



UNIVERSITI PUTRA MALAYSIA

***ANTIBACTERIAL ACTIVITY AND PHYTOCHEMICAL ANALYSIS OF
Garcinia mangostana L. LEAF EXTRACTS AGAINST Xanthomonas
oryzae
pv. oryzae AND Pseudomonas syringae pv. TOMATO***

QAMAR MOHAMMED NAJI

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By
QAMAR MOHAMMED NAJI

Thesis Submitted to the School of Graduate Studies, Universiti Putra
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Master of Science

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

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By

QAMAR MOHAMMED NAJI

November 2016

**Chairman : Associate Professor Kamaruzaman Sijam, PhD
Faculty : Agriculture**

The immense diversity of plant pathogens, which include viruses, bacteria, fungi, nematodes, and insects, approximates 7100 species. Among these, roughly 150 are bacterial species that cause diseases to plants. Bacterial plant diseases are most frequent and severe in tropical and subtropical places, where warm and humid conditions like Malaysia are ideal for bacterial growth. Indeed, consistent annual crop losses are recorded in all countries.

The problem of plant diseases is depended on chemical control as antibiotic. This chemical control is very expensive and yet not very effective against for many bacterial diseases. In this study, the leaf of mangosteen was used to prepare extract to bio-control for two types of plant pathogens namely *Pseudomonas syringe* pv. *tomato* and *Xanthomonas oryzae* pv. *Oryzae*.

The potential of mangosteen (*Garcinia mangostana* L.) leaf extract as a biological control agent against plant pathogenic bacteria which are responsible to decrease the quality and volume of crop production worldwide was assessed. Extract was obtained by maceration of the leaves using chloroform, n-hexane, and methanol. Crude extracts of about 1.45 % were derived using chloroform, 1.25 % using n-hexane, and 1.65 % using methanol leaf crude. Compared to chloroform and n-hexane, effective extraction of readily soluble compounds was observed in case of methanol, as the highest yield was collected from it.

For the in-vitro antibacterial activity, two plant pathogenic bacteria, namely *Pseudomonas syringe* pv. *tomato* and *Xanthomonas oryzae* pv. *oryzae* were acquired. Four different concentrations of 12.5, 25, 50, and 100 mg/mL were used through the cup-plate agar diffusion technique. Streptomycin sulphate at 30 µg/mL concentration was set as a positive control, whereas every respective solvent used in the leaf extraction was set as negative controls. The highest diameter value of inhibition zone was observed in *P. syringe* pv. *tomato* at all range of concentrations, followed by *X. oryzae* pv. *oryzae*.

Since only the methanol extract demonstrated antibacterial activity, it was the only solvent subjected in the assay for minimum bactericidal concentration (MBC) and minimum inhibitory concentration (MIC) determination. The least methanol extract concentration utilised in MIC assay was at 1.56 mg/mL, inhibiting *X. oryzae* pv. *oryzae* followed by *P. syrinx* pv. *tomato* at a concentration of 3.13 mg/mL. This assay indicated that methanol extract caused bactericidal impacts at concentrations of 1.56 mg/ml and 3.13 mg/ml for varying plant pathogenic bacteria species.

The least concentration of MBC noted was at 3.13 mg/mL against *X. oryzae* pv. *oryzae* and 6.25 mg/mL against *P. syrinx* pv. *tomato*. Mangosteen methanol leaf extracts' primary phytochemical screening indicated the existence of flavonoids, alkaloids, saponins, tannins, anthraquinones, terpenoids, and phenols. When visualised in the thin layer chromatography (TLC) profiling of methanol extract using acetone and n-hexane in a ratio of 6:4 (v/v) gave 11 maximum colourful bands.

Retention factor (Rf) values proved the presence of various active secondary metabolites within the methanol extract. Methanol extract's antibacterial activity was screened through the direct bioautography procedure. The intention was to determine the location of active bands on chromatograms developed for TLC profiling in the same manner. The most active Rf values that inhibited every tested plant pathogenic bacteria at the same location of Rf values were noted at 0.93, 0.86, 0.66, 0.46, 0.33 and 0.16. A comparison of the Rf values acquired was made with earlier studies utilising the same solvent system. Antibacterial impacts of the most effectual extract of mangosteen crude were supported by the existence of chemical components identified by Gas Chromatography-Mass Spectrometry (GC-MS).

The *G. mangostana* leaf extract was exhibited good potential to be used to inhibit growth of *Xanthomonas oryzae* pv. *oryzae* and *Pseudomonas syringae* pv. *tomato* invitro. The results showed that methanol extract demonstrated antibacterial activity when tested on the plant pathogenic bacteria in-vitro. On the other hand, chloroform and n-hexane did not exhibit any antibacterial activity against plant pathogenic bacteria, as there was no inhibition zone noted under these treatments. Cycloartenol, caryophyllene, docosane, and 4, 4-methylenebis (2, 6-di-tert-butylphenol) were noted as key compounds in the mangosteen leaf extract.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Master Sains

AKTIVITI ANTIBAKTERIA DAN ANALISIS PHYTOKIMIA *Garcinia mangostana* L. LEAF EKSTRAK TERHADAP *Xanthomonas oryzae* pv. *oryzae* dan *Pseudomonas syringae* pv. *TOMATO*

Oleh

QAMAR MOHAMMED NAJI

November 2016

Pengerusi : Profesor Madya Kamaruzaman Sijam, PhD
Fakulti : Pertanian

Kepelbagaiannya besar patogen tumbuhan, termasuk virus, bakteria, kulat, nematod dan serangga, lebih kurang 7100 spesies. Antaranya, kira-kira 150 spesies bakteria yang menyebabkan penyakit kepada tumbuhan. Penyakit tumbuhan bakteria yang paling kerap dan teruk di tempat-tempat tropika dan subtropika, di mana keadaan panas dan lembap seperti Malaysia adalah sesuai untuk pertumbuhan bakteria. Sesungguhnya, selaras kerugian tanaman tahunan direkodkan di semua negara.

Masalah penyakit tumbuhan adalah bergantung kepada kawalan kimia sebagai antibiotik. Ini kawalan kimia adalah sangat mahal tetapi tidak sangat berkesan terhadap pelbagai penyakit bakteria. Dalam kajian ini, daun manggis telah digunakan untuk menyediakan ekstrak untuk bio-kawalan untuk dua jenis patogen tumbuhan iaitu *Pseudomonas syringae* pv. *tomato* dan *Xanthomonas oryzae* pv. *Oryzae*. Potensi *Garcinia mangostana* sebagai agen kawalan biologi terhadap bakteria penyakit tumbuhan atau mikrob memudaratkan yang mengurangkan kualiti dan jumlah pengeluaran tanaman di seluruh dunia telah dinilai. Daun manggis diekstrak secara maksima menggunakan chloroform, n-heksana dan metanol.

Ekstrak mentah kira-kira 1.45% telah diperolehi menggunakan kloroform , 1.25 % menggunakan n- heksana, dan 1.65 % menggunakan metanol daun mentah. Berbanding dengan kloroform dan heksana, pengekstrakan sebatian mudah larut adalah lebih berkesan di dalam metanol, dengan hasil tertinggi dikumpulkan daripadanya. Bagi aktiviti anti-bakteria secara in-vitro, dua spesies berbeza bakteria penyakit tumbuhan, iaitu *Pseudomonas syringae* pv. *tomato* dan *Xanthomonas oryzae* pv . *oryzae* diperolehi. Empat kepekatan berbeza, 12.5, 25, 50 dan 100 mg/ml yang diperolehi melalui teknik cawan-plat serapan agar. Streptomycin sulfate pada kepekatan 30 µg/ml telah disetkan sebagai kawalan positif, dan setiap pelarut yang digunakan di dalam pengekstrakan daun disetkan sebagai kawalan negatif. Zon perencutan paling

tinggi diperhatikan pada *P. syrange* pv. *tomato* pada pelbagai julat kepekatan, diikuti oleh *X. oryzae* pv. *oryzae*. Oleh kerana hanya ekstrak methanol mempamerkan aktiviti anti-bakteria, hanya pelarut ini sahaja digunakan untuk menentukan percubaan kepekatan bakterisidal minimum (MBC) dan kepekatan perencatan minimum (MIC). Kepekatan ekstrak methanol yang paling sedikit digunakan di dalam MIC adalah pada 1.56 mg/ml, merentangkan *X. oryzae* pv. *oryzae*, diikuti oleh *P. syringae* pv. *tomato* pada kepekatan 3.13 mg/ml. percubaan ini menunjukkan ekstrak methanol menyebabkan kesan bakteriostatik pada kepekatan 1.56 mg/ml dan 3.13 mg/ml untuk pelbagai spesies bakteria penyakit tumbuhan.

Kepekatan yang paling sedikit untuk MBC telah dicatatkan pada 3.13 mg/ml terhadap *X. oryzae* pv. *oryzae* dan 6.25 mg/ml terhadap *P. syringae* pv. *tomato*. Pemeriksaan fitokimia utama bagi ekstrat metanol daun manggis telah menyatakan kewujudan flavonoid, alkaloid, saponin, tannin, anthraquinone, terpenoid dan phenol. Apabila diperhatikan, pemprofilan kromatografi lapisan nipis (TLC) bagi ekstrak metanol menggunakan aseton dan heksana dalam nisbah 6:4 (v/v) telah memberikan maksimum 11 jalur warna warni. Nilai Rf mengesahkan kehadiran pelbagai metabolit sekunder yang aktif di dalam ekstrak metanol.

Aktiviti anti-bakteria bagi ekstrak metanol telah dijalankan melalui prosedur bio-autografi secara langsung. Ianya bertujuan untuk menentukan lokasi jalur yang aktif pada kromatogram yang telah dibuat bagi pemprofilan TLC dalam perkara yang sama. Nilai faktor pengekalan yang merentangkan setiap bakteria penyakit tumbuhan pada lokasi Rf yang sama telah dicatatkan pada 0.93, 0.86, 0.66, 0.46, 0.33 dan 0.16. Suatu perbandingan nilai Rf yang diperolehi telah dibuat melalui kajian awal menggunakan sistem pelarut yang sama. Kesan anti-bakteria bagi ekstrak mentah manggis yang paling berkesan telah disokong melalui kehadiran komponen kimia yang dikenalpasti melalui GC-MS.

G. ekstrak daun mangostana dipamerkan potensi yang baik untuk digunakan untuk menghalang pertumbuhan *Xanthomonas oryzae* pv. *oryzae* dan *Pseudomonas syringae* pv. *tomato* invitro. Hasil kajian menunjukkan bahawa ekstrak metanol menunjukkan aktiviti anti-bakteria apabila diuji pada tumbuhan bakteria patogenik in-vitro. Sebaliknya, kloroform dan n-heksana tidak menunjukkan apa-apa aktiviti antibakteria terhadap tumbuhan bakteria patogenik, kerana tidak ada zon perencatan dinyatakan di bawah rawatan ini. Cycloartenol, caryophyllene, docosane, dan 4, 4-methylenebis (2, 6-di-tert-butylphenol) telah diperhatikan sebagai sebatian utama dalam ekstrak daun manggis.

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Kamaruzaman Sijam, PhD

Associate Professor

Agriculture

Universiti Putra Malaysia

(Chairman)

Khairulmazmi Ahmad, PhD

Senior Lecturer

Agriculture

Universiti Putra Malaysia

(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

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Signature:

Name of Chairman
of Supervisory
Committee:

Associate Professor Dr. Kamaruzaman Sijam

Signature:

Name of Member
of Supervisory
Committee:

Dr. Khairulmazmi Ahmad

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xii
LIST OF FIGURES	xiii
LIST OF ABBREVIATIONS	xiv
 CHAPTER	
1 INTRODUCTION	1
1.2 Problem Statements	2
1.3 Objectives	2
2 LITERATURE REVIEW	
2.1 Description of Mangosteen	4
2.2 Uses of Mangosteen	4
2.2.1 Traditional uses of mangosteen	4
2.2.2 Food uses of mangosteen	5
2.2.3 Cosmetic uses of mangosteen	5
2.3 Activity of Mangosteen in the Antioxidant	5
2.4 Activity of Mangosteen in the ntimicrobial	6
2.5 Chemical composition of mangosteen	7
2.6 Effect of plant pathogenic bacteria	7
2.7 Genus Xanthomonas	8
2.8 Genus Pseudomonas	9
2.9 Control practices of phytopathogenic bacteria	10
2.10 Biological Control	12
2.10.1 Biocontrol	12
2.10.2 Botanicals	12
2.11 Major Groups of Plant Secondary Metabolites	14
2.12 Thin-Layer Chromatography-Direct Bioautography	18
2.13 Gas Chromatography-Mass Selective (GC-MS)	18
3 IN-VITRO ANTIBACTERIAL ACTIVITIES OF LEAF CRUDE EXTRACT OF MANGOSTEEN AGAINST <i>Xanthomonas oryzae</i> pv. <i>oryzae</i> AND <i>Pseudomonas syringae</i> pv. tomato	
3.1 Introduction	19
3.2 Materials and methods	20
3.2.1 Plant materials	20
3.2.2 Preparation of Mangosteen leaf extracts	20
3.2.3 Preparation of bacterial culture	21
3.2.4 Preparation of cell suspension for	22

	antibacterial assay	
3.2.5	In-vitro evaluation of antibacterial activity	22
3.2.6	Determination of minimum inhibitory concentration (MIC)	23
3.2.7	Minimum bacterial concentration (MBC)	23
3.2.8	Statistical analysis	24
3.3	Results	24
3.3.1	Mangosteen extracts physical characteristics and yield percentage	24
3.3.2	In-vitro evaluation of antibacterial activity for mangosteen leaf extracts against plant pathogenic bacteria	25
3.3.3	The MIC and MBC	27
3.4	Discussion	29
3.5	Conclusion	30
4	DETERMINATION OF PHYTO COMPONENTS AND BIOAUTOGRAPHY OF <i>Garcinia mangostana L.</i> METHANOL LEAF EXTRACT	
4.1	Introduction	31
4.2	Materials and Methods	32
4.2.1	Preparation of mangosteen leaf extract	32
4.2.2	phytochemical screening	32
4.2.3	Qualitative Analysis by thin Layer Chromatography (TLC)	34
4.2.4	Bioautography	34
4.2.5	Determination of Phytocompounds using Gas Chromatography-Mass Spectrometry (GC-MS)	35
4.2.6	Identification of components by GC-MS	35
4.3	Results	36
4.3.1	Phytochemical Screening	36
4.3.2	TLC analysis of methanol crude extract	37
4.3.3	Direct TLC bioautography assay	38
4.3.4	Analysis of the phytocompounds from methanol crude extract using Gas Chromatography-Mass Spectrometry (GC-MS)	40
4.4	Discussion	42
4.5	Conclusion	44
5	SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	
5.1	Summary	45
5.2	General conclusion	47
5.3	Recommendations	48
REFERENCES		49
APPENDICES		62
BIODATA OF STUDENT		75
PUBLICATION		76

LIST OF TABLES

Table		Page
3.1	Identification and confirmation of plant pathogenic bacteria	22
3.2	Physical characteristics of mangosteen leaf extracts	24
3.3	The percentage of mass yield of different crude extracts of mangosteen leaf	24
3.4	Antibacterial activity of <i>G. mangostana</i> leaf crude extracts against pathogenic bacteria using agar well diffusion method	26
3.5	Minimal inhibitory concentration (MIC) and Minimal bactericidal concentration (MBC) of mangosteen methanol leaf extract against pathogenic bacteria	27
3.6	The LC50 and LC90 values of the methanol extract of Mangosteen against the selected bacteria	29
4.1	Phytochemical screening of mangosteen methanol leaf extract	36
4.2	The retention factor (Rf) for <i>G. mangostana</i> L. methanol leaf extract with different visualization technique	38
4.3	Retention factor (Rf) values of inhibition zone from <i>G. mangostana</i> L. methanol leaf extract against <i>P. syringae</i> and <i>X. oryzae</i>	39
4.4	Retention factor (Rf) values of inhibition zone from <i>G. mangostana</i> L. methanol leaf extract against <i>X. oryzae</i>	39
4.5	The major highest placated components exhibited in the methanol crude extract of mangosteen identified based on GC-MS analysis	41

LIST OF FIGURES

Table	Page
3.1 Preparation stages of crude extract of leaves <i>G. mangosteen</i> : collection young fresh leave (A), Dried leave (B), fine powder of leaves (C) and getting crude extract (D).	21
3.2 Pure bacterial culture (A) <i>Pseudomonas syringe</i> pv. <i>tomato</i> showing yellow-light and (B) <i>Xanthomonas oryzae</i> pv. <i>oryzae</i> growing on NA showing yellow pigment	22
3.3 Antibacterial activity of <i>G. mangostana</i> methanol leaf crude extract against <i>P. syringae</i> pv. <i>tomato</i> (A) and <i>X. oryzae</i> pv. <i>oryzae</i> (B). (1): Positive control; (2):100; (3):50; (4):25; (5):12.5 mg/ml and (6): negative control	26
3.4 Minimal inhibitory concentration (MIC) of mangosteen methanol leaf extracts against <i>Xanthomonas oryzae</i> pv. <i>oryzae</i> by macro-broth dilution method	27
3.5 Minimal inhibitory concentration (MIC) of mangosteen methanol leaf extracts against <i>Pseudomonas syringe</i> pv. <i>tomato</i> by macro-broth dilution method	28
3.6 The minimum bactericidal concentration (MBC) of mangosteen methanol leaf extract against <i>X. oryzae</i> and <i>P. syringe</i>	28
4.1 The positive results obtained for the phytochemical screening of mangosteen methanol leaf extract for detection: T1 (Cardiac glycosides), T2 (Alkaloids), T3 (Tannins), T4 (Saponins), T5 (Phenol), T6 (Terpenoids), T7 (Flavonoids), T8 (Anthraquinone).	36
4.2 TLC spot visualization under daylight (normal light) (A), short UV wavelength (245 nm) (B), and long UV wavelength (365 nm) (C).	37
4.3 Direct bioautography of <i>G. mangostana</i> L. methanol leaf extract against <i>P. syringae</i> (A), and <i>X. oryzae</i> (B). Inhibition zones were circled due to low light quality during the photography session.	39
4.4 GC-MS chromatogram analysis of methanol leaf extract of mangosteen	40

LIST OF ABBREVIATIONS

ANOVA	Analysis of Variance
cfu/ml	Colony forming unit per milliliter
CRD	Completely Randomized Design
cm	Centimeter
GC-MS	Gas chromatography mass spectrometry
TLC	Thin layer chromatography
TTC	2,3,5- Triphenylterazolium chloride
GML	<i>G. mangostana</i> L
Kg	Kilogram
LCB	lacto phenol-Cotton blue
LSD	least significant difference
M	Molar
MeOH	Methanol
MBC	Minimum Bactericidal Concentration
MIC	Minimum inhibitory concentration
MHA	Muller Hinton Agar
MHB	Muller Hinton Broth
µl	Microliter
Mm	Mill molar
MI	Millilitre
mL min-1	Millilitre per minute
µm	Micrometer
NA	Nutrient agar
NB	Nutrient broth
OD	Optical density
± SEM	Standard error of means
Temp	Temperature
v/v	Volume per volume
w/v	Weight per volume

CHAPTER 1

INTRODUCTION

1.1 Introduction

The rate of plant production worldwide is severely affected due to the presence of plant diseases. There are more than 200 types of diseases that affect plants, out of which 70% of the diseases are caused by fungal and bacterial infections (Janisiewicz et.al. 2001). The bacterial plant pathogens play havoc with the economy as they can destroy the economically relevant crops. They also affect the quality of the commercial crops. Hence, they need to be properly controlled. However, the major problem of effective bacterial pathogens controlled in the orchards is the unavailability of proper commercial antibacterial agents. The available antibiotics have a questionable efficacy and are banned in several countries, and they are also not very effective against many pathogenic agents.

Unfortunately, there are several reports that state the control of the fungal outbreaks is possible using chemicals compounds but, very few reports have stated the control of the bacterial diseases using nonchemicals. Hence, it is important to find alternative means for controlling the bacterial diseases, rather than using chemical means, and therefore, there is an increase in re-examining and improving several traditional practices and techniques and developing new practices for controlling bacterial crop diseases (Moazami, 2008).

The most popular botanical pesticides include the phenolic-rich plant extracts. The plant extracts which are rich in tannins have been widely used for the treatment of human infectious diseases (Tegegne, 2008). Moreover, they display antimicrobial activities against several bacterial species and the phytopathogenic fungi.

Mangosteen belonging to the Clusiaceae family is called as the “the queen of fruits”. This fruit is cultivated widely in the tropical rainforests of the Southeast Asian countries like Malaysia. The mangosteen pericarp has been used in traditional medicines for treating disorders like diarrhoea, dysentery, abdominal pain, suppuration, infected wounds, and chronic ulcers. There are reports in the literature which have stated that the leaf of mangosteen extracts contain antitumor, antiviral, antibacterial, anti-allergic, antioxidant, and anti-inflammatory activities (Pedraza-Chaverri et.al. 2008).

The xanthones can be extracted from the pericarp, heartwood, whole fruit, and leaves of GML. The mangosteen extracts were widely used in the medical field and pharmacology for the treatment of several animal and human diseases. The *X. oryzae* pv. *oryzae*, is a very destructive and globally found bacterium, which mainly infects the rice crop and can lead to epidemic infections among

the high-yielding cultivars present in the temperate and the tropical regions of Asia (Mew et al; 1993; Young et al., 2008). Also, the *Pseudomonas syringae* pv. *tomato* (Okabe) causes the bacterial speck disease in tomatoes and this is a major disease afflicting the tomato plant worldwide (Milijašević, 2005).

Three major experiments have been done to achieve the objectives of this study. The *in-vitro* antibacterial activity assay was used through the cup-plate agar diffusion technique to determine the efficacy of the *G. mangostana* L plant extract. (MIC) Minimal Inhibitory Concentration and Minimum Bactericidal Concentration (MBC). Assay were used to determine the lowest concentrations of the most effective plant extracts.

Bioautography procedure enables recognition of known antimicrobial compounds in extracts. Gas Chromatography Mass Spectrometry (GC-MS) is the best method used to division, classification, and quantification of organic volatile and semi-volatile compounds.

1.2 Problem Statements

Chemical control is a very expensive process, and moreover, it is also not very effective against certain pathogens. Hence, developing biological control measures can help to manage the crop diseases. However, the biological controlling measures should also have additional economic attributes for making them more commercially viable. Since a very long time, the natural bioactive components, also called as herbal medicines or botanical pesticides have been used to contain the spread of plant diseases and killing the pathogens as well. However, the increase in the use of chemical pesticides in the agricultural sector has decreased the application of the botanical pesticides. Though the chemical pesticides are very effective, they have harmful side effects e.g., they damage the environment and are toxic to human and animal health.

1.3 Objectives

This study aim to prepare extracts from Mangostana leaves for antibacterial activity for two types of bacterial which are *Xanthomonas oryzae* pv. *oryzae* and *Pseudomonas Syringae* pv. *tomato*. and focusing on the following objectives:

- 1) To screen antibacterial activity of crude leaf of the *G. mangostana* L extracts against *Xanthomonas oryzae* pv. *oryzae* and *Pseudomonas syringae* pv. *tomato*. and determine the Minimal Inhibitory Concentration (MIC) and Minimum Bactericidal Concentration (MBC) of the active crude extract against two plant pathogen bacteria in vitro.

- 2) To determine the phytochemical in the active crude extracts of leaf *G. mangostana L.* using Thin Layer Chromatography (TLC) determination of phytocompound constituents by using Gas Chromatography Mass Spectrometry GC-MS.

REFERENCES

- Al-Daihan, S., Al-Faham, M., Al-shawi, N., Almayman, R., Brnawi, A., & shafi Bhat, R. (2013). Antibacterial activity and phytochemical screening of some medicinal plants commonly used in Saudi Arabia against selected pathogenic microorganisms. *Journal of King Saud University-Science*. 25(2): 115-120.
- Al-Massarani, S. M., El Gamal, A. A., Al-Musayeib, N. M., Mothana, R. A., Basudan, O. A., Al-Rehaily, A. J., ... & Maes, L. (2013). Phytochemical, antimicrobial and antiprotozoal evaluation of *Garcinia mangostana* pericarp and α-mangostin, its major xanthone derivative. *Molecules*. 18 (9): 10599-10608.
- Arifullah, M., Vikram, P., Chiruvella, K. K., Shaik, M. M., & Ripain, I. H. B. A. (2014). A review on Malaysian plants used for screening of antimicrobial activity. *Annual Research & Review in Biology*. 4(13): 2088.
- Ashok, P. K., & Upadhyaya, K. (2012). Tannins are astringent. *Journal of Pharmacognosy and Phytochemistry*, 1(3):45-53.
- Asif, M. (2015). Pharmacological activities and phytochemistry of various plant containing coumarin derivatives. *Current Science*. 1(3): 77-90.
- Baharvand-Ahmadi, B., Bahmani, M., Eftekhari, Z., Jelodari, M., & Mirhoseini, M. (2016). Overview of medicinal plants used for cardiovascular system disorders and diseases in ethnobotany of different areas in Iran. *Journal of HerbMed Pharmacology*. 5(1):39-44.
- Basile, A., Giordano, S., López-Sáez, J. A., & Cobianchi, R. C. (1999). Antibacterial activity of pure flavonoids isolated from mosses. *Phytochemistry*, 52(8), 1479-1482.
- Basri, D. F., Tan, L. S., Shafiei, Z., & Zin, N. M. (2011). In vitro antibacterial activity of galls of *Quercus infectoria* Olivier against oral pathogens. *Evidence-Based Complementary and Alternative Medicine*. 2012: 1-7.
- Bernhoft, A., Siem, H., Bjertness, E., Meltzer, M., Flaten, T., Holmsen, E., & Aaseth, J. O. (2010). Bioactive compounds in plants—benefits and risks for man and animals. In *Proceedings from a Symposium Held at The Norwegian Academy of Science and Letters, Novus forlag, Oslo*.
- Bhandary, S. K., Kumari, N. S., Bhat, V. S., Sharmila, K. P., & Bekal, M. P. (2012). Preliminary phytochemical screening of various extracts of *Punica granatum* peel, whole fruit and seeds. *Journal of Health Science*. 2(4): 35-38.
- Bhatti, H. N., & Khera, R. A. (2014). Biotransformations of diterpenoids and triterpenoids: a review. *Journal of Asian natural products research*, 16(1), 70-104.

- Biradar, S. R., & Rachetti, B. D. (2013). Extraction of some secondary metabolites & thin layer chromatography from different parts of *Centella asiatica* L.(URB). American Journal of Life Science. 1(6): 243-247.
- Biresaw, G., & Mittal, K. L. (Eds.). (2014). *Surfactants in tribology* (Vol. 4). CRC Press. 597 pp.
- Blumenthal, M. (2009). African Natural Plant Products: A Foreword to the Science and Challenges. *African Natural Plant Products: New Discoveries and Challenges In Chemistry and Quality*. 3-5.
- Butsat, S., & Siriamornpun, S. (2016). Effect of solvent types and extraction times on phenolic and flavonoid contents and antioxidant activity in leaf extracts of *Amomum chinense* C. International Food Research Journal. 23(1).
- Chanarat, P., Chanarat, N., Fujihara, M., Nagumo, T., (1997). Immunopharmacological activity of polysaccharide from the pericarp of mangosteen *garcinia*: phagocytic intracellular killing activities. J. Med. Assoc. Thai. 80: S149– S154.
- Charernsriwilaiwat, N., Rojanarata, T., Ngawhirunpat, T., Sukma, M., & Opanasopit, P. (2013). Electrospun chitosan-based nanofiber mats loaded with *Garcinia mangostana* extracts. International journal of pharmaceutics. 452(1): 333-343.
- Cheenpracha, S., Jittonnom, J., Komek, M., Ritthiwigrom, T., & Laphookhieo, S. (2016). Acetylcholinesterase inhibitory activity and molecular docking study of steroid alkaloids from *Holarrhena pubescens* barks. *Steroids*.
- Chin, Y. W., & Kinghorn, A. D. (2008). Structural characterization, biological effects, and synthetic studies on xanthones from mangosteen (*Garcinia mangostana*), a popular botanical dietary supplement. *Mini-reviews in organic chemistry*. 5(4): 355.
- Chin, Y.W., Jung, H.A., Chai, H., Keller, W.J., Kinghorn, A.D. (2008). Xanthones with quinone reductase-inducing activity from the fruits of *Garcinia mangostana* (Mangosteen). *Phytochemistry*. 69: 754–758.
- Choma, I. M., & Grzelak, E. M. (2011). Bioautography detection in thin-layer chromatography. *Journal of Chromatography A*. 1218(19): 2684-2691.
- Chung, E. J., Khan, A., & Jeon, C. O. (2015). The Plant Pathology Journal: Open Access; *Bacillus oryzicola* sp. nov., an Endophytic Bacterium Isolated from the Roots of Rice with Antimicrobial, Plant Growth Promoting, and Systemic Resistance Inducing Activities in Rice. *The Plant Pathology Journal*. 31(2): 152-164.
- Cicerale, S., Lucas, L., & Keast, R. (2010). Biological activities of phenolic compounds present in virgin olive oil. *International Journal of Molecular Sciences*. 11(2): 458-479.

- Cragg, G. M., & Newman, D. J. (2013). Natural products: a continuing source of novel drug leads. *Biochimica et Biophysica Acta (BBA)-General Subjects*. 1830(6): 3670-3695.
- Cui, J., Hu, W., Cai, Z., Liu, Y., Li, S., Tao, W., & Xiang, H. (2010). New medicinal properties of mangostins: Analgesic activity and pharmacological characterization of active ingredients from the fruit hull of *Garcinia mangostana* L. *Pharmacology Biochemistry and Behavior*. 95(2): 166-172.
- Cunha, B. L. A., França, J. P. D., Moraes, A. A. D. F. S., Chaves, A. L. F., Gaiba, S., Fontana, R., ... & França, L. P. D. (2014). Evaluation of antimicrobial and antitumoral activity of *Garcinia mangostana* L.(mangosteen) grown in Southeast Brazil. *Acta Cirurgica Brasileira*. 29: 21-28.
- Dai, J., & Mumper, R. J. (2010). Plant phenolics: extraction, analysis and their antioxidant and anticancer properties. *Molecules*. 15(10): 7313-7352.
- Das, K., Tiwari, R. K. S., & Srivastava, D. K. (2010). Techniques for evaluation of medicinal plant products as antimicrobial agent: Current methods and future trends. *Journal of medicinal plants research*. 4(2): 104-111.
- Davies, J., & Davies, D. (2010). Origins and evolution of antibiotic resistance. *Microbiology and Molecular Biology Reviews*. 74(3): 417-433.
- DeAngelis, D. (2012). *Dynamics of nutrient cycling and food webs* (Vol. 9). Springer Science & Business Media.
- Degrassi, G., Devescovi, G., Bigirimana, J., & Venturi, V. (2010). *Xanthomonas oryzae* pv. *oryzae* XKK. 12 contains an AroQy chorismate mutase that is involved in rice virulence. *Phytopathology*. 100(3): 262-270.
- Dewanjee, S., Gangopadhyay, M., Bhattacharya, N., Khanra, R., & Dua, T. K. (2015). Bioautography and its scope in the field of natural product chemistry. *Journal of Pharmaceutical Analysis*. 5(2): 75-84.
- Dias, D. A., Urban, S., & Roessner, U. (2012). A historical overview of natural products in drug discovery. *Metabolites*. 2(2): 303-336.
- Dubey, N. K. (Ed.). (2010). *Natural products in plant pest management*. CABI. 293 pp.
- Dunn, W. B., Broadhurst, D., Begley, P., Zelena, E., Francis-McIntyre, S., Anderson, N., ... & Nicholls, A. W. (2011). Procedures for large-scale metabolic profiling of serum and plasma using gas chromatography and liquid chromatography coupled to mass spectrometry. *Nature protocols*. 6(7): 1060-1083.

- Dutta, S. (2015). Biopesticides: An ecofriendly approach for pest control. *Institutions*. 124: 60.
- Eldeen, I., Van Staden, J., & Elgorashi, E. (2010). Ethnopharmacology of some African trees. *VDM Verlag Dr. Müller Aktiengesellschaft & Co. KG Dudweiler Landstr.* 99: 66123.
- Fousia, S., Paplomatas, E. J., & Tjamos, S. E. (2015). *Bacillus subtilis QST 713 Confers Protection to Tomato Plants Against Pseudomonas syringae pv tomato and Induces Plant Defence-related Genes*. *Journal of Phytopathology*.
- Fowler, S., Wise, J., & Moran, V. (2013). *Concepts of Biology*. OpenStax College. 663 pp.
- Fry, W. E. (2012). Management with chemicals. *Plant Disease: An Advanced Treatise: How Disease Is Managed*. 213.
- Gaillot, C., Sirdaarta, J., & Cock, I. E. (2014). An examination of the antimicrobial and anticancer properties of mangosteen pericarp extracts. In *XXIX International Horticultural Congress on Horticulture: Sustaining Lives, Livelihoods and Landscapes (IHC2014)*: 1106 (231-238).
- Garrison, A. R., Morton, G. A., & Morton, J. C. (2013). *U.S. Patent No. 8,524,285*. Washington, DC: U.S. Patent and Trademark Office.
- Gaysinski, M., Ortalo-Magné, A., Thomas, O. P., & Culoli, G. (2015). Extraction, Purification, and NMR Analysis of Terpenes from Brown Algae. *Natural Products From Marine Algae: Methods and Protocols*. 207-223.
- Ghelardini, C., Galeotti, N., Mannelli, L. D. C., Mazzanti, G., & Bartolini, A. (2001). Local anaesthetic activity of β-caryophyllene. *II Farmaco*. 56(5): 387-389.
- Gibbons, S. (2012). An Introduction to Planar Chromatography and Its Application to Natural Products Isolation. *Natural Products Isolation*. 117-153.
- Gokaraju, G. R., Gokaraju, R. R., Golakoti, T., Chirravuri, V. R., & Bhupathiraju, K. (2014). *U.S. Patent No. 8,853,261*. Washington, DC: U.S. Patent and Trademark Office.
- Gopalakrishnan, G., Banumathi, B., Suresh, G., (1997). Evaluation of the antifungal activity of natural xanthones from the fruits of *Garcinia mangostana* and their synthetic derivatives. *J. Nat. Prod.* 60: 519–524.
- Goyal, S., Lambert, C., Cluzet, S., Mérillon, J. M., & Ramawat, K. G. (2012). Secondary metabolites and plant defence. In *Plant Defence: Biological Control*. 109-138.

- Grigoris, D., Venskutonis, P. R., Sivik, B., Sandahl, M., & Eskilsson, C. S. (2005). Comparison of different extraction techniques for isolation of antioxidants from sweet grass (*Hierochloe odorata*). *The Journal of supercritical fluids*. 33(3): 223-233.
- Gunatilleke, S., De Zoysa, N., Dassanayake, M. D., Gunatilleke, N., & Wijesundera, S. (1997). *A field guide to the common trees and shrubs of Sri Lanka* (p. 430). WHT Publication (Pvt.) Limited.
- Gutierrez-Orozco, F., & Failla, M. L. (2013). Biological activities and bioavailability of mangosteen xanthones: a critical review of the current evidence. *Nutrients*. 5(8): 3163-3183.
- Gutierrez-Orozco, F., Chitchumroonchokchai, C., Lesinski, G. B., Suksamarn, S., & Failla, M. L. (2013). α-Mangostin: Anti-inflammatory activity and metabolism by human cells. *Journal of agricultural and food chemistry*. 61(16): 3891-3900.
- Hasmida, M. N., Nur Syukriah, A. R., Liza, M. S., & Mohd Azizi, C. Y. (2014). Effect of different extraction techniques on total phenolic content and antioxidant activity of *Quercus infectoria* galls. *International Food Research Journal*. 21(3).
- Hata, E. M., Sijam, K., Ahmad, Z. A. M., Yusof, M. T., & Azman, N. A. (2015). In vitro Antimicrobial Assay of Actinomycetes in Rice Against *Xanthomonas oryzae* pv. *oryzicola* and as Potential Plant Growth Promoter. *Brazilian Archives of Biology and Technology*. 58(6): 821-832.
- Havsteen, B. (1983). Flavonoids, a class of natural products of high pharmacological potency. *Biochemical pharmacology*, 32(7), 1141-1148.
- Hemingway, R. W., & Laks, P. E. (Eds.). (2012). *Plant polyphenols: synthesis, properties, significance* (Vol. 59). Springer Science & Business Media.
- Hong, C. E., Kwon, S. Y., & Park, J. M. (2016). Biocontrol activity of *Paenibacillus polymyxa* AC-1 against *Pseudomonas syringae* and its interaction with *Arabidopsis thaliana*. *Microbiological Research*. 185:13–21.
- Horst, R. K. (2013). Bacterial diseases. In *Westcott's Plant Disease Handbook* (pp. 69-90). Springer Netherlands.
- Huang, Y. L., Chen, C. C., Chen, Y. J., Huang, R. L., & Shieh, B. J. (2001). Three xanthones and a benzophenone from *Garcinia mangostana*. *Journal of natural products*. 64(7): 903-906.
- Hübschmann, H. J. (2015). *Handbook of GC-MS: Fundamentals and Applications*. John Wiley & Sons.

- Hussain, M. S., Fareed, S., Ansari, S., Rahman, M. A., Ahmad, I. Z., & Saeed, M. (2012). Current approaches toward production of secondary plant metabolites. *Journal of Pharmacy and Bioallied Sciences*. 4(1): 10.
- Ibrahim, M. Y., Hashim, N. M., Mariod, A. A., Mohan, S., Abdulla, M. A., Abdelwahab, S. I., & Arbab, I. A. (2014). An updated review of its pharmacological properties. *Arabian Journal of Chemistry*.
- Ignat, I., Volf, I., & Popa, V. I. (2011). A critical review of methods for characterisation of polyphenolic compounds in fruits and vegetables. *Food Chemistry*. 126(4): 1821-1835.
- Janisiewicz, W.J., Tworkoski T.J., Kurtzman C.P. (2001). Biocontrol potential of *Metchnikowia pulcherrima* strains against blue mold of apple. *Phytopathology*. 91(11): 1098–1108.
- Jimenez-Garcia, S. N., Vazquez-Cruz, M. A., Guevara-Gonzalez, R. G., Torres-Pacheco, I., Cruz-Hernandez, A., & Feregrino-Perez, A. A. (2013). Current Approaches for Enhanced Expression of Secondary Metabolites as Bioactive Compounds in Plants for Agronomic and Human Health Purposes—a Review. *Polish Journal of Food and Nutrition Sciences*. 63(2): 67-78.
- Jones, J. B., Jackson, L. E., Balogh, B., Obradovic, A., Iriarte, F. B., & Momol, M. T. (2007). Bacteriophages for Plant Disease Control*. *Annu. Rev. Phytopathol.* 45: 245-262.
- Junqueira-Gonçalves, M. P., Yáñez, L., Morales, C., Navarro, M., A Contreras, R., & Zúñiga, G. E. (2015). Isolation and Characterization of Phenolic Compounds and Anthocyanins from Murta (*Ugni molinae* Turcz.) Fruits. Assessment of Antioxidant and Antibacterial Activity. *Molecules*. 20(4): 5698-5713.
- Kang, Y. B., Mallikarjuna, P. R., Fabian, D. A., Gorajana, A., Lim, C. L., & Tan, E. L. (2013). Bioactive molecules: Current trends in discovery, synthesis, delivery and testing. *International e-Journal of Science and Medical Education*. 7: S32-S46.
- Karthikeyan, R., Devadasu, C., & Srinivasa Babu, P. (2015). Isolation, Characterization, and RP-HPLC Estimation of P-Coumaric Acid from Methanolic Extract of Durva Grass (*Cynodon dactylon* Linn.) (Pers.). *International journal of analytical chemistry*.
- Kataria, S. (2011). Gas chromatography-mass spectrometry: applications. *International Journal of Pharmaceutical & Biological Archive*. 2(6).
- Kennedy, D. O., & Wightman, E. L. (2011). Herbal extracts and phytochemicals: plant secondary metabolites and the enhancement of human brain function. *Advances in Nutrition: An International Review Journal*. 2(1): 32-50.

- Khoddami, A., Wilkes, M. A., & Roberts, T. H. (2013). Techniques for analysis of plant phenolic compounds. *Molecules*. 18(2): 2328-2375.
- Kim, Y. S., Park, S. J., Lee, E. J., Cerbo, R. M., Lee, S. M., Ryu, C. H., ... & Ha, Y. L. (2008). Antibacterial Compounds from Rose Bengal-Sensitized Photooxidation of β -Caryophyllene. *Journal of food science*. 73(7): C540-C545.
- Kohanski, M. A., Dwyer, D. J., & Collins, J. J. (2010). How antibiotics kill bacteria: from targets to networks. *Nature Reviews Microbiology*. 8(6): 423-435.
- Kowalska, T., Sajewicz, M., & Sherma, J. (Eds.). (2015). Planar Chromatography-Mass Spectrometry. Chemical Rubber Company (CRC) press. 380pp.
- Kumar, S., & Pandey, A. K. (2013). Chemistry and biological activities of flavonoids: an overview. *The Scientific World Journal*. 1-16.
- Lahlou, M. (2013). The success of natural products in drug discovery. *Pharmacology & Pharmacy*. 4(3A): 17.
- Lim, T. K. (2012). Garcinia mangostana. In *Edible Medicinal And Non-Medicinal Plants* (pp. 83-108). Springer Netherlands.
- Louisa, M. (2013). Medicinal plants: source of new lead compounds in therapeutics. *Medical Journal of Indonesia*. 22(3): 127-8.
- Mahabusarakam, W., Proudfoot, J., Taylor, W., Croft, K., (2000). Inhibition of lipoprotein oxidation by prenylated xanthones derived from mangostin. *Free Radic. Res.* 33: 643–659.
- Manimekalai, I., Sivakumari, K., Ashok, K., & Rajesh, S. (2015). Phytochemical Profiling Of Mangosteen Fruit, Garcinia Mangostana. *World Journal of Pharmacy and Pharmaceutical Sciences*. 5(2): 221-252.
- Mansfield, J., Genin, S., Magori, S., Citovsky, V., Sriariyanum, M., Ronald, P., ... & Toth, I. A. N. (2012). Top 10 plant pathogenic bacteria in molecular plant pathology. *Molecular plant pathology*. 13(6): 614-629.
- Mariita, R. M., Orodho, J. A., Okemo, P. O., Kirimuhuza, C., Otieno, J. N., & Magadula, J. J. (2011). Methanolic extracts of *Aloe secundiflora* Engl. inhibits in vitro growth of tuberculosis and diarrhea-causing bacteria. *Pharmacognosy research*, 3(2), 95.
- Martínez, J. A. (2012). *Natural Fungicides obtained from plants*. INTECH Open Access Publisher.
- Mazid, M., Khan, T. A., & Mohammad, F. (2011). Role of secondary metabolites in defense mechanisms of plants. *Biology and medicine*. 3(2): 232-249.

- Mazimbaa, O., Nanab, F., Kuetec, V., & Singha, G. S. (2013). Xanthones and Anthranoids from the Medicinal Plants of Africa. *Medicinal Plant Research in Africa: Pharmacology and Chemistry*. 393.
- Mew, T., Alvarez, A., Leach, J. and Swings, J. (1993) Focus on bacterial blight of rice. *Plant Dis.* 77: 5–12.
- Meyer, J. J. M., & Dilika, F. (1996). Antibacterial activity of *Helichrysum pedunculatum* used in circumcision rites. *Journal of ethnopharmacology*. 53(1): 51-54.
- Mierziak, J., Kostyn, K., & Kulma, A. (2014). Flavonoids as important molecules of plant interactions with the environment. *Molecules*. 19(10): 16240-16265.
- Milijašević, S. (2005). Bacteriological Characteristics and Physiological Specialization of *P. syringae* pv. *tomato* (Okabe) Yang, Dye & Wilkie, a Pathogen of Tomato in Serbia. Master's Thesis, Faculty of Agriculture, University of Belgrade.
- Moazami, N. (2008). Biopesticide production. *Industrial biotechnology VI, Encyclopedia of life support systems (EOLSS)*, Eolss, Oxford, UK, 1-52.
- Mommaerts, V., & Smaghe, G. (2011). Pesticides in the Modern World: Pests Control and Pesticides Exposure and Toxicity Assessment.
- Momo, I. J., Kuete, V. I. C. T. O. R., Dufat, H. A. N. H., Michel, S. Y. L. V. I. E., & Wandji, J. E. A. N. (2011). Antimicrobial activity of the methanolic extract and compounds from the stem bark of *Garcinia lucida* Vesque (Clusiaceae). *Int J Pharm Pharm Sci.* 3: 215-7.
- Montanari, R. M., Barbosa, L. C., Demuner, A. J., Silva, C. J., Carvalho, L. S., & Andrade, N. J. (2011). Chemical composition and antibacterial activity of essential oils from verbenaceae species: alternative sources of (E)-caryophyllene and germacrene-D. *Química Nova*. 34(9): 1550-1555.
- Morebise, O. (2015). Medicinal plants of Dominica—Uses, chemical constituents, bioactivities and prospects. *Journal of Medicinal Plants*. 3(5): 144-154.
- Móricz, Á., Horváth, G., & Ott, P. (2015). Direct bioautographic detection of antibacterial components of clary sage and spearmint essential oils. *JPC-Journal of Planar Chromatography-Modern TLC*. 28(2): 173-177.
- Moses, T., Papadopoulou, K. K., & Osbourn, A. (2014). Metabolic and functional diversity of saponins, biosynthetic intermediates and semi-synthetic derivatives. *Critical reviews in biochemistry and molecular biology*. 49(6): 439-462.

- Mushore, J., & Matuvhunye, M. (2013). Antibacterial properties of mangifera indica on *Staphylococcus Aureus*. *African Journal of Clinical and Experimental Microbiology*. 14(2): 62-74.
- NIÑO-LIU, D. O., Ronald, P. C., & Bogdanove, A. J. (2006). *Xanthomonas oryzae* pathovars: model pathogens of a model crop. *Molecular Plant Pathology*. 7(5): 303-324.
- Obolskiy, D., Pischel, I., Siriwananametanon, N., & Heinrich, M. (2009). *Garcinia mangostana* L.: a phytochemical and pharmacological review. *Phytotherapy research*. 23(8): 1047-1065.
- Omojate Godstime, C., Enwa Felix, O., Jewo Augustina, O., & Eze Christopher, O. (2014). Mechanisms of antimicrobial actions of phytochemicals against enteric pathogens—a review. *J Pharm Chem Biol Sci*. 2(2): 77-85.
- Palakawong, C., Sophanodora, P., Pisuchpen, S., & Phongpaichit, S. (2010). Antioxidant and antimicrobial activities of crude extracts from mangosteen (*Garcinia mangostana* L.) parts and some essential oils. *Int Food Res J*. 17: 583-589.
- Pedraza-Chaverri, J., Cárdenas-Rodríguez, N., Orozco-Ibarra, M., & Pérez-Rojas, J. M. (2008). Medicinal properties of mangosteen (*Garcinia mangostana*). *Food and chemical toxicology*. 46(10): 3227-3239.
- Perez-Garcia, A., Romero, D. and Vicente, A. (2011). Plant Protection and Growth Stimulation by Microorganisms: Biotechnological Applications of Bacilli in Agriculture. *Current Opinion in Biotechnology*. 22(2): 187-193.
- Petrovska, B. B. (2012). Historical review of medicinal plants' usage. *Pharmacognosy reviews*. 6(11): 1.
- Phillipson, J. D. (2007). Phytochemistry and pharmacognosy. *Phytochemistry*. 68(22): 2960-2972.
- Postma, J., Schepers, R.W.A. and Schilder, M.T. (2010). Effect of successive cauliflower plantings and *Rhizoctonia solani* AG 2-1 inoculations on disease suppressiveness of a suppressive and a conducive soil. *Soil Biology and Biochemistry*. 42(5): 804-812.
- Prakash, D., & Sharma, G. (Eds.). (2014). *Phytochemicals of nutraceutical importance*. CABI. 376 pp.
- Priya, V., Jainu, M., Mohan, S. K., Saraswathi, P., & Gopan, S. C. (2010). Antimicrobial activity of pericarp extract of *Garcinia mangostana* Linn. *Int. J. Pharm. Sci. Res.* 1: 278-281.
- Rademaker, J.L.W., Louws, F.J., and Schultz, M.H. (2005). "A comprehensive species to strain taxonomic framework for *Xanthomonas*". *Phytopathology*

- Radulovic, N., Dekic, M., STOJANOVIC-RADIC, Z. O. R. I. C. A., & Palic, R. (2011). Chemical composition and antimicrobial activity of the essential oils of Geranium columbinum L. and G. lucidum L.(Geraniaceae). *Turkish Journal of Chemistry*. 35(3): 499-512.
- Ragasa, C. Y., Crisostomo, C. J. J., Garcia, K. D. C., & Shen, C. C. (2010). Antimicrobial xanthones from Garcinia mangostana L. *Philippine Scientist*. 47: 63-75.
- Rajakannu, S., Shankar, S., Perumal, S., Subramanian, S., & Dhakshinamoorthy, G. P. (2015). Biosynthesis of Silver Nanoparticles using Garcinia mangostana Fruit Extract and their Antibacterial, Antioxidant Activity. *Int. J. Curr. Microbiol. App. Sci.* 4(1): 944-952.
- Rajauria, G., & Abu-Ghannam, N. (2013). Isolation and partial characterization of bioactive fucoxanthin from Himanthalia elongata brown seaweed: a TLC-based approach. *International journal of analytical chemistry*.
- Razali, N, Mat-Junit S, Abdul-Muthalib AF, Subramaniam S, Abdul-Aziz A. (2012). Effects of various solvents on the extraction of antioxidant phenolics from the leaves, seeds, veins and skins of Tamarindus indica L. *Food Chemistry*.131(2): 441–448
- Reddy, P. P. (2016). Biorational Pest Management. In *Sustainable Crop Protection under Protected Cultivation* (pp. 99-108). Springer Singapore.
- Rigane, G., Ben Younes, S., Ghazghazi, H., & Ben Salem, R. (2013). Investigation into the biological activities and chemical composition of Calendula officinalis L. growing in Tunisia. *International Food Research Journal*. 20(6): 3001-3007.
- Rodriguez-Amaya, D. B. (2001). *A guide to carotenoid analysis in foods* (p. 65). Washington, DC: ILSI press.
- Sajewicz, M., Staszek, D., Waksmundzka-Hajnos, M., & Kowalska, T. (2012). Comparison of TLC and HPLC Fingerprints of Phenolic Acids and Flavonoids Fractions Derived from Selected Sage (*Salvia*) Species. *Journal of Liquid Chromatography & Related Technologies*. 35(10): 1388-1403.
- Sampah, M. E. S., Shen, L., Jilek, B. L., & Siliciano, R. F. (2011). Dose-response curve slope is a missing dimension in the analysis of HIV-1 drug resistance. *Proceedings of the National Academy of Sciences*, 108(18), 7613-7618.
- Sarwar, M. (2015). Information on Activities Regarding Biochemical Pesticides: An Ecological Friendly Plant Protection against Insects. *International Journal of Engineering and Advanced Research Technology*. 1(2): 27-31.
- Sasidharan, S., Chen, Y., Saravanan, D., Sundram, K. M., & Latha, L. Y. (2011). Extraction, isolation and characterization of bioactive compounds

- from plants' extracts. *African Journal of Traditional, Complementary and Alternative Medicines*. 8(1).
- Schmourlo, G., Mendonça-Filho, R. R., Alviano, C. S., & Costa, S. S. (2005). Screening of antifungal agents using ethanol precipitation and bioautography of medicinal and food plants. *Journal of Ethnopharmacology*. 96(3): 563-568.
- Seigler, D. S. (2012). *Plant secondary metabolism*. Springer Science & Business Media. 759 pp.
- Shan, T., Ma, Q., Guo, K., Liu, J., Li, W., Wang, F., & Wu, E. (2011). Xanthones from mangosteen extracts as natural chemopreventive agents: potential anticancer drugs. *Current molecular medicine*. 11(8): 666.
- Sherma, J. (2005). Thin-Layer Chromatography of pesticides-a review of applications for 2002-2004. *Acta Chromatographica*, 15: 5.
- Sikin, A. M., Zoellner, C., & Rizvi, S. S. (2013). Current intervention strategies for the microbial safety of sprouts. *Journal of Food Protection®*. 76(12): 2099-2123.
- Silby, M. W., Winstanley, C., Godfrey, S. A., Levy, S. B., & Jackson, R. W. (2011). Pseudomonas genomes: diverse and adaptable. *FEMS microbiology reviews*. 35(4): 652-680.
- Silva, M. T., Simas, S. M., Batista, T. G., Cardarelli, P., & Tomassini, T. C. (2005). Studies on antimicrobial activity, in vitro, of *Physalis angulata* L.(Solanaceae) fraction and physalin B bringing out the importance of assay determination. *Memorias do Instituto Oswaldo Cruz*. 100(7): 779-782.
- Singh, S., Srivastava, R., & Choudhary, S. (2010). Antifungal and HPLC analysis of the crude extracts of *Acorus calamus*, *Tinospora cordifolia* and *Celestrus paniculatus*. *Journal of Agricultural Technology*. 6(1): 149-158.
- Stanier, R.Y., Palleroni, N.J., Doudoroff, M. (1966). The aerobic pseudomonads: a taxonomic study. *J Gen Microbiol*. 43:159-271.
- Suksamrarn, S., Suwannapoch, N., Phakhodee, W., Thanuhiranlert, J., Ratananukul, P., Chimnoi, N., & Suksamrarn, A. (2003). Antimycobacterial activity of prenylated xanthones from the fruits of *Garcinia mangostana*. *Chemical and pharmaceutical bulletin*. 51(7): 857-859.
- Suleiman, M. M., McGaw, L. I., Naidoo, V., & Eloff, J. (2010). Detection of antimicrobial compounds by bioautography of different extracts of leaves of selected South African tree species. *African Journal of Traditional, Complementary and Alternative Medicines*. 7(1).

- Sultana, B., Anwar, F., & Ashraf, M. (2009). Effect of extraction solvent/technique on the antioxidant activity of selected medicinal plant extracts. *Molecules*. 14(6): 2167-2180.
- Sundaram, B.M., Gopalakrishnan, C., Subramanian, S., Shankaranarayanan, D., Kameswaran, L., (1983). Antimicrobial activities of *Garcinia mangostana*. *Planta Med*. 48: 59–60.
- Tadtong, S., Viriyaroj, A., Vorarat, S., Nimkulrat, S., & Suksamrarn, S. (2009). Antityrosinase and antibacterial activities of mangosteen pericarp extract. *J Health Res*. 23(2): 99-102.
- Taokaew, S., Nunkaew, N., Siripong, P., & Phisalaphong, M. (2014). Characteristics and anticancer properties of bacterial cellulose films containing ethanolic extract of mangosteen peel. *Journal of Biomaterials Science, Polymer Edition*. 25(9): 907-922.
- Taweechaisupapong, S., Singhara, S., Lertsatitthanakorn, P., & Khunkitti, W. (2010). Antimicrobial effects of *Boesenbergia pandurata* and *Piper sarmentosum* leaf extracts on planktonic cells and biofilm of oral pathogens. *Pak J Pharm Science*. 23(2): 224-231.
- Tegegne, G., Pretorius, J. C., & Swart, W. J. (2008). Antifungal properties of *Agapanthus africanus* L. extracts against plant pathogens. *Crop Protection*. 27(7): 1052-1060.
- Tiwari, V., Roy, R., & Tiwari, M. (2015). Antimicrobial active herbal compounds against *Acinetobacter baumannii* and other pathogens. *Frontiers in microbiology*. 6: 618.
- Tomlinson, T. R., & Akerele, O. (Eds.). (2015). *Medicinal plants: their role in health and biodiversity*. University of Pennsylvania press.
- Torrungruang, K., Vichienroj, P., & Chutimaworapan, S. (2013). Antibacterial activity of mangosteen pericarp extract against cariogenic *Streptococcus mutans*. *Chulalongkorn University Dental Journal-วารสาร ทันตแพทยศาสตร์ จุฬาลงกรณ์มหาวิทยาลัย*. 30(1): 1-10.
- Trease, G.E., Evans, W.C. (2002). *Pharmacognosy*, 15th ed. Springer, Berlin.
- Upadhyaya, S., Behera, J., & Tewari, S. N. (2012). Integrated management of foliar blast through ecofriendly formulated product, Oscext-e developed from *Ocimum sanctum* ethanolic extract. *Archives Of Phytopathology And Plant Protection*. 45(19): 2290-2300.
- Upaganlawar, A. B., & Badole, S. L. (2013). Mangosteen (*Garcinia mangostana* Linn.): Role in Prevention of Skin Disorders. In *Bioactive Dietary Factors and Plant Extracts in Dermatology* (pp. 451-457). Humana Press.

- Vidaver, A.K. and P.A. Lambrecht (2004). Bacteria as plant pathogens. The Plant Health Instructor. DOI: 10.1094/PHI-I-2004-0809-01.
- Villas-Boas S. G., Mas S., Akesson M., Smedsgaard J., Nielsen J. (2005). Mass spectrometry in metabolome analysis. *Mass Spectrom. Rev.* 24: 613–646.
- War, A. R., Paulraj, M. G., Ahmad, T., Buhroo, A. A., Hussain, B., Ignacimuthu, S., & Sharma, H. C. (2012). Mechanisms of plant defense against insect herbivores. *Plant signaling & behavior.* 7(10): 1306-1320.
- Weecharangsan, W., Opanasopit, P., Sukma, M., Ngawhirunpat, T., Sotanaphun, U., Siripong, P., (2006). Antioxidative and neuroprotective activities of extracts from the fruit hull of mangosteen (*Garcinia mangostana* Linn.). *Med. Princ. Pract.* 15: 281–287.
- Wilson, M., Campbell, H. L., Ji, P., Jones, J. B., & Cuppels, D. A. (2002). Biological control of bacterial speck of tomato under field conditions at several locations in North America. *Phytopathology.* 92(12): 1284-1292.
- Wu, L., Wu, H., Chen, L., Yu, X., Borri, R., & Gao, X. (2015). Difficidin and bacilysin from *Bacillus amyloliquefaciens* FZB42 have antibacterial activity against *Xanthomonas oryzae* rice pathogens. *Scientific reports.* 5.
- Xie, Z., Sintara, M., Chang, T., & Ou, B. (2015). Daily consumption of a mangosteen-based drink improves in vivo antioxidant and anti-inflammatory biomarkers in healthy adults: a randomized, double-blind, placebo-controlled clinical trial. *Food science & nutrition.* 3(4): 342-348.
- Yadeta, K. A., & Thomma, B. P. (2014). The xylem as battleground for plant hosts and vascular wilt pathogens. *Induced plant responses to microbes and insects.* 110.
- Yaduraju, N. T., & Mishra, J. S. (2004). Soil Solarization. In *Weed biology and management* (pp. 345-362). Springer Netherlands.
- Young, J.M., Park, D.C., Shearman, H.M. and Fargier, E. (2008) A multilocus sequence analysis of the genus *Xanthomonas*. *Syst. Appl. Microbiol.* 5: 366–377.
- Zorofchian, M. S., Abdul Kadir, H., Hassandarvish, P., Tajik, H., Abubakar, S., & Zandi, K. (2014). A review on antibacterial, antiviral, and antifungal activity of curcumin. *BioMed research international.* 2014.
- Zweig, G. (Ed.). (2013). *Principles, Methods, and General Applications: Analytical Methods for Pesticides, Plant Growth Regulators, and Food Additives* (Vol. 1). Elsevier.