



UNIVERSITI PUTRA MALAYSIA

***NON-DESTRUCTIVE ESTIMATION OF OIL PALM YIELD AND QUALITY
WITH FLUORESCENCE SENSING***

NORUL HUSNA BINTI ABU KASSIM

FP 2016 12



**NON-DESTRUCTIVE ESTIMATION OF OIL PALM YIELD AND QUALITY
WITH FLUORESCENCE SENSING**

By

NORUL HUSNA BINTI ABU KASSIM

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of Master of
Science**

January 2016



All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

Allhamdulillah, praises to Allah S.W.T, with His blessing and guidance, I have managed to complete this thesis on time. This thesis would not have been possible without the guidance and the help of several individuals who in one way or another contributed and extended their valuable assistance in the preparation and completion of this study. My utmost gratitude and appreciation goes to my beloved:

Mother, father and family members,

Lecturers and staff of UPM and UMB,

Research teammates,

Best friend and colleagues,

Former supervisor

and

others that have been contributed in whatever way to the successful completion of this thesis.

Thank you.

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the Degree of Master of Science

NON-DESTRUCTIVE ESTIMATION OF OIL PALM YIELD AND QUALITY WITH FLUORESCENCE SENSING

By

NORUL HUSNA BINTI ABU KASSIM

January 2016

Chair : Assoc. Prof. Siva Kumar Balasundram, PhD
Faculty : Agriculture

Fluorescence sensing is one of the proximal sensor techniques which use instruments operating very close to or in contact with the object of inquiry. It is an emerging approach in assessing and monitoring crop yield and quality. At present, ineffective Fresh Fruit Bunch (FFB) maturity assessment and untimely harvesting are problematic to the Malaysian oil palm industry. The current method of FFB assessment is extremely tedious and a time-consuming process that is destructive, prone to errors and inconsistent. Thus, implementation of an oil palm harvesting protocol that deploys a non-destructive sensor-based approach to assess FFB ripening is highly desirable. The objectives of this study were: i) to quantify the relationship between palm oil quality (DOBI) and fluorescence-sensed anthocyanin and flavonol contents across different palm ages, ii) to quantify the relationship between palm oil yield (OER) and fluorescence-sensed anthocyanin and flavonol contents across different palm ages, and iii) to identify the best-fitted model and validate the spatial distribution of palm oil quality, yield and fluorescence-sensed anthocyanin and flavonol contents across different palm ages. In this study, three 2-ha plots were demarcated based on different year of planting; 2002 (12 year old palms), 2005 (9 year old palms) and 2008 (6 year old palms). Sixty FFBs and their corresponding loose fruits were scanned for each plots using fluorescence sensor (Multiplex®). Overall, secondary metabolites such as anthocyanin and flavonol are proven to be a reliable indicator of palm oil quality and yield ($r=0.6$ of anthocyanin and $r=0.8$ of flavonol). However, fluorescence indices (Anth_B, Flav_B and NBI) show mixed correlation strengths with DOBI and OER across different palm ages. For the first objective, the secondary metabolites; anthocyanin and flavonol, were analysed to determine the relationship between Deterioration of Bleachability Index (DOBI) and secondary metabolites content. Results showed that the estimated strength of secondary metabolites was more pronounced toward oil quality in 6 and 9 year old palms (younger palms). For the second objective, the secondary were analyzed to determine the relationship between Oil Extraction Rate (OER) and secondary metabolites content. Results showed that the estimated strength of secondary metabolites was more pronounced toward oil yield in 9 and 12 year old palms (older palms). As a conclusion, both palm oil quality and oil yield

determinations using fluorescence indices are most applicable in matured 9 year palms ($r=0.63$ and 0.52 with DOBI; $r=0.65$ and 0.53 with OER, respectively). This indicates that palm oil quality and yield, particularly of 9 year old palms, are highly related to fluctuations in anthocyanin and flavonol contents in the fruit. According to Corley and Tinker 2003, oil palm has reached its maturity and their growth becomes more stable at 9-10 years after planting which is when their productivity is at its maximum. This could also infer that anthocyanin and flavonol contents in the 6 and 12 year old palms are more indicative of their metabolic process rather than their oil quality or oil yield. This observation is in agreement with Muller et al. (2013) who reported that during plant growth (particularly in 6 and 12 year palms), higher accumulation of anthocyanin and flavonol contents will act as a trade-off between the plant's primary and secondary metabolic system, which are not stable as compared in 9 year old palms. For the third objective, ten variables of spatial distribution of palm oil quality, palm oil yield and fluorescence-sensed secondary metabolites were modeled and validated for each age group. Majority of variables for all three age groups have a short effective range (<153 m) with a strong spatial dependence ($>99.7\%$) except for OER of 2008. Most of the tested variables could be had 99% variability that is explainable. Across different age groups, TFC was recorded with lowest R^2 values ranging between $0.03-0.07$ which due to secondary metabolism of flavonoid in palm oil meanwhile majority of the variables showing a moderate R^2 with value ranging between $0.5-0.7$ (Anth_B, Flav_B, Anth_Lf, Flav_Lf and TAC). CV values of variables in all age groups can be divided into two groups; low and moderate variability (CV= $7-49\%$ in 2002; $13-41\%$ in 2005; and $7.7-37.8\%$ and $52.7-73.4\%$ in 2008, respectively). This indicates that all variables are greatly being influenced and significantly associated with secondary metabolite concentrations in oil palm compared to spatial regularity. Fluorescence sensor has a promising potential in determining secondary metabolites in oil palm, however more research and future findings are needed to support and compliment this work.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Master Sains

PENILAIAN KUALITI DAN HASIL MINYAK SAWIT SECARA BUKAN MEMUSNAH DENGAN MENGGUNAKAN KAEDAH PENGESAN FLORESEN

Oleh

NORUL HUSNA BINTI ABU KASSIM

Januari 2016

Pengerusi : Prof. Madya Siva Kumar Balasundram, PhD
Fakulti : Pertanian

Pengesan floresen merupakan salah satu teknik pengesan jarak dekat yang menggunakan instrumen fungsi jarak dekat atau bersentuhan dengan objek yang dikesan. Pengesan floresen merupakan pendekatan yang kian mendapat perhatian dalam menganalisis dan mengawal hasil dan kualiti tanaman. Dewasa ini, ketidakcekapan dalam menganalisis tandan buah sawit dan penuaian pramatang merupakan masalah yang semakin menular terutamanya dalam industri kelapa sawit di Malaysia. Kaedah menganalisis tandan buah sawit sedia ada adalah sangat sukar dan merupakan proses yang mengambil masa yang lama sehinggakan ia akan merosakkan fizikal buah, terdedah kepada kesilapan dan tidak konsisten. Justeru, pelaksanaan terhadap protokol penuaian kelapa sawit yang menggunakan kaedah pengesan secara tidak-memusnahkan-fizikal buah dalam menganalisis kemasakan tandan buah sawit ini adalah sangat diperlukan. Objektif kajian ini ialah: i) untuk mengkaji hubungan antara kualiti minyak sawit (DOBI) dan kandungan antosianin dan flavonol yang dikesan floresen berdasarkan usia pokok yang berbeza, ii) untuk mengkaji hubungan antara hasil minyak sawit (OER) dan kandungan antosianin dan flavonol yang dikesan floresen berdasarkan usia pokok yang berbeza, dan iii) untuk mengenal pasti model yang paling bersesuaian dan mengesahkan ruang agihan daripada kualiti minyak sawit, hasil minyak sawit dan kandungan antosianin dan flavonol yang dikesan floresen berdasarkan usia pokok yang berbeza. Dalam kajian ini, tiga 2-ha plot telah ditanda dalam usia pokok yang berbeza berdasarkan tahun penanaman: 2002 (12 tahun), 2005 (9 tahun) dan 2008 (6 tahun). Enam puluh tandan buah sawit dan buah terlerainya telah diimbab bagi setiap plot dengan menggunakan pengesan floresen (Multiplex®). Secara keseluruhan, metabolit sekunder seperti antosianin dan flavonol telah terbukti sebagai petunjuk yang dipercayai bagi kualiti dan hasil minyak sawit ($r=0.6$ of antosianin and $r=0.8$ of flavonol). Namun, index floresen (Anth_B, Flav_B and NBI) menunjukkan kekuatan kolerasi yang berbeza-beza terhadap DOBI dan OER dalam usia pokok yang berbeza. Bagi objektif pertama, metabolit sekunder, antosianin dan flavonol, dianalisis untuk mengetahui hubungan antara index kemerosotan meluntur (DOBI) dan kandungan metabolit sekunder. Hasil kajian mendapati anggaran kekuatan metabolit sekunder adalah lebih menonjol terhadap kualiti minyak

sawit pada usia 6 dan 9 tahun (buah sawit yang lebih muda). Bagi objektif kedua, metabolit sekunder tersebut dianalisis untuk mengetahui hubungan antara kadar perahan minyak (OER) dan kandungan metabolit sekunder. Hasil kajian mendapati anggaran kekuatan metabolit sekunder adalah lebih menonjol terhadap hasil minyak sawit pada usia 9 dan 12 tahun (buah sawit yang lebih tua). Secara keseluruhannya, penentuan kedua-dua kualiti dan hasil minyak sawit menggunakan pengesan floresen adalah paling sesuai diaplikasi pada pokok sawit matang yang berusia 9 tahun ($r=0.63$ dan 0.52 untuk DOBI; $r=0.65$ and 0.53 untuk OER, masing-masing). Ini menunjukkan bahawa kualiti dan hasil minyak sawit, terutamanya pokok berusia 9 tahun adalah sangat berkait rapat dengan penurunan atau kenaikan kandungan antosianin dan flavonol di dalam buah. Menurut Corley dan Tinker (2003), kelapa sawit telah mencapai tempoh matang dan proses pertumbuhan mereka telah menjadi lebih stabil pada usia 9-10 tahun selepas ditanam dan dimana produktiviti mereka adalah pada tahap maksimum. Ini membuktikan bahawa kandungan antosianin dan flavonol pada usia 6 dan 12 tahun adalah lebih menunjukkan proses metabolik mereka berbanding kualiti atau hasil minyak sawit. Pemerhatian ini bertepatan dengan penemuan daripada Muller et al. (2013) yang melaporkan bahawa pengumpulan kandungan antosianin dan flavonol yang lebih tinggi ketika pertumbuhan pokok (terutamanya di usia 6 dan 12 tahun), akan bertindak sebagai proses penyeimbangan "trade-off" antara sistem metabolisme pertama dan sekunder pokok, yang mana tidak stabil berbanding pokok berusia 9 tahun. Bagi objektif ketiga, sepuluh pemboleh-ubah ruang agihan daripada kualiti minyak sawit, hasil minyak sawit dan kandungan antosianin dan flavonol yang dikesan floresen telah dimodelkan dan disahkan untuk setiap kumpulan usia pokok. Kebanyakan daripada ruang agihan kualiti minyak, hasil minyak dan metabolit sekunder yang telah dikesan mempunyai julat efektif (ER) yang pendek (<153 m) dan ruang pergantungan "spatial dependence" yang kuat ($>99.7\%$) kecuali dalam OER bertahun 2008. Hampir keseluruhan parameter yang dikaji mungkin mempunyai 99% pembolehubah yang boleh diterangkan. Dalam usia pokok yang berbeza-beza ini, TFC telah merekodkan nilai R^2 terendah bernilai 0.03 hingga 0.07 yang berkemungkinan berlaku disebabkan oleh metabolisme sekunder (flavonoid) di dalam minyak sawit, manakala kebanyakan parameter yang lain mempunyai nilai R^2 sederhana bernilai 0.5 hingga 0.7 (Anth_B, Flav_B, Anth_Lf, Flav_Lf dan TAC). Nilai CV setiap parameter boleh dibahagikan kepada dua kumpulan; variabiliti rendah dan sederhana (CV=7-49% untuk 2002; 13-41% untuk 2005; dan 7.7-37.8% dan 52.7-73.4% untuk 2008, masing-masing). Ini menunjukkan bahawa kesemua parameter sangat dipengaruhi dan berkait rapat dengan kekerapan ruang. Maka, pengesan floresen berpotensi dalam menentukan metabolit sekunder dalam buah kelapa sawit, namun lebih banyak penyelidikan dan penemuan pada masa hadapan yang diperlukan untuk menyokong kerja penyelidikan ini.

ACKNOWLEDGEMENTS

First and foremost, my utmost appreciation to my parents and family members, for all those encouragements through difficulties and becoming my attentive motivator whenever problem occurs. I am also extremely grateful to my supervisor, Assoc. Prof. Dr. Siva Kumar Balasundram, for his supervision, guidance, constructive criticism and support in order to guide me in completing this thesis. Under his guidance, I successfully overcome many difficulties and learned a lot of valuable lessons in becoming a better person.

I take this opportunity to earnestly acknowledge my co-supervisor, Prof. Dr. Tan Chin Ping and his PhD students for their priceless guidance, knowledge and help in facilitating this research. Not to be forgotten, my gratitude goes to the staff of Agrotechnology Department, Fats and Oil Laboratory as well as staff of United Malacca Berhad for providing necessary infrastructure and resources in order to accomplish my research work within the time provided. I would like to express my gratefulness to my colleagues especially my research team, postgraduate students of Agrotechnology Department and Master of Science students (Batch of 2013-2015) for our beautiful friendship and valued moment as students of Universiti Putra Malaysia. Last but not least, my deepest gratitude to United Malacca Berhad (UMB) plantation for funding this research.

I would like to thank all those people who made this thesis possible and expressing my heartiest appreciation to all those who contributed in many ways to the success of this study and made it as an unforgettable experience for me.

I certify that a Thesis Examination Committee has met on 12 January 2016 to conduct the final examination of Norul Husna binti Abu Kassim on her thesis entitled "Non-Destructive Estimation of Oil Palm Yield and Quality with Fluorescence Sensing" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Mohamed Hanafi bin Musa, PhD

Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Phebe Ding, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Internal Examiner)

Yuxin Miao, PhD

Associate Professor
China Agricultural University
China
(External Examiner)



ZULKARNAIN ZAINAL, PhD

Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 24 March 2016

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Siva K. Balasundram, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Tan Chin Ping, PhD

Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Norul Husna Binti Abu Kassim, GS 35989

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____
Name of
Chairman of
Supervisory
Committee: Siva K. Balasundram, PhD

Signature: _____
Name of
Member of
Supervisory
Committee: Tan Chin Ping, PhD

TABLE OF CONTENTS

	ABSTRACT	Page
	ABSTRAK	i
	ACKNOWLEDGEMENTS	iii
	APPROVAL	v
	DECLARATION	vi
	LIST OF TABLES	viii
	LIST OF FIGURES	xii
	LIST OF ABBREVIATIONS	xiii
	CHAPTER	xv
 	CHAPTER	
1	INTRODUCTION	1
1.1	Background of oil palm	1
1.2	Problem statement	1
1.3	Research objectives	2
1.4	Significance of study	2
2	LITERATURE REVIEW	3
2.1	Oil palm (<i>Elaeis guineensis</i> Jacq.)	3
2.1.1	The oil palm industry	3
2.1.2	Classification and morphology of oil palm	3
2.1.3	Physical and chemical properties of palm oil	4
2.1.4	Types and uses of palm oil	5
2.1.5	Palm oil quality determination	6
2.1.6	Palm oil yield determination	7
2.1.7	Secondary metabolites in oil palm	9
	2.1.7.1 Structure and biosynthesis of anthocyanin	10
	2.1.7.2 Structure and biosynthesis of flavonol	11
	2.1.7.3 Structure and biosynthesis of chlorophyll	11
	2.1.7.4 Phytochemical function and application of secondary metabolites	12
2.2	Fluorescence-sensing technology	14
2.2.1	Types of detection	14
2.2.2	Principles and mechanism of fluorescence sensor	15
2.2.3	Fluorescence sensor application on crops	19
2.3	Statistical and geostatistical analysis	21
2.3.1	Outlier test (Grubbs' test)	21
2.3.2	Descriptive statistics	22
2.3.3	Normality test	23
2.3.4	Correlation test	23
2.3.5	Variography	24
2.3.6	Spatial dependence test	27
2.3.7	Interpolation: Point kriging	27
2.3.8	Cross-validation test	28

3	MATERIALS AND METHODS	29
3.1	Experimental location	29
3.2	Field sampling	29
3.3	Non-destructive method of palm oil determination	31
3.3.1	Anthocyanin and flavonol indices	32
3.3.2	Chlorophyll index	33
3.3.3	Nitrogen balance index	34
3.4	Destructive method of palm oil determination	35
3.4.1	Deterioration of bleachability index	35
3.4.2	Oil extraction rate	35
3.4.3	Total anthocyanin content (TAC)	36
3.4.3.1	Sample preparation	36
3.4.3.2	TAC assay	36
3.4.4	Total flavonoid content (TFC)	37
3.4.4.1	Sample preparation	37
3.4.4.2	TFC assay	38
3.5	Statistical analysis	38
3.5.1	Outlier test (Grubbs' test)	38
3.5.2	Descriptive statistics	38
3.5.3	Normality test	38
3.5.4	Correlation test	38
3.6	Geostatistical analysis	39
3.6.1	Variography	39
3.6.2	Spatial dependence test	39
3.6.3	Interpolation: Point kriging	39
3.6.4	Cross-validation test	39
4	RESULTS AND DISCUSSION	40
4.1	Statistical analysis	40
4.1.1	Normality test	40
4.1.2	Descriptive statistics	41
4.1.3	Correlation test	47
4.2	Geostatistical analysis	49
4.2.1	Variography and spatial dependence test	49
4.2.2	Point kriging (Variability map)	58
4.2.3	Cross-validation test	65
4.3	Conclusion	68
5	SUMMARY, CONCLUSION AND RECOMMENDATION FOR FUTURE RESEARCH	69
	REFERENCES	71
	APPENDICES	80
	BIODATA OF STUDENT	90
	LIST OF PUBLICATIONS	91

LIST OF TABLES

Table		Page
1	Different types of Proximal sensor (based on signal affecting detection)	15
2	Types of theoretical semivariogram models ($a_0 > 0$, unless noted otherwise)	26
3	List of variables measured using non-destructive method (Multiplex® fluorescence sensor) and destructive method (laboratory)	31
4	Normality test (p value) of all variables across different age groups (6, 9 and 12 year old palms)	40
5	Descriptive statistics of statistical attributes of all variables in 2002 plot (12 year old palms)	43
6	Descriptive statistics of statistical attributes of all variables in 2005 plot (9 years old palms)	44
7	Descriptive statistics of statistical attributes of all variables in 2008 plot (6 years old palms)	46
8	Correlation between fluorescence indices (Anth_Lf and Flav_Lf) and measured concentrations of anthocyanin (TAC) and flavonoid (TFC) across different palm ages	47
9	Correlation of fluorescence indices (anthocyanin and flavonol) between different parts (bunch and loose fruit) across different palm ages	48
10	Correlation between fluorescence indices (Anth_B, Flav_B and NBI) and palm oil quality (DOBI) and yield (OER) attributes across different palm ages	48
11	Geostatistical attributes and spatial dependence values of all attributes across different palm ages	57
12	Cross validation test which consist of mean error values (ME), mean squared error (MSE) and standardized mean squared error (SMSE) across different palm age groups	67

LIST OF FIGURES

Figure		Page
1	Morphology of <i>Elaeis guineensis</i> palm tree (left) and morphology of pinnate leaf of oil palm (right)	4
2	Fresh Fruit Bunch (FFB) at ripe stage (left) and its fruitlets and their morphology (right)	5
3	Structure of flavonoid, in position of C ₆ -C ₃ -C ₆	9
4	Structure of anthocyanin within the carbon chain	10
5	Structure of flavonol within the carbon chain	11
6	Different color spectrum absorption between chlorophyll a and b	12
7	Absorption between chlorophyll pigments and malvidin-3O-glucoside (oenin) at different wavelength which results in different coloration of pigments	12
8	Focused proximity sensors where the source and the detector are focused to a point in front of the sensor	15
9	Electromagnetic spectra of various sources and wavelength range	16
10	Operating principle of proximity sensors	17
11	Technical specification and the components of the Multiplex [®] sensor	17
12	Mechanism of fluorescence excitation using the Multiplex [®] sensor	18
13	An isotropic semivariogram with principal attributes such as nugget, sill and range	25
14	Location of study area	29
15	Location of field plots in study area	30
16	Research design and sampling method	30
17	Specifications of using the Multiplex [®] fluorescence sensor to capture data from the FFB	31
18	Assessment of anthocyanin and flavonol indices on the bunch	32
19	Assessment of anthocyanin and flavonol indices on the loose fruits	32
20	Frond of the oil palm is assessed to quantify the chlorophyll index (SFR) of the plant	33
21	Cross section of leaf structure	34
22	Differential measurement of fluorescence emitted by chlorophyll in the leaf	34

23	Samples were left to stand until a yellowish layer of oil was formed, which was removed from the extract	36
24	Supernatant was filtered from a mixture of oil to get a clear fruit extract for assay	37
25	The Gaussian distribution, $f(x)$, also known as “bell-shaped” curve model consist of attributes of univariate statistics	41
26	Isotropic variograms of all variables in 2002 age group (12 year old palms): (a) Anth_B, (b) Flav_B, (c) DOBI, (d) OER, (e) Anth_Lf, (f) Flav_Lf, (g) TAC, (h) TFC, (i) SFR and (j) NBI	52
27	Isotropic variograms of all variables in 2005 age group (9 year old palms): (a) Anth_B, (b) Flav_B, (c) DOBI, (d) OER, (e) Anth_Lf, (f) Flav_Lf, (g) TAC, (h) TFC, (i) SFR and (j) NBI	54
28	Isotropic variograms of all variables in 2008 age group (6 year old palms): (a) Anth_B, (b) Flav_B, (c) DOBI, (d) OER, (e) Anth_Lf, (f) Flav_Lf, (g) TAC, (h) TFC, (i) SFR and (j) NBI	55
29	Spatial distribution of anthocyanin indices of 2002 age group: (a) Anth_B and (b) Anth_Lf	59
30	Spatial distribution of flavonol indices of 2002 age group: (a) Flav_B and (b) Flav_Lf	59
31	Spatial distribution of (a) TAC and (b) TFC of 2002 age group	60
32	Spatial distribution of (a) DOBI and (b) OER of 2002 age group	60
33	Spatial distribution of (a) SFR and (b) NBI of 2002 age group	60
34	Spatial distribution of anthocyanin indices of 2005 age group: (a) Anth_B and (b) Anth_Lf	61
35	Spatial distribution of flavonol indices of 2005 age group: (a) Flav_B and (b) Flav_Lf	61
36	Spatial distribution of (a) TAC and (b) TFC of 2005 age group	62
37	Spatial distribution of (a) DOBI and (b) OER of 2005 age group	62
38	Spatial distribution of (a) SFR and (b) NBI of 2005 age group	62
39	Spatial distribution of anthocyanin indices of 2008 age group: (a) Anth_B and (b) Anth_Lf	63
40	Spatial distribution of flavonol indices of 2008 age group: (a) Flav_B and (b) Flav_Lf	63
41	Spatial distribution of (a) TAC and (b) TFC of 2008 age group	64
42	Spatial distribution of (a) DOBI and (b) OER of 2008 age group	64
43	Spatial distribution of (a) SFR and (b) NBI of 2008 age group	64

LIST OF ABBREVIATIONS

FFB	Fresh Fruit Bunch
DOBI	Deterioration of Bleachability Index
OER	Oil Extraction Rate
Anth_B	Anthocyanin Index of Bunch
Flav_B	Flavonol Index of Bunch
Anth_LF	Anthocyanin Index of Loose Fruit
Flav_B	Flavonol Index of Loose Fruit
TAC	Total Anthocyanin Content of Loose Fruit
TFC	Total Flavonoid Content of Loose Fruit
SFR	Simple Fluorescence Ratio (Chlorophyll content)
NBI	Nitrogen Balance Index
CPO	Crude Palm Oil
RPO	Refined Palm Oil
PKO	Palm Kernel Oil
FFA	Free Fatty Acid
GMQ	Good Merchantable Quality
MPOB	Malaysian Palm Oil Board
MPOC	Malaysian Palm Oil Council
%F/B	Percentage of Fruit Per Bunch
M/F	Mesocarp to Fruit Ratio
S/F	Shell to Fruit Ratio
K/F	Kernel to Fruit Ratio
%O/M	Percentage of Oil to Mesocarp
%O/K	Percentage of Oil to Kernel
%O/B	Percentage of Oil to Bunch
O/DM	Oil to Dry Mesocarp
LED	Light Emitting Diode
IR	Infra-Red
UV	Ultra-Violet
GPS	Global Positioning System
N	Nitrogen

NNI	Nitrogen Nutrition Index
BRR	Blue-to-Red fluorescence Ratio
NDVI	Normalized Difference Vegetation Index
CV	Coefficient of Variation
ME	Mean Error
MSE	Mean Squared Error
SMSE	Standardized Mean Squared Error
SD	Standard Deviation
SEM	Standard Error of Mean
SES	Standard Error of Skewness
SEK	Standard Error of Kurtosis
C_0	Nugget
$C_0 + C$	Sill
A_0	Range
ER	Effective Range
DEM	Digital Elevation Model

CHAPTER 1

INTRODUCTION

1.1 Background of oil palm

Oil palm (*Elaeis guineensis* Jacq.) is a multipurpose tree crop of high productivity, yielding a total of 19.22 million tons of crude palm oil in 2013. At present, oil palm produces the highest yields per hectare among commercial oil crops. The oil palm industry has become a major source of oils and fats production worldwide and has led to a rapidly expanding world oils and fats industry, which now is based in the tropical areas of Asian regions, Africa and America. The origin of oil palm is believed to be in Africa. However, today the most productive regions of the oil palm industry are located in Malaysia and Indonesia, which produce most of the oil entering international trade.

1.2 Problem statement

The oil palm industry has continually faced the problem of a significant gap between actual yields and potential yields. Ineffective harvesting and poor maturity/grading assessment had been identified as one of determinants of these problems. The common traditional methods of oil palm maturity assessment followed by quality and yield assessments are destructive, costly, laborious, inconsistent and time-consuming. At present, Fresh Fruit Bunch (FFB) ripeness is determined based on the number or percentage of fruits detached from the bunch. Collection of detached fruits is costly when manually carried out, but it is highly necessary because they influence the oil yield.

Commercial plantations commonly carry out FFB harvesting based on the minimum ripeness criterion, which usually comprising 1-5 detached fruit per FFB. The current method of FFB human grading which is based on the following parameters: 1) bunch ripeness, 2) bunch abnormality, 3) bunch weight, 4) bunch contamination and 5) number of detached fruit, is prone to errors and inconsistencies. It is therefore desirable to implement a better harvesting and grading assessment practices while reducing the number of labor needed by introducing a sensory system that will quantify the FFB ripening stages, oil content and quality in a more effective way.

1.3 Research objectives

1. To quantify the relationship between palm oil quality and fluorescence-sensed anthocyanin and flavonol contents across different palm age groups.
2. To quantify the relationship between palm oil yield and fluorescence-sensed anthocyanin and flavonol contents across different palm age groups.
3. To identify the best-fitted model and validate the spatial distribution of palm oil quality, yield and fluorescence-sensed anthocyanin and flavonol contents across different palm age groups.

1.4 Significance of study

Fluorescence sensing can provide an alternative to the current practice of FFB assessment and grading, which is tedious, costly and based on destructive methods. Assessment and grading of FFB in real-time will reduce the time load for field operation and can potentially increase the efficiency of assessment, which can affect the quality of oil content in FFB. The quality of oil in FFBs that are delayed in processing after harvest usually decrease due to build-up of free fatty acids. FFB assessment and grading via fluorescence sensing can produce consistency, as compared to the standard manual approach of using human eyes. Importantly, fluorescence sensing provides a non-destructive means to measure oil yield and quality, which is more practical than the current destructive method of palm oil quality determination via Deterioration of Bleachability Index (DOBI) and palm oil yield via Oil Extraction Rate (OER). DOBI and OER assessments are commonly conducted using standard laboratory procedures which are costly and time-consuming.

REFERENCES

- Adir, N., Zer, H., Scholat, S. and Ohad, I. 2003. Photoinhibition – a historical perspective. *Photosynthetic Research*, 76: 343-370.
- Agati, G., Traversi, M. and Cerovic, Z. 2008. Chlorophyll fluorescence imaging for noninvasive assessment of anthocyanins in whole grape (*Vitis vinifera* L.) bunches. *Photochemistry and Photobiology*, 84: 1431-34.
- Andersen, O. M. and Jordheim, M. . 2006. The anthocyanins. In Andersen, O. M. and Markham, K. R. (Eds.). *Flavonoids: chemistry, biochemistry and applications*. CRC Press, Boca Raton, pp: 471-553.
- Andersen, O. M. 2001. Anthocyanins. *Encyclopedia of life sciences*. John Wiley and Sons, Ltd., ISBN: 0470016175.
- Anderson, P. J. 2011. Identifying commonly cultivated palms - in a resource for pests and diseases of cultivated palms. Florida Department of Agriculture and Consumer Service, Division of Plant Industry and Identification Technology Program, CPHST, PPQ, APHIS, USDA; Fort Collins, CO. Retrieved April 8 2015, from <http://idtools.org/id/palms/palmid/>.
- Atinmo, T. and Bakre, A. T. 2003. Palm fruit in traditional African food culture. *Asia Pacific Journal Clinical Nutrition*, 12(3): 350-354.
- Azmil, H. A. T. 2006. Standard reference materials - fatty acid compositions of palm oil, palm olein and palm stearin (set no. 2). MPOB Information Series, ISSN: 1511-7871.
- Balasundram, S. K., Husni, M. H. A. and Ahmed, O. H. 2008. Application of geostatistical tools to quantify spatial variability of selected soil chemical properties from a cultivated tropical peat. *Journal of Agronomy*, 7(1): 82-87.
- Balasundram, S. K., Mulla, D. J. and Robert, P. C. 2007. Spatial data calibration for site-specific phosphorus management. *International Journal of Agricultural Research*, 2(11): 888-899.
- Balasundram, S. K., Mulla, D. J., Robert, P. C. and Allan, D. L. 2006. Spatial variability of soil fertility variables influencing yield in oil palm (*Elaeis guineensis* Jacq.). *Asian Journal of Plant Science*, 5(2): 397-408.
- Basiron, Y. 2005. Palm oil. In Shahidi, F. (Eds.). *Bailey's Industrial Oil and Fat Products*, Sixth Edition, Six Volume Set. John Wiley and Sons, Inc., ISBN: 978-0-471-38460-1.
- Berger, K. G. 2006. Understanding oils and fats. *Global Oil Fat Business Magazine*, 3(4): 1-51.

- Bilger, W., Rolland, M. and Nybakken, L. 2001. UV-excited chlorophyll fluorescence as a tool for assessment of UV-protection by epidermis of plants. *Journal of Experimental Botany*, 52: 2007-14.
- Bockish, M. 1998. *Fats and oils handbook*. Champaign IL: AOCS Press, pp: 739-743.
- Buschmann, C., Langsdorf, G. and Lichtenthaler, H. K. 2000. Imaging of the blue, green, and red fluorescence emission of plants: an overview. *Photosynthetica*, 38: 483-491.
- Cartelat, A., Cerovic, Z. G., Goulas, Y., Meyer, S., Lelarge, C., Prioul, J. L., Barbottin, A., Jeuffroy, M. H., Gate, P., Agati, G. and Moya, I. 2005. Optically assessed contents of leaf polyphenolics and chlorophyll as indicators of nitrogen deficiency in wheat (*Triticum aestivum* L.). *Field Crops Research*, 91: 35-49.
- Carr, J. R. 2002. *Data visualization in the geosciences*. Prentice-Hall Inc., ISBN: 0-13-089706-X.
- Cambardella, C. A., Moorman, T. B., Parkin, T. B., Karlen, D. L., Novak, J. M., Turco, R. F. and Konopka, A. E. 1994. Field-scale variability of soil properties in Central Iowa soils. *Soil Science Society of America Journal*, 58(5): 1501-1511.
- Cerovic, Z. G., Moise, N., Agati, G., Lataouche, G., Ben Ghazlen, N. and Meyer, S. 2008. New portable optical sensors for the assessment of winegrape phenolic maturity based on berry fluorescence. *Journal of Food Composition and Analysis*, 21: 650-54.
- Cerovic, Z. G., Masdoumier, G., Ben Ghazlen, N., and Latouche, G. 2012. A new optical leaf-clip meter for simultaneous non-destructive assessment of leaf chlorophyll and epidermal flavonoids. *Physiologia Plantarum*, ISSN: 0031-931-7.
- Chang, Kang-tsung. 2012. *Introduction to geographic information systems*. Sixth edition. Mc-Graw Hill Companies, Inc., ISBN: 978-0-07-336931-0.
- Chalker-Scott, L. 1999. Environmental significance of anthocyanins in plant stress responses. *Photochemical Photobiology*, 70: 1-9.
- Corley, R. H. V. and Tinker, P. B. 2003. *The oil palm*. Fourth edition. Oxford: Blackwell Publishing, ISBN: 0-632-05212-0.
- Corley, R. H. V. 1973. In Wastie, R. L. and Earp, D. A. (Eds.). *Advances in oil palm cultivation*. Incorporated Society of Planters, Kuala Lumpur, pp: 37-49.
- Cowan, M. M. 1999. Plant products as antimicrobial agents. *Clinical Microbiology Reviews*, 12(4): 564-582.

- Cronquist, A. 1981. An integrated system of classification of flowering plants. Columbia University Press.
- Eric, G. and Badarudin, P. 2003. Fundamentals and applications of gesostatistics: for scientists, engineers and geographers. Penerbit Universiti Sains Malaysia, ISBN 983-861-260-X.
- Feynman, R. P. 2006. QED - The strange theory of light and matter. Princeton University Press, Princeton, ISBN: 9781400847464.
- Fraden, J. 2010. Handbook of modern sensors: physics, designs and applications. Fourth edition. Springer Science + Business Media, LLC, ISBN: 978-1-4419-6465-6.
- Friedli, G. L. 1992. Structure/activity comparison in the ability of some terpenoid food flavours to cause peroxisome proliferation (PhD). University of Surrey, United Kingdom.
- Foster, H. L., Tayeb Dolmat, M. and Gurmit, S. 1987. In proceedings of International Oil Palm Conference, PORIM, Kuala Lumpur, pp: 294-304.
- Fossen, T., Cabrita, L. and Andersen, O. M. 1998. Colour and stability of pure anthocyanins influenced by pH including the alkaline region. Food Chemistry, 63(4): 435-440.
- FORCE-A®. 2009. Multiplex Aromatic plants. Edition 2. In Proceedings of ITEIPMAI 2008. ORSAY Cedex - France.
- Force-a.eu. Multiplex® research. Retrieved December 2013, from <http://www.force-a.eu/an/multiplex.php>.
- Gee, P. T. 2005. Use of the deterioration of bleachability index (DOBI) to characterise the quality of crude palm oil. Crude palm oil and the DOBI, Keck Seng (M) Berhad, Masai Johor. Retrieved February 17 2014, from http://innoleague.com/Deterioration_of_Bleachability.pdf.
- Gibon, V., Ayala, J. V., Dijckmans, P., Maes, J. and Greyt, W. D. 2009. Future prospects for palm oil refining and modifications. OCL 16, pp: 193-200.
- Ghozlen, N., Moise, N., Lataouche, G., Martinon, V., Mercier, L., Besancon, E. and Cerovic, Z. G. 2010. Assessment of winegrape maturity using a new portable sensor: non-destructive quantification of anthocyanins. Journal of International Science Vigne Vin, 44: 1-8.
- Gerald, J. S. and Kenneth, R. M. 1998. Tautomerism of flavonol glucosides: relevance to plant UV protection and flower colour. Journal of Photochemistry and Photobiology A: Chemistry, 118(2): 99-105.

- Goulas, Y., Cerovic, Z. G., Cartelat, A., and Moya, I. 2004. Dualex: A new instrument for field measurements of epidermal UV-absorbance by chlorophyll fluorescence. *Applied Optics*, 43(2): 4488-4496.
- Gould, K., Davies, K. and Winefield, C. 2009. Anthocyanins: biosynthesis, functions and applications. Springer Science+Business Media, LLC, ISBN: 978-0-387-77334-6.
- Gould, K. S. and Lister, C. 2006. Flavanoid functions in plants. In Andersen, O. M. and Markham, K. R. (Eds.). *Flavonoid - Chemistry, Biochemistry and Application*. CRC Press, Boca Raton, pp: 397-442.
- Googleearth.com. 2014. Google earth®. Images of Peninsular Malaysia. Acquisition date of November 17, 2014. Retrieved on December 2014, from <https://mapsengine.google.com/10446176163891957399-047495316809535685774/mapview/?authuser>.
- Googleearth.com. 2015. Google earth®. Images of Malacca. Acquisition date of Jan 31, 2014. Retrieved on December 2014, from <https://mapsengine.google.com/10446176163891957399-047495316809535685774/mapview/?authuser=0>.
- Graphpad.com. 2015. GraphPad Software, Inc. 1995-2015. Retrieved on January 25, 2015, from [http://www.graphpad.com/guides/prism/6/statistics/index.htm?coefficient_of_variation_\(cv\).html](http://www.graphpad.com/guides/prism/6/statistics/index.htm?coefficient_of_variation_(cv).html).
- Hagen, S., Solhaug, K., Bengtsson, G., Borge, G. and Bilger, W. 2006. Chlorophyll fluorescence as a tool for non-destructive estimation of anthocyanins and total flavonoids in apples. *Postharvest Biology and Technology*, 41: 156-63.
- Hartley, C. W. S. 1988. The oil palm (*Elaeis guineensis* Jacq.). Longman Scientific and Technical Publication. John Wiley and Sons, NY, pp: 694-703.
- Harun, M. H. 2000. Yield and yield components and their physiology. In Yusof, B. (Ed.). *Advances in oil palm*. Malaysian Palm Oil Board, pp: 146-170.
- Hatier, J. H. and Gould, K. S. 2007. Black coloration in leaves of *Ophiopogon planiscapus* "Nigrescens". Leaf optics, chromaticity and internal light gradient. *Functional Plant Biology*, 34: 130-138.
- Hadi, S., Ahmad, D. and Akande, F. B. 2009. Determination of the bruise indexes of oil palm fruits. *Journal of Food Engineering*, 95: 322-326.
- Hazir, M. H. M. and Shariff, A. R. M. 2011. Oil palm physical and optical characteristics from two different planting materials. *Research Journal of Applied Sciences, Engineering and Technology*, 3: 953-62.

- Hazir, M. H. M, Shariff, A. R. M and Amirruddin, M. D. 2012. Determination of oil palm fresh fruit bunch ripeness-based on flavonoid and anthocyanin content. *Industrial Crops and Products*, 36: 466-75.
- Hughes, N. M., Neufeld, H. S. and Burkey, K. O. 2007. Coordination of anthocyanin decline and photosynthetic maturation in juvenile leaves of three deciduous tree species. *New Phytology*, 168: 575-587.
- Isaaks, E. H. and Srivastava, R. M. 1989. An introduction to applied geostatistic. Oxford University Press, New York.
- Ishak, W. I. W and Hudzari, R. M. 2010. Image-based modeling for oil palm fruit maturity prediction. *Journal of Food, Agriculture and Environment*, 8: 469-76.
- Jacquin, N. J. 1763. *Selectarum stirpium Americanarum historia*, 1: 277-282.
- Junaidah, M. T., Norrizah, A. R. and Zaliha, O. 2013. Effect of sterilization process on deterioration of bleaching index (DOBI) of crude palm oil (CPO) extracted from different degree of oil palm ripeness. *International Journal of Bioscience, Biochemistry and Bioinformatics*, 3(4): 322-327.
- Kermack, K. A. and Haldane, J. B. S. 1950. *Biometrika*, 37(30): 1-2.
- Kegen, Y., Ian, S. and Jay Guo, Y. 2009. Ground-based wireless positioning. *IEEE Press Digital and Mobile Communication*, ISBN: 978-0-470-74704-9.
- Konczak, I. and Zhang, W. 2004. Anthocyanins - more than nature's colors. *Journal of Biomedical Biotechnology*, 2004: 239-240.
- Kuntom, A., Siew, W. L., Tan, Y. A., Nor, A. I., Mohtar, Y., Tang, T. S. and Nuzul, A. I. 2004. *MPOB Test Methods – A compendium of test on palm oil products, palm kernel products, fatty acids, food related products and others*. MPOB, Bangi.
- Liu, M., Li, X. Q., Weber, C., Lee, C. Y., Brown, J. and Liu, R. H. 2002. Antioxidant and antiproliferative activities of raspberries. *Journal of Agricultural and Food Chemistry*, 50: 2926-2930.
- Liang, Z., Sang, M., Fan, P., Wu, B., Wang, L., Yang, S. and Li, S. 2011. CIELAB coordinates in response to berry skin anthocyanin and their composition in *Vitis*. *Journal of Food Science*, 76: 490-97.
- Longchamps, L., Khosla, R. and Westfall, D. G. 2012. Proximal sensing for early detection of nitrogen deficiency in corn for in-season precision nitrogen management. Colorado State University, Fort Collins, Colorado, USA. *Agronomy Journal*, 106(2): 511-518.

- Lorenzen, C. J. 1966. A method for the continuous measurement of *in vivo* chlorophyll concentration. Deep Sea Research and Oceanographic Abstracts, 13: 223-227.
- Malaysian Palm Oil Board. 2003. Oil palm fruit grading manual. Malaysian Palm Oil Board (MPOB). Ministry of Primary Industries, Malaysia, ISBN: 967-961-091-8.
- Marshall, G. F. and Stutz, G. E. 2012. Handbook of optical and laser scanning. Second edition. Taylor and Francis Group, LLC, ISBN: 978-1-4398-0879-5.
- Macheix, J.-J., Fleuriet, A. and Billot, J. 1990. Fruit phenolics. CRC Press. Boca Raton, ISBN: 9780849349683.
- McClure, J. W. 1975. Physiology and functions of flavonoids. In: Harborne, J. B., Mabry, T. J. and Mabry, H. (Eds.). The flavonoids. Chapman and Hall, London, pp: 970-1055.
- Meyer, S., Cartelat, A., Moya, I. and Cerovic, Z. G. 2003. Ultraviolet-induced fluorescence for plant monitoring: present state and prospects. Agronomy: Agriculture and Environment, 1(9): 543-578.
- Merzlyak, M. N. and Chivkunova, O. B. 2000. Light-stress-induced pigments changes and evidence for anthocyanin photoprotection in apples. Journal of Photochemical Photobiology B: Biology, 55: 155-163.
- Merken, H. M. and Beecher, G. R. 2000. Measurement of food flavonoids by high-performance liquid chromatography: a review. Journal of Agriculture Food Chemistry, 48: 577-99.
- Michael, N. D. 2009. Fundamentals of geographic information systems. Fourth edition. John Wiley and Sons, Inc., ISBN: 978-0-470-12906-7.
- Mishra, G. J., Reddy, M. N. and Rana, J. S. 2012. Isolation of flavonoid constituent from *Launaea procumbens* Roxb. by preparative HPTLC method. IOSR Journal of Pharmacy, 2: 2250-3013.
- Mohamad, S. 2007. On the fresh fruit bunch quality, ripeness standard and oil extraction rate. Retrieved on September 2007, from <http://www.mpob.gov.my>.
- Muller, V., Lankes, C., Schmitz-E, M., Noga, G. and Hunsche, M. 2013. Estimation of flavonoid and centelloside accumulation in leaves of *Centella asiatica* L. by multiparametric fluorescence measurements. Journal of Environment and Experimental Botany, 93: 27-34.
- Neill, S. O. and Gould, K. S. 1999. Optical properties of leaves in relation to anthocyanin concentration and distribution. Canadian Journal Botany, 77: 1777-1782.

- Neill, S. O. and Gould, K. S. 2003. Anthocyanins in leaves: light attenuators or antioxidants?. *Functional Plant Biology*, 30: 865-873.
- Ng, S. Y., Abdullah, S. and Chong, K. P. 2013. Phytochemical constituents from leaves of *Elaeis guineensis* and their antioxidant and antimicrobial activities. *International Journal of Pharmacy and Pharmaceutical Sciences*, 5(4): 137-140.
- Ng, S. B. 2009. Understanding palm oil - Malaysian Palm Oil Promotion Council (MPOC), Kuala Lumpur, Malaysia.
- Okoye, M. N., Okwuagwu, C. O. and Uguru M. I. 2009. Population improvement for fresh fruit bunch yield and yield components in oil palm (*Elaeis guineensis* Jacq.). *American-Eurasian Journal of Scientific Research*, 4(2): 59-63.
- Okoye, M. N., Okwuagwu, C. O. and Uguru, M. I. 2008. Biplot analysis of fresh fruit bunch yield and yield components of oil palm (*Elaeis guineensis* Jacq.). *Journal of Applied Bioscience*, 8(1): 288-303.
- Onwuka, G. I. and Akaerue, B. I. 2006. Evaluation of the quality of palm oil produced by different methods of processing. *Research Journal of Biological Sciences*, 1: 16-19.
- Orji, M. U. and Mbata, T. I. 2008. Effect of extraction methods on the quality and spoilage of Nigerian palm oil. *African Journal of Biochemistry Research*, 2(9): 192-196.
- Padilla, F. M., Pena-Fleitas, M. T., Gallardo, M. and Thompson, R. B. 2014. Evaluation of optical sensor measurements of canopy reflectance and of leaf flavonols and chlorophyll contents to assess crop nitrogen status of muskmelon (*Cucumis melo* L. cv. Tezac). *European Journal of Agronomy*, 58: 39-52.
- Patrick, F. D. 2011. Fundamental of sensor for engineering and science. Taylor and Francis Group, LLC, ISBN: 978-1-4398-6103-5.
- Pfündel, E. E., Ben Ghazlen, N., Meyer, S., and Cerovic, Z. G. 2007. Investigating UV screening in leaves by two different types of portable UV fluorimeters reveals in vivo screening by anthocyanins and carotenoids. *Photosynthesis Research*, 93: 205-221.
- Polo-Insfran, D. D., Brenes, C. H. and Talcott, S. T. 2004. Phytochemical composition and pigment stability of acai (*Euterpe oleraceae* Mart.). *Journal of Agriculture Food Chemistry*, 52: 1539-1545.
- Rao, V., Soh, A. C., Corley, R. H. V., Lee, C. H., Rajanaidu, N., Tan, Y. P., Chin, C. W., Lim, K. C., Tan, S. T., Lee, T. P. and Ngui, M. 1983. A critical reexamination of the method of bunch quality analysis in oil palm breeding. PORIM Occasional Paper, No. 9, pp: 28.

- Rafii, M. Y., Rajanaidu, N., Jalani, B. S. and Zakri, A. H. 2001. Genotype x environment interaction and stability analyses in oil palm (*Elaeis guineensis* Jacq.) progenies over six locations. *Journal of Oil Palm Research*, 13(1): 11-41.
- Sambanthamurthi, R., Sundram, K. and Tan, Y. A. 2000. Chemistry and biochemistry of palm oil. *Progress in Lipid Research*, 39: 507-558.
- Sas.com. 2015. SAS/STAT®. 9.3 User's Guide. SAS documentation 2011. Retrieved January 2015, from <http://support.sas.com/documentation/cdl/en/statug/63962/HTML/default/viewer.htm#titlepage.htm>.
- Schauss, A. G., Wu, X., Prior, R. L., Ou, B., Patel, D., Huang, D. and Kababick, J. P. 2006. Phytochemical and nutrient composition of the freeze-dried amazonian palm berry, *Euterpe oleraceae* Mart. (Acai). *Journal of Agricultural Food Chemistry*, 54: 8598-8603.
- Schaefer, H. M. and Rolshausen, G. 2005. Plants on red alert: do insect pay attention?. *Bio-essays*, 28: 65-71.
- Schwinn, K. E. and Davies, K. M. 2004. Flavanoids. In Davies, K. M. (Eds.). *Plant pigments and their manipulation. Annual Plant reviews*. Blackwell Publishing Oxford, pp: 92-149.
- Sivasothy, K., Ma, A. N., Maycock, J. H. and Koichiro, Y. 1992. Combined sterillisation-stripping process: a promising approach for oil quality improvement. *Malaysian Palm Oil Board (MPOB)*, pp: 59-74.
- Siew, W. L. and Monammed, N. 1992. Effect of fruit storage on the palm oil bleachability. *JAOCS*, 69(12).
- Simedarby.com. 2015. Sime Darby Plantation. Retrieved November 2013, from <http://www.simedarbyplantation.com/>.
- Smillie, R. M. and Hetherington, S. E. 1999. Photoabatement by anthocyanin shields photosynthetic systems from light stress. *Photosynthetica*, 36: 451-463.
- Solomon, S. 2010. *Sensors and control systems in manufacturing*. Second edition. The McGraw-Hill Companies, Inc., ISBN: 978-0-07-160572-4.
- Sparnaaij, L. D., Rees, A. R. and Chapas, L. C. 1963. Annual yield variation in the oil palm. *Journal of West African Institute for Oil Palm Research*, 4: 11-125.
- Standards and Industrial Research Institute of Malaysia (SIRIM). 2008. *Malaysian Standard (MS 814:2007)*. Kuala Lumpur, Malaysia.
- Stintzing, F. C. and Carle, R. 2004. Functional properties of anthocyanins and betalains in plants, food and in human nutrition. *Trends Food Science Technology*, 15: 19-38.

- Steyn, W. J., Wand, S. J. E., Holcroft, D. M. and Jacobs, G. 2002. Anthocyanins in vegetative tissues: a proposed unified function in photoprotection. *New Phytology*, 155: 349-361.
- Tang, T. S. and Pantzari, T. P. 2009. Pocketbook of palm oil uses. Malaysian Palm Oil Board (MPOB), ISBN: 978-967-961-151-9.
- Tay, P. Y., Tan, C. P., Faridah, A., Yim, H. S. and Ho, C. W. 2014. Assessment of extraction parameters on antioxidant capacity, polyphenol content, epigallocatechin gallate (EGCG), epicatechin gallate (ECG) and iriflophenone 3-c- β -glucoside of agarwood (*Aquilaria crassna*) young leaves. *Journal of Molecules*, 19(8): 12304-12319.
- Treutter, D. 2005. Significance of flavonoids in plants resistance and enhancement of their biosynthesis. *Plant Biology*, 7: 581-591.
- Van der Vossen, H. A. M. 1974. Plant resources of South East Asia. *Plant Resources of Tropical Africa (PROTA)*, ISBN: 10-9-7983-1600-2.
- Velayuthan, A. 1990. Local premium for better quality palm oil, why not?. *The Planter*, 66: 5-7.
- Wackernagel, H. 2003. *Multivariate geostatistics: an introduction with applications*. Third edition. Berlin Heidelberg New York, ISBN: 3-540-44142-5.
- Webster, R. and Oliver, M. A. 2007. *Geostatistics for environmental scientists*. Second Edition. John Wiley and Sons, Chichester, UK., ISBN-13: 978-0-470-02858-2.
- Wessels-Boer, J. G. 1965. *The indigenous palm of Surinam (PhD)*. University Leiden, Leiden.
- Wikipedia.com. 2015. Chlorophyll. Retrieved on March 2015, from <http://en.wikipedia.org>.
- Willson, M. F. and Whelan, C. J. 1990. The evolution of fruit color in fleshy-fruited plants. *American Naturalist*, 136: 790-809.
- Wroslad, R. E. 2004. Symposium 12: Interaction of natural colors with other ingredients. Anthocyanin pigments - Bioactivity and coloring properties. *Journal of Food Science*, 69: C419-C421.