



UNIVERSITI PUTRA MALAYSIA

***GROWTH, PHYTOCHEMICAL AND ANTIOXIDANT ACTIVITY OF
Orthosiphon stamineus BENTH. IN RESPONSE TO ORGANIC
AMENDMENT, FERTILIZER AND HARVEST DATE***

ESTHER YAP SHIAU PING

FP 2016 34



**GROWTH, PHYTOCHEMICAL AND ANTIOXIDANT ACTIVITY OF
Orthosiphon stamineus BENTH. IN RESPONSE TO ORGANIC AMENDMENT,
FERTILIZER AND HARVEST DATE**

By

ESTHER YAP SHIAU PING

**Thesis Submitted to the School of Graduate Studies,
Universiti Putra Malaysia, in Fulfillment of the Requirements for the Degree of
Master of Science**

June 2016

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

Dedicated to my beloved parents, Yap Lian Huat and Teoh Sok Em, my sister, Estina Yap Shiau Yih for their endless love, support, understandings, sacrifices, motivation, advice and encouragement.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

**GROWTH, PHYTOCHEMICAL AND ANTIOXIDANT ACTIVITY OF
Orthosiphon stamineus BENTH. IN RESPONSE TO ORGANIC AMENDMENT,
FERTILIZER AND HARVEST DATE**

By

ESTHER YAP SHIAU PING

June 2016

Chairperson : Siti Hajar Ahmad, PhD
Faculty : Agriculture

Orthosiphon stamineus have been identified by Malaysian Department of Agriculture with the potential to be developed as complementary and alternative medicine. *O. stamineus* acts as a diuretic agent and has nephroprotective, antifungal, antimicrobial and antipyretic properties. It contained chemical markers, such as sinensetin (SEN) and rosmarinic acid (RA), with beneficial effects on consumer's health. Malaysian soils are predominantly kaolinitic clays and sandy due to weathering and is unsuitable for plant growth. Thus, it is important to incorporate organic soil amendments before planting to improve the soil. The objective of the first experiment was to determine the rates of soil amendments (rice husk biochar (BC): 0, 5 and 10 t/ha and chicken manure (CM): 0, 2.5 and 5 t/ha) and harvesting week (HW) that could produce the maximum yield and phytochemicals. The experiment was conducted as a potted experiment using a randomized complete block design in a three factorial arrangement of treatments with four replications and three subsamples each. There was a significant quadratic increase in plant height as harvesting time was increased, irrespective of soil amendments. Plant fresh weights were significantly affected by interactions between BC \times CM \times HW, whereby BC 0 t/ha and CM 5 t/ha at week 8 after transplanting gave the highest yield among all the treatments. There was a significant linear relationship between BC \times HW for plant dry weight. The plant dry weight showed differences of 15%, 20% and 42% between week 6 and 8 at each BC rate. Both total phenolic and flavonoid contents were also affected by interactions between BC \times CM \times HW. 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical scavenging activity showed a significant quadratic response in the interaction between BC \times HW whereby week 7 produced maximum DPPH activity. Sinensetin was significantly reduced by 63% at week 4 as compared to week 8, irrespective of soil amendments. The combination of BC 5 t/ha, CM 2.5 t/ha and HW 8 produced highest biomass yield, as HW 8 gave 36% higher yield than week 6, although SEN yield was not significantly different between week 6 and 8. The second experiment was carried out to determine the optimum fertilizer rate, FR (0, 100, 200, 300 and 400 kg/ha N) and fertilizer type, FT (plant- and animal-based) that could produce the maximum yield and phytochemicals. The selected treatment combination (BC 5 t/ha and CM 2.5 t/ha at HW 8) from the first experiment was used for soil

preparation. The potted experiment was conducted using a RCBD in a two factorial arrangement of treatments with four replications and three subsamples each. Both fresh and dry weights showed a significant quadratic trend against FR, irrespective of FT applied, with optimum rates at 323 kg/ha N and 219 kg/ha N, respectively. Total flavonoid contents (TFC) and DPPH activity were significantly affected by interaction between FR \times FT. DPPH activity and TFC showed a quadratic trend for plant-based fertilizer with optimum rate at 218 kg/ha N and 244 kg/ha N, respectively. Chemical markers, RA and SEN, were also affected by interaction between FT \times FR. Plant-based fertilizer showed a reduction in both the chemical markers as fertilizer rates increased. However, for animal-based fertilizer, the trend was quadratic, whereby the concentrations of RA and SEN were decreased by 61% and 13%, respectively, with increasing fertilizer rate from 0 kg/ha N to 200 kg/ha N. A gradual decrease, thereafter an increase in response to fertilizer rate was seen. The results showed plant-based fertilizers at 300 kg/ha N is the optimum organic fertilizer order to achieve maximum yield and phytochemicals. In conclusion, the optimum yield of *O. stamineus* can be obtained by using 5 t/ha rice husk biochar and 2.5 t/ha chicken manure soil amendments, plant-based organic fertilizer of 300 kg/ha N and harvested at week 8 after transplanting.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PERTUMBUHAN, KANDUNGAN FITOKIMIA DAN AKTIVITI ANTIOKSIDA BAGI *Orthosiphon stamineus* BENTH. SEBAGAI TINDAK BALAS TERHADAP PERAPI ORGANIK, BAJA DAN MASA PENUAIAN

Oleh

ESTHER YAP SHIAU PING

Jun 2016

Pengerusi : Siti Hajar Ahmad, PhD
Fakulti : Pertanian

Orthosiphon stamineus telah dikenal pasti oleh Jabatan Pertanian Malaysia sebagai tumbuhan yang berpotensi untuk dibangunkan sebagai perubatan komplementari dan alternatif. *O. stamineus* bertindak sebagai agen diuretik dan melindungi buah pinggang, anti-kulat, anti-mikrob dan antipiretik. Ia mengandungi penanda kimia seperti sinensetin (SEN) dan asid rosmarinik (RA) yang memberi kesan baik terhadap kesihatan penggunanya. Sebahagian besar tanah di Malaysia terdiri daripada tanah liat kaolin dan berpasir yang disebabkan oleh proses luluhawa dan tidak sesuai untuk pertumbuhan pokok. Oleh itu, menggunakan perapi tanah adalah penting sebelum penanaman untuk membaikpulih keadaan tanah. Objektif kajian pertama adalah untuk menentukan kadar perapi tanah (sekam padi biochar (BC): 0, 5 dan 10 t/ha dan tahi ayam (CM): 0, 2,5 dan 5 t/ha) dan minggu penuaian (HW) yang dapat mengeluarkan hasil tanaman dan kandungan fitokimia yang maksimum. Kajian ini dijalankan sebagai eksperimen pasu yang menggunakan reka bentuk blok rawak lengkap dalam tiga susunan faktorial rawatan, dengan empat ulangan dan tiga sub-sampel setiap ulangan. Terdapat peningkatan kuadratik yang ketara dalam ketinggian tumbuhan seiring dengan peningkatan masa penuaian, tanpa dipengaruhi oleh aplikasi perapi tanah. Berat basah tanaman dipengaruhi dengan ketara oleh interaksi diantara BC \times CM \times HW, di mana BC 0 t / ha dan CM 5 t/ha pada minggu ke-8 selepas anak pokok dialih, memberikan hasil yang tertinggi di antara semua rawatan. Terdapat hubungan linear yang ketara antara BC \times HW untuk berat kering tanaman. Berat kering tanaman menunjukkan perbezaan pada kadar 15%, 20% dan 42% diantara minggu ke-6 dan 8 bagi setiap kadar BC. Kedua-dua jumlah kandungan fenolik dan flavonoid turut dipengaruhi oleh interaksi diantara BC \times CM \times HW. Aktiviti 2,2- difenil -1- pikrilhidrazil (DPPH) yang memerangkap radikal bebas menunjukkan tindak balas kuadratik yang ketara dalam interaksi antara BC \times HW manakala minggu ke-7 menghasilkan aktiviti DPPH yang maksimum. Kandungan SEN telah berkurang dengan ketara sebanyak 63% pada minggu ke-4 berbanding minggu ke-8, tanpa mengambil kira aplikasi perapi tanah. Gabungan BC 5 t/ha, CM 2.5 t/ha dan HW 8 menghasilkan hasil biojisim tertinggi, kerana HW 8 memberikan hasil 36% lebih tinggi daripada minggu ke-6, walaupun hasil SEN tidak jauh berbeza antara minggu ke-6 dan ke-8. Kajian kedua telah

dijalankan untuk menentukan kadar optimum baja, FR (0 , 100, 200 , 300 dan 400 kg/ha N) dan jenis baja, FT (berasaskan tumbuhan dan berasaskan haiwan) yang boleh mengeluarkan hasil dan kandungan fitokimia yang maksimum. Gabungan rawatan yang dipilih (BC 5 t/ha , CM 2.5 t/ha dan HW 8) daripada percubaan pertama telah digunakan untuk penyediaan tanah. Kajian di dalam pasu telah dijalankan menggunakan RCBD dalam rawatan dua susunan faktorial, dengan empat ulangan dan tiga sub-sampel. Kedua-dua berat basah dan kering menunjukkan tren kuadratik yang ketara berbanding FR, tanpa mengira aplikasi FT, dengan kadar optimum pada 323 kg/ha N dan 219 kg/ha N masing-masing. Jumlah kandungan flavonoid (TFC) dan aktiviti DPPH dipengaruhi dengan ketara oleh interaksi antara FR \times FT. Aktiviti DPPH dan TFC menunjukkan corak kuadratik untuk baja berasaskan tumbuhan dengan kadar optimum pada 218 kg/ha N dan 244 kg/ha N , masing-masing. Penanda kimia, RA dan SEN, turut dipengaruhi oleh interaksi antara FT \times FR. Baja yang berasaskan tumbuhan menunjukkan pengurangan dalam kedua-dua penanda kimia seiring dengan peningkatan kadar baja. Walau bagaimanapun, baja berasaskan haiwan telah menunjukkan ciri kuadratik, di mana kepekatan RA dan SEN telah menurun dengan kadar baja yang semakin meningkat diikuti dengan penurunan secara beransur-ansur, selepas itu meningkat sebagai tindak balas kepada kadar baja. Hasil kajian menunjukkan baja berasaskan tumbuhan pada 300 kg/ha N adalah kadar baja organik yang optimum untuk mencapai hasil dan kandungan fitokimia yang maksimum. Kesimpulannya, hasil optima *O. stamineus* boleh didapati dengan menggunakan 5 t/ha sekam padi biochar, 2.5 t/ha tahi ayam, baja berasaskan tumbuhan pada 300 kg/ha N dan dituai pada minggu 8 selepas pemindahan anak pokok.

ACKNOWLEDGEMENTS

First and foremost, I would like to offer my heartfelt appreciation and utmost gratitude to my supervisor, Associate Professor Dr. Siti Hajar Ahmad for her continuous support and invaluable guidance for my Master study, for her patience, motivation and enthusiasm. During my Master study, she provided advice and shared a lot of her expertise, research insight and ideas. I simply could not imagine having a better advisor and mentor for Master study. I believe that one of the main gains of my Master study was working with Associate Professor Dr. Siti Hajar Ahmad.

With a great deal of luck, I got an excellent Supervisory Committee. I owe an immense debt to the rest of my supervisory committee, Professor Datin Dr. Rosenani Abu Bakar, and Professor Dr. Khozirah Shaari for their encouragement, insightful comments and critical review. This thesis could not have been done without their strong supervision.

I am deeply indebted to my dear family Mr. Yap Lian Huat, Mrs Teoh Sok Em and Estina Yap Shiau Yih, who deserve special attention for their unconditional support.

I would like to thank UPM for providing Graduate Research Fellowship (GRF) and research facilities to conduct my Master study. I consider it an honor to work with all the administrative and technical staffs of the Faculty Agriculture especially Mr Azhar Othman. I would also like to say thank you very much to Miss Surisa Phornvillay, Miss Zahidah Ab Razak, Miss Tan Xue Yi, Miss Nur Indah Shukor, Miss Nor Elliza Tajidin, Miss Azimah Hamidon, Miss Munirah, Miss Bunga Raya Ketaren, Miss Najihah and Mr Mohd Zakwan Zamri who helped me during my Master study duration.

I certify that a Thesis Examination Committee has met on 30 June 2016 to conduct the final examination of Esther Yap Shiau Ping on her thesis entitled "Growth, Phytochemical and Antioxidant Activity of *Orthosiphon stamineus* Benth. in Response to Organic Amendment, Fertilizer and Harvest Date" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Uma Rani a/p Sinniah, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Siti Aishah binti Hassan, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Internal Examiner)

Zakaria Wahab, PhD

Professor
Universiti Malaysia Perlis
Malaysia
(External Examiner)



ZULKARNAIN ZAINAL, PhD

Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 28 September 2016

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree Master of Science. The members of Supervisory Committee were as follows:

Siti Hajar Binti Ahmad, PhD

Faculty of Agriculture
Universiti Putra Malaysia
(Chairperson)

Rosenani Binti Abu Bakar, PhD

Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Khozirah Shaari, PhD

Faculty of Science
Universiti Putra Malaysia
(Member)

BUJANG KIM HUAT, PHD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Esther Yap Shiau Ping; GS39029

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of the thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____

Name of Chairman of
Supervisory Committee:

Siti Hajar Binti Ahmad, PhD

Signature: _____

Name of Member of
Supervisory Committee:

Rosenani Abu Bakar, PhD

Signature: _____

Name of Member of
Supervisory Committee:

Khozirah Shaari, PhD

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiii
LIST OF FIGURES	xv
LIST OF APPENDICES	xvii
LIST OF ABBREVIATIONS	xix
CHAPTER	
1 GENERAL INTRODUCTION	1
2 LITERATURE REVIEW	
2.1 <i>Orthosiphon stamineus</i>	4
2.1.1 Morphology	5
2.1.2 Plant Propagation, Planting and Cultural Practices	7
2.1.3 Harvest Age and Postharvest Handling	8
2.2 Antioxidant Compound	9
2.2.1 Extraction	9
2.2.2 Total Phenolic Content (TPC)	9
2.2.3 Total Flavonoid Content (TFC)	10
2.2.4 Antioxidant Activity	11
2.3 Soil amendment	11
2.3.1 Rice husk biochar (BC)	12
2.3.2 Chicken manure (CM)	12
2.4 Organic fertilizer	13
3 IMPACT OF TYPES AND RATES OF ORGANIC AMENDMENTS, AND HARVESTING TIME ON YIELD, ANTIOXIDANT COMPOUNDS AND ACTIVITIES, AND CHEMICAL MARKER OF <i>Orthosiphon stamineus</i> Benth.	
3.1 Introduction	14
3.2 Materials and Methods	15
3.2.1 Media and plant preparation	15
3.2.2 Plant maintenance	16
3.2.3 Experimental design and data analysis	16
3.2.4 Harvesting and drying	16
3.2.5 Parameter evaluation	16
3.2.5.1 Plant height	16
3.2.5.2 Fresh and dry weights	16
3.2.6 Chemical analysis	16
3.2.6.1 Extraction	16
3.2.6.2 Total phenolic content determination	17
3.2.6.3 Total flavonoids content	17

	determination	
	3.2.6.4 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging capacity	17
	3.2.6.5 High-performance liquid chromatography (HPLC) Analysis for Sinensetin	18
3.2.7	Plant nutrient analysis (N, P, K)	18
3.2.8	Soil pH	18
3.3	Results and Discussion	19
3.3.1	Plant height, fresh and dry weights	19
3.3.2	Soil characteristics before and after treatment	24
3.3.3	Antioxidant compounds and activity	25
3.3.4	Correlation coefficients between fresh and dry weight, plant height, soil pH, total phenolic contents, total flavonoid contents and 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical scavenging capacity	31
3.3.5	Chemical marker	32
3.3.6	Phytochemicals and plant nitrogen uptake based on biomass yield and harvesting week	32
3.4	Conclusion	33
4	IMPACT OF ORGANIC FERTILIZERS ON ANTIOXIDANT COMPOUNDS AND ACTIVITIES, AND CHEMICAL MARKER OF <i>Orthosiphon stamineus</i> BENTH.	
4.1	Introduction	35
4.2	Materials and methods	36
4.2.1	Media and plant preparation	36
4.2.2	Plant maintenance	36
4.2.3	Experimental design and data analysis	36
4.2.4	Harvesting and drying	37
4.2.5	Parameter evaluation	37
	4.2.5.1 Plant height	37
	4.2.5.2 Fresh and dry weights	37
4.2.6	Chemical analysis for Rosmarinic Acid and Sinensetin	37
	4.2.6.1 Extraction	37
	4.2.6.2 Total phenolic content determination	38
	4.2.6.3 Total flavonoids content determination	38
	4.2.6.4 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging capacity	38
	4.2.6.5 High-performance liquid chromatography (HPLC) Analysis for Rosmarinic Acid and Sinensetin	39
4.2.7	Nutrient Analysis (N, P, K)	39
4.3	Results and Discussion	39
4.3.1	Plant height, fresh and dry weights	39
4.3.2	Antioxidant compounds and activity	44
4.3.3	Chemical markers	47
4.3.4	Plant nutrient uptake (nitrogen, phosphorous and	50

	potassium)	
4.3.5	Correlation coefficients between plant nitrogen, phosphorous, potassium, dry weight, total phenolic contents, total flavonoid contents, 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical scavenging capacity, rosmarinic acid, sinensetin and nitrogen content	51
4.3.6	Phytochemicals and plant nitrogen uptake based on biomass yield and fertilizer rates	54
4.4	Conclusions	55
5	SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	57
	REFERENCES	59
	APPENDICES	72
	BIODATA OF STUDENT	79
	LIST OF PULICATIONS	80

LIST OF TABLES

Table	Page	
3.1	Characteristics of chicken manure and rice husk biochar.	15
3.2	Gradient elution program used in separation of sinensetin.	18
3.3	Main and interaction effects of rice husk biochar rates, chicken manure and harvesting week on plant height, and fresh and dry weights and pH of <i>Orthosiphon stamineus</i> .	19
3.4	Response of <i>Orthosiphon stamineus</i> to treatments on plant nitrogen (N), phosphorous (P) and potassium (K) uptake at week 8 after transplanting.	24
3.5	Characteristics of soil before and after treatment.	25
3.6	Main and interaction effects of three rice husk biochar rates, three chicken manure rates and three harvesting weeks on total phenolic content, total flavonoid contents and DPPH free radical scavenging activity of <i>Orthosiphon stamineus</i> .	26
3.7	Correlation coefficients (r) between fresh weight, dry weight, plant height, soil pH, total phenolic contents (TPC), total flavonoid contents (TFC), 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical scavenging capacity sinensetin content of <i>Orthosiphon stamineus</i> grown in three rice husk biochar rates (0, 5 and 10 t/ha), three chicken manure rates (0, 2.5 and 5 t/ha) and at three harvesting weeks (4, 6 and 8 weeks after transplanting).	31
3.8	Main and interaction effects of biochar, chicken manure and harvesting time (4, 6 and 8 weeks after transplanting) on sinensetin (% w/w).	32
3.9	Effect of harvesting week on dry weight biomass and yield of sinensetin, total phenolic, total flavonoid, 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical scavenging activity and uptake of plant nitrogen of <i>Orthosiphon stamineus</i> .	33
4.1	Characteristics of organic plant-based and animal-based fertilizer.	36
4.2	Gradient elution program used in separation of sinensetin and rosmarinic acid.	39
4.3	Main and interaction effects of two fertilizer types and five fertilizer rates on fresh and dry weights, and plant height of <i>Orthosiphon stamineus</i> .	40
4.4	Main and interaction effects of two fertilizer types and five	44

fertilizer rates on total phenolic content, total flavonoid content and 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical scavenging activity of *Orthosiphon stamineus*.

- | | | |
|-----|---|----|
| 4.5 | Main and interaction effects of two fertilizer types and five fertilizer rates on rosmarinic acid (% w/w) and sinensetin (% w/w) of <i>Orthosiphon stamineus</i> . | 48 |
| 4.6 | Main and interaction effects of two fertilizer types and five fertilizer rates on plant nitrogen, phosphorous and potassium uptake of <i>Orthosiphon stamineus</i> . | 51 |
| 4.7 | Correlation coefficient (R) between plant nitrogen, phosphorous, potassium, fresh weight, dry weight, total phenolic contents (TPC), total flavonoid contents (TFC), 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical scavenging activity, rosmarinic acid, sinensetin and nitrogen content. | 53 |
| 4.8 | Effect of fertilizer rates on dry weight biomass and yield of sinensetin, rosmarinic acid, total phenolic, total flavonoid and uptake of plant nitrogen of <i>Orthosiphon stamineus</i> . | 55 |

LIST OF FIGURES

Figure		Page
2.1	Leaf morphology of white variety: rhomboid shape with acute apex and green veination on the adaxial (left) and abaxial surface (right).	6
2.2	<i>Orthosiphon stamineus</i> flower. A (calyx), B (corolla), C (stamen).	7
2.3	Developmental stages of <i>Orthosiphon stamineus</i> flowers. (A) Bottom flowers were fully opened, petals and sepals still had the same colour (white and green respectively); (B) Early unopened buds.	7
3.1	Relationship between harvesting week and plant height of <i>Orthosiphon stamineus</i> . Vertical bars = SE. The solid line indicates a significant quadratic regression trend at $P = 0.05$, ($n = 12$).	20
3.2	Interaction effects between harvesting week and three rates of rice husk biochar (BC) on fresh weights of <i>Orthosiphon stamineus</i> produced under produced under three rates of chicken manure (CM). Vertical line indicates pooled LSD at $P = 0.05$. $n = 12$. Soil pH at 0, 4, 6 and 8 weeks after transplanting were 4.59, 5.05, 5.15 and 5.16, respectively.	22
3.3	Relationship between harvesting week and dry weight and of <i>Orthosiphon stamineus</i> produced at different rice husk biochar rates (BC). Vertical bars = SE. Solid line indicates a significant linear regression trend at $P=0.05$, ($n=12$).	23
3.4	Interaction effects of harvesting week and rice husk biochar (BC) on total phenolic content of <i>Orthosiphon stamineus</i> produced under three rates of chicken manure (CM). Vertical line indicates pooled LSD at $P=0.05$, ($n=12$).	27
3.5	Interaction effects harvesting week and rice husk biochar (BC) on total flavonoid content of <i>Orthosiphon stamineus</i> produced under three rates of chicken manure (CM). Vertical line indicates pooled LSD at $P=0.05$. $n=12$.	29
3.6	Relationship between harvesting week and DPPH free radical scavenging activity and of <i>Orthosiphon stamineus</i> produced at three different rice husk biochar rates. Vertical bars = SE. Solid line indicates a significant quadratic regression trend at $P=0.05$.	30
4.1	Relationship between fertilizer rates and fresh weight of <i>Orthosiphon stamineus</i> . Vertical bars = SE. The solid line	41

indicates a significant quadratic regression trend at $P = 0.05$. $n = 12$. Blue line with arrows indicate maximum yield of 48 g/plant could be obtained at optimum fertilizer rate of 323 kg/ha N.

- 4.2 Relationship between fertilizer rates and dry weight of *Orthosiphon stamineus*. Vertical bars = SE. The solid line indicates a significant quadratic regression trend at $P=0.05$. $n = 12$. Blue line with arrows indicate maximum yield of 6.05 g/plant could be obtained at optimum fertilizer rate of 219 kg/ha N. 42
- 4.3 Plant height in the form of $y = A/(1+be^{-cx})$ of *Orthosiphon stamineus* during 56 days growing duration. 43
- 4.4 Growth rate of plant height in the form of $dy/dx = (Abce^{-cx})/(1+be^{-cx})^2$ of *Orthosiphon stamineus* during 56 days growing duration. 43
- 4.5 Relationship between fertilizer rates and total flavonoid content of *Orthosiphon stamineus*. Vertical bars = SE. The solid line indicates a significant quadratic regression trend at $P=0.05$, $n = 12$. Blue line with arrows indicate maximum total flavonoid contents of 25 mg QE/g DW could be obtained at optimum fertilizer rate of 244 kg/ha N. 45
- 4.6 Relationships between fertilizer rates and 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical scavenging activity of *Orthosiphon stamineus*. Vertical bars = SE. The solid line indicates a significant quadratic and linear regression trend at $P=0.05$, $n = 12$. Blue line with arrows indicate maximum DPPH free radical scavenging activity of 72.20% inhibition could be obtained at optimum fertilizer rate of 218 kg/ha N. 46
- 4.7 Relationship between fertilizer rates and rosmarinic acid of *Orthosiphon stamineus* produced under animal- and plant-based fertilizers. Vertical bars = SE. The solid lines indicate a significant regression trend at $P=0.05$, ($n = 12$). 48
- 4.8 Relationship between fertilizer rates and sinensetin of *Orthosiphon stamineus* produced under animal- and plant-based fertilizers. Vertical bars = SE. The solid line indicate a significant regression trend at $P=0.05$, ($n = 12$). 49
- 4.9 HPLC chromatogram of reference markers, rosmarinic acid and sinensetin of *Orthosiphon stamineus* plant extracts. 49
- 4.10 HPLC chromatogram of aqueous ethanol (70%) extract of *Orthosiphon stamineus* plant extracts. 50

LIST OF APPENDICES

Appendix		Page
1	One way ANOVA table showing the significant interaction effects of three rice husk biochar rates and two chicken manure rates and three harvesting week on plant height.	72
2	One way ANOVA table showing the significant interaction effects of three rice husk biochar rates and two chicken manure rates and three harvesting week on fresh weight.	72
3	One way ANOVA table showing the significant interaction effects of three rice husk biochar rates and two chicken manure rates and three harvesting week on dry weight.	73
4	One way ANOVA table showing the significant interaction effects of three rice husk biochar rates and two chicken manure rates and three harvesting week on pH.	73
5	One way ANOVA table showing the significant interaction effects of three rice husk biochar rates and two chicken manure rates and three harvesting week on total phenolic content.	74
6	One way ANOVA table showing the significant interaction effects of three rice husk biochar rates and two chicken manure rates and three harvesting week on total flavonoid content.	74
7	One way ANOVA table showing the significant interaction effects of three rice husk biochar rates and two chicken manure rates and three harvesting week on DPPH free radical scavenging activity.	75
8	One way ANOVA table showing the significant interaction effects of three rice husk biochar rates and two chicken manure rates and three harvesting week on sinensetin.	75
9	One way ANOVA table showing the significant interaction effects of two fertilizer types and five fertilizer rates on plant fresh weight.	75
10	One way ANOVA table showing the significant interaction effects of two fertilizer types and five fertilizer rates on plant dry weight.	76
11	One way ANOVA table showing the significant interaction effects of two fertilizer types and five fertilizer rates on plant height.	76

12	One way ANOVA table showing the significant interaction effects of two fertilizer types and five fertilizer rates on total phenolic contents.	76
13	One way ANOVA table showing the significant interaction effects of two fertilizer types and five fertilizer rates on total flavonoid contents.	77
14	One way ANOVA table showing the significant interaction effects of two fertilizer types and five fertilizer rates on DPPH free radical scavenging activity.	77
15	One way ANOVA table showing the significant interaction effects of two fertilizer types and five fertilizer rates on sinensetin.	77
16	One way ANOVA table showing the significant interaction effects of two fertilizer types and five fertilizer rates on rosmarinic acid.	78
17	One way ANOVA table showing the significant interaction effects of two fertilizer types and five fertilizer rates on plant nitrogen content.	78

LIST OF ABBREVIATIONS

TPC	Total phenolic contents
TFC	Total flavonoid contents
DPPH	2,2-diphenyl-1-picrylhydrazyl
w/w	Weight over weight
WHO	World Health Organization
RA	Rosmarinic acid
SEN	Sinensetin
BC	Rice husk biochar
t/ha	Tonnes per hectare
cm	Centimeter
kg	Kilogram
g	Gram
RCBD	Randomized complete block design
kg/ha	Kilogram per hectare
ANOVA	Analysis of variance
DMRT	Duncan multiple range test
LSD	Least significant differences
°C	Degree celsius
g/plant	Gram per plant
ml	Milliliter
FC	Folin-Ciocalteu
µl	Microliter
Na ₂ CO ₃	Sodium carbonate
nm	Nanometer
GAE	Gallic acid equivalent
NaNO ₃	Sodium nitrate
AlCl ₃	Aluminium chloride
NaOH	Sodium hydroxide
QE	Quecetin equivalent
µm	Micrometer
mm	Milimeter
kg/plant	Kilogram per plant
cm/plant	Centimeter per plant
mg	Miligram
g	Gram
DW	Dry weight
N	Nitrogen
CM	Chicken manure
NKEA	National Key Economic Area
PMF	Polymethoxylated flavone
IFOAM	International Federation of Organic Agriculture Movement
DOA	Department of Agriculture
mg/kg	miligram per kilogram
ALP	Alkaline phosphatase
AST	Aspartate aminotransferase
g/kg	gram per kilogram
USDA	United States Department of Agriculture
DHPL	3,4-dihydroxyphenyllactic acid
CEC	Cation exchange capacity

CHAPTER 1

GENERAL INTRODUCTION

1.1 Introduction

Orthosiphon stamineus has been identified as one of the high value product under the National Key Economic Area (NKEA) introduced by the Ministry of Agriculture and Agro-Based Industry (Anon, 2015). This is due to the high antioxidant compounds such as flavones, polyphenols, bioactive proteins and glycosides in the plant which gives the plant its diuretic, hepatoprotective, antifungal, antimicrobial and antidiabetic roles in human health (Hossain et al., 2008; Adam et al., 2009; Alshawsh et al., 2011; Alshawsh et al., 2012; Elsnoussi et al., 2012). However, common agricultural practices were unable to produce optimum antioxidant compounds in regards to maximum yield. This is because the herb producers' focus is on the amount of yield that they can produce in order to meet the demands from the industry. However, the amount of yield produced does not quantify the amount of antioxidant compounds in the plant. Thus, it is important to find the most suitable agricultural practices that can provide optimum yield and also antioxidant properties.

The two main chemical markers which make *O. stamineus* a valuable medicinal herb are rosmarinic acid (RA) and sinensetin (SEN). Rosmarinic acid, a phenolic acid present in plants as a secondary metabolite, is a water-soluble ester of caffeic acid and 3,4-dihydroxyphenyllactic acid, which can be found naturally in many plants, especially in the species of Boraginaceae and Lamiaceae (Petersen et al., 2003; Petersen, 2013). Rosmarinic acid has been reported to be an antioxidant, possess anti-inflammatory, antiapoptotic, antifibrotic, antimicrobial activity, neuroprotective and neurorescue effect (Jordan et al., 2012; Wang et al., 2012; Domitroric et al., 2013; Braidy et al., 2014). Sinensetin, a flavonoid is a rare polymethoxylated flavone (PMF) and is mostly found in citrus plants with 5-methoxy groups on the basic benzo- γ -pyrone skeleton with a carbonyl group at the C₄ position. Sinensetin has been reported to show anti-inflammatory activity, antioxidant potential and antiproliferative activity (Du and Chen, 2010; Jiang et al., 2012; Shin et al., 2012).

However, the amount of chemical compounds produced by herbs is irregular and insufficient due to poor soil condition and insufficient fertilization lack of water supply, irradiation and temperature (Hansen and Wold, 2010). Thus, it is often altered structurally to produce drugs that are potentially more active. Traditional medicine usages have been on the rise since the 1990s in many developed and developing countries (WHO, 2015). The reasons for this rise are due to lower costs of herbs as compared to synthetic drugs, the need for alternative treatments for drug-resistant pathogens and increasing popularity for products that are natural and environmental friendly (Bandaranayake, 2006). Thus, organic farming is the most likely answer to cultivating herbs for medicinal purposes. IFOAM (2008) defines organic agriculture as a production system that takes into consideration the health of soils, ecosystems and people, which discourage the use of inputs with adverse effects but relies on ecological processes, biodiversity and cycles adapted to local conditions. Chin et al. (2010)

reported that organic farming of tea plantation helps in increasing tea leaf growth by 23% and also major polyphenols by 38%. Organic inputs such as organic fertilizers and bio-organic fertilizers also showed an increase in fresh yield (20%), total phenolic content (45%) and total flavonoid content (51%) in sweet fennel (Salama et al., 2015). Organic fertilization also helps in the improvement of biomass yield and nutrient absorption in herbs at different levels of organic nitrogen fertilizers (Jennifer et al., 2015; Marcio et al., 2015).

The main step in organic farming is to ensure that the soil is healthy to be planted on. However, soils in Malaysia are lacking in organic matter and thus, soil amendments are needed to improve the soil characteristics. Soil amendments are defined as materials that are applied to the soil in order to correct major constraint other than low nutrient content in the soil (Anon, 2010). Organic amendments are used due to their organic matter content which acts as a carrier of utilizable energy and nutrients for the soil organisms. They also help to improve soil structure and porosity, increase water holding capacity of soils, improve aeration, reduce soil temperature fluctuations, storage of nutrients in exchangeable form and as provision of nutrients (Anon, 2010). Rice husk biochar and chicken manure are some examples of soil amendment, which can be used in agriculture. Both are used as a soil amendment because they are not only easily available and cheap, but also help to reduce heavy metals availability in the soil by increasing the soil pH (Masulili et al., 2010).

Besides organic farming, harvesting date or time is also crucial in ensuring the optimum biomass weight, phytochemical and antioxidant activities are achieved. Generally, crops that are for export purposes are usually harvested earlier as compared for consumption as fresh products or for local use, whereby quality attributes such as colour meets consumer preferences. Studies have been done on the impact of harvesting time on the phytochemical content of various crops. Lata et al. (2005) found that blueberries harvested in August had a lower content of phenolics (5%), flavonols (40%) and anthocyanins (2%) than those harvested in July. In another study in Hungary, the total phenolics in broccoli increased by 0.8% and antioxidant activity decreased by 21% when harvested during fall as compared to spring season (Pek et al., 2012). Lv et al. (2016) reported a decrease in oleanolic acid by 2% from the first to the third harvest in the year 2012 and the same trend can be seen in the year 2013 with a decrease of 13%. However, ursolic acid was inconsistent throughout the three harvests in the year 2012, but in the year 2013, a reduction of 9% was reported. These studies showed inconsistent results of phytochemicals and antioxidant activities in crops at different harvesting time. Harvesting time in most crops are planned in order to obtain the optimum levels of health-promoting compounds and thus, it is important to determine the suitable harvesting time in a particular crop.

Despite the high demand for quality and safe *O. stamineus* that are produced organically, not much research has been done to look at the effect of organic planting of *O. stamineus* on the yield, quality and safety. Therefore, the objectives for this study were to study the effects of (i) different types and rates of soil amendments for optimum yield, antioxidant compounds and activities, and chemical markers of *O. stamineus* and (ii) different types and rates of organic fertilizer for optimum yield, antioxidant compounds and activities, and chemical markers of *O. stamineus*. The

hypothesis of the study are (i) combination of rice husk biochar and chicken manure soil amendment at 5 t/ha and 2.5 t/ha, respectively will provide the optimum yield. (ii) harvesting week 8 will produce the highest fresh and dry weights. (iii) plant-based organic fertilizer at 300 kg/ha N will give optimum yield.



REFERENCES

- Abdullah, S., Ahmad, M. S., Shaari, A. R., Johar, H. M. and Mohd Noor, N. F. (2011). Drying characteristics and herbal metabolites composition of misai kucing (*Orthosiphon stamineus* Benth.) leaves. *International Conference on Food Engineering and Biotechnology IPCBEE*, 9, 305-309.
- Dardak, R., Abidin, A. Z. Z. and Ali, A. K. (2009). Consumers' perceptions, consumption and preference on organic product: Malaysian perspective. *Economic and Technology Management Review*, 4, 95 – 107.
- Adam, Y., Somchit, M. N., Sulaiman, M. R., Nasaruddin, A. A., Zuraini, A., Bustamam, A. A. and Z.A., Z. (2009). Diuretic properties of *Orthosiphon stamineus* Benth. *Journal of Ethnopharmacology*, 124, 154 – 158.
- Affendy, H. (2010). *Misai kucing: panduan lengkap jana pendapatan*. Shah Alam, Malaysia: Alaf 21 Sdn. Bhd.
- Affendy, H., Aminuddin, M., Azmy, M., Amizi, M. A., Assis, K. and Tamer, A. T. (2011). Effect of organic fertilizers application to the growth of *Orthosiphon stamineus* Benth. intercropped with *Hevea brasiliensis* Willd. and *Durio zibenthinus* Murr. . *International Journal of Agricultural Research*, 6, 180-187.
- Agyenim, B. S., Zickermann, J. and Kornahrens, M. (2006). Poultry manure effect on growth and yield of maize. *West Africa Journal of Applied Ecology*, 9, 1-11.
- Akowuah, G. A., Ismail, I., Norhayati, I. and Sadikun, A. (2005). The effects of different extraction solvents of varying polarities on polyphenols of *Orthosiphon stamineus* and evaluation of the free radical-scavenging activity. *Food Chemistry*, 93, 311-317.
- Akowuah, G. A. and Zhari, I. (2010). Effect of extraction temperature on stability of major polyphenols and antioxidant activity of *Orthosiphon stamineus* leaf. . *Journal of Herbs, Spices and Medicinal Plant*, 16, 160-166.
- Akowuah, G. A., Zhari, I., Norhayati, I., Sadikun, A. and Khamsah, S. M. (2004). Sinensetin, eupatorin, 3'-hydroxy-5,6,7,4'-tetramethoxyflavone and rosmarinic acid contents and antioxidative effect of *Orthosiphon stamineus* from Malaysia. *Food Chemistry*, 87, 559 – 566.
- Alizadeh, A., Khoshkhui, M., Javidnia, K., Firuzi, O., Tafazoli, E. and Khalighi, A. (2010). Effects of fertilizer on yield, essential oil composition, total phenolic content and antioxidant activity in *Satureja hortensis* L. (Lamiaceae) cultivated in Iran. *Journal of Medicinal Plants Research*, 4(1), 33-40.
- Almatar, M., Rahmat, Z. and Mohd Salleh, F. (2013). Preliminary morphological and anatomical study of *Orthosiphon stamineus*. *Indian Journal Pharmaceutical Biological Research*, 1, 1-6.

- Alphonse, L. M. J. and Nithiya, T. (2015). Effect of organic and inorganic fertilizer on growth, phenolic compounds and antioxidant activity of *Solanum nigrum* L. *World Journal of Pharmacy and Pharmaceutical Sciences*, 5, 808 – 822.
- Alshawsh, M. A., Abdulla, M. A., Ismail, S., Amin, Z. A., Qader, S. W., Hadi, H. A. and Harmal, N. S. (2012). Free radical scavenging, antimicrobial and immunomodulatory activities of *Orthosiphon stamineus*. *Molecules*, 17, 5385 – 5395.
- Alshawsh, M. A., Mahmood, A. A., Ismail, S. and Amin, Z. A. (2011). Hepatoprotective effects of *Orthosiphon stamineus* extract on thioacetamide-induced liver cirrhosis in rats. *Evidence-Based Complementary and Alternative Medicine*, 2011, 1-6.
- Aminah, A. and Anna, P. K. (2011). Influence of ripening stages on physicochemical characteristics and antioxidant properties of bitter melon (*Momordica charantia*). *International Food Research Journal*, 18(3), 895 – 900.
- Angelova, V. R., Akova, V. I., Artinova, N. S. and Ivanov, K. I. (2013). The effect of organic amendments on soil chemical characteristics. *Bulgarian Journal of Agricultural Science*, 19 (5), 958-971.
- Antonious, G. F., Kochhar, T. S. and Coolong, T. (2012). Yield, quality and concentration of seven heavy metals in cabbage and broccoli grown in sewage sludge and chicken manure amended soil. *Journal of Environmental Science and Health*, 47, 1955-1965.
- Antonious, G. F., Turley, E. T., Hill, R. R. and Snyder, J. C. (2014). Chicken manure enhanced yield and quality of field-grown kale and collard greens. *Journal of Environmental Science and Health*, 49, 299 – 304.
- Anonymous. (2010). Database on commercially available organic fertilizers and water-retaining products. Retrieved 1 December, 2015, from <http://www.fao.org/ag/agp/orgfert/intro.htm>
- Anonymous. (2014). Biochar introduction. Retrieved 13 September 2015, from <http://biochar-us.org/biochar-introduction>
- Anonymous. (2015). Malaysia Broiler Meat (Poultry) Production Annual Growth Rate. Retrieved 4 July 2016, from <http://www.indexmundi.com/agriculture/?country=my&commodity=broiler-meat&graph=production-growth-rate>
- Babalar, M., Mumivand, H., Hadian, J. and Tabatabaei, S. M. F. (2010). Effects of nitrogen and calcium carbonate on growth, rosmarinic acid content and yield of *Satureja hortensis* L. *Journal of Agricultural Science*, 2(3), 92-98.
- Bandaranayake, W. M. (2006). Quality control, screening, toxicity and regulation of herbal drugs. In I. Ahmad, F. Aqil and M. Owais (Eds.), *Modern*

phytomedicine. Turning medicinal plants into drugs (pp. 404). Weinheim: Wiley.

- Benard, C., Gautier, H., Bourgaud, F., Grasselly, D., Navez, B., Caris-Veyrat, C., Wiess, M. and Genard, M. (2009). Effects of low nitrogen supply on tomato (*Solanum lycopersicum*) fruit yield and quality with special emphasis on sugars, acids, ascorbate, carotenoids, and phenolic compounds. *Journal of Agricultural and Food Chemistry*, 57(10), 4112 - 4123.
- Bi, G., Evans, W. B., Spiers, J. M. and Witcher, A. L. (2010). Effects of organic and inorganic fertilizers on marigold growth and flowering. *HortScience*, 45(9), 1373 – 1377.
- Boroujerdnia, M. and Ansari, N. A. (2007). Effect of different levels of nitrogen fertilizer and cultivars on growth, yield and yield components of Romaine lettuce (*Lactuca sativa* L.). *Middle Eastern and Russian Journal of Plant Science and Biotechnology*, 1(2), 47 – 53.
- Bouajila, K. and Sanaa, M. (2011). Effects of organic amendments on soil physico-chemical and biological properties. *J. Mater. Environ. Sci.*, 2 (S1), 485-490.
- Braidy, N., Matin, A., Rossi, F., Chinain, M., Laurent, D. and Guillemin, G. J. (2014). Neuroprotective effects of rosmarinic acid on ciguatoxin in primary human neurons. *Neurotoxicity Research*, 25, 226 – 234.
- Bruno, B. C., Hubert, O., Mbeguie-A-Mbeguie, D., Pallet, D., Hiol, A., Reynes, M. and Poucheret, P. (2013). Effect of physiological harvest stages on the composition of bioactive compounds in Cavendish bananas. *Journal of Zhejiang University Science B.*, 14(4), 270 – 278.
- Bruno, O. D., Carlos, A. S., Fabio, S. H., Asuncion, R. and Miguel, A. S. (2010). Use of biochar as bulking agent for the composting of poultry manure: effect on organic matter degradation and humification. *Bioresource Technology*, 101, 1239 – 1246.
- Card, A., Whiting, D., Wilson, C., Reeder, J. and Goldhamer, D. (2015). Organic fertilizers. Retrieved from <http://www.cmg.colostate.edu/index.shtml>
- Cardarelli, M., Roupheal, Y., Rea, E., Lucini, L., Pellizzoni, M. and Colla, G. (2013). Effects of fertilization, arbuscular mycorrhiza, and salinity on growth, yield, and bioactive compounds of two *Aloe* species. *HortScience*, 48(5), 568 – 575.
- Chan, L. K. and Loo, P. S. (2006). Morphological similarities and differences between the two varieties of cat's whiskers (*Orthosiphon stamineus* Benth.) grown in Malaysia. *International Journal of Botany*, 2, 1-6.
- Chew, K. K., Khoo, M. Z., Ng, S. Y., Thoo, Y. Y., Wan Aida, W. M. and Ho, C. W. (2011). Effect of ethanol concentration, extraction time and extraction temperature on the recovery of phenolic compounds and antioxidant capacity

- of *Orthosiphon stamineus* extract. *International Food Research Journal*, 18(4), 1427 – 1435.
- Chin, F. S., Ho, T. Y., Chong, K. P., Mohamadu, B. J. and Nyet, K. W. (2010). Organic versus conventional farming of tea plantation. *Borneo Science*, 26, 19-26.
- Chin, J. H., Abas, H. H. and Sabariah, I. (2008). Toxicity study of *Orthosiphon stamineus* Benth (Misai Kucing) on Sprague Dawley rats. . *Tropical Biomedicine*, 25, 9-16.
- Colling, J., Stander, M. A. and Makunga, N. P. (2010). Nitrogen supply and abiotic stress influence canavanine synthesis and the productivity of in vitro regenerated *Sutherlandia frutescens* microshoots. *Journal of Plant Physiology*, 167, 1521-1524.
- Cooperband, L. (2002). Building soil organic matter with organic amendments (pp. 6). Wisconsin, United States: University of Wisconsin-Madison.
- Dai, Z., Meng, J., Muhammad, N., Liu, X., Wang, H., He, Y., Brookes, P. C. and Xu, J. (2013). The potential feasibility for soil improvement, based on the properties of biocharspyrolyzed from different feedstocks. *Journal Soil Sediments*, 13, 989 – 1000.
- Davis, J. G. and Whiting, D. (2013). Choosing a soil admendment. *Gardening series* Retrieved 13 September, 2015, from <http://extension.colostate.edu/docs/pubs/garden/07235.pdf>
- Dikinya, O. and Mufwanzala, N. (2010). Chickn manure-enhanced soil fertility and productivity: effects of application rates. *Journal of Soil Science and Environmental Management*, 1(3), 46 – 54.
- DOA. (2013). *Herbs and Spices Statistics*. Putrajaya, Malaysia: Department of Agriculture Malaysia.
- Doughari, J. H. (2014). *Phytochemicals: Extraction Methods, Basic Structures and Mode of Action as Potential Chemotherapeutic Agents*: INTECH Open Access Publisher.
- Du, Q. and Chen, H. (2010). The methoxyflavones in *Citrus reticulata* Blanco cv. Ponkan and their antiproliferative activity against cancer cells. *Food Chemistry*, 119, 567 – 572.
- Eaton, T. E., Cox, D. A. and Barker, A. V. (2013). Sustainable production of marigold and calibrachoa with organic fertilizers. *HortScience*, 48(5), 637 – 644.
- Eghdami, A. and Sadeghi, F. (2010). Determination of total phenolic and flavonoids contents in methanolic and aqueous extract of *Achillea millefolium*. *Organic Chemistry Journal*, 2, 81-84.

- Elsnoussi, A. H. M., Mohammad, J. A. S., Lee, F. A., Amirin, S., Sue, H. C., Soo, C. T., Mohd, Z. A. and Mun, F. Y. (2012). Potent α -glucosidase and α -amylase inhibitory activities of standardized 50% ethanolic extracts and sinensetin from *Orthosiphon stamineus* Benth as anti-diabetic mechanism. *BMC Complementary and Alternative Medicine*, 12, 176.
- Enders, A., Hanley, K., Whitman, T., Joseph, S. and Lehmann, J. (2012). Characterization of biochars to evaluate recalcitrance and agronomic performance. *Bioresource Technology*, 114, 644-653.
- Ezz El-Din, A. A., Hendawy, S. F., Aziz, E. E. and Omer, E. A. (2010). Enhancing growth, yield and essential oil of caraway plants by nitrogen and potassium fertilizers. *International Journal of Academic Research*, 2, 192-197.
- Mohd Farhan, M., Abdul Razak, S., Pin, K. Y. and Chuah, A. L. (2012). Antioxidant activity and phenolic content of different parts of *Orthosiphon stamineus* grown under different light intensities. *Journal of Tropical Forest Science*, 24(2), 173-177.
- Foad, S. R. A., Elham, F., Mohamed, B. K. A., Ismail, Z., Aman, S. A. M. and Amin, M. S. A. M. (2012). Marked antitumor activity of cat's whiskers tea (*Orthosiphon stamineus*) extract in orthotopic model of human colon tumor in nude mice. *Journal of Biochemical Technology*, 3, 170-176.
- Galende, M. A., Becerril, J. M., Barrutia, O., Artetxe, U., Garbisu, C. and Hernandez, A. (2014a). Field assessment of the effectiveness of organic amendments for aided phytostabilization of a Pb-Zn contaminated mine soil. *Journal of Geochemical Exploration*, 145, 181 – 189.
- Galende, M. A., Becerril, J. M., Gomez-Sagasti, M. T., Barrutia, O., Epelde, L., Garbisu, C. and Hernandez, A. (2014b). Chemical stabilization of metal-contaminated mine soil: early short-term soil-amendment interactions and their effects on biological and chemical parameters. *Water Air Soil Pollut*, 225, 1863.
- Gemma, C., Mesches, M. H., Sepesi, B., Choo, K., Holmes, D. B. and Bickford, P. C. (2002). Diets enriched in foods with high antioxidant activity reverse age-induced decreases in cerebellar β -adrenergic function and increases in proinflammatory cytokines. *The Journal of Neuroscience*, 22, 6114-6120.
- Giuffre, L., Giardina, E., Ciarlo, E., Rios, P. and Vella, L. (2015). A study on the effect of soil amendments and environmental conditions of *Stevia rebaudiana* in urban soils of Buenos Aires, Argentina. *Current Agriculture Research Journal*, 3(1), 7-13.
- Giaccio, G., De Sensale, G. R. and Zerbino, R. (2007). Failure mechanism of normal and high-strength concrete with rice-husk ash. *Cement and Concrete Composites*, 29, 566 – 574.

- Glab, T. and Gondek, K. (2008). Effect of organic amendments on morphometric properties of macropores in Stagnic Gleysol soil. *Polish J. of Environ. Stud.* 17(2), 209-214.
- Globinmed. (2015). *Orthosiphon stamineus*. Retrieved 30 August 2015 http://www.globinmed.com/index.php?option=com_content&view=article&id=79215:orthosiphon-stamineus&Itemid=139
- Hagos, H., Mengistu, L. and Mequanint, Y. (2014). Determining optimum harvest age of sugarcane varieties on the newly establishing sugar project in the tropical areas of Tendaho, Ethiopia. *Advances in Crop Science and Technology*, 2, 1 - 4.
- Hansen, M. and Wold, A. (2010). Contents of bioactive compounds in food plants as affected by traditional breeding and environmental factors. *Bioactive compounds in plants-benefits and risks for man and animals*, 212 - 222.
- Hassan, A., Mohamad, A., Abdu, A., Md Idrus, R. and Awang Besar, N. (2010). *Soil properties under Orthosiphon stamineus (Benth) intercropped with Duriozi benthus (Murr) and treated with various organic fertilizers*. Paper presented at the 19th World Congress of Soil Science, Soil Solutions for a Changing World, Brisbane, Australia.
- Himani, B., Seema, B., Bhole, N., Mayank, Y., Vinod, S. and Mamta, S. (2013). Misai kucing: a glimpse of maestro. *International Journal of Pharmaceutical Sciences Review and Research*, 22, 55-59.
- Hochmuth, G., Hochmuth, R. and Mylawarapu, R. (2009). Using composted poultry manure (litter) in mulched vegetable production. Retrieved from <http://edis.ifas.ufl.edu/ss506>
- Hossain, M. A., Ismail, Z., Rahman, A. and Sun, C. K. (2008). Chemical composition and anti-fungal properties of the essential oils and crude extracts of *Orthosiphon stamineus* Benth. *Industrial Crops and Products*, 27, 328 – 334.
- Hu, J., Lin, X., Wang, J., Dai, J., Chen, R., Zhang, J. and Wong, M. H. (2011). Microbial functional diversity, metabolic quotient, and invertase activity of a sandy loam soil as affected by long-term application of organic amendment and mineral fertilizer. *Journal Soils Sediments*, 11, 271 – 280.
- Ibrahim, M. H. and Jaafar, H. Z. E. (2011). The relationship of nitrogen and C/N ratio with secondary metabolites levels and antioxidant activities in three varieties of Malaysian kacip Fatimah (*Labisia pumila* Blume). *Molecules*, 16, 5514-5526.
- Ibrahim, M. H., Jaafar, H. Z. E., Karimi, E. and Ghasemzadeh, A. (2013). Impact of organic and inorganic fertilizers application on the phytochemical and antioxidant activity of kacip fatimah (*Labisia pumila* Benth). *Molecules*, 18, 10973 - 10988.

- IFOAM. (2008). Definition of Organic Agriculture Retrieved 18 November, 2015, from <http://www.ifoam.bio/en/organic-landmarks/definition-organic-agriculture>
- Jeffrey, L. S., Harold, P. C. and Vanessa, L. B. (2010). The effect of young biochar on soil respiration. *Soil Biology and Biochemistry*, 42, 2345-2347.
- Jennifer, B., Charles, L. C., Tessema, A., Valtcho, D. Z., Archana, G. and Carmen, S. F. B. (2015). Organic versus conventional fertilization effects on sweet basil (*Ocimum basilicum* L.) growth in a greenhouse system. *Industrial Crops and Products*, 74, 249-254.
- Jiang, Y., Wong, J. H., Fu, M., Ng, T. B., Liu, Z. K., Wang, C. R., Li, N., Qiao, W. T., Wen, T. Y. and Liu, F. (2015). Isolation of adenosine, iso-sinensetin and dimethylguanosine with antioxidant and HIV-1 protease inhibiting activities from fruiting bodies of *Cordyceps militaris*. *Phytomedicine*, 18, 189 – 193.
- Jones, D. L., Rousk, J., Edwards-Jones, G., DeLuca, T. H. and Murphy, D. V. (2012). Biochar-mediated changes in soil quality and plant growth in a three year field trial. *Soil Biology and Biochemistry*, 45, 113 – 124.
- Jordan, M. J., Lax, V., Rota, M. C., Loran, S. and Sotomayor, J. A. (2012). Relevance of carnolic acid, carnosol and rosmarinic acid concentrations in the in vitro antioxidant and antimicrobial activities of *Rosmarinic officinalis* (L.) methanolic extracts. *Journal of Agricultural and Food Chemistry*, 60, 9603 – 9608.
- Kang, M. K. and Juvik, J. A. (2013). Environmental stress and methyl jasmonate-mediated changes in flavonoid concentrations and antioxidant activity in broccoli florets and kale leaf tissues. *HortScience*, 48(8), 996 – 1002.
- Kassa, M. and Sorsa, Z. (2015). Effect of nitrogen and phosphorus fertilizer rates on yield and yield components of barley (*Hordeum vulgare* L.) varieties at Damot Gale district, Wolaita Zone, Ethiopia. *American Journal of Agriculture and Forestry*, 3(6), 271 – 275.
- Kim, D., Jeond, S. and Lee, C. (2003). Antioxidant capacity of phenolic phytochemicals from various cultivars of plums. *Food Chemistry*, 81, 321 – 326.
- Koes, R. E., Quattrocchio, R. and Mol, J. N. M. (1994). The flavonoid biosynthetic pathway in plants: function and evolution. *BioEssays*, 16, 123 – 132.
- Lata, B., Trampczynska, A. and Mike, A. (2005). Effect of cultivar and harvest date on thiols, ascorbate and phenolic compounds content in blueberries. *Acta Sci. Pol., Hortorum Cultus*, 4(1), 163 - 171.
- Laird, D., Fleming, P., Wang, B., Horton, R. and Karlen, D. (2010). Biochar impact on nutrient leaching from Midwestern agricultural soil. *Geoderma*, 158, 436 – 442.

- Lee, J. (2010). Effect of application methods of organic fertilizer on growth, soil chemical properties and microbial densities in organic bulb production. *Scientia Horticulturae*, 124, 299 – 305.
- Lee, P. C., Guo, H. Y., Huang, C. C. and Chan, C. F. (2008). Chemical composition of leaf essential oils of *Syzygium samarangense* (BL.) Merr. et Perry cv. Pink at three maturity stages. *International Journal of Applied Research in Natural Products*, 9(1), 9 – 13.
- Liu, C. W., Sung, Y., Chen, B. C. and Lai, H. Y. (2014). Effects of nitrogen fertilizers on the growth and nitrate content of lettuce (*Lactuca sativa* L.). *International Journal of Environment Reseach and Public Health*, 11(4), 4427 – 4440.
- Liu, L., Chen, H., Peng, C., Liang, W. and Huang, Q. (2009). Immobilization and phytotoxicity of Cd in contaminated soil amended with chicken manure compost. *Journal of Hazardous Materials*, 163, 563-567.
- Liu, Z. and Zhang, F. S. (2009). Removal of lead from water using biochars prepared from hydrothermal liquefaction of biomass. *Journal of Hazardous Materials*, 167, 933 – 939.
- Lopez-Velez, M., Martinez-Martinez, F. and Del Valle-Ribes, C. (2003). The study of phenolic compounds as natural antioxidants in wine. *Critical Reviews in Food Science and Nutrition*, 43, 233 – 244.
- Lu, S. G., Sun, F. F. and Zong, Y. T. (2014). Effect of rice husk biochar and coal fly ash on some physical properties of expansive clayey soil (vertisol). *Catena*, 114, 37-44.
- Lv, Y., Tahir, I. I. and Olsson, M. E. (2016). Ursolic and oleanolic acid in 'aroma' apple peel as affected by rootstock, harvest maturity, and storage method. *HortScience*, 51(4), 349 - 355.
- Ma, Z., Li, S. and Zhang, M. (2010). Light intensity affects growth, photosynthetic capability, and total flavonoid accumulation of *Anoectochilus* plants. *HortScience*, 45(6), 863 – 867.
- Marais, J. P. J., Deavours, B., Dixon, R. A. and Ferreira, D. (2006). The stereochemistry of flavonoids. In E. Grotewold (Ed.), *The science of flavonoids*. New York, USA: Springer Science and Business Media.
- Marcio, C., Maira, U., Daniel, V. M., Giuseppina, L. and Lin, C. M. (2015). Effect of organic fertilization on biomass production and bioactive compounds in *Passiflora incarnata* L. *International Journal of Phytocosmetics and Natural Ingredients*, 2, 11.
- Masarirambi, M. T., Hlawe, M. M., Oseni, O. T. and Sibiyi, T. E. (2010). Effects of organic fertilizers on growth, yield, quality and sensory evaluation of red

lettuce (*Lactuca sativa* L.) 'Veneza Roxa'. *Journal of Agriculture and Biology of North America*, 1(6), 1319 – 1324.

- Masayu, H., Zamree, M. S., Pin, K. Y. and Mohd Shahidan, M. A. (2012). *Penghasilan daun teh misai kucing berkualiti tinggi*. Selangor, Malaysia: Institut Penyelidikan Perhutanan Malaysia.
- Masulili, A., Wani, H. O. and Syechfani, M. S. (2010). Rice husk biochar for rice based cropping system in acid soil 1. The characteristics of rice husk biochar and its influence on the properties of acid sulphate soils and rice growth in West Kalimantan, Indonesia. *Journal of Agricultural Science*, 2, 39 – 47.
- Matyssek, R., Schnyder, H., Obwald, W., Ernst, D., Munch, J. C. and Prezsch, H. (2012). *Growth and defence in plants* (Vol. 220). New York: Springer.
- Mediani, A., Abas, F., Khatib, A., Tan, C. P., Ismail, I. S., Shaari, K., Ismail, A. and Lajis, N. H. (2015). Phytochemical and biological features of *Phyllanthus niruri* and *Phyllanthus urinaria* harvested at different growth stages revealed by ¹H NMR-based metabolomics. *Industrial Crops and Products*, 77, 602 – 613.
- Melero-Vara, J. M., López-Herrera, C. J., Prados-Ligero, A. M., Vela-Delgado, M. D., Navas-Becerra, J. A. and Basallote-Ureba, M. J. (2011). Effects of soil amendment with poultry manure on carnation Fusarium wilt in greenhouses in southwest Spain. *Crop Protection*, 30, 970 – 976.
- Milla, O. V., Rivera, E. B., Huang, W. J., Chien, C. C. and Wang, Y. M. (2013). Agronomic properties and characterization of rice husk and wood biochars and their effect on the growth of water spinach in a field test. *Journal of Soil Science and Plant Nutrition*, 13(2), 251 – 266.
- Mohamed, E. A. H., Mohammad, J. A. S., Lee, F. A., Amirin, S., Sue, H. C., Soo, C. T., Mohd Zaini, A. and Mun, F. Y. (2012). Potent α -glucosidase and α -amylase inhibitory activities of standardized 50% ethanolic extracts and sinensetin from *Orthosiphon stamineus* Benth as anti-diabetic mechanism. *BMC Complementary and Alternative Medicine*, 12, 176.
- Nantitanon, W., Yotsawimonwat, S. and Okonogi, S. (2010). Factors influencing antioxidant activities and total phenolic content of guava leaf extract. *LWT – Food Science and Technology*, 43, 1095 – 1103.
- Nguyen, P. M. and Niemeyer, E. D. (2008). Effects of nitrogen fertilization on the phenolic composition and antioxidant properties of basil (*Ocimum basilicum* L.). *Journal of Agricultural and Food Chemistry*, 56, 8685 - 8691.
- Nur Faezah, O., Siti Aishah, H., Umi Kalsom, Y., Nur Ashikin, P. A., Puteri Edaroyati, M. W. and Uma Rani, S. (2012). Phenolic, flavonoids, antioxidant activity and cyanogenic glycosides of organic and mineral-based fertilized cassava tubers. *Molecules*, 17, 2378 – 2387.

- Osakabe, N., Yasuda, A., Natsume, M., Sanbongi, C., Kato, Y., Osawa, T. and Yoshikawa, T. (2002). Rosmarinic acid, a major polyphenolic component of *Perilla frutescens*, reduces lipopolysaccharide (LPS)-induced liver injury in D-galactosamine (D-GalN)- sensitized mice. *Free Radical Biology and Medicine*, 33, 798 – 806.
- Pek, Z., Daood, H., Nagyne, M. G., Berki, M., Tothne, M. M., Nemenyi, A. and Helyes, L. (2012). Yield and phytochemical compounds of broccoli as affected by temperature, irrigation, and foliar sulfur supplementation. *HortScience*, 47(11), 1646 - 1652.
- Pereira, A. P., Ferreira, I. C. F. F., Marcelino, F., Valentao, P., Andrade, P. B., Seabra, R., Estevinho, L., Bento, A. and Pereira, J. A. (2007). Phenolic compounds and antimicrobial activity of olive (*Olea europaea* L. Cv. Cobrançosa) leaves. *Molecules*, 12, 1153 – 1162.
- Petersen, M. (2013). Rosmarinic acid : new aspects. *Phytochemistry Reviews*, 12, 207-227.
- Petersen, M. and Simmonds, M. S. J. (2003). Molecules of interest : Rosmarinic acid. *Phytochemistry*, 62, 121-125.
- Pincemail, J., Kevers, C., Tabart, J., Defraigne, J. O. and Dommes, J. (2012). Cultivars, culture conditions, and harvest time influence phenolic and ascorbic acid contents and antioxidant capacity of strawberry (*Fragaria x ananassa*). *Journal of Food Science*, 77, C205 - C210.
- Rajkovich, S., Enders, A., Hanley, K., Hyland, C., Zimmerman, A. R. and Lehmann, J. (2012). Corn growth and nitrogen nutrition after additions of biochars with varying properties to a temperate soil. *Biology and Fertility of Soils*, 48, 271 – 284.
- Ramesh, K., Manohar, S. and Rajeshkumar, S. (2014). Nephroprotective activity of ethanolic extract of *Orthosiphon stamineus* leaves on ethylene glycol induced urolithiasis in albino rats. *International Journal Of PharmTech Research*, 6, 403-408.
- Rees, F., Simonnot, M. O. and Morel, J. L. (2014). Short-term effects of biochar on soil heavy metal mobility are controlled by intra-particle diffusion and soil pH increase. *European Journal of Soil Science*, 65, 149 – 161.
- Relf, D. and Ball, E. (2009). Propagation by cutting, layering and division. 426(002), 1-6. Retrieved from <https://pubs.ext.vt.edu/426/426-002/426-002.html>
- Roslan, A. F. (2015). *The effect of planting distance and mulching on the growth performance of misai kucing (Orthosiphon stamineus)*. Bachelor Degree, Universiti Putra Malaysia, Selangor, Malaysia.

- Sajap, A. S. and Tan, L. P. (2010). The lace bug *Cochlochila bullita* (Stål) (Heteroptera: Tingidae), a potential pest of *Orthosiphon stamineus* Benthham (Lamiales: Lamiaceae) in Malaysia. *Insecta Mundi*, 136, 1-5.
- Salama, Z. A., El Baz, F. K., Gaafar, A. A. and Zaki, M. F. (2015). Antioxidant activities of phenolics, flavonoids and vitamin C in two cultivars of fennel (*Foeniculum vulgare* Mill.) in responses to organic and bio-organic fertilizers. *Journal of the Saudi Society of Agricultural Sciences*, 14(1), 91-99. doi: 10.1016/j.jssas.2013.10.004
- Sánchez-Campillo, M., Gabaldon, J. A., Castillo, J., Benavente-García, O., Del Bano, M. J., Alcaraz, M., Vicente, V., Alvarez, N. and Lozano, J. A. (2009). Rosmarinic acid, a photo-protective agent against UV and other ionizing radiations. *Food and Chemical Toxicology*, 47, 386 – 392.
- Schijlen, E. G. W. M., Ric de Vos, C. H., Van Tunen, A. J. and Bovy, A. G. (2004). Modification of flavonoid biosynthesis in crop plants. *Phytochemistry*, 65, 2631 – 2648.
- Selma, M. V., Martinez-Sanchez, A., Allende, A., Ros, M., Hernandez, M. T. and Gil, M. I. (2010). Impact of organic soil amendments on phytochemicals and microbial quality of rocket leaves (*Eruca sativa*). *Journal of Agricultural and Food Chemistry*, 58, 8331 – 8337.
- Sharma, S. and Kumar, R. (2012). Effect of nitrogen on growth, biomass and oil composition of clary sage (*Salvia sclarea* Linn.) under mid hills of north western Himalayas. *Journal of Natural Products and Resources*, 3(1), 79-83.
- Shin, H. S., Kang, S. I., Yoon, S. A., Ko, H. C. and Kim, S. J. (2012). Sinensetin attenuates LPS-induced inflammation by regulating the protein level of I κ B- α . *Bioscience, Biotechnology and Biochemistry*, 76, 847 – 849.
- Shukla, S., Mehta, A., Mehta, P. and Bajpai, V. K. (2012). Antioxidant ability and total phenolic content of aqueous leaf extract of *Stevia rebaudiana* Bert. . *Experimental and Toxicologic Pathology*, 64, 807-811.
- Sinkovic, L., Demsar, L., Znidarcic, D., Vidrih, R., Hribar, J. and Treutter, D. (2015). Phenolic profiles in leaves of chicory cultivars (*Cichorium intybus* L.) as influenced by organic and mineral fertilizers. . *Food Chemistry*, 166, 507 – 513.
- Sreelatha, S. and Padma, P. R. (2009). Antioxidant activity and total phenolic content of *Moringa oleifera* leaves in two stages of maturity. *Plant Foods for Human Nutrition*, 64, 303 – 311.
- Srivalli, B. and Khanna-Chopra, R. (2001). Induction of new isoforms of superoxide dismutase and catalase enzymes in the flag leaf of wheat during monocarpic senescence. *Biochemical and Biophysical Research Communications*, 288, 1037 – 1042.

- Stagnari, F., Galieni, A. and Pisante, M. (2015). Shading and nitrogen management affect quality, safety and yield of greenhouse-grown leaf lettuce. *Scientia Horticulturae*, 192, 70 - 79.
- Stewart, A. J., Chapman, W., Jenkins, G. I., Graham, I., Martin, T. and Crozier, A. . (2001). The effect of nitrogen and phosphorus deficiency on flavonol accumulation in plant tissues. *Plant, Cell and Environment*, 24(11), 1189 - 1197.
- Streubel, J. D., Collins, H. P., Gracia-Perez, M., Tarara, J., Granatstein, D. and Kruger, C. E. (2011). Influence of contrasting biochar types on five soils at increasing rates of application. *Soil Science Society of America*, 75(4), 1402 – 1413.
- Taghizadeh-Toosi, A., Clough, T. J., Sherlock, R. R. and Condon, L. M. (2012). Biochar adsorbed ammonia is bioavailable. *Plant Soil*, 350, 57-69.
- Tavarini, S. and Angelini, L. G. (2013). Stevia rebaudiana Bertoni as a source of bioactive compounds: the effect of harvest time, experimental site and crop age on steviol glycoside content and antioxidant properties. *Journal of the Science of Food and Agriculture*, 93(9), 2121 - 2129.
- Uchida, R. (2000). Chapter 3: Essential nutrients for plant growth: nutrient functions and deficiency symptoms. In J. A. Silvia and R. S. Uchida (Eds.), *Plant Nutrient Management in Hawaii's Soils: Approaches for Tropical and Subtropical Agriculture*. Honolulu; University of Hawaii.
- United Nation. (2015). UN Comtrade Database. Retrieved 10 February, 2016, from <http://comtrade.un.org/>
- Vagts, T. (2005). Nitrogen fertilizers and soil pH. Retrieved 12 December, 2015, from https://www.extension.iastate.edu/nwcrops/fertilizer_and_soil_ph.htm
- Varela, M. O., Rivera, E. B., Huang, W. J., Chien, C. C. and Wang, Y. M. (2013). Agronomic properties and characterization of rice husk and wood biochars and their effect on the growth of water spinach in field test. *Journal of Soil Science and Plant Nutrition*, 13(2), 251 – 266.
- Walker, D. J., Clemente, R., Roig, A. and Bernal, M. P. (2003). The effects of soil amendments on heavy metal bioavailability in two contaminated Mediterranean soils. *Environmental Pollution*, 122, 303 – 312.
- Wang, J., Xu, H., Jiang, H., Du, X., Sun, P. and Xie, J. (2012). Neurorescue effect of rosmarinic acid on 6-hydroxydopamine-lesioned nigral dopamine neurons in rat model of parkinson's disease. *Journal of Molecular Neuroscience*, 47, 113 – 119.
- Wang, J., Zhang, M., Z., X., Liu, P. and Pan, G. (2011). Effects of biochar addition on N₂O and CO₂ emissions from two paddy soils. *Biological and Fertility of Soils*, 47, 887 – 896.

- WHO. (2015). Traditional medicine. Retrieved 30 August 2015
http://www.who.int/topics/traditional_medicine/en/
- Widowati, U. W. H., Guritno, B. and Soehono, L. A. (2012). The effect of biochar on the growth and N fertilizer requirement of maize (*Zea mays* L.) in green house experiment. *Journal of Agricultural Science*, 4(5), 255 – 262.
- Winkel-Shirley, B. (2002). Biosynthesis of flavonoids and effects of stress. *Current Opinion in Plant Biology*, 5, 218 – 223.
- Wong, C. C. and Chen, C. P. (1998). Malaysia: Country pasture/forage resources profiles. Retrieved from
<http://www.fao.org/ag/agp/agpc/doc/counprof/malaysia.htm#2.2>
- Xu, C. and Mou, B. (2016). Responses of spinach to salinity and nutrient deficiency in growth, physiology and nutritional value. *Journal of American Society of Horticultural Science*, 141(1), 12-21.
- Yam, M. F., Ang, L. F., Basir, R., Salman, I. M., Ameer, O. Z. and Asmawi, M. Z. (2009). Evaluation of the anti-pyretic potential of *Orthosiphon stamineus* Benth standardized extract. *Inflammopharmacology*, 17, 50-54.
- Yao, Y., Gao, B., Zhang, M., Inyang, M. and Zimmerman, A. R. (2012). Effect of biochar amendment on sorption and leaching of nitrate, ammonium, and phosphate in a sandy soil. *Chemosphere*, 89, 1467 – 1471.
- Zaharah, A. and Salbiah, H. (2005). Populasi tanaman misai kucing di tanah bris. *Buletin Teknologi Tanaman*, 2, 55 – 59.
- Zamani, Z., Zeinali, H., Sinaki, J. M. and Madani, H. (2014). Effect of nitrogen and phosphorous fertilizers on the yield and secondary metabolites of medicinal plant *Rubia tinctorum* L. under salinity conditions. *Iranian Journal of Plant Physiology*, 4(2), 949-955.
- Zhen, M. (2013). Organic food still not taken seriously despite sector's strong potential. Retrieved 10 February, 2016, from
<http://www.businesscircle.com.my/organic-food-still-not-taken-seriously-despite-sectors-strong-potential/>
- Zheng, H., Wang, Z., Deng, X., Herbert, S. and Xing, B. (2013). Impacts of adding biochar on nitrogen retention and bioavailability in agricultural soil. *Geoderma*, 206, 32 – 39.