

UNIVERSITI PUTRA MALAYSIA

GROWTH PERFORMANCE, GUT MORPHOLOGY AND IMMUNE RESPONSE OF BROILER CHICKENS FED LOW PROTEIN DIETS SUPPLEMENTED WITH LYSINE AND METHIONINE

NURHAZIRAH BINTI SHAZALI

ITA 2015 14



GROWTH PERFORMANCE, GUT MORPHOLOGY AND IMMUNE RESPONSE OF BROILER CHICKENS FED LOW PROTEIN DIETS SUPPLEMENTED WITH LYSINE AND METHIONINE

By

NURHAZIRAH BINTI SHAZALI

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirements for the Master of Science

May 2015

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree in Master of Science

GROWTH PERFORMANCE, GUT MORPHOLOGY AND IMMUNE RESPONSE OF BROILER CHICKENS FED LOW PROTEIN DIETS SUPPLEMENTED WITH LYSINE AND METHIONINE

By

NURHAZIRAH SHAZALI

May 2015

Chairman: Professor Loh Teck Chwen, PhD

Faculty: Institute of Tropical Agriculture

Protein is one of the important nutrients in feedstuff that needs to be met for the basic nutrient requirement of animals. The ideal protein concept contains all amino acids in the exact amount and proportion in order to maintain and fulfill the chicken's requirements and at the same time reduce the feed cost. The present study was conducted to evaluate the effect of feeding a low crude protein diet with lysine and methionine supplementation on broiler performance. Two experiments were conducted in this study. In the first experiments, a total of 288 Cobb500 broiler chickens were used with 8 dietary treatments. The chickens were offered with a starter diet (21% to 18% crude protein) and finisher diet (18% to 15% crude protein) supplemented with three commercial amino acids (L-Lysine, DL-Methionine and L-Threonine). The amino acids in the starter and finisher diets from the different treatment groups were adjusted to similar levels. In continuation from the first experiment, the optimum level of the crude protein diet was used whilst the level of methionine and lysine was manipulated to the high, normal and low levels in the diets. Three hundred Cobb 500 chickens were used in this study with 10 treatments. The chickens were offered a starter diet that consisted of 1.4%, 1.2%, 1.0% lysine and 0.51%, 0.46%, 0.41% methionine in 19% crude protein; whereas the finisher diet contained 1.25%, 1.05%, 0.85% lysine and 0.48% 0.43%, 0.38% methionine in 16% crude protein diets. In the first experiment, reducing dietary crude protein by 2% with amino acid supplementation had a better growth performance, carcass composition, gut morphology and microflora than birds fed with commercial diet. The second experiment reported that high lysine and normal methionine levels in the diets had a greater growth performance, breast meat yield and liver weight as well as lower feed conversion ratio and abdominal fats. Increased methionine levels in the low crude protein diets showed a higher final body weight, weight gain, breast meat yield, liver weight and lower feed conversion ratio, abdominal fat. In conclusion, increasing 0.2% lysine level and maintaining methionine level supplementation in a dietary crude protein reduction by 2% is optimal for maximizing growth performance, absorptive capacity, and immune response in broiler chickens. In addition, it has been widely accepted that the case of the dietary protein level is an economic decision to be made by industrial companies to increase the cost effective benefits.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia Sebagai memenuhi keperluan untuk ijazah Master Sains

PRESTASI PERTUMBUHAN, MORFOLOGI USUS DAN TINDAKBALAS IMUNISASI AYAM PEDAGING DIBERI MAKANAN DIET RENDAH PROTEIN DENGAN PENAMBAHAN LYSINE DAN METHIONINE

Oleh

NURHAZIRAH SHAZALI

Mei 2015

Pengerusi: Profesor Loh Teck Chwen, PhD

Fakulti: Institut Pertanian Tropika

Protein merupakan salah satu nutrien penting dalam pemakanan ternakan sebagai nutrien asas haiwan tersebut. Konsep protein ideal mengandungi semua asid amino dalam jumlah dan nisbah yang tepat untuk mengekalkan dan memenuhi keperluan asas ayam dan pada masa yang sama ia mengurangkan kos makanan. Oleh yang demikian, kajian ini dijalankan untuk menilai kesan pemberian makanan yang mengandungi rendah protein kasar dengan penambahan asid amino terhadap prestasi ayam daging. Dua ujikaji telah dijalankan di dalam kajian ini. Dalam ujikaji pertama, sejumlah 288 ayam daging Cobb500 telah dibahagikan kepada 8 jenis diet. Ayam disajikan dengan diet permulaan (21% hingga 18% protein kasar) dan diet pengakhiran (18% hingga 15% protein kasar) yang ditambah dengan tiga jenis asid amino (L-Lysine, DL-Methionine dan L-Threonine). Asid amino dalam diet permulaan dan diet pengakhiran dalam kumpulan diet yang berbeza telah diselaraskan pada tahap yang sama. Daripada kesinambungan ujikaji pertama, diet yang mengandungi protein kasar yang optimum telah digunakan manakala kepekatan methionine dan lysine telah dimanipulasi kepada kepekatan tinggi, serdahana, dan rendah dalam diet tersebut. 300 ekor ayam Cobb500 telah digunakan dalam kajian ini dengan 10 jenis diet. Ayam telah diberikan diet permulaan yang terdiri daripada 1.4%, 1.2%, 1.0% lysine dan 0.51%, 0.46%, 0.41% methionine dalam 19% protein kasar; manakala diet pengakhiran mengandungi 1.25%, 1.05%, 0.85% lysine dan 0.48%, 0.43%, 0.38% methionine dalam 16% diet protein kasar. Dalam ujikaji pertama, pengurangan protein kasar dalam diet sehingga 2% ditambahkan dengan asid amino menunjukkan prestasi pertumbuhan, komposisi karkas, morfologi usus dan mikroflora yang lebih baik berbanding dengan ayam memakan makanan komersial. Ujikaji kedua melaporkan bahawa paras lysine yang tinggi dan methionine yang normal dalam diet memperolehi prestasi pertumbuhan, hasil daging dibahagian dada, berat hati yang lebih bagus serta nisbah penukaran makanan dan lemak abdomen yang lebih rendah. Peningkatan paras methionine dalam diet rendah protein kasar menunjukkan tinggi berat badan akhir, penambahan berat badan, hasil daging dibahagian dada, berat hati dan rendah nisbah penukaran makanan, lemak dibahagian abdomen berbanding kawalan positif. Kesimpulannya, peningkatan sebanyak 0.2% paras lysine dan pengekalan paras methionine ditambah dalam diet protein kasar yang dikurangkan sebanyak 2% adalah optimum bagi memaksimumkan



prestasi pertumbuhan, keupayaan menyerap, dan tindak balas imun dalam ayam daging. Di samping itu, ia telah diterima dengan meluas bahawa kes tahap protein pemakanan adalah satu keputusan ekonomi yang akan dibuat oleh syarikat-syarikat industri untuk meningkatkan faedah kos efektif.



Ċ,

ACKNOWLEDGEMENTS

I owe a debt of gratitude to those who are so generous with their time and expertise. There are so many people whom I deeply respect and whose efforts are amazing during my studies.

Firstly, I would like to attribute my deepest appreciation and sincere gratitude toProfessor Dr. Loh Teck Chwen, chairperson of the supervisory committee for granting Graduate Research Assistantship and his beneficial advice, guidance, scientific critism and invaluable suggestion during the study. He gets my highest amount of praise for his professionalism and dedicated role as a supervisor. He also gives me the freedom to make choice and to think without boundaries to achieve good results in my studies.

Besides, I would also extend my gratefulness and appreciation to members of the supervisory committee: Dr Anjas Asmara Samsuddin and Associate Professor. Dr. Foo Hooi Ling for their guidance, concern, patience, inspiration, support and constructive comments throughout the course of the study. The caring and easy going characteristics make me feel comfortable to approach them.

Deep appreciation is also extended to School of Graduate Studies, Universiti Putra Malaysia for granting Graduate Research Fellowship and providing excellent service and support. Specially thanks to Associate Professor. Dr. Foo Hooi Ling for giving me permission to use her laboratory in Biotechnology and Biomolecular Science 3 and Institute of Bioscience to analyse my samples. I wish to thank all lecturers and staffs from the Department of Animal Science, Faculty of Agriculture, UniversityPutra Malaysia for their support.

Sincere thanks and appreciation is also extended to my institution, Institute of Tropical Agriculture, UPM for the supports. Finally, I am deeply grateful to all my family and friends who gave me endless love and support me when I happened to be down and depressed. Special thanks go to En. Shazali B. Abu Hassan, Pn. Zaleha Abdullah, Nurmaya Rafeah Bt. Shazali and also my friends in high places.



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master Science. The members of the Supervisory Committee were as follows:

Loh Teck Chwen, PhD Professor, Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Anjas Asmara @ Ab. Hadi Bin Samsudin, PhD Senior Lecturer, Faculty of Agriculture Universiti Putra Malaysia (Member)

> BUJANG BIN KIM HUAT, PhD Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- This thesis is my original work;
- Quotations, illustrations and citations have been duly referenced;
- This thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- Intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- Written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- There is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Date:	and the second second

Declaration by Members of Supervisory Committee

This is to confirm that:

- The research conducted and the writing of this thesis was under our supervision;
- Supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

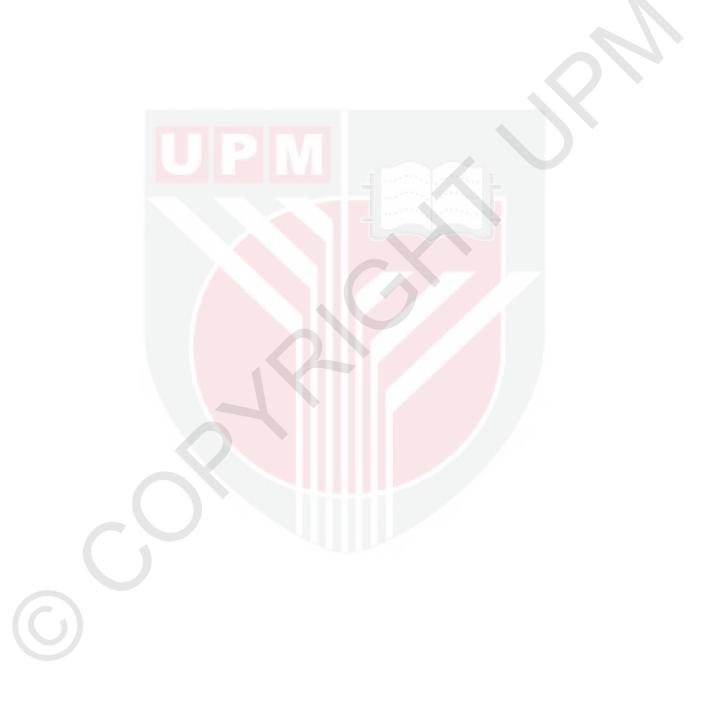
Signature: Name of Chairman of Supervisory Committee:	
Signature: Name of Member of Supervisory Committee:	

TABLE OF CONTENTS

		Page
	STRACT	1
	STRAK	ii
	CKNOWLEDGEMENTS	iv
	PROVAL ECLARATION	v vii
	ST OF TABLES	xii
	ST OF FIGURES	xiv
	ST OF ABBREVIATIONS	XV
СН	IAPTER	
1.	INTRODUCTION	1
2.	LITERATURE RIVIEW	
	2.1. Amino Acid in Poultry Feed	3
	2.1.1. Interactions, Imbalances, and Antagonisms of Amino Acids	5
	2.1.2. Limiting Amino Acids For Poultry	6
	2.2. The Ideal Protein Concept For Broiler	7
	2.3. Low Protein Diet in Poultry Feed2.4. Amino Acids and Crude Protein Enhance Growth Performance	9
	2.4. Amino Acids and Crude Protein Ennance Growth Performance 2.5. Amino Acids and Crude Protein Improve Immune System	10 11
	2.6. Amino Acids and Crude Protein Enhance Gut Microflora and Morphology	12
	2.0. Annual reads and crude Protein Emilance out interoriora and interprotogy	12
3.	GENERAL MATERIALS AND METHODS	
	3.1. Feeding Trial	
	3.1.1. Animal Feed Formulation	14
	3.1.2. Animal Housing System and Managements	14
	3.1.3. Data Collections and Samplings	15
	3.2. Nutrient Analyses 3.2.1. Dry Matter	16
	3.2.2. Crude Protein	16
	3.2.3. Crude Fat	16
	3.2.4. Crude Fiber	17
	3.2.5. Amino Acid Analysis	17
	3.2.5.1. Acid Hydrolysis	17
	3.2.5.2. Perfomic Acid Oxidation	18
	3.2.5.3. Alkaline Hydrolysis	18
	3.2.5.4. High Pressure Liquid Chromatography	
	(HPLC) Analysis	18
4.	GROWTH PERFORMANCE, APPARENT ILEAL DIGESTIBILITY,	
	INTESTINAL MORPHOLOGY, VOLATILE FATTY ACID AND IMMUNE	
	RESPONSE OF BROILER CHICKENS FED DIFFERENT LEVELS OF	
	DIETARY CRUDE PROTEIN WITH LYSINE, METHIONINE AND	
	THREONINE SUPPLEMENTATION 4.1. Introduction	19
	4.1. Introduction 4.2. Materials and Methods	17
	4.2.1. Birds and Experimental Design	19
	4.2.1. Data and Sample Collection	23
	4.2.3. Carcass Characteristic	23
	4.2.4. Apparent Digestibility	23
	4.2.4.1. Titanium Oxide Analysis	23

		4.2.4.2. Digestibility	24
	4	.2.5. Faecal Lactic Acid Bacteria (LAB) and Enterobacteriaceae (ENT)	
		Count	24
	4	.2.6. Small Intestine Morphology	24
		.2.7. Volatile Fatty Acid Analysis	24
		.2.8. Plasma Immunity Status of Broiler	25
		.2.9. Data Analysis	25
	4.3. F		
	4	.3.1. Growth Performance	25
	4	.3.2. Carcass Characteristic	29
	4	.3.3. Nutrient Analyses and Apparent Digestibility	31
		.3.4. Faecal Lactic Acid Bacteria, <i>Enterobacteriaceae</i> Count and VFA	
		Profile	36
	4	.3.5. Small Intestinal Morphology	38
		.3.6. Plasma Immunoglobulin Concentrations	40
		Discussion	
	4	.4.1. Growth Performance	40
	4	.4.2. Carcass Characteristic	41
	4	.4.3. Feed And Amino acid Digestibility	42
	4	.4.4. Faecal Lactic Acid Bacteria And Enterobacteriaceae Count	43
	4	.4.5. Volatile Fatty Acid Analysis	43
	4	.4.6. Small Intestine Morphology	44
		.4.7. Broiler Plasma Immunoglobulin Concentration	45
	4.5. C	Conclusion	45
5.	EFFE	CTS OF LYSINE AND METHIONINE SUPLEMENTATION IN LOW	
	CRUI	DE PROTEIN DIET ON GROWTH PERFORMANCE, APPARENT	
	ILEA	L DIGESTIBILIT <mark>Y, INTESTINAL MORPHOLO</mark> GY AND IMMUNE	
	RESP	ONSE OF BROILER CHICKENS	
	51 L	ntroduction	
			46
	5.2. N	Interials and Methods	46
	5.2. N	Interials and Methods Interials and Experimental Design	47
	5.2. N	Atterials and Methods .2.1. Birds and Experimental Design .2.2. Data and Sample Collection	47 50
	5.2. N	Atterials and Methods .2.1. Birds and Experimental Design .2.2. Data and Sample Collection .2.3. Data Analysis	47
	5.2. M 5.3. F	Aaterials and Methods 2.1. Birds and Experimental Design 2.2. Data and Sample Collection 2.3. Data Analysis tesult	47 50 50
	5.2. N 5.3. F	Atterials and Methods .2.1. Birds and Experimental Design .2.2. Data and Sample Collection .2.3. Data Analysis .esult .3.1. Growth Performance	47 50 50 50
	5.2. N 5 5.3. F 5 5	Atterials and Methods .2.1. Birds and Experimental Design .2.2. Data and Sample Collection .2.3. Data Analysis essult .3.1. Growth Performance .3.2. Carcass Characteristic	47 50 50 50 52
	5.2. M 5.3. F	Atterials and Methods .2.1. Birds and Experimental Design .2.2. Data and Sample Collection .2.3. Data Analysis .esult .3.1. Growth Performance .3.2. Carcass Characteristic .3.3. Nutrient and Apparent Digestibility	47 50 50 50 52 54
	5.2. M 5.3. F	Atterials and Methods .2.1. Birds and Experimental Design .2.2. Data and Sample Collection .2.3. Data Analysis esult .3.1. Growth Performance .3.2. Carcass Characteristic .3.3. Nutrient and Apparent Digestibility .3.4. Faecal Lactic Acid Bacteria and Enterobacteriaceae Count	47 50 50 50 52 54 57
	5.2. M 5.3. F 5.3. F	Atterials and Methods .2.1. Birds and Experimental Design .2.2. Data and Sample Collection .2.3. Data Analysis result .3.1. Growth Performance .3.2. Carcass Characteristic .3.3. Nutrient and Apparent Digestibility .3.4. Faecal Lactic Acid Bacteria and <i>Enterobacteriaceae</i> Count .3.5. Small Intestine Morphology	47 50 50 50 52 54 57 59
	5.2. N 5.3. F 5.3. F	Atterials and Methods .2.1. Birds and Experimental Design .2.2. Data and Sample Collection .2.3. Data Analysis .esult .3.1. Growth Performance .3.2. Carcass Characteristic .3.3. Nutrient and Apparent Digestibility .3.4. Faecal Lactic Acid Bacteria and Enterobacteriaceae Count .3.5. Small Intestine Morphology .3.6. Plasma Immunoglobulin Concentration	47 50 50 50 52 54 57
	5.2. M 5.3. F 5.4. L	Atterials and Methods .2.1. Birds and Experimental Design .2.2. Data and Sample Collection .2.3. Data Analysis tesult .3.1. Growth Performance .3.2. Carcass Characteristic .3.3. Nutrient and Apparent Digestibility .3.4. Faecal Lactic Acid Bacteria and Enterobacteriaceae Count .3.5. Small Intestine Morphology .3.6. Plasma Immunoglobulin Concentration Discussion	47 50 50 52 54 57 59 61
	5.2. N 5.3. F 5.4. I	Atterials and Methods .2.1. Birds and Experimental Design .2.2. Data and Sample Collection .2.3. Data Analysis tesult .3.1. Growth Performance .3.2. Carcass Characteristic .3.3. Nutrient and Apparent Digestibility .3.4. Faecal Lactic Acid Bacteria and Enterobacteriaceae Count .3.5. Small Intestine Morphology .3.6. Plasma Immunoglobulin Concentration Discussion .4.1. Growth Performance	47 50 50 52 54 57 59 61 62
	5.2. N 5.3. F 5.4. I	Atterials and Methods .2.1. Birds and Experimental Design .2.2. Data and Sample Collection .2.3. Data Analysis esult .3.1. Growth Performance .3.2. Carcass Characteristic .3.3. Nutrient and Apparent Digestibility .3.4. Faecal Lactic Acid Bacteria and Enterobacteriaceae Count .3.5. Small Intestine Morphology .3.6. Plasma Immunoglobulin Concentration Discussion .4.1. Growth Performance .4.2. Carcass Characteristic	47 50 50 52 54 57 59 61 62 62
	5.2. N 5.3. F 5.4. E	Atterials and Methods .2.1. Birds and Experimental Design .2.2. Data and Sample Collection .2.3. Data Analysis esult .3.1. Growth Performance .3.2. Carcass Characteristic .3.3. Nutrient and Apparent Digestibility .3.4. Faecal Lactic Acid Bacteria and Enterobacteriaceae Count .3.5. Small Intestine Morphology .3.6. Plasma Immunoglobulin Concentration Discussion .4.1. Growth Performance .4.2. Carcass Characteristic .4.3. Apparent Digestibility	47 50 50 52 54 57 59 61 62 62 63
	5.2. N 5.3. F 5.4. I	Atterials and Methods 2.1. Birds and Experimental Design 2.2. Data and Sample Collection 2.3. Data Analysis esult 3.1. Growth Performance 3.2. Carcass Characteristic 3.3. Nutrient and Apparent Digestibility 3.4. Faecal Lactic Acid Bacteria and Enterobacteriaceae Count 3.5. Small Intestine Morphology 3.6. Plasma Immunoglobulin Concentration Discussion .4.1. Growth Performance 4.2. Carcass Characteristic .4.3. Apparent Digestibility .4.4. Faecal Lactic Acid Bacteria and Enterobacteriaceae Count	47 50 50 52 54 57 59 61 62 62 63 64
	5.2. N 5.3. F 5.4. L	 Atterials and Methods 2.1. Birds and Experimental Design 2.2. Data and Sample Collection 2.3. Data Analysis Analysis Assessive Assessive<td>47 50 50 52 54 57 59 61 62 62 63</td>	47 50 50 52 54 57 59 61 62 62 63
	5.2. N 5.3. F 5.4. L	Atterials and Methods .2.1. Birds and Experimental Design .2.2. Data and Sample Collection .2.3. Data Analysis result .3.1. Growth Performance .3.2. Carcass Characteristic .3.3. Nutrient and Apparent Digestibility .3.4. Faecal Lactic Acid Bacteria and Enterobacteriaceae Count .3.5. Small Intestine Morphology .3.6. Plasma Immunoglobulin Concentration Discussion .4.1. Growth Performance .4.2. Carcass Characteristic .4.3. Apparent Digestibility .4.4. Faecal Lactic Acid Bacteria and Enterobacteriaceae Count .4.5. Small Intestine Morphology .4.4. Faecal Lactic Acid Bacteria and Enterobacteriaceae Count .4.5. Small Intestine Morphology .4.6. Plasma Immunoglobulin Concentration	47 50 50 52 54 57 59 61 62 62 63 64 64 64 65
	5.2. N 5.3. F 5.4. L	 Atterials and Methods 2.1. Birds and Experimental Design 2.2. Data and Sample Collection 2.3. Data Analysis Analysis Assessive Assessive<td>47 50 50 52 54 57 59 61 62 62 63 64 64</td>	47 50 50 52 54 57 59 61 62 62 63 64 64
	5.2. N 5.3. F 5.4. E 5.5. C	 Atterials and Methods 2.1. Birds and Experimental Design 2.2. Data and Sample Collection 2.3. Data Analysis Analysis Analysis Analysis Carcass Characteristic Carcass Characteristic Nutrient and Apparent Digestibility Carcass Characteria and <i>Enterobacteriaceae</i> Count Small Intestine Morphology Analysis Apparent Digestibility Apparent Digesti	47 50 50 52 54 57 59 61 62 62 63 64 64 64 65
6.	5.2. N 5.3. F 5.4. I 5.5. C GENH	 Atterials and Methods 2.1. Birds and Experimental Design 2.2. Data and Sample Collection 2.3. Data Analysis esult 3.1. Growth Performance 3.2. Carcass Characteristic 3.3. Nutrient and Apparent Digestibility 3.4. Faecal Lactic Acid Bacteria and <i>Enterobacteriaceae</i> Count 3.5. Small Intestine Morphology 3.6. Plasma Immunoglobulin Concentration Discussion 4.1. Growth Performance 4.2. Carcass Characteristic 4.3. Apparent Digestibility 4.4. Faecal Lactic Acid Bacteria and <i>Enterobacteriaceae</i> Count 4.5. Small Intestine Morphology 4.6. Plasma Immunoglobulin Concentration Conclusion 	47 50 50 52 54 57 59 61 62 62 63 64 64 64 65 65
6.	5.2. M 5.3. F 5.4. I 5.5. C GENH 6.1. C	 Atterials and Methods 2.1. Birds and Experimental Design 2.2. Data and Sample Collection 2.3. Data Analysis assult 3.1. Growth Performance 3.2. Carcass Characteristic 3.3. Nutrient and Apparent Digestibility 3.4. Faecal Lactic Acid Bacteria and <i>Enterobacteriaceae</i> Count 3.5. Small Intestine Morphology 3.6. Plasma Immunoglobulin Concentration biscussion 4.1. Growth Performance 4.2. Carcass Characteristic 4.3. Apparent Digestibility 4.4. Faecal Lactic Acid Bacteria and <i>Enterobacteriaceae</i> Count 4.5. Small Intestine Morphology 4.6. Plasma Immunoglobulin Concentration conclusion 	47 50 50 52 54 57 59 61 62 62 63 64 64 64 65 65
6.	5.2. M 5.3. F 5.3. F 5.4. E 5.5. C GENH 6.1. C 6.2. C	 Atterials and Methods 2.1. Birds and Experimental Design 2.2. Data and Sample Collection 2.3. Data Analysis esult 3.1. Growth Performance 3.2. Carcass Characteristic 3.3. Nutrient and Apparent Digestibility 3.4. Faecal Lactic Acid Bacteria and <i>Enterobacteriaceae</i> Count 3.5. Small Intestine Morphology 3.6. Plasma Immunoglobulin Concentration Discussion 4.1. Growth Performance 4.2. Carcass Characteristic 4.3. Apparent Digestibility 4.4. Faecal Lactic Acid Bacteria and <i>Enterobacteriaceae</i> Count 4.5. Small Intestine Morphology 4.6. Plasma Immunoglobulin Concentration Conclusion 	47 50 50 52 54 57 59 61 62 62 63 64 64 64 65 65

REFERENCES BIODATA OF STUDENT LIST OF PUBLICATIONS 70 86 87



LIST OF TABLES

Table 2.1	List of essential and non-essential amino acids	Page 3
2.2	NRC (1994) requirement for crude protein and the most rate limiting amino acids for broilers.	8
2.3	Dietary amino acid (% of diet) requirements for high-yielding broilers (Dozier et al., 2008)	8
2.4	The ideal protein patterns based on the original experiments	9
3.1	Cobb 500 recommended broiler feed formulation	15
4.1	Starter feed formulation and composition of different levels of dietary crude protein diet with lysine, methionine and threonine supplementation	21
4.2	Finisher feed formulation and composition of different levels of dietary crude protein with lysine, methionine and threonine supplementation	22
4.3	Growth performance of birds fed different levels of dietary crude protein supplemented with lysine, methionine and threonine at 3 and 6 week of age	27
4.4	The Cumulative Growth performance of birds fed different levels of dietary crude protein supplemented with lysine, methionine and threonine	28
4.5	Effects of carcass characteristic of broiler fed different levels of dietary crude protein with lysine, methionine and threonine supplementation	30
4.6	Nutrient analysis of starter diets (Means ± SE)	32
4.7	Nutrient analysis of finisher diets (Means ± SE)	33
4.8	Nutrient Contents (dry matter basis) of digesta of birds fed different levels of dietary crude protein with lysine, methionine and threonine supplementation at 6 weeks of age	34
4.9	Apparent digestibility of birds fed different levels of dietary crude protein with lysine, methionine and threonine supplementation	35
4.10	Faecal microflora count and volatile fatty acid concentration of birds fed different levels of dietary crude protein with lysine, methionine and threonine supplementation	37
4.11	Villus Height and crypt depth in small intestine of birds fed different levels of dietary crude with lysine, methionine and threonine supplementation	39
5.1	Starter feed formulation and composition of different levels of lysine and methionine supplementation in low crude protein diet	48
5.2	Finisher feed formulation and composition of different levels of lysine and methionine supplementation in low crude protein diet	49
5.3	Growth performance of bird fed different levels of lysine and methionine supplementation in low crude protein diet at 21 days and 42 days of ages	51

- 5.4 Carcass characteristic of broiler fed different levels of lysine and methionine in low crude protein diet at 42 days old
- 5.5 Nutrient content of low crude protein diet supplemented with different levels of lysine and methionine

Nutrient Contents (dry matter basis) of digesta and apparent digestibility of bird
fed low crude protein diet supplemented with different levels of lysine and methionine

- 5.7 Faecal microflora count of birds fed with different levels of lysine and methionine supplementation in low crude protein diet
- 5.8 Villus Height and crypt depth in small intestine of birds fed with different levels of lysine and methionine supplementation in low crude protein diet

60



53

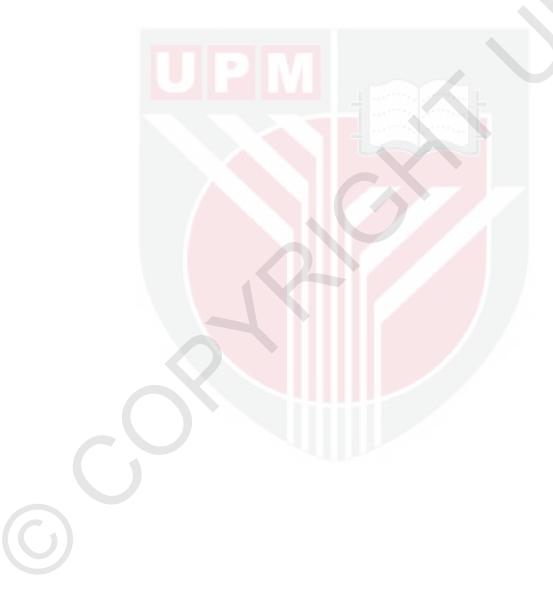
55

58

56

LIST OF FIGURES

Figure 2.1	Roles of AA in nutrition and whole body homeostasis. (Adopted from Wu et al., 2010)	Page 4
4.1	Plasma immunoglobulin concentration of broiler fed different level of crude protein supplemented with essential amino acids	40
5.1	Plasma immunoglobulin concentration of broiler fed different level of lysine and methionine supplemented in low crude protein diet	61



LIST OF ABBREVIATIONS

°C	Degree Celsius
μL	Microliter
μm	Micrometer
4MV	4-methyl-valeric acid
AA	Amino Acid
ACN	Acetonitrile
ADG	Average Daily Gain
ANOVA	Analysis Of Variance
AOAC	Association of Analytical Communities
ATP	Adenosine Triphosphate
BW	Body Weight
CF	Crude Fiber
CFU	Colony forming units
cm	Centimeter
-COOH	Carboxyl Group
CP	Crude Protein
CDD	Columbian Plymouth Rock
DAD	
	Diode Array Detector
dH ₂ O	Deionized water
DM	Dry Matter
DNA	Deoxyribonucleic acid
EAA	Essential Amino Acid
EDTA	Ethylene Diaminetetraacetic Acid
EE	Ether Extract
ENT	Enterobacteriaceae
FCR	Feed Conversion Ratio
FI	Feed Intake
FLD	Fluorescence Detector
FMOC	Fluorenylmethoxycarbonyl chloride
g	Gram
GDP	Gross Domestic Product
GLC	Gas Liquid Chromatography
h	Hours
Н	Hydrogen Atom
H ₂ O ₂	Hydrogen Peroxide
H_2SO_4	Hydrogen Sulfuric Acid
HBr	Hydrogen Bromide
HCL	Hydrochloric Acid
HPLC	High Pressure Liquid Chromatography
IBD	Infectious Bursal Disease
IgG	Immunoglobulin G
IgG-HRP	Immunoglobulin G Horseradish Peroxidase
-	e
IgM	Immunoglobulin M Immunoglobulin M Hammadish Damasi daga
IgM-HRP	Immunoglobulin M Horseradish Peroxidase
Kcal	Kilo Calorie
LAB	Lactic Acid Bacteria
LC	Liquid Chromatography
Log_{10} CFU	Logarithm at base of 10
Lys	Lysine
M	Molarity
ME	Metabolizable Energy
MeOH	Methanol
Met	Methionine
mg	Milligrams
mL	Milliliter
mM	Milli Molar
mm	Millimeter
mRNA	Messenger RNA
	5

MRS – agar	Lactobacillus – Agar De Man, Rogosa and Shape
MRS	Man Rogosa Sharpe
N	Nitrogen
Na ₂ HPO ₄	Di-Sodium Hydrogen Phosphate
NaOH	Sodium hydroxide
ND	Not Determine
NEAA	Non-Essential Amino Acid
NH	New Hampshire
-NH ₂	Amino Group
NH ₃	Ammonia
-	nanometer
nm OPA	
P P	O - phthaldialdehyde Significant laval
r R	Significant level
	Variable Group Round Bottom Flask
RBF RNA	Round Bottom Flask Ribonucleic acid
rpm	Round per minute
SAS	Statistical analysis system
SBM	Soybean Meal
SEM	Standard Error Means
TiO ₂	Titanium Oxide
tRNA	Transfer RNA
Ub	Ubiquitin
UPM	University Putra Malaysia
UPP	Proteasome pathway
v/v	Volume Versus Volume
VFA	Volatile Fatty Acid
w/v	Weight Versus Volume
WG	Weight Gain

G

CHAPTER 1

INTRODUCTION

Agriculture is one of the largest sectorin Malaysian economy and it contributed about 7% to the gross domestic products (GDP) in year 2013 (Abidin *et al.*, 2014). Considering overall performance of the agricultural sector in the year 2012, livestock sector contributed about 11.7%. Among the entire livestock subsector, poultry industry is the most viable industry in Malaysia due to its advancedcommercialization and integration in the production systems (Economic report, 2013).The rapid progress of this sector was partly due to advance technology, genetic improvement and structural changes in the industry.This enhances the private sector to plays an important role in the research and development divisions.

Poultry industry in Malaysia has been self-sufficient since 1984. On the whole, the poultry subsector contributed about of RM10.73 billion, includingpoultry meatworth RM7.07 billion and eggsworthRM3.66 billion. Other than that, 76.14% of the ex-farm output valuealso contributed to the industry (DVS, 2012/2013).Since 1984, the production in the country is more than sufficient to meet domestic demands. In addition, this subsector provides opportunities for poultry products to be exported all over the world and it is expected to integrate and consolidate further to become more efficient and more productive in order to capitalize the growing global export market. However, this subsector is still dependent on imported inputs such as breeding stock and feed grain especially energy and protein sources, feedstuff and animal vaccines that makes it uncompetitive to the changes in world price of such inputs. In the recent years, farmers encountered numerous obstacles due to fluctuation of feed price in the poultry industry (Lepleaideur, 2004). As a result, they are forced to tolerate the increased cost of productionwhich contributes about 75% of feed cost per gram (Conolly, 2012). From the total cost of the feed, 95% is used to meet energy and protein requirements, while 3 to 4% for major mineral, trace mineral and vitamin requirements. Furthermore, 1 to 2% is used to spend for various feed additives. Feed cost is expected to continue in an upward swing for another 20 years due to a shortage of foodstuffs and biofuel policies (Baker, 2009).

Protein is one of the important nutrients in feedstuff and it is necessary to fulfill the basic nutrient requirement of animals (NRC, 1994). Concentratedproteins include soybean meals and other oilseed meals, cottonseed meals and animal protein sources. Since protein, in general, is one of the most expensive feed ingredients, targeted rations are used inindustries to decrease the quantity of protein in the diet of the broiler chickens. The chickens will be in need ofloweramount of protein as they age. Nevertheless, it may not be cost-effective to have different diets for starters, growers, and finishers for small-scale producers.

In poultry diet, different rations are often used to formulate the broiler's diet andit depends on the production stage of the broiler chickens. Proteins in starter rations are higheras it containsexpensive feed ingredients to fulfill the broiler's requirement. However, protein content for grower and finisher rations can be lower since older broiler chickens have lower protein consumption. Starter diets contains of 24% protein, while grower and finisher diet consists of 20% and 18% protein, respectively (Cheeke, 1991). It is well known for more than 20 years, whereby there are limitations to the amount of free amino acids that can replace the intact protein in order to achieve the

maximum weight gain and feed efficiency of the broiler chicks. A vital example is the usage of corn and soybean since the limiting order in corn are lysine, threonine, tryptophan, arginine, valine and isoleucine, sulfur amino acid, phenylalanine, tyrosine, and histidine. Interestingly, the order of limiting amino acids in soybean meal waslysine and valine, threonine, sulfur amino acid, and histidine (Baker, 2009).

Protein is known for its sources of amino acids and an ideal protein concept contains all amino acids in the exact amount and ideal proportion. This helps to maintain and fulfill the chicken requirement at the same time reduce the feed cost (Baker, 2009). However, lately it has been reported that the sulfur amino acids which comprise of methionine and cysteine are limiting in broiler diets (Fatufe and Rodehutscord, 2005). Methionine plays a vital role to maintain proper amino acids balance in the body that stimulates growth, maximizes the carcass yield, reduces carcass fatness and promotes efficient feed intake and reduce production costs (Lamme et al., 2005; Bunchasak, 2009). The second that limiting the amino acid in the broiler chicken's diets is lysine beside methionine, and it is mainly involved in protein deposition such as high lean meat deposition (Bellaver et al., 2002). Unlike lysine, threonine is not only used for protein deposition, but it plays several other vital functions. Threonine is the third that limiting the amino acid in broiler diets and is particularly involved in the maintenance processes and the synthesis of immune protein to the broiler. These limited amino acids should be supplemented in low crude protein diet since the protein level is reduced and the available amino acids in the diet are reduced as well. We carry out this study by putting forward the hypothesis that feeding diet withlow crude protein that is supplemented by optimum level of lysine and methionine will support optimum nutrient requirement of the broiler chickens, leads to improved performance of growth, digestibility of nutrient, immune response, morphology of the intestine, and intestinal microflora population of the broiler chickens. Therefore, this study embarks with the objectives:

- i. To determine the optimum reduction level of crude protein in the diet while supplemented with lysine, methionine and threonine on growth performance, nutrient digestibility, intestinal morphology, microflora count and volatile fatty acid profile of the broiler chickens.
- ii. To identify the optimum level of lysine and methionine supplementation in the low crude protein diet on growth performance, nutrient digestibility, intestinal morphology, microflora count and immunity profile of the broiler chickens.

REFERENCES

- AOAC International. (2005). Official Methods of Analysis of AOACInternational. 18th Edn., Gaithersburg,MD.
- Abbasi, M.A., Mahdavi, A.H., Samie, A.H. and Jahanian, R. (2014).Effects of different levels of dietary crude protein and threonine on performance, humoral immune responses and intestinal morphology of broiler chicks. Brazilian Journal of Poultry science. 16:35-44.
- Abidin, Z.Z., Zainalabidin M., Latif, I.A., Rezai,G. and Sharifudi, J. (2014). Comparing the comparative advantage and profitability of poultry subsector in peninsularMalaysia:an application of policy analysis matrix. Proceeding paper in International Agriculture Conference 2014.
- Akimov, M. and Bezuglov, V. (2012). Methods of Protein DigestiveStability Assay -State of the Art, New Advances in the Basic and Clinical Gastroenterology, Prof. Tomasz Brzozowski (Ed.), InTech. Pp: 211-234.
- Alberts, B., Johnson, A., Lewis, J., Raff, M., Roberts, K. and Walter, P.(2002). Molecular Biology of the Cell. 4th edition. New York: Garland Science.
- Aletor, V.A, Hamid, I.I., Nieb, E. and Pfeffer, E. (2000).Low-proteinaminoacid supplemented diets in broilers chickens: effects on performance,carcass characteristics,whole-bodycomposition and efficiencies of nutrient utilization.Journal of Science Food Agriculture. 80:547-554.
- Al-Masri, S. A. (2014). Biological effect of high plant protein legumes intake on hyperuricemic rats. Life Science Journal.11(1):325-331.
- Almquist, H.J. (1952). Utilization of amino acids by chicks. Archives of Biochemistry and Biophysics. 59:197-202.
- Asyifah , M.N., Abd-Aziz, S., Phang, L.Y. and Azlian, M.N. (2012). Brown rice as a potential feedstuff for poultry. Journal Applied Poultry Research. 21:103–110.
- Austic, R.E. (1986). Biochemical description of nutritional effects. In: Nutrient Requirements of Poultry and Nutritional Research. Fisher C. and Boorman, K.N. Eds.Butterworth's.London, England.
- Austic, R.E. and Scott, R.L. (1975). Involvement of foodintake in the lysine-arginine antagonism in chicks. Journal of Nutrition. 105:1122-1131.
- Baker, D. H. (2009). Advances in protein-amino acid nutrition of poultry. Amino Acids. 37:29-41.
- Baker, D. H. and Han, Y. (1994). Ideal amino acid profile for chicks during the first three weeks post hatching. Poultry Science. 73:1441-1447.

- Baker, D.H. (1997). Ideal amino acid profiles for swine and poultry and their applications in feed formulation. Bio Kyowa Technical Review 9. Biokyowa Publication Corporation, Chesterfield. Pp 1–24.
- Baker, D.H., Batal, A.B., Parr, T.M., Augspurger, N.R. and Parsons, C.M. (2002). Ideal ratio (relative to lysine) of tryptophan, threonine, isoleucine, and valine for chicksduring the second and third weeks post hatch. Poultry Science. 81:458-494.
- Baker, D.H., Fernandez, S.R., Webel, D.M. and Parsons. C.M., (1996). Sulfur amino acid requirement and cysteine replacement value of broiler chicks during the period three to six weeks post hatching. Poultry Science. 75(6):737-742.
- Baker, D.H., Parsons, C.M., Fernandez, S., Aoyagi, S. and Han, Y. (1993). Digestible amino acid requirements of broiler chickens based upon ideal protein considerations. Arkansas Nutrition Conference. Pp. 22-32.
- Ball, R.O. (2001). Threonine requirement and the interaction between threonine intake and gut mucins in pigs. Symposium of the 2001 Degussa Banff Pork Seminar, Canada.
- Banerjee, S., Melesse, A., Dotamo, E., Berihun, K. and Beyan, M. (2013). Effect of feeding different dietary protein levels with Iso-Caloric ration on nutrients intake and growth performances of dual-purpose koekoeck chicken breeds. International Journal of Applied Poultry Research. 2:27-32.
- Batal, A.B. and Parsons, C.M. (2002). Effects of age on nutrient digestibility in chicks fed different diets. Poultry Science. 81: 400-407.
- Bellaver, C. A., Guidoni, A. L., Brum, P. A. R. de and Rosa, P. S. (2002). Estimates of requirements of metabolizable lysine and energy in broilers from 1 to 21 age, usingmultivariate canonical variable. Revista Brasileira de Zootecnia 31:71-78.
- Bender, D.A. (2005). Amino Acids. In: A Dictionary of Food and Nutrition. 3rd Ed. Oxford University Press, USA.
- Berri, C., Besnard, J., and Relandeau, C. (2008). Increasing dietary lysine increases final pH and decreases drip loss of broiler breast meat. Poultry Science. 87:480-484
- Berg, J.M., Tymoczko, J.L. and Stryer, L. (2002). Biochemistry. 5th edition. New York: Freeman, W. H.; Section 30.2, Each Organ Hasa Unique Metabolic Profile.Available from: <u>http://www.ncbi.nlm.nih.gov/books/NBK22436/</u>.
- Bikker, P., Dirkzwager, A., Fledderus, J., Trevisi, P., Hue rou-Luron, I. le, Lalle's, J. P.and Awati, A. (2006). The effect of dietary protein and fermentable carbohydrateslevelson growth performance and intestinal characteristics innewly weaned piglets. Journal of Animal Science. 84:3337-3345.

- Boorman, K.N. and Burgess, A.D. (1986). Responses to amino acids. In: Nutrien Requirements of Poultry and Nutritional Research.Fisher, C. and Boorman, K.N.Eds. Butterworth's. London,England.
- Bowmaker, J. E., and Gous, R. M. (1991). The response of broiler breeder hens to dietary lysine and methionine. British Poultry Science. 32:1069–1088.
- Bregendahl, K, Sell J.L. and Zimmerman, D.R. (2002). Effect of low protein diets on growth performance and body composition of broiler chicks. Poultry Science. 81:1156-1167.
- Bunchasak, C. (2009). Role of dietary methionine in poultry production. Journal of Poultry Science. 46:169–179.
- Burrack, K.S. and Morrison, T.E. (2014). The role of myeloid cell activation and arginine metabolism in the pathogenesis of virus-induced diseases. Frontiers in Immunology. 5:428.
- Carew, L. B., Evarts, K. G. and Alster, A. (1998). Growth, feed intake and plasma thyroid hormone levels in chicks fed dietary excesses of essential amino acids. Poultry Science. 77(2): 295-298.
- Cengiz, O., Onol, G., Sevim, O., Ozturk, M., Sari, M. and Daskiran, M.(2008). Influence of excessive lysine and /or methionine supplementation on growth performance and carcass traits in broiler chicks. Revue de Medecine Veterinaire. 159(4):230-236.
- Cheeke, P. R. (1991). Applied Animal Nutrition: Feeds and Feeding. Macmillan, New York, NY.504.
- Chen, Y. P., Chen, X., Zhang, H. and Zhou Y. M. (2014). Effects of dietary concentrations of methionine on growth and oxidative status of broiler chickens with different hatching weight. British Poultry Science. 54(4):531-537.
- Cheng, T.K., Hamre, M.L. and Coon, C.N. (1997). Responses of broilers to dietary protein levels and amino acid supplementation to low protein diets at various environmental temperatures. Journal of Applied Poultry Research. 6: 18-33.
- Cherian, G., Orr, A., Burke, I. C. and Pan, W. (2013). Feeding Artemisia annual alters digesta pH and muscle lipid oxidation products in broiler chickens. Poultry Science. 92:1085–1090.
- Choe, D.W., Loh, T.C., Foo, H.L., Bejo, M.H. and Awis, Q.S. (2012). Egg production, faecal, pH and microbial population, small intestine morphology, and plasma and yolk cholesterol in laying hens given liquid metabolites produced by *Lactobacillus plantarum* strains. British Poultry Science. 1:106-115.
- Church, D.C. (1991). The nutrients, their metabolism, and feeding standards. In: Livestock Feeds and Feeding. 3rd Ed. D.C.Church, Ed. Prentice Hall. Englewood Cliffs, New Jersey.

- Ciftci, I. and Ceylan, N. (2004). Effects of dietary threonine and crude protein on growth performance, carcass and meat composition of broiler chickens. British Poultry Science.45:280-289.
- Conde-Aguilera, J.A., Cobo-Ortega, C., Tesseraud, S., Lessire, M., Mercier, Y. and Milgen, J. (2013). Changes in body composition in broilers by sulfur amino aciddeficiency during growth. Poultry Science. 92:1266-1275.
- Conolly, A. (2012). Pushing the boundaries performance and profitability. In: Seminar presentation of Poultry International Magazine, Mark Clement. January (2012). Pp.18.
- Corzo, A., Fritts, C.A., Kidd, M.T. and Kerr, B.J. (2005). Response of broiler chicks to essential and non-essential amino acid supplementation of low crude protein diets. Animal Feed Science and Technology. 118:319-327.
- D.V.S. (2012/2013). Department of Veterinary Service, Malaysia.
- D'Mello, J.P.F. and Lewis, D. (1970). Amino acid interactions in chick nutrition. 2. Interrelationships between leucine, isoleucine, and valine. British Poultry Science. 11: 313-323.
- Dahiya, J.P., Hoehler, D., VanKessel, A.G. and Drew, M.D. (2007). Effect of different dietary methionine sources on intestinal microbial populations in broiler chickens. Poultry Science. 86:2358–2366.
- Dari, R.L., Penz, A.M., Kessler, AM.and Jost, H.C. (2005). Use of digestible amino acids and the concept of ideal protein in feed formulation for broilers. Journal of Applied Poultry Research. 14:195-203.
- de Beer, M. (2011).Current trends in broiler breeder nutrition. Proc. III Int. symposium.Nutritional Requirements Poultry and Swine, Vicosa, MG, Brasil. (CD).
- Dekker, J., Aelmans, P.H. and Strous, G.J. (1991). The oligomeric structure of rat and human gastric mucins. Biochem Journal. 277:423–427.
- Deng, P. and Zhongtang, Y. (2014). Intestinal microbiome of poultryand its interaction with host and diet. Gut Microbes. 5(1):108-119.
- Dozier, III W.A., Corzo, A., Kidd, M.T. and Schilling, M.W. (2008). Digestible lysine requirements of male and female broilers from forty-nine to sixty-three days of age. Poultry Science. 87:1385–1391.

Economic Report, (2012/2013).3rd Economic Performance and Prospect.

Edmonds, M.S., Parsons, C.M. and Baker, D.H. (1985). Limiting amino acids in low protein corn-soybean meal diets fed to growing chicks. Poultry Science. 64:1519-1526.

- Fancher, B.I. and Jensen, L.S. (1989). Male broiler performance during the starting and growing periods as affected by dietary protein, essential amino acids and potassium levels. Poultry Science. 68:1385-1395.
- Farkhoy, M., Modirsanei, M., Ghavidel, O., Sadegh, M. and Jafarnejad, S. (2012). Evaluation of Protein Concentration and LimitingAmino Acids Including Lysine andMet + Cys in Prestarter Diet on Performance of Broilers Veterinary Medicine International.
- Farrell, David, J. (2000). A Comparison of Total & Digestible Amino Acids in Diets for Broilers and Layers.Rural Industries Research and Development Corporation.98/124
- Fatufe, A. A. and Rodehutscord, M. (2004). Response to lysine intake in composition of body weight gain and efficiency of lysine utilization of growing male chickens from two genotypes. Poultry Science. 83:1314-1324.
- Fatufe, A. A. and Rodehutscord, M. (2005). Growth, body composition, and marginal efficiency of methionine utilization are affected by nonessential amino acid nitrogen supplementation in male broiler chicken. Poultry Science. 84 (10):1584-1592.
- Fernandez, S.R., Aoyagi, S. Han, Y. Parsons, C.M. and Baker, D.H. (1994). Limiting order of amino acids in corn and soybean meal for growth of the chick. PoultryScience. 73:1887-1896.
- Foo H.L., Loh T.C., Law F.L., Lim Y.Z., Kufli C.N. and Rusul G. (2003) Effects of feeding *Lactobacillus plantarum* I-UL4 isolated from Malaysian Tempeh on growth performance, faecal flora and lactic acid bacteria and plasma cholesterol concentrations in post weaning rats. Food Science and Biotechnology. 4:403-408.
- Franco, J., Murakami, A., Natali, M., Garcia, E. and Furlan, A. (2006). Influenceof delayed placement and dietary lysine levels on small intestine morphometric andperformance of broilers. Revista Brasileira de Ciência Avícola. Pp: 8.
- Franklin, M.A., Mathew A.G., Vickers J.R., and Clift, R.A. (2002). Characterization of microbial populations and volatile fatty acid compositions in the jejunum, ileum, and cecum of pigs weaned at 17 vs. 24 days of age. Journal of Animal Science. 80:2904–2910.
- Fuller, M. F. (2004). The encyclopedia of farm animal nutrition. C.A.B. international Publishing, UK.
- Fuller, M.F. and Reeds, P.J. (1998). Nitrogen cycling in the gut. Annual Review of Nutrition. 18:385–411.
- Furlan, R.L., FariaFilho, D.E. de, Rosa, P.S. and Macari, M. (2004). Does low-protein diet improve broiler performance under heat stress conditions? Brazilian Journal of Poultry Science. 1.6:71 – 79.

- Galassi, G., Malagutti, L., Colombini, S., Rapetti, L. and Crovetto, G. M. (2011). Effects of benzoic acid on nitrogen, phosphorus and energy balance and on ammonia emission from slurries in the heavy pig. Italian Journal of Animal Science. 10:e38.
- Garcia, A.R., Batal, A.B. and Baker, D.H. (2006). Variations in the digestible lysine requirement of broiler chickens due to sex, performance parameters, rearing environment, and processing yield characteristics. Poultry Science. 85:498–504.
- Geraert, P.A. and Mercier Y. (2010). Amino acids: beyond the building blocks! Antony: ADISSEO France SAS.
- Gholamhasan, V., Masoud, T., Shahab, B., Heyder F. and Saeed H. (2011). Effects of Different Levels of Lysine on Small Intestinal Villous Morphology in Starter Diet of Broiler Chickens. Global Veterinaria 7(6): 523-526.
- Glick, B., Taylor, R.L., Martin, D.E., Watabe, M., Day, E. J. and Thompson, D. (1983).Calorie-proteindeficiencies and the immune response of the chicken. II. Cell-mediated immunity. Poultry Science 62:1889-18893.
- Golian, A., AamiAzghadi, M. and Pilevar, M (2010). Influence of various of energy and proteinon performance and humoral immune responses in broiler chick. Global Veterinarian. 4 (5):434-440.
- Gomide, E.M., Rodrigues, P.B., Naves, L.P., Bernardino, V.M.P., Santos, L.M. and Garcia, A.A.P. (2012) Diets with reduced levels of nutrients supplemented with phytase and amino acids for broilers. Ciência e AgrotecnologiaLavras. 36:100-107.
- Gorman, I. and Balnave D. (1995). The effect of dietary lysine and Methionine on the growth characteristics and breast meat yield of Australian broiler chickens. Australian Journal of Agricultural Research. 46:1569-1577.
- Gous, R.M. and T.R. Morris. (2005). Nutritional interventions in alleviating the effect of high temperatures in broiler production. World's Poultry Science Journal. 61:463-475.
- Grimble, R. F. (2006). The effects of sulfur amino acid intake on immune function in humans. Journal of Nutrition.136:1660–1665.
- Gu, X. and Li, D. (2004). Effect of dietary crude protein level on villous morphology, 821 immune status and histo-chemistry parameters of digestive tract in weaning822 piglets. AnimalFeed Science Technology. 114:113-126.
- Han, I.K. and Lee, J.H. (2000). The role of synthetic amino acids in monogastric animalproduction. Asian-Australia Journal Animal Science. 13: 543-560.
- Han, Y. and Baker, D. H. (1994). Digestible lysine requirement of male and female broiler chicks during the period three to six weeks post-hatching. Poultry science. 73(11):1739–1745.

- Han, Y., Suzuki, H., Parsons, C.M. and Baker, D.H. (1992). Amino acid fortification of a low-protein corn and soybean meal diet for chicks. Poultry Science. 71:1168-1178.
- Harper, A.E. (1956). Amino acid imbalances, toxicities, and antagonism. Nutrition Reviews. 14:225-227.
- Harper, A.E. (1958). Balance and imbalance of amino acids. Annals of the New York Academy of Sciences.69:1025-1041.
- Hesabi, A., Nasiri, H. and Birjandi, M. (2008). Effect of supplemental methionine and lysine on performance and carcass yield characteristics in broiler chicks. Journal of Animal Science. 8:26-30.
- Hickling, D., Guenter, W. and Jackson, M. E., (1990). The effects of dietary methionineand lysine on broiler chicken performance and breast meat yield. Canadian Journal of Animal Science. 70:673–678.
- Holsheimer, J.P. and Janssen, W.M.M.A. (1991). Limiting amino acids in low-protein maize±soyabean meal diets fed to broilers from 3±7 weeks of age. British Poultry Science. 32:151-158.
- Htoo, J. K., Araiza, B. A., Sauer, W. C., Rademacher, M., Zhang, Y., Cervantes, M. andZijlstra, R. T. (2007). Effect of dietary protein content on ileal amino acid digestibility, growth performance, and formation of microbial metabolites in ileal and caecal digesta of early- weaned pigs. Journal of Animal Science. 85:3303-3312.
- Hughes, R. J. (2005). An integrated approach to understanding gut function and gut health of chickens. Asia Pacific Journal Clinical Nutrition. 14:S27.
- Hurwitz, S., Sklan, D., Talpaz, H. and Plavnik, I. (1998). The effect of dietary protein level on the lysine and arginine requirements of growing chickens. Poultry Science.77:689-696.
- Hurwitz, S., Weiselberg, M., Eisner, U., Bartov, I., Riesenfeld, G., Sharvit, M., Niv, A. and Bornstein, S. (1980). The energy requirements and performance of growing chickensand turkeysas affected by environmental temperature. Poultry Science. 59:2290-2299.
- Jackson, A.A. (1989). Optimizing amino acid and protein supply and utilization in the newborn. Proceedings of the Nutrition Society.48:293–301.
- Jahanian, R. (2010). Effects of dietary threonine on performance and immune competence of starting broiler chicks.2nd International Veterinary Poultry Congress; Feb 21-21; Tehran. Iran. Pp.200.
- Jankowski, J., Magdalena K. and Zenon Z. (2014). Nutritional andimmunomodulatory function of methionine in poultry diets a review. Annual Animal Science. 14(1):17-31.

- Jiang, Q., Waldroup, P.W. and Fritts, C.A. (2005). Improving the utilization of diets low in crude protein for broiler chicken evaluation of special amino acid supplementation to diets low in crude protein. International Journal of Poultry Science. 4 (3):115-122.
- Kadim, I. T., Moughan, P. J. and Ravindran, V. (2002). Ileal amino acid digestibility assay for the growing meat chicken—Comparison of ileal and excreta amino acid digestibility in the chicken. British Poultry Science. 43:588–597.
- Khatri, I.A., Forstner, G.G. and Forstner, F. (1998). Susceptibility of the cysteine-rich N-terminal and C-terminal ends of rat intestinal mucin Muc (2) to proteolytic cleavage. Biochemistry Journal. 331:323–330.
- Kidd, M.T., Corzo, A., Hoehler, D., Miller E.R. and Dozier III W.A. (2005). Broiler responsiveness (Ross x 708) to diets varying in amino acid density. Poultry Science.84: 1389-1396.
- Kidd, M.T., Kerr, B.J., Allard, J.P., Rao, S.K. and Halley, J.T. (2000). Limiting amino acid responses in commercial broilers. Journal Applied Poultry Research. 9: 223-233.
- Kim, S.W., Mateo, R.D., Yin, Y.L. and Wu, G. (2007). Functional amino acids and fatty acids for enhancingproduction performance of sows and piglets. Asian-Australian Journal of Animal Science. 20:295-306.
- Kirk, C. K. (2000). Comparative avian nutrition. 2nd ed. C.A.B.I. Publishing, C.A.B. Inteernational, Wallingford, UK. Pp:25-26.
- Koenen, M.E., Boonstra-Blom, A.G. and Jeurissen, S.H.M. (2002). Immunological differences between layer and broiler-type chickens. Veterinary Immunology and Immunopathology. 89:47-56.
- Laudadio, V., Dambrosioa, A., Normannob, G., Khanc, R.U., Nazd, S, Rowghanie, E. andTufarellia, V. (2012). Effect of reducing dietary protein level on performance responses and some microbiological aspects of broiler chickens under summer environmental conditions. Avian Biology Research. 5(2):88– 92.
- Law, G., Adjiri-Awere, A., Pencharz, P.B. and Ball, R.O. (2000). Gut mucusin piglets are dependent upon dietary threonine. Advances in Pork Production. Proceedings of the Banff Pork Seminar.
- Le,T. T., Wiele, V.T., Do, T.N.H., Debyser, G., Struijs, K., Devreese, B., Dewettinck, K. and Camp, V.J. (2011). Stability of milk fat globule membrane proteins towardhuman enzymatic gastrointestinal digestion. Journal Dairy Science,95:2307–2318.
- Lemme, A., Kozłowski, K., Jankowski, J., Petri, A. and Zduńczyk, Z. (2005). Responses of 36 to 63 day old BUT Big 6 turkey toms to graded dietary methionine and cysteine levels. Journal Animal Feed Science. 14(1):139–142.

- Lepleaideur, M. (2004). Poultry farming-A disease called competition. In: Singapore Magazine. 114: 4–5.
- Li, P., Yin, Y.L., Li, D., Kim, S.W. and Wu, G. (2007). Amino acids and immune function: a review. British Journal of Nutrition. 98:237-252.
- Loh, T.C., Choe, D.W., Foo, H.L., Sazili, A.Q. and Bejo, M.H. (2014). Effects of feeding different postbiotic metabolite combinations produced by *Lactobacillus plantarum* strains on egg quality and production performance, faecal parameters and plasma cholesterol in laying hens. BMC Veterinary Research. 10:149.
- Loh, T.C., Foo, H.L., Tan, S.H., Goh, Y.M., Shukriyah, M.H. and Kufli, C.N. (2003). Effects of fermented products on performance, faecal pH, *Enterobacteriaceae* and lactic acid bacteria counts and interrelationships and plasma cholesterol concentration in rats. Journal Animal Feed Science. 12:633–644.
- Loh, T.C., Law, F.L., Foo, H.L., Goh, Y.M. and Zulkifli, I. (2009a). Effects of feeding fermented fish on egg cholesterol content in hens. Animal Science Journal. 80:27–33.
- Loh, T.C., Chong, S.W., Foo, H.L. and Law, F.L.(2009b). Effects on growth performance, faecal microflora and plasma cholesterol after supplementation ofspray-dried metabolite to postweaning rats. Czech Journal of Animal Science. 54(1):10–16.
- Henderson, J.H., Ricker, R.D., Bidlingmeyer, B.A. and Woodward, C.(2000). Rapid, accurate and reproducible HPLC analysis of amino acids. Amino acid analysis using Zorbax Eclipse AAA columns and the Agilent 1100 HPLC. Agilent Technologies. 10 Pages.
- Loh, T.C., Thanh, N.T., Foo, H.L., Bejo, M.H. and Azhar, B. K. (2010). Feeding Of Different Levels Of Metabolite Combinations Produced By Lactobacillus Plantarum On Growth Performance, Fecal Microflora, Volatile Fatty Acids And Villi Height In Broilers. Animal Science Journal. 81:205–214.
- Lopez, G. and Leeson, S. (1994). Egg weight and offspring performance of older broiler breeders fed low-protein diets. Journal of Applied Poultry Research. 3:164–170.
- Lu, S.C. (2009). Regulation of glutathione synthesis. Molecular Aspects of Medicine.30:42-59.
- Mack, S., Bercovici, D., DeGroate, G., Leclercq, B., Lippens, M., Pack, M., Schutte, J.B. and Van Cauwenberghe, S. (1999). Ideal aminoacid profile and dietary lysine specification for broiler chickens of 20 to 40 days of age. British Poultry Science. 40:257–265.
- Maiorka, A., Dahlke, F., Penz, A.M. and Kessler, A.M. (2005). Diets formulated on total or digestible amino acid basis with different energy levels and physical form on broiler performance. Brazilian Journal Poultry Science. 7(1):47-50.

- Mateo, R.D., Wu, G., Bazer, F.W., Park, J.C., Shinzato, I. and Kim, S.W. (2007). Dietary L-arginine supplementation enhances the reproductive performance of gilts. Journal of Nutrition.137:652–656.
- Matsushita, K., Takahashi, K. and Akiba, Y. (2007). Effects of adequate or marginal excess of dietary methionine hydroxyl analogue free acid on growth performance, edible meat yields and inflammatory response in female broiler chickens. Journal of Poultry Science. 44:265-272.
- Matthews, D.E. (1999). Proteins and amino acids. In: Shils ME, Olson JA, Shike M, vRossAC, eds. Modern Nutrition in Health and Disease, 9th ed. Baltimore: Williams and Wilkins. Pp:11–48.
- McGill, E.R. (2009). Effects of low crude protein diets with amino acid supplementation on broiler performance in the starter period. Master Thesis of Faculty of the Graduate School University of Missouri-Columbia.
- Mohn, S., Gillis, A.M., Moughan, P.J. and Lange, C.F. (2000). Influence of dietary lysine and energy intakes on body protein deposition and lysine utilization in the growing pig. Journal of Animal Science. 78:1510–9.
- Monirujjaman, M.d. and Ferdouse, A. (2014). Metabolic and Physiological Roles of Branched-Chain Amino Acids. Advances in Molecular Biology. 2014:6.
- Moran, E.T.Jr. (2007). Nutrition of the developing embryo and hatchling. Poultry Science.86:1043–1049.
- Moran, E.T.Jr., Bushong, R.D. and Bilgili, S.F. (1992). Reducing dietary crude protein for broilers while satisfying amino acid requirements by least-cost formulation: Live performance, litter composition, and yield of fast-food carcass cuts at six weeks. Poultry Science. 71:1687-1694.
- Mtimuni, J.P. (1995). Ration Formulation and Feed Guides. Likuni Press and Publishing House, Lilongwe, Malawi, Pp:1
- Munro, H.N. (1979). Glutamic Acid: Advances in Biochemistry and Physiology, edited by Filer, L.J., Jr. Raven Press, New York Pp:55-69.
- Murillo, M. G. and Jensen, L. S. (1976). Methionine requirements of poultry 9th revised end. National Academic Press, Washington, DC.
- Nahm, K.H. (2007). Feed formulation to reduce N excretion and ammonia emission from poultry manure. Bio-recourse Technology. 98:2282-2300.
- Nalle, C.L., Ravindran, G. and Ravindran, V. (2010). Nutritional value of faba beans (Vicia faba L.) for broilers: Apparent metabolisable energy, ileal amino acid digestibility and production performance. Animal Feed Science Technology. 156:104-111
- Namroud, N.F., Shivazad, M. and Zaghari, M. (2008). Effects of fortifying low crude protein diet with crystalline amino acids on performance, blood ammonia

level, and excretacharacteristics of broiler chicks. Poultry Science. 87:2250-2258.

- Nasr, J. and Kheiri, F. (2012). Effect of lysine level of diets formulated based on total ordigestible amino acid on broiler carcass composition. Revista Brasileira deCiência Avícola. 14(4):233-304.
- NRC (1994). Nutrient Requirements of Poultry. 9th Review Edition National Academy Press Washington DC
- Newell-McGloughlin, M. (2008). Nutritionally Improved Agricultural Crops. Plant Physiology.147(3), 939–953.
- Neto, M. Garcia, Pesti, G. M. and Bakalli, R. I. (2000). Influence of Dietary Protein Level on the Broiler Chicken's Response to Methionine and Betaine Supplements. Poultry Science. 79:1478–1484.
- Nyachoti, C. M., Omogbenigun, F. O., Rademacher, M. and Blank, G. (2006). Performance responses and indicators of gastrointestinal health in early weaned pigs fed low- protein amino acid-supplemented diets. Journal Animal Science. 84:125-134.
- O'Dell, B.L. and Savage, J.E. (1966). Arginine-lysine antagonism in the chick and its relationship to dietary rations. Journal of Nutrition. 90:364-370.
- Oxenboll, K.M., Pontoppidan, K. and Fru-Nji, F. (2011). Use of a Protease in Poultry Feed Offers Promising Environmental Benefits. International Journal of Poultry Science 10 (11): 842-848.
- Oyedeji, J.O., Olasupo, O.O., Ekunwe, P. A. and Okugbo, O. T. (2005). Response of Broilers to Alternative Dietary Crude Protein Regimen. International Journal Poultry Science. 4:360-364.
- Panda, A.K., Rama Rao, S.V., Raju, M.V.L.N., Lavanya, G., Pradeep K.R. E. and Shyam, S.G. (2011). Early Growth Response of Broilers to Dietary Lysine at Fixed Ratio to Crude Protein and Essential Amino Acids. Asian-Aust.Journal of Animal Science. 24(11):1623 – 1628.
- Park B.C. (2006). Amino Acid Imbalance-Biochemical Mechanism and NutritionalAspects.Asian-Australian Journal of Animal Science. 19(9):1361 - 1368.
- Pascual, M., Hugas, M., Badiola, J.I., Monfort, J.M. and Garriga, M. (1999). *L. salivarius* CTC2197 prevents *Salmonella enteritidis* colonization in chickens. Applied and Environmental Microbiology. 65(11):4981-4986.
- Perry, T.W., Cullison, A.E. and Lowrey, R.S. (2004). Feeds and Feeding. 6th Ed. Pearson Education, Inc. Upper Saddle River, New Jersey.
- Pluske, J.R., Black, B., Pethick, D.W., Mullan, B.P. and Hampson, D.J. (2003). Effects of different sources of dietary fiber in diets on performance, digesta

characteristics and antibiotic treatment of pigs after weaning. Animal Feed Science Technology. 107:129–142.

- Pond, W.G., Church, D.C. and Pond, K.R. (1995). Basic Animal Nutrition and Feeding.4th Ed. John Wiley and Sons, Inc. Canada.
- Praharaj, N.k., Dungnington E.A., Gross, W.B. and Siegel, P.B. (1997). Dietary effects on immune response of fast growing chicks to inoculation of sheep erythrocytesand *Echerichia coli*. Poultry Science. 51:975-985.
- Rama, R.S.V., Praharaj, N. K., Reddy, M. R. and Sridevi, B. (1999). Immune competence, resistance to Escherichia coli and growth in male broiler parent chicks fed different levels of crude protein. Veterinary Research Communications. 23:323-336.
- Rasekh, J., Thaler, A.M., Engeljohn, D.L. and Pihkala, N.H. (2005). Food safety and inspection service policy for control of poultry contaminated by digestive tract contents: a review. Journal of Applied Poultry Research. 14:603–611.
- Ravindran, V. and Bryden, W. L. (1999). Amino acid availability in poultryIn vitro and in vivo measurements," Australian Journal of Agricultural Research. 50(5):889–908.
- Razaei, M., Nassiri, M., Reza, H., Pour, J. and Kermanshahi, H. (2004). The effects of dietary protein and lysine levels in broiler performance, carcass characteristics and N excretion. International Journal of Poultry Science. 3 (2):148-152.
- Rosebrough, R. W., Mitchell, A. D. and Mcmurtry, J. P. (1995). Carry- over effects ofdietary crude protein and triiodothyronine (T3) in broiler chickens. British Journal of Nutrition. 75:573-581.
- Roth, F.X., Gruber, K. and Kirchgessner, M. (2001). The ideal dietary amino acid pattern for broiler chicks of age 7 to 28 days. Archiv fur Geflugelkunde. 5:199-206.
- Ruth, M. and Catherine, J. Field. (2013). The immune modifying effects of amino acids on gut-associated lymphoid tissue. Journal of Animal Science and Biotechnology. 4:27.
- Rao S. V. R., Raju, M.V.L.N., Panda, A.K., Saharia, P. and Sunder, G.S. (2011). Effect of Supplementing Betaine on Performance, Carcass Traits and Immune Responses in Broiler Chicken Fed Diets Containing Different Concentrations of Methionine. Asian-Australian Journal of Animal Science. 24(5):662-669.
- S.A.S., (1998). SAS® User Guide: Statistic SAS Institute Inc. Cary, NC.
- Samadi, and Liebert, F. (2006). Estimation of nitrogen maintenance requirements and potential for nitrogen deposition in fast-growing chickens depending on age and sex. Poultry Science. 85:1421-1429.

- Schaar,t M. W., Schierbeek, H., Van der Schoor, S. R. D., Stoll, B., Burrin, D. G., Reeds, P. G. and Van Goudoever, J. B. (2005). Threonine utilization is high in the intestine of piglets. The Journal of Nutrition. 135:765-770.
- Short, F. J., Gorton, P. and Wiseman, J. (1996). Determination of titanium dioxide added as inert maker in chicken digestibility studies. Animal Feed Science Technology. 59:215-221.
- Si, J., Fritte, C. A., Burnham, D. J. and Waldroup, P. W. (2001).Relationship of dietary lysine level to the concentration of all essential amino acids in broiler diets, Poultry Science. 80(10):1472–1479.
- Si, J., Fritts, C. A., Waldroup, P. W. and Burnham, D. J. (2004). Effects of tryptophan to large neutral amino acid ratios and overall amino acid levels on utilization of diets low in crude protein by broilers. Journal of Applied Poultry Research. 13:570–578.
- Sibbald, I. and Wolynetz, M. S. (1986). Effects of dietary lysine and feed intake on energy utilization and tissue synthesis by broiler chicks. Poultry Science. 65:98.
- Sklan, D. and Plavnik, I. (2002). Interactions between dietary crude protein and essential amino acid intake on performance in broilers. British Poultry Science. 43(3):442-449.
- Smith, T.K. and Austic, R.E. (1978). The branched-chain amino acid antagonism in chicks. Journal of Nutrition. 108:1180-1191.
- Snel, J., Harmsen, H.J.M., Van der Wilen, P.W.J.J. and Williams, B.J.(2002). Dietary strategies to influence the gastrointestinal microflora of young animals, and its potential to improve intestinal health. in Nutrition and Health of the Gastrointestinal Tract, ed. M. C. Blok, H. A. Vahl, L. de Lange, A. E. Van de Braak, G. Hemke, and Hessing, Wageningen Academic Publisher, Wageningen, The Netherlands. Pp:37-69.
- Sritiawthai, E., Sakulthai, S., Sakdee, J., Bunchasak, C., Kaewtapee, C. and Poeikhampha, T. (2013). Effect of protein level and dietary energy on production, intestinal morphology, and carcass yield of meat duck during starter phase of 14 days. Journal of Applied Science. 13(2):315-320.
- Sterling, K.G., Vedenov, D.V., Pesti, G.M. and Bakalli, R.I. (2005). Economically optimal dietary crude protein and lysine levels for starting broiler chicks. Poultry Science. 84:29-36.
- Sterling, K.G., Pesti, G. M. and Bakalli, R. I. (2006). Performance of different broiler genotypes fed diets with varying levels of dietary crude protein and lysine. Poultry Science. 85:1045-1054.
- Stoll, B., Henry, J., Reeds, P.J., Yu, H., Jahoor, F. and Burrin, D.G. (1998). Catabolism dominates the first-pass intestinal metabolism of dietary essential amino acids in milk protein-fed piglets. Journal of Nutrition. 128:606–614.

- Takahashi, K., Konashi, S., Akiba, Y. and Horiguchi, M. (1994). The effect of dietary methionine and dispensable amino acid supplementation on abdominal fat deposition in male broiler. Journal Animal Science Technology. 65(3):244-250.
- Takahashi, K., Shinichi, Y. and Akiba, Y. (1995). Effect of dietary protein concentration on responses to Escherichia coli endotoxin in broiler chickens. British Journal Nutrition. 74: 173-182.
- Taverner, M.R., Hume, I.D. and Farrell, D.J. (1981). Availability to pigs of amino acids in cereal grains. 1. Endogenous levels of amino acids in ileal digesta and faecesof pigs given cereal diets. British Journal Nutrition. 46:149–158.
- Tesseraud, S., Le Bihan- Duval, S., Peresson, R., Michel, J. and Chagneau A. M. (1999). Response of chick lines selected on carcass quality to dietary lysine supply: live performanceandmuscle development. Poultry Science. 78:80-84.
- Tsukahara, T., Koyama, H., Okada, M. and Ushida, K. (2002). Stimulation of butyrate production by gluconic acid in batch culture of pig cecal digesta and identification of butyrate-producing bacteria. Journal of Nutrition. 132:2229–2234.
- United State Environmental Protection Agency. (2013). Literature review of contaminants in livestock and poultry manure and implications for water quality. EPA 820-R-13-002.
- Uni Z., Noy, Y. and David, S. (1999). Posthatch Development of Small Intestinal Function in the Poultry. Poultry Science. 78:215–222.
- Uzu, G. (1982). Limit of reduction of the protein level in broiler feeds. Poultry Science.61: 1557-1558.
- Vieira, S. L., Lemme, A., Goldenberg, D. B. and Brugalli, I. (2004). Responses of growing broilers to diets with increased sulfur amino acids to lysine ratios at twodietary protein levels, Poultry Science. 83(8):1307–1313.
- Waguespack, A. M., Powell, S., Bidner, T. D. and Southern, L. L. (2009). The glycine plus serine requirement of broiler chicks fed low-crude protein, corn-soybean meal diets. Journal of Applied Poultry Research. 18:761–765.
- Waldroup, P.W., Jiang, Q. and Fritts, C.A. (2005). Effects of supplementing broiler diets low in crude protein with essential and nonessential amino acids. International Journal of Poultry Science. 4(6):425-431.
- Waldroup, P.W., Kersey, J.H. and Fritts, C.A. (2002). Influence of branched-chain amino acid balance in broiler diets. International Journal of Poultry Science.1(5): 136-144.
- Waldroup, P.W., Mitchell, R.J., Payne, J.R. and Hazen, K.R. (1976). Performance of chicks fed diets formulated to minimize excess levels of essential amino acids. Poultry Science. 55: 243-253.

- Wallis, I.R., (1999). Dietary supplements of methionine increase breast meat yielded and decrease abdominal fat in growing broiler chickens. Australian Journal of Experimental Agriculture. 39:131-141.
- Wang, X., Qiao, S.Y., Liu, M. and Ma, Y.X. (2006). Effects of graded levels of true ileal digestible threonine on performance, serum parameters and immune function of 10-25kg pigs. Animal Feed Science and Technology 129:264-278.
- Widyaratne, G., and Drew, P. M. D. (2011). Effects of protein level and digestibility on the growth and carcass characteristics of broiler chickens. Poultry Science. 90: 595-603.
- Williams, P.E.V. (1995). Digestible amino acids for non-ruminant animals: theory and recent challenges. Animal Feed Technology. 53 (2):173-187.
- Wilson, H. R. (1997). Effects of maternal nutrition on hatchability. Poultry Science. 76:134–143.
- Wu, G. (1998) Intestinal mucosal amino acid catabolism. The Journal of Nutrition. 128:1249-1252.
- Wu, G. (2009). Amino acids: metabolism, functions, and nutrition. Amino Acids. 37:117.
- Wu, G., Bazer, F.W., Burghardt, R.C., Johnson, G.A., Kim, S.W., Knabe, D.A., Li, X.L., Satterfield M.C., Smith S.B. and Spencer T.E. (2010). Functional amino acids in swine nutrition and production. In Dynamics in Animal Nutrition. Edited by Doppenberg Journal van der Aar P. The Netherlands: Wageningen Academic Publishers. Pp:69–98.
- Wu, G., Sabina, A.M. and Knabe, D.A. (1996). Dietary glutamine supplementation prevents jejunal atrophy in weaned pigs. Journal of Nutrition. 126:2578.
- Wu, G., Wu, Z.L., Dai, Z.L., Yang, Y., Wang, W.W., Liu, C., Wang, B., Wang, J.J. and Yin, Y.L. (2013). Dietary requirements of nutritionally nonessential amino acids by animals and humans. Amino Acids. 44:1107–1113.
- Xie, C., Zhang, S., Zhang, G., Zhang, F., Chu, L. and Qiao, S. (2013). Estimation of the Optimal Ratio of Standardized Ileal Digestible Threonine to Lysine for Finishing Barrows Fed Low Crude Protein Diets. Asian Australas. Journal of Animal Science. 26 (8):1172-1180.
- Yao, K., Yin, Y.L., Chu, W.Y., Liu, Z.Q., Deng, D., Li, T.J., Huang, R.L., Zhang, J.S., Tan, B.E., Wang, W., and Wu, G. (2008). Dietary arginine supplementation increases mTOR signaling activity in skeletal muscle of neonatal pigs. Journal ofNutrition. 138:867–872.
- Yeh, Y.Y. and Leveeille, G.A. (1969). Effect of dietary protein on hepatic lipogenesis in the growing chick. Journal of Nutrition. 98: 356-366.

- Yuan, J., Karimi, A., Zornes, S., Goodgame, S., Mussini F., Lu C. and Waldroup, P.
 W. (2012). Evaluation of the role of glycine in low-protein amino acidsupplemented diets. Journal of Applied Poultry Research. 21:726–737.
- Zafarian, R., Faradonbeh, O.P. and Bagheri, H. 2011. Better feeding for better performance in broilers: A review. Research opinions in animal and veterinary sciences. 1(12):781-786.
- Zarate, A.J., Moran, E.T. Jr. and Burnham, D.J. (2003). Exceeding essential amino acid requirements and improving their balance as a means to minimize heat stress in broilers. Journal of Applied Poultry Research. 12:37-44.
- Ziegler, T.R., Evans, M.E., Fernandez-Estivariz, C. and Jones, D.P. (2003). Trophic and cytoprotective nutrition for intestinal adaptation, mucosal repair, and barrierfunction. Annual Review Nutrition. 23:229–261.