



**UNIVERSITI PUTRA MALAYSIA**

***EFFECTS OF POSTBIOTIC AND INULIN SUPPLEMENTS ON GROWTH  
PERFORMANCE, GUT MORPHOLOGY, GENE EXPRESSION AND  
FECAL  
CHARACTERISTICS OF BROILER CHICKENS***

**KARWAN YASEEN KARREM**

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By

**KARWAN YASEEN KARREM**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia in Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

**June 2016**

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## DEDICATION

*I would like to dedicate this work to those who taught, motivated and helped me throughout my study. This work is also dedicated to my dearest wife, Dlgash Nooraldin Kareem without her support and love, I could not have accomplished many of the things that I have accomplished.*



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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the Degree of Doctor of Philosophy

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By

**KARWAN YASEEN KAREEM**

**May 2016**

**Chairman : Professor Loh Teck Chwen, PhD**  
**Faculty : Agriculture**

Antibiotic growth promoters (AGP) are effective in modulating the gut microflora and promoting growth performance in birds. Nonetheless, prolonged use of AGP can cause antibiotic resistance and residual effects, which could imperil the health of birds and human consumers. Prebiotics, probiotics and their combinations are potent replacements for AGP. Nonetheless, commercial probiotics have consistently failed to meet the anticipated capability and their efficacy is uncertain. In addition, the use of probiotics could lead to the occurrence of antibiotics gene resistance that could be transferred between organisms. This justifies the need to search for suitable and safe alternative to probiotics. Postbiotic, a metabolite of probiotics exhibits probiotic effect without living cells. Despite the efficacies of postbiotics and prebiotics, the synergistic effects of prebiotic and postbiotic combinations have not been elucidated. This study examined the influence of postbiotic produced by strains of *Lactobacillus plantarum*, and inulin combinations on growth performance, gut morphology, meat quality, gene expression and fecal characteristics of broiler chickens. The study was partitioned into three experiments. The first experiment examined the inhibitory activity of postbiotic produced by strains of *Lactobacillus plantarum* using reconstituted media supplemented with inulin. Postbiotics were produced by six strains of *L. plantarum* (RG11, RG14, RI11, UL4, TL1 and RS5) using reconstituted media supplemented with different levels of inulin (0, 0.2, 0.4, 0.6, 0.8, and 1.0) yielding 36 combinations. The modified inhibitory activity (MAU) of the produced postbiotics were tested against indicator microorganism, *Pediococcus acidilactici* and pathogenic microorganisms; *Listeria monocytogenes*, *Salmonella enterica*, Vancomycin-Resistant Enterococci (VRE) and *Escherichia coli*. The combination of postbiotic and inulin had higher ( $P < 0.05$ ) MAU than postbiotic alone against all indicator organisms except *P. acidilactici*, and *E. coli*. The RI11 + 0.8% Inulin, RG14 + 0.8% Inulin and RG14 + 0% Inulin had significantly ( $P < 0.05$ ) higher MAU/mL against *P. acidilactici* than other treatments. The RI11 + 0.8% Inulin and RG14 + 0.4% Inulin had a significantly ( $P < 0.05$ ) higher MAU/mL against VRE. The MAU/mL against *L. monocytogenes* was greater in RI11 + 1.0% Inulin, RI11 + 0.6% Inulin and RI11 + 0.8% Inulin. The combinations of RS5 + 1.0% Inulin, RS5 + 0.8% Inulin and RS5 + 0.6% Inulin had greater MAU/mL

against *S. enterica*; whereas in *E. coli*, the inhibitory activity had higher activity that can only be found in RS5 + 0.8% Inulin. The combination of postbiotics and inulin had higher optical density and lower pH, which corresponds to increased inhibitory activity against indicator organisms. The results of this study showed that postbiotics and inulin combinations inhibit the proliferation of pathogenic bacteria. Four combinations of postbiotic and inulin with the highest MAU were used as a feed additive in the second experiment.

The second experiment examined the influence of postbiotics on growth performance, apparent ileal digestibility (AID), meat quality and gene expression in broiler chickens. A total of 280, one day-old male, Cobb broiler chicks were randomly assigned to 8 treatment groups. The treatments include; basal diet (negative control), basal diet+ neomycin and oxytetracycline (positive control), (T1) basal diet+0.3% postbiotic RI11, (T2) basal diet+0.3% postbiotic RG14, (T3) basal diet+0.3% postbiotic RI11+0.8% inulin, (T4) basal diet+0.3% postbiotic RI11+1.0% inulin, (T5) basal diet+0.3% postbiotic RG14+0.8% inulin, and (T6) basal diet+0.3% postbiotic RG14+1.0% inulin. Birds fed postbiotic and inulin combinations had greater ( $P < 0.05$ ) final body weight, body weight gain and feed efficiency compared with the control birds. Similarly, supplementation of postbiotic and inulin combinations increased ( $P < 0.05$ ) the population of faecal lactic acid bacteria and villi height of small intestine and lowered faecal *Enterobacteriaceae*. Birds fed postbiotics and inulin combinations had lower ( $P < 0.05$ ) drip loss and higher ( $P < 0.05$ ) lightness of *Pectoralis major* muscle compared with the control birds. Birds fed diets supplemented with postbiotic and inulin combinations had greater ( $P < 0.05$ ) percentage of tibia bone Ca and P, faecal acetic acid, propionic acid and total organic acid compared with the control birds. The liver of birds fed T4 and T6 had higher Insulin-like growth factor 1 (IGF-I) expression compared with other treatments while T6 had higher growth hormone receptor (GHR) mRNA expression compared with other treatments. The supplementation of postbiotics and inulin combination had beneficial effect on total body weight, feed efficiency, mucosal architecture, and expression of IGF-I and GHR mRNA in broiler chickens. The third experiment examined the effect of graded levels of RG14 and 1% inulin on growth performance, meat quality and gene expression in broiler chickens. A total of 216, one day old male Cobb chicks were randomly assigned to six dietary treatments. The treatment were basal diet (negative control), basal diet+ neomycin and oxytetracycline (positive control), (T1) basal diet+0.15% postbiotic RG14+1.0% inulin (T2) basal diet+0.3% postbiotic RG14+1.0% inulin (T3) basal diet+0.45% postbiotic RG14+1.0% inulin, (T4) basal diet+0.6% postbiotic RG14+1.0% inulin. Supplementation of 0.15 and 0.45 % RG14 with 1% inulin improved final body weight, weight gain and meat quality of broiler chickens compared with other treatments. The supplementation of postbiotic and inulin combinations increased the concentration of acetic acid, ileal cytokine expression, plasma immunoglobulin IgG and IgM, and population of total bacteria and beneficial bacteria and reduced the population of *Enterobacteria* and *E. coli* compared with the control diet. The combination of 0.15% RG14 with 1% inulin is potential replacement for antibiotic growth promoter (AGP) in the poultry industry.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

**KESAN MAKANAN TAMBAHAN POSBIOTIK DAN INSULIN KE ATAS PRESTASI TUMBESARAN, MORFOLOGI USUS, EKSPRESI GEN DAN CIRI-CIRI TINJA AYAM PEDAGING.**

Oleh

**KARWAN YASEEN KAREEM**

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Penggalak Tumbesaran Antibiotik (AGP) diakui berkesan dalam memodulasi mikroflora usus dan menggalakkan prestasi tumbesaran ayam. Walau bagaimanapun, penggunaan AGP yang berpanjangan boleh menyebabkan rintangan antibiotik dan kesan sampingan, yang mampu membahayakan kesihatan ayam dan manusia yang memakannya. Prebiotik, probiotik dan kombinasinya merupakan pengganti AGP yang berpotensi. Namun, probiotik komersil seringkali gagal memenuhi keupayaan yang diharapkan dan keberkesannya adalah tidak menentu. Tambahan pula, penggunaan probiotik boleh membawa kepada berlakunya rintangan gen antibiotik yang boleh berpindah sesama organisma. Ini menunjukkan bahawa perlunya untuk mencari alternatif kepada probiotik yang selamat dan sesuai. Posbiotik yang merupakan suatu metabolit bagi probiotik menunjukkan kesan probiotik tanpa kehadiran sel-sel hidup. Meskipun posbiotik dan prebiotik adalah berkesan, kesan sinergi di antara kombinasi prebiotik dan posbiotik belum pernah diperjelaskan. Kajian ini menguji pengaruh posbiotik yang dihasilkan oleh strain *Lactobacillus plantarum*, dan kombinasi inulin ke atas prestasi tumbesaran, morfologi usus, kualiti daging, ekspresi gen dan ciri-ciri tinja pada ayam pedaging. Kajian ini dibahagikan kepada tiga eksperimen. Eksperimen pertama mengkaji aktiviti perencatan posbiotik yang dihasilkan oleh strain *Lactobacillus plantarum* menggunakan media yang dibentuk semula dan ditambah dengan inulin. Posbiotik telah dihasilkan dari enam strain *L. plantarum* (RG11, RG14, RI11, UL4, TL1 and RS5) menggunakan media yang dibentuk semula dengan aras inulin yang berbeza (0, 0.2, 0.4, 0.6, 0.8, dan 1.0) menghasilkan sejumlah 36 kombinasi. Aktiviti Perencatan Terubah suai (MAU) bagi posbiotik yang dihasilkan telah diuji dengan mikroorganisma indikator, *Pediococcus acidilactici* dan mikroorganisma patogen; *Listeria monocytogenes*, *Salmonella enterica*, Vancomycin-Resistant Enterococci (VRE) dan *Escherichia coli*. Kombinasi di antara posbiotik dan inulin menunjukkan MAU yang lebih tinggi berbanding posbiotik melawan kesemua organisma indikator secara tunggal, kecuali *P. acidilactici*, dan *E. coli*. Kombinasi RI11 + 0.8% Inulin, RG14 + 0.8% Inulin dan RG14 + 0% Inulin mempunyai ( $P < 0.05$ ) MAU/mL yang ketara lebih tinggi melawan *P. acidilactici* berbanding rawatan-rawatan lain. RI11 + 0.8% Inulin RG14 + 0.4% Inulin menunjukkan ( $P < 0.05$ )

MAU/mL yang ketara lebih tinggi melawan VRE. MAU/mL melawan *L. monocytogenes* adalah lebih tinggi dalam RI11 + 1.0% Inulin, RI11 + 0.6% Inulin dan RI11 + 0.8% Inulin. Kombinasi RS5 + 1.0% Inulin, RS5 + 0.8% Inulin dan RS5 + 0.6% Inulin mempunyai MAU/mL yang lebih tinggi secara ketara melawan *S. enterica*; manakala dalam *E. coli*, aktiviti perencatan menunjukkan aktiviti yang lebih tinggi yang hanya boleh ditemukan dalam RS5 + 0.8% Inulin. Kombinasi di antara posbiotik dan inulin mempunyai ketumpatan optik yang lebih tinggi dan pH yang lebih rendah, yang merujuk kepada peningkatan aktiviti perencatan melawan organisma indikator. Keputusan kajian ini menunjukkan bahawa penambahan posbiotik dan inulin merencatkan pembiakan bakteria patogenik. Empat kombinasi posbiotik dan inulin dengan MAU tertinggi telah digunakan sebagai additif makanan dalam eksperimen kedua. Eksperimen kedua menguji pengaruh posbiotik ke atas prestasi tumbesaran, kebolehcernaan ketara ileum(AID), kualiti daging dan ekspresi gen dalam ayam pedaging. Sejumlah 280 anak ayam pedaging Cobb yang berumur sehari, telah dibahagikan kepada 8 kumpulan rawatan secara rawak. Rawatan-rawatannya termasuklah; diet asas(kawalan negatif), diet asas+ neomisin dan oksitetrasiklin (kawalan positif), (T1) diet asas+0.3% posbiotik RI11, (T2) diet asas+0.3% posbiotik RG14, (T3) diet asas+0.3% posbiotik RI11+0.8% inulin, (T4) diet asas+0.3% posbiotik RI11+1.0% inulin, (T5) diet asas+0.3% posbiotik RG14+0.8% inulin, dan (T6) diet asas+0.3% posbiotik RG14+1.0% inulin. Makanan ayam dengan kombinasi posbiotik dan inulin menunjukkan ( $P < 0.05$ ) berat badan akhir, penambahan berat badan dan keberkesanan makanan yang lebih tinggi berbanding ayam kawalan. Demikian juga, penambahan posbiotik inulin telah meningkatkan ( $P < 0.05$ ) populasi asid laktik yang dihasilkan bakteria dalam tinja dan ketinggian vilus dalam usus kecil dan menurunkan *Enterobacteriaceae* dalam tinja. Ayam yang diberi makan kombinasi posbiotik dan inulin mempunyai kehilangan titis yang lebih rendah ( $P < 0.05$ ) dan keringanan otot *Pectoralis major* yang lebih tinggi ( $P < 0.05$ ) berbanding ayam kawalan. Ayam yang memakan diet yang ditambahkan dengan kombinasi posbiotik dan inulin mempunyai perstusan Ca dan P dalam tulang tibia, asid asetik dalam tinja, asid propionik dan jumlah asid organik yang lebih tinggi ( $P < 0.05$ ) berbanding ayam kawalan. Hati bagi ayam yang telah diberi makan T4 dan T6 mempunyai ekspresi faktor tumbesaran seiras-insulin 1 (IGF-I) berbanding rawatan-rawatan lain manakala T6 mempunyai ekspresi reseptor hormon tumbesaran (GHR) mRNA yang lebih tinggi berbanding rawatan-rawatan lain. Penambahan kombinasi posbiotik dan insulin mempunyai kesan berfaedah ke atas berat badan keseluruhan, keberkesanan makanan, seni bina mukosa, dan ekspresi IGF-I dan GHR mRNA dalam ayam pedaging. Eksperimen ketiga mengkaji kesan aras-aras RG14 dan 1% inulin berperingkat ke atas prestasi tumbesara, kualiti makanan dan ekspresi gen dalam ayam pedaging. Sejumlah 216, ayam jantan baka Cobb yang berusia sehari telah dibahagikan secara rawak kepada enam rawatan pemakanan. Rawatan tersebut adalah diet asal (kawalan negatif), diet asas+ neomisin dan oksitetrasiklin (kawalan positif), (T1) diet asas+0.15% postiotik RG14+1.0% inulin (T2) diet asas+0.3% posbiotik RG14+1.0% inulin (T3) diet asas+0.45% posbiotik RG14+1.0% inulin, (T4) diet asas+0.6% posbiotik RG14+1.0% inulin. Penambahan 0.15 dan 0.45 % RG14 dengan 1% inulin telah meningkatkan berat badan akhir, penambahan berat badan dan kualiti daging ayam pedaging berbanding rawatan-rawatan lain. Penambahan kombinasi posbiotik dan inulin telah meningkatkan kepekatan asid asetik, ekspresi sitokin ileum, immunoglobulin plasma IgG dan IgM, dan populasi jumlah bakteria dan bakteria berfaedah dan menurunkan populasi Enterobakteria dan bakteria *E. coli* berbanding diet kawalan. Kombinasi 0.15% RG14 dengan 1% inulin telah dikenal pasti sebagai



pengganti bagi Penggalak Tumbesaran Antibiotik (AGP) yang berpotensi dalam industri poltri.



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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirements for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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## LIST OF ABBREVIATIONS

μL	Microliter
μm	Micrometer
a*	Yellowness
AGP	Antibiotic growth promoter
AID	Apparent ileal digestibility
ATP	Adenosine triphosphate
b*	Redness
BHA	Butylated hydroxyanisole
BMC	Bone mineral density
BW	Body weight
BWG	Body weight gain
cDNA	Complementary-DNA
CFU	Colony forming units
Cm	Centimeter
CP	Crude protein
CPO	Crude palm oil
CRD	Complete randomized design
CT	Cycle threshold
DC	Dendritic cell
DM	Dry matter
DNA	Deoxyribonucleic acid
DP	Degree of polymerization
EE	Ether extract
ENT	Enterobacteriaceae
FCR	Feed conversion ratio
FI	Feed intake
FOS	Fructooligosaccharide
G	Gram
GARS	Generally recognized as safe
GHR	Growth hormone receptor
GI	Gastrointestinal
GIT	Gastrointestinal tract
GLM	General linear model
GOS	Galactooligosaccharide
H <sub>2</sub> O <sub>2</sub>	Hydrogen peroxide
H <sub>2</sub> SO <sub>4</sub>	Sulphuric acid
HDL-C	High density lipoprotein cholesterol
IFN-γ	Interferon gamma
IgA	Immunoglobulin A
IGF-I	Insulin-like growth factor 1
IgG	Immunoglobulin G
IgM	Immunoglobulin M
IL	Interleukin
IMO	Isomaltooligosaccharide
kg	Kilogram
L	Liter

L*	Lightness
LAB	Lactic acid bacteria
LDL-C	Low-density lipoprotein cholesterol
M	Molar
MAU	Modified arbitrary unit
mL	Milliliter
Mm	Millimeter
mM	Millimolar
MOS	Mannan oligosaccharide
mRNA	Messenger ribonucleic acid
MRS	de-Mann Rogosa Sharp
N <sup>-</sup> CL	Sodium chloride
OD	Optical density
OF	Oligofructose
PK <sub>a</sub>	The dissociation constant
PSE	Pale, soft, exudative
RNA	Ribonucleic acid
Rpm	Rounds per minute
SAS	Statistical analysis system
SCFA	Short chain fatty acid
SOS	Soya oligosaccharide
TCHOL	Total cholesterol
TG	Triglyceride
Th1	T-helper 1
TiO <sub>2</sub>	Titanium dioxide
TLR	Toll-like receptor
v/v	Volume versus volume
VFA	Volatile fatty acid
VLDL-C	Very low-density lipoprotein cholesterol
VRE	Vancomycin-Resistant Enterococci
w/v	Weight versus volume
WHC	Water holding capacity
XOS	Xylooligosaccharide

## CHAPTRE ONE

### GENERAL INTRODUCTION

Poultry production is an active and dynamic industry that is central to the wellbeing of many people and an indispensable part of the economy in many countries (FAO, 2014). The main targets of poultry production are high growth rate, optimum performance and feed efficiency at least cost. Meeting these targets is premised solely on the genetic potential of the birds, environmental conditions, quality of the diets, and outbreak of diseases. Aside the aforementioned-factors, gut health has recently been the subject of intense studies in poultry production (Rinttilä and Apajalahti, 2013).

The gut plays a vital role in mediating the uptake and use of nutrients by the animals and a major site of potential exposure to environmental pathogens (Yegani and Korver, 2008). Thus, a healthy and well-functioning gut is germane for optimum performance and health of birds. When the gut function and health are impaired, digestion and absorption of nutrients are affected and the health and performance of birds will be compromised (Sugiharto, 2014).

Besides being responsible for the absorption of nutrients from the lumen, intestinal mucosa of broiler chicken plays an important role in providing an effective barrier between the hostile luminal content and the host internal tissues. In this respect, intestinal mucosa is an important determinant of gut health and performance of chicken (Rinttilä and Apajalahti, 2013).

Among the factors responsible for the gut health and performance of chicken, commensal microbiota in the gut seem to have pivotal roles as they help to direct the development of gut structure and morphology, modulate the immune responses, offer protection from luminal pathogens and aid the digestion and utilization of nutrients (Sugiharto, 2014). In order to support the intestinal mucosal barrier functions, the dynamic balance between the mucus layer, epithelial cells, microbiota and immune cells in the intestine is of importance (Schenk and Mueller, 2008).

Various factors associated with diet and infectious disease agents affect this dynamic balance, and subsequently affecting the health status and production performance of the chicken (Yegani and Korver, 2008). Decreased incidence of disease may be related to changes in immune regulation through cytokine secretion. Thus, there is evidence of correlation between the composition of the colonizing microbiota and variations in immunity (O'Hara and Shanahan, 2006). The dependence of nutritional and growth hormones on hepatic IGF-I production has been demonstrated (Moriyama, 1995; Shamblott *et al.*, 1995). Moreover, amongst the genes influencing growth, IGF-I has been demonstrated as an indicator of growth rate in chicken (Beccavin *et al.*, 2001; Jones and Clemmons, 1995). The overall nutritional status of the animal modulates the ability of hepatic tissue to respond to growth hormones (Beckman, 2011).

Antibiotics growth promoters (AGP) have been widely used in the poultry industry for decades to maintain the balance of ecosystem in the gut and to promote growth performance of birds (Huyghebaert *et al.*, 2011). Nonetheless, the usage of antibiotics as feed additives for long periods in poultry diets can lead to antibiotic resistance (Shazali *et al.*, 2014) and high residue levels in animal products.

Genes encoding for this resistance have the potential to be transferred to other formerly susceptible bacteria, therefore posing a threat to animal and human health (Montagne *et al.*, 2003). The use of AGP in poultry production has been banned or restricted in many countries due to public concern (Ohimain and Ofongo, 2012). The need to optimize gut health and growth performance in birds to produce a safe and quality products to consumers has created an impetus to find alternatives for the usage of antibiotics in poultry production (Kleter and Marvin, 2009; Sugiharto, 2014). Some of these additives are prebiotics, probiotics, symbiotics and postbiotics (metabolic products by probiotic) (Alloui *et al.*, 2013; Loh *et al.*, 2014).

Prebiotics such as inulin (Rebolé *et al.*, 2010) are effective in controlling pathogens such as *Escherichia coli* and *Salmonella* and in stimulating the growth of *Bifidobacteria* and *Lactobacilli*, thus promoting health and performance of animals (Bogusławska-Tryk *et al.*, 2012; Jung *et al.*, 2008). Probiotics are beneficial bacteria capable of colonizing the host digestive system, increasing the natural flora and preventing colonization of pathogenic organisms (Loh *et al.*, 2014). In spite of beneficial effects of probiotics, the major problem with their application is that some probiotics have antibiotic resistance genes, especially those encoded by plasmids, which can be transferred between organisms (Marteau and Shanahan, 2003). This justifies the need to search for suitable and safe alternative to probiotics.

As a substitute to probiotics, metabolite products synthesized from probiotic known as postbiotics could be used. Postbiotics exhibit probiotic effects without living cells (Loh *et al.*, 2010; Thanh *et al.*, 2009; Thu *et al.*, 2011). The efficacy of metabolites produced from *L. plantarum* to inhibit the proliferation of pathogenic bacteria such as *Listeria monocytogenes*, *Salmonella typhimurium*, *Escherichia coli* and Vancomycin Resistant *Enterococci* has been documented (Loh *et al.*, 2010). Furthermore, improvements in growth performance, faecal lactic acid bacteria and villus height were observed when metabolite combinations were added to the feed of broilers (Thanh *et al.*, 2009), laying hens (Choe *et al.*, 2012) and pigs (Thu *et al.*, 2011).

Synbiotics, the combination of prebiotics and probiotics may possibly produce greater beneficial effects on gut health than individual application of probiotics and prebiotics (Abdel-Fattah and Fararh, 2009; Mookiah *et al.*, 2014; Hamasalim, 2016). Synbiotics exerts synergistic and/or additive effects in the improvement of gut health and growth performance in livestock (Nekoubin and Sudagar, 2012; Al-Baadani *et al.*, 2016; Hamasalim, 2016).



Despite the beneficial roles of synbiotics, there is a paucity of information on the synergistic effects of the combination of postbiotics and prebiotics on gut health, growth performance, meat quality, gene expression and faecal characteristics of broiler chickens. Thus, the current study was initiated to examine such effects with the following hypothesis and objectives.

#### Hypothesis statements

1. The combinations of inulin and postbiotics produced by *Lactobacillus plantarum* would exhibit inhibitory activity against pathogenic bacteria.
2. The combinations of postbiotics and inulin would alter gut morphology, promote nutrient digestibility, growth performance and meat quality in broiler chickens.
3. Different levels of postbiotics and inulin would alter faecal bacteria profile, gene expression and cytokine expression in broiler chickens.

#### Objectives

The main objective of this research was to examine the effects of combinations of postbiotics and inulin on gut health, growth performance, gene expression and faecal characteristics of broiler chickens.

The specific objectives of this study were:

1. To determine the modified inhibitory activity of postbiotics produced by *Lactobacillus plantarum* RG11, RG14, RI11, UL4, TL1 and RS5 using reconstituted media supplemented with different levels of inulin.
2. To examine the impact of dietary postbiotics and inulin combinations on gut morphology, nutrient digestibility, faecal bacteria, growth performance, meat quality and hepatic IGF-I and GHR mRNA expressions in broiler chickens.
3. To determine the effect of feeding different levels of postbiotic RG14 and 1% inulin on growth performance, gut morphology, faecal bacteria, and cytokine expression in broiler chickens.

## REFERENCES

- Abdel-Fattah, F. A., & Fararh, K. M. (2009). Effect of Dietary Supplementation of Probiotic, Prebiotic and Synbiotic on Performance, Carcass Characteristics, Blood Picture and Some Biochemical Parameters in Broiler Chickens. *Benha Veterinary Medicine Journal*, *19* (2): 9-23.
- Abdel-Raheem, S. M., & Abd-Allah, S. M. (2011). The effect of single or combined dietary supplementation of mannan oligosaccharide and probiotics on performance and slaughter characteristics of broilers. *International Journal of Poultry Science*, *10*, 854-862.
- Abdel-Raheem, S. M., Abd-Allah, S. M., & Hassanein, K. M. (2012). The effects of prebiotic, probiotic and synbiotic supplementation on intestinal microbial ecology and histomorphology of broiler chickens. *International Journal for Agro Veterinary and Medical Sciences*, *6*, 277-289.
- Abdulla, N. R., Mohd Zamri, A. N., Sabow, A. B., Kareem, K. Y., Nurhazirah, S., Ling, F. H., & Chwen Loh, T. (2015). Physico-chemical properties of breast muscle in broiler chickens fed probiotics, antibiotics or antibiotic-probiotic mix. *Journal of Applied Animal Research*, 1-7. DOI: 10.1080/09712119.2015.1124330
- Abul, K. A. (2006). Properties and Overview of Immune Response. Page 4-11. In : Cellular and Molecular Immunology. 5th Edition. K. A. Abul, A. H. Lichman, and S. Pilli, eds. Philadelphia, PA. Saunders Elsevier.
- Adams, M. (1990). Topical aspects of fermented foods. *Trends in Food Science and Technology*, *1*, 140-144.
- Adams, M., & Hall, C. (1988). Growth inhibition of food-borne pathogens by lactic and acetic acids and their mixtures. *International Journal of Food Science and Technology*, *23*, 287-292.
- Adeyemi, K. D., & Sazili, A. Q. (2014). Efficacy of Carcass Electrical Stimulation in Meat Quality Enhancement: A Review. *Asian-Australasian Journal of Animal Sciences*, *27* (3), 447.
- Adil, S., & Magray, S. (2012). Impact and manipulation of gut microflora in poultry: a review. *Journal of Animal and Veterinary Advances*, *11*, 873-877.
- Adil, S., Banday, T., Ahmad Bhat, G., Salahuddin, M., Raquib, M., & Shanaz, S. (2011). Response of broiler chicken to dietary supplementation of organic acids. *Journal of Central European Agriculture*, *12* (3), 498-508.
- Adil, S., Banday, T., Bhat, G.A., Mir, M.S., & Rehman, M., 2010. Effect of dietary supplementation of organic acids on performance, intestinal histomorphology, and serum biochemistry of broiler chicken. *Veterinary Medicine International* **2010**, 1-7

- Ahmad, I. (2006). Effect of probiotics on broilers performance. *International Journal of Poultry Science*, **5**, 593-597.
- Ahmed, A.M., & Hamed, D.M. (2012). Improvement of chicken meat quality using probiotics feed supplement. *Journal of Food Processing and Technology* 2012, **3**: 10
- Akoy, R. A. M., 2015. The effects of probiotics, prebiotics and synbiotics on gut flora, immune function and blood characteristics of broilers. PhD Thesis, University of Plymouth. UK
- Aksu, M. I., Karaoglu, M., Esenbuga, N., Kaya, M., Macit, M., & Ockerman, H. (2005). Effect of a dietary probiotic on some quality characteristics of raw broiler drumsticks and breast meat. *Journal of Muscle Foods* **16**, 306-317.
- Al-Baadani, H. H., Abudabos, A. M., Al-Mufarrej, S. I., & Alzawqari, M. (2016). Effects of dietary inclusion of probiotics, prebiotics and synbiotics on intestinal histological changes in challenged broiler chickens. *South African Journal of Animal Science*, **46**(2), 157-165.
- Alexopoulos, C., Georgoulakis, I., Tzivara, A., Kyriakis, C., Govaris, A., & Kyriakis, S. (2004). Field Evaluation of the Effect of a Probiotic containing *Bacillus licheniformis* and *Bacillus subtilis* Spores on the Health Status, Performance, and Carcass Quality of Grower and Finisher Pigs. *Journal of Veterinary Medicine Series A*, **51**, 306-312.
- Al-Ghazzewi, F. H., & Tester, R. F. (2012). Efficacy of cellulase and mannanase hydrolysates of konjac glucomannan to promote the growth of lactic acid bacteria. *Journal of the Science of Food and Agriculture*, **92** (11), 2394-2396.
- Ali, F. H. (2010). " Probiotics Feed Supplement" to Improve Quality of Broiler Chicken Carcasses. *World Journal of Dairy & Food Sciences*, **5**, 93-99.
- Alkhalaf, A., Alhaj, M., & Al-Homidan, I. (2010). Influence of probiotic supplementation on immune response of broiler chicks. *Egyptian Poultry Science*, **30**, 271-280.
- Allen, C., Fletcher, D., Northcutt, J., & Russell, S. (1998). The relationship of broiler breast color to meat quality and shelf-life. *Poultry Science*, **77**, 361-366.
- Alloui, M. N., Szczurek, W., & Świątkiewicz, S. (2013). The Usefulness of Prebiotics and Probiotics in Modern Poultry Nutrition: a Review/Przydatność prebiotyków i probiotyków w nowoczesnym żywieniu drobiu–przeгляд. *Annals of Animal Science*, **13** (1), 17-32.
- Al-Sheraji, S. H., Ismail, A., Manap, M. Y., Mustafa, S., Yusof, R. M., & Hassan, F. A. (2013). Prebiotics as functional foods: A review. *Journal of Functional Foods*, **5** (4), 1542-1553.
- Alvarado, C., & Sams, A. (2004). Early postmortem injection and tumble marination effects on broiler breast meat tenderness. *Poultry Science*, **83**, 1035-1038.

- Alvarado, C., Richards, M., O'Keefe, S., & Wang, H. (2007). The effect of blood removal on oxidation and shelf life of broiler breast meat. *Poultry science*, **86**, 156-161.
- Alvarez-Olmos, M.I., & Oberhelman, R.A. (2001). Probiotic agents and infectious diseases: a modern perspective on a traditional therapy. *Clinical Infectious Diseases*, **32**, 1567-1576.
- Alzueta C., Rodriguez M. L., Ortiz L. T., Rebole A., & Trevino J. (2010). Effects of inulin on growth performance, nutrient digestibility and metabolisable energy in broiler chickens. *British Poultry Science*, **51**, 393-398
- Amit-Romach, E., Sklan, D., & Uni, Z. (2004). Microflora ecology of the chicken intestine using 16S ribosomal DNA primers. *Poultry Science*, **83**, 1093-1098.
- Andino, A., Zhang, N., Diaz-Sanchez, S., Yard, C., Pendleton, S., & Hanning, I. (2014). Characterization and specificity of probiotics to prevent salmonella infection in mice. *Functional Foods in Health and Disease*, **4**, 370-380.
- AOAC, (1995). Association of Official Analytical Chemists. Washington, DC.
- Apajalahti, J., & Kettunen, A. (2006). Microbes of the chicken gastrointestinal tract. *Avian Gut Function in Health and Disease*, **8**, 124-137.
- Apata, D. (2008). Growth performance, nutrient digestibility and immune response of broiler chicks fed diets supplemented with a culture of *Lactobacillus bulgaricus*. *Journal of the Science of Food and Agriculture*, **88**, 1253-1258.
- Aristides, L. G., Paião, F. G., Murate, L. S., Oba, A., & Shimokomaki, M. (2012). The Effects of Biotic Additives on Growth Performance and Meat Qualities in Broiler Chickens. *International Journal of Poultry Science*, **11**, 599-604.
- Aryana, K. J., & McGrew, P. (2007). Quality attributes of yogurt with *Lactobacillus casei* and various prebiotics. *LWT-Food Science and Technology*, **40**, 1808-1814.
- Ashayerizadeh, A., Dabiri, N., Ashayerizadeh, O., Mirzadeh, K., Roshanfekar, & H., Mamooee, M. (2009). Effect of dietary antibiotic, probiotic and prebiotic as growth promoters, on growth performance, carcass characteristics and hematological indices of broiler chickens. *Pakistan Journal of Biological Science*, **12**, 52-57.
- Awad, W. A., Ghareeb, K., Abdel-Raheem, S., & Böhm, J. (2009). Effects of dietary inclusion of probiotic and synbiotic on growth performance, organ weights, and intestinal histomorphology of broiler chickens. *Poultry Science*, **88** (1), 49-56.
- Bahman, N., J. B. Liang, & M. F. Jahromi. (2012). Correlation coefficients between different methods of expression of bacterial quantification using real time PCR. *International Journal of Molecular Science*, **13**:2119-2132.

- Bai, A. P., Ouyang, Q., Zhang, W., Wang, C. H., & Li, S. F. (2004). Probiotics inhibit TNF-alpha-induced interleukin-8 secretion of HT29 cells. *World Journal of Gastroenterology*, **10**, 455-457.
- Bai, S., Wu, A., Ding, X., Lei, Y., Bai, J., Zhang, K., & Chio, J. (2013). Effects of probiotic-supplemented diets on growth performance and intestinal immune characteristics of broiler chickens. *Poultry Science*, **92**, 663-670.
- Barbut, S. (1993). Colour measurements for evaluating the pale soft exudative (PSE) occurrence in turkey meat. *Food Research International*, **26**, 39-43.
- Barnett, E., & Nordin, B. (1960). The radiological diagnosis of osteoporosis: a new approach. *Clinical Radiology* **11**, 166-174.
- Bartosch, S., Fite, A., Macfarlane, G. T., & McMurdo, M. E. (2004). Characterization of bacterial communities in feces from healthy elderly volunteers and hospitalized elderly patients by using real-time PCR and effects of antibiotic treatment on the fecal microbiota. *Applied and Environmental Microbiology*, **70** (6), 3575-3581.
- Battula, V., Schilling, M., Vizzier-Thaxton, Y., Behrends, J., Williams, J., & Schmidt, T. (2008). The effects of low-atmosphere stunning and deboning time on broiler breast meat quality. *Poultry Science*, **87**, 1202-1210.
- Baurhoo, B., Ferket, P., & Zhao, X. (2009). Effects of diets containing different concentrations of mannanoligosaccharide or antibiotics on growth performance, intestinal development, cecal and litter microbial populations, and carcass parameters of broilers. *Poultry Science*, **88**, 2262-2272.
- Baurhoo, B., Phillip, L., & Ruiz-Feria, C. (2007). Effects of purified lignin and mannan oligosaccharides on intestinal integrity and microbial populations in the ceca and litter of broiler chickens. *Poultry Science*, **86**, 1070-1078.
- Beccavin, C., Chevalier, B., Cogburn, L., Simon, J., & Duclos, M. (2001). Insulin-like growth factors and body growth in chickens divergently selected for high or low growth rate. *Journal of Endocrinology*, **168**, 297-306.
- Beckman, B. R. (2011). Perspectives on concordant and discordant relations between insulin-like growth factor 1 (IGF1) and growth in fishes. *General and Comparative Endocrinology*, **170**, 233-252.
- Beckman, B. R., Larsen, D. A., Moriyama, S., Lee-Pawlak, B., & Dickhoff, W. W. (1998). Insulin-like growth factor-I and environmental modulation of growth during smoltification of spring chinook salmon (*Oncorhynchus tshawytscha*). *General and Comparative Endocrinology*, **109**, 325-335.
- Beckman, B. R., Shimizu, M. G., B. A., Parkins, P. J., & Cooper, K. A. (2004). The effect of temperature change on the relations among plasma IGF-1, 41-kDa IGFBP, and growth rate in postsmolt coho salmon. *Aquaculture*, **241**, 601-619.

- Berndt, A., Wilhelm, A., Jugert, C., Pieper, J., Sachse, K., & Methner, U. (2007). Chicken cecum immune response to *Salmonella enterica* serovars of different levels of invasiveness. *Infection and Immunity*, **75**, 5993-6007.
- Beski, S. S. M., & Al-Sardary, S. Y. T. (2015). Effects of dietary supplementation of probiotic and synbiotic on broiler chickens hematology and intestinal integrity. *International Journal of Poultry Science*, **14** (1), 31-36.
- Bielecka, M., Biedrzycka, E., & Majkowska, A. (2002). Selection of probiotics and prebiotics for synbiotics and confirmation of their in vivo effectiveness. *Food Research International*, **35**, 125-131.
- Biggs P., Parsons C. M., & Fahey G. C. (2007). Effects of Several oligosaccharides on growth performance, nutrient digestibilities, and caecal microbial populations in young chicks. *Poultry Science*, **86**, 2327-2336
- Bjerrum, L., Engberg, R., Leser, T. D., Jensen, B. B., Finster, K., & Pedersen, K. (2006). Microbial community composition of the ileum and cecum of broiler chickens as revealed by molecular and culture-based techniques. *Poultry Science*, **85**, 1151-1164.
- Bogusławska-Tryk, M., Piotrowska, A., & Burlikowska, K. (2012). Dietary fructans and their potential beneficial influence on health and performance parameters in broiler chickens. *Journal of Central European Agriculture*, **13** (2), 272-291.
- Bouhnik, Y., Raskine, L., Simoneau, G., Vicaut, E., Neut, C., Flourié, B., Brouns, F., & Bornet, F. R. (2004). The capacity of nondigestible carbohydrates to stimulate fecal bifidobacteria in healthy humans: a double-blind, randomized, placebo-controlled, parallel-group, dose-response relation study. *The American Journal of Clinical Nutrition*, **80**, 1658-1664.
- Boulanger, N., Bulet, P., & Lowenberger, C. (2006). Antimicrobial peptides in the interactions between insects and flagellate parasites. *Trends Parasitology*, **22**, 262-268.
- Bowdish, D. M., Davidson, D. J., & Hancock, R. (2005). A re-evaluation of the role of host defence peptides in mammalian immunity. *Current Protein and Peptide Science*, **6**, 35-51.
- Bozkurt, M., Alçiçek, A., Çabuk, M., Küçükylmaz, K., & Çatli, A. (2009). Effect of an herbal essential oil mixture on growth, laying traits, and egg hatching characteristics of broiler breeders. *Poultry Science*, **88**, 2368-2374.
- Brashears, M.M., Amezcua, A., & Jaroni, D. (2005). Lactic acid bacteria and their uses in animal feeding to improve food safety. *Advances in Food and Nutrition Research*, **50**, 1-31.
- Breidt, F., & Fleming, H. P. (1997). Using lactic acid bacteria to improve the safety of minimally processed fruits and vegetables. *Food Technology*, **51**, 44-51.

- Brisbin, J. T., Gong, J., & Sharif, S. (2008). Interactions between commensal bacteria and the gut-associated immune system of the chicken. *Animal Health Research Reviews*, **9**, 101-110.
- Brogden, K. A. (2009). Antimicrobial peptides: pore formers or metabolic inhibitors in bacteria? *Nat Rev Microbiol* 2005;3:238–50. derived host defense peptides. *Federation of American Societies for Experimental Biology Journal*, **23**, 13–20.
- Brunser, O., & Gotteland, M. (2010). Probiotics and Prebiotics in Human Health: An Overview. *Bioactive Foods in Promoting Health: Probiotics and Prebiotics*, 73-74.
- Bry, L., Falk, P. G., Midtvedt, T., & Gordon, J. I. (1996). A model of host-microbial interactions in an open mammalian ecosystem. *Science*, **273**, 1380-1383.
- Buchanan, N., Hott, J., Cutlip, S., Rack, A., Asamer, A., & Moritz, J. (2008). The effects of a natural antibiotic alternative and a natural growth promoter feed additive on broiler performance and carcass quality. *The Journal of Applied Poultry Research*, **17**, 202-210.
- Burkholder, K., Thompson, K., Einstein, M., Applegate, T., & Patterson, J. (2008). Influence of stressors on normal intestinal microbiota, intestinal morphology, and susceptibility to Salmonella enteritidis colonization in broilers. *Poultry Science*, **87**, 1734-1741.
- Butaye, P., Devriese, L. A., & Haesebrouck, F. (2003). Antimicrobial growth promoters used in animal feed: effects of less well known antibiotics on gram-positive bacteria. *Clinical Microbiology Reviews*, **16**, 175-188.
- Carr, F. J., Chill, D., & Maida, N. (2002). The lactic acid bacteria: a literature survey. *Critical Reviews in Microbiology*, **28**, 281-370.
- Cashman, K. (2002). Probiotics and calcium bioavailability. Probiotics and prebiotics: where are we going, 149-174.
- Castanon, J. (2007). History of the use of antibiotic as growth promoters in European poultry feeds. *Poultry Science*, **86**, 2466-2471.
- Castellini, C., Mugnai, C., & Dal Bosco, A. (2002). Effect of organic production system on broiler carcass and meat quality. *Meat Science*, **60**, 219-225.
- Çataloluk, O., & Gogebakan, B. (2004). Presence of drug resistance in intestinal lactobacilli of dairy and human origin in Turkey. *FEMS Microbiology Letters*, **236**, 7-12.
- Cavitt, L., Meullenet, J., Gandhapuneni, R., Youm, G., & Owens, C. (2005). Rigor development and meat quality of large and small broilers and the use of Allo-Kramer shear, needle puncture, and razor blade shear to measure texture. *Poultry science*, **84**, 113-118.

- Cengiz, O., Koksall, B., Tatli, O., Sevim, O., Avci, H., Epikmen, T., Beyaz, D., Buyukyork, S., Boyacioglu, M., & Uner, A. (2012). Influence of dietary organic acid blend supplementation and interaction with delayed feed access after hatch on broiler growth performance and intestinal health. *Veterinary Medicine*, **57**, 515-528.
- Cetin, N., Güçlü, B., & Cetin, E. (2005). The effects of probiotic and mannanoligosaccharide on some haematological and immunological parameters in turkeys. *Journal of Veterinary Medicine Series A*, **52**, 263-267.
- Chambers, J. R., & Gong, J. (2011). The intestinal microbiota and its modulation for Salmonella control in chickens. *Food Research International*, **44**, 3149-3159.
- Charalampopolus D., & Rastall R. A. (2009). Prebiotics and probiotics science and echnology. Springer Verlag, New York, **1**, 516.
- Charalampopoulos, D., & Rastall, R. A. (2009). *Prebiotics and probiotics science and technology*. Springer, New York.
- Chartrin, P., Meteau, K., Juin, H., Bernadet, M., Guy, G., Larzul, C., Remignon, H., Mourot, J., Duclos, M., & Baéza, E. (2006). Effects of intramuscular fat levels on sensory characteristics of duck breast meat. *Poultry Science*, **85**, 914-922.
- Chee, S., Iji, P., Choct, M., Mikkelsen, L., & Kocher, A. (2010). Characterisation and response of intestinal microflora and mucins to manno-oligosaccharide and antibiotic supplementation in broiler chickens. *British Poultry Science* **51**, 368-380.
- Chen, W. L., Liang, J. B., Jahromi, M. F., Abdullah, N., Ho, Y. W., & Tufarelli, V. (2014). Enzyme Treatment Enhances Release of Prebiotic Oligosaccharides from Palm Kernel Expeller. *BioResources*, **10** (1), 196-209.
- Chen, X., Ma, Q., Tang, M., & Ji, C. (2007). Development of breast muscle and meat quality in Arbor Acres broilers, Jingxing 100 crossbred chickens and Beijing fatty chickens. *Meat Science*, **77**, 220-227.
- Chiba, L. I. (2007). Digestive physiology. *Animal Nutrition*, 29-53.
- Chichlowski, M., Croom, J., McBride, B., Daniel, L., Davis, G., & Koci, M. (2007). Direct-fed microbial PrimaLac and salinomycin modulate whole-body and intestinal oxygen consumption and intestinal mucosal cytokine production in the broiler chick. *Poultry Science*, **86**, 1100-1106.
- Chichlowski, M., Croom, J., McBride, B., Havenstein, G., & Koci, M. (2007b). Metabolic and physiological impact of probiotics or direct-fed-microbials on poultry: a brief review of current knowledge. *International Journal of Poultry Science*, **6**, 694-704.



- Cho, I. J., Lee, N. K., & Hahm, Y. T. (2009). Characterization of *Lactobacillus* spp. isolated from the feces of breast-feeding piglets. *Journal of Bioscience and Bioengineering*, **108**, 194-198.
- Choct, M. (2009). Managing gut health through nutrition. *British Poultry Science*, **50**, 9-15.
- Choe, D., Foo, H., Loh, T., Hair-Bejo, M., & Awis, Q. (2013). Inhibitory property of metabolite combinations produced from *Lactobacillus plantarum* strains. *Pertanika Journal of Tropical Agricultural Science*, **36**, 79-87.
- Choe, D., Loh, T., Foo, H., Hair-Bejo, M., & Awis, Q. (2012). Egg production, faecal pH and microbial population, small intestine morphology, and plasma and yolk cholesterol in laying hens given liquid metabolites produced by *Lactobacillus plantarum* strains. *British Poultry Science*, **53**, 106-115.
- Christensen, H. R., Frøkiær, H., & Pestka, J. J. (2002). Lactobacilli differentially modulate expression of cytokines and maturation surface markers in murine dendritic cells. *The Journal of Immunology*, **168**, 171-178.
- Cintas, L., Casaus, M., Herranz, C., Nes, I., & Hernández, P. (2001). Review: bacteriocins of lactic acid bacteria. *Food Science and Technology International* **7**, 281-305.
- Coleman, R. A., & Korver, D. R. (2004). Amino acid requirements of broilers: relationships with growth and meat quality. Proceedings of the 16th Annual Australian Poultry Science Symposium Sydney Australia.
- Coppa, G., Zampini, L., Galeazzi, T., & Gabrielli, O. (2006). Prebiotics in human milk: a review. *Digestive and Liver Disease*, **38**, S291-S294.
- Cornforth, D. (1994). Color—its basis and importance, Quality attributes and their measurement in meat, poultry and fish products. Springer, US, 34-78.
- Corzo, A., Schilling, M., Loar, R., Jackson, V., Kin, S., & Radhakrishnan, V. (2009). The effects of feeding distillers dried grains with solubles on broiler meat quality. *Poultry Science*, **88**, 432-439.
- Cotter, P. D., Hill, C., & Ross, R. P. (2005). Bacteriocins: developing innate immunity for food. *Nature Reviews Microbiology*, **3**, 777-788.
- Coudray, C., Demigné, C., & Rayssiguier, Y. (2003). Effects of dietary fibers on magnesium absorption in animals and humans. *The Journal of Nutrition*, **133**, 1-4.
- Cox, C. M., & Dalloul, R. A. (2014). Immunomodulatory role of probiotics in poultry and potential in ovo application. *Beneficial microbes*, **6**(1), 45-52.

- Crittenden, R., Martinez, N., & Playne, M. (2003). Synthesis and utilisation of folate by yoghurt starter cultures and probiotic bacteria. *International Journal of Food Microbiology*, **80**, 217-222.
- Cross, M. L., Ganner, A., Teilab, D., & Fray, L. M. (2004). Patterns of cytokine induction by gram-positive and gram-negative probiotic bacteria. *FEMS Immunology & Medical Microbiology*, **42**, 173-180.
- da Rocha, A. P., Abreu, R. D., Marques da Costa, M. D. C. M., de Oliveira, G. J. C., Albinati, R. C. B., da Paz, A. S., & Pedreira, T. M. (2010). Prebiotics, organic acids and probiotics in feed for broilers. *Journal of Animal Health and Production*, **11** (3).
- da Silva, B. R., de Freitas, V. A. A., Nascimento-Neto, L. G., Carneiro, V. A., Arruda, F. V. S., de Aguiar, A. S. W., & Teixeira, E. H. (2012). Antimicrobial peptide control of pathogenic microorganisms of the oral cavity: a review of the literature. *Peptides*, **36** (2), 315-321.
- da Silva, W. T. M., Nunes, R. V., Pozza, P. C., dos Santos Pozza, M. S., Appelt, M. D., & Eyng, C. (2011). Inulin evaluation and probiotic for broilers. *Animal Sciences*, **33** (1), 19-24.
- Dalić, D., Deschamps, A., & Richard-Forget, F. (2010). Lactic acid bacteria–Potential for control of mould growth and mycotoxins: A review. *Food Control*, **21**, 370-380.
- Dalloul, R. A., Lillehoj, H. S., Tamim, N. M., Shellem, T. A., & Doerr, J. A. (2005). Induction of local protective immunity to *Eimeria acervulina* by a *Lactobacillus*-based probiotic. *Comparative Immunology, Microbiology and Infectious Diseases*, **28**, 351-361.
- Dalloul, R., Lillehoj, H., Shellem, T., & Doerr, J. (2003). Enhanced mucosal immunity against *Eimeria acervulina* in broilers fed a *Lactobacillus*-based probiotic. *Poultry Science*, **82**, 62-66.
- Das, L., Bhaumik, E., Raychaudhuri, U., & Chakraborty, R. (2012). Role of nutraceuticals in human health. *Journal of Food Science and Technology*, **49**, 173-183.
- de Faria, D. E., Henrique, A. P. F., Neto, R. F., Medeiros, A. A., Junqueira, O. M., & de Faria Filho, D. E. (2009). Alternatives to the use of antibiotics as promoters of growth for chickens cut: 1. probiotics. *Brazilian Animal Science*, **10**(1), 18-28.
- De Preter, V., Hamer, H. M., Windey, K., & Verbeke, K. (2011). The impact of pre and/or probiotics on human colonic metabolism: Does it affect human health?. *Molecular Nutrition and Food Research*, **55**(1), 46-57.
- De Vries, M. C., Vaughan, E. E., Kleerebezem, M., & de Vos, W. M. (2006). *Lactobacillus plantarum* survival, functional and potential probiotic properties in the human intestinal tract. *International Dairy Journal*, **16**, 1018-1028.

- Denbow, M. D. (2003). Factors Affecting Meat Quality. URL (<http://www.poultryscience.org/pba/1952-2003/2000/2000%20Denbow.pdf>). Accessed on February 7th, 2010.
- Deng, W., Dong, X. F., Tong, J. M., & Zhang, Q. (2012). The probiotic *Bacillus licheniformis* ameliorates heat stress-induced impairment of egg production, gut morphology, and intestinal mucosal immunity in laying hens. *Poultry Science*, **91**, 575–582.
- Department of Standards Malaysia (2009). MS1500: 2009 (1st revision) Halal food production, preparation, handling and storage-general guideline (pp. 1–13).
- Di Giacinto, C., Marinaro, M., Sanchez, M., Strober, W., & Boirivant, M. (2005). Probiotics ameliorate recurrent Th1-mediated murine colitis by inducing IL-10 and IL-10-dependent TGF- $\beta$ -bearing regulatory cells. *The Journal of Immunology*, **174**, 3237-3246.
- Dibner, J., & Richards, J. (2005). Antibiotic growth promoters in agriculture: history and mode of action. *Poultry Science*, **84**, 634-643.
- Dibner, J., Richards, J., Kitchell, M., & Quiroz, M. (2007). Metabolic challenges and early bone development. *The Journal of Applied Poultry Research*, **16**, 126-137.
- Diop, M. B., Dubois-Dauphin, R., Tine, E., Ngom, A., Destain, J., & Thonart, P. (2007). Bacteriocin producers from traditional food products. *Biotechnology, Agronomy, society and Environnement*, **11**, 275-281.
- Doyle, J. (1979). Toxic and essential elements in bone—a review. *Journal of Animal Science*, **49**, 482-497.
- Drider, D., Fimland, G., Héchard, Y., McMullen, L. M., & Prévost, H. (2006). The continuing story of class IIa bacteriocins. *Microbiology and Molecular Biology Reviews*, **70**, 564-582.
- Dunkley, K. D., Dunkley, C. S., Njongmeta, N. L., Callaway, T. R. Hume, M. E. Kubena, L. F. Nisbet, D. J. & Ricke, S. C. (2007a). Comparison of in vitro fermentation and molecular microbial profiles of high-fiber feed substrates (HFFS) incubated with chicken cecal inocula. *Poultry Science*, **86**, 801–810.
- Dunkley, K. D., McReynolds, J. L. Hume, M. E. Dunkley, C. S. Callaway, T. R. Kubena, L. F. Nisbet, D. J. & Ricke, S. C. (2007c.) Molting in *Salmonella Enteritidis*-challenged laying hens fed alfalfa crumbles. II. Fermentation and microbial ecology response. *Poultry Science*, **86**, 2101–2109.
- Dunkley, K., Callaway, T., Chalova, V., McReynolds, J., Hume, M., Dunkley, C., Kubena, L., Nisbet, D., & Ricke, S. (2009). Foodborne *Salmonella* ecology in the avian gastrointestinal tract. *Anaerobe*, **15**, 26-35.

- Elrayeh, A. S., & Yildiz, G. (2012). Effects of inulin and beta-glucan supplementation in broiler diets on growth performance, serum cholesterol, intestinal length, and immune system. *Turkish Journal of Veterinary and Animal Sciences*, **36**, 388-394.
- Engberg, R. M., Hedemann, M. S., Leser, T., & Jensen, B. B. (2000). Effect of zinc bacitracin and salinomycin on intestinal microflora and performance of broilers. *Poultry Science*, **79**, 1311-1319.
- Falaki, M., Shargh, M. S., Dastar, B., & Zerehdaran, S. (2011). Effect of different levels of probiotic and prebiotic on performance and carcass characteristics of broiler chickens. *Journal of Animal and Veterinary Advances*, **10**, 378-384.
- Fan, Y., Croom, J., Christensen, V. L., Black, B. L., Bird, A. R., Daniel, L. R., McBride, B. W., & Eisen, E. J. (1997). Jejunal glucose uptake and oxygen consumption in turkey poult selected for rapid growth. *Poultry Science*, **76**, 1738-1745.
- FAO, IFAD, and WFP. (2014). The state of food insecurity in the world 2014. Strengthening the enabling environment for food security and nutrition. Rome: FAO.
- Farnell, M., Donoghue, A., De Los Santos, F.S., Blore, P., Hargis, B., Tellez, G., & Donoghue, D. (2006). Upregulation of oxidative burst and degranulation in chicken heterophils stimulated with probiotic bacteria. *Poultry Science*, **85**, 1900-1906.
- Faseleh Jahromi, M., Liang, J. B., Mohamad, R., Goh, Y. M., Shokryazdan, P., & Ho, Y. W. (2013). Lovastatin-enriched rice straw enhances biomass quality and suppresses ruminal methanogenesis. *BioMed Research International*, **2013**.
- Ferket, P., Parks, C., & Grimes, J. (2002). Benefits of dietary antibiotic and mannanoligosaccharide supplementation for poultry, *Multi-State Poultry Meeting*, **14**, 1-22.
- Fernandez, X., Forslid, A., & Tornberg, E. (1994). The effect of high post-mortem temperature on the development of pale, soft and exudative pork: Interaction with ultimate pH. *Meat Science*, **37**, 133-147.
- Ferreira, C. L., Salminen, S., Grzeskowiak, L., Brizuela, M. A., Sanchez, L., Carneiro, H., & Bonnet, M. (2011). Terminology concepts of probiotic and prebiotic and their role in human and animal health. *Journal of Animal Health*, **33**, 137-139.
- Fioramonti, J., Theodorou, V., & Bueno, L. (2003). Probiotics: what are they? What are their effects on gut physiology? *Best Practice and Research Clinical Gastroenterology*, **17**, 711-724.
- Fletcher, D. (1999). Broiler breast meat color variation, pH, and texture. *Poultry Science*, **78**, 1323-1327.
- Fletcher, D. (2002). Poultry meat quality. *World's Poultry Science Journal*, **58**, 131-145.

- Fletcher, D., & Smith, D. (2006). The relationship between breast muscle color variation and meat functionality, Proc. XII Eur. Poult. Conf., Verona, Italy. University Bologna, Bologna, Italy, 1-4.
- Flickinger, E., & Fahey, G. (2002). Pet food and feed applications of inulin, oligofructose and other oligosaccharides. *British Journal of Nutrition*, **87**, S297-S300.
- Flickinger, E., Schreijen, E., Patil, A., Hussein, H., Grieshop, C., Merchen, N., & Fahey, G. (2003). Nutrient digestibilities, microbial populations, and protein catabolites as affected by fructan supplementation of dog diets. *Journal of Animal Science*, **81**, 2008-2018.
- Foley, S., & Lynne, A. (2008). Food animal-associated challenges: Pathogenicity and antimicrobial resistance. *Journal of animal science*, **86**, E173-E187.
- Foligne, B., Nutten, S., Grangette, C., Dennin, V., Goudercourt, D., Poiret, S., Dewulf, J., Brassart, D., Mercenier, A., & Pot, B. (2007). Correlation between in vitro and in vivo immunomodulatory properties of lactic acid bacteria. *World Journal of Gastroenterology*, **13**, 236-43.
- Foo, H., Lim, Y., Loh, T., Saleh, N., Raha, A., & Rusul, G. (2005). Characterization of bacteriocin produced by *Lactobacillus plantarum* I-UL4 isolated from Malaysian fermented tapioca, Tapai Ubi. Proceeding of 4th NIZO Dairy Conference. Papendal, Netherland. 15-17 June. pp. 33.
- Foo, H., Loh, T., Lai, P., Lim, Y., Kufli, C., & Rusul, G. (2003a). Effects of adding *Lactobacillus plantarum* I-UL4 metabolites in drinking water of rats. *Pakistan Journal of Nutrition*, **2** (5), 283-288.
- Foo, H., Loh, T., Law, F., Lim, Y., Kufli, C., & Rusul, G. (2003b). Effects of feeding *Lactobacillus plantarum* I-UL4 isolated from Malaysian Tempeh on growth performance, faecal flora and lactic acid bacteria and plasma cholesterol concentrations in postweaning rats. *Food Science and Biotechnology*, **12**, 403-408.
- Fooks, L. J., & Gibson, G. R. (2002). In vitro investigations of the effect of probiotics and prebiotics on selected human intestinal pathogens. *FEMS Microbiology Ecology*, **39**, 67-75.
- Forshell, L. P., & Wierup, M. (2006). *Salmonella* contamination: a significant challenge to the global marketing of animal food products. *Scientific and Technical Review of International Office of Epizootics*, **25**, 541-554.
- Frahm, E., & Obst, U. (2003). Application of the fluorogenic probe technique (TaqMan PCR) to the detection of Enterococcus spp. and *Escherichia coli* in water samples. *Journal of Microbiological Methods*, **52**(1), 123-131.

- Franklin, M. A., Mathew, A. G., Vickers, J. R. & Clift, R. A. (2002). Characterization of microbial populations and volatile fatty acid concentrations in the jejunum, ileum, and cecum of pigs weaned at 17 vs 24 days of age. *Journal of Animal Science*, **80**, 2904- 2910.
- Friedewald, W. T., Levy, R. I., & Fredrickson, D. S. (1972). Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. *Clinical Chemistry*, **18**, 499-502.
- Fukata, T., Sasai, K., Miyamoto, T., & Baba, E. (1999). Inhibitory effects of competitive exclusion and fructooligosaccharide, singly and in combination, on *Salmonella* colonization of chicks. *Journal of Food Protection*, **62**, 229-233.
- Fuller, M.F., 2004. The Encyclopedia of Farm Animal Nutrition. London UK.
- Gabriel, I., Lessire, M., Mallet, S., & Guillot, J. (2006). Microflora of the digestive tract: critical factors and consequences for poultry. *World's Poultry Science Journal*, **62**, 499-511.
- Gadd, J. (1997). Life without antibiotic digestive enhancers, Biotechnology in the feed industry. Proceedings Alltechs 13th Annual Symposium, Nicholasville, Kentucky, USA, 277-291.
- Gaggia, F., Mattarelli, P., & Biavati, B. (2010). Probiotics and prebiotics in animal feeding for safe food production. *International Journal of Food Microbiology* **141**, S15-S28.
- Ganguly, S. (2013). Supplementation of prebiotics, probiotics and acids on immunity in poultry feed: a brief review. *World's Poultry Science Journal*, **69**, 639-648.
- Garcia, V., Catala-Gregori, P., Hernandez, F., Megias, M., & Madrid, J. (2007). Effect of formic acid and plant extracts on growth, nutrient digestibility, intestine mucosa morphology, and meat yield of broilers. *The Journal of Applied Poultry Research*, **16**, 555-562.
- Gaskins, H., Collier, C., & Anderson, D. (2002). Antibiotics as growth promotants: mode of action. *Animal Biotechnology*, **13**, 29-42.
- Gebbink, G., Sutton, A., Richert, B., Patterson, J., Nielsen, J., Kelly, D., Verstegen, M., Williams, B., Bosch, M., & Cobb, M. (1999). Effects of addition of fructooligosaccharide (FOS) and sugar beet pulp to weanling pig diets on performance, microflora and intestinal health. Access, 53-59.
- Ghanem, K., Badawy, I., & Abdel-Salam, A. (2004). Influence of yoghurt and probiotic yoghurt on the absorption of calcium, magnesium, iron and bone mineralization in rats. *Milchwissenschaft* **59**, 472-475.

- Ghasemi, H., & Taherpour, K. (2013). Comparative effects of probiotic, prebiotic and synbiotic supplements on performance, jejunal morphology, serum lipid profile and antibody response of broiler chicks. *Journal of Livestock Science and Technologies*, **2**, 21-28.
- Gibson, G. R., & Roberfroid, M. B. (1995). Dietary modulation of the human colonic microbiota: introducing the concept of prebiotics. *Journal of nutrition*, **125**, 1401-1412.
- Gibson, G. R., Beatty, E. R., Wang, X., & Cummings, J. H. (1995). Selective stimulation of bifidobacteria in the human colon by oligofructose and inulin. *Gastroenterology*, **108**, 975-982.
- Gibson, G. R., Probert, H. M., Van Loo, J., Rastall, R. A., & Roberfroid, M. B. (2004). Dietary modulation of the human colonic microbiota: updating the concept of prebiotics. *Nutrition Research Reviews*, **17**, 259-275.
- Gibson, G., & Wang, X. (1994). Regulatory effects of bifidobacteria on the growth of other colonic bacteria. *Journal of Applied Bacteriology*, **77**, 412-420.
- Goh, Y. S., Grant, A. J., Restif, O., McKinley, T. J., Armour, K. L., Clark, M. R., & Mastroeni, P. (2011). Human IgG isotypes and activating Fcγ receptors in the interaction of *Salmonella enterica* serovar Typhimurium with phagocytic cells. *Immunology*, **133**, 74-83.
- Gómez-Verduzco, G., Cortes-Cuevas, A., López-Coello, C., Ávila-González, E., & Nava, G. M. (2009). Dietary supplementation of mannan-oligosaccharide enhances neonatal immune responses in chickens during natural exposure to *Eimeria spp.* *Acta Veterinaria Scandinavica* **51**, 11
- Gong, J., Forster, R. J., Yu, H., Chambers, J. R., Sabour, P. M., Wheatcroft, R., & Chen, S. (2002). Diversity and phylogenetic analysis of bacteria in the mucosa of chicken ceca and comparison with bacteria in the cecal lumen. *FEMS Microbiology Letters* **208**, 1-7.
- Gong, J., Si, W., Forster, R. J., Huang, R., Yu, H., Yin, Y., Yang, C., & Han, Y. (2007). 16S rRNA gene-based analysis of mucosa-associated bacterial community and phylogeny in the chicken gastrointestinal tracts: from crops to ceca. *FEMS Microbiology Ecology*, **59**, 147-157.
- Groom, G. (1990). Factors affecting poultry meat quality. *Options Mediterraneees*, **7**, 205-210.
- Gulewicz, P., Ciesiolka, D., Frias, J., Vidal-Valverde, C., Frejnagel, S., Trojanowska, K., & Gulewicz, K. (2000). Simple method of isolation and purification of  $\alpha$ -galactosides from legumes. *Journal of Agricultural and Food Chemistry*, **48**, 3120-3123.

- Haghighi, H. R., Gong, J., Gyles, C. L., Hayes, M. A., Sanei, B., Parvizi, P., Gisavi, H., Chambers, J. R., & Sharif, S. (2005). Modulation of antibody-mediated immune response by probiotics in chickens. *Clinical and Diagnostic Laboratory Immunology*, **12**, 1387-1392.
- Hajati, H., & Rezaei, M. (2010). The application of prebiotics in poultry production. *International Journal of Poultry Science*, **9**, 298-304.
- Hamasalim, H. J. (2016). Synbiotic as Feed Additives Relating to Animal Health and Performance. *Advances in Microbiology*, **6**(4), 288-302.
- Hammes, W. P., & Hertel, C. (2006). The genera *Lactobacillus* and *Carnobacterium*. *Prokaryotes*, **4**, 320-403.
- Hancock, R., & Viola, R. (2001). The use of micro-organisms for L-ascorbic acid production: current status and future perspectives. *Applied Microbiology and Biotechnology*, **56**, 567-576.
- Havenaar, R., & Spanhaak, S. (1994). Probiotics from an immunological point of view. *Current Opinion in Biotechnology*, **5**, 320-325.
- Hayes, M., Ross, R. P., Fitzgerald, G. F., & Stanton, C. (2007a). Putting microbes to work: dairy fermentation, cell factories and bioactive peptides. Part I: overview. *Biotechnology Journal*, **2**, 426-434.
- Hayes, M., Stanton, C., Fitzgerald, G. F., & Ross, R. P. (2007b). Putting microbes to work: dairy fermentation, cell factories and bioactive peptides. Part II: bioactive peptide functions. *Biotechnology Journal*, **2**, 435-449.
- Herfel, T. M., Jacobi, S. K., Lin, X., Fellner, V., Walker, D. C., Jouni, Z. E., & Odle, J. (2011). Polydextrose enrichment of infant formula demonstrates prebiotic characteristics by altering intestinal microbiota, organic acid concentrations, and cytokine expression in suckling piglets. *The Journal of Nutrition*, **141**(12), 2139-2145.
- Hernández, F., García, V., Madrid, J., Orengo, J., Catalá, P., & Megias, M. (2006). Effect of formic acid on performance, digestibility, intestinal histomorphology and plasma metabolite levels of broiler chickens. *British Poultry Science*, **47**, 50-56.
- Hinton, M., & Linton, A. (1988). Control of *Salmonella* infections in broiler chickens by the acid treatment of their feed. *Veterinary Record*, **123**, 416-421.
- Hiroshi, Y. T. A., & Khane, R. (2004). Bacteriocins produced by lactic acid bacteria and their use for food preservation. *Tohoku Journal of Agricultural Research*, **55**, 51-55.
- Holásková, E., Galuszka, P., Frébort, I., & Öz, M. T. (2015). Antimicrobial peptide production and plant-based expression systems for medical and agricultural biotechnology. *Biotechnology Advances*, **33**(6), 1005-1023.



- Holyoak, C. D., Stratford, M., McMullin, Z., Cole, M. B., Crimmins, K., & Brown, A. J. P. (1996). Activity of the membrane H<sup>+</sup>-ATPase and optimal glycolytic flux required for rapid adaptation and growth in the presence of weak acid preservative sorbic acid. *Applied Environmental Microbiology*, **62**, 3158–3164.
- Honikel, K. O. (1998). Reference methods for the assessment of physical characteristics of meat. *Meat Science*, **49**, 447-457.
- Hosono, A., Ozawa, A., Kato, R., Ohnishi, Y., Nakanishi, Y., Kimura, T., & Nakamura, R. (2003). Dietary fructooligosaccharides induce immunoregulation of intestinal IgA secretion by murine Peyer's patch cells. *Bioscience, Biotechnology, and Biochemistry*, **67**(4), 758-764.
- Hossain, M. A., & Rahman, A. (2011). Chemical composition of bioactive compounds by GC-MS screening and antifungal properties of the crude extracts of cabbage samples. *Asian Journal of Biotechnology*. **3** (1), 68-76.
- Houshmand, M., Azhar, K., Zulkifli, I., Bejo, M. H. Meiman-dipour, A., & Kamyab, A. (2012). Effects of non-antibiotic feed additives on performance, tibial dyschondroplasia incidence and tibia characteristics of broilers fed low-calcium diets. *Poultry Science*, **91**, 393-401.
- Houshmand, M., Azhar, K., Zulkifli, I., Bejo, M. H., Meimandipour, A., & Kamyab, A. (2011). Effects of non-antibiotic feed additives on performance, tibial dyschondroplasia incidence and tibia characteristics of broilers fed low-calcium diets. *Journal of Animal Physiology and Animal Nutrition*, **95**, 351-358.
- Huang, M. K., Choi, Y., Houde, R., Lee, J. W., Lee, B., & Zhao, X. (2004). Effects of *Lactobacilli* and an acidophilic fungus on the production performance and immune responses in broiler chickens. *National Research Council Canada* **83**, 788-795.
- Huang, R. L., Deng, Z. Y., Yang, C. b., Yin, Y. L., Xie, M. Y., Wu, G. Y., Li, T. J., Li, L. L., Tang, Z. R., & Kang, P. (2007). Dietary oligochitosan supplementation enhances immune status of broilers. *Journal of the Science of Food and Agriculture*, **87**, 153-159.
- Huyghebaert, G., Ducatelle, R., & Van Immerseel, F. (2011). An update on alternatives to antimicrobial growth promoters for broilers. *The Veterinary Journal*, **187**, 182-188.
- Ibrahim, S. M., & Desouky, S. G. (2009). Effect of antimicrobial metabolites produced by lactic acid bacteria (Lab) on quality aspects of frozen tilapia (*Oreochromis niloticus*) filets. *Journal of Fish and Marine Sciences*, **1** (1), 40-45.
- Igarashi M, Liyama Y, Kato R, Tomita M, & Asami N, I., E. (1994). Effect of *Bifidobacterium longum* and lactulose on the strength of bone in ovariectomized osteoporosis model rats. *Bifidus*, **7**, 139-147.

- Ishimwe, N., Daliri, E. B., Lee, B. H., Fang, F., & Du, G. (2015). The perspective on cholesterol lowering mechanisms of probiotics. *Molecular Nutrition & Food Research*, **59**, 94-105.
- Isolauri, E., Sütas, Y., Kankaanpää, P., Arvilommi, H., & Salminen, S. (2001). Probiotics: effects on immunity. *The American Journal of Clinical Nutrition*, **73**, 444s-450s.
- Izadi, H., Arshami, J., Golian, A., & Raji, M. R. (2013). Effects of chicory root powder on growth performance and histomorphometry of jejunum in broiler chicks. *Veterinary Research Forum*, **4** (3) 169-174.
- Jack, R. W., Tagg, J. R., & Ray, B. (1995). Bacteriocins of gram-positive bacteria. *Microbiological Reviews*, **59**, 171-200.
- Jacobasch, G., Schmiedl, D., Kruschewski, M., & Schmehl, K. (1999). Dietary resistant starch and chronic inflammatory bowel diseases. *International Journal of Colorectal Disease*, **14**, 201-211.
- Jadhav, D. Y., Sahoo, A. K., Ghosh, J. S., Ranveer, R. C., & Mali, A. M. (2010). Phytochemical detection and in vitro evaluation of Tamarind fruit pulp for potential antimicrobial activity. *International Journal of Tropical Medicine*, **5** (3), 68-72.
- Jama, N., Muchenje, V., Chimonyo, M., Strydom, P., Dzama, K., & Raats, J. (2008). Cooking loss components of beef from Nguni, Bonsmara and Angus steers. *African Journal of Agriculture Research*, **3**, 416-420.
- Janardhana, V., Broadway, M. M., Bruce, M. P., Lowenthal, J. W., Geier, M. S., Hughes, R. J., & Bean, A. G. (2009). Prebiotics modulate immune responses in the gut-associated lymphoid tissue of chickens. *The Journal of nutrition*, **139**, 1404-1409.
- Janczyk, P., Halle, B., & Souffrant, W. B. (2009). Microbial community composition of the crop and ceca contents of laying hens fed diets supplemented with *Chlorella vulgaris*. *Poultry Science*, **88**, 2324-2332.
- Janeway, C. A. (2001). Antigen Recognition by B cells and T Cell Receptors. Pages 245. In: Immunobiology. C. A. Janeway, J. P. Travers, M. Walport, and M. J. Shlomchik, eds. New York, Garland Science.
- Jayaraman, S., Thangavel, G., Kurian, H., Mani, R., Mukkalil, R., & Chirakkal, H. (2013). *Bacillus subtilis* PB6 improves intestinal health of broiler chickens challenged with *Clostridium perfringens*-induced necrotic enteritis. *Poultry Science*, **92**, 370-374.
- Jeurissen, S., Lewis, F., van der Klis, J. D., Mroz, Z., Rebel, J., & Huurne, A. (2002). Parameters and techniques to determine intestinal health of poultry as constituted by immunity, integrity, and functionality. *Current Issues in Intestinal Microbiology*, **3**, 1-14.

- Jones, J. I., & Clemmons, D. R. (1995). Insulin-Like Growth Factors and Their Binding Proteins: Biological Actions. *Endocrine reviews*, **16**, 3-34.
- Józefiak, D., Kaczmarek, S., & Rutkowski, A. (2008). A note on the effects of selected prebiotics on the performance and ileal microbiota of broiler chickens. *Journal of Animal and Feed Sciences*, **17**, 392-397.
- Jukna, C., Jukna, V., & Simkus, A. (2005). The effect of probiotics and phytobiotics on meat properties and quality in pigs. *Veterinary Medicine and Zootechnics*, **29**, 80-84.
- Jung, K. H., Yoo, S. K., Moon, S. K., & Lee, U. S. (2007). Furfural from pine needle extract inhibits the growth of a plant pathogenic fungus, *Alternaria mali*. *Mycobiology*, **35** (1), 39-43.
- Jung, S., Houde, R., Baurhoo, B., Zhao, X., & Lee, B. (2008). Effects of galacto-oligosaccharides and a *Bifidobacteria lactis*-based probiotic strain on the growth performance and fecal microflora of broiler chickens. *Poultry science*, **87**, 1694-1699.
- Kabir, S. L., Rahman, M., Rahman, M., Rahman, M., & Ahmed, S. (2004). The dynamics of probiotics on growth performance and immune response in broilers. *International Journal of Poultry Science*, **3**, 361-364.
- Kabir, S., Rahman, M., & Rahman, M. (2005). Potentiation of probiotics in promoting microbiological meat quality of broilers. *Journal of the Bangladesh Society Agriculture Science Technology*, **2**, 93-96.
- Kandler, O., & Weiss N. (1986). Regular non-sporing Gram positive rods. Pages 1208–1234 in Bergey's Manual of Systematic Bacteriology, Vol. 2, Sneath, P.H.A., M. E. Sharpe, and J. G. Holt, William and Wilkins, Baltimore, MD.
- Kandler, O., (1983). Carbohydrate metabolism in lactic acid bacteria. *Antonie van Leeuwenhoek*, **49**, 209-224.
- Karaoglu, M., Aksu, M., Esenbuga, N., Kaya, M., Macit, M., & Durdag, H. (2004). Effect of dietary probiotic on the pH and colour characteristics of carcasses, breast fillets and drumsticks of broilers. *Animal Science*, **78**, 253-259.
- Karaoglu, M., Aksu, M., Esenbuga, N., Macit, M., & Durdağ, H. (2006). pH and colour characteristics of carcasses of broilers fed with dietary probiotics and slaughtered at different ages. *Asian-Australasian Journal of Animal Sciences*, **19**, 605-610.
- Keeney, K. M., & Finlay, B. B. (2011). Enteric pathogen exploitation of the microbiota-generated nutrient environment of the gut. *Current Opinion in Microbiology*, **14**, 92-98.
- Kelly, G. (2009). Inulin-type prebiotics: a review.(Part 2). Alternative medicine review: *Journal of Clinical Therapeutic*, **14**, 36-55.

- Khalafalla, F. A., Zahran, D., & Mosa, A. (2011). Influence of feed additives in quality of broiler carcasses. *The Journal of World's Poultry Research*, **2**, 40-47.
- Khan, A. R., Yousaf, M. S., Rehman, H., Zaneb, H., Pasha, T. N., Fatima, N., & Afzal, M. (2012c). Response of maternally isolated rock pigeons (*Columba livia domestica*) to different dietary concentrations of mannan-oligosaccharide. *Poultry Science*, **91** (7), 1598-1603.
- Khan, S., Hasan, S., & Sardar, R. (2011). Effect of dietary supplementation of probiotic on the performance of F1 crossbred (Rhode Island red male× Fayoumi female) cockerels. *Journal of Animal Physiology and Animal Nutrition*, **95**, 523-532.
- Kidd, M. (2004). Nutritional modulation of immune function in broilers. *Poultry Science*, **83**, 650-657.
- Kim, C. H., Shin, K. S., Woo, K. C., & Paik, I. K. (2009). Effect of dietary oligosaccharides on the performance, intestinal microflora and serum immunoglobulin contents in laying hens. *Korean Journal of Poultry Science*, **36**, 125-131.
- Kim, G. B., Seo, Y., Kim, C., & Paik, I. (2011). Effect of dietary prebiotic supplementation on the performance, intestinal microflora, and immune response of broilers. *Poultry Science*, **90**, 75-82.
- Kishi, M., Fukaya, M., Tsukamoto, Y., Nagasawa, T., Takehana, K., & Nishizawa, N. (1999). Enhancing effect of dietary vinegar on the intestinal absorption of calcium in ovariectomized rats. *Bioscience, Biotechnology, and Biochemistry*, **63**, 905-910.
- Kjos N. P., Overland M., Fauske A. K., & Sorum H. (2010). Feeding chicory inulin to entire male pigs during the last period before slaughter reduces skatole in digesta and backfat. *Livestock Science*, **134**, 143-145.
- Klaver, F., & Van Der Meer, R. (1993). The assumed assimilation of cholesterol by *Lactobacilli* and *Bifidobacterium bifidum* is due to their bile salt-deconjugating activity. *Applied and Environmental Microbiology*, **59**, 1120-1124.
- Kleter, G. A., & Marvin, H. J. (2009). Indicators of emerging hazards and risks to food safety. *Food and Chemical Toxicology*, **47**, 1022-1039.
- Klosterbuer, A., Roughead, Z.F., & Slavin, J. (2011). Benefits of dietary fiber in clinical nutrition. *Nutrition in Clinical Practice*, **26**, 625-635.
- Kochi, M., Takeuchi, S., Mizutani, T., Mochizuki, K., Matsumoto, Y., & Saito, Y. (1980). Antitumor activity of benzaldehyde. *Cancer Treatment Reports*, **64** (1), 21-3.
- Koenen, M., Kramer, J., Van Der Hulst, R., Heres, L., Jeurissen, S., & Boersma, W. (2004). Immunomodulation by probiotic *Lactobacilli* in layer-and meat-type chickens. *British Poultry Science*, **45**, 355-366.

- Kolida, S., & Gibson, G. R. (2011). Synbiotics in health and disease. *Annual Review of Food Science and Technology*, **2**, 373-393.
- Konca, Y., Kirkpınar, F., & Mert, S. (2009). Effects of mannan-oligosaccharides and live yeast in diets on the carcass, cut yields, meat composition and colour of finishing turkeys. *Asian-Australian Journal of Animal Science*, **22**, 550-556.
- Kotula, K. L., & Wang, Y. (1994). Characterization of broiler meat quality factors as influence by feed withdrawal time. *The Journal of Applied Poultry Research*, **3**, 103-110.
- Krag, L., Thomsen, L. E., & Iburg, T. (2006). Pathology of *Trichuris suis* Infection in Pigs Fed an Inulin-and a Non-inulin-containing Diet. *Journal of Veterinary Medicine Series A*, **53**(8), 405-409.
- Král, M., Angelovičova, M., Alfaig, E., Bučko, O., & Walezycka, M. (2014). Influence of *Bacillus subtilis* and acetic acid on Cobb500 intestinal microflora. *Scientific Papers Animal Science and Biotechnologies*, **47**, 22-25.
- Kruger, M. C., Brown, K. E., Collett, G., Layton, L., & Schollum, L. M. (2003). The effect of fructooligosaccharides with various degrees of polymerization on calcium bioavailability in the growing rat. *Experimental Biology and Medicine*, **228**, 683-688.
- Kudoh, K., Shimizu, J., Ishiyama, A., Wada, M., Takita, T., Kanke, Y., & Innami, S. (1999). Secretion and excretion of immunoglobulin A to cecum and feces differ with type of indigestible saccharides. *Journal of Nutritional Science and Vitaminology*, **45**, 173-181.
- Kunnen, S., & Van Eck, M. (2012). Lecithin: cholesterol acyltransferase: old friend or foe in atherosclerosis? *Journal of lipid research*, **53**, 1783-1799.
- Kuroda, K., Caputo, G. A., & DeGrado, W. F. (2009). The role of hydrophobicity in the antimicrobial and hemolytic activities of polymethacrylate derivatives. *Chemistry European Journal*, **15**, 1123-1133.
- Kyriakis, S., Tsiolyiannis, V., Vlemmas, J., Sarris, K., Tsinas, A., Alexopoulos, C., & Jansegers, L. (1999). The effect of probiotic LSP 122 on the control of post-weaning diarrhoea syndrome of piglets. *Research in veterinary science*, **67**, 223-228.
- Labioui, H., Elmoualdi, L., El Yachioui, M., & Ouhssine, M. (2005). Sélection de souches de bactéries lactiques antibactériennes. *Bulletin-societe de pharmacie de bordeaux*, **144**, 237.
- Lagier, J. C., Million, M., Hugon, P., Armougom, F., & Raoult, D. (2012). Human gut microbiota: repertoire and variations. *Frontiers in cellular and infection microbiology*, 25-43. doi: 10.3389/fcimb.2012.00136.

- Lammers, K. M., Brigidi, P., Vitali, B., Gionchetti, P., Rizzello, F., Caramelli, E., Matteuzzi, D., & Campieri, M. (2003). Immunomodulatory effects of probiotic bacteria DNA: IL-1 and IL-10 response in human peripheral blood mononuclear cells. *FEMS Immunology & Medical Microbiology*, **38**, 165-172.
- Lampromsuk, P., Bunchasak, C., & Kaewtapee, C. (2012). Effect of supplementing acidifiers and organic zinc in diet on growth performances and gut conditions of pigs. *Journal of Applied Sciences*, **12**, 553.
- Langen, L. V., Mirjam, A. C., & Dieleman, L. A. (2009). Prebiotics in chronic intestinal inflammation. *Inflammatory Bowel Diseases*, **15** (3), 454-462.
- Lawrie, R., & Ledward, D. (2006). The eating qualities of meat. *Lawrie's Meat Science*, 7th edition, Cambridge, England, 1-14.
- Lawson, M. A. (2004). The role of integrin degradation in post-mortem drip loss in pork. *Meat Science*, **68**, 559-566.
- Le Bihan-Duval, E., Millet, N., & Remignon, H. (1999). Broiler meat quality: effect of selection for increased carcass quality and estimates of genetic parameters. *Poultry Science*, **78**, 822-826.
- Lee, K., Lillehoj, H. S., & Siragusa, G. R. (2010). Direct-fed microbials and their impact on the intestinal microflora and immune system of chickens. *The journal of Poultry Science*, **47**, 106-114.
- Lee, Y. K., & Salminen, S. (2009). *Handbook of probiotics and prebiotics*. John Wiley & Sons. Singapore.
- Lei, X., Piao, X., Ru, Y., Zhang, H., Péron, A., & Zhang, H. (2015). Effect of *Bacillus amyloliquefaciens*-based direct-fed microbial on performance, nutrient utilization, intestinal morphology and cecal microflora in broiler chickens. *Asian-Australasian Journal of Animal Sciences*, **28**, 239.
- Lei, X., Ru, Y., & Zhang, H. (2014). Effect of *Bacillus amyloliquefaciens*-based direct-fed microbials and antibiotic on performance, nutrient digestibility, cecal microflora, and intestinal morphology in broiler chickens. *The Journal of Applied Poultry Research*, doi: 10.3382/japr.2014-00965
- Leroy, F., & De Vuyst, L. (2000). Natural antimicrobial systems. CRC Press, London, 589-610.
- Leroy, F., & De Vuyst, L. (2004). Lactic acid bacteria as functional starter cultures for the food fermentation industry. *Trends in Food Science & Technology*, **15**, 67-78.
- Levrat, M. A., Rémésy, C., & Demigné, C. (1991). High propionic acid fermentations and mineral accumulation in the cecum of rats adapted to different levels of inulin. *The Journal of Nutrition*, **121**, 1730-1737.

- Li, L. L., Hou, Z. P., Li, T. J., Wu, G. Y., Huang, R. L., Tang, Z. R., Yang, C. B., Gong, J., Yu, H., & Kong, X. F. (2008). Effects of dietary probiotic supplementation on ileal digestibility of nutrients and growth performance in 1 to 42-day-old broilers. *Journal of the Science of Food and Agriculture*, **88**, 35-42.
- Li, X., Piao, X., Kim, S., Liu, P., Wang, L., Shen, Y., Jung, S., & Lee, H. (2007). Effects of chito-oligosaccharide supplementation on performance, nutrient digestibility, and serum composition in broiler chickens. *Poultry science*, **86**, 1107-1114.
- Liasi, S., Azmi, T., Hassan, M., Shuhaimi, M., Rosfarizan, M., & Ariff, A. (2009). Antimicrobial activity and antibiotic sensitivity of three isolates of lactic acid bacteria from fermented fish product, Budu. *Malaysian Journal of Microbiology*, **5**, 33-37.
- Lilly, R. A. (2010). The effect of dietary amino acid density in broiler on carcass characteristics and meat quality. Master Thesis, Mississippi State University, USA.
- Lima Filho, J., Vieira, E., & Nicoli, J. (2000). Antagonistic effect of *Lactobacillus acidophilus*, *Saccharomyces boulardii* and *Escherichia coli* combinations against experimental infections with *Shigella flexneri* and *Salmonella enteritidis* sub sp. typhimurium in gnotobiotic mice. *Journal of Applied Microbiology*, **88**, 365-370.
- Lin, J., Hunkapiller, A. A., Layton, A. C., Chang, Y. J., & Robbins, K. R. (2013). Response of intestinal microbiota to antibiotic growth promoters in chickens. *Foodborne Pathogens and Disease*, **10**, 331-337.
- Lin, P. P., Hsieh, Y. M., & Tsai, C. C. (2009). Antagonistic activity of *Lactobacillus acidophilus* RY2 isolated from healthy infancy feces on the growth and adhesion characteristics of enteroaggregative *Escherichia coli*. *Anaerobe*, **15** (4), 122-6.
- Liu, X., Yan, H., Lv, L., Xu, Q., Yin, C., Zhang, K., Wang, P., & Hu, J. (2012). Growth performance and meat quality of broiler chickens supplemented with *Bacillus licheniformis* in drinking water. *Asian-Australasian Journal of Animal Sciences*, **25**, 682.
- Livak, K. J., & Schmittgen, T. D. (2001). Analysis of relative gene expression data using real-time quantitative PCR and the 2- $\Delta\Delta$ CT method. *Methods*, **25**, 402-408.
- Loddi, M. M., Gonzales, E., Takita, T. S., Mendes, A. A., & Roça, R. d. O. (2000). Effect of the use of probiotic and antibiotic on the performance, yield and carcass quality of broilers. *Revista Brasileira de Zootecnia*, **29**, 1124-1131.
- Loddi, M., Maraes, V., Nakaghi, I., Tucci, F., Hannas, M., & Ariki, J. (2004). Mannan oligosaccharide and organic acids on performance and intestinal morphometric characteristics of broiler chickens, proceedings of the 20th annual symposium. Supplement, 1: 45.

- Loh G., Eberhard M., Brunner R.M., Hennig U., Kuhla S., Kleessen B., Metges C.C. (2006). Inulin alters the intestinal microbiota and short-chain fatty acid concentrations in growing pigs regardless of their basal diet. *Journal of Nutrition*, **136**, 1198-1202.
- Loh, T. C., Choe, D. W., Foo, H. L., Sazili, A. Q., & Bejo, M. H. (2014). Effects of feeding different postbiotic metabolite combinations produced by *Lactobacillus plantarum* strains on egg quality and production performance, faecal parameters and plasma cholesterol in laying hens. *BMC Veterinary Research*, **10**(1):149.
- Loh, T. C., Foo, H. L., Tan, S. H., Goh, Y. M., Shukriah, M. H., & Kufli, C. N. (2003b). Effects of fermented products on performance, faecal pH, *Enterobacteriaceae* and lactic acid bacteria counts and relationships, and plasma cholesterol concentration in rats. *Journal of Animal Feed Science*, **12**, 633-644.
- Loh, T. C., Lee, T., Foo, H. L., Law, F., & Ajion, M. (2008). Growth performance and fecal microflora of rats offered metabolites from lactic acid bacteria. *Journal of Applied Animal Research*, **34**, 61-64.
- Loh, T. C., Thanh, N. T., Foo, H. L., HAIR-BEJO, M., & Azhar, B. K. (2010). Feeding of different levels of metabolite combinations produced by *Lactobacillus plantarum* on growth performance, fecal microflora, volatile fatty acids and villi height in broilers. *Animal Science Journal*, **81**, 205-214.
- Loh, T. C., Thanh, N., Foo, H. L., & Hair-Bejo, M., (2013). Effects of Feeding metabolite combinations from *Lactobacillus plantarum* on plasma and breast meat lipids in broiler chickens. *Revista Brasileira de Ciência Avícola*, **15**, 307-316.
- Loh, T. C., Thu, T. V., Foo, H. L., & Bejo, M. H. (2013). Effects of different levels of metabolite combination produced by *Lactobacillus plantarum* on growth performance, diarrhoea, gut environment and digestibility of postweaning piglets. *Journal of Applied Animal Research*, **41**(2), 200-207.
- Loh, T., Foo, H., Lee, K., Lim, Y., & Kufli, C., (2003a). Effect of fermented fruits on the growth performance, shedding of *Enterobacteriaceae* and *Lactobacilli* in post-weaning pigs. *Asian-Australasian Journal of Animal Sciences*, **16**, 1656-1660.
- Loh, T., Law, F., Foo, H., Goh, Y., & Zulkifli, I. (2007). Effects of feeding a fermented product on egg production, faecal microflora and faecal pH in laying hens. *Journal of Animal and Feed Sciences*, **16**, 452.
- Lopez, H. W., Coudray, C., Levrat-Verny, M. A., Feillet-Coudray, C., Demigné, C., & Révész, C. (2000). Fructooligosaccharides enhance mineral absorption and counteract the deleterious effects of phytic acid on mineral homeostasis in rats. *The Journal of nutritional biochemistry*, **11**, 500-508.



- Lott, B., Reece, F., & Drott, J. (1980). Effect of preconditioning on bone breaking strength. *Poultry Science*, **59**, 724-725.
- Luciano, G., Monahan, F. J., Vasta, V., Biondi, L., Lanza, M., & Priolo, A. (2009). Dietary tannins improve lamb meat colour stability. *Meat Science*, **81**, 120-125.
- Lutz, T., & Scharrer, E. (1991). Effect of short-chain fatty acids on calcium absorption by the rat colon. *Experimental Physiology*, **76**, 615-618.
- Lynch, M. B., Sweeney, T., Callan, J. J., & O' Doherty, J. V. (2007). The effect of dietary crude protein concentration and inulin supplementation on nitrogen excretion and intestinal microflora from finisher pigs. *Livestock Science*, **109**, 204-207.
- Macfarlane, G., Gibson, G., & Cummings, J. (1992). Comparison of fermentation reactions in different regions of the human colon. *Journal of Applied Bacteriology*, **72**, 57-64.
- Macfarlane, G., Macfarlane, S., & Gibson, G. (1998). Validation of a three-stage compound continuous culture system for investigating the effect of retention time on the ecology and metabolism of bacteria in the human colon. *Microbial Ecology*, **35**, 180-187.
- Macfarlane, S., Macfarlane, G., & Cummings, J. T. (2006). Review article: prebiotics in the gastrointestinal tract. *Alimentary Pharmacology & Therapeutics* **24**, 701-714.
- Macpherson, A. J., & Uhr, T. (2004). Induction of protective IgA by intestinal dendritic cells carrying commensal bacteria. *Science*, **303**, 1662-1665.
- Madrigal, L., & Sangronis, E. (2007). La inulina y derivados como ingredientes claves en alimentos funcionales. *Latin American Archives Nutrition*, **57**, 387-396.
- Maiorano, G., Sobolewska, A., Cianciullo, D., Walasik, K., Elminowska-Wenda, G., Sławińska, A., Tavaniello, S., Żylińska, J., Bardowski, J., & Bednarczyk, M. (2012). Influence of in ovo prebiotic and synbiotic administration on meat quality of broiler chickens. *Poultry Science*, **91**, 2963-2969.
- Markovic, R. (2005). The effect of different growth promoters in broiler nutrition on performance and health status Master Thesis. Faculty of Veterinary Medicine 2005, University of Belgrade, Belgrade, Serbia.
- Marteau, P., & Shanahan, F. (2003). Basic aspects and pharmacology of probiotics: an overview of pharmacokinetics, mechanisms of action and side-effects. *Best Practice and Research Clinical Gastroenterology*, **17** (5), 725-740.
- Marten B, Scholz-Ahrens & KE, J., S. (2004). Präbiotika und Synbiotika aber nicht Probiotika steigern die Calciumresorption und -retention bei ovariekomierten Ratten [abstract]. *Proceedings of the Nutrition Society*, **6**, 71.

- Matsuzaki, T. (1998). Immunomodulation by treatment with *Lactobacillus casei* strain Shirota. *International Journal of Food Microbiology*, **41**, 133-140.
- Matsuzaki, T., & Chin, J. (2000). Modulating immune responses with probiotic bacteria. *Immunology and Cell Biology*, **78**, 67-73.
- McHan, F., & Shotts, E. B. (1993). Effect of short-chain fatty acids on the growth of *Salmonella typhimurium* in an in vitro system. *Avian Diseases*, 396-398.
- McKay, D. M., & Perdue M. H. (1993). Intestinal epithelial function: The case for immune physiological regulation. *Digestive Diseases and Science*, **38**, 1377-1387.
- Menconi, A., Kuttappan, V., Hernandez-Velasco, X., Urbano, T., Matté, F., Layton, S., Kallapura, G., Latorre, J., Morales, B., & Prado, O. (2014). Evaluation of a commercially available organic acid product on body weight loss, carcass yield, and meat quality during preslaughter feed withdrawal in broiler chickens: A poultry welfare and economic perspective. *Poultry Science*, **93**, 448-455.
- Merrifield, D.L., & Ringo, E. (2014). Aquaculture nutrition: gut health, probiotics and prebiotics. Plymouth. UK
- Miles, R. D., Butcher, G. D., Henry, P. R. & Littell, R. C. (2006) Effect of antibiotic growth promoters on broiler performance, intestinal growth parameters and quantitative morphology. *Poultry Science*, **85**, 476-485.
- Mineo, H., Hara, H., Kikuchi, H., Sakurai, H., & Tomita, F. (2001). Various indigestible saccharides enhance net calcium transport from the epithelium of the small and large intestine of rats in vitro. *The Journal of Nutrition*, **131**, 3243-3246.
- Miremadi, F., Ayyash, M., Sherkat, F., & Stojanovska, L. (2014). Cholesterol reduction mechanisms and fatty acid composition of cellular membranes of probiotic *Lactobacilli* and *Bifidobacteria*. *Journal of Functional Foods*, **9**, 295-305.
- Moghadam, M. S., Foo, H. L., Leow, T. C., Rahim, R. A., & Loh, T. C. (2010). Novel bacteriocinogenic *Lactobacillus plantarum* strains and their differentiation by sequence analysis of 16S rDNA, 16S-23S and 23S-5S intergenic spacer regions and randomly amplified polymorphic DNA analysis. *Food Technology and Biotechnology*, **48**, 476-483.
- Momose, Y., Hirayama, K., & Itoh, K. (2008). Effect of organic acids on inhibition of *Escherichia coli* O157:H7 colonization in gnotobiotic mice associated with infant intestinal microbiota. *Antonie van Leeuwenhoek*, **93**, 141-9.
- Montagne, L., Pluske, J. R., & Hampson, D. J. (2003). A review of inter-actions between dietary fiber and the intestinal mucosa, and their consequences on digestive health in young non-ruminant animals. *Animal Feed Science and Technology*, **108**, 95-117.

- Mookiah, S., Sieo, C. C., Ramasamy, K., Abdullah, N., & Ho, Y. W. (2014). Effects of dietary prebiotics, probiotic and synbiotics on performance, caecal bacterial populations and caecal fermentation concentrations of broiler chickens. *Journal of the Science of Food and Agriculture*, **94** (2), 341-348.
- Moriyama, S., (1995). Increased plasma insulin-like growth factor-I (IGF-I) following oral and intraperitoneal administration of growth hormone to rainbow trout, *Oncorhynchus mykiss*. *Growth regulation*, **5**, 164-167.
- Mountzouris, K., Tsitrsikos, P., Palamidi, I., Arvaniti, A., Mohnl, M., Schatzmayr, G., & Fegeros, K. (2010). Effects of probiotic inclusion levels in broiler nutrition on growth performance, nutrient digestibility, plasma immunoglobulins, and cecal microflora composition. *Poultry Science*, **89**, 58-67.
- Muir, W., Bryden, W., & Husband, A. (2000). Immunity, vaccination and the avian intestinal tract. *Developmental and Comparative Immunology*, **24**, 325-342.
- Mulder, I. E., Schmidt, B., Stokes, C. R., Lewis, M., Bailey, M., Aminov, R. I., Prosser, J. I., Gill, B. P., Pluske, J. R., & Mayer, D. (2009). Environmentally-acquired bacteria influence microbial diversity and natural innate immune responses at gut surfaces. *BMC Biology*, **7**(1), 79.
- Muramatsu, T., Nakajima, S., & Okumura, J. (1994). Modification of energy metabolism by the presence of the gut microflora in the chicken. *British Journal of Nutrition*, **71**, 709-717.
- Murray, A. C. (1995). The evaluation of muscle quality. In quality and grading of carcasses of meat animals. New York, 234.
- Mutuş, R., Kocabağlı, N., Alp, M., Acar, N., Eren, M., & Gezen, Ş. (2006). The effect of dietary probiotic supplementation on tibial bone characteristics and strength in broilers. *Poultry science*, **85**, 1621-1625.
- Nabizadeh, A. (2012). The effect of inulin on broiler chicken intestinal microflora, gut morphology, and performance. *Journal of Animal and Feed Science*, **21**, 725-734.
- Nam, H. M., Srinivasan, V., Gillespie, B. E., Murinda, S. E., & Oliver, S. P. (2005). Application of SYBR green real-time PCR assay for specific detection of *Salmonella* spp. in dairy farm environmental samples. *International Journal of Food microBiology*, **102** (2), 161-171.
- Nankervis, L., Matthews, S., & Appleford, P. (2000). Effect of dietary non-protein energy source on growth, nutrient retention and circulating insulin-like growth factor I and triiodothyronine levels in juvenile barramundi, *Lates calcarifer*. *Aquaculture*, **191**, 323-335.
- Navidshad, B., Liang, J. B., & Jahromi, M. F. (2012). Correlation coefficients between different methods of expressing bacterial quantification using real time PCR. *International Journal of Molecular Sciences*, **13** (2), 2119-2132.

- Nayebpor, M., Farhomand, P., & Hashemi, A. (2007). Effects of different levels of direct fed microbial (Primalac) on growth performance and humoral immune response in broiler chickens. *Journal of Animal and Veterinary Advances*, **6**, 1308-1313.
- Nekoubin, H., & Sudagar, M. (2012). Assessment of the Effects of symbiotic (Biomim Imbo) via supplementation with Artificial Diet (with Different protein levels) on Growth Performance and Survival Rate in Grass carp (*Ctenopharyngodon idella*). *World Journal of Zoology*, **7**(3), 236-240.
- Nes, I. F., Diep, D. B., Håvarstein, L. S., Brurberg, M. B., Eijsink, V., & Holo, H. (1996). Biosynthesis of bacteriocins in lactic acid bacteria. *Antonie van Leeuwenhoek*, **70**, 113-128.
- Newman, K. (1994). *Mannan oligosaccharides: Natural Polymers with Significant Impact on the Gastrointestinal Microflora and the Immune System*. Pages 167-174 in: *Biotechnology in the Feed Industry*. Nottingham, UK.
- Northcutt J. K. (1997). Factors affecting poultry meat quality. Bulletin 1157 1997. [Accessed nov. 18, 2006]. Available from: URL: <http://pubs.caes.uga.edu>.
- Nunes, R. V., Scherer, C., Pozza, P. C., Eyng, C., Bruno, L. D. G., & Vieites, F. M. (2012). Use of probiotics to replace antibiotics for broilers. *Revista Brasileira de Zootecnia*, **41** (10), 2219-2224.
- Ofek, I., Mirelman, D., & Sharon, N. (1977). Adherence of *Escherichia coli* to human mucosal cells mediated by mannose receptors. *Nature*, **265**, 623-625.
- Offer, G., & Knight, P. (1988). Structural basis of water-holding in meat. 2. Drip losses In: *Developments in Meat Science*. Washington, USA.
- O'Hara, A. M., & Shanahan, F. (2006). The gut flora as a forgotten organ. *EMBO Reports*, **7**, 688-693.
- Ohashi, Y., Hiraguchi, M., & Ushida, K. (2006). The composition of intestinal bacteria affects the level of luminal IgA. *Bioscience, Biotechnology, and Biochemistry*, **70** (12), 3031-3035.
- Ohimain, E. I., Ofongo, R. T. (2012). The effect of probiotic and prebiotic feed supplementation on chicken health and gut microflora: a review. *International Journal of Animal and Veterinary Advance*, **4**, 135-143.
- Ondruška, L., Chrástínová, L., Rafay, J., Pospíšilová, D., & Parkányi, V. (2012). Effect of humic substances and probiotics on growth performance and meat quality of rabbits. *Potravinárstvo*, **6**, 39-41.
- Ooi, L. G., & Liang, M. T. (2010). Cholesterol-lowering effects of probiotics and prebiotics: a review of in vivo and in vitro findings. *International Journal of Molecular Sciences*, **11**, 2499-2522.

- Orla-jensen, S. (1919). The lactic acid bacteria. Den Konglige Danske Videnskablige Selskabog Skrifter Andr., 184-196.
- Ortiz, L., Rodriguez, M., Alzueta, C., Rebole, A., & Trevino, J. (2009). Effect of inulin on growth performance, intestinal tract sizes, mineral retention and tibial bone mineralisation in broiler chickens. *British Poultry Science*, **50**, 325-332.
- Otto, G., Roche, R., Looft, H., Thoelking, L., Henning, M., Plastow, G. S., & Kalm, E. (2006). Drip loss of case-ready meat and of premium cuts and their association with earlier measured sample drip loss, meat quality and carcass traits in pigs. *Meat Science*, **72**, 680–687.
- Ozturk, E., Ocak, N., Turan, A., Erener, G., Altop, A., & Cankaya, S. (2012). Performance, carcass, gastrointestinal tract and meat quality traits, and selected blood parameters of broilers fed diets supplemented with humic substances. *Journal of the Science of Food and Agriculture*, **92**, 59-65.
- Pan, D. D., Zeng, X. Q., & Yan, Y. T. (2011). Characterisation of *Lactobacillus fermentum* SM-7 isolated from koumiss, a potential probiotic bacterium with cholesterol-lowering effects. *Journal of the Science of Food and Agriculture*, **91**, 512-518.
- Partanen, K. H., & Mroz, Z. (1999). Organic acids for performance enhancement in pig diets. *Nutrition Research Reviews*, **12**, 117-145.
- Parvaneh, K., Jamaluddin, R., Karimi, G., & Erfani, R. (2014). Effect of probiotics supplementation on bone mineral content and bone mass density. *The Scientific World Journal*, **2014**, 1-6.
- Patten, J., & Waldroup, P. (1988). Use of organic acids in broiler diets. *Poultry Science*, **67**, 1178-1182.
- Patterson, P., Cook, M., Crenshaw, T., & Sunde, M. L. (1986). Mechanical properties of the tibiotarsus of broilers and poults loaded with artificial weight and fed various dietary protein levels. *Poultry Science*, **65**, 1357-1364.
- Paul Ross, R., Morgan, S., & Hill, C. (2002). Preservation and fermentation: past, present and future. *International Journal of Food Microbiology*, **79**, 3-16.
- Pelicano, E. R. L., De Souza, P., De Souza, H., Oba, A., Norkus, E., Kodawara, L., & De Lima, T. (2003). Effect of different probiotics on broiler carcass and meat quality. *Revista Brasileira de Ciência Avícola*, **5**, 207-214.
- Perdigon, G., Maldonado, G. C., Valdez, J., & Medici, M. (2002). Interaction of lactic acid bacteria with the gut immune system. *European Journal of Clinical Nutrition*, **56**, S21-26.
- Pereira, D. I., & Gibson, G. R. (2002). Cholesterol assimilation by lactic acid bacteria and bifidobacteria isolated from the human gut. *Applied and Environmental Microbiology*, **68**, 4689-4693.

- Petracci, M., & Cavani, C. (2012). Muscle growth and poultry meat quality issues. *Nutrients*, **4**, 1-12.
- Petracci, M., Betti, M., Bianchi, M., & Cavani, C. (2004). Color variation and characterization of broiler breast meat during processing in Italy. *Poultry Science*, **83**, 2086-2092.
- Piva, A., Panciroli, A., Meola, E., & Formigoni, A. (1996). Lactitol enhances short-chain fatty acid and gas production by swine cecal microflora to a greater extent when fermenting low rather than high fiber diets. *The Journal of Nutrition*, **126** (1), 280-289.
- Ponnampalam E., Jayasooriya D., Gill H., Pluske J., & Dunshea F. (2009). Nutritional manipulation of iron level in finisher pigs and fresh pork. Final Report (3A-108) prepared for the Co-operative Research Centre for an Internationally Competitive Pork Industry. Werribee, Australia.
- Qiao, M., Fletcher, D., Northcutt, J., & Smith, D. (2002). The relationship between raw broiler breast meat color and composition. *Poultry Science*, **81**, 422-427.
- Qiao, M., Fletcher, D., Smith, D., & Northcutt, J. (2001). The effect of broiler breast meat color on pH, moisture, water-holding capacity, and emulsification capacity. *Poultry Science*, **80**, 676-680.
- Qiu, R., Croom, J., Ali, R. A., Ballou, A. L., Smith, C., Ashwell, C. M., Hassan, H. M., Chiang, C. C., & Koci, M. D. (2012). Direct fed microbial supplementation repartitions host energy to the immune system. *Journal of Animal Science*, **90**, 2639–2651
- Racevičiūtė-Stupelienė, A., Šašytė, V., Gružasuskas, R., & Šimkus, A. (2007). Influence of probiotic preparation yeasture-w on the productivity and meat quality of broiler chickens. *Biotechnology in Animal Husbandry*, **23**, 543-550.
- Rakoff-Nahoum, S., Paglino, J., Eslami-Varzaneh, F., Edberg, S., & Medzhitov, R. (2004). Recognition of commensal microflora by toll-like receptors is required for intestinal homeostasis. *Cell*, **118**, 229-241.
- Rasmussen, A., & Andersson, M. (1996). New method for determination of drip loss in pork muscles, Proceedings 42<sup>nd</sup>. The International Congress of Meat Science and Technology, 286-287.
- Rastall, R. (2004). Bacteria in the gut: friends and foes and how to alter the balance. *The Journal of Nutrition*, **134**, 2022S-2026S.
- Rath, N., Huff, G., Huff, W., & Balog, J. (2000). Factors regulating bone maturity and strength in poultry. *Poultry Science*, **79** (7), 1024-1032.

- Rebolé, A., Ortiz, L., Rodríguez, M. L., Alzueta, C., Treviño, J., & Velasco, S. (2010). Effects of inulin and enzyme complex, individually or in combination, on growth performance, intestinal microflora, cecal fermentation characteristics, and jejunal histomorphology in broiler chickens fed a wheat-and barley-based diet. *Poultry Science*, **89**, 276-286.
- Rehman H., Hellweg P., Taras D., & Zentek J. (2008). Effects of dietary inulin on the intestinal short chain fatty acids and microbial ecology in broiler chickens as revealed by denaturing gradient gel electrophoresis. *Poultry Science*, **87**, 783-789
- Rehman, H. U., Vahjen, W., Awad, W. A., & Zentek, J. (2007). Indigenous bacteria and bacterial metabolic products in the gastrointestinal tract of broiler chickens. *Archives of Animal Nutrition*, **61**, 319-335.
- Rehman, H., Vahjen, W., Kohl-Parisini, A., Ijaz, A., & Zentek, J. (2009). Influence of fermentable carbohydrates on the intestinal bacteria and enteropathogens in broilers. *World's Poultry Science Journal*, **65**, 75-90.
- Remesy, C., Levrat, M., Gamet, L., & Demigné, C. (1993). Cecal fermentations in rats fed oligosaccharides (inulin) are modulated by dietary calcium level. *American Journal of Physiology-Gastrointestinal and Liver Physiology*, **264**, G855-G862.
- Rescigno, M., Urbano, M., Valzasina, B., Francolini, M., Rotta, G., Bonasio, R., Granucci, F., Kraehenbuhl, J. P., & Ricciardi-Castagnoli, P. (2001). Dendritic cells express tight junction proteins and penetrate gut epithelial monolayers to sample bacteria. *Nature Immunology*, **2**, 361-367.
- Rezaei, S., Jahromi, M. F., Liang, J. B., Zulkifli, I., Farjam, A. S., Laudadio, V., & Tufarelli, V. (2015). Effect of oligosaccharides extract from palm kernel expeller on growth performance, gut microbiota and immune response in broiler chickens. *Poultry Science*, **94** (10), 2414-2420.
- Riesenfeld, A. (1972). Metatarsal robusticity in bipedal rats. *American Journal of Physical Anthropology*, **36**, 229-233.
- Rinttilä, T., & Apajalahti, J. (2013). Intestinal microbiota and metabolites—Implications for broiler chicken health and performance<sup>1</sup>. *The Journal of Applied Poultry Research*, **22**, 647-658.
- Roberfroid, M. (2004). Inulin-type fructans: functional food ingredients. Washington, USA.
- Roberfroid, M. (2007). Prebiotics: the concept revisited. *The Journal of Nutrition*, **137**, 830S-837S.
- Roberfroid, M. B. (2002). Functional foods: concepts and application to inulin and oligofructose. *British Journal of Nutrition*, **87**(S2), S139-S143.

- Robinson, R. K. (1995). The potential of inulin as a functional ingredient. *British Food Journal*, **97**, 30-32.
- Rodríguez, E., González, B., Gaya, P., Nuñez, M., & Medina, M. (2000). Diversity of bacteriocins produced by lactic acid bacteria isolated from raw milk. *International Dairy Journal*, **10**, 7-15.
- Rodríguez, J., Martínez, M., Horn, N., & Dodd, H. (2003). Heterologous production of bacteriocins by lactic acid bacteria. *International Journal of Food Microbiology*, **80**, 101-116.
- Rodríguez, M. L., Rebole, A., Velasco, S., Ortiz, L. T., Tre-vino, J., & Alzueta, C. (2012). Wheat- and barley-based diets with or without additives influence broiler chicken performance, nutrient digestibility and intestinal microflora. *Journal of the Science of Food and Agriculture*, **92**, 184-190.
- Ros, E. (2000). Intestinal absorption of triglyceride and cholesterol. Dietary and pharmacological inhibition to reduce cardiovascular risk. *Atherosclerosis*, **151**, 357-379.
- Ross, R. P., Morgan, S., & Hill, C. (2002). Preservation and fermentation: past, present and future. *International Journal of Food Microbiology*, **79**, 3-16.
- Rosyidah, M. R., Loh, T. C., Foo, H., Cheng, X., & Bejo, M. H. (2011). Effects of feeding metabolites and acidifier on growth performance, faecal characteristics and microflora in broiler chickens. *Journal of Animal and Veterinary Advances*, **10**, 2758-2764.
- Rudel, L. L., & Morris, M. D. (1973). Determination of cholesterol using o-phthalaldehyde. *Journal of Lipid Research*, **14** (3), 364-366.
- Ruiz, J., Guerrero, L., Arnau, J., Guardia, M., & Esteve-Garcia, E. (2001). Descriptive sensory analysis of meat from broilers fed diets containing vitamin E or  $\beta$ -carotene as antioxidants and different supplemental fats. *Poultry Science*, **80**, 976-982.
- Russell, J. (1992). Another explanation for the toxicity of fermentation acids at low pH: anion accumulation versus uncoupling. *Journal of Applied Microbiology*, **73**, 363-370.
- Russell, M. W., Reinholdt, J., & Kilian, M. (1989). Anti-inflammatory activity of human IgA antibodies and their Fab  $\alpha$  fragments: Inhibition of IgG-mediated complement activation. *European Journal of Immunology*, **19**, 2243-2249.
- Saengkerdsub, S., Anderson, R. C., Wilkinson, H. H., Kim, W. K., Nisbet, D. J., & Ricke, S. C. (2007a). Identification and quantification of methanogenic archaea in adult chicken ceca. *Applied Environmental Microbiology*, **73**, 353-356.



- Saengkerdsub, S., Herrera, P., Woodward, C. L., Anderson, R. C., Nisbet, D. J., & Ricke, S. C. (2007b). Detection of methane and quantification of methanogenic archaea in faeces from young broiler chickens using real-time PCR. *Letters in Applied Microbiology*, **45**, 629-634.
- Saengkerdsub, S., Kim, W. K., Anderson, R. C., Nisbet, D. J., & Ricke, C. S. (2006). Effects of nitrocompounds and feedstuffs on in vitro methane production in chicken cecal contents and rumen fluid. *Anaerobe*, **12**, 85-92.
- Saki, A. A., Harcini, R. N., Rahmatnejad, E., & Salary, J. (2012). Herbal additives and organic acids as antibiotic alternatives in broiler chickens diet for organic production. *African Journal of Biotechnology*, **11**, 2139-2145.
- Salma, U., Miah, A., Maki, T., Nishimura, M., & Tsujii, H. (2007). Effect of dietary *Rhodobacter capsulatus* on cholesterol concentration and fatty acid composition in broiler meat. *Poultry Science*, **86**, 1920-1926.
- Salminen, S., & Isolauri, E. (2004). Identification of probiotics and prebiotics with anti-allergenic properties. *Allergic Diseases and the Environment*, **53**, 251-265.
- Samanta A. K., Jayapal N., Senani S., Kolte A. P., & Sridhar M. (2013). Prebiotic inulin: Useful dietary adjuncts to manipulate the livestock gut microflora. *Brazilian Journal of Microbiology*, **44**, 1-14.
- Sams, A. (1999). Looking for solutions pale meat, poor yield. *Broiler Industry*, **62**, 26-30.
- Santos, A., San Mauro, M., & Diaz, D. M. (2006). Prebiotics and their long-term influence on the microbial populations of the mouse bowel. *Food Microbiology*, **23**, 498-503.
- SAS Institute Inc, (2014). SAS/STAT® User's Guide Version 9.4 SAS Institute Inc, Cary, North Carolina, USA.
- Savadogo, A., Ouattara, A. C., Bassole, H. I., & Traore, S. A. (2006). Bacteriocins and lactic acid bacteria-a minireview. *African Journal of Biotechnology* **5** (9).
- Savage, A. W. J., Warriss, P. D., & Jolley, P. D. (1990). The amount and composition of the proteins in drip from stored pig meat. *Meat Science*, **27**, 289-303.
- Savell, J., Cross, H., Francis, J., Wise, J., Hale, D., Wilkes, D., & Smith, G. (1989). National consumer retail beef study: Interaction of trim level, price and grade on consumer acceptance of beef steaks and roasts. *Journal of Food Quality*, **12**, 251-274.
- Savković, T., Tojagić, S., & Jakanović, M. (2005). Effect of probiotics on production performance and meat quality of fattening chicks. *Biotechnology in Animal Husbandry*, **21**, 135-139.

- Sazili, A., Parr, T., Sensky, P., Jones, S., Bardsley, R., & Buttery, P. (2005). The relationship between slow and fast myosin heavy chain content, calpastatin and meat tenderness in different ovine skeletal muscles. *Meat Science*, **69**, 17-25.
- Schenk, M., & Mueller, C. (2008). The mucosal immune system at the gastrointestinal barrier. *Best Practice and Research Clinical Gastroenterology*, **22**, 391-409.
- Schilling, M., Radhakrishnan, V., Thaxton, Y., Christensen, K., Thaxton, J., & Jackson, V. (2008). The effects of broiler catching method on breast meat quality. *Meat Science*, **79**, 163-171.
- Scholz-Ahrens, K. E., Ade, P., Marten, B., Weber, P., Timm, W., Açil, Y., Glüer, C. C., & Schrezenmeir, J. (2007). Prebiotics, probiotics, and synbiotics affect mineral absorption, bone mineral content, and bone structure. *The Journal of nutrition*, **137**, 838S-846S.
- Scholz-Ahrens, K.E., & Schrezenmeir, J. (2002). Inulin, oligofructose and mineral metabolism-experimental data and mechanism. *British Journal of Nutrition*, **87**, S179-S186.
- Schultz, M., Linde, H. J., Lehn, N., Zimmermann, K., Grossmann, J., Falk, W., & Schölmerich, J. (2003). Immunomodulatory consequences of oral administration of *Lactobacillus rhamnosus* strain GG in healthy volunteers. *Journal of dairy research*, **70**, 165-173.
- Scott, K. P., Gratz, S. W., Sheridan, P. O., Flint, H. J., & Duncan, S. H. (2013). The influence of diet on the gut microbiota. *Pharmacological Research*, **69**(1), 52-60.
- Senkoylu, N., Samli, H., Kanter, M., & Agma, A. (2007). Influence of a combination of formic and propionic acids added to wheat-and barley-based diets on the performance and gut histomorphology of broiler chickens. *Acta Veterinaria Hungarica*, **55**, 479-490.
- Shabani, R., Nosrati, M., Javandel, F., Gothbi, A., & Kioumarsi, H. (2012). The effect of probiotics on growth performance of broilers. *Annals of Biological Research*, **3**, 5450-5452.
- Shamblott, M. J., Cheng, C. M., Bolt, D., & Chen, T. T. (1995). Appearance of insulin-like growth factor mRNA in the liver and pyloric ceca of a teleost in response to exogenous growth hormone. *Proceedings of the National Academy of Sciences*, **92**, 6943-6946.
- Shane, S. (2005). Antibiotic alternatives in turkey production. *World Poultry*, **19**, 14-15.
- Shazali, N., Foo, H. L., Loh, T. C., Choe, D. W., & Abdul Rahim, R. (2014). Prevalence of antibiotic resistance in lactic acid bacteria isolated from the faeces of broiler chicken in Malaysia. *Gut pathogens*, **6** (1), 1

- Shi, N. (2010). Effects of bacillus licheniformis, xylooligosaccharides and lactic acid on intestinal villi and intestinal microflora in broilers. MSc thesis, Henan Polytechnic University, China.
- Shi, X., & Noblet, J. (1993). Contribution of the hindgut to digestion of diets in growing pigs and adult sows: effect of diet composition. *Livestock Production Science*, **34**, 237-252.
- Shin, H. S., Chung, M. J., Kim, J. E., Lee, K. O., & Ha, N. J. (2009). Lactic acid bacteria affect serum cholesterol levels, harmful fecal enzyme activity, and fecal water content. *Lipids in Health and Disease*, **8** (1), 1.
- Short, F., Gorton, P., Wiseman, J., & Boorman, K. (1996). Determination of titanium dioxide added as an inert marker in chicken digestibility studies. *Animal Feed Science and Technology*, **59**, 215-221.
- Sifour, M., Tayeb, I., Haddar, H. O., Namous, H., & Aissaoui, S. (2012). Production and characterization of bacteriocin of *Lactobacillus plantarum* F12 with inhibitory activity against *Listeria monocytogenes*. *The Online Journal of Science and Technology*, **2**(1).
- Silverstein, J. T., Shearer, K. D., Dickhoff, W. W., Plisetskaya, E. M. (1998). Effects of growth and fatness on sexual development of chinook salmon (*Oncorhynchus tshawytscha*) parr. *Canadian Journal of Fisheries and Aquatic Sciences*, **55**, 2376-2382.
- Skinner, J. T., Izat, A. L., & Waldroup, P. W. (1991). Research note: Fumaric acid enhances performance of broiler chickens. *Poultry Science*, **70**, 1444-1447.
- Sobolewska, S., & Grela, E. R. (2014). The effect of inulin extraction method or powder from inulin-producing plants in fattener diets on performance, carcass traits and meat quality. *Annals of Animal Science*, **14** (4), 911-920.
- Sohail, M. U., Hume, M. E., Byrd, J. A., Nisbet, D. J., Ijaz, A., Sohail, A., Shabbir, M. Z., & Rehman, H. (2012). Effect of supplementation of prebiotic mannan-oligosaccharides and probiotic mixture on growth performance of broilers subjected to chronic heat stress. *Poultry Science*, **91**, 2235-2240.
- Solomon, M., Laack, R., & Eastridge, J. (1998). Biophysical basis of pale, soft, exudative (PSE) pork and poultry muscle: a review. *Journal of Muscle Foods*, **9**, 1-11.
- Sommer, F., & Bäckhed, F. (2013). The gut microbiota-masters of host development and physiology. *Nature Reviews Microbiology*, **11** (4), 227-238.
- Spring, P., Wenk, C., Dawson, K., & Newman, K. (2000). The effects of dietary mannaoligosaccharides on cecal parameters and the concentrations of enteric bacteria in the ceca of salmonella-challenged broiler chicks. *Poultry Science*, **79**, 205-211.

- Steed, H., & Macfarlane, S. (2009). Mechanisms of prebiotic impact on health, prebiotics and probiotics science and technology, New Yourk, 135-161.
- Steiner, T. (2006). Managing gut health: natural growth promoters as a key to animal performance. Nottingham University Press.
- Sugiharto, S. (2014). Role of nutraceuticals in gut health and growth performance of poultry. *Journal of the Saudi Society of Agricultural Sciences*, 1-13. doi:10.1016/j.jssas.2014.06.001.
- Sunvold, G. D., Hussein, H. S., Fahey Jr., J. C., Merchen, N. R., & Reinhart, G. A. (1995). In vitro fermentation of cellulose, beet pulp, citrus pulp, and citrus pectin using fecal inoculum from cats, dogs, horses, humans, and pigs and ruminal fluid from cattle. *Journal of Animal Science*, **73**, 3639-3648.
- Suo, H. Q., Lin, L. U., Xu, G. H., Lin, X. I. A. O., Chen, X. G., Xia, R. R., & Luo, X. G. (2015). Effectiveness of dietary xylo-oligosaccharides for broilers fed a conventional corn-soybean meal diet. *Journal of Integrative Agriculture*, **14** (10), 2050-2057.
- Swanson, K. S., Grieshop, C. M., Flickinger, E. A., Bauer, L. L., Chow, J., Wolf, B. W., Garleb, K. A., & Fahey, G. C. (2002). Fructooligosaccharides and *Lactobacillus acidophilus* modify gut microbial populations, total tract nutrient digestibilities and fecal protein catabolite concentrations in healthy adult dogs. *The Journal of nutrition*, **132**, 3721-3731.
- Świątkiewicz, S., & Arczewska-Włosek, A. (2012). Bone quality characteristics and performance in broiler chickens fed diets supplemented with organic acids. *Czech Journal of Animal Science*, **57**, 193-205.
- Świątkiewicz, S., Koreleski, J., & Arczewska, A. (2010b). Effect of organic acids and prebiotics on bone quality in laying hens fed diets with two levels of calcium and phosphorus. *Acta Veterinaria Brno*, **79**, 185-193.
- Świątkiewicz, S., Koreleski, J., & Arczewska-Włosek, A. (2011). Effect of inulin and oligofructose on performance and bone characteristics of broiler chickens fed on diets with different concentrations of calcium and phosphorus. *British Poultry Science*, **52**, 483-491.
- Tagg, J., & McGiven, A. (1971). Assay system for bacteriocins. *Applied microbiology*, **21** (5), 943.
- Taheri, H., Moghadam, M. K., Kakebaveh, M., & Harakinezhad, T. (2014). Growth performance and immune response of broiler chickens fed diets supplemented with probiotic and (or) prebiotic preparations. *Journal of Livestock Science and Technologies*, **2**, 1-8.
- Tancharoenrat, P., Ravindran, V., Zaefarian, F., & Ravindran, G. (2014). Digestion of fat and fatty acids along the gastrointestinal tract of broiler chickens. *Poultry Science*, **93**, 371-379.

- Teo, A., & Tan, H. M. (2007). Evaluation of the performance and intestinal gut microflora of broilers fed on corn-soy diets supplemented with *Bacillus subtilis* PB6 (CloSTAT). *The Journal of Applied Poultry Research*, **16**, 296-303.
- Thanh, N., Loh, T. C., Foo, H. L., Hair-Bejo, M., & Azhar, B. (2009). Effects of feeding metabolite combinations produced by *Lactobacillus plantarum* on growth performance, faecal microbial population, small intestine villus height and faecal volatile fatty acids in broilers. *British poultry science*, **50**, 298-306.
- Thanh, T. N., Loh, T. C., Foo, H. L., Bejo, M. H., & Kasim, A. (2010). Inhibitory activity of metabolites produced by strains of *Lactobacillus plantarum* isolated from Malaysian fermented food. *International Journal of Probiotics and Prebiotics*, **5**, 37-44.
- Thompson, J. L., & Hinton, M. (1997). Antibacterial activity of formic and propionic acids in the diet of hens on salmonellas in the crop. *British poultry science*, **38**, 59-65.
- Thu, T., Chwen, L.T., Foo, H., Halimatun, Y., & Bejo, M. (2010). Effects of metabolite combinations produced by *Lactobacillus plantarum* on plasma cholesterol and fatty acids in piglets. *American Journal of Animal and Veterinary Sciences*, 233-236.
- Thu, T., Loh, T. C., Foo, H., Yaakub, H., & Bejo, M. (2011). Effects of liquid metabolite combinations produced by *Lactobacillus plantarum* on growth performance, faeces characteristics, intestinal morphology and diarrhoea incidence in postweaning piglets. *Tropical animal health and production*, **43**, 69-75.
- Todorov, S. D., & Dicks, L. M. (2005). Pediocin ST18, an anti-listerial bacteriocin produced by *Pediococcus pentosaceus* ST18 isolated from boza, a traditional cereal beverage from Bulgaria. *Process Biochemistry*, **40**, 365-370.
- Todorov, S. D., Vaz-Velho, M., & Gibbs, P. (2004). Comparison of two methods for purification of plantaricin ST31, a bacteriocin produced by *Lactobacillus plantarum* ST31. *Brazilian Journal of Microbiology*, **35**, 157-160.
- Touchette, K. J., Carroll, J. A., Allee, G. L., Matteri, R. L., Dyer, C. J., Beausang, L. A., & Zannelli, M. E. (2002). Effect of spray-dried plasma and lipopolysaccharide exposure on weaned pigs: I. Effects on the immune axis of weaned pigs. *Journal of Animal Science*, **80**, 494-501.
- Tsai, C. C., Lin, P. P., & Hsieh, Y. M. (2008). Three *Lactobacillus* strains from healthy infant stool inhibit enterotoxigenic *Escherichia coli* grown in vitro. *Anaerobe*, **14** (2), 61-7.
- Tsilingiri, K., Barbosa, T., Penna, G., Caprioli, F., Sonzogni, A., Viale, G., & Rescigno, M. (2012). Probiotic and postbiotic activity in health and disease: comparison on a novel polarised ex-vivo organ culture model. *Gut*, *Gut* doi:10.1136/gutjnl-2011-30097

- Uni, Z. (1999). Functional development of the small intestine in domestic birds: cellular and molecular aspects. *Poultry and Avian Biology Reviews*, **10**, 167-179.
- Uni, Z., Noy, Y., & Sklan, D. (1995). Posthatch changes in morphology and function of the small intestines in heavy- and light-strain chicks. *Poultry Science*, **74**, 1622-1629
- Uraipan, S., & Hongpattarakere, T. (2015). Antagonistic characteristics against food-borne pathogenic bacteria of lactic acid bacteria and bifidobacteria isolated from feces of healthy thai infants. *Jundishapur journal of microbiology*, **8**(6), e18246.
- Vahdatpour, T., Ebrahimnezhad, Y., & Vahdatpour, S. (2014). Effects of dietary functional additives on characteristics and minerals of tibia bone and blood parameters of japanese quails (*coturnix coturnix japonica*). *International Journal of Plant, Animal and Environmental Sciences*, **4** (2), 690-695
- Vamanu, E., & Vamanu, A. (2010). The influence of prebiotics on bacteriocin synthesis using the strain *Lactobacillus paracasei* CMGB16. *African Journal of Microbiology Research*, **4**, 534-537.
- van der Wielen, P. W., Biesterveld, S., Notermans, S., Hofstra, H., Urlings, B. A., & van Knapen, F. (2000). Role of volatile fatty acids in development of the cecal microflora in broiler chickens during growth. *Applied and Environmental Microbiology*, **66**, 2536-2540.
- Van Immerseel, F., Cauwerts, K., Devriese, L., Haesebrouck, F., & Ducatelle, R. (2002). Feed additives to control *Salmonella* in poultry. *World's Poultry Science Journal*, **58**, 501-513.
- Van Immerseel, F., Russell, J., Flythe, M., Gantois, I., Timbermont, L., Pasmans, F., Haesebrouck, F., & Ducatelle, R. (2006). The use of organic acids to combat *Salmonella* in poultry: a mechanistic explanation of the efficacy. *Avian Pathology*, **35**, 182-188.
- Van Laack, R., Liu, C. H., Smith, M., & Loveday, H. (2000). Characteristics of pale, soft, exudative broiler breast meat. *Poultry Science*, **79**, 1057-1061.
- Van Laack, R., Stevens, S., & Stalder, K. (2001). The influence of ultimate pH and intramuscular fat content on pork tenderness and tenderization. *Journal of Animal Science*, **79**, 392-397.
- Van Loo, J. (2007). How chicory fructans contribute to zootechnical performance and well-being in livestock and companion animals. *The Journal of Nutrition*, **137**, 2594S-2597S.
- Van Loo, J., Coussement, P., De Leenheer, L., Hoebregs, H., & Smits, G. (1995). On the presence of inulin and oligofructose as natural ingredients in the western diet. *Critical Reviews in Food Science and Nutrition*, **35**, 525-552.

- Van Meer, G., Voelker, D. R., & Feigenson, G. W. (2008). Membrane lipids: where they are and how they behave. *Nature Reviews Molecular Cell Biology*, **9**(2), 112-124.
- Van Thu, T., Foo, H. L., Loh, T. C., & Bejo, M. H. (2013). Inhibitory activity and organic acid concentrations of metabolite combinations produced by various strains of *Lactobacillus plantarum*. *African Journal of Biotechnology*, **10**, 1359-1363.
- Velasco, S., Ortiz, L., Alzueta, C., Rebolé, A., Treviño, J., & Rodriguez, M. (2010). Effect of inulin supplementation and dietary fat source on performance, blood serum metabolites, liver lipids, abdominal fat deposition, and tissue fatty acid composition in broiler chickens. *Poultry Science*, **89**, 1651-1662.
- Vilà, B., Esteve-Garcia, E., & Brufau, J. (2010). Probiotic micro-organisms: 100 years of innovation and efficacy; modes of action. *World's Poultry Science Journal*, **66**, 369-380.
- Viniegra Gonzalez, G., & Gomez, J. (1984). Lactic acid production by pure and mixed bacterial cultures.
- Vispo, C., & Karasov, W. H. (1997). The interaction of avian gut microbes and their host: an elusive symbiosis. *Gastrointestinal microbiology*. USA, 116-155.
- Vogt, H., Mathes, S., & Harnisch, S. (1982). The effect of organic acids on productivity of broilers. *Archives fur Geflugelkunde*, **46**, 223-227.
- Von Mollendorff, J., Todorov, S., & Dicks, L. (2006). Comparison of bacteriocins produced by lactic-acid bacteria isolated from boza, a cereal-based fermented beverage from the Balkan Peninsula. *Current Microbiology*, **53**, 209-216.
- Waite, J. G., Jones, J. M., & Yousef, A. E. (2009). Isolation and identification of spoilage microorganisms using food-based media combined with rDNA sequencing: ranch dressing as a model food. *Food Microbiology*, **26**, 235-239.
- Waldenstedt, L. (2006). Nutritional factors of importance for optimal leg health in broilers: A review. *Animal Feed Science and Technology*, **126**, 291-307.
- Wang, C. Y., Lin, P. R., Ng, C. C., & Shyu, Y. T. (2010). Probiotic properties of Lactobacillus strains isolated from the feces of breast-fed infants and Taiwanese pickled cabbage. *Anaerobe*, **16** (6), 578-85.
- Wang, J., Zhang, H., Chen, X., Chen, Y., & Bao, Q. (2012). Selection of potential probiotic lactobacilli for cholesterol-lowering properties and their effect on cholesterol metabolism in rats fed a high-lipid diet. *Journal of Dairy Science*, **95**, 1645-1654.
- Wang, R. F., Cao, W. W., & Cerniglia, C. E. (1996). PCR detection and quantitation of predominant anaerobic bacteria in human and animal fecal samples. *Applied and Environmental Microbiology*, **62** (4), 1242-1247.

- Wang, Y. (2009). Prebiotics: Present and future in food science and technology. *Food Research International*, **42**, 8-12.
- Willard, M. D., Simpson, R. B., Cohen, N. D., Clancy, J.S. (2000). Effects of dietary fructooligosaccharide on selected bacterial populations in feces of dogs. *American Journal of Veterinary Research*, **61**, 820-825.
- Williams, B., Solomon, S., Waddington, D., Thorp, B., & Farquharson, C. (2000). Skeletal development in the meat-type chicken. *British Poultry Science*, **41**, 141-149.
- Williams, P., & Losa, R. (2001). The use of essential oils and their compounds in poultry nutrition. *World Poultry*, **17**, 14-15.
- Woo, K., Kim, C., & Paik, I. (2007). Effects of supplementary immune modulators (MOS, Lectin) and organic acid mixture (organic acid F, organic acid G) on the performance, profile of leukocytes and erythrocytes, small intestinal microflora and immune response in laying hens. *Journal of Animal Science and Technology*, **49**, 481-490.
- Xu, Z.R., Hu C.H., Xia M.S., Zhan X.A., Wang M.D. (2003). Effects of dietary fructooligosaccharides on digestive enzyme activities, intestinal microflora and morphology of male broilers. *Poultry Science* **82**, 1030-1036
- Yalçinkaya, İ., Guengoer, T., Başalan, M., & Erdem, E. (2008). Mannan oligosaccharides (MOS) from *Saccharomyces cerevisiae* in broilers: Effects on performance and blood biochemistry. *Turkish Journal of Veterinary and Animal Sciences*, **32**, 43-48.
- Yang, C. M., Cao, G. T., Ferket, P. R., Liu, T. T., Zhou, L., Zhang, L., & Chen, A. G. (2012). Effects of probiotic, *Clostridium butyricum*, on growth performance, immune function, and cecal microflora in broiler chickens. *Poultry Science*, **91**(9), 2121-2129.
- Yang, C., & Chen, T. (1993). Effects of refrigerated storage, pH adjustment, and marinade on color of raw and microwave cooked chicken meat. *Poultry Science*, **72**, 355-362.
- Yang, E., Fan, L., Jiang, Y., Doucette, C., & Fillmore, S. (2012). Antimicrobial activity of bacteriocin-producing lactic acid bacteria isolated from cheeses and yogurts. *AMB Express*, **2**, 1-12.
- Yang, Y., Iji, P., & Choct, M. (2007). Effects of different dietary levels of mannanoligosaccharide on growth performance and gut development of broiler chickens. *Asian Australasian Journal of Animal Sciences*, **20**, 1084.
- Yang, Y., Iji, P., & Choct, M. (2009). Dietary modulation of gut microflora in broiler chickens: a review of the role of six kinds of alternatives to in-feed antibiotics. *World's Poultry Science Journal*, **65**, 97-114.



- Yasuda, K., Ronecker, K. R., Miller, D., Welch, R. M., & Lei, X. G. (2006). Supplemental dietary inulin affects the bioavailability of iron in corn and soybean meal to young pigs. *Journal of Nutrition*, **136**, 3033-3038.
- Yegani, M., & Korver, D. (2008). Factors affecting intestinal health in poultry. *Poultry science*, **87**, 2052-2063.
- Yin, Y., Tang, Z., Sun, Z., Liu, Z., Li, T., Huang, R., Ruan, Z., Deng, Z., Gao, B., & Chen, L. (2008). Effect of galacto-mannan-oligosaccharides or chitosan supplementation on cytoimmunity and humoral immunity in early-weaned piglets. *Asian-Australasian Journal of Animal Sciences*, **21**, 723.
- Younes, H., Coudray, C., Bellanger, J., Demigné, C., Rayssiguier, Y., & Rémésy, C. (2001). Effects of two fermentable carbohydrates (inulin and resistant starch) and their combination on calcium and magnesium balance in rats. *British Journal of Nutrition*, **86**, 479-485.
- Zacharof, M., & Lovitt, R. (2012). Bacteriocins produced by lactic acid bacteria a Review Article. *APCBEE Procedia*, **2**, 50-56.
- Zhang, A., Lee, B., Lee, S., Lee, K., An, G., Song, K., & Lee, C. (2005). Effects of yeast (*Saccharomyces cerevisiae*) cell components on growth performance, meat quality, and ileal mucosa development of broiler chicks. *Poultry Science*, **84**, 1015-1021.
- Zhao, P. Y., Wang, J. P., Kim, I. H. (2013). Evaluation of dietary fructan supplementation on growth performance, nutrient digestibility, meat quality, fecal microbial flora, and fecal noxious gas emission in finishing pigs. *Journal of Animal Science*, **91**, 5280-5286.
- Zhou, X., Wang, Y., Gu, Q., & Li, W. (2010). Effect of dietary probiotic, *Bacillus coagulans*, on growth performance, chemical composition, and meat quality of Guangxi Yellow chicken. *Poultry science*, **89**, 588-593.
- Zhuang, H. Y. (2007). Effects of xylooligosaccharides on growth performance, meat quality and PRKAG3 gene expression in growing-finishing pig. MSc thesis, Chinese Academy of Agricultural Sciences, China.
- Zhuang, S., Jiang, F. B., Jia, Z. X., & Yan, R. (2015). *Clostridium butyricum* can be used as a potential alternative for the antibiotic in Cherry Valley ducks. *Journal of Animal and Plant Sciences*, **25** (5), 1227-1232.
- Ziaie, H., Bashtani, M., Torshizi, M. K., Naeemipour, H., Farhangfar, H., & Zeinali, A. (2011). Effect of antibiotic and its alternatives on morphometric characteristics, mineral content and bone strength of tibia in ross broiler chickens. *Global Veterinary*, **7**, 315-322.

Zulkifli, I., Abdullah, N., Azrin, N. M., & Ho, Y. (2000). Growth performance and immune response of two commercial broiler strains fed diets containing *Lactobacillus* cultures and oxytetracycline under heat stress conditions. *British Poultry Science*, **41**, 593-597.

Zuppa, A. A., Sindico, P., Orchi, C., Carducci, C., Cardiello, V., Catenazzi, P., & Romagnoli, C. (2010). Safety and efficacy of galactogogues: substances that induce, maintain and increase breast milk production. *Journal of Pharmacy & Pharmaceutical Sciences*, **13**, 162-174.

