

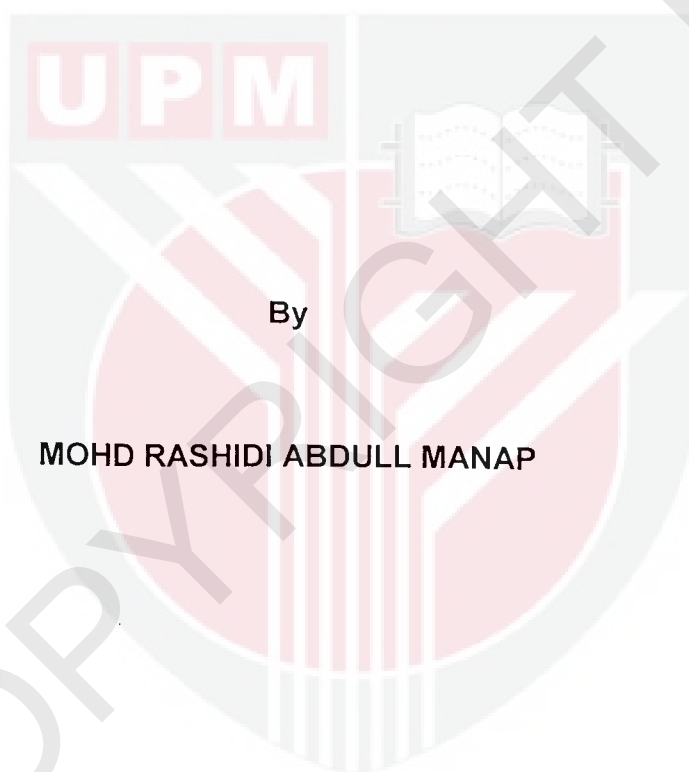


***SOLUTION PHASE SYNTHESIS AND CHARACTERIZATION OF PEPTIDYL
CHEMOSENSORS FOR Pb(II) AND As(III) DETECTION***

MOHD RASHIDI ABDULL MANAP

FS 2012 24

**SOLUTION PHASE SYNTHESIS AND CHARACTERIZATION OF PEPTIDYL
CHEMOSENSORS FOR Pb(II) AND As(III) DETECTION**



By

MOHD RASHIDI ABDULL MANAP

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfillment of the Requirements for the Degree of
Master of Science (Analytical Chemistry)**

July 2012

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

SOLUTION PHASE SYNTHESIS AND CHARACTERIZATION OF PEPTIDYL CHEMOSENSORS FOR Pb(II) AND As(III) DETECTION

By

MOHD RASHIDI ABDULL MANAP

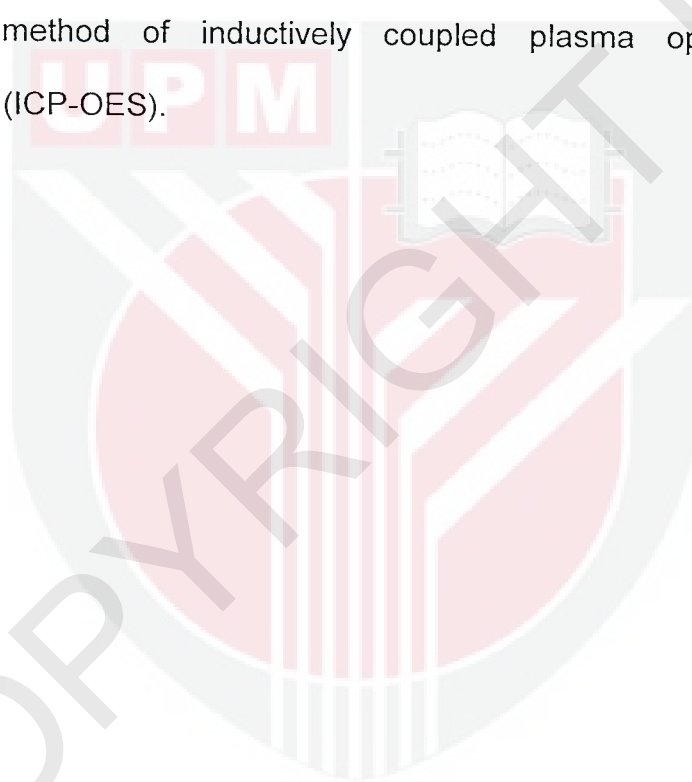
July 2012

Chairman: Nor Azah Yusof, PhD

Faculty: Faculty of Science, UPM.

A new fluorescent dipeptide Dansyl-Ala-Val-OMe (**5**) has been synthesized and characterized. Peptide **5** was synthesized based on sulfonation reaction between dipeptide and 5-(dimethylamino)naphthalene-1-sulfonyl chloride (Dns-Cl) with molecular weight of m/z 435.2. Peptide **5** was obtained as a greenish solid in 80% yield. This compound was in a form of diastereomer in a 1:5 mixture ratio and it was not separated. Peptide **5** was characterized by melting point, IR, NMR, UPLC, GC-MS and HR-MS. Meanwhile another fluorescent dipeptide, Dansyl-Ala-Gly-OH (**6**) was purchased. The fluorescence intensity of peptide **5** was measured at pH 5 with excitation and emission wavelengths of 289 nm and 487 nm, respectively where as the excitation and emission wavelengths of peptide **6** at pH 12 were 331 nm and 524 nm, respectively. Fluorescence difference measurement was used for measurement. Effect of amount of reagent, different pH, dynamic range, reproducibility, interference and validation were studied. 4.0 μM of peptide **5** in a mixture of methanol:water (4:1) was observed to be the optimum concentration for detection of Pb(II). Meanwhile 1.5 μM of peptide **6** in 100% aqueous solution was needed for optimum detection of As(III). The working

medium of peptide **5** is at pH 5, meanwhile peptide **6** at pH 12. Limit of detection (LOD) for detection of Pb(II) and As(III) were calculated to be 0.08 μM and 1.86 μM respectively. The RSD of the reproducibility were 2.03% and 9.66% for 4.00 μM Pb(II) and 7.84 μM As(III). The presence of foreign ions at 1:1 molar ratio of interfering ion: studied metals did not affect the determination of Pb(II) and As(III) based on the low percentage of interference. The developed detection systems were validated with the established method of inductively coupled plasma optical emission spectrometry (ICP-OES).



© COPYRIGHT

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**SINTESIS FASA LARUTAN
DAN PENCIRIAN PEPTIDA PNDARFLUOR PENGESAN KIMIA
UNTUK PENGESANAN Pb (II) DAN As(III)**

Oleh

MOHD RASHIDI ABDULL MANAP

Julai 2012

Pengerusi: Nor Azah Yusof, PhD

Fakulti: Fakulti Sains,UPM.

Satu dipeptida pendarfluor baru iaitu Dansyl-Ala-Val-OMe (**5**) telah disintesis dan dicirikan. Peptida **5** telah disintesis berdasarkan tindak balas persulfonan antara dipeptida dan 5 - (dimetilamino) naftalena-1-sulfonil klorida (Dns-Cl) dengan berat molekul m/z 435.2. Peptida **5** telah diperolehi sebagai pepejal kehijauan dalam 80% hasil. Peptida **5** adalah dalam bentuk diastereomer dalam nisbah campuran 1:5 dan ia tidak diasingkan. Peptida **5** dicirikan menggunakan takat lebur, NMR, UPLC, GC-MS dan HR-MS. Sementara itu, satu lagi dipeptida pendarfluor, Dansyl-Ala-Gly-OH (**6**) telah dibeli. Keamatan pendarfluor peptida **5** adalah diukur pada pH 5 dengan pengujaan dan pelepasan jarak gelombang 289 nm dan 487 nm, masing-masing di mana panjang gelombang pengujaan dan pancaran peptida **6** pada pH 12 ialah 331 nm dan 524 nm, masing-masing. Perbezaan pengukuran pendarfluor telah digunakan untuk pengukuran. Kesan jumlah reagen, perbezaan pH, julat dinamik, keboleholangan gangguan dan kajian pengesanan telah di kaji. Untuk jumlah reagen, 4 μ M peptida **5** dalam campuran metanol: air (4:1) adalah kepekatan optimum untuk mengesan

Pb(II). Sementara itu, hanya 1.5 μM peptida **6** dalam 100% larutan akueus diperlukan untuk pengesanan optimum As(III). Medium kerja peptida **5** ialah pH 5 dan peptida **6** ialah pH 12. Had pengesanan (LOD) untuk pengesanan Pb(II) dan As(III) telah ditentukan; 0.08 μM dan 1.86 μM masing-masing. RSD kebolehlulangan gangguan ialah 2.03% dan 9.66% untuk 4.00 μM Pb(II) dan 7.84 μM As(III). Kehadiran ion asing pada nisbah 1:1 molar ion pengganggu atau Pb(II) dan peptida **6**-As(III) ion pengganggu tidak menjejaskan penentuan Pb(II) dan As(III) berdasarkan peratusan rendah gangguan. Sistem pengesanan yang dibangunkan telah disahkan dengan kaedah induktif plasma berganding aruhan spektroskopi pemancaran optik (ICP-OES) yang sedia ada.

ACKNOWLEDGEMENTS

First of all, all praise is to the almighty where by His Will I managed to accomplish this research work. My special thanks to my supervisory committee members namely, Prof. Dr. Faujan Ahmad, Assoc. Prof. Dr. Nor Azah Yusof and Dr.Siti Mariam Mohd Nor for all the supervision and fruitful discussions which has guided me throughout this research work.

Last but not least, Ramizah and Abu Rayyan for the unconditional supports which forced me to accomplish this research work.

I certify that a Thesis Examination Committee has met on 30 July 2012 to conduct the final examination of Mohd Rashidi bin Abdull Manap on his thesis entitled "Solution Phase Synthesis and Characterization of Peptidyl Chemosensors for Pb (II) and As (III) Detection" in accordance with the Universities and University College Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Mohamed Ibrahim bin Mohamed Tahir, PhD

Senior Lecturer
Faculty of Science
Universiti Putra Malaysia
(Chairman)

Emilia binti Abd Malek, PhD

Senior Lecturer
Faculty of Science
Universiti Putra Malaysia
(Internal Examiner)

Bimo Ario Tejo, PhD

Senior Lecturer
Faculty of Science
Universiti Putra Malaysia
(Internal Examiner)

Wan Aini Wan Ibrahim, PhD

Professor
Faculty of Science
Universiti Teknologi Malaysia
(External Examiner)



SEOW HENG FONG, PhD
Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

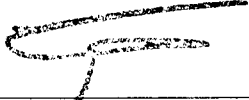
Date: 27 August 2012

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Nor Azah Yusof, PhD
Associate Professor
Faculty of Science
Universiti Putra Malaysia
(Chairman)

Faujan H. Ahmad, PhD
Professor
Faculty of Science
Universiti Putra Malaysia
(Member)

Siti Mariam Mohd Nor, PhD
Lecturer
Faculty of Science
Universiti Putra Malaysia
(Member)



BUJANG BIN KIM HUAT, PhD
Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 13 SEP 2012

DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.



MOHD RASHIDI ABDULL MANAP

Date: 30 May 2012

LIST OF ABBREVIATIONS

app d	apparently doublet (spectral)
dd	double of doublet (spectral)
Boc	<i>tert</i> -butoxycarbonyl
br	broad
br s	broad singlet
°C	degrees Celsius
cm ⁻¹	wavenumber(s)
COSY	correlation spectroscopy
δ	chemical shift in parts per million downfield from tetramethylsilane
d	doublet (spectral)
dansyl	5-(dimethylamino)-1-naphthalenesulfonyl
DCC	<i>N,N'</i> -dicyclohexylcarbodiimide
DCM	dichloromethane
DEPT	distortionless enhancement by downfield from tetramethylsilane
EDTA	ethylenediaminetetraacetic acid
equiv	equivalent
FT	Fourier transform
g	gram(s)
GC	gas chromatography
HMBC	Heteronuclear multiple bond correlation
HMQC	Heteronuclear multiple quantum correlation
HPLC	High-performance liquid chromatography
HRMS	High-resolution mass spectrometry
Hz	hertz
IR	infrared
<i>J</i>	coupling constant (in NMR)
L	liter(s)
LOD	Limit of detection
LOQ	Limit of quantification
μ	micro
m	multiplet (spectral); mili
M	molar (moles per liter)
M ⁺	parent molecular ion
Me	methyl
MHz	megahertz
mL	milliliter
mM	millimolar (millimoles per liter)
mol	mole(s); molecular (as in mol wt)
mp	melting point
MS	mass spectrometry
MW	mol wt molecular weight
<i>m/z</i>	mass-to-charge ratio
NMR	Nuclear magnetic resonance
NOESY	Nuclear overhauser effect spectroscopy
ppb	part(s) per billion
ppm	part(s) per million

rt	room temperature
s	singlet (spectral)
t	triplet (spectral)
temp	temperature
TLC	thin-layer chromatography
TMS	trimethylsilyl; tetramethylsilane
UPLC	Ultra-performance liquid chromatography
t_R	retention time (in chromatography)
UV	ultraviolet
vis	visible



LIST OF TABLES

	Page
Table 1. ICP-OES condition	32
Table 2. Comparison ^1H and ^{13}C of dipeptide 3 in CDCl_3 with reference	42
Table 3. ^1H and ^{13}C NMR data of peptide 5 in CDCl_3 (500 MHz)	74
Table 4. Solubility of peptide 5 in various solvents at 25.0 °C and 60.0 °C	80
Table 5. Determination of Pb(II) in electroplating waste using peptide 5	90
Table 6. Determination of As(III) in electroplating waste using peptide 6	100
Table 7. Comparison of Pb(II) detection limit obtained in the present study with the reported fluorophores	100
Table 8. Comparison of As(III) detection limit obtained in the present study with the reported fluorophores	101
Table 9. Fluorescence data at 487 nm and statistical analysis of (5) for pH study	121
Table 10. Fluorescence intensity at 487 nm of (5) for amount of reagent	122
Table 11. Statistical analysis of (5) for amount of reagent	122
Table 12. Fluorescence data at 487 nm and statistical analysis of (5) for dynamic range	123
Table 13. Fluorescence data at 487 nm and statistical analysis of (5) for reproducibility	124
Table 14. Fluorescence data at 524 nm and of (5) for Interference Study	125
Table 15. Statistical analysis of (5)- for Interference Study	125
Table 16. Percentage interference of (5)	126
Table 17. ICP-OES data of (5) for Pb(II)	127

Table 18. Statistical analysis at 487 nm of (5) for Validation Test	127
Table 19. Fluorescence data at 524 nm and statistical analysis of (6) for pH study	129
Table 20. Fluorescence intensity at 524 nm of (6) for amount of reagent	130
Table 21. Statistical analysis of (6) for amount of reagent	130
Table 22. Fluorescence data at 524 nm and statistical analysis of (6) for dynamic range	131
Table 23. Fluorescence data at 524 nm and statistical analysis of (6) for reproducibility	132
Table 24. Fluorescence data at 524 nm and of (6) for Interference Study	133
Table 25. Statistical analysis of (6) for Interference Study	133
Table 26. Percentage Interference of (6)	134
Table 27. ICP-OES data of (6) for As(III)	135
Table 28. Statistical analysis at 524 nm of (6) for Validation Test	135

LIST OF FIGURES

	Page
Figure 1. Fabrication of MPA-Gly-Gly-His gold electrode and complexation with Cu(II)	10
Figure 2. Structure of Ionophore I and Ionophore II	12
Figure 3. Dansyl chloride or 5-(dimethylamino)naphthalene-1-sulfonyl chloride	19
Figure 4. Sulfonamide bond	20
Figure 5. Dansyl derivative	22
Figure 6. Molecular structure of dipeptide 3	42
Figure 7. ^{13}C of dipeptide 3 in CDCl_3	45
Figure 8. ^1H spectrum of dipeptide 3 in CDCl_3	46
Figure 9. Expansion of ^1H spectrum of dipeptide 3 in between 0.58 ppm to 1.87 ppm	47
Figure 10. Expansion of ^1H spectrum of dipeptide 3 in between 3.39 ppm to 7.24 ppm	48
Figure 11. FT-IR spectrum of dipeptide 3	50
Figure 12. GCMS spectrum of dipeptide 3	51
Figure 13. FTIR spectrum of peptide 4	53
Figure 14. DIMS of hydrochloride salt 4	54
Figure 15. Expansion of ^1H spectrum showing diastereomer of peptide 5 at δ_{H} 8.47 ppm and δ_{H} 8.57 ppm	57
Figure 16. Expansion of ^1H spectrum of peptide 5 in Dansyl region	58
Figure 17. ^{13}C DEPT 90 and DEPT 135 spectra of peptide 5	59

Figure 18. Expansion of ^{13}C spectrum of peptide 5 in Dansyl region	60
Figure 19. Molecular structure of peptide 5	61
Figure 20. COSY (—) correlations of peptide 5	62
Figure 21. Expansion of HMQC spectrum of peptide 5 in Dansyl region	63
Figure 22. FTIR spectrum of peptide 5	64
Figure 23. COSY spectrum of peptide 5	65
Figure 24. Expansion of COSY spectrum of peptide 5 in Dansyl region	66
Figure 25. Selected HMBC (—) correlations of peptide 5	68
Figure 26. HMBC spectrum of peptide 5	69
Figure 27. HMQC spectrum of peptide 5	70
Figure 28. Expansion of HMQC spectrum of peptide 5 at δ_{H} 2.81 ppm	71
Figure 29. ^{13}C spectrum of peptide 5 in CDCl_3	72
Figure 30. ^1H spectrum of peptide 5 in CDCl_3	73
Figure 31. Molecular structure of peptide 5	74
Figure 32. GCMS of peptide 5	76
Figure 33. Selective fragmentation ion of 5-Dimethylamino-naphthalene-1-sulfonic acid diethylamide	77
Figure 34. GCMS of 5-Dimethylamino-naphthalene-1-sulfonic acid diethylamide	78
Figure 35. UPLC Chromatogram of 100 mg/L of peptide 5 at a) 289 nm and b) 327 nm in methanol	79
Figure 36. UV absorption spectrum of peptide 5 (10 μM , 10 mM HEPES, pH 5) in methanol/water mixture (4:1)	82

Figure 37. Fluorescence emission spectrum of peptide 5 (10 μM , 10 mM HEPES, pH 5) in methanol/water mixture (4:1). Excitation at 289 nm	83
Figure 38. Fluorescence emission spectrum of peptide 5 (10 μM , 10 mM HEPES, pH 5) with addition of metal ions (10 μM) in methanol/water mixture (4:1)	84
Figure 39. Fluorescence intensity of peptide 5 (10 μM , 10 mM HEPES) at different pH values	86
Figure 40. Fluorescence difference of peptide 5 (1, 2, 4, 12, 18 μM) with 10 μM of Pb(II) at pH 5	87
Figure 41. Calibration plot of peptide 5 (4 μM , 4 mM HEPES, pH 5) at different concentrations of Pb(II)	88
Figure 42. Effect of interfering ions on detection Pb(II) using peptide 5	89
Figure 43. Molecular structure of peptide 6	91
Figure 44. UV absorption spectrum of peptide 6 (10 μM , 10 mM HEPES, pH 5) in water	92
Figure 45. Fluorescence emission spectra of water and peptide 6 (10 μM , 10 mM HEPES, pH 5) in water. Excitation at 331 nm	93
Figure 46. Fluorescence emission spectrum of peptide 6 (10 μM , 10 mM HEPES, pH 5) with addition of metal ions (10 μM) in water	94
Figure 47. Fluorescence intensity of peptide 6 (5 μM , 5 mM HEPES) at different pH values	95
Figure 48. Fluorescence difference of peptide 6 (1 μM to 3 μM) with 10 μM of As(III) at pH 12	96

Figure 49. Calibration plot of peptide **6** (1.5 μM , 1.5 mM HEPES, pH 12) at different concentrations of As(III) 97

Figure 50. Effect of interfering ions on detection As(III) using peptide **6** 99



LIST OF SCHEMES

	Page
Scheme 1. General formation of amide bond	17
Scheme 2. Fragmentation ion of dipeptide 3	51
Scheme 3. Fragmentation ion of hydrochloride salt 4	54
Scheme 4. Dansylation	55
Scheme 5. Fragmentation ion of peptide 5	76
Scheme 6. Proposed binding mode of peptide 5 with Pb(II)	85
Scheme 7. Proposed binding mode of peptide 6 with As(III)	94

LIST OF APPENDICES

	Page
Appendix 1. GCMS of dipeptide 3	112
Appendix 3. DIMS of hydrochloride salt of 4	116
Appendix 4. HRMS of peptide 5	117
Appendix 5. UPLC-TUV parameters of peptide 5	118
Appendix 6. ^1H spectrum of Dansyl Chloride	119
Appendix 7. DIMS of Dansyl Chloride	126

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF ABBREVIATIONS	ix
LIST OF TABLES	xi
LIST OF FIGURES	xiii
LIST OF SCHEMES	xvii
LIST OF APPENDICES	xviii
CHAPTER	
INTRODUCTION	1
Research Problem	1
Objectives	3
LITERATURE REVIEW	4
Heavy Metals in the Environment	4
Amino acids and peptides in Metal Ion Analysis	5
Short Peptide	8
Cyclic Peptide	11
Oligopeptide	12
Synthesis of Peptide <i>via</i> Solution Phase	15
Sulfonation of Fluorescent Peptide	19
Compound Profiling	24
Dansyl Fluorescent for Metal Ion Detection	25
MATERIALS AND METHODOLOGY	28
General	28
Synthesis	32
General	32
Characterization	34
RESULTS AND DISCUSSION	41
Synthesis of dipeptide, Boc-Ala-Val-OMe (3)	41
Deprotection of Boc group HCl. H ₂ N-Ala-Val-OMe (4)	52
Synthesis of dansyl-dipeptide, Dansyl-Ala-Val-OMe (5)	55
UV Absorption Spectrum	81
Emission Spectrum	82
Metal Screening	83

pH Study	85
Peptide Concentration	86
Dynamic Range	87
Reproducibility	88
Interference	89
Validation	90
Dansyl-Ala-Gly-OH (6)	90
UV Absorption Spectrum	91
Emission Spectra	92
Metal Screening	93
pH Study	95
Peptide Concentration	96
Dynamic Range	97
Reproducibility	98
Interference	98
Validation	99
Comparison with previous works	100
CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	102
REFERENCES/BIBLIOGRAPHY	104
APPENDICES	112
BIODATA OF STUDENT	137
LIST OF PUBLICATIONS	138

CHAPTER 1

INTRODUCTION

Research Problem

Heavy metal ions such as As(III), Pb(II), Zn(II), Ni(II) and Hg(II) are easily found in the environment especially in industrial waste, agriculture and mining industry. These inorganic chemicals are usually present at low concentration in the environment either free or complexed with organic or inorganic ligand (Pesavento et al., 2009).

Unlike organic contaminants, these inorganic chemicals have high solubility in the aquatic environments. As a result, it can easily penetrate the food chain and stored in living organism. Moreover, these inorganic chemicals are non-biodegradable in nature, and on top of that it has long biological half life for elimination from body. It results in negative major problems to living organisms (Chen et al., 2008). Fu and Wang (2011) reported on a few heavy metal ions such as Zn(II), Ni(II), Cd(II) and Pb(II) which affect human's health such as vomiting, kidney failure and skin dermatitis.

Electronic configuration of metal ions are different from each other based on its oxidation state. Based on this properties, some of the metals can give spectroscopic due to its electronic configuration. However, some of them can not give spectroscopic signal.

For example, Zn(II) can not be detected directly using common analytic techniques such as UV-Vis spectroscopy or nuclear magnetic resonance (NMR) due to its $3d^{10}4s^0$ electronic configuration (Jiang and Guo, 2004). Thus the fluorescence method stands out as a method of choice. Fluorescent method offers high sensitivity compared to other methods.

Dansyl groups were widely used as fluorophore due to its intense absorption in the UV region and emits in the visible range with high fluorescence quantum yield. In addition, the dansyl molecule can undergo sulfonation reaction and it is suitable for the synthesis of peptide derivative (Chao et al., 2011). There were many fluorescent peptides that have been synthesized *via* Solid Phase Peptide Synthesis (SPPS) and used as a chemosensor (Joshi et al., 2007; White and Holcombe, 2007). Solution phase offers fast preparation of a small sequence of peptide and fast synthesis, meanwhile SPPS is most suitable for bulky and large scale of peptide.

It has been described that peptide formed complex with metal ion (Chekmeneva et al., 2006). There are a great number of potential donor atoms through the peptide backbone such as on nitrogen atom, oxygen atom, side chain, carbonyl on terminal or amide hydrogen. Owing to its great structural variety, peptide is suitable for complex formation. Metal binding is achieved with a high degree of selectivity using peptide motifs. Glycine and valine were chosen because of the non polar side chains. Glycine side chain is weakly hydrophobic meanwhile

valine side chain is strongly hydrophobic. This properties make the chemosensor works in different medium of solvents.

Therefore, the idea of this research is to synthesize a peptide which consists of a fluorophore which acts as a recognition molecule in metal ions detection. The fluorescent peptide will be synthesized *via* solution phase method.

Two different fluorescent peptides (synthesized and purchased) were tested as recognition molecule for analysis in different mediums (mixture solvent and single solvent). Several parameters were studied such as selectivity, pH, reagent concentration, dynamic range, reproducibility, interference and validation.

Objectives

The objectives of this research are:

- 1)To synthesize a fluorescent dipeptide *via* solution phase method
- 2)To characterize the synthesized fluorescent dipeptide by spectroscopic methods
- 3)To characterize fluorescent dipeptides for metal ion detection

REFERENCES/BIBLIOGRAPHY

- Adam, V., Hanustiak, P., Krizkova, S., Beklova, M., Zehnalek, J., Trnkova, L., Horna, A., Sures, B., & Kizek, R. (2007). Palladium biosensor. *Electroanalysis*, 19(18), 1909-1914.
- Addy, M. E., Steinman, G., & Mallette, M. F. (1973). Selectivity in aqueous peptide synthesis by 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide: A model for primordial processes. *Biochimica Et Biophysica Acta (BBA) - Protein Structure*, 295(2), 385-395.
- Adelaju, S. B., Young, T. M., Jagner, D., & Batley, G. E. (1999). Constant current cathodic stripping potentiometric determination of arsenic on a mercury film electrode in the presence of copper ions. *Analytica Chimica Acta*, 381(2-3), 207-213.
- Aguilar, A. D., Forzani, E. S., Li, X., Tao, N., Nagahara, L. A., Amlani, I., & Tsui, R. (2005). Chemical sensors using peptide-functionalized conducting polymer nanojunction arrays. *Applied Physics Letters*, 87(19), 193108-1-193108-3.
- Akta Kualiti Alam Sekeliling* 1974. (2009). Retrieved 07/17, 2012, from [http://www.doe.gov.my/files/multimedia141/EFLUEN PERINDUSTRI AN 2009 1.pdf](http://www.doe.gov.my/files/multimedia141/EFLUEN_PERINDUSTRI_AN_2009_1.pdf)
- Anari, M. R., Bakhtiar, R., Zhu, B., Huskey, S., Franklin, R. B., & Evans, D. C. (2002). Derivatization of ethinylestradiol with dansyl chloride to enhance electrospray ionization: Application in trace analysis of ethinylestradiol in rhesus monkey plasma. *Analytical Chemistry*, 74(16), 4136-4144.
- Barakat, M. A. (2011). New trends in removing heavy metals from industrial wastewater. *Arabian Journal of Chemistry*, 4(4), 361-377.
- Bartzatt, R. (2001). Fluorescent labeling of drugs and simple organic compounds containing amine functional groups, utilizing dansyl chloride in Na₂CO₃ buffer. *Journal of Pharmacological and Toxicological Methods*, 45(3), 247-253.
- Bi, X., Agarwal, A., & Yang, K. (2009). Oligopeptide-modified silicon nanowire arrays as multichannel metal ion sensors. *Biosensors and Bioelectronics*, 24(11), 3248-3251.
- Bok, J. H., Kim, H. J., Lee, J. W., Kim, S. K., Choi, J. K., Vicens, J., & Kim, J. S. (2006). Selective metal detection in an unsymmetrical 1,3-alternate calix[4]biscrown chemosensor. *Tetrahedron Letters*, 47(8), 1237-1240.

- Bowman, L., Spencer, D., Muntele, C., Muntele, I., & Ila, D. (2007). Cyclic voltammetry and RBS study of paint components. *Nuclear Instruments and Methods in Physics Research Section B: Beam Interactions with Materials and Atoms*, 261(1-2), 557-560.
- Cardona, C. M., Alvarez, J., Kaifer, A. E., McCarley, T. D., Pandey, S., Baker, G. A., Bonzagni, N. J., & Bright, F. V. (2000). Dendrimers functionalized with a single fluorescent dansyl group attached "Off Center"; Synthesis and photophysical studies. *Journal of the American Chemical Society*, 122(26), 6139-6144.
- Chao, H., Su, B., Li, C., Lam, C., & Feng, X. (2011). Mono-, di-, and tri-nuclear gold(I) complexes with dansyl moiety: Synthesis, characterization, and photophysical properties. *Inorganic Chemistry Communications*, 14(9), 1436-1439.
- Chapman, D. I., Chapman, J. R., & Clark, J. (1972). Synthesis and mass spectra of dansyl derivatives of some physiologically important β -phenylalkylamine derivatives. *International Journal of Biochemistry*, 3(13), 66-72.
- Chekmeneva, E., Díaz-Cruz, J. M., Ariño, C., & Esteban, M. (2006). Identification of heavy metal complexes of a hexapeptide inhibitor of the human immunodeficiency virus integrase protein by using a voltammetric approach. *Analytical Biochemistry*, 348(2), 252-258.
- Chen, T., Liu, X., Zhu, M., Zhao, K., Wu, J., Xu, J., & Huang, P. (2008). Identification of trace element sources and associated risk assessment in vegetable soils of the urban-rural transitional area of Hangzhou, China. *Environmental Pollution*, 151(1), 67-78.
- Chersi, A., Giommi, S., & Rosanò, L. (2000). Selective 'in synthesis' labeling of peptides with biotin and rhodamine. *Biochimica Et Biophysica Acta (BBA) - General Subjects*, 1474(2), 196-200.
- Chow, E., Wong, E. L. S., Böcking, T., Nguyen, Q. T., Hibbert, D. B., & Gooding, J. J. (2005). Analytical performance and characterization of MPA-Gly-Gly-His modified sensors. *Sensors and Actuators B: Chemical*, 111-112, 540-548.
- Clarke, R. J., & Oprysa, A. (2004). Fluorescence and light scattering. *Journal of Chemical Education*, 81(5), 705-706.
- Costa, S. P. G., Oliveira, E., Lodeiro, C., & Raposo, M. M. M. (2007). Synthesis, characterization and metal ion detection of novel fluoroionophores based on heterocyclic substituted alanines. *Sensors*, 7(10), 2096-2114.

- Cruces, B. C., Segura Carretero, A., Merino Boyle, E., & Fernández Gutiérrez, A. (1999). The use of dansyl chloride in the spectrofluorimetric determination of the synthetic antioxidant butylated hydroxyanisole in foodstuffs. *Talanta*, 50(5), 1099-1108.
- Dabrowski, A., Hubicki, Z., Podkoscielny, P., & Robens, E. (2004). Selective removal of the heavy metal ions from waters and industrial wastewaters by ion-exchange method. *Chemosphere*, 56(2), 91-106.
- Dahiya, R., & Kumar A. (2008). Synthetic and biological studies on a cyclopolypeptide of plant origin. *Journal of Zhejiang University SCIENCE B*, 9(5), 391-400.
- Davis, B. A. (1978). Dansylation of amines, phenolic and catecholic amines and amino acids in aprotic solvents. *Journal of Chromatography A*, 151(2), 252-255.
- Deo, S., & Godwin, H. A. (2000). A selective, ratiometric fluorescent sensor for Pb^{2+} . *Journal of the American Chemical Society*, 122(1), 174-175.
- Fontana, E., & Pignatti, A. (2002). Synthesis of Brostallicin (PNU-166196A) labelled with 2H and ^{14}C . *Journal of Labelled Compounds and Radiopharmaceuticals*, 45(11), 899-909.
- Forzani, E. S., Zhang, H., Chen, W., & Tao, N. (2005). Detection of heavy metal ions in drinking water using a high-resolution differential surface plasmon resonance sensor. *Environmental Science & Technology*, 39(5), 1257-1262.
- Frei, H. M., Frei, R. W., & Hutzinger, O. (1973). Determination of hydroxybiphenyls as dansyl derivatives. *Journal of Chromatography A*, 79, 209-216.
- Fu, F., & Wang, Q. (2011). Removal of heavy metal ions from wastewaters: A review. *Journal of Environmental Management*, 92(3), 407-418.
- Gadd, G. M. (1992). Metals and microorganisms: A problem of definition. *FEMS Microbiology Letters*, 100(1-3), 197-203.
- Ghaneolhosseini, H., Tjarks, W., & Sjöberg, S. (1998). Synthesis of novel boronated acridines and spermidines as possible agents for BNCT. *Tetrahedron*, 54(15), 3877-3884.
- Godwin, H. A., & Berg, J. M. (1996). A fluorescent zinc probe based on metal-induced peptide folding. *Journal of the American Chemical Society*, 118(27), 6514-6515.

- Gros, C., & Labouesse, B. (1969). Study of the dansylation reaction of amino acids, peptides and proteins. *European Journal of Biochemistry*, 7(4), 463-470.
- Hassan, S. S. M., Elnemma, E. M., & Mohamed, A. H. K. (2005). Novel potentiometric copper (II) selective membrane sensors based on cyclic tetrapeptide derivatives as neutral ionophores. *Talanta*, 66(4), 1034-1041.
- Hayman, A. R., & Gray, D. O. (1987). O-acetyethanolamine a natural product from the leguminosae. *Phytochemistry*, 26(3), 839-841.
- Hepel, M., Dallas, J., & Noble, M. D. (2008). Interactions and reactivity of Hg(II) on glutathione modified gold electrode studied by EQCN technique. *Journal of Electroanalytical Chemistry*, 622(2), 173-183.
- Hioki, H., Kinami, H., Yoshida, A., Kojima, A., Kodama, M., Takaoka, S., Ueda, K., & Katsu, T. (2004). Synthesis of N-substituted cyclic triglycines and their response to metal ions. *Tetrahedron Letters*, 45(5), 1091-1094.
- Honour, J. W., Lim, C. K., & Mitchell, F. L. (1983). Metabolic and drug profiling. *Journal of Pharmaceutical and Biomedical Analysis*, 1(2), 127-142.
- Hsu, S., Hsieh, H., Chen, C., Tseng, C., Huang, S., Huang, C., Huang, Y., Radashevsky, V., & Lin, S. (2011). Tungsten and other heavy metal contamination in aquatic environments receiving wastewater from semiconductor manufacturing. *Journal of Hazardous Materials*, 189(1-2), 193-202.
- Hu, Z., Lin, C., Wang, X., Ding, L., Cui, C., Liu, S., & Lu, H. Y. (2010). Highly sensitive and selective turn-on fluorescent chemosensor for Pb²⁺ and Hg²⁺ based on a rhodamine-phenylurea conjugate. *Chemical Communications*, (21), 3765-3767.
- Jensen, O. E., Lawesson, S. D., Bardi, R., Piazzesi, A. M., & Toniolo, C. (1985). Studies on amino acids and peptides VIII : Synthesis and crystal structure of two monothiated analogues of Boc-Gly-S-Ala-Aib-OMe. *Tetrahedron*, 41(23), 5595-5606.
- Jiang, P., & Guo, Z. (2004). Fluorescent detection of zinc in biological systems: recent development on the design of chemosensors and biosensors. *Coordination Chemistry Reviews*, 248(1-2), 205-229.
- Joshi, B. P., Cho, W., Kim, J., Yoon, J., & Lee, K. (2007). Design, synthesis, and evaluation of peptidyl fluorescent probe for Zn²⁺ in aqueous solution. *Bioorganic & Medicinal Chemistry Letters*, 17(23), 6425-6429.

- Joshi, B. P., & Lee, K. (2008). Synthesis of highly selective fluorescent peptide probes for metal ions: Tuning selective metal monitoring with secondary structure. *Bioorganic & Medicinal Chemistry*, 16(18), 8501-8509.
- Kaneko, E., Isoe, J., Iwabuchi, T., Hoshi, S., Akatsuka, K., & Yotsuyanagi, T. (2002). In-vessel extraction using a microtube and its application to the fluorimetric determination of trace lead. *Analyst*, 127(2), 219-222.
- Kohtani, M., Jarrold, M. F., Wee, S., & O'Hair, R. A. J. (2004). Metal ion interactions with polyalanine peptides. *The Journal of Physical Chemistry B*, 108(19), 6093-6097.
- Koneswaran, M., & Narayanaswamy, R. (2009). L-Cysteine-capped ZnS quantum dots based fluorescence sensor for Cu²⁺ ion. *Sensors and Actuators B: Chemical*, 139(1), 104-109.
- Krzysiak, A. J., Scott, S. A., Hicks, K. A., Fierke, C. A., & Gibbs, R. A. (2007). Evaluation of protein farnesyltransferase substrate specificity using synthetic peptide libraries. *Bioorganic & Medicinal Chemistry Letters*, 17(20), 5548-5551.
- Lee, M. H., Kim, H. J., Yoon, S., Park, N., & Kim, J. S. (2008). Metal ion induced FRET OFF-ON in tren/dansyl-appended Rhodamine. *Organic Letters*, 10(2), 213-216.
- Li, B., Bemish, R., Buzon, R. A., Chiu, C. K., Colgan, S. T., Kissel, W., Le, T., Leeman, K. R., Newell, L., & Roth, J. (2003). Aqueous phosphoric acid as a mild reagent for deprotection of the *t*-butoxycarbonyl group. *Tetrahedron Letters*, 44(44), 8113-8115.
- Liu, Q., Yan, X., Guo, J., Wang, D., Li, L., Yan, F., & Chen, L. (2009). Spectrofluorimetric determination of trace nitrite with a novel fluorescent probe. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 73(5), 789-793.
- Lohani, C. R., Kim, J. M., & Lee, K. (2011). Two dansyl fluorophores bearing amino acid for monitoring Hg²⁺ in aqueous solution and live cells. *Tetrahedron*, 67(22), 4130-4136.
- Mahoney, M. E., Oliver, A., Einarsdóttir, O., & Konopelski, J. P. (2009). Synthesis of a cyclic pentapeptide mimic of the active site his-tyr cofactor of cytochrome c oxidase. *The Journal of Organic Chemistry*, 74(21), 8212-8218.
- Majid, S., Rhazi, M. E., Amine, A., & Brett, C. M. A. (2002). An amperometric method for the determination of trace mercury(II) by formation of complexes with tyrosine. *Analytica Chimica Acta*, 464(1), 123-133.

- Marce, M., Brown, D.S., Capell, T., Figueras, X., Tiburcio, A.F. (1995). Rapid high-performance liquid chromatographic method for the quantitation of polyamines as their dansyl derivatives: Application to plant and animal tissues. *Journal of Chromatography B: Biomedical Applications*, 666(2), 329-335.
- Mays, D. E., & Hussam, A. (2009). Voltammetric methods for determination and speciation of inorganic arsenic in the environment—A review. *Analytica Chimica Acta*, 646(1-2), 6-16.
- Metivier, R., Leray, I., & Valeur, B. (2004). Photophysics of calixarenes bearing two or four dansyl fluorophores: charge, proton and energy transfers. *Photochemical & Photobiological Sciences*, 3(4), 374-380.
- Meybeck, M., Lestel, L., Bonté, P., Moilleron, R., Colin, J. L., Rousselot, O., Hervé, D., de Pontevès, C., Grosbois, C., & Thévenot, D. R. (2007). Historical perspective of heavy metals contamination (Cd, Cr, Cu, Hg, Pb, Zn) in the Seine River basin (France) following a DPSIR approach (1950–2005). *Science of the Total Environment*, 375(1–3), 204-231.
- Morse, D., & Horecker, B. L. (1966). Thin-layer chromatographic separation of DNS-amino acids. *Analytical Biochemistry*, 14(3), 429-433.
- Ozay, O., Ekici, S., Baran, Y., Kubilay, S., Aktas, N., & Sahiner, N. (2010). Utilization of magnetic hydrogels in the separation of toxic metal ions from aqueous environments. *Desalination*, 260(1-3), 57-64.
- Parker, K. J., Kumar, S., Pearce, D. A., & Sutherland, A. J. (2005). Design, synthesis and evaluation of a fluorescent peptidyl sensor for the selective recognition of arsenite. *Tetrahedron Letters*, 46(41), 7043-7045.
- Pesavento, M., Alberti, G., & Biesuz, R. (2009). Analytical methods for determination of free metal ion concentration, labile species fraction and metal complexation capacity of environmental waters: A review. *Analytica Chimica Acta*, 631(2), 129-141.
- Reshetova, O. S., Onoprienko, V. V., Rozynov, B. V., & Kozmin, Y. P. (1987). Analysis of N-dansyl peptide methyl esters by means high performance liquid chromatography and mass spectrometry. *Bioorganicheskaya Khimiya*, 13(3), 320-337.
- Sanagi, M. M., Ling, S. L., Nasir, Z., Hermawan, D., Ibrahim, W. A., & Abu Naim, A. (2009). Comparison of signal-to-noise, blank determination, and linear regression methods for the estimation of detection and quantification limits for volatile organic compounds by gas chromatography. *Journal of AOAC International*, 92(6), 1833-1838.

- Satofuka, H., Amano, S., Atomi, H., Takagi, M., Hirata, K., Miyamoto, K., & Imanaka, T. (1999). Rapid method for detection and detoxification of heavy metal ions in water environments using phytochelation. *Journal of Bioscience and Bioengineering*, 88(3), 287-292.
- Shibnev, V. A., & Mar'yash, L. I. (1973). Synthetic analogs of histones and protamines. *Russian Chemical Bulletin*, 22(2), 413-417.
- Sigel, H., Da Costa, C. P., & Martin, R. B. (2001). Interactions of lead(II) with nucleotides and their constituents. *Coordination Chemistry Reviews*, 219-221(0), 435-461.
- So, J., Kang, S., & Kim, B. (1998). Lipase-catalyzed synthesis of peptides containing d-amino acid. *Enzyme and Microbial Technology*, 23(3), 211-215.
- Srinivasa, G. S., & Govil, P. (2008). Distribution of heavy metals in surface water of Ranipet industrial area in Tamil Nadu. *Environmental Monitoring And Assessment India* 136(1-3), 197-207.
- Tan, W. T., Ooi, L. L., & Lim, E. B. (2001). Voltammetric studies of Cadmium study ion at the mercury electrode in the presence of L-cysteine. *Malaysian Journal of Chemistry*, 3(1), 0013-0019.
- Tapuhi, Y., Schmidt, D. E., Lindner, W., & Karger, B. L. (1981). Dansylation of amino acids for high-performance liquid chromatography analysis. *Analytical Biochemistry*, 115(1), 123-129.
- Valdivielso, A. M., García, L. M. T., & Herranz, R. (2008). Improved synthesis of the PAR-1 thrombin receptor antagonist RWJ-58259. *ARKIVOC*, 0, 287-294.
- Vidal, J. L. (1981). $[\text{Rh}_{10}\text{As}(\text{CO})_{22}]^{3-}$. Example of encapsulation of arsenic by transition-metal carbonyl clusters as illustrated by the structural study of the benzyltriethylammonium salt. *Inorganic Chemistry*, 20(1), 243-249.
- Vivian C, E., & Todd C, H. (2012). A sensitive and selective fluorescence Sensor for the detection of arsenic(III) in organic media. *Inorganic Chemistry Communications*, 51(3), 1213-1215.
- Wang, G., Li, C., Li, J., & Jia, X. (2009). Catalyst-free water-mediated N-Boc deprotection. *Tetrahedron Letters*, 50(13), 1438-1440.
- White, B. R., & Holcombe, J. A. (2007). Fluorescent peptide sensor for the selective detection of Cu^{2+} . *Talanta*, 71(5), 2015-2020.

- White, B. R., Stackhouse, B. T., & Holcombe, J. A. (2009). Magnetic $\gamma\text{-Fe}_2\text{O}_3$ nanoparticles coated with poly-L-cysteine for chelation of As(III), Cu(II), Cd(II), Ni(II), Pb(II) and Zn(II). *Journal of Hazardous Materials*, 161(2-3), 848-853.
- Wolf, C., Villalobos, C. N., Cummings, P. G., Kennedy-Gabb, S., Olsen, M. A., & Trescher, G. (2005). Elucidation of the presence and location of t-Boc protecting groups in amines and dipeptides using on-column H/D exchange HPLC/ESI/MS. *Journal of the American Society for Mass Spectrometry*, 16(4), 553-564.
- Yang, W., Chow, E., Willett, G. D., Hibbert, D. B., & Gooding, J. J. (2003). Exploring the use of the tripeptide Gly-Gly-His as a selective recognition element for the fabrication of electrochemical copper sensors. *Analyst*, 128, 712-718.
- Zen, J., Chung, H., & Ilangoan, G. (1999). Selective voltammetric determination of lead(II) on partially quaternized poly(4-vinylpyridine)/mercury film electrodes. *Electroanalysis*, 11(2), 108-113.
- Zheng, Y., Gattás, A. K. M., Konka, V., & Leblanc, R. M. (2002). A dansylated peptide for the selective detection of copper ions. *Chemical Communications*, (20), 2350-2351.
- Zheng, Y., Huo, Q., Kele, P., Andreopoulos, F. M., Pham, S. M., & Leblanc, R. M. (2001). A new fluorescent chemosensor for copper ions based on tripeptide glycyl-histidyl-lysine (GHK). *Organic Letters*, 3(21), 3277-3280.