



UNIVERSITI PUTRA MALAYSIA

***SCREENING, DESIGN, SYNTHESIS AND BIOACTIVITY OF INHIBITORS
AGAINST PROTEIN ARGININE DEIMINASE TYPE IV***

TEO CHIAN YING

FS 2015 72



**SCREENING, DESIGN, SYNTHESIS AND BIOACTIVITY OF INHIBITORS
AGAINST PROTEIN ARGININE DEIMINASE TYPE IV**

By

TEO CHIAN YING

**Thesis Submitted to School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

January 2015

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Doctor of Philosophy

SCREENING, DESIGN, SYNTHESIS AND BIOACTIVITY OF INHIBITORS AGAINST PROTEIN ARGININE DEIMINASE TYPE IV (PAD4)

By

TEO CHIAN YING

January 2015

Chair: Professor Mohd Basyaruddin Abdul Rahman, PhD

Faculty: Science

Protein Arginine Deiminase IV (PAD4) is a new target for rheumatoid arthritis (RA) therapeutic. It catalyses the citrullination process in human body that produces citrullinated protein which is believed to be the root cause of RA. Inhibitor for the enzyme can be a drug for the treatment of RA. The main objective of this research is to search for potent inhibitors for PAD4. Human PAD4 has been successfully expressed in *Escherichia coli* BL21 (DE3). PAD4 expressed from pET32b vector with the use of auto-induction media introduced by Studier gave higher solubility and activity compared to conventional induction system. PAD4 was successfully purified using affinity chromatography technique with purity of 90%, approximately. Several compounds have been identified as potential inhibitors for PAD4 by structure-based virtual screening using Ligand Discovery at Edinburgh University (LIDAEUS) programme. Three compounds out from 22 top-ranked water-soluble compounds showed significant inhibition to PAD4 and their IC_{50} values were ranged from 1.49 ± 0.03 to 2.96 ± 0.01 mM. Binding affinities of the compounds to PAD4 obtained from molecular docking were -7.49, -7.27 and -6.00 kcal/mol. The structures of the three compounds showed no resemblance with previously discovered PAD4 inhibitors, nor with existing drugs for RA treatment. Ultrafast Shape Recognition (USR) was utilized in searching of compounds possess similar molecular shape with previous reported inhibitor, streptonigrin. Five compounds out of the selected 37 compounds inhibited enzymatic activity of PAD4 significantly, with more than 10% inhibition at 100 μ M. The best compound, (2*E*)-*N*-{(2*R*)-1-[(furan-2-ylmethyl)(methyl)amino]-1-oxopropan-2-yl}-3-(4-methoxyphenyl) prop-2-enamide, is a moderate inhibitor for PAD4 with IC_{50} value of 362.67 ± 4.13 μ M. The structure of the compound showed no resemblance with the parent compound. Interestingly, furan ring in USR discovered compound was able to enter the active site cleft of PAD4. Four peptide-based inhibitors incorporated with non-standard amino acid containing furan ring (X) with sequence based-on PAD4 natural substrate, nucleophosmin were designed and synthesized using solid phase technique. Circular dichroism spectroscopy showed that the structure of the designed peptide-based inhibitors was predominantly unordered. The IC_{50} value of the best peptide-based

inhibitors with sequence KSIXDTP is $243.2 \pm 2.4 \mu\text{M}$ which is lower than compounds obtained from LIDAEUS and USR. Kinetic studies revealed that it inhibited PAD4 reversibly and competitively. Three dimensional structure of the best peptide-based inhibitor was further elucidated using nuclear magnetic resonance spectroscopy. Molecular docking of the peptide-based inhibitor suggested favourable interaction between the peptide with PAD4 with binding affinity of -5.4 kcal/mol . Inhibitors containing furan ring in the structure show high potential in inhibiting PAD4 and the structure of the inhibitors discovered in this research can be modified to be a better drug candidate for treatment of rheumatoid arthritis.

Abstrak tesis yang dikemukakan kepada Senat of Universiti Putra Malaysia
Sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

SARINGAN, REKA BENTUK, SINTESIS DAN BIOAKTIVITI PERENCAT TERHADAP PROTEIN ARGININA DEIMINASE JENIS 4

Oleh

TEO CHIAN YING

Januari 2015

Pengerusi: Profesor Mohd Basyaruddin Abdul Rahman, PhD

Faculti: Sains

Protein Arginina Deiminase jenis IV (PAD4) adalah sasaran baru untuk terapeutik artritis reumatoid (RA). Ia merupakan pemangkin proses pensitrulinaan yang menghasilkan protein mengandungi sitrulina yang dipercayai menjadi punca kepada RA. Perencat enzim ini boleh menjadi ubat untuk rawatan RA. Objektif utama kajian ini adalah mencari perencat untuk PAD4. PAD4 manusia telah berjaya dihasilkan dalam *Escherichia coli* BL21 (DE3). PAD4 yang dihasilkan dari vektor pET32b dengan penggunaan media auto-induksi yang diperkenalkan oleh Studier mempunyai kelarutan dan aktiviti yang lebih tinggi berbanding dengan sistem induksi konvensional. PAD4 telah berjaya ditulenkan dengan menggunakan teknik kromatografi afiniti dengan 90% ketulenan. Beberapa sebatian telah dikenalpasti sebagai perencat berpotensi untuk PAD4 oleh program *Ligand Discovery at Edinburgh University* (LIDAEUS) berasaskan struktur PAD4. Dua puluh dua sebatian paling bagus yang larut air telah dipilih untuk pemeriksaan potensi perencatan mereka terhadap PAD4. Tiga sebatian menunjukkan perencatan yang ketara kepada PAD4 dan nilai-nilai IC_{50} mereka adalah di antara $1.49 \pm 0.03 - 2.96 \pm 0.01$ mM. Tenaga penambatan untuk sebatian tersebut dengan PAD4 yang diperolehi daripada *docking* molekul adalah -7.49, -7.27 dan -6.00 kcal/ mol. Struktur tiga sebatian itu tidak menunjukkan persamaan dengan perencat PAD4 yang ditemui sebelum ini atau dengan dadah yang sedia ada untuk rawatan RA. *Ultrafast Shape recognition* (USR) telah digunakan untuk mencari sebatian yang mempunyai bentuk molekul yang menyerupai satu perencat yang dilaporkan sebelum ini, iaitu *streptonigrin*. Lima sebatian daripada 37 sebatian terpilih merencat aktiviti enzim PAD4 dengan ketara, lebih daripada 10% perencatan pada 100 μ M. Sebatian yang terbaik, (2E)-N-{(2R)-1-[(furan-2-ilmetil)(metil)amino]-1-oxopropan-2-il}-3-(4-metoksilfenil)prop-2-enamida, adalah perencat sederhana untuk PAD4 dengan nilai IC_{50} 362.67 ± 4.13 . Struktur sebatian tidak menunjukkan persamaan dengan sebatian induknya. Gelang furan di sebatian yang ditemui dengan USR dapat memasuki rekahan tapak aktif PAD4. Empat perencat berasaskan peptida yang diperbadankan dengan asid amino tidak semula jadi yang mengandungi gelang furan (X) dengan urutan berasaskan substrat semula jadi PAD4 iaitu nucleophosmin telah direka dan

disintesis dengan menggunakan teknik fasa pepejal. Spektroskopi dikroisme bulatan menunjukkan bahawa struktur perencat berasaskan peptida yang direka adalah kebanyakannya tidak tertib. Nilai IC_{50} perencat berasaskan peptida yang terbaik dengan urutan KSIXDTP ialah $243.2 \pm 2.4 \mu M$ iaitu lebih rendah daripada sebatian yang diperolehi daripada LIDAEUS dan USR. Kajian kinetik menunjukkan perencat tersebut merencat PAD4 secara berbalik dan kompetitif. Struktur tiga dimensi bagi perencat berasaskan peptida yang terbaik telah dijelaskan selanjutnya dengan menggunakan spektroskopi resonans magnet nukleus. *Docking* molekul untuk perencat berasaskan peptida mencadangkan kewujudan interaksi tersesuaian antara peptida tersebut dengan PAD4 dengan tenaga penambatan -5.4 kcal/mol . Perencat yang mengandungi gelang furan dalam strukturnya menunjukkan potensi yang tinggi dalam merencat PAD4 dan struktur perencat yang ditemui dalam kajian ini boleh diubah suai untuk menjadi calon dadah yang lebih baik untuk rawatan artritis reumatoid.

ACKNOWLEDGEMENTS

I would like to express my deep gratitude to my former and current supervisory committee chairs, Dr. Bimo Ario Tejo and Prof. Mohd. Basyaruddin Abdul Rahman, for the valuable guidance and advices. Their encouragement and useful critiques of the research really help me in completing this study. Without their knowledge and assistance this study would not have been successful.

I would like to thank my supervisory committee members, Dr. Adam Leow and Prof. Abu Bakar Salleh, for their insightful comments and suggestions through this research.

I would like to acknowledge Kuok Foundation for the financial support they have provided to me in order to make this thesis possible.

Special thanks also go to my lab mates in Lab 140 (Biotech 2), Lab 248 and 105 (Chemistry Department) especially Izzuddin Ahmad Nadzirin, Foong Pik Mun, Zalikha Ibrahim, Hafizah Kamarudin, and Ely for sharing the literature and invaluable assistance. In particular, I would like to express my appreciation to Azira Muhamad for her patience and assistance in this research.

Finally, an honourable mention goes to my beloved family and friends for their support and encouragement through the duration of study.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Mohd Basyaruddin Abdul Rahman, PhD

Professor
Faculty of Science
Universiti Putra Malaysia
(Chairman)

Abu Bakar Salleh, PhD

Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

Adam Leow Thean Chor, PhD

Senior Lecturer
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

Bimo Ario Tejo, PhD

Department of Biotechnology
Surya University
Indonesia
(Member)

BUJANG KIM HUAT, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____

Date: _____

Name and Matric No.: _____

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____

Name of
Chairman of
Supervisory
Committee:

Signature: _____

Name of
Member of
Supervisory
Committee:

Signature: _____

Name of
Member of
Supervisory
Committee:

Signature: _____

Name of
Member of
Supervisory
Committee:

TABLE OF CONTENTS

	ABSTRACT	Page i
	ABSTRAK	iii
	ACKNOWLEDGEMENT	v
	APPROVAL	vi
	DECLARATION	viii
	LIST OF TABLES	xii
	LIST OF FIGURES	xiv
	LIST OF APPENDICES	xvii
	LIST OF ABBREVIATIONS	xviii
CHAPTER		
1	INTRODUCTION	1
2	LITERATURE REVIEW	3
	2.1 Rheumatoid Arthritis	3
	2.2 Autoantibodies Associated with RA	4
	2.3 Anti-citrullinated Protein Autoantibodies and RA	5
	2.4 Protein Arginine Deiminase	6
	2.4.1 Family of Protein Arginine Deiminase	6
	2.4.2 Expression of PAD4	8
	2.5 PAD4 and RA	9
	2.6 Catalytic Activity of PAD4	11
	2.7 Current Treatments Available For RA	15
	2.7.1 First-line Drugs for RA	15
	2.7.2 Second-line Drugs for RA	15
	2.7.3 Tumor Necrosis Factor (TNF) Inhibitors	18
	2.8 Current Inhibitors of PAD4	19
	2.9 Drug Design and Virtual Screening	23
	2.9.1 Ligand Discovery at Edinburgh University (LIDAEUS)	24
	2.9.2 Ultrafast Shape Recognition (USR)	25
	2.10 Peptides as Drugs	26
	2.11 NMR of Peptides	29
3	MATERIALS AND METHODS	31
	3.1 Overview	31
	3.2 Materials	32
	3.3 Cloning, Expression and Purification of PAD4	34
	3.3.1 Preparation of Culture Media and Buffer Solution	34

3.3.2	Preparation of Sample cDNA on Disc	34
3.3.3	Preparation of Competent Cells	34
3.3.4	Chemical Transformation	35
3.3.5	Cloning	35
3.3.6	Agarose Gel Electrophoresis	36
3.3.7	Plasmid Extraction	36
3.3.8	Protein Expression	37
3.3.9	Preparation of Non-inducing and Auto-inducing Media	37
3.3.10	Protein Expression Using Non-inducing and Auto-inducing Media	37
3.3.11	Preparation of Crude PAD4	37
3.3.12	Protein Purification	38
3.3.13	Sodium Dodecyl Sulfate Polyacrylamide Gel Electrophoresis (SDS-PAGE)	39
3.3.14	Citrulline Colorimetric Assay	39
3.3.15	Protein Concentration Determination	40
3.4	High Throughput Virtual Screening	41
3.4.1	Structure-based Virtual Screening	41
3.4.2	Ligand-based Virtual Screening	41
3.5	Peptide-based PAD4 Inhibitors	42
3.5.1	Peptide Design	42
3.5.2	Binding of Peptides to PAD4	44
3.5.3	Peptide Synthesis	44
3.5.4	Kaiser Test	47
3.5.5	Cleavage of the Peptide from the Resin	47
3.5.6	Peptide Analysis and Purification	48
3.5.7	Molecular Weight Determination	49
3.6	Experiments of PAD4 Inhibitors	50
3.6.1	PAD4 Inhibitors Rapid screening	50
3.6.2	Determination of IC ₅₀	50
3.6.3	Molecular Docking	51
3.7	Studies of Peptide-based Inhibitors	51
3.7.1	Biophysical Studies	51
3.7.2	Kinetic Studies	52

4	RESULTS AND DISCUSSION	55
4.1	PAD4 Expression using Commercial plasmid	55
4.2	Cloning of PAD4 into pET-32b	55
4.3	PAD4 Expression in Auto-induction System	59
4.4	Purification of PAD4	61
4.5	High-throughput Virtual Screening using LIDAEUS	64
4.6	Inhibitory Activity of Hits from LIDAEUS	66

4.7	Molecular Docking Analysis of Hits from LIDAEUS	69
4.8	Ligand-based Virtual Screening using USR	73
4.9	In Vitro Enzyme Assay with Inhibitors Identified by USR	74
4.10	Molecular Docking Analysis of Compound 5	78
4.11	Design of Peptide-based PAD4 Inhibitors	81
4.12	Synthesis and Purification of Peptide-based Inhibitors	86
4.13	Inhibitory Activity of Peptide-based Inhibitors	89
4.14	Inhibition Characteristics of Inh Dap	91
4.15	Three Dimensional Studies of Peptide-based Inhibitors	93
4.15.1	Secondary Structure Estimation of Peptide-based Inhibitors	93
4.15.2	Three Dimensional Studies of Inh Dap by NMR	94
4.16	Molecular Docking Analysis of Inh Dap	101
5	CONCLUSION AND RECOMMENDATIONS	105
5.1	Conclusion	105
5.2	Recommendations for future research	106
	REFERENCES	107
	APPENDICES	132
	BIODATA OF STUDENT	160
	LIST OF PUBLICATIONS	161

LIST OF TABLES

Table	Page
3.1 Materials and their manufacturers.	32
3.2 Thermal cycling conditions for gradient PCR.	36
3.3 Sequences of designed peptides.	43
3.4 General cleavage cocktails for rink amide AM resin.	48
3.5 Gradient elution system for peptide analysis.	49
3.6 Gradient elution system for peptide purification.	49
3.7 Gradient elution system of LC/MS.	50
4.1 Purification table of PAD4.	64
4.2 Compound ID, rank, structure, and binding affinity of 22 compounds tested for their inhibitory activity against PAD4.	66
4.3 Summary of residues involved in the interactions with compound 9, 10 and 59.	72
4.4 Yield and purity of crude peptide-based inhibitors.	88
4.5 Expected and experimental molecular weight of peptide-based inhibitors.	89
4.6 IC ₅₀ values of the designed peptide-based inhibitors.	91
4.7 Summary of secondary structure estimation of peptide-based inhibitors.	94
4.8 Structural statistics for the simulated Inh Dap peptide ensembles.	99
4.9 List of conformations selected from each top 10 cluster with their energy.	100

LIST OF FIGURES

Figures	Page
2.1 Post-translational conversion of peptidylarginine into peptidylcitrulline catalyzed by calcium ion dependent protein arginine deiminase (PAD).	5
2.2 Structural changes induced by citrullination.	5
2.3 Ribbon representation of PAD4-substrate complex.	11
2.4 Structure of the C-terminal domain of PAD4.	12
2.5 Electrostatic surface potentials of Ca ²⁺ -free PAD4 (left), Ca ²⁺ -bound PAD4 (middle) and substrate complex (right).	12
2.6 Catalytic mechanism of PAD4.	14
2.7 Chemical structure of methotrexate.	16
2.8 Chemical Structure of Leflunomide.	17
2.9 Chemical structures of selected inhibitors of PAD4.	20
2.10 Chemical structure of benzoylarginine amide.	21
2.11 Two proposed mechanisms of PAD4 inactivation by Cl-amidine and F-amidine.	22
2.12 LIDAEUS matches ligands to the site points generated on the binding pocket	24
2.13 Chemical structure of potent aminopyrimidine CDK2 inhibitors discovered by LIDAEUS.	25
2.14 Structure of octreotide and goreselin acetate.	28
2.15 Overlay of 10 NMR restrained MD simulation structures of a cyclic peptide.	30
3.1 Overall workflow.	31
3.2 Affinity chromatography.	38
3.3 6xHis protein tag binds to Ni-sepharose resin via coordinate bonds.	39
3.4 Chemical structures of amino acids substituted arginine in peptide-based inhibitors.	43
3.5 General protocol of solid phase peptide synthesis.	45
3.6 Synthesis of peptide-based inhibitors.	47
4.1 PAD4 expressed from vector pReceiver-B13.	56
4.2 PCR products.	56
4.3 Double digested vector and purified PCR product.	57
4.4 <i>E.coli</i> grown on LB agar plate after ligation.	57
4.5 PCR products generated from extracted plasmids.	58
4.6 Double digestion of extracted plasmids.	59
4.7 PAD4 expressed using Studier media.	60
4.8 Specific activity of PAD4 expressed from conventional and auto-induction systems.	61
4.9 Chromatogram of PAD4 purification.	62
4.10 Activity of every fraction of protein after purification.	63
4.11 Coomassive-stained SDS-PAGE of PAD4 before and after purification.	63
4.12 Structure of PAD4 [PDB: 1WDA].	64
4.13 Site points map generated by LIDAEUS in the binding pocket of PAD4.	65

4.14	Percentage of remaining activity of PAD4 in the presence of 22 compounds.	68
4.15	Correlation plot between Autodock binding affinity and the inhibitory activity of the compounds.	69
4.16	Concentration - response graphs of (a) compound 9, (b) 10 and (c) 59 against PAD4.	70
4.17	Binding conformations of (a) compounds 9, (b) 10 and (c) 59 in the PAD4 binding pocket.	71
4.18	Binding pocket of PAD4 with two accessible tunnels labeled as "front door" and "back door" [PDB:1WDA].	73
4.19	Structure of streptonigrin.	74
4.20	Percentage of remaining activity of PAD4 in the presence of 37 compounds.	75
4.21	Structures of compounds that exhibit significant inhibition to PAD4.	76
4.22	Concentration-response graph of Compound 5 against PAD4.	77
4.23	Binding conformation of Compound 5 inside the PAD4 binding pocket.	79
4.24	Binding conformation of streptonigrin inside the PAD4 binding pocket.	80
4.25	Sequence of human nucleophosmin. (UniProt accession no. P06748).	81
4.26	Screening of peptide substrates of PAD4.	82
4.27	Screening of shortened Pep5 as substrate of PAD4.	83
4.28	Overall design strategy for peptide-based inhibitors.	83
4.29	General structure of designed peptide-based inhibitors.	84
4.30	Structures of (a) designed peptide-based inhibitors, (b) BAA, a synthetic substrate and (c) F-amidine, one of the reported inhibitors of PAD4.	85
4.31	Kaiser test results during synthesis of branched peptides.	86
4.32	Chromatogram of Inh Lys.	87
4.33	Mass spectrum of purified Peak 2 of Inh Lys.	88
4.34	Concentration – response graphs of peptide-based inhibitors.	90
4.35	Rapid dilution time course experiment of Inh Dap.	92
4.36	Lineweaver Burk plot of various concentration of Inh Dap.	92
4.37	Michaelis-Menten plot of various concentration of Inh Dap.	93
4.38	Far UV CD spectroscopy of peptide-based inhibitors.	94
4.39	1D proton NMR spectrum of the HN region of Inh Dap.	95
4.40	TOCSY spectrum of Inh Dap.	96
4.41	Partial NOESY spectrum showing the connectivity between residues.	97
4.42	Chemical shift index (CSI) for the residues in the sequence of Inh Dap.	98
4.43	Summary of the interresidue NOEs and $^3J_{\text{NH-HC}\alpha}$ coupling constants of each residue.	98
4.44	Backbone RMSD of 2000 trajectories generated during MD simulation.	99
4.45	Heavy atoms alignment for 10 NMR restrained MD simulation conformations of Inh Dap.	100

4.46	Binding conformation of Inh Dap inside the PAD4 binding pocket.	102
4.47	Conformation of Inh Dap after binding to PAD4.	103



LIST OF APPENDICES

Appendix		Page
A	Amino acid sequence of human protein arginine deiminase type 4.	132
B	Recipes of media, buffers and reagents.	133
C1	Citrulline standard curve.	135
C2	Bradford standard curve.	136
D	Calculation of PAD4 activity.	137
E	Amount of N-terminal protected amino acid required in each coupling process.	138
F	Distance restraints applied in structure calculations of Inh Dap.	139
G	Torsion restraints applied in structure calculations of Inh Dap.	141
H	Rank, compound ID, binding affinity and predicted LogP value of compounds identified by LIDAEUS and AutoDock 4.	142
I	Compound ID and structure of the compounds similar to streptonigrin identified by USR.	145
J	Structures of Pep1 to Pep8.	149
K1	HPLC chromatograms and LCMS spectra of shorten peptides.	151
K2	HPLC chromatograms and LCMS spectra of peptide-based inhibitors.	153
L1	Two-dimensional ^1H NOESY spectrum.	155
L2	Two-dimensional ^1H - ^{13}C HSQC-TOCSY spectrum.	156
L3	One-dimensional ^1H spectrum.	157
M	List of chemical shifts of ^{13}C and ^1H of Inh Dap.	159

LIST OF ABBREVIATIONS

ABPP	Activity-Based Protein Profiling
ACN	Acetonitrile
ACPA	anticitrullinated protein autoantibody
AKA	anti-keratin antibodies
APF	anti-perinuclear factor
Ala/A	alanine
Arg/R	arginine
Asn/N	asparagine
Asp/D	aspartic acid
BAA	N- α -benzoyl arginine amide
BAEE	N- α -benzoyl-L-arginine ethyl ester
BMO	2, 3-Butanedione monoxime
BSA	Bovine serum albumin
BzADMA	benzoyl-N ^G -asymmetrical dimethyl-Arg
bp	base pairs
CCP	cyclic citrullinated peptide
CD	circular dichroism
CSI	Chemical shift index
Cys/C	cystein
DCM	Dichloromethane
DIEA	N, N – diisopropylethylamine
DMARD	Disease Modifying Anti-Rheumatic Drug
DMF	Dimethylformamide
DMSO	Dimethyl sulfoxide
DTT	Dithiothreitol
Dab	L-2,4-diaminobutyric acid
Dap	L-2,3-diaminopropionic acid
Dfa	Dap-Fal
EDTA	Ethylenediaminetetraacetic acid disodium salt dihydrate
Fal	L-2-furylalanine
Fmoc	fluorenylmethyloxycarbonyl
Gln/Q	glutamine
Glu/E	glutamic acid
Gly/G	glycine
HCTU	2-(6-Chloro-1H-benzotriazole-1-yl)-1, 1, 3, 3-tetramethylaminium hexafluorophosphate
HTS	High-Throughput Screening
His/H	histidine
IC ₅₀	half maximal inhibitory concentration
IPTG	Isopropyl β -D-1-thiogalactopyranoside
Ile/I	isoleucine
LB	Luria-bertani
LIDAEUS	Ligand Discovery At Edinburgh University
LPPS	Liquid phase peptide synthesis
Lys/K	L-lysine
MTX	Methotrexate
Met/M	methionine
Mtt	Methyltrityl

NLS	nuclear localization signal
NMF	natural moisturizing factor
NMP	N-methyl-2-pyrrolidone
NSAID	Non-Steroidal Anti-Inflammatory Drug
Orn	L-orthinine
PAD	protein arginine deiminase
PAD4	protein arginine deiminase type IV
Pro/P	proline
RA	rheumatoid arthritis
RF	Rheumatoid factor
RFA	rhodamine conjugated F-amidine
SE	Shared epitope
SNP	single-nucleotide polymorphism
SPPS	solid phase peptide synthesis
Ser/S	serine
TEMED	N, N, N', N'-tetramethylethylenediamine
TFA	trifluoroacetic acid
TIS	Triisopropylsilane
TNF	Tumor Necrosis Factor
Thr/T	threonine
Trp/W	tryptophan
USR	Ultrafast Shape Recognition
vHTS	virtual High-Throughput Screening
Val/V	valine

CHAPTER 1

INTRODUCTION

The immune system protects us against diseases and infections. Substances which are foreign and harmful to our body are called antigens. Examples of antigens are viruses, cancer cells and blood or tissues from other person or species. When antigens enter our body, our immune system will produce antibodies to destroy the harmful substances. Autoimmune disease occurs when our immune system attacks body's own tissues by mistake. It cannot differentiate between healthy body's tissues and antigens. Rheumatoid arthritis (RA) is an autoimmune disease where the immune system attacks synovial membrane around joints. RA is a disease which lasts for a long period of time, and it may affect many tissues and organs, but principally attacks joints. The attacked joints will become swollen, stiff, red and painful. This will lead to joint destruction and functional disability.

About 0.5-1.0% of the adult population is affected by RA (Wegner *et al.*, 2010). It is the second most common type of arthritis that most often starts after 40 years of age and before 60 years of age (Knuckley *et al.*, 2008). Similar to other autoimmune diseases, such as multiple sclerosis and type-1 diabetes, the etiology of the disease is still unknown. The treatment of the disease is only to reduce the symptoms of the disease, but not to cure the disease. Most of the autoimmune diseases are still not curable.

As an autoimmune disease, there are many autoantibodies that react with various autoantigens are detectable in the sera of RA patients. RA associated autoantibodies are useful in diagnosis of the disease. Diagnosis at the early stage of the disease can prevent irreversible joint damage, reducing signs and symptoms of erosion and improving physical function (Conrad *et al.*, 2010). Among the autoantibodies, anticitrullinated protein autoantibody (ACPA) has been documented as a highly specific marker for RA and has the most valuable diagnosis and prognosis potential for RA (van Boekel *et al.*, 2002).

There are five highly related calcium-dependent protein arginine deiminases (PADs), PAD1 to PAD4 and PAD6, exist in humans. Among the isozymes, PAD4 has emerged as a potential therapeutic target for the treatment of RA (Jones *et al.*, 2009). By inhibiting the enzymatic activity of PAD4, it is believed that the development of RA can be suppressed and subsequently the disease can be cured.

Several drugs are available for treating RA. They can be generally divided into three groups: Non-Steroidal Anti-Inflammatory Drugs (NSAIDs), corticosteroidals, and Disease Modifying Anti-Rheumatic Drugs (DMARDs) (Masumoto *et al.*, 1998). Although the existing drugs can treat the disease, they do bring many side effects to the patients. For examples, long term use of NSAIDs will lead to gastrointestinal disturbance such as ulcers and bleeding while corticosteroidals such as prednisone will cause weight gain, increase blood pressure and blood sugar (Masumoto *et al.*, 1998).

The drug that is now considered as the most commonly used drug for RA patients is methotrexate, an examples of DMARDs. The drug will also cause diarrhea, nausea and fatigue (Masumoto *et al.*, 1998). Several strong inactivators of PAD4 have been reported, such as o-F-amidine and Thr-Asp-F-amidine (TDFA) (Causey *et al.*, 2011; Jones *et al.*, 2012). However they inactivate PAD4 irreversibly. Introduction of the inactivators in human body will cause PAD4 to lose its functions permanently and this may lead to other health problems.

Searching for effective drugs for a disease in a short period of time is crucial in curing the disease or even in saving the patients' lives. Yet drug discovery is a time consuming and complex process. The process from discovery to approval of a new drug may take more than 10 years and cost hundreds of millions of Dollars (Morgan *et al.*, 2011). However practice of bioinformatics techniques generally accelerates the drug discovery process and also reduces the overall risk and cost throughout the process (Katara, 2013). Bioinformatics helps in the gene identification for druggable targets and discovery of new targets for new drugs (Chen & Chen, 2008; Katara *et al.*, 2011). Bioinformatics approach supports rational drug discovery and subsequently encourages the application of computer-aided drug design (CADD) in drug development. The first drug approved with the aid of bioinformatics tools is Relenza, an inhibitor of neuraminidase for influenza treatment (MacConnachie, 1999). With the use of bioinformatics tools, more and more drugs were discovered later. For instances, with the use of structure-based drug design, Indinavir was discovered for human immunodeficiency virus (HIV) infection treatment (Askin *et al.*, 1994).

Peptide-based drugs attract attention from public for their attractive advantages such as high specificity and low toxicity in comparison to conventional small molecule drugs. As peptide is a natural substance, introduction of peptide-based drug to human body is believed to have less negative impact compared to small molecules. Strong binding between the peptide-based inhibitor to the receptor can be achieved by structure modification of the existing natural amino acids. Combination of the CADD techniques and peptide-based drug design can be a fascinating method in designing PAD4 inhibitors with strong binding and low toxicity properties.

Since current therapies for rheumatoid arthritis are not completely effective in treating the disease, new drugs with minimum side effects need to be discovered. PAD4 inhibitors were searched by different approaches in this research. The objectives of this study are:

1. To express and purify human PAD4 using recombinant DNA technology.
2. To screen and evaluate inhibitors of PAD4 by structure-based virtual screening.
3. To screen and evaluate inhibitors of PAD4 by ligand-based virtual screening.
4. To design and synthesize peptide-based inhibitors of PAD4.
5. To elucidate inhibition characteristics and structure of PAD4 inhibitor.

REFERENCES

- Acharya, C., Coop, A., Polli, J. E., & Mackerell, A. D., Jr. (2011). Recent advances in ligand-based drug design: relevance and utility of the conformationally sampled pharmacophore approach. *Current Computer-Aided Drug Design*, 7(1), 10-22.
- Aho, K., Koskenvuo, M., Tuominen, J., & Kaprio, J. (1986). Occurrence of rheumatoid arthritis in a nationwide series of twins. *Journal of Rheumatology*, 13(5), 899-902.
- Aletras, A., Barlos, K., Gatos, D., Koutsogianni, S., & Mamos, P. (1995). Preparation of the very acid-sensitive Fmoc-Lys(Mtt)-OH. Application in the synthesis of side-chain to side-chain cyclic peptides and oligolysine cores suitable for the solid-phase assembly of MAPs and TASP. *International Journal of Peptide and Protein Research*, 45(5), 488-496.
- Amblard, M., Fehrentz, J.-A., Martinez, J., & Subra, G. (2006). Methods and protocols of modern solid phase peptide synthesis. *Molecular Biotechnology*, 33(3), 239-254.
- Andrade, M., Chacon, P., Merelo, J., & Moran, F. (1993). Evaluation of secondary structure of proteins from UV circular dichroism spectra using an unsupervised learning neural network. *Protein Engineering*, 6(4), 383-390.
- Anzilotti, C., Pratesi, F., Tommasi, C., & Migliorini, P. (2010). Peptidylarginine deiminase 4 and citrullination in health and disease. *Autoimmunity Reviews*, 9(3), 158-160.
- Aparoy, P., Reddy, K. K., & Reddanna, P. (2012). Structure and ligand based drug design strategies in the development of novel 5- LOX inhibitors. *Current Medicinal Chemistry*, 19(22), 3763-3778.
- Arai, T., Kusubata, M., Kohsaka, T., Shiraiwa, M., Sugawara, K., & Takahara, H. (1995). Mouse uterus peptidylarginine deiminase is expressed in decidual cells during pregnancy. *Journal of Cellular Biochemistry*, 58(3), 269-278.
- Arita, K., Hashimoto, H., Shimizu, T., Nakashima, K., Yamada, M., & Sato, M. (2004). Structural basis for Ca(2+)-induced activation of human PAD4. *Nature Structural & Molecular Biology*, 11(8), 777-783.
- Arita, K., Hashimoto, H., Shimizu, T., Yamada, M., & Sato, M. (2003). Crystallization and preliminary X-ray crystallographic analysis of human peptidylarginine deiminase V. *Acta Crystallographica Section D: Biological Crystallography*, 59(Pt 12), 2332-2333.

- Arita, K., Shimizu, T., Hashimoto, H., Hidaka, Y., Yamada, M., & Sato, M. (2006). Structural basis for histone N-terminal recognition by human peptidylarginine deiminase 4. *Proceedings of the National Academy of Sciences of the United States of America*, 103(14), 5291-5296.
- Asaga, H., Nakashima, K., Senshu, T., Ishigami, A., & Yamada, M. (2001). Immunocytochemical localization of peptidylarginine deiminase in human eosinophils and neutrophils. *Journal of Leukocyte Biology*, 70(1), 46-51.
- Asaga, H., Yamada, M., & Senshu, T. (1998). Selective deimination of vimentin in calcium ionophore-induced apoptosis of mouse peritoneal macrophages. *Biochemical and Biophysical Research Communications*, 243(3), 641-646.
- Askin, D., Eng, K. K., Rossen, K., Purick, R. M., Wells, K. M., Volante, R. P. (1994). Highly diastereoselective reaction of a chiral, non-racemic amide enolate with (S)-glycidyl tosylate. Synthesis of the orally active HIV-1 protease inhibitor L-735,524. *Tetrahedron Letters*, 35(5), 673-676.
- Badiani, K. (2012). Peptides as drugs. *Spring*, 4(2), 84-90.
- Baeten, D., Peene, I., Union, A., Meheus, L., Sebbag, M., Serre, G. (2001). Specific presence of intracellular citrullinated proteins in rheumatoid arthritis synovium: relevance to antifilaggrin autoantibodies. *Arthritis & Rheumatology*, 44(10), 2255-2262.
- Ballester, P. J., Finn, P. W., & Richards, W. G. (2009). Ultrafast shape recognition: evaluating a new ligand-based virtual screening technology. *Journal of Molecular Graphics*, 27(7), 836-845.
- Ballester, P. J., Mangold, M., Howard, N. I., Robinson, R. L., Abell, C., Blumberger, J. (2012). Hierarchical virtual screening for the discovery of new molecular scaffolds in antibacterial hit identification. *Journal of the Royal Society Interface*, 9(77), 3196-3207.
- Ballester, P. J., & Richards, W. G. (2007a). Ultrafast shape recognition for similarity search in molecular databases. *Proceedings of the Royal Society A: Mathematical, Physical and Engineering Science*, 463(2081), 1307-1321.
- Ballester, P. J., & Richards, W. G. (2007b). Ultrafast shape recognition to search compound databases for similar molecular shapes. *Journal of Computational Chemistry*, 28(10), 1711-1723.
- Ballester, P. J., Westwood, I., Laurieri, N., Sim, E., & Richards, W. G. (2010). Prospective virtual screening with Ultrafast Shape Recognition: the identification of novel inhibitors of arylamine N-acetyltransferases. *Journal of the Royal Society Interface*, 7(43), 335-342.
- Barany, G., & Merrifield, R. B. (1977). A new amino protecting group removable by reduction. Chemistry of the dithiasuccinoyl (Dts) function. *Journal of the American Chemical Society*, 99(22), 7363-7365.

- Basus, V. J. (1989). Proton nuclear magnetic resonance assignments. *Methods Enzymology*, 177, 132-149.
- Bathon, J. M., Martin, R. W., Fleischmann, R. M., Tesser, J. R., Schiff, M. H., Keystone, E. C. (2000). A comparison of etanercept and methotrexate in patients with early rheumatoid arthritis. *New England Journal of Medicine* 343(22), 1586-1593.
- Beddell, C. R., Goodford, P. J., Norrington, F. E., Wilkinson, S., & Wootton, R. (1976). Compounds designed to fit a site of known structure in human haemoglobin. *British Journal of Pharmacology*, 57(2), 201-209.
- Blanco, F. J., Rivas, G., & Serrano, L. (1994). A short linear peptide that folds into a native stable [beta]-hairpin in aqueous solution. *Nature Structural and Molecular Biology*, 1(9), 584-590.
- Böhm, H.-J., Flohr, A., & Stahl, M. (2004). Scaffold hopping. *Drug Discovery Today: Technologies*, 1(3), 217-224.
- Bolzan, A. D., & Bianchi, M. S. (2001). Genotoxicity of streptonigrin: a review. *Mutation Research*, 488(1), 25-37.
- Bradford, M. M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry*, 72, 248-254.
- Bray, B. L. (2003). Large-scale manufacture of peptide therapeutics by chemical synthesis. *Nature Reviews Drug Discovery*, 2(7), 587-593.
- Brown, N., & Jacoby, E. (2006). On scaffolds and hopping in medicinal chemistry. *Mini Reviews in Medicinal Chemistry*, 6(11), 1217-1229.
- Brown, T. A. (1990). *Gene cloning: an introduction*. : Chapman and Hall.
- Burr, M. L., Naseem, H., Hinks, A., Eyre, S., Gibbons, L. J., Bowes, J. (2010). PADI4 genotype is not associated with rheumatoid arthritis in a large UK Caucasian population. *Annals of the Rheumatic Diseases*, 69(4), 666-670.
- Carpino, L. A., & Han, G. Y. (1972). 9-Fluorenylmethoxycarbonyl amino-protecting group. *The Journal of Organic Chemistry*, 37(22), 3404-3409.
- Case, D. A., Darden, T. A., Cheatham, T. E., Simmerling, C. L., Wang, J., Duke, R. E. (2012). AMBER 12: University of California, San Francisco.

- Causey, C. P., Jones, J. E., Slack, J. L., Kamei, D., Jones, L. E., Subramanian, V. (2011). The development of N- α -(2-Carboxyl)benzoyl-N5-(2-fluoro-1-iminoethyl)-l-ornithine Amide (o-F-amidine) and N- α -(2-Carboxyl)benzoyl-N5-(2-chloro-1-iminoethyl)-l-ornithine Amide (o-Cl-amidine) as second generation protein arginine deiminase (PAD) inhibitors. *Journal of Medicinal Chemistry*, 54(19), 6919-6935.
- Chang, X., Yamada, R., Sawada, T., Suzuki, A., Kochi, Y., & Yamamoto, K. (2005a). The inhibition of antithrombin by peptidylarginine deiminase 4 may contribute to pathogenesis of rheumatoid arthritis. *Rheumatology*, 44(3), 293-298.
- Chang, X., Yamada, R., Suzuki, A., Kochi, Y., Sawada, T., & Yamamoto, K. (2005b). Citrullination of fibronectin in rheumatoid arthritis synovial tissue. *Rheumatology (Oxford)*, 44(11), 1374-1382.
- Chang, X., Yamada, R., Suzuki, A., Sawada, T., Yoshino, S., Tokuhira, S. (2005c). Localization of peptidylarginine deiminase 4 (PADI4) and citrullinated protein in synovial tissue of rheumatoid arthritis. *Rheumatology*, 44(1), 40-50.
- Chang, X., Zhao, Y., Sun, S., Zhang, Y., & Zhu, Y. (2009). The expression of PADI4 in synovium of rheumatoid arthritis. *Rheumatology International*, 29(12), 1411-1416.
- Chapuy-Regaud, S., Sebbag, M., Baeten, D., Clavel, C., Foulquier, C., De Keyser, F. (2005). Fibrin deimination in synovial tissue is not specific for rheumatoid arthritis but commonly occurs during synovitides. *Journal of Immunology*, 174(8), 5057-5064.
- Chavanas, S., Méchin, M.-C., Nachat, R., Adoue, V., Coudane, F., Serre, G. (2006). Peptidylarginine deiminases and deimination in biology and pathology: Relevance to skin homeostasis. *Journal of Dermatological Science*, 44(2), 63-72.
- Chavanas, S., Méchin, M.-C., Takahara, H., Kawada, A., Nachat, R., Serre, G. (2004). Comparative analysis of the mouse and human peptidylarginine deiminase gene clusters reveals highly conserved non-coding segments and a new human gene, PADI6. *Gene*, 330(0), 19-27.
- Chen, Y. P., & Chen, F. (2008). Identifying targets for drug discovery using bioinformatics. *Expert Opinion on Therapeutic Targets*, 12(4), 383-389.
- Cheung, M.-S., Maguire, M. L., Stevens, T. J., & Broadhurst, R. W. (2010). DANGLE: a Bayesian inferential method for predicting protein backbone dihedral angles and secondary structure. *Journal of Magnetic Resonance*, 202(2), 223-233.
- Choy, E. H., & Panayi, G. S. (2001). Cytokine pathways and joint inflammation in rheumatoid arthritis. *New England Journal of Medicine*, 344(12), 907-916.

- Closse, A., Haefliger, W., Hauser, D., Gubler, H. U., Dewald, B., & Baggiolini, M. (1981). 2, 3-Dihydrobenzofuran-2-ones: a new class of highly potent antiinflammatory agents. *Journal of Medicinal Chemistry*, 24(12), 1465-1471.
- Cohen, S. (1977). A strategy for the chemotherapy of infectious disease. *Science*, 197(4302), 431-432.
- Cohen, S., Cannon, G. W., Schiff, M., Weaver, A., Fox, R., Olsen, N. (2001). Two-year, blinded, randomized, controlled trial of treatment of active rheumatoid arthritis with leflunomide compared with methotrexate. *Arthritis & Rheumatism*, 44(9), 1984-1992.
- Conrad, K., Roggenbuck, D., Reinhold, D., & Dorner, T. (2010). Profiling of rheumatoid arthritis associated autoantibodies. *Autoimmunity Reviews*, 9(6), 431-435.
- Criswell, L. A., Saag, K. G., Mikuls, T. R., Cerhan, J. R., Merlino, L. A., Lum, R. F. (2006). Smoking interacts with genetic risk factors in the development of rheumatoid arthritis among older Caucasian women. *Annals of the Rheumatic Diseases*, 65(9), 1163-1167.
- Cunnane, G., Doran, M., & Bresnihan, B. (2003). Infections and biological therapy in rheumatoid arthritis. *Best Practice & Research Clinical Rheumatology*, 17(2), 345-363.
- Das, K., Butler, G. H., Kwiatkowski, V., Clark, A. D., Jr., Yadav, P., & Arnold, E. (2004). Crystal structures of arginine deiminase with covalent reaction intermediates; implications for catalytic mechanism. *Structure*, 12(4), 657-667.
- de Groot, N. S., Espargaró, A., Morell, M., & Ventura, S. (2008). Studies on bacterial inclusion bodies. *Future Microbiology*, 3(4), 423-435.
- de Rosa, L., Diana, D., Basile, A., Russomanno, A., Isernia, C., Turco, M. C. (2014). Design, structural and biological characterization of a VEGF inhibitor beta-hairpin-constrained peptide. *European Journal of Medicinal Chemistry*, 73, 210-216.
- DeLano, W. L. (2002). The PyMOL Molecular Graphics System, Version 1.1r, Schrödinger, LLC.
- Despres, N., Boire, G., Lopez-Longo, F. J., & Menard, H. A. (1994). The Sa system: a novel antigen-antibody system specific for rheumatoid arthritis. *Journal of Rheumatology*, 21(6), 1027-1033.
- Edwards, C. M. B., Cohen, M. A., & Bloom, S. R. (1999). Peptides as drugs. *QJM*, 92(1), 1-4.

- El-Gabalawy, H. S., & Wilkins, J. A. (2004). Anti-Sa antibodies: prognostic and pathogenetic significance to rheumatoid arthritis. *Arthritis Research & Therapy*, 6(2), 86-89.
- Elliott, M. J., Maini, R. N., Feldmann, M., Kalden, J. R., Antoni, C., Smolen, J. S. (1994). Randomised double-blind comparison of chimeric monoclonal antibody to tumour necrosis factor alpha (cA2) versus placebo in rheumatoid arthritis. *Lancet*, 344(8930), 1105-1110.
- Emsley, P., Lohkamp, B., Scott, W. G., & Cowtan, K. (2010). Features and development of Coot. *Acta Crystallographica Section D*, 66(4), 486-501.
- Esposito, G., Vitale, A. M., Leijten, F. P. J., Strik, A. M., Koonen-Reemst, A. M. C. B., Yurttas, P. (2007). Peptidylarginine deiminase (PAD) 6 is essential for oocyte cytoskeletal sheet formation and female fertility. *Molecular and Cellular Endocrinology*, 273(1-2), 25-31.
- Fan, L. Y., Wang, W. J., Wang, Q., Zong, M., Yang, L., Zhang, H. (2008). A functional haplotype and expression of the PADI4 gene associated with increased rheumatoid arthritis susceptibility in Chinese. *Tissue Antigens*, 72(5), 469-473.
- Farrera-Sinfreu, J., Royo, M., & Albericio, F. (2002). Undesired removal of the Fmoc group by the free ϵ -amino function of a lysine residue. *Tetrahedron Letters*, 43(43), 7813-7815.
- Felson, D. T., Anderson, J. J., & Meenan, R. F. (1990). The comparative efficacy and toxicity of second-line drugs in rheumatoid arthritis. Results of two metaanalyses. *Arthritis & Rheumatology*, 33(10), 1449-1461.
- Felson, D. T., Anderson, J. J., & Meenan, R. F. (1992). Use of short-term efficacy/toxicity tradeoffs to select second-line drugs in rheumatoid arthritis. A metaanalysis of published clinical trials. *Arthritis & Rheumatology*, 35(10), 1117-1125.
- Forslind, K., Ahlmen, M., Eberhardt, K., Hafstrom, I., & Svensson, B. (2004). Prediction of radiological outcome in early rheumatoid arthritis in clinical practice: role of antibodies to citrullinated peptides (anti-CCP). *Annals of the Rheumatic Diseases*, 63(9), 1090-1095.
- Frisch, M. J., Trucks, G. W., Schlegel, H. B., Scuseria, G. E., Robb, M. A., Cheeseman, J. R. (2009). Gaussian 09, Revision A.02. Wallingford CT.
- Galkin, A., Lu, X., Dunaway-Mariano, D., & Herzberg, O. (2005). Crystal structures representing the Michaelis complex and the thiouronium reaction intermediate of *Pseudomonas aeruginosa* arginine deiminase. *Journal of Biological Chemistry*, 280(40), 34080-34087.

- Gangwal, R. P., Dhoke, G. V., Damre, M. V., Khandelwal, K., & Sangamwar, A. T. (2013). Structure-based virtual screening and molecular dynamic simulation studies to identify novel cytochrome bc1 inhibitors as antimalarial agents. *Journal of Computational Medicine*, 2013, 9.
- Geppert, T., Bauer, S., Hiss, J. A., Conrad, E., Reutlinger, M., Schneider, P. (2012). Immunosuppressive small molecule discovered by structure-based virtual screening for inhibitors of protein–protein interactions. *Angewandte Chemie International Edition*, 51(1), 258-261.
- Gerstein, A. S. (2001). *Molecular biology problem solver: a laboratory guide*. : John Wiley & Sons Ltd.
- Girbal-Neuhausser, E., Durieux, J.-J., Arnaud, M., Dalbon, P., Sebbag, M., Vincent, C. (1999). The epitopes targeted by the rheumatoid arthritis-associated antifilaggrin autoantibodies are posttranslationally generated on various sites of (pro)filaggrin by deimination of arginine residues. *The Journal of Immunology*, 162(1), 585-594.
- Goldbach-Mansky, R., Lee, J., McCoy, A., Hoxworth, J., Yarboro, C., Smolen, J. S. (2000). Rheumatoid arthritis associated autoantibodies in patients with synovitis of recent onset. *Arthritis Research*, 2(3), 236-243.
- Goodsell, D. S., Morris, G. M., & Olson, A. J. (1996). Automated docking of flexible ligands: applications of AutoDock. *Journal of Molecular Recognition*, 9(1), 1-5.
- Grabski, A., Mehler, M., & Drott, D. (2005). The Overnight Express Autoinduction System: High-density cell growth and protein expression while you sleep. *Nature Methods*, 2(3), 233-235.
- Greenfield, N. J. (2007). Using circular dichroism spectra to estimate protein secondary structure. *Nature protocols*, 1(6), 2876-2890.
- Guo, Q., & Fast, W. (2011). Citrullination of inhibitor of growth 4 (ING4) by peptidylarginine deiminase 4 (PAD4) disrupts the interaction between ING4 and p53. *Journal of Biological Chemistry*, 286(19), 17069-17078.
- Hagiwara, T., Hidaka, Y., & Yamada, M. (2005). Deimination of histone H2A and H4 at arginine 3 in HL-60 granulocytes. *Biochemistry*, 44(15), 5827-5834.
- Hagiwara, T., Nakashima, K., Hirano, H., Senshu, T., & Yamada, M. (2002). Deimination of arginine residues in nucleophosmin/B23 and histones in HL-60 granulocytes. *Biochemical and Biophysical Research Communications*, 290(3), 979-983.
- Hall, P. R., Malone, L., Sillerud, L. O., Ye, C., Hjelle, B. L., & Larson, R. S. (2007). Characterization and NMR solution structure of a novel cyclic pentapeptide inhibitor of pathogenic hantaviruses. *Chemical Biology & Drug Design*, 69(3), 180-190.

- Hann, M. M., & Oprea, T. I. (2004). Pursuing the leadlikeness concept in pharmaceutical research. *Current Opinion in Chemical Biology*, 8(3), 255-263.
- Haraoui, B., & Bykerk, V. (2007). Etanercept in the treatment of rheumatoid arthritis. *Journal of Therapeutics and Clinical Risk Management*, 3(1), 99-105.
- Harney, S. M., Meisel, C., Sims, A. M., Woon, P. Y., Wordsworth, B. P., & Brown, M. A. (2005). Genetic and genomic studies of PADI4 in rheumatoid arthritis. *Rheumatology (Oxford)*, 44(7), 869-872.
- Hassfeld, W., Steiner, G., Graninger, W., Witzmann, G., Schweitzer, H., & Smolen, J. S. (1993). Autoantibody to the nuclear antigen RA33: a marker for early rheumatoid arthritis. *British Journal of Rheumatology*, 32(3), 199-203.
- Hassfeld, W., Steiner, G., Hartmuth, K., Kolarz, G., Scherak, O., Graninger, W. (1989). Demonstration of a new antinuclear antibody (anti-RA33) that is highly specific for rheumatoid arthritis. *Arthritis & Rheumatology*, 32(12), 1515-1520.
- Hayem, G., Chazerain, P., Combe, B., Elias, A., Haim, T., Nicaise, P. (1999). Anti-Sa antibody is an accurate diagnostic and prognostic marker in adult rheumatoid arthritis. *Journal of Rheumatology*, 26(1), 7-13.
- Hehemann, J.-H., Correc, G., Barbeyron, T., Helbert, W., Czjzek, M., & Michel, G. (2010). Transfer of carbohydrate-active enzymes from marine bacteria to Japanese gut microbiota. *Nature*, 464(7290), 908-912.
- Hidaka, Y., Hagiwara, T., & Yamada, M. (2005). Methylation of the guanidino group of arginine residues prevents citrullination by peptidylarginine deiminase IV. *FEBS Letters*, 579(19), 4088-4092.
- Hoppe, B., Haupl, T., Gruber, R., Kiesewetter, H., Burmester, G. R., Salama, A. (2006). Detailed analysis of the variability of peptidylarginine deiminase type 4 in German patients with rheumatoid arthritis: a case-control study. *Arthritis Research & Therapy*, 8(2), R34.
- Hornak, V., Abel, R., Okur, A., Strockbine, B., Roitberg, A., & Simmerling, C. (2006). Comparison of multiple Amber force fields and development of improved protein backbone parameters. *Proteins*, 65(3), 712-725.
- Howe, C. (2007). *Gene cloning and manipulation*: Cambridge University Press.
- Hoyer, J., Schatzschneider, U., Schulz-Siegmund, M., & Neundorff, I. (2012). Dimerization of a cell-penetrating peptide leads to enhanced cellular uptake and drug delivery. *Beilstein Journal of Organic Chemistry*, 8, 1788-1797.

- Huizinga, T. W., Amos, C. I., van der Helm-van Mil, A. H., Chen, W., van Gaalen, F. A., Jawaheer, D. (2005). Refining the complex rheumatoid arthritis phenotype based on specificity of the HLA-DRB1 shared epitope for antibodies to citrullinated proteins. *Arthritis & Rheumatology*, 52(11), 3433-3438.
- Humm, A., Fritsche, E., Steinbacher, S., & Huber, R. (1997). Crystal structure and mechanism of human L-arginine:glycine amidinotransferase: a mitochondrial enzyme involved in creatine biosynthesis. *EMBO J*, 16(12), 3373-3385.
- Ishida-Yamamoto, A., Senshu, T., Eady, R. A. J., Takahashi, H., Shimizu, H., Akiyama, M. (2002). Sequential reorganization of cornified cell keratin filaments involving filaggrin-mediated compaction and keratin 1 deimination. *Journal of Investigative Dermatology* 118(2), 282-287.
- Iwamoto, T., Ikari, K., Nakamura, T., Kuwahara, M., Toyama, Y., Tomatsu, T. (2006). Association between PADI4 and rheumatoid arthritis: a meta-analysis. *Rheumatology (Oxford)*, 45(7), 804-807.
- J.A. Joule, K. Mills, & Smith, G. F. (1995). *Heterocyclic Chemistry*: Chapman & Hall, London.
- Jia, D., Jurkowska, R. Z., Zhang, X., Jeltsch, A., & Cheng, X. (2007). Structure of Dnmt3a bound to Dnmt3L suggests a model for de novo DNA methylation. *Nature*, 449(7159), 248-251.
- Jones, J. E., Causey, C., Knuckley, B., Slack-Noyes, J. L., & Thompson, P. R. (2009). Protein arginine deiminase 4 (PAD4): Current understanding and future therapeutic potential. *Current opinion in drug discovery & development*, 12(5), 616.
- Jones, J. E., Slack, J. L., Fang, P., Zhang, X., Subramanian, V., Causey, C. P. (2012). Synthesis and screening of a haloacetamide containing library to identify PAD4 selective inhibitors. *ACS Chemical Biology*, 7(1), 160-165.
- Kabsch, W., & Sander, C. (1983). Dictionary of protein secondary structure: pattern recognition of hydrogen-bonded and geometrical features. *Biopolymers*, 22(12), 2577-2637.
- Kameda, H., Ueki, Y., Saito, K., Nagaoka, S., Hidaka, T., Atsumi, T. (2010). Etanercept (ETN) with methotrexate (MTX) is better than ETN monotherapy in patients with active rheumatoid arthritis despite MTX therapy: a randomized trial. *Modern Rheumatology*, 20(6), 531-538.
- Kang, C. P., Lee, H. S., Ju, H., Cho, H., Kang, C., & Bae, S. C. (2006). A functional haplotype of the PADI4 gene associated with increased rheumatoid arthritis susceptibility in Koreans. *Arthritis and Rheumatism*, 54(1), 90-96.

- Kapitany, A., Szabo, Z., Lakos, G., Aleksza, M., Vegvari, A., Soos, L. (2008). Associations between serum anti-CCP antibody, rheumatoid factor levels and HLA-DR4 expression in Hungarian patients with rheumatoid arthritis. *Israel Medical Association Journal*, 10(1), 32-36.
- Katara, P. (2013). Role of bioinformatics and pharmacogenomics in drug discovery and development process. *Network Modeling Analysis in Health Informatics and Bioinformatics*, 2(4), 225-230.
- Katara, P., Grover, A., Kuntal, H., & Sharma, V. (2011). In silico prediction of drug targets in *Vibrio cholerae*. *Protoplasma*, 248(4), 799-804.
- Katti, S. K., LeMaster, D. M., & Eklund, H. (1990). Crystal structure of thioredoxin from *Escherichia coli* at 1.68 Å resolution. *Journal of Molecular Biology*, 212(1), 167-184.
- Kavanaugh, A., St Clair, E. W., McCune, W. J., Braakman, T., & Lipsky, P. (2000). Chimeric anti-tumor necrosis factor- α monoclonal antibody treatment of patients with rheumatoid arthritis receiving methotrexate therapy. *Journal of Rheumatology*, 27(4), 841-850.
- Kearney, P. L., Bhatia, M., Jones, N. G., Yuan, L., Glascock, M. C., Catchings, K. L. (2005). Kinetic characterization of protein arginine deiminase 4: a transcriptional corepressor implicated in the onset and progression of rheumatoid arthritis. *Biochemistry*, 44(31), 10570-10582.
- Kelley, L. A., Gardner, S. P., & Sutcliffe, M. J. (1996). An automated approach for clustering an ensemble of NMR-derived protein structures into conformationally related subfamilies. *Protein Engineering*, 9(11), 1063-1065.
- Kelly, S. M., & Price, N. C. (2000). The use of circular dichroism in the investigation of protein structure and function. *Current protein and peptide science*, 1(4), 349-384.
- Knuckley, B., Bhatia, M., & Thompson, P. R. (2007). Protein arginine deiminase 4: evidence for a reverse protonation mechanism. *Biochemistry*, 46(22), 6578-6587.
- Knuckley, B., Jones, J. E., Bachovchin, D. A., Slack, J., Causey, C. P., Brown, S. J. (2010). A fluopol-ABPP HTS assay to identify PAD inhibitors. *Chemical communications (Cambridge, England)*, 46(38), 7175-7177.
- Knuckley, B., Luo, Y., & Thompson, P. R. (2008). Profiling Protein Arginine Deiminase 4 (PAD4): a novel screen to identify PAD4 inhibitors. *Bioorganic & Medicinal Chemistry*, 16(2), 739-745.
- Kolb, W. P., Kolb, L. M., & Podack, E. R. (1979). C1q: isolation from human serum in high yield by affinity chromatography and development of a highly sensitive hemolytic assay. *Journal of Immunology*, 122(5), 2103-2111.

- Kremer, J. M. (1997). Safety, efficacy, and mortality in a long-term cohort of patients with rheumatoid arthritis taking methotrexate: followup after a mean of 13.3 years. *Arthritis & Rheumatology*, 40(5), 984-985.
- Kroot, E. J., de Jong, B. A., van Leeuwen, M. A., Swinkels, H., van den Hoogen, F. H., van't Hof, M. (2000). The prognostic value of anti-cyclic citrullinated peptide antibody in patients with recent-onset rheumatoid arthritis. *Arthritis & Rheumatology*, 43(8), 1831-1835.
- Kubilus, J., & Baden, H. P. (1983). Purification and properties of a brain enzyme which deiminates proteins. *Biochimica et Biophysica Acta*, 745(3), 285-291.
- Kuntz, I. D., Blaney, J. M., Oatley, S. J., Langridge, R., & Ferrin, T. E. (1982). A geometric approach to macromolecule-ligand interactions. *Journal of Molecular Biology*, 161(2), 269-288.
- Kupchan, S. M., Eakin, M. A., & Thomas, A. M. (1971). Tumor inhibitors. 69. Structure-cytotoxicity relationships among the sesquiterpene lactones. *Journal of Medicinal Chemistry*, 14(12), 1147-1152.
- Kurutz, J. W., & Lee, K. Y. (2002). NMR structure of lung surfactant peptide SP-B(11-25). *Biochemistry*, 41(30), 9627-9636.
- LaVallie, E. R., Lu, Z., Diblasio-Smith, E. A., Collins-Racie, L. A., & McCoy, J. M. (2000). Thioredoxin as a fusion partner for production of soluble recombinant proteins in *Escherichia coli*. *Methods Enzymology*, 326, 322-340.
- Laemmli, U. K. (1970). Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature*, 227(5259), 680-685.
- Lam, G. (2006). Round 4: Citrullinated proteins, peptidylarginine deiminase (PAD), and rheumatoid arthritis. Retrieved 19/02/2014, from <http://www.hopkinsarthritis.org/physician-corner/rheumatology-rounds/round-4-citrullinated-proteins-peptidylarginine-deiminase-pad-and-rheumatoid-arthritis/#ref1>
- Landre-Beauvais, A. J. (2001). The first description of rheumatoid arthritis. Unabridged text of the doctoral dissertation presented in 1800. *Joint Bone Spine*, 68(2), 130-143.
- Laskowski, R. A., Macarthur, M. W., Moss, D. S., & Thornton, J. M. (1993). {PROCHECK}: a program to check the stereochemical quality of protein structures. *Journal of Applied Crystallography*, 26, 283-291.
- Lax, R. (2010). The future of peptide development in the pharmaceutical industry. *PharManufacturing: The International Peptide Review*, 10-15.
- Lee, C. C., Wong, D. W., & Robertson, G. H. (2001). An *E. coli* expression system for the extracellular secretion of barley alpha-amylase. *Journal of Protein Chemistry*, 20(3), 233-237

- Lee, C. H., Huang, H. C., & Juan, H. F. (2011). Reviewing ligand-based rational drug design: the search for an ATP synthase inhibitor. *International Journal of Molecular Sciences*, 12(8), 5304-5318.
- Lee, D. M., & Weinblatt, M. E. (2001). Rheumatoid arthritis. *Lancet*, 358(9285), 903-911.
- Legrand, B., Mathieu, L., Lebrun, A., Andriamanarivo, S., Lisowski, V., Masurier, N. (2014). Thiazole-based γ -building blocks as reverse-turn mimetic to design a gramicidin S analogue: conformational and biological evaluation. *Chemistry – A European Journal*, 20(22), 6713-6720.
- Leone, M., Barile, E., Dahl, R., & Pellicchia, M. (2011). Design and NMR studies of cyclic peptides targeting the N-terminal domain of the protein tyrosine phosphatase YopH. *Chemical Biology & Drug Design*, 77(1), 12-19.
- Li, D., & Elbert, D. L. (2002). The kinetics of the removal of the N-methyltrityl (Mtt) group during the synthesis of branched peptides. *The Journal of Peptide Research*, 60(5), 300-303.
- Li, S., McGuire, M. J., Lin, M., Liu, Y.-H., Oyama, T., Sun, X. (2009). Synthesis and characterization of a high-affinity $\alpha\beta 6$ -specific ligand for in vitro and in vivo applications. *Molecular Cancer Therapeutics*, 8(5), 1239-1249.
- Lien, S., & Lowman, H. B. (2003). Therapeutic peptides. *Trends in Biotechnology*, 21(12), 556-562.
- Lim, S. V., Rahman, M. B., & Tejo, B. A. (2011). Structure-based and ligand-based virtual screening of novel methyltransferase inhibitors of the dengue virus. *BMC Bioinformatics*, 12 Suppl 13, S24.
- Lipinski, C. A., Lombardo, F., Dominy, B. W., & Feeney, P. J. (2001). Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. *Advanced Drug Delivery Reviews*, 46(1-3), 3-26.
- Lipsky, P. E., van der Heijde, D. M., St Clair, E. W., Furst, D. E., Breedveld, F. C., Kalden, J. R. (2000). Infliximab and methotrexate in the treatment of rheumatoid arthritis. Anti-tumor necrosis factor trial in rheumatoid arthritis with concomitant therapy study group. *New England Journal of Medicine*, 343(22), 1594-1602.
- Loffet, A. (2002). Peptides as drugs: is there a market? *Journal of Peptide Science*, 8(1), 1-7.
- Luo, Y., Arita, K., Bhatia, M., Knuckley, B., Lee, Y. H., Stallcup, M. R. (2006a). Inhibitors and inactivators of protein arginine deiminase 4: functional and structural characterization. *Biochemistry*, 45(39), 11727-11736.

- Luo, Y., Knuckley, B., Bhatia, M., Pellechia, P. J., & Thompson, P. R. (2006b). Activity-based protein profiling reagents for protein arginine deiminase 4 (PAD4): synthesis and in vitro evaluation of a fluorescently labeled probe. *Journal of the American Chemical Society*, 128(45), 14468-14469.
- Luo, Y., Knuckley, B., Lee, Y. H., Stallcup, M. R., & Thompson, P. R. (2006c). A fluoroacetamide-based inactivator of protein arginine deiminase 4: design, synthesis, and in vitro and in vivo evaluation. *Journal of the American Chemical Society*, 128(4), 1092-1093.
- MacConnachie, A. M. (1999). Zanamivir (Relenza®) — A new treatment for influenza. *Intensive and Critical Care Nursing*, 15(6), 369-370.
- MacGregor, A. J., Snieder, H., Rigby, A. S., Koskenvuo, M., Kaprio, J., Aho, K. (2000). Characterizing the quantitative genetic contribution to rheumatoid arthritis using data from twins. *Arthritis and Rheumatism*, 43(1), 30-37.
- Maini, R. N., Breedveld, F. C., Kalden, J. R., Smolen, J. S., Davis, D., Macfarlane, J. D. (1998). Therapeutic efficacy of multiple intravenous infusions of anti-tumor necrosis factor alpha monoclonal antibody combined with low-dose weekly methotrexate in rheumatoid arthritis. *Arthritis & Rheumatology*, 41(9), 1552-1563.
- Masson-Bessière, C., Sebbag, M., Durieux, J. J., Nogueira, L., Vincent, C., Girbal-Neuhauser, E. (2000). In the rheumatoid pannus, anti-filaggrin autoantibodies are produced by local plasma cells and constitute a higher proportion of IgG than in synovial fluid and serum. *Clinical & Experimental Immunology*, 119(3), 544-552.
- Masson-Bessiere, C., Sebbag, M., Girbal-Neuhauser, E., Nogueira, L., Vincent, C., Senshu, T. (2001). The major synovial targets of the rheumatoid arthritis-specific antifilaggrin autoantibodies are deiminated forms of the alpha- and beta-chains of fibrin. *Journal of Immunology*, 166(6), 4177-4184.
- Mastronardi, F. G., Wood, D. D., Mei, J., Rajmakers, R., Tseveleki, V., Dosch, H.-M. (2006). Increased citrullination of histone H3 in multiple sclerosis brain and animal models of demyelination: a role for tumor necrosis factor-induced peptidylarginine deiminase 4 translocation. *The Journal of Neuroscience*, 26(44), 11387-11396.
- Masumoto, A. K., Bathon, J., & Bingham III, C. O. (1998, 24 September 2013). Rheumatoid arthritis treatment. Retrieved December, 2010, from http://www.hopkins-arthritis.org/arthritis-info/rheumatoid-arthritis/rheum_treat.html
- Matsuura, H., Saxena, G., Farmer, S. W., Hancock, R. E., & Towers, G. H. (1996). Antibacterial and antifungal polyine compounds from *Glehnia littoralis* ssp. *leiocarpa*. *Planta Med*, 62(3), 256-259.

- Mauser, H., & Guba, W. (2008). Recent developments in de novo design and scaffold hopping. *Current Opinion in Drug Discovery Development*, 11(3), 365-374.
- McInnes, C. (2007). Recent advances in the structure-guided design of protein kinase inhibitors. In Atta-Ur_Rahman, G. W. Caldwell, M. I. Choudhary, & M. R. Player (Eds.), *Frontiers in drug design and discovery* (Vol. 3, pp. 145-170): Bentham Science Publishers.
- Mechin, M. C., Enji, M., Nachat, R., Chavanas, S., Charveron, M., Ishida-Yamamoto, A. (2005). The peptidylarginine deiminases expressed in human epidermis differ in their substrate specificities and subcellular locations. *Cellular and Molecular Life Sciences*, 62(17), 1984-1995.
- Menard, H. A., Lapointe, E., Rochdi, M. D., & Zhou, Z. J. (2000). Insights into rheumatoid arthritis derived from the Sa immune system. *Arthritis Research*, 2(6), 429 - 432.
- Meotti, F. C., Silva, D. O., Dos Santos, A. R., Zeni, G., Rocha, J. B., & Nogueira, C. W. (2003). Thiophenes and furans derivatives: a new class of potential pharmacological agents. *Environ Toxicol Pharmacol*, 15(1), 37-44.
- Merrifield, R. B. (1963). Solid Phase Peptide Synthesis. I. The synthesis of a tetrapeptide. *Journal of the American Chemical Society*, 85(14), 2149-2154.
- Merrifield, R. B. (1964). Solid-Phase Peptide Synthesis. 3. An improved synthesis of bradykinin. *Biochemistry*, 3, 1385-1390.
- Mewar, D., & Wilson, A. G. (2006). Autoantibodies in rheumatoid arthritis: a review. *Biomedicine & Pharmacotherapy*, 60(10), 648-655.
- Meyer, O., Labarre, C., Dougados, M., Goupille, P., Cantagrel, A., Dubois, A. (2003). Anticitrullinated protein/peptide antibody assays in early rheumatoid arthritis for predicting five year radiographic damage. *Annals of the Rheumatic Diseases*, 62(2), 120-126.
- Mladenovic, V., Domljan, Z., Rozman, B., Jajic, I., Mihajlovic, D., Dordevic, J. (1995). Safety and effectiveness of leflunomide in the treatment of patients with active rheumatoid arthritis. Results of a randomized, placebo-controlled, phase II study. *Arthritis & Rheumatology*, 38(11), 1595-1603.
- Mohanan, S., Cherrington, B. D., Horibata, S., McElwee, J. L., Thompson, P. R., & Coonrod, S. A. (2012). Potential role of peptidylarginine deiminase enzymes and protein citrullination in cancer pathogenesis. *Biochemistry Research International*, 2012, 895343.
- Mor-Vaknin, N., Punturieri, A., Sitwala, K., & Markovitz, D. M. (2003). Vimentin is secreted by activated macrophages. *Nature Cell Biology*, 5(1), 59-63.

- Morand, E. F., McCloud, P. I., & Littlejohn, G. O. (1992). Life table analysis of 879 treatment episodes with slow acting antirheumatic drugs in community rheumatology practice. *Journal of Rheumatology*, 19(5), 704-708.
- Moreland, L. W., Margolies, G., Heck, L. W., Jr., Saway, A., Blosch, C., Hanna, R. (1996). Recombinant soluble tumor necrosis factor receptor (p80) fusion protein: toxicity and dose finding trial in refractory rheumatoid arthritis. *Journal of Rheumatology*, 23(11), 1849-1855.
- Morgan, S., Grootendorst, P., Lexchin, J., Cunningham, C., & Greyson, D. (2011). The cost of drug development: a systematic review. *Health Policy*, 100(1), 4-17.
- Moscarello, M. A., Wood, D. D., Ackerley, C., & Boulias, C. (1994). Myelin in multiple sclerosis is developmentally immature. *Journal of Clinical Investigation*, 94(1), 146-154.
- Muhamad, A., Ho, K. L., A. Rahman, M. B., Uhrin, D., & Tan, W. S. (2013). Solution structure and in silico binding of a cyclic peptide with Hepatitis B surface antigen. *Chemical Biology & Drug Design*, 81(6), 784-794.
- Murray-Rust, J., Leiper, J., McAlister, M., Phelan, J., Tilley, S., Santa Maria, J. (2001). Structural insights into the hydrolysis of cellular nitric oxide synthase inhibitors by dimethylarginine dimethylaminohydrolase. *Nature Structural and Molecular Biology*, 8(8), 679-683.
- Nachat, R., Mechin, M. C., Charveron, M., Serre, G., Constans, J., & Simon, M. (2005a). Peptidylarginine deiminase isoforms are differentially expressed in the anagen hair follicles and other human skin appendages. *Journal of Investigative Dermatology*, 125(1), 34-41.
- Nachat, R., Mechin, M. C., Takahara, H., Chavanas, S., Charveron, M., Serre, G. (2005b). Peptidylarginine deiminase isoforms 1-3 are expressed in the epidermis and involved in the deimination of K1 and filaggrin. *Journal of Investigative Dermatology*, 124(2), 384-393.
- Nakayama-Hamada, M., Suzuki, A., Kubota, K., Takazawa, T., Ohsaka, M., Kawaida, R. (2005). Comparison of enzymatic properties between hPADI2 and hPADI4. *Biochemical and Biophysical Research Communications*, 327(1), 192-200.
- Nielen, M. M., van Schaardenburg, D., Reesink, H. W., van de Stadt, R. J., van der Horst-Bruinsma, I. E., de Koning, M. H. (2004). Specific autoantibodies precede the symptoms of rheumatoid arthritis: a study of serial measurements in blood donors. *Arthritis & Rheumatology*, 50(2), 380-386.
- Nienhuis, R. L., & Mandema, E. (1964). A new serum factor in patients with rheumatoid arthritis; the antiperinuclear factor. *Annals of the Rheumatic Disease*, 23, 302-305.

Nordqvist, C. (2010, April 2013). All About Rheumatoid Arthritis. Retrieved 11 March, 2014, from <http://www.medicalnewstoday.com/info/rheumatoid-arthritis/treatment-for-rheumatoid-arthritis.php>

Number Of Therapeutic Peptides In Clinical Study Has Nearly Doubled Since The 1990s. (2009). Retrieved from: http://csdd.tufts.edu/news/complete_story/ir_may_09_pr

Okutani, R., Itoh, Y., Hirata, H., Kasahara, T., Mukaida, N., & Kawai, T. (1992). Simple and high-yield purification of urine protein 1 using immunoaffinity chromatography: evidence for the identity of urine protein 1 and human Clara cell 10-kilodalton protein. *Journal of Chromatography*, 577(1), 25-35.

Oleson, J. J., Calderella, L. A., Mjos, K. J., Reith, A. R., Thie, R. S., & Toplin, I. (1961). The effects of streptonigrin on experimental tumors. *Antibiotic and Chemotherapy (Northfield Ill)*, 11, 158-164.

Oliver, J. E., & Silman, A. J. (2006). Risk factors for the development of rheumatoid arthritis. *Scandinavian Journal of Rheumatology*, 35(3), 169-174.

Payne, D. J., Gwynn, M. N., Holmes, D. J., & Pompliano, D. L. (2007). Drugs for bad bugs: confronting the challenges of antibacterial discovery. *Nature Reviews Drug Discovery*, 6(1), 29-40.

Phillips, K., Husni, M. E., Karlson, E. W., & Coblyn, J. S. (2002). Experience with etanercept in an academic medical center: are infection rates increased? *Arthritis & Rheumatology*, 47(1), 17-21.

Pichereau, C., & Allary, C. (2005). Therapeutic peptides under the spotlight. *EBR - European Biopharmaceutical Review*(WINTER), 88-93.

Pincus, T., Marcum, S. B., & Callahan, L. F. (1992). Longterm drug therapy for rheumatoid arthritis in seven rheumatology private practices: II. Second line drugs and prednisone. *Journal of Rheumatology*, 19(12), 1885-1894.

Porter, C. J., & Wilce, J. A. (2007). NMR analysis of G7-18NATE, a nonphosphorylated cyclic peptide inhibitor of the Grb7 adapter protein. *Peptide Science*, 88(2), 174-181.

Powell, B., Crocker, L., & Rogers, G. (1992). Hair follicle differentiation: expression, structure and evolutionary conservation of the hair type II keratin intermediate filament gene family. *Development*, 114(2), 417-433.

Pritzker, L. B., & Moscarello, M. A. (1998). A novel microtubule independent effect of paclitaxel: the inhibition of peptidylarginine deiminase from bovine brain. *Biochimica et Biophysica Acta*, 1388(1), 154-160.

Provencher, S. W., & Glockner, J. (1981). Estimation of globular protein secondary structure from circular dichroism. *Biochemistry*, 20(1), 33-37.

- Rantapaa-Dahlqvist, S., de Jong, B. A., Berglin, E., Hallmans, G., Wadell, G., Stenlund, H. (2003). Antibodies against cyclic citrullinated peptide and IgA rheumatoid factor predict the development of rheumatoid arthritis. *Arthritis & Rheumatology*, 48(10), 2741-2749.
- Rawat, A., & Kumar, D. (2013). NMR investigations of structural and dynamics features of natively unstructured drug peptide - salmon calcitonin: implication to rational design of potent sCT analogs. *Journal of Peptide Science*, 19(1), 33-45.
- Reddy, A. S., Pati, S. P., Kumar, P. P., Pradeep, H. N., & Sastry, G. N. (2007). Virtual screening in drug discovery -- a computational perspective. *Current Protein & Peptide Science*, 8(4), 329-351.
- Reetz, M. T., Carballeira, J. D., & Vogel, A. (2006). Iterative saturation mutagenesis on the basis of B factors as a strategy for increasing protein thermostability. *Angewandte Chemie International Edition*, 45(46), 7745-7751.
- Reichert, J. (2010). Development trends for peptide therapeutics (2010 Report Summary) (pp. 1-10): Peptide Therapeutics Foundation.
- Reparon-Schuijt, C. C., van Esch, W. J. E., van Kooten, C., Schellekens, G. A., de Jong, B. A. W., van Venrooij, W. J. (2001). Secretion of anti-citrulline-containing peptide antibody by B lymphocytes in rheumatoid arthritis. *Arthritis & Rheumatism*, 44(1), 41-47.
- Rink, H. (1987). Solid-phase synthesis of protected peptide fragments using a trialkoxy-diphenyl-methylester resin. *Tetrahedron Letters*, 28(33), 3787-3790.
- Rogers, G., Winter, B., McLaughlan, C., Powell, B., & Nesci, T. (1997). Peptidylarginine deiminase of the hair follicle: characterization, localization, and function in keratinizing tissues. *Journal of Investigative Dermatology*, 108(5), 700-707.
- Rogers, G. E., Harding, H. W., & Llewellyn-Smith, I. J. (1977). The origin of citrulline-containing proteins in the hair follicle and the chemical nature of trichohyalin, an intracellular precursor. *Biochimica et Biophysica Acta*, 495(1), 159-175.
- Roque, A. C. A., & Lowe, C. R. (2008). Affinity chromatography: history, perspectives, limitations and prospects. In M. Zachariou (Ed.), *Affinity Chromatography: Methods and Protocols* (Vol. 421, pp. 1-15): Humana Press.
- Rothschild, B. M., Coppa, A., & Petrone, P. P. (2004). "Like a virgin": Absence of rheumatoid arthritis and treponematosi, good sanitation and only rare gout in Italy prior to the 15th century. *Reumatismo*, 56(1), 61-66.
- Rush, T. S., 3rd, Grant, J. A., Mosyak, L., & Nicholls, A. (2005). A shape-based 3-D scaffold hopping method and its application to a bacterial protein-protein interaction. *Journal of Medicinal Chemistry*, 48(5), 1489-1495.

- Saiki, R., Gelfand, D., Stoffel, S., Scharf, S., Higuchi, R., Horn, G. (1988). Primer-directed enzymatic amplification of DNA with a thermostable DNA polymerase. *Science*, 239(4839), 487-491.
- Sany, J., Anaya, J. M., Lussiez, V., Couret, M., Combe, B., & Daures, J. P. (1991). Treatment of rheumatoid arthritis with methotrexate: a prospective open longterm study of 191 cases. *Journal of Rheumatology*, 18(9), 1323-1327.
- Sarduy, E. S., Muñoz, A. C., Trejo, S. A., & Chavéz Planes, M. d. l. A. (2012). High-level expression of Falcipain-2 in *Escherichia coli* by codon optimization and auto-induction. *Protein Expression and Purification*, 83(1), 59-69.
- Sax, B., Dick, F., Tanner, R., & Gosteli, J. (1992). 4-Methyltrityl (Mtt): a new protecting group for the side chain protection of Asn and Gln in solid-phase peptide synthesis. *Peptide Research*, 5(4), 245-246.
- Schellekens, G. A., de Jong, B. A., van den Hoogen, F. H., van de Putte, L. B., & van Venrooij, W. J. (1998). Citrulline is an essential constituent of antigenic determinants recognized by rheumatoid arthritis-specific autoantibodies. *Journal of Clinical Investigation*, 101(1), 273-281.
- Schellekens, G. A., Visser, H., de Jong, B. A., van den Hoogen, F. H., Hazes, J. M., Breedveld, F. C. (2000). The diagnostic properties of rheumatoid arthritis antibodies recognizing a cyclic citrullinated peptide. *Arthritis & Rheumatology*, 43(1), 155-163.
- Schnecke, V., & Bostrom, J. (2006). Computational chemistry-driven decision making in lead generation. *Drug Discovery Today*, 11(1-2), 43-50.
- Schneider, G., Neidhart, W., Giller, T., & Schmid, G. (1999). "Scaffold-Hopping" by topological pharmacophore search: a contribution to virtual screening. *Angewandte Chemie International Edition in English*, 38(19), 2894-2896.
- Sebbag, M., Moinard, N., Auger, I., Clavel, C., Arnaud, J., Nogueira, L. (2006). Epitopes of human fibrin recognized by the rheumatoid arthritis-specific autoantibodies to citrullinated proteins. *European Journal of Immunology*, 36(8), 2250-2263.
- Senshu, T., Akiyama, K., Ishigami, A., & Nomura, K. (1999). Studies on specificity of peptidylarginine deiminase reactions using an immunochemical probe that recognizes an enzymatically deiminated partial sequence of mouse keratin K1. *Journal of Dermatological Science*, 21(2), 113-126.
- Sharp, J. T., Strand, V., Leung, H., Hurley, F., & Loew-Friedrich, I. (2000). Treatment with leflunomide slows radiographic progression of rheumatoid arthritis: results from three randomized controlled trials of leflunomide in patients with active rheumatoid arthritis. Leflunomide Rheumatoid Arthritis Investigators Group. *Arthritis & Rheumatology*, 43(3), 495-505.

- Shirai, H., Blundell, T. L., & Mizuguchi, K. (2001). A novel superfamily of enzymes that catalyze the modification of guanidino groups. *Trends Biochem Sci*, 26(8), 465-468.
- Shoichet, B. K. (2004). Virtual screening of chemical libraries. *Nature*, 432(7019), 862-865.
- Silman, A. J., MacGregor, A. J., Thomson, W., Holligan, S., Carthy, D., Farhan, A. (1993). Twin concordance rates for rheumatoid arthritis: results from a nationwide study. *British Journal of Rheumatology*, 32(10), 903-907.
- Simmons, K. J., Chopra, I., & Fishwick, C. W. (2010). Structure-based discovery of antibacterial drugs. *Nature Reviews Microbiology*, 8(7), 501-510.
- Simms, R. W., Kwok, C. K., Anderson, L. G., Erlandson, D. M., Greene, J. M., Kelleher, M. (1996). Guidelines for monitoring drug therapy in rheumatoid arthritis: American College of Rheumatology Ad Hoc Committee on Clinical Guidelines. *Arthritis and Rheumatism*, 39(5), 723-731.
- Skriner, K., Sommergruber, W. H., Tremmel, V., Fischer, I., Barta, A., Smolen, J. S. (1997). Anti-A2/RA33 autoantibodies are directed to the RNA binding region of the A2 protein of the heterogeneous nuclear ribonucleoprotein complex. Differential epitope recognition in rheumatoid arthritis, systemic lupus erythematosus, and mixed connective tissue disease. *Journal of Clinical Investigation*, 100(1), 127-135.
- Slack, J. L., Causey, C. P., Luo, Y., & Thompson, P. R. (2011). Development and use of clickable activity based protein profiling agents for protein arginine deiminase 4. *ACS Chemical Biology*, 6(5), 466-476.
- Smith, L. J., Bolin, K. A., Schwalbe, H., MacArthur, M. W., Thornton, J. M., & Dobson, C. M. (1996). Analysis of main chain torsion angles in proteins: prediction of NMR coupling constants for native and random coil conformations. *Journal of Molecular Biology*, 255(3), 494-506.
- Smolen, J. S., Han, C., Bala, M., Maini, R. N., Kalden, J. R., van der Heijde, D. (2005). Evidence of radiographic benefit of treatment with infliximab plus methotrexate in rheumatoid arthritis patients who had no clinical improvement: a detailed subanalysis of data from the anti-tumor necrosis factor trial in rheumatoid arthritis with concomitant therapy study. *Arthritis & Rheumatology*, 52(4), 1020-1030.
- Smolen, J. S., Kalden, J. R., Scott, D. L., Rozman, B., Kvien, T. K., Larsen, A. (1999). Efficacy and safety of leflunomide compared with placebo and sulphasalazine in active rheumatoid arthritis: a double-blind, randomised, multicentre trial. European Leflunomide Study Group. *Lancet*, 353(9149), 259-266.
- Sreerama, N., Venyaminov, S. Y., & Woody, R. W. (1999). Estimation of the number of α -helical and β -strand segments in proteins using circular dichroism spectroscopy. *Protein Science*, 8(2), 370-380.

- Sreerama, N., & Woody, R. W. (1993). A self-consistent method for the analysis of protein secondary structure from circular dichroism. *Analytical Biochemistry*, 209(1), 32-44.
- Sreerama, N., & Woody, R. W. (2000). Estimation of Protein Secondary Structure from Circular Dichroism Spectra: Comparison of CONTIN, SELCON, and CDSSTR Methods with an Expanded Reference Set. *Analytical Biochemistry*, 287(2), 252-260.
- St. Clair, E. W., van der Heijde, D. M. F. M., Smolen, J. S., Maini, R. N., Bathon, J. M., Emery, P. (2004). Combination of infliximab and methotrexate therapy for early rheumatoid arthritis: A randomized, controlled trial. *Arthritis & Rheumatism*, 50(11), 3432-3443.
- Steen, J., Uhlén, M., Hober, S., & Ottosson, J. (2006). High-throughput protein purification using an automated set-up for high-yield affinity chromatography. *Protein Expression and Purification*, 46(2), 173-178.
- Steiner, G., & Smolen, J. (2002). Autoantibodies in rheumatoid arthritis and their clinical significance. *Arthritis Research*, 4 Suppl 2, S1-5.
- Stensland, M. E., Pollmann, S., Molberg, O., Sollid, L. M., & Fleckenstein, B. (2009). Primary sequence, together with other factors, influence peptide deimination by peptidylarginine deiminase-4. *Biological Chemistry*, 390(2), 99-107.
- Stevenson, C. L. (2009). Advances in peptide pharmaceuticals. *Current Pharmaceutical Biotechnology*, 10(1), 122-137.
- Stone, E. M., Schaller, T. H., Bianchi, H., Person, M. D., & Fast, W. (2005). Inactivation of two diverse enzymes in the amidinotransferase superfamily by 2-chloroacetamide: dimethylargininase and peptidylarginine deiminase†. *Biochemistry*, 44(42), 13744-13752.
- Storey, G. D. (2001). Alfred Baring Garrod (1819-1907). *Rheumatology*, 40(10), 1189-1190.
- Strand, V., Cohen, S., Schiff, M., Weaver, A., Fleischmann, R., Cannon, G. (1999). Treatment of active rheumatoid arthritis with leflunomide compared with placebo and methotrexate. Leflunomide Rheumatoid Arthritis Investigators Group. *Archives of Internal Medicine*, 159(21), 2542-2550.
- Studier, F. W. (2005). Protein production by auto-induction in high density shaking cultures. *Protein Expression and Purification*, 41(1), 207-234.
- Stupp, S., & Guler, M. (2005). Branched peptide amphiphiles, related epitope compounds and self assembled structures thereof: Google Patents.
- Subramaniam, S., Mehrotra, M., & Gupta, D. (2008). Virtual high throughput screening (vHTS)--a perspective. *Bioinformation*, 3(1), 14-17.

- Suzuki, A., Yamada, R., Chang, X., Tokuhira, S., Sawada, T., Suzuki, M. (2003). Functional haplotypes of PADI4, encoding citrullinating enzyme peptidylarginine deiminase 4, are associated with rheumatoid arthritis. *Nature Genetics*, 34(4), 395-402.
- Suzuki, A., Yamada, R., & Yamamoto, K. (2007). Citrullination by peptidylarginine deiminase in rheumatoid arthritis. *Annals of the New York Academy of Sciences*, 1108, 323-339.
- Takahara, H., Okamoto, H., & Sugawara, K. (1985). Specific modification of the functional arginine residue in soybean trypsin inhibitor (Kunitz) by peptidylarginine deiminase. *Journal of Biological Chemistry*, 260(14), 8378-8383.
- Takahara, H., Okamoto, H., & Sugawara, K. (1986). Affinity chromatography of peptidylarginine deiminase from rabbit skeletal muscle on a column of soybean trypsin inhibitor (Kunitz)-Sepharose. *Journal of Biochemistry*, 99(5), 1417-1424.
- Takizawa, Y., Suzuki, A., Sawada, T., Ohsaka, M., Inoue, T., Yamada, R. (2006). Citrullinated fibrinogen detected as a soluble citrullinated autoantigen in rheumatoid arthritis synovial fluids. *Annals of the Rheumatic Diseases*, 65(8), 1013-1020.
- Tanikawa, C., Espinosa, M., Suzuki, A., Masuda, K., Yamamoto, K., Tsuchiya, E. (2012). Regulation of histone modification and chromatin structure by the p53-PADI4 pathway. *Nature Communications*, 3, 676.
- Tarcsa, E., Marekov, L. N., Andreoli, J., Idler, W. W., Candi, E., Chung, S. I. (1997). The fate of trichohyalin. Sequential post-translational modifications by peptidyl-arginine deiminase and transglutaminases. *Journal of Biological Chemistry*, 272(44), 27893-27901.
- Tarcsa, E., Marekov, L. N., Mei, G., Melino, G., Lee, S. C., & Steinert, P. M. (1996). Protein unfolding by peptidylarginine deiminase. Substrate specificity and structural relationships of the natural substrates trichohyalin and filaggrin. *Journal of Biological Chemistry*, 271(48), 30709-30716.
- Taylor, P., Blackburn, E., Sheng, Y. G., Harding, S., Hsin, K. Y., Kan, D. (2008). Ligand discovery and virtual screening using the programme LIDAEUS. *British Journal of Pharmacology*, 153 Suppl 1, S55-67.
- Tejo, B. A., & Siahaan, T. J. (2009). Solution structure of a novel T-cell adhesion inhibitor derived from the fragment of ICAM-1 receptor: cyclo(1,8)-Cys-Pro-Arg-Gly-Gly-Ser-Val-Cys. *Biopolymers*, 91(8), 633-641.
- Tighe, H., & Carson, D. A. (1997). *Rheumatology* (5 ed.): Philadelphia, PA: WB Saunders.

- Tresadern, G., Bemporad, D., & Howe, T. (2009). A comparison of ligand based virtual screening methods and application to corticotropin releasing factor 1 receptor. *Journal of Molecular Graphics and Modelling*, 27(8), 860-870.
- Trott, O., & Olson, A. J. (2010). AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading. *Journal of Computational Chemistry*, 31(2), 455-461.
- Tsuchida, M., Takahara, H., Minami, N., Arai, T., Kobayashi, Y., Tsujimoto, H. (1993). cDNA nucleotide sequence and primary structure of mouse uterine peptidylarginine deiminase. Detection of a 3'-untranslated nucleotide sequence common to the mRNA of transiently expressed genes and rapid turnover of this enzyme's mRNA in the estrous cycle. *European Journal of Biochemistry*, 215(3), 677-685.
- Tugwell, P., Wells, G., Strand, V., Maetzel, A., Bombardier, C., Crawford, B. (2000). Clinical improvement as reflected in measures of function and health-related quality of life following treatment with leflunomide compared with methotrexate in patients with rheumatoid arthritis: sensitivity and relative efficiency to detect a treatment effect in a twelve-month, placebo-controlled trial. Leflunomide Rheumatoid Arthritis Investigators Group. *Arthritis & Rheumatology*, 43(3), 506-514.
- Ukkonen, K., Veijola, J., Vasala, A., & Neubauer, P. (2013). Effect of culture medium, host strain and oxygen transfer on recombinant Fab antibody fragment yield and leakage to medium in shaken E. coli cultures. *Microbial Cell Factories*, 12(1), 73.
- Union, A., Meheus, L., Humbel, R. L., Conrad, K., Steiner, G., Moereels, H. (2002). Identification of citrullinated rheumatoid arthritis-specific epitopes in natural filaggrin relevant for antifilaggrin autoantibody detection by line immunoassay. *Arthritis & Rheumatology*, 46(5), 1185-1195.
- Urano, Y., Watanabe, K., Sakaki, A., Arase, S., Watanabe, Y., Shigemi, F. (1990). Immunohistochemical demonstration of peptidylarginine deiminase in human sweat glands. *American Journal of dermatopathology*, 12(3), 249-255.
- van Boekel, M. A., Vossenaar, E. R., van den Hoogen, F. H., & van Venrooij, W. J. (2002). Autoantibody systems in rheumatoid arthritis: specificity, sensitivity and diagnostic value. *Arthritis Research*, 4(2), 87-93.
- van der Helm-van Mil, A. H., Wesoly, J. Z., & Huizinga, T. W. (2005). Understanding the genetic contribution to rheumatoid arthritis. *Current Opinion in Rheumatology*, 17(3), 299-304.
- van der Helm-van Mil, A. H. M., Verpoort, K. N., Breedveld, F. C., Huizinga, T. W. J., Toes, R. E. M., & de Vries, R. R. P. (2006). The HLA-DRB1 shared epitope alleles are primarily a risk factor for anti-cyclic citrullinated peptide antibodies and are not an independent risk factor for development of rheumatoid arthritis. *Arthritis & Rheumatism*, 54(4), 1117-1121.

- van Gaalen, F. A., van Aken, J., Huizinga, T. W., Schreuder, G. M., Breedveld, F. C., Zanelli, E. (2004). Association between HLA class II genes and autoantibodies to cyclic citrullinated peptides (CCPs) influences the severity of rheumatoid arthritis. *Arthritis & Rheumatology*, 50(7), 2113-2121.
- van Stokkum, I. H., Spoelder, H. J., Bloemendal, M., van Grondelle, R., & Groen, F. C. (1990). Estimation of protein secondary structure and error analysis from circular dichroism spectra. *Analytical biochemistry*, 191(1), 110-118.
- Varney, M. D., Marzoni, G. P., Palmer, C. L., Deal, J. G., Webber, S., Welsh, K. M. (1992). Crystal-structure-based design and synthesis of benz[cd]indole-containing inhibitors of thymidylate synthase. *Journal of Medicinal Chemistry*, 35(4), 663-676.
- Vlieghe, P., Lisowski, V., Martinez, J., & Khrestchatisky, M. (2010). Synthetic therapeutic peptides: science and market. *Drug Discovery Today*, 15(1-2), 40-56.
- von Itzstein, M., Wu, W.-Y., Kok, G. B., Pegg, M. S., Dyason, J. C., Jin, B. (1993). Rational design of potent sialidase-based inhibitors of influenza virus replication. *Nature*, 363(6428), 418-423.
- Vossenaar, E. R., Radstake, T. R. D., van der Heijden, A., van Mansum, M. A. M., Dieteren, C., de Rooij, D.-J. (2004a). Expression and activity of citrullinating peptidylarginine deiminase enzymes in monocytes and macrophages. *Annals of the Rheumatic Diseases*, 63(4), 373-381.
- Vossenaar, E. R., Smeets, T. J. M., Kraan, M. C., Raats, J. M., Van Venrooij, W. J., & Tak, P. P. (2004b). The presence of citrullinated proteins is not specific for rheumatoid synovial tissue. *Arthritis & Rheumatism*, 50(11), 3485-3494. D
- Vossenaar, E. R., Zendman, A. J., van Venrooij, W. J., & Pruijn, G. J. (2003). PAD, a growing family of citrullinating enzymes: genes, features and involvement in disease. *Bioessays*, 25(11), 1106-1118.
- Vranken, W. F., Boucher, W., Stevens, T. J., Fogh, R. H., Pajon, A., Llinas, M. (2005). The CCPN data model for NMR spectroscopy: development of a software pipeline. *Proteins*, 59(4), 687-696.
- Waler, E. (1940). On the occurrence of a factor in human serum activating the specific agglutination of sheep blood corpuscles. . *Acta Pathologica Microbiologica Scandinavica*, 17, 172.
- Wakimoto, T., Kondo, H., Nii, H., Kimura, K., Egami, Y., Oka, Y. (2011). Furan fatty acid as an anti-inflammatory component from the green-lipped mussel *Perna canaliculus*. *Proceedings of the National Academy of Sciences*, 108(42), 17533-17537.

- Wallace, A. C., Laskowski, R. A., & Thornton, J. M. (1995). LIGPLOT: a programme to generate schematic diagrams of protein-ligand interactions. *Protein Engineering*, 8(2), 127-134.
- Wang, Y., Li, M., Stadler, S., Correll, S., Li, P., Wang, D. (2009). Histone hypercitrullination mediates chromatin decondensation and neutrophil extracellular trap formation. *Journal of Cell Biology*, 184(2), 205-213.
- Wang, Y., Li, P., Wang, S., Hu, J., Chen, X. A., Wu, J. (2012). Anticancer peptidylarginine deiminase (PAD) inhibitors regulate the autophagy flux and the mammalian target of rapamycin complex 1 activity. *Journal of Biological Chemistry*, 287(31), 25941-25953.
- Wang, X., Wang, M., Tong, Y., Shan, L., & Wang, J. (2006). Probing the folding capacity and residual structures in 1-79 residues fragment of staphylococcal nuclease by biophysical and NMR methods. *Biochimie*, 88(10), 1343-1355.
- Watanabe, K., Akiyama, K., Hikichi, K., Ohtsuka, R., Okuyama, A., & Senshu, T. (1988). Combined biochemical and immunochemical comparison of peptidylarginine deiminases present in various tissues. *Biochimica et Biophysica Acta*, 966(3), 375-383.
- Wegner, N., Lundberg, K., Kinloch, A., Fisher, B., Malmstrom, V., Feldmann, M. (2010). Autoimmunity to specific citrullinated proteins gives the first clues to the etiology of rheumatoid arthritis. *Immunological Reviews*, 233(1), 34-54.
- Weinblatt, M. E., Kaplan, H., Germain, B. F., Block, S., Solomon, S. D., Merriman, R. C. (1994). Methotrexate in rheumatoid arthritis. A five-year prospective multicenter study. *Arthritis & Rheumatology*, 37(10), 1492-1498.
- Weinblatt, M. E., Kremer, J. M., Coblyn, J. S., Maier, A. L., Helfgott, S. M., Morrell, M. (1999). Pharmacokinetics, safety, and efficacy of combination treatment with methotrexate and leflunomide in patients with active rheumatoid arthritis. *Arthritis & Rheumatology*, 42(7), 1322-1328.
- Weinblatt, M. E., Maier, A. L., Fraser, P. A., & Coblyn, J. S. (1998). Longterm prospective study of methotrexate in rheumatoid arthritis: conclusion after 132 months of therapy. *Journal of Rheumatology*, 25(2), 238-242.
- Weinblatt, M. E., Polisson, R., Blotner, S. D., Sosman, J. L., Aliabadi, P., Baker, N. (1993). The effects of drug therapy on radiographic progression of rheumatoid arthritis. Results of a 36-week randomized trial comparing methotrexate and auranofin. *Arthritis & Rheumatology*, 36(5), 613-619.
- Westermann, J.-C., & Craik, D. J. (2008). NMR in peptide drug development. In L. Otvos (Ed.), *Peptide-Based Drug Design* (Vol. 494, pp. 87-113): Humana Press.

- Whitmore, L., & Wallace, B. A. (2004). DICHROWEB, an online server for protein secondary structure analyses from circular dichroism spectroscopic data. *Nucleic Acids Research*, 32(Web Server issue), W668-673.
- Whitmore, L., & Wallace, B. A. (2008). Protein secondary structure analyses from circular dichroism spectroscopy: methods and reference databases. *Biopolymers*, 89(5), 392-400.
- Wilson, W. L., Labra, C., & Barrist, E. (1961). Preliminary observations on the use of streptonigrin as an antitumor agent in human beings. *Antibiot Chemother*, 11, 147-150.
- Wishart, D. S., Sykes, B. D., & Richards, F. M. (1992). The chemical shift index: a fast and simple method for the assignment of protein secondary structure through NMR spectroscopy. *Biochemistry*, 31(6), 1647-1651.
- Wu, S. Y., McNae, I., Kontopidis, G., McClue, S. J., McInnes, C., Stewart, K. J. (2003). Discovery of a novel family of CDK inhibitors with the program LIDAEUS: structural basis for ligand-induced disordering of the activation loop. *Structure*, 11(4), 399-410.
- Wuthrich, K. (1986). *NMR of Protein and Nucleic Acids*. New York: Wiley Interscience.
- Yang, Y., Moir, E., Kontopidis, G., Taylor, P., Wear, M. A., Malone, K. (2007). Structure-based discovery of a family of synthetic cyclophilin inhibitors showing a cyclosporin-A phenotype in *Caenorhabditis elegans*. *Biochemical and Biophysical Research Communications*, 363(4), 1013-1019.
- Young, B. J., Mallya, R. K., Leslie, R. D., Clark, C. J., & Hamblin, T. J. (1979). Anti-keratin antibodies in rheumatoid arthritis. *British Medical Journal*, 2(6182), 97-99.
- Yu, X., Zhao, X., Zhu, L., Zou, C., Liu, X., Zhao, Z. (2013). Discovery of novel inhibitors for human farnesyltransferase (hFTase) via structure-based virtual screening. *Medicinal Chemistry Communications*, 4(6), 962-971.
- Zauhar, R. J., Moyna, G., Tian, L., Li, Z., & Welsh, W. J. (2003). Shape signatures: a new approach to computer-aided ligand- and receptor-based drug design. *Journal of Medicinal Chemistry*, 46(26), 5674-5690.
- Zeni, G., Lüdtke, D. S., Nogueira, C. W., Panatieri, R. B., Braga, A. L., Silveira, C. C. (2001). New acetylenic furan derivatives: synthesis and anti-inflammatory activity. *Tetrahedron Letters*, 42(51), 8927-8930.
- Zhang, X., Gamble, M. J., Stadler, S., Cherrington, B. D., Causey, C. P., Thompson, P. R. (2011). Genome-wide analysis reveals PADI4 cooperates with Elk-1 to activate c-Fos expression in breast cancer cells. *PLoS Genetics*, 7(6), e1002112.