



UNIVERSITI PUTRA MALAYSIA

***OPTIMIZATION OF SEED AND SEEDLING PRODUCTION IN HEMPEDU
BUMI (*Andrographis paniculata* (Burm. F.) Wall. Ex Nees)***

WONG KIAN JOO

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By

WONG KIAN JOO

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfillment of the Requirements for the Degree of Master of Science**

March 2016

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

**OPTIMIZATION OF SEED AND SEEDLING PRODUCTION IN HEMPEDU
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March 2016

Chairman : Associate Professor Thohirah Lee Abdullah, PhD
Faculty : Agriculture

The propagation of *Andrographis paniculata* is generally through seed but the germination is poor due to seed dormancy. Therefore, seven seed dormancy breaking treatments namely water soaking for 24 hours, sand paper scarification, hot water soaking at 40°C for 10 minutes, hot water soaking at 50°C for 10 minutes, hot water soaking at 60°C for 5 minutes and combination of sand paper scarification, hot water soaking (60°C for 5 minutes) and a control were investigated to determine the best seed dormancy breaking treatments. The percentage of seed germination scarified using sand paper was 91%. Seeds took the shortest time of 3.5 days to reach 50 % germination after scarified and followed with hot water soaking at 60°C for 5 minutes.

Maturity at harvest and storage are important that will affect the seed quality. Currently no study has been carried out to study the effect of maturity and storage on seed quality dan germination. The effects of two maturity stages (grey purple capsule and grey brown capsule) and four storage periods (0, 3, 6 and 9 months) with and without sand paper scarification were investigated to determine the seed germinability. Freshly harvested seeds from grey brown capsule had lower 1000 seed weight, lower seed moisture and higher seed dry matter of 1 gram seed compared to seeds from grey purple capsules. However, maturity stage and storage period did not affect 1000 seed weight, seed moisture and seed dry matter of 1 gram seed after the capsules were slow dried in desiccator. Germination percentage was recorded higher for seeds from grey brown capsule rather than grey purple capsule. Germination improved after 6 months of storage without scarification. None of the untreated seeds from different maturity stages and storage periods had more than 50 % germination. Seeds from grey brown capsule reached 50 % germination faster than seeds from grey purple capsule. Time to 50 % germination was significantly reduced after 6 months of storage.

The effects of three seedling tray cell volume (43 cm³, 30 cm³ and 25 cm³) and seven growing media (mixed soil, cocopeat, 4 cocopeat : 1 rice husk biochar, 3 cocopeat : 2 rice husk biochar, 4 cocopeat : 1 EFB compost, 3 cocopeat : 2 EFB compost and 4 cocopeat : 3 rice husk biochar : 3 EFB compost) were investigated to determine a suitable growing media and seedling tray for the production of organically grown

seedlings *A. paniculata*. Seedling height, fresh weight, total leaf area and dry weight were increased when the cell volume increased from 43 cm³ to 30 cm³ then reduced at cell with 25 cm³. Root shoot ratio did not differ significantly between seedling tray cell volumes. Root length and root volume increased significantly when the number of cell in tray increased. Seedlings grown in media cocopeat : biochar : EFB compost in ratio 4:3:3 (CBE) had the highest seedling height, fresh weight, total leaf area, dry weight, root length and root volume. There were significant interaction effects between seedling tray cell volume and growing media on seedling height, fresh weight, total leaf area, root length and root volume. Quality seedlings can be obtained in cell with 30 cm³ with media CBE.

Light duration is one of the environmental factors important that can influence flowering, seed fertility and seed production. However, no photoperiodic treatments has been studied for this plant. Therefore, the effects of four short day treatments (12 hours, 10 hours, 8 hours and 6 hours) was investigated to determine the suitable short day treatment for seed production. Plant height increased significantly when the day length provided becomes shorter. Short day treatments did not affect secondary branches and dry weight of plants. The heaviest plants of 172.70 g in fresh weight were grown in 6 hours day length. Prolonged days to flowering and days to mature capsule formation were observed when day length was reduced from 12 hours to 6 hours. *A. paniculata* grown in 12 hours (control) recorded the highest number of flower of 459.50, number of capsule of 359.67 and harvest index of 9.78 % respectively. The highest 1000 seed weight of 0.146 g was recorded from the plants grown in 12 hours. However, short day treatments did not influence the seed moisture content, seed dry matter of 1 gram seed and seed germinability.

It is recommended that the seedling and seed production of *A. paniculata* can optimized by planting sand paper scarified seeds from grey brown capsule on combination media of cocopeat:biochar:EFB compost in 30 cm³ seedling tray. *A. paniculata* is not a short day plants.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

PENAMBAHBAIKKAN BIJI BENIH DAN PENGELUARAN ANAK BENIH
(*Andrographis paniculata* (Burm. F.) Wall. Ex Nees)

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Pembiakan *Andrographis paniculata* biasanya melalui biji benih tetapi percambahan tidak memuaskan disebabkan oleh kedormanan biji benih. Oleh itu, tujuh kaedah rawatan pemecahan kedormanan biji benih iaitu rendaman air selama 24 jam, pelepasan dengan kertas pasir, rendaman air panas bersuhu 40°C selama 10 minit, rendaman air panas bersuhu 50°C selama 10 minit, rendaman air panas bersuhu 60°C selama 5 minit dan kombinasi pelepasan kertas pasir dan rendaman air panas bersuhu 60°C selama 5 minit dan satu kawalan tanpa rawatan telah dikaji. Peratusan percambahan biji benih hempedu bumi selepas dilelaskan dengan kertas pasir adalah 91 %. Biji benih menggunakan masa yang paling singkat iaitu 3.5 hari untuk mencapai 50 % percambahan selepas menggunakan kaedah pelepasan kertas pasir lalu direndam dalam air panas bersuhu 60°C selama 5 minit.

Kematang semasa penuaian dan penyimpanan adalah penting dan akan menjejaskan kualiti biji benih. Tiada kajian dilakukan untuk kesan kematangan dan penyimpanan ke atas kualiti percambahan biji benih. Kesan dua peringkat kematangan (kapsul ungu kelabu dan kapsul perang kelabu) dan empat tempoh penyimpanan (0, 3, 6 dan 9 bulan) dengan dan tanpa pelepasan kertas pasir telah dikaji untuk menentukan kemampuan percambahan biji benih. Biji benih dari kapsul perang kelabu yang baru dituai telah merekod berat 1000 biji benih, kelembapan biji benih yang lebih rendah dan isi kering biji benih untuk 1 gram biji benih yang lebih tinggi berbanding dengan biji benih dari kapsul perang ungu. Manakala, peringkat kematangan dan tempoh penyimpanan tidak menjejaskan berat 1000 biji benih, kelembapan biji benih dan isi kering biji benih untuk 1 gram biji benih selepas dikeringkan dalam dessicator. Peratus percambahan adalah tinggi bagi biji benih dari kapsul perang kelabu berbanding dengan biji benih dari kapsul perang ungu. Percambahan bertambah selepas penyimpanan 6 bulan. Biji benih yang tidak dilelaskan daripada peringkat kematangan dan tempoh penyimpanan yang berbeza tidak bercambah lebih daripada 50 % percambahan. Masa yang lebih singkat untuk 50 % percambahan telah diperhatikan dalam biji benih kapsul perang kelabu selepas pelepasan. Masa untuk 50 % percambahan telah dikurangkan dengan ketara selepas 6 bulan penyimpanan.

Untuk tujuan penanaman organik bagi menghasilkan ubat-ubatan, kesan tiga isipadu sel dulang (berisipadu 43 cm³, 30 cm³ dan 25 cm³) dan tujuh media pertumbuhan (tanah campuran, habuk sabut kelapa, 4 habuk sabut kelapa : 1 biochar sekam padi, 3 habuk sabut kelapa : 2 biochar sekam padi, 4 habuk sabut kelapa : 1 kompos tandan kosong kelapa sawit, 3 habuk sabut kelapa : 2 kompos tandan kosong kelapa sawit dan 4 habuk sabut kelapa : 3 biochar sekam padi : 3 kompos tandan kosong kelapa sawit) telah dikaji untuk menentukan media pertumbuhan dan dulang anak benih yang sesuai bagi pertumbuhan anak benih hempedu bumi. Ketinggian anak benih, berat basah, jumlah luas permukaan daun dan berat kering meningkat dari dulang bersel 43 cm³ ke dulang bersel 30 cm³ tetapi menurun pada dulang bersel 25 cm³. Nisbah akar kepada pucuk tidak mempunyai perbezaan ketara antara saiz dulang. Pajang akar dan isi padu akar menunjukkan peningkatan ketara apabila nombor sel dalam dulang bertambah. Biji benih ditanam dalam media habuk sabut kelapa : biochar : kompos tandan kosong kelapa sawit dalam nisbah 4:3:3 (CBE) mempunyai ketinggian anak benih, berat basah, jumlah luas permukaan daun, berat kering, panjang akar dan isi padu akar yang paling tinggi. Terdapat kesan interaksi yang ketara antara saiz dulang dan media pertumbuhan terhadap ketinggian anak benih, berat basah, jumlah luas permukaan daun, panjang akar dan isi padu akar. Anak benih yang ditanam dalam dulang bersel 30 cm³ dengan media CBE menghasilkan anak benih yang lebih berkualiti.

Tempoh cahaya adalah salah satu factor persekitaran yang penting untuk berbunga, kesuburan biji benih dan pengeluaran benih. Walau bagaimanapun, tiada kajian dilakukan ke atas tumbuhan ini. Empat kesan rawatan hari siang singkat (12 jam, 10 jam, 8 jam dan 6 jam) telah dikajikan untuk menentukan rawatan hari siang singkat yang sesuai untuk pengeluaran biji benih. Ketinggian tumbuhan meningkat dengan ketara apabila kepanjangan hari siang yang diberikan menjadi pendek. Rawatan hari siang singkat tidak menjejaskan dahan sekunder dan berat kering tumbuhan. Berat basah paling tinggi direkodkan daripada hempedu bumi yang telah ditanam dalam 6 jam panjang hari siang, iaitu 172.70 g. Pemanjangan hari untuk pembungaan dan hari untuk pembentukan kapsul yang matang didapati apabila panjang hari siang dipendekan daripada 12 jam kepada 6 jam. Hempedu bumi ditanam dalam 12 jam (kawalan) mencatatkan bilangan bunga, bilangan kapsul dan indeks menuai yang paling tinggi dengan masing-masing 459.50, 359.67 dan 9.78 %. Berat 1000 biji benih yang paling tinggi, 0.146 g telah direkodkan daripada tumbuhan yang ditanam dalam 12 jam. Rawatan hari siang singkat tidak menjejaskan kandungan kelembapan, isi keing biji benih daripada 1 gram biji benih dan kemampuan percambahan biji benih.

Ia adalah disyorkan bahawa pengeluaran anak benih dan benih *A. paniculata* boleh dioptimumkan dengan menanam benih dari kapsul perang kerabu yang telah dilelaskan dengan kertas pasir dalam dulang bersaiz 30 cm³ dengan gabungan media habuk sabut kelapa:biochar:kompos tandan kosong kelapa sawit. *A. paniculata* bukan tanaman hari pendek.

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LIST OF ABBREVIATIONS

%	Percentage
°C	Degree Celsius
ABA	Absciscic Acid
ANOVA	Analysis of Variance
ATP	Adenosine Triphosphate
C	Carbon
Ca	Calcium
CEC	Cation Exchange Capacity
cm	Centimeter
cm ²	Square Centimeter
cm ³	Cubic Centimeter
CRD	Completely Randomized Design
DN	Day Neutral
EC	Electrical Conductivity
EFB	Empty Fruit Bunches
<i>et al.</i>	And friends
g	Gram
g ⁻¹	Per Gram
GA ₃	Gibberellic Acid
g cm ⁻³	Gram per Cubic Centimeter
GP	Germination Percentage
h	hour
H ₂ O	Water
IAA	Indole-3-acetic Acid
IBA	Indole-3-butyric Acid
ISTA	International Seed Testing Association
K	Potassium
KCl	Potassium Chloride
kg	Kilogram
KNO ₃	Potassium Nitrate
LD	Long Day
LSD	Least Significant Difference
N	Nitrogen
NADPH	Nicotinamide Adenine Dinucleotide Phosphate
M	Molar
meq	Milliequivalents
Mg	Magnesium
MGT	Mean Germination Time

min	Minute
ml	Milliliter
mm	Millimeter
mM	Millimolar
mS/cm	Millisiemens per Centimeter
n.d.	No date
NaHClO ₃	Sodium Hypochlorite
NI	Night Interruption
No.	Number
P	Phosphorus
pH	Measurement of Acidity/Alkalinity
ppm	Part Per Million
RCBD	Randomized Complete Block Design
rpm	Revolutions per Minute
SAS	Statistical Analysis System
SD	Short Day
T ₅₀	Time to 50% Germination
US\$	United States dollar

CHAPTER 1

INTRODUCTION

Malaysia has a suitable climate for the growth of various herbal plants. There are more than 2000 plant species with medicinal value found in Malaysia with commercialization potential. Malaysia Agricultural Research and Development Institute (MARDI) has reported that an annual gross profit from herbal related products was more than RM 5.4 billion (Ahmad and Othman, 2013). However, Malaysian herbal and medicinal plants industry is still lagging behind in terms of Research and Development (R&D) when compared to China and also other ASEAN countries. Furthermore, the knowledge on plant part uses, preparation methods and other traditional knowledge is important for the development of herbal industry in Malaysia (Ahmad and Othman, 2013).

In Malaysia, some popular medicinal plants like male aphrodisiac *Eurycoma longifolia* Jack. (tongkat ali), *Labisia pumila* (kacip fatima), *Orthosiphon stamineus* (misai kucing), *Andrographis paniculata* (hempedu bumi), *Phyllanthus niruri* (dukung anak), *Centella asiatica* (pegaga) and *Momordica charantia* (peria) are under extensive research (Jamal, 2006). Tongkat ali, kacip fatima, misai kucing, hempedu bumi and dukung anak were listed in the agriculture sector under National Key Economic Areas (NKEAs) as main herbal plants to focus on (Ganesan, 2011). The Malaysian Economic Transformation Programmed through NKEA Agriculture sector has identified *A. paniculata* as one of the high value herbal crops that need to be commercially exploited for a new source of economic growth in the herbal industry. There is extensive knowledge of herbs for treatment of various ailments and the recognition of biomedical values in the global scenario (Noorhanin *et al.*, 2013).

A. paniculata (Family: Acanthaceae) commonly known as hempedu bumi (bile of earth) in Malaysia or 'King of Bitters' due to its extremely bitter taste in all parts of the plant. The bitter taste of this plant comes from chemical substances called diterpene lactones which have medicinal benefits. This medicinal plant grows well in tropical Asian countries (Parashar *et al.*, 2011; Valdiani *et al.* 2012). This plant is widely used for treating fever, liver diseases, diabetes, snake bite, bronchitis and a variety of ailments (Parashar *et al.*, 2011). A kilogram of good quality *A. paniculata* dried leaves can be priced for US\$5, while one kilogram of purified active compounds, andrographolide and its derivatives, are sold for US\$100,000 from specialist chemical suppliers (Chulabhorn, 2005). Shamar *et al.* (2008) reported that the estimated consumption of this plant to be 250 tons annually. Since this plant is propagated through seed, high seed quality is needed to ensure crop stand and yield.

The propagation of *A. paniculata* is generally through seeds. However, the main issue of *A. paniculata* is due to its low germination. Generally, this indicates the presence of seed dormancy. It is a survival strategy for plants to disperse the seeds without germination under unfavorable condition. Seed dormancy is an internal condition that

prevents seeds from germination under favorable condition (appropriate temperature regime, adequate moisture, normal atmosphere and light requirement in some cases) (Schmidt, 2007; Batlla & Benech-Arnold, 2010). Some plant species have developed regulatory mechanisms to delay germination under the conditions where seedlings can hardly survive. Depending on species of plants, seed dormancy can be imposed by the envelopes (seed coat, endosperm, etc.), embryo, or both factors together (Debeaujon *et al.*, 2000). Talei *et al.*, (2012) mentioned that the *A. paniculata* seeds seriously stagnated during dormant stage after imbibition. Therefore, there is a need to study the seed dormancy of *A. paniculata* to improve the germination.

Seed germination and emergence depends on the seed properties such as seed quality. High quality seed lots can improve crop yield. One of the main factor to determine the seed quality is the seed maturation. Generally, seed quality parameters are associated with the harvesting stage of the seed crop. High quality seed with maximum dry weight, high viability and vigor can be obtained if the seed crop harvested at physiological maturity. For storage purposes, fully matured seeds are desirable to be collected, rather than the immature seeds (Eskandari, 2012; Kamotho *et al.*, 2014). Therefore, it is important to know the relationship between seed harvesting stage and seed quality. However, there is little information of seed maturity on relation to seed quality for *A. paniculata*.

Seed storage as *ex situ* germplasm is important for long term conservation of plant genetic resources. Depending on the storage method and duration, drying and long term storage may reduce the seed germination or eventually kill the seeds (Kamotho *et al.*, 2014). On the other hand, the biochemical processes in the seeds after dispersed from the mother plant and experienced dry conditions also have a pronounced effect on seed dormancy and germination depend on seed crop species. During dry storage, the seed dormancy is gradually lost is due to the process called after-ripening. Hence, it is important to study the storage effect on *A. paniculata* seeds since the seeds are normally stored after harvest the matured plant by the farmer (Donohue, 2015).

For large scale plantation, direct sowing in the field is commonly practice. But the production of seedlings in containers is also gaining popularity recently compared to field production due to easy marketing, transportation and rapid product rotation (Aklibasinda *et al.*, 2011). Choosing the most suitable container size and growing media is important for plant growth as it will influence the transplant performance and production (Ingram, *et al.*, 1993). With the changes in container size, plants may undergo morphological and physiological changes in the response on root growth. A proper growing media should provide biological, chemical and physical requirements for plants.

Besides the container size and growing media, some other factors like photoperiod also important which will physiologically influence the plant growth. However, there are few studies on the factors affecting production of the seeds of *A. paniculata*. Seed production can be affected by different photoperiods. The plant is widely found and well-adapted to tropical and subtropical Asia, south-east Asia and India, suggesting it is a short-day plant (Chang, 1986). To produce seeds successfully in Malaysia, there is a

need to determine the photoperiod effects on the seed production of *A. paniculata*. The objectives of this study were:

1. To determine the best seed dormancy breaking treatments for germinating *A. paniculata*.
2. To determine the seed maturity stages and storage periods of *A. paniculata* for highest seed quality production.
3. To determine the germination tray cell volume and growing media types for the best seedling growth of *A. paniculata*.
4. To determine the best photoperiod for the highest growth and seed production of *A. paniculata*.



REFERENCES

- Abad, M., Noguera, P., Puchades, R., Maquieira, A., Noguera, V. 2002. Physico-chemical and chemical properties of some coconut coir dusts for use as a peat substitute for containerised ornamental plants. *Bioresource Technology* 82: 241–245.
- Abdalla, J.A. 2005. *Genetic Variation and Anticancer activity of Andrographis paniculata Germplasm from Malaysia*, Master Thesis, Universiti Putra Malaysia.
- Abdullah, N., Sulaiman, F. 2013. The oil palm wastes in Malaysia. In *Biomass Now - Sustainable Growth and Use*, ed. M.D. Matovic. Retrieved at 17 April 2015. Available from: <http://www.intechopen.com/books/biomass-now-sustainable-growth-and-use/the-oil-palm-wastes-in-malaysia>
- Ahmad, I., Ahmad, T., Gulfam, A., Saleem, M. 2012. Growth and flowering of gerbera as influenced by various horticultural substrates. *Pakistan Journal of Botany*. 44: 291–299.
- Ahmad, S., Othman, N. 2013. Strategic planning, issues, prospects and the future of the Malaysian herbal industry. *International Journal of Academic Research in Accounting, Finance and Management Sciences* 3: 91–102.
- Akbar, S. 2011. *Andrographis paniculata*: A review of pharmacological activities and clinical effects. *Alternative Medicine Review* 16: 66–77.
- Aklibasinda, M., Tunc, T., Bulut, Y., Sahin, U. 2011. Effects of different growth media on scotch pine (*Pinus sylvestris*) production. *The Journal of Animal and Plant Sciences* 21(3): 535-541.
- Al-Menaie, H.S., Al-Ragam, O., Al-Dosery, N., Zalzaleh, M., Mathew, M., Suresh, N. 2012. Effect of pot size on plant growth and multiplication of water lilies (*Nymphaea* sp). *American-Eurasian Journal of Agricultural & Environmental Sciences* 12(2): 148–153.
- Animesh, D., Benoy, G.K., Aninda, M., Priyanka, D.K., Sandip. H. 2012. An overview on *Andrographis paniculata* (Burm.F.) Nees. *International Journal of Research in Ayurveda and Pharmacy* 3(6): 752-760.
- Aphalo, P., Rikala, R. 2003. Field performance of silver-birch planting-stock grown at different spacing and in containers of different volume. *New Forests* 25: 93–108.
- Arenas, M., Vavrina, C.S., Cornell, J.A., Hanlon, E. A., Hochmuth, G.J. 2002. Coir as an alternative to peat in media for tomato plant production. *HortScience* 37: 309-312.
- Arp, W.J. 1991 Effects of source–sink relations on photosynthesis acclimation to elevated CO₂. *Plant, Cell & Environment* 14: 869–875.

- Asiah, A., Mohd. Razi, I., Mohd. Khanif, Y., Marziah, M., Shaharuddin, M. 2004. Physical and chemical properties of coconut coir dust and oil palm empty fruit bunch and the growth of hybrid heat tolerant cauliflower plant. *Pertanika Journal of Tropical Agricultural Science* 27: 121–133.
- Balasubramanian, J., Narayanan, N. 2012. A review on collection, cultivation, and medical value of herba *Andrographidis*. *Discocery life* 2: 6.
- Bar-Tal, A., Feigin, A., Sheinfeld, S., Rosenberg, R., Sternbaum, B., Rylski, I., and Pressman, E. 1995. Root restriction and N-NO₃ solution concentration effects on nutrient uptake, transpiration and dry matter production of tomato. *Scientia Horticulturae* 63: 195-208.
- Baskin, C.C., Baskin, J.M. 1998. Seeds - Ecology, biogeography, and evolution of dormancy and germination. San Diego, CA: *Academic Press*. 666 p.
- Baskin, C.C., Baskin, J.M. 2008. Advances in understanding seed dormancy at the whole-seed level: An ecological, biogeographical and phylogenetic perspective. *Acta Botanica Yunnainca* 30: 279–294.
- Baskin, J.M., Baskin, C.C. 2004. A classification system for seed dormancy. *Seed Science Research* 14: 1-16.
- Batlla, D., Benech-Arnold, R.L. 2010. Predicting changes in dormancy level in natural seed soil banks. *Plant Molecular Biology* 73: 3–13.
- Berjak, P., Pammenter, N.W. 2002. Orthodox and recalcitrant seeds. *Tropical tree seed manual. Agricultural handbook* 721: 137-147.
- Bewley, J. 1997. Seed germination and dormancy. *The Plant Cell* 9: 1055–1066.
- Bewley, J.D., Bradford, K.J., Hilhorst, H.W.M., Nonogaki, H. 2013. Germination. In *Seeds: Physiology of Development, Germination and Dormancy*. Volume 3. pp 133-181. New York: Springer.
- Bewley, J.D., Black, M. 1994. Seeds - Physiology of development and germination. New York: *Plenum Press* 445 p.
- Bhattacharya, J., Khuspe, S. 2001. In vitro and in vivo germination of papaya (*Carica papaya* L.) seeds. *Scienti Horticulturae* 91: 39–49.
- Bhattacharya, S., Puri, S., Jamwal, A., Sharma, S. 2012. Studies on seed germination and seedling growth in Kalmegh (*Andrographis paniculata* Wall. Ex Nees) under abiotic stress conditions. *International Journal of Science, Environment and Technology* 1(3): 197–204.

- Bilderback, T.E., Fonteno, W.C. 1991. Effects of container geometry and media physical properties on air and water volumes in containers. *Journal of Environmental Horticulture* 5: 180-182.
- Carrera, E., Holman, T., Medhurst, A., Dietrich, D., Footitt, S., Theodoulou, F.L., Holdsworth. 2008. Seed after-ripening is a discrete developmental pathway associated with specific gene networks in Arabidopsis. *The Plant Journal* 53: 214-224
- Ceunen, S., Geuns, J.M.C. 2012. Influence of photoperiodism on the spatio-temporal accumulation of steviol glycosides in *Stevia rebaudiana* (Bertoni). *Plant Science* 198: 72-82.
- Chan, K.Y., Van Zwieten, L., Meszaros, I., Downie, A., Joseph, S. 2008: Using poultry litter biochars as soil amendments. *Soil Research* 46: 437-444.
- Chan, K.Y., Van Zwieten, L., Meszaros, I., Downie, D., Joseph, S. 2007. Agronomic values of green waste biochars as a soil amendments. *Soil Research*. 45, 629-634.
- Chang, H.M. 1986. But PPH eds. Pharmacology and Applications of Chinese Materia Medica. Singapore, World Scientific, Vol 1 : 918-928.
- Chauhan, J.S., Y.K. Tomar., N.I. Singh, S. Ali., A.Badoni., Debarati., and A. Rana. 2009. Assessment of complete substratum for *Andrographis paniculata* standard seed germination testing. *Journal of American Science* 5: 70-75.
- Chen, C.L., Tsai, Y.J., Sung, J.M. 2010. Photoperiod effects on flowering and seed setting of *Hypericum perforatum*. *Experimental Agriculture* 46(3): 393-400.
- Chin, H.F., Krishnapillay, B., Stanwood, P.C. 1989. Seed moisture: recalcitrant vs. orthodox seeds. In *Seed moisture*, ed. Stanwood, P.C., McDonald, M.B, pp. 15-22.
- Christopher, T.B.S., Jamal, T. 2006. Soil physics analysis volume 1. Selangor: Universiti Putra Malaysia Press.
- Chulabhorn, H.R.H.P. 2005. Promoting herbal drugs: Thailand. In *Examples of the Development of Pharmaceutical Products from Medicinal Plants*. Volume 10. pp 45-56. Cornell University: UNDP.
- Chung, L.C., Chung, J.T., Jih, M.S. 2010. Photoperiod effects on flowering and seed setting of *Hypericum perforatum*. *Experimental Agriculture* 46(3): 393-400.
- Contreras, S., Bennett, M.A., Metzger, J.D., Tay, D. 2008. Maternal light environment during seed development affects lettuce seed weight, germinability, and storability. *HortScience* 43(3): 845-852.
- Csizinszky, A.A., Schuster, D.J. 1993. Impact of insecticide schedule, N and K rates, and transplant container size on cabbage yield. *HortScience* 28: 299-301.

- Čupić, T., Popović, S., Grljušić, S., Tucak, M., Andrić, L. 2005. Effect of storage time on alfalfa seed quality. *Journal Central European of Agriculture* 6(1): 65–68.
- Debeaujon, I., Léon-Kloosterziel, K.M., Koornneef, M. 2000. Influence of the testa on seed dormancy, germination, and longevity in arabidopsis. *Plant Physiology* 122: 403–414.
- Demir, I., Ashirov, A., Mavi, K. 2008. Effect of seed production environment and time of harvest on tomato (*Lycopersicon esculentum*) seedling growth. *Research Journal of Seed Science* 1(1), 1–10.
- Dezfuli, P.M., Sharif-zadeh, F., Janmohammadi, M. 2008. Influence of priming techniques on seed germination behavior of maize inbred lines (*Zea mays* L.). *ARP Journal of Agricultural and Biological Science* 3(3): 22-25.
- Di Benedetto, a.H., Klasman, R., 2004. The effect of plug cell volume on the post-transplant growth for *Impatiens walleriana* pot plant. *European Journal of Horticultural Science* 69(2): 82–86.
- Dole, J.M. and Wilkins, H.F. 2005. *Floriculture: Principles and Species*, 2nd edition. Pearson Prentice Hall, Upper Saddle River, New Jersey
- Dole, J.M. and Wilkins, H.T. 1999. *Floriculture principles and species*. Prentice Hall, Upper Saddle River, N.J.
- Donohue, K. 2009. Completing the cycle: Maternal effects as the missing link in plant life histories. *Philosophical transactions of the Royal Society of London. Series B, Biological Sciences* 364: 1059–1074.
- Dubik, S.P., Krizek, D.T., and Stimart, D.P. 1990. Influence of root zone restriction on mineral element concentration, water potential, chlorophyll concentration, and partitioning of assimilate in spreading euonymus (*E. Kiautschovica* Loes. 'Sieboldiana'). *Journal of Plant Nutrition* 13: 677-699.
- Dufault, R.J. and Waters, L. Jr. 1985. Container size influences broccoli and cauliflower transplant growth but not yield. *HortScience* 20: 682-684.
- Ekpong, B., Sukprakarn, S., 2008. Seed physiological maturity in dill (*Anethum graveolens* L.). *Kasetsart Journal: Natural Science* 6: 1–6.
- Ellis, R.H., Roberts, E.H. 1981. The quantification of ageing and survival in orthodox seeds. *Seed Science and Technology* 9: 373–409.
- Ellis, R.H., Hong, T.D., Roberts, E.H. 1990. An intermediate category of seed storage behaviour. *Journal of Experimental Botany* 41: 1167-1174.
- Ennen, R.D. 2011. *Earlier harvest and drying of soybean seed within intact pods maintains seed quality*, Master Degree Thesis, Iowa State University.

- Evans, M.R., Konduru, S., Stamps, R.H. 1996. Source variation in physical and chemical properties of coconut coir dust. *HortScience* 31: 965–967.
- Fischer, D., Glaser, B. 2012. Synergisms between Compost and Biochar for Sustainable Soil Amelioration. In *Management of Organic Waste*, ed. Kumar, S. & Bharti, A., pp 167-198. Croatia: InTech.
- Foley, M.E. 2001. Seed dormancy: an update on terminology, physiological genetics, and quantitative trait loci regulating germinability. *Weed Science* 49: 305-317.
- Fornes, F., Belda, R.M., Abad, M., Noguera, P., Puchades, R., Maquieira, a., Noguera, V. 2003. The microstructure of coconut coir dusts for use as alternatives to peat in soilless growing media. *Australian Journal of Experimental Agriculture* 43: 1171–1179.
- Franklin, K.A., Whitelam, G.C. 2004. Light signals, phytochromes and cross-talk with other environmental cues. *Journal of Experimental Botany* 55: 271-276.
- Frąszczak, B. 2012. The effect of changes in diurnal temperature and photoperiod on growth and yielding of garden dill grown in pots. *Acta Scientiarum Polonorum Hortorum Cultus* 11(2): 217–228.
- Frąszczak, B., Kałużewicz, A., Krzesiński, W., Lisiecka, J., Spizewski, T. 2011. Effect of differential temperature and photoperiod on growth of *Ocimum basilicum* *Zemdirbyste-Agriculture* 98: 375–382.
- Ganesan, S. (2011). Growing our herbal industry. In *The Star* Malaysia: The Star.
- Gardner F.P., Pearce R.B., Mitchel R. L. 2008 Physiology of crop plant. Herawati Susilo (translator). Jakarta: UI Press.
- Ghosh, B.K., Datta, A.K., Das, A., Mandal, A., 2012. Induced macromutation in *Andrographis paniculata* (Burm. F.) nees. *International Journal of Research in Ayurveda and Pharmacy* 3: 604–610.
- Glaser, B., Birk, J.J. 2011. State of the scientific knowledge on properties and genesis of anthropogenic dark earths in Central Amazonia (terra preta de Índio). *Geochimica Et Cosmochimica Acta* 82: 39-51
- Glaser, B., Haumaier, L., Guggenberger, G., Zech, W. 2001. The “Terra Preta” phenomenon: A model for sustainable agriculture in the humid tropics. *Naturwissenschaften* 88: 37–41.
- Glaser, B., Lehmann, J., Zech, W. 2002. Ameliorating physical and chemical properties of highly weathered soils in the tropics with charcoal – a review. *Biology and Fertility of Soils* 35: 219-230.

- Goverdarica-Lucic, A., Perkovic, G., Rahimic, A. 2014. Influence of substrate and cell volume of containers on the quality of basil. In *Proceeding of Fifth International Scientific Agricultural Symposium*, ed. Kovacevic, D. pp 75–78. Mediterranean Agronomic Institute of Bari.
- Graber, E.R., Harel, Y.M., Kolton, M., Cytryn, E., Silber, A., David, D.R., Tsechansky, L., Borenshtein, M., Elad, Y. 2010. Biochar impact on development and productivity of pepper and tomato grown in fertigated soilless media. *Plant and Soil* 337(1): 481–496.
- Grappin, P., Bouinot, D., Sotta, B., Miginiac, E. and Julien, M. 2000. After-ripening of tobacco seeds. *Planta* 210: 279–285.
- Gutterman, Y. 1978. Seed coat permeability as a function of photoperiodical treatment of the mother plants during seed maturation in the desert annual plant *Trigonella arabica* Del. *Journal of Arid Environments* 1: 141–144.
- Gutterman, Y. 2000. Maternal effects on seeds during development. In *Seeds: The Ecology of Regeneration in Plant Communities*, ed. M. Fenner, 2nd Edition pp. 59–84. Oxon: CAB International
- Gutterman, Y. and Porath, D. 1975. Influences of photoperiodism and light treatments during fruits storage on the phytochrome and on the germination of *Cucumis prophetarum* L. and *Cucumis sativus* L. seeds. *Oecologia* 18: 37–45.
- Eskandari, H. 2012. Seed quality variation of crop plants during seed development and maturation. *International Journal of Agronomy and Plant Production* 3(11): 557–560.
- Hamman, B., Koning, G., Lok, K.H. 2003. Homeopathically prepared gibberellic acid and barley seed germination. *Homeopathy* 92: 140–144.
- Haug, R.T. 1993. The practical handbook of compost engineering. Florida: Lewis Publishers.
- Hay, F., Probert, R., Coomber, S. 1997. Development of desiccation tolerance and longevity in seeds from detached capsules of foxglove (*Digitalis purpurea* L.). *Annals of Botany* 79: 419–427.
- Heikal, M.M. 1981. The effect of osmotic stress on seed germination of flax, sesame and onion seeds. *Biologia Plantarum* 24(2): 124–129.
- Hochmuth, G., Hochmuth, R. 2012. Open-field soilless culture of vegetables. pp 1–5. Florida: University of Florida.
- Hong, T.D., Linington, S., Ellis, R.H. 1996. Seed storage behaviour: a compendium. UK: International Plant Genetic Resources Institute.

- Hossain, S., Urbi, Z., Sule, A., Rahman, K.M.H. 2014. *Andrographis paniculata* (Burm . f.) Wall. ex Nees : A review of ethnobotany , phytochemistry , and pharmacology. *The Scientific World Journal* 2014: 1-29.
- Hsu, C.C., Chen, C.L., Chen, J.J., Sung, J.M. 2003. Accelerated aging-enhanced lipid peroxidation in bitter gourd seeds and effects of priming and hot water soaking treatments. *Scientia Horticulture*. 98: 201–212.
- Hussain, A., Iqbal, K., Aziem, S., Mahato, P., Negi, A.K. 2014. A review on the science of growing crops without soil (soilless culture) – A novel alternative for growing crops. *International Journal of Agriculture and Crop Sciences* 7(11): 833–842.
- Iersel, M.V. 1997. Roots restriction effects on growth and development of salvia (*Salvia splendens*). *HortScience* 32(7): 1186-1190.
- Iglesias-Fernández, R., Matilla, A. 2009. After-ripening and sensitivity to GAs. *Journal of Experimental Botany* 60: 1645–1661.
- International Seed Testing Association (ISTA). 1985. International rules for seed testing. *Seed Science and Technology*. 13(2): 299-355.
- Ingram, D.L., Henley, R., Yeager, T.H. 1993. Growth media for container grown ornamental plants. Florida: Florida Cooperative Extension Service.
- Ismail, M.R., Noor, K.M. 1996. Growth, water relations and physiological processes of starfruit (*Averrhoa carambola* L) plants under root growth restriction. *Scientia Horticulturae* 66: 51-58.
- Jackson, S.D. 2008. Plant responses to photoperiod. *New Phytologist*. 181: 517-31.
- Jamal, J.A. 2006. Malay traditional medicine: An overview of scientific and technological progress. *Tech Monitor* 37–49.
- Jarillo, J. a, Olmo, I., Gómez-Zambrano, a., Lázaro, a, López-González, L., Miguel, E., Narro-Diego, L., Sáez, D., Piñeiro, M. 2008. Review. Photoperiodic control of flowering time. *Spanish Journal of Agricultural Research* 6: 221–244.
- Jarukamjorn, K., Nemoto, N. 2008. Pharmacological aspects of *Andrographis paniculata* on health and its major diterpenoid constituent andrographolide. *Journal of Health Science* 54(4): 370–381.
- Johnson, H. 1980. Soilless culture of greenhouse vegetables. California: Cooperative Extension.
- Kamotho, G., Mathenge, P., Muasya, R., Dulloo, M. 2014. Effects of maturity stage, dessication and storage period on seed quality of cleome (*Cleome gynandra* L.). *Research Desk* 3(1): 419–433.

- Kandil, A., Sharief, A., Sheteiwy, M. 2013. Effect of seed storage periods, conditions and materials on germination of some soybean seed cultivars. *American Journal of Experimental Agriculture* 3(4): 1020–1043.
- Kantolic, A.G., Peralta, G.E., and Slafer, G.A. 2013. Seed number responses to extended photoperiod and shading during reproductive stages in indeterminate soybean. *European Journal of Agronomy* 51: 91–100.
- Kermode, A.R. 1995. Regulatory mechanisms in the transition from seed development to germination: Interactions between the embryo and the seed development. In *Seed Development and Germination*, ed. G. Galili and J. Kigel, pp. 273–332. New York: Marcel Dekker, Inc.
- Khatun, A., Kabir, G., Bhuiyan, M. 2009. Effect of harvesting stages on the seed quality of lentil (*Lens culinaris* L.) during storage. *Bangladesh Journal of Agricultural Research* 34(4): 565–576.
- Konduru, S., Evans, M.R., Stamps, R.H. 1999. Coconut husk and processing effects on chemical and physical properties of coconut coir dust. *HortScience* 34: 88–90.
- Kraus, H.T., Extension, V.C., Mikkelsen, R.L., Warren, S.L. 2000. Container Substrate temperatures affect mineralization of composts. *HortScience* 35(1): 16–18.
- Kumar, B., Verma, S.K., Singh, H.P. 2011. Effect of temperature on seed germination parameters in Kalmegh (*Andrographis paniculata* Wall. ex Nees.). *Industrial Crops and Products* 34(1): 1241–1244.
- Kumar, R.N., Chakraborty, S., Kumar N.J.I. 2012. Influence of light and development stages on active principles of *Andrographis paniculata* (Burm. f.) Wall. ex Nees. *Indian Journal of Scientific Research* 3(1): 91–95.
- Kumar, R.N., Chakraborty, S., Kumar, N.J.I. 2010. Methods to break seed dormancy of *Andrographis paniculata* (Burm. f. Nees): An Important Medicinal Herb of Tropical Asia. *Advances in Bioresearch* 1(2): 35–39.
- Kumari, A., Lal, R.K., Singh, K.L.B. 2012. Comparative study of seed germination and seed vigour test in *Andrographis paniculata* (Acanthaceae). *Botanica Serbica* 36(1): 49–52.
- Laird, D.A., Fleming, P.D., Davis, D.D., Horton, R., Wang, B., Karlen, D.L. 2010. Impact of biochar amendments on the quality of a typical Midwestern agricultural soil. *Geoderma* 158: 443–449.
- Larson, R.A. 1992. Introduction to Floriculture, 2nd edition. California: Academic Press Inc, San Diego.
- Latimer, J.G. 1991. Container size and shape influence growth and landscape performance of marigold seedlings. *HortScience* 26: 124–126.

- Lattoo, S.K., Khan, S., Dhar, A.K., Choudhary, D.K., Gupta, K.K., Sharma, P.R. 2006. Genetics and mechanism of induced male sterility in *Andrographis paniculata* (Burm. f.) Nees and its significance. *Current Science* 91: 515–519.
- Lehmann, J. 2007. Bio-energy in the black. *Frontiers in Ecology and the Environment* 5(7): 381–387.
- Lehmann, J., Gaunt, J., Rondon, M. 2006. Biochar sequestration in terrestrial ecosystems—a review. *Mitigation and Adaptation Strategies for Global Change* 11: 403–427.
- Lehmann, J., Joseph, S. 2009. Biochar for environmental management: An introduction. *Science and Technology* 1: 1–12.
- Lehmann, J., Rondon, M. 2006. Bio-Char soil management on highly weathered soils in the humid tropics. In *Biological Approaches to Sustainable Soil Systems*, ed. Uphoff, N., Ball, A.S., Palm, C., Fernandes, E., Pretty, J., Herren, H., Sanchez, P., Husson, O., Sanginga, N., Laing, M., Thies, J., pp. 517–530. London: CRC Press.
- Leubner-Metzger, G. 2003. Functions and regulation of β -1,3-glucanases during seed germination, dormancy release and after-ripening. *Seed Science Research* 13: 17–34.
- Liu, J., Schulz, H., Brandl, S., Miehtke, H., Huwe, B., Glaser, B. 2012. Short-term effect of biochar and compost on soil fertility and water status of a Dystric Cambisol in NE Germany under field conditions. *Journal of Plant Nutrition and Soil Science* 175: 698–707.
- Lopez, R.G., Runkle, E.S., Heins, R.D., Whitman, C.M. 2003. Temperature and Photoperiodic Effects on Growth and Flowering of *Zygopetalum Redvale* ‘Fire Kiss’ Orchids. *Elegant Science in Floriculture* 624: 155–162.
- Mahesha, C.R., Channaveeraswami, A.S., Kurdikeri, M.B., Shekhargouda, M., Merwade, M.N. 2001. Storability of sunflower seeds harvested at different maturity dates. *Seed Research* 29(1): 98–102.
- Marsh, D.B., Paul, K.B. 1988. Influence of container type and cell size on cabbage transplant development and field performance. *HortScience* 23: 310–311.
- Maynard, E.T., Vavrina, C.S., Scott, W.D. 1996. Containerized muskmelon transplants: Cell volume effects on pretransplant development and subsequent yield. *HortScience* 31: 58–61.
- Mazumdar, B.C. 2005. Photoperiodism and Vernalisation in Plants. pp. 40–44. New Delhi: Daya Publishing House.
- McClellan, T., Deenik, J., Uehara, G., Antal, M. 2007. Effects of flashed carbonized macadamia nutshell charcoal on plant growth and soil chemical properties. November 6, 2007, ASA-CSSA-SSA International Annual Meetings, New

- Orleans, Louisiana. Retrieved at 15 January 2015. Available at <http://acs.confex.com/crops/2007am/techprogram/P35834.HTM>.
- McDonald MB. 1999. Seed deterioration: physiology, repair and assessment. *Seed Science Technology* 27: 177–237.
- McKenzie, H. R. *Soil pH and Plant Nutrients*. Government of Alberta. Agri-Facts: Alberta. 2003.
- McLaughlin, H., Anderson, P.S., Shields, F.E., Reed, T.B. 2009. All biochars are not created equal, and how to tell them apart. In *Proceedings of North American Biochar Conference*, Boulder, Colorado, August 2009, International Biochar Initiative: Boulder. 2009.
- Megat Johari, M.M., Ahmad Jusoh, Abdul Halim, G. 1990. Land application. In *Management and Utilization of Oil Palm Wastes. A Review*, pp. 25-33. Selangor: Malindo Publisher.
- Methods Manual: Soil Testing in India*. 2011. Retrieved from <http://agricoop.nic.in/dacdivision/mmsoil280311.pdf>.
- Milla, O.V., Rivera, E.B., Huang, W.J., Chien, C.C., Wang, Y.M. 2013. Agronomic properties and characterization of rice husk and wood biochars and their effect on the growth of water spinach in a field test. *Journal of Soil Science and Plant Nutrition* 13(2): 251–266.
- Mitrovic, A., Giba, Z., Culafic, L. 2007. The photoperiodic control of growth and development of *Chenopodium rubrum* L. plants in vitro. *Archives of Biological Science Belgrade* 59(3): 203–208.
- Mitrovic, A., Zivanovic, B., Culafic, L. 2002. Maternal effect on *Chenopodium rubrum* L. seeds: seed size, germination, growth and flowering in vitro. *Ekologija* 37: 53–58.
- Mohammadi, H., Soltani, A., Sadeghipour, H.R., Zeinali, E. 2011. Effects of seed aging on subsequent seed reserve utilization and seedling growth in soybean. *International Journal of Plant Production* 5(1): 65-70.
- Mousavi, S.R., Rezaei, M., Mousavi, A. 2011. A General Overview on Seed Dormancy and Methods of Breaking It. *Advances in Environmental Biology* 5: 3333–3337.
- Mubvuma, M.T., Mapanda, S., Mashonjowa, E. 2013. Effect of storage temperature and duration on germination of moringa seeds (*Moringa oleifera*). *Greener Journal of Agricultural Sciences* 3(5): 427–432.
- Nesmith, D.S., Duval, J.R. 1998. The effect of container size. *Horttechnology* 8(4): 495–498.

- Nikolaeva, M.G. 1977. Factors controlling the seed dormancy pattern. In: Khan AA ed., *The physiology and biochemistry of seed dormancy and germination*. Amsterdam, North-Holland, 51-74.
- Niranjan, A., Tewari, S.K., Lehri, A. 2010. Biological activities of Kalmegh (*Andrographis paniculata* Nees) and its active principles-A review. *Indian Journal of Natural Products and Resources* 1(2): 125–135.
- Noguera, P., Abad, M., Noguera, V., Puchades, R., Maquieira, A. 2000. Coconut coir-waste, a new and viable ecologically friendly peat substitute. *Acta Horticulturae* 517: 279-286.
- Noguera, P., Abad, M., Puchades, R., Noguera, V., Maquieira, A., Martinez, J. 1997. Physical and chemical properties of coir waste and their relation to plant growth. *Acta Horticulturae* 450: 365-373.
- Noorhanin, D., Edaroyati, P. M. W. & Aishah, S. H. (2013). Response of Different Media Ratio on Growth and Biomass Production of *Andrographis paniculata* (Hempedu Bumi) Grown under Soilless Culture System. *Trans. Malaysian Soc. Plant Physiology* 21: 12-18.
- Northup, J.I. 2013. *Biochar as a replacement for perlite in greenhouse soilless substrates*, Master Thesis, Iowa State University.
- Olasoji, J.O., Aluko, A.O., Adeniyi, O.N., Olanipekun, S.O., Olosunde, A.A., Okoh, J.O. 2011. Effect of time of harvest on physiological maturity and kenaf (*Hibiscus cannabinus*) seed quality. *African Journal of Plant Science* 6(10): 282–289.
- Parashar, R., Upadhyay, A., Singh, J., Diwedi, S.K., Khan, N.A. 2011. Morpho-Physiological evaluation of *Andrographis paniculata* at different growth stages. *World Journal of Agricultural Sciences* 7(2): 124–127.
- Passioura, J.B. 2006. The perils of pot experiments. *Functional Plant Biology* 33, 1075–1079.
- Peterson, T.A., Reinsel, M.D., Krizek, D.T. 1991a. Tomato (*Lycopersicon esculentum* Mill. cv 'Better Bush') plant response to root restriction. Alteration of plant morphology. *Journal of Experimental Botany* 42: 1233-1240.
- Peterson, T.A., Reinscl, M.D., Krizek, D.T. 1991b. Tomato (*Lycopersicon esculentum* Mill. cv 'Better Bush') plant response to root restriction. Root respiration and ethylene generation. *Journal of Experimental Botany* 42: 1241-1249.
- Pietikäinen, J., Kiikkilä, O., Fritze, H. 2000. Charcoal as a habitat for microbes and its effect on the microbial community of the underlying humus. *Oikos* 89: 231-242.

- Pourrat, Y., Jacques, R. 1978. The influence of photoperiodic conditions received by the mother plant on morphological and physiological characteristics of *Chenopodium polyspermum* L. seeds. *Plant Science Letters* 4: 273–279.
- Pradhan, B.K., Badola, H.K., 2012. Effect of storage conditions and storage periods on seed germination in eleven populations of *Swertia chirayita*: A critically endangered medicinal herb in himalaya. *The Scientific World Journal* 2012: 1–9.
- Prasad, M. 1997. Physical, chemical and microbiological properties of coir (cocopeat). *Acta Horticulturae* 40: 21–27.
- Puustjarvi, V., Robertson, R.A. 1975. Physical and chemical properties. In *Peat in Horticulture*, ed. Robinson, D.W. and Lamb, J.G.D., pp. 23–38. New York: Academics.
- Rao, N. K., Roberts, E. H., Ellis, R. H. 1987. Loss of viability in lettuce seeds and the accumulation of chromosome damage under different storage conditions. *Annals of Botany* 60(1): 85–96.
- Raviv, M., Lieth, J.H. 2008. Significance of soilless culture in agriculture. In *Soilless culture: theory and practice*, 1st edition, ed. Raviv, M., Lieth, J.H., pp. 1–11. Amsterdam: Elsevier.
- Rawat, R., Vashistha, D.P. 2011. Seed germination improvement in *Andrographis paniculata*. *International Journal of Medicinal and Aromatic Plants* 1(3): 348–350.
- Rayburn, A.P., Davidson, J.B., Schupp, E.W. 2013. Effect of storage time, site and floral morph on seed germination of the threatened distylous primrose *Primula cusickiana* var. *maguirei*. *Plant Species Biology* 28: 101–108.
- Reeve, J. R., Carpenter-Boggs, L., Reganold, J. P., York, A. L., Brinton, W. F. 2010. Influence of biodynamic preparations on compost development and resultant compost extracts on wheat seedling growth. *Bioresource Technology* 101: 5658–5666.
- Robbins, J.A., Evans, M.R. 2001. Growing Media for Container Production in a Greenhouse or Nursery Part II – Physical and Chemical Properties. Fayetteville: University of Arkansas Cooperative Extension Service Printing Services. Retrieved from <http://www.uaex.edu/publications/pdf/FSA-6098.pdf>
- Rohimin, F.W. 2013. *Seed Dormancy Breaking Techniques and Seed Production of Andrographis paniculata as Influenced by Photoperiod*, Bachelor Degree Thesis, Universiti Putra Malaysia.

- Sacandé, M., Buitink, J., Hoekstra, F.A. 2000. A study of water relations in neem (*Azadirachta indica*) seed that is characterized by complex storage behaviour. *Journal of Experimental Botany* 51: 635–43.
- Samsuddin, M.F., Saud, H.M., Ismail, M.R., Omar, M.H., Habib, S.H., Saikat, M., Bhuiyan, H., Kausar, H. 2014. Effect of different combinations of coconut coir dust and compost on rice grown under soilless culture. *Journal of Food, Agriculture & Environment* 12(2): 1280-1283.
- Samarah, N.H. 2006. Effect of air-drying immature seeds in harvested pods on seed quality of common vetch (*Vicia sativa* L.). *New Zealand Journal of Agriculture Research* 49(3): 331–339.
- Saraswathy, S., Manavalan, R. S. A., Vadivel, E., Manian, K., Subramanian, S. 2004. Studies on seed germination in Kalmegh (*Andrographis paniculata* Nees.). *South India Horticulture* 52(1/6): 286-290.
- Schmidt, L. 2007. Seed storage. In *Tropical Forest Seed, Tropical Forestry*, ed. Heidelberg D.C., pp 143-197. New York: Springer.
- Schnelle, R.A., Barrett, J.E., Clark, D.G. 2006. High temperature delay of floral initiation in modern poinsettia cultivars. V International Symposium on Artificial Lighting in Horticulture. ISHS Acta Horticulture 711: 273-278.
- Schulz, H., Glaser, B. 2012. Effects of biochar compared to organic and inorganic fertilizers on soil quality and plant growth in a greenhouse experiment. *Journal of Plant Nutrition and Soil Science* 175(3): 410–422.
- Sharma, A., Shanker, C., Tyagi, L.K., Singh, M., Rao, C.V. 2008. Herbal medicine for market potential in India: An overview. *Academic Journal of Plant Sciences* 1(2): 26-36.
- Sicelakuphila, S., Kamau Wahome, P. 2010. Effects of application of short-days at different periods of the day on growth and flowering in chrysanthemum (*Dendranthema grandiflorum*). *Journal of Agriculture and Social Science* 10(6): 39–42.
- Simic, A., Vuckovic, S., Sokolovic, D., Stanisavljevic, R., Mandic, V., Duronic, G. 2012. Response of Italian ryegrass seed crop to spring nitrogen application in the first harvest year. *African Journal of Biotechnology* 11(26): 6826-6831.
- Simpson, G.M. 1990. Seed dormancy in grasses. New York: *Cambridge University Press*. 297 p.
- Singh, D., Singh, J.P., Tewari, A. 2011. Influence of photoperiod and flowering on tuber yield of potato cultivar Kufri Pukharaj in Tarai region of Uttarakhand. *Pantnagar Journal of Research* 9: 315–316.

- Sirtautas, R., Samuolienė, G., Brazaitytė, A., Duchovskis, P., 2011. Temperature and photoperiod effects on photosynthetic indices of radish (*Raphanus sativus* L.). *Zemdirbyste-Agriculture* 98(1): 57–62.
- Siva, G., Sivakumar, S., Premkumar, G., Baskaran, P., Senthilkumar, T., Jayabalan, N. 2014. Enhanced seed germination of *Psoralea corylifolia* L. by heat treatment. *World Journal of Agricultural Research* 2(4): 151–154.
- Smith, J.L. & Collins, H.P. 2007. Composting. In *Soil Microbiology, Ecology, and Biochemistry* (3rd edition), ed. Paul, E.A., pp. 483–486. Burlington: Academic Press.
- Sohi, S., Lopez-Capel, E., Krull, E., Bol, R. *Biochar, climate change and soil: A review to guide future research*. Commonwealth Scientific and Industrial Research Organisation (CSIRO) Land and Water Science. CSIRO: Canberra. 2009
- Sønsteby, A., Heide, O.M. 2009. Effects of photoperiod and temperature on growth and flowering in the annual (primocane) fruiting raspberry (*Rubus idaeus* L.) cultivar 'Polka'. *Journal of Horticultural Science & Biotechnology* 84(4): 439–446.
- Sopha, G.A., Widodo, W.D., Poerwanto, R., Endah, R. 2014. Photoperiod and gibberellins effect on True shallot seed formation. *Advances in Agriculture & Botany-International Journal of the Bioflux Society* 6(1): 70–76.
- Spann, T.M. 2001. *Environmental influences on flower bud initiation in Vaccinium species*, Master Thesis, University of Florida.
- Spann, T.M., Williamson, J.G., Darnell, R.L. 2003. Photoperiodic effects on vegetative and reproductive growth of *Vaccinium darrowi* and *V. corymbosum* interspecific hybrids. *HortScience*, 38(2): 192–195.
- Spann, T.M., Williamson, J.G., Darnell, R.L. 2004. Photoperiod and temperature effects on growth and carbohydrate storage in Southern highbush blueberry interspecific hybrid. *Journal of the American Society for Horticultural Science* 129(3): 294–298.
- Steiner, C., Teixeira, W. G., Lehmann, J., Nehls, T., de Macedo, J. L. V., Blum, W. E. H., Zech, W. (2007): Long term effects of manure, charcoal and mineral fertilization on crop production and fertility on a highly weathered Central Amazonian upland soil. *Plant and Soil* 291: 275–290.
- Suhaimi, M.Y., Mohamad, a. M., Mahamud, S., Khadzir, D. 2012. Effects of substrates on growth and yield of ginger cultivated using soilless culture. *Journal of Tropical Agriculture and Food Science* 40(2): 159–168.
- Sutisna A. 2010 Improvement of lily flowering by GA3 and long photoperiode application. *Buletin Teknik Pertanian* 15(1):19–23.

- Tabatabaei, S.A. 2013. The effects of storage on germination characteristics of barley seeds. *International Journal of Agronomy and Plant Production* 4(12): 3337–3343.
- Talei, D., Valdiani, A., Abdullah, M.P., Hassan, S.A. 2012. A rapid and effective method for dormancy breakage and germination of King of Bitters (*Andrographis paniculata* Nees.) seeds. *Maydica* 57(2): 98-105.
- Vaknin, Y., Dudai, N., Murkhovsky, L., Gelfandbein, L., Fischer, R., Degani, A. 2009. hook. (Lemon-Scented Gum). *Journal of Herbs, Spices & Medicinal Plants* 15(2): 164–176.
- Valdiani, A., Kadir, M.A., Tan, S.G., Talei, D., Abdullah, M.P., Nikzad, S. 2012. Nain-e havandi *Andrographis paniculata* present yesterday, absent today: A plenary review on underutilized herb of Iran's pharmaceutical plants. *Molecular Biology Reports* 39: 5409–5424.
- Vavrina, C.S. *An introduction to the production of containerized vegetable transplants*. University of Florida. University of Florida, Cooperative Extension Service, Institute of Food and Agricultural Sciences: Florida. 1995.
- Will R.E., Teskey R.O. 1997. Effect of elevated carbon dioxide concentration and root restriction on net photosynthesis, water relations and foliar carbohydrate status of loblolly pine seedlings. *Tree Physiology* 17: 655–661.
- Wira, a B., Razi, I.M., Jamil, Z.A. 2011. Composts as additives in coconut coir dust culture for growing rockmelon (*Cucumis melo* L.). *Journal of Tropical Agriculture and Food Science* 39(2): 229–237.
- Woods, W. I., Mann, C.C. 2000. The good earth: Did people improve the Amazon basin? *Science* 287, 788.
- Yadav, O.P., Verma, P.K., Tyagi, C.S., Singh, S. 2008. Seed treatment to improve germination in liquorice (*Glycyrrhiza glabra* L.). *Seed Research* 36, 76-77.
- Yamato, M., Okimori, Y., Wibowo, I.F., Anshori, S., Ogawa, M. 2006. Effects of the application of charred bark of acacia mangium on the yield of maize, cowpea and peanut, and soil chemical properties in South Sumatra, Indonesia. *Journal of Soil Science and Plant Nutrition* 52: 489-495.
- Yang, Z., Hammer, G., van Oosterom, E., Rochais, D., Deifel, K. 2010. Effects of pot size on growth of maize and sorghum plants, In *Proceedings of the 1st Australian Summer Grains Conference*, Gold Coast, Australia, June 21-24, 2010, ed. George_Jaeggli, B and Jordan, D.J. The Grains Research and Development Corporation: Gold Coast. 2010.

Zheng, Z.L., Yang, Z., Jang, J.C., Metzger, J.D. 2006. Phytochromes AI and BI have distinct functions in the photoperiodic control of flowering in the obligate long-day plant *Nicotiana sylvestris*. *Plant, Cell & Environment* 29: 1673-1685.

Zhu L.N., Wang S.P., Yang T.Y., Zhang C.X., Xu W.P. 2006. Vine growth and nitrogen metabolism of 'Fujiminori' grapevines in response to root restriction. *Scientia Horticulturae* 107: 143-149.

Zohary, M. 1972. *Flora Palestina*, Part II – *Text*. Israel Academy of Sciences and Humanities, Jerusalem, pp. 489.

