



UNIVERSITI PUTRA MALAYSIA

***CHEMOPREVENTIVE POTENTIAL OF WATER AND METHANOLIC
EXTRACTS OF *Clinacanthus nutans* (Burm.f) . LINDAU ON COLON
CANCER In Vitro AND In Vivo***

MARIAMU NYAKO KURA

IB 2015 30



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By

MARIAMU NYAKO KURA

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of Master of Science**

September 2015

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DEDICATION

I dedicate this MSc thesis and give special thanks to my parents Vice Admiral Murtala H. Nyako and Hajiya Zainab M. Nyako. Their constant love, support and encouragement have been a blessing throughout my life.



Abstract of thesis presented to the senate of Universiti Putra Malaysia in fulfilment of the requirement for the Degree of Master of Science

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EXTRACTS OF *Clinacanthus nutans* (Burm.f) . LINDAU ON COLON
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By

Mariam Nyako Kura

September 2015

**Chairman : Prof Maznah binti Ismail PhD
Faculty : Institute of Bioscience**

Chemoprevention is a promising approach in controlling the occurrence of colorectal cancer (CRC). In this study the effects of extracts of *Clinacanthus nutans* (Cn) on colorectal cancer *in vitro* and *in vivo* was investigated. Cn Leaf and stem were extracted using sonication. Cytotoxicity testing of the extracts on the human colon adenocarcinoma cell line HT- 29 was determined. For the *in vivo* study, male Sprague Dawley rats were randomly assigned to six groups. Rats in treatment groups were administered 100mg/kg 1, 2- dimethylhydrazine (DMH) by gavage twice in the first week. The extracts were administered for eight weeks. After sacrifice, full blood count and histological analysis of the colons and livers of experimental animals was then carried out. The extracts were also analyzed qualitatively for the presence of certain phytochemicals.

IC₅₀ values of 771.28±0.35µg/ml, 911.8±0.06µg/ml and 622.63±0.2µg/ml were observed for the water, hot water and methanolic extracts of the leaves of Cn after 72h treatment respectively. The number of aberrant crypt foci and crypt multiplicity significantly decreased in the colons of rats in the 500mg/kg water extract treated group. The 500mg/kg water extract also significantly decreased hepatic steatosis and significantly increased mRNA expression of SOD1 antioxidant gene in the liver of rats. The extracts induced a certain amount of toxicity in blood of treated rats (anemia, decreased total White blood cell counts, decreased thrombocyte counts and decreased monocyte counts). Phytochemical screening of the extracts of also revealed the presence of saponins, tannins, alkaloids, flavonoids, carbohydrates, phenols and diterpenes. Though it induced toxicity in the blood of treated rats, the water extract of the leaf showed the most chemopreventive potential among the 12 extracts of *Clinacanthus nutans* studied therefore this merits the further study of this extract for clinical trials.

Abstrak tesis yang dikemukakan kepada senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains.

**POTENSI KEMOPREVENTIF DARIPADA EKSTRAK AIR DAN METANOL
DARIPADA *Clinacanthus nutans* (Burm.f). LINDAU PADA KANSER KOLON
In Vitro DAN *In Vivo***

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Pencegahan kimo adalah satu pendekatan yang menjanjikan harapan dalam mengawal kejadian kanser kolorektal (CRC). Dalam kajian ini kesan ekstrak *Clinacanthus nutans* (Cn) ke atas kanser kolorektal *in vitro* dan *in vivo* telah dikaji. Daun dan batang Cn telah diekstrak menggunakan kaedah sonikasi. Ujian sitotoksik ekstrak ke atas sel kolon adenokarsinoma manusia HT-29 telah ditentukan. Untuk kajian *in vivo*, tikus jantan Sprague Dawley dibahagikan kepada enam kumpulan secara rawak. Tikus dalam kumpulan rawatan telah diberikan 100mg / kg 1, 2- dimethylhydrazine (DMH) secara gavage sebanyak dua kali dalam minggu pertama. Ekstrak telah diberikan selama lapan minggu. Selepas tikus dibunuh, kiraan darah lengkap dan analisis histologi daripada kolon dan hati haiwan eksperimen telah dijalankan. Ekstrak juga dianalisis secara kualitatif bagi menentukan kehadiran fitokimia tertentu. Nilai IC_{50} untuk ekstrak air, ekstrak air panas dan ekstrak metanol daripada daun Cn selepas rawatan selama 72 jam masing-masing adalah $771.28 \pm 0.35 \mu\text{g/ml}$, $911.8 \pm 0.06 \mu\text{g/ml}$ dan $622.63 \pm 0.2 \mu\text{g/ml}$. Bilangan "abberant crypt foci" dan "crypt multiplicity" menurun dengan ketara dalam kolon tikus yang dirawat menggunakan 500mg/kg ekstrak air. 500mg/kg ekstrak air juga menurunkan steatosis hati dan meningkatkan ekspresi mRNA gen antioksidan SOD1 dalam hati dengan signifikan. Ekstrak Cn menyebabkan kesan toksik dalam darah tikus yang dirawat (anemia, pengurangan jumlah bilangan sel darah putih, pengurangan bilangan thrombocyte dan pengurangan bilangan monosit). Penyaringan fitokimia ke atas ekstrak juga mendedahkan kehadiran saponin, tanin, alkaloid, flavonoid, karbohidrat, fenol dan diterpenes. Walaupun menyebabkan kesan toksik dalam darah tikus yang dirawat, ekstrak air daun Cn menunjukkan kesan pencegahan kimo yang paling berpotensi antara 12 ekstrak *Clinacanthus nutans* yang dikaji, justeru memberi merit ke atas kajian lanjut ekstrak ini bagi ujian klinikal.

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I certify that a Thesis Examination Committee has met on 4 September 2015 to conduct the final examination of Mariamu Nyako Kura on her thesis entitled "Chemopreventive Potential of Water and Methanolic Extracts of *Clinacanthus nutans* (Burm.f.) Lindau on Colon Cancer *In Vitro* and *In Vivo*" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

AC	Aberrant crypts
ACF	Aberrant crypt foci
AO	Acridine orange
AOM	Azoxymethane
Apc	Adenomatous polyposis coli
ATCC	American type culture collection
ANOVA	Analysis of variance
BA	Bile acids
BMI	Body mass index
CD	Crohn' s disease
Cn	<i>Clinacanthus nutans</i>
CRC	Colorectal cancer
CVD	Cardiovascular disease
DMSO	Dimethyl sulfoxide
DMEM	Dulbecco's modified eagle's medium
DMH	1, 2- dimethylhydrazine
DOC	Deoxycholic acid
DPPH	1, 1- diphenyl- 2- picrylhydrazyl
DSS	Dextrane sodium sulphate
EGCG	Epigallocatechin- 3- gallate
EPIC	European prospective investigation into cancer and nutrition
FAP	Familial adenomatous polyposis
FBS	Fetal bovine serum
FDA	Food and drug administration
5- Fu	5- Fluorouracil
FTIR	Fourier transform infrared spectroscopy
Gpx1	Glutathione peroxidase 1
GSH	Glutathione
HCA	Heterocyclic amines
HCL	Hydrochloric acid
Hmox1	Heme oxygenase (decycling) 1

HNPCC	Hereditary non- polyposis colon cancer
HSV	Herpes simplex virus
HFD	High fat diet
IACUC	Institutional Animal Care and Use Committee
IC ₅₀	50% inhibitory concentration
IFD	Inflammatory bowel disease
IR	Infrared spectroscopy
Keap1	Kelch- like ECH- associated protein 1
LCA	Lithocholic acid
M1	250mg/ kg 80% methanol extract
M2	500mg/ kg 80% methanol extract
MAM	Formmethylazoxymethanol
MNNG	N- methyl- N'- nitro- N- nitrosoguanidine
MNU	Methylnitrosourea
MTT	3- (4, 5- dimethylthiazol- 2- yl)- 2, 5, diphenyl tetrazolium bromide assay
NC	Normal/ negative control
NCD	Non- communicable chronic disease
NO	Nitric oxide
Nrf2	Nuclear factor erythroid 2- related factor 2
NSAIDS	Non- steroidal anti- inflammatory drugs
Nqo1	NAD(P)H dehydrogenase (quinone) 1
OD	Optical density
PBS	Phosphate buffered saline
PCR	Polymerase chain reaction
PI	Propidium iodide
PS	Phosphatidyl serine
ROns	Reactive oxygen/ nitrogen species
ROS	Reactive oxygen species
RONS	Reactive nitrogen species
RT	Reverse transcription
SD	Standard deviation
Sod1	Superoxide dismutase 1
Sod2	Superoxide dismutase 2
TE	Tris- EDTA

UC	Ulcerative colitis
VDR	Vitamin D receptor
VZU	Varicella- zoster virus
W1	250mg/ kg water extract
W2	500mg/ kg water extract
WHO	World Health Organization



CHAPTER 1

INTRODUCTION

1.1 General introduction

The Non- Communicable diseases (NCDs) comprising mainly of diabetes, obesitycardiovascular disease and the cancers are the leading cause of death globally. The World Health Organization (WHO) estimates that 36 million of the 57 million deaths that occurred in 2008 were due to the NCDs (Alwan et al., 2011). Four main behavioral risk factors have been recognized as the causes of NCDs, a combination of changes in lifestyle and dietary patterns, including the harmful use of alcohol, unhealthy diet, tobacco use and a sedentary lifestyle. The public health sectors in nations worldwide have been under continuous strain due to the increase in the morbidity and mortality brought on by these diseases. Available data demonstrates that almost 80% of deaths caused by NCDs occur in low- and middle-income countries. Malaysia is no exception to this trend. Poor nutritional choices and a sedentary lifestyle have also increased the morbidity and mortality of these diseases in Malaysia (Ministry of Health Malaysia., 2010; Alwan et al., 2011).

Colorectal cancer (CRC) is caused by modifiable and non- modifiable risk factors. Non- modifiable risk factors include genetic predisposition, age, male sex and previous colonic polyps. Modifiable risk factors are mostly associated with a western lifestyle consisting of a diet rich in processed and red meats, sugars, refined starches, saturated and trans- fatty acids; and poor in whole grains, fiber, fruits, vegetables, calcium, vitamin D and fish oils. A high body mass Index, visceral adiposity, alcohol consumption and physical inactivity among other risk factors are strongly implicated in the etiology of CRC. Malaysian government strategies to fight the increasing burden of this disease on the public health sector have not produced the desired impact so other alternative strategies need to be explored (Bhalla et al., 2009; MOH Malaysia., 2010).

Chemoprevention involves the use of phytochemicals, dietary bioactives, pharmacological or even whole plant extracts to block or arrest the cellular and molecular events that lead to neoplastic transformation (Neerghen et al, 2010). Preclinical testing is an important part of the anti- cancer drug development process. This includes both *in vitro* (the use of human cancer cell lines) and *in vivo* techniques (using animal models) (Cree, 2011).

Human cancer cells lines are important tools in cancer research as they are commercially available and exhibit unrestrained growth which makes them well suited for *in vitro* studies. They are also relevant models for the *in vivo* situation because they show epigenetic changes that are compatible with those found in primary carcinomas. The HT29 (human colon adenocarcinoma cell line) is one of those human cancer cell lines (Ahmad et al., 2013; Gillet et al., 2013). Chemopreventive studies involving animal models to test the efficacy of many compounds in inhibiting the initiation and progression of the disease with some

compounds showing good efficacy is a promising strategy in the fight against colorectal cancer (CRC) (Takayama et al, 2009).

Experimental models like the 1,2-dimethylhydrazine (DMH) model are histologically, morphologically and anatomically similar to human colonic epithelial neoplasms (Perse et al, 2011). DMH and its metabolite azoxymethane (AOM) induce colon cancer through a multistep process similar to that observed in human colon cancer. There are three stages to the carcinogenesis process: initiation, promotion and progression. The initiation stage, which this study was focused on, involves the formation of a mutated, preneoplastic cell from a genotoxic event (Femia and Caderni., 2008; Klaunig and Kamendulis., 2004).

Aberrant crypt foci (ACF) are preneoplastic lesions of the colonic mucosa. Large numbers of ACF are commonly observed in familial adenomatous polyposis and sporadic CRC patients. ACF are also one of the early morphological changes observed in DMH induced CRC in rodents. The DMH model is a good model to use when trying to understand the molecular alterations that arise in human CRC (Perse et al, 2011).

Equilibrium between cell proliferation and death needs to be maintained within tissues or the result will be carcinogenesis. The carcinogenesis process involves endogenous and exogenous factors influencing DNA damage, cell growth and cell death. An important role for reactive oxygen species (ROS) and reactive nitrogen species (RNS) in the cancer process is supported by experimental evidence. An increase in ROS/ RNS production either through physiological modification or exposure to a chemical carcinogen contributes to the multistep carcinogenesis process. This could be through modification of gene expression or genotoxic effects resulting in oxidative DNA adducts being formed (Klaunig and Kamendulis., 2004). Under normal physiological conditions, cells have the ability to counterbalance the production of ROS/ RNS with antioxidants. Endogenous cellular antioxidant defenses are mainly enzymatic and include catalase, super oxide dismutase and glutathione peroxidase (Klaunig and Kamendulis., 2004).

For centuries plants and humans have sustained each other. In every culture there is definable plant knowledge about medicinal, ceremonial and edible plants. Modern conventional medicine is derived mostly from work done by the first botanists who were physicians that kept personnel herb gardens for treating the sick (Warber et al, 2006). Malaysia is blessed with untapped floral and faunal biodiversity. It also has a rich tapestry of traditional healing techniques and plants that are yet underutilized. Plant phytochemicals are receiving much attention due to their huge potential in reducing cancer risk (Neergheen et al., 2010; Yadav and Agarwala., 2011; Sengottuvelan et al., 2006; Russo et al., 2010). The public is disillusioned with conventional medication due to problems regarding side effects and have fueled an intense interest in herbal preparations.

Plants contain phytochemicals such as terpenoids, phenolic acids, tannins, flavonoids, alkaloids and other metabolites that are rich in antioxidant activity. Studies have shown many of these antioxidant phytochemicals possess anti-inflammatory, antitumor and anticarcinogenic effects. Reduced risk of cancers and

other chronic diseases has been associated with the ingestion of natural antioxidants (Aiyegoro and Okoh., 2010).

Clinacanthus nutans (Cn) is a small shrub found throughout south-east Asia. Its English name is Sabah snake grass and its Malay name is Belalai gajah. It is reported to have anti- inflammatory, immune modulatory, antimicrobial, hepatoprotective and antioxidant effects (Chelyn et al., 2014; Yong et al., 2013).

1.2 Statement of research problem

Colorectal cancer is the fourth most common cancer among both sexes in south-east Asia and the second most common cancer among both sexes in Malaysia. Mortality rates for the disease are unacceptably high. In Malaysia, the disease has a 5- year prevalence rate of 12.2% and 10.6% mortality for both sexes (WHO/IARC, 2012).

It is doubtful that incidence and mortality rates will change because current therapies are unsatisfactory, associated with severe side effects and unsuccessful remission rates. There is an urgent need in finding alternatives to current therapies and an even greater need for prevention. Chemoprevention is an alternative that is strongly being advocated. Chemoprevention as a strategy has been used successfully in reducing the incidence and mortality associated with cardiovascular disease. Defined as the use of natural or pharmacological agents to reverse or delay carcinogenesis at an early stage, chemoprevention is gaining popularity both among clinicians and the general public (Steward and Brown., 2013).

Phytochemicals from both dietary and non- dietary (medicinal plants or herbs) sources are currently been studied for their potential benefits against an array of cancers. The Federal Drug law enforcement Agency (FDA) in the US has several drugs under consideration as potential chemopreventive agents; some are presently undergoing clinical trials. Some positive results have been attained for non-steroidal anti- inflammatory drugs (NSAIDs) in colorectal adenoma and carcinoma prevention, vitamin C for bladder cancer and finasteride for prostate cancer (Russo, 2007).

Clinacanthus nutans is easily available in Malaysia and research on its anti-colorectal cancer effects have yet to be determined. Therefore due to the public health burden of colorectal cancer in Malaysia there is a strong need to find readily accessible and inexpensive medicinal plants that could possibly help in the fight against this disease.

1.3 Significance of study

The focus of this study was to investigate the effects of extracts of *Clinacanthus nutans* (Cn) on cytotoxicity of CRC cells *in vitro* and also to investigate the ability of the extracts to stop the initiation stage of DMH induced cancer *in vivo*. This study will endeavor to investigate the anti- colon cancer effects of the leaf and stem of Cn. Due to Cn been readily available and inexpensive; it could be beneficial in the fight against colorectal cancer in Malaysia.

1.4 General objective

To study the chemopreventive potential of different extracts of the stem and leaves of Cn.

1.4.1 Specific objectives

1. To determine the cytotoxic effect of extracts of *Clinacanthus nutans* leaves and stems on HT29 and NIH/3t3 cells.
2. To determine the ability of candidate extracts to inhibit the formation of aberrant crypt foci in a
DMH induced CRC model in rats.
3. To determine the mechanistic basis for the anti-oxidative effect of extracts.
4. To determine phytochemicals present in the extracts of Cn.

REFERENCES

- Abd Ghafar, S. A., Yazan, L. S., Md Tahir, P. and Ismail, M. Kenaf seed supercritical fluid extract reduces aberrant crypt foci formation in azoxymethane-induced rats. *Experimental and Toxicologic Pathology* 64: 247–251 (2012).
- Aggarwal, B. B., Danda, D., Gupta, S. and Gehlot, P. Models for prevention and treatment of cancer: Problems vs Promises. *Biochemical pharmacology* 78: 1083- 1094 (2009).
- Ahmed, D., Eide, P. W., Eilertsen, I. A., Danielsen, S. A., Eknæs, M., Hektoen, M., Lind, G.E. and Lothe, R. A. Epigenetic and genetic features of 24 colon cancer cell lines *Oncogenesis*. 2, e71 (2013).
- Ajouz, H., Mukherji, D. and Shamseddine, A. Secondary bile acids: an underrecognized cause of colon cancer. *World Journal of Surgical Oncology* 12: 164 (2014).
- Akhdara, H., Loyerb, P., Raucha, C., Corlub, A., Guillouzo, A. and Morela, F. Involvement of Nrf2 activation in resistance to 5-fluorouracil in human colon cancer HT-29 cells. *European Journal of Cancer* 45: 2219 –2227 (2009).
- Aleksandrova, K., Pischon, T., Buijsse, B., May, A. M., Peeters, P. H., Bueno de-Mesquita, B. H., Jenab, M., Fedirko, V., Dahm, C. C., Siersema, P. D., Freisling, H., Ferrari, P., Overvad, K., Tjønneland, A., Trichopoulou, A., Lagiou, P., Naska, A., Pala, V., Mattiello, A., Ohlsson, B., Jirstro, M. K., Key, T. J., Khaw, K., Riboli, E. and Boeing, H. Adult weight change and risk of colorectal cancer in the European Prospective Investigation into Cancer and Nutrition. *European Journal of Cancer* 49: 3526–3536 (2013).
- Alwan A., Armstrong, T., Bettcher, D., Branca, F., Chisholm, D., Ezzati, M., Garfield, R., MacLean, D., Mathers, C., Mendis, S., Poznyak, V., Riley, L., Tang, K. C. and Wild C. *Global status report on non- communicable diseases 2010*. World Health Organization Publication (2011).
- Alwan, A., Armstrong, T., Cowan, M. and Riley, L., WHO: *Non- communicable diseases country profiles 2011*. World Health Organization Publication.
- American Cancer Society. *Colorectal cancer facts and figures 2011- 2013*.
- Andersen V, Kopp T.I, Tjønneland A and Vogel U. No Association between HMOX1 and Risk of Colorectal Cancer and No Interaction with Diet and Lifestyle Factors in a Prospective Danish Case-Cohort Study. *International Journal of Molecular Sciences* 16(1): 1375–1384 (2015).

- Barrera, G. Oxidative Stress and Lipid Peroxidation Products in Cancer Progression and Therapy. *International Scholarly Research Network (ISRN) Oncology* Article ID 137289: 1- 21 (2012).
- Bernstein, C., Payne, C. M. and Bernstein, H. Bile Acids: Promoters or Carcinogens in Colon Cancer? *Journal of Carcinogenesis and Mutagenesis* 2 (2): 1-2 (2011).
- Bhalla, V. and Bhalla, M. A. Risk factors for colorectal cancer. *Oncology News* 4: 84– 85 (2009).
- Bird, R. P. and Good, C. K. The significance of aberrant crypt foci in understanding the pathogenesis of colon cancer. *Toxicological letters* 112-113: 395- 402 (2000).
- Brenner H, Kloor M and Pox C. P. Colorectal cancer. www.thelancet.com November 11: 1- 13 (2013).
- Cavina, C., Holzhaeusera, D., Scharfb, G., Constablea, A., Huberb, W. and Schilter, B. Cafestol and kahweol, two coffee specific diterpenes with anticarcinogenic activity. *Food and Chemical Toxicology* 40: 1155–1163 (2002).
- Chen, Y. Q., Edwards, I. J., Kridel, S. J., Todd, T. and Berquin, I. M. Dietary fat-gene interactions in cancer. *Cancer metastasis reviews* 26: 535- 551 (2007).
- Chidambara, M. K. N. C., Jayaprakasha, G. K. and Patil, B. S. The natural alkaloid berberine targets multiple pathways to induce cell death in cultured human colon cancer cells. *European Journal of Pharmacology* 688 (1- 3): 14- 21 (2012).
- Cho, E., Lee, J. E., Rimm, E. B., Fuchs, C. S. and Giovannucci, E. L. Alcohol consumption and the risk of colon cancer by family history of colorectal cancer. *American Journal of Clinical Nutrition* 95: 413–9 (2012).
- Chung, M., Lim, T. G. and Lee, K. W. Molecular mechanisms of chemopreventive phytochemicals against gastroenterological cancer development. *World Journal of Gastroenterology* 19 (7): 984-993 (2013).
- Coates, J. Interpretation of infrared spectra, a practical approach. In R. A. Meyers, editor. *Encyclopedia of Analytical Chemistry*. Chichester: John Wiley and sons Ltd; 2000. p. 10815- 10837.
- Coxon K. M, Duggan J, Cordeiro M. F and Moss S. E. In *Cancer Cell Culture Methods and Protocols*. 2nd Edition. Humana press: New York, New York; 2011. p. 293- 308.
- Cree I. A. Principles of cancer cell culture. *Methods in Molecular Biology* 731: 13-26 (2011).

- Cseke L. J, Kirakosyan A, Kaufman P. B, Warber S. L, Duke J. A and Brielmann H. L. Natural Products from Plants. Florida: Taylor and Francis; 2006. 632p.
- Dampawan P, Huntrakul C and Reutrakul V. Constituents of *Clinacanthus nutans* and the crystal structure of LUP-20(29)-ENE-3-ONE. *Journal of Science Society of Thailand* 3: 14–26 (1977).
- Do Q. D, Angkawijaya A. E, Tran-Nguyen P. L, Huynh L. H, Soetaredjo F. E, Ismadji S, Ju Y. Effect of extraction solvent on total phenol content, total flavonoid content, and antioxidant activity of *Limnophila aromatic*. *Journal of Food and Drug Analysis* 22(3): 296–302 (2014).
- Donohue, C. L., Doyle, S. L. and Reynolds, J. V. Visceral adiposity, insulin resistance and cancer risk. *Diabetology and metabolic syndrome* 3: 1-13 (2011).
- Erbach M, Mehnert H and Schnell O. Diabetes and the risk for colorectal cancer. *Journal of diabetes and its complications* 26: 50- 55 (2012).
- Fedirko V, Tramacere I, Bagnardi V, Rota M, Scotti L, Islami F, Negri E. K, Straif K, Romieu I, La Vecchia C, Boffetta P and Jenab M. Alcohol drinking and colorectal cancer risk: an overall and dose– response meta-analysis of published studies. *Annals of Oncology* 22: 1958–1972 (2011).
- Femia A. P. and Caderni G. Rodent models of colon carcinogenesis for the study of chemopreventive activity of natural products. *Planta Medica* 74: 1602- 1607 (2008).
- Feskanich D, Ma J, Fuchs C. S, Kirkner G. J, Hankinson S. E, Hollis B. W and Giovannucci E. L. Plasma Vitamin D Metabolites and Risk of Colorectal Cancer in Women. *Cancer Epidemiology, Biomarkers and Prevention* 13(9): 1502- 1508 (2004).
- Flis S. and Splawinski J. Inhibitory effects of 5- Fluorouracil and oxaliplatin on human colorectal cancer cell survival are synergistically enhanced by sulindac sulfide. *Anticancer research* 29: 435- 442 (2009.)
- Frezza E. E, Wachtel M. S and Chriva- Internati M. Influence of obesity on the risk of developing colon cancer. *Gut* 55: 285- 291 (2006).
- Gali-Muhtasib H. U, Younes I. H, Karchesy J. J and El-Sabba M. E. Plant Tannins Inhibit the Induction of Aberrant Crypt Foci and Colonic Tumors by 1, 2 Dimethylhydrazine in Mice. *Nutrition and Cancer* 39 (1): 108– 116 (2001).
- Gill C. I. R and Rowland I. R. Diet and cancer: assessing the risk. *British journal of nutrition* 88 (1): S73- S87 (2002).
- Gillet J, Varma S and Gottesman M. M. The Clinical Relevance of Cancer Cell Lines. *Journal of the National cancer institute* 105 (7): 452- 458 (2013).

- Giovannucci E. Commentary: Vitamin D and colorectal cancer— twenty-five years later. *International Journal of Epidemiology* 35: 224– 225 (2006).
- Goh K. L, Quek K. F, Yeo G. T. S, Hilmi I. N, Lee C. K, Hasnida N, Aznan M, Kwan K. L and Ong K. T. Colorectal cancer in Asians: a demographic and anatomic survey in Malaysian patients undergoing colonoscopy. *Alimentary Pharmacology & Therapeutics* 22: 859–864 (2005).
- Goodman S. R editor. Regulation of gene expression. In *Medical Cell Biology* 2nd edition. Lippincott- Raven Publishers: Philadelphia, PA; 1998. p. 165-193.
- Guo Y, Chen Y, Chiu W, Liao H and Lin S. Soy Saponins Mediate the Progression of Colon Cancer in Rats by Inhibiting the Activity of β -Glucuronidase and the Number of Aberrant Crypt Foci but Not Cyclooxygenase-2 Activity. *ISRN Oncology*, Volume 2013, Article ID 645817, 9 pages, doi:10.1155/2013/645817.
- Gunter M. J and Leitzmann M. F. Obesity and colorectal cancer: epidemiology, mechanisms and candidate genes. *Journal of nutritional biochemistry* 17: 145- 156 (2006).
- Hagland H. R and Søreide K. Cellular metabolism in colorectal carcinogenesis: Influence of lifestyle, gut microbiome and metabolic pathways. *Cancer Letters*. 356 (2), pp. 273- 280 (2015).
- Hamanishi T, Furuta H, Kato H, Doi A, Tamai M, Shimomura H, Sakagashira S, Nishi M, Sasaki H, Sanke T and Nanjo K. Functional Variants in the Glutathione Peroxidase-1(GPx-1) Gene Are Associated With Increased Intima-Media Thickness of Carotid Arteries and Risk of Macrovascular Diseases in Japanese Type 2 Diabetic Patients. *Diabetes* 53: 2455- 2460 (2004).
- Hatziantonioua S, Dimasb K, Georgopoulos A, Sotiriadouc N and Demetzos C. Cytotoxic and antitumor activity of liposome-incorporated sclareol against cancer cell lines and human colon cancer xenografts. *Pharmacological Research* 53: 80–87 (2006).
- Hirose Y, Kuno T, Yamada Y, Sakata K, Katayama M, Yoshida K, Qiao Z, Hata K, Yoshimi N and Mori H. Azoxymethane-induced beta-catenin-accumulated crypts in colonic mucosa of rodents as an intermediate biomarker for colon carcinogenesis. *Carcinogenesis* 24 (1): 107–111 (2003).
- Houghton P, Fang R, Techatanawat I, Steventon G, Hylands P. J and Lee C. C. The sulphorhodamine (SRB) assay and other approaches to testing plant extracts and derived compounds for activities related to reputed anticancer activity. *Methods* 42: 377– 387 (2007)^(a).

- Houghton P. J, Howes M. J, Lee C. C and Steventon G. Uses and abuses of in vitro tests in Ethnopharmacology: visualizing an elephant. *Journal of Ethnopharmacology* 110: 391- 400 (2007)^(b).
- Imam M. U and Ismail M. Effects of Brown Rice and White Rice on Expression of Xenobiotic Metabolism Genes in Type 2 Diabetic Rats. *International Journal of Molecular Sciences* 13: 8597-8608 (2012).
- Imam M. U and Ismail M. Nutrigenomic effects of germinated brown rice and its bioactives on hepatic gluconeogenic genes in type 2 diabetic rats and HEPG2 cells. *Molecular Nutrition and Food research* 00: 1-11 (2013).
- Inger, T. Aberrant crypt foci in the colo-rectal mucosa as reliable markers of tumor development: With special reference to histomorphological and immunohistochemical characterization of aberrant crypt foci in rats. Ph.D. Thesis. *Institute of Toxicology, Department of General Toxicology, National Food Agency of Denmark* (1996).
- Ishino K, Mutoh M, Totsuka Y and Nakagama H. Metabolic syndrome: A novel high- risk state for colorectal cancer. *Cancer letters* 334: 56- 61 (2013).
- Jacobs E. J, Connell C. J, Patel A. V, Chao A, Rodriguez C, Seymour J, McCullough M. L, Calle E. E and Thun M. J. Vitamin C and Vitamin E Supplement Use and Colorectal Cancer Mortality in a Large American Cancer Society Cohort. *Cancer Epidemiology, Biomarkers and Prevention* 10: 17-23 (2001).
- Jaramillo M. C and Zhang D. D. The emerging role of the Nrf2–Keap1 signaling pathway in cancer. *Genes and Development* 27: 2179-2191 (2013).
- Jones W. P and Kinghorn D. Extraction of plant secondary metabolites. In *Natural products isolation, Methods in Molecular Biology*. Humana press: New York, New York; 2012, 864: 341- 366.
- Joyner P. M, Waters A. L, Williams R. B, Powell D. R, Janakiram N. B, Chinthalapally V, Rao C. V and Cichewicz R. H. Briarane Diterpenes Diminish COX-2 Expression in Human Colon Adenocarcinoma Cells. *Journal of Natural Products* 74: 857– 861 (2011).
- Kang M, Kobayashi A, Wakabayashi N, Kim S and Yamamoto M. Scaffolding of Keap1 to the actin cytoskeleton controls the function of Nrf2 as key regulator of cytoprotective phase 2 genes. *Proceedings of the National Academy of Sciences* 101(7): 2046–2051 (2004).
- Kannen V, Zanette D. L, Fernandes C. R, Ferreira F. R, Marini T, Carvalho M. C, Brandão M. L, Junior J. E, Mauad F. M, Silva Jr W. A, Stopper H and Garcia S.B. High fat diet causes an imbalance in the colonic serotonergic system promoting adipose tissue enlargement and dysplasia in rats. *Toxicology Letters* 213: 135– 141 (2012).

- Kanuri G and Bergheim I. In Vitro and In Vivo models of Non- alcoholic fatty liver disease (NAFLD). *International Journal of Molecular Sciences* 14: 11963-11980 (2013).
- Kasdagly M, Radhakrishnan S, Reddivari L, Rao V. D. N and Vanamala J. Colon carcinogenesis: influence of western diet- induced obesity and targeting stem cells using dietary bioactive compounds. *Nutrition* 30 (11- 12): 1242-1256 (2014).
- Karthishwaran K, Mirunalini S, Dhamodharan G, Krishnaveni M and Arulmozhi V. Phytochemical investigation of Methanolic Extract of the leaves of *Pergularia daemia*. *Journal of Biological Sciences* 10 (3): 242- 246 (2010).
- Kitagishi Y, Kobayashi M and Matsuda S. Protection against Cancer with Medicinal Herbs via Activation of Tumor Suppressor. *Journal of Oncology* Article ID 236530, pp. 1- 7 (2012).
- Kobaek- Larsen M, Thorup I, Diederichsen A, Fenger C and Hoitinga M. R. Review of colorectal cancer and its metastases in rodent models: comparative aspects with those in humans. *Comparative medicine* 50 (1): 16- 26 (2000).
- Kozoni B, Tsioulis G, Shiff S and Rigas B. The effect of lithocholic acid on proliferation and apoptosis during the early stages of colon carcinogenesis: differential effect on apoptosis the presence of a colon carcinogen. *Carcinogenesis* 21(5): 999- 1005 (2000).
- Law P. C, Auyeung K. K, Chan L. Y and Ko J. K. Astragalus saponins downregulate vascular endothelial growth factor under cobalt chloride-stimulated hypoxia in colon cancer cells. *BMC Complementary and Alternative Medicine* (12): 160 (2012).
- Leopoldo A. S, Sugizaki M. M, Lima-Leopoldo A. P, Ferreira do Nascimento A, Luvizotto R. A. M, Salome de Campos D. H., Okoshi K, Pai- Silva M. D, Padovani C. R and Cicogna A. C. Cardiac remodeling in a rat model of diet- induced obesity. *Canadian Journal of Cardiology* 26(8): 423- 429 (2010).
- Li A. P. Preclinical in vitro screening assays for drug-like properties. *Drug Discovery Today: Technologies* 2(2): 179- 185 (2005).
- Magaji B. A, Law C, Roslani A. C, Moy F. M, Blazeby J. M, Sagap I and Zakaria J. Health-related quality of life among colorectal cancer patients in Malaysia: a study protocol. *BMC Cancer* 12: 384 (2012).
- Mahaira L. G, Tsimplouli C, Sakellaridis N, Alevizopoulos K, Demetzos C, Han Z, Pantazis P and Dimas K. The labdane diterpene sclareol (labd-14-ene-8, 13-diol) induces apoptosis in human tumor cell lines and suppression of tumor growth in vivo via a p53-independent mechanism of action. *European Journal of Pharmacology* 666: 173–182 (2011).

- Md Zamri N. D, Imam M. U, Abd Ghafar S. A and Ismail M. Antioxidative Effects of Germinated Brown Rice-Derived Extracts on H₂O₂-Induced Oxidative Stress in HepG2 Cells. *Evidence-Based Complementary and Alternative Medicine* Article ID 371907: pp. 1- 11 (2014).
- Melecchi M. I. S, Pe'eres V. F, Dariva C. Zini C. A, Abad F. C, Martinez M. M and Caramaño E. B. Optimization of the sonication extraction method of *Hibiscus tiliaceus* L. flowers. *Ultrasonic Sonochemistry* 13: 242–250 (2006).
- Ministry of Health Malaysia (MOH). *Malaysian dietary guidelines*. National coordinating committee on food and nutrition (2010).
- Moghimi-Dehkordi B and Safaee A. An overview of colorectal cancer survival rates and prognosis in Asia. *World Journal of Gastrointestinal Oncology* 4(4): 71-75 (2012).
- Mollah M. L, Park D. K and Park H. *Cordyceps militaris* grown on germinated soybean induces G2/ M cell cycle arrest through down regulation of cyclin B1 and Cdc25c in human colon cancer HT- 29 cells. *Evidence-Based Complementary and Alternative Medicine* 2012: 1-7 (2012).
- Nairooz S, Ibrahim S. H, Omar S. M. M and Affan M. Structural Changes of the Colonic Mucosa Induced by Orlistat: Experimental Study. *Egyptian Journal of Histology* 33 (4): 635 – 648 (2010).
- Neergheen V. S, Bahorun T, Taylor E. W, Jen L and Aruoma O. I. Targeting specific cell signaling transduction pathways by dietary and medicinal phytochemicals in cancer chemoprevention. *Toxicology* 278: 229- 241 (2010).
- Nobili S, Lippi D, Witort E, Donnini M, Bausi, L, Mini E and Capaccioli S. Natural compounds for cancer treatment and prevention. *Pharmacological Research* 59: 365–378 (2009).
- Ortiz L. M. G, Tillhon M, Parks M., Dutto I, Prosperi E, Savio M, Arcamone A. G, Buzzetti F, Lombardi P and Scovassi A. I. Multiple Effects of Berberine Derivatives on Colon Cancer Cells. *BioMed Research International* Article ID 924585, 12 pages (2014).
- Pannangpetch P, Laupattarakasem P, Kukongviriyapan V, Kukongviriyapan U, Kongyingyoes B and Aromdee C. Antioxidant activity and protective effect against oxidative hemolysis of *Clinacanthus nutans* (Burm. F) Lindau. *Songklanakarin Journal of Science and technology* 29(1): 1-9 (2007).
- Park J, Ku J and Park S. 2004. Isolation and Culture of Colon Cancer Cell Lines. In *Cancer Cell Culture Methods in Molecular Medicine*. Volume 88. Totowa, New Jersey: Humana press, 2004: 79-92.

- Park E. J and Pezzuto J. M. Botanicals in cancer chemoprevention. *Cancer and Metastasis Reviews* 21: 231–255 (2002).
- Prieto-Hontoria P. L, Perez-Matute P, Fernandez-Galilea M, Bustos M, Martinez J. A and Moreno-Aliaga M. J. Role of obesity-associated dysfunctional adipose tissue in cancer: a molecular nutrition approach. *Biochimica et Biophysica Acta* 1807: 664–678 (2011).
- Perse M and Cerar A. Morphological and molecular alterations in 1, 2-dimethylhydrazine and azoxymethane induced colon carcinogenesis in rats. *Journal of Biomedicine and Biotechnology* 2011: 1- 14 (2011).
- Peterlik M. Role of bile acid secretion in human colorectal cancer. *Wien Med Wochenschr* 158(19–20): 539–541 (2008).
- Picchi M. G, Marques de Mattos A, Barbosa M. R, Duarte C. P, Gandini M, Portari G. V and Jordao A. A. A high- fat diet as a model of fatty liver disease in rats. *Acta Cirurgica Brasileira* 26(2): 25- 30 (2011).
- Pourhoseingholi M. A. Increased burden of colorectal cancer in Asia. *World Journal of Gastrointestinal Oncology* 4(4): 68-70 (2012).
- Raju J, Patlolla J. M, Swamy M. V and Rao C. V. Diosgenin, a Steroid Saponin of *Trigonella foenum graecum* (Fenugreek), Inhibits Azoxymethane-Induced Aberrant Crypt Foci Formation in F344 Rats and Induces apoptosis in HT-29 Human Colon Cancer Cells. *Cancer Epidemiology, Biomarkers & Prevention* 13: 1392-1398 (2004).
- Ramaswamy S, Huang H and Ramarao B. V. Separation and Purification Technologies in Biorefineries. West Sussex : J Wiley; 2013. 608p.
- Ramos S. Cancer chemoprevention and chemotherapy: Dietary polyphenols and signaling pathways. *Molecular Nutrition and Food Research* 52: 507 – 526 (2008).
- Raymond L. J, Deth R. C and Ralston N. V. C. Potential Role of Selenoenzymes and Antioxidant Metabolism in relation to Autism Etiology and Pathology. *Autism Research and Treatment* Article ID 164938: 15 pages (2014).
- Reddy B. S. Novel Approaches to the Prevention of Colon Cancer by Nutritional Manipulation and Chemoprevention. *Cancer, Epidemiology, Biomarkers and Prevention* 9: 239-247 (2000).
- Remmen H. V, Ikeno Y, Hamilton M, Pahlavani M, Wolf N, Thorpe S. R, Alderson N. L, Baynes J. W, Epstein C. J, Huang T, Nelson J, Strong R and Richardson A. Life-long reduction in MnSOD activity results in increased DNA damage and higher incidence of cancer but does not accelerate aging. *Physiological Genomics* 16(1): 29- 37 (2003).

- Russo M, Tedesco I, Oacomino G, Palumbo R, Galano G and Russo G. L. Dietary phytochemicals in chemoprevention of cancer. *Current Medicinal Chemistry - Immunology Endocrine & Metabolic Agents* 5: 61–72 (2005).
- Russo M, Spagnuolo C, Tedesco I and Russo G. L. Phytochemicals in cancer prevention and therapy: Truth or dare? *Toxins* 2: 517- 551 (2010).
- Russo G. L. Ins and outs of dietary phytochemicals in cancer chemoprevention. *Biochemical pharmacology* 74: 533– 544 (2007).
- Saccon R. A, Bunton- Stasyshyn R. K. A, Fisher E. M. C and Fratta P. Is SOD1 loss of function involved in amyotrophic lateral sclerosis? *Brain* 136: 2342-2358 (2013).
- Sakdarat S, Shuyprom A, Ayudhya T. D. N, Waterman P. G and Karagianis G. Chemical composition investigation of the *Clinacanthus nutans* Lindau leaves. *Thai Journal of Phytopharmacy* 13(2): 13- 24 (2006).
- Sakdarat S, Shuyprom A, Pientong C, Ekalaksananan T and Thongchai S. Bioactive constituents from the leaves of *Clinacanthus nutans* Lindau. *Bioorganic and medicinal chemistry* 17: 1857- 1860 (2009).
- Senedese J. M, Alves J. M, Lima I. M. S, Andrade E. A. P, Furtado R. A, Bastos J. K and Tavares D. C. Chemopreventive effect of *Copaifera langsdorffii* leaves hydroalcoholic extract on 1,2-dimethylhydrazine-induced DNA damage and preneoplastic lesions in rat colon. *BMC Complementary and Alternative Medicine* 13: 1–8 (2013).
- Sengottuvelan M, Viswanathan P and Nalini N. Chemopreventive effect of trans-resveratrol – a phytoalexin against colonic aberrant crypt foci and cell proliferation in 1, 2– dimethylhydrazine induced colon carcinogenesis. *Carcinogenesis* 27(5): 1038- 1046 (2006).
- Sengottuvelan M and Nalini N. Dietary supplementation of resveratrol suppresses colonic tumour incidence in 1, 2-dimethylhydrazine-treated rats by modulating biotransforming enzymes and aberrant crypt foci development. *British Journal of Nutrition* 96: 145–153 (2006).
- Shankar M. K and Kiran B. R. Medicinal plants used as an antidiabetic drug in pharmaceutical industry and their conservation: an overview. *International Research Journal of Pharmacy* 3(10): 65- 71 (2012).
- Shureiqi I, Reddy P and Brenner D. E. Chemoprevention: general perspective. *Critical Reviews in Oncology/ Hematology* 33: 157- 167 (2000).
- Siegel D, Gustafson D. L, Dehn D. L, Han J. Y, Boonchoong P, Berliner, L. J and Ross D. NAD(P)H: Quinone Oxidoreductase 1: Role as a Superoxide Scavenger. *Molecular Pharmacology* 65: 1238–1247 (2004).

- Slattery M. L, Edwards S. L, Anderson K and Caan B. Vitamin E and Colon Cancer: Is There an Association? *Nutrition and Cancer* 50(3): 201-206 (1998).
- Sporn M. B and Suh N. Chemoprevention: an essential approach to controlling cancer. *Nature Reviews: Cancer* 2: 537- 543 (2002).
- Steward W. P and Brown K. Cancer chemoprevention: a rapidly evolving field. *British Journal of Cancer* 109: 1–7 (2013).
- Sung J. J. Y, Lau J. Y. W, Goh K. L and Leung W. K. Increasing incidence of colorectal cancer in Asia: implications for screening. *Lancet Oncology* 6: 871-6 (2005).
- Sung J. J. Y, Lau J. Y. W, Young G. P, Sano Y, Chiu H. M, Byeon J. S, Yeoh K. G, Goh K. L, Sollano J, Rerknimitr R, Matsuda T, Wu K. C, Ng S, Leung S. Y, Makharia G, Chong V. H, Ho K. Y, Brooks D, Lieberman D. A and Chan F. K. L. Asia Pacific consensus recommendations for colorectal cancer screening. *Gut* 57: 1166–1176 (2008).
- Takayama T, Goji T, Taniguchi T and Inoue A. Chemoprevention of colorectal cancer – experimental and clinical aspects (Review). *Journal of Medical Investigation* 56: 1-5 (2009).
- Teshima K, Kanako T, Ohtan K, Kasai R, Lhieochaiphant S, Pichensoonthon C and Yamasaki K. Sulfur-containing glucosides from *Clinacanthus nutans*. *Phytochemistry* 488: 831–835 (1998).
- Thun M. J, DeLancey J. O, Center M. M, Jemal A and Ward E. M. The global burden of cancer: priorities for prevention. *Carcinogenesis* 31(1): 100–110 (2010).
- Tiwari P, Kumar B, Kaur M, Kaur G and Kaur H. Phytochemical screening and Extraction: A Review. *Internationale Pharmaceutica Scientia* 1(1): 98-106 (2011).
- Treyzon L, Ohning G and Heber D. Colon cancer. In Heber D, Blackburn G. L, Go V. L. W and Wilner J, editors. *Nutritional Oncology*. 2nd ed. Massachusetts: Elsevier; 2006. p. 423- 435.
- Tsai C. Y, Chen Y. H, Chien Y. W, Huang W. H and Lin S. H. Effect of soy saponin on the growth of human colon cancer cells. *World Journal of Gastroenterology* 16 (27): 3371-3376 (2010).
- Tudek B, Bird R. P and Bruce W. R. Foci of aberrant crypts in the colons of mice and rats exposed to carcinogens associated with foods. *Cancer research* 49: 1236- 1240 (1989).
- Tuntiwachwuttikul P, Pootaeng-on Y, Phansa P and Taylor W. C. Cerebrosides and a monoacylmonogalactosylglycerol from *Clinacanthus nutans*. *Chemical and Pharmaceutical Bulletin* 52: 27–32 (2004).

- Valdés A, García-Cañas V, Rocamora-Reverte L, Gómez-Martínez A, Ferragut J. A and Cifuentes A. Effect of rosemary polyphenols on human colon cancer cells: transcriptomic profiling and functional enrichment analysis. *Genes Nutrition* 8(1): 43–60 (2013).
- Vayghan H. J, Ghadimi S. S and Nourazarian A. R. Preventive and Therapeutic Roles of Ginseng - Focus on Colon cancer. *Asian Pacific Journal of Cancer Prevention*. 15 (2): 585-588 (2014).
- Wactawski-Wende J, Kotchen J. M, Anderson G. L, Assaf A. R, Brunner R. L, O'Sullivan M. J, Margolis K. L, Ockene J. K, Phillips L, Pottern L, Prentice R. L, Robbins J, Rohan T. E, Sarto G. E, Sharma S, Stefanick M. L, Van Horn L, Wallace R. B, Whitlock E, Bassford T, Beresford S. A. A, Black H. R, Bonds D. E, Brzyski R. G, Caan B, Chlebowski R. T, Cochrane B, Garland C, Gass M, Hays J, Heiss G, Hendrix S. L, Howard B. V, Hsia J, Hubbell F. A, Jackson R. D, Johnson K. C, Judd H, Kooperberg C. L, Kuller L. H, LaCroix A. Z, Lane D. S, Langer R. D, Lasser N. L, Lewis C. E, Limacher M. C and Manson J. E. Calcium plus Vitamin D Supplementation and the Risk of Colorectal Cancer. *The New England journal of medicine* 354: 684-96 (2006).
- Warenius H, Kyritsi L, Grierson I, Howarth A, Seabra L, Jones M, Thomas C, Browning P and Whit R. *Anticancer Research* 29: 1933-1942 (2009).
- Watson A. J. M. 2006. An overview of apoptosis and the prevention of colorectal cancer. *Critical reviews in oncology/ hematology* 57: 107- 121 (2006).
- World Health Organization (WHO) cancer fact sheet N°297 Updated February 2014. <http://www.who.int/mediacentre/factsheets/fs297/en/>.
- World Health Organization (WHO) International agency for research on cancer (IARC). *Globacon 2012*: Estimated cancer incidence, mortality and prevalence worldwide in 2012.
- Willett W. C. Diet and Cancer. *The Oncologist* 5: 393- 404 (2000).
- Xiao H. H. X, Simi B, Ju J, Jiang H, Reddy B. S and Yang C. S. Green tea polyphenols inhibit colorectal aberrant crypt foci (ACF) formation and prevent oncogenic changes in dysplastic ACF in azoxymethane-treated F344 rats. *Carcinogenesis* 29 (1): 113–119 (2008).
- Xiu W. P, Akowuah G. A and Jin H C. Acute oral toxicity study of *Clinacanthus nutans* in mice. *International journal of pharmaceutical sciences and research* 3 (11): 4202- 4205 (2012).
- Xu Y, Fang F, Dhar S. K, Bosch A, St. Clair W. H, Kasarskis E. J and St. Clair D.K. Mutations in the SOD2 Promoter Reveal a Molecular Basis for an Activating Protein 2-Dependent dysregulation of Manganese Superoxide Dismutase Expression in Cancer Cells. *Molecular Cancer Research* 6 (12): 1881–93 (2008).

- Yadav R. N. S and Agarwala M. Phytochemical analysis of some medicinal plants. *Journal of Phytology* 3 (12): 10- 14 (2011).
- Yaffe P. B, Power C. M. R, Doucette C. D, Walsh M and Hoskin D. W. Piperine, an alkaloid from black pepper, inhibits growth of human colon cancer cells via G1 arrest and apoptosis triggered by endoplasmic reticulum stress. *Molecular Carcinogenesis* doi: 10.1002/mc.22176 (2014).
- Yong Y. K, Tan J. J, Teh S. S, Mah S. H, Ee G. C. L, Chiong H. S and Ahmad Z. Clinacanthus nutans Extracts Are Antioxidant with Antiproliferative Effect on Cultured Human Cancer Cell Lines. *Evidence-Based Complementary and Alternative Medicine* Article ID 462751: 1- 8 (2013).
- Yusoff H. M, Daud N, Noor N. M and Abdul Rahim A. Participation and Barriers to Colorectal Cancer Screening in Malaysia. *Asian Pacific Journal of Cancer Prevention* 13: 3983-3987 (2012).
- Yu Z, Li W and Liu F. Inhibition of proliferation and induction of apoptosis by genistein in colon cancer HT- 29 cells. *Cancer letters* 215: 159- 166 (2004).
- Yuann J. P, Wang J, Jian H, Lin C and Liang J. Effects of *Clinacanthus nutans* (Burm.f) Lindau leaf extracts on protection of plasmid DNA from riboflavin photoreaction. *MC- Transaction on Biotechnology* 4(1): 45- 58 (2012).
- Yusoff H. M, Daud N, Noor N. M and Abdul Rahim A. Participation and Barriers to Colorectal Cancer Screening in Malaysia. *Asian Pacific Journal of Cancer Prevention* 13: 3983-3987 (2012).
- Zanders M. M, Vissers P. A, Haak H. R and Van de Poll-Franse L. V. Colorectal cancer, diabetes and survival: epidemiological insights. *Diabetes and Metabolism* 40 (2): 120– 127 (2014).
- Zips D, Thames H. D and Baumann M. New anticancer agents: in vitro and in vivo evaluation. *In vivo* 19: 1- 8 (2005).