



UNIVERSITI PUTRA MALAYSIA

***IN- VITRO ANTIBACTERIAL ACTIVITY AN D PHYTOCHEMICAL
SCREENING OF BIOACTIVE COMPOUNDS FROM POMEGRANATE
(*Punica granatum L.*) CRUDE PEEL EXTRACTS***

AYAD ISMAEL KHALEEL

FP 2016 40



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By

AYAD ISMAEL KHALEEL

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfillment of the Requirements for the Degree of Master of Science**

May 2016

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DEDICATION

This Thesis is dedicated to

The most precious people in my life;

My Father and mother

My brother and sisters

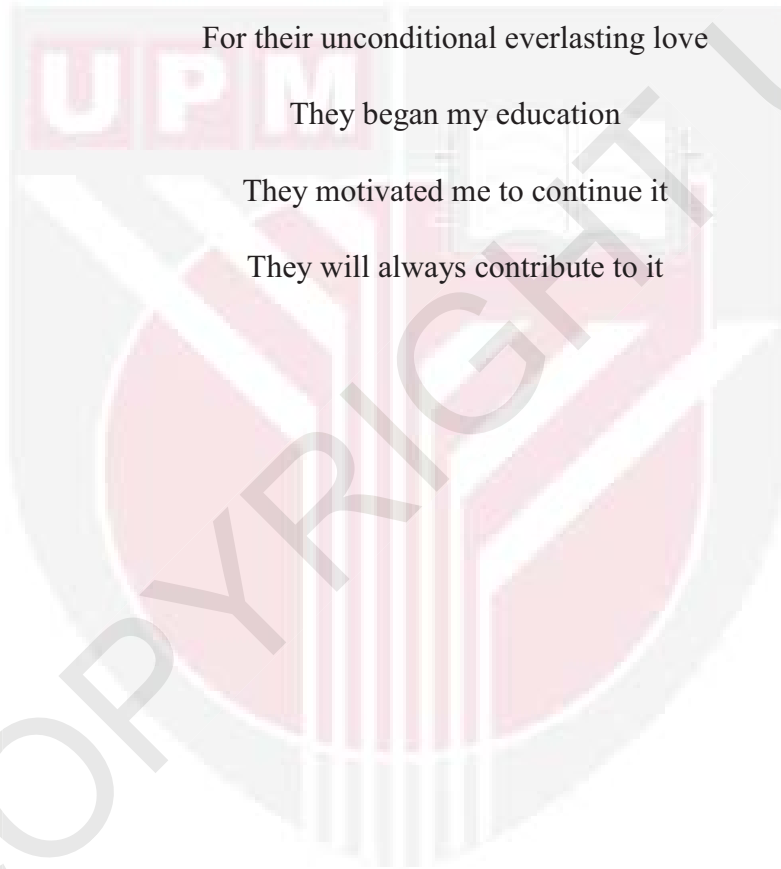
My wife and kids

For their unconditional everlasting love

They began my education

They motivated me to continue it

They will always contribute to it



Abstract of thesis presented to the senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

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By

AYAD ISMAEL KHALEEL

May 2016

Chairman : Associate Professor Kamaruzaman Bin Sijam, PhD
Faculty : Agriculture

Plant pathogenic bacteria are recognized to be harmful microbes able to decrease the quantity and quality of crop production in agriculture. *Punica granatum* L. peel was screened for its potential use as a biological control agent for plant pathogenic bacteria. *P. granatum* peel was successfully extracted using n-hexane, methanol and ethyl acetate. The highest percentage of crude extracts was obtained from ethyl acetate 1.37% followed by methanol crude extracts 1.17% and n-Hexane 0.89%. The highest yield obtained by ethyl acetate showed that ethyl acetate extracted more compounds that are readily soluble to methanol and n-hexane. For *in-vitro* antibacterial activity, three different species of plant pathogenic bacteria were used namely *Erwinia carotovorum* subsp. *carotovorum*, *Ralstonia solanacearum*, and *Xanthomonas gardneri*. For all crude extracts, four different concentrations of 25, 50, 100 and 200 mg/mL were used in cup-plate agar diffusion method. Streptomycin sulfate at concentration 30 µg/mL was used as positive control while each respective solvent used for peel extraction was used as a negative control. The results obtained from *in vitro* studies showed only ethyl acetate extract possessed antibacterial activity tested on the plant pathogenic bacteria. Methanol and n-hexane did not show any antibacterial activity against plant pathogenic bacteria selected where no inhibition zones were recorded. *R. solanacearum* recorded the highest diameter of inhibition zone for all ranges of concentrations introduced followed by *P. carotovorum* subsp. *carotovorum* and *X. gardneri*. For the minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC), only the ethyl acetate extract was subjected to the assay as only ethyl acetate extract exhibited antibacterial activity. The minimum concentration of ethyl acetate extract that was able to inhibit plant pathogenic bacteria was recorded at a concentration of 3.12 mg/mL, which inhibited *R. solanacearum* and *P. carotovorum* subsp. *carotovorum*, followed by *X. gardneri* at concentration of 6.25 mg/mL. For the minimum bactericidal concentration (MBC), the results showed that at the concentration of 12.5 mg/mL, the extract was still capable of killing the pathogenic bacteria, *R. solanacearum*, and *P. caratovora* sub.sp. *caratovora* while for the bacteria *X. gardneri*, the concentration that was able to kill the bacteria was 25 mg/mL. The

qualitative estimation of phytochemical constituents within *P. granatum* ethyl acetate peel extracts revealed the presence of tannins, flavonoids, phenols alkaloid, Saponins, and terpenoids. The thin layer chromatography (TLC) profiling of ethyl acetate extract using hexane, ethyl acetate, and chloroform with ratio 5:3:2 (v/v) gave eleven maximum colorful bands when visualized under short UV wavelength (254nm), six bands under long UV wavelength (365nm) and eight bands in daylight (normal light) with different retention factors, and R_f values that provined the presence of various secondary metabolites within ethyl acetate extract. The antibacterial activity of ethyl acetate peel extract was also screened through direct bioautography technique in order to detect the location of the progressive band on chromatograms developed in the same substance for TLC profiling. The recorded active R_f values that inhibited all selected plant pathogenic bacteria at the same R_f values location were 0.45, 0.83 and 0.92. The GC-MS spectrum range affirmed the vicinity of 292 different components with diverse retention times and chemical structure eleven elements in the high peak chosen were (DMSO; n-Hexadecanoic acid; 9,12-Octadecadienoic acid (Z,Z); cis-Vaccenic acid; Octadecanoic acid; Pentanoic acid; cis-9-Hexadecenoic acid; Tetradecanoic acid; 2-Heptenoic acid; Octanal diethyl acetal; Glycerin). The results obtained from this study suggest that *P. granatum* L ethyl acetate peel extracts have the potential to be industrialized as a novel bactericide.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

AKTIVITI ANTIBAKTERIA *IN-VITRO* DAN PENYARINGAN KOMPOUN BIOAKTIF FITOKIMIA DARI EKSTRAK KULIT MENTAH DELIMA (*Punica granatum* L.)

Oleh,

AYAD ISMAEL KHALEEL

Mei 2016

Pengerusi : Profesor Madya Kamaruzaman Bin Sijam, PhD
Fakulti : Pertanian

Bakteria patogenik tumbuhan diketahui umum sebagai mikrob berbahaya yang dapat mengurangkan kuantiti dan kualiti hasil pertanian. Kulit *Punica granatum* L. telah disaring potensinya sebagai agen kawalan biologi bakteria patogenik tumbuhan. Kulit *P. granatum* L. ini telah berjaya diekstrak menggunakan n-heksana, metanol dan ethyl acetat. Hasil peratusan tertinggi adalah 1.37% melalui ekstrak mentah ethyl acetat, diikuti oleh ekstrak mentah methanol, 1.17% dan n-heksana, 0.89%. Hasil tertinggi diperolehi daripada ethyl acetate menunjukkan ethyl acetat mengekstrak lebih sebatian bahan mudah larut berbanding metanol dan n-heksana. Bagi aktiviti antibakteria *in-vitro*, tiga jenis spesies bakteria patogenik tumbuhan telah digunakan iaitu *Pectobacterium carotovorum* subsp. *carotovorum*, *Ralstonia solanacearum*, dan *Xanthomonas gardneri*. Bagi kesemua ekstrak mentah, empat kepekatan berbeza iaitu 25, 50, 100 dan 200 mg/ml telah diuji menggunakan kaedah resapan plat-cawan. Streptomisin sulfat pada kepekatan 30 µg/ml telah digunakan sebagai kawalan positif, manakala setiap bahan pelarut pengekstrakan kulit digunakan sebagai kawalan negatif. Keputusan yang diperolehi daripada kajian *in-vitro* ini menunjukkan hanya ekstrak ethyl acetat mempunyai aktiviti anti-bakteria terhadap bakteria patogenik tumbuhan yang diuji. Metanol dan n-heksana tidak menunjukkan sebarang aktiviti antibakteria terhadap bakteria patogenik tumbuhan terpilih di mana tiada zon perencatan telah direkodkan. *Ralstonia solanacearum* merekodkan diameter perencatan tertinggi untuk semua julat kepekatan yang diuji dan diikuti oleh *P. carotovorum* subsp. *carotovorum* dan *X. gardneri*. Untuk kepekatan perencatan minima (MIC) dan kepekatan minima bakterisidal (MBC), hanya ekstrak ethyl acetat yang tertakluk kepada ujian kerana hanya ekstrak etil asetat menunjukkan aktiviti anti-bakteria. Kepekatan minima ekstrak ethyl acetat yang mampu untuk merencat bakteria patogenik tumbuhan dicatatkan pada kepekatan 3.12 mg/ml, untuk merencat *R. solanacearum* dan *P. carotovorum* subsp. *carotovorum* dan *X. gardneri* pula pada kepekatan 6.25 mg/ml. Untuk kepekatan minima bakterisidal (MBC), keputusan menunjukkan bahawa pada kepekatan 12.5 mg/ml, ekstrak itu masih mampu membunuh bakteria patogenik, *R. solanacearum* dan *P. carotovorum* subsp. *carotovorum* manakala bagi bakteria *X. gardneri*,

kepekatan yang mampu membunuh bakteria adalah 25 mg/ml. Anggaran kualitatif juzuk fitokimia dalam *P. granatum* L. ekstrak kulit ethyl acetat mendedahkan kehadiran tanin, flavonoid, fenol alkaloid, saponin dan terpenoid. Profil kromatografi lapisan nipis (TLC) bagi ekstrak ethyl acetat menggunakan heksana, ethil acetat, dan kloroform dengan nisbah 5:3:2 (v/v) memberi 11 jalur yang berwarna-warni maksim apabila digambarkan di bawah gelombang UV ringkas (254 nm), enam jalur di bawah gelombang UV panjang (365 nm) dan lapan jalur di bawah cahaya putih (cahaya biasa) dengan faktor pengekalakan berbeza dan nilai Rf yang menunjukkan kehadiran pelbagai metabolit sekunder di dalam ekstrak ethyl acetat. Aktiviti antibakteria ekstrak kulit ethyl acetat juga telah disaring melalui teknik bioautography terus untuk mengesan lokasi jalur progresif pada kromatogram terbentuk dengan menggunakan bahan yang sama melalui pemprofil TLC. Direkodkan nilai Rf aktif yang merencatkan semua bakteria patogenik tumbuhan berada di nilai Rf yang sama iaitu 0.45, 0.83 dan 0.92. Julat spektrum GC-MS mengesahkan sebanyak 292 komponen berbeza dalam masa tahanan yang pelbagai dan 11 struktur kimia mempunyai puncak tinggi telah dipilih adalah (DMSO; n-Hexadecanoic asid; asid 9,12-Octadecadienoic (Z, Z); cis- asid Vaccenic; asid Octadecanoic; asid Pentanoic; cis-9-Hexadecenoic asid; asid Tetradecanoic; asid 2-Heptenoic; Octanal diethyl asetal; Glycerin). Keputusan yang diperolehi daripada kajian ini mencadangkan bahawa ekstrak kulit *P. granatum* L ethyl acetat mempunyai potensi untuk dikilangkan sebagai baktetsidal yang baharu.

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I certify that a Thesis Examination Committee has met on 10 Mei 2016 to conduct the final examination of Ayad Ismael Khaleel on his thesis entitled "*In-Vitro* Antibacterial Activity and Phytochemical Screening of Bioactive Compounds from Pomegranate (*Punica granatum* L.) Crude Peel Extracts" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

PG _E	<i>Punica granatum</i> L. ethyl acetate peel extracts
PG _N	<i>Punica granatum</i> L. n-hexane peel extracts
PG _M	<i>Punica granatum</i> L. methanol peel extracts
%	Percent
ANOVA	Analysis Of Variance
DCSO	dimethyl sulfoxide
FeCl ₃	Ferric chloride
H ₂ SO ₄	Sulfuric acid
HCl	Hydrochloric acid
cm	Centimeter
INT	2-(4-iodophenyl)-3-(4-nitrophenyl)-5-phenyl-2H-tetrazolium chloride
MIC	Minimum inhibitory concentration
MBC	Minimum bactericidal concentration
MHA	Mueller Hinton Agar
MHB	Mueller Hinton Broth
MTC	3-(4,5-dimethylthiazolyl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium
MTT	3-(4,5-dimethylthiazolyl)-2,5-diphenyl-2H-tetrazolium bromide
FAO	Food and Agriculture Organization of the United Nations
g	Gram
n	Number of replicates
NA	Nutrient Agar
kb	Kilo base
L	Liter
LSD	Least Significant Difference
M	Molarity
min	Minute
mm	millimeter
NaOH	Sodium hydroxide
°C	Degree
pH	potential Hydrogen
SAS	Statistical Analysis System
sec	Second
UPM	Universiti Putra Malaysia
UV	Ultra violet

V	Volts
NH ₃	Ammonia
O.D.	Optical density
Pv.	Pathovar
R _f	Retention factor
rpm	Revolutions per minute
S.D	Standard deviation
Subsp.	Subspecies
TLC	Thin layer chromatography
TTC	2,3,5-triphenyltetrazolium
XTT	2,3-bis-(2-methoxy-4-nitro-5-sulfoheny)-2H-tetrazolium-5-carboxanilide
GCMS	Gas chromatography–mass spectrometry
IC ₅₀	Inhibitory Concentration 50% of bacteria
IC ₉₀	Inhibitory Concentration 90% of bacteria

CHAPTER 1

INTRODUCTION

1.1 Background

The history of human civilization offers an abundance of evidence that an amazing range of plants and plant materials have been used as medicinal agents and numerous natural products acquired from medicinal plants either as a crude extract or as purified products have been applied in disease control. Owing to the presence of various medicinal properties in these medicinal plants and plant parts, they have been extensively used for the extraction of natural drugs. They constitute credible sources for a huge number of modern drugs, several of which are based on their traditional folk homoeopathic efficacy. The World Health Organization (WHO) has stated that medicinal plants are the best source for obtaining a variety of therapeutic agents, and several medicinal plants have been employed as sources of medicine in daily life for treatment of various types of ailments globally (Alo et al., 2012). Clinical microbiologists have widely used medicinal plants for the screening of new therapeutic agents (Ashokkumar & Rajkumar, 2010). A great range of biotic molecules referred to as secondary metabolites are produced by plants (Dash et al., 2011), thereby making them a rich source for diverse forms of medicine. Additionally, the primary advantages of using these naturally derived products include safety for human consumption, environmentally friendly, and economical in treating microbial infections when they are used (Al-Zubaydi & Al-Hmdany, 2009).

Recently, much attention has been paid to the use and search for natural products. Extracts of plant origin are usually rich in a broad range of active compounds (Al-Zubaydi & Al-Hmdany, 2009) with the potential to be developed as natural antibacterial agents (Bhardwaj & Laura, 2009); (Dash et al., 2011) Plant extracts have been demonstrated to have antibacterial properties in *in-vitro* studies (Ghosh et al., 2008; Rios & Recio, 2005) Plant-based products have been shown to be safer than synthetic products, and they have gained more acceptance as they are non-toxic to humans, biodegradable and non-polluting. Additionally, they are less phytotoxic and possess greater systemic effect. These properties have led to plants being further investigated for their effectiveness in combatting diverse kinds of microbes in many fields of study (Abdollahzadeh et al., 2011).

The use of hazardous chemicals to control the spread of disease is of great concern to those who are aware of the dangers of the toxic effects on food crops. The consequences of short-term or long-term toxicity caused by toxic chemicals can affect human health negatively. In addition, it also may environmental pollution, particularly of water, soil and the air. This may could lead to serious ecological damage that would impact both humans and a whole range of living things.

Several studies have been conducted to address this issue in attempts for finding solutions of the problems associated with compare to harmful chemicals in agriculture. In recent years, biological control methods have been studied as a viable and safer alternative to compare to harmful chemicals as they have been proven to be effective, inexpensive and easily available. Biological control methods are also easily manipulated from natural resources and at the same time are friendly to the environment and ensure that the agricultural product safe for human consumption. Pathogens such as fungi, viruses and bacteria have been identified being capable of causing plant diseases that may lead to deterioration of yield in terms of a decline in quality and quantity of crops. Add to all that, it causes economic losses to countries that largely depend on agricultural produce both for food production and for export.

Biological control measures were therefore introduced to the agriculture industry owing to increased demand for safer crop products as more people are becoming aware of the harmful and toxic effects of chemical pesticides on crops. Biological control of plant pests and pathogens is continually inspiring research and development in diverse fields where plant pathogens act as a class of targets of biological control that are designed to limit other pests including insects, parasitic nematodes and weed (Gardener & Fravel, 2002). Among the diverse, the use of plant extract as a biological control measure has gained considerable acceptance as a way to develop safe pesticides or antibiotics taken from nature in the form of phytochemicals to control pathogenic microbes desired by scientists in diverse fields.

P. granatum (pomegranate) is one of the oldest known edible fruits. It has been commonly used in traditional medicine in America, Asia, Africa and Europe for the treatment of different types of diseases. In addition to its older historical purposes, pomegranate is used in several systems of medicine for a variety of ailments (Olapour & Najafzadeh, 2010). In Ayurvedic medicine, pomegranate is viewed as “a pharmacy unto itself” and is used as an antiparasitic agent, a “blood tonic,” and to heal aphthae, and ulcers (Jurenka, 2008). Pomegranate (*P. granatum* L.) is native to the Mediterranean region and has been used extensively in the folk medicine of many countries. In India, it is used in the form of juice, concentrate, canned beverage, wine, jam, and jelly (Tripathi et al., 2014). Fresh juice contains a small amount of pectin, ascorbic acid, and flavonoids. The soluble polyphenolic content of pomegranate juice (0.2-1.0%) includes anthocyanins, catechins, ellagic tannins, and gallic and ellagic acids (Aviram et al., 2000). Previous work carried out in the laboratory showed high antioxidant activities of the methanolic extracts of pomegranate peel in various in vitro models (Singh et al., 2002).

Although several studies have attempted using natural plant extract to protect crop plants from plant pathogens, the use of *P. granatum* L. peel extracts against plant pathogenic bacteria, and its MIC and MBC values have not been investigated and there are no previous studies done for the selected bacteria. In view of the increasing quest for safer biological control measures against plant pathogens, investigating the effect of *P. granatum* L. peel extracts as a plant pathogenic bacteria are imperative, and is therefore the primary objective of this study.

1.2 Research Objectives

In order to meet the requirements for safety and non-hazardous pesticides that can be used to control plant pathogenic bacteria, *P. granatum* L. peel was used and tested to fulfill the research objectives, which were:

1. To evaluate the efficacy of *P. granatum* L. peel extracts against plant pathogenic bacteria *in vitro* and determine the Minimum Inhibitory Concentration (MIC), Minimum Bactericidal Concentration (MBC) of the extracts.
2. To determine the chemical constituents and identify the bioactive compounds of *P. granatum* L. peel extracts that exhibited the strongest antibacterial effect against plant pathogenic bacteria by using phytochemical screening test, Direct TLC bioautography assay and Gas Chromatography- Mass Spectrometry (GC-MS) Analysis.

REFERENCES

- A.Akpuaka¹, M.M. Ekwenchi¹, D.A. Dashak¹, A. D. (2013). Biological Activities of Characterized Isolates of n-Hexane Extract of Azadirachta Indica A.Juss (Neem) Leaves. *Journal of Chemical Information and Modeling*, 53(6), 160. <http://doi.org/10.1017/CBO9781107415324.004>
- Abdollahzadeh, S., Mashouf, R., Mortazavi, H., Moghaddam, M., Roozbahani, N., & Vahedi, M. (2011). Antibacterial and antifungal activities of punica granatum peel extracts against oral pathogens. *Journal of Dentistry (Tehran, Iran)*, 8(1), 1–6. Retrieved from <http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=3184731&tool=pmcentrez&rendertype=abstract>
- Aerts, M. M. L., Hogenboom, A. C., & Brinkman, U. A. T. (1995). Analytical strategies for the screening of veterinary drugs and their residues in edible products. *Journal of Chromatography B: Biomedical Applications*, 667(1), 1–40. [http://doi.org/10.1016/0378-4347\(95\)00021-A](http://doi.org/10.1016/0378-4347(95)00021-A)
- Afaq, F., Saleem, M., Krueger, C. G., Reed, J. D., & Mukhtar, H. (2005). Anthocyanin- and hydrolyzable tannin-rich pomegranate fruit extract modulates MAPK and NF- κ B pathways and inhibits skin tumorigenesis in CD-1 mice. *International Journal of Cancer*, 113(3), 423–433. <http://doi.org/10.1002/ijc.20587>
- Ahmad, I., & Aqil, F. (2007). *In vitro* efficacy of bioactive extracts of 15 medicinal plants against ES β L-producing multidrug-resistant enteric bacteria. *Microbiological Research*, 162, 264–275. <http://doi.org/10.1016/j.micres.2006.06.010>
- Aidah, N., Abdullah, N., Oskoueian, E., Sio, C. C., & Saad, W. Z. (2014). Membrane-active antibacterial compounds in methanolic extracts of Jatropha curcas and their mode of action against staphylococcus aureus S1434 and escherichia coli E216. *International Journal of Agriculture and Biology*, 16(4), 723–730.
- Alabri, T. H. A., Al Musalami, A. H. S., Hossain, M. A., Weli, A. M., & Al-Riyami, Q. (2014). Comparative study of phytochemical screening, antioxidant and antimicrobial capacities of fresh and dry leaves crude plant extracts of *Datura metel L.* *Journal of King Saud University - Science*, 26(3), 237–243. <http://doi.org/10.1016/j.jksus.2013.07.002>
- Alo, M. N., Anyim, C., Igwe, J. C., Elom, M., & Uchenna, D. S. (2012). Antibacterial activity of water, ethanol and methanol extracts of *Ocimum gratissimum*, *Vernonia amygdalina* and *Aframomum melegueta*. *Advances in Applied Science Research*, 3(2), 844–848.
- Al-Zubaydi SR., Al-Hmdany MA., R. S. (2009). Antibacterial effect of some medicinal plant extracts against some pathogenic bacterial strains. *Journal of Duhok University.*, 12(1), 244–249.
- Amorati, R., & Valgimigli, L. (2012). Modulation of the antioxidant activity of

phenols by non-covalent interactions. *Organic & Biomolecular Chemistry*, 10, 4147. <http://doi.org/10.1039/c2ob25174d>

- Anandhan, S. (2011). Biorestraining potentials of marine macroalgae collected from Rameshwaram, Tamil nadu. *Jresearchbiology.Com*, 1(5), 385–392. Retrieved from <http://jresearchbiology.com/Documents/RA0100.pdf>
- Anne K. Vidaver and Patricia A. Lambrecht. (2004). Bacteria as Plant Pathogens. Retrieved October 13, 2015, from <http://www.apsnet.org/edcenter/intropp/pathogengroups/pages/bacteria.aspx>
- Applicable, N. (2003). Rajiv Gandhi University Of Health Sciences, Karnataka. *British Pharmacopoeia*, 1, 4. Retrieved from http://www.rguhs.ac.in/cdc/onlinecdc/uploads/04_P007_2249.doc
- Araud-Razou, I., Vasse, J., Montrozier, H., Etchebar, C., & Trigalet, A. (1998). Detection and visualization of the major acidic exopolysaccharide of *Ralstonia solanacearum* and its role in tomato root infection and vascular colonization. *European Journal of Plant Pathology*, 104(8), 795–809. <http://doi.org/10.1023/A:1008690712318>
- Ashokkumar & Rajkumar (2010). Phytochemical screening and antimicrobial activity from five Indian medicinal plants against human pathogens.
- Aviram, M., Dornfeld, L., Rosenblat, M., Volkova, N., Kaplan, M., Coleman, R., ... Fuhrman, B. (2000). Pomegranate juice consumption reduces oxidative stress, atherogenic modifications to LDL, and platelet aggregation: studies in humans and in atherosclerotic apolipoprotein E-deficient mice. *The American Journal of Clinical Nutrition*, 71(5), 1062–76. [http://doi.org/10.1016/S0021-9150\(00\)80502-X](http://doi.org/10.1016/S0021-9150(00)80502-X)
- Avrova, A. O., Hyman, L. J., Toth, R. L., & Toth, I. K. (2002). Application of amplified fragment length polymorphism fingerprinting for taxonomy and identification of the soft rot bacteria *Erwinia carotovora* and *Erwinia chrysanthemi*. *Applied and Environmental Microbiology*. <http://doi.org/10.1128/AEM.68.4.1499>
- Báidez, A. G., Gómez, P., Del Río, J. A., & Ortuño, A. (2006). Antifungal capacity of major phenolic compounds of *Olea europaea* L. against *Phytophthora megasperma* Drechsler and *Cylindrocarpon destructans* (Zinssm.) Scholten. *Physiological and Molecular Plant Pathology*, 69(4-6), 224–229. <http://doi.org/10.1016/j.pmpp.2007.05.001>
- Bakhru, H. K. (1992). *Herbs that Heal: Natural Remedies for Good Health*. Retrieved from <https://books.google.com/books?hl=ar&lr=&id=C0D3z66O8Q8C&pgis=1>
- Ballhorn, D. J., Kautz, S., Heil, M., & Hegeman, A. D. (2009). Cyanogenesis of Wild Lima Bean (*Phaseolus lunatus* L.) Is an Efficient Direct Defence in Nature. *PLoS ONE*, 4(5), e5450. <http://doi.org/10.1371/journal.pone.0005450>

- Baris, O., Gulluce, M., Sahin, F., Ozer, H., & Kilic, H. (2006). Biological activities of the essential oil and methanol extract of *Achillea Biebersteinii* Afan. *Asteraceae) Turk J ...*, 30, 65–73. Retrieved from ;*TURKISH JOURNAL OF BIOLOGY*;2006;30;-;65;73
- Basri, D. F., Tan, L. S., Shafiei, Z., & Zin, N. M. (2012). In Vitro Antibacterial Activity of Galls of *Quercus infectoria* Olivier against Oral Pathogens. *Evidence-Based Complementary and Alternative Medicine : eCAM*, 2012, 632796. <http://doi.org/10.1155/2012/632796>
- Basu, A., & Penugonda, K. (2009). Pomegranate juice: A heart-healthy fruit juice. *Nutrition Reviews*, 67(1), 49–56. <http://doi.org/10.1111/j.1753-4887.2008.00133.x>
- Batista, R., De Jesus Silva Júnior, A., & De Oliveira, A. B. (2009). Plant-derived antimalarial agents: New leads and efficient phytomedicines. part II. non-alkaloidal natural products. *Molecules*, 14(8), 3037–3072. <http://doi.org/10.3390/molecules14083037>
- Belkacem, N., Djaziri, R., Lahfa, F., El-Haci, I. A., & Boucherit, Z. (2014). Phytochemical screening and in vitro antioxidant activity of various *Punica granatum* L. Peel extracts from Algeria: A comparative study. *Phytotherapie*, 12(6), 372–379. <http://doi.org/10.1007/s10298-014-0850-x>
- Beltran De Heredia, J., Torregrosa, J., Dominguez, J. R., & Peres, J. a. (2001). Kinetic model for phenolic compound oxidation by Fenton's reagent. *Chemosphere*, 45(1), 85–90. [http://doi.org/10.1016/S0045-6535\(01\)00056-X](http://doi.org/10.1016/S0045-6535(01)00056-X)
- Bhardwaj, S. K., & Laura, J. S. (2009). Antibacterial activity of some plant-extracts against plant pathogenic bacteria *Xanthomonas campestris* pv . *campestris*. *Indian J. Agric. Res*, 43(1), 26–31.
- Biedermann, M., & Grob, K. (2015). Comprehensive two-dimensional gas chromatography for characterizing mineral oils in foods and distinguishing them from synthetic hydrocarbons. *Journal of Chromatography. A*, 1375, 146–53. <http://doi.org/10.1016/j.chroma.2014.11.064>
- Biswas, K., Chattopadhyay, I., Banerjee, R. K., & Bandyopadhyay, U. (2002). Biological activities and medicinal properties of neem (*Azadirachta indica*). *Current Science*, 82(11), 1336–1345.
- Bononi, M., & Tateo, F. (2009). Identification of diisobutyl phthalate (DIBP) suspected as possible contaminant in recycled cellulose for take-away pizza boxes. *Packaging Technology and Science*, 22(1), 53–58. <http://doi.org/10.1002/pts.805>
- Braga, L. C., Shupp, J. W., Cummings, C., Jett, M., Takahashi, J. a., Carmo, L. S., ... Nascimento, a. M. a. (2005). Pomegranate extract inhibits *Staphylococcus aureus* growth and subsequent enterotoxin production. *Journal of Ethnopharmacology*, 96, 335–339. <http://doi.org/10.1016/j.jep.2004.08.034>
- Buddenhagen, I., & Kelman, A. (1964). Biological and physiological aspects of bacterial wilt caused by *Pseudomonas solanacearum*. *Annual Review of*

- Caporale, L. H. (1995). Chemical ecology: a view from the pharmaceutical industry. *Proceedings of the National Academy of Sciences of the United States of America*, 92(1), 75–82. <http://doi.org/10.1073/pnas.92.1.75>
- Celik, I., Temur, A., & Isik, I. (2009). Hepatoprotective role and antioxidant capacity of pomegranate (*Punica granatum*) flowers infusion against trichloroacetic acid-exposed in rats. *Food and Chemical Toxicology: An International Journal Published for the British Industrial Biological Research Association*, 47(1), 145–9. <http://doi.org/10.1016/j.fct.2008.10.020>
- Chanthaphon, S., Chanthachum, S., & Hongpattarakere, T. (2008). Antimicrobial activities of essential oils and crude extracts from tropical Citrus spp. Against food-related microorganisms. *Songklanakarin Journal of Science and Technology*, 30(SUPPL. 1), 125–131.
- Chauhan, J. S., & Chauhan, D. (2001). Flavonoid Diglycoside from *Punica granatum*. *Pharmaceutical Biology*. <http://doi.org/10.1076/phbi.39.2.155.6254>
- Chávez, J. H., Leal, P. C., Yunes, R. A., Nunes, R. J., Barardi, C. R. M., Pinto, A. R., ... Zanetti, C. R. (2006). Evaluation of antiviral activity of phenolic compounds and derivatives against rabies virus. *Veterinary Microbiology*, 116(1-3), 53–9. <http://doi.org/10.1016/j.vetmic.2006.03.019>
- Choma, I. (2005). The Use of Thin-Layer Chromatography with Direct Bioautography for Antimicrobial Analysis. *Journal of Chromatography. A*. Retrieved from <http://www.chromatographyonline.com/use-thin-layer-chromatography-direct-bioautography-antimicrobial-analysis>
- Choma, I. M., & Grzelak, E. M. (2011). Bioautography detection in thin-layer chromatography. *Journal of Chromatography. A*, 1218(19), 2684–91. <http://doi.org/10.1016/j.chroma.2010.12.069>
- Cloete, T. E. (2003). Resistance mechanisms of bacteria to antimicrobial compounds. *International Biodeterioration and Biodegradation*, 51, 277–282. [http://doi.org/10.1016/S0964-8305\(03\)00042-8](http://doi.org/10.1016/S0964-8305(03)00042-8)
- Cohen, M. F., Sakihama, Y., & Yamasaki, H. (2001). Roles of Plant Flavonoids in Interactions with Microbes: from protection against pathogens to the mediation of mutualism. *Developmental Plant Physiology*.
- Croteau, R., & Martinkus, C. (1979). Metabolism of Monoterpenes. *Plant Physiology*, 64(2), 169–175.
- Daami-remadi, M., Phytopathologie, L. De, Régional, C., Biologique, A., & De, I. S. A. (2007). First Report of *Sclerotium rolfsii* Causing Atypical Soft Rot on Potato Tubers in Tunisia, 2(1), 59–62.
- Dai, J., & Mumper, R. J. (2010). Plant phenolics: Extraction, analysis and their antioxidant and anticancer properties. *Molecules*, 15(10), 7313–7352. <http://doi.org/10.3390/molecules15107313>

- Dar, P., Ahmed, D., Waqas, U., Saeed, R., & Chaudhery, R. (2014). Comparative Analysis of Antimicrobial Potential of Peel and Mesocarp of *Lagenaria Siceraria* Fruit Extracts in, 3, 100–105.
- Das, K., Tiwari, R. K. S., & Shrivastava, D. K. (2010). Techniques for evaluation of medicinal plant products as antimicrobial agents: current methods and future trends. *Journal of Medicinal Plants Research*, 4(2), 104–111. <http://doi.org/10.5897/JMPR09.030>
- Dash, B. K., Faruquee, H. M., Biswas, S. K., Alam, M. K., Sisir, S. M., & Prodhana, U. K. (2011). Antibacterial and Antifungal Activities of Several Extracts of *Centella asiatica* L. against Some Human Pathogenic Microbes. *Life Sciences and Medicine Research*, 2011(L), 1–5. Retrieved from <http://astonjournals.com/lsmr>
- Devienne, K. F., & Raddi, M. S. G. (2002). Screening for antimicrobial activity of natural products using a microplate photometer. *Brazilian Journal of Microbiology*, 33(2), 166–168. <http://doi.org/10.1590/S1517-83822002000200014>
- DiSilvestro, R. A., DiSilvestro, D. J., & DiSilvestro, D. J. (2009). Pomegranate extract mouth rinsing effects on saliva measures relevant to gingivitis risk. *Phytotherapy Research : PTR*, 23(8), 1123–1127. <http://doi.org/10.1002/ptr.2759>
- Donia, M., & Hamann, M. T. (2003). Marine natural products and their potential applications as anti-infective agents. *Lancet Infectious Diseases*, 3(6), 338–348. [http://doi.org/10.1016/S1473-3099\(03\)00655-8](http://doi.org/10.1016/S1473-3099(03)00655-8)
- Dufkova, V., Cabala, R., & Sevcik, V. (2012). Determination of C-5-C-12 perfluoroalkyl carboxylic acids in river water samples in the Czech Republic by GC-MS after SPE preconcentration. *Chemosphere*, 87(5), 463–469. <http://doi.org/DOI 10.1016/j.chemosphere.2011.12.029>
- Duraipandiyan, V., Ayyanar, M., & Ignacimuthu, S. (2006). Antimicrobial activity of some ethnomedicinal plants used by Paliyar tribe from Tamil Nadu, India. *BMC Complementary and Alternative Medicine*, 6, 35. <http://doi.org/10.1186/1472-6882-6-35>
- Egharevba, Henry Omoregie and Kunle, F. O. (2010). Preliminary Phytochemical and Proximate Analysis of the leaves of *Piliostigma thonningii* (Schumach.) Milne-Redhead. Retrieved November 26, 2015, from <http://www.ethnoleaflets.com/leaflets/eghokufo>
- Ehiabhi, S., Omachonu, M., & Adeola, I. (2010). Phytochemical and Pharmacognostic Investigation of Antidiabetic *Scoparia dulcis* Linn Scrophulariaceae Whole Plant Grown in Nigeria. *Researcher*, 2(6), 7–16.
- El-Toumy, S. a a, & Rauwald, H. W. (2002). Two ellagitannins from *Punica granatum* heartwood. *Phytochemistry*, 61(02), 971–974. [http://doi.org/10.1016/S0031-9422\(02\)00435-1](http://doi.org/10.1016/S0031-9422(02)00435-1)
- Fengchun, H., Liu, X., & Chen, H. (1997). Medicine for treatment of infectious oral diseases. *Chinese Patent 1145793A.[Links]*, 1997. Retrieved from

http://scholar.google.com/scholar?q=Medicine+for+treatment+of+infectious+oral+diseases.&btnG=&hl=ar&as_sdt=0%2C5#0

- Fiori, G. M. L., Fachin, A. L., Correa, V. S. C., Bertoni, B. W., Juliatti, S., Amui, S. F., França, S. C. and Pereira, A. M. S. (2013). Antimicrobial Activity and Rates of Tannins in *Stryphnodendron adstringens* Mart . Accessions Collected in the Brazilian Cerrado. *American Journal of Plant Sciences*, 4(November), 2193–2198.
- Gardan, L. (2003). Elevation of three subspecies of *Pectobacterium carotovorum* to species level: *Pectobacterium atrosepticum* sp. nov., *Pectobacterium betavascularum* sp. nov. and *Pectobacterium wasabiae* sp. nov. *International Journal of Systematic and Evolutionary Microbiology*, 53(2), 381–391. <http://doi.org/10.1099/ijs.0.02423-0>
- Gardener, B. B. M. & Fravel, D. R. (2002). Biological Control of Plant Pathogens: Research, Commercialization, and Application in the USA. *Plant Health Progress, Online*(May 2002), 1–18. <http://doi.org/10.1094/PHP-2002-0510-01-RV.Biological>
- Gaspar, L., Oliveira, A. P., Silva, L. R., & Paula, B. (2012). Metabolic and biological prospecting of, 22(2), 350–358.
- Gershenzon, J., & Dudareva, N. (2007). The function of terpene natural products in the natural world. *Nature Chemical Biology*, 3(7), 408–414. <http://doi.org/10.1038/nchembio.2007.5>
- Ghasemzadeh, A., Jaafar, H. Z. E., & Rahmat, A. (2011). Effects of solvent type on phenolics and flavonoids content and antioxidant activities in two varieties of young ginger (*Zingiber officinale* Roscoe) extracts. *Journal of Medicinal Plants Research*, 5(7), 1147–1154. <http://doi.org/10.3390/molecules15064324>
- Ghosh, A., Das, B. K., Roy, A., Mandal, B., & Chandra, G. (2008). Antibacterial activity of some medicinal plant extracts. *Journal of Natural Medicines*, 62(2), 259–262. <http://doi.org/10.1007/s11418-007-0216-x>
- Gil, M. I., Tomás-Barberán, F. A., Hess-Pierce, B., Holcroft, D. M., & Kader, A. A. (2000). Antioxidant Activity of Pomegranate Juice and Its Relationship with Phenolic Composition and Processing. *Journal of Agricultural and Food Chemistry*, 48(10), 4581–4589. <http://doi.org/10.1021/jf000404a>
- Girard, J. E. (2010). *Principles Of Environmental Chemistry*. Retrieved from <http://www.amazon.com/Principles-Environmental-Chemistry-James-Girard/dp/0763759392>
- Gracia-Garza, J. A., Blom, T. J., Brown, W., & Allen, W. (2002). Pre- and post-plant applications of copper-based compounds to control *Erwinia* soft rot of calla lilies. *Canadian Journal of Plant Pathology*, 24(3), 274–280. Retrieved from <Go to ISI>://BIOSIS:PREV200300014161
- Grierson, D. S., & Afolayan, A. J. (1999). Antibacterial activity of some indigenous plants used for the treatment of wounds in the Eastern Cape, South Africa. *Journal of Ethnopharmacology*, 66, 103–106. <http://doi.org/10.1016/S0378->

- Grigonis, D., Venskutonis, P. R., Sivik, B., Sandahl, M., & Eskilsson, C. S. (2005). Comparison of different extraction techniques for isolation of antioxidants from sweet grass (*Hierochloë odorata*). *The Journal of Supercritical Fluids*, 33(3), 223–233. <http://doi.org/10.1016/j.supflu.2004.08.006>
- Gulluce, M., Sahin, F., Sokmen, M., Ozer, H., Daferera, D., Sokmen, A., ... Ozkan, H. (2007). Antimicrobial and antioxidant properties of the essential oils and methanol extract from *Mentha longifolia* L. ssp. *longifolia*. *Food Chemistry*, 103(4), 1449–1456. <http://doi.org/10.1016/j.foodchem.2006.10.061>
- Halama, P., & Van Haluwin, C. (2004). Antifungal activity of lichen extracts and lichenic acids. *BioControl*, 49(1), 95–107.
- Hayward, A. C. (1991). Biology and epidemiology of bacterial wilt caused by *pseudomonas solanacearum*. *Annual Review of Phytopathology*, 29, 65–87. <http://doi.org/10.1146/annurev.py.29.090191.000433>
- Heftmann, E., Ko, S. T., & Bennett, R. D. (1966). Response of steroids to sulfuric acid in thin-layer chromatography. *Journal of Chromatography*, 21(3), 490–4. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/5945063>
- Hegnauer, R. (1988). Biochemistry, distribution and taxonomic relevance of higher plant alkaloids. *Phytochemistry*, 27(8), 2423–2427. [http://doi.org/10.1016/0031-9422\(88\)87006-7](http://doi.org/10.1016/0031-9422(88)87006-7)
- Hema R., S. K. and K. A. (2011). GC/MS Determination of Bioactive Components of *Murraya koenigii*. *Journal of American Science*, 7(1), 2009–2012.
- Hemingway, R. W., Karchesy, J. J., & Branham, S. J. (Eds.). (1989). *Chemistry and Significance of Condensed Tannins*. Boston, MA: Springer US. <http://doi.org/10.1007/978-1-4684-7511-1>
- Herbivores: Their Interactions with Secondary Plant Metabolites: Ecological and Evolutionary Processes.* (n.d.). Retrieved from <https://books.google.com/books?hl=ar&lr=&id=g6BlbAfDveQC&pgis=1>
- Hossain, M. A., Shah, M. D., Gnanaraj, C., & Iqbal, M. (2011). In vitro total phenolics, flavonoids contents and antioxidant activity of essential oil, various organic extracts from the leaves of tropical medicinal plant *Tetragium* from Sabah. *Asian Pacific Journal of Tropical Medicine*, 4(9), 717–721. [http://doi.org/10.1016/S1995-7645\(11\)60180-6](http://doi.org/10.1016/S1995-7645(11)60180-6)
- Hsouna, A. Ben, Trigui, M., Mansour, R. Ben, Jarraya, R. M., Damak, M., & Jaoua, S. (2011). Chemical composition, cytotoxicity effect and antimicrobial activity of *Ceratonia siliqua* essential oil with preservative effects against *Listeria* inoculated in minced beef meat. *International Journal of Food Microbiology*, 148(1), 66–72. <http://doi.org/10.1016/j.ijfoodmicro.2011.04.028>
- Huang, H. C., Chang, T. Y., Chang, L. Z., Wang, H. F., Yih, K. H., Hsieh, W. Y., & Chang, T. M. (2012). Inhibition of melanogenesis Versus antioxidant properties of essential oil extracted from leaves of *vitex negundo* linn and

chemical composition analysis by GC-MS. *Molecules*, 17, 3902–3916. <http://doi.org/10.3390/molecules17043902>

- Hussin, N. M., Muse, R., Ahmad, S., Ramli, J., Mahmood, M., Sulaiman, M. R., ... Aziz, K. N. K. (2009). Antifungal activity of extracts and phenolic compounds from *Barringtonia racemosa* L. (Lecythidaceae). *African Journal of Biotechnology*, 8(12), 2835–2842. <http://doi.org/10.5897/AJB09.450>
- Inderjit, & Keating, K. I. (1999). Allelopathy: Principles, Procedures, Processes, and Promises for Biological Control. *Advances in Agronomy*, 67(C), 141–231.
- Islam, N., Parveen, S. a., Nakazawa, N., Marston, a., & Hostettmann, K. (2003). Bioautography with the Fungus *Valsa ceratosperma* in the Search for Antimycotic Agents. *Pharmaceutical Biology*, 41(8), 637–640. <http://doi.org/10.1080/13880200390502621>
- Ismail, T., Sestili, P., & Akhtar, S. (2012). Pomegranate peel and fruit extracts: A review of potential anti-inflammatory and anti-infective effects. *Journal of Ethnopharmacology*, 143(2), 397–405. <http://doi.org/10.1016/j.jep.2012.07.004>
- James, A. T., & Martin, A. J. P. (1952). Gas-liquid partition chromatography. A technique for the analysis of volatile materials. *The Analyst*, 77(921), 915–931. <http://doi.org/10.1007/BF01520304>
- Jassim, S. A. A. 1998. (1998, November 24). Antiviral or antifungal composition comprising an extract of pomegranate rind or other plants and method of use. Retrieved from <http://www.google.com/patents/US5840308>
- Jia, C., & Zia, C. (1998). A fungicide made from Chinese medicinal herb extract. *Chinese Patent CN*, 1998. Retrieved from http://scholar.google.com/scholar?q=Fungicide+made+from+Chinese+medicinal+herb+extract+&btnG=&hl=ar&as_sdt=0%2C5#0
- Jorgensen, J. H., & Ferraro, M. J. (2009). Antimicrobial susceptibility testing: a review of general principles and contemporary practices. *Clinical Infectious Diseases: An Official Publication of the Infectious Diseases Society of America*, 49(11), 1749–1755. <http://doi.org/10.1086/647952>
- Joshi, B., Sah, G. P., Basnet, B. B., Bhatt, M. R., Sharma, D., Subedi, K., ... Malla, R. (2011). Phytochemical extraction and antimicrobial properties of different medicinal plants: *Ocimum sanctum* (Tulsi), *Eugenia caryophyllata* (Clove), *Achyranthes bidentata* (Datiwan) and *Azadirachta indica* (Neem). *J Microbiol Antimicrob*, 3(1), 1–7. Retrieved from [http://www.academicjournals.org/JMA/PDF/pdf2011/January/Joshi et al.pdf](http://www.academicjournals.org/JMA/PDF/pdf2011/January/Joshi%20et%20al.pdf)
- Joyce, L. F., Downes, J., Stockman, K., & Andrew, J. H. (1992). Comparison of five methods, including the PDM Epsilon meter test (E test), for antimicrobial susceptibility testing of *Pseudomonas aeruginosa*. *Journal of Clinical Microbiology*, 30(10), 2709–2713.
- Jurenka, J. S. (2008). Therapeutic applications of pomegranate (*Punica granatum* L.): a review. *Alternative Medicine Review: A Journal of Clinical Therapeutic*, 13(2), 128–144.

- Kado, C. I. (2006). *Erwinia* and Related Genera, 443–450. <http://doi.org/10.1007/0-387-30746-x>
- Kamal, J., & Bano, A. (2008). Potential allelopathic effects of sunflower (*Helianthus annuus* L.) on microorganisms. *African Journal of Biotechnology*, 7(22).
- Kamatou, G. P. P., Makunga, N. P., Ramogola, W. P. N., & Viljoen, a. M. (2008). South African *Salvia* species: A review of biological activities and phytochemistry. *Journal of Ethnopharmacology*, 119(3), 664–672. <http://doi.org/10.1016/j.jep.2008.06.030>
- Khanam, Z., Wen, C. S., & Bhat, I. U. H. (2014). Phytochemical screening and antimicrobial activity of root and stem extracts of wild *Eurycoma longifolia* Jack (Tongkat Ali). *Journal of King Saud University - Science*, 27(1), 23–30. <http://doi.org/10.1016/j.jksus.2014.04.006>
- Kianbakht, S., & Jahaniani, F. (2003). Evaluation of antibacterial activity of *Tribulus terrestris* L. growing in Iran. *Iranian Journal of Pharmacology & Therapeutics*, 2(1), 22–24. Retrieved from <http://scholar.google.com/scholar?hl=en&btnG=Search&q=intitle:Evaluation+of+Antibacterial+Activity+of+Tribulus+terrestris+L.+Growing+in+Iran#0>
- Kleiman, R., & Payne-Wahl, K. L. (1984). Fatty acid composition of seed oils of the meliaceae, including one genus rich in cis-vaccenic acid. *Journal of the American Oil Chemists' Society*, 61(12), 1836–1838. <http://doi.org/10.1007/BF02540810>
- Kondo, T., Yoshida, K., Nakagawa, A., Kawai, T., Tamura, H., & Goto, T. (1992). Structural Basis Of Blue-Color Development In Flower Petals From *Commelina-Communis*. *Nature*, 358(6386), 515–518.
- Krasteva, I., Nikolova, I., Danchev, N., & Nikolov, S. (2004). Phytochemical analysis of ethyl acetate extract from *Astragalus corniculatus* Bieb. and brain antihypoxic activity. *Acta Pharmaceutica (Zagreb, Croatia)*, 54, 151–156.
- Kulkarni, A. P., & Aradhya, S. M. (2005). Chemical changes and antioxidant activity in pomegranate arils during fruit development. *Food Chemistry*, 93(2), 319–324. <http://doi.org/10.1016/j.foodchem.2004.09.029>
- Kumar Bhandary, S., Kumari, S. N., Bhat, V. S., & Prasad Bekal, M. (2012). Preliminary phytochemical screening of various extracts of *punica granatum* peel, whole fruit and seeds 1 2 3. *Nitte University Journal of Health Science PRELIMINARY PHYTOCHEMICAL -Satheesh Kumar Bhandary NUJHS*, 2(4), 2249–7110.
- Kumar, S., Samydurai, P., Ramakrishnan, R., & Nagarajan, N. (2014). Innovare Academic Sciences Gas chromatography and mass spectrometry analysis of bioactive constituents of *adiantum capillus-veneris* l.
- Kumar Sharma, A., Gangwar, M., Tilak, R., Nath, G., Sudhir Kumar Sinha, A., Bhusan Tripathi, Y., & Kumar, D. (2012). Comparative in vitro Antimicrobial and Phytochemical Evaluation of Methanolic Extract of Root, Stem and Leaf of *Jatropha curcas* Linn. *Pharmacognosy Journal*, 4(30), 34–

40. <http://doi.org/10.5530/pj.2012.30.7>

- Kunle, O., Okogun, J., Egamana, E., Emojevwe, E., & Shok, M. (2003). Antimicrobial activity of various extracts and carvacrol from *Lippia multiflora* leaf extract. *Phytomedicine: International Journal of Phytotherapy and Phytopharmacology*, 10(1), 59–61. <http://doi.org/10.1078/094471103321648674>
- Kvasenkov, O., Lomachinski, V., & Goren’Kov, E. (1999). Method of producing beverages on juice base. *Russian Patent 2129396C1, 1999*. Retrieved from http://scholar.google.com/scholar?q=Kvasenkov%2C+O.+I.%3B+Lomachinski%2C+V.+A.%3B+Goren%E2%80%99Kov%2C+Eh.+S.+Method+of+producing+beverages+on+juice+base&btnG=&hl=ar&as_sdt=0%2C5#0
- Langfield, R. D., Scarano, F. J., Heitzman, M. E., Kondo, M., Hammond, G. B., & Neto, C. C. (2004). Use of a modified microplate bioassay method to investigate antibacterial activity in the Peruvian medicinal plant *Peperomia galioides*. *Journal of Ethnopharmacology*, 94(2-3), 279–281. <http://doi.org/10.1016/j.jep.2004.06.013>
- Lansky, E. P., & Newman, R. a. (2007). *Punica granatum* (pomegranate) and its potential for prevention and treatment of inflammation and cancer. *Journal of Ethnopharmacology*, 109(2), 177–206. <http://doi.org/10.1016/j.jep.2006.09.006>
- Lau, A.-J., Holmes, M. J., Woo, S.-O., & Koh, H.-L. (2003). Analysis of adulterants in a traditional herbal medicinal product using liquid chromatography–mass spectrometry–mass spectrometry. *Journal of Pharmaceutical and Biomedical Analysis*, 31(2), 401–406. [http://doi.org/10.1016/S0731-7085\(02\)00637-4](http://doi.org/10.1016/S0731-7085(02)00637-4)
- Lavanya, G. & Brahma Prakash, G. P. (2011). Phytochemical screening and antimicrobial activity of compounds from selected medicinal and aromatic plants *International Journal of Science and Nature*, 2(2), 287–291. Retrieved from <http://www.slideshare.net/drshkrp/citation-paper-ijns-vol2225>
- LeBoeuf, J., Cuppels, D., Dick, J., Pitblado, R., Loewen, S. and Celetti, M. (2011). Bacterial Diseases Of Tomato: Bacterial Spot, Bacterial Speck, Bacterial Canker. Retrieved from <http://www.omafra.gov.on.ca/english/crops/facts/05-069.htm>
- Lee, J.-C., Kim, H.-R., Kim, J., & Jang, Y.-S. (2002). Antioxidant property of an ethanol extract of the stem of *Opuntia ficus-indica* var. *saboten*. *Journal of Agricultural and Food Chemistry*, 50(22), 6490–6. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/12381138>
- Lelliott and Dickey, (1984). (n.d.). Lelliott, R.A. and Dickey, R.S. (1984) Genus VII. *Erwinia*. In: Krieg, N.R. and Holt J.G., Eds., *Bergey’s Manual of Systematic Bacteriology*, Williams & Wilkins, Baltimore, 469-476. |Reference|Scientific Research Publish. Retrieved October 13, 2015, from <http://www.ljemail.org/reference/ReferencesPapers.aspx?ReferenceID=147559>

- Lim, J.-A., Jee, S., Lee, D. H., Roh, E., Jung, K., Oh, C., & Heu, S. (2013). Biocontrol of *Pectobacterium carotovorum* subsp. *carotovorum* using bacteriophage PP1. *Journal of Microbiology and Biotechnology*, 23(8), 1147–53. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/23727798>
- Lourens, A. C. U., Reddy, D., Başer, K. H. C., Viljoen, A. M., & Van Vuuren, S. F. (2004). In vitro biological activity and essential oil composition of four indigenous South African *Helichrysum* species. *Journal of Ethnopharmacology*, 95(2-3), 253–258. <http://doi.org/10.1016/j.jep.2004.07.027>
- Mahdihassan, S. (1984). Outline of the beginnings of alchemy and its antecedents. *The American Journal of Chinese Medicine*, 12(1-4), 32–42. <http://doi.org/10.1142/S0192415X84000039>
- Mahesh, B., & Satish, S. (2008). Antimicrobial Activity of Some Important Medicinal Plant Against Plant and Human Pathogens. *World Journal of Agricultural Sciences*, 4, 839–843. Retrieved from [http://idosi.org/wjas/wjas4\(s\)/7.pdf](http://idosi.org/wjas/wjas4(s)/7.pdf)
- Mansfield, J., Genin, S., Magori, S., Citovsky, V., Sriariyanum, M., Ronald, P., ... Foster, G. D. (2012). Top 10 plant pathogenic bacteria in molecular plant pathology. *Molecular Plant Pathology*, 13(6), 614–629. <http://doi.org/10.1111/j.1364-3703.2012.00804.x>
- March, J. G., Simonet, B. M., Grases, F., Muñoz, J. A., & Valiente, M. (2003). Determination of trace amounts of oxalate in renal calculi and related samples by gas chromatography-mass spectrometry. *Chromatographia*, 57(11-12), 811–817. <http://doi.org/10.1007/BF02491770>
- Maruthupandian, A., & Mohan, V. R. (2011). GC-MS analysis of some bioactive constituents of *Pterocarpus marsupium* Roxb., 3(3), 1652–1657.
- Maskovic, P., Manojlovic, N., Mandic, A., Misan, A., Milovanovic, I., Radojkovic, M., ... Solujic, S. (2012). Phytochemical screening and biological activity of extracts of plant species *Halacsysa sendtneri* (Boiss.) Dörfel. *Chemical Industry*, 66(1), 43–51. <http://doi.org/10.2298/HEMIND110828068M>
- Matsumoto, I., & Kuhara, T. (1996). A new chemical diagnostic method for inborn errors of metabolism by mass spectrometry—rapid, practical, and simultaneous urinary metabolites analysis. *Mass Spectrometry Reviews*, 15(1), 43–57. [http://doi.org/10.1002/\(SICI\)1098-2787\(1996\)15:1<43::AID-MAS3>3.0.CO;2-](http://doi.org/10.1002/(SICI)1098-2787(1996)15:1<43::AID-MAS3>3.0.CO;2-)
- Mazid, M., Khan, T. a., & Mohammad, F. (2011). Role of secondary metabolites in defense mechanisms of plants. *Biology and Medicine*, 3(2), 232–249. <http://doi.org/citeulike-article-id:10435815>
- McGoodwin Family Website Home Page. (n.d.). Retrieved October 4, 2015, from <http://www.mcgoodwin.net/>
- Mcgoodwin, M. (2008). The Physiology Of Higher Plants An Outline. *Energy Conversion*, (Mcm), 1–138.

- Meléndez, P. a., & Capriles, V. a. (2006). Antibacterial properties of tropical plants from Puerto Rico. *Phytomedicine*, 13(4), 272–276. <http://doi.org/10.1016/j.phymed.2004.11.009>
- Mendoza, M. T. (1998). What's New in Antimicrobial Susceptibility Testing? *Philippine Journal of Microbiology and Infectious Diseases*, 27(3), 113–115. Retrieved from https://www.researchgate.net/publication/237559281_What's_New_in_Antimicrobial_Susceptibility_Testing
- Mertens, L., Struelens, M. J., Nonhoff, C., Auwera, P. V. a N. D. E. R., Serruys, E., Pour, G., ... Bordet, I. J. (1995). Evaluation of Rapid ATB Staph for 5-Hour Antimicrobial Susceptibility Testing of Staphylococcus aureus, 33(9), 2395–2399.
- Mert-Türk, F. (2006). Saponins versus plant fungal pathogens. *J. Cell Mol. Biol*, (5), 13–17. Retrieved from <http://jcmb.halic.edu.tr/pdf/5-1/saponins.pdf>
- Meyer, J. J., & Afolayan, a J. (1995). Antibacterial activity of Helichrysum aureonitens (Asteraceae). *Journal of Ethnopharmacology*, 47(2), 109–111. [http://doi.org/http://dx.doi.org/10.1016/0378-8741\(95\)01261-B](http://doi.org/http://dx.doi.org/10.1016/0378-8741(95)01261-B)
- Miguel, G., Fontes, C., Antunes, D., Neves, A., & Martins, D. (2004). Anthocyanin Concentration of “Assaria” Pomegranate Fruits During Different Cold Storage Conditions. *Journal of Biomedicine & Biotechnology*, 2004(5), 338–342. <http://doi.org/10.1155/S1110724304403076>
- Mori, A., Nishino, C., Enoki, N., & Tawata, S. (1987). Antibacterial activity and mode of action of plant flavonoids against Proteus vulgaris and Staphylococcus aureus. *Phytochemistry*, 26(8), 2231–2234. <http://doi.org/10.1016/S0031>
- Mori-Okamoto, J., Otawara-Hamamoto, Y., Yamato, H., & Yoshimura, H. (2004). Pomegranate extract improves a depressive state and bone properties in menopausal syndrome model ovariectomized mice. *Journal of Ethnopharmacology*, 92(1), 93–101. <http://doi.org/10.1016/j.jep.2004.02.006>
- Muñoz-Olivas, R. (2004). Screening analysis: An overview of methods applied to environmental, clinical and food analyses. *TrAC - Trends in Analytical Chemistry*, 23(3), 203–216. [http://doi.org/10.1016/S0165-9936\(04\)00318-8](http://doi.org/10.1016/S0165-9936(04)00318-8)
- Mushore, J., & Matuvhunye, M. (2013). Antibacterial properties of Mangifera indica on Staphylococcus aureus. *African Journal of Clinical and Experimental Microbiology*, 14(2), 62–74. <http://doi.org/10.4314/ajcem.v14i2.4>
- Nasrine, S., El-darier, S. M., & El-taher, H. M. (2011). Allelopathic Effect from some Medicinal Plants and Their Potential Uses as control of weed. *Environment and Chemistry*, 24, 15–22.
- Naziri, Z., Rajaian, H., & Firouzi, R. (2012). Antibacterial effects of Iranian native sour and sweet pomegranate (Punica granatum) peel extracts against various pathogenic bacteria. *Iranian Journal of Veterinary Research*, 13(41).

- Ncube, N. S., Afolayan, A. J., Okoh, A. I., Ncube, N. S., Afolayan, A. J., Afolayan, A. J., ... Okoh, A. I. (2008). Assessment techniques of antimicrobial properties of natural compounds of plant origin : current methods and future trends. *African Journal of Biotechnology*, 7(12), 1797–1806. Retrieved from <Go to ISI>://000257358700002
- Nostro, a, Germanò, M. P., D'angelo, V., Marino, a, & Cannatelli, M. a. (2000). Extraction methods and bioautography for evaluation of medicinal plant antimicrobial activity. *Letters in Applied Microbiology*, 30(5), 379–384. <http://doi.org/10.1046/j.1472-765x.2000.00731.x>
- Obadoni, B. O., & Ochuko, P. O. (2002). Phytochemical Studies And Comparative Efficacy Of The Crude Extracts Of Some Haemostatic Plants In Edo And Delta States Of Nigeria. *Global Journal of Pure and Applied Sciences*, 8(2), 203–208. <http://doi.org/10.4314/gjpas.v8i2.16033>
- Obeidat, M. (2011). Antimicrobial activity of some medicinal plants against multidrug resistant skin pathogens. *Journal of Medicinal Plants Research*, 5(16), 3856.
- Odjakova, M., & Hadjiivanova, C. (2001). The complexity of pathogen defense in plants. *Bulgarian Journal of Plant Physiology*, 27, 101–109. <http://doi.org/10.1002/lt.22131>
- Olapour, S., & Najafzadeh, H. (2010). Evaluation Analgesic , Anti-Inflammatory and Antiepileptic Effect of Hydro Alcoholic Peel Extract of *Punica granatum* (pomegranate). *Asian Journal of Medical Sciences*, 2(6), 266–270.
- Omoriege, H., Nkiruka, O. P., Sabo, M., Koma, S., & Ibumeh, J. (2010). Phytochemical Analysis and Antimicrobial Activity of *Punica granatum L.* (fruit bark and leaves). *New York Science Journal*, 3(12), 91–98.
- Patel, J. M. (2010). A Review of Potential Health Benefits of Flavonoids. *Lethbridge Undergraduate Research Journal*, 3(2), 2–6.
- Patel, R., Patel, A., Vaghasiya, D., & Nagee, A. (2012). Antimicrobial Evaluation of Hibiscus rosa-sinensis Plant Extracts against Some Pathogenic Bacteria. *Bulletin of Environmental and Scientific Research*, 1(3), 14–17. Retrieved from <http://www.besr.org.in>
- Pereira, J. A., Oliveira, I., Sousa, A., Valentão, P., Andrade, P. B., Ferreira, I. C. F. R., ... Estevinho, L. (2007). Walnut (*Juglans regia L.*) leaves: Phenolic compounds, antibacterial activity and antioxidant potential of different cultivars. *Food and Chemical Toxicology*, 45(11), 2287–2295. <http://doi.org/10.1016/j.fct.2007.06.004>
- Pérombelon, M. C. M. (2002). Potato diseases caused by soft rot erwinias: An overview of pathogenesis. *Plant Pathology*. <http://doi.org/10.1046/j.0032-0862.2001.Short title.doc.x>
- Perombelon, M. C. M., & Kelman, A. (1980). Ecology of the Soft Rot Erwinias. *Annual Review of Phytopathology*, 18(1), 361–387. <http://doi.org/10.1146/annurev.py.18.090180.002045>

- Poyrazoğlu, E., Gökmen, V., & Artık, N. (2002). Organic Acids and Phenolic Compounds in Pomegranates (*Punica granatum* L.) Grown in Turkey. *Journal of Food Composition and Analysis*, 15, 567–575. <http://doi.org/10.1006/jfca.2002.1071>
- Price, K. R., Johnson, I. T., & Fenwick, G. R. (1987). The chemistry and biological significance of saponins in foods and feedingstuffs. *Critical Reviews in Food Science and Nutrition*, 26(1), 27–135. <http://doi.org/10.1080/10408398709527461>
- Prior P. et al., 1998. (2013). *Bacterial Wilt Disease: Molecular and Ecological Aspects*. Springer Science & Business Media. Retrieved from <https://books.google.com/books?id=run7CAAAQBAJ&pgis=1>
- Putnam, A. R. (1988). Allelochemicals from Plants as Herbicides on JSTOR. Retrieved February 9, 2016, from http://www.jstor.org/stable/3987390?seq=1#page_scan_tab_contents
- Rajan, S., Mahalakshmi, S., Deepa, V., Sathya, K., Shajitha, S., & Thirunalasundari, T. (2011). Antioxidant potentials of *Punica granatum* fruit rind extracts. *International Journal of Pharmacy and Pharmaceutical Sciences*, 3(3).
- Rajeswari, G., Murugan, M., & Mohan, V. R. (2012). GC-MS analysis of bioactive components of *Hugonia mystax* L. (Linaceae). *Research Journal of Pharmaceutical, Biological and Chemical Sciences*, 3(4), 301–308.
- Rao, K. B. (2013). *In vitro* Evaluation of antibacterial activity of five indigenous plants extract against five bacterial pathogens of human, 5.
- Reddy, M. K., Gupta, S. K., Jacob, M. R., Khan, S. I., & Ferreira, D. (2007a). Antioxidant, antimalarial and antimicrobial activities of tannin-rich fractions, ellagitannins and phenolic acids from *Punica granatum* L. *Planta Medica*, 73(5), 461–7. <http://doi.org/10.1055/s-2007-967167>
- Reddy, M. K., Gupta, S. K., Jacob, M. R., Khan, S. I., & Ferreira, D. (2007b). Antioxidant, antimalarial and antimicrobial activities of tannin-rich fractions, ellagitannins and phenolic acids from *Punica granatum* L. *Planta Medica*, 73(5), 461–467. <http://doi.org/10.1055/s-2007-967167>
- Reigosa, M. J., Pedrol, N., & Gonzalez, L. (2006). *Allelopathy: A physiological process with ecological implications. Allelopathy: A Physiological Process with Ecological Implications*. <http://doi.org/10.1007/1-4020-4280-9>
- Rios, J. L., & Recio, M. C. (2005). Medicinal plants and antimicrobial activity. *Journal of Ethnopharmacology*, 100(1-2), 80–84. <http://doi.org/10.1016/j.jep.2005.04.025>
- Rispail, N., Morris, P., & Webb, K. J. (2005). Phenolic Compounds: Extraction and Analysis. In *Lotus japonicus Handbook* (pp. 349–354).
- Rizvi, S. J. H., & Rizvi, V. (Eds.). (1992). *Allelopathy*. Dordrecht: Springer Netherlands. <http://doi.org/10.1007/978-94-011-2376-1>
- Rosenthal, G. A. (1991). The biochemical basis for the deleterious effects of 1-

canavanine. *Phytochemistry*, 30(4), 1055–1058.
[http://doi.org/10.1016/S0031-9422\(00\)95170-7](http://doi.org/10.1016/S0031-9422(00)95170-7)

- Runyoro, D. K. B., Matee, M. I. N., Ngassapa, O. D., Joseph, C. C., & Mbwambo, Z. H. (2006). Screening of Tanzanian medicinal plants for anti-Candida activity. *BMC Complementary and Alternative Medicine*, 6, 11. <http://doi.org/10.1186/1472-6882-6-11>
- Sadik, M. S., & Asker, M. M. S. (2014). Antioxidant and antitumor activities of Pomegranate (*Punica granatum*) peel extracts, 2.
- Salada, J., Balala, L., & Vasquez, E. (2015). Phytochemical and Antibacterial Studies of *Lantana camara* L. Leaf Fraction and Essential Oil. *Ijsrp.Org*, 5(3), 1–5. Retrieved from <http://www.ijsrp.org/research-paper-0315/ijsrp-p3920.pdf>
- Salie, F., Eagles, P. F. K., & Leng, H. M. J. (1996). Preliminary antimicrobial screening of four South African Asteraceae species. *Journal of Ethnopharmacology*, 52(1), 27–33. [http://doi.org/10.1016/0378-8741\(96\)01381-5](http://doi.org/10.1016/0378-8741(96)01381-5)
- Scalbert, A. (1991). Antimicrobial properties of tannins.pdf. *Phytochemistry*.
- Schäfer, H., & Wink, M. (2009). Medicinally important secondary metabolites in recombinant microorganisms or plants: progress in alkaloid biosynthesis. *Biotechnology Journal*, 4(12), 1684–703. <http://doi.org/10.1002/biot.200900229>
- Schaller, A., & Ryan, C. A. (1996). Systemin--a polypeptide defense signal in plants. *BioEssays : News and Reviews in Molecular, Cellular and Developmental Biology*, 18(1), 27–33. <http://doi.org/10.1002/bies.950180108>
- Scheuerell, S., & Mahaffee, W. (2002). Compost tea: Principles and prospects for plant disease control. *Compost Science & Utilization*, 10(4), 313–338. Retrieved from http://apps.webofknowledge.com.proxy.library.cornell.edu/full_record.do?product=UA&search_mode=GeneralSearch&qid=120&SID=2DeKI57IjfNd2a7JkIB&page=1&doc=1
- Schmourlo, G., Mendonça-Filho, R. R., Alviano, C. S., & Costa, S. S. (2005). Screening of antifungal agents using ethanol precipitation and bioautography of medicinal and food plants. *Journal of Ethnopharmacology*, 96(3), 563–568. <http://doi.org/10.1016/j.jep.2004.10.007>
- Schubert, S. Y., Lansky, E. P., & Neeman, I. (1999). Antioxidant and eicosanoid enzyme inhibition properties of pomegranate seed oil and fermented juice flavonoids. *Journal of Ethnopharmacology*, 66(1), 11–17. [http://doi.org/10.1016/S0378-8741\(98\)00222-0](http://doi.org/10.1016/S0378-8741(98)00222-0)
- Schwartz, A. R., Potnis, N., Timilsina, S., Wilson, M., Patane, J., Martins, J. J., ... Staskawicz, B. J. (2015). Phylogenomics of *Xanthomonas* field strains infecting pepper and tomato reveals diversity in effector repertoires and identifies determinants of host specificity. *Frontiers in Microbiology*, 6, 535. <http://doi.org/10.3389/fmicb.2015.00535>

- Sciences, B., Siemens, D. H., Garner, S. H., Mitchell-Olds, T., & Callaway, R. M. (2002). Cost of defense in the context of plant competition: Brassica rapa may grow and defend. *Ecology*, 83(2), 505–517. <http://doi.org/10.2307/2680031>
- Seanego, C. T., & Ndip, R. N. (2012). Identification and antibacterial evaluation of bioactive compounds from Garcinia kola (Heckel) seeds. *Molecules (Basel, Switzerland)*, 17(6), 6569–84. <http://doi.org/10.3390/molecules17066569>
- Seeram, N., Lee, R., Hardy, M., & Heber, D. (2005). Rapid large scale purification of ellagitannins from pomegranate husk, a by-product of the commercial juice industry. *Separation and Purification Technology*, 41, 49–55. <http://doi.org/10.1016/j.seppur.2004.04.003>
- Seeram, N. P., Adams, L. S., Henning, S. M., Niu, Y., Zhang, Y., Nair, M. G., & Heber, D. (2005). In vitro antiproliferative, apoptotic and antioxidant activities of punicalagin, ellagic acid and a total pomegranate tannin extract are enhanced in combination with other polyphenols as found in pomegranate juice. *Journal of Nutritional Biochemistry*, 16(6), 360–367. <http://doi.org/10.1016/j.jnutbio.2005.01.006>
- Sheikh, N., Kumar, Y., Mishra, A. K., & Pfoze, L. (2013). Journal of Medicinal Plants Studies Phytochemical screening to validate the ethnobotanical importance of root tubers of Dioscorea species of. *Journal of Medicinal Plant Studies*, 1(6), 62–69.
- Silva, M. T., Simas, S. M., Batista, T. G., & Tomassini, P. C. & T. C. (2005). Studies on antimicrobial activity, in vitro, of Physalis angulata L. (Solanaceae) fraction and physalin B bringing out the importance of assay determination. *Memórias Do Instituto Oswaldo Cruz*, 100(7), 779–782. Retrieved from <http://www.bioline.org.br/request?oc05163>
- Simms, E. (1992). Costs of plant resistance to herbivory. In *Plant resistance to herbivores and pathogens: ecology, evolution, and genetics* (pp. 392–425).
- Singh, R. P., Chidambara Murthy, K. N., & Jayaprakasha, G. K. (2002a). Studies on the Antioxidant Activity of Pomegranate (Punica granatum) Peel and Seed Extracts Using in Vitro Models. *Journal of Agricultural and Food Chemistry*, 50(1), 81–86. <http://doi.org/10.1021/jf010865b>
- Singh, R. P., Chidambara Murthy, K. N., & Jayaprakasha, G. K. (2002b). Studies on the antioxidant activity of pomegranate (Punica granatum) peel and seed extracts using in vitro models. *Journal of Agricultural and Food Chemistry*, 50, 81–86. <http://doi.org/10.1021/jf010865b>
- Singh, R. P., Chidambara Murthy, K. N., & Jayaprakasha, G. K. (2002c). Studies on the antioxidant activity of pomegranate (Punica granatum) peel and seed extracts using in vitro models. *Journal of Agricultural and Food Chemistry*, 50(1), 81–86.
- Singh, S., Srivastava, R., & Choudhary, S. (2010). Antifungal and HPLC analysis of the crude extracts of Acorus calamus, Tinospora cordifolia and Celestrus paniculatus. *Journal of Agricultural Technology*, 6(61), 149–158.

- Snijder, R. C., Lindhout, P., & Van Tuyl, J. M. (2004). Genetic control of resistance to soft rot caused by *Erwinia carotovora* subsp. *carotovora* in *Zantedeschia* spp. (Araceae), section *Aestivae*. *Euphytica*, *136*(3), 319–325. <http://doi.org/10.1023/B:EUPH.0000032734.83569.f4>
- Stankovic, M. S., Niciforovic, N., Mihailovic, V., Topuzovic, M., & Solujic, S. (2012). Antioxidant activity, total phenolic content and flavonoid concentrations of different plant parts of *Teucrium polium* L. subsp. *polium*. *Acta Societatis Botanicorum Poloniae*, *81*(2), 117–122. <http://doi.org/10.5586/asbp.2012.010>
- Stewart, G. S., Jassim, S. A., Denyer, S. P., Newby, P., Linley, K., & Dhir, V. K. (1998). The specific and sensitive detection of bacterial pathogens within 4 h using bacteriophage amplification. *Journal of Applied Microbiology*, *84*(5), 777–783. <http://doi.org/10.1046/j.1365-2672.1998.00408.x>
- Suleiman, M. M., McGaw, L. J., Naidoo, V., & Eloff, J. N. (2010). Detection of antimicrobial compounds by bioautography of different extracts of leaves of selected south african tree species. *African Journal of Traditional, Complementary and Alternative Medicines*, *7*(1), 64–78. <http://doi.org/10.4314/ajtcam.v7i1.57269>
- swamy, K. M., Pokharen, N., & Dahal, S. (2011). Phytochemical and antimicrobial studies of leaf extract of *Euphorbia neriifolia*. *Journal of Medicinal Plants Research*, *5*(24), 5785–5788.
- Tafesh, A., Najami, N., Jadoun, J., Halahlh, F., Riepl, H., & Azaizeh, H. (2011). Synergistic antibacterial effects of polyphenolic compounds from olive mill wastewater. *Evidence-Based Complementary and Alternative Medicine : eCAM*, *2011*(Mic), 431021. <http://doi.org/10.1155/2011/431021>
- Taweechaisupapong, S., Singhara, S., Lertsatitthanakorn, P., & Khunkitti, W. (2010). Antimicrobial effects of *Boesenbergia pandurata* and *Piper sarmentosum* leaf extracts on planktonic cells and biofilm of oral pathogens. *Pakistan Journal of Pharmaceutical Sciences*, *23*(2), 224–231.
- Tayel, A. a., & El-Tras, W. F. (2010). Anticandidal activity of pomegranate peel extract aerosol as an applicable sanitizing method. *Mycoses*, *53*, 117–122. <http://doi.org/10.1111/j.1439-0507.2008.01681.x>
- Tripathi, J., Chatterjee, S., Gamre, S., Chattopadhyay, S., Variyar, P. S., & Sharma, A. (2014). Analysis of free and bound aroma compounds of pomegranate (*Punica granatum* L.). *LWT - Food Science and Technology*, *59*(1), 461–466. <http://doi.org/10.1016/j.lwt.2014.05.055>
- Tsuchiya, H., & Iinuma, M. (2000). Reduction of membrane fluidity by antibacterial sophoraflavanone G isolated from *Sophora exigua*. *Phytomedicine*, *7*(2), 161–165. [http://doi.org/10.1016/S0944-7113\(00\)80089-6](http://doi.org/10.1016/S0944-7113(00)80089-6)
- Unica, O. P., & Is, G. L. (2003). Antioxidant and Antibacterial Activities of *Punica granatum* Peel Extracts. *Food Microbiology and Safety*, *68*(4), 1473–1477.
- Valgas, C., Souza, S. M. De, Smânia, E. F. a., & Artur, S. J. (2007). Screening Methods to Determine Antibacterial Activity of Natural Products. *Brazilian*

Journal of Microbiology, 38, 369–380. <http://doi.org/10.1590/S1517-83822007000200034>

- Vasconcelos, L. C. D. S., Sampaio, F. C., Sampaio, M. C. C., Pereira, M. D. S. V., Higino, J. S., & Peixoto, M. H. P. (2006). Minimum inhibitory concentration of adherence of *Punica granatum* L.(pomegranate) gel against *S. mutans*, *S. mitis* and *C. albicans*. *Brazilian Dental Journal*, 17, 223–227. <http://doi.org/10.1590/S0103-64402006000300009>
- Vasconcelos, L. C. D. S., Sampaio, M. C. C., Sampaio, F. C., & Higino, J. S. (2003). Use of *Punica granatum* as an antifungal agent against candidosis associated with denture stomatitis Verwendung von *Punica granatum* als Antimykotikum gegen Candidose in Verbindung mit Zahnprothesen-Stomatitis. *Mycoses*, 46(May 2002), 192–196. <http://doi.org/10.1046/j.1439-0507.2003.00884.x>
- Vasse, J., Frey, P., & Trigalet, A. (1995). Microscopic studies of intercellular infection and protoxylem invasion of tomato roots by *Pseudomonas solanacearum*. *Molecular Plant-Microbe Interactions: MPMI (USA)*. Retrieved from <http://agris.fao.org/agris-search/search.do?recordID=US9549032>
- Vasudha Pai et al, 2011. (2011). Evaluation of the antimicrobial activity of *Punica granatum* peel against the enteric pathogens : An invitro study, 1(2), 57–62.
- Vermerris, W., & Nicholson, R. L. (2008). Phenolic Compound Biochemistry. *Phenolic Compound Biochemistry*, 151–190. <http://doi.org/10.1007/978-1-4020-5164-7>
- Vidal, A., Fallarero, A., Peña, B. R., Medina, M. E., Gra, B., Rivera, F., ... Vuorela, P. M. (2003). Studies on the toxicity of *Punica granatum* L. (Punicaceae) whole fruit extracts. *Journal of Ethnopharmacology*, 89(2-3), 295–300. <http://doi.org/10.1016/j.jep.2003.09.001>
- Waldee, E. L., & others. (1945). Comparative studies of some peritrichous phytopathogenic bacteria. *Iowa State College Journal of Science*, 19(4), 435–484.
- Waller, G. R. (1996). Saponins: Chemistry and Pharmacology of Natural Products. *Journal of the American Chemical Society*, 118(35), 8509–8509. <http://doi.org/10.1021/ja9553056>
- Whittaker, R. H., & Feeny, P. P. (1971). Allelochemicals: Chemical Interactions between Species. *Science*, 171(3973), 757–770. <http://doi.org/10.1126/science.171.3973.757>
- Wink M.1999. (n.d.). Biochemistry of plant secondary metabolism. Retrieved October 20, 2015, from <http://www.slideshare.net/Hariezt24/biochemistry-of-plant-secondary-metabolism>
- Wong, S.-L., Chang, H.-S., Wang, G.-J., Chiang, M. Y., Huang, H.-Y., Chen, C.-H., ... Chen, I.-S. (2011). Secondary Metabolites from the Roots of *Neolitsea daibuensis* and Their Anti-inflammatory Activity. *Journal of Natural Products*, 74(12), 2489–2496. <http://doi.org/10.1021/np100874f>

- Wright, P. J. (1998). A soft rot of calla (*Zantedeschia spp.*) caused by *Erwinia carotovora subspecies carotovora*. *New Zealand Journal of Crop and Horticultural Science*, 26(4), 331–334. <http://doi.org/10.1080/01140671.1998.9514072>
- Wuyts, N., De Waele, D., & Swennen, R. (2006). Extraction and partial characterization of polyphenol oxidase from banana (*Musa acuminata* Grande naine) roots. *Plant Physiology and Biochemistry: PPB / Societe Francaise de Physiologie Vegetale*, 44(5-6), 308–314. <http://doi.org/10.1016/j.plaphy.2006.06.005>
- Yang, R. Y., Lin, S., & Kuo, G. (2008). Content and distribution of flavonoids among 91 edible plant species. *Asia Pacific Journal of Clinical Nutrition*, 17(SUPPL. 1), 275–279.
- Zahin, M., Aqil, F., & Ahmad, I. (2010). Broad spectrum antimutagenic activity of antioxidant active fraction of *Punica granatum* L. peel extracts. *Mutation Research - Genetic Toxicology and Environmental Mutagenesis*, 703(2), 99–107. <http://doi.org/10.1016/j.mrgentox.2010.08.001>
- Zaidan, M. R. S., Noor Rain, a, Badrul, a R., Adlin, a, Norazah, a, & Zakiah, I. (2005). In vitro screening of five local medicinal plants for antibacterial activity using disc diffusion method. *Tropical Biomedicine*, 22(2), 165–70. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/16883283>
- Zhang, J., Zhan, B., Yao, X., Gao, Y., & Shong, J. (1995). [Antiviral activity of tannin from the pericarp of *Punica granatum* L. against genital Herpes virus in vitro]. *Zhongguo Zhong Yao Za Zhi*, 20(9), 556–8, 576, inside backcover. Retrieved from http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=PubMed&dopt=Citation&list_uids=8679095
- Zheng, C. J., Yoo, J.-S., Lee, T.-G., Cho, H.-Y., Kim, Y.-H., & Kim, W.-G. (2005). Fatty acid synthesis is a target for antibacterial activity of unsaturated fatty acids. *FEBS Letters*, 579(23), 5157–5162. <http://doi.org/10.1016/j.febslet.2005.08.028>
- Zito, P., Sajeve, M., Bruno, M., Maggio, A., Rosselli, S., Formisano, C., & Senatore, F. (2010). Essential oil composition of stems and fruits of *Caralluma europaea* N.E.Br. (apocynaceae). *Molecules*, 15(2), 627–638. <http://doi.org/10.3390/molecules15020627>