



**UNIVERSITI PUTRA MALAYSIA**

***CYTOTOXICITY OF THYMOQUINONE AND THYMOQUINONE-LOADED  
NANOSTRUCTURED LIPID CARRIER ON  
CERVICAL CANCER CELLS (SiHa AND HeLa)***

**NG WEI KEAT**

**IB 2015 37**



**CYTOTOXICITY OF THYMOQUINONE AND THYMOQUINONE-LOADED  
NANOSTRUCTURED LIPID CARRIER ON  
CERVICAL CANCER CELLS (SiHa AND HeLa)**

**By**

**NG WEI KEAT**

**Thesis Submitted to the School of Graduate Studies,  
Universiti Putra Malaysia, in Fulfilment of the Requirements for the  
Degree of Doctor of Philosophy**

**July 2015**

## **COPYRIGHT**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirements for the degree of Doctor of Philosophy

**CYTOTOXICITY OF THYMOQUINONE AND THYMOQUINONE-LOADED  
NANOSTRUCTURED LIPID CARRIER ON  
CERVICAL CANCER CELLS (SiHa AND HeLa)**

By

**NG WEI KEAT**

**July 2015**

**Chairman : Latifah Saiful Yazan, PhD**

**Faculty : Institute of Bioscience**

Cancer is a life-threatening disease and a major leading cause of death worldwide. Drug resistance and adverse effects impede the success of chemotherapy. Hence, searching for anti-cancer bioactive compounds with low toxicity from natural products is now in demand. Thymoquinone (TQ), the bioactive compound of *Nigella sativa*, has been reported to exert anti-cancer properties. Nevertheless, TQ exhibits poor oral bioavailability. Therefore, nanostructured lipid carrier (NLC), provides an alternative delivery system for TQ to overcome the limitations. The objective of the study was to determine the cytotoxicity and mechanisms of action of TQ and TQ-NLC towards cervical cancer cells (SiHa and HeLa). The cytotoxicity of TQ towards SiHa and HeLa was determined by MTT assay. Cell cycle and mode of cell death was performed by using flowcytometry. The expression of p53, Bcl-2 and Bax, and caspases was studied by using Western blot and ELISA, respectively. Thymoquinone-loaded nanostructured lipid carrier (TQ-NLC) was synthesized by high pressure homogenization method. Physicochemical characteristics of TQ-NLC were determined. Cytotoxicity of TQ-NLC\_4 towards SiHa and HeLa cells was evaluated by MTT assay. *In vitro* drug release kinetic, gastrointestinal digestion, absorption and bioavailability studies of TQ-NLC\_4 were performed. Result shows that TQ was cytotoxic towards SiHa and HeLa cells by inducing cell cycle arrest and apoptosis in the cells. Elevation of Bax to Bcl-2 ratio and expression of caspase-3 and -9 were noted in SiHa cells while in HeLa cells, elevation of Bax to Bcl-2 ratio and expression of caspase-3, -8 and -9 were noted. Physicochemical analysis revealed that average diameter of TQ-NLC\_4 was less than 100 nm. TQ-NLC\_4 was found stable up to 24 months. High EE and DLC of TQ-NLC\_4 were achieved. TQ-NLC\_4 was cytotoxic towards SiHa and HeLa cells. TQ was released from NLC in a zero-order manner. Degradation and aggregation of TQ-NLC\_4 occurred in simulated intestinal fluid. *In vitro* absorption and bioavailability studies indicated that bioavailability of TQ-NLC\_4 was high. In conclusion, TQ was cytotoxic against the cervical cancer cells by modulating the apoptotic pathways. NLC conferred drug controlled release, protection and enhanced bioavailability to TQ. Hence, TQ-NLC\_4 may be a promising anti-cancer agent against cervical cancer.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**KESAN SITOTOKSIK TIMOKUINON DAN PEMBAWA LIPID  
BERSTRUKTUR NANO DIMUAT TIMOKUINON TERHADAP SEL KANSER  
SERVIKS MANUSIA (SiHa DAN HeLa)**

Oleh

**NG WEI KEAT**

**Julai 2015**

**Pengerusi : Latifah Saiful Yazan, PhD**

**Fakulti : Institut Biosains**

Kanser merupakan sejenis penyakit yang mengancam nyawa dan punca kematian utama di seluruh dunia. Oleh itu, pencarian sebatian bioaktif anti-kanser dengan ketoksikan yang rendah atau tiada ketoksikan daripada produk semulajadi menjadi tumpuan. Timokuinon (TQ), sebatian bioaktif daripada *Nigella sativa*, telah dilaporkan mempunyai ciri anti-kanser. Namun demikian, TQ mempunyai keterbiosediaan oral yang rendah. Oleh itu, sistem penghantaran ubatan koloid berasaskan lipid seperti pembawa lipid berstruktur nano (NLC) menyediakan sistem penghantaran alternatif untuk TQ bagi menyelesaikan masalah tersebut. Objektif kajian ini adalah untuk menentukan kesitotoksikan dan mekanisme aksi TQ dan TQ-NLC terhadap sel kanser serviks (SiHa dan HeLa). Kesitotoksikan TQ ke atas SiHa dan HeLa ditentukan oleh asai MTT. Analisis kitaran sel dan cara kematian dilaksanakan dengan menggunakan sitometer aliran. Kesan TQ ke atas pengekspresan p53, Bcl-2 dan Bax, dan caspase dikaji dengan pemblotan Western dan ELISA, masing-masing. Pembawa lipid berstruktur nano dimuat timokuinon (TQ-NLC) disintesis dengan kaedah penghomogenan tekanan tinggi. Ciri-ciri fizikokimia TQ-NLC ditentukan. Kesitotoksikan TQ-NLC terhadap sel SiHa dan HeLa ditentukan dengan asai MTT. Kinetik pembebasan TQ-NLC<sub>4</sub> secara *in vitro*, pencernaan dalam gastrousus, penyerapan dan keterbiosediaan TQ-NLC secara *in vitro* telah dijalankan. Keputusan menunjukkan TQ adalah sitotoksik terhadap sel SiHa dan HeLa dengan mengaruhkan penahanan kitaran cell dan apoptosis. Peningkatan dalam pengekspresan p53, caspase-3 dan -9, tetapi penurunan pengekspresan Bcl-2 dalam sel SiHa telah dikesan. Walau bagaimanapun, dalam sel HeLa, peningkatan nisbah Bax kepada Bcl-2, dan pengekspresan caspase-3, -8 dan -9 telah dikesan. Analisis fizikokimia menunjukkan bahawa TQ-NLC<sub>4</sub> mempunyai purata diameter zarah kurang daripada 100 nm. TQ-NLC<sub>4</sub> didapati stabil sehingga 24 bulan (2 tahun). EE dan DLC TQ-NLC<sub>4</sub> yang tinggi telah dicapai. TQ dibebaskan daripada NLC mengikut kinetik tertib sifar. Simulasi pencernaan gastrousus menunjukkan bahawa pencernaan dan pengumpulan TQ-NLC<sub>4</sub> berlaku di dalam SIF. Kajian penyerapan dan keterbiosediaan TQ secara *in vitro* menunjukkan bahawa TQ-NLC<sub>4</sub> mempunyai keterbiosediaan yang tinggi. Kesimpulannya, TQ adalah sitotoksik terhadap sel kanser serviks tersebut dengan memodulasikan laluan apoptosis. NLC memberi TQ pembebasan yang terkawal, menyediakan perlindungan dan peningkatan keterbiosediaan. Oleh demikian, TQ-NLC<sub>4</sub> adalah agen anti-kanser serviks yang berpotensi.

## ACKNOWLEDGEMENT

This thesis represents not only my work at the keyboard, but also the result of many experiences I have encountered at Universiti Putra Malaysia, particularly Institute of Bioscience, from tonnes of astonishing individuals whom I wish to acknowledge.

First and foremost, I offer my profoundest gratitude to my supervisor, Associate Professor Dr. Latifah Saiful Yazan, who has supported me throughout my thesis with her patience, motivation, support and enthusiasm since the days I began working on my project. She has supported me, not only academically, but also emotionally, through the rough road to finish this thesis. During the most difficult time when writing this thesis, she gave me moral support. Without her, this thesis would not have been completed.

I will also forever be thankful to my co-supervisors, Professor Dr. Rasedee Abdullah and Professor Dr. Maznah Ismail for their kindness in providing me the research materials and guiding me throughout my years of study. I would also like to express my sincere gratitude to Professor Dr. Johnson Stanslas for his willingness and kindness to share his invaluable research knowledge, idea and suggestions, which certainly helped me to improve my experimental design.

It will never and forever be enough for me to thank my beloved parents, Ng Chin Fatt and Tan Chin Hua, and my dearly beloved wife, Chew Yuan Peng. I would like to dedicate this thesis to them for their endless love, encouragement, support and understanding. Without them, this work would certainly have faltered. Thanks a lot to all my siblings, Dr. Ng Wei Chun, Ng Sin Yee and Ng Sin Ju, for their continuous inspiration.

Next, I would like to express my sincere appreciation to all the professional staff in the Institute of Bioscience, particularly Laboratory of Molecular Biomedicine (MOLEMED) and Laboratory of Vaccines & Immunotherapeutics (VAKSIN), Dr. Yeap Swee Keong, Dr. Tan Sheau Wei, Mrs. Nancy Liew, Mrs. Norhaszalina Md. Isa, Ms. Arba'ah Md. Salleh, Mrs. Norhafiza Azwa Ghozali, Ms. Norsharina Ismail, Ms. Norhayati Yusuf and Mr. Chan Kim Wei. Thanks for lending your hand in guiding me how to operate the equipments and devices.

Dozens of people have helped and taught me immensely throughout my PhD journey. Special thanks to my comrades and friends, Dr. How Chee Wun, Dr. Agustono Wibowo, Dr. Foo Jhi Biau, Hisyam Abd. Hamid, Noreen Husain, Norsyafini Ishak, Zulfahmi Said, Noraina Muhamad Zakuan, Armania Nurdin, Thor Yin Sim, Ong Yong Sze, Wan Nor Hafiza, Goh Su Hua, Kavitha, Kuan Wen Bin, and Yap Huan Yong. Without all of you, the journey will be lonely and empty. I definitely will miss all the moments we worked, played, and have fun and crazy times together.

I am sincerely grateful to my dearest mentor, Mr. Goh Boon Hoe and his wife, Mdm. Bernice Chan. Thanks for giving me hearty support and encouragement and motivation when I was depressed. Not to forget, I would like to express my sincere appreciation to my fellow colleagues from Sunway College A-level Department, particularly the late Chong Soo Sin, for continuously motivating me, and telling me not to give up my study.

Finally, I would like to thank Univeriti Putra Malaysia, for providing Research University Grant Scheme (RUGS) and Graduate Research Fellowship (GRF), which not only allowed me to undertake this research, but also giving me the opportunity to attend conferences, exhibitions and workshops.





© COPYRIGHT UPM



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

**Latifah Saiful Yazan, PhD**

Associate Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Chairman)

**Maznah Ismail, PhD**

Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Internal member)

**Rasedee Abdullah, PhD**

Professor  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Internal member)

---

**BUJANG BIN KIM HUAT, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No.: NG WEI KEAT (GS21291)

## Declaration by supervisory committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: \_\_\_\_\_

Name of Chairman of

Supervisory Committee:

Assoc. Prof. Dr. Latifah Saiful Yazan

Signature: \_\_\_\_\_

Name of

Member of Supervisory

Committee:

Prof. Dr. Maznah Ismail

Signature: \_\_\_\_\_

Name of

Member of Supervisory

Committee:

Prof. Dr. Rasedee Abdullah

## TABLE OF CONTENTS

|                              |                                                        | Page |
|------------------------------|--------------------------------------------------------|------|
| <b>ABSTRACT</b>              |                                                        | i    |
| <b>ABSTRAK</b>               |                                                        | ii   |
| <b>ACKNOWLEDGEMENT</b>       |                                                        | iii  |
| <b>APPROVAL</b>              |                                                        | v    |
| <b>DECLARATION</b>           |                                                        | vii  |
| <b>LIST OF TABLES</b>        |                                                        | xiv  |
| <b>LIST OF FIGURES</b>       |                                                        | xv   |
| <b>LIST OF APPENDICES</b>    |                                                        | xix  |
| <b>LIST OF ABBREVIATIONS</b> |                                                        | xx   |
| <br>                         |                                                        |      |
| <b>CHAPTER</b>               |                                                        |      |
| <br>                         |                                                        |      |
| <b>1</b>                     | <b>INTRODUCTION</b>                                    | 1    |
| 1.1                          | Background                                             | 1    |
| 1.2                          | Objective                                              | 4    |
| 1.2.1                        | General Objective                                      | 4    |
| 1.2.2                        | Specific Objectives                                    | 4    |
| 1.3                          | Hypotheses                                             | 4    |
| <br>                         |                                                        |      |
| <b>2</b>                     | <b>LITERATURE REVIEW</b>                               | 5    |
| 2.1                          | Cancer                                                 | 5    |
| 2.1.1                        | Carcinogenesis                                         | 6    |
| 2.1.2                        | Cancer Incidence                                       | 8    |
| 2.2                          | Cervical Cancer                                        | 10   |
| 2.2.1                        | Cervix                                                 | 10   |
| 2.2.2                        | Cervical Carcinogenesis                                | 11   |
| 2.2.3                        | Cervical Cancer Incidence                              | 13   |
| 2.2.4                        | Prevention of Cervical Cancer                          | 14   |
| 2.2.4.1                      | Human Papillomavirus Vaccines                          | 14   |
| 2.2.4.2                      | Papanicolaou Smear Test                                | 15   |
| 2.2.5                        | Treatment of Cervical Cancer                           | 15   |
| 2.3                          | Natural Product as a Source of Chemotherapeutic Agents | 17   |
| 2.4                          | <i>Nigella sativa</i>                                  | 18   |
| 2.4.1                        | Cytotoxicity of <i>Nigella sativa</i>                  | 20   |
| 2.4.2                        | Anti-cancer Effects of <i>Nigella sativa</i>           | 21   |
| 2.5                          | Thymoquinone                                           | 23   |
| 2.5.1                        | Cytotoxicity of Thymoquinone                           | 24   |
| 2.5.2                        | Anti-cancer Effects of Thymoquinone                    | 27   |
| 2.6                          | Cell Cycle                                             | 28   |
| 2.7                          | Mode of Cell Death                                     | 30   |
| 2.8                          | Regulation of Apoptosis                                | 31   |
| 2.8.1                        | Regulatory Protein of Apoptosis                        | 32   |
| 2.8.1.1                      | p53 Tumour Suppressor Protein                          | 32   |
| 2.8.1.2                      | Bcl-2 Family Proteins                                  | 32   |

|          |         |                                                                                                         |           |
|----------|---------|---------------------------------------------------------------------------------------------------------|-----------|
|          | 2.8.1.3 | Caspases Family                                                                                         | 33        |
|          | 2.8.2   | Apoptotic Pathways                                                                                      | 33        |
|          | 2.8.2.1 | Intrinsic Pathway                                                                                       | 33        |
|          | 2.8.2.2 | Extrinsic Pathway                                                                                       | 34        |
| 2.9      |         | Colloidal Drug Delivery System                                                                          | 35        |
|          | 2.9.1   | Lipid-based Nanocarriers                                                                                | 35        |
|          | 2.9.1.1 | Liposome                                                                                                | 35        |
|          | 2.9.1.2 | Solid Lipid Nanoparticle                                                                                | 36        |
|          | 2.9.1.3 | Nanostructured Lipid Carrier                                                                            | 37        |
|          | 2.9.2   | Polymeric Nanoparticles                                                                                 | 37        |
|          | 2.9.3   | Metal-based Nanoparticles                                                                               | 38        |
| 2.10     |         | Applications of Colloidal Drug Delivery System in Cancer Therapy                                        | 38        |
| 2.11     |         | Nano-formulation of Thymoquinone                                                                        | 40        |
| <b>3</b> |         | <b>CYTOTOXICITY OF THYMOQUINONE FROM <i>Nigella sativa</i> TOWARDS SELECTED CANCER CELL LINES</b>       | <b>41</b> |
|          | 3.1     | Introduction                                                                                            | 41        |
|          | 3.2     | Materials and Methods                                                                                   | 42        |
|          | 3.2.1   | Cell Lines                                                                                              | 42        |
|          | 3.2.2   | Chemicals and Reagents                                                                                  | 42        |
|          | 3.2.3   | Cell Culture                                                                                            | 42        |
|          | 3.2.4   | Cell Counting                                                                                           | 42        |
|          | 3.2.5   | Treatment                                                                                               | 43        |
|          | 3.2.6   | Determination of Cytotoxicity of Thymoquinone                                                           | 43        |
|          |         | 3.2.6.1 MTT Assay                                                                                       | 43        |
|          |         | 3.2.6.2 Trypan Blue Dye Exclusion Test                                                                  | 44        |
|          | 3.2.7   | Cell Morphological Studies                                                                              | 44        |
|          | 3.2.8   | Cell Cycle Analysis                                                                                     | 44        |
|          | 3.2.9   | Determination of the Mode of Cell Death                                                                 | 45        |
|          | 3.2.10  | Statistical Analysis                                                                                    | 45        |
|          | 3.3     | Results                                                                                                 | 45        |
|          | 3.3.1   | Cytotoxicity of Thymoquinone and Cisplatin                                                              | 45        |
|          | 3.3.2   | Cell Morphological Changes                                                                              | 50        |
|          | 3.3.3   | Cell Cycle Arrest Induced by Thymoquinone                                                               | 55        |
|          | 3.3.4   | Mode of Cell Death Induced by Thymoquinone                                                              | 62        |
|          | 3.4     | Discussion                                                                                              | 66        |
|          | 3.5     | Conclusion                                                                                              | 68        |
| <b>4</b> |         | <b>APOPTOTIC PATHWAYS INVOLVED IN THE HUMAN CERVICAL CARCINOMA CELL LINES TREATED WITH THYMOQUINONE</b> | <b>69</b> |
|          | 4.1     | Introduction                                                                                            | 69        |
|          | 4.2     | Materials and Methods                                                                                   | 70        |
|          | 4.2.1   | Chemicals and Reagents                                                                                  | 70        |
|          | 4.2.2   | Cell Culture                                                                                            | 70        |
|          | 4.2.3   | Determination of the Effects of Thymoquinone on the Level of Expression of Apoptotic-related Proteins   | 71        |

|          |                                                                                                             |                                                                                                                           |           |
|----------|-------------------------------------------------------------------------------------------------------------|---------------------------------------------------------------------------------------------------------------------------|-----------|
|          | 4.2.3.1                                                                                                     | Preparation of Cell Lysate                                                                                                | 71        |
|          | 4.2.3.2                                                                                                     | Detection of the Expression of Caspase-3, -8 and -9                                                                       | 71        |
|          | 4.2.3.3                                                                                                     | Detection of the Expression of p53, Bax and Bcl-2                                                                         | 71        |
|          | 4.2.4                                                                                                       | Statistical Analysis                                                                                                      | 72        |
| 4.3      | Results                                                                                                     |                                                                                                                           | 72        |
|          | 4.3.1                                                                                                       | Expression of Caspase-3, -8 and -9                                                                                        | 72        |
|          | 4.3.2                                                                                                       | Expression of p53, Bax and Bcl-2                                                                                          | 77        |
| 4.4      | Discussion                                                                                                  |                                                                                                                           | 79        |
| 4.5      | Conclusion                                                                                                  |                                                                                                                           | 80        |
| <b>5</b> | <b>DEVELOPMENT AND PHYSICOCHEMICAL CHARACTERISATION OF THYMOQUINONE-LOADED NANOSTRUCTURED LIPID CARRIER</b> |                                                                                                                           | <b>81</b> |
| 5.1      | Introduction                                                                                                |                                                                                                                           | 81        |
| 5.2      | Materials and Methods                                                                                       |                                                                                                                           | 82        |
|          | 5.2.1                                                                                                       | Chemicals and Reagents                                                                                                    | 82        |
|          | 5.2.2                                                                                                       | Determination of the Solubility of TQ in Lipid Matrix                                                                     | 82        |
|          | 5.2.3                                                                                                       | Preparation and Synthesis of Thymoquinone-loaded Nanostructured Lipid Carrier                                             | 82        |
|          | 5.2.3.1                                                                                                     | Preparation of Lipid Matrices                                                                                             | 82        |
|          | 5.2.3.2                                                                                                     | Preparation of Aqueous Surfactant Mixture                                                                                 | 83        |
|          | 5.2.3.3                                                                                                     | Synthesis of Thymoquinone-loaded Nanostructured Lipid Carrier                                                             | 83        |
|          | 5.2.4                                                                                                       | Measurement of the Particle Average Diameter and Polydispersity Index of Thymoquinone-loaded Nanostructured Lipid Carrier | 83        |
|          | 5.2.5                                                                                                       | Measurement of the Zeta Potential of Thymoquinone-loaded Nanostructured Lipid Carrier                                     | 83        |
|          | 5.2.6                                                                                                       | Determination of Encapsulation Efficiency and Drug Loading Capacity of Thymoquinone-loaded Nanostructured Lipid Carrier   | 85        |
|          | 5.2.7                                                                                                       | Morphological Imaging of Thymoquinone-loaded Nanostructured Lipid Carrier by Transmission Electron Microscopy             | 85        |
|          | 5.2.8                                                                                                       | Determination of the Stability of Thymoquinone-loaded Nanostructured Lipid Carrier                                        | 85        |
|          | 5.2.9                                                                                                       | Determination of the Thermal Behaviour of Thymoquinone-loaded Nanostructured Lipid Carrier                                | 86        |
|          | 5.2.10                                                                                                      | Statistical Analysis                                                                                                      | 86        |
| 5.3      | Results                                                                                                     |                                                                                                                           | 86        |
|          | 5.3.1                                                                                                       | Solubility of Thymoquinone in Lipid Phase                                                                                 | 86        |

|          |                                                                                                              |           |
|----------|--------------------------------------------------------------------------------------------------------------|-----------|
| 5.3.2    | Physicochemical Characteristic of Thymoquinone-loaded Nanostructured Lipid Carrier                           | 86        |
| 5.3.3    | Morphology of Thymoquinone-loaded Nanostructured Lipid Carrier                                               | 89        |
| 5.3.4    | Stability of Thymoquinone-loaded Nanostructured Lipid Carrier                                                | 89        |
| 5.3.5    | Thermal Behaviour of Thymoquinone-loaded Nanostructured Lipid Carrier                                        | 89        |
| 5.4      | Discussion                                                                                                   | 92        |
| 5.5      | Conclusion                                                                                                   | 94        |
| <b>6</b> | <b>CYTOTOXICITY, RELEASE PROFILE AND BIOAVAILABILITY OF THYMOQUINONE-LOADED NANOSTRUCTURED LIPID CARRIER</b> | <b>95</b> |
| 6.1      | Introduction                                                                                                 | 95        |
| 6.2      | Materials and Methods                                                                                        | 96        |
| 6.2.1    | Chemicals and Reagents                                                                                       | 96        |
| 6.2.2    | Cell Culture                                                                                                 | 96        |
| 6.2.3    | Determination of Cytotoxicity of Thymoquinone-loaded Nanostructured Lipid Carrier                            | 97        |
| 6.2.4    | Development of Dissolution Medium for Thymoquinone                                                           | 97        |
| 6.2.5    | <i>In vitro</i> Drug-release of Thymoquinone-loaded Nanostructured Lipid Carrier                             | 98        |
| 6.2.6    | Mathematical Modelling and Release Kinetics of Thymoquinone-loaded Nanostructured Lipid Carrier              | 99        |
| 6.2.7    | <i>In Vitro</i> Digestion Assay of Thymoquinone-loaded Nanostructured Lipid Carrier                          | 100       |
| 6.2.7.1  | Simulated Gastric Digestion of Thymoquinone-loaded Nanostructured Lipid Carrier                              | 100       |
| 6.2.7.2  | Simulated Intestinal Digestion of Thymoquinone-loaded Nanostructured Lipid Carrier                           | 100       |
| 6.2.8    | <i>In Vitro</i> Absorption and Bioavailability Study of Thymoquinone-loaded Nanostructured Lipid Carrier     | 101       |
| 6.2.9    | Statistical Analysis                                                                                         | 102       |
| 6.3      | Results                                                                                                      | 102       |
| 6.3.1    | Cytotoxicity of Thymoquinone-loaded Nanostructured Lipid Carrier                                             | 102       |
| 6.3.2    | Solubility of Thymoquinone in Various Types of Dissolution Medium                                            | 103       |
| 6.3.3    | <i>In Vitro</i> Release Profile of Thymoquinone-loaded Nanostructured Lipid Carrier                          | 104       |
| 6.3.4    | Release Kinetic of Thymoquinone-loaded Nanostructured Lipid Carrier                                          | 106       |

|          |                                                                                                                                         |            |
|----------|-----------------------------------------------------------------------------------------------------------------------------------------|------------|
| 6.3.5    | <i>In Vitro</i> Digestion of Thymoquinone-loaded Nanostructured Lipid Carrier in Simulated Gastric Fluid and Simulated Intestinal Fluid | 107        |
| 6.3.6    | <i>In Vitro</i> Absorption and Bioavailability of Thymoquinone-loaded Nanostructured Lipid Carrier                                      | 110        |
| 6.4      | Discussion                                                                                                                              | 111        |
| 6.5      | Conclusion                                                                                                                              | 114        |
| <b>6</b> | <b>GENERAL DISCUSSION, CONCLUSION AND FUTURE WORK</b>                                                                                   | <b>115</b> |
| 7.1      | General Discussion                                                                                                                      | 115        |
| 7.2      | Conclusion                                                                                                                              | 118        |
| 7.3      | Future Work                                                                                                                             | 118        |
|          | <b>REFERENCES</b>                                                                                                                       | <b>119</b> |
|          | <b>APPENDIX</b>                                                                                                                         | <b>163</b> |
|          | <b>BIODATA OF STUDENT</b>                                                                                                               | <b>168</b> |
|          | <b>LIST OF PUBLICATIONS</b>                                                                                                             | <b>169</b> |



## LIST OF TABLES

| Table |                                                                                                                                                                            | Page |
|-------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------|------|
| 2.1   | Carcinogens and their respective cancer                                                                                                                                    | 7    |
| 2.2   | Type of fatty acid isolated from <i>N. sativa</i> seed oil with its respective composition                                                                                 | 18   |
| 2.3   | Nutrients and minerals found in <i>N. sativa</i> seed oil                                                                                                                  | 19   |
| 2.4   | Differences between apoptosis and necrosis                                                                                                                                 | 31   |
| 2.5   | Applications of colloidal drug delivery system in clinical trial or in the market for cancer therapy                                                                       | 39   |
| 2.6   | Thymoquinone nano-formulation and their respective biological activities                                                                                                   | 40   |
| 3.1   | Cytotoxicity of TQ towards SiHa and HeLa cells at various incubation times as determined by using MTT assay and trypan blue dye exclusion test as reflected by IC50 values | 49   |
| 5.1   | Composition of the lipid matrix and aqueous surfactant mixture, and the high pressure homogenisation condition for the synthesis of TQ-NLC                                 | 84   |
| 5.2   | Physicochemical characteristics of different formulation of TQ-NLC after recrystallization at 25°C for 24 hours                                                            | 87   |
| 5.3   | Physicochemical characteristics of TQ-NLC_4 at different storage time as determined by zeta sizer and HPLC                                                                 | 90   |
| 6.1   | Composition of the solutions used in the development of dissolution medium for TQ                                                                                          | 98   |
| 6.2   | Linearised equations of the kinetic models used for the analysis of TQ-NLC_4 release data                                                                                  | 99   |
| 6.3   | Cytotoxicity of TQ-NLC_4 and TQ towards various cell lines at different time point as reflected by the IC50 value determined using MTT assay                               | 102  |
| 6.4   | Solubility of TQ in various types of dissolution medium as determined by HPLC                                                                                              | 104  |
| 6.5   | Linear regression of TQ-NLC_4 release kinetic by mathematical modelling using various drug kinetic models                                                                  | 106  |

## LIST OF FIGURES

| Figure |                                                                                                                        | Page |
|--------|------------------------------------------------------------------------------------------------------------------------|------|
| 2.1    | Two emerging hallmarks and enabling characteristics added to the previous list of six hallmarks of cancer in 2011      | 6    |
| 2.2    | Genetic and epigenetic alteration during tumour initiation, promotion and progression during multistage carcinogenesis | 8    |
| 2.3    | Worldwide estimated new cancer cases in 2012 by gender (age-standardised rate per 100,000 population)                  | 9    |
| 2.4    | Anatomy of the cervix uteri                                                                                            | 11   |
| 2.5    | Involvement of hr-HPV in cervical carcinogenesis, where the untreated lesions progress to invasive cervical cancer     | 12   |
| 2.6    | Chemical structure of six approved platinum-based anti cancer agents for treatment of cervical cancer                  | 15   |
| 2.7    | Mechanism of cisplatin in treatment of cancer                                                                          | 16   |
| 2.8    | <i>Nigella sativa</i> (A) flower and (B) black seeds                                                                   | 18   |
| 2.9    | Chemical structure of thymoquinone, C <sub>10</sub> H <sub>12</sub> O <sub>2</sub>                                     | 23   |
| 2.10   | Hallmarks of cancer which are considered as promising targets for TQ except “avoiding the immune system”               | 24   |
| 2.11   | Overview of cell cycle                                                                                                 | 28   |
| 2.12   | Involvement of cyclin and CDKs in cell cycle                                                                           | 29   |
| 2.13   | Comparison of apoptotic and necrotic cell death process                                                                | 30   |
| 2.14   | Apoptosis is activated via the intrinsic (or mitochondrial) and extrinsic (or death receptors) pathways                | 34   |
| 2.15   | Structure of a bilaminar liposome                                                                                      | 36   |
| 2.16   | Structure of a solid lipid nanoparticle                                                                                | 36   |
| 2.17   | Crystallization of SLN and NLC after synthesis. SLN shows high crystallinity but NLC shows imperfect matrix            | 37   |
| 2.18   | Structure of a cyclodextrin polymeric nanoparticle                                                                     | 38   |

|      |                                                                                                                                                                                                    |    |
|------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|----|
| 3.1  | Cytotoxicity of TQ towards various selected cancer cell lines as determined by MTT assay at 72 hours as reflected by IC50 values                                                                   | 46 |
| 3.2  | Cytotoxicity of TQ towards SiHa and HeLa cells as determined by MTT assay at various incubation times as reflected by IC50 values                                                                  | 47 |
| 3.3  | Cytotoxicity of TQ towards SiHa and HeLa cells as determined by trypan blue dye exclusion test at various incubation times as reflected by IC50 values                                             | 48 |
| 3.4  | Cytotoxicity of TQ and cisplatin towards cervical cancer cells (SiHa and HeLa) and non-cancerous normal cells (Vero and 3T3-L1) as determined by MTT assay at 72 hours as reflected by IC50 values | 50 |
| 3.5  | Morphological changes of SiHa cells treated with various concentrations of TQ at different incubation time viewed under an inverted light microscope (200× magnification)                          | 51 |
| 3.6  | Close up view of SiHa cells (a) control and (b) treated with 10 µM of TQ at 72 hours                                                                                                               | 52 |
| 3.7  | Morphological changes of HeLa cells treated with various concentrations of TQ at different incubation time, viewed under an inverted light microscope (200× magnification)                         | 53 |
| 3.8  | Close up view of HeLa cells (a) control and (b) treated with 15 µM of TQ at 72 hours                                                                                                               | 54 |
| 3.9  | Effects of TQ on cell cycle progression of SiHa cells at different incubation time as analyzed by a flowcytometer                                                                                  | 56 |
| 3.10 | Cell cycle distribution of SiHa cells treated with various concentrations of TQ at 24 hours                                                                                                        | 57 |
| 3.11 | Cell cycle distribution of SiHa cells treated with various concentrations of TQ at 48 hours                                                                                                        | 58 |
| 3.12 | Effects of TQ on cell cycle progression of HeLa cells different incubation time as analyzed by a flowcytometer                                                                                     | 59 |
| 3.13 | Cell cycle distribution of HeLa cells treated with various concentrations of TQ at 24 hours                                                                                                        | 60 |
| 3.14 | Cell cycle distribution of HeLa cells treated with various concentrations of TQ at 48 hours                                                                                                        | 61 |

|      |                                                                                                                                                                                                                                                                                                                                                                                                         |    |
|------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|----|
| 3.15 | Flowcytometric analysis of TQ-treated and -untreated SiHa cells using the Annexin V-FITC/PI dual-labelling technique at different incubation time                                                                                                                                                                                                                                                       | 63 |
| 3.16 | Percentage of viable (Annexin V <sup>-</sup> /PI <sup>-</sup> ), early apoptotic (Annexin V <sup>+</sup> /PI <sup>-</sup> ), late apoptotic and necrotic (Annexin V <sup>+</sup> /PI <sup>+</sup> ) and cell debris (Annexin V <sup>-</sup> /PI <sup>+</sup> ) SiHa cells treated with TQ at different incubation time analyzed by a flowcytometer using the Annexin V-FITC/PI dual-labelling technique | 64 |
| 3.17 | Flowcytometric analysis of TQ-treated and -untreated HeLa cells using the Annexin V-FITC/PI dual-labelling technique at different incubation time                                                                                                                                                                                                                                                       | 65 |
| 3.18 | Percentage of viable (Annexin V <sup>-</sup> /PI <sup>-</sup> ), early apoptotic (Annexin V <sup>+</sup> /PI <sup>-</sup> ), late apoptotic and necrotic (Annexin V <sup>+</sup> /PI <sup>+</sup> ) and cell debris (Annexin V <sup>-</sup> /PI <sup>+</sup> ) HeLa cells treated with TQ at different incubation time analyzed by a flowcytometer using the Annexin V-FITC/PI dual-labelling technique | 66 |
| 4.1  | Effect of TQ on the expression of caspase-3, -8 and -9 in SiHa cells after 24 hours as determined by Caspase colorimetric protease assay sampler kit                                                                                                                                                                                                                                                    | 73 |
| 4.2  | Effect of TQ on the expression of caspase-3, -8 and -9 in SiHa cells after 48 hours as determined by Caspase colorimetric protease assay sampler kit                                                                                                                                                                                                                                                    | 74 |
| 4.3  | Effect of TQ on the expression of caspase-3, -8 and -9 in HeLa cells after 24 hours as determined by Caspase colorimetric protease assay sampler kit                                                                                                                                                                                                                                                    | 75 |
| 4.4  | Effect of TQ on the expression of caspase-3, -8 and -9 in HeLa cells after 48 hours as determined by Caspase colorimetric protease assay sampler kit                                                                                                                                                                                                                                                    | 76 |
| 4.5  | Effect of TQ on the expression of p53, Bax and Bcl-2 in SiHa cells at different incubation time as determined by the Western blot analysis                                                                                                                                                                                                                                                              | 77 |
| 4.6  | Effect of TQ on the expression of p53, Bax and Bcl-2 in HeLa cells at different incubation time as determined by the Western blot analysis                                                                                                                                                                                                                                                              | 78 |
| 5.1  | Optical appearance of (A) TQ-NLC <sub>4</sub> and (B) blank NLC after re-crystallisation at 25°C for 24 hours                                                                                                                                                                                                                                                                                           | 88 |

|     |                                                                                                            |     |
|-----|------------------------------------------------------------------------------------------------------------|-----|
| 5.2 | Morphology of TQ-NLC_4 visualised under a transmission electron microscope (magnification 150000×)         | 89  |
| 5.3 | Thermogram of the tested materials recorded as a function of temperature from 25 to 70°C                   | 91  |
| 6.1 | Effects of blank NLC on the cell viability as determined by MTT assay after 72 hours                       | 103 |
| 6.2 | <i>In vitro</i> drug release profile of TQ-NLC_4 and pure TQ solution                                      | 105 |
| 6.3 | Linear regression of TQ-NLC_4 release kinetic by using Korsmeyer–Peppas equation                           | 106 |
| 6.4 | Average diameter of TQ-NLC_4 after incubation in SGF and SIF as determined by zeta sizer                   | 107 |
| 6.5 | Release profile of TQ from TQ-NLC_4 in SGF and SIF                                                         | 108 |
| 6.6 | <i>In vitro</i> release of different form of TQ after 3 hours of simulated digestion as determined by HPLC | 109 |
| 6.7 | <i>In vitro</i> absorption and bioavailability of various forms of TQ as reflected by the $P_{app}$ values | 110 |

## LIST OF APPENDICES

| <b>Table</b> |                                               | <b>Page</b> |
|--------------|-----------------------------------------------|-------------|
| A            | Experimental Design                           | 163         |
| B1           | Radio immunoprecipitation assay (RIPA) buffer | 164         |
| B2           | Resolving gel (12%)                           | 164         |
| B3           | Stacking gel (4%)                             | 164         |
| B4           | 10X transfer buffer                           | 165         |
| B5           | 1X transfer buffer                            | 165         |
| B6           | 10X running buffer                            | 165         |
| B7           | 1X running buffer                             | 165         |
| B8           | 10X Tris-buffered saline (TBS)                | 166         |
| B9           | 1X Tris-buffered saline-Tween 20 (TBST)       | 166         |
| C1           | Chromatogram of TQ                            | 167         |

## LIST OF ABBREVIATIONS

|           |                                   |
|-----------|-----------------------------------|
| DLC       | drug loading capacity             |
| DSC       | differential scanning calorimetry |
| EE        | encapsulation efficiency          |
| ELISA     | enzyme-linked immunosorbent assay |
| GIT       | gastrointestinal tract            |
| GRAS      | generally recognized as safe      |
| HPO       | hydrogenated palm oil             |
| MWCO      | molecular weight cut-off          |
| NLC       | nanostructured lipid carriers     |
| $P_{app}$ | permeability coefficient          |
| PDI       | polydispersity index              |
| $R^2$     | correlation coefficient           |
| SGF       | simulated gastric fluid           |
| SIF       | simulated intestinal fluid        |
| SLN       | solid lipid nanoparticle          |

# CHAPTER 1

## INTRODUCTION

### 1.1 Background

Malignant tumour or cancer is a family of genetic and epigenetic diseases in which a single cell in the body overrides the normal controls of mitosis and cell division (Richards & Hawley, 2011; Riley & Anderson, 2011). It is characterized by a deregulated proliferation of cells with the consequences of an abnormal increase of the cell number in particular organs, invasion of the surrounding tissues, formation of metastasis and eventually choking off the life (Schwab, 2011).

Globally, vast effort and innumerable researches have been conducted to improve the miserable outcomes of cancer. Nonetheless, cancer remains as one of the most hazardous and fast propagating diseases of the current century. In the past 30 years, the overall mortality rates from cancer have shown no significant decline (Ameta *et al.*, 2012; Zhao *et al.*, 2010). In 2004, cancer stayed as the second leading cause of death worldwide (following heart diseases), accounted for 12.6% of the total death cases. Globally, one in eight deaths is due to cancer and cancer causes more deaths than total death cases caused by AIDS, tuberculosis and malaria (American Cancer Society, 2011; Jemal *et al.*, 2011). In 2012, malignant neoplasms, which accounted for 14.9% of the total death cases (8.2 million death cases) have overtaken the ischemic heart disease (7.4 million death cases) as the first leading cause of death worldwide, ahead of other causes of death such as stroke (6.7 million death cases), chronic obstructive pulmonary disease (3.1 million death cases) and AIDS (1.5 million death cases) (Ferlay *et al.*, 2015; WHO, 2014; Ferlay *et al.*, 2013).

According to the report of the GLOBOCAN project (GLOBOCAN-2012), there were 14.06 million estimated new cancer cases reported worldwide (increase by 10.24% as compared to year 2008), of which 6.05 million (43.03%) and 8.01 million (56.97%) cases occurred in economically developed countries and economically developing countries, respectively, in 2012. Meanwhile, a total of 8.20 million cancer deaths (about 22,500 cancer deaths per day) were estimated worldwide, with 2.88 million in economically developed countries and 5.32 million in economically developing countries. Due to the growth and aging of the population, it is expected that the global cancer burden is going to increase to 23.98 million new cancer cases and 14.63 million cancer deaths by 2035, if the current trends continue. Worldwide, lung (12.42 million cases, 16.8%), prostate (1.09 million cases, 14.8%), colorectal (7.46 million cases, 10.1%), stomach (6.31 million cases, 8.5%) and liver cancer (5.54 million cases, 7.5%) were the five most commonly diagnosed cancer among men, while breast (16.71 million cases, 25.1%), colorectal (6.14 million cases, 9.2%), lung (5.83 million cases, 8.8%), cervix uteri (5.28 million cases, 7.9%) and corpus uteri cancer (3.20 million cases, 4.8%) among women (Ferlay *et al.*, 2015; Ferlay *et al.*, 2013).



In 2012, cancer (11.64%) was ranked as the fourth leading cause of death in the Ministry of Health Malaysia Hospitals after cardiovascular diseases (24.69%), diseases of pulmonary circulation (18.80%) and infectious and parasitic diseases (17.17%) (Ministry of Health Malaysia, 2013). A total of 37,426 new cancer cases (increase by 105.4% as compared to year 2007) were reported in the year 2012, comprising 18,125 males (48.43%) and 19,301 females (51.57%) in Malaysia. In the meantime, a total of 21,678 cancer deaths were estimated in Malaysia, of which 11,281 (52.04%) and 10,397 (47.96%) cases occurred in male and female, respectively. Lung cancer (17.88%) was the most frequently diagnosed cancer among Malaysian males in 2012, followed by colorectal (14.14%), nasopharynx (8.20%), prostate (6.54%) and stomach cancer (6.49%). Meanwhile, the five most commonly reported female cancers were breast (28.03%), cervix uteri (11.11%), colorectal (10.24%), lung (6.03%) and ovarian cancer (5.69%) (Ferlay *et al.*, 2013; Zainal Ariffin & Nor Saleha, 2011).

High rate of mitosis and uncontrolled proliferation of the cervical cells result in the development of cervical cancer (Chang *et al.*, 2010b). Primarily, due to the introduction of Papanicolaou (PAP) smear screening programme, the cervical carcinoma incidences and mortality rates have gradually declined. Nevertheless, it remains one of the third most prevalent gynaecologic malignancies and major causes of cancer death in women around the world (Zagouri *et al.*, 2012). Cervical squamous cell carcinoma, which is more common than cervical adenocarcinoma, accounts approximately for 80% of all invasive cervical neoplasia (Katanyoo *et al.*, 2012). Due to the high oncogenic human papilloma virus (HPV) infection rates, the absence of screening programmes and the lack of access to affordable HPV vaccination programmes, cervical cancer causes a major health problem in the developing countries (Diaz-Padilla *et al.*, 2013).

Approximately, 528,000 new cases (83,000 cases in developed countries and 445,000 cases in developing countries) and 266,000 cancer deaths (36,000 deaths in developed countries and 230,000 deaths in developing countries) were attributed to cervical cancer worldwide in 2012. Cancer of the cervix was the second most commonly diagnosed cancer (11.11% of total female cancers) among women in Malaysia in 2012. There were a total of 2,145 new cases (increase by 153.2% as compared to year 2007) of cervical neoplasia diagnosed in Malaysia, with the age-standardized incidence rate (ASR) of 15.6 per 100,000 population (Ferlay *et al.*, 2013; Zainal Ariffin & Nor Saleha, 2011).

Cis-diamminedichloroplatinum(II) or cisplatin-based chemoradiotherapy is the commonest, well known and promising regime used to manage cervical cancer. Despite of its capability of destroying the rapidly growing cancer cells, it damages the non-targeted, proliferating normal healthy cells as well. Cisplatin shows undesirable adverse-effects that include mild myelosuppression, ototoxicity and neurotoxicity. However, the most significant dose-limiting toxicity caused by cisplatin is renal dysfunction (Singh *et al.*, 2013b; Jiang *et al.*, 2012). Moreover, emergence of multidrug resistance (MDR) in cervical cancer patients hinders the success of the cisplatin-based chemoradiotherapy (Lo & Wang, 2013).

Natural products, including plants, have become the most significant natural resources for the discovery of anti-cancer compounds (Kitdamrongtham *et al.*, 2013; Zhang *et al.*, 2013b; Zhao *et al.*, 2013). It is believed that natural products or bioactive compounds derived from natural products usually exhibit low toxicity, inexpensive and are well tolerated by the human body (Greenlee, 2012; Wu *et al.*, 2011). Due to the advance in analytical chemistry and chemical biology, characterizations of commonly used herbs or

herbal formulation from natural products provide the possible discoveries of bioactive ingredients or entities (Li & Zhang, 2013). Since 1981 to 2010, over 74.8% of the approved drugs were derived from natural products (Newman & Cragg, 2012; Nobili *et al.*, 2009). The success stories of vinca alkaloids (vinblastine and vincristine) from *Catharanthus roseus* (Madagascar periwinkle) as well as paclitaxel (Taxol®) from *Taxus brevifolia* (Pacific Yew tree) encourage a continuous research and discovery on potential anti-cancer candidates from natural products (Zhang *et al.*, 2013b; Manosroi *et al.*, 2012).

*Nigella sativa* (also known as black seed or *habbatus sauda*) appears as one of the important herbs among various medicinal plants. Majority of the biological activities of *Nigella sativa* are associated with the presence of thymoquinone (TQ), the major bioactive compound found in the seeds of the plant (Ahmad *et al.*, 2013). *N. sativa* and TQ are well known with their biological activities that include anti-cancer (Peng *et al.*, 2013; Mahmoud & Torchilin, 2012; Woo *et al.*, 2011), anti-oxidant (Bourgou *et al.*, 2012; Umar *et al.*, 2012), anti-inflammatory (Alemi *et al.*, 2013; Chehl *et al.*, 2009), hepatoprotective (Zafeer *et al.*, 2012; Yildiz *et al.*, 2008) and renal protective (Saleem *et al.*, 2012; Ulu *et al.*, 2012; Yildiz *et al.*, 2010) properties.

Among all the route of administrations, non-invasive oral route is the most preferred route for chemotherapy. Nevertheless, oral administration of 50% of the drug compounds is hampered by the lipophilic property of the drugs. In fact, among the newest drug candidates, 40% exhibit low water solubility, resulting in poor oral bioavailability (Dey *et al.*, 2012; Kattaboyna *et al.*, 2009). Most of the time, these poor water solubility anticancer candidates are dissolved in surfactants or in organic solvents such as dehydrated alcohol which cause adverse side effects to the patients. These side effects include neurotoxicity, nephrotoxicity and cardiotoxicity (Mognetti *et al.*, 2012). Similar trends are expected from TQ as it is lipophilic in nature with relatively low water solubility (Khader *et al.*, 2009). Therefore, oral delivery of TQ will be limited by the solubility-related poor oral bioavailability (Pathan *et al.*, 2011).

In order to overcome the low solubility and low bioavailability of the active anticancer compounds, lipid based nano-drug delivery systems particularly solid lipid nanoparticles (SLNs) and nanostructured lipid carriers (NLCs) are utilised as potential carriers for hydrophobic and lipophilic drugs (Zhuang *et al.*, 2010). Both SLNs and NLCs confer many advantages including capability of increasing the bioavailability of poorly soluble compounds, providing protection for sensitive active compounds and facilitating controlled release of drugs (Müller *et al.*, 2000). NLC is the second-generation of lipid based nanoparticles which are developed from SLN. NLC overcomes the limitations of SLN, which include low drug loading capacity and drug expulsion during storage (Fang *et al.*, 2008).

Although has been documented to exhibit cytotoxic effects in several cancer cell lines, the cytotoxicity and mechanisms of action of TQ from *Nigella sativa* towards cervical cancer cells (HeLa and SiHa) have not been investigated yet. Furthermore, the formulation and characterisation of thymoquinone-loaded nanostructure lipid carrier (TQ-NLC) as well as its cytotoxicity have not been performed before.

## 1.2 Objective

### 1.2.1 General Objective

The general objective of the study was to determine the cytotoxicity of TQ and TQ-NLC towards cervical cancer cells (SiHa and HeLa).

### 1.2.2 Specific Objectives

The specific objectives were:

- (1) to determine the cytotoxicity of TQ towards cervical cancer cells (SiHa and HeLa) and the mode of cell death induced by TQ,
- (2) to study the mechanisms of action underlying cytotoxicity of TQ,
- (3) to formulate and synthesize TQ-NLC,
- (4) to characterize the physicochemical properties, drug release profile and *in vitro* bioavailability of TQ-NLC and
- (5) to evaluate the cytotoxicity and mode of cell death induced by TQ-NLC.

## 1.3 Hypotheses

TQ will be cytotoxic towards all the selected cancer cell lines by inducing cell cycle arrest and apoptosis. The induction of apoptosis by TQ will involve p53-dependent intrinsic pathway.

TQ-NLC will be formulated. NLC will confer drug controlled release, protection, and enhanced bioavailability and cytotoxicity to TQ.

## REFERENCES

- Abel, E.L., & DiGiovanni, J. (2011). Multistage carcinogenesis. In T.M. Penning (Ed.), *Chemical Carcinogenesis* (pp. 27–51). New York, NY: Humana Press.
- Acharya, B.R., Chatterjee, A., Ganguli, A., Bhattacharya, S., & Chakrabarti, G. (2014). Thymoquinone inhibits microtubule polymerization by tubulin binding and causes mitotic arrest following apoptosis in A549 cells. *Biochimie*, *97*, 78–91.
- Adams, J.M., & Cory, S. (2007). The Bcl-2 apoptotic switch in cancer development and therapy. *Oncogene*, *26*(9), 1324–1337.
- Adey, A., Burton, J.N., Kitzman, J.O., Hiatt, J.B., Lewis, A.P., Martin, B.K., Qiu, R., Lee, C., & Shendure, J. (2013). The haplotype-resolved genome and epigenome of the aneuploid HeLa cancer cell line. *Nature*, *500*(7461), 207–211.
- Aditya, N.P., Macedo, A.S., Doktorovova, S., Souto, E.B., Kim, S., Chang, P.S., & Ko, S. (2014). Development and evaluation of lipid nanocarriers for quercetin delivery: A comparative study of solid lipid nanoparticles (SLN), nanostructured lipid carriers (NLC), and lipid nanoemulsions (LNE). *LWT-Food Science & Technology*, *59*(1), 115–121.
- Aditya, N.P., Shim, M., Lee, I., Lee, Y.J., Im, M.H., & Ko, S. (2013). Curcumin and genistein coloaded nanostructured lipid carriers: *in vitro* digestion and antiprostata cancer activity. *Journal of Agricultural & Food Chemistry*, *61*(8), 1878–1883.
- Ahmad, A., Husain, A., Mujeeb, M., Khan, S.A., Najmi, A.K., Siddique, N.A., Damanhour, Z. A., & Anwar, F. (2013). A review on therapeutic potential of *Nigella sativa*: A miracle herb. *Asian Pacific Journal of Tropical Biomedicine*, *3*(5), 337–352.
- Ait Mbarek, L., Ait Mouse, H., Elabbadi, N., Bensalah, M., Gamouh, A., Aboufatima, R., Benharref, A., Chait, A., Kamal, M., Dalal, A., & Ziad, A. (2007). Antitumor properties of blackseed (*Nigella sativa* L.) extracts. *Brazilian Journal of Medical & Biological Research*, *40*(6), 839–847.
- Akhondian, J., Kianifar, H., Raoofzadeh, M., Moayedpour, A., Toosi, M.B., & Khajedaloe, M. (2011). The effect of thymoquinone on intractable pediatric seizures (pilot study). *Epilepsy Research*, *93*(1), 39–43.
- Alam, S., Khan, Z., Mustafa, G., Kumar, M., Islam, F., Bhatnagar, A., & Ahmad, F.J. (2012). Development and evaluation of thymoquinone-encapsulated chitosan nanoparticles for nose-to-brain targeting: a pharmacoscintigraphic study. *International Journal of Nanomedicine*, *7*, 5705–5718.
- Albanese, A., & Chan, W.C.W. (2011). Effect of gold nanoparticle aggregation on cell uptake and toxicity. *ACS Nano*, *5*(7), 5478–5489.

- Alemi, M., Sabouni, F., Sanjarian, F., Haghbeen, K., & Ansari, S (2013). Anti-inflammatory effect of seeds and callus of *Nigella sativa* L. extracts on mix glial cells with regard to their thymoquinone content. *AAPS PharmSciTech*, 14(1), 160–167.
- Alenzi, F.Q., Lotfy, M., & Wyse, R. (2010). Swords of cell death: caspase activation and regulation. *Asian Pacific Organization for Cancer Prevention*, 11(2), 271–280.
- Alexis, F., Rhee, J.W., Richie, J.P., Radovic-Moreno, A.F., Langer, R., & Farokhzad, O.C. (2008). New frontiers in nanotechnology for cancer treatment. *Urologic Oncology*, 26(1), 74–85.
- Alhebshi, A.H., Gotoh, M., & Suzuki, I. (2013). Thymoquinone protects cultured rat primary neurons against amyloid  $\beta$ -induced neurotoxicity. *Biochemical & Biophysical Research Communications*, 433(4), 362–367.
- Alhosin, M., Abusnina, A., Achour, M., Sharif, T., Muller, C., Peluso, J., Chataigneau, T., Lugnier, C., Schini-Kerth, V.B., Bronner, C., & Fuhrmann, G. (2010). Induction of apoptosis by thymoquinone in lymphoblastic leukemia Jurkat cells is mediated by a p73-dependent pathway which targets the epigenetic integrator UHRF1. *Biochemical Pharmacology*, 79(9), 1251–1260.
- Ali, B.H., & Blunden, G. (2003). Pharmacological and toxicological properties of *Nigella sativa*. *Phytotherapy Research*, 17(4), 299–305.
- Ali, Z., Ferreira, D., Carvalho, P., Avery, M.A., & Khan, I.A. (2008). Nigellidine-4-*O*-sulfite, the first sulfated indazole-type alkaloid from the seeds of *Nigella sativa*. *Journal of Natural Products*, 71(6), 1111–1112.
- Alkam, Y., Mitomi, H., Nakai, K., Himuro, T., Saito, T., Takahashi, M., Arakawa, A., Yao, T., & Saito, M. (2013). Protein expression and methylation of DNA repair genes *hMLH1*, *hMSH2*, *MGMT* and *BRCA1* and their correlation with clinicopathological parameters and prognosis in basal-like breast cancer. *Histopathology*, 63(5), 713–725.
- Al-Naqeep, G.N., Ismail, M.M., Al-Zubairi, A.S., & Esa, N.M. (2009) Nutrients composition and minerals content of three different samples of *Nigella sativa* L. cultivated in Yemen. *Asian Journal of Biological Sciences*, 2(2), 43–48.
- Al-Qurashi, A.R., Akhtar, N., Al-Jabre, S., AL-Akloby, O., & Randhawa, M.A. (2007). Anti-fungal activity of thymoquinone and amphotericin B against *Aspergillus niger*. *Scientific Journal of King Faisal University*, 8(1), 143–149.
- Amara, F., Muzi-Falconi, M., & Plevani, P. (2013). Cell cycle checkpoints. In W. Dubitzky, O. Wolkenhauer, K.H. Cho, H. Yokota (Eds.), *Encyclopedia of Systems Biology* (pp. 254–259). New York: Springer New York.
- American Cancer Society (2011). *Global Cancer Facts & Figures 2nd Edition*. Atlanta: American Cancer Society, Inc.



- American Cancer Society (2014). *Cancer Facts & Figures 2014*. Atlanta: American Cancer Society, Inc.
- Ameta, K.L., Rathore, N.S., & Kumar, B. (2012). Synthesis and *in vitro* anti-breast cancer activity of some novel 1,5-benzothiazepine derivatives. *Journal of the Serbian Chemical Society*, 77(6), 725–731.
- An, M.J., Cheon, J.H., Kim, S.W., Kim, E.S., Kim, T.I., & Kim, W.H. (2009). Guggulsterone induces apoptosis in colon cancer cells and inhibits tumor growth in murine colorectal cancer xenografts. *Cancer Letters*, 279(1), 93–100.
- Ang, W.H., Myint, M., & Lippard, S.J. (2010). Transcription inhibition by platinum-DNA cross-links in live mammalian cells. *Journal of the American Chemical Society*, 132(21), 7429–7435.
- Ao, L., Liu, J.Y., Liu, W.B., Gao, L.H., Hu, R., Fang, Z.J., Zhen, Z.X., Huang, M.H., Yang, M.S. & Cao, J. (2010). Comparison of gene expression profiles in BALB/c 3T3 transformed foci exposed to tumor promoting agents. *Toxicology in Vitro*, 24(2), 430–438.
- Aomori, T., Makino, H., Sekizuka, M., Hashita, T., Araki, T., Iizuka, K., Nakamura, T., & Yamamoto, K. (2012). Effect of ethanol in Paclitaxel injections on the ethanol concentration in exhaled breath. *Drugs in R&D*, 12(3), 165–170.
- Arafa, el-S.A., Zhu, Q., Shah, Z.I., Wani, G., Barakat, B.M., Racoma, I., El-Mahdy, M.A., & Wani, A.A. (2011). Thymoquinone up-regulates PTEN expression and induces apoptosis in doxorubicin-resistant human breast cancer cells. *Mutation Research*, 706(1–2), 28–35.
- Araújo, J., Garcia, M.L., Mallandrich, M., Souto, E.B., & Calpena, A.C. (2012). Release profile and transscleral permeation of triamcinolone acetonide loaded nanostructured lipid carriers (TA-NLC): *in vitro* and *ex vivo* studies. *Nanomedicine*, 8(6):1034–1041.
- Arepalli, S.K., Sridhar, V., Rao, J.V., Kennady, P.K., & Venkateswarlu, Y. (2009). Furano- sesquiterpene from soft coral, *Sinularia kavarittiensis*: induces apoptosis via the mitochondrial-mediated caspase-dependent pathway in THP-1, leukemia cell line. *Apoptosis*, 14(5), 729–740.
- Arifin, D.Y., Lee, L.Y., & Wang, C.H. (2006). Mathematical modeling and simulation of drug release from microspheres: Implications to drug delivery systems. *Advanced Drug Delivery Reviews*, 58(12-13), 1274–1325.
- Artursson, P., & Karlsson, J. (1991). Correlation between oral drug absorption in humans and apparent drug permeability coefficients in human intestinal epithelial (Caco-2) cells. *Biochemical & Biophysical Research Communications*, 175(3), 880–885.
- Aswathy, S., Quereshi, M.A., Kurian, B., & Leelamoni, K. (2012). Cervical cancer screening: Current knowledge & practice among women in a rural population of Kerala, India. *Indian Journal of Medical Research*, 136(2), 205–210.

- Aulisa, L., Forraz, N., McGuckin, C., & Hartgerink, J.D. (2009). Inhibition of cancer cell proliferation by designed peptide amphiphiles. *Acta Biomaterialia*, 5(3), 842–853.
- Awad, E.M. (2005). *In vitro* decreases of the fibrinolytic potential of cultured human fibrosarcoma cell line, HT1080, by *Nigella sativa* oil. *Phytomedicine*, 12(1–2), 100–107.
- Aydn, R.S.T., & Pulat, M. (2012). 5-Fluorouracil encapsulated chitosan nanoparticles for pH-stimulated drug delivery: Evaluation of controlled release kinetics. *Journal of Nanomaterials*, 2012. Doi: 10.1155/2012/313961.
- Aziz, N.A., Majdina, H., Hassan, Y., Zulkifly, H.H., Wahab, M.S.A., Aziz, M.S.A., Yahaya, N., & AbdulRazzaq, H.A. (2014). Assessment of the Halal status of respiratory pharmaceutical products in a hospital. *Procedia-Social & Behavioral Sciences*, 121, 158–165.
- Babu, A., Templeton, A.K., Munshi, A., & Ramesh, R. (2013). Nanoparticle-based drug delivery for therapy of lung cancer: progress and challenges. *Journal of Nanomaterials*, 2013. Doi: 10.1155/2013/863951.
- Badary, O.A., & Gamal El-Din, A.M. (2001). Inhibitory effects of thymoquinone against 20-methylcholanthrene-induced fibrosarcoma tumorigenesis. *Cancer Detection & Prevention*, 25(4), 362–368.
- Badary, O.A., Abdel-Naim, A.B., Abdel-Wahab, M.H., & Hamada, F.M. (2000). The influence of thymoquinone on doxorubicin-induced hyperlipidemic nephropathy in rats. *Toxicology*, 143(3), 219–226.
- Badr, G., Mahmoud, M.H., Farhat, K., Waly, H., Al-Abdin, O.Z., & Rabah, D.M. (2013). Maternal supplementation of diabetic mice with thymoquinone protects their offspring from abnormal obesity and diabetes by modulating their lipid profile and free radical production and restoring lymphocyte proliferation via PI3K/AKT signaling. *Lipids in Health & Disease*, 12(37). Doi: 10.1186/1476-511X-12-37.
- Balaha, M.F., Tanaka, H., Yamashita, H., Abdel Rahman, M.N., & Inagaki N. (2012). Oral *Nigella sativa* oil ameliorates ovalbumin-induced bronchial asthma in mice. *International Immunopharmacology*, 14(2), 224–231.
- Barba, A.A., d'Amore, M., Chirico, S., Lamberti, G., & Titomanlio, G. (2009). A general code to predict the drug release kinetics from different shaped matrices. *European Journal of Pharmaceutical Sciences*, 36(2-3), 359–368.
- Barnes, M.A., McMullen, M.R., Roychowdhury, S., Pisano, S.G., Liu, X., Stavitsky, A.B., Bucala, R., & Nagy, L.E. (2013). Macrophage migration inhibitory factor contributes to ethanol-induced liver injury by mediating cell injury, steatohepatitis, and steatosis. *Hepatology*, 57(5), 1980–1991.
- Barrasa, J.I., Olmo, N., Lizarbe, M.A., & Turnay, J. (2013). Bile acids in the colon, from healthy to cytotoxic molecules. *Toxicology in Vitro*, 27(3), 964–977.

- Baskić, D., Popović, S., Ristić, P., & Arsenijević, N.N. (2006). Analysis of cycloheximide-induced apoptosis in human leukocytes: fluorescence microscopy using annexin V/propidium iodide versus acridin orange/ethidium bromide. *Cell Biology International*, 30(11), 924–932.
- Bawa, R., Fung, S.Y., Shiozaki, A., Yang, H., Zheng, G., Keshavjee, S., & Liu, M. (2012). Self-assembling peptide-based nanoparticles enhance cellular delivery of the hydrophobic anticancer drug ellipticine through caveolae-dependent endocytosis. *Nanomedicine: Nanotechnology, Biology & Medicine*, 8(5), 647–654.
- Bawarski, W.E., Chidlow, E., Bharali, D.J., & Mousa, S.A. (2008). Emerging nanopharmaceuticals. *Nanomedicine: Nanotechnology, Biology & Medicine*, 4(4), 273–282.
- Beck, R.C., Pohlmann, A.R., Hoffmeister, C., Gallas, M.R., Collnot, E., Schaefer, U.F., Guterres, S.S., & Lehr, C.M. (2007). Dexamethasone-loaded nanoparticle-coated microparticles: correlation between *in vitro* drug release and drug transport across Caco-2 cell monolayers. *European Journal of Pharmaceutics & Biopharmaceutics*, 67(1), 18–30.
- Beckerman, R., & Prives, C. (2010). Transcriptional regulation by p53. *Cold Spring Harbor Perspectives in Biology*, 2. Doi: 10.1101/cshperspect.a000935.
- Beesoo, R., Neergheen-Bhujun, V., Bhagooli, R., & Bahorun, T. (2014). Apoptosis inducing lead compounds isolated from marine organisms of potential relevance in cancer treatment. *Mutation Research-fundamental & Molecular Mechanisms of Mutagenesis*, 768, 84–97.
- Behl, C., & Ziegler, C. (2014). Cell cycle: the life cycle of a cell. In C. Behl, & C. Ziegler (Eds.), *Cell Aging: Molecular Mechanisms and Implications for Disease* (pp. 3–14). Berlin: Springer Berlin Heidelberg.
- Beija, M., Salvayre, R., Lauth-de Viguerie, N., & Marty, J.D. (2012). Colloidal systems for drug delivery: from design to therapy. *Trends in Biotechnology*, 30(9), 485–496.
- Benamira, M., Johnson, K., Chaudhary, A., Bruner, K., Tibbetts, C., & Marnett, L.J. (1995). Induction of mutations by replication of malondialdehyde-modified M13 DNA in *Escherichia coli*: determination of the extent of DNA modification, genetic requirements for mutagenesis, and types of mutations induced. *Carcinogenesis*, 16(1), 93–99.
- Bharali, D.J., & Mousa, S.A. (2010). Emerging nanomedicines for early cancer detection and improved treatment: current perspective and future promise. *Pharmacology & Therapeutics*, 128(2), 324–335.



- Bhattacharya, S., Ahir, M., Patra, P., Mukherjee, S., Ghosh, S., Mazumdar, M., Chattopadhyay, S., Das, T., Chattopadhyay, D., & Adhikary, A. (2015). PEGylated-thymoquinone-nanoparticle mediated retardation of breast cancer cell migration by deregulation of cytoskeletal actin polymerization through miR-34a. *Biomaterials*, *51*, 91–107.
- Bhattacharyya, G.S. (2010). Oral systemic therapy: Not all "win-win". *Indian Journal of Medical & Paediatric Oncology*, *31*(1), 1–3.
- Bhumkar, D.R., Joshi, H.M., Sastry, M., & Pokharkar, V.B. (2007). Chitosan reduced gold nanoparticles as novel carriers for transmucosal delivery of insulin. *Pharmaceutical Research*, *24*(8):1415–1426.
- Biswas, S., Dodwadkar, N.S., Deshpande, P.P., & Torchilin, V.P. (2012). Liposomes loaded with paclitaxel and modified with novel triphenylphosphonium-PEG-PE conjugate possess low toxicity, target mitochondria and demonstrate enhanced antitumor effects *in vitro* and *in vivo*. *Journal of Controlled Release*, *159*(3), 393–402.
- Böing, A.N., Stap, J., Hau, C.M., Afink, G.B., Ris-Stalpers, C., Reits, E.A., Sturk, A., van Noorden, C.J., & Nieuwland, R. (2013). Active caspase-3 is removed from cells by release of caspase-3-enriched vesicles. *Biochimica et Biophysica Acta*, *1833*(8), 1844–1852.
- Bondì, M.L., Azzolina, A., Craparo, E.F., Botto, C., Amore, E., Giammona, G., & Cervello, M. (2014). Entrapment of an EGFR inhibitor into nanostructured lipid carriers (NLC) improves its antitumor activity against human hepatocarcinoma cells. *Journal of Nanobiotechnology*, *12*(21). Doi: 10.1186/1477-3155-12-21.
- Bonneau, B., Prudent, J., Popgeorgiev, N., & Gillet, G. (2013). Non-apoptotic roles of Bcl-2 family: the calcium connection. *Biochimica et Biophysica Acta*, *1833*(7), 1755–1765.
- Bourgou, S., Pichette, A., Marzouk, B., & Legault, J. (2012). Antioxidant, anti-inflammatory, anticancer and antibacterial activities of extracts from *Nigella sativa* (Black Cumin) plant parts. *Journal of Food Biochemistry*, *36*(5): 539–546.
- Boya, P., & Kroemer, G. (2008). Lysosomal membrane permeabilization in cell death. *Oncogene*, *27*, 6434–6451.
- Bradford, M.M. (1976). A rapid and sensitive method for quantitation of microgram quantities of protein utilizing the principle of protein–dye binding. *Analytical Biochemistry*, *72*(1–2), 248–254.
- Broadhead, M.L., Dass, C.R., & Choong, P.F.M. (2009). Cancer cell apoptotic pathways mediated by PEDF: prospects for therapy. *Trends in Molecular Medicine*, *15*(10), 461–467.
- Burdock, G.A., & Carabin, I.G. (2004). Generally recognized as safe (GRAS): history and description. *Toxicology Letters*, *150*(1), 3–18.

- Burz, C., Berindan-Neagoe, I., Balacescu, O., & Irimie, A. (2009). Apoptosis in cancer: key molecular signaling pathways and therapy targets. *Acta Oncologica*, 48(6), 811–821.
- Buy, J.N., & Ghossain, M. (2013). Embryology, anatomy, and histology of the cervix. In J.N. Buy, and M. Ghossain, (Eds) *Gynecological Imaging* (pp. 649–652). Berlin: Springer Berlin Heidelberg.
- Caracciolo, G. (2014). Liposome-protein corona in a physiological environment: Challenges and opportunities for targeted delivery of nanomedicines. *Nanomedicine: Nanotechnology, Biology & Medicine*, 11, 543–557.
- Carlomagno, N., Duraturo, F., Rizzo, G., Cremone, C., Izzo, P., & Renda, A. (2009). Carcinogenesis. In A. Renda (Eds.), *Multiple Primary Malignancies* (pp. 51–61). Milan: Springer-Verlag Italia.
- Carneiro, P., Fernandes, M.S., Figueiredo, J., Caldeira, J., Carvalho, J., Pinheiro, H., Leite, M., Melo, S., Oliveira, P., Simões-Correia, J., Oliveira, M.J., Carneiro, F., Figueiredo, C., Paredes, J., Oliveira, C., & Seruca, R. (2012). E-cadherin dysfunction in gastric cancer--cellular consequences, clinical applications and open questions. *FEBS Letters*, 586(18), 2981–2989.
- Carosati, E., Sforza, G., Pippi, M., Marverti, G., Ligabue, A., Guerrieri, D., Piras, S., Guitoli, G., Luciani, R., Costi, M.P., & Cruciani, G. (2010). Ligand-based virtual screening and ADME-tox guided approach to identify triazolo-quinolines as folate cycle inhibitors. *Bioorganic & Medicinal Chemistry Letters*, 18(22), 7773–7785.
- CDC (2012). Sexually transmitted diseases (STDs): HPV vaccine information for young women - fact sheet. Retrieved September 6, 2014, from the Centers for Disease Control and Prevention website: <http://www.cdc.gov/std/hpv/stdfact-hpv-vaccine-young-women.htm>.
- Cepeda, V., Fuertes, M.A., Castilla, J., Alonso, C., Quevedo, C., & Pérez, J.M. (2007). Biochemical mechanisms of cisplatin cytotoxicity. *Anti-Cancer Agents in Medicinal Chemistry*, 7(1), 3–18.
- Chaieb, K., Koudhi, B., Jrah, H., Mahdouani, K., & Bakhrouf, A. (2011). Antibacterial activity of thymoquinone, an active principle of *Nigella sativa* and its potency to prevent bacterial biofilm formation. *BMC Complementary & Alternative Medicine*, 11(29). Doi: 10.1186/1472-6882-11-29.
- Chang, H.R., Huang, H.P., Kao, Y.L., Chen, S.L., Wu, S.W., Hung, T.W., Lian, J.D., & Wang, C.J. (2010a). The suppressive effect of Rho kinase inhibitor, Y-27632, on oncogenic Ras/RhoA induced invasion/migration of human bladder cancer TSGH cells. *Chemico-Biological Interactions*, 183(1), 172–180.
- Chang, W.C., Hsieh, C.H., Hsiao, M.W., Lin, W.C., Hung, Y.C., & Ye, J.C. (2010b). Caffeic acid induces apoptosis in human cervical cancer cells through the mitochondrial pathway. *Taiwanese Journal of Obstetrics & Gynecology*, 49(4), 419–424.

- Chaudhary, A., Tiwari, N., Jain, V., & Singh, R. (2011). Microporous bilayer osmotic tablet for colon-specific delivery. *European Journal of Pharmaceutics & Biopharmaceutics*, 78(1), 134–140.
- Chehl, N., Chipitsyna, G., Gong, Q., Yeo, C.J., & Arafat, H.A. (2009). Anti-inflammatory effects of the *Nigella sativa* seed extract, thymoquinone, in pancreatic cancer cells. *HPB (Oxford)*, 11(5), 373–381.
- Chen, C.C., Tsai, T.H., Huang, Z.R., & Fang, J.Y. (2010). Effects of lipophilic emulsifiers on the oral administration of lovastatin from nanostructured lipid carriers: physicochemical characterization and pharmacokinetics. *European Journal of Pharmaceutics & Biopharmaceutics*, 74(3), 474–482.
- Chen, W.C., Ma, B., Mao, C., & Wu, T.C. (2011). Molecular pathogenesis, detection and clinical management of pre-invasive cervical lesions. In R.C. Fitzgerald (Ed.), *Pre-Invasive Disease: Pathogenesis and Clinical Management* (pp. 437–466). New York, NY: Springer New York.
- Cho, E.S., Lee, K.W., & Lee, H.J. (2008). Cocoa procyanidins protect PC12 cells from hydrogen-peroxide-induced apoptosis by inhibiting activation of p38 MAPK and JNK. *Mutation Research/Fundamental & Molecular Mechanisms of Mutagenesis*, 640(1–2), 123–130.
- Choi, I.K., Sung, H.J., Lee, J.H., Kim, J., & Seo, J.H. (2012). The relationship between *Helicobacter pylori* infection and the effects of chemotherapy in patients with advanced or metastatic gastric cancer. *Cancer Chemotherapy & Pharmacology*, 70(4), 555–558.
- Chu, E., & Sartorelli, A.C. (2004). Cancer chemotherapy. In B.G. Katzung (9th Eds.), *Basic & Clinical Pharmacology* (pp. 898–930). New York, NY: McGraw-Hill Global Education Holdings.
- Compton, C.C., Byrd, D.R., Garcia-Aguilar, J., Kurtzman, S.H., Olawaiye, A., & Washington, M.K. (2012). Cervix uteri. In C.C. Compton, D.R. Byrd, J. Garcia-Aguilar, S.H. Kurtzman, A. Olawaiye, & M.K. Washington (Eds.), *AJCC Cancer Staging Atlas* (pp. 463–475). New York, NY: Springer New York.
- Coutts, A.S., & La Thangue, N. (2006). The p53 response during DNA damage: impact of transcriptional cofactors. *Biochemical Society Symposia*, 73, 181–189.
- Csika'sz-Nagy, A. (2013). Cell cycle. In W. Dubitzky, O. Wolkenhauer, K.H. Cho, H. Yokota (Eds.), *Encyclopedia of Systems Biology* (pp. 220–231). New York: Springer New York.
- Dahan, A., & Hoffman, A. (2008). Rationalizing the selection of oral lipid based drug delivery systems by an *in vitro* dynamic lipolysis model for improved oral bioavailability of poorly water soluble drugs. *Journal of Controlled Release*, 129(1), 1–10.

- Dash, S., Murthy, P.N., Nath, L., & Chowdhury, P. (2010). Kinetic modelling on drug release from controlled drug delivery systems. *Acta Poloniae Pharmaceutica-Drug Research*, 67(3), 217–223.
- Davidson, S. (2011). Treatment for advanced cervical cancer: impact on quality of life. *Critical Reviews in Oncology/Hematology*, 79(1), 24–30.
- Dawidowicz, A.L., Rado, E., Wianowska, D., Mardarowicz, M., & Gawdzik, J. (2008). Application of PLE for the determination of essential oil components from *Thymus vulgaris* L. *Talanta*, 76(4), 878–884.
- Desai, K.G.H., & Park, H.J. (2004). Solubility studies on valdecoxib in the presence of carriers, cosolvents, and surfactants. *Drug Development Research*, 62(1), 41–48.
- Desai, N., Trieu, V., Damascelli, B., & Soon-Shiong, P. (2009). SPARC expression correlates with tumor response to albumin-bound paclitaxel in head and neck cancer patients. *Translational Oncology*, 2(2), 59–64.
- Devi, P.U. (2005). Basics of carcinogenesis. *Health Administrator*, 17(1), 16–24.
- Devraj, R., Williams, H.D., Warren, D.B., Mullertz, A., Porter, C.J., & Pouton, C.W. (2013). *In vitro* digestion testing of lipid-based delivery systems: calcium ions combine with fatty acids liberated from triglyceride rich lipid solutions to form soaps and reduce the solubilization capacity of colloidal digestion products. *International Journal of Pharmaceutics*, 441(1-2), 323–333.
- Dewson, G., & Kluck, R. (2010). Bcl-2 family-regulated apoptosis in health and disease. *Cell Health & Cytoskeleton*, 2, 9–22.
- Dey, S., Jha, S.K., Malakar, J., & Gangpodhyay, A. (2012). Improvement of bioavailability of poorly soluble drugs through self emulsifying drug delivery system. *Journal of PharmaSciTech*, 1(2), 6–11.
- Dhanalakshmi, J., Divakar, R., & Amy, M.P. (2015). To study the potential of microbial lipid based nanostructured lipid carriers for topical drug delivery applications. *International Journal of ChemTech Research*, 7(2), 795–799.
- Diaz-Padilla, I., Monk, B.J., Mackay, H.J., & Oaknin, A. (2013). Treatment of metastatic cervical cancer: Future directions involving targeted agents. *Critical Reviews in Oncology/Hematology*, 85(3), 303–314.
- Dilshad, A., Abulkhair, O., Nemenqani, D., & Tamimi, W. (2012). Antiproliferative properties of methanolic extract of *Nigella sativa* against the MDA-MB-231 cancer cell line. *Asian Pacific Journal of Cancer Prevention*, 13(11), 5839–5842.
- Ding, L., Liu, B., Qi, L., Zhou, Q., Hou, Q., Li, J., & Zhang, Q. (2009). Anti-proliferation, cell cycle arrest and apoptosis induced by a natural xanthone from *Gentianopsis paludosa* Ma, in human promyelocytic leukemia cell line HL-60 cells. *Toxicology in Vitro*, 23(3), 408–417.

- Dixon, K., & Kopras, E. (2004). Genetic alterations and DNA repair in human carcinogenesis. *Seminars in Cancer Biology*, 14(6), 441–448.
- Dua, J.S., Rana, A.C., & Bhandari, A.K. (2012). Liposome: methods of preparation and applications. *International Journal of Pharmaceutical Studies & Research*, 3(2), 14–20.
- Earnshaw, W.C., Martins, L.M., & Kaufmann, S.H. (1999). Mammalian caspases: structure, activation, substrates, and functions during apoptosis. *Annual Review of Biochemistry*, 68, 383–424.
- Eferl, R., & Wagner, E.F. (2003). AP-1: a double-edged sword in tumorigenesis. *Nature Reviews Cancer*, 3(11), 859–868.
- Effenberger, K., Breyer, S., & Schobert, R. (2010). Terpene conjugates of the *Nigella sativa* seed-oil constituent thymoquinone with enhanced efficacy in cancer cells. *Chemistry & Biodiversity*, 7(1), 129–139.
- Einstein, M.H. (2008). Acquired immune response to oncogenic human papillomavirus associated with prophylactic cervical cancer vaccines. *Cancer Immunology, Immunotherapy*, 57(4), 443–451.
- Ekamparam, P., Sathali, A.A.H., & Priyanka, K. (2012). Solid Lipid Nanoparticles: A review. *Scientific Reviews & Chemical Communications*, 2(1), 80-102.
- El-Aziz, M.A., Hassan, H.A., Mohamed, M.H., Meki, A.R., Abdel-Ghaffar, S.K., & Hussein, M.R. (2005). The biochemical and morphological alterations following administration of melatonin, retinoic acid and *Nigella sativa* in mammary carcinoma: an animal model. *International Journal of Experimental Pathology*, 86(6), 383–396.
- El-Dakhkhny, M. (1963). Studies on the chemical constitution of Egyptian *N. sativa* L. seeds. *Planta Medica*, 11(4), 465–470.
- Elgendya, E.M., & Al-Ghamdy, H. (2007). Thermal and photooxidation reactions of the steroids:  $\beta$ -sitosterol, stigmasterol and diosgenin. *Taiwan Pharmaceutical Journal*, 59(3), 113–132.
- Eliaš, J., Dimitrio, L., Clairambault, J., & Natalini, R. (2014). The p53 protein and its molecular network: modelling a missing link between DNA damage and cell fate. *Biochimica et Biophysica Acta*, 1844(1), 232–247.
- Elkholi, R., Floros, K.V., & Chipuk, J.E. (2011). The role of BH3-only proteins in tumor cell development, signaling, and treatment. *Genes & Cancer*, 2(5), 523–537.
- El-Massik, M.A., Darwish, I.A., Hassan, E.E., & El-Khordagui, L.K. (1996). Development of a dissolution medium for glibenclamide. *International Journal of Pharmaceutics*, 140(1), 69–76.
- Elmore, S. (2007). Apoptosis: a review of programmed cell death. *Toxicologic Pathology*, 35(4), 495–516.



- Emmanuel, S. (2010). *Predictive in vitro dissolution tools: application during formulation development* (Doctoral dissertation). Retrieved December 6, 2014, from HAL Dissertations and Theses. (HAL Id: tel-00719613).
- Erdem, T., Bayindir, T., Filiz, A., Iraz, M., & Selimoglu, E. (2012). The effect of resveratrol on the prevention of cisplatin ototoxicity. *European Archives of Oto-Rhino-Laryngology*, 269(10), 2185–2188.
- Fahmy, H.M., Noor, N.A., Mohammed, F.F., Elsayed, A.A., & Radwan, N.M. (2014). *Nigella sativa* as an anti-inflammatory and promising remyelinating agent in the cortex and hippocampus of experimental autoimmune encephalomyelitis-induced rats. *Journal of Basic & Applied Zoology*, 67(5), 182–195.
- Fang, J.Y., Fang, C.L., Liu, C.H., & Su, Y.H. (2008). Lipid nanoparticles as vehicles for topical psoralen delivery: Solid lipid nanoparticles (SLN) versus nanostructured lipid carriers (NLC). *European Journal of Pharmaceutics & Biopharmaceutics*, 70(2), 633–640.
- Fang, X., Shao, L., Zhang, H., & Wang, S. (2005). CHMIS-C: a comprehensive herbal medicine information system for cancer. *Journal of Medicinal Chemistry*, 48(5), 1481–1488.
- Farah, I.O. (2005). Assessment of cellular responses to oxidative stress using MCF-7 breast cancer cells, black seed (*N. Sativa* L.) extracts and H<sub>2</sub>O<sub>2</sub>. *International Journal of Environmental Research & Public Health*, 2(3-4), 411–419.
- Farah, I.O., & Begum, R.A. (2003). Effect of *Nigella sativa* (*N. sativa* L.) and oxidative stress on the survival pattern of MCF-7 breast cancer cells. *Biomedical Sciences Instrumentation*, 39, 359–364.
- Farah, N., Benghuzzi, H., Tucci, M., & Cason, Z. (2005). The effects of isolated antioxidants from black seed on the cellular metabolism of A549 cells. *Biomedical Sciences Instrumentation*, 41, 211–216.
- Fatouros, D.G., Bergenstahl, B., & Mullertz, A. (2007a). Morphological observations on a lipid-based drug delivery system during *in vitro* digestion. *European Journal of Pharmaceutical Sciences*, 31(2), 85–94.
- Fatouros, D.G., Deen, G.R., Arleth, L., Bergenstahl, B., Nielsen, F.S., Pedersen, J.S., & Mullertz, A. (2007b). Structural development of self nano emulsifying drug delivery systems (SNEDDS) during *in vitro* lipid digestion monitored by small-angle X-ray scattering. *Pharmaceutical Research*, 24(10), 1844–1853.
- FDA (2014). FDA drug safety communication: FDA warns that cancer drug docetaxel may cause symptoms of alcohol intoxication after treatment. Retrieved December 6, 2014, from the U.S. Food and Drug Administration website: <http://www.fda.gov/Drugs/DrugSafety/ucm401752.htm>.
- Fera, D., Schultz, D.C., Hodawadekar, S., Reichman, M., Donover, P.S., Melvin, J., Troutman, S., Kissil, J.L., Hury, D.M., & Marmorstein, R. (2012). Identification and characterization of small molecule antagonists of pRb inactivation by viral oncoproteins. *Chemistry & Biology*, 19(4), 518–528.

- Ferlay, J., Soerjomataram, I., Dikshit, R., Eser, S., Mathers, C., Rebelo, M., Parkin, D.M., Forman, D., & Bray, F. (2015). Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *International Journal of Cancer*, 136(5), E359–386.
- Ferlay, J., Soerjomataram, I., Ervik, M., Dikshit, R., Eser, S., Mathers, C., Rebelo, M., Parkin, D.M., Forman, D., & Bray, F. (2013). GLOBOCAN 2012 v1.0, Cancer Incidence and Mortality Worldwide: IARC Cancer Base No. 11. Retrieved December 11, 2014, from the International Agency for Research on Cancer (IARC) website: [http://globocan.iarc.fr/Pages/fact\\_sheets\\_cancer.aspx](http://globocan.iarc.fr/Pages/fact_sheets_cancer.aspx).
- Ferrari, E., Lazzari, S., Marverti, G., Pignedoli, F., Spagnolo, F., & Saladini, M. (2009). Synthesis, cytotoxic and combined cDDP activity of new stable curcumin derivatives. *Bioorganic & Medicinal Chemistry Letters*, 17(8), 3043–3052.
- Ferreres, F., Pereira, D.M., Valentão, P., Oliveira, J.M., Faria, J., Gaspar, L., Sottomayor, M., & Andrade, P.B. (2010). Simple and reproducible HPLC-DAD-ESI-MS/MS analysis of alkaloids in *Catharanthus roseus* roots. *Journal of Pharmaceutical & Biomedical Analysis*, 51(1), 65–69.
- Ford Versypt, A.N., Pack, D.W., & Braatz, R.D. (2013). Mathematical modeling of drug delivery from autocatalytically degradable PLGA microspheres—a review. *Journal of Controlled Release*, 165(1), 29–37.
- Fulda, S., & Debatin, K.M. (2006). Extrinsic versus intrinsic apoptosis pathways in anticancer chemotherapy. *Oncogene*, 25(34), 4798–4811.
- Fulda, S., & Pervaiz, S. (2010). Apoptosis signaling in cancer stem cells. *The International Journal of Biochemistry & Cell Biology*, 42(1), 31–38.
- Fulda, S., Gorman, A.M., Hori, O., & Samali, A. (2010). Cellular stress responses: cell survival and cell death. *International Journal of Cell Biology*, 2010. Doi: 10.1155/2010/214074
- Gabizon, A., Bradbury, M., Prabhakar, U., Zamboni, W., Libutti, S., & Grodzinski, P. (2014). Cancer nanomedicines: closing the translational gap. *Lancet*, 384(9961), 2175–2176.
- Gali-Muhtasib, H.U., Abou Kheir, W.G., Kheir, L.A., Darwiche, N., & Crooks, P.A. (2004a). Molecular pathway for thymoquinone-induced cell-cycle arrest and apoptosis in neoplastic keratinocytes. *Anti-Cancer Drugs*, 15(4), 389–399.
- Gali-Muhtasib, H.U., Diab-Assaf, M., Boltze, C., Al-Hmaira, J., Hartig, R., Roessner, A., & Schneider-Stock, R. (2004b). Thymoquinone extracted from black seed triggers apoptotic cell death in human colorectal cancer cells via a p53-dependent mechanism. *International Journal of Oncology*, 25(4), 857–866.
- Gali-Muhtasib, H.U., Roessner, A., & Schneider-Stock, R. (2006). Thymoquinone: a promising anti-cancer drug from natural sources. *International Journal of Biochemistry & Cell Biology*, 38(8), 1249–1253.

- Ganea, G.M., Fakayode, S.O., Losso, J.N., van Nostrum, C.F., Sabliov, C.M., & Warner, I.M. (2010). Delivery of phytochemical thymoquinone using molecular micelle modified poly(D, L lactide-co-glycolide) (PLGA) nanoparticles. *Nanotechnology*, 21(28). Doi: 10.1088/0957-4484/21/28/ 285104.
- García-Tuñón, I., Ricote, M., Ruiz, A., Fraile, B., Paniagua, R., & Royuela, M. (2006). Cell cycle control related proteins (p53, p21, and Rb) and transforming growth factor beta (TGFbeta) in benign and carcinomatous (*in situ* and infiltrating) human breast: implications in malignant transformations. *Cancer Investigation*, 24(2), 119–125.
- Garcini, L.M., Galvan, T., & Barnack-Tavlaris, J.L. (2012). The study of human papillomavirus (HPV) vaccine uptake from a parental perspective: A systematic review of observational studies in the United States. *Vaccine*, 30(31), 4588–4595.
- Garland, S.M., Hernandez-Avila, M., Wheeler, C.M., Perez, G., Harper, D.M., Leodolter, S., Tang, G.W., Ferris, D.G., Steben, M., Bryan, J., Taddeo, F.J., Railkar, R., Esser, M.T., Sing, H.L., Nelson, M., Boslego, J., Sattler, C., Barr, E., & Koutsky, L.A. (2007). Quadrivalent vaccine against human papillomavirus to prevent anogenital diseases. *New England Journal of Medicine*, 356(19), 1928–1943.
- Garrett, D. A., Failla, M. L., & Sarama, R. J. (1999). Development of an *in vitro* digestion method to assess carotenoid bioavailability from meals. *Journal of Agricultural & Food Chemistry*, 47(10), 4301–4309.
- Gartel, A.L., & Radhakrishnan, S.K. (2005). Lost in transcription: p21 repression, mechanisms, and consequences. *Cancer Research*, 65(10), 3980–3985.
- Garud, A., Singh, D., & Garud, N. (2012). Solid lipid nanoparticles (SLN): Method, characterization and applications. *International Current Pharmaceutical Journal*, 1(11): 384–393.
- Gauvin-Bialecki, A., & Marodon, C. (2008). Essential oil of *Ayapana triplinervis* from Reunion Island: A good natural source of thymohydroquinone dimethyl ether. *Biochemical Systematics & Ecology*, 36(11), 853–858.
- Ghosh, P., Han, G., De, M., Kim, C.K., & Rotello, V.M. (2008). Gold nanoparticles in delivery applications. *Advanced Drug Delivery Reviews*, 60(11), 1307–1315.
- Giménez-Cassina, A., & Danial, N.N. (2015). Regulation of mitochondrial nutrient and energy metabolism by BCL-2 family proteins. *Trends in Endocrinology & Metabolism*, 26(4), 165–175.
- Ginger, V.A.T., & Yang, C.C. (2011). Functional anatomy of the female sex organs. In J.P. Mulhall, L. Incrocci, I. Goldstein, & R. Rosen (Eds.), *Cancer and Sexual Health* (pp. 13–23). New York, NY: Humana Press.



- Giordano, A., & Galderisi, U. (2010). Short introduction to the cell cycle. In A. Giordano, U. Galderisi (Eds.), *Cell Cycle Regulation and Differentiation in Cardiovascular and Neural Systems* (pp. 3–14). New York: Springer New York.
- Girardini, J.E., Marotta, C., & Del Sal, G. (2014). Disarming mutant p53 oncogenic function. *Pharmacological Research*, 79, 75–87.
- Gmeiner, W.H., Jennings-Gee, J., Stuart, C.H., & Pardee, T.S. (2015). Thymineless death in F10-treated AML cells occurs via lipid raft depletion and Fas/FasL colocalization in the plasma membrane with activation of the extrinsic apoptotic pathway. *Leukemia Research*, 39(2), 229–235.
- Gokulakrishnan, R., Ravikumar, S., & Raj, J.A. (2012). *In vitro* antibacterial potential of metal oxide nanoparticles against antibiotic resistant bacterial pathogens. *Asian Pacific Journal of Tropical Disease*, 2(5), 411–413.
- Grassi, M., Lamberti, G., Cascone, S., & Grassi, G. (2011). Mathematical modeling of simultaneous drug release and *in vivo* absorption. *International Journal of Pharmaceutics*, 418(1), 130–141.
- Greenlee, H. (2012). Natural products for cancer prevention. *Seminars in Oncology Nursing*, 28(1), 29–44.
- Guo, H.X., & Shi, Y.P. (2009). A novel zein-based dry coating tablet design for zero-order release. *International Journal of Pharmaceutics*, 370(1–2), 81–86.
- Gupta, S., Kesarla, R., & Omri, A. (2013). Formulation strategies to improve the bioavailability of poorly absorbed drugs with special emphasis on self-emulsifying systems. *ISRN Pharmaceutics*, 2013. Doi: 10.1155/2013/848043.
- Gurung, R.L., Lim, S.N., Khaw, A.K., Soon, J.F., Shenoy, K., Mohamed Ali, S, Jayapal, M., Sethu, S., Baskar, R., & Hande, M.P. (2010). Thymoquinone induces telomere shortening, DNA damage and apoptosis in human glioblastoma cells. *PLoS One*, 5(8). Doi:10.1371/journal.pone.0012124.
- Han, Y.H., & Park, W.H. (2009). Growth inhibition in antimycin A treated-lung cancer Calu-6 cells via inducing a G1 phase arrest and apoptosis. *Lung Cancer*, 65(2), 150–160.
- Han, Y.K., Lee, J.H., Park, G.Y., Chun, S.H., Han, J.Y., Kim, S.D., Lee, J., Lee, C.W., Yang, K., & Lee, C.G. (2013). A possible usage of a CDK4 inhibitor for breast cancer stem cell-targeted therapy. *Biochemical & Biophysical Research Communications*, 430(4), 1329–1333.
- Hanahan, D., & Weinberg, R.A. (2000). The hallmarks of cancer. *Cell*, 100(1), 57–70.
- Hanahan, D., & Weinberg, R.A. (2011). Hallmarks of cancer: the next generation. *Cell*, 144(5), 646–674.
- Handayani, T., Sakinah, S., Nallappan, M., & Pihie, A.H. (2007). Regulation of p53-, Bcl-2- and caspase-dependent signaling pathway in xanthorrhizol-induced apoptosis of HepG2 hepatoma cells. *Anticancer Research*, 27(2), 965–971.

- Harper, D.M. (2008). Impact of vaccination with Cervarix (trade mark) on subsequent HPV-16/18 infection and cervical disease in women 15-25 years of age. *Gynecologic Oncology*, 110(3 Suppl 1), S11–17.
- Harper, J.V., & Brooks, G. (2005). The mammalian cell cycle. In T. Humphrey, & G. Brooks (Eds), *Cell cycle control* (pp. 113–153). Totowa: Humana Press.
- Harzallah, H.J., Kouidhim, B., Flamini, G., Bakhrouf, A., & Mahjoub, T. (2011). Chemical composition, antimicrobial potential against cariogenic bacteria and cytotoxic activity of Tunisian *Nigella sativa* essential oil and thymoquinone. *Food Chemistry*, 129(4), 1469–1474.
- Hassan, M.I., Mabrouk, G.M., Shehata, H.H., & Aboelhussein, M.M. (2012). Antineoplastic effects of bee honey and *Nigella sativa* on hepatocellular carcinoma cells. *Integrative Cancer Therapies*, 11(4), 354–363.
- Hayat, K., Asim, M.B., Nawaz, M., Li, M., Zhang, L., & Sun, N. (2011). Ameliorative effect of thymoquinone on ovalbumin-induced allergic conjunctivitis in Balb/c mice. *Current Eye Research*, 36(7), 591–598.
- Henken, F.E., Oosterhuis, K., Öhlschläger, P., Bosch, L., Hooijberg, E., Haanen, J.B., & Steenbergen, R.D. (2012). Preclinical safety evaluation of DNA vaccines encoding modified HPV16 E6 and E7. *Vaccine*, 30(28), 4259–4266.
- Hernandez-Flores, G., Ortiz-Lazareno, P.C., Lerma-Diaz, J.M., Dominguez-Rodriguez, J.R., Jave-Suarez, L.F., Aguilar-Lemarroy Adel, C., de Celis-Carrillo, R., del Toro-Arreola, S., Castellanos-Esparza, Y.C., & Bravo-Cuellar, A. (2011). Pentoxifylline sensitizes human cervical tumor cells to cisplatin-induced apoptosis by suppressing NF-kappa B and decreased cell senescence. *BMC Cancer*, 11(483). Doi:10.1186/1471-2407-11-483.
- Herrup, K., & Yang, Y. (2007). Cell cycle regulation in the postmitotic neuron: oxymoron or new biology? *Nature Reviews Neuroscience*, 8(5), 368–378.
- Hill, M.M., Adrain, C., Duriez, P.J., Creagh, E.M., & Martin, S.J. (2004). Analysis of the composition, assembly kinetics and activity of native Apaf-1 apoptosomes. *EMBO Journal*, 23(10), 2134–2145.
- Hoff, P.M., & Machado, K.K. (2012). Role of angiogenesis in the pathogenesis of cancer. *Cancer Treatment Reviews*, 38(7), 825–833.
- Honary, S., & Zahir, F. (2013). Effect of zeta potential on the properties of nano-drug delivery systems-A review (part 2). *Tropical Journal of Pharmaceutical Research*, 12(2), 265–273.
- Hong, S.S., Kim, S.H., & Lim, S.J. (2015). Effects of triglycerides on the hydrophobic drug loading capacity of saturated phosphatidylcholine-based liposomes. *International Journal of Pharmaceutics*, 483(1-2), 142–150.

- Hosseinzadeh, H., Parvardeh, S., Nassiri-Asl, M., & Mansouri, M.T. (2005). Intracerebroventricular administration of thymoquinone, the major constituent of *Nigella sativa* seeds, suppresses epileptic seizures in rats. *Medical Science Monitor*, 11(4), 106–110.
- Houtgraaf, J.H., Versmissen, J., & van der Giessen, W.J. (2006). A concise review of DNA damage checkpoints and repair in mammalian cells. *Cardiovascular Revascularization Medicine*, 7(3), 165–172.
- How, C.W., Rasedee, A., & Abbasalipourkabar R. (2011). Physicochemical properties of nanostructured lipid carriers as colloidal carrier system stabilized with polysorbate 20 and polysorbate 80. *African Journal of Biotechnology*, 10(9), 1684–1689.
- How, C.W., Rasedee, A., Manickam, S., & Rosli, R. (2013). Tamoxifen-loaded nanostructured lipid carrier as a drug delivery system: characterization, stability assessment and cytotoxicity. *Colloids & Surfaces B: Biointerfaces*, 112, 393–399.
- Huang, X., Han, Y., Wang, Y., Cao, M., & Wang, Y. (2008). Aggregation properties of cationic gemini surfactants with dihydroxyethylamino headgroups in aqueous solution. *Colloids and Surfaces A: Physicochemical & Engineering Aspects*, 325(1–2), 26–32.
- Hubatsch, I., Ragnarsson, E. G. E., & Artursson, P. (2007). Determination of drug permeability and prediction of drug absorption in Caco-2 monolayers. *Nature Protocols*, 2(9), 2111–2119.
- Iddamaldeniya, S.S., Thabrew, M.I., Wickramasinghe, S.M., Ratnatunge, N., & Thammitiyagodage, M.G. (2006). A long-term investigation of the anti-hepatocarcinogenic potential of an indigenous medicine comprised of *Nigella sativa*, *Hemidesmus indicus* and *Smilax glabra*. *Journal of Carcinogenesis*, 5(11). Doi:10.1186/1477-3163-5-11.
- Inci, M., Davarci, M., Inci, M., Motor, S., Yalcinkaya, F.R., Nacar, E., Aydin, M., Sefil, N.K., & Zararsiz, I. (2013). Anti-inflammatory and antioxidant activity of thymoquinone in a rat model of acute bacterial prostatitis. *Human & Experimental Toxicology*, 32(4), 354–361.
- Intemann, K., & de Melo-Martín, I. (2010). Social values and scientific evidence: the case of the HPV vaccines. *Biology & Philosophy*, 25(2), 203–213.
- Irigaray, P., & Belpomme, D. (2010). Basic properties and molecular mechanisms of exogenous chemical carcinogens. *Carcinogenesis*, 31(2), 135–148.
- Jafri, S.H., Glass, J., Shi, R., Zhang, S., Prince, M., & Kleiner-Hancock, H. (2010). Thymoquinone and cisplatin as a therapeutic combination in lung cancer: *In vitro* and *in vivo*. *Journal of Experimental & Clinical Cancer Research*, 29(87). Doi: 10.1186/1756-9966-29-87.

- Jaggi, A.S., & Singh, N. (2012). Mechanisms in cancer-chemotherapeutic drugs-induced peripheral neuropathy. *Toxicology*, 291(1-3), 1–9.
- Jain, M., Kasetty, S., Khan, S., & Desai, A. (2014). An insight to apoptosis. *Journal of Research and Practice in Dentistry*, 2014. Doi: 10.5171/2014.372284.
- Jain, P.K., El-Sayed, I.H., & El-Sayed, M.A. (2007). Au nanoparticles target cancer. *Nano Today*, 2(1), 18–29.
- Jakopec, S., Dubravcic, K., Polanc, S., Kosmrlj, J., & Osmak, M. (2006). Diazene JK-279 induces apoptosis-like cell death in human cervical carcinoma cells. *Toxicology in Vitro*, 20(2), 217–226.
- Jamal, J.A., Abd. Ghafar, Z., & Husain, K. (2011). Medicinal plants used for postnatal care in Malay traditional medicine in the Peninsular Malaysia. *Pharmacognosy Journal*, 3(24), 15–24.
- Jawahar, N., & Meyyanathan, S.N. (2012). Polymeric nanoparticles for drug delivery and targeting: A comprehensive review. *International Journal of Health & Allied Sciences*, 1(4), 217–223.
- Jemal, A., Bray, F., Center, M.M., Ferlay, J., Ward, E., & Forman, D. (2011), Global cancer statistics. *CA: A Cancer Journal for Clinicians*, 61(2), 69–90.
- Jenning, V., Thunemann, A.F., & Gohla, S.H. (2000). Characterization of a novel solid lipid nanoparticle carrier system based on binary mixtures of liquid and solid lipids. *International Journal of Pharmaceutics*, 199, 167–177.
- Ji, L. (2011). Cisplatin. In M. Schwab (Ed.), *Encyclopedia of Cancer* (pp. 869–872). Berlin: Springer Berlin Heidelberg.
- Jiang, C.Z., Han, I., & Choung, S. (2012). Decursin mediated protection on Cisplatin-induced nephrotoxicity in SD rats and BDF1 Mice. *Journal of Northeast Agricultural University*, 19(1), 50–56.
- Joerger, A.C., & Fersht, A.R. (2007). Structural biology of the tumor suppressor p53 and cancer-associated mutants. *Advances in Cancer Research*, 97, 1–23.
- Joshi, M., & Patravale, V. (2008). Nanostructured lipid carrier (NLC) based gel of celecoxib. *International Journal of Pharmaceutics*, 346(1-2), 124–132.
- Jost, P.J., Grabow, S., Gray, D., McKenzie, M.D., Nachbur, U., Huang, D.C., Bouillet, P., Thomas, H.E., Borner, C., Silke, J., Strasser, A., & Kaufmann, T. (2009). XIAP discriminates between Type I and Type II FAS-induced apoptosis. *Nature*, 460(7258), 1035–1039.

- Joura, E.A., Leodolter, S., Hernandez-Avila, M., Wheeler, C.M., Perez, G., Koutsky, L.A., Garland, S.M., Harper, D.M., Tang, G.W., Ferris, D.G., Steben, M., Jones, R.W., Bryan, J., Taddeo, F.J., Bautista, O.M., Esser, M.T., Sings, H.L., Nelson, M., Boslego, J.W., Sattler, C., Barr, E., & Paavonen, J. (2007). Efficacy of a quadrivalent prophylactic human papillomavirus (types 6, 11, 16, and 18) L1 virus-like-particle vaccine against high-grade vulval and vaginal lesions: a combined analysis of three randomised clinical trials. *Lancet*, 369(9574), 1693–1702.
- Junyapraserta, V.B., & Morakula, B. (2015). Nanocrystals for enhancement of oral bioavailability of poorly water-soluble drugs. *Asian Journal of Pharmaceutical Sciences*, 10(1), 13–23.
- Kakadia, P.G., & Conway, B.R. (2014). Solid lipid nanoparticles: A potential approach for dermal drug delivery. *American Journal of Pharmacological Sciences*, 2(5A), 1–7.
- Kanasty, R., Dorkin, J.R., Vegas, A., & Anderson, D. (2013). Delivery materials for siRNA therapeutics. *Nature Materials*, 12, 967–977.
- Kandouz, M., Alachkar, A., Zhang, L., Dekhil, H., Chehna, F., Yasmeen, A., & Al Moustafa, A.E. (2010). *Teucrium polium* plant extract inhibits cell invasion and motility of human prostate cancer cells via the restoration of the E-cadherin/catenin complex. *Journal of Ethnopharmacology*, 129(3), 410–415.
- Kantari, C., & Walczak, H. (2011). Caspase-8 and Bid: caught in the act between death receptors and mitochondria. *Biochimica et Biophysica Acta*, 1813(4), 558–563.
- Kanter, M., Coskun, O., & Uysal, H. (2006). The antioxidative and antihistaminic effect of *Nigella sativa* and its major constituent, thymoquinone on ethanol-induced gastric mucosal damage. *Archives of Toxicology*, 80(4), 217–224.
- Katanyoo, K., Sanguanrungririkul, S., & Manusirivithaya, S. (2012). Comparison of treatment outcomes between squamous cell carcinoma and adenocarcinoma in locally advanced cervical cancer. *Gynecologic Oncology*, 125(2), 292–296.
- Katteboina, S., Chandrasekhar, P.V.S.R., & Balaji, S. (2009). Approaches for the development of solid self-emulsifying drug delivery systems and dosage forms. *Asian Journal of Pharmaceutical Sciences*, 4(4), 240–253.
- Kawabata, Y., Wada, K., Nakatani, M., Yamada, S., & Onoue, S. (2011). Formulation design for poorly water-soluble drugs based on biopharmaceutics classification system: basic approaches and practical applications. *International Journal of Pharmaceutics*, 420(1), 1–10.
- Kawakami, K., Oda, N., Miyoshi, K., Funaki, T., & Ida, Y. (2006). Solubilization behavior of a poorly soluble drug under combined use of surfactants and cosolvents. *European Journal of Pharmaceutical Sciences*, 28(1–2), 7–14.
- Kelly, C., Jefferies, C., & Cryan, S.A. (2011). Targeted liposomal drug delivery to monocytes and macrophages. *Journal of Drug Delivery*, 2011. Doi: 10.1155/2011/727241.



- Khader, M., Bresgen, N., & Eckl, P.M. (2009). *In vitro* toxicological properties of thymoquinone. *Food & Chemical Toxicology*, 47(1), 129–133.
- Khadka, P., Ro, J., Kim, H., Kim, I., Kim, J.T., Kim, H., Cho, J.M., Yun, G., & Lee, J. (2015). Pharmaceutical particle technologies: An approach to improve drug solubility, dissolution and bioavailability. *Asian Journal of Pharmaceutical Sciences*, 9(6), 304–316.
- Khairuzzaman, A., Ahmed, S.U., Savva, M., & Patel, N.K. (2006) Zero-order release of aspirin, theophylline and atenolol in water from novel methylcellulose glutarate matrix tablets. *International Journal of Pharmaceutics*, 318(1–2), 15–21.
- Khan, K.H., Blanco-Codesido, M., & Molife, L.R. (2014). Cancer therapeutics: Targeting the apoptotic pathway. *Critical Reviews in Oncology/Hematology*, 90(3), 200–219.
- Khan, N., & Sultana, S. (2005). Inhibition of two stage renal carcinogenesis, oxidative damage and hyperproliferative response by *Nigella sativa*. *European Journal of Cancer Prevention*, 14(2), 159–168.
- Khan, N., Sharma, S., & Sultana, S. (2003). *Nigella sativa* (black cumin) ameliorates potassium bromate-induced early events of carcinogenesis: diminution of oxidative stress. *Human & Experimental Toxicology*, 22(4), 193–203.
- Khattab, M.M., & Nagi, M.N. (2007). Thymoquinone supplementation attenuates hypertension and renal damage in nitric oxide deficient hypertensive rats. *Phytotherapy Research*, 21(5), 410–414.
- Khurana, S., Jain, N.K., & Bedi, P.M. (2013). Development and characterization of a novel controlled release drug delivery system based on nanostructured lipid carriers gel for meloxicam. *Life Sciences*, 93(21), 763–772.
- Kim, H., Kim, K., Choi, J., Heo, K., Baek, H.J., Roeder, R.G., & An, W. (2012). p53 requires an intact C-terminal domain for DNA binding and transactivation. *Journal of Molecular Biology*, 415(5), 843–854.
- Kim, H., Tu, H.C., Ren, D., Takeuchi, O., Jeffers, J.R., Zambetti, G.P., Hsieh, J.J., & Cheng, E.H. (2009). Stepwise activation of BAX and BAK by tBID, BIM, and PUMA initiates mitochondrial apoptosis. *Molecular Cell*, 36(3), 487–99.
- Kitdamrongtham, W., Manosroi, A., Akazawa, H., Gidado, A., Stienrut, P., Manosroi, W., Lohcharoenkal, W., Akihisa, T., & Manosroi, J. (2013). Potent anti-cervical cancer activity: Synergistic effects of Thai medicinal plants in recipe N040 selected from the MANOSROI III database. *Journal of Ethnopharmacology*, 149(1), 288–296.
- Klein, T.J., & Glazer, P.M. (2010). The tumor microenvironment and DNA repair. *Seminars in Radiation Oncology*, 20(4), 282–287.
- Knowlton, C., & Mackay, M.K. (2013). Papanicolaou Smear. In L.W. Brady, & T.E. Yaeger (Eds.), *Encyclopedia of Radiation Oncology* (pp. 933–939). Berlin: Springer Berlin Heidelberg.

- Koka, P.S., Mondal, D., Schultz, M., Abdel-Mageed, A.B., & Agrawal, K.C. (2010). Studies on molecular mechanisms of growth inhibitory effects of thymoquinone against prostate cancer cells: role of reactive oxygen species. *Experimental Biology & Medicine*, 235(6),751–760.
- Koornstra, J.J., de Jong, S., Hollema, H., de Vries, E.G., & Kleibeuker, J.H. (2003). Changes in apoptosis during the development of colorectal cancer: a systematic review of the literature. *Critical Reviews in Oncology/Hematology*, 45(1), 37–53.
- Korany, N.S., & Ezzat, B.A. (2011). Prophylactic effect of green tea and *Nigella sativa* extracts against fenitrothion-induced toxicity in rat parotid gland. *Archives of Oral Biology*, 56(11):1339–1346.
- Kovacevic, A., Savic, S., Vuleta, G., Müller, R.H., & Keck, C.M. (2011). Polyhydroxy surfactants for the formulation of lipid nanoparticles (SLN and NLC): effects on size, physical stability and particle matrix structure. *International Journal of Pharmaceutics*, 406(1-2), 163–172.
- Koyuncuoglu, M., Okyay, E., Saatli, B., Olgan, S., Akin, M., & Saygili, U. (2012). Tumor budding and E-Cadherin expression in endometrial carcinoma: are they prognostic factors in endometrial cancer? *Gynecologic Oncology*, 125(1), 208–213.
- Krishnaiah, Y.S.R. (2010). Pharmaceutical technologies for enhancing oral bioavailability of poorly soluble drugs. *Journal of Bioequivalence & Bioavailability*, 2(2), 28–36.
- Kroemer, G., Galluzzi, L., Vandenabeele, P., Abrams, J., Alnemri, E.S., Baehrecke, E.H., Blagosklonny, M.V., El-Deiry, W.S., Golstein, P., Green, D.R., Hengartner, M., Knight, R.A., Kumar, S., Lipton, S.A., Malorni, W., Nuñez, G., Peter, M.E., Tschopp, J., Yuan, J., Piacentini, M., Zhivotovsky, B., & Melino, G. (2009). Classification of cell death: recommendations of the Nomenclature Committee on Cell Death 2009. *Cell Death & Differentiation*, 16(1), 3–11.
- Kumara, S.S., & Huat, B.T. (2001). Extraction, isolation and characterisation of antitumor principle, alpha-hederin, from the seeds of *Nigella sativa*. *Planta Medica*, 67(1), 29–32.
- Kumari, A., Yadav, S.K., & Yadav, S.C. (2010). Biodegradable polymeric nanoparticles based drug delivery systems. *Colloids & Surfaces B: Biointerfaces*, 75(1), 1–18.
- Kundu, J., Chun, K., Aruoma, O.I., & Kundu, J.K. (2014). Mechanistic perspectives on cancer chemoprevention/chemotherapeutic effects of thymoquinone. *Mutation Research-fundamental & Molecular Mechanisms of Mutagenesis*, 768, 22–34.
- Kushwaha, A.K., Vuddanda, P.R., Karunanidhi, P., Singh, S.K., & Singh, S. (2013). Development and evaluation of solid lipid nanoparticles of raloxifene hydrochloride for enhanced bioavailability. *BioMed Research International*, 2013. Doi: 10.1155/2013/584549.



- Kuwana, T., Mackey, M.R., Perkins, G., Ellisman, M.H., Latterich, M., Schneiter, R., Green, D.R., & Newmeyer, D.D. (2002). Bid, Bax, and lipids cooperate to form supramolecular openings in the outer mitochondrial membrane. *Cell*, *111*(3), 331–342.
- Kvitek, L., Panacek, A., Soukupova, J., Kolar, M., Vecerova, R., Pucek, R., Holecova, M., & Zboril, R. (2008). Effect of surfactants and polymers on stability and antibacterial activity of silver nanoparticles (NPs). *Journal of Physical Chemistry C*, *112*(15), 5825–5834.
- Kyrgiou, M., & Shafi, M.I. (2014). Colposcopy and cervical intraepithelial neoplasia. *Obstetrics, Gynaecology & Reproductive Medicine*, *24*(7), 204–214.
- Lai, C.S., Mas, R.H., Nair, N.K., Majid, M.I., Mansor, S.M., & Navaratnam, V. (2008). *Typhonium flagelliforme* inhibits cancer cell growth *in vitro* and induces apoptosis: an evaluation by the bioactivity guided approach. *Journal of Ethnopharmacology*, *118*(1), 14–20.
- Lang, M., Borgmann, M., Oberhuber, G., Evstatiev, R., Jimenez, K., Dammann, K.W., Jambrich, M., Khare, V., Campregher, C., Ristl, R., & Gasche, C. (2013). Thymoquinone attenuates tumor growth in Apc<sup>Min</sup> mice by interference with Wnt-signaling. *Molecular Cancer*, *12*(41). Doi:10.1186/1476-4598-12-41.
- Lau, C., Mooiman, K.D., Maas-Bakker, R.F., Beijnen, J.H., Schellens, J.H., & Meijerman, I. (2013). Effect of Chinese herbs on CYP3A4 activity and expression *in vitro*. *Journal of Ethnopharmacology*, *149*(2), 543–549.
- Lee, G., Lee, J.H., Ham, K.K., Lee, H., Kim, H., Lee, H., Hong, M., Shin, M., & Bae, H. (2012). Cisplatin induced nephrotoxicity is inhibited by *Taxilli Ramulus*. *Molecular & Cellular Toxicology*, *8*(3), 311–315.
- Lee, J.C., Lee, K.Y., Son, Y.O., Choi, K.C., Kim, J., Truong, T.T., & Jang, Y.S. (2005). Plant-originated glycoprotein, G-120, inhibits the growth of MCF-7 cells and induces their apoptosis. *Food & Chemical Toxicology*, *43*(6), 961–968.
- Lee, J.E., Kang, J.S., Shin, I.C., Lee, S.J., Hyun, D.H., Lee, K.S., & Koh, H.C. (2011a). Fluazinam-induced apoptosis of SH-SY5Y cells is mediated by p53 and Bcl-2 family proteins. *NeuroToxicology*, *32*(6), 702–710.
- Lee, J.H., & Yeo, Y. (2015). Controlled drug release from pharmaceutical nanocarriers. *Chemical Engineering Science*, *125*, 75–84.
- Lee, J.J., Huang, J., England, C.G., McNally, L.R., & Frieboes, H.B. (2013). Predictive modeling of *in vivo* response to gemcitabine in pancreatic cancer. *PLoS Computational Biology*, *9*(9). Doi: 10.1371/journal.pcbi.1003231.
- Lee, K., Lee, A.Y., Kwon, Y.K., & Kwon, H. (2011b). Suppression of HPV E6 and E7 expression by BAF53 depletion in cervical cancer cells. *Biochemical & Biophysical Research Communications*, *412*(2), 328–333.

- Lei, L., Liu, X., Shen, Y.Y., Liu, J.Y., Tang, M.F., Wang, Z.M., Guo, S.R., & Cheng, L. (2011). Zero-order release of 5-fluorouracil from PCL-based films featuring trilayered structures for stent application. *European Journal of Pharmaceutics & Biopharmaceutics*, 78(1), 49–57.
- Lei, X., Liu, M., Yang, Z., Ji, M., Guo, X., & Dong, W. (2012). Thymoquinone prevents and ameliorates dextran sulfate sodium-induced colitis in mice. *Digestive Diseases & Sciences*, 57(9), 2296–2303.
- Leone, L.M., & Roberts, S.C. (2013). Accessing anti-cancer natural products by plant cell culture. In F.E. Koehn (Ed.), *Natural Products and Cancer Drug Discovery* (pp. 193–211). New York, NY: Springer New York.
- Levy, M.Y., & Benita, S. (1990). Drug release from submicronized o/w emulsion: a new *in vitro* kinetic evaluation model. *International Journal of Pharmaceutic*, 66(1–3), 29–37.
- Li, H.L., Chen, D.D., Li, X.H., Zhang, H.W., Lu, Y.Q., Ye, C.L., & Ren, X.D. (2002). Changes of NF- $\kappa$ B, p53, Bcl-2 and caspase in apoptosis induced by JTE-522 in human gastric adenocarcinoma cell line AGS cells: role of reactive oxygen species. *World Journal of Gastroenterology*, 8(3), 431–435.
- Li, J., & Yuan, J. (2008). Caspases in apoptosis and beyond. *Oncogene*, 27, 6194–6206.
- Li, S., & Zhang, B. (2013). Traditional Chinese medicine network pharmacology: Theory, methodology and application. *Chinese Journal of Natural Medicines*, 11(2), 0110–0120.
- Li, Y., Hu, M., & McClements, D.J. (2011). Factors affecting lipase digestibility of emulsified lipids using an *in vitro* digestion model: proposal for a standardised pH-stat method. *Food Chemistry*, 126(2), 498–505.
- Li, Z., Wang, J., Jiang, B., Zhang, X., An, L., & Bao, Y. (2007). Benzobijuglone, a novel cytotoxic compound from *Juglans mandshurica*, induced apoptosis in HeLa cervical cancer cells. *Phytomedicine*, 14(12), 846–852.
- Liao, T.T., Shi, Y.L., Jia, J.W., Jia, R.W., & Wang, L. (2010). Sensitivity of morphological change of Vero cells exposed to lipophilic compounds and its mechanism. *Journal of Hazardous Materials*, 179(1–3), 1055–1064.
- Lim, T.K. (2013). *Nigella sativa*. In T.K. Lim (Ed.), *Edible Medicinal And Non-Medicinal Plants* (pp. 506–567). Netherlands: Springer Netherlands.
- Lin, K., Doolan, K., Hung, C.F., & Wu, T.C. (2010a). Perspectives for preventive and therapeutic HPV vaccines. *Journal of the Formosan Medical Association*, 109(1), 4–24.
- Lin, K., Roosinovich, E., Ma, B., Hung, C.F., & Wu, T.C. (2010b). Therapeutic HPV DNA vaccines. *Immunologic Research*, 47(1–3), 86–112.

- Lin, X., Liu, M., Hu, C., & Liao, D.J. (2010c). Targeting cellular proapoptotic molecules for developing anticancer agents from marine sources. *Current Drug Targets*, *11*(6), 708–715.
- Lin, Y.K., Huang, Z.R., Zhuo, R.Z., & Fang, J.Y. (2010d). Combination of calcipotriol and methotrexate in nanostructured lipid carriers for topical delivery. *International Journal of Nanomedicine*, *5*, 117–128.
- Lindsay, J., Esposti, M.D., & Gilmore, A.P. (2011). Bcl-2 proteins and mitochondria-specificity in membrane targeting for death. *Biochimica et Biophysica Acta*, *1813*(4), 532–539.
- Liu, J., Yang, L., Zhang, J., Zhang, J., Chen, Y., Li, K., Li, Y., Li, Y., Yao, L., & Guo, G. (2012). Knock-down of NDRG2 sensitizes cervical cancer HeLa cells to cisplatin through suppressing Bcl-2 expression. *BMC Cancer*, *12*(370). Doi:10.1186/1471-2407-12-370.
- Liu, X., & Zhu, X.Z. (1999). Roles of p53, c-Myc, Bcl-2, Bax and caspases in glutamate-induced neuronal apoptosis and the possible neuroprotective mechanism of basic fibroblast growth factor. *Molecular Brain Research*, *71*(2), 210–216.
- Llambi, F., & Green, D.R. (2011). Apoptosis and oncogenesis: give and take in the BCL-2 family. *Current Opinion in Genetics & Development*, *21*(1), 12–20.
- Lo, Y.L., & Wang, W. (2013). Formononetin potentiates epirubicin-induced apoptosis via ROS production in HeLa cells *in vitro*. *Chemico-Biological Interactions*, *205*(3), 188–197.
- Lockshin, R.A., & Zakeri, Z. (2007). Cell death in health and disease. *Journal of Cellular & Molecular Medicine*, *11*, 1214–1224.
- Long, M., & Chen, Y. (2009). Dissolution testing of solid products. In Y. Qiu, Y. Chen, G.G.Z. Zhang, L. Liu, & W.R. Porter, (Ed.) *Developing Solid Oral Dosage Forms: Pharmaceutical Theory And Practice* (pp. 319–340). USA: Elsevier Inc.
- Loo, C.H., Basri, M., Ismail, R., Lau, H., Tejo, B., Kanthimathi, M., Hassan, H., & Choo, Y. (2013). Effect of compositions in nanostructured lipid carriers (NLC) on skin hydration and occlusion. *International Journal of Nanomedicine*, *8*, 13–22.
- Lozano-Kühne, J. (2013). Cancer. In W. Dubitzky, O. Wolkenhauer, K.H. & Cho, H. Yokota (Eds.), *Encyclopedia of Systems Biology* (p. 172). Berlin: Springer Berlin Heidelberg.
- Luan, J., Zhang, D., Hao, L., Li, C., Qi, L., Guo, H., Liu, X., & Zhang, Q. (2013). Design and characterization of Amoitone B-loaded nanostructured lipid carriers for controlled drug release. *Drug Delivery*, *20*(8), 324–330.
- Luan, J., Zheng, F., Yang, X., Yu, A., & Zhai, G. (2015). Nanostructured lipid carriers for oral delivery of baicalin: *In vitro* and *in vivo* evaluation. *Colloids & Surfaces A*, *466*, 154–159.

- Lukas, B., Schmiderer, C., Franz, C., & Novak, J. (2009). Composition of essential oil compounds from different Syrian populations of *Origanum syriacum* L. (Lamiaceae). *Journal of Agricultural & Food Chemistry*, 57(4), 1362–1365.
- Lynch, B., & Friedenreich, C.M. (2012). Cancer, Prevention. In F.C. Mooren (Ed.), *Encyclopedia of Exercise Medicine in Health and Disease* (pp. 148–150). Berlin: Springer Berlin Heidelberg.
- Mabrouk, G.M., Moselhy, S.S., Zohny, S.F., Ali, E.M., Helal, T.E., Amin, A.A., & Khalifa, A.A. (2002). Inhibition of methylnitrosourea (MNU) induced oxidative stress and carcinogenesis by orally administered bee honey and *Nigella* grains in *Sprague dawley* rats. *Journal of Experimental & Clinical Cancer Research*, 21(3), 341–346.
- MacDonald, P.M. (2011). Cervix. In S. Goldstein, & J.A. Naglieri (Eds.), *Encyclopedia of Child Behavior and Development* (p. 329). US: Springer US.
- Mackay, M.K., & Knowlton, C.A. (2013). Uterine cervix. In L.W. Brady, & T.E. Yaeger (Eds.), *Encyclopedia of Radiation Oncology* (pp. 933–939). Berlin: Springer Berlin Heidelberg.
- Magaldi, T.G., Almstead, L.L., Bellone, S., Prevatt, E.G., Santin, A.D., & DiMaio, D. (2012). Primary human cervical carcinoma cells require human papillomavirus E6 and E7 expression for ongoing proliferation. *Virology*, 422(1), 114–124.
- Magdy, M.A., Hanan, E., & Nabila, E. (2012). Thymoquinone: Novel gastroprotective mechanisms. *European Journal of Pharmacology*, 697(1–3), 126–131.
- Mahmoud, S.S., & Torchilin, V.P. (2012). Hormetic/cytotoxic effects of *Nigella sativa* seed alcoholic and aqueous extracts on MCF-7 breast cancer cells alone or in combination with doxorubicin. *Cell Biochemistry & Biophysics*, 25(7), 1392–1398.
- Mahmoudvand, H., Sepahvand, A., Jahanbakhsh, S., Ezatpour, B., & Ayatollahi Mousavi, S.A. (2014). Evaluation of antifungal activities of the essential oil and various extracts of *Nigella sativa* and its main component, thymoquinone against pathogenic dermatophyte strains. *Journal of Medical Mycology*, 24(4), e155–e161.
- Majdalawieh, A.F., Hmaidan, R., & Carr, R.I. (2010). *Nigella sativa* modulates splenocyte proliferation, Th1/Th2 cytokine profile, macrophage function and NK anti-tumor activity. *Journal of Ethnopharmacology*, 131(2), 268–275.
- Malam, Y., Loizidou, M., & Seifalian, A.M. (2009). Liposomes and nanoparticles: nanosized vehicles for drug delivery in cancer. *Trends in Pharmacological Sciences*, 30(11), 592–599.
- Manhart, L.E., Burgess-Hull, A.J., Fleming, C.B., Bailey, J.A., Haggerty, K.P., & Catalano, R.F. (2011). HPV vaccination among a community sample of young adult women. *Vaccine*, 29(32), 5238–5244.

- Manosroi, J., Sainakham, M., Manosroi, W., & Manosroi, A. (2012). Anti-proliferative and apoptosis induction activities of extracts from Thai medicinal plant recipes selected from MANOSROI II database. *Journal of Ethnopharmacology*, *141*(1), 451–459.
- Mansour, M.A., Nagi, M.N., El-Khatib, A.S., & Al-Bekairi, A.M. (2002). Effects of thymoquinone on antioxidant enzyme activities, lipid peroxidation and DT-diaphorase in different tissues of mice: a possible mechanism of action. *Cell Biochemistry and Function*, *20*(2), 143–151.
- Marczak, A., Denel-Bobrowska, M., Rogalska, A., Łukawska, M., & Oszczapowicz, I. (2015). Cytotoxicity and induction of apoptosis by formamidineoxorubicins in comparison to doxorubicin in human ovarian adenocarcinoma cells. *Environmental Toxicology & Pharmacology*, *39*(1), 369–383.
- Marín-García, J. (2011). Signaling in the heart. In J. Marín-García (Ed.), *Cell-Cycle Signaling, Epigenetics, and Nuclear Function* (pp. 21–30). US: Springer US.
- Marnett, L.J. (2000). Oxyradicals and DNA damage. *Carcinogenesis*, *21*(3), 361–370.
- Marques, E.F., & Silva, B.F.B. (2013). Surfactants, phase behavior. In T. Tadros, (Ed.) *Encyclopedia of Colloid and Interface Science* (pp. 1290–1333). Berlin: Springer-Verlag Berlin Heidelberg.
- Marsik, P., Kokoska, L., Landa, P., Nepovim, A., Soudek, P., & Vanek, T. (2005). *In vitro* inhibitory effects of thymol and quinones of *Nigella sativa* seeds on cyclooxygenase-1- and -2-catalyzed prostaglandin E2 biosyntheses. *Planta Medica*, *71*(8), 739–742.
- Martin, L., Coffey, M., Lawler, M., Hollywood, D., & Marignol, L. (2010). DNA mismatch repair and the transition to hormone independence in breast and prostate cancer. *Cancer Letters*, *291*(2), 142–149.
- Martínez-Mesa, J., Werutsky, G., Campani, R.B., Wehrmeister, F.C., & Barrios, C.H. (2013). Inequalities in Pap smear screening for cervical cancer in Brazil. *Preventive Medicine*, *57*(4), 366–371.
- Matthaus, B., & Ozcan, M.M. (2011). Fatty acids, tocopherol, and sterol contents of some *Nigella* species seed oil. *Czech Journal of Food Sciences*, *29*(2), 145–150.
- Maznah, I. (1999). The use of Caco-2 cells as an *in vitro* method to study bioavailability of iron. *Malaysian Journal of Nutrition*, *5*(1), 31–45.
- McClements, D.J. (2012). Crystals and crystallization in oil-in-water emulsions: implications for emulsion-based delivery systems. *Advances in Colloid & Interface Science*, *174*, 1–30.
- McDonald, J. (2012). Cancer screening. In S. Loue, & M. Sajatovic (Ed.), *Encyclopedia of Immigrant Health* (pp. 358–361). New York: Springer New York.



- McIlwain, D.R., Berger, T., Mak, T.W. (2013). Caspase functions in cell death and disease. *Cold Spring Harbor Perspectives in Biology*, 5(4). Doi: 10.1101/cshperspect.a008656.
- Medina-Arana, V., Delgado, L., Bravo, A., Martín, J., Fernández-Peralta, A.M., & González-Aguilera, J.J. (2012). Tumor spectrum in Lynch syndrome, DNA mismatch repair system and endogenous carcinogens. *Journal of Surgical Oncology*, 106(1), 10–16.
- Mendes, A.I., Silva, A.C., Catita, J.A., Cerqueira, F., Gabriel, C., & Lopes, C.M. (2013). Miconazole-loaded nanostructured lipid carriers (NLC) for local delivery to the oral mucosa: improving antifungal activity. *Colloids & Surfaces B: Biointerfaces*, 111, 755–763.
- Miao, R., Wei, J., Lv, M., Cai, Y., Du, Y., Hui, X., & Wang, Q. (2011). Conjugation of substituted ferrocenyl to thiadiazine as apoptosis-inducing agents targeting the Bax/Bcl-2 pathway. *European Journal of Medicinal Chemistry*, 46(10), 5000–5009.
- Mignani, S., El Kazouli, S., Bousmina, M., & Majoral, J.P. (2013). Expand classical drug administration ways by emerging routes using dendrimer drug delivery systems: a concise overview. *Advanced Drug Delivery Reviews*, 65(10), 1316–1330.
- Milrot, E., Jackman, A., Kniazhanski, T., Gonen, P., Flescher, E., & Sherman, L. (2012). Methyl jasmonate reduces the survival of cervical cancer cells and downregulates HPV E6 and E7, and survivin. *Cancer Letters*, 319(1), 3–38.
- Ministry of Health Malaysia. (2013). *Health Facts 2012*. Kuala Lumpur: Health Informatics Centre and Planning Division, Ministry of Health Malaysia.
- Mirza, A., & Mithal, N. (2011). Alcohol intoxication with the new formulation of docetaxel. *Journal of Clinical Oncology*, 23(8), 560–561.
- Mishra, B., Patel, B.B., & Tiwari, S. (2010). Colloidal nanocarriers: a review on formulation technology, types and applications toward targeted drug delivery. *Nanomedicine: Nanotechnology, Biology & Medicine*, 6(1), 9–24.
- Mitsuhashi, T., & Takahashi, T. (2009). Cell cycle. In M.D. Binder, N. Hirokawa, & U. Windhorst (Eds.), *Encyclopedia of Neuroscience* (pp. 588–591). New York: Berlin: Springer Berlin Heidelberg.
- Mjelle, R., Hegre, S.A., Aas, P.A., Slupphaug, G., Drabløs, F., Sætrum, P., & Krokan, H.E. (2015). Cell cycle regulation of human DNA repair and chromatin remodeling genes. *DNA Repair*, 30, 53–67.
- Mognetti, B., Barberis, A., Marino, S., Berta, G., Francia, S., Trotta, F., & Cavalli, R. (2012). *In vitro* enhancement of anticancer activity of paclitaxel by a cremophor free cyclodextrin-based nanosponge formulation. *Journal of Inclusion Phenomena & Macrocyclic Chemistry*, 74(1-4), 201–210.

- Mohamed, A., & Mahfoodh, A.S.M. (2006). Solubilization of naphthalene and pyrene by sodium dodecyl sulfate (SDS) and polyoxyethylenesorbitan monooleate (Tween 80) mixed micelles. *Colloids & Surfaces A: Physicochemical & Engineering Aspects*, 87(1–3), 44–50.
- Montoro, P., Maldini, M., Piacente, S., Macchia, M., & Pizza, C. (2010). Metabolite fingerprinting of *Camptotheca acuminata* and the HPLC-ESI-MS/MS analysis of camptothecin and related alkaloids. *Journal of Pharmaceutical & Biomedical Analysis*, 51(2), 405–415.
- Moraco, A.H., & Kornfeld, H. (2014). Immunity to *Mycobacterium tuberculosis*. *Seminars in Immunology*, 26(6), 497–511.
- Morikawa, T., Ninomiya, K., Xu, F., Okumura, N., Matsuda, H., Muraoka, O., Hayakawa, T., & Yoshikawa, M. (2013). Acylated dolabellane-type diterpenes from *Nigella sativa* seeds with triglyceride metabolism-promoting activity in high glucose-pretreated HepG2 cells. *Phytochemistry Letters*, 6(2), 198–204.
- Morikawa, T., Xu, F., Kashima, Y., Matsuda, H., Ninomiya, K., & Yoshikawa, M. (2004). Novel dolabellane-type diterpene alkaloids with lipid metabolism promoting activities from the seeds of *Nigella sativa*. *Organic Letters*, 6(6), 869–872.
- Mosmann, T. (1983). Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *Journal of Immunological Methods*, 65(1–2), 55–63.
- Mountzios, G., Soultati, A., Pectasides, D., Pectasides, E., Dimopoulos, M.A., & Papadimitriou, C.A. (2013). Developments in the systemic treatment of metastatic cervical cancer. *Cancer Treatment Reviews*, 39(5), 430–443.
- Muchow, M., Maincent, P., & Muller, R.H. (2008). Lipid nanoparticles with a solid matrix (SLN, NLC, LDC) for oral drug delivery. *Drug Development & Industrial Pharmacy*, 34(12), 1394–1405.
- Muller, P.A., & Vousden, K.H. (2013). p53 mutations in cancer. *Nature Cell Biology*, 15(1), 2–8.
- Müller, R.H., Mader, K., & Gohla, S. (2000). Solid lipid nanoparticles (SLN) for controlled drug delivery - A review of the state of the art. *European Journal of Pharmaceutics & Biopharmaceutics*, 50(1), 161–177.
- Müller, R.H., Radtke, M., & Wissing, S.A. (2002). Solid lipid nanoparticles (SLN) and nanostructured lipid carriers (NLC) in cosmetic and dermatological preparations. *Advanced Drug Delivery Reviews*, 54(Suppl 1), S131–S155.
- Müller, R.H., Radtke, M., & Wissing, S.A. (2004). Solid lipid nanoparticles and nanostructured lipid carriers. In H.S. Nalwa, (Ed.) *Encyclopedia of Nanoscience and Nanotechnology* (pp. 43–56). Los Angeles, USA: American Scientific Publishers.



- Müller, R.H., Rühl, D., & Runge, S.A. (1996). Biodegradation of solid lipid nanoparticles as a function of lipase incubation time. *International Journal of Pharmaceutics*, 144(1), 115–121.
- Munagala, R., Kausar, H., Munjal, C., & Gupta, R.C. (2011). Withaferin A induces p53-dependent apoptosis by repression of HPV oncogenes and upregulation of tumor suppressor proteins in human cervical cancer cells. *Carcinogenesis*, 32(11), 1697–1705.
- Munnia, A., Bonassi, S., Verna, A., Quaglia, R., Pelucco, D., Ceppi, M., Neri, M., Buratti, M., Taioli, E., Garte, S., & Peluso, M. (2006). Bronchial malondialdehyde DNA adducts, tobacco smoking, and lung cancer. *Free Radical Biology & Medicine*, 41(9), 1499–1505.
- Murakami, A., Fukushima, C., Yoshidomi, K., Sueoka, K., Nawata, S., Yokoyama, Y., Tsuchida, S., Ismail, E., Al-Mulla, F., & Sugino, N. (2011). Suppression of carbonyl reductase expression enhances malignant behaviour in uterine cervical squamous cell carcinoma: carbonyl reductase predicts prognosis and lymph node metastasis. *Cancer Letter*, 311(1), 77–84.
- Nag, O.K., & Awasthi, V. (2013). Surface engineering of liposomes for stealth behavior. *Pharmaceutics*, 2013(5), 542–569.
- Nagi, M.N., Al-Shabanah, O.A., Hafez, M.M., & Sayed-Ahmed, M.M. (2011). Thymoquinone supplementation attenuates cyclophosphamide-induced cardiotoxicity in rats. *Journal of Biochemical & Molecular Toxicology*, 25(3), 135–142.
- Newman, D.J., & Cragg, G.M. (2012). Natural products as sources of new drugs over the 30 years from 1981 to 2010. *Journal of Natural Products*, 75(3), 311–335.
- Niccolai, L.M., Mehta, N.R., & Hadler, J.L. (2011). Racial/ethnic and poverty disparities in human papillomavirus vaccination completion. *American Journal of Preventive Medicine*, 41(4), 428–433.
- Nigam, M., Singh, N., Ranjan, V., Zaidi, D., Sharma, R., Nigam, D., Gupta, D.K., Sundaram, S., & Balapure, A.K. (2010). Centchroman mediated apoptosis involves cross-talk between extrinsic/intrinsic pathways and oxidative regulation. *Life Sciences*, 87(23-26), 750–758.
- Nijhuis, E.R., Nijman, H.W., Oien, K.A., Bell, A., ten Hoor, K.A., Reesink-Peters, N., Boezen, H.M., Hollema, H., & van der Zee, A.G.J. (2007). Loss of MSH2 protein expression is a risk factor in early stage cervical cancer. *Journal of Clinical Pathology*, 60(7), 824–830.
- Nikoletopoulou, V., Markaki, M., Palikaras, K., & Tavernarakis, N. (2013). Crosstalk between apoptosis, necrosis and autophagy. *Biochimica et Biophysica Acta (BBA)*, 1833(12), 3448–3459.

- Ningthoujam, S.S., Talukdar, A.D., Potsangbam, K.S., & Choudhury, M.D. (2012). Challenges in developing medicinal plant databases for sharing ethnopharmacological knowledge. *Journal of Ethnopharmacology*, *141*(1), 9–32.
- Nishiyama, N., & Kataoka, K. (2006). Current state, achievements, and future prospects of polymeric micelles as nanocarriers for drug and gene delivery. *Pharmacology & Therapeutics*, *112*(3):630–648.
- Nnamani, P.O., Hansen, S., Windbergs, M., & Lehr, C.M. (2014). Development of artemether-loaded nanostructured lipid carrier (NLC) formulation for topical application. *International Journal of Pharmaceutics*, *477*(1-2), 208–217.
- Noack, A., Oidtmann, J., Kutza, J., & Mader, K. (2012). *In vitro* digestion of curcuminoid-loaded lipid nanoparticles. *Journal of Nanoparticle Research*, *14*(9), 1–19.
- Nobili, S., Lippi, D., Witort, E., Donnini, M., Bausi, L., Mini, E., & Capaccioli, S. (2009). Natural compounds for cancer treatment and prevention. *Pharmacological Research*, *59*(6), 365–378.
- Norbury, C.J., & Hickson, I.D. (2001). Cellular responses to DNA damage. *Annual Review of Pharmacology & Toxicology*, *41*, 367–401.
- Oishi, M., Nakaogami, J., Ishii, T., & Nagasaki, Y. (2006). Smart PEGylated gold nanoparticles for the cytoplasmic delivery of siRNA to induce enhanced gene silencing. *Chemistry Letters*, *35*(9), 1046–1047.
- Oliveira, P.A., Colaço, A., Chaves, R., Guedes-Pinto, H., De-La-Cruz P, L.F., & Lopes, C. (2007). Chemical carcinogenesis. *Annals of the Brazilian Academy of Sciences*, *79*(4), 593–616.
- Ong, H.C., & Nordiana, M. (1999). Malay ethno-medico botany in Machang, Kelantan, Malaysia. *Fitoterapia*, *70*(5), 502–513.
- Oun, R., & Wheate, N.J. (2013). Platinum anticancer drugs. In R.H. Kretsinger, V.N. Uversky, & E.A. Permyakov (Eds), *Encyclopedia of Metalloproteins* (pp. 1710–1714). New York, NY: Springer New York.
- Ozawa, T. (1995). Mechanism of somatic mitochondrial DNA mutations associated with age and diseases. *Biochimica et Biophysica Acta (BBA)*, *1271*(1), 177–189.
- Padhye, S., Banerjee, S., Ahmad, A., Mohammad, R., & Sarkar, F.H. (2008). From here to eternity - the secret of Pharaohs: Therapeutic potential of black cumin seeds and beyond. *Cancer Therapy*, *6*(b), 495–510.
- Pandit-Taskar, N. & Ma, W. (2013). Gynecologic Cancers. In H.W. Strauss, G. Mariani, D. Volterrani, & S.M. Larson (Eds.), *Nuclear Oncology: Pathophysiology and Clinical Applications* (pp. 591–620). New York, NY: Springer New York.

- Pantsyrnaya, T., Blanchard, F., Delaunay, S., Goergen, J.L., Guédon, E., Guseva, E., & Boudrant, J. (2011). Effect of surfactants, dispersion and temperature on solubility and biodegradation of phenanthrene in aqueous media. *Chemosphere*, 83(1), 29–33.
- Pardeike, J., Hommoss, A., & Müller, R.H. (2009). Lipid nanoparticles (SLN, NLC) in cosmetic and pharmaceutical dermal products. *International Journal of Pharmaceutics*, 366(1-2), 170–184.
- Pardeike, J., Weber, S., Matsko, N., & Zimmer, A. (2012). Formation of a physical stable delivery system by simply autoclaving nanostructured lipid carriers (NLC). *International Journal of Pharmaceutics*, 439(1-2), 22–27.
- Pari, L., & Sankaranarayanan, C. (2009). Beneficial effects of thymoquinone on hepatic key enzymes in streptozotocin-nicotinamide induced diabetic rats. *Life Sciences*, 85(23–26), 830–834.
- Parrish, A.B., Freel, C.D., & Kornbluth, S. (2013). Cellular mechanisms controlling caspase activation and function. *Cold Spring Harbor Perspectives in Biology*, 5(6). Doi: 10.1101/cshperspect.a008672.
- Patel, A.R., & Velikov, K.P. (2011). Colloidal delivery systems in foods: A general comparison with oral drug delivery. *LWT-Food Science & Technology*, 44(9), 1958–1964.
- Patel, D., Dasgupta, S., Dey, S., Ramani, Y.R., Ray, S., & Mazumder, B. (2012). Nanostructured lipid carriers (NLC)-based gel for the topical delivery of aceclofenac: preparation, characterization, and *in vivo* evaluation. *Scientia Pharmaceutica*, 80(3), 749–764.
- Pathan, S.A., Jain, G.K., Zaidi, S.M.A., Akhter, S., Vohora, D., Chander, P., Kole, P.L., Ahmad, F.J., & Khar, R.K. (2011). Stability-indicating ultra-performance liquid chromatography method for the estimation of thymoquinone and its application in biopharmaceutical studies. *Biomedical Chromatography*, 25(5), 613–620.
- Peng, L., Liu, A., Shen, Y., Xu, H.Z., Yang, S.Z., Ying, X.Z., Liao, W., Liu, H.X., Lin, Z.Q., Chen, Q.Y., Cheng, S.W., & Shen, W.D. (2013). Antitumor and anti-angiogenesis effects of thymoquinone on osteosarcoma through the NF- $\kappa$ B pathway. *Oncology Reports*, 29(2), 571–578.
- Pereira, D., & Garey, S.L. (2013). Cancer, cervical. In M.D. Gellman & J. R. Turner (Ed.), *Encyclopedia of Behavioral Medicine* (pp. 309–310). New York: Springer New York.
- Petanidis, S., Hadzopoulou-Cladaras, M., & Salifoglou, A. (2013). Cadmium modulates *H-ras* expression and caspase-3 apoptotic cell death in breast cancer epithelial MCF-7 cells. *Journal of Inorganic Biochemistry*, 121, 100–107.
- Pichayakorn, W., Suksaeree, J., Boonme, P., Amnuait, T., Taweepreda, W., & Ritthidej, G.C. (2012). Nicotine transdermal patches using polymeric natural rubber as the matrix controlling system: Effect of polymer and plasticizer blends. *Journal of Membrane Science*, 411–412, 81–90.

- Pietras, K., & Östman, A. (2010). Hallmarks of cancer: interactions with the tumor stroma. *Experimental Cell Research*, 316(8), 1324–1331.
- Pišlar, A.H., Zidar, N., Kikelj, D., & Kos, J. (2014). Cathepsin X promotes 6-hydroxydopamine-induced apoptosis of PC12 and SH-SY5Y cells. *Neuropharmacology*, 82, 121–131.
- Pojarová, M., Kaufmann, D., Gastpar, R., Nishino, T., Reszka, P., Bednarski, P.J., & von Angerer, E. (2007). [(2-Phenylindol-3-yl)methylene]propanedinitriles inhibit the growth of breast cancer cells by cell cycle arrest in G(2)/M phase and apoptosis. *Bioorganic & Medicinal Chemistry*, 15(23), 7368–7379.
- Qi, F., Li, A., Inagaki, Y., Gao, J., Li, J., Kokudo, N., Li, X.K., & Tang, W. (2010). Chinese herbal medicines as adjuvant treatment during chemo- or radio-therapy for cancer. *BioScienceTrends*, 4(6), 297–307.
- Rabik, C.A., & Dolan, M.E. (2007). Molecular mechanisms of resistance and toxicity associated with platinating agents. *Cancer Treatment Reviews*, 33(1), 9–23.
- Radogna, F., Dicato, M., & Diederich, M. (2015). Cancer-type-specific crosstalk between autophagy, necroptosis and apoptosis as a pharmacological target. *Biochemical Pharmacology*, 94(1):1–11.
- Raemy, E., & Martinou, J.C. (2014). Involvement of cardiolipin in tBID-induced activation of BAX during apoptosis. *Chemistry & Physics of Lipids*, 179, 70–74.
- Raghunandhakumar, S., Paramasivam, A., Senthilraja, S., Naveenkumar, C., Asokkumar, S., Binuclara, J., Jagan, S., Anandakumar, P., & Devaki, T. (2013). Thymoquinone inhibits cell proliferation through regulation of G1/S phase cell cycle transition in N-nitrosodiethylamine-induced experimental rat hepatocellular carcinoma. *Toxicology Letters*, 223(1), 60–72.
- Rahman, H.S., Rasedee, A., How, C.W., Abdul, A.B., Zeenathul, N.A., Othman, H.H., Saeed, M.I., & Yeap, S.K. (2013). Zerumbone-loaded nanostructured lipid carriers: preparation, characterization, and antileukemic effect. *International Journal of Nanomedicine*, 8, 2769–2781.
- Rajkamal, G., Suresh, K., Sugunadevi, G., Vijayaanand, M.A., & Rajalingam, K. (2010). Evaluation of chemopreventive effects of Thymoquinone on cell surface glycoconjugates and cytokeratin expression during DMBA induced hamster buccal pouch carcinogenesis. *BMB Reports*, 43(10), 664–669.
- Rajput, S., Kumar, B.N., Dey, K.K., Pal, I., Parekh, A., & Mandal, M. (2013). Molecular targeting of Akt by thymoquinone promotes G1 arrest through translation inhibition of cyclin D1 and induces apoptosis in breast cancer cells. *Life Sciences*, 93(21), 783–790.
- Rajsekhar, S., & Kuldeep, B. (2011). Pharmacognosy and pharmacology of *Nigella sativa*-A review. *International Research Journal of Pharmacy*, 2(11), 36–39.

- Randhawa, M.A., & Alghamdi, M.S. (2011). Anticancer activity of *Nigella sativa* (black seed)-a review. *American Journal of Chinese Medicine*, 39(6), 1075–1091.
- Ravindran, J., Nair, H.B., Sung, B., Prasad, S., Tekmal, R.R., & Aggarwal, B.B. (2010). Thymoquinone poly (lactide-co-glycolide) nanoparticles exhibit enhanced anti-proliferative, anti-inflammatory, and chemosensitization potential. *Biochemical Pharmacology*, 79(11), 1640–1647.
- Ray, R.S., Rana, B., Swami, B., Venu, V., & Chatterjee, M. (2006). Vanadium mediated apoptosis and cell cycle arrest in MCF7 cell line. *Chemico-Biological Interactions*, 163(3), 239–247.
- Ren, D., Tu, H.C., Kim, H., Wang, G.X., Bean, G.R., Takeuchi, O., Jeffers, J.R., Zambetti, G.P., Hsieh, J.J., & Cheng, E.H. (2010). BID, BIM, and PUMA are essential for activation of the BAX- and BAK-dependent cell death program. *Science*, 330(6009), 1390–1393.
- Renzi, D., Valtolini, M., & Forster, R. (1993). The evaluation of a multi-endpoint cytotoxicity assay system. *ATLA-Alternative to Laboratory Animals*, 21(1), 89–96.
- Richards, J.E., & Hawley, R.S. (2011). The multiple-hit hypothesis: how genes play a role in cancer. In J.E. Richards, & R.S. Hawley, (3<sup>rd</sup> Eds.), *The Human Genome* (pp. 343–366). Massachusetts: Elsevier Inc.
- Riedl, S.J., & Salvesen, G.S. (2007). The apoptosome: signalling platform of cell death. *Nature Reviews Molecular Cell Biology*, 8(5), 405–413.
- Righi, A., Gambarotti, M., Benini, S., Gamberi, G., Cocchi, S., Picci, P., & Bertoni, F. (2015). MDM2 and CDK4 expression in periosteal osteosarcoma. *Human Pathology*, 46(4), 549–553.
- Riley, L.B., & Anderson, D.W. (2011). Cancer epigenetics. In T. Tollefsbol, (Ed.), *Handbook of Epigenetics: The New Molecular and Medical Genetics* (pp. 521–534). Massachusetts: Elsevier Inc.
- Riley, T., Sontag, E., Chen, P., & Levine, A. (2008). Transcriptional control of human p53-regulated genes. *Nature Reviews Molecular Cell Biology*, 9(5), 402–412.
- Roepke, M., Diestel, A., Bajbouj, K., Walluscheck, D., Schonfeld, P., Roessner, A., Schneider-Stock, R., & Gali-Muhtasib, H.U. (2007). Lack of p53 augments thymoquinone-induced apoptosis and caspase activation in human osteosarcoma cells. *Cancer Biology & Therapy*, 6(2), 160–169.
- Roos, W.P., & Kaina, B. (2006). DNA damage-induced cell death by apoptosis. *Trends in Molecular Medicine*, 12(9), 440–450.
- Rubasinghege, G., Lentz, R.W., Park, H., Scherer, M.M. & Grassian, V.H. (2010). Nanorod dissolution quenched in the aggregated state. *Langmuir*, 26(3), 1524–1527.



- Sachdeva, V., & Kataria, M.K. (2013). Enhancement of bioavailability through increase in drug permeation, stability and retention time. *International Research Journal of Pharmacy*, 4(5), 59–66.
- Saleem, U., Ahmad, B., Rehman, K., Mahmood, S., Alam, M., & Erum, A. (2012). Nephro-protective effect of vitamin C and *Nigella sativa* oil on gentamicin associated nephrotoxicity in rabbits. *Pakistan Journal of Pharmaceutical Sciences*, 25(4), 727–730.
- Saleh, H.S. (2014). Can visual inspection with acetic acid be used as an alternative to Pap smear in screening cervical cancer? *Middle East Fertility Society Journal*, 19(3), 187–191.
- Salim, E.I. (2010). Cancer chemopreventive potential of volatile oil from black cumin seeds, *Nigella sativa* L., in a rat multi-organ carcinogenesis bioassay. *Oncology Letters*, 1(5), 913–924.
- Salome, A.C., Godswill, C.O., & Ikechukwu, I.O. (2013). Kinetics and mechanisms of drug release from swellable and non Swellable matrices: A review. *Research Journal of Pharmaceutical, Biological & Chemical Sciences*, 4(2), 97–103.
- Salomi, M.J., Nair, S.C., & Panikkar, K.R. (1991). Inhibitory effects of *Nigella sativa* and saffron (*Crocus sativus*) on chemical carcinogenesis in mice. *Nutrition & Cancer*, 16(1), 67–72.
- Salvia-Trujillo, L., Qian, C., Martín-Belloso, O., & McClements, D.J. (2013). Influence of particle size on lipid digestion and  $\beta$ -carotene bioaccessibility in emulsions and nanoemulsions. *Food Chemistry*, 141(2), 1472–1480.
- Samarakoon, S.R., Thabrew, I., Galhena, P.B., & Tennekoon, K.H. (2012). Modulation of apoptosis in human hepatocellular carcinoma (HepG2 cells) by a standardized herbal decoction of *Nigella sativa* seeds, *Hemidesmus indicus* roots and *Smilax glabra* rhizomes with anti- hepatocarcinogenic effects. *BMC Complementary & Alternative Medicine*, 12(25). Doi: 10.1186/1472-6882-12-25.
- Samarakoon, S.R., Thabrew, I., Galhena, P.B., De Silva, D., & Tennekoon, K.H. (2010). A comparison of the cytotoxic potential of standardized aqueous and ethanolic extracts of a polyherbal mixture comprised of *Nigella sativa* (seeds), *Hemidesmus indicus* (roots) and *Smilax glabra* (rhizome). *Pharmacognosy Research*, 2(6), 335–342.
- Sanna, V., Pala, N., & Sechi, M. (2014). Targeted therapy using nanotechnology: focus on cancer. *International Journal of Nanomedicine*, 9, 467–483.
- Sarnes, A., Kovalainen, M., Häkkinen, M.R., Laaksonen, T., Laru, J., Kiesvaara, J., Ilkka, J., Oksala, O., Rönkkö, S., Järvinen, K., Hirvonen, J., & Peltonen, L. (2014). Nanocrystal-based per-oral itraconazole delivery: superior *in vitro* dissolution enhancement versus Sporanox® is not realized in *in vivo* drug absorption. *Journal of Controlled Release*, 180, 109–116.

- Sawant, K.K., & Dodiya, S.S. (2008). Recent advances and patents on solid lipid nanoparticles. *Recent Patents on Drug Delivery & Formulation*, 2(2), 120–135.
- Schafmayer, C., Buch, S., Egberts, J.H., Brosch, M., El-Sharawy, A., Conring, M., Koschnick, M., Koschnick, M., Schwiedernoch, S., Katalinic, A., Kremer, B., Folsch, U.R., Krawczak, M., Fandrich, F., Schreiber, S., Tepel, J., & Hampe, J. (2007). Genetic investigation of DNA-repair pathway genes *PMS2*, *MLH1*, *MSH2*, *MSH6*, *MUTYH*, *OGG1* and *MTH1* in sporadic colon cancer. *International Journal of Cancer*, 121(3), 555–558.
- Schmitt, M., & Pawlita, M. (2011). The HPV transcriptome in HPV16 positive cell lines. *Molecular & Cellular Probes*, 25(2-3), 108–113.
- Schneider-Stock, R., Fakhoury, I.H., Zaki, A.M., El-Baba, C.O., & Gali-Muhtasib, H.U. (2014). Thymoquinone: fifty years of success in the battle against cancer models. *Drug Discovery Today*, 19(1), 18–30.
- Schöler, N., Zimmermann, E., Katzfey, U., Hahn, H., Müller, R.H., & Liesenfeld, O. (2000). Effect of solid lipid nanoparticles (SLN) on cytokine production and the viability of murine peritoneal macrophages. *Journal of Microencapsulation*, 17(5):639–650.
- Scholz, M., Lipinski, M., Leupold, M., Luftmann, H., Harig, L., Ofir, R., Fischer, R., Prüfer, D., & Müller, K.J. (2009). Methyl jasmonate induced accumulation of kalopanaxsaponin I in *Nigella sativa*. *Phytochemistry*, 70(4), 517–522.
- Schwab, M. (2011). Cancer. In M. Schwab, (Ed.) *Encyclopedia of Cancer* (pp. 605–607). Berlin: Springer Berlin Heidelberg.
- Schwartzmann, G., Brondani da Rocha, A., Berlinck, R.G., & Jimeno, J. (2001). Marine organisms as a source of new anticancer agents. *Lancet Oncology*, 2(4), 221–225.
- Sedaghat, R., Roghani, M., & Khalili, M. (2014). Neuroprotective effect of thymoquinone, the *Nigella sativa* bioactive compound, in 6-hydroxydopamine-induced hemi-parkinsonian rat model. *Iranian Journal of Pharmaceutical Research*, 13(1), 227–234.
- Selvamuthukumar, S., & Velmurugan, R. (2012). Nanostructured lipid carriers: a potential drug carrier for cancer chemotherapy. *Lipids in Health and Disease*, 11(159). Doi: 10.1186/1476-511X-11-159.
- Sethi, G., Ahn, K.S., & Aggarwal, B.B. (2008). Targeting nuclear factor-kappa B activation pathway by thymoquinone: role in suppression of antiapoptotic gene products and enhancement of apoptosis. *Molecular Cancer Research*, 6(6), 1059–1070.
- Severino, P., Andreani, T., Macedo, A.S., Fangueiro, J.F., Santana, M.H., Silva, A.M., & Souto, E.B. (2011). Current state-of-art and new trends on lipid nanoparticles (SLN and NLC) for oral drug delivery. *Journal of Drug Delivery*, 2012. Doi: 10.1155/2012/750891.



- Shafi, G., Hasan, T.N., & Syed, N.A. (2008). Methanolic extract of *Nigella sativa* seeds is potent clonogenic inhibitor of PC3 cells. *International Journal of Pharmacology*, 4(6), 477–481.
- Shafi, G., Munshi, A., Hasan, T.N., Alshatwi, A.A., Jyothy, A., & Lei, D.K. (2009). Induction of apoptosis in HeLa cells by chloroform fraction of seed extracts of *Nigella sativa*. *Cancer Cell International*, 9(29). Doi:10.1186/1475-2867-9-29.
- Shaheen, B.S. (2009). *Nigella sativa*. Retrieved from University of Wisconsin-La website: [http://bioweb.uwlax.edu/bio203/s2009/shaheen\\_baya/Habitat.htm](http://bioweb.uwlax.edu/bio203/s2009/shaheen_baya/Habitat.htm).
- Shi, L.L., Cao, Y., Zhu, X.Y., Cui, J.H., & Cao, Q.R. (2014). Optimization of process variables of zanamivir-loaded solid lipid nanoparticles and the prediction of their cellular transport in Caco-2 cell model. *International Journal of Pharmaceutics*, 478(1), 60–69.
- Shoieb, A.M., Elgayyar, M., Dudrick, P.S., Bell, J.L., & Tithof P.K. (2003). *In vitro* inhibition of growth and induction of apoptosis in cancer cell lines by thymoquinone. *International Journal of Oncology*, 22(1), 107–113.
- Shu, K.X., Li, B., & Wu, L.X. (2007). The p53 network: p53 and its downstream genes. *Colloids and Surfaces B: Biointerfaces*, 55(1), 10–18.
- Siegel, J., Kvítek, O., Ulbrich, P., Kolská, Z., Slepíčka, P., & Švorčík, V. (2012). Progressive approach for metal nanoparticle synthesis. *Materials Letters*, 89, 47–50.
- Siegel, R.A., & Rathbone, M.J. (2012). Overview of controlled release mechanisms. In J. Siepmann, R.A. Siegel, & M.J. Rathbone, (Ed) *Fundamentals and Applications of Controlled Release Drug Delivery* (pp. 19–43). US: Controlled Release Society.
- Siepmann, J., & Siepmann, F. (2008). Mathematical modeling of drug delivery. *International Journal of Pharmaceutics*, 364(2), 328–343.
- Silva, A.C., Amaral, M.H., González-Mira, E., Santos, D., & Ferreira, D. (2012). Solid lipid nanoparticles (SLN)-based hydrogels as potential carriers for oral transmucosal delivery of risperidone: preparation and characterization studies. *Colloids and Surfaces B: Biointerfaces*, 93, 241–248.
- Singh, A, Ahmad, I., Akhter, S., Jain, G.K., Iqbal, Z., Talegaonkar, S., & Ahmad, F.J. (2013a). Nanocarrier based formulation of Thymoquinone improves oral delivery: stability assessment, *in vitro* and *in vivo* studies. *Colloids & Surfaces B: Biointerfaces*, 102, 822–832.
- Singh, M., Bhui, K., Singh, R., & Shukla, Y. (2013b). Tea polyphenols enhance cisplatin chemosensitivity in cervical cancer cells via induction of apoptosis. *Life Sciences*, 93(1), 7–16.
- Singh, N. (2007). Apoptosis in health and disease and modulation of apoptosis for therapy: An overview. *Indian Journal of Clinical Biochemistry*, 22(2), 6–16.

- Singhvi, G., & Singh, M. (2011). Review: *In vitro* drug release characterization models. *International Journal of Pharmaceutical Studies & Research*, 2(1), 77–84.
- Slade, B.A., Leidel, L., Vellozzi, C., Woo, E.J., Hua, W., Sutherland, A., Izurieta, H., Ball, R., Miller, N., Miles Braun, M., Markowitz, L.E., & Iskander, J. (2009). Postlicensure safety surveillance for quadrivalent human papillomavirus recombinant vaccine. *Journal of the American Medical Association*, 302(7), 750–757.
- Sonnenschein, C., & Soto, A.M. (2013). Neoplasm. In W. Dubitzky, O. Wolkenhauer, K.H. Cho, H. Yokota (Eds.), *Encyclopedia of Systems Biology* (pp. 1506–1507). Berlin: Springer Berlin Heidelberg.
- Sotiriadis, A., Dagklis, T., Siamanta, V., Chatzigeorgiou, K., & Agorastos, T. (2012). Increasing fear of adverse effects drops intention to vaccinate after the introduction of prophylactic HPV vaccine. *Archives of Gynecology & Obstetrics*, 285(6), 1719–1724.
- Souto, E.B., Wissing, S.A., Barbosa, C.M., & Müller, R.H. (2004). Development of a controlled release formulation based on SLN and NLC for topical clotrimazole delivery. *International Journal of Pharmaceutics*, 278(1), 71–77.
- Sriamornsak, P., Thirawong, N., Weerapol, Y., Nunthanid, J., & Sungthongjeen, S. (2007). Swelling and erosion of pectin matrix tablets and their impact on drug release behavior. *European Journal of Pharmaceutical Sciences*, 67(1), 211–219.
- Stoka, V., Turk, V., & Bredesen, D.E. (2006). Differential regulation of the intrinsic pathway of apoptosis in brain and liver during ageing. *FEBS Letters*, 580(15), 3739–3745.
- Suddek, G.M., Ashry, N.A., & Gameil, N.M. (2013). Thymoquinone attenuates cyclophosphamide-induced pulmonary injury in rats. *Inflammopharmacology*, 21(6), 427–435.
- Sun, B., Geng, S., Huang, X., Zhu, J., Liu, S., Zhang, Y., Ye, J., Li, Y., & Wang, J. (2011). Coleusin factor exerts cytotoxic activity by inducing G0/G1 cell cycle arrest and apoptosis in human gastric cancer BGC-823 cells. *Cancer Letters*, 301(1), 95–105.
- Sutton, K.M., Greenshields, A.L., & Hoskin, D.W. (2014). Thymoquinone, a bioactive component of black caraway seeds, causes G1 phase cell cycle arrest and apoptosis in triple-negative breast cancer cells with mutant p53. *Nutrition & Cancer*, 66(3), 408–418.
- Swamy, S.M., & Tan, B.K. (2000). Cytotoxic and immunopotentiating effects of ethanolic extract of *Nigella sativa* L. Seeds. *Journal of Ethnopharmacology*, 70(1), 1–7.
- Tadros, T. (2013). Drug delivery. In T. Tadros (Ed.), *Encyclopedia of Colloid and Interface Science* (pp. 339–340). Berlin: Springer Berlin Heidelberg.

- Tamatani, T., Azuma, M., Motegi, K., Takamaru, N., Kawashima, Y., & Bando, T. (2007). Cepharranthin-enhanced radiosensitivity through the inhibition of radiation-induced nuclear factor-kappaB activity in human oral squamous cell carcinoma cells. *International Journal of Oncology*, *31*(4), 761–768.
- Tamjidi, F., Shahedi, M., Varshosaz, J., & Nasirpour, A. (2013). Nanostructured lipid carriers (NLC): A potential delivery system for bioactive food molecules. *Innovative Food Science & Emerging Technologies*, *19*, 29–43.
- Tariq, M. (2008). *Nigella sativa* seeds: folklore treatment in modern day medicine. *Saudi Journal of Gastroenterology*, *14*(3), 105–106.
- Tascilar, M., de Jong, F.A., Verweij, J., & Mathijssen, R.H. (2006). Complementary and alternative medicine during cancer treatment: beyond innocence. *Oncologist*, *11*(7), 732–741.
- Taylor, R.C., Cullen, S.P., & Martin, S.J. (2008). Apoptosis: controlled demolition at the cellular level. *Nature Reviews Molecular Cell Biology*, *9*(3), 231–241.
- Teeranachaideekul, V., Boonme, P., Souto, E.B., Müller, R.H., & Junyaprasert, V.B. (2008). Influence of oil content on physicochemical properties and skin distribution of Nile red-loaded NLC. *Journal of Controlled Release*, *128*(2), 134–141.
- Teeranachaideekul, V., Souto, E.B., Junyaprasert, V.B., & Müller, R.H. (2007). Cetyl palmitate-based NLC for topical delivery of Coenzyme Q10 - Development, physicochemical characterization and *in vitro* release studies. *European Journal of Pharmaceutics & Biopharmaceutics*, *67*, 141–148.
- Thabrew, M.I., Mitry, R.R., Morsy, M.A., & Hughes, R.D. (2005). Cytotoxic effects of a decoction of *Nigella sativa*, *Hemidesmus indicus* and *Smilax glabra* on human hepatoma HepG2 cells. *Life Sciences*, *77*(12), 1319–1330.
- Thangam, R., Sathuvan, M., Poongodi, A., Suresh, V., Pazhanichamy, K., Sivasubramanian, S., Kanipandian, N., Ganesan, N., Rengasamy, R., Thirumurugan, R., & Kannan, S. (2014). Activation of intrinsic apoptotic signaling pathway in cancer cells by *Cymbopogon citratus* polysaccharide fractions. *Carbohydrate Polymers*, *107*, 138–150.
- Thatipamula, R.P., Palem, C.R., Gannu, R., Mudragada, S., & Yamsani, M.R. (2011). Formulation and *in vitro* characterization of domperidone loaded solid lipid nanoparticle and nanostructured lipid carrier. *DARU*, *19*(1), 23–32.
- Tian, Z., Yi, Y., Yuan, H., Han, J., Zhang, X., Xie, Y., Lu, Y., Qi, J., & Wu, W. (2013). Solidification of nanostructured lipid carriers (NLCs) onto pellets by fluid-bed coating: Preparation, *in vitro* characterization and bioavailability in dogs. *Powder Technology*, *247*, 120–127.
- Tjalma, W.A., & Depuydt, C.E. (2013). Cervical cancer screening: which HPV test should be used-L1 or E6/E7? *European Journal of Obstetrics & Gynecology and Reproductive Biology*, *170*(1), 45–46.

- Tjalma, W.A., Weyler, J.J., Bogers, J.J., Pollefliet, C., Baay, M., Goovaerts, G.C., Vermorken, J.B., van Dam, P.A., van Marck, E.A., & Buytaert, P.M. (2001). The importance of biological factors (bcl-2, bax, p53, PCNA, MI, HPV and angiogenesis) in invasive cervical cancer. *European Journal of Obstetrics & Gynecology and Reproductive Biology*, 97(2), 223–230.
- Torre, G.L., de Waure, C., Chiaradia, G., Mannocci, A., & Ricciardi, W. (2007). HPV vaccine efficacy in preventing persistent cervical HPV infection: A systematic review and meta-analysis. *Vaccine*, 25(50), 8352–8358.
- Torres, M.P., Ponnusamy, M.P., Chakraborty, S., Smith, L.M., Das, S., Arafat, H.A., & Batra, S.K. (2010). Effects of thymoquinone in the expression of mucin 4 in pancreatic cancer cells: implications for the development of novel cancer therapies. *Molecular Cancer Therapeutics*, 9(5), 1419–1431.
- Trang, N.T., Wanner, M.J., Phuong le, V.N., Koomen, G.J., & Dung, N.X. (1993). Thymoquinone from *Eupatorium ayapana*. *Planta Medica*, 59(1), 99.
- Tripathi, A. Gupta, R., & Saraf, S. (2010). PLGA nanoparticles of anti tubercular drug: drug loading and release studies of a water insoluble drug. *International Journal of PharmTech Research*, 2(3), 2116–2123.
- Trump, B.F., Berezsky, I.K., Chang, S.H., & Phelps, P.C. (1997). The pathways of cell death: oncosis, apoptosis, and necrosis. *Toxicologic Pathology*, 25(1), 82–88.
- Ullah, I., Baloch, M.K., & Durrani, G.F. (2012). Solubility of lidocaine in ionic, nonionic and zwitterionic surfactants. *Journal of Solution Chemistry*, 41(2), 215–222.
- Ullah, I., Baloch, M.K., Ullah, I., & Mustaqeem, M. (2014). Enhancement in aqueous solubility of mefenamic acid using micellar solutions of various surfactants. *Journal of Solution Chemistry*, 43(8), 1360–1373.
- Ulu, R., Dogukan, A., Tuzcu, M., Gencoglu, H., Ulas, M., Ilhan, N., Muqbil, I., Mohammad, R.M., Kucuk, O., & Sahin, K. (2012). Regulation of renal organic anion and cation transporters by thymoquinone in cisplatin induced kidney injury. *Food & Chemical Toxicology*, 50(5), 1675–1679.
- Umar, S., Zargan, J., Umar, K., Ahmad, S., Katiyar, C.K., & Khan, H.A. (2012). Modulation of the oxidative stress and inflammatory cytokine response by thymoquinone in the collagen induced arthritis in Wistar rats. *Chemico-Biological Interactions*, 197(1), 40–46.
- Uprit, S., Kumar Sahu, R., Roy, A., & Pare, A. (2013). Preparation and characterization of minoxidil loaded nanostructured lipid carrier gel for effective treatment of alopecia. *Saudi Pharmaceutical Journal*, 21(4), 379–385.
- USP (2008). *United States Pharmacopeia (USP 31-NF 26) 711, 1088, 1092, 1225*. Rockville: United States Pharmacopoeial Convention, Inc.

- van Aken, G.A., Bomhof, E., Zoet, F.D., Verbeek, M., & Oosterveld, A. (2011). Differences in *in vitro* gastric behaviour between homogenized milk and emulsions stabilised by Tween 80, whey protein, or whey protein and caseinate. *Food Hydrocolloid*, 25(4), 781–788.
- Van Cruchten, S., & Van Den Broeck, W. (2002). Morphological and biochemical aspects of apoptosis, oncosis and necrosis. *Anatomia, Histologia, Embryologia*, 31(4), 214–223.
- van der Velden, L.A., Manni, J.J., Ramaekers, F.C., & Kuijpers, W. (1999). Expression of intermediate filament proteins in benign lesions of the oral mucosa. *European Archives of Oto-Rhino-Laryngology*, 256(10), 514–519.
- Varshosaz, J., Tabbakhian, M., & Mohammadi, M.Y. (2010). Formulation and optimization of solid lipid nanoparticles of buspirone HCl for enhancement of its oral bioavailability. *Journal of Liposome Research*, 20(4), 286–296.
- Veluthoor, S., Kelsey, R.G., González-Hernández, M.P., Panella, N., Dolan, M., & Karchesy, J. (2011). Composition of the heartwood essential oil of incense cedar (*Calocedrus decurrens* Torr.). *Holzforschung*, 65(3), 333–336.
- Venkatachallam, S.K.T., Pattekhan, H., Divakar, S., & Kadimi, U.S. (2010). Chemical composition of *Nigella sativa* L. seed extracts obtained by supercritical carbon dioxide. *Journal of Food Science & Technology*, 47(6), 598–605
- Verma, P., Thakur, A.S., Deshmukh, K., Jha, A.K., & Verma, S. (2010). Routes of drug administration. *International Journal of Pharmaceutical Studies & Research*, 1(1), 54–59.
- Vermes, I., Haanen, C., Steffens-Nakken, H., & Reutellingsperger, C. (1995). A novel assay for apoptosis Flow cytometric detection of phosphatidylserine expression on early apoptotic cells using fluorescein labelled Annexin V. *Journal of Immunological Methods*, 184(1), 39–51.
- Vijayan, V., Reddy, K.R., Sakthivel, S., & Swetha, C. (2013). Optimization and characterization of repaglinide biodegradable polymeric nanoparticle loaded transdermal patches: *in vitro* and *in vivo* studies. *Colloids & Surfaces B: Biointerfaces*, 111, 150–155.
- Villa, L.L. (2012). Cervical cancer in Latin America and the Caribbean: the problem and the way to solutions. *Cancer Epidemiology, Biomarkers & Prevention*, 21(9), 1409–1413.
- Vivek, R., Thangam, R., Nipunbabu, V., Ponraj, T., & Kannan, S. (2014). Oxaliplatin-chitosan nanoparticles induced intrinsic apoptotic signaling pathway: a "smart" drug delivery system to breast cancer cell therapy. *International Journal of Biological Macromolecules*, 65, 289–297.
- von Schwarzenberg, K., & Vollmar, A.M. (2013). Targeting apoptosis pathways by natural compounds in cancer: marine compounds as lead structures and chemical tools for cancer therapy. *Cancer Letters*, 332(2), 295–303.



- Vucic, D., Dixit, V.M., & Wertz, I.E. (2011). Ubiquitylation in apoptosis: a post-translational modification at the edge of life and death. *Nature Reviews Molecular Cell Biology*, 12(7), 439–452.
- Wachmann, K., Pop, C., van Raam, B.J., Drag, M., Mace, P.D., Snipas, S.J., Zmasek, C., Schwarzenbacher, R., Salvesen, G.S., & Riedl, S.J. (2010). Activation and specificity of human caspase-10. *Biochemistry*, 49(38), 8307–8315.
- Wain, G. (2010). The human papillomavirus (HPV) vaccine, HPV related diseases and cervical cancer in the post-reproductive years. *Maturitas*, 65(3), 205–209.
- Walsh, K. R., Zhang, Y.C., Vodovotz, Y., Schwartz, S.J., & Failla, M.L. (2003). Stability and bioaccessibility of isoflavones from soy bread during *in vitro* digestion. *Journal of Agricultural & Food Chemistry*, 51(16), 4603–4609.
- Wang, K. (2014). Molecular mechanisms of liver injury: apoptosis or necrosis. *Experimental & Toxicologic Pathology*, 66(8), 351–356.
- Wang, P., Reed, M., Wang, Y., Mayr, G., Stenger, J.E., Anderson, M.E., Schwedes, J.F., & Tegtmeyer, P. (1994). p53 domains: structure, oligomerization, and transformation. *Molecular and Cellular Biology*, 14(8), 5182–5191.
- Wang, R., Billone, P.S., & Mullett, W.M. (2013). Nanomedicine in action: an overview of cancer nanomedicine on the market and in clinical trials. *Journal of Nanomaterials*, 2013. Doi: 10.1155/2013/629681.
- Wang, S., Yu, H., & Wickliffe, J.K. (2011). Limitation of the MTT and XTT assays for measuring cell viability due to superoxide formation induced by nano-scale TiO<sub>2</sub>. *Toxicology In Vitro*, 25(8), 2147–2151.
- Wani, M.C., Taylor, H.L., Wall, M.E., Coggon, P., & McPhail, A.T. (1971). Plant antitumor agents. VI. The isolation and structure of taxol, a novel antileukemic and antitumor agent from *Taxus brevifolia*. *Journal of the American Chemical Society*, 93(9), 2325–2327.
- Warren, L., Buck, C.A., & Tuszynski, G.P. (1978). Glycopeptide changes and malignant transformation: A possible role for carbohydrate in malignant behavior. *Biochimica et Biophysica Acta*, 516(1), 97–127.
- Weber, G.F. (2007). Theories of carcinogenesis. In G.F. Weber (Ed.), *Molecular Mechanisms of Cancer* (pp. 7–28). Netherlands: Springer Netherlands.
- Wei, J., Yam, W., Li, X., Ding, Y., & Tai, H. (2010). Thromboxane receptor  $\alpha$  mediates tumor growth and angiogenesis via induction of vascular endothelial growth factor expression in human lung cancer cells. *Lung Cancer*, 69(1), 26–32.
- Westphal, D., Dewson, G., Czabotar, P.E., & Kluck, R.M. (2011). Molecular biology of Bax and Bak activation and action. *Biochimica et Biophysica Acta*, 1813(84), 521–531.



- WHO. (2014). The top 10 causes of death. Retrieved December 11, 2014, from the World Health Organisation website: <http://www.who.int/mediacentre/factsheets/fs310/en/>.
- Wicki, A., Witzigmann, D., Balasubramanian, V., & Huwyler, J. (2015). Nanomedicine in cancer therapy: challenges, opportunities, and clinical applications. *Journal of Controlled Release*, *200*, 138–157.
- Willenberg, I., Michael, M., Wonik, J., Bartel, L.C., Empl, M.T., & Schebb, N.H. (2015). Investigation of the absorption of resveratrol oligomers in the Caco-2 cellular model of intestinal absorption. *Food Chemistry*, *167*, 245–250.
- Williams, H.D., Trevaskis, N.L., Charman, S.A., Shanker, R.M., Charman, W.N., Pouton, C.W., & Porter, C.J. (2013). Strategies to address low drug solubility in discovery and development. *Pharmacological Reviews*, *65*(1), 315–499.
- Willis, S.N., Fletcher, J., Kaufmann, T., van Delft, M.F., Chen, L., Czabotar, P.E., Ierino, H., Lee, E.F., Fairlie, W.D., Bouillet, P., Strasser, A., Kluck, R.M., Adams, J.M., & Huang, D.C. (2007). Apoptosis initiated when BH3 ligands engage multiple Bcl-2 homologs, not Bax or Bak. *Science*, *315*(5813), 856–859.
- Woo, C.C., Hsu, A., Kumar, A.P., Sethi, G., & Tan, K.H. (2013). Thymoquinone inhibits tumor growth and induces apoptosis in a breast cancer xenograft mouse model: the role of p38 MAPK and ROS. *PLoS One*, *8*(10). Doi: 10.1371/journal.pone.0075356.
- Woo, C.C., Loo, S.Y., Gee, V., Yap, C.W., Sethi, G., Kumar, A.P., & Tan, K.H. (2011). Anticancer activity of thymoquinone in breast cancer cells: possible involvement of PPAR- $\gamma$  pathway. *Biochemical Pharmacology*, *82*(5), 464–475.
- Woodman, C.B., Collins, S.I., & Young, L.S. (2007). The natural history of cervical HPV infection: unresolved issues. *Nature Reviews Cancer*, *7*(1), 11–22.
- Wu, C.P., Ohnuma, S., & Ambudkar, S. (2011). Discovering natural product modulators to overcome multidrug resistance in cancer chemotherapy. *Current Pharmaceutical Biotechnology*, *12*(4), 609–620.
- Xu, M.Y., & Kim, Y.S. (2014). Antitumor activity of glycyrol via induction of cell cycle arrest, apoptosis and defective autophagy. *Food & Chemical Toxicology*, *74*, 311–319.
- Xu, X., Ho, W., Zhang, X., Bertrand, N., & Farokhzad, O. (2015). Cancer nanomedicine: from targeted delivery to combination therapy. *Trends in Molecular Medicine*, *21*(4), 223–232.
- Yamamoto, M., Taguchi, K., Baba, H., Endo, K., Kohnoe, S., Okamura, T., & Maehara, Y. (2006). Loss of protein expression of hMLH1 and hMSH2 with double primary carcinomas of the stomach and colorectum. *Oncology Reports*, *16*(1), 41–47.
- Yang, F., Teves, S.S., Kemp, C.J., & Henikoff, S. (2014). Doxorubicin, DNA torsion, and chromatin dynamics. *Biochimica et Biophysica Acta*, *1845*(1), 84–89.

- Yang, X., Zhao, L., Almasry, L., Garamus, V.M., Zou, A., Willumeit, R., & Fan, S. (2013). Preparation and characterization of 4-dedimethylamino sancycline (CMT-3) loaded nanostructured lipid carrier (CMT-3/NLC) formulations. *International Journal of Pharmaceutics*, 450(1-2), 225–234.
- Yao, M., McClements, D.J., & Xiao, H. (2015). Improving oral bioavailability of nutraceuticals by engineered nanoparticle-based delivery systems. *Current Opinion in Food Science*, 2, 14–19.
- Yap, T.A., Molife, L.R., & de Bono, J.S. (2008). CDK inhibitors as anticancer agents. In W. Dai (Ed.), *Checkpoint Responses in Cancer Therapy* (pp. 135–162). Totowa: Humana Press.
- Yetimallar, H., Kasap, B., Cukurova, K., Yildiz, A., Keklik, A., & Soylu, F. (2012). Cofactors in human papillomavirus infection and cervical carcinogenesis. *Archives of Gynecology & Obstetrics*, 285(3), 805–810.
- Yi, T., Cho, S.G., Yi, Z., Pang, X., Rodriguez, M., Wang, Y., Sethi, G., Aggarwal, B.B., & Liu, M. (2008). Thymoquinone inhibits tumor angiogenesis and tumor growth through suppressing AKT and extracellular signal-regulated kinase signaling pathways. *Molecular Cancer Therapeutics*, 7(7), 1789–1796.
- Yildiz, F., Coban, S., Terzi, A., Ates, M., Aksoy, N., Cakir, H., Ocak, A.R., & Bitiren, M. (2008). *Nigella sativa* relieves the deleterious effects of ischemia reperfusion injury on liver. *World Journal of Gastroenterology*, 14(33), 5204–5209.
- Yildiz, F., Coban, S., Terzi, A., Savas, M., Bitiren, M., Celik, H., & Aksoy N. (2010). Protective effects of *Nigella sativa* against ischemia-reperfusion injury of kidneys. *Renal Failure*, 32(1), 126–131.
- Yin, X., Li, H., Guo, Z., Wu, L., Chen, F., de Matas, M., Shao, Q., Xiao, T., York, P., He, Y., & Zhang, J. (2013). Quantification of swelling and erosion in the controlled release of a poorly water-soluble drug using synchrotron X-ray computed microtomography. *AAPS Journal*, 15(4), 1025–1034.
- Yu, S.H., Tang, D.W., Hsieh, H.Y., Wu, W.S., Lin, B.X., Chuang, E.Y., Sung, H.W., & Mi, F.L. (2013). Nanoparticle-induced tight-junction opening for the transport of an anti-angiogenic sulfated polysaccharide across Caco-2 cell monolayers. *Acta Biomaterialia*, 9(7), 7449–7459.
- Yunokawa, M., Katsumata, N., Yamamoto, H., Kodaira, M., Yonemori, K., Shimizu, C., Ando, M., Tamura, K., & Fujiwara, Y. (2013). A pilot feasibility study for cisplatin plus S-1 for the treatment for advanced or recurrent cervical cancer. *Cancer Chemotherapy & Pharmacology*, 71(5), 1369–1374.
- Zafeer, M.F., Waseem, M., Chaudhary, S., & Parvez, S. (2012). Cadmium-induced hepatotoxicity and its abrogation by thymoquinone. *Journal of Biochemical & Molecular Toxicology*, 26(5), 199–205.

- Zagouri, F., Sergentanis, T.N., Chrysikos, D., Filipits, M.F., & Bartsch, R. (2012). Molecularly targeted therapies in cervical cancer. A systematic review. *Gynecologic Oncology*, 126(2), 291–303.
- Zainal Ariffin, O., & Nor Saleha, I.T. (2011). *Malaysia Cancer Statistics – Data and Figure 2007*. Kuala Lumpur: National Cancer Registry, Ministry of Health Malaysia.
- Zaridah, S. (2014). A review of cervical cancer research in Malaysia. *Medical Journal of Malaysia*, 69, 33–41.
- Zein, S., Awada, S., Al-Hajje, A., Rachidi, S., Salameh, P., & Kanaan, H. (2012). Variation of thymol, carvacrol and thymoquinone production from wild and cultivated *Origanum syriacum* of South Lebanon. *Journal of Medicinal Plants Research*, 6(9), 1692–1696.
- Zhang, Y., Zhang, Y.J., Zhao, H.Y., Zhai, Q.L., Zhang, Y., & Shen, Y.F. (2013a). The impact of R213 mutation on p53-mediated p21 activity. *Biochimie*, 99, 215–218.
- Zhang, Y.F., Xiao, S., Sun, L., Ge, Z., Fang, F., Zhang, W., Wang, Y., & Cheng, Y. (2013b). Rapid screening of bioactive compounds from natural products by integrating 5-channel parallel chromatography coupled with on-line mass spectrometry and microplate based assays. *Analytica Chimica Acta*, 777, 49–56.
- Zhao, C.R., Gao, Z.H., & Qua, X.J. (2010). Nrf2-ARE signaling pathway and natural products for cancer chemoprevention. *Cancer Epidemiology*, 34(5), 523–533.
- Zhao, R., Gao, X., Cai, Y., Shao, X., Jia, G., Huang, Y., Qin, X., Wang, J., & Zheng, X. (2013). Antitumor activity of *Portulaca oleracea* L. polysaccharides against cervical carcinoma *in vitro* and *in vivo*. *Carbohydrate Polymers*, 96(2), 376–383.
- Zhao, Z., & Lee, C.C. (2010). Circadian clock, cell cycle and cancer: circadian rhythm and cell growth regulation. In U. Albrecht (Ed.), *The Circadian Clock* (pp. 139–155). New York: Springer New York.
- Zhou, X., Zhang, X., Ye, Y., Zhang, T., Wang, H., Ma, Z., & Wu, B. (2015). Nanostructured lipid carriers used for oral delivery of oridonin: An effect of ligand modification on absorption. *International Journal of Pharmaceutics*, 479(2), 391–398.
- Zhu, G., Song, L., & Lippard, S.J. (2013). Visualizing inhibition of nucleosome mobility and transcription by cisplatin-DNA interstrand crosslinks in live mammalian cells. *Cancer Research*, 73(14), 4451–4460.
- Zhuang, C.Y., Li, N., Wang, M., Zhang, X.N., Pan, W.S., Peng, J.J., Pan, Y.S., & Tang, X. (2010). Preparation and characterization of vinpocetine loaded nanostructured lipid carriers (NLC) for improved oral bioavailability. *International Journal of Pharmaceutics*, 394(1-2), 179–185.

Zribi, I., Omezzine, F., & Haouala, R. (2014). Variation in phytochemical constituents and allelopathic potential of *Nigella sativa* with developmental stages. *South African Journal of Botany*, 94, 255–262.

zur Mühlen, A., Schwarz, C., & Mehnert, W. (1998). Solid lipid nanoparticles (SLN) for controlled drug delivery-drug release and release mechanism. *European Journal of Pharmaceutics & Biopharmaceutics*, 45(2), 149–155.

