

PHYSICOCHEMICAL CHARACTERISATION OF EMULSIFIED AGARWOOD OIL USING GEL ELECTROPHORESIS

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FK 2017 78



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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the Degree of Doctor of Philosophy

PHYSICOCHEMICAL CHARACTERISATION OF EMULSIFIED AGARWOOD OIL USING GEL ELECTROPHORESIS

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Quality of the agarwood oil is very subjective and the market price of the oil mainly depends on the quality itself. GC-MS is one of the expensive analytical techniques that is used to determine the quality of the agarwood oil based on the major chemical compound detected. There is no simple and cheaper method established to determine the quality of the agarwood oil based on their physicochemical and electrical properties. Therefore, this dissertation describes three interdependent studies from the beginning in which sample preparation till sample quality were characterised using gel electrophoresis. The first study examined the role of non-ionic surfactant and its critical aggregation concentration (CAC) (0.0167% v/v) on the emulsification process of agarwood oil. The physicochemical properties of emulsified agarwood oil were studied with focus on their droplet size, surface charge, and chemical constituents. Each emulsified droplet consisted of a specific zeta potential value that represented its specific bioactive compounds that were trapped inside the droplets and responded to a specific molecular mass and electrophoretic mobility. Thus, gel electrophoresis technique was performed in the second study to separate the emulsion produced. Due to the nano size (80-100 nm) of emulsified droplet produced at CAC condition, SDSpolyacrylamide gel electrophoresis technique, which is an in-situ, reliable, and sustainable technique, was successfully performed to separate the emulsified oils based on their mobility through the gel in terms of molecular size. The third study used the mechanism and technique studied previously to determine the quality of two inoculated oil emulsions. GC-MS result reported that the high concentration of major compounds present in naturally inoculated agarwood oil led to a large molecular weight. To obtain a vivid visible band in the gel electrophoresis, the optimisation of the gel formulation was conducted to control the necessary pore size with the optimised formulation at 21% resolving gel with 48% acrylamide concentration and 3.33% cross-linker. Electrophoretic mobility of droplet through gel matrix was the main factor in evaluating the quality of emulsified oils in terms of their molecular weight. The high quality oil with large molecular weight (211 Da) migrated slower than the low quality oil with small molecular weight (195 Da) when both samples were subjected to gel electrophoresis.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

PENCIRIAN FIZIKOKIMIA MINYAK GAHARU TEREMULSI MENGGUNAKAN PRINSIP DAN TEKNIK ELEKTROFORESIS GEL

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Kualiti minyak gaharu sangat subjektif dan harga pasaran minyak bergantung terutamanya pada kualiti minyak itu sendiri. GC-MS adalah salah satu teknik beranalisis yang mahal yang digunakan untuk menentukan kualiti minyak gaha<mark>ru berdasarkan sebatian kimia utama y</mark>ang dikesan. Tiada kaedah yang lebih mudah dan murah untuk menentukan kualiti minyak gaharu berdasarkan sifat fizikokimia dan elektrik minyak tersebut. Oleh itu, disertasi ini menerangkan tiga kajian berkaitan penyediaan sampel dan pencirian kualiti sampel menggunakan elektroforesis gel. Kajian pertama mengkaji peranan surfaktan bukan ionik dan kepekatan agregat kritikal (CAC) (0.0167% v/v) terhadap proses pengemulsian minyak gaharu. Ciri-ciri fizikokimia emulsi minyak gaharu dikaji berdasarkan factor-faktor saiz titisan, caj permukaan, dan unsur-unsur kimia. Setiap titisan emulsi mempunyai nilai zeta tentu masing-masing dan mewakili sebatian bioaktif tertentu yang terperangkap di dalam titisan tersebut dan mempunyai jisim molekul dan mobiliti elektroforetik tertentu. Oleh itu, teknik elektroforesis gel dilakukan dalam kajian kedua untuk memisahkan emulsi yang dihasilkan. Oleh kerana saiz nano (80-100 nm) titisan emulsi yang dihasilkan pada keadaan CAC, teknik elektroforesis gel menggunakan SDS-poliakrilamida yang merupakan teknik in-situ, boleh dipercayai, dan mampan, berjaya dilakukan untuk memisahkan minyak teremulsi berdasarkan saiz. Kajian ketiga menggunakan mekanisme dan teknik yang dikaji sebelum ini untuk menentukan kualiti dua emulsi minyak yang dihasilkan. Hasil GC-MS melaporkan bahawa kepekatan tinggi sebatian utama yang terdapat dalam minyak gaharu terinokulasi secara semula jadi menyumbang kepada berat molekul yang besar. Untuk memperoleh jalur yang jelas dalam elektroforesis gel, pengoptimuman formulasi gel telah dijalankan bagi mengawal saiz liang yang diperlukan dengan rumusan yang optimum pada 21% gel pemisahan dengan kepekatan akrilamida sebanyak 48% dan 3.33% bahan pemautsilang. Mobiliti elektroforetik titisan melalui matriks gel merupakan faktor utama dalam menilai kualiti minyak emulsi dari segi berat molekul. Minyak yang berkualiti tinggi dengan berat molekul yang besar (211 Da) bergerak dengan lebih perlahan daripada minyak berkualiti rendah dengan berat molekul yang kecil (195 Da) apabila kedua-dua sampel dianalisis menggunakan elektroforesis gel.



ACKNOWLEDGEMENTS

Completion of this doctoral dissertation was possible with the support of several people. I would like to express my sincere gratitude to all of them. Firstly, I would like to express my deep and sincere gratitude to my supervisor, Dr. Mohd Nazli Naim for the valuable guidance, scholarly inputs, and consistent encouragement that I received throughout the research work. Thank you for allowing me to grow as a research scientist. Your advices on both research and my career have been priceless.

I would also like to thank my supervisor committee members, Associate Professor Dr. Noor Fitrah, Dr. Rabitah Zakaria, and Dr. Noraini Ahmad for serving as my supervisor committee members. I am much thankful for your patience and advices during my PhD work. Much gratitude to my final year students Tey Chia Ying, Ooi Chew Lei, and Too Jun Hao for helping me to complete this work. Your cooperation and inputs allowed me to complete this work successfully.

My sincere appreciation also goes to Assoc. Prof. Dr. Wuled Lenggoro for giving me the opportunity to conduct the attachment program in Tokyo University of Agriculture and Technology, Japan. Collaboration work from the Lenggoro laboratory members was an important contribution to this body of work.

The thesis would not have come to a successful completion without the help that I received from the laboratory technicians. I would like to thank Mr. Raman Morat, Mr. Mohd. Zahiruddin Daud, Mrs. Siti Hajar Zakaria, and Mr. Shahrulrizal Zakaria for helping me to complete this work.

I also would like to thank all my friends who supported me in writing, and incentivised me to strive towards my goal. Finally, I wish to express my sincere thanks to my family for their love, understanding, encouragements, and never-ending support.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS AND NOMENCLATURES

Abbreviations

o/w Oil-in-water

w/o Water-in-oil

HLB Hydrophilic-Lipophilic Balance

IFT Interfacial tension

CMC Critical micelle concentration

CAC Critical aggregation concentration

TEM Transmission electron microscopy

GC-MS Gas chromatography-mass spectrometry

FTIR Fourier transform infrared spectroscopy

PAGE Polyacrylamide gel electrophoresis

SDS Sodium dodecyl sulfate

SDS-PAGE Sodium dodecyl sulfate polyacrylamide gel electrophoresis

NMR Nuclear magnetic resonance

DL Double layer

TAE Tris-acetate-EDTA

TBE Tris-borate-EDTA

DNA Deoxyribonucleic acid

APS Ammonium persulfate

TEMED Tetramethylethylenediamine

HCl Hydrochloric acid

NaOH Sodium hydroxide

ANOVA Analysis of variance

Nomenclatures

Symbol	Description	Unit
F_E	Electric force	N
Q	Charge	С
E	Electric field	N C ⁻¹
F_f	Frictional force	N
f	Frictional coefficient	dimensionless
R	Particle's radius	m
R_f	Relative mobility	dimensionless
M_W	Molecular weight	gmol ⁻¹
M_a	Average molecular weight	g mol-1 / Da
%T	Total concentration of the acrylamide	%
	monomer	
%C	Proportion of the cross-linker bis-crylamide	%

Greek letter

Symbol	Description	Unit
v	Velocity of the particle	ms ⁻¹
η	Viscosity	Pas
$arepsilon_o$	Permittivity of free space	$C^2N^{-1}m^{-2}$
Δρ	Densities difference	kgm ⁻³
μ_e	Electrophoretic mobility	cm ² /Vs
ζ	Zeta potential	mV
$arepsilon_r$	Dielectric constant of the dispersion	dimensionless
σ	Centrifugal force	kgms ⁻²
ω	Interfacial tension	mNm ⁻¹
λ	Wavelength	nm

CHAPTER 1

INTRODUCTION

1.1 Background of study

Aquilaria malaccensis, one of the species in the genus Aquilaria, produces resinimpregnated heartwood that is fragrant and highly prized in international trading (Soehartono & Newton, 2000; Chakrabarty et al., 1994). Formation of resin is due to the physiological reactions of the tree against pathogenic fungal infections from Aspergillus sp., Fusarium bulbiferum, and Penicillium sp. (Dias et al., 2015; Mohamed et al., 2010). There are many names associated with the resinous wood, including agarwood, aloeswood, eaglewood, gaharu, chenxiang, jinkoh, oud, kalamabak, and so on, depending on the tree-growing region. Agarwood is divided into several grades in the market, based on their fragrance strength and longevity, resin content, wood density, and colour (Husni et al., 2013; Barden et al., 2000). The high-grade woods are used in incense mixtures, while the low-grade woods are extracted to produce agarwood oil used in perfumery (Gibson, 1977). However, the high-grade agarwood which grow naturally is hardly available. Furthermore, the resin which is produced naturally by fungal infection happens slowly and very infrequently. As a result, the price of agarwood has tremendously increased in the past few decades and range from a few US dollars per kilo for the lowest quality to over thirty thousand US dollars for the highest quality agarwood and its oil (Akter et al., 2013).

Agarwood oil, which is produced from the resin by a common extraction process such as hydro-distillation, is widely used in aromatherapy as an anxiety treatment or for stress and depression release. Based on the literature, agarwood oil is often graded by trained panels according to the individual perception on basic specifications such as composition, colour, and aroma. However, this method is limited in terms of subjectivity, time consumption, large labour expenses, and poor reproducibility (Ismail et al., 2014). Currently, many studies have been carried out to classify and analyse the quality of agarwood oil based on its chemical composition (Pripdeevech et al., 2011; Mailina et al., 2009; Ishihara et al., 1993b). Several analytical methods such as electronic nose, gas chromatography-mass spectrometry (GC-MS), and nuclear magnetic resonance (NMR) are widely used for analysing and identifying the active compounds (Hidayat et al., 2010; Brage et al., 2007; Pourmortazavi & Hajimirsadeghi, 2007; Mohd. Rosli, 2006). However, the equipment and maintenance costs for each technique are particularly high. Moreover, a highly trained person is required to operate those instruments. The techniques also contribute to samples' destruction. High operating pressure, large organic solvents quantities, high cost standard compounds, and long column preparation time will be required to achieve efficient separation when using chromatography technique (Lord & Bralley, 2008; Lioy, 1991).

In the Southeast Asia region, such as Malaysia, the major chemical compounds of Aquilaria malaccensis oil consist of 4-phenyl-2-butanone, α-bulnesene, αguaiene, agarospirol, ledene oxide-(II), elemol, and γ-eudesmol (Tajuddin et al., 2013). Unfortunately, the chemical components that are responsible for the unique aromatic characteristics of agarwood oil are lipophilic and very sensitive to the environmental factors such as oxygen, temperature, pH, and light, which can shorten their shelf life as well as reduce the quality of the agarwood oil. Due to the high sensitivity of the compounds to the environment, plus the stock depletion in the wild forest environment, a shortage of supply occurs, resulting in annually increasing price of agarwood oil (Lim & Noorainie, 2010). As a solution to the increasing price, active ingredients of the agarwood oil are diluted in a solvent, in which an aqueous solution is more preferable compared to an organic solvent. Oil-in-water emulsification processes allow the delivery of active compounds in aqueous solution and enhance the stability of chemically unstable compounds (Chime et al., 2014). However, the emulsified oil produced may be stable for a few hours to a number of years only, depending on the emulsification process parameter. In order to produce a stable emulsion without coalescence, encapsulation of the emulsified oil with a surfactant or emulsifier is necessary (Mason et al., 2006). The surfactant provides a thin layer at the oil/water interface that enhances droplet protection and provides extra stability in the colloidal system due to electrostatic or steric repulsion (Vaclavik & Christian, 2014). During the preparation of emulsified oil, the addition of a surfactant at critical aggregation concentration (CAC) also assists droplet break-up by reducing the interfacial tension (IFT) value to the maximum and preventing the immediate re-coalescence of newly-formed droplets by rapid adsorption and stabilisation of the newly formed interface of the oil droplet, which facilitates droplet deformation (Mason et al., 2006; Briceno et al., 2001). Furthermore, encapsulation of the bioactive compounds in aqueous media is a viable and efficient approach to increase the physical stability of the bioactive compounds, protecting them against environmental factors such as oxygen and light, facilitating colloidal-based delivery system, improving bioavailability, and increasing the shelf life of the oil. Other than the advantages stated earlier, the emulsified oil droplet in aqueous media also possesses a unique value in terms of electrical properties. The electrical properties are measured in terms of zeta potential value which is defined as electric potential at location of slipping plane in the electrical double layer and commonly used to measure the stability of emulsion against creaming and coalescence.

Gel electrophoresis is a powerful, yet simple, convenient, and inexpensive technique used for separating charged macromolecules such as nucleic acids and proteins through the gel matrix according to their sizes and charges when an electric current is applied (Wrolstad et al., 2005). A small and high-charge molecule will migrate faster through the gel matrix than a large and lowcharge molecule. The electrophoretic mobility of the molecules throughout the gel matrix is described by sieving mechanism (Stellwagen, 1985). Two main types of gels commonly used are agarose and polyacrylamide (PA) gels. Agarose gel is used for separating large-size molecules and has a relatively low resolving power, whereas polyacrylamide gel has a greater resolving power for small-size molecules. Likewise, the average pore size is typically 200–500 nm for agarose gels, and 5–100 nm for polyacrylamide gels (Viovy, 2000). However, currently, there is no study on the assessment and evaluation of the quality of the agarwood oil based on its physicochemical and electrical properties by using gel electrophoresis. Therefore, based on the preliminary approach, sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) was chosen to assess the quality of emulsified agarwood oil droplets produced at CAC by using SDS-PAGE based on their relative mobility either in terms of their size, molecular weight, and/or zeta potential value.

1.2 Problem statements

Agarwood species has been classified as endangered species due to the increasing demand of the agarwood and its extracted valuable essential oil (Lim & Noorainie, 2010). It has been used in cultural and religious activities and for medicinal purposes for hundreds of years in the Middle East, China, and Japan. Currently, agarwood oil is also used as ingredients in perfume, essence, and others products such as soap and shampoo, whereby these products are marketed at prices about ten times more expensive than the common brands of toiletries products. With advancing technology, it is expected that in the future, more new products derived from agarwood oil will be sold in the market. However, serious depletion of the wild resource in the global market leads to a reduced supply of agarwood oil. In this regard, a sustainable effort of emulsification was applied in the handling of the extracted agarwood oil. Emulsification allows delivery of active materials in green solvent, water, which is an inexpensive and innocuous solvent. Emulsification not only helps to reduce the size of bulk oil down to nano size, the process also maintains the optimum concentration of the oil's active ingredient and allows the dilution of the expensive functional materials to be effectively suspended in water, and thus the appropriate dosage of active ingredient can be controlled. However, the emulsion produced is inherently unstable due to the immiscible properties of oil and water, which separate into two layers very quickly after the mixing process. Thus, a surfactant was added to encapsulate agarwood oil within the colloidal system (Wrolstad et al., 2005).

The next problem is creating an in-situ, inexpensive, and sustainable technique to evaluate and assess the quality of essential oil extracted from agarwood. There are many grades of agarwood oil, and the highest quality of the agarwood oil in terms of major chemical compound is extremely expensive. As a matter of fact, the quoted prices for pure agarwood oil, which is one of the most valuable natural products in the world, in average, is 236 dollar per tola (Agarwood and Oud Oil Quarterly Market Price Report - Sep 2015). Conventionally, the grading method of agarwood is based on its physical properties such as aroma, colour, and density, which is very subjective, time consuming, associated with expensive labour cost, and lacks repeatability (Ali et al., 2012; Hidayat et al., 2010; Hung et al., 2014). Currently, many studies have been carried out to analyse the quality of agarwood oil based on its chemical compositions (Ishihara et al., 1993b; Mailina et al., 2009; Pripdeevech et al., 2011). Several analytical methods such as electronic nose, gas chromatography-mass spectrometry (GC-MS), and nuclear magnetic resonance (NMR) are widely used to identify the active compounds (Brage et al., 2007; Hidayat et al., 2010; Mohd Rosli, 2006; Pourmortazavi & Hajimirsadeghi, 2007). However, the equipment and maintenance costs for each technique are particularly high and a highly trained person is required to operate those instruments. Furthermore, quality assessing or evaluating system based on physicochemical and electrical properties of agarwood oil has not yet been extensively studied.

1.3 Objectives of study

The general objective of this study is to assess the quality of agarwood oils using gel electrophoresis via emulsification mechanism based on their relative mobility through gel matrix while the specific objectives of this study are:

- 1. To investigate the ultrasonic parameters and surfactant critical aggregate concentration (CAC) to obtain stable emulsion in terms of interfacial tension, droplet size, and zeta potential, as well as under effect of storage and pH;
- 2. To characterize the physical properties of the emulsified agarwood oil in terms of droplet size, molecular weight, and/or charge using gel electrophoresis separation technique;
- 3. To assess the relative mobility as the quality indicator of emulsified agarwood oil.

1.4 Scope of study

The scope of this study is essentially to compare and assess the quality of commercial agarwood oil (as-received) with different resin inoculation techniques (naturally or artificially) through gel electrophoresis based on their

relative mobility either in terms of their droplet size, molecular weight, and/or zeta potential. Prior to electrophoresis separation, several parameters such as ultrasonication power and time, and surfactant concentration were manipulated to produce the most stable emulsion. Fine and monodispersed oil droplets are preferred in most cases because a homogeneous emulsion can be produced easily and each emulsified droplet is able to entrap an equal amount of bioactive compounds. The effects of pH and stability during sample preparation were also controlled and investigated. The critical aggregation concentration (CAC) was determined at which the emulsified droplets were screened uniformly with diffused counter-ions in aqueous media, so that each commercial oil sample has its own specific zeta potential value based on the oil chemical constituents or oil quality. A particular concern was given to the optimisation of gel pore size by controlling their acrylamide and bis-acrylamide percentage. The visible vivid bands that appeared were then used for the oil quality comparison. As a result, the droplet that responded to a specific molecular mass and electrophoretic mobility can be separated using SDS-PAGE.

1.5 Rationale and significant

Due to the problem statements stated above, especially the analytical cost of the GC-MS technique, the concern of finding an in-situ, inexpensive, and sustainable green technique that can be used to evaluate the quality of agarwood oils based on their physicochemical and electrical properties as well as major compounds are the crucial parts of this study. Prior to the separation, emulsification mechanism for sample preparation in terms of size, molecular weight, and/ or charge will be investigated. The discovery of such technique or any research leading to that discovery would concrete all steps towards solving the problems stated above. It is believed that the technique can be more widely used for the separation of other oils, and provides a new insight into how electrophoresis can be applied in emulsion science.

1.6 Hypothesis

Here, the hypothesis proposed is that the addition of surfactant Tween 80 not only stabilises the emulsified oil, but also creates a thin layer that protects the oil droplets against various pH conditions. It is also hypothesised that SDS coats the emulsified droplets with a uniform negative charge, and the intrinsic charges of emulsified droplets become negligible when compared to the negative charges contributed by SDS. Thus, SDS-coated droplets possess a uniform charge density, which is the same net charge per unit weight. Therefore, there will be no differential migration based on charge and the separation mobility is hypothesised to be based on the molecular weight of the droplets. The zeta potential values in the system were also hypothesised

to be derived originally from the individual oil droplet charge and were independent from non-ionic surfactant (Tween 80) effects. It is speculated that zeta potential values represented the specific bioactive compounds trapped in the droplets, and responded to a specific molecular mass and electrophoretic mobility so that separation using SDS-PAGE can be performed. High-quality emulsified oil with greater major chemical compounds that contribute to the aromatic characteristics is hypothesised to have larger molecular weight. Thus, high-quality emulsified oil migrates slower through the gel medium when an electric field is applied.



REFERENCES

- Adelina, N., Harum, F., Schmidt, L. H., & Jøker, D. (2004). Aquilaria malaccensis Lam. *Seed Leaflet*, 103, 1–2.
- Affandi, M. M. R. M. M., Julianto, T., & Majeed, A. (2011). Development and stability evaluation of astaxanthin nanoemulsion. *Asian Journal of Pharmaceutical and Clinical Research*, 4(1), 142–148.
- Agarwood. (2008). Retrieved December 3, 2016, from https://en.wikipedia.org/w/index.php?title=Agarwood&oldid=752164 327
- Agarwood and Oud Oil Quarterly Market Price Report Sep 2015. (2015). Singapore: Sustainable Asset Management.
- Agarwood insence buds. (2014). Retrieved December 3, 2016, from http://specialfoodvietnam.com/products/agarwood-insence-buds/384.html
- Agarwood Market Demand. (2010). Retrieved January 28, 2016, from http://www.gaharuonline.com/articles/agarwood_market_demand.ht
- Ahmad, K., Ho, C. C., Fong, W. K., & Toji, D. (1996). Properties of palm oil-inwater emulsions stabilized by nonionic emulsifiers. *Journal of Colloid and Interface Science*, 181(2), 595–604.
- Ahmed, E. M. (2015). Hydrogel: Preparation, characterization, and applications: A review. *Journal of Advanced Research*, 6(2), 105–121.
- Ahmed, H. (2004). *Principles and reactions of protein extraction, purification, and characterization*. Boca Raton: CRC Press.
- Aidar, E., Sigaud-Kutner, T. C. S., Nishihara, L., Schinke, K. P., Braga, M. C. C., Farah, R. E., & Kutner, M. B. B. (1997). Marine phytoplankton assays: Effects of detergents. *Marine Environmental Research*, 43(1), 55–68.
- Akrarapholchote, S. (2008). Krisna leaf tea. First News and MEdia Group, Bangkok, pp 66-67.
- Al-Sabagh, A. M., Emara, M. M., Noor El-Din, M. R., & Aly, W. R. (2011). Formation of water-in-diesel oil nano-emulsions using high energy method and studying some of their surface active properties. *Egyptian Journal of Petroleum*, 20(2), 17–23.

- Alaimo, M. H., & Kumosinski, T. F. (1997). Investigation of hydrophobic interactions in colloidal and biological systems by molecular dynamics simulations and NMR spectroscopy. *Langmuir*, 13(7), 2007–2018.
- Alberts, B., Wilson, J., & Hunt, T. (2008). *Molecular biology of the cell* (5th ed.). New York: Garland Science.
- Ali, N. A. M., Ismail, N., & Taib, M. N. (2012). Analysis of agarwood oil (Aquilaria Malaccensis) based on GC-MS data. In *IEEE 8th International Colloquium on Signal Processing and its Applications* (pp. 470–473). 23-25 March 2012, Malacca: IEEE.
- Allen, T. (2003). *Powder Sampling and Particle Size Determination*. New York: Elsevier Science.
- ALOthman, Z. (2012). A Review: Fundamental Aspects of Silicate Mesoporous Materials. *Materials*, 5(12), 2874–2902.
- Analyses of proposals to amend the CITES appendices. (1994). IUCN Species Survival Commission and the TRAFFIC Network for the Ninth Meeting of the Conference of the Parites to CITES. Gland, Switzerland.
- Argenta, D. F., de Mattos, C. B., Misturini, F. D., Koester, L. S., Bassani, V. L., Simões, C. M. O., & Teixeira, H. F. (2014). Factorial design applied to the optimization of lipid composition of topical antiherpetic nanoemulsions containing isoflavone genistein. *International Journal of Nanomedicine*, 9, 4737–4747.
- Ariyaprakai, S. (2007). Mechanisms of oil transport by micelles in oil-in-water emulsions. Ann Arbor: ProQuest.
- Ariyaprakai, S., & Dungan, S. R. (2007). Solubilization in monodisperse emulsions. *Journal of Colloid and Interface Science*, 314(2), 673–82.
- Artificial inoculation. (2012). Retrieved December 20, 2016, from https://gaharujinkou.wordpress.com/cultivated-agarwood/inoculation-method/
- Atkinson, A. (2015). Essential Oils for beauty, wellness, and the home: 100 natural, non-toxic recipes for the beginner and beyond. New York: Skyhorse Publishing, Inc.
- Aveyard, R., & Haydon, D. A. (1973). An introduction to the principles of surface chemistry. London: Cambridge University Press.
- Azema, N. (2006). Sedimentation behaviour study by three optical methods granulometric and electrophoresis measurements, dispersion optical analyser. *Powder Technology*, 165(3), 133–139.

- Baglimieri, C., Cenciarini, J., Fernex, F., Pucci, R., & Vassiere, R. (1980). Problems of storage of various substances found in the interstitial waters of the surface sediments of the French continental shelf. *Prog Water Technol*, 12(1), 79–87.
- Bagwe, R. P., Kanicky, J. R., Palla, B. J., Patanjali, P. K., & Shah, D. O. (2001). Improved drug delivery using microemulsions: rationale, recent progress, and new horizons. *Critical Reviews in Therapeutic Drug Carrier Systems*, 18(1), 77–140.
- Bandoni, A. L. (2000). Los recursos vegetales aromáticos en Latinoamérica: su aprovechamiento industrial para la producción de aromas y sabores. La Plata: CYTED.
- Barden, A., Anak, N. A., Mulliken, T., & Song, M. (2000). Heart of the Matter: Agarwood use and trade and CITES implementation for Aquilaria malaccensis. Cambridge: TRAFFIC International.
- Barril, P., & Nates, S. (2012). Introduction to agarose and polyacrylamide gel electrophoresis matrices with respect to their detection sensitivities. In S. Magdeldin (Ed.), *Gel Electrophoresis Principles and Basics* (pp. 1–14). Rijeka: InTech.
- Becher, P. (2001). *Emulsions: theory and practice* (3rd ed.). Washington: American Chemical Society.
- Beniwal, B. S. (1989). Silvical characteristics of Aquilaria agallocha Roxb. *Indian Forester*, 115(1), 17–21.
- Bera, A., Mandal, A., & Guha, B. B. (2014). Synergistic effect of surfactant and salt mixture on interfacial tension reduction between crude oil and water in enhanced oil recovery. *Journal of Chemical & Engineering Data*, 59(1), 89–96.
- Besheer, A., Vogel, J., Glanz, D., Kressler, J., Groth, T., & Mäder, K. (2009). Characterization of PLGA nanospheres stabilized with amphiphilic polymers: hydrophobically modified hydroxyethyl starch vs pluronics. *Molecular Pharmaceutics*, 6(2), 407–415.
- Besnard, L., Protat, M., Malloggi, F., Daillant, J., Cousin, F., Pantoustier, N., Guenoun, P., Perrin, P. (2014). Breaking of the Bancroft rule for multiple emulsions stabilized by a single stimulable polymer. *Soft Matter*, *10*, 7073–7087.
- Bhowmik, G., & Bose, S. (2011). *Analytical Techniqs in Biotechnology (A complete laboratory manual)*. New Delhi: Tata McGraw-Hill Education.

- Binks, B. P. (1998). *Modern Aspects of Emulsion Science*. Cambridge: Royal Society of Chemistry.
- BioRad Laboratories. (2012). A guide to polyacrylamide gel electrophoresis and detection. *Bulletin 6040 Rev B*. USA. Retrieved from http://www.biorad.com/webroot/web/pdf/lsr/literature/Bulletin_6040.pdf
- Blanchette, R. A. (2006). Sustainable agarwood production in Aquilaria trees. Retrieved from http://forestpathology.cfans.umn.edu/agarwood.htm
- Borzenkov, M., & Hevus, O. (2014). Surface Active Monomers: Synthesis, Properties, and Application. New York: Springer.
- Bouchemal, K., Briançon, S., Perrier, E., & Fessi, H. (2004). Nano-emulsion formulation using spontaneous emulsification: solvent, oil and surfactant optimisation. *International Journal of Pharmaceutics*, 280(1–2), 241–51.
- Brage, M. E. M., Meireles, M., & Angela, A. (2007). Accelerated solvent extraction and fractioned extraction to obtain the curcuma longa volatile oil and oleoresin. *Journal of Food Process Engineering*, 30(4), 501–521.
- Briceno, M. I., Salager, J. L., & Bracho, C. L. (2001). Heavy hydrocarbon emulsions making use of the state of the art in formulation engineering. In J. Sjöblom (Ed.), *Encyclopedic Handbook of Emulsion Technology* (pp. 455–495). New York: CRC Press.
- Buckton, G. (2000). *Interfacial Phenomena in Drug Delivery and Targeting*. CRC Press.
- Burkill, I. H., Birtwistle, W., Foxworthy, F. W., Scrivenor, J. B., & Watson, J. G. (1966). A dictionary of the economic products of the Malay peninsula. (Vol. II(I-Z)). London: Published on behalf of the governments of the Straits settlements and Federated Malay states by the Crown agents for the colonies.
- Caballero, B., Finglas, P. M., & Toldra, F. (2015). *Encyclopedia of food and health*. Waltham: Academic Press.
- Carroll, B. J. (1986). Solubilisation of two-component oil mixtures by micellar surfactant solutions. *Journal of the Chemical Society, Faraday Transactions 1: Physical Chemistry in Condensed Phases, 82*(10), 3205.
- Chakrabarty, K., Kumar, A., & Menon, V. (1994). *Trade in agarwood*. New Delhi: TRAFFIC-India.
- Chen, B. H., Miller, C. A., & Garrett, P. R. (1997). Rates of solubilization of triolein into nonionic surfactant solutions. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 128(1–3), 129–143.

- Chen, H. Q., Wei, J. H., Yang, J. S., Zhang, Z., Yang, Y., Gao, Z. H., Sui, C., & Gong, B. (2012). Chemical constituents of agarwood originating from the endemic genus Aquilaria plants. *Chemistry & Biodiversity*, 9(2), 236–250.
- Chetpattananondh, P. (2012). Overview of the agarwood oil industry. In *International Federation of Essential Oils and Aroma Trades (IFEAT)* (pp. 131–138). 4-8 November 2012, Singapore: IFEAT.
- Chime, S. A., Kenechukwu, F. C., & Attama, A. A. (2014). Nanoemulsions-advances in formulation, characterization and applications in drug delivery. In A. D. Sezer (Ed.), *Application of Nanotechnology in Drug delivery* (p. 552). Rijeka: InTech.
- Chitre, T., Bhutada, P., Nandakumar, K., Somani, R., Miniyar, P., Mundhada, Y., Gore, S., & Jain, K. (2007). Analgesik and anti-imflamatory activity of heartwood of Aquilaria agallocha in laboratory animal. *Pharmacologyonline* 1, 288–298.
- Choi, S. J., Decker, E. A., Henson, L., Popplewell, L. M., & McClements, D. J. (2010). Inhibition of citral degradation in model beverage emulsions using micelles and reverse micelles. *Food Chemistry*, 122(1), 111–116.
- Christian, G. D. (2016). *Analytical Chemistry* (7th ed.). New York: Content Technologies Inc.
- Chua, L. S. L. (2008). Agarwood (Aquilaria Malaccensis) in Malaysia. NDF Workshop case studies, Mexico: Forest Research Institute Malaysia.
- Cifuentes, A., & Poppe, H. (1997). Behavior of peptides in capillary electrophoresis: Effect of peptide charge, mass and structure. *Electrophoresis*, 18(12–13), 2362–2376.
- Cooper, T. G. (1977). The tools of biochemistry. New York: Wiley.
- Copeland, R. A. (1994). *Methods for protein analysis: A practical guide for laboratory protocols.* New York: Chapman and Hall.
- Cserháti, T., Forgács, E., & Oros, G. (2002). Biological activity and environmental impact of anionic surfactants. *Environment International*, 28(5), 337–348.
- Dahham, S. S., Tabana, Y. M., Ahmed Hassan, L. E., Khadeer Ahamed, M. B., Abdul Majid, A. S., & Abdul Majid, A. M. S. (2016). In vitro antimetastatic activity of agarwood (Aquilaria crassna) essential oils against pancreatic cancer cells. *Alexandria Journal of Medicine*, 52(2), 141–150.
- Das, S., & Dash, H. R. (2014). *Microbial biotechnology- A laboratory manual for bacterial systems*. Rourkela: Springer.

- Davey, J., & Lord, J. M. (2003). Essential cell biology volume 1: Cell structure. New York: Oxford University Press.
- de Morais, J. M., dos Santos, O. D. H., Delicato, T., & da Rocha-Filho, P. A. (2006). Characterization and evaluation of electrolyte influence on canola oil/water nano-emulsion. *Journal of Dispersion Science and Technology*, 27(7), 1009–1014.
- de Villiers, M. M., Aramwit, P., & Kwon, G. S. (2008). *Nanotechnology in drug delivery*. New York: Springer Science & Business Media.
- Dehnul oud (Agarwood oil). (2010). Retrieved December 3, 2016, from http://www.oudline.com/dehnul_oud.php
- Destombe, C., Wattier, R., Bulke, D., & Valero, M. (1998). *Delimitation of species and population genetic structure in Gracilaria verrucosa: consequences for cultivation*. France: Lille Univ., Villeneuve d'Ascq. Lab. de Genetique et Evolution des Populations Vegetales.
- Dey, J., Sultana, N., Kumar, S., Aswal, V. K., Choudhury, S., & Ismail, K. (2015). Controlling the aggregation of sodium dodecylsulphate in aqueous poly(ethylene glycol) solutions. *RSC Adv.*, *5*(91), 74744–74752.
- Dias, A. A. P., Fernando, K. M. E. P., & Subasinghe, S. M. C. U. P. (2015). Optimisation of fungal inoculum for inoculation of Gyrinops walla for inducing agarwood formation. *Proceedings of 20th International Forestry and Environment Symposium*. 16-17 October 2015, Sri Lanka: Department of Forestry and Environmental Science, University of Sri Jayewardenepura.
- Divan, A., & Royds, J. (2013). *Tools and techniques in biomolecular science*. Oxford: Oxford University Press.
- Donovan, D. G., & Puri, R. K. (2004). Learning from traditional knowledge of non-timber forest products: Penan Benalui and the autoecology of Aquilaria in Indonesian Borneo. *Ecology and Society*, 9(3), 3.
- Dynamic light scattering. (2010). Retrieved December 3, 2016, from https://en.wikipedia.org/wiki/Dynamic_light_scattering
- Dynamic light scattering: an introduction in 30 minutes. (n.d.). Retrieved December 3, 2016, from www.malvern.com.
- Eid, A. M. M., Elmarzugi, N. A., & El-Enshasy, H. A. (2013). Preparation and evaluation of olive oil nanoemulsion using sucrose monoester. *International Journal of Pharmacy and Pharmaceutical Sciences*, 5(SUPPL 3), 434–440.
- El-Mansi, E. M. T., Bryce, C. F. A., Demain, A. L., & Allman, A. R. (2006).

- Fermentation microbiology and biotechnology. (2nd ed.). New York: CRC Press.
- Fernandez, P., André, V., Rieger, J., & Kühnle, A. (2004). Nano-emulsion formation by emulsion phase inversion. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 251(1–3), 53–58.
- Fisher, J. P., Mikos, A. G., Bronzino, J. D., & Peterson, D. R. (2012). *Tissue engineering: principles and practices*. Boca Raton: CRC Press.
- Florence, A. T., & Attwood, D. (2015). *Physicochemical principles of pharmacy: In manufacture, mormulation and clinical use* (6th ed.). London: Pharmaceutical Press.
- Freifelder, D. (1985). *Principles of physical chemistry: With applications to the biological sciences* (2nd ed.). University of Michigan: Jones and Bartlett Publishers.
- Fukuzumi, H. (2012). Studies on structures and properties of TEMPO-oxidized cellulose nanofibril films (PhD's thesis). University of Tokyo.
- García, C. D., Chumbimuni-Torres, K. Y., & Carrilho, E. (2013). Capillary electrophoresis and microchip capillary electrophoresis: principles, applications, and limitations. Hoboken: John Wiley & Sons.
- Garrett, R. H., & Grisham, C. M. (2012). *Biochemistry* (5th ed.). California: Brooks/Cole, Cengage Learning.
- Gaus, H. J., Beck-Sickinger, A. G., & Bayer, E. (1993). Optimization of capillary electrophoresis of mixtures of basic peptides and comparison with HPLC. *Analytical Chemistry*, 65(10), 1399–1405.
- Geed, S. R., Singh, R. P., & Rai, B. N. (2014). Recent development of extraction processes and extraction of essential oil from Coriander by clean technology. *International Journal of Engineering and Advanced Technology*, 3(6), 58–63.
- Ghosh, T. K., & Jasti, B. R. (2004). Theory and practice of contemporary pharmaceutics. (Vol. 23). Boca Raton: CRC Press.
- Ghosh, V., Mukherjee, A., & Chandrasekaran, N. (2013). Ultrasonic emulsification of food-grade nanoemulsion formulation and evaluation of its bactericidal activity. *Ultrasonics Sonochemistry*, 20(1), 338–344.
- Ghoshal, S. (2009). Fundamentals of bioanalytical techniques and instrumentation. New Delhi: PHI Learning Pvt. Ltd.
- Gibson, I. A. S. (1977). The role of fungi in the origin of oleoresin deposits

- (agaru) in the wood of Aquilaria agallocha Roxb. Bano Biggyan Patrika, 6(1), 16–26.
- Gilbert, J. C., & Martin, S. F. (2010). *Experimental organic chemistry: A miniscale and microscale approach* (5th ed.). Boston: Cengage Learning.
- Glasel, J. A., & Deutscher, M. P. (1995). *Introduction to biophysical methods for protein and nucleic acid research*. New York: Academic Press.
- Gokhale, S. ., Gattani, S. G., & Bakliwal, S. R. (2007). Practical pharmaceutics for Rajiv Gandhi Universtiry of health sciences, Karnataka (4th ed.). Shivaji Nagar: Nirali Prakashan.
- Gratzfeld, J., & Tan, B. (2008). Agarwood saving a precious and threatened resource. *Botanic Gardens Conservation International*, 5(1), 27–29.
- Grossman, P. D., & Colburn, J. C. (2012). Capillary electrophoresis: theory and practice. California: Academic Press.
- Guilliatt, A. M. (2002). Agarose and polyacrylamide gel electrophoresis. *Methods in Molecular Biology*, 187, 1–11.
- Guo, L., Colby, R. H., Lin, M. Y., & Dado, G. P. (2001). Micellar structure changes in aqueous mixtures of nonionic surfactants. *Journal of Rheology*, 45(5), 1223–1243.
- Gupta, P. K. (2008). *Molecular Biology and Genetic Engineering*. New Delhi: Rastogi Publications.
- Gupta, P. K., & Cannon, J. B. (2000). Emulsions, microemulsions, and lipid-based drug delivery systems for drug solubilization and delivery. In R. Liu (Ed.), *Water-Insoluble Drug Formulation* (pp. 169–212). Boca Raton: CRC Press.
- Hajar, A. (2013). *Agarwood: the most expensive wood fragrance*. Abu Hajar. Retrieved from https://books.google.com/books?id=L6rY-kSfNd4C&pgis=1
- Handa, S. S., Khanuja, S. P. S., Longo, G., & Rakesh, D. D. (2008). *Extraction technologies for medicinal and aromatic plants*. Trieste: International centre for science and high technology.
- Hansen, E. (2000). Stranger in the forest: on foot across Borneo. New York: Vintage.
- Harrison, R. G., Todd, P., Rudge, S. R., & Petrides, D. P. (2015). *Bioseparations science and engineering: topics in chemical engineering* (2nd ed.). New York: Oxford University Press.

- Hartfelder, K., Bitondi, M. M. G., Brent, C. S., Guidugli-Lazzarini, K. R., Simões, Z. L. P., Stabentheiner, A., Tanaka, E. D., & Wang, Y. (2013). Standard methods for physiology and biochemistry research in Apis mellifera. *Journal of Apicultural Research*, 52(1), 1–48.
- Hashim, Y. Z. H. Y., Ismail, N. I., & Abbas, P. (2014). Analysis of chemical compounds of agarwood oil from different species by gas chromatography mass spectrometry (GCMS). *International Islamic University Malaysia Engineering Journal*, 15(1), 55–60.
- Hasnida, H. N., Aziah, M. Y., Salbiah, M., Fadhilah, Z., & Haliza, I. (2001). Multiplication of shoots from in vitro germinated seedlings of Eurycoma longifolia and Aquilaria malaccensis. In *Tropical forestry research in the new millennium: meeting demands and challenges. Proceedings of The International Conference on Forestry and Forest Products Reearch (CFFPR 2001)* (pp. 269–276). 1-3 October 2001, Kuala Lumpur: Forest Research Institute Malaysia (FRIM).
- Heller, M. J. (1992). Methods for the electrophoretic separation of nucleic acids and other linear macromolecules in gel media with restrictive pore diameters. Google Patents.
- Heuveling Van Beek, H., & Phillips, D. (1999). *Agarwood: trade and CITES implementation in Southeast Asia*. Malaysia: TRAFFIC Southeast Asia.
- Hidayat, W., Shakaff, A. Y. M., Ahmad, M. N., & Adom, A. H. (2010). Classification of agarwood oil using an electronic nose. *Sensors*, 10(5), 4675–4685.
- High quality agarwood tea for better health & life. (2009). Retrieved January 27, 2016, from http://yuricorpagarwoodtea.blogspot.my/2009/10/agarwood-tea-for-enhanced-health-living.html
- Hill, J. W., McCreary, T. W., & Kolb, D. K. (2016). *Chemistry for changing times* (13th ed.). New York: Content Technology Inc.
- History and health of gaharu. (2014). Retrieved August 27, 2017, from https://vietnamagarwood.wordpress.com/tag/benefits-of-agarwood-tea/
- Hou, D. (1960). Thymelaeaceae. Flora Malesiana-Series I, Spermatophyta, 6(1), 1-48.
- Hsu, J. P., & Nacu, A. (2003). Behavior of soybean oil-in-water emulsion stabilized by nonionic surfactant. *Journal of Colloid and Interface Science*, 259(2), 374–381.

- Hung, C. H., Lee, C. Y., Yang, C. L., & Lee, M. R. (2014). Classification and differentiation of agarwoods by using non-targeted HS-SPME-GC/MS and multivariate analysis. *Analytical Methods*, 6(18), 7449–7456.
- Hunter, R. J. (1981). *Zeta Potential in Colloid Science. Principles and applications*. New York: Academic Press.
- Husni, S. S., Mailina, J., Sahrim, L., Majid, J. A., & Faridz, Z. M. (2013). Classification of agarwood (gaharu) by resin content. *Journal of Tropical Forest Science*, 25(2), 213–219.
- Husnu Can Baser, K., & Buchbauer, G. (2015). *Handbook of essential oils: Science, technology, and applications* (2nd ed.). Boca Raton: CRC Press.
- Introduction to Polyacrylamide Gels. (n.d.). Retrieved December 3, 2016, from http://www.bio-rad.com/en-my/applications-technologies/introduction-polyacrylamide-gels
- Ishihara, M., Tsuneya, T., & Uneyama, K. (1993a). Components of the agarwood smoke on heating. *Journal of Essential Oil Research*, 5(4), 419–423.
- Ishihara, M., Tsuneya, T., & Uneyama, K. (1993b). Components of the volatile concentrate of agarwood. *Journal of Essential Oil Research*, 5(3), 283–289.
- Ismail, N., Mohd Ali, N., Jamil, M., Rahiman, M. H. F., Tajuddin, S. N., & Taib, M. N. (2014). A review study of agarwood oil and its quality analysis. *Jurnal Teknologi*, 68(1), 37–42.
- Ismail, N., Nor Azah, M. A., Jamil, M., Rahiman, M. H. F., Tajuddin, S. N., & Taib, M. N. (2013). Analysis of high quality agarwood oil chemical compounds by means of SPME / GC-MS and Z-score technique. *Malaysian Journal of Analytical Sciences*, 17(3), 403–413.
- Izquierdo, P., Esquena, J., Tadros, T. F., Dederen, J. C., Feng, J., Garcia-Celma, M. J., Azemar, N., & Solans, C. (2004). Phase behavior and nano-emulsion formation by the phase inversion temperature method. *Langmuir*: *The ACS Journal of Surfaces and Colloids*, 20(16), 6594–6598.
- Jacobs, C., Kayser, O., & Müller, R. H. (2000). Nanosuspensions as a new approach for the formulation for the poorly soluble drug tarazepide. *International Journal of Pharmaceutics*, 196(2), 161–164.
- Jain, N., Trabelsi, S., Guillot, S., McLoughlin, D., Langevin, D., Letellier, P., & Turmine, M. (2004). Critical aggregation concentration in mixed solutions of anionic polyelectrolytes and cationic surfactants. *Langmuir*, 20(20), 8496–8503.
- Jamahseri, N. F., Najwa, M., Rodhi, M., Zulkarnain, H., Husain, N. C., Fakhri,

- A., & Masruddin, S. (2014). FTIR analysis of phenolic compound as pancreatic lipase inhibitor from inoculated aquilaria malaccensis. *The Malaysian Journal of Analytical Sciences*, 18(3), 683–689.
- Josephine, M. (2013). Back to basics with essential oils. New York: BookBaby.
- Joshi, H. C., Pandey, I. P., Kumar, A., & Gar, N. (2012). A study of various factors determining the stability of molecules. *Advances in Pure and Applied Chemistry*, *1*(1), 7–11.
- Kabalnov, A. S., Pertzov, A. V., & Shchukin, E. D. (1987). Ostwald ripening in two-component disperse phase systems: Application to emulsion stability. *Colloids and Surfaces*, 24(1), 19–32.
- Kendrew, J. (1994). The encylopaedia of molecular biology. Oxford: Blackwell Science Ltd.
- Kessler, P. J. A., & Sidiyas, K. (1994). *Trees of the Balikpapan-Samarinda area*, East *Kalimantan*, *Indonesia: A manual to 280 selected trees* (*Tropenbos Kalimantan series*). Wageningen: The Tropenbos Foundation.
- Khalil, A. S., Rahim, A. A., Taha, K. K., & Abdallah, K. B. (2013). Characterization of methanolic extracts of agarwood leaves. *Journal of Applied and Industrial Sciences*, 1(3), 78–88.
- Klinkesorn, U., Sophanodora, P., Chinachoti, P., & McClements, D. . (2004). Stability and rheology of corn oil-in-water emulsions containing maltodextrin. *Food Research International*, *37*(9), 851–859.
- Krotz, A., & Ravikumar, V. (2003). Purification of oligonucleotides. U.S.:

 United States Patent. Retrieved from http://www.google.com/patents/US20030153742
- Kumar, V., Koshy, A. S., Prabeesh, E., & Rema, A. K. (2008). *Biochemistry for nursing and healthcare*. New Delhi: BI Publications Pvt Ltd.
- Lafrankie, J. V. (1994). population dynamics of some tropical trees that yield non-timber forest products. *Economic Botany*, 48(3), 301–309.
- le Maire, M., Champeil, P., & Møller, J. V. (2000). Interaction of membrane proteins and lipids with solubilizing detergents. *Biochimica et Biophysica Acta*, 1508, 86–111.
- Lee, M. S. (2012). *Mass spectrometry handbook*. New Jersey: John Wiley & Sons.
- Leong, T. S. H., Wooster, T. J., Kentish, S. E., & Ashokkumar, M. (2009). Minimising oil droplet size using ultrasonic emulsification. *Ultrasonics Sonochemistry*, 16(6), 721–727.

- Lerman, L. S., & Frisch, H. L. (1982). Why does the electrophoretic mobility of DNA in gels vary with the length of the molecule? *Biopolymers*, 21(5), 995–997.
- Lewis, L. A. (2010). Cyanoacrylate Fuming Method. In R. Ramotowski (Ed.), Lee and Gaensslen's Advances in Fingerprint Technology (3rd ed., p. 263). New York: CRC Press.
- Lieleg, O., Lieleg, C., Bloom, J., Buck, C. B., & Ribbeck, K. (2012). Mucin Biopolymers As Broad-Spectrum Antiviral Agents. *Biomacromolecules*, 13(6), 1724–1732.
- Lioy, P. J. (1991). *Human exposure assessment for airborne pollutants: advances and opportunities.* Washington: National Academy of Sciences.
- Lim, H. F., Mamat, M. P., & Chang, Y. S. (2007). Production, use and trade of gaharu in peninsular Malaysia. In *International Economic Conference on Trade & Industry* (pp. 1–10). 3-5 December 2007, Penang: Faculty of Economics, Universiti Utara Malaysia.
- Lim, T. W., & Noorainie, A. A. (2010). Wood for the Trees: a review of the agarwood (gaharu) trade in Malaysia. Petaling Jaya: TRAFFIC Southeast Asia.
- Littler, M. M., & Littler, D. S. (1986). *Handbook of Phycological Methods: Ecological Field Methods: Macroalgae*. (Vol. 4). New York: Cambridge University Press.
- Liu, D. (2009). Handbook of nucleic acid purification. Boca Raton: CRC Press.
- Lodish, H., Berk, A., Zipursky, S. L., Matsudaira, P., Baltimore, D., & Darnell, J. (2000). Purifying, Detecting, and Characterizing Proteins. In *Molecular cell biology* (4th ed., p. 1084). New York: W.H. Freeman.
- López-Montilla, J. C., Herrera-Morales, P. E., Pandey, S., & Shah, D. O. (2002). Spontaneous emulsification: mechanisms, physicochemical aspects, modeling, and applications. *Journal of Dispersion Science and Technology*, 23(1–3), 219–268.
- Lord, R. S., & Bralley, J. A. (2008). Laboratory evaluations for integrative and functional medicine. Duluth: Metametrix Institute.
- Lu, K. C. (1983). The cultivation of the "incense tree" (Aquilaria sinensis). *Journal of the Hong Kong Branch of the Royal Asiatic Society*, 23, 247–249.
- Luning Prak, D. J., Abriola, L. M., Weber, W. J., Bocskay, K. A., & Pennell, K. D. (2000). Solubilization rates of n-alkanes in micellar solutions of nonionic surfactants. *Environmental Science & Technology*, 34(3), 476–482.

- Maali, A., & Hamed Mosavian, M. T. (2013). Preparation and application of nanoemulsions in the last decade (2000–2010). *Journal of Dispersion Science and Technology*, 34(1), 92–105.
- Macrae, R., Robinson, R. K., & Sadler, M. J. (1993). Encyclopaedia of food science, food technology, and nutrition: dressings and mayonnaise-fruits of tropical climates. San Diego: Academic Press.
- Mailina, J., Abd. Majid, J., Saidatul Husni, S., Chang, Y. S., Nor Azah, M. A., Nor Hasnida, H., Abu said, A., & Mohamad Nasir, M. A. (2009). Profiles of selected supreme agarwood oils from Malaysia. In *Herbal Globalisation : A New Paradigm for Malaysian Herbal Industry : Proceedings of the Seminar on Medicinal and Aromatic Plants (MAPS 2008)* (pp. 393–398). 21–22 October 2008, Kuala Lumpur: Forest Research Institute Malaysia (FRIM).
- Malik, M. A., Wani, M. Y., & Hashim, M. A. (2012). Microemulsion method: A novel route to synthesize organic and inorganic nanomaterials. *Arabian Journal of Chemistry*, 5(4), 397–417.
- Manning, F. S., & Thompson, R. E. (1995). *Oilfield processing of petroleum volume* 2: *crude oil*. (R. E. Thompson, Ed.). Tulsa: PennWell Books.
- Manz, A., Pamme, N., & Iossifidis, D. (2004). *Bioanalytical chemistry*. London: Imperial College Press.
- Marshak, D. R. (1996). *Techniques in protein chemistry vii*. San Diego: Academic Press.
- Mason, T. G., Wilking, J. N., Meleson, K., Chang, C. B., & Graves, S. M. (2006). Nanoemulsions: formation, structure, and physical properties. *Journal of Physics:* Condensed Matter, 18(41), R635–R666. http://doi.org/10.1088/0953-8984/18/41/R01
- Mat, N., Rahman, S. A., Ngah, N., Mahmud, K., Rosni, A., & Rahim, K. A. (2012). Growth and mineral nutrition of Aquilaria malaccensis (karas) in two habitats as affected by different cultural practices. *Journal of Nuclear and Related Technologies*, 9(1), 6–16.
- McClatchey, K. D. (2002). *Clinical laboratory medicine* (2nd ed.). Philadelphia: Lippincott Williams & Wilkins.
- McClements, D. J. (1999a). *Food emulsions: principles, practice, and techniques*. Boca Raton: CRC Press.
- McClements, D. J. (1999b). *Hydrophobic interactions, in food emulsions: principles, practice, and techniques.* Boca Raton: CRC Press.
- McClements, D. J. (2005). Food Emulsions: Principles, Practice and Techniques

- (2nd ed.). Boca Raton: CRC Press.
- McClements, D. J., Decker, E. A., & Weiss, J. (2007). Emulsion-based delivery systems for lipophilic bioactive components. *Journal of Food Science*, 72(8), R109-124.
- McClements, D. J., & Li, Y. (2010). Structured emulsion-based delivery systems: Controlling the digestion and release of lipophilic food components. *Advances in Colloid and Interface Science*, 159(2), 213–228.
- Meleson, K. (2008). The formation and stability of nanoemulsions. Ann Arbor: ProQuest.
- Mengual, O., Meunier, G., Cayré, I., Puech, K., & Snabre, P. (1999). TURBISCAN MA 2000: Multiple light scattering measurement for concentrated emulsion and suspension instability analysis. *Talanta*, 50(2), 445–456.
- Meyer, S., Berrut, S., Goodenough, T. I. J., Rajendram, V. S., Pinfield, V. J., & Povey, M. J. W. (2006). A comparative study of ultrasound and laser light diffraction techniques for particle size determination in dairy beverages. *Measurement Science and Technology*, 17(2), 289–297.
- Meyers, R. A. (1987). Encyclopedia of physical science and technology (2nd ed.). New York: Academic Press.
- Mishra, R. K., Soni, G. C., & Mishra, R. P. (2014). A review article: on nanoemulsion. *World Journal of Pharmacy and Pharmaceutical Sciences*, 3(9), 258–274.
- Module 1.2: Agarose gel electrophoresis. (2007). Retrieved December 3, 2016, from https://ocw.mit.edu/courses/biological-engineering/20-109-laboratory-fundamentals-in-biological-engineering-fall-2007/labs/mod1_2/
- Mohamed, R., Jong, P. L., & Zali, M. S. (2010). Fungal diversity in wounded stems of Aquilaria malaccensis. *Fungal Diversity*, 43(1), 67–74.
- Mohd Rosli, R. (2006). Extraction of gaharu essential oil using microwave assisted extraction (MAE) (Degree's thesis). University College of Engineering & Technology Malaysia.
- Morales, D., Gutiérrez, J. M., García-Celma, M. J., & Solans, Y. C. (2003). A study of the relation between bicontinuous microemulsions and oil/water nano-emulsion formation. *Langmuir*, 19(18), 7196–7200.
- Müller, R. H. (1991). Colloidal carriers for controlled drug delivery and targeting: modification, characterization and in vivo distribution. Boca Raton: CRC Press.

- Nadia, S. (2012). Extraction performance study of gaharu using microwave extraction method (Degree's thesis). Universiti Malaysia Pahang.
- Naef, R. (2011). The volatile and semi-volatile constituents of agarwood, the infected heartwood of Aquilaria species: a review. *Flavour and Fragrance Journal*, 26(2), 73–87.
- Nakajima, H. (1997). Microemulsion in cosmetics. In C. Solans & H. Kunieda (Eds.), *Industrial ap-plication of microemulsion, surfactant science series volume* 66 (pp. 175–197). New York: Marcel Dekker.
- Nakanishi, T., Yamagata, E., Yoneda, K., Nagashima, T., Kawasaki, I., Yoshida, T., Mori, H., & Miura, I. (1984). Three fragrant sesquiterpenes of agarwood. *Phytochemistry*, 23(9), 2066–2067.
- Ng, L. T., Chang, Y. S., & Kadir, A. A. (1997). A review on agar (gaharu) producing Aquilaria species. *Journal of Tropical Forest Products*, 2(2), 272–285.
- Ngan, C. L., Basri, M., Lye, F. F., Fard Masoumi, H. R., Tripathy, M., Karjiban, R. A., & Abdul-Malek, E. (2014). Comparison of process parameter optimization using different designs in nanoemulsion-based formulation for transdermal delivery of fullerene. *International Journal of Nanomedicine*, 9, 4375–4386.
- Ninfa, A. J., Ballou, D. P., & Benore, M. (2010). Fundamental laboratory approaches for biochemistry and biotechnology (2nd ed.). Hoboken: John Wiley & Sons, Inc.
- Nor Azah, M. A., Chang, Y. S., Mailina, J., Abu Said, A., Abd Majid, J., Saidatul Husni, S., Nor Hasnida, H., & Nik Yasmin, Y. (2008). Comparison of chemical profiles of selected gaharu oils from peninsular Malaysia. *The Malaysian Journal of Analytical Sciences*, 12(2), 338–340.
- Norzafneza, M. A., Saripa Salbiah, S., Hasimah, A., Ramli, I., Mohd Aspollah, H., Mat Ropi, M., & Abdul Hamid, A. (2011). Chemical constituents of leaves from Aquilaria crassna Pierre (karas) and their biological activities. In M. Mastura (Ed.), Harnessing the Tropical Herbal Heritage: Recent Advances in R&D and Commercialization. Proceedings of the Seminar on Medicinal and Aromatic Plants. (pp. 111–120). 3-4 August 2010, Kuala Lumpur: Forest Research Institute Malaysia (FRIM).
- Odokuma, L. O., & Okpokwasili, G. C. (1997). Seasonal influences of the organic pollution monitoring of the New Calabar River, Nigeria. *Environmental Monitoring and Assessment*, 45(1), 43–56.
- Okugawa, H., Ueda, R., Matsumoto, K., Kawanishi, K., & Kato, A. (1993). Effect of Jinkoh-eremol and Agarospirol from Agarwood on the Central

- Nervous System in Mice. Planta Medica, 59(1), 32-36.
- Olabisi, O., & Adewala, K. (1997). *Handbook of thermoplastics. Plastic Engineering* (Vol. 41). New York: CRC Press.
- Öztekin, N., & Erim, F. B. (2013). Determination of critical aggregation concentration in the poly-(vinylpyrrolidone)–sodium dodecyl sulfate system by capillary electrophoresis. *Journal of Surfactants and Detergents*, 16(3), 363–367.
- Parvinzadeh, M., & Hajiraissi, R. (2008). Macro- and Microemulsion Silicone Softeners on Polyester Fibers: Evaluation of Different Physical Properties. *Journal of Surfactants and Detergents*, 11(4), 269–273.
- Pasaribu, G., Waluyo, T. K., & Pari, G. (2015). Analysis of chemical compounds distinguisher for agarwood qualities. *Indonesian Journal of Forestry Research*, 2(1), 1–7.
- Paul Kan. (2011). World Cultures Festival 2011 Enchanting Arts of Asia. Trinity International Limited. Retrieved from http://www.3trinity.hk/press/agarwood/1/
- Peng, C. S., Rahim, K. A., & Awang, M. R. (2014). Histology study of Aquilaria malaccensis and the a resin formation under light microscope. *Journal of Agrobiotechnology*, *5*, 77–83.
- Persoon, G. A. (2007). Agarwood: the life of a wounded tree. *IIAS Newsletter*, 45, 24–25.
- Pitman, V. (2004). *Aromatherapy: a practical approach*. Cheltenham: Nelson Thornes.
- Polyacrylamide gel electrophoresis. (n.d.). Retrieved December 3, 2016, from https://en.wikipedia.org/wiki/Polyacrylamide_gel_electrophoresis
- Pornpunyapat, J., Chetpattananondh, P., & Tongurai, C. (2011). Mathematical modeling for extraction of essential oil from Aquilaria Crassna by hydrodistillation and quality of agarwood oil. *Bangladesh Journal of Pharmacology*, 6(1), 18–24.
- Pourmortazavi, S. M., & Hajimirsadeghi, S. S. (2007). Supercritical fluid extraction in plant essential and volatile oil analysis. *Journal of Chromatography*. *A*, 1163(1–2), 2–24.
- Preedy, V. R. (2105). Essential oils in food preservation, flavor and safety. London: Academic Press.
- Pripdeevech, P., Khummueng, W., & Park, S. K. (2011). Identification of odor-

- active components of agoarwood essential oils from Thailand by solid phase microextraction-GC/MS and GC-O. *Journal of Essential Oil Research*, 23(4), 46–53.
- Qi, S. Y. (1995). Aquilaria species: in vitro Culture and the production eaglewood (agarwood). In Y. P. S. Bajaj (Ed.), *Medicinal and Aromatic Plants VIII.* (Vol. 33, pp. 36–46). New York: Springer.
- Rahaman, M. N. (2003). *Ceramic processing and sintering* (2nd ed.). New York: Marcel Dekker Inc.
- Reddy, P. R., & Raju, N. (2012). Gel-electrophoresis and its applications. In S. Magdeldin (Ed.), *Gel Electrophoresis Principles and Basics* (pp. 2–32). Rijeka: InTech.
- Reddy, S. R., & Fogler, H. S. (1981). Emulsion stability: Determination from turbidity. *Journal of Colloid And Interface Science*, 79(1), 101–104.
- Rédei, G. P. (1998). Genetics manual: current theory, concepts, terms. Singapore: World Scientific.
- Reid, R. (1999). *Peptide and protein drug analysis*. (Vol. 12). New York: Marcel Dekker Inc.
- Rhein, L. D., Schlossman, M., O'Lenick, A., & Somasundaran, P. (2006). Surfactants in personal care products and decorative cosmetics, surfactant science series volume 135 (3rd ed.). Boca Raton: CRC Press.
- Ridaoui, H., Jada, A., Vidal, L., & Donnet, J. B. (2006). Effect of cationic surfactant and block copolymer on carbon black particle surface charge and size. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 278(1–3), 149–159.
- Riddick, T. M. (1968). Control of colloid stability through zeta potential: with a closing chapter on its relationship to cardiovascular disease. (Vol. 1). Staunton: Zeta Meter Inc.
- Robinson, J. W., Skelly Frame, E. M., & Frame II, G. M. (2004). *Undergraduate instrumental analysis* (6th ed.). New York: Marcel Dekker.
- Rocha, A. J. S., Gomes, V., Ngan, P. V., Passos, M. J. A. C. R., & Furia, R. R. (2007). Effects of anionic surfactant and salinity on the bioenergetics of juveniles of Centropomus parallelus (Poey). *Ecotoxicology and Environmental Safety*, 68(3), 397–404.
- Rodrigues, L. R. (2015). Microbial surfactants: Fundamentals and applicability in the formulation of nano-sized drug delivery vectors. *Journal of Colloid and Interface Science*, 449, 304–316.

- Principles and Basics (pp. 33-40). Rijeka: InTech.
- Yoneda, K., Yamagata, E., Nakanishi, T., Nagashima, T., Kawasaki, I., Yoshida, T., Mori, H., & Miura, I. (1984). Sesquiterpenoids in two different kinds of agarwood. *Phytochemistry*, 23(9), 2068–2069.
- Yoswathana, N., Eshiaghi, M. N., & Jaturapornpanich, K. (2012). Enhancement of essential oil from agarwood by subcritical water extraction and pretreatments on hydrodistillation. *International Journal of Chemical, Molecular, Nuclear, Materials and Metallurgical Engineering*, 6(5), 453–459.
- Zainuddin, N. A. S., Ku Halim, K. H., Musa, M., & Rodhi, M. N. M. (2014). Effect of hydrochloric acid (HCl) treatment on FTIR profile of agarwood waste from hydrodistillation process. *Applied Mechanics and Materials*, 575, 165–169.
- Zhang, H. (2007). Developments and applications of electrophoresis and small molecule laser desorption ionization mass spectrometry. Ames: ProQuest.
- Zhou, M., Wang, H., Suolangjiba, Kou, J., & Yu, B. (2008). Antinociceptive and anti-inflammatory activities of Aquilaria sinensis (Lour.) Gilg. Leaves extract. *Journal of Ethnopharmacology*, 117(2), 345–350.
- Zimm, B. H., & Levene, S. D. (1992). Problems and prospects in the theory of gel electrophoresis of DNA. *Quarterly Reviews of Biophysics*, 25(2), 171–204.

- Rosen, M. J. (2004). *Surfactants and interfacial phenomena* (3rd ed.). Hoboken: John Wiley & Sons.
- Rosen, M. J., & Kunjappu, J. T. (2012). *Surfactants and Interfacial Phenomena* (4th ed.). Hoboken: John Wiley & Sons.
- Sadgopal. (1960). Exploratory studies in the development of essential oils and their constituents in aromatic plants. Part I. Oil of agarwood. *Soap, Perfumery and Cosmetics, London, 33*(1), 41–6.
- Salager, J. L. (2002). Surfactant types and uses. Version 2. *Teaching Aid in Surfactant Science & Engineering*. Merida, Venezuela: Laboratory of Formulation, Interfaces, Rheology and Processes, Universidad De Los Andes.
- Sampaio, B. L., Edrada-Ebel, R., & Da Costa, F. B. (2016). Effect of the environment on the secondary metabolic profile of Tithonia diversifolia: a model for environmental metabolomics of plants. *Scientific Reports*, 6(29265).
- Sanjeewani, N. A., & Sakeena, M. H. F. (2013). Formulation and characterization of virgin coconut oil (VCO) based emulsion. *International Journal of Scientific and Research Publications*, 3(12), 1–6.
- Sarapardeh, A. H., Ayatollahi, S., Ghazanfari, M. H., & Masihi, M. (2014). Experimental determination of interfacial tension and miscibility of the CO2-crude oil system; temperature, pressure, and composition effects. *Journal of Chemical and Engineering Data*, 59(1), 61–69.
- Sarmento, B., & Neves, J. das. (2012). *Chitosan-based systems for biopharmaceuticals: delivery, targeting, and polymer therapeutics*. Chichester: John Wiley & Sons.
- Schippmann, U. (2001). *Medicinal plants significant trade study : CITES project S* 109, *Plants Committee document PC9 9.1.3 (rev.)*. Bonn: German Federal Agency for Nature Conservation.
- Sereshti, H., Izadmanesh, Y., & Samadi, S. (2011). Optimized ultrasonic assisted extraction-dispersive liquid-liquid microextraction coupled with gas chromatography for determination of essential oil of Oliveria decumbens Vent. *Journal of Chromatography*. *A*, 1218(29), 4593–4598.
- Shah, L. K., & Amiji, M. M. (2006). Intracellular delivery of saquinavir in biodegradable polymeric nanoparticles for HIV/AIDS. *Pharmaceutical Research*, 23(11), 2638–2645.
- Sharma, M. K., & Shah, D. O. (1985). Introduction to macro- and microemulsions. In *Macro- and Microemulsions: theoy and applications*. (Vol.

- 272, pp. 1-18). Washington: American Chemical Society.
- Sharma, N., Bansal, M., Visht, S., Sharma, P., & Kulkarni, G. (2010). Nanoemulsion: a new concept of delivery system. *Chronicles of Young Scientists*, 1(2), 2–6.
- Sharon. (2011). Differences between vegetable oils and essential oils. Retrieved January 29, 2016, from http://naturalhealthezine.com/differences-between-vegetable-oils-and-essential-oils/
- Shi, Q., & Jackowski, G. (1998). One-dimensional polyacrylamide gel electrophoresis. In Hames B.D. (Ed.), *Gel Electrophoresis of Proteins A Practical Approach* (3rd ed., pp. 1–52). New York: Oxford University Press.
- Shimada, Y., Tominaga, T., T., K., & Kiyosawa, S. (1982). Studies on the agarwood (Jinko). I. Structures of 2-(2-phenylethyl) chromone derivatives. *Chemical & Pharmaceutical Bulletin*, 30(10), 3791–3795.
- Shourie, A., & Chapadgaonkar, S. S. (2005). *Bioanalytical techniques*. New Delhi: The Energy and Resources Institute (TERI).
- Siffert, B., Jada, A., & Letsango, J. E. (1994). Location of the shear plane in the electric double layer in an organic medium. *Journal of Colloid and Interface Science*, 163(2), 327–333.
- Siwach, P., & Singh, N. (2007). *Molecular biology: principles and practices*. New Delhi: Laxmi Publications.
- Skaria, B. P. (2007). *Aromatic Plants: Horticulture science series*. (K. V. Peter, Ed.) (Vol. 1). New Delhi: New India Publishing.
- Smith, I. (1960). *Chromatographic and electrophoretic techniques: zone electrophoresis* (2nd ed.). London: Heinemann Medical.
- Smith, R. J., Lotya, M., & Coleman, J. N. (2010). The importance of repulsive potential barriers for the dispersion of graphene using surfactants. *New Journal of Physics*, 12(12), 11.
- Snelder, D. J., & Lasco, R. D. (2008). Smallholder tree growing for rural development and environmental services: lessons from Asia. (P. K. R. Nair, Ed.), Advances in Agroforestry. (Vol. 5). New York: Springer Science & Business Media.
- Soehartono, T., & Mardiastuti, A. (2002). *CITES implementation in Indonesia*. (Vol. xvii). Jakarta: Nagao Natural Environment Foundation.
- Soehartono, T., & Newton, A. C. (2000). Conservation and sustainable use of tropical trees in the genus Aquilaria I. Status and distribution in

- Indonesia. *Biological Conservation*, 96(1), 83–94.
- Soehartono, T., & Newton, A. C. (2002). The gaharu trade in Indonesia: Is it sustainable? *Economic Botany*, 56(3), 271–284.
- Sokima, S. (2008). Comparison Between Chemical Compounds in Gaharu Smoke (Burning) and Gaharu Oil (Hidrodistillation) (Degree's thesis). Universiti Malaysia Pahang.
- Solans, C., Izquierdo, P., Nolla, J., Azemar, N., & Garciacelma, M. (2005). Nano-emulsions. *Current Opinion in Colloid & Interface Science*, 10(3–4), 102–110.
- Spinning drop method. (n.d.). Retrieved December 3, 2016, from https://en.wikipedia.org/wiki/Spinning_drop_method
- Stellwagen, N. C. (1985). Effect of the electric field on the apparent mobility of large DNA fragments in agarose gels. *Biopolymers*, 24(12), 2243–2255.
- Tadros, T. F. (2013). Emulsion formation, stability, and rheology. In *Emulsion Formation and Stability* (pp. 1–75). Weinheim, Germany: Wiley-VCH Verlag GmbH & Co. KGaA.
- Tadros, T. F., Vandamme, A., Levecke, B., Booten, K., & Stevens, C. V. (2004). Stabilization of emulsions using polymeric surfactants based on inulin. *Adv Colloid Interface Sci.*, 20(108–109), 207–226.
- Tadros, T., Izquierdo, P., Esquena, J., & Solans, C. (2004). Formation and stability of nano-emulsions. *Advances in Colloid and Interface Science*, 108–109, 303–318.
- Tajuddin, S. N., Muhamad, N. S., Yarmo, M. A., & Yusoff, M. M. (2013). Characterization of the chemical constituents of agarwood oils from Malaysia by comprehensive two dimensional gas chromatography–time-of-flight mass spectrometry. *Mendeleev Communications*, 23(1), 51–52.
- Tajuddin, S. N., & Yusoff, M. M. (2010). Chemical composition of volatile oils of Aquilaria malaccensis (Thymelaeaceae) from Malaysia. *Natural Product Communications*, *5*(12), 1965–1968.
- Takemoto, H., Ito, M., Shiraki, T., Yagura, T., & Honda, G. (2008). Sedative effects of vapor inhalation of agarwood oil and spikenard extract and identification of their active components. *Journal of Natural Medicines*, 62(1), 41–6.
- Tambe, D. E., & Sharma, M. M. (1993). Factors controlling the stability of colloid-stabilized emulsions. *Journal of Colloid and Interface Science*, 157(1), 244–253.

- Tang, Y., & Stratton, C. W. (2012). *Advanced techniques in diagnostic microbiology* (2nd ed.). New York: Springer Science & Business Media.
- Taurozzi, J. S., Hackley, V. A., & Wiesner, M. R. (2012). Protocol for preparation of nanoparticle dispersions from powdered material using ultrasonic disruption, Version 1.1. Gaithersburg: NIST special publication 1200-2.
- Tawan, C. . (2004). Thymelaeaceae. In E. Soepadmo, L. G. Saw, & R. C. K. Chung (Eds.), *Tree Flora of Sabah and Sarawak.* (Vol. 5, pp. 433–484). Kuala Lumpur: Forest Research Institute Malaysia.
- Taylor, P., & Ottewill, R. H. (1994). The formation and ageing rates of oil-in-water miniemulsions. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 88(2–3), 303–316.
- Tcholakova, S., Denkov, N. D., Ivanov, I. B., & Campbell, B. (2006). Coalescence stability of emulsions containing globular milk proteins. *Advances in Colloid and Interface Science*, 123–126, 259–293.
- Thawatchai, S. (2007). Taxonomy, geography and ecology of Aquilaria Lamk. (Thymelaeaceae: Aquilarioideae) in continental Asia. In 2nd International Agarwood Conference. 4-11 March 2007, Bangkok: The Tropical Rainforest Project.
- The chemistry of essential oils the lowdown and upside of the chemical components and constituents. (n.d.). Retrieved January 29, 2016, from http://www.essentialoils.co.za/components.htm
- Tian, K. (2011). Electrophoresis. In *Soft-Matter*. Retrieved from http://soft-matter.seas.harvard.edu/index.php/Electrophoresis
- Tomar, R. S. (2010). *Molecular markers and plant biotechnology*. New Delhi: New India Publishing.
- Tran, Q. Le, Tran, Q. K., Kouda, K., Nguyen, N. T., Maruyama, Y., Saiki, I., & Kadota, S. (2003). A survey on agarwood in Vietnam. *Journal of Traditional Medicines*, 20(3), 124–131.
- Tresset, G. (2009). The multiple faces of self-assembled lipidic systems. *PMC Biophysics*, 2(1), 3.
- Troy, D. B., & Beringer, P. (2006). *Remington: the science and practice of pharmacy*. (D. B. Troy & P. Beringer, Eds.) (21st ed., Vol. 1). Philadelphia: Lippincott Williams & Wilkins.

- Tsai, C. S. (2007). *Biomacromolecules: introduction to structure, function and informatics*. Hoboken: John Wiley & Sons, Inc.
- Turner, S. R., Siano, D. B., & Bock, J. (1985). Microemulsion process for producing acrylamide-alkyl acrylamide copolymers. U.S.: United States Patent. Retrieved from https://www.google.com/patents/US4521580
- Uli Kozok. (2013). Meet the Batak Manuscript Collection Scope Notes. Retrieved December 3, 2016, from http://lib.typepad.com/scrc/2013/11/guest-post-meet-the-batak-manuscript-collection-.html
- Vaclavik, V., & Christian, E. W. (2014). Essentials of food science (4th ed.). New York: Springer Science & Business Media.
- Viovy, J. L. (2000). Electrophoresis of DNA and other polyelectrolytes: physical mechanisms. *Reviews of Modern Physics*, 72(3), 813–872.
- Vispute, M., Nazim, S., Khan, T., Shaikh, S. (2013). An overview on exploring nasal microemulsion for treatment of CNS disorders. *International Journal of Pharmaceutical Sciences and Research*, 4(4), 1294–1311.
- Vyas, S. P., & Kholi, D. V. (2002). *Methods in biotechnology and bioengineering:* for pharmaceutical & other biosciences. New Delhi: CBS Publishers and Distributors.
- Vyas, T. K., Shahiwala, A., & Amiji, M. M. (2008). Improved oral bioavailability and brain transport of Saquinavir upon administration in novel nanoemulsion formulations. *International Journal of Pharmaceutics*, 347(1–2), 93–101.
- Walstra, P. (1996). Emulsion stability. In P. Becher (Ed.), In Encyclopedia of Emulsion Technology, Vol. 4. (pp. 1–62). New York: Marcel Dekker.
- Wang, H., Wang, M., Ge, X., Liu, H., & Zhang, Z. (2008). Radiation miniemulsion polymerization system with HTPB or its derivative as the costabilizer. *Colloid and Polymer Science*, 286(8–9), 1039–1047.
- Ward, A. J. I. (1989). The kinetics of oil solubilisation in aqueous surfactant solutions. In *Proceedings of the Royal Irish Academy. Section B: Biological, Geological, and Chemical Science.* (Vol. 89B, pp. 375–382). Dublin: Royal Irish Academy.
- Wasan, K. M. (2006). Role of lipid excipients in modifying oral and parenteral drug delivery: basic principles and biological examples. Hoboken: John Wiley & Sons.
- West, C. C., & Harwell, J. H. (1992). Surfactants and subsurface remediation.

- Environmental Science & Technology, 26(12), 2324–2330.
- Whitaker, J. R., & Tannenbaum, S. R. (1977). *Food proteins*. Connecticut: Avi Publishing Company.
- White, B., Banerjee, S., O'Brien, S., Turro, N. J., & Herman, I. P. (2007). Zeta-potential measurements of surfactant-wrapped individual single-walled carbon nanotubes. *Journal of Physical Chemistry C*, 111(37), 13684–13690.
- Whitmore, T. (1973). *Tree flora of Malaya: a manual for foresters.* (Vol. 2). Kuala Lumpur: Longman Malaysia.
- Wilson, I. D., Adlard, E. R., Cooke, M., & Poole, C. F. (2000). *Encyclopedia of separation science*. (Vol. 5). London: Academic Press.
- Wilson, K., & Walker, J. (2005). *Principles and Techniques of Biochemistry and Molecular Biology* (7th ed.). New York: Cambridge University Press.
- Wrolstad, R. E., Acree, T. E., Decker, E. A., Penner, M. H., Reid, D. S., Schwartz, S. J., Shoemaker, C. F., Smith, D. M., & Sporns, P. (2005). *Handbook of food analytical chemistry, water, proteins, enzymes, lipids, and carbohydrates*. (Vol. 1). New Jersey: John Wiley & Sons.
- Xavier, J. A. (2011). Effect of varying surfactant concentration on interfacial tension in tertiary recovery of crude oil (Master's thesis). Dalhousie University.
- Yaacob, S. (1999). *Agarwood: Trade and CITES Implementation in Malaysia*. Malaysia: TRAFFIC Southeast Asia.
- Yagura, T., Shibayama, N., Ito, M., Kiuchi, F., & Honda, G. (2005). Three novel diepoxy tetrahydrochromones from agarwood artificially produced by intentional wounding. *Tetrahedron Letters*, 46(25), 4395–4398.
- Yamada, I. (1995). Aloeswood forest and the maritime world. *Japanese Journal of Southeast Asian Studies*, 33(3), 463–468.
- Yan, M., Li, B., & Zhao, X. (2010). Determination of critical aggregation concentration and aggregation number of acid-soluble collagen from walleye pollock (Theragra chalcogramma) skin using the fluorescence probe pyrene. *Food Chemistry*, 122(4), 1333–1337.
- Yip, Y. T. (2014). Sniffing out illegal trade. *The Star Online*. Retrieved from http://www.thestar.com.my/news/community/2014/04/15/sniffing-out-illegal-trade-authorities-believe-syndicates-are-behind-sale-of-the-endangered-agarwo/
- Ylmaz, M., Ozic, C., & Gok, L. (2012). Principles of nucleic acid separation by agarose gel electrophoresis. In S. Magdeldin (Ed.), Gel Electrophoresis -