



UNIVERSITI PUTRA MALAYSIA

***PROCESS DEVELOPMENT FOR HIGHER YIELD PRODUCTION OF
DIACYLGLYCEROL OIL VIA PARTIAL HYDROLYSIS***

PHUAH ENG TONG

IB 2015 36



**PROCESS DEVELOPMENT FOR HIGHER YIELD PRODUCTION OF
DIACYLGLYCEROL OIL VIA PARTIAL HYDROLYSIS**

By

PHUAH ENG TONG

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of
Philosophy**

December 2015

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Doctor of Philosophy

PROCESS DEVELOPMENT FOR HIGHER YIELD PRODUCTION OF DIACYLGLYCEROL OIL VIA PARTIAL HYDROLYSIS

By

PHUAH ENG TONG

December 2015

Chair : Lai Oi-Ming, PhD
Faculty : Institute of Bioscience

The disclosure of diacylglycerol (DAG) oil to replace the conventional edible oils has received increasing interest among researchers and food manufacturers owing to its anti-obesity properties. Distinct processing approaches have been proposed to produce DAG-enriched oil in which enzymatic partial hydrolysis outstands other methods due to its inexpensive raw materials and single-step hydrolytic reaction involved. In present work, single-factor optimization of partial hydrolysis for DAG production from refined, bleached, deodorized palm oil (RBDPO) catalysed by immobilized *Rhizomucor miehei* lipase (Lipozyme RMIM) was carried out in batch system. Effects of four operating parameters namely temperature, enzyme dosage, water content and agitation speed were investigated. Optimum production conditions for palm based-DAG are as follows: temperature = 55°C, enzyme dosage = 10-wt%, water content = 5-wt% and agitation speed = 500 rpm. A DAG yield of 31-wt% was obtained after 6 h of reaction. The partial hydrolysis reaction was found to conform to Ping-Pong Bi-Bi with substrate inhibition mechanism. The optimum operating conditions were then applied to the lab-scale packed bed system.

Packed bed reactor (PBR) is an effective reactor configuration because it enables reusability of the enzyme particles besides enhancing its operational stability. However, mass transfer limitation remains a key challenge in packed bed column system, especially at large scale. A dimensionless mathematical mass transfer model of Colburn factor, J_D , which is a function of Reynolds (Re) and Schmidt (Sc) numbers, was therefore developed to simulate mass transfer phenomena of the reaction mixture in PBR during enzymatic partial hydrolysis reaction. The results revealed that the mass transfer correlation of $J_D = 0.92(Re)^{0.2}$ was able to predict the experimental data accurately. In addition, response surface methodology (RSM) was employed to optimize the process variables

namely packed bed height and substrate flow rates on DAG production in PBR. Quadratic models were successfully developed for both DAG and unhydrolyzed triacylglycerol (TAG) with insignificant lack of fit ($P>0.05$). Optimum conditions for DAG synthesis were evaluated to be 10 cm packed bed height and 3.8 ml/min flow rate with 29-wt% DAG being reported. Immobilized enzyme can be reused up to 10 times without significant loss in enzymatic activity.

The present study also investigated the production efficiency using columns with different length-to-diameter ratios (L/D ratio) to determine the most potential process setup for industrial DAG manufacturing. Practical design issues such as operating temperature, substrate flow rate and reaction time were evaluated with respect to various packed bed column configurations. A column dimension with L/D ratio of two was determined to be the most suitable bed column design for lipase-mediated partial hydrolysis reaction. The optimal reaction temperature, substrate flow rate and residence time for the production of DAG in packed bed column dimension of two were found to be 55°C, 5 ml/min and 5.8 min, respectively. Under these operating conditions, a maximal DAG content of 35-wt% was obtained within the first 2 h. Since scientific knowledge is lacking in the employment of PBR for the production of DAG-enriched oil *via* enzyme-catalysed partial hydrolysis, the findings of the study would facilitate the design of a pilot-scale fixed bed reactor system for lipase-mediated partial hydrolysis to obtain DAG-enriched oil as functional oil without constraints.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

PEMBANGUNAN PROSES BAGI PENGHASILAN MINYAK DIASILGLISERIDA YANG TINGGI MELALUI HIDROLISIS SEPARA

Oleh

PHUAH ENG TONG

Disember 2015

Pengerusi: Lai Oi-Ming, PhD
Fakulti: Institut Biosains

Penemuan minyak diasilgliserida (DAG) untuk menggantikan minyak makan konvensional telah mendapatkan perhatian di kalangan penyelidik dan pengeluar makanan disebabkan oleh sifat-sifat anti-obesiti minyak tersebut. Pelbagai kaedah pemprosesan telah dicadangkan untuk menghasilkan minyak DAG di mana kaedah hidrolisis separa dengan menggunakan enzim lipase bersefahaman dengan cara-cara pemprosesan lain kerana kaedah tersebut melibatkan bahan-bahan mentah yang murah dan memerlukan satu langkah hidrolisis sahaja. Dalam projek ini, kaedah hidrolisis separa dioptimumkan dalam sistem batch bagi menghasilkan DAG daripada minyak kelapa sawit yang telah ditapis, diluntur warna dan dinyahbau (RBDPO) dengan menggunakan lipase tersekat-gerak daripada *Rhizomucor miehei* (Lipozyme RMIM). Kesan empat parameter operasi iaitu suhu, dos enzim, kandungan air dan kelajuan pergolakan telah dikaji. Keadaan penghasilan DAG yang optimum adalah seperti berikut: suhu = 55°C, dos enzim = 10-wt%, kandungan air = 5-wt% and kelajuan pergolakan = 500 rpm. Sebanyak 31-wt% DAG dapat dihasilkan selepas 6 jam. Tindak balas hidrolisis separa didapati mematuhi mekanisme Ping-Pong Bi-Bi dengan perencatan substrat. Keadaan operasi optimum kemudiannya digunakan dalam reaktor lapisan terpadat berskala makmal.

Reaktor lapisan terpadat (PBR) merupakan reaktor yang efisien kerana reaktor tersebut membolehkan kebolegunaan enzim lipase selain meningkatkan kestabilan operasinya. Walau bagaimanapun, fenomena pemindahan jisim dalam PBR telah menjadi cabaran utama terutamanya dalam reaktor berskala besar. Oleh itu, model matematik tidak berdimensi bagi pemindahan jisim iaitu faktor Colburn, J_D yang merupakan gabungan nombor Reynolds (Re) dan nombor Schmidt (Sc) telah dicadangkan bagi mensimulasikan pemindahan

jisim dalam sistem PBR semasa reaksi hidrolisis separa berlaku. Model ini telah diperiksa dengan pelbagai nilai n dan keputusan menunjukkan bahawa korelasi pemindahan jisim $J_D = 0.92(Re)^{-0.2}$ dapat meramalkan data eksperimen dengan tepat. Di samping itu, kaedah gerak balas permukaan (RSM) telah digunakan untuk mengoptimumkan kedua-dua pembolehubah tidak bersandar iaitu ketinggian lapisan turus dan kadar aliran substrat bagi memaksimumkan hasil DAG dalam sistem PBR. Model kuadratik digunakan untuk mewakili kedua-dua pembolehubah bergerak balas iaitu DAG(y) dan triasilgliserida (TAG) yang tidak bertindak balas ($_{(un)}$ TAG) dengan kekurangan penyesuaian yang tidak ketara ($P > 0.05$). Keadaan optimum untuk mensintesis DAG adalah 10 cm ketinggian lapisan turus dan 3.8 ml/min kadar aliran substrat dengan 29-wt% DAG dilaporkan. Lipase tersekat-gerak boleh digunakan semula sehingga 10 kali tanpa kehilangan aktiviti enzim yang ketara.

Selain itu, kecekapan penghasilan DAG dengan menggunakan ruangan lapisan turus berbeza yang mempunyai nisbah tinggi lapisan turus kepada diameter dalaman ruangan lapisan turus (nisbah L/D) yang berbeza telah dikaji bagi menentukan ruangan lapisan turus yang paling berpotensi untuk menghasilkan DAG di industri. Isu-isu praktikal seperti suhu operasi, kadar aliran substrat dan masa tindak balas telah dinilai bagi setiap konfigurasi ruang padat. Dimensi ruangan dengan L/D nisbah dua merupakan reka bentuk ruang lapisan turus yang paling sesuai untuk reaksi hidrolisis separa yang dimungkinkan oleh enzim lipase. Keadaan optimum bagi menghasilkan DAG adalah 55°C suhu operasi, 5.0 ml/min kadar aliran substrat dan 5.8 min masa kediaman dalam PBR. Di bawah keadaan operasi tersebut, sebanyak 35-wt% DAG dapat diperolehi dalam tempoh 2 jam pertama. Oleh kerana pengetahuan saintifik tentang penggunaan sistem PBR untuk menghasilkan minyak DAG melalui hidrolisis separa dengan bantuan enzim masih kurang, hasil penyelidikan ini dipercayai akan memudahkan kerja perekabentuk reaktor berskala besar untuk memproses minyak DAG melalui kaedah hidrolisis separa tanpa kekangan.

ACKNOWLEDGEMENTS

I would like to express his deepest gratitude and appreciation to Prof Dr Lai Oi-Ming of the Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia for her supervision and constant support. I would also like to acknowledge his supervisory committee members, Prof Dr Thomas Choong Shean-Yaw of the Faculty of Engineering, Universiti Putra Malaysia, Prof Dr Tan Chin-Ping of the Faculty of Food Science and Technology, Universiti Putra Malaysia and Dr Lo Seong-Koon of the Wilmar (Shanghai) Biotechnology Research & Development Centre for their guidance and stimulating suggestions.

I would also like to extend his appreciation to his team mates, Mr Tang Teck-Kim and Ms Lee Yee-Ying for their assistance in the laboratory experiment and manuscripts preparation. Last but not least, I appreciatively thanks Mr Vijay Krishnan and Ms Sri Nesa from Sime Darby, Banting for their kind technical assistance. Financial support of this work by Sime Darby Research Sdn. Bhd. is also gratefully acknowledged.

I certify that a Thesis Examination Committee has met on (date of viva voce) to conduct the final examination of (student's name) on his (her) thesis entitled ("Title of Thesis") in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the (insert the name of relevant degree).

Members of the Thesis Examination Committee were as follows:

Name of Chairperson, PhD

Title (e.g., Professor/Associate Professor/Ir; omit if irrelevant)

Name of Faculty

Universiti Putra Malaysia

(Chairman)

Name of Examiner 1, PhD

Title (e.g., Professor/Associate Professor/Ir; omit if irrelevant)

Name of Faculty

Universiti Putra Malaysia

(Internal Examiner)

Name of Examiner 2, PhD

Title (e.g., Professor/Associate Professor/Ir; omit if irrelevant)

Name of Faculty

Universiti Putra Malaysia

(Internal Examiner)

Name of External Examiner, PhD

Title (e.g., Professor/Associate Professor/Ir; omit if irrelevant)

Name of Department and/or Faculty

Name of Organisation (University/Institute)

Country

(External Examiner)

(Insert name of current Deputy Dean)

(E.g. XXXXX XXXX, PhD)

Professor and Deputy Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Lai Oi Ming, PhD

Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Chairman)

Thomas Choong Shean Yaw, PhD

Professor, Ir
Faculty of Engineering
Universiti Putra Malaysia
(Member)

Tan Chin Ping, PhD

Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Member)

Lo Seong Koon, PhD

Biotechnology Research & Development Centre
Wilmar International Ltd. (Shanghai)
(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____
Name of Chairman
of Supervisory
Committee: Professor Dr. Lai Oi Ming

Signature: _____
Name of Member of
Supervisory
Committee: Professor Dr. Thomas Choong Shean Yaw

Signature: _____
Name of Member of
Supervisory
Committee: Professor Dr. Tan Chin Ping

Signature: _____
Name of Member of
Supervisory
Committee: Dr. Lo Seong Koon

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiii
LIST OF FIGURES	xv
LIST OF ABBREVIATIONS	xix
 CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	4
2.1 Diacylglycerol	4
2.1.1 Introduction	4
2.2.1 Nutrition and health effects	5
2.2 Chemical route for diacylglycerol production	7
2.3 Enzymatic route for diacylglycerol synthesis	12
2.3.1 Enzymatic esterification	12
2.3.2 Enzymatic glycerolysis	20
2.3.3 Enzymatic partial hydrolysis	29
2.4 Types of reactor	35
2.4.1 Batch stirred tank reactor	35
2.4.2 Continuous stirred tank reactor	35
2.4.3 Bubble column reactor	36
2.4.4 Packed bed reactor	36
2.4.5 Fluidized bed reactor	38
2.4.6 Membrane reactor	39
2.5 Response surface methodology	40
2.6 Mass transfer phenomena	42
3 OPTIMISATION AND KINETIC STUDY ON PARTIAL HYDROLYSIS OF PALM OIL CATALSED BY RHIZOMUCOR MIEHEI LIPASE	44
3.1 Introduction	44
3.2 Materials and methods	45
3.2.1 Materials	45
3.2.2 Methods	46
3.2.2.1 Optimal conditions for partial hydrolysis	46
3.2.2.2 Effect of enzyme particle suspension	46
3.2.2.3 Analysis of diacylglycerol content by RP-HPLC	46
3.2.2.4 Rheology	47

	3.2.2.5 Microscopic evaluation of enzyme particles and shear effect	47
	3.2.2.6 Statistical analysis	48
3.3	Results and discussion	48
3.3.1	Optimization of lipase RMIM-catalysed partial hydrolysis for DAG production	48
	3.3.1.1 Effect of speed rate	48
	3.3.1.2 Effect of enzyme load	51
	3.3.1.3 Effect of temperature	52
	3.3.1.4 Effect of water content	54
3.3.2	Viscosity and rheological properties	55
3.3.3	Surface morphology of Lipozyme RMIM	56
3.3.4	Mass transfer study	57
3.3.5	Kinetic study of lipase RMIM-catalysed partial hydrolysis	58
3.4	Summary	60
4	OPTIMISATION OF LIPASE-CATALYSED PARTIAL HYDROLYSIS FOR DIACYLGLYCEROL PRODUCTION IN PACKED BED REACTOR VIA RESPONSE SURFACE METHODOLOGY	61
4.1	Introduction	61
4.2	Materials and methods	62
4.2.1	Materials	62
4.2.2	Experimental design	62
4.2.3	Partial hydrolysis reaction in packed bed reactor	62
4.2.4	Analysis of diacylglycerol content	65
4.2.5	Mass transfer studies	65
4.3	Results and discussion	66
4.3.1	Model fitting and statistical analysis	66
4.3.2	Main effects and interaction between parameters	70
4.3.3	Optimization of partial hydrolysis and model verification	74
4.3.4	Stability of immobilized lipase in packed bed reactor	76
4.3.5	Mass transfer investigation	76
4.4	Summary	78
5	MASS TRANSFER STUDIES ON SOLVENT-FREE LIPASE-CATALYSED PARTIAL HYDROLYSIS OF PALM OIL IN PACKED BED REACTOR	79
5.1	Introduction	79
5.2	Materials and methods	80
5.2.1	Materials	80
5.2.2	Lipase-catalysed partial hydrolysis in packed bed reactor	80
5.2.3	Analysis of diacylglycerol content	81
5.2.4	External mass transfer model description	81
5.3	Results and discussion	87

5.4	Summary	92
6	EFFECTS OF BED COLUMN DIMENSION ON LIPASE-CATALYSED PARTIAL HYDROLYSIS OF PALM OIL FOR DIACYLGLYCEROL PRODUCTION IN PACKED BED REACTOR	93
6.1	Introduction	93
6.2	Materials and methods	94
6.2.1	Materials	94
6.2.2	Lipase-catalysed partial hydrolysis in packed bed reactor	95
6.2.3	Analysis of diacylglycerol content	95
6.2.4	Statistical analysis	95
6.3	Results and discussion	96
6.3.1	Effect of temperature	96
6.3.2	Effect of substrate flow rate	100
6.3.3	Effect of residence time / linear flow rate	104
6.4	Summary	107
7	CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	108
	REFERENCES	110
	APPENDICES	122
	BIODATA OF STUDENT	123
	LIST OF PUBLICATIONS	124

LIST OF TABLES

Table	Page
2.1 Relative contribution of acylglycerides in chosen edible oils	4
2.2 Enzymatic esterification reaction for diacylglycerol (DAG) production	14
2.3 Enzymatic glycerolysis reaction for diacylglycerol (DAG) production	21
2.4 Enzymatic partial hydrolysis reaction for diacylglycerol (DAG) production	30
3.1 Results of ANOVA for different reaction parameters on initial reaction rate	49
3.2 Apparent viscosity and activation energy for the reaction mixture before and after enzymatic reaction under optimum conditions	56
4.1 Independent variables: coded and real value in center composite rotatable design (CCRD)	62
4.2 Summary from experimental studies conducted in batch system	63
4.3 Experimental design results for partial hydrolysis of palm oil	67
4.4 ANOVA table for DAG(y) and (un)TAG	68
4.5 Regression coefficients and P-values for DAG(y) wt%	70
4.6 Regression coefficients and P-values for (un)TAG wt%	72
4.7 Optimization of partial hydrolysis and model verification by Chi-squared test	75
4.8 Weight percentage of DAG(y) and (un)TAG in ten continuous productions	76
5.1 Summary from experimental studies conducted in packed bed system	81

5.2	Observed reaction rate constants (k_p) at different flow rates	88
5.3	The percentage deviation of calculated values of k_p from the experimental values at different n	91
5.4	Effects of external mass transfer and overall substrate utilization rate in enzyme particle on the apparent reaction rate	91
6.1	Column length, column inner diameter and length-to-diameter (L/D) ratio for different bed column dimensions	95
6.2	Production of DAG using packed bed reactor with different sizes	105
6.3	Calculated Sherwood Number (Sh) for lipase-catalyzed partial hydrolysis in three columns with different column length-to-inner diameter (L/D) ratios	106

LIST OF FIGURES

Figure		Page
2.1	Illustration of chemical structure of different isomeric forms of diacylglycerol. Adapted from Lo et al. (2008).	5
2.2	Illustration of metabolism pathways for (A) Triacylglycerol (TAG) and (B) Diacylglycerol (DAG): DGAT, diacylglycerol acyltransferase; FFA, free fatty acid; 2-MAG, 2-monoacylglycerol; 1(3)-MAG, 1-monoacylglycerol or 3-monoacylglycerol; MGAT, monoacylglycerol acyltransferase; MTP, microsomal triglyceride transfer protein. Adapted from Yanai et al. (2007).	6
2.3	Illustration of chemical glycerolysis for diacylglycerol (DAG) and monoacylglycerol (MAG) production in industrial scale. Adapted from Sonntag (1982).	10
2.4	Schematic representation of the Ping-Pong Bi-Bi mechanism for lipase-catalysed esterification reaction: E: enzyme (lipase); FFA: free fatty acid; G: glycerol; H ₂ O: water; DAG: diacylglycerol; E.FFA: binary complex of lipase and FFA; E*.H ₂ O: intermediate complex of lipase and water; E*: modified enzyme complex; E*.G: binary complex of lipase and glycerol; E.DAG: intermediate complex of lipase and DAG; E-G: binary complex of lipase and glycerol. Adapted from Duan et al. (2010).	20
2.5	Schematic representation of the lipase-catalysed glycerolysis which follows a series of glycerolysis, hydrolysis and esterification reactions: FFA: free fatty acid; G: glycerol; H ₂ O: water; TAG: triacylglycerol; DAG: diacylglycerol; MAG: monoacylglycerol. Adapted from Valério et al. (2009).	26
2.6	Schematic representation of ordered-sequential Bi-Bi mechanism for lipase-catalysed glycerolysis reaction: E: enzyme (lipase); FFA: free fatty acid; G: glycerol; H ₂ O: water; TAG: triacylglycerol; DAG: diacylglycerol; MAG: monoacylglycerol; A x B x C: enzyme-substrate complex. . Adapted from Voll et al. (2011).	28
2.7	Schematic representation of the lipase-catalysed partial hydrolysis which follows a series of hydrolysis or re-esterification and acyl migration reactions. Adapted from Wang et al. (2010).	33

2.8	Schematic representation of the lipase-catalysed partial hydrolysis which follows ordered-sequential Bi-Bi mechanism considering a sequence of hydrolysis or re-esterification, solubility of water in oil phase and inhibition effect of high concentration of enzyme: E: enzyme (lipase); FFA: free fatty acid; G: glycerol; H ₂ O: water; H ₂ O ^{ins} : concentration of water not solubilised in the oil phase; H ₂ O ^t : total water concentration in the system; TAG: triacylglycerol; DAG: diacylglycerol; MAG: monoacylglycerol; A x B x C: enzyme-substrate complex. Adapted from Voll et al. (2012).	34
2.9	Schematic illustration of the lipase-catalysed hydrolysis and esterification reaction in membrane reactor (MR). Adapted from van der Padt (1993).	41
2.10	Illustration of mass transport and chemical reactions interactions in a symmetric slab of immobilised enzyme. The mass transfer phenomena is summarized in the top section and the parameters indicated in the bottom section can be used to describe quantitatively the mass transfer effects. Adapted from Bailey and Ollis (1986).	43
3.1	Effect of the speed rate on the initial rate based on DAG released. Reactions were carried out with 5% (w/w) water and 10% (w/w) Lipozyme RMIM at 55°C.	50
3.2	Volume fraction of the enzyme particles of the sample taken at the middle sampling point of the flask where ϕ_s is the average solid fraction.	50
3.3	Effect of the enzyme concentration on the initial rate based on DAG released. Reactions were carried out with 5% (w/w) water with speed rate of 500 rpm at 55°C.	52
3.4	Effect of the temperature on the initial rate based on DAG released. Reactions were carried out with 5% (w/w) water and 10% (w/w) Lipozyme RM IM with speed rate of 500 rpm.	53
3.5	Relationship between initial reaction rates based on DAG synthesis and reaction temperature (K) during partial hydrolysis of palm oil.	54
3.6	Effect of the water concentration on the initial rate based on DAG released. Reactions were carried out with 10% (w/w) Lipozyme RMIM with speed rate of 500 rpm at 55°C.	55

3.7	Light microscopic evaluation of Lipozyme RM IM particles under speed rate effect: (A) Enzyme particles after reaction at 100 rpm. (B) Enzyme particles after reaction at 500 rpm.	57
3.8	Comparison between the modelled (—) and the experimental data (■) based on the initial reaction rate based on DAG synthesis. Reactions were carried out with 5% (w/w) water and 10% (w/w) Lipozyme RMIM with speed rate of 500 rpm at 55°C.	59
4.1	Diagram of packed-bed reactor used for enzymatic partial hydrolysis reaction.	64
4.2	Predicted versus actual ($\text{DAG}_{(y)}$).	69
4.3	Predicted versus actual ($_{(un)}\text{TAG}$).	70
4.4	Three dimension response surface contour plot indicating the effect of interaction between packed bed height (A) and substrate flow rate (B) on $\text{DAG}_{(y)}$.	72
4.5	Three dimension response surface contour plot indicating the effect of interaction between packed bed height (A) and substrate flow rate (B) on $_{(un)}\text{TAG}$.	73
4.6	Variation in packed bed height and substrate flow rate for (A) reaction limited model and (B) mass transfer limited model.	77
5.1	Diacylglycerol content as a function of time at different substrate flow rates: (a) 1 ml min^{-1} (—■—); (b) 3 ml min^{-1} (—■—); (c) 5 ml min^{-1} (—■—); (d) 10 ml min^{-1} (—■—) and (e) 15 ml min^{-1} (—■—).	87
5.2	Overall rate of reaction at different flow rates: (a) 1 ml min^{-1} (—■—); (b) 3 ml min^{-1} (—■—) and (c) 5 ml min^{-1} (—■—).	88
5.3	Plots of $1/k_p$ against $1/G^n$ for (a) $n = 0.1$; (b) $n = 0.3$; (c) $n = 0.5$; (d) $n = 0.7$; (e) $n = 0.8$ and (f) $n = 0.9$.	90
6.1	Effects of temperatures on the TAG concentration and DAG yield produced by enzymatic partial hydrolysis as a function of reaction time for column dimension of seven: (a) 45°C (—■—); (b) 55°C (—■—); (c) 65°C (—■—) and (d) 75°C (—■—). The reaction was performed at substrate flow rate of 3 ml min^{-1} and water content of 5-wt% based on oil weight.	97

- 6.2 Effects of temperatures on the TAG concentration and DAG yield produced by enzymatic partial hydrolysis as a function of reaction time for column dimension of two: (a) 45°C (—◆—); (b) 55°C (—■—); (c) 65°C (—▲—) and (d) 75°C (—✕—). The reaction was performed at substrate flow rate of 3 ml min⁻¹ and water content of 5-wt% based on oil weight. 98
- 6.3 Effects of temperatures on the TAG concentration and DAG yield produced by enzymatic partial hydrolysis as a function of reaction time for column dimension of one: (a) 45°C (—◆—); (b) 55°C (—■—); (c) 65°C (—▲—) and (d) 75°C (—✕—). The reaction was performed at substrate flow rate of 3 ml min⁻¹ and water content of 5-wt% based on oil weight. 99
- 6.4 Effects of substrate flow rate on the TAG concentration and DAG yield produced by enzymatic partial hydrolysis as a function of reaction time for column dimension of seven: (a) 1 ml min⁻¹ (—■—); (b) 3 ml min⁻¹ (—▲—); (c) 5 ml min⁻¹ (—✕—); (d) 10 ml min⁻¹ (—◆—) and (e) 15 ml min⁻¹ (—●—). The reaction was performed at reaction temperature of 55°C and water content of 5-wt% based on oil weight. 101
- 6.5 Effects of substrate flow rate on the TAG concentration and DAG yield produced by enzymatic partial hydrolysis as a function of reaction time for column dimension of two: (a) 1 ml min⁻¹ (—■—); (b) 3 ml min⁻¹ (—▲—); (c) 5 ml min⁻¹ (—✕—); (d) 10 ml min⁻¹ (—◆—) and (e) 15 ml min⁻¹ (—●—). The reaction was performed at reaction temperature of 55°C and water content of 5-wt% based on oil weight. 102
- 6.6 Effects of substrate flow rate on the TAG concentration and DAG yield produced by enzymatic partial hydrolysis as a function of reaction time for column dimension of one: (a) 1 ml min⁻¹ (—■—); (b) 3 ml min⁻¹ (—▲—); (c) 5 ml min⁻¹ (—✕—); (d) 10 ml min⁻¹ (—◆—) and (e) 15 ml min⁻¹ (—●—). The reaction was performed at reaction temperature of 55°C and water content of 5-wt% based on oil weight. 103

LIST OF ABBREVIATIONS

E_a	Activation energy
[TOMA].[Tf2N]	trioctylmethylammonium bis
1(3)-MAG	(trifluoromethylsulphonyl)imide
2M2B	1-monoacylglycerol or 3-monoacylglycerol
2-MAG	2-methyl-2-butanol
A x B x C	2-monoacylglycerol
AAD	Enzyme-substrate complex
	Absolute average deviation
	Cocosalkyl pentaethoxy methyl ammonium
Ammoeng 102	methylsulfate
ANOVA	One-way analysis of variance
AOT	Sodium (bis-2-ethyl-hexyl) sulfosuccinate
BCR	Bubble column reactor
BR	Batch reactor
CA	Caprylic acid
CCRD	Centre composite rotatable design
CO ₂	Carbon dioxide
CSTR	Continuous stirred tank reactor
DA	Capric acid
DAG	Diacylglycerol
DGAT	Diacylglycerol acyltransferase
E	Enzyme
E*	Modified enzyme complex
ELSD	Evaporative light scattering detector
EPA	Eicosapentaenoic acid
FBR	Fluidized bed reactor
FDA	Food and Drug Administration
FFA	Free fatty acid
G	Glycerol
GRAS	Generally recognized as safe
H ₂ O	Water
HPLC	High performance liquid chromatography
IL	Ionic liquid
L/D ratio	Column length-to-inner diameter ratio
LA	Lauric acid
Lipozyme RMIM	<i>Rhizomucor miehei</i> lipase
Lipozyme TLIM	<i>Thermomyces lanuginosus</i> lipase
LOA	Linoleic acid
MAG	Monoacylglycerol
ME	Methyl esters
MGAT	Monoacylglycerol acyltransferase
MR	Membrane reactor
MTBE	<i>tert</i> -butyl methyl ether
MTP	Microsomal triglyceride transfer protein
OA	Oleic acid
PA	Palmitic acid
PBR	Packed bed reactor

PODD	Palm oil deodorized distillate
PUFA	Polyunsaturated fatty acid
RBDPO	Refined, bleached, and deodorized palm oil
Re	Reynolds number
ROFA	Rapeseed oil fatty acids
RSM	Response surface methodology
Sc	Schmidt number
SC-CO ₂	Supercritical carbon dioxide
TAG	Triacylglycerol
WHO	World Health Organization
ΔP	Pressure drop



CHAPTER 1

INTRODUCTION

Dietary fats and oils are known to be the key nutrients essential for sustaining life. In addition to providing energy for daily activities, the roles of lipids as fundamental building blocks for healthy cells, carriers for fat soluble vitamins, organ protector and body insulator are well documented. Moreover, the incorporation of dietary fats into food enhances its sensory and textural properties. However, strong evidence demonstrates that increased dietary energy intake especially fat-dense food coupled with inadequate physical activities is the main culprit that leads to the overwhelming incidences of obesity (Golay & Bobbioni, 1997; Astrup, 2005). Recent reports reveal that obesity prevalence rates are rising at an alarming rate and remain to be a critical global epidemic (WHO 2014). Tremendous research studies indicate a strong positive correlation between obesity and adverse health effects such as heart disease, cancer, diabetes mellitus, hypertension besides mental trauma and physical discomfort (Lavie et al. 2009; Artham et al. 2011; Louie et al. 2013). The consequences are potential decline in life expectancy, early retirement, widespread discrimination and increased cost of health care system that burdens the government and economic growth. In United States, obesity-related medical expenses have seen a drastic increase over the years with USD 86 billion of aggregate health care cost being recorded in year 2008 (Finkelstein et al. 2009). A recent published research report also indicates that the prevalence of obesity is rather severe in Malaysia as compared to other Asian countries with 45.3% of its population being overweight and the obese population is forecasted to skyrocket in the next decade (Ng et al. 2014). Although World Health Organization (WHO) advises and limits the dietary fats consumption to 30% of the total calorie intake in order to impart positive effects on human health, the recommended action may sacrifice the mouthfeel quality in fat-based food.

With heightened health consciousness, the disclosure of functional diacylglycerol (DAG)-oil has therefore drawn increasing attention of researchers and food manufacturers to replace the conventional edible oil or triacylglycerol (TAG) oil. Previous literatures clearly pointed out that DAG-enriched oil is capable of inhibiting the accumulation of visceral fat and suppressing the blood serum TAG besides increasing the rate of β -oxidation of fatty acids which translates into potent anti-obesity properties (Flickinger & Matsuo, 2003; Teramoto et al., 2004). Apart from that, DAG with exposing hydrophilic group within the molecular structure, exhibits excellent emulsifying capability and has been widely used as emulsifier together with monoacylglycerol (MAG) in food, cosmetic and pharmaceutical products (Shimada & Ohashi, 2003; Masui et al., 2001; Nakajima, 2004). Foreseeing the increasing demand for DAG-oil, Kao Corporation (Japan) began commercializing the functional edible oil under product name of "Healthy Econa Cooking Oil" in early 1999. Sales of this functional edible oil accounts for 80%

of premium oil which constitute around 14% of the total Japanese edible oil market worth ¥10 billion (Sakaguchi, 2001).

Strategy for DAG-oil production includes both chemical and enzyme-assisted approaches, in which the latter exhibits several advantages over chemical method namely, reduced energy consumption, improved selectivity and yield. Enzymatic partial hydrolysis reaction outperforms other methods because of the low cost reactants and single hydrolytic step involved (Lai et al. 2006; Cheong et al. 2007; Lo et al, 2008). Malaysia, being one of the largest palm oil producer and exporter, contributes nearly 17.7 million tonnes, accounting for 11% of the global fats and oils production and becomes a dominant player in the palm oil trade with 44% of the market share (MPOC 2014). To broaden the commercial use and functionality of palm oil in order to stay competitive in edible oils and fats market, production of DAG-enriched oil from conventional palm oil *via* lipase-catalysed partial hydrolysis is indeed a necessity.

To date, literature on the kinetic study of partial hydrolysis reaction and production of palm-based DAG is limited. As such, various operating parameters namely temperature, agitation speed, water content and enzyme load for the production of palm-based DAG in batch stirred reactor system were investigated in present work. In addition, this work aimed to develop a kinetic model to describe the reaction mechanism of partial hydrolysis as well as to provide information on the optimum processing conditions. Analysis of reaction kinetics has great potential because the mathematical model generated are capable of simulating the complex reaction under different conditions and thereby improving the reaction conditions (Fedosov et al. 2013). Although batch stirred reactor could be used to produce DAG *via* enzyme-catalysed partial hydrolysis, abrasion of the matrix particles under mechanical stirring force should be paid attention. The optimum conditions determined were then applied to packed bed reactor (PBR) system owing to its higher reaction rate and enhanced stability of particulate catalysts in PBR system (Phuah et al. 2015). Two important operating variables namely packed bed height and substrate flow rate were evaluated and optimized by response surface methodology (RSM). The application of RSM as a statistical techniques based on the fit of a polynomial model to the experimental data enables evaluation of the effects of multiple operating parameters, alone or in combination, on response variables and prediction of reaction performance accurately (Xu et al. 1998).

Although PBR is a preferred bioreactor configuration, dominance of external mass transfer resistance during enzymatic reaction remains to be major hindrance for fixed bed system especially at large scale. External mass transport limitation exists when the rate of diffusional transport of substrate through the external film of the enzyme particles is the rate determining step which is caused by low substrate flow rate (Chew et al. 2008; McCabe et al. 2005; Kasaini & Mbaya 2009; Murty et al. 2005). Therefore, the mass transfer phenomena in PBR were investigated and represented with external mass transfer model. The development of the mathematical model would enable

evaluation of mass transfer coefficients in fixed bed system under different operating conditions. The effects of different bed column designs with distinct column length-to-inner diameter ratios (L/D ratio) were also studied as the column dimensions determine both linear fluid flow rate and external mass transfer coefficient even with constant residence time and substrate flow rate, thereby affecting the efficiency of the packed bed system. In summary, the objectives of this study were as follows:

1. To optimize the reaction parameters for the production of DAG-oil in batch reactor and to evaluate its kinetic mechanism.
2. To optimize the operating parameters on the production yield of DAG in packed bed reactor.
3. To develop a mass transfer model to predict the reactor performance and to simulate the partial hydrolysis reaction in packed bed system.
4. To evaluate the effects of different bed column designs on the production efficiency of DAG.

REFERENCES

- Abro, S., Pouilloux, Y., & Barrault, J. (1997). Selective synthesis of monoglycerides from glycerol and oleic acid in the presence of solid catalysts. In A. B. H.U. Blaser, & R. Prins (Eds.), *Studies in Surface Science and Catalysis* (Vol. Volume 108, pp. 539-546): Elsevier.
- Akoh, C. C. (1998). Fat replacers. *Food Technology*, 52(3), 47 – 53.
- Akoh, C. C., & Decker E. A. (1995). Lipid-based fat substitutes. *Critical Reviews in Food Science and Nutrition*, 35(5), 405 – 430.
- Arcos, J. A., & Otero, C. (1996). Enzyme, medium, and reaction engineering to design a low-cost, selective production method for mono- and dioleoylglycerols. *Journal of the American Oil Chemists' Society*, 73(6), 673-682.
- Arcos, J. A., Otero, C., & Hill, C. G., Jr. (1998). Rapid enzymatic production of acylglycerols from conjugated linoleic acid and glycerol in a solvent-free system. *Biotechnology Letters*, 20(6), 617-621.
- Artham, S., Lavie, C., De Schutter, A., Ventura, H., & Milani, R. (2011). Obesity, Age, and Cardiac Risk. *Current Cardiovascular Risk Reports*, 5(2), 128-137.
- Astrup, A. (2005). The role of dietary fat in obesity. *Seminars in Vascular Medicine*, 5(1), 40 – 47.
- Awadallak, J. A., Voll, F., Ribas, M. C., da Silva, C., Filho, L. C., & da Silva, E. A. (2013). Enzymatic catalyzed palm oil hydrolysis under ultrasound irradiation: Diacylglycerol synthesis. *Ultrasonics Sonochemistry*, 20(4), 1002-1007.
- Babicz, I., Leite, S. G. F., de Souza, R. O. M. A., & Antunes, O. A. C. (2010). Lipase-catalyzed diacylglycerol production under sonochemical irradiation. *Ultrasonics Sonochemistry*, 17(1), 4-6.
- Bailey, J. E., & Ollis, D. F. (1986). Biochemical engineering fundamentals. New York, USA, McGraw-Hill.
- Balcão, V. M., Paiva, A. L., & Xavier Malcata, F. (1996). Bioreactors with immobilized lipases: state of the art. *Enzyme and Microbial Technology*, 18 (6) 392 – 416.
- Baş, D., & Boyacı, İ. H. (2007). Modeling and optimization I: Usability of response surface methodology. *Journal of Food Engineering*, 78(3), 836 – 845.
- Beg, Q. K., Saxena, R. K., & Gupta, R. (2002). Kinetic constants determination for an alkaline protease from *Bacillus mojavensis* in a bioreactor. *Process Biochemistry*, 39, 203-209.
- Bellot, J. C., Choisnard, L., Castillo, E., & Marty, A. (2001). Combining solvent engineering and thermodynamic modeling to enhance selectivity during monoglyceride synthesis by lipase-catalyzed esterification. *Enzyme and Microbial Technology*, 28(4–5), 362-369.
- Berger, M., Laumen, K., & Schneider, M. (1992). Enzymatic esterification of glycerol I. Lipase-catalyzed synthesis of regioisomerically pure 1,3-sn-diacylglycerols. *Journal of the American Oil Chemists' Society*, 69(10), 955-960.
- Blanch, H. W., & Clark, D. S. (1997). Biochemical Engineering, New York, USA, Marcel Dekker, Inc.

- Calderbank, P. H. (1958). Physical rate processes in industrial fermentation. Part I: The interfacial area in gas-liquid contacting with mechanical agitation. *Transactions of IChemE, Part A, Chemical Engineering Research and Design*, 36, 443 – 463.
- Cheirsilp, B., Kaewthong, W., & H-Kittikun, A. (2007). Kinetic study of glycerolysis of palm olein for monoacylglycerol production by immobilized lipase. *Biochemical Engineering Journal*, 35(1), 71-80.
- Cheong, L.-Z., Tan, C.-P., Long, K., Affandi Yusoff, M. S., Arifin, N., Lo, S.-K., et al. (2007). Production of a diacylglycerol-enriched palm olein using lipase-catalyzed partial hydrolysis: Optimization using response surface methodology. *Food Chemistry*, 105(4), 1614-1622.
- Cheong, L.-Z., Zhang, H., Xu, Y., & Xu, X. (2009). Physical Characterization of Lard Partial Acylglycerols and Their Effects on Melting and Crystallization Properties of Blends with Rapeseed Oil. *Journal of Agricultural and Food Chemistry*, 57(11), 5020-5027.
- Chew, Y. H., Chua, L. S., Cheng, K. K., Sarmidi, M. R., Aziz, R. A., & Lee, C. T. (2008). Kinetic study on the hydrolysis of palm olein using immobilized lipase. *Biochemical Engineering Journal*, 39(3), 516 – 520.
- Chilton, T. H., & Colburn, A. P. (1934). Mass Transfer (Absorption) Coefficients Prediction from Data on Heat Transfer and Fluid Friction. *Industrial and Engineering Chemistry*, 26(11), 1183 – 1187.
- Chong, F., Tey, B., Dom, Z., Cheong, K., Satiawihardja, B., Ibrahim, M., et al. (2007). Rice bran lipase catalyzed esterification of palm oil fatty acid distillate and glycerol in organic solvent. *Biotechnology and Bioprocess Engineering*, 12(3), 250-256.
- Choo, Y. M., Puah, C. W., Ma, A. N., & Basiron, Y. (2006). Production of edible oil with high diglyceride content. European Patent no. EP20060250474 (in English).
- Coker, A. K., & Kayode, C. A. (2001). Modeling of Chemical Kinetics and Reactor Design, Houston, USA, Gulf Publishing Company.
- Coteron, A., Martinez, M., & Aracil, J. (1998). Reactions of olive oil and glycerol over immobilized lipases. *Journal of the American Oil Chemists' Society*, 75(5), 657-660.
- Damstrup, M. L., Abildskov, J., Kiil, S., Jensen, A. D., Sparsø, F. V., & Xu, X. (2006). Evaluation of Binary Solvent Mixtures for Efficient Monoacylglycerol Production by Continuous Enzymatic Glycerolysis. *Journal of Agricultural and Food Chemistry*, 54(19), 7113-7119.
- Dickey, D. S., & Fasano, J. B. (2004). Handbook of Industrial Mixing: Science and Practice, New Jersey, USA, John Wiley and Sons, Inc.
- Duan, Z.-Q., Du, W., & Liu, D.-H. (2010). Novozym 435-catalyzed 1,3-diacylglycerol preparation via esterification in t-butanol system. *Process Biochemistry*, 45(12), 1923-1927.
- Duan, Z.-Q., Fang, X.-L., Wang, Z.-Y., Bi, Y.-H., & Sun, H. (2015). Sustainable process for 1,3-diolein synthesis catalyzed by immobilized lipase from *Penicillium expansum*. *ACS Sustainable Chemistry and Engineering*, 3(11), 2804-2808.
- Fedosov, S. N., Brask, J., Pedersen, A. K., Nordblad, M. Woodley J. M., & Xu, X. (2013). Kinetic model of biodiesel production using immobilized lipase *Candida antarctica* lipase B. *Journal of Molecular Catalysis B: Enzymatic*, 85-86, 156-168.

- Feltes, M., Vladimir Oliveira, J., Treichel, H., Block, J., de Oliveira, D., & Ninow, J. (2010). Assessment of process parameters on the production of diglycerides rich in omega-3 fatty acids through the enzymatic glycerolysis of fish oil. *European Food Research and Technology*, 231(5), 701-710.
- Fiametti, K. G., Sychoski, M. M., Cesaro, A. D., Furigo Jr, A., Bretanha, L. C., Pereira, C. M. P., et al. (2011). Ultrasound irradiation promoted efficient solvent-free lipase-catalyzed production of mono- and diacylglycerols from olive oil. *Ultrasonics Sonochemistry*, 18(5), 981-987.
- Finkelstein, E. A., Trogon, J. G., Cohen, J. W., & Dietz, W. (2009). Annual Medical Spending Attributable To Obesity: Payer-And Service-Specific Estimates. *Health Affairs*, 28(5), 822-831.
- Flickinger, B., & Matsuo, N. (2003). Nutritional characteristics of DAG oil. *Lipids*, 38(2), 129-132.
- Fregolente, P. B. L., Pinto, G. M. F., Wolf-Maciel, M. R., & Filho, R. M. (2010). Monoglyceride and diglyceride production through lipase-catalyzed glycerolysis and molecular distillation. *Applied Biochemistry and Biotechnology*, 160(7), 1879 - 1887.
- Fureby, A. M., Tian, L., Adlercreutz, P., & Mattiasson, B. (1997). Preparation of diglycerides by lipase-catalyzed alcoholysis of triglycerides. *Enzyme and Microbial Technology*, 20 (3) 198 – 206.
- Garzon-Aburbeh, A., Poupaert, J. H., Claesen, M., & Dumont, P. (1986). A lymphotropic prodrug of L-dopa: synthesis, pharmacological properties and pharmacokinetic behavior of 1,3-dihexadecanoyl-2-[(S)-2-amino-3-(3,4-dihydroxyphenyl)propanoyl]propane-1,2,3-triol. *Journal of Medicinal Chemistry*, 29(5), 687-691.
- Garzon-Aburbeh, A., Poupaert, J. H., Claesen, M., Dumont, P., & Atassi, G. (1983). 1,3-Dipalmitoylglycerol ester of chlorambucil as a lymphotropic, orally administrable antineoplastic agent. *Journal of Medicinal Chemistry*, 26(8), 1200-1203.
- Giacometti, J., Giacometti, F., Milin, Č., & Vasić-Rački, Đ. a. (2001). Kinetic characterisation of enzymatic esterification in a solvent system: adsorptive control of water with molecular sieves. *Journal of Molecular Catalysis B: Enzymatic*, 11(4–6), 921-928.
- Golay, A. & Bobbioni, E. (1997). The role of dietary fat in obesity. *International Journal of Obesity and Related Metabolic Disorders: Journal of the International Association for the Study of Obesity*, 21(3), 2 – 11.
- Goldberg, M., Thomas, D., & Legoy, M.-D. (1990). The control of lipase-catalysed transesterification and esterification reaction rates. *European Journal of Biochemistry*, 190(3), 603-609.
- Gonçalves, J. C. S., Razzouk, C., Poupaert, J. H., & Dumont, P. (1989). High-performance liquid chromatography of chlorambucil prodrugs structurally related to lipids in rat plasma. *Journal of Chromatography B: Biomedical Sciences and Applications*, 494(0), 389-396.
- Gonçalves, K., Sutili, F., Leite, S., de Souza, R. O. M. A., & Leal, I. (2012). Palm oil hydrolysis catalyzed by lipases under ultrasound irradiation – The use of experimental design as a tool for variables evaluation. *Ultrasonics Sonochemistry*, 19(2), 232-236.

- Guedes de Carvalho, J. R. F., Delgado, J. M. P. Q., & Alves, M. A. (2004). Mass transfer between flowing fluid and sphere buried in packed bed of inerts. *American Institute of Chemical Engineers*, 50, 65 - 74.
- Gunstone, F. D. (1999). Enzymes as biocatalysts in the modification of natural lipids. *Journal of the Science of Food and Agriculture*, 79(12), 1535-1549.
- Guo, Z., & Sun, Y. (2007). Solvent-free production of 1,3-diglyceride of CLA: Strategy consideration and protocol design. *Food Chemistry*, 100(3), 1076-1084.
- Halim, S. F. A., Kamaruddin, A. H., & Fernando, W. J. N. (2009). Continuous biosynthesis of biodiesel from waste cooking palm oil in a packed bed reactor: Optimization using response surface methodology (RSM) and mass transfer studies, *Bioresource Technology*, 100 (2), 710 – 716.
- Hamam, F., & Budge, S. (2010). Structured and Specialty Lipids in Continuous Packed Column Reactors: Comparison of Production Using One and Two Enzyme Beds. *Journal of the American Oil Chemists' Society*, 87(4), 385 – 394.
- Ison, A. P., Macrae, A. R., Smith, C. G., & Bosley, J. (1994). Mass transfer effects in solvent-free fat interesterification reactions: influences on catalyst design. *Biotechnology and Bioengineering*, 43(2), 122 - 130.
- Jacobs, L., Lee, I., & Poppe, G. (2003). Chemical Process for the Production of 1,3-Diglyceride Oils. U.S. Patent 2003104109.
- Kahveci, D., Guo, Z., Özçelik, B., & Xu, X. (2010). Optimisation of enzymatic synthesis of diacylglycerols in binary medium systems containing ionic liquids. *Food Chemistry*, 119(3), 880-885.
- Kamphuis, M. M., Mela, D. J., & Westerterp-Plantenga, M. S. (2003). Diacylglycerols affect substrate oxidation and appetite in humans. *The American Journal of Clinical Nutrition*, 77(5), 1133-1139.
- Kantarci, N., Borak, F., & Ulgen, K. O. (2005). Bubble column reactors. *Process Biochemistry*, 40, 2263 - 2283.
- Kasaini, H & Mbaya R. K. (2009). Continuous adsorption of pt ions in a batch reactor and packed-bed column. *Hydrometallurgy*, 97, 111-118.
- Kathiravan, M. N., Rani, K. R., Karthick, R., & Muthukumar, K. (2010). Mass transfer studies on the reduction of Cr(VI) using calcium alginate immobilized *Bacillus* sp. in packed bed reactor. *Bioresource Technology*, 101(3), 853 – 858.
- Katsuragi, Y., Yasukawa, T., Matsuo, N., Flickinger, B. D., Tokimitsu, I., & Matlock, M. G. (2008). *Diacylglycerol Oil* (2nd ed.), Illinois, USA, AOCS Press.
- Klaewkla, R., Arend, M., & Hoelderich, W. F. (2011). A review of mass transfer controlling the reaction rate in heterogeneous catalytic systems. In H. Nakajima (Eds.), *Mass Transfer – Advanced Aspects* (pp. 667-684): InTech.
- Koizumi, Y., Mukai, K., Murakawa, K., & Yamane, T. (1987). Scale-up of Microporous Hydrophobic Membrane Bioreactor with respect to Continuous Glycerolysis of Fat by Lipase Bioreactor for Enzymic Reaction of Fat and Fatty Acid Derivatives, Part X. *Journal of Japan Oil Chemists' Society*, 36(8), 561-564.

- Kondo, H., Hase, T., Murase, T., & Tokimitsu, I. (2003). Digestion and assimilation features of dietary DAG in the rat small intestine. *Lipids*, 38(1), 25 – 30.
- Kosugi, Y., Tanaka, H., & Tomizuka, N. (1990). Continuous hydrolysis of oil by immobilized lipase in a countercurrent reactor. *Biotechnology and Bioengineering*, 36(6), 617-622.
- Kristensen, J., Xu, X., & Mu, H. (2005). Diacylglycerol synthesis by enzymatic glycerolysis: Screening of commercially available lipases. *Journal of the American Oil Chemists' Society*, 82(5), 329-334.
- Krüger, R. L., Valério, A., Balen, M., Ninow, J. L., Vladimir Oliveira, J., de Oliveira, D., et al. (2010). Improvement of mono and diacylglycerol production via enzymatic glycerolysis in tert-butanol system. *European Journal of Lipid Science and Technology*, 112(8), 921-927.
- Kwon, S. J., Han, J. J., & Rhee, J. S. (1995). Production and in situ separation of mono- or diacylglycerol catalyzed by lipases in n-hexane. *Enzyme and Microbial Technology*, 17(8), 700-704.
- Laddu, D., Dow, C., Hingle, M., Thomson, C., & Going, S. (2011). A Review of Evidence-Based Strategies to Treat Obesity in Adults. *Nutrition in Clinical Practice*, 26(5), 512 – 525.
- Lai, O. M., Yusoff, M. S. A., Lo, S. K., Long, K., Tan, C. P., Lim, J. Y., et al. (2006). Process for the production of diacylglycerol. PCT International Application No. PCT/MY2006/000034 (in English).
- Lai, O. M., Yusoff, M. S. A., Lo, S. K., Long, K., Tan, C. P., Tahiruddin, S., et al. (2007). Production of acylglycerol esters. PCT International Application No. PCT/MY2007/000025 (in English).
- Lavie, C. J., Milani, R. V., & Ventura, H. O. (2009). Obesity and Cardiovascular Disease: Risk Factor, Paradox, and Impact of Weight Loss. *Journal of the American College of Cardiology*, 53(21), 1925-1932.
- Li, D., Qin, X., Wang, J., Yang, B., Wang, W., Huang, W., & Wang, Y. (2015). Hydrolysis of soybean oil to produce diacylglycerol by a lipase from *Rhizopus oryzae*. *Journal of Molecular Catalysis B: Enzymatic*, 115, 43-50.
- Liao, H.-F., Tsai, W.-C., Chang, S.-W., & Shieh, C.-J. (2003). Application of solvent engineering to optimize lipase-catalyzed 1,3-diglycerols by mixture response surface methodology. *Biotechnology Letters*, 25(21), 1857-1861.
- Lin, S. W., & Yoo, C. K. (2009). Short-path distillation of palm olein and characterization of products. *European Journal of Lipid Science and Technology*, 111(2), 142-147.
- Liu, N., Wang, Y., Zhao, Q., Zhang, Q., & Zhao, M. (2011). Fast synthesis of 1,3-DAG by Lecitase® Ultra-catalyzed esterification in solvent-free system. *European Journal of Lipid Science and Technology*, 113(8), 973 - 979.
- Lo, S.-K., Baharin, B. S., Tan, C. P., & Lai, O. M. (2004). Diacylglycerols from Palm Oil Deodoriser Distillate. Part 1 – Synthesis by Lipase-catalysed Esterification. *Food Science and Technology International*, 10(3), 149 - 156.

- Lo, S.-K., Cheong, L.-Z., Arifin, N., Tan, C.-P., Long, K., Yusoff, M. S. A., et al. (2007). Diacylglycerol and Triacylglycerol as Responses in a Dual Response Surface-Optimized Process for Diacylglycerol Production by Lipase-Catalyzed Esterification in a Pilot Packed-Bed Enzyme Reactor. *Journal of Agricultural and Food Chemistry*, 55(14), 5595 – 5603.
- Lo, S.-K., Tan, C.-P., Long, K., Yusoff, M. S., & Lai, O.-M. (2008). Diacylglycerol Oil—Properties, Processes and Products: A Review. *Food and Bioprocess Technology*, 1(3), 223-233.
- Lortie, R., Trani, M., & Ergun, F. (1993). Kinetic study of the lipase-catalyzed synthesis of triolein. *Biotechnology and Bioengineering*, 41(11), 1021-1026.
- Louie, S. M., Roberts, L. S., & Nomura, D. K. (2013). Mechanisms linking obesity and cancer. *Biochimica et Biophysica Acta (BBA) - Molecular and Cell Biology of Lipids*, 1831(10), 1499-1508.
- Lue, B.-M., Guo, Z., & Xu, X. (2007). Lipid processing in ionic liquids. *Lipid Technology*, 19(9), 204-207.
- Maki, K. C., Davidson, M. H., Tsushima, R., Matsuo, N., Tokimitsu, I., Umporowicz, D. M., Dicklin, M. R., Foster, G. S., Ingram, K. A., Anderson, B. D., Frost, S. D. & Bell, M. (2002). Consumption of diacylglycerol oil as part of a reduced-energy diet enhances loss of body weight and fat in comparison with consumption of a triacylglycerol control oil. *American Journal of Clinical Nutrition*, 76 (6) 1230 – 1236.
- Malcata, F. X., Reyes, H. R., Garcia, H. S., Hill Jr, C. G., & Amundson, C. H. (1992). Kinetics and mechanisms of reactions catalysed by immobilized lipases. *Enzyme and Microbial Technology*, 14(6), 426 – 446.
- Martinez, C. E., Vinay, J. C., Brieva, R., Hill, C. G., Jr., & Garcia, H. S. (2005). Preparation of mono- and diacylglycerols by enzymatic esterification of glycerol with conjugated linoleic acid in hexane. *Applied Biochemistry and Biotechnology*, 125(1), 63-75.
- Marty, A., Dossat, V., & Condoret, J.-S. (1997). Continuous operation of lipase-catalyzed reactions in nonaqueous solvents: Influence of the production of hydrophilic compounds. *Biotechnology and Bioengineering*, 56(2), 232-237.
- Masui, K., Yasunaga, K., Nishide, T., Nakajima, Y. & Yasukawa, T. (2001). Physiochemical and frying characteristics of oil high in diacylglycerol. *Proceedings of the 2001 PIPOC International Palm Oil Congress*, Kuala Lumpur, 65 – 73.
- Matos, L. M. C., Leal, I. C. R., & de Souza, R. O. M. A. (2011). Diacylglycerol synthesis by lipase-catalyzed partial hydrolysis of palm oil under microwave irradiation and continuous flow conditions. *Journal of Molecular Catalysis B: Enzymatic*, 72(1–2), 36-39.
- McCabe, W. L., Smith, J. C., & Harriott, P. (2005). Unit operations of chemical engineering (7th ed.), New York, USA, McGraw-Hill.
- Meng, X. H., Sun, P. L., Yang, K., He, R. J., & Mao, Z. G. (2005). Synthesis of diacylglycerol using immobilised regiospecific lipase in continuously operated fixed bed reactors. *Chinese Journal of Biotechnology*, 21(3), 425 – 429.

- Monte Blanco, S. F. M., Santos, J. S., Feltes, M. M. C., Dors, G., Licodiedoff, S., Lerin, L. A., de Oliveira, D., Ninow, J. L., & Furigo Jr., A. (2015). Optimization of diacylglycerol production by glycerolysis of fish oil catalyzed by Lipozyme TLIM with Tween 65. *Bioprocess and Biosystems Engineering*, 38, 2379-2388.
- Mozammel Hoo, M., Yamane, T., Shimizu, S., Funada, T., & Ishida, S. (1984). Continuous synthesis of glycerides by lipase in a microporous membrane bioreactor. *Journal of the American Oil Chemists' Society*, 61(4), 776-781.
- MPOC (2014). Malaysian Palm Oil Industry. [http://www.mpoc.org.my/Malaysian Palm Oil Industry.aspx](http://www.mpoc.org.my/Malaysian_Palm_Oil_Industry.aspx) (Accessed 20 Sept 2014)
- Murty, V. R., Bhat, J., & Muniswaran, P. K. A. (2002). Hydrolysis of oils by using immobilized lipase enzyme: A review. *Biotechnology and Bioprocess Engineering*, 7(2), 57-66.
- Murty V. R., Bhat, J., & Muniswaran, P. K. A. (2004). Mass transfer effects in immobilized lipase bed reactor during hydrolysis of rice bran oil. *Chemical and Biochemical Engineering Quarterly*, 18(2), 177 – 182.
- Murty V. R., Bhat, J., & Muniswaran, P. K. A. (2005). External mass transfer effects during the hydrolysis of rice bran oil in immobilized lipase packed bed reactor. *Chemical and Biochemical Engineering Quarterly*, 19(1), 57 – 61.
- Myers, R. H., & Montgomery, D. C. (2002). Response surface methodology: process and product optimisation using designed experiments. Eds. Myers, R. H. And D. C. Montgomery, New York: John Wiley and Sons.
- Nagao, T., Watanabe, H., Goto, N., Onizawa, K., Taguchi, H., Matsuo, N., Yasukawa, T., Tsushima, R., Shimasaki, H., Itakura, H. (2000). Dietary diacylglycerol suppresses accumulation of body fat compared to triacylglycerol in men in a double-blind controlled trial. *The Journal of Nutrition*, 130, 792 – 797.
- Nakajima, Y. (2004). Water-retaining ability of diacylglycerol. *Journal of the American Oil Chemists' Society*, 81(10), 907-912.
- Nath, S., & Chand, S. (1996). Mass transfer and biochemical reaction in immobilized cell packed bed reactors: Correlation of experiment with theory. *Journal of Chemical Technology and Biotechnolgy*, 66(3), 286 – 292.
- Ng, M., Fleming, T., Robinson, M., Thomson, B., Graetz, N., Margono, C., et al. (2014). Global, regional, and national prevalence of overweight and obesity in children and adults during 1980-2013: a systematic analysis for the Global Burden of Disease Study 2013. *The Lancet*, 384(9945), 766 – 781.
- Noor, I. M., Hasan, M., & Ramachandran, K. B. (2003). Effect of operating variables on the hydrolysis rate of palm oil by lipase. *Process Biochemistry*, 39(1), 13-20.
- Noureddini, H., & Medikonduru, V. (1997). Glycerolysis of fats and methyl esters. *Journal of the American Oil Chemists' Society*, 74(4), 419-425.
- Onyike, C. U., Crum, R. M., Lee, H. B., Lyketsos, C. G., & Eaton, W. W. (2003). Is Obesity Associated with Major Depression? Results from the Third National Health and Nutrition Examination Survey. *American Journal of Epidemiology*, 158(12), 1139 – 1147.

- Padmini, P., Iyengar, K. K. S., & Baradarajan, A. (1995). Hydrolysis of ricebran oil in a fluidised-bed recycle reactor using immobilised lipase on nylon-6. *Journal of Chemical Technology & Biotechnology*, 64(1), 31-34.
- Pawongrat, R., Xu, X., & H-Kittikun, A. (2007). Synthesis of monoacylglycerol rich in polyunsaturated fatty acids from tuna oil with immobilized lipase AK. *Food Chemistry*, 104(1), 251-258.
- Phuah, E.-T., Lai, O.-M., Choong, T. S.-Y., Tan, C.-P., & Lo, S.-K. (2012). Kinetic study on partial hydrolysis of palm oil catalyzed by Rhizomucor miehei lipase. *Journal of Molecular Catalysis B: Enzymatic*, 78(0), 91-97.
- Phuah, E.-T., Tang, T. K., Lee, Y. Y., Choong, T. S. Y., Tan, C. P. & Lai, O. M. (2015). Review on the current state of diacylglycerol production using enzymatic approach. *Food and Bioprocess Technology*, 8(6), 1169 – 1186.
- Plou, F. J., Barandiarán, M., Calvo, M. V., Ballesteros, A., & Pastor, E. (1996). High-yield production of mono- and di-oleylglycerol by lipase-catalyzed hydrolysis of triolein. *Enzyme and Microbial Technology*, 18(1), 66-71.
- Pouilloux, Y., Métayer, S., & Barrault, J. (2000). Synthesis of glycerol mono-octadecanoate from octadecanoic acid and glycerol. Influence of solvent on the catalytic properties of basic oxides. *Comptes Rendus de l'Académie des Sciences - Series IIC - Chemistry*, 3(7), 589-594.
- Rahman, N. K., Kamaruddin, A. H., & Uzir, M. H. (2011). Enzymatic synthesis of farnesyl laurate in organic solvent: initial water activity, kinetics mechanism, optimization of continuous operation using packed bed reactor and mass transfer studies. *Bioprocess and Biosystems Engineering*, 34(6), 687 – 699.
- Rana, D. S., Theodore, K., Naidu, G. S. N., & Panda, T. (2003). Stability and kinetics of β -1,3-glucanase from *Trichoderma harzianum*. *Process Biochemistry*, 39, 149-155.
- Remonato, D., Santin, C. M. T., Valério, A., Lerin, L., Batistella, L., Ninow, J. L., de Oliveira, J. V., de Oliveira, D. (2015). Lipase-catalyzed glycerolysis of soybean and canola oils in a free organic solvent system assisted by ultrasound. *Applied Biochemistry and Biotechnology*, 176, 850-862.
- Rendón, X., López-Munguía, A., & Castillo, E. (2001). Solvent engineering applied to lipase-catalyzed glycerolysis of triolein. *Journal of the American Oil Chemists' Society*, 78(10), 1061-1066.
- Ribarova, F., Tsanev, R., & Tsanev, T. (2000). Food fat replacers. Advantages and disadvantages. *Khranitel'novkusova Promishlenost*, 49(9), 7 – 8.
- Romero, M. D., Calvo, L., Alba, C., & Daneshfar, A. (2007). A kinetic study of isoamyl acetate synthesis by immobilized lipase-catalyzed acetylation in *n*-hexane. *Journal of Biotechnology*, 127(2), 269 – 277.
- Rosu, R., Yasui, M., Iwasaki, Y., & Yamane, T. (1999). Enzymatic synthesis of symmetrical 1,3-diacylglycerols by direct esterification of glycerol in solvent-free system. *Journal of the American Oil Chemists' Society*, 76(7), 839-843.
- Rovito, B. J., & Kittrell J. R. (1973). Film and pore diffusion studies with immobilized glucose oxidase. *Biotechnology and Bioengineering*, 15(1), 143 – 161.

- Rudkowska, I., Roynette, C. E., Demonty, I., Vanstone, C. A., Jew, S., & Jones, P. J. H. (2005). Diacylglycerol: Efficacy and Mechanism of Action of an Anti-Obesity Agent. *Obesity Research*, 13(11), 1864-1876.
- Sakaguchi, H. (2001). Marketing a healthy oil. *Oils & Fats International*, 16, 18 – 19.
- Shaikh, M., & Huang, X. (2012). Organic Ionic Liquids: Ultimate Green Solvents in Organic Synthesis. In A. Mohammad, & D. Inamuddin (Eds.), *Green Solvents II* (pp. 473-491): Springer Netherlands.
- Shimada, A. & Ohashi, K. (2003). Interfacial and emulsifying properties of diacylglycerol. *Food Science and Technology Research*, 9(2), 142 – 147.
- Sonntag, N. V. (1982). Glycerolysis of fats and methyl esters — Status, review and critique. *Journal of the American Oil Chemists Society*, 59(10), 795A-802A.
- Sugiura, M., Yamaguchi, H., & Yamada, N. (2002). Preparation process of diglyceride. US Patent no. US6361980 (in English).
- Sun, S., Shan, L., Liu, Y., Jin, Q., Song, Y., & Wang, X. (2009). Solvent-free enzymatic synthesis of feruloylated diacylglycerols and kinetic study. *Journal of Molecular Catalysis B: Enzymatic*, 57(1), 104 – 108.
- Tada, N. (2004). Physiological actions of diacylglycerol outcome. *Current Opinion in Clinical Nutrition & Metabolic Care*, 7(2), 145-149.
- Tada, N., Watanabe, H., Matsuo, N., Tokimitsu, I., & Okazaki, M. (2001). Dynamics of postprandial remnant-like lipoprotein particles in serum after loading of diacylglycerols. *Clinica Chimica Acta*, 311(2), 109-117.
- Taguchi, H., Omachi, T., Nagao, T., Matsuo, N., Tokimitsu, I., & Itakura, H. (2002). Dietary diacylglycerol suppresses high fat diet-induced hepatic fat accumulation and microsomal triacylglycerol transfer protein activity in rats. *The Journal of Nutritional Biochemistry*, 13(11), 678-683.
- Tai, H. P., & Brunner, G. (2011). Mono- and di-acylglycerol synthesis in CO₂-expanded acetone. *The Journal of Supercritical Fluids*, 59(0), 87-91.
- Takase, H. (2007). Metabolism of diacylglycerol in humans. *Asia Pacific journal of clinical nutrition*, 16, 398 - 403.
- Tan, T., & Yin, C. (2005). The mechanism and kinetic model for glycerolysis by 1,3 position specific lipase from *Rhizopus arrhizus*. *Biochemical Engineering Journal*, 25(1), 39-45.
- Tatterson, G. B. (1994). Scale-up and design for industrial mixing processes, New York, USA, McGraw-Hill.
- Temelli, F., King, J., & List, G. (1996). Conversion of oils to monoglycerides by glycerolysis in supercritical carbon dioxide media. *Journal of the American Oil Chemists' Society*, 73(6), 699-706.
- Tepe, O., & Dursun, A. Y. (2008). Combined effects of external mass transfer and biodegradation rates on removal of phenol by immobilized *Ralstonia eutropha* in a packed bed reactor. *Journal of Hazardous Materials*, 151(1), 9 – 16.
- Teramoto, T., Watanabe, H., Ito, K., Omata, Y., Furukawa, T., Shimoda, K., Hoshino, M., Nagao, T. & Naito, S. (2004). Significant effects of diacylglycerol on body fat and lipid metabolism in patients on hemodialysis. *Clinical Nutrition*, 23(5), 1122 – 1126.

- Thoméo, J. C., & Grace, J. R. (2004). Heat transfer in packed beds: experimental evaluation of one-phase water flow. *Brazilian Journal of Chemical Engineering*, 21, 13 – 22.
- Torres, C., Lin, B., & Hill, C., Jr. (2002). Lipase-catalyzed glycerolysis of an oil rich in eicosapentaenoic acid residues. *Biotechnology Letters*, 24(9), 667-673.
- Tripathi, V., Trivedi, R., & Singh, R. P. (2006). Lipase-Catalyzed Synthesis of Diacylglycerol and Monoacylglycerol from Unsaturated Fatty Acid in Organic Solvent System. *Journal of Oleo Science*, 55(2), 65-69.
- Valério, A., Krüger, R. L., Ninow, J., Corazza, F. C., de Oliveira, D., Oliveira, J. V., et al. (2009). Kinetics of Solvent-Free Lipase-Catalyzed Glycerolysis of Olive Oil in Surfactant System. *Journal of Agricultural and Food Chemistry*, 57(18), 8350-8356.
- Valério, A., Rovani, S., Treichel, H., de Oliveira, D., & Oliveira, J. V. (2010). Optimization of mono and diacylglycerols production from enzymatic glycerolysis in solvent-free systems. *Bioprocess and Biosystems Engineering*, 33(7), 805-812.
- van der Padt, A. (1993). Enzymatic acylglycerol synthesis in membrane reactor systems. Wageningen Agricultural University.
- Voll, F., Krüger, R. L., de Castilhos, F., Filho, L. C., Cabral, V., Ninow, J., et al. (2011). Kinetic modeling of lipase-catalyzed glycerolysis of olive oil. *Biochemical Engineering Journal*, 56(3), 107-115.
- Voll, F. A. P., Zanette, A. F., Cabral, V. F., Dariva, C., De Souza, R. O. M. A., Filho, L. C., et al. (2012). Kinetic Modeling of Solvent-Free Lipase-Catalyzed Partial Hydrolysis of Palm Oil. *Applied Biochemistry and Biotechnology*, 168(5), 1121-1142.
- Waldinger, C., & Schneider, M. (1996). Enzymatic esterification of glycerol III. Lipase-catalyzed synthesis of regioisomerically pure 1,3-diacylglycerols and 1 (3)-monoacylglycerols derived from unsaturated fatty acids. *Journal of the American Oil Chemists' Society*, 73(11), 1513 – 1519.
- Wang, W., Li, T., Ning, Z., Wang, Y., Yang, B., & Yang, X. (2011). Production of extremely pure diacylglycerol from soybean oil by lipase-catalyzed glycerolysis. *Enzyme and Microbial Technology*, 49(2), 192-196.
- Wang, X., Han, Z., Zou, W., Yang, W., Jin, Q., & Wang, X. (2015). Preparation of 1,3-diolein by irreversible acylation. *Journal of the American Oil Chemists' Society*, 92, 185-191.
- Wang, Y., Zhao, M., Ou, S., Song, K., & Han, X. (2009a). Preparation of diacylglycerol-enriched palm olein by phospholipase A1-catalyzed partial hydrolysis. *European Journal of Lipid Science and Technology*, 111(7), 652-662.
- Wang, Y., Zhao, M., Ou, S., Xie, L., & Tang, S. (2009b). Preparation of a diacylglycerol-enriched soybean oil by phospholipase A₁ catalyzed hydrolysis. *Journal of Molecular Catalysis B: Enzymatic*, 56(2–3), 165-172.
- Wang, Y., Zhao, M., Ou, S., & Song, K. (2009c). Partial hydrolysis of soybean oil by phospholipase A₁ to produce diacylglycerol-enriched oil. *Journal of Food Lipids*, 16(1), 113 – 132.

- Wang, Y., Zhao, M., Song, K., Wang, L., Tang, S., & Riley, W. W. (2010). Partial hydrolysis of soybean oil by phospholipase A1 (Lecitase Ultra). *Food Chemistry*, 121(4), 1066-1072.
- Watanabe, H., Onizawa, K., Taguchi, H., Kobori, M., Chiba, H., Naito, S., et al. (1997). Nutritional Characterization of Diacylglycerols in Rats. *Journal of Japan Oil Chemists' Society*, 46(3), 301-307.
- Watanabe, T., Shimizu, M., Sugiura, M., Sato, M., Kohori, J., Yamada, N., et al. (2003). Optimization of reaction conditions for the production of DAG using immobilized 1,3-regiospecific lipase lipozyme RM IM. *Journal of the American Oil Chemists' Society*, 80(12), 1201-1207.
- Watanabe, T., Sugiura, M., Sato, M., Yamada, N., & Nakanishi, K. (2005). Diacylglycerol production in a packed bed bioreactor. *Process Biochemistry*, 40(2), 637 – 643.
- Weber, N., & Mukherjee, K. D. (2004). Solvent-free lipase-catalyzed preparation of diacylglycerols. *Journal of Agricultural and Food Chemistry*, 52(17), 5347 - 5353.
- WHO (2014). Obesity and overweight. <http://www.who.int/mediacentre/factsheets/fs311/en/> (Accessed 20 Sept 2014)
- Willis, W. M., & Marangoni, A. G. (2008). Enzymatic Interesterification. In *Food Lipids: Chemistry, Nutrition and Biotechnology (3rd ed.)* (pp. 807 - 840, Food Science and Technology): CRC Press
- Wyatt, V. T., Bush, D., Lu, J., Hallett, J. P., Liotta, C. L., & Eckert, C. A. (2005). Determination of solvatochromic solvent parameters for the characterization of gas-expanded liquids. *The Journal of Supercritical Fluids*, 36(1), 16-22.
- Xu, X. (2000). Enzyme bioreactors for lipid modifications. *Inform*, 11, 1004 – 1012.
- Xu, X. (2003). Engineering of enzymatic reactions and reactors for lipid modification and synthesis. *European Journal of Lipid Science and Technology*, 105(6), 289-304.
- Xu, X., Balchen, S., Høy, C. E., & Adler-Nissen, J. (1998). Production of specific-structured lipids by enzymatic interesterification in a pilot continuous enzyme bed reactor. *Journal of the American Oil Chemists' Society*, 75(11), 1573-1579.
- Xu, X., Skands A. R. H., Høy, C. E., Mu, H., Balchen, S., & Adler-Nissen, J. (1999). Production of specific-structured lipids by enzymatic interesterification: Elucidation of acyl migration by response surface design. *Journal of the American Oil Chemists' Society*, 75(9), 1179-1186.
- Xue, X., Mu, H., Høy, C. E., & Adler-Nissen, J. (1999). Production of specifically structured lipids by enzymatic interesterification in a pilot enzyme bed reactor: process optimization by response surface methodology. *Lipid / Fett* 101(6), 207 – 214.
- Yamada, Y., Shimizu, M., Sugiura, M., & Yamada, N. (1999). Process for producing diglycerides. PCT International Application No. WO9909119 (in English).
- Yamane, T., Kang, S., Kawahara, K., & Koizumi, Y. (1994). High-yield diacylglycerol formation by solid-phase enzymatic glycerolysis of hydrogenated beef tallow. *Journal of the American Oil Chemists' Society*, 71(3), 339-342.

- Yanai, H., Tomono, Y., Ito, K., Furutani, N., Yoshida, H., & Tada, N. (2007). Diacylglycerol oil for the metabolic syndrome. *Nutrition Journal*, 6(43), 1 - 6.
- Yang, T., Rebsdorf, M., Engelrud, U., & Xu, X. (2005). Monoacylglycerol synthesis via enzymatic glycerolysis using a simple and efficient reaction system. *Journal of Food Lipids*, 12(4), 299-312.
- Yang, T., Zhang, H., Mu, H., Sinclair, A., & Xu, X. (2004). Diacylglycerols from butterfat: Production by glycerolysis and short-path distillation and analysis of physical properties. *Journal of the American Oil Chemists' Society*, 81(10), 979-987.
- Yeoh, C. M., Choong, T. S. Y., Abdullah, L. C., Yunus, R., & Siew, W. L. (2009). Influence of silica gel in production of diacylglycerol via enzymatic glycerolysis of palm olein. *European Journal of Lipid Science and Technology*, 111(6), 599-606.
- Yeoh, C. M., Phuah, E. T., Tang, T. K., Siew, W. L., Abdullah, L. C., & Choong, T. S. Y. (2014). Molecular distillation and characterization of diacylglycerol-enriched palm olein. *European Journal of Lipid Science and Technology*, 116(12), 1654 – 1663.
- Yesiloglu, Y., & Kilic, I. (2004). Lipase-catalyzed esterification of glycerol and oleic acid. *Journal of the American Oil Chemists' Society*, 81(3), 281 – 284.
- Yuan, X., Liu, J., Zeng, G., Shi, J., Tong, J., & Huang, G. (2008). Optimization of conversion of waste rapeseed oil with high FFA to biodiesel using response surface methodology. *Renewable Energy*, 33(7), 1678 – 1684.
- Zaks, A., & Klibanov, A. M. (1988a). The effect of water on enzyme action in organic media. *Journal of Biological Chemistry*, 263(17), 8017 - 8021.
- Zaks, A., & Klibanov, A. M. (1988b). Enzymatic catalysis in nonaqueous solvents. *Journal of Biological Chemistry*, 263(7), 3194-3201.
- Zhao, T. T., Kim, B. H., Hong, S. I., Yoon, S. W., Kim, C. T., Kim, Y. & Kim, I. H. (2012). Lipase-catalysed production of pinolenic acid concentrate from pine nut oil using a recirculating packed bed reactor. *Journal of Food Science*, 77(2), 267 – 271.
- Zhong, N., Gui, Z., Xu, L., Huang, J., Hu, K., Gao, Y., et al. (2013). Solvent-free enzymatic synthesis of 1, 3-Diacylglycerols by direct esterification of glycerol with saturated fatty acids. *Lipids in Health and Disease*, 12(65).
- Zhong, N., Li, B., Xu, X., Cheong, L.-Z., Xu, Z., & Li, L. (2011). Low-Temperature Chemical Glycerolysis: An Evaluation of Substrates Miscibility on Reaction Rate. *Journal of the American Oil Chemists' Society*, 88(7), 1077-1079.
- Zhong, N., Li, L., Xu, X., Cheong, L.-Z., Zhao, X., & Li, B. (2010). Production of diacylglycerols through low-temperature chemical glycerolysis. *Food Chemistry*, 122(1), 228-232.
- Zhong, N., Li, L., Xu, X., Cheong, L., Li, B., Hu, S., et al. (2009). An Efficient Binary Solvent Mixture for Monoacylglycerol Synthesis by Enzymatic Glycerolysis. *Journal of the American Oil Chemists' Society*, 86(8), 783-789.