKERS FOR
GESTATIONAL DIABETES MELLITUS**

By

PUSHPA GANDI SANGARAN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Master of Science**

September 2012



This thesis is dedicated to my spiritual master Bhoghar for his unwavering support, understanding, strength, love, constant encouragement and for inspiring me with a belief in my own abilities.

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

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PUSHPA GANDI SANGARAN

September 2012

Chairman: Professor Rozita Rosli, PhD

Faculty : Medicine and Health Sciences

Gestational Diabetes Mellitus (GDM) is defined as glucose intolerance that is detected during pregnancy. It is caused by the decreased sensitivity of insulin with increasing gestation week. Identifying this group of women is important in not only preventing perinatal morbidity but also improving long term outcomes for the mothers and their children. The usual approach in diagnosing GDM is to screen GDM patients during the second trimester of gestation through an oral glucose tolerance test (OGTT). There is potential harm and high costs may be involved in screening, especially given the high false-positive rate. However, there is a need to diagnose GDM as early as possible to minimize the undesirable effects in the development of the fetus.

In this exploratory study, two dimensional gel electrophoresis (2DGE) and Liquid Chromatography Mass Spectrometry (LC/MS/MS) were used in the quest for new potential biomarkers for the disease. The purpose of this study was to identify a set of potential protein markers which are differentially expressed during the first, second or third trimester of pregnancy. Once identified and validated, the candidate biomarkers for GDM could provide

opportunities for rapid and low-cost screening. For this study, five millilitres of fresh blood was drawn for each trimester of pregnancy from five normal pregnant women and five GDM patients at Hospital Kuala Lumpur and Klinik Kesihatan Seri Kembangan. The plasma samples were used for 2DGE, where proteins are separated first by characteristic isoelectric point using isoelectrofocusing, and then by protein molecular weight, using sodium dodecylsulfate-polyacrylamide gel electrophoresis (SDS-PAGE). The 2D gels were stained using Colloidal Coomassie Blue stain and scanned by using a high resolution imager. The images were quantitatively analyzed using Nonlinear Progenesis SameSpots Software to identify protein spots that demonstrate significant differences in expression between the GDM samples and with normal pregnant samples. Spots demonstrating significant differences were manually picked using sterile procedure and identified by LC/MS/MS analysis through a service from the Proteomics Centre at Medical University of South Carolina.

In this study, 11 distinct proteins were identified which were expressed differentially in the plasma of normal pregnant women and plasma from GDM patients. The proteins were Alpha-1-Antitrypsin, Albumin, Antithrombin III, Fibrinogen, Apolipoprotein A1, Apolipoprotein E, Alpha 1B-Glycoprotein, Complement Factors, Angiotensinogen, Transferrin, and Vitamin D binding protein. Among these proteins, Antithrombin III, Apolipoprotein E and Vitamin D binding protein could be candidates to screen the GDM patients. Further studies are recommended to validate these candidate biomarkers by using ELISA and western blotting.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Sarjana Sains

**PENGENALPASTIAN PENANDA PROTEIN BERPOTENSI BAGI DIABETES
MELLITUS SEMASA KEHAMILAN**

Oleh

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Diabetes mellitus semasa kehamilan (GDM) ialah ketaktoleranan glukosa yang dikesan semasa kehamilan. Ini disebabkan oleh kemerosotan kesensitifan insulin yang berlaku sepanjang tempoh kehamilan. Kepentingan mengenal pasti kumpulan wanita yang terbabit dengan keadaan ini adalah penting bukan sahaja bagi menghindari mereka daripada kematian semasa kelahiran malahan juga bagi memperbaiki kesan jangka panjang terhadap para ibu dan anak mereka. Pendekatan bagi mendiagnosis GDM biasanya ialah dengan menyaring penghidap GDM semasa trimester kedua kehamilan melalui ujian ketoleranan glukosa secara oral (OGTT). Penyaringan mungkin menyebabkan kesan yang buruk dan kos yang tinggi, terutamanya kerana kaedah ini melibatkan kadar positif palsu yang tinggi. Walau bagaimanapun, GDM perlu didiagnosis seawal yang mungkin bagi meminimumkan kesan kurang baik dalam perkembangan fetus.

Dalam kajian tinjauan ini, elektroforesis gel dua dimensi (2DGE) dan spektrometri massa kromatografi cecair (LC/MS/MS) telah digunakan bagi menentukan beberapa penanda bio

berpotensi penyakit ini. Tujuan kajian ini adalah bagi mengenal pasti satu set penanda protein berpotensi yang diekspresi secara berbeza-beza semasa trimester pertama, kedua dan ketiga kehamilan. Calon penanda bio GDM yang telah dikenal pasti dan disahkan boleh digunakan dalam penyaringan yang pantas dan berkos rendah. Bagi kajian ini, lima milliliter darah diambil daripada lima wanita hamil yang normal dan lima penghidap GDM bagi setiap trimester kehamilan di Kuala Lumpur dan Klinik Kesihatan Seri Kembangan. Sampel plasma digunakan bagi 2DGE, di mana protein diasingkan melalui ciri-ciri titik isoelektrik menggunakan pemfokusan isoelektrik, dan kemudian secara berat molekul protein menggunakan elektroforesis gel dodesilsulfat poliakrilamida (SDS-PAGE). Gel 2D tersebut telah diwarnakan dengan pewarna biru Coomassie berkolid dan diimbas dengan menggunakan pengimbas beresolusi tinggi. Imej dianalisis secara kuantitatif menggunakan perisian Nonlinear Progenesis SameSpots bagi mengenal pasti bintik protein yang menunjukkan perbezaan signifikan ekspresi antara sampel GDM dengan sampel kehamilan normal. Bintik yang menunjukkan perbezaan signifikan dipilih dengan tangan secara steril dan dikenal pasti melalui analisis LC/MS/MS oleh perkhidmatan Proteomics Centre di Medical University of South Carolina.

Sebelas protein berlainan telah dikenal pasti di mana semuanya terekspresi secara berbeza-beza di dalam plasma wanita hamil yang normal dan plasma wanita dengan GDM. Protein-protein itu ialah Alpha-1-Antitrypsin, Albumin, Antithrombin III, Fibrinogen, Apolipoprotein A1, Apolipoprotein E, Alpha 1B-Glycoprotein, Faktor Pelengkap, Angiotensinogen, Transferrin, dan protein penambat Vitamin D. Antara protein itu, Antithrombin III, Apolipoprotein E dan protein penambat Vitamin D berpotensi menjadi calon bagi menyaring penghidap GDM. Kajian lanjutan disyorkan bagi mengesahkan calon penanda bio dengan menggunakan ELISA dan mendapan Western.

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I certify that an Examination Committee has met on-----to conduct the final examination of Pushpa Gandhi Sangaran on her Master of Science thesis entitled 'Identification of potential protein markers for Gestational Diabetes Mellitus' in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Pertanian Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the degree of Master of Science.

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DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously and is not concurrently submitted for any other degree at Universiti Putra Malaysia or at any other institutions.

PUSHPA GANDI SANGARAN

Date : 27 September 2012



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LIST OF ABBREVIATIONS

1,25(OH)-D	Calcitriol
2D-DIGE	2-Dimensional-Fluorescence Difference Gel Electrophoresis
2D-PAGE	2-Dimensional-Polyacrlamide Gel Electrophoresis
³² P	Phosphorus
3T3-L1	Mouse embryonic fibroblast-adipose like cell line) Nuclear Lysate
A1BG	Alpha 1B-Glycoprotein
AAT	Alpha-1-Antitrypsin
ALB	Albumin
ACOG	American College of Obstetricians and Gynaecologists
ADA	American Diabetes Association
Apo A1	Apolipoprotein A1
Apo E	Apolipoprotein E
AT3	Antithrombin III
BLAST	Basic Local Alignment Search Tool
BMI	Body Mass Index
BSA	Bovine Serum Albumin
CCB	Coomassie Brilliant Blue

CHAPS	3-[(3cholamidopropyl) dimethylammonio]-1-propanesulfonate
Cm	centimetre
DTT	Dithiothreitol
ELISA	Enzyme Linked Immunosorbent Assay
ESI-MS	Electrospray Ionization Mass Spectrometry
G	gravity
G	Gram
GCK	Glucokinase
GDM	Gestational Diabetes Mellitus
HCl	Hydrochloric acid
hCS	human Chorionic Sommatomammotropin
HDL	High Density Lipoprotein
HNF4a	Hepatocyte Nuclear Factor-4a
hPL	human Placental Lactogen
IDL	Intermediate Density Lipoprotein
IEF	Isoelectric Focusing
IGF2	Insulin-Like Growth Factor-2
INSR	Insulin Receptor
INS-VNTR	Insulin gene
IPG	Immobilised pH Gradient

IRS	Insulin Receptor Substrate
kDa	kiloDalton
Kg	kilogram
L	Litre
LC	Liquid Chromatography
LC/MS/MS	Liquid Chromatography Mass Spectrometry
LDL	Low Density Lipoproteins
LPL	Lipoprotein Lipase
M	Molar
MAC	Membrane Attacks Complex
MALDI-TOF	Matrix Assisted Laser Desorption Time Of Flight
MAP	mean arterial pressure
MS	Mass Spectrometry
mg/dl	milligram per decilitre
ml	millilitre
mM	miliMolar
MODY	Maturity Onset Diabetes of the Young
MSAFP	Maternal Serum Alpha Fetoprotein
mW	molecular Weight
NCBI	National Centre for Biotechnology Information

Ng	nanogram
NGT	Normal Glucose Tolerance
Nm	nanometer
OGTT	Oral Glucose Tolerance Test
p value	probability
PAI-1	Plasminogen Activator Inhibitor-1
pH	Hydrogen Ion concentration
pI	Isoelectric point
PPAR γ	Peroxisome Proliferator-Activated Receptor gamma
PTM	Post-Translational Modifications
RAAS	Renin-Angiotensin Aldosterone System
Rpm	revolutions per minute
SDS	Sodium Dodecyl Sulphate
SDS-PAGE	Sodium Dodecyl Sulphate Polyacrylamide Gel Electrophoresis
T1DM	Type 1 Diabetes Mellitus
TCA	Trichloroacetic acid
2-DGE	Two-Dimensional Gel Electrophoresis
T2DM	Type 2 Diabetes Mellitus
USPSTF	U.S. Preventive Service Task Force
μ l	Microlitre

UPM	University Putra Malaysia
V	Voltage
V/hr	Voltage per hour
v/v	volume concentration
VDBP	Vitamin D Binding Protein
VDR	Vitamin D Receptor
VDRE	Vitamin D Response Element
VLDL	Very Low Density Lipoprotein
w/v	weight per volume
WHO	World Health Organisation
A	Alpha
B	Beta
Γ	Gamma

CHAPTER 1

INTRODUCTION

1.1 Gestational Diabetes Mellitus

Gestational Diabetes Mellitus (GDM) is defined as any glucose intolerance with onset or first recognition during pregnancy (Buchanan & Xiang, 2005). GDM affects 3 to 14% of pregnancies depending on the population studied. For instance, GDM affects about 14% of the pregnant population in the United States (Jovanovic & Pettitt, 2001). In Malaysia, according to the statistics provided by Hospital Kuala Lumpur in the year 2008, GDM was the most common medical complication and metabolic disorder during pregnancy. There were 3325 new cases of GDM in 2007 while in 2008, 3898 cases were diagnosed (Che Minah, Staff Nurse of Hospital Kuala Lumpur, pers. comm. 20 December 2009).

Generally, the pathophysiology of GDM is still unclear but it is understood that GDM results when the insulin levels are insufficient to meet insulin demands during pregnancy. Placental secretion of hormones such as human Placental Lactogen (hPL), estrogen, progesterone and cortisol is a major contributor to the insulin resistant state seen in pregnancy (Butte, 2000). As pregnancy progresses, the increased placental hormones lead to insulin resistance which prevents glucose from entering the cells and remains in the bloodstream and elevates blood glucose levels to cause GDM. The peak of these hormones is seen from the 26th to 33rd week of gestation (Barros *et al.*, 2006).

The presence of GDM has implications for both the mother and her baby. Prenatal morbidity includes macrosomia, shoulder dystocia, neonatal metabolic abnormalities, congenital malformations and neonatal death which in turn may generate subsequent complications. Long term outcomes for the offspring may include obesity and diabetes independent of genetic factors. For the mother, uncontrolled blood sugar level in expectant mother can lead to preeclampsia, hypertension and cardiovascular diseases during pregnancy. Complications by GDM could lead to caesarean delivery which could add to the length of hospitalisation and there is an increased risk of obvious type 2 diabetes mellitus (T2DM) after pregnancy (Metzenger *et al.*, 2007).

The risk factors that contribute to GDM are advancing age, where the age of the expectant mother is more than 35 years old, being overweight or obese, diabetes in their first degree relatives, previous history of GDM and prior delivery to a macrosomic infant (Cho *et al.*, 2007).

Screening for glucose intolerance during pregnancy provides an opportunity to offer management to those women diagnosed with GDM and thus reducing maternal morbidity and prenatal morbidity and mortality. The usual approach to detecting GDM is to screen all pregnant women by measuring their plasma glucose by oral glucose tolerance test at 24 to 28 weeks gestation according to World Health Organisation (WHO) guidelines (Maresh, 2005).

Insulin therapy is the most common treatment used to control the blood sugar level among GDM patients. The dose of insulin depends and is tailored to meet each patient's requirement. Self monitoring of blood glucose, diet and exercise are also

recommended to GDM patients in order to keep their blood glucose range at their optimum level (Crowther *et al.*, 2005).

1.2 Problem Statement

The main purpose of identifying GDM is to detect women at risk of adverse prenatal outcomes and reduce the serious neonatal complications. It is well recognised that women with previous history of GDM have an increased risk to develop T2DM, which is a major cause of morbidity and mortality and has economic implications as well. The American Diabetes Association (ADA), the American College of Obstetricians and Gynaecologists (ACOG) and the World Health Organisation recommend screening most pregnant women for GDM between 24 and 28 weeks of gestation and screening high risk pregnant women at the first antenatal visit. However, in the year of 2003, the U.S. Preventive Service Task Force (USPSTF) concluded that there was insufficient evidence to advise for or against routinely screening all pregnant women since using clinical risk factors failed to diagnose one third to one half of patients with GDM (Hillier *et al.*, 2008).

Even in light of these guidelines, the current diagnostic criteria for GDM remain controversial since the screening approach, prevalence, clinical management and impact on maternal and infant outcomes lack sufficient facts and evidence (Jovanovic & Pettitt, 2001). Many investigators stress the importance of establishing a new criteria for screening that are specific to pregnancy and validated by pregnancy outcomes.

Aside from the lack of international agreement on the diagnosis of GDM, controversy abounds as to the relevance and timing of screening, which would influence management of GDM and affect prenatal outcomes.

1.3 Significance of the study

A major focus of human genetics research is the identification of the molecular basis of diseases as to improve diagnostic capabilities and therapeutic options. Screening for a disease is recommended if the disease is common and clinically important. In addition, also of are about the potential harm and cost of screening especially given the high rates of false-positive or false negative results. In the case of GDM, the purpose of screening is not to diagnose the disease but to identify those at risk as early as reasonably possible for whom a diagnostic test may be offered. Thus, this would provide an opportunity for better health management of those women diagnosed with GDM in the early trimesters.

In order to achieve this goal, the usefulness of a biomarker in screening for complications in pregnancies relies on its detection in maternal blood during the early stages of pregnancy (Shankar *et al.*, 2004). These biomarkers could potentially be used as indicators of the effects or progression of GDM. Therefore, a better understanding of the changes that occur at the molecular level in GDM will facilitate early detection of disease. Thus far, no studies have assessed the utility of multiple biomarker screening for GDM in Malaysia.

Proteomic analysis based on mass spectrometry is a powerful tool for the identification of disease-related biomarkers (Shankar *et al.*, 2004). With proteomics, profiling of human plasma proteome becomes more feasible in searching for disease related biomarkers in the field of reproduction. The attention of several researchers has focused on the examination of the global protein expression in body fluids in order to provide better diagnosis for GDM and pregnancy related complications.

In the present study, a combined approach based on two-dimensional gel electrophoresis (2-DGE) and tandem mass spectrometry (MS), was used in order to determine differentially expressed proteins for GDM by comparing the protein expression patterns of maternal plasma from normal pregnant women and GDM patients during the different weeks of gestation and to identify a set of GDM specific biomarkers.

Comparative proteome profiling of normal and GDM patients' plasma will contribute significantly to our understanding of the physiology and pathophysiology of GDM. This could translate into identification of disease markers as well as for monitoring of disease progression.

Possibly the most important outcome from this differential proteomic approach is that identified markers could be used to screen patients in the early trimester to reduce the consequences of GDM. In addition, this approach could be the basis in providing a better understanding of GDM especially with regard to underlying molecular pathways involved in GDM progression and potentially lead to novel therapeutic approaches for GDM (Sparre *et al.*, 2005).

1.4 Rationale of this study

Identification of differentially expressed proteins in plasma from pregnant woman obtained in early trimester periods as candidate biomarkers for GDM and may potentially provide opportunities for the early detection and treatment of GDM which in turn would help reduce foetal and maternal morbidity and reduce unfavourable outcomes.

1.5 Objectives

1.5.1 General objective:

To identify potential protein markers for GDM.

1.5.2 Specific objectives:

1. To compare the plasma protein profiles for the first, second and third trimesters between GDM patients and normal pregnant women.
2. To compare differentially expressed proteins in plasma between GDM patients and normal pregnant women.
3. To identify potential protein markers in plasma for GDM.

REFERENCES

- Abdullah, N. M., Kachman, M., Walker, A., Hawley, A. E., Wroblewski, S. K., Myers, D. D., et al. (2009). Microparticle surface proteins are associated with experimental venous thrombosis: a preliminary study. *Clinical and applied thrombosis/hemostasis: official journal of the International Academy of Clinical and Applied Thrombosis/Hemostasis*, 15(2), 201.
- Aberg, A., L. Westbom, et al. (2001). "Congenital malformations among infants whose mothers had gestational diabetes or preexisting diabetes." *Early human development* 61(2): 85-95.
- Adamson, J. W. (2008). The anemia of inflammation/malignancy: mechanisms and management. *ASH Education Program Book*, 2008(1), 159-165.
- Aebersold, R. and M. Mann (2003). "Mass spectrometry-based proteomics." *Nature* 422(6928): 198-207.
- Agarwal, M. M., Dhatt, G. S., & Shah, S. M. (2010). Gestational Diabetes Mellitus. *Diabetes Care*, 33(9), 2018-2020.
- Ahima, R. S., & Flier, J. S. (2000). Adipose tissue as an endocrine organ. *Trends in Endocrinology & Metabolism*, 11(8), 327-332.
- Aird, W. (2007). Vascular bed-specific thrombosis. *Journal of Thrombosis and Haemostasis*, 5, 283-291.
- Alaupovic, P. (1996). [2] Significance of apolipoproteins for structure, function, and classification of plasma lipoproteins. *Methods in enzymology*, 263, 32-60.
- Alvarez, J. A., & Ashraf, A. (2009). Role of vitamin D in insulin secretion and insulin sensitivity for glucose homeostasis. *International journal of endocrinology*, 2010.
- Artal, R. A. R., R. B. C. R. B. Catanzaro, et al. (2007). "A lifestyle intervention of weight-gain restriction: diet and exercise in obese women with gestational diabetes mellitus." *Applied physiology, nutrition, and metabolism* 32(3): 596-601.
- Barbour, L. A., C. E. McCurdy, et al. (2007). "Cellular mechanisms for insulin resistance in normal pregnancy and gestational diabetes." *Diabetes care* 30(Supplement 2): S112.
- Barros, R., Machado, U. F., & Gustafsson, J. A. (2006). Estrogen receptors: new players in diabetes mellitus. *Trends in molecular medicine*, 12(9), 425-431.

- Ben-Haroush, A., Yogeve, Y., & Hod, M. (2004). Epidemiology of gestational diabetes mellitus and its association with type 2 diabetes. *Diabetic Medicine*, 21(2), 103-113.
- Berger, H., J. Crane, et al. (2002). "Screening for gestational diabetes mellitus." *Journal of obstetrics and gynaecology Canada: JOGC= Journal d'obstétrique et gynécologie du Canada: JOGC* 24(11): 894.
- Bjorhall, K., T. Miliotis, et al. (2005). "Comparison of different depletion strategies for improved resolution in proteomic analysis of human serum samples." *Proteomics* 5(1): 307-317.
- Blumenstein, M., McMaster, M. T., Black, M. A., Wu, S., Prakash, R., Cooney, J., et al. (2009). A proteomic approach identifies early pregnancy biomarkers for preeclampsia: novel linkages between a predisposition to preeclampsia and cardiovascular disease. *Proteomics*, 9(11), 2929-2945.
- Bouillon, R., Okamura, W. H., & Norman, A. W. (1995). Structure-function relationships in the vitamin D endocrine system. *Endocrine reviews*, 16(2), 200-257.
- Brantly, M., Nukiwa, T., & Crystal, R. G. (1988). Molecular basis of alpha-1-antitrypsin deficiency. *The American journal of medicine*, 84(6), 13-31.
- Brody, S. C., R. Harris, et al. (2003). "Screening for gestational diabetes: a summary of the evidence for the US Preventive Services Task Force." *Obstetrics & Gynecology* 101(2): 380.
- Brown, M. A. (1989). Sodium balance in pregnancy. *Fetal and Maternal Medicine Review*, 1(02), 193-212.
- Buchanan, T. A., & Xiang, A. H. (2005). Gestational diabetes mellitus. *J Clin Invest*, 115(3), 485-491.
- Buhl, R., & Farmer, S. G. (2005). Future directions in the pharmacologic therapy of chronic obstructive pulmonary disease. *Proceedings of the American Thoracic Society*, 2(1), 83-93.
- Butte, N. F. (2000). Carbohydrate and lipid metabolism in pregnancy: normal compared with gestational diabetes mellitus. *The American journal of clinical nutrition*, 71(5), 1256S.
- Carlin, A., & Alfirevic, Z. (2008). Physiological changes of pregnancy and monitoring. *Best Practice & Research Clinical Obstetrics & Gynaecology*, 22(5), 801-823.

- Carrell, R. W., Jeppsson, J. O., Laurell, C. B., Brennan, S. O., Owen, M. C., Vaughan, L., et al. (1982). Structure and variation of human alpha 1-antitrypsin. *Nature*, 298(5872), 329.
- Carroll, M. C. (2004). The complement system in regulation of adaptive immunity. *Nature immunology*, 5(10), 981-986.
- Catalano, P. M. and J. P. Kirwan (2001). "Maternal factors that determine neonatal size and body fat." *Current diabetes reports* 1(1): 71-77.
- Catalano, P. M., J. P. Kirwan, et al. (2003). "Gestational diabetes and insulin resistance: role in short-and long-term implications for mother and fetus." *The Journal of nutrition* 133(5): 1674S.
- Chauhan, S. P., D. J. West, et al. (2000). "Antepartum detection of macrosomic fetus: clinical versus sonographic, including soft-tissue measurements." *Obstetrics & Gynecology* 95(5): 639.
- Che Minah, Staff Nurse of Hospital Kuala Lumpur, pers. comm. 20 December 2009
- Chen, X., & Scholl, T. O. (2005). Oxidative stress: changes in pregnancy and with gestational diabetes mellitus. *Current diabetes reports*, 5(4), 282-288.
- Cho, A. R., Kyeung, K. S., Park, M. A., Lee, Y. M., & Jeong, E. H. (2007). Risk Factors of Gestational Diabetes Mellitus. *Korean Journal of Perinatology*, 18(4), 329-337.
- Christakos, S., Raval-Pandya, M., Wernyj, R. P., & Yang, W. (1996). Genomic mechanisms involved in the pleiotropic actions of 1, 25-dihydroxyvitamin D3. *Biochemical Journal*, 316(Pt 2), 361.
- Christine, A., & LUMSDEN, M. A. (2001). Diabetes and the maternal resistance vasculature. *Clinical Science*, 101, 719-729.
- Cedazo Mínguez, A. (2007). Apolipoprotein E and Alzheimer's disease: molecular mechanisms and therapeutic opportunities. *Journal of cellular and molecular medicine*, 11(6), 1227-1238.
- Cerneca, F., Ricci, G., Simeone, R., Malisano, M., Alberico, S., & Guaschino, S. (1997). Coagulation and fibrinolysis changes in normal pregnancy. Increased levels of procoagulants and reduced levels of inhibitors during pregnancy induce a hypercoagulable state, combined with a reactive fibrinolysis. *European Journal of Obstetrics & Gynecology and Reproductive Biology*, 73(1), 31-36.
- Clifton Bligh, R., McElduff, P., & McElduff, A. (2008). Maternal vitamin D deficiency, ethnicity and gestational diabetes. *Diabetic medicine*, 25(6), 678-684.

- Coakley, F. V., & Schwartz, L. H. (2001). *Imaging of hepatocellular carcinoma: a practical approach*.
- Cosson, E., M. Benchimol, et al. (2006). "Universal rather than selective screening for gestational diabetes mellitus may improve fetal outcomes." *Diabetes & metabolism* 32(2): 140-146.
- Coustan, D. R. (2004). "Delivery: timing, mode, and management." *Diabetes in women: adolescence, pregnancy, and menopause*: 433.
- Coakley, R. J., Taggart, C., O'Neill, S., & McElvaney, N. G. (2001). [alpha] 1-Antitrypsin Deficiency:: Biological Answers to Clinical Questions. *The American journal of the medical sciences*, 321(1), 33-41.
- Crowther, C. A., Hiller, J. E., Moss, J. R., McPhee, A. J., Jeffries, W. S., & Robinson, J. S. (2005). Effect of treatment of gestational diabetes mellitus on pregnancy outcomes. *New England Journal of Medicine*, 352(24), 2477-2486.
- Danilenko-Dixon, D. R., J. T. Van Winter, et al. (1999). "Universal versus selective gestational diabetes screening: application of 1997 American Diabetes Association recommendations." *American journal of obstetrics and gynecology* 181(4): 798-802.
- Darde, V. M., M. G. Barderas, et al. (2007). "Depletion of High-Abundance Proteins in Plasma by Immunoaffinity Subtraction for Two-Dimensional Difference Gel Electrophoresis Analysis." *Methods in Molecular Biology- Clifton then Totowa* - 357: 351.
- Davison, J., & Lindheimer, M. (1989). 8 Volume homeostasis and osmoregulation in human pregnancy. *Baillière's clinical endocrinology and metabolism*, 3(2), 451-472.
- DeLuca, H. F. (2004). Overview of general physiologic features and functions of vitamin D. *The American journal of clinical nutrition*, 80(6), 1689S.
- DeLuca, H. F., & Zierold, C. (1998). Mechanisms and functions of vitamin D. *Nutrition reviews*, 56, S4-S10.
- Dempsey, J. C., C. L. Butler, et al. (2004). "A case-control study of maternal recreational physical activity and risk of gestational diabetes mellitus." *Diabetes research and clinical practice* 66(2): 203-215.
- Di Cianni, G., L. Volpe, et al. (2003). "Prevalence and risk factors for gestational diabetes assessed by universal screening." *Diabetes research and clinical practice* 62(2): 131-137.
- Diamandis, E. P. (2004). Analysis of serum proteomic patterns for early cancer diagnosis: drawing attention to potential problems. *Journal of the National Cancer Institute*, 96(5), 353-356.

- Eapen, R. S., D. G. Rowland, et al. (1998). "Effect of prenatal diagnosis of critical left heart obstruction on perinatal morbidity and mortality." *American journal of perinatology* 15: 237-242.
- Ember, J. A., & Hugli, T. E. (1997). Complement factors and their receptors. *Immunopharmacology*, 38(1-2), 3.
- Erkan, D. (2002). The relation between antiphospholipid syndrome-related pregnancy morbidity and non-gravid vascular thrombosis: a review of the literature and management strategies. *Current rheumatology reports*, 4(5), 379-386.
- Fountoulakis, M. (2001). Proteomics: current technologies and applications in neurological disorders and toxicology. *Amino Acids*, 21(4), 363-381
- Franz, M. J., J. P. Bantle, et al. (2002). "Evidence-based nutrition principles and recommendations for the treatment and prevention of diabetes and related complications." *Diabetes care* 25(1): 148.
- Friedman, D. B., S. Hoving, et al. (2009). "Isoelectric focusing and two-dimensional gel electrophoresis." *Methods in enzymology* 463: 515-540.
- Freinkel, N. and B. E. Metzger (1979). "Pregnancy as a tissue culture experience: the critical implications of maternal metabolism for fetal development."
- Gabbe, S. G. and C. R. Graves (2003). "Management of diabetes mellitus complicating pregnancy." *Obstetrics & Gynecology* 102(4): 857.
- Galli, M., & Barbui, T. (2003). Antiphospholipid antibodies and pregnancy. *Best Practice & Research Clinical Haematology*, 16(2), 211-225.
- Georgiou, H. M., M. Lappas, et al. (2008). "Screening for biomarkers predictive of gestational diabetes mellitus." *Acta diabetologica* 45(3): 157-165.
- Giustarini, D., Dalle-Donne, I., Tsikas, D., & Rossi, R. (2009). Oxidative stress and human diseases: origin, link, measurement, mechanisms, and biomarkers. *Critical reviews in clinical laboratory sciences*, 46(5-6), 241-281.
- Goldman, A. S., L. Baker, et al. (1985). "Hyperglycemia-induced teratogenesis is mediated by a functional deficiency of arachidonic acid." *Proceedings of the National Academy of Sciences of the United States of America* 82(23): 8227.
- Gorg, A., W. Weiss, et al. (2004). "Current two dimensional electrophoresis technology for proteomics." *Proteomics* 4(12): 3665-3685.
- Graves, S. W. (2007). Sodium regulation, sodium pump function and sodium pump inhibitors in uncomplicated pregnancy and preeclampsia. *Front Biosci*, 12, 2438-2446.

- Gravett, M. G., Novy, M. J., Rosenfeld, R. G., Reddy, A. P., Jacob, T., Turner, M., et al. (2004). Diagnosis of intra-amniotic infection by proteomic profiling and identification of novel biomarkers. *JAMA: the journal of the American Medical Association*, 292(4), 462-469.
- Greer, I. A. (1999). Thrombosis in pregnancy: maternal and fetal issues. *The Lancet*, 353(9160), 1258-1265.
- Greenough, C., Jenkins, R. E., Kitteringham, N. R., Pirmohamed, M., Park, B. K., & Pennington, S. R. (2004). A method for the rapid depletion of albumin and immunoglobulin from human plasma. *Proteomics*, 4(10), 3107-3111.
- Gunton, J. E., A. McElduff, et al. (2000). "Outcome of pregnancies complicated by pre gestational diabetes mellitus." *Australian and New Zealand Journal of Obstetrics and Gynaecology* 40(1): 38-43.
- Gygi, S. P., G. L. Corthals, et al. (2000). "Evaluation of two-dimensional gel electrophoresis-based proteome analysis technology." *Proceedings of the National Academy of Sciences* 97(17): 9390.
- Heng, Y. J., Di Quinzio, M. K. W., Permezel, M., Ayhan, M., Rice, G. E., & Georgiou, H. M. Temporal proteomic analysis of human cervicovaginal fluid with impending term labor. *Journal of proteome research*, 9(3), 1344-1350.
- Henschen, A., Lottspeich, F., Kehl, M., & Southan, C. (1983). Covalent structure of fibrinogen. *Annals of the New York Academy of Sciences*, 408(1), 28-43.
- Herrera, E. (2000). Metabolic adaptations in pregnancy and their implications for the availability of substrates to the fetus. *European journal of clinical nutrition*, 54, S47.
- Hillier, T. A., Vesco, K. K., Pedula, K. L., Beil, T. L., Whitlock, E. P., & Pettitt, D. J. (2008). Screening for gestational diabetes mellitus: a systematic review for the US Preventive Services Task Force. *Annals of internal medicine*, 148(10), 766.
- Holick, M. F. (2007). Vitamin D deficiency. *New England journal of medicine*, 357(3), 266-281.
- Holmer, E., Lindahl, U., Bäckström, G., Thunberg, L., Sandberg, H., Söderström, G., et al. (1980). Anticoagulant activities and effects on platelets of a heparin fragment with high affinity for antithrombin. *Thrombosis Research*, 18(6), 861-869.
- Holt, R., J. Goddard, et al. (2003). "A postnatal fasting plasma glucose is useful in determining which women with gestational diabetes should undergo a postnatal oral glucose tolerance test." *Diabetic medicine* 20(7): 594-598.

- Huang, Y., Krein, P. M., Muruve, D. A., & Winston, B. W. (2002). Complement factor B gene regulation: synergistic effects of TNF- α and IFN- γ in macrophages. *The Journal of Immunology*, 169(5), 2627-2635.
- Issaq, H. J. and T. D. Veenstra (2008). "Two-dimensional polyacrylamide gel electrophoresis (2D-PAGE): advances and perspectives." *Biotechniques* 44(5): 697.
- Janeway Jr, C. A., & Medzhitov, R. (2002). Innate immune recognition. *Annual review of immunology*, 20(1), 197-216.
- Janus, E. D., Phillips, N. T., & Carrell, R. W. (1985). SMOKING, LUNG FUNCTION, AND [alpha] 1-ANTITRYPSIN DEFICIENCY. *The Lancet*, 325(8421), 152-154.
- Jiang, X. S., H. Zhou, et al. (2004). "A high-throughput approach for subcellular proteome." *Molecular & Cellular Proteomics* 3(5): 441-455.
- Jolly, M. C., N. J. Sebire, et al. (2003). "Risk factors for macrosomia and its clinical consequences: a study of 350,311 pregnancies." *European Journal of Obstetrics & Gynecology and Reproductive Biology* 111(1): 9-14.
- Jovanovic, L., & Pettitt, D. J. (2001). Gestational diabetes mellitus. *JAMA: The Journal of the American Medical Association*, 286(20), 2516.
- Jovanovic Peterson, L. and C. M. Peterson (1996). "Review of Gestational Diabetes Mellitus and Low calorie Diet and Physical Exercise as Therapy." *Diabetes/metabolism reviews* 12(4): 287-308.
- Kaaja, R. J. and I. A. Greer (2005). "Manifestations of chronic disease during pregnancy." *JAMA: the journal of the American Medical Association* 294(21): 2751.
- Kamath, U., G. Rao, et al. (1998). "Erythrocyte indicators of oxidative stress in gestational diabetes." *Acta Paediatrica* 87(6): 676-679.
- Kim, C., K. M. Newton, et al. (2002). "Gestational diabetes and the incidence of type 2 diabetes." *Diabetes care* 25(10): 1862.
- Kjos, S. L., U. Schaefer-Graf, et al. (2001). "A randomized controlled trial using glycemic plus fetal ultrasound parameters versus glycemic parameters to determine insulin therapy in gestational diabetes with fasting hyperglycemia." *Diabetes care* 24(11): 1904.
- Kjos, S. L., & Buchanan, T. A. (1999). Gestational diabetes mellitus. *New England Journal of Medicine*, 341(23), 1749-1756.

- Kobori, H., Harrison-Bernard, L. M., & Navar, L. G. (2001). Expression of angiotensinogen mRNA and protein in angiotensin II-dependent hypertension. *Journal of the American Society of Nephrology*, 12(3), 431-439.
- Kolialexi, A., Tsangaris, G. T., Papantoniou, N., Anagnostopoulos, A. K., Vougas, K., Bagiokos, V., et al. (2008). Application of proteomics for the identification of differentially expressed protein markers for Down syndrome in maternal plasma. *Prenatal diagnosis*, 28(8), 691-698.
- Korc, M. (2003). Diabetes mellitus in the era of proteomics. *Molecular & Cellular Proteomics*, 2(6), 399.
- Koukkou, E., Watts, G., & Lowy, C. (1996). Serum lipid, lipoprotein and apolipoprotein changes in gestational diabetes mellitus: a cross-sectional and prospective study. *Journal of clinical pathology*, 49(8), 634-637.
- Kovacs, C. S. (2008). Vitamin D in pregnancy and lactation: maternal, fetal, and neonatal outcomes from human and animal studies. *The American journal of clinical nutrition*, 88(2), 520S-528S.
- Kragh-Hansen, U. (1990). Structure and ligand binding properties of human serum albumin. *Danish medical bulletin*, 37(1), 57.
- Krane, N. K., & Hamrahan, M. (2007). Pregnancy: kidney diseases and hypertension. *Am J Kidney Dis*, 49(2), 336-345.
- Kühl, C. (1991). "Insulin secretion and insulin resistance in pregnancy and GDM. Implications for diagnosis and management." *Diabetes* 40: 18.
- Langer, O., D. L. Conway, et al. (2000). "A comparison of glyburide and insulin in women with gestational diabetes mellitus." *therapy* 343: 1134-8.
- Lappas, M., K. Yee, et al. (2005). "Release and regulation of leptin, resistin and adiponectin from human placenta, fetal membranes, and maternal adipose tissue and skeletal muscle from normal and gestational diabetes mellitus-complicated pregnancies." *Journal of endocrinology* 186(3): 457.
- Lenz, T., Sealey, J. E., & Tewksbury, D. A. (1993). Regional distribution of the angiotensinogens in human placentae. *Placenta*, 14(6), 695-699.
- Lilley, K. S., A. Razzaq, et al. (2002). "Two-dimensional gel electrophoresis: recent advances in sample preparation, detection and quantitation." *Current opinion in chemical biology* 6(1): 46-50.
- Lockitch, G., & Gamer, P. (1997). Clinical biochemistry of pregnancy. *Critical reviews in clinical laboratory sciences*, 34(1), 67-139.

- Lomas, D., & Parfrey, H. (2004). α 1-Antitrypsin deficiency: 4: Molecular pathophysiology. *Thorax*, 59(6), 529-535.
- MacFarlane, C., & Tsakalagos, N. (1988). The extended Pedersen hypothesis. *Clinical physiology and biochemistry*, 6(2), 68.
- Macones, G. A., J. Peipert, et al. (2005). "Maternal complications with vaginal birth after cesarean delivery: a multicenter study." *American journal of obstetrics and gynecology* 193(5): 1656-1662.
- Mahley, R. W., Innerarity, T. L., Rall Jr, S. C., & Weisgraber, K. H. (1984). Plasma lipoproteins: apolipoprotein structure and function. *Journal of lipid research*, 25(12), 1277-1294.
- Maresh, M. (2005). *Screening for gestational diabetes mellitus*.
- Martinelli, I., Bucciarelli, P., & Mannucci, P. M. (2010). Thrombotic risk factors: basic pathophysiology. *Critical care medicine*, 38, S3.
- Mathieu, C., Gysemans, C., Giulietti, A., & Bouillon, R. (2005). Vitamin D and diabetes. *Diabetologia*, 48(7), 1247-1257.
- Mellor, A., & Munn, D. (2000). Immunology at the maternal-fetal interface: lessons for T cell tolerance and suppression. *Annual review of immunology*, 18(1), 367-391.
- Mensink, R. P., Zock, P. L., Kester, A. D. M., & Katan, M. B. (2003). Effects of dietary fatty acids and carbohydrates on the ratio of serum total to HDL cholesterol and on serum lipids and apolipoproteins: a meta-analysis of 60 controlled trials. *The American journal of clinical nutrition*, 77(5), 1146-1155.
- Metzger, B. E., Buchanan, T. A., Coustan, D. R., de Leiva, A., Dunger, D. B., Hadden, D. R., et al. (2007). Summary and recommendations of the fifth international workshop-conference on gestational diabetes mellitus. *Diabetes care*, 30(Supplement 2), S251.
- Milman, N. (2006). Iron and pregnancy—a delicate balance. *Annals of hematology*, 85(9), 559-565.
- Monaghan, J. M., Godber, I. M., Lawson, N., Kaur, M., Wark, G., Teale, D., et al. (2004). Longitudinal changes of insulin-like growth factors and their binding proteins throughout normal pregnancy. *Annals of clinical biochemistry*, 41(3), 220-226.
- Morris, N. H., Eaton, B. M., & Dekker, G. (1996). Nitric oxide, the endothelium, pregnancy and pre eclampsia. *BJOG: An International Journal of Obstetrics & Gynaecology*, 103(1), 4-15.

- Nanda, S., Savvidou, M., Syngelaki, A., Akolekar, R., & Nicolaides, K. H. (2011). Prediction of gestational diabetes mellitus by maternal factors and biomarkers at 11 to 13 weeks. *Prenatal diagnosis*, 31(2), 135-141.
- Ong, S. E. and A. Pandey (2001). "An evaluation of the use of two-dimensional gel electrophoresis in proteomics." *Biomolecular engineering* 18(5): 195-205.
- Osorio, J. H. (2003). "Carbohydrate metabolism during pregnancy." *Rev Colomb Obstet Ginecol* 54(2): 97-106.
- O'Sullivan, J. B. (1982). "Body weight and subsequent diabetes mellitus." *JAMA: the journal of the American Medical Association* 248(8): 949.
- Pangburn, M., & Rawal, N. (2002). Structure and function of complement C5 convertase enzymes. *Biochemical Society Transactions*, 30(Pt 6), 1006.
- Palomer, X., González-Clemente, J., Blanco-Vaca, F., & Mauricio, D. (2008). Role of vitamin D in the pathogenesis of type 2 diabetes mellitus. *Diabetes, Obesity and Metabolism*, 10(3), 185-197.
- Palomer, X., González Clemente, J., Blanco Vaca, F., & Mauricio, D. (2007). Role of vitamin D in the pathogenesis of type 2 diabetes mellitus. *Diabetes, Obesity and Metabolism*, 10(3), 185-197.
- Patz Jr, E. F., Campa, M. J., Gottlin, E. B., Kusmartseva, I., Guan, X. R., & Herndon II, J. E. (2007). Panel of serum biomarkers for the diagnosis of lung cancer. *Journal of Clinical Oncology*, 25(35), 5578-5583.
- Pinter, E., E. Reece, et al. (1986). "Arachidonic acid prevents hyperglycemia-associated yolk sac damage and embryopathy." *American journal of obstetrics and gynecology* 155(4): 691.
- Plominsky, Á. M., Soto-Liebe, K., & Vasquez, M. (2009). Optimization of 2D-PAGE protocols for proteomic analysis of two nonaxenic toxin-producing freshwater cyanobacteria: *Cylindrospermopsis raciborskii* and *Raphidiopsis* sp. *Letters in applied microbiology*, 49(3), 332-337.
- Qiu, C., Sorensen, T. K., Luthy, D. A., & Williams, M. A. (2004). A prospective study of maternal serum C-reactive protein (CRP) concentrations and risk of gestational diabetes mellitus. *Paediatric and perinatal epidemiology*, 18(5), 377-384.
- Quinlan, G. J., Martin, G. S., & Evans, T. W. (2005). Albumin: biochemical properties and therapeutic potential. *Hepatology*, 41(6), 1211-1219.
- Radaelli, T., A. Varastehpour, et al. (2003). "Gestational diabetes induces placental genes for chronic stress and inflammatory pathways." *Diabetes* 52(12): 2951.

- Rao, P. V., Lu, X., Standley, M., Pattee, P., Neelima, G., Girishes, G., et al. (2007). Proteomic identification of urinary biomarkers of diabetic nephropathy. *Diabetes Care*, 30(3), 629-637.
- Ray, J. G., M. J. Vermeulen, et al. (2005). "Cardiovascular health after maternal placental syndromes (CHAMPS): population-based retrospective cohort study." *The Lancet* 366(9499): 1797-1803.
- Retnakaran, R., A. J. G. Hanley, et al. (2005). "Adiponectin and beta cell dysfunction in gestational diabetes: pathophysiological implications." *Diabetologia* 48(5): 993-1001.
- Riley, J. K. (2008). Trophoblast immune receptors in maternal-fetal tolerance. *Immunological investigations*, 37(5-6), 395-426.
- Rosengren, A. T., J. M. Salmi, et al. (2003). "Comparison of PDQuest and Progenesis software packages in the analysis of two-dimensional electrophoresis gels." *Proteomics* 3(10): 1936-1946.
- Roth-Kleiner, M., B. P. Wagner, et al. (2003). "Respiratory distress syndrome in near-term babies after caesarean section." *Swiss Medical Weekly* 133(19-20): 283-288.
- Salhab, W. A., M. H. Wyckoff, et al. (2004). "Initial hypoglycemia and neonatal brain injury in term infants with severe fetal acidemia." *Pediatrics* 114(2): 361.
- Samuel, S., & Sitrin, M. D. (2008). Vitamin D's role in cell proliferation and differentiation. *Nutrition reviews*, 66, S116-S124.
- Scholten, R. R., Sep, S., Peeters, L., Hopman, M. T. E., Lotgering, F. K., & Spaanderman, M. E. A. Prepregnancy low-plasma volume and predisposition to preeclampsia and fetal growth restriction. *Obstetrics & Gynecology*, 117(5), 1085.
- Seshadri, K. G., Tamilselvan, B., & Rajendran, A. (2011). Role of Vitamin D in Diabetes. *Journal of Endocrinology and Metabolism*, 1(2), 47-56.
- Shankar, R., Cullinane, F., Brennecke, S. P., & Moses, E. K. (2004). Applications of proteomic methodologies to human pregnancy research: A growing gestation approaching delivery *Proteomics*, 4(7), 1909-1917.
- Sears, P., & Wong, C. H. (1998). Enzyme action in glycoprotein synthesis. *Cellular and molecular life sciences*, 54(3), 223-252.
- Sibai, B. M., S. Caritis, et al. (2000). "Risks of preeclampsia and adverse neonatal outcomes among women with pregestational diabetes mellitus." *American journal of obstetrics and gynecology* 182(2): 364-369.

- Simpson, J. (1995). Noninvasive screening for prenatal genetic diagnosis. *Bulletin of the World Health Organization*, 73(6), 799.
- Sonoda, J., Pei, L., & Evans, R. M. (2008). Nuclear receptors: decoding metabolic disease. *FEBS letters*, 582(1), 2-9.
- Sparre, T., Larsen, M. R., Heding, P. E., Karlsen, A. E., Jensen, O. N., & Pociot, F. (2005). Unraveling the pathogenesis of type 1 diabetes with proteomics: present and future directions. *Molecular & Cellular Proteomics*, 4(4), 441.
- Stoller, J. K., & Aboussouan, L. S. (2005). [alpha] 1-antitrypsin deficiency. *The Lancet*, 365(9478), 2225-2236.
- Sundsten, T., & Ortsäter, H. (2009). Proteomics in diabetes research. *Molecular and Cellular Endocrinology*, 297(1), 93-103.
- Sutton, A. L. M., & MacDonald, P. N. (2003). Vitamin D: more than a "bone-a-fide" hormone. *Molecular Endocrinology*, 17(5), 777-791.
- Tahara, Y., & Shima, K. (1995). Kinetics of HbA1c, glycated albumin, and fructosamine and analysis of their weight functions against preceding plasma glucose level. *Diabetes care*, 18(4), 440.
- Teegarden, D., & Donkin, S. S. (2009). Vitamin D: emerging new roles in insulin sensitivity. *Nutrition research reviews*, 22(1), 82.
- Toglia, M. R., & Weg, J. G. (1996). Venous thromboembolism during pregnancy. *New England Journal of Medicine*, 335(2), 108-114.
- Turok, D. K., S. D. Ratcliffe, et al. (2003). "Management of gestational diabetes mellitus." *American family physician* 68(9): 1767.
- Van den Broek, N., & Letsky, E. (2001). Pregnancy and the erythrocyte sedimentation rate. *British Journal of Obstetrics and Gynaecology*, 108(11), 1164-1167.
- Vihinen, M. (2001). Bioinformatics in proteomics. *Biomolecular engineering*, 18(5), 241-248.
- Vlodavsky, I., & Friedmann, Y. (2001). Molecular properties and involvement of heparanase in cancer metastasis and angiogenesis. *Journal of Clinical Investigation*, 108(3), 341-348.
- Wang, H. and S. Hanash (2005). "Intact protein based sample preparation strategies for proteome analysis in combination with mass spectrometry." *Mass spectrometry reviews* 24(3): 413-426.
- Whitaker, R. C., M. S. Pepe, et al. (1998). "Gestational diabetes and the risk of offspring obesity." *Pediatrics* 101(2): e9.

- Weinstock, M. (2008). "The long-term behavioural consequences of prenatal stress." *Neuroscience & Biobehavioral Reviews* 32(6): 1073-1086.
- Weis, W. I., & Drickamer, K. (1996). Structural basis of lectin-carbohydrate recognition. *Annual review of biochemistry*, 65(1), 441-473.
- Wolf, P. L. (1999). Biochemical diagnosis of liver disease. *Indian Journal of clinical biochemistry*, 14(1), 59-90.
- Xiong, X., L. Saunders, et al. (2001). "Gestational diabetes mellitus: prevalence, risk factors, maternal and infant outcomes." *International Journal of Gynecology & Obstetrics* 75(3): 221-228.
- Yaghmaei, M., Hashemi, M., Shikhzadeh, A., Mokhtari, M., Niazi, A., & Ghavami, S. (2009). Serum trypsin inhibitory capacity in normal pregnancy and gestational diabetes mellitus. *Diabetes research and clinical practice*, 84(3), 201-204.
- Yocum, A. K. and A. M. Chinnaiyan (2009). "Current affairs in quantitative targeted proteomics: multiple reaction monitoring "mass spectrometry." *Briefings in Functional Genomics & Proteomics* 8(2): 145-157.
- Zhang, C., Qiu, C., Hu, F. B., David, R. M., Van Dam, R. M., Bralley, A., et al. (2008). Maternal plasma 25-hydroxyvitamin D concentrations and the risk for gestational diabetes mellitus. *PLoS One*, 3(11), e3753.