



**UNIVERSITI PUTRA MALAYSIA**

***PHOTOLITHOGRAPHIC MODIFICATION OF POLYETHYLENE GLYCOL  
SILANE MONOLAYER FOR DEVELOPMENT OF LABEL-FREE DENGUE  
BIOSENSOR***

**NOR ZIDA BINTI ROSLY**

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BIOSENSOR**

By

**NOR ZIDA BINTI ROSLY**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfilment of the Requirements for the Degree of Master of Science**

**December 2015**

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

## **PHOTOLITHOGRAPHIC MODIFICATION OF POLYETHYLENE GLYCOL SILANE MONOLAYER FOR DEVELOPMENT OF LABEL-FREE DENGUE BIOSENSOR**

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**NOR ZIDA BINTI ROSLY**

**December 2015**

**Chairman : Shahrul Ainiah Alang Ahmad, PhD**  
**Faculty : Science**

Dengue disease becomes critical global health issue after millions were infected worldwide every year, which lead to fatality due to late detection. Hence, a new technique has been explored to improve the performance of the detection in term of time, price, sensitivity and selectivity. In order to develop the diagnostic devices, electron beam (e-beam) or ultra-violet (UV) lithography were used to fabricate small-sized patterns to provide surfaces with high sensitivity and selectivity to biomolecules. For this particular project, polyethylene glycol (PEG)-silane monolayer, which was the best-known antifouling polymer was used as a lithographic template due to its non-polar, non-toxic and non-immunogenic properties of the surface. The monolayer formed was thoroughly characterized with contact angle, atomic force microscopy (AFM), and X-ray photoelectron spectroscopy (XPS). The AFM and contact angle data showed a uniform surface and hydrophilic properties of PEG-silane monolayer obtained which confirmed by the XPS. The e-beam and UV irradiations were subsequently performed, which generated the aldehyde functional groups. These offers conjugation sites for the immobilization of DNA dengue. Further studies were done to verify the presence of aldehyde functionality by testing with reducing agent and 2-amino-1,1,1-trifluoroethane (TFEA). The TFEA test showed an increased value of contact angle from 42° to approximately 74°, suggesting the hydrophobicity of surface that corresponds to the bonding of carbon atom to three fluorine atoms was formed. The reduction test studied the reduction aldehyde to alcohols which resulted in the decrease of contact angle values. The properties of gold nanoparticles (AuNPs) on particles sizes, shape, morphology and elemental composition were investigated which was utilized as a colorimetric probe for the determination dengue-DNA. For the DNA hybridization, target DNA was attached directly to probe DNA that had been immobilized on irradiated modified surfaces in which an amine-terminus (or N-terminus) from the DNA bound with aldehyde on modified surfaces. Gold enhancement process was introduced for naked eye detection after inducing electrostatic interaction between positively charged AuNPs and negatively charged target DNA to probe DNA. Control experiments were performed with mismatch DNA sequences to confirm the selectivity of the sensor. The effect of target DNA concentration was studied in the hybridization of DNA. The results obtained indicate an efficient and selective device have been successfully developed for label-free dengue detection.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

## **PENGUBAHSUAIAN FOTOLITOGRAFI POLIETILINA GLIKOL SILANA EKALAPIS UNTUK PEMBANGUNAN LABEL BEBAS DENGGI BIOSENSOR**

Oleh

**NOR ZIDA BINTI ROSLY**

**Disember 2015**

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Penyakit denggi menjadi isu kesihatan global yang kritikal selepas berjuta-juta manusia dijangkiti di seluruh dunia setiap tahun, yang membawa kepada kematian disebabkan pengesanan yang lewat. Oleh itu, satu teknik baru telah diterokai untuk meningkatkan prestasi pengesanan dari segi masa, harga, kepekaan dan pemilihan. Dalam usaha untuk membangunkan diagnostik yang sebegini, alur elektron (e-beam) dan litografi sinar ultra-lembayung (UV) telah digunakan untuk fabrikasi corak bersaiz kecil yang mampu menghasilkan permukaan dengan sensitiviti yang tinggi dan pengecaman biomolekul tertentu. Dalam kajian ini khususnya, polietilena glikol (PEG)-silana ekalapis, yang merupakan polimer antifouling yang paling terkenal digunakan sebagai templat lithografi kerana sifatnya yang tidak berkutub, tidak beracun dan tidak imunogenik. Ekalapis yang terbentuk dengan sempurna dicirikan dengan ukuran sudut sentuh, mikroskopi daya atom (AFM), dan spektroskopi fotoelektron sinar-X (XPS). Data daripada AFM dan sudut sentuh menunjukkan permukaan ekalapis yang seragam dan hidrofilik ekalapisan PEG-silana diperolehi dan ini disahkan oleh XPS. E-beam dan penyinaran UV kemudiannya dilaksanakan, telah menjana kumpulan berfungsi aldehid. Kumpulan ini menyediakan laman konjugasi untuk imobilisasi denggi DNA. Selanjutnya ujian telah dilakukan untuk mengesahkan kehadiran kumpulan berfungsi aldehid dengan 2-amino-1,1,1-trifluoroethane (TFEA) dan ujian penurunan. Ujian TFEA menunjukkan terdapat peningkatan sudut sentuh daripada 42 ° kepada kira-kira 74 °, mencadangkan kehidrofobikan permukaan yang sepadan dengan ikatan atom karbon yang mempunyai tiga atom fluorin. Ujian penurunan dikaji menunjukkan aldehid menurun kepada alkohol yang menghasilkan penurunan bacaan sudut sentuh. Sifat-sifat nanopartikel emas (AuNPs) ke atas saiz zarah, bentuk, morfologi dan komposisi unsur telah dikaji yang digunakan sebagai kajian kolorimetrik bagi menentukan denggi DNA. Untuk penghibridan DNA, sasaran DNA telah diikat langsung dengan prob DNA yang telah diimobilisasi pada permukaan radiasi diubahsuai di mana amina-terminal (atau N-terminal) daripada DNA terikat dengan aldehid pada permukaan yang diubahsuai. Proses peningkatan emas telah diperkenalkan untuk pemerhatian secara mata kasar selepas menjalankan interaksi elektrostatik antara AuNPs bercas positif dan DNA bercas negatif terhadap DNA prob. Eksperimen kawalan telah dijalankan dengan urutan DNA tidak sepadan untuk mengesahkan pemilihan sensor. Kesan kepekatan sasaran DNA telah dikaji dalam penghibridan DNA. Keputusan yang diperolehi menunjukkan kejayaan peranti yang cekap dan selektif dibangunkan pada label bebas denggi diagnostik.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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- the research conducted and the writing of this thesis was under our supervision;
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## LIST OF ABBREVIATIONS

$\mu$ CP	Microcontact printing
AFM	Atomic force microscopy
Au	Gold
AuNP	Gold nanoparticles
DF	Dengue fever
DHF	Dengue hemorrhagic fever
DNA	Deoxyribonucleic acid
DPN	Dip-pen nanolithography
DSS	Dengue shock syndrome
e-beam	Electron beam
FE-SEM	Field emission scanning electron microscope
FTIR	Fourier transform infrared spectroscopy
OEG	Oligo ethylene glycol
PDMS	Polydimethylsiloxane
PEG	Poly(ethylene glycol)
PEG-silane	2-[Methoxy(polyethyleneoxy)propyl] trimethoxysilane
RNA	Ribonucleic acid
RT-PCR	Reverse-transcription-polymerase chain reaction
SAMs	Self-assembled monolayers
SiO <sub>2</sub>	Silicon dioxide
TEM	Transmission electron microscopy
UV	Ultraviolet



## CHAPTER 1

### INTRODUCTION

#### 1.1 Background

Biosensor technologies have been well developed due to demands of devices with high sensitivity, fast detection, small and easy to use. Sensing of biomolecule (antibodies, enzymes, nucleic acids) and biological systems (receptors, cells) can be performed through transducer (e.g optical, electrochemical, mass sensitive devices) (Wang, 2000). The chemical and physical transducers translate the event into important electrical signal such as light, current and frequency. There are two types of biosensor; classified by the nature of recognition event. The first type is bioaffinity devices which use a surface confined ligand partner (e. g antibody, oligonucleotide) for the attachment of target analyte. Otherwise, biocatalytic devices based on enzyme for recognizing the target substrate (Junhui *et al.*, 1997).

Biosensors apply self-assembled monolayers (SAMs) as interfaces between solid surfaces and biomolecules. Self-assembled monolayers (SAMs) are formed when amphiphilic surfactant molecules are adsorbed spontaneously onto solid surface of interest by specific interaction of surface chemistry. The adsorption of molecules on substrate can be performed in variety of solvents (e.g polar or non polar) which allow a greater flexibility in molecular design, therefore, providing surfaces properties that can be modified and controlled. Monolayers have been witnessed as successful model surfaces in biosensor's research since homogenous on thin films can be formed.

Over the past few decades, a lot of scientific studies have witnessed dramatic changes for detection of deoxyribonucleic acid (DNA) sequence, genomic analyses and early diagnosis for critical diseases. At that time, DNA hybridization biosensor was introduced by recognition of nucleic acid on transducer surfaces towards the goal for a rapid, simple and inexpensive devices (Junhui *et al.*, 1997). DNA biosensor can be fabricated using various strategies including electrical, optical and mechanical.

Optical strategy especially fluorescent method shows a great potential due to high sensitivity and easy accessibility to commercialized equipment (Csa *et al.*, 2000). However, this method requires expensive equipment and complex steps for label analytes (fluorophores) which are not suitable for shorter detection time. Therefore, label free naked eye detection is developed as an alternative to fluorescence technique. The detection is significantly simplified by simply using by naked eye or with a flat-scanner (so called scanometric detection). The technique used metal enhancement as a signal amplification process to detect conjugation between AuNPs and DNAs. The method shows a great potential to be developed since the analysis used instrument (e.g flat scanner) which is cost-effective and enhancement process takes a few minutes to operate.

## 1.2 Dengue Virus

Dengue diagnostic devices has been rapidly developed in order to meet the requirement of worldwide demand due to the increase of death in tropical and sub-tropical regions, mostly in urban and semi-urban countries, leading to approximately 50-100 million infections per year. The infection occurs when the four serotypes (DEN-1, DEN-2, DEN-3, DEN-4) transmitted in human body by *Aedes aegypti* mosquito, members of the Flavivirus family. In recent years, number of cases for Dengue fever (DF), Dengue hemorrhagic fever (DHF) and Dengue shock syndrome (DSS) are getting increased among children in some Asian and Latin American country (Weaver & Reisen, 2010).

So far, infection with dengue virus has no apparent symptoms or clinical signs since it can be confused with other vector borne viral and parasitic diseases such as influenza, chikungunya, malaria and zika viruses (Zhang *et al.*, 2010). There is no available vaccine and therapy for the treatment of dengue infection, to date, the early prevention is the only way to control the disease caused by dengue virus. Hence, laboratory diagnoses are required to identify the disease rapidly and reliably, thus treat the disease at early stage of infection.

Currently, the techniques used for diagnosis of dengue infections are through virological detection and serological test. Virological test allows the identification of specific viral molecules. Meanwhile, serological test is to identify specific dengue antibodies such as IgG and IgM. However, the sufficient amounts of antibodies are not always presence in the body at early stage of infection and there are available at least 5 days after onset of illness. Additionally, the test should be confirmed with two or more serum sample since it has similarity with other Flaviviruses.

Recently, a new technique, reverse transcription polymerase chain reaction (RT-PCR) was created with high sensitivity, low risk of contamination and rapid real time assays as compared to serological test. The technique used agarose gel electrophoresis to detect dengue viral RNA. However, the process is time-consuming and hazardous. The summary of the available techniques are shown in Table 1.

**Table 1: Advantages and limitations of current dengue diagnostics (Peeling *et al.*, 2010)**

Diagnostic tests	Advantages	Limitations
Viral isolation	<ul style="list-style-type: none"> <li>- confirmed infection</li> <li>- specific</li> <li>- serotypes detection</li> </ul>	<ul style="list-style-type: none"> <li>- needs acute sample (0-5 days post onset)</li> <li>- needs expertise and appropriate equipments</li> <li>- takes more than 1 week</li> <li>- does not differentiate between primary and secondary infection</li> <li>- expensive</li> </ul>
RNA detection	<ul style="list-style-type: none"> <li>- confirmed infection</li> <li>- sensitive and specific</li> <li>- serotypes and genotypes detection</li> <li>- results in 24-48 hours</li> </ul>	<ul style="list-style-type: none"> <li>- potential false-positives owing to contamination</li> <li>- needs acute sample (0-5 days post onset)</li> <li>- needs expertise and positive laboratory equipment</li> <li>- does not differentiate between primary and secondary infection</li> </ul>
Antigen detection		
Clinical specimens (for example, using blood in an NS1 assay)	<ul style="list-style-type: none"> <li>- confirmed infection</li> <li>- easy to perform</li> <li>- less expensive than virus isolation and RNA detection</li> </ul>	<ul style="list-style-type: none"> <li>- not sensitive as virus isolation or RNA detection</li> </ul>
Tissues from fatal cases ( for example in immunohistochemistry)	<ul style="list-style-type: none"> <li>- confirmed infection</li> </ul>	<ul style="list-style-type: none"> <li>- not sensitive as virus isolation or RNA detection</li> </ul>
Serological tests		
IgM or IgG seroconversion	<ul style="list-style-type: none"> <li>- confirmed infection</li> <li>- least expensive</li> <li>- easy to perform</li> </ul>	<ul style="list-style-type: none"> <li>- IgM levels can be low in secondary infections</li> <li>- confirmation needs two or more serum samples</li> </ul>
IgM detection (single sample)	<ul style="list-style-type: none"> <li>- probable Dengue cases detection</li> <li>- useful for surveillance, tracking outbreaks and monitoring effectiveness of interventions</li> </ul>	<ul style="list-style-type: none"> <li>- IgM levels can be low in second infections</li> </ul>

### 1.3 Surface lithography

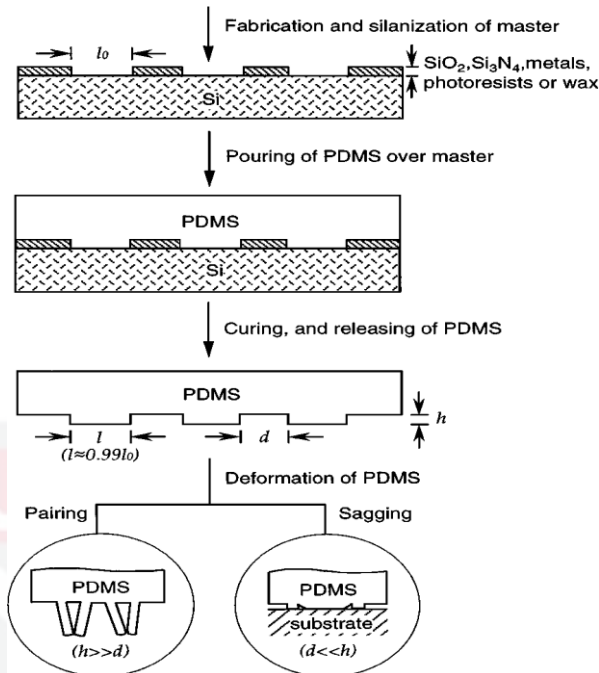
Lithography has been widely used as it offers potential applications in a wide variety of fields, including biosensor, biomedical, bioelectronics and fundamental studies of cell biology. It enables the creation of patterns down to smaller (micrometer to nanometer) scale precision, which provides surfaces with high sensitivity and specific biomolecular recognition. Patterning is important for modification of surfaces and it is useful to build functional nanostructures completely or partially on the exposed surfaces (Smith *et al.*, 2004).

Lithography is performed to functionalize surfaces either by removing particular adsorbates (Mulder *et al.*, 2001), placement of adsorbates (Buoninsegni *et al.*, 1998), or by the particular reaction of adsorbates (Nan Li & Ho, 2008). Once the adsorbates are placed on sub-monolayer regions, the remaining surface with the exposed area can interact with a new adsorbate. It creates adsorbates with various groups on the same adsorbent.

Currently, there are many techniques that exist to create patterns onto chip substrate such as contact lithography (dip pen lithography (DPN), atomic force microscope (AFM), microcontact printing ( $\mu$ CP), scanning-near field photolithography, soft lithography, electron beam (e-beam) lithography (Agarwal *et al.*, 2003; Demers *et al.*, 2002; Mendes *et al.*, 2004; Whitesides *et al.*, 2001; Zhang *et al.*, 2004). Most of the techniques used both physical adsorption and covalent reaction.

#### Microcontact printing ( $\mu$ CP)

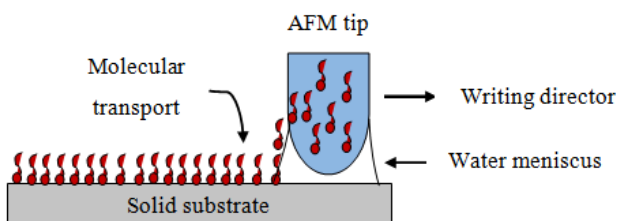
Microcontact printing ( $\mu$ CP) has proven a valuable technique for patterning DNA (Lange *et al.*, 2004), proteins and cells (Turner & Shain, 1998). The technique was pioneered by Whitesides and the group where adsorbates were transferred to a surface by using flexible, polymeric stamps with pattern reliefs (usually made from polydimethylsiloxane, PDMS) (Whitesides *et al.*, 2001). The stamps are dipped in ink (alkanethiol) and dried before brought to contact with a surface like gold. However, one major limitation of such  $\mu$ CP based approaches is the uncontrollable contamination of the stamp due to changes in environment condition (humidity, pH) during the biomolecules transferring process (Tan *et al.*, 2004).



**Figure 1: Microcontact printing by PDMS stamp on silicon surfaces (Whitesides *et al.*, 2001)**

### Dip-pen nanolithography (DPN)

Dip-pen nanolithography (DPN) is a technique that performed under ambient condition without using any large electron magnetic field and shear effect. The technique was introduced by Piner and co-workers as new scanning probe lithography for fabricating thiol molecules on Au surfaces (Piner *et al.*, 1999). The technique used AFM tip where was first coated with chemical reagents by immersing the cantilever in a solution or by evaporation. The coated tip was then transporting chemical reagents to nanoscopic regions on substrate of interest in one step, which is an advantage over serial techniques. Although the technique is simple but it is much more expensive due to powerful instrument for transporting chemical reagents from AFM tip to substrate.

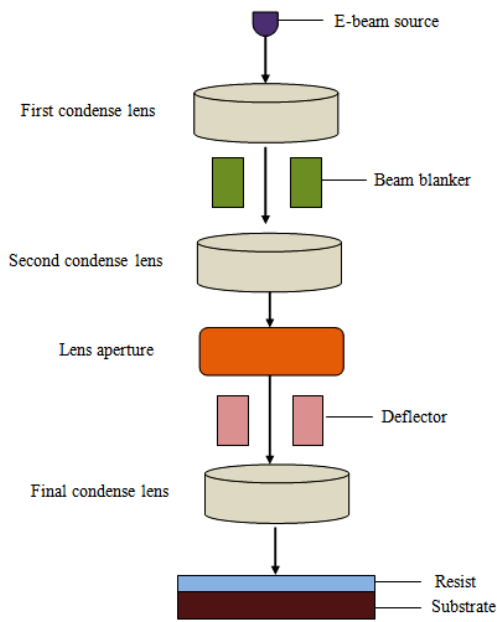


**Figure 2: Schematic illustration of dip-pen nanolithography (DPN) process**

### **Electron beam (e-beam) lithography**

Electron beam (e-beam) lithography has been exploited as a maskless technique for patterning biomolecules with featured sizes ranging from several micrometers to nanometer, in addition to the possibility of designing the arbitrary shapes on the surfaces. In general, the patterning is performed by first coating the substrate with a polymeric thin film or resist and then exposing it to the e-beam (Glezos *et al.*, 2002).

The high energy electrons present in the e-beam causes the changes in the chemical bonding by cross-linking or degrading the elements available at the surface. The exposed substrate is then rinsed with a developer solution to remove the unattached soluble portion of the resist. The technique is likely preferred for fabricating surfaces since it gives high resolution small scale patterns (Kolodziej & Maynard, 2012). Recently, the used of scanning electron microscope (SEM) equipped with the technique enable to scan electron beam spot within desired area, thus generate nanoscale pattern.



**Figure 3: Schematic illustration of electron beam (e-beam) lithography process**

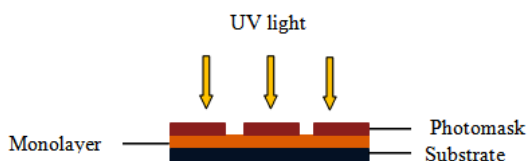
### Photolithography

Photolithography is another option for fabrication of monolayer surfaces. It has been employed to create a three-dimensional topography of resist layer and can provide chemical attachment on modified surfaces. The conventional technique utilizes an exposure of photo-resist layer to ultraviolet (UV) through photomask with opaque features (e.g. Cu grid). Commonly, UV light with wavelength 193-436 nm is performed in order to affect the terminal end group at the designated regions of substrate. The technique becomes attractive due to high resolution (sub-micrometer line width) and excellent control of feature dimensions. It also has a big potential to be used in industrial exposure tools due to the high quality and high resolution pattern such as deep UV ( $\lambda < \sim 250$  nm) (Dulcey *et al.*, 1991).

The principle of this method is by irradiate the top of monolayer by UV light through the photomask which ables to create pattern on surfaces. It has been demonstrated on various materials including alkylthiolates (Montague *et al.*, 2007) and phosphonic acids as well as films of Au (Tizazu *et al.*, 2009), polymers (Hurley *et al.*, 2010) and metallic nanoparticles (Sun *et al.*, 2006). Light plays an important tool for surface immobilization due to big success in attachment of biomolecules (Blawas & Reichert, 1998; Yang *et al.*, 2006). The main purpose for irradiating monolayer under UV light is to generate functional groups such as aldehyde and carboxylic acid that available for immobilization of biomolecules.



The UV light have ability to shorten the alkyl chain and dissociate the oxygen molecules and create active oxygen species such as oxygen free radical thus oxidize the alkyl chain ( Xue & Yang, 2010). Therefore, UV lithography method is well established technique and can be used repeatedly on the same substrate with different protein to create multiple protein patterns (Lee *et al.*, 2003).



**Figure 4: Schematic illustration of photolithography process**

#### 1.4 Problem statements

Dengue is a serious tropical disease that causes death to human if it is not treated promptly. Therefore, many strategies have been developed to detect dengue for the requirement of rapid detection, high sensitivity and selectivity, inexpensive and portable devices. However, the challenge dealing with dengue viruses is the unstable structure of virus and the differences among the four serotypes (DEN-1, DEN-2, DEN-3, DEN-4), causing the dengue viruses can not be controlled. Moreover, the existing of conventional techniques for dengue detection is focusing on specific anti-Dengue antibodies which are developed in response to Dengue viruses. Unfortunately, these antibodies produce at least 5 days after the onset of illness, whose the process is time-consuming for early detection (Vijayakumar *et al.*, 2005). The advancement today's technology provides new opportunities for improving the performance of biosensors in term of time, price, sensitivity and selectivity which enable it to be used as portable device.

Optical biosensors are powerful technological developments that have a great potential for the direct, real-time and label-free detection of biomolecules especially nucleic acid. Optical biosensors are high specification, sensitivity, inexpensive, rapid and easy for users. The development of such sensors should be given high attention to improve the bio-recognition where the researchers require developing a new technique that can give a well respond to the selective biomolecules element such as nucleic acid without any interference from non-specific proteins element.

#### 1.5 Objective of the study

The main objective of this study is to develop naked eye detection of DNA-dengue biosensor on modified 2-[Methoxy(polyethyleneoxy)propyl] trimethoxysilane (PEG-silane) monolayer. The detection facility incorporates the use of AuNPs to form Au/DNA



composites. The following specific objectives are designed to achieve the main objective:

- i. To prepare and characterize PEG-silane monolayer on silicon oxide or glass surfaces.
- ii. To identify the reactive functional groups formed and hence the surface chemistry by exposing e-beam or UV onto SAMs surfaces
- iii. To immobilize and hybridize dengue DNA on photo-modified surfaces.
- iv. To make the targeted spot visible by Au enhancement process.
- v. To identify the response of various target base length and various target DNA concentrations in hybridization process by naked eye detection.





## REFERENCES

- Adden, N., Gamble, L. J., Castner, D. G., Hoffmann, A., Gross, G., & Forschung, B. (2006). Phosphonic Acid Monolayers for Binding of Bioactive Molecules to Titanium Surfaces, *Langmuir*, 22 (19), 8197–8204.
- Agarwal, G., Naik, R. R., Stone, M. O., Base, W. A. F., & Management, T. (2003). Immobilization of Histidine-Tagged Proteins on Nickel by Electrochemical Dip Pen Nanolithography, *Journal of the American Chemical Society*, 125 (14), 7408–7412.
- Ahmad, S. A., Hucknall, A., Chilkoti, A., & Leggett, G. J. (2010). Protein Patterning by UV-Induced Photodegradation of Poly ( oligo ( ethylene glycol ) methacrylate ) Brushes, *Langmuir : The ACS Journal of Surfaces and Colloids*, 26 (13), 9937–9942.
- Ahner, T. T., Delissen, F., & Sokolov, S. (2010). Mechanism of Gold Nanoparticle Formation in the Classical Citrate Synthesis Method Derived from Coupled In Situ XANES and SAXS Evaluation. *Journal of the American Chemical Society*, 132 (4), 1296–1301.
- Alcantar, N. A., Aydil, E. S., & Israelachvili, J. N. (2000). Polyethylene Glycol-coated Biocompatible Surfaces. *Journal of Biomedical Materials Research*, 51(3), 343–351.
- Alexandre, I., Hamels, S., Dufour, S., Collet, J., Zammateo, N., De Longueville, F., Gala, J. –L., Remacle, J. (2001). Colorimetric Silver Detection of DNA Microarrays. *Analytical Biochemistry*, 295, 1-8
- Alonso, J. M., Reichel, A., Piehler, J., & del Campo, A. (2008). Photopatterned Surfaces for Site-specific and Functional Immobilization of Proteins. *Langmuir : The ACS Journal of Surfaces and Colloids*, 24(2), 448–57.
- Ariza, M. J., Rodríguez-Castellón, E., Rico, R., Benavente, J., Muñoz, M., & Oleinikova, M. (2000). X-Ray Photoelectron Spectroscopy Analysis of Di-(2-ethylhexyl) Phosphoric Acid Activated Membranes. *Journal of Colloid and Interface Science*, 226(1), 151–158.
- Bai, L., Tan, L., Chen, L., Liu, S., & Wang, Y. (2014). Preparation and Characterizations of Poly(2-methyl-2-oxazoline) Based Antifouling Coating by Thermally Induced Immobilization. *Journal of Material Chemistry B*, 2, 7785–7794.
- Baker, M. V., & Watling, J. D. (1997). Functionalization of Alkylsiloxane Monolayers via Free-Radical Bromination. *Langmuir*, 13, 2027–2032.

- Bishop, L. M., Yeager, J. C., Chen, X., Wheeler, J. N., Torelli, M. D., Benson, M. C., & Hamers, R. J. (2012). A Citric Acid-derived Ligand for Modular Functionalization of Metal Oxide Surfaces Via “click” Chemistry. *Journal of Surfaces and Colloids*, 28 (2), 1322–1329.
- Bjorling, M., Karlstriim, G., & Linse, P. (1991). Conformational Adaption of Poly(ethylene oxide). *Journal of the American Chemical Society*, 95 (17), 6706–6709.
- Blawas, A. S., & Reichert, W. M. (1998). Protein Patterning. *Biomaterials*, 19, 595–609.
- Brough, B., Christman, K. L., Wong, T. S., Kolodziej, C. M., Forbes, J. G., Wang, K., Maynard, H. D., Ho, C.-M. (2007). Surface Initiated Actin Polymerization from Top-down Manufactured Nanopatterns. *Soft Matter*, 3, 541.
- Brown, W. H. (2000). Introduction to Organic Chemistry (Second Edition), *Aldehydes and Ketones* (pp. 324-325), A Division of Harcourt College Publishers.
- Buoninsegni, F. T., Herrero, R., & Moncelli, M. R. (1998). Alkanethiol Monolayers and Alkanethiol Phospholipid Bilayers Supported by Mercury: An Electrochemical Characterization. *Journal of Electroanalytical Chemistry*, 452, 33–42.
- Cao, Y. C., Jin, R., Thaxton, C. S., & Mirkin, C. A. (2005). A Two-color-change, Nanoparticle-based Method for DNA Detection. *Talanta*, 67, 449–455.
- Cecchet, F., De Meersman, B., Demoustier-Champagne, S., Nysten, B., & Jonas, A. M. (2006). One Step Growth of Protein Antifouling Surfaces: Monolayers of Poly(ethylene oxide) (PEO) Derivatives on Oxidized and Hydrogen-passivated Silicon Surfaces. *Langmuir: The ACS Journal of Surfaces and Colloids*, 22(3), 1173–1181.
- Chaki, N. K., & Vijayamohan, K. (2002). Self-assembled Monolayers as a Tunable Platform for Biosensor Applications. *Biosensors & Bioelectronics*, 17, 1–12.
- Chen, M.-S., Dulcey, C. S., Chrisey, L. A., & Dressick, W. J. (2006). Deep-UV Photochemistry and Patterning of (Aminoethylaminomethyl)phenethylsiloxane Self-Assembled Monolayers. *Advanced Functional Materials*, 16, 774–783.
- Chupa, J. A., Xu, S., Fischetti, R. F., Strongin, R. M., Mccauley, J. P., Smith, A. B., & Blasie, J. K. (1993). A Monolayer of C60 Tethered to The Surface of an Inorganic Substrate: Assembly and Structure, *Journal of the American Chemical Society*, 115, 4383–4384.
- Cras, J. J., Rowe-Taitt, C. A., Nivens, D. A., & Ligler, F. S. (1999). Comparison of Chemical Cleaning Methods of Glass in Preparation for Silanization. *Biosensors and Bioelectronics*, 14, 683–688.

- Crowe, J. A., Genzer, J., & Carolina, N. (2005). Creating Responsive Surfaces with Tailored Wettability Switching Kinetics and Reconstruction Reversibility. *Journal of the American Chemical Society*, 127, 17610–17611.
- Csa, A., Ko, J. M., & Fritzsche, W. (2000). Chip-Based Optical Detection of DNA Hybridization by Means of Nanobead Labeling. *Analytical Chemistry*, 72(24), 6025–6029.
- Demers, L. M., Ginger, D. S., Park, S.-J., Li, Z., Chung, S.-W., & Mirkin, C. A. (2002). Direct Patterning of Modified Oligonucleotides on Metals and Insulators by Dip-pen Nanolithography. *Science*, 296, 1836–1838.
- Devaraj, N. K., & Collman, J. P. (2007). Copper Catalyzed Azide-Alkyne Cycloadditions on Solid Surfaces: Applications and Future Directions. *QSAR & Combinatorial Science*, 26(11-12), 1253–1260.
- Dulcey, C. S., Georger, J. H., Krauthamer, V., Stenger, D. A, Fare, T. L., & Calvert, J. M. (1991). Deep UV Photochemistry of Chemisorbed Monolayers: Patterned Coplanar Molecular Assemblies. *Science*, 252, 551–554.
- Glezos, N., Misiakos, K., Kakabakos, S., Petrou, P., & Terzoudi, G. (2002). Electron Beam Patterning of Biomolecules. *Biosensors & Bioelectronics*, 17, 279–282.
- Graham, R. L., Colin, D. B., Biebuyck, H. A., Laibinis, P. E., & Whitesides, G. M. (1993). Damage to CF<sub>3</sub>CONH-Terminated Organic Self-Assembled Monolayers (SAMs) on Al, Ti, Cu, and Au by Al K $\alpha$  X-rays Is Due Principally to Electrons. *The Journal of Physical Chemistry*, 97(37), 9456–9464.
- Grandbois, M., Beyer, M., Rief, M., Clausen-Schaumann, H., & Gaub, H. (1999). How Strong Is a Covalent Bond? *Science*, 283, 1727–1730.
- Haensch, C., Ott, C., Hoepfner, S., & Schubert, U. S. (2008). Combination of Different Chemical Surface Reactions for the Fabrication of Chemically Versatile Building Blocks onto Silicon Surfaces. *Langmuir*, 24, 10222–10227.
- Harnett, C. K., Satyalakshmi, K. M., & Craighead, H. G. (2000). Low-energy Electron-beam Patterning of Amine-functionalized Self-Assembled Monolayers. *Applied Physics Letters*, 76(17), 2466.
- Heise, A., Stamm, M., Rauscher, M., Duschner, H., & Menzel, H. (1998). Mixed Silane Self Assembled Monolayers and Their In Situ Modification. *Thin Solid Films*, 29, 199–203.
- Herzer, N., Hoepfner, S., & Schubert, U. S. (2010). Fabrication of Patterned Silane Based Self-assembled Monolayers by Photolithography and Surface Reactions on Silicon-oxide Substrates. *Chemical Communications*, 46, 5634–5652.

- Hurley, C. R., Ducker, R. E., Leggett, G. J., & Ratner, B. D. (2010). Fabrication of Submicrometer Biomolecular Patterns by Near-field Exposure of Plasma-polymerized Tetraglyme Films. *Langmuir*, 26(12), 10203–10209.
- Ide, M. S., & Davis, R. J. (2014). The Important Role of Hydroxyl on Oxidation Catalysis by Gold Nanoparticles. *American Chemical Society*, 47(3), 825–833
- Junhui, Z., Hong, C. U. I., & Ruifu, Y. (1997). DNA Based Biosensors. *Biotechnology Advances*, 15(1), 43–58.
- Kasthuri, J., Veerapandian, S., & Rajendiran, N. (2009). Biological Synthesis of Silver and Gold Nanoparticles Using Apiin as Reducing Agent. *Colloids and Surfaces. B, Biointerfaces*, 68, 55–60.
- Kim, S. K., Cho, H., Jeong, J., Kwon, J. N., Jung, Y., & Chung, B. H. (2010). Label-free and Naked Eye Detection of PNA/DNA Hybridization using Enhancement of Gold Nanoparticles. *Chemical Communications (Cambridge, England)*, 46, 3315–3317.
- Kirby, B. J., & Jr, E. F. H. (2004). Review Zeta Potential of Microfluidic Substrates : 1 . Theory , Experimental Techniques , and Effects on Separations. *Electrophoresis*, 25 , 187–202.
- Kolodziej, C. M., & Maynard, H. D. (2012). Electron-Beam Lithography for Patterning Biomolecules at the Micron and Nanometer Scale. *Chemistry of Materials*, 24, 774–780.
- Konradi, R., Pidhatika, B., Mühlebach, A., & Textor, M. (2008). Poly-2-methyl-2-oxazoline: a Peptide-like polymer for Protein-repellent Surfaces. *Langmuir*, 24, 613–616.
- Kumar, A., Biebuyck, H. A., & Whitesides, G. M. (1994). Patterning Self -Assembled Monolayers: Applications in Materials Science, *Langmuir*, 10(5), 1498–1511.
- Labadie, G. R., & Poulter, C. D. (2006). Regio- and Chemoselective Covalent Immobilization of Proteins through Unnatural Amino Acids. *Journal of the American Chemical Society*, 128, 9274–9275.
- Lan, S., Veiseh, M., & Zhang, M. (2005). Surface Modification of Silicon and Gold-patterned Silicon Surfaces for Improved Biocompatibility and Cell Patterning Selectivity. *Biosensors & Bioelectronics*, 20, 1697–1708.
- Lange, S. A., Benes, V., Kern, D. P., Ho, J. K. H., & Morgenstelle, A. Der. (2004). Microcontact Printing of DNA Molecules. *Analytical Chemistry*, 76(6), 1641–1647.

- Lee, C.-S., Lee, S.-H., Park, S.-S., Kim, Y.-K., & Kim, B.-G. (2003). Protein Patterning on Silicon-based Surface Using Background Hydrophobic Thin Film. *Biosensors & Bioelectronics*, 18, 437–444.
- Li, N., & Ho, C. (2008). Patterning Functional Proteins with High Selectivity for Biosensor Applications. *Journal of the Association for Laboratory Automation*, 13, 237–242.
- Li, N., & Ho, C.-M. (2008). Photolithographic Patterning of Organosilane Monolayer for Generating Large Area Two-dimensional B Lymphocyte Arrays. *Lab on a Chip*, 8, 2105–2112.
- Ligler, F. S., & Washington, F. (1992). New Approach To Producing Patterned Biomolecular Assemblies. *Journal of the American Chemical Society*, 114, 4432–4433.
- Liu, H., & Bhushan, B. (2003). Nanotribological Characterization of Molecularly Thick Lubricant Films for Applications to MEMS/NEMS by AFM. *Ultramicroscopy*, 97, 321–40.
- Lofas, S., & Johnsson, B. (1990). A Novel Hydrogel Matrix on Gold Surfaces in Surface Plasmon Resonance Sensors for Fast and Efficient Covalent Immobilization of Ligands. *Journal of the Chemical Society*, 1526–1528.
- Lou, S., Ye, J., Li, K., & Wu, A. (2012). A Gold Nanoparticle-based Immunochromatographic Assay: the Influence of Nanoparticulate Size. *The Analyst*, 137, 1174–1181.
- Mahanta, C., & Bora, U. (2012). The Synthesis of Gold Nanoparticles Using Amaranthus Spinous Leaf Extract and Study of Their Optical. *Advances in Materials Physics and Chemistry*, 2, 275–281.
- Malikova, N., Pastoriza-santos, I., Schierhorn, M., Kotov, N. A., & Liz-marza, L. M. (2002). Layer-by-Layer Assembled Mixed Spherical and Planar Gold Nanoparticles : Control of Interparticle Interactions. *Langmuir*, 18, 3694–3697.
- Martínez, J. C., A. Chequer, N., L. González, J., & Cordova, T. (2013). Alternative Metodology for Gold Nanoparticles Diameter Characterization Using PCA Technique and UV-VIS Spectrophotometry. *Nanoscience and Nanotechnology*, 2(6), 184–189.
- Mendes, P. M., Jacke, S., Critchley, K., Plaza, J., Chen, Y., Nikitin, K., Palmer, R. E., Preece, J. A., Evans, S.D., & Fitzmaurice, D. (2004). Gold Nanoparticle Patterning of Silicon Wafers Using Chemical E-Beam Lithography. *Langmuir*, 20, 3766–3768.
- Montague, M., Ducker, R. E., Chong, K. S. L., Manning, R. J., Rutten, F. J. M., Davies, M. C., & Leggett, G. J. (2007). Fabrication of Biomolecular Nanostructures by



- Scanning Near-Field Photolithography of Oligo(ethylene glycol)-terminated Self-Assembled Monolayers. *Nano Letters*, 6(1), 7328–7337.
- Mulder, W. H., Jose, J., & Andreu, R. (2001). A Kinetic Model for the Reductive Desorption of Self-Assembled Thiol Monolayers. *Langmuir*, 17, 3273–3280.
- Nam, J.-M., Thaxton, C. S., & Mirkin, C. A. (2003). Nanoparticle-Based Bio-Bar Codes for the Ultrasensitive Detection of Proteins. *Science*, 301, 1884–1886.
- Netzer, L. & Sagiv, J. (1983). A New Approach to Construction of Artificial Monolayer Assemblies. *Journal of the American Chemical Society*, 105, 674–676.
- Newton, L., Slater, T., Clark, N., & Vijayaraghavan, A. (2013). Self Assembled Monolayers (SAMs) on Metallic Surfaces (Gold and Graphene) for Electronic Applications. *Journal of Materials Chemistry C*, 1, 376-393.
- Onclin, S., Mulder, A., Huskens, J., Ravoo, B. J., & Reinhoudt, D. N. (2004). Molecular Printboards : Monolayers of -Cyclodextrins on Silicon Oxide Surfaces. *Langmuir*, 20, 5460–5466.
- Papra, A., Gadegaard, N., & Larsen, N. B. (2001). Characterization of Ultrathin Poly (ethylene glycol ) Monolayers on Silicon Substrates. *Langmuir*, 17(5), 1457–1460.
- Peeling, R. W., Artsob, H., Pelegriño, J. L., Buchy, P., Cardosa, M. J., Devi, S., Enria, D. A., Farrar, J., Gubler, D. J., Guzman, M. G., Halstead, S. B., Hunsperger, E., Kliks, S., Margolis, H. S., Nathanson, C. M., Nguyen V. C., Rizzo, N., Vazquez, S., & Yoksan, S. (2010). Evaluation of diagnostic tests: dengue. *Nature Reviews Microbiology*, 8(12), 30–37.
- Petrovykh, D. Y., Kimura-suda, H., Tarlov, M. J., & Whitman, L. J. (2004). Quantitative Characterization of DNA Films by X-ray Photoelectron Spectroscopy. *Langmuir*, 20(2), 429–440
- Piner, R. D., Zhu, J., Xu, F., & Hong, S. (1999). “Dip-Pen” Nanolithography. *Science*, 283, 661–664.
- Ptasińska, S., Stypczyńska, A., Nixon, T., Mason, N. J., Klyachko, D. V, & Sanche, L. (2008). X-ray Induced Damage in DNA Monitored by X-ray Photoelectron Spectroscopy. *The Journal of Chemical Physics*, 129(6), 1-6.
- Pujari, S. P., Scheres, L., Marcelis, A. T. M., & Zuilhof, H. (2014). Covalent Surface Modification of Oxide Surfaces. *Angewandte Chemie*, 53, 6322–6356.
- Qin, D., Xia, Y., & Whitesides, G. M. (2010). Soft Lithography for Micro- and Nanoscale Patterning. *Nature Protocols*, 5(3), 491–502.



- Ron, A., Lee, G. H., Amar, L., Ghassemi, S., & Hone, J. (2011). Adjacent Assembly of Self-Assembled Monolayers for the Construction of Selective Bio-Platforms. *Sensors and Actuators B: Chemical*, 159, 75–81.
- Sagiv, J. (1980). Organized Monolayers by Adsorption. 1. Formation and Structure of Oleophobic Mixed Monolayers on Solid Surfaces. *Journal of the American Chemical Society*, 399, 92–98.
- Schlapak, R., Armitage, D., Saucedo-Zeni, N., Hohage, M., & Howorka, S. (2007). Dense Passivating Poly(ethylene glycol) Films on Indium Tin Oxide Substrates. *Langmuir*, 23, 10244–53.
- Seshadri, K., Froyd, K., Parikh, A. N., Allara, D. L., Lercel, M. J., Craighead, H. G. (1996). Electron-Beam-Induced Damage in Self-Assembled Monolayers. *Journal of Physical Chemistry*, 100(39), 15900–15909.
- Sfez, R., De-zhong, L., Turyan, I., Mandler, D., & Yitzchaik, S. (2001). Polyaniline Monolayer Self-Assembled on Hydroxyl-Terminated Surfaces. *Langmuir*, 17, 2556–2559.
- Shaik, S., Mkize, L., Khumalo, M., & Singh, N. (2014). Tetradenia Riparia-Mediated Synthesis of Nano-Gold Particles. *Journal of Nanomaterials and Biostructures*, 9(2), 567–573.
- Sharma, S., Johnson, R. W., & Desai, T. A. (2004). Evaluation of the Stability of Nonfouling Ultrathin Poly(ethylene glycol) Films for Silicon-Based Microdevices. *Langmuir*, 20, 348–356.
- Sharma, S., Johnson, R. W., & Desai, T. A. (2004). XPS and AFM Analysis of Antifouling PEG Interfaces for Microfabricated Silicon Biosensors. *Biosensors & Bioelectronics*, 20, 227–239.
- Sharma, S., Popat, K. C., & Desai, T. A. (2002). Controlling Nonspecific Protein Interactions in Silicon Biomicrosystems with Nanostructured Poly(ethylene glycol) Films. *Langmuir*, 18, 8728–8731.
- Shirtcliffe, N. J., Mchale, G., Atherton, S., & Newton, M. I. (2010). An Introduction to Superhydrophobicity. *Advances in Colloid and Interface Science*, 161(1-2), 124–138.
- Singh, S., Zack, J., Kumar, D., Srivastava, S. K., Saluja, D., Khan, M. A., & Singh, P. K. (2010). DNA Hybridization on Silicon Nanowires. *Thin Solid Films*, 519(3), 1151–1155.
- Skoda, M. W. A., Schreiber, F., Jacobs, R. M. J., Webster, J. R. P., Wolff, M., Dahint, R., Schwendel, D., & Grunze, M. (2009). Protein Density Profile at the Interface of Water with Oligo(ethylene glycol) Self-Assembled Monolayers. *Langmuir*, 25, 4056–4064.

- Smith, R. K., Lewis, P. A., & Weiss, P. S. (2004). Patterning Self-Assembled Monolayers. *Progress in Surface Science*, 75, 1–68.
- Statz, A. R., Meagher, R. J., Barron, A. E., & Messersmith, P. B. (2005). New Peptidomimetic Polymers for Antifouling Surfaces. *Journal of the American Chemical Society*, 127, 7972–7973.
- Storhoff, J. J., Marla, S. S., Bao, P., Hagenow, S., Mehta, H., Lucas, A., Garimella, V., Patno, T., Buckingham, W., Cork, W., & Müller, U. R. (2004). Gold Nanoparticle-Based Detection of Genomic DNA Targets on Microarrays Using a Novel Optical Detection System. *Biosensors and Bioelectronics*, 19, 875–883.
- Sun, S., Mendes, P., Critchley, K., Diegoli, S., Hanwell, M., Evans, S. D., Leggett, G. J., Preece, J. A., & Richardson. (2006). Fabrication of Gold Micro- and Nanostructures by Photolithographic Exposure of Thiol-Stabilized Gold Nanoparticles. *Nano Letters*, 6(3), 345–350.
- Tan, J. L., Tien, J., & Chen, C. S. (2002). Microcontact Printing of Proteins on Mixed Self-Assembled Monolayers. *Langmuir*, 18, 519–523.
- Tanaka, M., Mochizuki, A., Shiroya, T., Motomura, T., Shimura, K., Onishi, M., & Okahata, Y. (2002). Study on Kinetics of Early Stage Protein Adsorption on Poly(2-methoxyethylacrylate) (PMEA) Surface. *Colloids and Surfaces A*, 203, 195–204.
- Taton, T. A. (2000). Scanometric DNA Array Detection with Nanoparticle Probes. *Science*, 289, 1757–1760.
- Tizazu, G., Adawi, A. M., Leggett, G. J., & Lidzey, D. G. (2009). Photopatterning, Etching, and Derivatization of Self-Assembled Monolayers of Phosphonic Acids on the Native Oxide of Titanium. *Langmuir*, 25(18), 10746–10753.
- Todd, S. J., Scurr, D. J., Gough, J. E., Alexander, M. R., & Ulijn, R. V. (2009). Enzyme-Activated RGD Ligands on Functionalized Poly(ethylene glycol) Monolayers: Surface Analysis and Cellular Response. *Langmuir*, 25(13), 7533–7539.
- Toworfe, G. K., Composto, R. J., Shapiro, I. M., & Ducheyne, P. (2006). Nucleation and Growth of Calcium Phosphate on Amine-, Carboxyl- and Hydroxyl-Silane Self-Assembled Monolayers. *Biomaterials*, 27, 631–642.
- Truong, N. P., Whittaker, M. R., Mak, C. W., & Davis, T. P. (2015). The Importance of Nanoparticle Shape in Cancer Drug Delivery. *Expert Opinion on Drug Delivery*, 12(1) 129–142.
- Turner, J. N., & Shain, W. (1998). Patterned Protein Layers on Solid Substrates by Thin Stamp Microcontact Printing. *Langmuir*, 14, 741–744.

- Ulman, A. (1996). Formation and Structure of Self-Assembled Monolayers. *Chemical Reviews*, 96(4), 1533–1554.
- Vijayakumar, T. S., Chandy, S., Sathish, N., Abraham, M., Abraham, P., & Sridharan, G. (2005). Is Dengue Emerging as a Major Public Health Problem? *The Indian Journal of Medical Research*, 121, 100–107.
- Wang, J. (2000). From DNA Biosensors to Gene Chips. *Nucleic Acids Research*, 28(16), 3011–3016.
- Wang, J., Polsky, R., & Xu, D. (2001). Silver-Enhanced Colloidal Gold Electrochemical Stripping Detection of DNA Hybridization. *Langmuir*, 17(19), 5739–5741.
- Watts, J. F., & Wolstenholme J. (2003). An Introduction to Surface Analysis by XPS and AES (pp. 324–325), John Wiley & Sons, Inc.
- Weaver, S. C., & Reisen, W. K. (2010). Present and Future Arboviral Threats. *Antiviral Research*, 85(2), 328–345.
- Whitesides, G. M., Ostuni, E., Takayama, S., Jiang, X., & Ingber, D. E. (2001). Soft Lithography in Biology and Biochemistry. *Annual Review of Biomedical Engineering*, 3, 335–373.
- Xue, C., & Yang, K. (2007). Chemical Modifications of Inert Organic Monolayers with Oxygen Plasma for Biosensor Applications. *Langmuir*, 23(10), 5831–5835.
- Xue, C.-Y., & Yang, K.-L. (2010). One-step UV Lithography for Activation of Inert Hydrocarbon Monolayers and Preparation of Protein Micropatterns. *Journal of Colloid and Interface Science*, 344, 48–53.
- Yang, W., Xue, H., Li, W., Zhang, J., & Jiang, S. (2009). Pursuing “Zero” Protein Adsorption of Poly(carboxybetaine) from Undiluted Blood Serum and plasma. *Langmuir*, 25(19), 11911–11916.
- Yang, Z., Frey, W., Oliver, T., & Chilkoti, A. (2000). Light-Activated Affinity Micropatterning of Proteins on Self-Assembled Monolayers on Gold. *Langmuir*, 16, 1751–1758.
- Yu, Q., Zhang, Y., Wang, H., Brash, J., & Chen, H. (2011). Anti-Fouling Bioactive Surfaces. *Acta Biomaterialia*, 7, 1550–1557.
- Zhang, G.-J., Tanii, T., Funatsu, T., & Ohdomari, I. (2004). Patterning of DNA Nanostructures on Silicon Surface by Electron Beam Lithography of Self-Assembled Monolayer. *Chemical Communications*, 7, 786–787.
- Zhang, G.-J., Zhang, L., Huang, M. J., Luo, Z. H. H., Tay, G. K. I., Lim, E.-J. A., Kang, T.G., & Chen, Y. (2010). Silicon Nanowire Biosensor for Highly Sensitive and

Rapid Detection of Dengue Virus. *Sensors and Actuators B: Chemical*, 146, 138–144.

Zhang, M., Desai, T., & Ferrari, M. (1998). Proteins and Cells on PEG Immobilized Silicon Surfaces. *Biomaterials*, 26, 953–960.

Zhu, X., Jun, Y., Staarup, D. R., Major, R. C., Danielson, S., Boiadjev, V., Gladfelter, W.L., Bunker, B.C., & Guo, A. (2001). Grafting of High-Density Poly ( Ethylene Glycol ) Monolayers on Si ( 111 ). *Langmuir*, 17(25), 7798–7803.

