



UNIVERSITI PUTRA MALAYSIA

***PRETREATMENT OF LIGNOCELLULOSIC MATERIALS BY DEEP
EUTECTIC SOLVENTS FOR ENHANCED ENZYMATIC HYDROLYSIS***

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By

SYARILADA ZULKEFLI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of Requirement for the Degree of Master of Science**

May 2015

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

PRETREATMENT OF LIGNOCELLULOSIC MATERIALS BY DEEP EUTECTIC SOLVENTS FOR ENHANCED ENZYMATIC HYDROLYSIS

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The potential of deep eutectic solvent (DES) for the process of oil palm biomass and cellulose conversion into glucose was investigated. DES has been studied extensively in various fields such as organic synthesis, biocatalysis, dissolution, electrochemistry and extraction process. Owing to its cheap and 'green' properties, we are interested in exploring the potential of DES for biomass processing. In order to achieve optimized glucose conversion via enzymatic hydrolysis, biomass must undergo pretreatment in order to break down the structure and this can enhance the access of cellulase to cellulose matrix which usually preferred amorphous or unorganized cellulose structure.

Herein, choline chloride:glycerol (ChCl:Gly), choline chloride:ethylene glycol (ChCl:EG), ethylammonium chloride:glycerol (EAC:Gly), ethylammonium chloride:ethylene glycol (EAC:EG) and choline chloride:urea (ChCl:U) with 1:2 molar ratio for all DESs were used as solvent for pretreatment of oil palm biomass and cellulose. The dissolution process was done by heating and stirring 5% (w/w) oil palm biomasses which were trunk (OPT), frond (OPF) and empty fruit bunch (EFB) in ChCl:Gly, ChCl:EG, EAC:Gly, EAC:EG and ChCl:U at 100 °C for 48 h and cellulose for 24 h at the same temperature. The chemical composition for pretreated oil palm biomass was investigated and it was found that EAC:EG gave major removal of hemicellulose in OPT, while significant lignin reduction was shown in all DESs for OPT. The highest percentage of dissolution for oil palm biomass was recorded by OPT pretreated in EAC:EG with 55.6% while the lowest dissolution recorded by OPF pretreated in ChCl:Gly with only 20%. The highest percentage of dissolution for cellulose was also recorded when pretreated in EAC:EG with 22.9% while ChCl:EG gives the lowest dissolution with only 7.6%. The regenerated or pretreated oil palm biomass was recovered by adding distilled water/ethanol to the pretreated sample. Recovered pretreated samples were analysed by optical microscopy for physical structure study. The result shows that EAC:EG provides better media for dissolution process as the solvent produced more small fragments which indicates that dissolution occurred for oil palm biomass and cellulose. From FTIR analysis, EAC:EG was recorded to be the best solvent for disruption of hydroxyl group in the biomass material while ChCl:U recorded the best solvent in removing lignin for OPT.

The screening of parameter study was carried out for enzymatic hydrolysis in order to determine the optimum conversion for the substrate pretreated in DES. Study of few crucial factors affecting the reaction rate such as hydrolysis media, enzyme loading and substrate concentration was conducted. The highest glucose release among pretreated oil palm biomass was when hydrolysing OPT pretreated in EAC:EG with 60% conversion. Cellulose hydrolysis recorded high amount of glucose release within range of 77-95% conversion. The optimisation study was carried out by using OPT pretreated in EAC:EG, not the cellulose because it considered to already achieved high conversion rate. The optimum condition was obtained by using 15 mg/ml as substrate concentration, 50 FPU/g and 100 CBU/mL as enzyme loading for Celluclast 1.5 L and Novozyme 188 respectively at 50 °C for 24 h with 73.5% glucose conversion.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Sarjana Sains

PRARAWATAN BAHAN LIGNOSELLULOSIK OLEH PELARUT EUTEKTIK UNTUK PENINGKATAN HIDROLISIS BERENZIM

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Potensi pelarut eutektik bagi proses penukaran biomass kelapa sawit dan selulosa kepada glukosa telah dikaji. Kajian terhadap pelarut eutektik sedang giat dijalankan di dalam beberapa bidang seperti sintesis organik, biopemangkinan, pemelarutan, elektrokimia dan proses pengekstrakan. Oleh kerana ia murah dan bersifat 'hijau', kami tertarik untuk mendalami potensi pelarut eutektik bagi pemprosesan hasil buangan. Untuk mencapai penukaran glukosa yang optima melalui hidrolisis berenzim, hasil buangan harus menjalani prarawatan untuk menguraikan strukturnya dan ini boleh meningkatkan laluan untuk selulosa ke matrik selulosa dimana kebiasaannya lebih cenderung kepada amorfos ataupun struktur selulosa yang tidak teratur.

Disini, kolin klorida:gliserol (ChCl:Gly), kolin klorida:etilena glikol (ChCl:EG), etilammonia klorida:gliserol (EAC:Gly), etilammonia klorida:etilena glikol (EAC:EG) dan kolin klorida:urea (ChCl:U) dengan kadar molar 1:2 untuk semua DES telah digunakan sebagai pelarut untuk prarawatan biomass kelapa sawit dan selulosa. Proses pemelarutan telah dilakukan dengan memanaskan dan mengacau 5% (b/b) biomass kelapa sawit iaitu batang pokok kelapa sawit, pelepah pokok kelapa sawit dan tandan buah kosong kelapa sawit, pada suhu 100°C untuk 48 jam dan selulosa untuk 24 jam pada suhu yang sama. Komposisi kimia dalam bahan pra-rawat biomass kelapa sawit telah dikaji dan mendapati EAC:EG memberikan penyingkiran hemiselulosa yang banyak dalam OPT, sementara penyingkiran lignin yang besar telah ditunjukkan oleh semua DES untuk OPT. Peratus pemelarutan tertinggi untuk biomass kelapa sawit adalah direkodkan oleh OPT diprarawat di dalam EAC:EG dengan 55.6%, sementara pemelarutan terendah direkodkan oleh OPF diprarawat di dalam ChCl:Gly dengan hanya 20%. Peratusan pemelarutan paling tinggi bagi selulosa adalah direkodkan apabila diprarawat di dalam EAC:EG dengan 22.9%, sementara ChCl:EG memberikan peratusan pemelarutan paling rendah dengan hanya 7.6%.

Penghasilan semula atau biomass kelapa sawit yang telah diprarawat diambil dengan menambahkan air suling/etanol pada sampel prarawat. Sampel yang diprarawat telah diambil untuk dianalisa oleh mikroskop optik untuk kajian struktur fizikal. Hasil menunjukkan bahawa EAC:EG menyediakan media yang lebih baik untuk proses pemelarutan dimana pelarut itu menghasilkan lebih banyak fragmentasi kecil,

menunjukkan bahawa pemelarutan berlaku untuk biomass kelapa sawit dan selulosa. Daripada analisis FTIR, EAC:EG merekodkan sebagai pelarut terbaik untuk penggangguan kumpulan hidroksil dalam bahan biomass sementara CHCl_3 merekodkan pelarut terbaik dalam penyingkiran lignin untuk OPT.

Kajian saringan parameter telah dijalankan untuk hidrolisis berenzim bagi mengenalpasti penukaran yang optimum untuk substrat prarawat di dalam DES. Kajian terhadap beberapa faktor penting yang mempengaruhi kadar tindak balas seperti media hidrolisis, muatan enzim dan kepekatan substrat telah dijalankan. Penukaran glukosa yang tertinggi di kalangan hasil buangan kelapa sawit yang telah diprarawat adalah apabila menghidrolisis OPT yang diprarawat di dalam EAC:EG dengan penukaran sebanyak 60%.

Hidrolisis selulosa merekodkan kandungan pembebasan glukosa yang tinggi iaitu antara 77-95%. Kajian optimum diteruskan dengan menggunakan OPT yang telah diprarawat di dalam EAC:EG, bukan selulosa kerana ia dianggap sudah mencapai kadar penukaran yang tinggi. Keadaan optimum telah diperolehi dengan menggunakan 15 mg/ml sebagai kepekatan substrat, 50 FPU/g dan 100 CBU/g sebagai muatan enzim untuk Celluclast 1.5 L dan Novozyme 188, pada suhu 50°C untuk 24 jam dengan 73.5% penukaran glukosa.

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The thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirements of the degree of Master of Science. The members of the Supervisory Committee were as follows:

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TABLE OF CONTENTS

ABSTRACT	Page i
ABSTRAK	iii
ACKNOWLEDGEMENT	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xii
LIST OF FIGURES	xiii
LIST OF APPENDICES	xv
LIST OF ABBREVIATIONS	xvi

CHAPTER

1. INTRODUCTION	
1.1 Background of Study	1
1.2 Problem Statement	2
1.3 Aim and Objectives	3
2. LITERATURE REVIEW	
2.1 Sources of Biomass	4
2.2 Oil Palm Biomass	5
2.2.1 Current Utilization of Oil Palm Biomass	7
2.2.2 Chemical Composition of Biomass	8
2.3 Pretreatment of Oil Palm Biomass	11
2.3.1 Physical Pretreatment	11
2.3.2 Biological Pretreatment	12
2.3.3 Chemical Pretreatment	12
2.4 Regenerated Cellulose-rich Solid from Biomass	15
2.5 Swelling and Dissolution of Oil Palm Biomass and Cellulose	15
2.6 Deep Eutectic Solvent (DES)	17
2.6.1 Background	17
2.6.2 Physico-chemical Properties of DES	18
2.6.2.1 Depression of Freezing Point	18
2.6.2.2 Polarity	20
2.6.2.3 Viscosity	21
2.6.3 Application of DESs	22
2.7 Hydrolysis of Lignocellulosic Materials	24
3. MATERIALS AND METHODS	
3.1 Materials	27
3.2 Method	27
3.2.1 Preparation of Deep Eutectic Solvent (DES)	27
3.2.2 Determination of Moisture Content	28
3.2.3 Determination of Extractive Content	28
3.2.4 Determination of Holocellulose Content	29

3.2.5	Determination of α -cellulose Content	29
3.2.6	Determination of Lignin Content	30
3.3	Dissolution of Oil Palm Biomass	30
3.3.1	Regeneration of Dissolved Materials	31
3.4	Observation of Dissolution Mechanism using Optical Microscopy	31
3.5	Fourier Transform Infrared Spectroscopy (FTIR)	31
3.6	Enzymatic Hydrolysis	31
3.7	High Performance Liquid Chromatography	32
4.	RESULTS AND DISCUSSION	
4.1	Chemical Composition of Oil Palm Biomass	33
4.2	Dissolution of Oil Palm Biomass	35
4.2.1	Physical Changes on Lignocellulosic Materials	35
4.2.2	Solubility	36
4.3	Observation of Swelling and Dissolution using Optical Microscopy	40
4.3.1	Effect of Different Ammonium Salt of DESs	40
4.3.2	Effect of Different Incubation Time	42
4.4	Fourier Transform-Infrared Spectroscopy (FTIR) Analysis	47
4.5	Enzymatic Hydrolysis	53
4.5.1	Effect of Different DESs Pretreatment	53
4.5.2	Effect of Different Hydrolysis Media	55
4.5.3	Effect of Different Enzyme Loading	56
4.5.4	Effect of Substrate Concentration	58
5.	CONCLUSIONS AND RECOMMENDATIONS FOR FUTURE RESEARCH	60
5.1	Recommendations and Suggestions	61
	REFERENCES	62
	BIODATA OF STUDENT	85
	LIST OF PUBLICATIONS	86

LIST OF TABLES

Table		Page
1	Land area of crops plantation and annual production in Malaysia for year 2007 (DOA, 2009)	5
2	Quantity of different biomass produced for year 2007 in Malaysia (Goh et al., 2010)	7
3	Chemical composition of different types of lignocellulosic biomass	9
4	Effect of different chemical pretreatment on sugar release	13
5	Effect of biomass dissolution on sugar release	14
6	List of reported DES where T_f =freezing point, T_m =melting point and T_g =glass transition temperature. HBA=hydrogen bond acceptor, HBD=hydrogen bond donor.	19
7	Polarity value of previous DES reported according to scale of $E_T(30)$ and normalized scale E_T^N	21
8	Viscosity of several DES from previous report	22
9	DESs used in this study	28
10	Percentage of chemical composition of untreated and pretreated Oil palm biomass determined by TAPPI method	
11	Viscosity of DESs used in this study is determined by Viscometer (Brookfield) at 34°C	47
12	The percentage of relative change of oil palm biomass in DESs	49
13	Percentage relative changes for cellulose in different DESs	51

LIST OF FIGURES

Figure		Page
1	Biomass from logging tree residues; a) woodchips and b) tree barks	4
2	Lignocellulosic biomass from agricultural crop of a) sugarcane and b) wheat	5
3	Oil palm plantation	6
4	The composition of oil palm fruit (Sime Darby Plantation, 2013)	6
5	Types of oil palm biomass produced annually. a) frond, b) trunk and c) empty fruit bunch	8
6	Hydrogen bond linkage within cellulose matrix caused it to exist naturally in crystalline form (Wang <i>et al.</i> , 2012)	9
7	Lignin is built up by complex polyaromatic compounds of sinapyl alcohol, coniferyl alcohol and p-coumaryl alcohol. All 3 different alcohol are held together generating different types of linkage within lignin matrix	10
8	Illustration of lignocellulosic biomass pretreatment proposed by Mosier <i>et al.</i> , (2005)	11
9	Mechanism of acid catalysed for cellulose hydrolysis	24
10	Skeletal structure of ammonium salt used in DESs	35
11	a) 5% (w/w) of fiber in DES was heated at 100°C for 24 h. b) Mixture of sulphuric acid and lignin extracted from oil palm fiber	36
12	Percentage dissolution of lignocellulosic materials in different DESs carried out at 100°C in 24 h by heating in an oil bath	37
13	Overall mechanism between DES and a) cellulose and xylan (hemicellulose) and b) coniferyl alcohol (lignin)	38
14	OPT in a) ChCl:Gly, b) ChCl:EG and c) ChCl:U at t=24 h. The fibres show an interaction of homogenous swelling without dissolution for each solvent	40
15	OPT in a) EAC:Gly and b) EAC:EG at t=24 h. The fibres also have the same interaction as the choline chloride-based DES; homogenous swelling with no dissolution.	41
16	Cellulose in a) ChCl:Gly, b) ChCl:EG, c) EAC:Gly, d) EAC:EG and e) ChCl:U at t=24 h.	42
17	OPT in ChCl:Gly at a) t=0 h, b) t=3 h, c) t=5 h, d) t=16 h and e) t=24 h	43
18	OPT in EAC:Gly at a) t=0h, b) t=3 h, c) t=5 h, d) t=16 h and e) t=24 h	44
19	a) Homogenous swelling of wood fibre from Cuissinat and Navard (2008) study, b) OPT in EAC:EG at 100°C for 48 h without stirring, and c) OPT treated in EAC:EG for 20 h and was observed under 20x magnification.	45
20	Flat rings appeared in the dissolution mixture. Images were observed at 20x magnification.	46
21	Schematic diagram proposed to describe the origin of flat rings exists in the dissolution mixture (Jardeby <i>et al.</i> , 2005)	46
22	IR spectrum of raw a) OPF and b) cellulose	48
23	FT-IR spectrum of a) raw OPT, treated in b) ChCl:Gly, c) ChCl:EG, d) EAC:Gly, e) EAC:EG and f) ChCl:U.	49
24	FTIR spectrum of a) raw OPF treated in b) ChCl:Gly, c) ChCl:EG, d) EAC:Gly, e) EAC:EG and f) ChCl:U.	51

25	FTIR spectrum of a) raw EFB, treated in b) ChCl:Gly, c) ChCl:EG, d) EAC:Gly, e) EAC:EG and f) ChCl:U.	52
26	FTIR spectrum of a) raw cellulose, treated in b) ChCl:Gly, c) ChCl:EG, d) EAC:Gly, e) EAC:EG and f) ChCl:U.	52
27	Enzymatic hydrolysis of pretreated OPT, OPF, EFB and cellulose in different DESs.	54
28	Enzymatic hydrolysis of OPT pretreated in EAC:EG in different DESs concentration as hydrolysis media.	55
29	Enzymatic hydrolysis of OPT pretreated in EAC:EG with different enzyme loading of Celluclast 1.5 L.	57
30	Enzymatic hydrolysis of OPT treated in EAC:EG with different enzyme loading of Novozyme 188.	58
31	Enzymatic hydrolysis of OPT treated in EAC:EG with different substrate concentration.	59



LIST OF APPENDICES

Appendix		Page
A.1	Calibration curve for concentration of glucose	75
B.1	FTIR spectrum of untreated OPT	76
B.2	FTIR spectrum of untreated OPF	77
B.3	FTIR spectrum of untreated EFB	78
B.4	FTIR spectrum of untreated α -cellulose	79
B.5	FTIR spectrum of treated in ChCl:Gly	80
B.6	FTIR spectrum of OPT in ChCl:EG	81
B.7	FTIR spectrum of OPT treated in EAC:Gly	82
B.8	FTIR spectrum of OPT treated in EAC:EG	83
B.9	FTIR spectrum of OPT in ChCl:U	84

LIST OF ABBREVIATIONS

DES	Deep eutectic solvent
HBA	Hydrogen bond acceptor
HBD	Hydrogen bond donor
LTTM	Low transition temperature mixture
ChCl	Choline chloride
Gly	Glycerol
EAC	Ethylammonium chloride
EG	Ethylene glycol
U	Urea
IL	Ionic liquid
OPT	Oil palm trunk
OPF	Oil palm frond
EFB	Empty fruit bunch
FT-IR	Fourier Transform Infrared Spectroscopy
HPLC	High Performance Liquid Chromatography
h	Hour
min	Minute
rpm	Rotation per minute
g	gram
FPU	Filter Paper Unit
CBU	Cellobiase Unit

CHAPTER 1

INTRODUCTION

1.1 Background of Study

Deep eutectic solvent (DES) is a novel solvent which is a combination of two components giving the solvent a very low melting point compared to its constituting components (Abbott *et al.*, 2003). It is non-flammable, biodegradable, less toxic and inexpensive, which can be suitable for many applications. Generally, the components involve hydrogen bond donor (alcohols, amides, amino acids, monosaccharides) and ammonium salt (choline chloride, betaine, ethylammonium chloride, tetra butylammonium chloride) in certain molar ratio. Variety of combinations make the solvent more attractive as changing the pairing will alter the physical or chemical properties of DES. This can be advantageous because by changing the properties of DES will provide wider range of applications by satisfying the requirements in certain process.

Many works have been done in exploring the potential of DES which includes metal dissolution, extraction and purification, electrochemistry, biocatalysis, organic synthesis and biomass processing. However, little work was done in exploring the potential of DES in biomass processing. Francisco *et al.*, (2012) worked on cellulose, starch and lignin dissolution by using LTTMs mixture and showed that most of the solvents can dissolve lignin such as malic acid:proline, lactic acid:betaine, lactic acid:choline chloride and lactic acid:histidine, from wheat straw. However, cellulose and starch did not give promising result as the percentage of dissolution was very low. From this report, it opens up more room to improve the LTTMs or DESs as the solvent for lignocellulosic biomass pretreatment. Thus, in this study, DESs was utilised for pretreatment of lignocellulosic biomass with aim to deconstruct the highly crystalline cellulose and remove or dissolve the lignin and hemicellulose to achieve optimum conversion of fermentable sugar.

There are 2 types of bioethanol, first generation (FGB) and second generation bioethanol (SGB). Currently, bioethanol is produced from food source such as starch and sugar-based feedstock which is categorized as FGB (Tan *et al.*, 2010). However, debate arises on the feedstock of FGB where competition between human-fuel supplies should be avoided. Therefore, SGB can be the alternative to FGB because it can be produced from lignocellulosic biomass. Biomass or wastes that can be converted into valuable end-products will contribute huge benefits on environment and able to avoid using human food source as feedstock for fuel production. Nevertheless, the lignocellulosic biomass is a complex material consists of highly crystalline cellulose, hemicellulose and lignin that could hinder the efficiency of the production (Silverstein *et al.*, 2007). Thus, lignocellulosic biomass must undergo few processes in order to obtain bioethanol.

Pretreatment process is the crucial step to break down the highly crystalline cellulose within the lignocellulosic biomass. The most common pretreatment process of lignocellulosic biomass usually involves the physical and chemical pretreatment. However, the pretreatment process for lignocellulosic biomass is still in research scale. Extensive studies have been done on both physical and chemical pretreatment where usually involves steam explosion and alkali pretreatment respectively (Holm and Lassi, 2011). By changing the crystalline structure to amorphous, enzyme or microbe will easily hydrolyse the polysaccharide into monosaccharide (glucose or xylose) via enzymatic hydrolysis. Enzymatic route is the preferred method as it can be carried out in mild condition and this could reduce consumption of energy, cost and corrosive chemicals. Due to the extensive development of these processes, there are many types of pretreatment and hydrolysis has been introduced. Chemical pretreatment by using ionic liquid (IL) has attracted many chemists since it provides good quality of solvent for lignocellulosic biomass. However, IL has few limitations regarding its toxicity and cost to synthesise where it may not be efficient for large-scale process. On the other hand, DES has low toxicity and can be produced using cheap material such as choline chloride. Therefore, the potential of DES in biomass processing is enormous.

1.2 Problem Statement

In this study, the concern is to achieve access of cellulase and cellobiase to cellulose from the oil palm biomass in order to obtain high yield of glucose. In lignocellulosic biomass, cellulose is hindered by heteropolymer of hemicellulose and lignin. Removal of lignin and hemicellulose can assist the access of enzymes on cellulose. The highly crystalline cellulose structure is also one of the issues where although removal of lignin and hemicellulose can enhance the enzymatic hydrolysis, the cellulose must be in amorphous or less packed structure in order for enzymes to be effective.

Thus, the lignocellulosic biomass should undergo pretreatment process to break or open up the cell wall and to decrease the recalcitrant by using physical, chemical or biological pretreatment. However, the common pretreatment process uses harsh chemicals such as sodium hydroxide (NaOH) solution that could cause the reactor to corrode. IL is reported the new cellulose solvent for the last few decades and researches are extensively carried out. Unfortunately, IL is very costly in terms of its preparation and materials, thus it is not suitable for practical scale. DES is one of the ionic solvent where it is cheap, biodegradable and environmental friendly. However, limited work was done on DES related to biomass processing. In order to fill the gap, DESs were used in this study to pretreat the lignocellulosic biomass prior to enzymatic hydrolysis.

1.2 Aim and Objectives

The aim of this study is to explore the potential of DES in pretreatment and enzymatic hydrolysis of oil palm biomass and cellulose. The objectives were set as follows:

- 1) To pretreat oil palm biomass and cellulose in DES and determine the physico-chemical changes of the pretreated materials.
- 2) To enhance the enzymatic hydrolysis of oil palm biomass by pretreatment using DES.



REFERENCES

- Abbott, A.P., Barron, J.C., Ryder, K.S. and Wilson, D. (2007). Eutectic-based ionic liquids with metal-containing anions and cations. *Chemistry-A European Journal*, 13: 6495-6501.
- Abbott, A.P., Boothby, D., Capper, G., Davies, D.L. and Rasheed, R.K. (2004). Deep eutectic solvents formed between choline chloride and carboxylic acid: versatile alternatives to ionic liquids. *Journal of the American Chemical Society*, 126: 9142-9147.
- Abbott, A.P., Capper, G. and Gray, S. (2006). Design of improved deep eutectic solvents using hole theory. *Chemical Physics and Physical Chemistry*, 7: 803-806.
- Abbott, A.P., Capper, G., Davies, D.L., Rasheed, R.K. and Tambyrajah, V. (2003). Novel solvent properties of choline chloride/urea mixtures. *Chemical Communications*, 70-71.
- Abbott, A.P., Capper, G., Davies, D.L., Rasheed, R.K. and Tambyrajah, V. (2002). Quarternary ammonium zinc- or tin-containing ionic liquids: water insensitive, recyclable catalysts for Diels-Alder reactions. *Green Chemistry*, 4: 24-26.
- Abbott, A.P., Frisch, G., Hartley, J. and Ryder, K.S. (2011). Processing of metals and metal oxides using ionic liquid. *Green Chemistry*, 13: 471-482.
- Abbott, A.P., Harris, R.C. and Ryder, K.S. (2007a). Application of hole theory to define ionic liquids by their transport properties, *Journal of Physical Chemistry B*, 111:4910-4913.
- Abbott, A.P., Harris, R.C., Ryder, K.S., D'Agostino, C., Gladden L.F. and Mantle, M.D. (2011a). Glycerol eutectics as sustainable solvent system. *Green Chemistry*, 13; 82-91.
- Agbor, V.B., Cicek, N., Sparling, R., Berlin, A. and Levin, D.B. (2011). Biomass pretreatment: Fundamentals toward application. *Biotechnology Advances*, 29: 675-685.
- Akpınar, O., Erdogan, K., Bakir, U. and Yilmaz, L. (2010). Comparison of acid and enzymatic hydrolysis of tobacco stalk and xylan for preparation of xylooligosaccharides. *LWT-Food Science and Technology*, 43: 119-125.
- Atwater, C. (2000). Ullman's encyclopedia of industrial chemistry, Wiley-VCH Verlag GmbH & co. KGaA.
- Bahcegul, E., Apaydin, S., Haykir, N.I., Tatli, E. and Bakir, U. (2012). Different ionic liquids favour different lignocellulosic biomass particle sizes during pretreatment to function efficiently. *Green Chemistry*, 14: 1896-1903.
- Balat, M., Balat, H. and Oz, C. (2008). Progress in bioethanol processing. *Progress in Energy and Combustion Science*, 34: 551-573.

- Banerjee, G., Car, S., Scott-Craig, J.S., Borrusch, M.S., Bongers, M. and Walton, J.D. (2010). Synthetic multi-component enzyme mixtures for deconstruction of lignocellulosic biomass. *Bioresource Technology*, 101: 9097-9105.
- Basiron, Y. and Simeh, M.D. (2005). Vision 2020—the palm oil phenomenon. *Oil Palm Industry Economic Journal* 5: 1–10.
- Brandt, A., Grasvik, J., Hallett, J.P. and Welton, T. (2013). Deconstruction of lignocellulosic biomass with ionic liquids. *Green Chemistry*, 15: 550-583.
- Brodeur, G., Yau, E., Badal, K., Collier, J., Ramachandran, K.B. and Ramakrishnan, S. (2011). Chemical and physicochemical pretreatment of lignocellulosic biomass: A review. *Enzyme Research*, 2011: 1-17.
- Carere, C.R., Sparling, R., Cicek, N. and Levin, D.B. (2008). Third generation biofuels via direct cellulose fermentation. *International Journal of Molecular Sciences*, 9: 1342-1360.
- Chandra, R.P., Bura, R., Mabee, W.E., Berlin, A., Pan, X. and Saddler, J.N. (2007). Substrate pretreatment: The key to effective enzymatic hydrolysis of lignocellulosics? *Advances in Biochemical Engineering/Biotechnology*, 108: 67-93.
- Chen, M., Zhao, J. and Xia, L. (2009). Comparison of four different chemical pretreatments of corn stover for enhancing enzymatic digestibility. *Biomass and Bioenergy*, 33: 1381-1385.
- Cheng, J.J. and Timilsina, G.R. (2011). Status and barriers of advanced biofuel technologies: a review. *Renewable Energy* 36:3541-3549.
- Chew, T.L. and Bhatia, S. (2008). Catalytic processes towards the production of biofuels in a palm oil and oil palm biomass-based biorefinery. *Bioresource Technology* 99: 7911-7922.
- Chiew, Y.L. and Shimada, S. (2013). Current state and environmental impact assessment for utilizing oil palm empty fruit bunches for fuel, fiber and fertilizer: A case study of Malaysia. *Biomass and Bioenergy*, 51: 109-124.
- Ciolacu, D., Kovac, J. and Kokol, V. (2010). The effect of the cellulose-binding domain from *Clostridium cellulovorans* on the supramolecular structure of cellulose fibres. *Carbohydrate Research*, 345: 621-630.
- Conde-Mejia, C., Jimenez-Gutierrez, A. and El-Halwagi, M. (2011). A comparison of pretreatment methods for bioethanol production from lignocellulosic materials. *Process Safety and Environmental Protection*, 90: 189-202.
- Cuissinat, C. and Navard, P. (2006) Swelling and dissolution of cellulose part I: Free floating cotton and wood fibres in N-Methylmorpholine-N-oxide-water mixtures. *Macromolecular Symposium*, 244: 1-18.

- Cuissinat, C. and Navard, P. (2006a). Swelling and dissolution of cellulose part II: Free floating cotton and wood fibres in NaOH-water-additives systems. *Macromolecular Symposium*, 244: 19-30.
- Cuissinat, C., Navard, P. and Heinz, T. (2008). Swelling and dissolution of cellulose: Part V: cellulose derivatives fibres in aqueous systems and ionic liquids. *Cellulose* 15:75-80.
- Cuissinat, C., Navard, P. and Heinze, T. (2008a). Swelling and dissolution of cellulose. Part IV: Free floating cotton and wood fibres in ionic liquid. *Carbohydrate Polymers*, 72: 590-596.
- D'Agostino, C., Harris, R.C., Abbott, A.P., Gladden, L.F. and Mantle, M.D. (2011). Molecular motion and ion diffusion in choline chloride based deep eutectic solvents studied by ^1H pulsed field gradient NMR spectroscopy. *Phys. Chem. Chem. Phys.*
- Dadi, A., Varanasi, S. and Schall, C.A. (2006) Enhancement of cellulose saccharification kinetics using an ionic liquid pretreatment step. *Wiley Periodicals Inc.*
- Dai, Y., Spronsen, J.v., Witkamp, G.-J., Verpoorte, R. and Choi, Y.H. (2013). Ionic liquids and deep eutectic solvents in natural products research: mixture of solids as extraction solvents. *Journal of Natural Products*, 76: 2162-2173.
- De Santi, V., Cardellini, F. Brinchi, L and Germani, R. (2012). Novel Bronsted acidic deep eutectic solvents as reaction media for esterification of carboxylic acid with alcohols. *Tetrahedron Letters*, 53: 5151-5155.
- Delgenes, J.P., Penaud, V. and Moletta, R. (2002). Pretreatment for the enhancement of anaerobic digestion of solid waster chapter 8. In *Biomethanization of the organic fraction of municipal solid waste*, IWA publishing. 201–228.
- DOA (Department of Agriculture Malaysia), (2009). Data of planting area and production of all commodity plants according to plants type (2003–2008). <<http://www.doa.gov.my/statistik/komoditi03-08.htm>>. Retrieved on June 2014.
- Dominguez de Maria, P. and Maugeri, P. (2011) Ionic liquids in biotransformations: from proof-of-concept to emerging deep-eutectic-solvents. *Current Opinion in Chemical Biology* 15:220-225.
- Dyk, J.S.V. and Pletschke, B.I. (2012). A review of lignocellulose bioconversion using enzymatic hydrolysis and synergistic cooperation between enzymes-Factors affecting enzymes, conversion and synergy. *Biotechnology Advances*, 30: 1458-1480.
- El-Zawawy, W.K., Ibrahim, M.M., Abdel-Fattah, Y.R., Soliman, N.A. and Mahmoud, M.M. (2011). Acid and enzyme hydrolysis to convert pretreated lignocellulosic materials into glucose for ethanol production. *Carbohydrate Polymers*, 84: 865-871.

- Eriksson, K.-E.L., Blanchette, R.A. Ander, P. (1990). Microbial and enzymatic degradation of wood and wood components. Springer-Verlag, Berlin, Germany. 416.
- Feng, L. and Chen, Z.-I. (2008). Research progress on dissolution and functional modification of cellulose in ionic liquids. *Journal of Molecular Liquids* 142:1-5.
- Fort, D.A., Remsing, R.C., Swatlotski, R.P. Moyna, P., Moyna, G. and Rogers, R.D. (2006). Can ionic liquids dissolve wood? Processing and analysis of lignocellulosic materials with 1-n-butyl-3-methylimidazolium chloride. *Green Chemistry*, 9: 63-69.
- Francisco, M., Bruinhorst, A. and Kroon, M.C. (2012). New natural and renewable low transition temperature mixtures (LTTMs): screening as solvents for lignocellulosic biomass processing. *Green Chemistry*, 14: 2153-2157.
- Ghose, T.K. (1987). Measurement of cellulase activities. *Pure and Applied Chemistry*, 59: 257-268.
- Goh, C.S., Tan, K.T., Lee, K.T. and Bhatia, S. (2010). Bio-ethanol from lignocellulose: Status, perspectives and challenges in Malaysia. *Bioresource Technology*, 101: 4834-4841.
- Gorke, J.T., Srienc, F. and Kazlauskas, R.J. (2008). Hydrolase-catalyzed biotransformations in deep eutectic solvents, *Chemical Communications*, 1235-1237.
- Gurmit, S., Lim Kim, H., Teo, L. and David Lee, K. (1999). Oil palm and the environment a Malaysian perspective. *Malaysian Oil Palm Growers Council*, 1: 253.
- Ha, S.H., Mai, N.L., An, G. and Koo, Y.-M. (2011). Microwave-assisted pretreatment of cellulose in ionic liquid for accelerated enzymatic hydrolysis. *Bioresource Technology* 102: 1214-1219.
- Hall, M., Bansal, P., Lee, J.H., Realff, M.J. and Bommarius, A.S. (2011). Biological pretreatment of cellulose: Enhancing enzymatic hydrolysis rate using cellulose-binding domains from cellulases. *Bioresource Technology* 102: 2910-2915.
- Hall, M., Bansal, P., Lee, J.H., Realff, M.J., and Bommarius, A.S. (2010). Cellulose crystallinity-A key predictor of the enzymatic hydrolysis rate. *FEBS Journal* 277: 1571-1582.
- Hallac, B.B. and Ragauskas, A.J. (2011). Analyzing cellulose degree of polymerization and its relevancy to cellulosic ethanol. *Biofuels, Bioproducts and Biorefining* 5: 215-225.
- Han, J. and Rowell, J. (1997). Chemical composition of fibers [Chapter 5]. In: Rowell Roger M, Young Raymond A, Rowell Judith K, editors. Paper and composites from agro-based resources.

- Harris, R.C. (2008). Physical properties of alcohol based deep eutectic solvents. University of Leicester.
- Harun, R. and Danquah, M.K. (2011). Enzymatic hydrolysis of microalgal for bioethanol production. *Chemical Engineering Journal* 168: 1079-1084.
- Hassan, M.A. and Shirai, Y. (2003). Palm biomass utilization in Malaysia for the production of bioplastic. <www.biomass-asia-workshop.jp/presentation_files/21_AliHassan.pdf>
- Hayyan, A., Mjalli, F.S., AlNashef, I.M., Al-Wahaibi, Y.M., Al-Wahaibi, T. and Hashim, M.A. (2013). Glucose-based deep eutectic solvents: physical properties. *Journal of Molecular Liquids*, 178: 137-141.
- Hendriks, A.T.W.M. and Zeeman, G. (2009). Pretreatments to enhance the digestibility of lignocellulosic biomass. *Bioresource Technology* 100:10-18.
- Holm, J. and Lassi, U. Ionic liquid in the pretreatment of lignocellulosic biomass. (2011). University of Oulu, Department of Chemistry, Finland 24: 545-560.
- Howard, R.L., Abotsi, E, Jansen van Rensburg, E.L. and Howard, S. (2003). Lignocellulose biotechnology: issues of bioconversion and enzyme production. *African Journal of Biotechnology*, 2: 606-619.
- Hsu, T.-C., Guo, G.-L., Chen, W.-H. and Hwang, W.-S. (2010). Effect of dilute acid pretreatment of rice straw on structural properties and enzymatic hydrolysis. *Bioresource Technology* 101: 4907-4913.
- Imperato, G., Hoger, S., Lenoir, D. and Konig, B. (2006). Low melting sugar–urea–salt mixtures as solvents for organic reactions—estimation of polarity and use in catalysis. *Green Chemistry*, 8: 1051-1055.
- Jacobsen, W.E. and Wyman, C.E. (2000). Cellulose and hemicellulose hydrolysis models for applications to current and novel pretreatment process. *Applied Biochemistry and Biotechnology*, 84-86: 81-96.
- Jana Holm and Ulla Lassi (2011). Ionic Liquids in the Pretreatment of Lignocellulosic Biomass, *Ionic Liquids: Applications and Perspectives*, Alexander Kokorin (Ed.), ISBN:978-953-307-248-7.
- Jardebey, K., Germgard, U., Kreutz, B., Heinze, T., Heinze, U. and Lennholm, U. (2005). The influence of fibre wall thickness on the undissolved residuals in CMC solutions. *Cellulose*, 12: 167-175.
- Jiang, F., Zhu, Q., Ma, D., Liu, X. and Han, X. (2011). Direct conversion and NMR observation of cellulose to glucose and 5-hydroxymethylfurfural (HMF) catalysed by acidic ionic liquids. *Journal of Molecular Catalysis A: Chemical*, 334: 8-12.
- Jimenez, J., Dominguez, J.M., Castillon, M.P. and Acerbal, C. (1995). Thermoinactivation of cellobiohydrolase I from reesei QM 9414. *Carbohydrate Research*, 268: 257-266.

- Jung, Y. H., Kim, I. J., Han, J., Choi, I., and Kim, K. H., (2011) Aqueous ammonia pretreatment of oil palm empty fruit bunches for ethanol production, *Bioresource Technology* 102: 9806-9809.
- Jung, Y.H., Kim, I.J., Kim, J.J., Oh, K.K., Han, J.-I., Choi, I.-G. and Kim, K.H. (2011). Ethanol production from oil palm trunks treated with aqueous ammonia and cellulase. *Bioresource Technology* 102:7307-7312.
- Kaiser, E.W. and Hauser, C.R. (1967). Influence of metallic cation in carbonyl addition reactions of certain nitriles and sulfones with benzophenone. *Journal of the American Chemical Society*, 89: 4566-4567.
- Karki, B., Maurer, D., Kim, T.H. and Jung, S. (2011). Comparison and optimization of enzymatic saccharification of soybean fibers recovered from aqueous extractions. *Bioresource Technology*, 102: 1228-1233.
- Katmandu, Nepal. (2006). Workshop on utilization of biomass for renewable energy. http://www.apo-tokyo.org/biomassboiler/D1_downloads/presentations/Nepal_Program_DEC2006/Country_Papers/Malaysia_CP.doc
- Ke, J., Laskar, D.D. and Chen, S. (2011). Biodegradation of hardwood lignocellulosics by the western poplar clearwing borer, *Parenthrene robinaie*. *Biomacromolecules*, 12: 1610-1620.
- Kelly-Yong, T.L., Lee, K.T., Mohamed, A.R. and Bhatia, S. (2007). Potential of hydrogen from oil palm biomass as a source of renewable energy worldwide. *Energy Policy*, 35: 5692-5701.
- Khalil, H.P.A., Ismail, H., Rozman, H.D. and Ahmad, M.N. (2001). The effect of acetylation on interfacial shear strength between plant fiber and various matrices. *European Polymer Journal*, 35: 1037-1045.
- Kilpelainen, I., Xie, H., King, A., Granstrom, M., Heikkinen, S., Dimitris, S. and Argyropoulos. (2007). Dissolution of wood in ionic liquids. *Journal Agricultural and Food Chemistry* 55: 9142-9148
- Kim, K.H. and Hong, J. (2001). Supercritical CO₂ pretreatment of lignocellulose enhances enzymatic cellulose hydrolysis. *Bioresource Technology* 77: 139-144.
- Kondo, T. and Sawatari, C. (1996). A fourier transform infra-red spectroscopic analysis of the character of hydrogen bonds in amorphous cellulose. *Polymer*, 37: 393-397.
- Krassig, H., Schurz, J., Steadman, R.G., Schliefer, K., Albrecht, W., Mohrig, M. and Schlosser, H. (2007). Cellulose. In: Ullmann's Encyclopedia of Industrial Chemistry, Wiley-VHC, Weinheim.
- Kumar, P., Barrett, D.M., Delwiche, M.J. and Stroeve, P. (2009). Methods for pretreatment of lignocellulosic biomass for efficient hydrolysis and biofuel production. *Ind. Eng. Chem. Res.* 48: 3713-3729.

- Kumar, R., Singh, S., and Singh, O.V. (2008). Bioconversion of lignocellulosic biomass: biochemical and molecular perspectives. *Journal of Industrial Microbiol Biotechnology*, 35: 377-391.
- Kuo, C.-H. and Lee, C.-K. (2009). Enhancement of enzymatic saccharification of cellulose by cellulose dissolution pretreatments. *Carbohydrate Polymers*, 77: 41-46.
- Kupiainen, L. (2012). Dilute acid catalysed hydrolysis of cellulose-extension to formic acid. (Doctoral Dissertation). Graduate School in Chemical Engineering, University of Oulu.
- Lenihan, P., Orozco, A., O'Neill, E., Ahmad, M.N.M., Rooney, D.W. and Walker, G.W. (2010). Dilute acid hydrolysis of lignocellulosic biomass. *Chemical Engineering Journal*, 156: 395-403.
- Levine, S.E., Fox, J.M., Blanch, H.W. and Clark, D.S. (2010). A mechanistic model of the enzymatic hydrolysis of cellulose. *Biotechnology and Bioengineering*, 107: 37-51.
- Li, M.-F., Fan, Y.-M., Xu, F., Sun, R.-C. and Zhang, X.-L. (2010) Cold sodium hydroxide/urea based pretreatment of bamboo for bioethanol production: Characterization of the cellulose rich fraction. *Industrial Crops and Products* 32: 551-559.
- Li, M.-F., Sun, S.-N., Xu, F. and Sun, R.-C. (2011). Cold NaOH/urea aqueous dissolved cellulose for benzylation: Synthesis and characterization. *European Polymer Journal*, 47: 1817-1826.
- Lim, S., and Teong, K.T. (2010). Recent trends, opportunities and challenges of biodiesel in Malaysia: An overview. *Renewable and Sustainable Energy Reviews*, 14: 938-954.
- Lindberg, D., Revenga, M. and Widersten, M. (2010). Deep eutectic solvents (DESs) are viable cosolvents for enzyme-catalyzed epoxide hydrolysis. *Journal of Biotechnology*, 147: 169-171.
- Liu, C.F., Sun, R.C., Zhang, A.P. and Ren, J.L. (2006). Preparation of sugarcane bagasse cellulosic phthalate using an ionic liquid as reaction medium. *Carbohydrate Polymers*, 68:17-25.
- Maugeri, Z. and Dominguez de Maria, P. (2012). Novel choline chloride-based deep-eutectic solvents with renewable hydrogen bond donors: levulinic acid and sugar-based polyols. *Royal Science Chemistry Advances*, 2: 421-425.
- Maugeri, Z. and Maria, P.D. (2011). Novel choline-chloride-based deep-eutectic-solvents with renewable hydrogen bond donors: levulinic acid and sugar-based polyols. *Royal Science Chemistry Advances*, 2: 421-425.
- Mekhilef, S., Saidur, R., Safari, A. and Mustafa, W.E.S.B. (2011). Biomass energy in Malaysia: Current state and prospects. *Renewable and Sustainable Energy Reviews* 15:3360-3370.

- Mekhilef, S., Siga, S. and Saidur, R. (2011). A review on palm oil biodiesel as a source of renewable fuel. *Renewable and Sustainable Energy Reviews*, 15: 1937-1949.
- Mielenz, J.R. (2001). Ethanol production from biomass: technology and commercialization status. *Current Opinion in Microbiology*, 4: 324-329.
- Misson, M., Haron, R., AhmadKamaroddin, M.F. and Nor Aishah, S.A. (2009). Pretreatment of empty palm fruit bunch for production of chemicals via catalytic pyrolysis. *Bioresource Technology* 100: 2867-2873.
- Mohamad Husin, Ridzuan Ramli, Anis Mokhtar, Wan Hasamudin Wan Hassan, Kamaruddin Hassan, Ropandi Mamat and Aztimar Abdul Aziz. (2005). Research and development of oil palm biomass in wood based industries. Malaysian oil palm board bulletin/palm oil developments 36/information series.
- Mohan, D., Pittman, C.U., and Steele, P.H., (2006). Pyrolysis of wood/biomass for bio-oil: a critical review, *Energy Fuels*, 848-889.
- Moigne, N. and Navard, P. (2010). Dissolution mechanism of wood cellulose fibres in NaOH/water. *Cellulose*, 17: 31-45.
- Mosier, N., Wyman, C., Dale, B., Elander, R., Lee, Y., Hotzapple, M., and Ladish, M. (2005). Features of promising technologies for pretreatment of lignocellulosic biomass. *Bioresource Technology* 96: 673-686.
- MPOB (Malaysian Palm Oil Board), (2009). A summary on the performance of the malaysian oil palm industry – 2008. <<http://econ.mpob.gov.my/economy/Performance-130109.htm>>.
- Nagel, M.C.V., Koschella, A., Voiges, K., Mischnick, P. and Heinze, T. (2010). Homogenous methylation of wood pulp cellulose dissolved in LiOH/urea/H₂O. *European Polymer Journal*, 46: 1726-1735.
- Nakashima, K., Yamaguchi, K., Taniguchi, N., Arai, S., Yamada, R., Katahira, S., Ishida, N., Takahashi, H., Ogino, C. and Kondo, A. (2011). Direct bioethanol production from cellulose by the combination of cellulase-displaying yeast and ionic liquid pretreatment. *The Royal Society of Chemistry*, 13: 2948-2953.
- Naser, J., Mjalli, F., Jibril, B., Al-Hatmi, S. and Gano, Z. (2013). Potassium carbonate as a salt for deep eutectic solvents. *International Journal of Chemical Engineering and Applications*, 4:114-118.
- Nguyuen, T.-A. D., Kim, K., Han, S.J., Cho, H.Y., Kim, J. W., Park, S.M., Park, J.C. and Sim, S.J. (2010). Pretreatment of rice straw with ammonia and ionic liquid for lignocellulose conversion to fermentable sugars, *Bioresource Technology*, 101:7432-7438.
- Nigam, P.S. and Singh, A. (2010). Production of liquid biofuels from renewable resources. *Prog. Energy Combust. Sci.*
- Palm oil facts and figures fact sheets, Sime Darby. (2014).

- Palonen, H. (2004) Role of lignin in the enzymatic hydrolysis of lignocellulosic. Espoo Publications 520.
- Pandey, K.K. (1999). A study of chemical structure of soft and hardwood and wood polymers by FTIR spectroscopy. *Journal of Applied Polymer Science*, 71: 1969-1975.
- Park, S., Baker, J.O., Himmel, M.E., Parilla, P.A. and Johnson, D.K. (2010). Cellulose crystallinity index: measurement techniques and their impact on interpreting cellulose performance. *Biotechnology for Biofuels*, 3: 10.
- Qiu, Z., Aita, G.M. and Walker, M.S. (2012). Effect of ionic liquid pretreatment on the chemical composition, structure and enzymatic hydrolysis of energy cane bagasse. *Bioresource Technology* 117:251-256.
- Rahman, S.H.A., Choudhury, J.P., Ahmad, A.L. and Kamaruddin, A.H. (2007). Optimization studies on acid hydrolysis of oil palm empty fruit bunch fiber for production of xylose. *Bioresource Technology* 98:554-559.
- Rass-Hansen, J., Falsig, H., Jorgensen, B. and Christensen, C.H. (2007). Perspective bioethanol: fuel or feedstock? *J. Chem. Technol. Biotechnol.* 82:329–333.
- Reichardt, C. (1994). Solvatochromic dyes as solvent polarity indicators. *Chemical Reviews*, 94: 2319-2358.
- Reichardt, C. (2005). Polarity of ionic liquids determined empirically by means of solvatochromic pyridinium *N*-phenolate betaine dyes. *Green Chemistry*, 7: 339-351.
- Rosche, C.M., Dibble, C.J. and Stickel, J.J. (2009). Laboratory-scale method for enzymatic saccharification of lignocellulosic biomass at high-solids loading. *Biotechnology for Biofuels*, 2:28.
- Ruan, D., Zhang, L., Mao, Y., Zeng, M. and Li, X. (2004). Microporous membranes prepared from cellulose in NaOH/thiourea aqueous solution. *Journal of Membrane Science*, 241: 265-274.
- Ruß, C. and König, K. (2012). Low melting mixtures in organic synthesis-an alternative to ionic liquids? *Green Chemistry*, 14: 2969.
- Saeman, J.F. (1945). Kinetics of wood saccharification-Hydrolysis of cellulose and decomposition of sugars in dilute acid at high temperature. *Industrial and Engineering Chemistry*, 37: 43-52.
- Saha, B.C., Iten, L.B., Cotta, M.A. and Wu, Y.V. (2005). Dilute acid pretreatment, enzymatic saccharification and fermentation of wheat straw to ethanol. *Process Biochemistry* 40:3693-3700.
- Sarkar, N., Ghosh, S.K., Bannerjee, S. and Aikat, K. (2012). Bioethanol production from agricultural wastes: An overview. *Renewable Energy* 37: 19-27.

- Shahbaz, K., Mjalli, F.S., Hashim, M.A. and AlNashef, I.A. (2011). Prediction of deep eutectic solvent densities at different temperature. *Thermochimica Acta* 515: 67-72.
- Shamsuri, A.A. and Abdullah, D.K. (2010). Ionic liquids: Preparation and limitations. *Makara, Sains*, 14: 101-106.
- Shinoj, S., Visvanathan, R., Panigrahi, S. and Kochubabu, M. (2011). Oil palm fiber (OPF) and its composites: A review. *Industrial Crops and Products*, 33: 7-22.
- Shuit, S.H., Tan, K.T., Lee, K.T. and Kamaruddin, A.H. (2009). Oil palm biomass as a sustainable energy source: a Malaysian case study. *Energy*, 34: 1225–1235.
- Sills, D.L. and Gossett, J.M. (2012). Using FTIR to predict saccharification from enzymatic hydrolysis of alkali pretreated biomasses. *Biotechnology and Bioenergy*, 109: 353-362.
- Silverstein, R.A., Chen, Y., Sharma-Shivappa, R.R., Boyette, M.D. and Osborne, J. (2007). A comparison of chemical pretreatment methods for improving saccharification of cotton stalks. *Bioresource Technology* 98:3000-3011.
- Singh, B.S., Lobo, H.R. and Shankarling, G.S. (2012). Choline chloride based deep eutectic solvents: magical catalytic system for carbon-carbon bond formation in the rapid synthesis of β -hydroxy functionalized derivatives. *Catalysis Communications*, 24: 70-74.
- Singh, P., Suman, A., Tiwari, P., Arya, N., Gaur, A., Shrivastava, A.K. (2008). Biological pretreatment of sugarcane trash for its conversion to fermentable sugars. *World Journal of Microbiology and Biotechnology* 24:667-673.
- Sumathi, S., Chai, S.P. and Mohamed, A.R. (2008). Utilization of oil palm as a source of renewable energy in Malaysia. *Renewable and Sustainable Energy Reviews*, 12: 2404-2421.
- Sun, N., Rahman, M. Qin, Y., Maxim, M.L., Rodriguez, H. and Rogers, R.D. (2009). Complete dissolution and partial delignification of wood in the ionic liquid 1-ethyl-3-methylimidazolium acetate. *Green Chemistry*, 11: 646-655.
- Sun, Y., and Cheng, J., (2002), Hydrolysis of lignocellulosic materials for ethanol production: A review, *Bioresource Technology*, 83(1): 1-11.
- Suppan, P. and Ghoneim, N. (1997). Solvatochromism. *Royal Science of Chemistry, Cambridge*.
- Swatlotski, R.P., Spear, S.K., Holbrey, J.D. and Rogers, R.D. (2002). Dissolution of cellulose with ionic liquids. *Journal of American Chemistry Society*, 124: 4974-4975.
- Tadesse, H. and Luque, R. (2011). Advances on biomass pretreatment using ionic liquids: an overview. *The Royal Society of Chemistry*

- Taherzadeh, M.J. and Karimi, K. (2007). Enzyme-based hydrolysis processes for ethanol from lignocellulosic material: A review. *Bioresource* 2(4): 707-738.
- Takacs, E., Wojnarovits, L., Foldvary, C., Hargittai, P., Borsa, J. and Sajo, I. (2000). Effect of combined gamma-irradiation and alkali treatment on cotton- cellulose. *Radiation Physics and Chemistry*, 57: 399-403.
- Tan, H.T. and Lee, K.T. (2012). Understanding the impact of ionic liquid pretreatment on biomass and enzymatic hydrolysis. *Chemical Engineering Journal* 183: 448-458.
- Tan, K.T., Lee, K.T. and Mohamed A.R. (2008). Role of energy policy in renewable energy accomplishment: The case of second-generation bioethanol. *Energy policy*, 36: 3360-3365.
- Tang, B. and Row, K.H. (2013). Recent developments in deep eutectic solvents in chemical sciences. *Monatshefte fur Chemie-Chemical Monthly*, 144:1427-1454.
- TAPPI (US Technical Association of Pulp and Paper Industry), Acid Insoluble Lignin in Wood and Pulp, T 222 om-88, 1988.
- TAPPI (US Technical Association of Pulp and Paper Industry), Alcohol-Benzene and Dichloromethane Solubles in Wood and Pulp, T 204 om-88, 1988.
- TAPPI (US Technical Association of Pulp and Paper Industry), Alpha-, Beta- and Gamma- Cellulose in Pulp, T 203 os-74, 1988.
- TAPPI (US Technical Association of Pulp and Paper Industry), Preparation of Wood for Chemical Analysis, T-264 om-88, 1988.
- Tucker, M.P., Nguyen, Q.A., Eddy, F.P., Kadam, K.L., Gedvilas, L.M. and Webb, J.D. (2001). Fourier transform infrared quantitative analysis of sugars and lignin in pretreated softwood solid residues. *Applied Biochemistry and Biotechnology*, 91: 51-61.
- Uju, Shoda, Y., Nakamoto, A., Goto, M., Tokuhara, W., Noritake, Y., Katahira, S., Ishida, N., Nakashima, K., Ogino, C., and Kamiya, N., (2011) Short time ionic liquid pretreatment on lignocellulosic biomass to enhance enzymatic saccharification, *Bioresource Technology*
- Umikalsom, M.S., Ariff, A.B. and Karim, M.I.A. (1998). Saccharification of oil palm empty fruit bunch fiber using cellulase of *Chaetomium globosum*. *Journal of Agriculture and Food Chemistry* 46:3359-3364.
- Vancov, T., Alston, A.-S., Brown, T. and McIntosh, S. (2012). Use of ionic liquid in converting lignocellulosic material to biofuels. *Renewable Energy*, 45: 1-6.
- Ververis, C., Georgio, K., Christodoulakis, N., Santas, P. and Santas, R. (2004). Fiber dimensions, lignin and cellulose content of various plant materials and their solubility for paper production. *Industrial Crops and Products* 19:245-254.

- Wan, C. and Li, W. (2012). Fungal pretreatment of lignocellulosic biomass. *Biotechnology Advances*, 30: 1447-1457.
- Wang, H. Gurau, G. and Rogers, R.D. (2011) Ionic liquid processing of cellulose. *Chem. Sov. Rev.* 41:1519-1537.
- Wang, Z., Keshwani, D.R., Redding, A.P. and Cheng, J.J. (2010). Sodium hydroxide pretreatment and enzymatic hydrolysis of coastal Bermuda grass. *Bioresource Technology*, 101: 3583-3585.
- Wanrosli, W.D., Zainuddin, Z., Law, K.N. and Asro, R. (2007). Pulp from oil palm fronds by chemical processes. *Industrial Crops and Products* 25:89-94.
- Wei, W., Wu, S. and Liu, L. (2012). Enzymatic saccharification of dilute acid pretreated eucalyptus chips for fermentable sugar production. *Bioresource Technology*, 110: 302-307.
- Wildenborg, T. and Lokhorst, A. (2005). Introduction on CO₂ geological storage-classification of storage options, *Oil Gas Science Technology-Review*, 60: 513-515.
- William, E.K. and Holtzapple, M.T. (2000). Using lime pretreatment to facilitate the enzymic hydrolysis of corn stover. *Biomass and Bioenergy*, 18: 189-199.
- Wise, L.E., Murphy, M. and D'Addieco, A.A. (1946). Chlorite holocellulose, its fractionation and bearing on summative wood analysis and on studies on the hemicelluloses. *Paper Trade Journal*, 122: 35-43.
- Wu, Y., Sasaki, T., Kazushi, K., Seo, T. and Kensuke, S. (2008). Interactions between spiropyrans and room-temperature ionic liquids: photochromism and solvatochromism. *The Journal of Physical Chemistry B*, 112: 7530-7536.
- Xiao, W., Yin, W., Xia, S. and Ma, P. (2012). The study of factors affecting the enzymatic hydrolysis of cellulose after ionic liquid pretreatment. *Carbohydrate Polymers*, 87: 2019-2023.
- Xu, Z., Wang, Q., Jiang, Z., Yang, X.-X. and Ji, Y. (2007). Enzymatic hydrolysis of pretreated soybean straw. *Biomass and Bioenergy*, 31: 162-167.
- Xu, Z., Wang, Q., Jiang, Z.H., Yang, X.-x. and Ji, Y. (2007). Enzymatic hydrolysis of pretreated soybean straw. *Biomass and Bioenergy* 31:162-167.
- Yang, B. and Wyman, C.E. (2008). Pretreatment: the key to unlocking the low-cost cellulosic ethanol. *Biofuels, Bioproducts and Biorefining* 2:26-40.
- Yeh, A.-I., Huang, Y.-C. and Chen, S.H. (2010). Effect of particle size on the rate of enzymatic hydrolysis of cellulose. *Carbohydrate Polymers*, 79: 192-199.
- Yusoff, S. (2006). Renewable energy from palm oil-innovation on effective utilization of waste. *Journal of Cleaner Production*, 14: 87-93.

- Zhang, Q., Chang, J., Wang, T. and Xu, Y. (2007). Review of biomass pyrolysis oil properties and upgrading research. *Energy Conversion and Management*, 48: 87-92.
- Zhang, Q., Vigier, K.D.E., Royer, S. and Jerome, F. (2012). Deep eutectic solvents: syntheses, properties and applications. *Chemical Society Reviews*, 41: 7108-7146.
- Zhang, X., Yu, H., Huang, H. and Liu, Y. (2007). Evaluation of biological pretreatment with white rot fungi for the enzymatic hydrolysis of bamboo culms. *International Biodeterioration and Biodegradation* 60:19-164.
- Zhang, Y.-H.P. and Lynd, L.R. (2004). Toward an aggregated understanding of enzymatic hydrolysis of cellulose: Noncomplexed cellulase systems. *Biotechnology and Bioengineering*, 88:797-824.
- Zhao, H., Baker, G.A. and Holmes, S. (2011). New eutectic ionic liquids for lipase activation and enzymatic preparation of biodiesel. *Organic Biomolecular Chemistry*, 9: 1908-1916.
- Zhao, H., Jones, C.L., Baker, G.A., Xia, S., Olubajo, O. and Person, V.N. (2009). Regenerating cellulose form ionic liquids for an accelerated enzymatic hydrolysis. *Journal of Biotechnology*, 139: 47-54.
- Zhao, H., Zhang, C. and Crittle, T.D. (2013). Choline-based deep eutectic solvents for enzymatic preparation of biodiesel from soybean oil. *Journal of Molecular Catalysis B: Enzymatic*, 85-86: 243-247.
- Zhao, X., Xang, L. and Liu, D. (2012). Biomass recalcitrance. Part 1: the chemical compositions and physical structures affecting the enzymatic hydrolysis of lignocellulose. *Biofuels, Bioproducts and Biorefining*, 6:465-482.
- Zhu, J.Y., Gleisner, R., Scott, C.T., Luo, X.L. and Tian, S. (2011). Higher titer ethanol production from simultaneous enzymatic saccharification and fermentation of aspen at high solids: A comparison between SPORL and dilute acid pretreatment. *Bioresource Technology*, 102: 8921-8929.