



UNIVERSITI PUTRA MALAYSIA

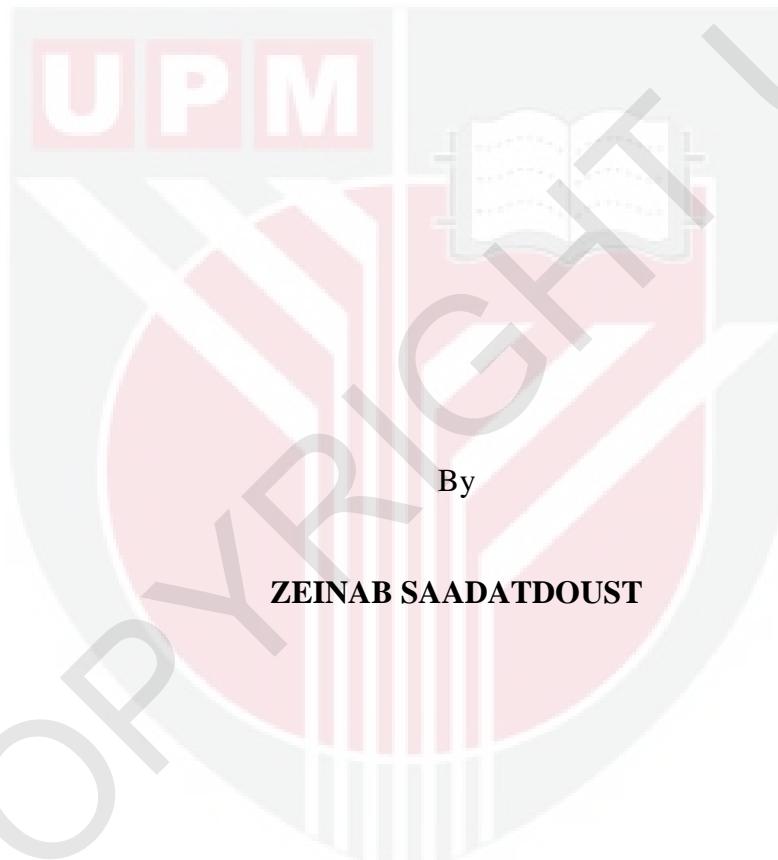
EFFECTS OF COCOA POLYPHENOL RICH DIET IN PREVENTION OF COLITIS-ASSOCIATED COLON CANCER

ZEINAB SAADATDOUST

FPSK(M) 2015 78



**EFFECTS OF COCOA POLYPHENOL RICH DIET IN PREVENTION OF
COLITIS-ASSOCIATED COLON CANCER**



Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfillment of the Requirements for the Degree of Master of Science

November 2015

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright© Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment
of the requirement for the Degree of Master of Science

**EFFECTS OF COCOA POLYPHENOL RICH DIET IN PREVENTION OF
COLITIS-ASSOCIATED COLON CANCER**

By

ZEINAB SAADATDOUST

November 2015

Chairman : Associate Professor Norhaizan Mohd Esa, PhD
Faculty : Medicine and Health Sciences

Colorectal cancer (CRC) is the second highest mortality and the third most diagnosed in both men and women. Colitis-associated cancer is a subtype of CRC that is associated with inflammatory bowel disease. Cocoa has a rich source of polyphenols and inhibits the cancer cell proliferation and decrease the risk of different type of cancers, cardiovascular disease and diabetes. This study was aimed to determine the anti-cancer effects of cocoa rich diets on dextran sulfate sodium (DSS) and azoxymethane (AOM)-induced colitis associated cancer in BALB/c mice. Natural Forastero cocoa powder was used for this study and diets were prepared from an AIN-93G formulation. The 5% and 10% cocoa diets are produced by adding 50 g/kg and 100 g/kg cocoa to AIN-93G at the expense of starch, cellulose and casein. The total polyphenol content of the cocoa powder was determined. Cocoa rich diet was modified to supplement 1 g and 2 g of polyphenols per kg of diet respectively. Total number of 50 female BALB/c mice (*Mus musculus*) weighing 25-30 g were divided into 5 different groups and each group consist of 10 mice. Data are presented as mean (n = 10 mice per group). The mice in groups 2, 3 and 4 were initiated by a single intraperitoneal (*i.p.*) injection of AOM (10 mg/kg body weight). Starting 1 week after the injection, 2% DSS in drinking water was administrated to mice of group 2, 3 and 4 for 7 days and 14 days and followed by normal drinking water for the recovery period. Totally 3 cycles of 2% DSS were treated. Group 1 (control) and Group 2 received AIN-93G diet, group 3 and 4 were treated with cocoa diet of 5% and 10%, respectively. Group 5 treated with 10% of cocoa diet alone to assess the toxicity of cocoa. On day 62 of the experiment, all mice were sacrificed and the entire colon and rectum were processed for histopathology examination and further evaluation.

Pro-inflammatory mediators and cytokines were measured by enzyme-linked immunohistochemical assay; real-time-PCR and western blot analysis. The tissue samples were examined for ultrastructural changes in experimental mice by Transmission Electron Microscopy. Change in colon length in mouse model of colitis-associated cancer was significantly improved in animals receiving cocoa-

enriched diet. Spleen weight was significantly decreased in animals treated with cocoa diet ($P<0.05$). Colon tumor number was increased upon DSS/AOM administration and fed with cocoa-enriched diet showed reduced number of tumors/mice. Cocoa diet modulates histological alterations caused by AOM/DSS. Control and cocoa diet alone treated group of mice shown normal architecture of microvilli. AOM/DSS treated mice showing the invasive gland in the submucosa layer of a large size adenoma. Treatment 5% and 10% of cocoa-enriched diet showed small polyps in the muscular layer.

In AOM/DSS group of animal, increased expressions of iNOS and COX-2 was observed by immunohistochemistry. However, treatment with 5% and 10% cocoa diet showed decreased expression of iNOS and COX-2, whereas control and cocoa alone groups showed fewer positive expressions. Deregulation of the JAK/STAT3 signaling pathway has also been implicated in colorectal tumorigenesis resulting in accumulation of cytokines and growth factors, Janus kinases.. Therefore, the potential of polyphenols in cocoa powder in targeting key components of the STAT3 signaling pathway along with iNOS and COX2 as a rational for cancer drug discovery was demonstrated. Colon tumors were further analyzed the mRNA levels of pro-inflammatory cytokines such as IL-6, TNF- α , IL-1 β , IL-17 by RT-PCR and Bcl-xL, Bax, Caspase 3 and Caspase 8 at protein levels by Western Blot analysis. It was shown that administration of cocoa significantly down-regulated inflammatory factors in colon cancer animal as compared with control (no cocoa treatment) ($p<0.05$).

In summary, the ability of cocoa to prevent the development of the colon carcinogenesis was demonstrated by lower tumor incidence, number and size of DSS/AOM-treated mice. In this present study, shown that after cocoa-enriched diet, the colitis presented a statistical improvement and tumors burden decreased significantly, this was accompanied by lower activity of p-STAT3 Y705 , decreased expressions of COX-2 and iNOS, lower expression of cytokines in the colons of CAC mice. We suggest that the chemopreventive effect of cocoa enriched diet on colitis-associated carcinogenesis could be mediated mainly through the IL-6/STAT3 pathway. Taken together, the present data provide evidence that cocoa polyphenols would offer a natural approach to improve individual health status including the prevention of colonic inflammation with no toxicity.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Master Sains

KESAN DIET KOKO KAYA POLIFENOL DALAM MENGHALANG KANSER KOLON BERKAIT KOLITIS

Oleh

ZEINAB SAADATDOUST

November 2015

Pengerusi : Profesor Madya Norhaizan Mohd Esa, PhD
Fakulti : Perubatan dan Sains Kesihatan

Kanser kolorektal (CRC) adalah mortaliti yang kedua tertinggi dan ketiga paling banyak didiagnosis di kalangan lelaki dan juga wanita. Kanser berkait kolitis adalah subjenis CRC yang berkaitan dengan penyakit radang usus. Ia sukar dirawati dan mempunyai mortaliti yang tinggi. Di tahun kebelakangan ini, penggunaan bahan semulajadi untuk rawatan pelbagai penyakit keradangan di kalangan pesakit yang mengalami kesan sampingan akibat rawatan dadah, semakin meningkat. Polifenol tumbuhan yang merupakan komponen utama di dalam diet manusia mempunyai pelbagai kesan kebaikan. Polifenol telah difikirkan mempunyai ciri-ciri anti-oksidatif dan anti-keradangan tersendiri. Koko adalah sumber yang kaya dengan polifenol dan menghalang proliferasi sel kanser dan mengurangkan risiko pelbagai jenis kanser, penyakit kardiovaskular dan diabetes. Kajian ini bertujuan untuk menentukan kesan anti-kanser diet koko kaya polifenol terhadap mencit BALB/c yang diaruh kanser kolon berkait kolitis oleh dekstran sulfat sodium (DSS) dan azoksimetana (AOM).

Serbuk koko ‘Natural Forastero’ digunakan untuk kajian ini dan diet disediakan dengan formulasi AIN-93G. Diet koko 5% dan 10% dihasilkan dengan menambahkan 50 g/kg dan 100 g/kg koko ke dalam diet AIN-93G dengan mengambilkira kanji, selulosa dan kasein. Kandungan jumlah polifenol serbuk koko ditentukan dengan kaedah spektrofotometrik *Folin – Ciocalteu* menggunakan epicatechin sebagai piawaian. Diet koko kaya polifenol diubahsuai untuk memberikan 1 g dan 2 g polifenol per kg diet, masing-masing sebagai suplemen. Sejumlah 50 ekor mencit BALB/c (*Mus musculus*) betina dengan berat 25-30 g dibahagikan kepada 5 kumpulan berlainan dan setiapnya mengandungi 10 ekor mencit. Data dibentangkan sebagai min (n = 10 ekor mencit setiap kumpulan). Mencit di kumpulan 2, 3 dan 4 diberi satu suntikan awal *intraperitoneal* (i.p.) AOM (10 mg/kg berat badan). Bermula satu minggu selepas suntikan, 2% DSS dalam air minuman diberi kepada mencit di kumpulan 2, 3 dan 4 selama 7 hari dan 14 hari, diikuti dengan air minuman biasa untuk tempoh pemulihan. Sejumlah 3 kitaran 2% DSS diberi. Kumpulan 1 (kawalan) dan kumpulan 2 menerima diet AIN-93G, kumpulan 3 dan 4 dirawat dengan diet koko 5% dan 10%, masing-masing.

Kumpulan 5 diberi diet koko 10% sahaja, untuk menilai ketoksikan koko. Pada hari ke 62 ujikaji, mencit dibunuh dan keseluruhan kolon, rektum diproses untuk penilaian histopatologi dan penilaian lain seterusnya.

Pengantara pro-keradangan dan sitokin diukur dengan asai *enzyme-linked immunohistochemical, real-time-PCR* dan analisis *Western Blot*. Sampel tisu dari ujikaji mencit ditentukan perubahan ultrastruktur menggunakan *Transmission Electron Microscopy*. Perubahan panjang kolon di dalam model mencit kanser berkait kolitis diperbaiki dengan signifikan pada kumpulan yang menerima diet kaya koko. Berat limpa berkurangan dengan signifikan pada mencit yang dirawat dengan diet koko ($p<0.05$). Bilangan tumor kolon meningkat dengan pemberian DSS/AOM dan dengan memberi diet koko kaya polifenol menunjukkan penurunan bilangan tumor/mencit. Diet koko, modulasi perubahan histologi yang disebabkan oleh AOM/DSS. Mencit dalam kumpulan kawalan (normal) dan yang diberi diet koko sahaja menunjukkan struktur microvili yang normal. Mencit yang diberi AOM/DSS menunjukkan kelenjar yang invasif di dalam lapisan submukosa adenoma bersaiz besar. Rawatan dengan diet kaya koko, 5% dan 10% menunjukkan polip yang kecil di dalam lapisan berotot.

Analisis immunohistokimia *cyclooxygenase-2* (COX-2) dan *inducible nitric oxide synthase* (iNOS) dilakukan ke atas haiwan kumpulan kawalan dan ujikaji. Bagi haiwan dalam kumpulan yang diberi AOM/DSS, didapati ekspresi iNOS dan COX-2 meningkat. Walaubagaimanapun, rawatan dengan diet koko 5% dan 10% menunjukkan ekspresi iNOS dan COX-2 adalah lebih rendah. Manakala kumpulan kawalan (normal) dan menerima koko sahaja menunjukkan ekspresi yang sedikit. Penyah-regulasi tapakjalan isyarat JAK/STAT3 juga mempunyai implikasi didalam tumorigenesis kolorektal yang mengakibatkan pengumpulan sitokin dan faktor tumbesaran, Janus kinases. Tambahan pula, ekspresi enzim berkait dengan keradangan seperti iNOS dan COX-2, ditunjukkan memainkan peranan yang penting di dalam perkembangan tumor kolon. Oleh itu, potensi polifenol di dalam serbuk koko dalam mensasarkan komponen utama tapak jalan isyarat STAT3 bersama iNOS dan COX2 sebagai penemuan ubat kanser yang rasional, telah ditunjukkan. Tumor kolon dianalisis lebih lanjut diperangkat mRNA sitokin pro-keradangan seperti IL-6, TNF- α , IL-1 β , IL-17 melalui *RT-PCR* and protein Bcl-xL, Bax, Caspase 3 dan Caspase 8 dianalisis dengan *Western Blot*. Keputusan menunjukkan bahawa pemberian koko *down-regulate* faktor keradangan di dalam haiwan yang mempunyai kanser kolon dengan signifikan ($p<0.05$) berbanding dengan kumpulan kawalan yang tidak menerima rawatan koko. Ini mencadangkan bahawa kesan chemopreventif diet kaya koko terhadap karsinogenesis berkait kolitis mungkin diperantarai oleh tapak jalan IL-6/STAT3.

Kesimpulannya, kebolehan koko untuk mencegah perkembangan karsinogenesis kolon pada mencit yang diberi dengan DSS/AOM ditunjukkan melalui pelemahan keradangan kolorektal dan pengurangan kejadian, bilangan dan saiz tumor. Kajian yang dijalankan ini juga menunjukkan bahawa selepas diet kaya koko diberikan, keadaan kolitis adalah lebih baik dengan sifnifikan dan ini disertai dengan aktiviti p-STAT3 Y705 yang lebih rendah, pengurangan ekspresi COX-2 dan iNOS, merendahkan ekspresi sitokin pro-radangan pada kolon mencit CAC. Kami mencadangkan kesan chemopreventif diet kaya koko keatas karsinogenesis berkait kolitis, mungkin diperantarai oleh tapak jalan IL-6/STAT3. Keseluruhannya, data

yang diperolehi ini membuktikan bahawa polifernol koko menawarkan pendekatan semulajadi untuk memperbaiki status kesihatan individu termasuklah menghalang keradangan kolon tanpa memberikan kesan toksik dan juga mempunyai potensi sebagai diet yang menghalang keradangan kolon dan perkembangan kanser berkaitan.



ACKNOWLEDGEMENTS

Firstly, I would like to express my special appreciation to my main supervisor Assoc. Prof. Dr. Norhaizan Mohd Esa, She has been a wonderful mentor for me. I would like to thank you for encouraging my research. Her advice on both research as well as on my career have been priceless. I am forever thankful to her for all the opportunities that she has given me to excel in my field of research.

I also would like to thank my committee member Prof. Amin Ismail for his guidance and for his timely and unconditional support.

Special thanks to Dr. Ashok Kumar Pandurangan for being there, whenever I had questions or I was in need, his expert guidance has been the most helpful and appreciated. I also want to thank him for allowing me to grow as a research scientist and for his billions comments and suggestions.

Also, I would like to take this opportunity to acknowledge my best friends, especially Maryam Kheirollahpour for being there during my critical preparation for thesis submission. Thank you

Lastly, I want to give my most heartfelt thanks to my sister, Ms Zohre Saadatdoust and the rest of my family for all the endless encouragement, love and support.

This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Norhaizan Mohd Esa, PhD

Associate Professor

Faculty of Medicine and Health Science

Universiti Putra Malaysia

(Chairman)

Amin Ismail, PhD

Professor

Faculty of Medicine and Health Science

Universiti Putra Malaysia

(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: _____

Name of Chairman
of Supervisory
Committee: Associate Professor Dr. Norhaizan Mohd Esa

Signature: _____

Name of Member
of Supervisory
Committee: Professor Dr. Amin Ismail

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	vi
APPROVAL	vii
DECLARATION	ix
LIST OF TABLES	xv
LIST OF FIGURES	xvi
LIST OF ABBREVIATIONS	xviii
 CHAPTER	
1 INTRODUCTION	1
1.1 Research Background	1
1.2 Problem Statement	2
1.3 Significance of Study	2
1.4 Objectives	4
1.4.1 General Objective	4
1.4.2 Specific Objectives	4
2 LITERATURE REVIEW	5
2.1 Cancer	5
2.1.1 Definition of Cancer	5
2.1.2 Mutation and Carcinogenesis	6
2.2 Colon Carcinogenesis	7
2.2.1 Colon Cancer Symptoms	7
2.2.2 Colon Cancer Prevalence	8
2.2.3 Risk Factors Involved in Colon Cancer	8
2.2.3.1 A Personal History of Colorectal Cancer or Polyps	9
2.2.3.2 Age	9
2.2.3.3 Diet	9
2.2.3.4 Alcohol	9
2.2.3.5 Obesity	9
2.2.3.6 Diabetes	9
2.2.3.7 Smoking	9
2.2.3.8 Inflammatory Intestinal Conditions	10
2.2.3.9 A Sedentary Lifestyle	10
2.2.3.10 Growth Hormone Disorder	10
2.2.3.11 Radiation Therapy for Cancer	10
2.2.3.12 Inherited syndromes:	10
2.3 Description of Inflammation	10
2.3.1 Acute Inflammation	11
2.3.2 Chronic Inflammation	12
2.3.3 Inflammation Bowl Disease	13
2.3.4 Ulcerative Colitis	14
2.4 Colitis Associated Colon Cancer:	15

2.4.1	Stage of Colon Cancer in Ulcerative Colitis	16
2.5	Immune System	18
2.5.1	JAK/STAT3 Signalling and Colon Cancer	18
2.5.2	Proinflammatory Cytokines	20
2.5.3	IL-6 and STAT3	21
2.5.4	Interleukin-17	22
2.5.5	TNF alpha	22
2.6	Apoptosis	23
2.6.1	Apoptosis Signaling Pathway	24
2.6.2	Apoptosis and Tumorigenesis	24
2.6.3	The Bcl-2 family of apoptotic regulators	25
2.7	NF-kB and Colorectal Cancers:	25
2.8	Cyclooxygenases and Colon Cancer	26
2.9	Role of Nitric Oxide in Colon Cancer	27
2.9.1	Inhibitors of iNOS in colon cancer	28
2.10	Chemical Carcinogenesis	28
2.10.1	Dimethyl Hydrazine (DMH) and Azoxymethane (AOM)	28
	2.10.1.1 Dextran Sulfate Sodium (DSS)	30
2.11	Animals Model to Study of Colon Cancer	30
2.12	Natural Antioxidant	31
2.13	Polyphenol as a Bio-Active Compound	32
2.13.1	Metabolism and Bioavailability of Polyphenols	33
	2.13.1.1 Main Factors Affecting the:Bioavailability of the Polyphenols	33
2.13.2	Polyphenols and Carcinogenesis:	35
2.13.3	Effect of Polyphenols on Inflammatory Mediators	37
2.14	Cocoa (<i>Theobroma Cacao L</i>)	37
2.14.1	Background of Cocoa:	37
2.14.2	Bioavailability of Phenolic from Cocoa:	38
	2.14.2.1 Gastric Degradation of Cocoa Phenolics.	38
	2.14.2.2 Intestinal Absorption of Phenols from Cocoa.	38
2.14.3	Cocoa Polyphenols and Their Potential Benefits for Human Health	39
2.14.4	Phenolic from Cocoa in Cancer Prevention:	39
	2.14.4.1 Anti-proliferative Effects:	40
	2.14.4.2 Effects of Cocoa Phenolics on the Immune System	40
3	METHODOLOGY	41
3.1	Materials	41
3.1.1	Cocoa Powder	41
3.1.2	Chemicals and Reagents	41
3.1.3	Instrument	41
3.2	Methods	42
3.2.1	Determination of Total Polyphenols Content in Cocoa Powder	42
	3.2.1.1 Sample Preparation	42
	3.2.1.2 Total Phenolic Assay (TPC)	42

3.2.2	Preparation of Cocoa Powder Diet	43
3.2.3	<i>In Vivo Study:</i>	44
3.2.3.1	Animals	44
3.2.3.2	Ethical Approval	44
3.2.3.3	Carcinogen Induction	44
3.2.3.4	Experimental Design	44
3.2.3.5	Body Weight Measurement:	46
3.2.3.6	Cage Side Observation	46
3.2.3.7	Toxicity Study	46
3.2.3.7.1	Analysis of kidney and liver Function by Clinical Chemistry Analysis	46
3.2.3.7.2	Pathology	46
3.2.3.8	Histological Analysis of Colon	46
3.2.3.9	Transmission Electron Microscopy of Colonic Tissue	47
3.2.3.10	Immunohistochemistry	47
3.2.3.11	Gene expression	48
3.2.3.11.1	Extraction of total RNA	48
3.2.3.11.2	Quantification of RNA Samples	49
3.2.3.11.3	Quantitative Real-time Polymerase Chain Reaction Analysis	49
3.2.3.11.3.1	Specific Primer for qRT PCR	49
3.2.3.11.3.2	Real-Time PCR Reactions	50
3.2.3.11.3.3	Real-Time PCR Reactions	50
3.2.3.11.3.4	Plate Setup and qRT-PCR Reactions	51
3.2.3.11.3.5	Real-Time PCR Data Analysis	51
3.2.3.12	Western Blot Analysis	52
3.2.3.12.1	Preparation of Tissue Sample	52
3.2.3.12.2	Quantification of Protein Samples	52
3.2.3.12.3	Sodium Dodecyl Sulfate-Polyacrylamide Gel Electrophoresis (SDS-PAGE)	52
3.2.3.12.4	Western Blotting	54
3.2.3.12.5	Densitometry	54
3.2.4	Statistical analysis	54
4	RESULTS AND DISCUSSION	55
4.1	Determination of Total Polyphenols Compound	55
4.2	Food Consumption	57
4.3	General Observations	57

4.4	Toxicity Evaluation of Cocoa Powder at Dietary Level of 10%	60
4.4.1	Cage Observation	61
4.4.2	Clinical Chemistry Analysis	62
4.4.3	Histopathology Analysis	63
4.5	Tumor Burden	65
4.6	Histopathology of Colonic Neoplasms Developed in Mice	66
4.7	Effect of Cocoa Rich Diet on Ultrastructure Changes of Colon	68
4.8	Cocoa Attenuates the RT-PCR Expression of Pro-Inflammatory Cytokines	70
4.9	Cocoa Reduced the Expression of NF- κ B	71
4.9.1	Effect of Cocoa Diet on The Expression of iNOS During AOM/DSS-Induced CAC	73
4.9.2	Reduction of the Immunohistochemical Expression of iNOS by Cocoa	73
4.10	Cocoa Reduced the RT-PCR Expression of iNOS	74
4.11	Effect of Cocoa Diet on the Expression of Cyclooxygenase-2(COX2) During AOM/DSS-Induced CAC	75
4.11.1	Down-regulation of COX-2 Expression in AOM/DSS Induced CAC by cocoa	75
4.11.2	Reduction of the RT-PCR Expression of COX-2 by Cocoa	77
4.12	Down-Modulatory Effects of Cocoa on iNOS and COX2 in AOM/DSS Induced Mice CAC	77
4.13	Effect of Cocoa Diet on the Expression of P-STAT3 Y705 During AOM/DSS-Induced CAC	78
4.13.1	Cocoa Reduced the Expression of P-STAT3 Y705 in AOM/DSS Induced CAC	78
4.13.2	Reduction of P-STAT3 Y705 Protein Expression After Treatment With Cocoa Diet In AOM/DSS Induced CAC In Mice.	79
4.14	Effect Of Cocoa Rich Diet on The Apoptosis Related Genes Such As Bcl-xL, Bax, Caspase 3 In AOM/DSS Induced CAC in Mice.	81
5	SUMMARY AND RECOMMENDATION	85
5.1	Conclusion	85
5.2	Limitations	86
5.3	Recommendation	86
REFERENCES		88
APPENDICES		117
BIODATA OF STUDENT		122
LIST OF PUBLICATIONS		123

LIST OF TABLES

Table	Page
2.1 Environmental Factor Which Can Regulate Colorectal Cancer Hazard	8
2.2 Main Features of Colitis-Associated Cancer and Colorectal Cancer	18
2.3 Main Factors Affecting the Bioavailability of Dietary Polyphenols in Humans.	33
3.1 Composition of the Experimental Control and Cocoa-Rich Diets	43
3.2 List of Primers Used	50
3.3 qRT-PCR reaction Mix	51
3.4 Real-Time PCR Cycling Protocol	51
3.5 Resolving Gel (10%) Mixture	53
3.6 Stacking Gel (6%) Mixture	53
4.1 Daily Food Intake of Mice	57
4.2 Organ Weights of Mice Treated with 10% Cocoa-Rich Diet for 62 Days.	62
4.3 Biochemical Parameters of Mice Treated with Cocoa Rich Diet for 62 days.	63
4.4 Incidence of Colonic Neoplasm	65

LIST OF FIGURES

Figure		Page
2.1	The Stage of the Carcinogenic Process	7
2.2	The Role of Chronic Inflammation in Tumor Initiation	14
2.3	Colitis-associated Cancer (CAC) and Colorectal Cancer (CRC) Mechanisms Development	17
2.4	Regulation of Intracellular STAT3	19
2.5	Cytokines and Transcription Factors Involved in Tumor Promotion	21
2.6	The Arachidonic Acid Cascade	26
2.7	Immunohistochemical Analysis of iNOS in the Tumor of Colon Cancer Induced with Azoxymethane	28
2.8	Macroscopic Observation of Colon Tumors	29
2.9	Mechanism of Induction of Colon Cancer by Azoxymethane	30
2.10	Classification of Polyphenols	32
2.11	The Absorption of Dietary Polyphenols in Human	35
2.12	Multistage Model of Carcinogenesis and Potential Effect of Polyphenols on Cancer Progression	36
3.1	Experimental Protocol for Colitis-Associated Colon Carcinogenesis Model	45
4.1	Standard Curve for Different Concentration of Epicatechin ($\mu\text{g/ml}$)	56
4.2	Change in Body Weight of Experimental Mice	58
4.3	Effect of Cocoa Rich Diet on Colon Length in Control and Treatment Mice	59
4.4	Effect of Cocoa Rich Diet on Spleen Weight in Control and Treatment Mice	60
4.5	Kidney of Control Rat Showing Normal Glomeruli and Renal Tubules	64

4.6	Bright Field Image of an H&E Stained Section Showing Normal Liver Architecture, and Components of Basic Liver Lobules, with Central Venua	64
4.7	Colon Reduces Tumor Number and Size	66
4.8	Histopathological Analysis of Colon in Control and Experimental Groups of Animals	67
4.9	Transmission Electron Microscopic Analysis of Colon Tissue in Control and Experimental Mice	69
4.10	Cocoa Attenuates Pro-Inflammatory Cytokines	70
4.11	Cocoa Reduced the Expressions of NF- κ B	72
4.12	Cocoa Reduced the Expression of iNOS in AOM/DSS Induced CAC	74
4.13	Expression of iNOS at mRNA Levels of AOM/DSS Induced the CAC	75
4.14	Cocoa Reduce the Expression of COX-2 in AOM/DSS Induced CAC	76
4.15	Expression of COX-2 at mRNA Levels of AOM/DSS Induced CAC after Treatment with Cocoa	77
4.16	Cocoa Reduced the Expression of p-STAT3 Y705	79
4.17	The Effect of Cocoa on Protein Level of p-STAT3 Y705 in AOM/DSS Induced CAC in Mice	80
4.18	Effect of Cocoa on Bcl-xL, Bax	82
4.19	Effect of Cocoa on Apoptotic Protein Levels of Caspase-3 in AOM/DSS Induce CAC in Mice	83
5.1	Summarizing Diagram Illustrating Possible Mechanism by which Cocoa Rich Diet Exerts an Anti-Cancer Effects in Colon Cancer.	86

LIST OF ABBREVIATIONS

Abbreviation	Full Term
%	Percentage
°C	Degree Celsius
ΔΔCt	Delta-delta threshold cycle
ΔΨm	Mitochondrial membrane potential
μg	Microgram
ALT	Alanine aminotransferase
AST	Aspartate aminotransferase
AOM	Azoxymethane
Bcl-2	B-cell lymphomomal/leukemia
BSA	Bovine serum albumin
CAC	Colitis associated colon cancer
COX-2	Cyclooxygenase-2
CRC	Colorectal cancer
Chol	Cholesterol
Creat	Creatinine
DNA	Deoxyribionucleic Acid
DSS	Dextran sulfate sodium
dH2O	Distilled Water
FBS	Foetal Bovine Serum
H&E	Hematoxylin and Eosin
IL-1β	Interleukin 1 beta
IL-6	Interleukin 6
IL-17	Interleukin 17
iNOS	Inducible isoform of nitric oxid synthase
kDa	Kilo dalton
kb	Kilo base pair
mg	Miligram
mg/dl	Miligram/decilitre
mg/kg	Miligram/kilogram
mRNA	Messenger Ribonucleic Acid
NaCl	Sodium chloride
NAOH	Sodium Hydroxide
NF-κB	Nuclear Factor κB
NOAEL	No-observable adverse effect levels
PBS	Phosphate Bovine Serum,
PCR	Polymerase chain Reaction

SDS-PAGE	Sodium-dodecyl Sulfate Polyacrylamide Gel
PG	Prostaglandin
PGE2	Prostaglandin endoperoxide synthase2
p-STAT3	Phosphorylated STAT3
PVDF	Polyvinylidene difluoride
qPCR	Quantitative Real –time polymerase chain reaction
qRT-PCR	Quantitative real-time reverse transcriptase polymerase Chain Reaction
RNA	Ribonucleic Acid
SD	Standard Deviation
TBA	Thiobarbituric Acid
TBST	Tris Buffer saline and twin 20
TEM	Transmission Electron Microscopy
TEMED	Tetramethylethylenediamine
TNF	Tumor Necrosis factor
TNF- α	Tumor Necrosis factor -alpha
WHO	World Health Organization

CHAPTER ONE

INTRODUCTION

1.1 Research Background

Colon cancer is the third most common cancer and the second leading cause of cancer death among both men and women in the United States (Zeller et al., 2008). It is responsible for around 700,000 deaths per year, worldwide (Lim & Halimah, 2003). Colon cancer, also known as colorectal cancer (CRC), is currently one of the most common cancers in Malaysia. According to Ministry of health, colorectal cancer is the second disease causing death after heart disease in Malaysia, and contributes 9.23% of total death cases (Balraj & Ruhana, 2007). Epidemiological and experimental studies have shown that patients with inflammatory bowel disease (IBD) are at a greater risk of developing CRC than the general population, and colitis-associated cancer (CAC) is the major cause of death in inflammatory IBD patients (Eaden et al., 2001).

CAC is the type of colon cancer which is preceded by clinically detectable IBD, such as Crohn's disease (Caprioli et al., 2008) or ulcerative colitis (UC) (Feagins et al., 2009; Rubin et al., 2012; Saleh & Trinchieri, 2010). UC increases cumulative risk of CAC by up to 18–20 %, while CD by up to 8 % after 30 years of active disease (Canavan et al., 2006; Eaden et al., 2001; Rubin et al., 2012). One of the important underlying etiologies of carcinogenesis in the colon is inflammation (Fran Balkwill & Mantovani, 2001). The microenvironment of chronic intestinal inflammation facilitates cell proliferation, migration and angiogenesis, thereby promotes tumor development, growth and progression (Boland et al., 2005). In particular, the incidence of colitis-associated cancer (CAC) approaches to ~40% in patients with colitis (Munkholm, 2003).

Polyphenols are phytochemicals derived from phenylalanine and contain an aromatic ring with a reactive hydroxyl group (Signorelli & Ghidoni, 2005). Polyphenols have gained much interest recently due to its antioxidant capacity and possible benefits to human health such as anti-carcinogenic, anti-atherogenic, anti-ulcer, anti-thrombotic, anti-inflammatory, immune modulating, anti-microbial, vasodilatory and analgesic effects (Hii et al., 2009). Furthermore, polyphenols, which constitute the active substances found in many medicinal plants, modulate the activity of a wide range of enzymes and cell receptors (Keen, 2001). Indeed, polyphenols have been reported to interfere with cancer initiation, promotion, and progression, acting as a strong chemopreventive agent (Araújo et al., 2011).

Cocoa (*Theobroma cacao L.*) consist rich source of polyphenols and cocoa beans has 6-8% total polyphenol (Ali et al., 2014). A reported by Vinson et al., (1999) had found phenolic compounds in cocoa are higher than 23 types of vegetables and several types of fruits. In this study the effect of combination of polyphenols on

colitis associated cancer was investigated since mixture of polyphenols may target overlapping and complementary phases of the carcinogenic process (de Kok et al., 2008) thus increasing the efficacy and potency of the chemopreventive effect.

1.2 Problem Statement

Colorectal cancer is a major cause of morbidity and mortality throughout the world. It accounts for over 9% of all cancer incidences. It is the third most common cancer worldwide and the fourth most common cause of death. It affects men and women almost equally, with just over 1 million new cases recorded in 2002 (Botteri et al., 2008). Countries with the highest incidence rates include Australia, New Zealand, Canada, the United States, and parts of Europe. The countries with the lowest risk include China, India, and parts of Africa and South America (Society, 2009). Worldwide, colorectal cancer represents 9.4% of all cancer incident in men and 10.1% in women. In Malaysia, colorectal cancer is the second most common cancer in males and females. A total of 2,246 cases were diagnosed in 2007 and reported to NCR, represent 12.3 % of all cases reported (Balraj & Ruhana, 2007).

Current treatment of colorectal cancer generally employs surgical resection combined with radiation or chemotherapy with synergistic cytotoxic drugs. Anti-inflammatory and also chemotherapeutic drugs have important effect for inflammation and cancer but they have a number of side-effects that can limit their efficacy (Wang et al., 2011). Moreover, the anti-inflammatory drugs in 2005 was cost around \$ 31.1 billion and this number is increaseable to reach \$ 47.8 in 2010 (Kearney et al., 2006). Hence, finding a suitable treatment for oxidative stress- and inflammation- related diseases with minimal or no side effects is still warranted.

Polyphenol has attracted many attentions because of its significant role in human health. Accumulating evidence showing that these compounds possess a high number of protective biologic properties such as antioxidant, anti-carcinogenic, anti-inflammatory (Scalbert et al., 2005), anti-allergic, anti-diarrheal, antiulcer, antibiotic (Howells et al., 2007) antilipidemic, vasorelaxing and antithrombotic properties (Scalbert et al., 2005). Because of these effects, polyphenols may confer protection against pathologies with very high incidence and mortality in occidental countries: cardiovascular and neurodegenerative diseases and cancer (Jemal et al., 2008). In relation to cancer, numerous case-control (Johnson, 2007) and animal and cell culture studies have corroborated a protector role of polyphenols and of foods and drinks that contain them (especially fruits and vegetables) in distinct cancer types (e.g., breast, lung, colon, stomach, esophagus, larynx, and oral cavity) (Jemal et al., 2008). Hence, examining the potent effect of Malaysian cocoa extract will enlarge using of cocoa as anti-oxidative or as anti-inflammatory agent.

1.3 Significance of Study

In the continuing effort to reduce the public health burden of cancer there is a constant search for more effective cancer treatment and increased interest in the

concept of prevention, as a promising approach to the control of cancer. A major target in current research is to identify cancer reduction strategies based on dietary modification including looking at natural sources that may have anticancer properties. Besides that, selective destruction of tumor cells without damaging normal cells is an important goal for cancer treatment (Keen, 2001).

Since the current treatment such as radiotherapy, chemotherapy and drugs possess unwanted side effects, the move to use potential bioactive compounds as the alternative should be made. Besides, in order to maintain full health as well as a broad range of nutraceutical compounds that has been demonstrated to have remarkable therapeutic properties, the opportunities to develop an alternative compound from the local source is the main reason why this study should be done and carried out successfully (Keen, 2001).

Cocoa is a rich source of bioactive compounds with potential chemopreventive ability but there are not enough studies that support its effectiveness in animal models of colon carcinogenesis. Rodríguez-Ramiro et al., (2011) have examined effect of cocoa rich diet on early levels of bowel tumorigenesis for the first time *in vivo*, so my research will provide the first *in vivo* evidence that cocoa-rich diet may inhibit the advance stage of colon carcinogenesis.

As compared with other flavonoid-containing foods, cocoa products exhibit a high concentration of procyanidins that are poorly absorbed in the intestine and consequently its beneficial effects would be more focused on the gastrointestinal tract where they may have an important local function neutralizing oxidants. Despite these evidences, the efficacy of cocoa against CRC initiation and development *in vivo* remains largely unexamined. Moreover, the effect of polyphenol from cocoa to inhibit the proliferation of colon cancer cell line (*in vitro*) has also been shown by Carnesecchi et al., (2002). However, we do not know whether it has similar effect in the body due to low bioavailability of polyphenol in our digestion system. So, this study was conducted *in vivo* to confirm this finding and also to know the possible mechanism how polyphenol acts as an anti-cancer agent.

In addition, Malaysia is one of the main cocoa-based product producer in the world and the biggest in Asia. However, Malaysian cocoa-based markets preferred cocoa beans/powder from African origins because cocoa are lacking in flavor. Although lack of flavor quality, a group of research based at University Putra Malaysia (UPM) has reported that Malaysian cocoa beans and its derived products could contribute toward decreasing of chronic diseases risk factors (Amin, Koh, et al., 2004; Othman et al., 2007). Through this study, it could be provide information that cocoa has a good potential in decreasing the risk of colon cancer. So, if the result from this study is able to show that Malaysian cocoa beans are able to prevent and treatment of colon cancer the beans then can be used as an alternative ways to reduce the incidence of colon cancer and hence boost the value of Malaysian cocoa beans. Therefore, in the present study, the chemopreventive activity of cocoa will be evaluated in the mouse model of azoxymethane (AOM)/ dextran sulfate sodium (DSS)-

induced colitis-associated cancer.

1.4 Objectives

1.4.1 General Objective

To investigate the protective effect of cocoa polyphenol rich diet from Malaysian cocoa powder on colitis associated colon cancer *in vivo*.

1.4.2 Specific Objectives

1. To determine the total amount of polyphenol and to evaluate chronic toxicity of Malaysian cocoa powder at dietary levels of 10%.
2. To analyze the effect of cocoa rich diet on incidence of tumor neoplasm, Histological alterations, Inflammatory mediators (iNOS and COX-2), pro-inflammatory cytokines ($TNF-\alpha$, $IL-1\beta$, $IL-6$, $IL-17$), and STAT3⁵ signaling pathway during AOM/DSS-induced colon cancer.
3. To quantify the expression of apoptotic proteins (Bax, Bcl-xL, caspase-3 and -8) induced by cocoa in colitis associated cancer.

REFERENCES

- Abraham, C., & Cho, J. (2009). Interleukin-23/Th17 pathways and inflammatory bowel disease. *Inflammatory Bowel Diseases*, 15(7), 1090-1100.
- Adachi, M., Kurotani, R., Morimura, K., Shah, Y., Sanford, M., Madison, B., Young, H. (2006). Peroxisome proliferator activated receptor γ in colonic epithelial cells protects against experimental *Inflammatory Bowel Disease*. *Gut*, 55(8), 1104-1113.
- Adams, J. M., & Cory, S. (2007). Bcl-2-regulated apoptosis: mechanism and therapeutic potential. *Current Opinion in Immunology*, 19(5), 488-496.
- Aggarwal, B. B., Kunnumakkara, A. B., Harikumar, K. B., Gupta, S. R., Tharakan, S. T., Koca, C., Sung, B. (2009). Signal transducer and activator of transcription-3, inflammation, and cancer. *Annals of the New York Academy of Sciences*, 1171(1), 59-76.
- Ahnen, D. J. (1999). Tissue markers of colon cancer risk. *Gastrointestinal Endoscopy*, 49(3), S50-S59.
- Alford, C. E., King, T. E., & Campbell, P. (1991). Role of transferrin, transferrin receptors, and iron in macrophage listericidal activity. *The Journal of Experimental Medicine*, 174(2), 459-466.
- Ali, F., Ismail, A., & Kersten, S. (2014). Molecular mechanisms underlying the potential antobesity-related diseases effect of cocoa polyphenols. *Molecular Nutrition and Food Research*, 58(1), 33-48.
- Ali, F., Ranneh, Y., Ismail, A., & Esa, N. M. (2013). Identification of phenolic compounds in polyphenols-rich extract of Malaysian cocoa powder using the HPLC-UV-ESI—MS/MS and probing their antioxidant properties. *Journal of Food Science and Technology*, 1-9.
- Ambs, S., Merriam, W. G., Bennett, W. P., Felley-Bosco, E., Ogunfusika, M. O., Oser, S. M., Harris, C. C. (1998). Frequent nitric oxide synthase-2 expression in human colon adenomas: implication for tumor angiogenesis and colon cancer progression. *Cancer Research*, 58(2), 334-341.
- Amin, I., Faizul, H., & Azli, R. (2004). Effect of cocoa powder extract on plasma glucose levels in hyperglycaemic rats. *Nutrition and Food Science*, 34(3), 116-121.
- Amin, I., Koh, B., & Asmah, R. (2004). Effect of cacao liquor extract on tumor marker enzymes during chemical hepatocarcinogenesis in rats. *Journal of Medicinal Food*, 7(1), 7-12.

- Andrä, J., Gutzmann, T., Garidel, P., & Brandenburg, K. (2006). Invited review: Mechanisms of endotoxin neutralization by synthetic cationic compounds. *Journal of Endotoxin Research*, 12(5), 261-277.
- Andújar, I., Recio, M., Giner, R., & Ríos, J. (2012). Cocoa polyphenols and their potential benefits for human health. *Oxidative Medicine And Cellular Longevity*, 2012.
- Andújar, I., Recio, M. C., Giner, R. M., Cienfuegos-Jovellanos, E., Laghi, S., Muguerza, B. a., & Ríos, J. L. (2011). Inhibition of ulcerative colitis in mice after oral administration of a polyphenol-enriched cocoa extract is mediated by the inhibition of STAT1 and STAT3 phosphorylation in colon cells. *Journal of Agricultural and Food Chemistry*, 59(12), 6474-6483.
- Araújo, J. R., Gonçalves, P., & Martel, F. (2011). Chemopreventive effect of dietary polyphenols in colorectal cancer cell lines. *Nutrition Research*, 31(2), 77-87.
- Aron, P. M., & Kennedy, J. A. (2008). Flavan-3-ols: Nature, occurrence and biological activity. *Molecular Nutrition and Food Research*, 52(1), 79-104.
- Arulselvan, P., Wen, C.-C., Lan, C.-W., Chen, Y.-H., Wei, W.-C., & Yang, N.-S. (2012). Dietary administration of scallion extract effectively inhibits colorectal tumor growth: cellular and molecular mechanisms in mice. *PloS One*, 7(9), e44658.
- Ashokkumar, P., & Sudhandiran, G. (2008). Protective role of luteolin on the status of lipid peroxidation and antioxidant defense against azoxymethane-induced experimental colon carcinogenesis. *Biomedicine and Pharmacotherapy*, 62(9), 590-597.
- Ashokkumar, P., & Sudhandiran, G. (2011). Luteolin inhibits cell proliferation during Azoxymethane-induced experimental colon carcinogenesis via Wnt/β-catenin pathway. *Investigational New Drugs*, 29(2), 273-284.
- Askling, J., Dickman, P. W., Karlén, P., Broström, O., Lapidus, A., Löfberg, R., & Ekbom, A. (2001). Family history as a risk factor for colorectal cancer in inflammatory bowel disease. *Gastroenterology*, 120(6), 1356-1362.
- Asquith, M., & Powrie, F. (2010). An innately dangerous balancing act: intestinal homeostasis, inflammation, and colitis-associated cancer. *The Journal of Experimental Medicine*, 207(8), 1573-1577.
- Atreya, I., & Neurath, M. F. (2008). Immune cells in colorectal cancer: prognostic relevance and therapeutic strategies. *Expert Review of Anticancer Therapy*, 8(4), 561-572.
- Atreya, R., Mudter, J., Finotto, S., Müllberg, J., Jostock, T., Wirtz, S., Becker, C. (2000). Blockade of interleukin 6 trans signaling suppresses T-cell resistance against apoptosis in chronic intestinal inflammation: evidence in crohn disease and experimental colitis in vivo. *Nature Medicine*, 6(5), 583-588.

- Atreya, R., & Neurath, M. (2008). Signaling molecules: the pathogenic role of the IL-6/STAT-3 trans signaling pathway in intestinal inflammation and in colonic cancer. *Current Drug Targets*, 9(5), 369-374.
- Augustyniak, A., Bartosz, G., Cipak, A., Duburs, G., Horáková, L. U., Luczaj, W., . Skrzypkiewska, E. (2010). Natural and synthetic antioxidants: an updated overview. *Free Radical Research*, 44(10), 1216-1262.
- Auletta, C. S. (1995). Acute, subchronic and chronic toxicology. *Handbook of Toxicology*, 51-162.
- Axelsson, L. G., Landström, E., & Bylund-Fellenius, A. C. (1998). Experimental colitis induced by dextran sulphate sodium in mice: beneficial effects of sulphasalazine and olsalazine. *Alimentary Pharmacology and Therapeutics*, 12(9), 925-934.
- Balkwill, F. (2006). TNF- α in promotion and progression of cancer. *Cancer and Metastasis Reviews*, 25(3), 409-416.
- Balkwill, F. (2009). Tumour necrosis factor and cancer. *Nature Reviews Cancer*, 9(5), 361-371.
- Balkwill, F., & Mantovani, A. (2001). Inflammation and cancer: back to Virchow? *The Lancet*, 357(9255), 539-545.
- Balraj, P., & Ruhana, S. (2007). PTEN mutation studies in Malaysian colorectal cancer patients. *Asia Pacific Journal of Molecular Biology and Biotechnology*, 15(1), 23-25.
- Bamba, S., Andoh, A., Ban, H., Imaeda, H., Aomatsu, T., Kobori, A., Inatomi, O. (2012). The severity of dextran sodium sulfate-induced colitis can differ between dextran sodium sulfate preparations of the same molecular weight range. *Digestive Diseases and Sciences*, 57(2), 327-334.
- Bannenberg, G. L., Chiang, N., Ariel, A., Arita, M., Tjonahen, E., Gotlinger, K. H., Serhan, C. N. (2005). Molecular circuits of resolution: formation and actions of resolvins and protectins. *The Journal of Immunology*, 174(7), 4345-4355.
- Barnes, P. J. (2002). Cytokine modulators as novel therapies for asthma. *Annual Review of Pharmacology and Toxicology*, 42(1), 81-98.
- Bates, R. C., & Mercurio, A. M. (2003). Tumor necrosis factor- α stimulates the epithelial-to-mesenchymal transition of human colonic organoids. *Molecular Biology of The Cell*, 14(5), 1790-1800.
- Batista Jr, M., Santos, R., Cunha, L., Mattos, K., Oliveira, E., Seelaender, M., & Costa Rosa, L. (2006). Changes in the pro-inflammatory cytokine production and peritoneal macrophage function in rats with chronic heart failure. *Cytokine*, 34(5), 284-290.

- Baud, V., & Karin, M. (2009). Is NF- κ B a good target for cancer therapy? Hopes and pitfalls. *Nature Reviews Drug Discovery*, 8(1), 33-40.
- Baumgart, D. C., & Carding, S. R. (2007). Inflammatory bowel disease: cause and immunobiology. *The Lancet*, 369(9573), 1627-1640.
- Becker, C., Fantini, M., Wirtz, S., Nikolaev, A., Lehr, H., Galle, P., Neurath, M. (2004). IL-6 Signaling Promotes Tumor Growth in Colorectal Cancer. *Cell Cycle*, 4(2), 220-223.
- Becker, C., Fantini, M. C., Schramm, C., Lehr, H. A., Wirtz, S., Nikolaev, A., Huber, S. (2004). TGF- β Suppresses Tumor Progression in Colon Cancer by Inhibition of IL-6*trans*-Signaling. *Immunity*, 21(4), 491-501.
- Beigel, F., Friedrich, M., Probst, C., Sotlar, K., Göke, B., Diegelmann, J., & Brand, S. (2014). Oncostatin M mediates STAT3-dependent intestinal epithelial restitution via increased cell proliferation, decreased apoptosis and upregulation of SERPIN family members. *PloS One*, 9(4), e93498.
- Bellingan, G. J., Caldwell, H., Howie, S., Dransfield, I., & Haslett, C. (1996). In vivo fate of the inflammatory macrophage during the resolution of inflammation: inflammatory macrophages do not die locally, but emigrate to the draining lymph nodes. *The Journal of Immunology*, 157(6), 2577-2585.
- Benetou, V., Orfanos, P., Lagiou, P., Trichopoulos, D., Boffetta, P., & Trichopoulou, A. (2008). Vegetables and fruits in relation to cancer risk: evidence from the Greek EPIC cohort study. *Cancer Epidemiology Biomarkers and Prevention*, 17(2), 387-392.
- Blokchina, O., Virolainen, E., & Fagerstedt, K. V. (2003). Antioxidants, oxidative damage and oxygen deprivation stress: a review. *Annals of Botany*, 91(2), 179-194.
- Boivin, G. P., Washington, K., Yang, K., Ward, J. M., Pretlow, T. P., Russell, R., Dove, W. F. (2003). Pathology of mouse models of intestinal cancer: consensus report and recommendations. *Gastroenterology*, 124(3), 762-777.
- Boland, C., Luciani, M., Gasche, C., & Goel, A. (2005). Infection, inflammation, and gastrointestinal cancer. *Gut*, 54(9), 1321-1331.
- Bollrath, J., Phesse, T. J., von Burstin, V. A., Putoczki, T., Bennecke, M., Bateman, T., Schwitalla, S. (2009). gp130-mediated Stat3 activation in enterocytes regulates cell survival and cell-cycle progression during colitis-associated tumorigenesis. *Cancer Cell*, 15(2), 91-102.
- Bonniaud, P., Margetts, P. J., Ask, K., Flanders, K., Gauldie, J., & Kolb, M. (2005). TGF- β and Smad3 signaling link inflammation to chronic fibrogenesis. *The Journal of Immunology*, 175(8), 5390-5395.

- Boots, A. W., Haenen, G. R., & Bast, A. (2008). Health effects of quercetin: from antioxidant to nutraceutical. *European Journal of Pharmacology*, 585(2), 325-337.
- Botteri, E., Iodice, S., Raimondi, S., Maisonneuve, P., & Lowenfels, A. B. (2008). Cigarette smoking and adenomatous polyps: a meta-analysis. *Gastroenterology*, 134(2), 388-395. e383.
- Bouchier-Hayes, L., Lartigue, L., & Newmeyer, D. D. (2005). Mitochondria: pharmacological manipulation of cell death. *Journal of Clinical Investigation*, 115(10), 2640.
- Bowman, T., Garcia, R., Turkson, J., & Jove, R. (2000). STATs in oncogenesis. *Oncogene*, 19(21), 2474-2488.
- Brand, S. (2009). Crohn's disease: Th1, Th17 or both? The change of a paradigm: new immunological and genetic insights implicate Th17 cells in the pathogenesis of Crohn's disease. *Gut*, 58(8), 1152-1167.
- Bravo, L. (1998). Polyphenols: chemistry, dietary sources, metabolism, and nutritional significance. *Nutrition Reviews*, 56(11), 317-333.
- Bromberg, J. (2002). Stat proteins and oncogenesis. *The Journal of Clinical Investigation*, 109(109 (9)), 1139-1142.
- Brown, J. R., & DuBois, R. N. (2005). COX-2: a molecular target for colorectal cancer prevention. *Journal of Clinical Oncology*, 23(12), 2840-2855.
- Buettner, R., Mora, L. B., & Jove, R. (2002). Activated STAT signaling in human tumors provides novel molecular targets for therapeutic intervention. *Clinical Cancer Research*, 8(4), 945-954.
- Calvert, P. M., & Frucht, H. (2002). The genetics of colorectal cancer. *Annals of Internal Medicine*, 137(7), 603-612.
- Canavan, C., Abrams, K., & Mayberry, J. (2006). Meta-analysis: colorectal and small bowel cancer risk in patients with Crohn's disease. *Alimentary Pharmacology and Therapeutics*, 23(8), 1097-1104.
- Caprioli, F., Sarra, M., Caruso, R., Stolfi, C., Fina, D., Sica, G., Monteleone, G. (2008). Autocrine regulation of IL-21 production in human T lymphocytes. *The Journal of Immunology*, 180(3), 1800-1807.
- Carnesecchi, S., Langley, K., Exinger, F., Gosse, F., & Raul, F. (2002). Geraniol, a component of plant essential oils, sensitizes human colonic cancer cells to 5-fluorouracil treatment. *Journal of Pharmacology and Experimental Therapeutics*, 301(2), 625-630.

- Carnésecchi, S., Schneider, Y., Lazarus, S. A., Coehlo, D., Gossé, F., & Raul, F. (2002). Flavanols and procyanidins of cocoa and chocolate inhibit growth and polyamine biosynthesis of human colonic cancer cells. *Cancer Letters*, 175(2), 147-155.
- Catlett-Falcone, R., Landowski, T. H., Oshiro, M. M., Turkson, J., Levitzki, A., Savino, R., Nuñez, G. (1999). Constitutive activation of Stat3 signaling confers resistance to apoptosis in human U266 myeloma cells. *Immunity*, 10(1), 105-115.
- Chen, C.-Y., Peng, W.-H., Tsai, K.-D., & Hsu, S.-L. (2007). Luteolin suppresses inflammation-associated gene expression by blocking NF- κ B and AP-1 activation pathway in mouse alveolar macrophages. *Life Sciences*, 81(23), 1602-1614.
- Chen, R., Alvero, A., Silasi, D., Kelly, M., Fest, S., Visintin, I., Mor, G. (2008). Regulation of IKK β by miR-199a affects NF- κ B activity in ovarian cancer cells. *Oncogene*, 27(34), 4712-4723.
- Cheynier, V. (2005). Polyphenols in foods are more complex than often thought. *The American Journal of Clinical Nutrition*, 81(1), 223S-229S.
- Chiarle, R., Simmons, W. J., Cai, H., Dhall, G., Zamo, A., Raz, R., Inghirami, G. (2005). Stat3 is required for ALK-mediated lymphomagenesis and provides a possible therapeutic target. *Nature Medicine*, 11(6), 623-629.
- Cianchi, F., Cortesini, C., Fantappiè, O., Messerini, L., Schiavone, N., Vannacci, A., Marzocca, C. (2003). Inducible nitric oxide synthase expression in human colorectal cancer: correlation with tumor angiogenesis. *The American Journal of Pathology*, 162(3), 793-801.
- Classen, A., Lloberas, J., & Celada, A. (2009). Macrophage activation: classical vs. alternative *Macrophages and Dendritic Cells* (pp. 29-43): Springer.
- Co-operation, O. f. E., & Development. (1995). Guideline for the testing of chemicals: Repeated dose 28-day oral toxicity study in rodents 407: Organisation for Economic Co-operation and Development Paris.
- Cooper, H. S., Murthy, S., Shah, R., & Sedergran, D. (1993). Clinicopathologic study of dextran sulfate sodium experimental murine colitis. *Laboratory Investigation; a Journal of Technical Methods and Pathology*, 69(2), 238-249.
- Corner, J., & Bailey, C. D. (2009). *Cancer Nursing: Care in Context*: John Wiley & Sons.
- Corvinus, F. M., Orth, C., Moriggl, R., Tsareva, S. A., Wagner, S., Pfitzner, E. B., Zatloukal, K. (2005). Persistent STAT3 activation in colon cancer is associated with enhanced cell proliferation and tumor growth. *Neoplasia*, 7(6), 545-555.

- Coussens, L. M., & Werb, Z. (2002). Inflammation and cancer. *Nature*, 420(6917), 860-867.
- Cui, X., Jin, Y., Hofseth, A. B., Pena, E., Habiger, J., Chumanovich, A., Singh, U. P. (2010). Resveratrol suppresses colitis and colon cancer associated with colitis. *Cancer Prevention Research*, 3(4), 549-559.
- D'Archivio, M., Filesi, C., Di Benedetto, R., Gargiulo, R., Giovannini, C., & Masella, R. (2007). Polyphenols, dietary sources and bioavailability. *Annali-Istituto Superiore di Sanita*, 43(4), 348.
- D'Archivio, M., Filesi, C., Vari, R., Scazzocchio, B., & Masella, R. (2010). Bioavailability of the polyphenols: status and controversies. *International Journal of Molecular Sciences*, 11(4), 1321-1342.
- Dai, Y., Jiao, H., Teng, G., Wang, W., Zhang, R., Wang, Y., Qiao, L. (2014). Embelin reduces colitis-associated tumorigenesis through limiting IL-6/STAT3 signaling. *Molecular Cancer Therapeutics*, 13(5), 1206-1216.
- Daniel, T., Thobe, B. M., Chaudry, I. H., Choudhry, M. A., Hubbard, W. J., & Schwacha, M. G. (2007). Regulation of the postburn wound inflammatory response by $\gamma\delta$ T-cells. *Shock*, 28(3), 278-283.
- de Kok, T. M., van Breda, S. G., & Manson, M. M. (2008). Mechanisms of combined action of different chemopreventive dietary compounds. *European Journal of Nutrition*, 47(2), 51-59.
- Decolongne, N., Kolb, M., Margetts, P. J., Menetrier, F., Artur, Y., Garrido, C., Bonniaud, P. (2007). TGF- β 1 induces progressive pleural scarring and subpleural fibrosis. *The Journal of Immunology*, 179(9), 6043-6051.
- Degterev, A., Boyce, M., & Yuan, J. (2003). A decade of caspases. *Oncogene*, 22(53), 8543-8567.
- Demma, J., Gebre-Mariam, T., Asres, K., Ergetie, W., & Engidawork, E. (2007). Toxicological study on Glinus lotoides A traditionally used taenicial herb in Ethiopia. *Journal of Ethnopharmacology*, 111(3), 451-457.
- Deng, L., Zhou, J.-F., Sellers, R. S., Li, J.-F., Nguyen, A. V., Wang, Y., Pollard, J. W. (2010). A novel mouse model of inflammatory bowel disease links mammalian target of rapamycin-dependent hyperproliferation of colonic epithelium to inflammation-associated tumorigenesis. *The American Journal of Pathology*, 176(2), 952-967.
- Derry, M. M., Raina, K., Balaiya, V., Jain, A. K., Shrotriya, S., Huber, K. M., Agarwal, C. (2013). Grape seed extract efficacy against azoxymethane-induced colon tumorigenesis in A/J mice: interlinking miRNA with cytokine signaling and inflammation. *Cancer Prevention Research*, 6(7), 625-633.

- Dillinger, T. L., Barriga, P., Escárcega, S., Jimenez, M., Lowe, D. S., & Grivetti, L. E. (2000). Food of the gods: cure for humanity? A cultural history of the medicinal and ritual use of chocolate. *The Journal of Nutrition*, 130(8), 2057S-2072S.
- Donehower, L. A., Harvey, M., Slagle, B. L., McArthur, M. J., Montgomery Jr, C. A., Butel, J. S., & Bradley, A. (1992). Mice deficient for p53 are developmentally normal but susceptible to spontaneous tumours. *Nature*, 356(6366), 215-221.
- Doucas, H., & Berry, D. P. (2006). Basic principles of the molecular biology of cancer I. *Surgery (Oxford)*, 24(2), 43-47.
- Dybing, E., Doe, J., Groten, J., Kleiner, J., O'Brien, J., Renwick, A., Walker, R. (2002). Hazard characterisation of chemicals in food and diet: dose response, mechanisms and extrapolation issues. *Food and Chemical Toxicology*, 40(2), 237-282.
- Eaden, J., Abrams, K., & Mayberry, J. (2001). The risk of colorectal cancer in ulcerative colitis: a meta-analysis. *Gut*, 48(4), 526-535.
- Eaton, D. L., & Klaassen, C. D. (1996). Principles of toxicology. *Casarett and Doull's Toxicology: The Basic Science of Poisons*, 1, 13-48.
- Efeyan, A., & Serrano, M. (2007). p53: guardian of the genome and policeman of the oncogenes. *Cell Cycle*, 6(9), 1006-1010.
- Eheman, C., Henley, S. J., Ballard-Barbash, R., Jacobs, E. J., Schymura, M. J., Noone, A. M., Kohler, B. A. (2012). Annual Report to the Nation on the status of cancer, 1975-2008, featuring cancers associated with excess weight and lack of sufficient physical activity. *Cancer*, 118(9), 2338-2366.
- Ekbom, A., Helmick, C., Zack, M., & Adami, H.-O. (1990). Ulcerative colitis and colorectal cancer: a population-based study. *New England Journal of Medicine*, 323(18), 1228-1233.
- Elmore, S. (2007). Apoptosis: a review of programmed cell death. *Toxicologic Pathology*, 35(4), 495-516.
- Epstein, F. H., Barnes, P. J., & Karin, M. (1997). Nuclear factor- κ B—a pivotal transcription factor in chronic inflammatory diseases. *New England Journal of Medicine*, 336(15), 1066-1071.
- Fang, J., Seki, T., Tsukamoto, T., Qin, H., Yin, H., Liao, L., Maeda, H. (2013). Protection from inflammatory bowel disease and colitis-associated carcinogenesis with 4-vinyl-2-, 6-dimethoxyphenol (canolol) involves suppression of oxidative stress and inflammatory cytokines. *Carcinogenesis*, bgt309.

- Feagins, L. A., Souza, R. F., & Spechler, S. J. (2009). Carcinogenesis in IBD: potential targets for the prevention of colorectal cancer. *Nature Reviews Gastroenterology and Hepatology*, 6(5), 297-305.
- Ferlay, J., Shin, H. R., Bray, F., Forman, D., Mathers, C., & Parkin, D. M. (2010). Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. *International Journal of Cancer*, 127(12), 2893-2917.
- Fernandes-Alnemri, T., Takahashi, A., Armstrong, R., Krebs, J., Fritz, L., Tomaselli, K. J., Salveson, G. (1995). Mch3, a novel human apoptotic cysteine protease highly related to CPP32. *Cancer Research*, 55(24), 6045-6052.
- Ferrero-Miliani, L., Nielsen, O., Andersen, P., & Girardin, S. (2007). Chronic inflammation: importance of NOD2 and NALP3 in interleukin-1 β generation. *Clinical and Experimental Immunology*, 147(2), 227-235.
- Fiala, E. S., Sohn, O. S., & Hamilton, S. R. (1987). Effects of chronic dietary ethanol on in vivo and in vitro metabolism of methylazoxymethanol and on methylazoxymethanol-induced DNA methylation in rat colon and liver. *Cancer Research*, 47(22), 5939-5943.
- Forrester, K., Ambs, S., Lupold, S. E., Kapust, R. B., Spillare, E. A., Weinberg, W. C., Tzeng, E. (1996). Nitric oxide-induced p53 accumulation and regulation of inducible nitric oxide synthase expression by wild-type p53. *Proceedings of the National Academy of Sciences*, 93(6), 2442-2447.
- Fuhrken, P. G., Chen, C., Miller, W. M., & Papoutsakis, E. T. (2007). Comparative, genome-scale transcriptional analysis of CHRF-288-11 and primary human megakaryocytic cell cultures provides novel insights into lineage-specific differentiation. *Experimental Hematology*, 35(3), 476-489. e423.
- Gao, S. P., Mark, K. G., Leslie, K., Pao, W., Motoi, N., Gerald, W. L., Clarkson, B. (2007). Mutations in the EGFR kinase domain mediate STAT3 activation via IL-6 production in human lung adenocarcinomas. *The Journal of Clinical Investigation*, 117(12), 3846-3856.
- Ghobrial, I. M., Witzig, T. E., & Adjei, A. A. (2005). Targeting apoptosis pathways in cancer therapy. *CA: a Cancer Journal for Clinicians*, 55(3), 178-194.
- Gonthier, M.-P., Verny, M.-A., Besson, C., Rémesy, C., & Scalbert, A. (2003). Chlorogenic acid bioavailability largely depends on its metabolism by the gut microflora in rats. *The Journal of Nutrition*, 133(6), 1853-1859.
- Gonzalez, R., Ballester, I., Lopez-Posadas, R., Suarez, M., Zarzuelo, A., Martinez-Augustin, O., & Medina, F. S. D. (2011). Effects of flavonoids and other polyphenols on inflammation. *Critical Reviews in Food Science and Nutrition*, 51(4), 331-362.

- Gosslau, A., Jao, E., Li, D., Huang, M. T., Ho, C. T., Evans, D., Chen, K. Y. (2011). Effects of the black tea polyphenol theaflavin-2 on apoptotic and inflammatory pathways in vitro and in vivo. *Molecular Nutrition and Food Research*, 55(2), 198-208.
- Granado-Serrano, A. B., Martín, M. A., Bravo, L., Goya, L., & Ramos, S. (2009). A diet rich in cocoa attenuates nitrosodimethylamine-induced liver injury in rats. *Food and Chemical Toxicology*, 47(10), 2499-2506.
- Grivennikov, S., Karin, E., Terzic, J., Mucida, D., Yu, G.-Y., Vallabhapurapu, S. Eckmann, L. (2009). IL-6 and Stat3 are required for survival of intestinal epithelial cells and development of colitis-associated cancer. *Cancer Cell*, 15(2), 103-113.
- Grivennikov, S. I. (2013). Inflammation and colorectal cancer: colitis-associated neoplasia. Paper presented at the Seminars in immunopathology.
- Grivennikov, S. I., Kuprash, D. V., Liu, Z. G., & Nedospasov, S. A. (2006). Intracellular signals and events activated by cytokines of the tumor necrosis factor superfamily: from simple paradigms to complex mechanisms. *International Review of Cytology*, 252, 129-161.
- Guzik, T., Mangalat, D., & Korbut, R. (2006). Adipocytokines novel link between inflammation. *Journal of Physiology Pharmacology*, 4, 505-528.
- Hagemann, T., Robinson, S. C., Schulz, M., Trümper, L., Balkwill, F. R., & Binder, C. (2004). Enhanced invasiveness of breast cancer cell lines upon co-cultivation with macrophages is due to TNF- α dependent up-regulation of matrix metalloproteases. *Carcinogenesis*, 25(8), 1543-1549.
- Hammerstone, J. F., Lazarus, S. A., Mitchell, A. E., Rucker, R., & Schmitz, H. H. (1999). Identification of procyanidins in cocoa (*Theobroma cacao*) and chocolate using high-performance liquid chromatography/mass spectrometry. *Journal of Agricultural and Food Chemistry*, 47(2), 490-496.
- Hanauer, S. B. (2006). Inflammatory bowel disease: epidemiology, pathogenesis, and therapeutic opportunities. *Inflammatory Bowel Diseases*, 12(5), S3-S9.
- Hanif, R., Pittas, A., Feng, Y., Koutsos, M. I., Qiao, L., Staiano-Coico, L., Rigas, B. (1996). Effects of nonsteroidal anti-inflammatory drugs on proliferation and on induction of apoptosis in colon cancer cells by a prostaglandin-independent pathway. *Biochemical Pharmacology*, 52(2), 237-245.
- Harrington, K. J. (2008). Biology of cancer. *Medicine*, 36(1), 1-4.
- Harris, T. J., Gross, J. F., Yen, H.-R., Xin, H., Kortylewski, M., Albesiano, E., Maris, C. H. (2007). Cutting edge: An in vivo requirement for STAT3 signaling in TH17 development and TH17-dependent autoimmunity. *The Journal of Immunology*, 179(7), 4313-4317.

- Hassan, F. A., Ismail, A., Abdulhamid, A., & Azlan, A. (2011). Identification and quantification of phenolic compounds in bambangan (*Mangifera pajang* Kort.) peels and their free radical scavenging activity. *Journal of Agricultural and Food Chemistry*, 59(17), 9102-9111.
- Heikkilä, K., Ebrahim, S., & Lawlor, D. A. (2008). Systematic review of the association between circulating interleukin-6 (IL-6) and cancer. *European Journal of Cancer*, 44(7), 937-945.
- Heywood, R. (1983). Long-term toxicity. *Animals and Alternatives in Toxicity Testing*, 79-89.
- Hii, C., Law, C., Suzannah, S., & Cloke, M. (2009). Polyphenols in cocoa (*Theobroma cacao L.*). Paper presented at the *Asian Journal of Food and Agro-Industry*.
- Howells, L. M., Moiseeva, E. P., Neal, C. P., Foreman, B. E., Andreadi, C. K., HUDSON, E., & MANSON, M. M. (2007). Predicting the physiological relevance of in vitro cancer preventive activities of phytochemicals1. *Acta Pharmacologica Sinica*, 28(9), 1274-1304.
- Huang, Y.-T., Wen, C.-C., Chen, Y.-H., Huang, W.-C., Huang, L.-T., Lin, W.-C. Hsiao, P.-W. (2013). Dietary uptake of Wedelia chinensis extract attenuates dextran sulfate sodium-induced colitis in mice. *PloS One*, 8(5), e64152.
- Hussain, A. I., Anwar, F., Hussain Sherazi, S. T., & Przybylski, R. (2008). Chemical composition, antioxidant and antimicrobial activities of basil (*Ocimum basilicum*)essential oils depends on seasonal variations. *Food Chemistry*, 108(3), 986-995.
- Hutfless, S. M., Ding, X., Girotra, S., & Ding, E. L. (2006). Chocolate and prevention of cardiovascular disease: a systematic review. *Nutrition and Metabolism*, 3: 2
- Hyun, Y. S., Han, D. S., Lee, A. R., Eun, C. S., Youn, J. H., & Kim, H.-Y. (2012). Role of IL-17A in the development of colitis-associated cancer. *Carcinogenesis*, 33 (4): 931-6.
- Igney, F. H., & Krammer, P. H. (2002). Death and anti-death: tumour resistance to apoptosis. *Nature Reviews Cancer*, 2(4), 277-288.
- Iimuro, Y., Gallucci, R. M., Luster, M. I., Kono, H., & Thurman, R. G. (1997). Antibodies to tumor necrosis factor alfa attenuate hepatic necrosis and inflammation caused by chronic exposure to ethanol in the rat. *Hepatology*, 26(6), 1530-1537.
- Itzkowitz, S. H., & Yio, X. (2004). Inflammation and cancer IV. Colorectal cancer in inflammatory bowel disease: the role of inflammation. *American Journal of Physiology-Gastrointestinal and Liver Physiology*, 287(1), G7-G17.

- Ivanov, V. N., Bhoumik, A., & Ronai, Z. e. (2003). Death receptors and melanoma resistance to apoptosis. *Oncogene*, 22(20), 3152-3161.
- Jackson, J. R., Seed, M., Kircher, C., Willoughby, D., & Winkler, J. (1997). The codependence of angiogenesis and chronic inflammation. *The Federation of American Societies for Experimental Biology Journal*, 11(6), 457-465.
- Jalil, A. M. M., & Ismail, A. (2008). Polyphenols in cocoa and cocoa products: is there a link between antioxidant properties and health? *Molecules*, 13(9), 2190-2219.
- Jemal, A., Siegel, R., Ward, E., Hao, Y., Xu, J., Murray, T., & Thun, M. J. (2008). Cancer statistics, 2008. *CA: A Cancer Journal for Clinicians*, 58(2), 71-96.
- Johnson, I. T. (2007). Phytochemicals and cancer. *Proceedings of the Nutrition Society*, 66(02), 207-215.
- Jump, R. L., & Levine, A. D. (2004). Mechanisms of natural tolerance in the intestine. Implications for inflammatory bowel disease. *Inflammatory Bowel Diseases*, 10(4), 462-478.
- Kargman, S. L., O'Neill, G. P., Vickers, P. J., Evans, J. F., Mancini, J. A., & Jothy, S. (1995). Expression of prostaglandin G/H synthase-1 and-2 protein in human colon cancer. *Cancer Research*, 55(12), 2556-2559.
- Karin, M., Lawrence, T., & Nizet, V. (2006). Innate immunity gone awry: linking microbial infections to chronic inflammation and cancer. *Cell*, 124(4), 823-835.
- Karin, M., & Lin, A. (2002). NF- κ B at the crossroads of life and death. *Nature Immunology*, 3(3), 221-227.
- Keane, M. P., & Strieter, R. M. (2000). Chemokine signaling in inflammation. *Critical Care Medicine*, 28(4), N13-N26.
- Kearney, P. M., Baigent, C., Godwin, J., Halls, H., Emberson, J. R., & Patrono, C. (2006). Do selective cyclo-oxygenase-2 inhibitors and traditional non-steroidal anti-inflammatory drugs increase the risk of atherothrombosis? Meta-analysis of randomised trials. *British Medical Journal*, 332(7553), 1302-1308.
- Keen, C. L. (2001). Chocolate: food as medicine/medicine as food. *Journal of the American College of Nutrition*, 20(sup5), 436S-439S.
- Kenny, T. P., Keen, C. L., Schmitz, H. H., & Gershwin, M. E. (2007). Immune effects of cocoa procyanidin oligomers on peripheral blood mononuclear cells. *Experimental Biology and Medicine*, 232(2), 293-300.

- Kim, H., & Keeney, P. (1983). Method of Analysis for (-)-Epicatechin in Cocoa Beans by High Performance Liquid Chromatography. *Journal of Food Science*, 48(2), 548-551.
- Kim, J. S., Lee, H. J., Lee, M. H., Kim, J., Jin, C., & Ryu, J.-H. (2006). Luteolin inhibits LPS-stimulated inducible nitric oxide synthase expression in BV-2 microglial cells. *Planta Medica*, 72(1), 65-68.
- Kim, S. W., Kim, H. M., Yang, K. M., Kim, S. A., Kim, S. K., An, M. J., Kim, W. H. (2010). Bifidobacterium lactis inhibits NF- κ B in intestinal epithelial cells and prevents acute colitis and colitis-associated colon cancer in mice. *Inflammatory Bowel Diseases*, 16(9), 1514-1525.
- Kitchens, R. L., Thompson, P. A., Munford, R. S., & O'Keefe, G. E. (2003). Acute inflammation and infection maintain circulating phospholipid levels and enhance lipopolysaccharide binding to plasma lipoproteins. *Journal of Lipid Research*, 44(12), 2339-2348.
- Klaunig, J. E., & Kamendulis, L. M. (2004). The role of oxidative stress in carcinogenesis. *Annual Review of Pharmacology and Toxicology*, 44, 239-267.
- Kobayashi, T., Okamoto, S., Hisamatsu, T., Kamada, N., Chinen, H., Saito, R., Koganei, K. (2008). IL23 differentially regulates the Th1/Th17 balance in ulcerative colitis and Crohn's disease. *Gut*, 57(12), 1682-1689.
- Korn, T., Bettelli, E., Gao, W., Awasthi, A., Jäger, A., Strom, T. B., Kuchroo, V. K. (2007). IL-21 initiates an alternative pathway to induce proinflammatory TH17 cells. *Nature*, 448(7152), 484-487.
- Korn, T., Bettelli, E., Oukka, M., & Kuchroo, V. K. (2009). IL-17 and Th17 Cells. *Annual Review of Immunology*, 27, 485-517.
- Krajewska, M., Moss, S. F., Krajewski, S., Song, K., Holt, P. R., & Reed, J. C. (1996). Elevated expression of Bcl-X and reduced Bak in primary colorectal adenocarcinomas. *Cancer Research*, 56(10), 2422-2427.
- Krakauer, T., Li, B. Q., & Young, H. A. (2001). The flavonoid baicalin inhibits superantigen-induced inflammatory cytokines and chemokines. *Federation European Biochemical Societies Letters*, 500(1), 52-55.
- Kraus, S., & Arber, N. (2009). Inflammation and colorectal cancer. *Current Opinion in Pharmacology*, 9(4), 405-410.
- Krishnan, K., Ruffin IV, M. T., & Brenner, D. E. (2000). Chemoprevention for colorectal cancer. *Critical Reviews in Oncology/Hematology*, 33(3), 199-219.

- Kroemer, G., El-Deiry, W., Golstein, P., Peter, M., Vaux, D., Vandenabeele, P., Knight, R. (2005). Classification of cell death: recommendations of the Nomenclature Committee on Cell Death. *Cell Death and Differentiation*, 12, 1463-1467.
- Kruglov, A. A., Kuchmiy, A., Grivennikov, S. I., Tumanov, A. V., Kuprash, D. V., & Nedospasov, S. A. (2008). Physiological functions of tumor necrosis factor and the consequences of its pathologic overexpression or blockade: mouse models. *Cytokine and Growth Factor Reviews*, 19(3), 231-244.
- Kuno, T., Hatano, Y., Tomita, H., Hara, A., Hirose, Y., Hirata, A., Tanaka, T. (2012). Organomagnesium suppresses inflammation-associated colon carcinogenesis in male Crj: CD-1 mice. *Carcinogenesis*, bgs348.
- Kurosawa, T., Itoh, F., Nozaki, A., Nakano, Y., Katsuda, S.-i., Osakabe, N., Itakura, H. (2005). Suppressive effect of cocoa powder on atherosclerosis in Kurosawa and Kusanagi-hypercholesterolemic rabbits. *Journal of Atherosclerosis and Thrombosis*, 12(1), 20-28.
- Kusaba, T., Nakayama, T., Yamazumi, K., Yakata, Y., Yoshizaki, A., Inoue, K., Sekine, I. (2006). Activation of STAT3 is a marker of poor prognosis in human colorectal cancer. *Oncology Reports*, 15(6), 1445-1451.
- Kusaba, T., Nakayama, T., Yamazumi, K., Yakata, Y., Yoshizaki, A., Nagayasu, T., & Sekine, I. (2005). Expression of p-STAT3 in human colorectal adenocarcinoma and adenoma; correlation with clinicopathological factors. *Journal of Clinical Pathology*, 58(8), 833-838.
- Lakatos, P. L., & Lakatos, L. (2008). Risk for colorectal cancer in ulcerative colitis: changes, causes and management strategies. *World Journal of Gastroenterology: WJG*, 14(25), 3937.
- Lala, P. K., & Chakraborty, C. (2001). Role of nitric oxide in carcinogenesis and tumour progression. *The Lancet Oncology*, 2(3), 149-156.
- Lassmann, S., Schuster, I., Walch, A., Göbel, H., Jütting, U., Makowiec, F., Werner, M. (2007). STAT3 mRNA and protein expression in colorectal cancer: effects on STAT3-inducible targets linked to cell survival and proliferation. *Journal of Clinical Pathology*, 60(2), 173-179.
- Lee, K. W., Kim, Y. J., Lee, H. J., & Lee, C. Y. (2003). Cocoa has more phenolic phytochemicals and a higher antioxidant capacity than teas and red wine. *Journal of Agricultural and Food Chemistry*, 51(25), 7292-7295.
- Lejeune, F. J. (2002). Clinical use of TNF revisited: improving penetration of anti-cancer agents by increasing vascular permeability. *The Journal of Clinical Investigation*, 110(4), 433-435.

- Levy, B. D., Clish, C. B., Schmidt, B., Gronert, K., & Serhan, C. N. (2001). Lipid mediator class switching during acute inflammation: signals in resolution. *Nature Immunology*, 2(7), 612-619.
- Levy, D. E., & Lee, C.-k. (2002). What does Stat3 do? *The Journal of Clinical Investigation*, 109(9), 1143-1148.
- Ley, K., Laudanna, C., Cybulsky, M. I., & Nourshargh, S. (2007). Getting to the site of inflammation: the leukocyte adhesion cascade updated. *Nature Reviews Immunology*, 7(9), 678-689.
- Li, H., Wu, W. K. K., Li, Z. J., Chan, K. M., Wong, C. C. M., Ye, C. G., Wang, M. (2010). 2, 3', 4, 4', 5'-Pentamethoxy-trans-stilbene, a resveratrol derivative, inhibits colitis-associated colorectal carcinogenesis in mice. *British Journal of Pharmacology*, 160(6), 1352-1361.
- Li, Q., Zhao, H., Zhang, Z., Liu, Z., Pei, X., Wang, J., Li, Y. (2009). Long-term administration of green tea catechins prevents age-related spatial learning and memory decline in C57BL/6 J mice by regulating hippocampal cyclic amp-response element binding protein signaling cascade. *Neuroscience*, 159(4), 1208-1215.
- Lim, G., & Halimah, Y. (2003). Second report of the national cancer registry. *Cancer Incidence in Malaysia*, 1-141.
- Liu, S.-C., Lin, J.-T., Wang, C.-K., Chen, H.-Y., & Yang, D.-J. (2009). Antioxidant properties of various solvent extracts from lychee (Litchi chinensisSonn.) flowers. *Food Chemistry*, 114(2), 577-581.
- Livak, K. J., & Schmittgen, T. D. (2001). Analysis of Relative Gene Expression Data Using Real-Time Quantitative PCR and the $2^{-\Delta\Delta CT}$ Method. *Methods*, 25(4), 402-408.
- Mackenzie, G. G., Adamo, A. M., Decker, N. P., & Oteiza, P. I. (2008). Dimeric procyanidin B2 inhibits constitutively active NF- κ B in Hodgkin's lymphoma cells independently of the presence of I κ B mutations. *Biochemical Pharmacology*, 75, 1461-1471.
- Mackenzie, G. G., Delfino, J. M., Keen, C. L., Fraga, C. G., & Oteiza, P. I. (2009). Dimeric procyanidins are inhibitors of NF- κ B-DNA binding. *Biochemical Pharmacology*, 78(9), 1252-1262.
- Manach, C., Williamson, G., Morand, C., Scalbert, A., & Rémésy, C. (2005). Bioavailability and bioefficacy of polyphenols in humans. I. Review of 97 bioavailability studies. *The American Journal of Clinical Nutrition*, 81(1), 230S-242S.
- Mannaioni, P., Di Bello, M., & Masini, E. (1997). Platelets and inflammation: role of platelet-derived growth factor, adhesion molecules and histamine. *Inflammation Research*, 46(1), 4-18.

- Mantovani, A., Allavena, P., Sica, A., & Balkwill, F. (2008). Cancer-related inflammation. *Nature*, 454(7203), 436-444.
- Marin, V., Montero-Julian, F. A., Grès, S., Boulay, V., Bongrand, P., Farnarier, C., & Kaplanski, G. (2001). The IL-6-soluble IL-6R α autocrine loop of endothelial activation as an intermediate between acute and chronic inflammation: an experimental model involving thrombin. *The Journal of Immunology*, 167(6), 3435-3442.
- Marletta, M. A. (1994). Nitric oxide synthase: aspects concerning structure and catalysis. *Cell*, 78(6), 927-930.
- Matsumoto, D., Hwang, H. S., & Yamada, H. (2010). Cultural differences in the relative contributions of face and context to judgments of emotions. *Journal of Cross-Cultural Psychology*, 0022022110387426.
- McGeachy, M. J., & Cua, D. J. (2008). Th17 cell differentiation: the long and winding road. *Immunity*, 28(4), 445-453.
- McGeer, E. G., Klegeris, A., & McGeer, P. L. (2005). Inflammation, the complement system and the diseases of aging. *Neurobiology of Aging*, 26(1), 94-97.
- Meng, C. C., Jalil, A. M. M., & Ismail, A. (2009). Phenolic and theobromine contents of commercial dark, milk and white chocolates on the Malaysian market. *Molecules*, 14(1), 200-209.
- Middleton, S., Shorthouse, M., & Hunter, J. (1993). Increased nitric oxide synthesis in ulcerative colitis. *The Lancet*, 341(8843), 465-466.
- Mitsuyama, K., Toyonaga, A., Sasaki, E., Ishida, O., Ikeda, H., Tsuruta, O., Tanikawa, K. (1995). Soluble interleukin-6 receptors in inflammatory bowel disease: relation to circulating interleukin-6. *Gut*, 36(1), 45-49.
- Mokhtar, M., Azli, R., Maleyki, A., Ismail, A., Abdul Ghani, N., Hamid, M., Mansor, S. M. (2008). Hypoglycaemic properties of Malaysian cocoa (*Theobroma Cacao*) polyphenols-rich extract. *International Food Research Journal*, 15(3).
- Moshi, M. J., van den Beukel, C., Hamza, O. J., Mbwambo, Z. H., Nondo, R. O., Masimba, P. J., Verweije, P. (2008). Brine shrimp toxicity evaluation of some Tanzanian plants used traditionally for the treatment of fungal infections. *African Journal of Traditional, Complementary and Alternative Medicines*, 4(2), 219-225.
- Motomura, M., Kwon, K. M., Suh, S.-J., Lee, Y.-C., Kim, Y.-K., Lee, I.-S., Kim, C.-H. (2008). Propolis induces cell cycle arrest and apoptosis in human leukemic U937 cells through Bcl-2/Bax regulation. *Environmental Toxicology and Pharmacology*, 26(1), 61-67.

- Mudter, J., & Neurath, M. F. (2007). IL-6 signaling in inflammatory bowel disease: Pathophysiological role and clinical relevance. *Inflammatory Bowel Diseases*, 13(8), 1016-1023.
- Mukaida, N., Matsumoto, T., Yokoi, K., Harada, A., & Matsushima, K. (1998). Inhibition of neutrophil-mediated acute inflammatory injury by an antibody against interleukin-8 (IL-8). *Inflammation Research*, 47(3), 151-157.
- Mukinda, J. T., & Syce, J. (2007). Acute and chronic toxicity of the aqueous extract of *Artemisia afra*. in rodents. *Journal of Ethnopharmacology*, 112(1), 138-144.
- Muller, W. A. (2003). Leukocyte-endothelial-cell interactions in leukocyte transmigration and the inflammatory response. *Trends in Immunology*, 24(6), 326-333.
- Munkholm, P. (2003). Review article: the incidence and prevalence of colorectal cancer in inflammatory bowel disease. *Alimentary Pharmacology and Therapeutics*, 18(s2), 1-5.
- Murad, F. (1996). Signal transduction using nitric oxide and cyclic guanosine monophosphate. *Journal of American Medicine Association*, 276(14), 1189-1192.
- Natrah, M., Ezat, S. W., Syed, M., Rizal, A. M., & Saperi, S. (2012). Quality of life in Malaysian colorectal cancer patients: a preliminary result. *Asian Pacific Journal of Cancer Prevention*, 13, 957-962.
- Neuman, M. G. (2007). Immune dysfunction in inflammatory bowel disease. *Translational Research*, 149(4), 173-186.
- Nowak, J., Weylandt, K. H., Habbel, P., Wang, J., Dignass, A., Glickman, J. N., & Kang, J. X. (2007). Colitis-associated colon tumorigenesis is suppressed in transgenic mice rich in endogenous n-3 fatty acids. *Carcinogenesis*, 28(9), 1991-1995.
- Obici, S., Otobone, F. J., Sela, V. R. d. S., Ishida, K., Silva, J. C. d., Nakamura, C. V., Audi, E. A. (2008). Preliminary toxicity study of dichloromethane extract of *Kielmeyera coriacea*. stems in mice and rats. *Journal of Ethnopharmacology*, 115(1), 131-139.
- Ohkusa, T. (1985). Production of experimental ulcerative colitis in hamsters by dextran sulfate sodium and changes in intestinal microflora. *Nihon Shokakibyo Gakkai zasshi= The Japanese Journal of Gastro-Enterology*, 82(5), 1327.
- Okayasu, I., Hatakeyama, S., Yamada, M., Ohkusa, T., Inagaki, Y., & Nakaya, R. (1990). A novel method in the induction of reliable experimental acute and chronic ulcerative colitis in mice. *Gastroenterology*, 98(3), 694-702.

- Okayasu, I., Ohkusa, T., Kajiura, K., Kanno, J., & Sakamoto, S. (1996). Promotion of colorectal neoplasia in experimental murine ulcerative colitis. *Gut*, 39(1), 87-92.
- Okayasu, I., Yamada, M., Mikami, T., Yoshida, T., Kanno, J., & Ohkusa, T. (2002). Dysplasia and carcinoma development in a repeated dextran sulfate sodium-induced colitis model. *Journal of Gastroenterology and Hepatology*, 17(10), 1078-1083.
- Ortega, N., Reguant, J., Romero, M.-P., Macia, A., & Motilva, M.-J. (2009). Effect of fat content on the digestibility and bioaccessibility of cocoa polyphenol by an in vitro digestion model. *Journal of Agricultural and Food Chemistry*, 57(13), 5743-5749.
- Osakabe, N., Yamagishi, M., Sanbongi, C., Natsume, M., Takizawa, T., & Osawa, T. (1998). The antioxidative substances in cacao liquor. *Journal of Nutritional Science and Vitaminology*, 44(2), 313-321.
- Othman, A., Ismail, A., Abdul Ghani, N., & Adenan, I. (2007). Antioxidant capacity and phenolic content of cocoa beans. *Food Chemistry*, 100(4), 1523-1530.
- Oviasogie, P., Okoro, D., & Ndiokwere, C. (2009). Determination of total phenolic amount of some edible fruits and vegetables. *African Journal of Biotechnology*, 8(12).
- Pan, M. H., Lai, C. S., Wu, J. C., & Ho, C. T. (2011). Molecular mechanisms for chemoprevention of colorectal cancer by natural dietary compounds. *Molecular Nutrition and Food Research*, 55(1), 32-45.
- Pandurangan, A., Ananda Sadagopan, S., & Dharmalingam, P. (2014). Inhibitory effect of luteolin on azoxymethane-induced colon carcinogenesis: involvement of iNOS and COX-2. *Pharmacognosy Magazine*. 10(38 supp), S306-310.
- Pandurangan, A. K., Ananda Sadagopan, S. K., Dharmalingam, P., & Ganapasm, S. (2013). Luteolin, a bioflavonoid inhibits Azoxymethane-induced colorectal cancer through activation of Nrf2 signaling. *Toxicology Mechanisms and Methods*, 24(1), 13-20.
- Pandurangan, A. K., Dharmalingam, P., Anandasadagopan, S., & Ganapasm, S. (2012). Effect of luteolin on the levels of glycoproteins during azoxymethane-induced colon carcinogenesis in mice. *Asian Pacific Journal of Cancer Prevention*, 13(4), 1569-1573.
- Pandurangan, A. K., Dharmalingam, P., Sadagopan, S. K. A., Ramar, M., Munusamy, A., & Ganapasm, S. (2013). Luteolin induces growth arrest in colon cancer cells through involvement of Wnt/β-catenin/GSK-3β signaling. *Journal of Environmental Pathology, Toxicology and Oncology*, 32(2), 131-139.

- Pandurangan, A. K., & Esa, N. M. (2013). Dietary non-nutritive factors in targeting of regulatory molecules in colorectal cancer: an update. *Asian Pacific Journal of Cancer Prevention*, 14(10), 5543-5552.
- Pandurangan, A. K., & Esa, N. M. (2014). Signal Transducer and Activator of Transcription 3-A Promising Target in Colitis-Associated Cancer. *Asian Pacific Journal of Cancer Prevention*, 15(2), 551-560.
- Pardal, R., Clarke, M. F., & Morrison, S. J. (2003). Applying the principles of stem-cell biology to cancer. *Nature Reviews Cancer*, 3(12), 895-902.
- Perše, M., & Cerar, A. (2012). Dextran sodium sulphate colitis mouse model: traps and tricks. *BioMed Research International*, 2012.
- Perwez Hussain, S., & Harris, C. C. (2007). Inflammation and cancer: an ancient link with novel potentiels. *International Journal of Cancer*, 121(11), 2373-2380.
- Pitot, H. (1986). Fundamental of oncology. Marcer Dekker. Inc, New York.
- Pokorný, J., & Korczak, J. (2001). Preparation of natural antioxidants. *Antioxidants in food: practical application*. Cambridge England: Woodhead Publishing Limited, 311-341.
- Popivanova, B. K., Kitamura, K., Wu, Y., Kondo, T., Kagaya, T., Kaneko, S., Mukaida, N. (2008a). Blocking TNF- $\tilde{\alpha}$ in mice reduces colorectal carcinogenesis associated with chronic colitis. *The Journal of Clinical Investigation*, 118(2), 560-570.
- Popivanova, B. K., Kitamura, K., Wu, Y., Kondo, T., Kagaya, T., Kaneko, S., Mukaida, N. (2008b). Blocking TNF- α in mice reduces colorectal carcinogenesis associated with chronic colitis. *The Journal of Clinical Investigation*, 118(2), 560-570.
- Porcheray, F., Viaud, S., Rimaniol, A. C., Leone, C., Samah, B., Dereuddre-Bosquet, N., Gras, G. (2005). Macrophage activation switching: an asset for the resolution of inflammation. *Clinical and Experimental Immunology*, 142(3), 481-489.
- Qu, Z., Liebler, J. M., Powers, M. R., Galey, T., Ahmadi, P., Huang, X.-N., Rosenbaum, J. T. (1995). Mast cells are a major source of basic fibroblast growth factor in chronic inflammation and cutaneous hemangioma. *The American Journal of Pathology*, 147(3), 564.
- Ramos, S. (2007). Effects of dietary flavonoids on apoptotic pathways related to cancer chemoprevention. *The Journal of Nutritional Biochemistry*, 18(7), 427-442.

- Ramos, S. (2008). Cancer chemoprevention and chemotherapy: dietary polyphenols and signalling pathways. *Molecular Nutrition and Food Research*, 52(5), 507-526.
- Rao, C., Kawamori, T., Hamid, R., Simi, B., Gambrell, B., & Reddy, B. (1998). Chemoprevention of colon cancer by iNOS specific and non-specific inhibitors: a safer colon cancer chemopreventive strategy. Paper presented at the Proc. Am. Assoc. *Cancer Research*.
- Rasmussen, S. E., Frederiksen, H., Struntze Krogholm, K., & Poulsen, L. (2005). Dietary proanthocyanidins: occurrence, dietary intake, bioavailability, and protection against cardiovascular disease. *Molecular Nutrition and Food Research*, 49(2), 159-174.
- Rawel, H., & Kulling, S. (2007). Nutritional contribution of coffee, cacao and tea phenolics to human health. *Journal für Verbraucherschutz und Lebensmittelsicherheit*, 2(4), 399-406.
- Raza, M., Al-Shabanah, O., El-Hadiyah, T., & Al-Majed, A. (2002). Effect of prolonged vigabatrin treatment on hematological and biochemical parameters in plasma, liver and kidney of Swiss albino mice. *Scientia Pharmaceutica*, 70(2), 135-145.
- Reddy, B. S. (2004). Studies with the azoxymethane–rat preclinical model for assessing colon tumor development and chemoprevention. *Environmental and Molecular Mutagenesis*, 44(1), 26-35.
- Reddy, B. S., Wang, C. X., Kong, A.-N., Khor, T. O., Zheng, X., Steele, V. E., Rao, C. V. (2006). Prevention of azoxymethane-induced colon cancer by combination of low doses of atorvastatin, aspirin, and celecoxib in F 344 rats. *Cancer Research*, 66(8), 4542-4546.
- Reid, K. E., Olsson, N., Schlosser, J., Peng, F., & Lund, S. T. (2006). An optimized grapevine RNA isolation procedure and statistical determination of reference genes for real-time RT-PCR during berry development. *BMC Plant Biology*, 6(1), 27.
- Rios, L. Y., Bennett, R. N., Lazarus, S. A., Rémesy, C., Scalbert, A., & Williamson, G. (2002). Cocoa procyanidins are stable during gastric transit in humans. *The American Journal of Clinical Nutrition*, 76(5), 1106-1110.
- Rodríguez-Ramiro, I., Ramos, S., López-Oliva, E., Agis-Torres, A., Bravo, L., Goya, L., & Martín, M. A. (2013). Cocoa polyphenols prevent inflammation in the colon of azoxymethane-treated rats and in TNF- α -stimulated Caco-2 cells. *British Journal of Nutrition*, 110(02), 206-215.

- Rodríguez-Ramiro, I., Ramos, S., López-Oliva, E., Agis-Torres, A., Gómez-Juaristi, M., Mateos, R., Martín, M. Á. (2011). Cocoa-rich diet prevents azoxymethane-induced colonic preneoplastic lesions in rats by restraining oxidative stress and cell proliferation and inducing apoptosis. *Molecular Nutrition and Food Research*, 55(12), 1895-1899.
- Romier-Crouzet, B., Van De Walle, J., During, A., Joly, A., Rousseau, C., Henry, O., Schneider, Y.-J. (2009). Inhibition of inflammatory mediators by polyphenolic plant extracts in human intestinal Caco-2 cells. *Food and Chemical Toxicology*, 47(6), 1221-1230.
- Rose-John, S., Mitsuyama, K., Matsumoto, S., Thaiss, W. M., & Scheller, J. (2009). Interleukin-6 trans-signaling and colonic cancer associated with inflammatory bowel disease. *Current Pharmaceutical Design*, 15(18), 2095-2103.
- Rosly, S., Shanmugavelu, S., Murugaiyah, M., Hadijah, H., Ahmad Tarmizi, S., Noridayusni, Y., & Subramaniam, K. (2011). Subchronic Oral Toxicity Study of Morinda citrifolia (Mengkudu) in Sprague Dawley Rats. *Pertanika Journal of Tropical Agricultural Science*, 34(2), 341-349.
- Rubin, D. C., Shaker, A., & Levin, M. S. (2012). Chronic intestinal inflammation: inflammatory bowel disease and colitis-associated colon cancer. *Frontiers In Immunology*, 3.
- Ruddon, R. W. (2007). *Cancer Biology*: Oxford University Press.
- Ruzaidi, A., Amin, I., Nawalyah, A., Hamid, M., & Faizul, H. (2005). The effect of Malaysian cocoa extract on glucose levels and lipid profiles in diabetic rats. *Journal of Ethnopharmacology*, 98(1), 55-60.
- Ruzaidi, A. M. M., Abbe, M. M. J., Amin, I., Nawalyah, A. G., & Muhajir, H. (2008). Protective effect of polyphenol-rich extract prepared from Malaysian cocoa (*Theobroma cacao*) on glucose levels and lipid profiles in streptozotocin-induced diabetic rats. *Journal of the Science of Food and Agriculture*, 88(8), 1442-1447.
- Saha, A., Kuzuhara, T., Echigo, N., Suganuma, M., & Fujiki, H. (2010). New role of (-)-epicatechin in enhancing the induction of growth inhibition and apoptosis in human lung cancer cells by curcumin. *Cancer Prevention Research*, 3(8), 953-962.
- Sakamoto, K., Maeda, S., Hikiba, Y., Nakagawa, H., Hayakawa, Y., Shibata, W., Omata, M. (2009). Constitutive NF-κB activation in colorectal carcinoma plays a key role in angiogenesis, promoting tumor growth. *Clinical Cancer Research*, 15(7), 2248-2258.

- Saleh, H. A., Jackson, H., & Banerjee, M. (2000). Immunohistochemical expression of bcl-2 and p53 oncoproteins: correlation with Ki67 proliferation index and prognostic histopathologic parameters in colorectal neoplasia. *Applied Immunohistochemistry and Molecular Morphology*, 8(3), 175-182.
- Saleh, M., & Trinchieri, G. (2010). Innate immune mechanisms of colitis and colitis-associated colorectal cancer. *Nature Reviews Immunology*, 11(1), 9-20.
- Sanbongi, C., Osakabe, N., Natsume, M., Takizawa, T., Gomi, S., & Osawa, T. (1998). Antioxidative polyphenols isolated from Theobroma cacao. *Journal of Agricultural and Food Chemistry*, 46(2), 454-457.
- Sasaki, R., Nishimura, N., Hoshino, H., Isa, Y., Kadokawa, M., Ichi, T., Ashida, H. (2007). Cyanidin 3-glucoside ameliorates hyperglycemia and insulin sensitivity due to downregulation of retinol binding protein 4 expression in diabetic mice. *Biochemical Pharmacology*, 74(11), 1619-1627.
- Scalbert, A., Manach, C., Morand, C., Rémesy, C., & Jiménez, L. (2005). Dietary polyphenols and the prevention of diseases. *Critical Reviews In Food Science and Nutrition*, 45(4), 287-306.
- Scalbert, A., & Williamson, G. (2000). Dietary intake and bioavailability of polyphenols. *The Journal of Nutrition*, 130(8), 2073S-2085S.
- Scheller, J., Ohnesorge, N., & Rose-John, S. (2006). Interleukin-6 Trans-Signalling in Chronic Inflammation and Cancer. *Scandinavian Journal of Immunology*, 63(5), 321-329.
- Schmidt, H. H., & Walter, U. (1994). NO at work. *Cell*, 78(6), 919-925.
- Seibel, J., Molzberger, A. F., Hertrampf, T., Laudenbach-Leschowski, U., & Diel, P. (2009). Oral treatment with genistein reduces the expression of molecular and biochemical markers of inflammation in a rat model of chronic TNBS-induced colitis. *European Journal of Nutrition*, 48(4), 213-220.
- Selmi, C., Cocchi, C. A., Lanfredini, M., Keen, C. L., & Gershwin, M. E. (2008). Chocolate at heart: The anti-inflammatory impact of cocoa flavanols. *Molecular Nutrition and Food Research*, 52(11), 1340-1348.
- Selmi, C., Mao, T. K., Keen, C. L., Schmitz, H. H., & Gershwin, M. E. (2006). The anti-inflammatory properties of cocoa flavanols. *Journal of Cardiovascular Pharmacology*, 47, S163-S171.
- Serhan, C. N., Jain, A., Marleau, S., Clish, C., Kantarci, A., Behbehani, B., Petasis, N. A. (2003). Reduced inflammation and tissue damage in transgenic rabbits overexpressing 15-lipoxygenase and endogenous anti-inflammatory lipid mediators. *The Journal of Immunology*, 171(12), 6856-6865.
- Serhan, C. N., & Savill, J. (2005). Resolution of inflammation: the beginning programs the end. *Nature Immunology*, 6(12), 1191-1197.

- Shafie, N., Mohd Esa, N., & Ithnin, H. (2013). Prophylactic Inositol Hexaphosphate (IP6) inhibits colon cancer through involvement of Wnt/β-catenin and COX-2 pathway. *BioMed Research International*, 2013, 681027.
- Shafie, N. H., Esa, N. M., Ithnin, H., Saad, N., & Pandurangan, A. K. (2013). Pro-Apoptotic Effect of Rice Bran Inositol Hexaphosphate (IP6) on HT-29 Colorectal Cancer Cells. *International Journal of Molecular Sciences*, 14(12), 23545-23558.
- Shafie, N. H., Mohd Esa, N., Ithnin, H., Md Akim, A., Saad, N., & Pandurangan, A. K. (2013). Preventive Inositol Hexaphosphate Extracted from Rice Bran Inhibits Colorectal Cancer through Involvement of Wnt/β-Catenin and COX-2 Pathways. *BioMed Research International*, 2013.
- Shan, G., & Li, J. (2002). Study of apoptosis in human liver cancers. *World Journal of Gastroenterology*, 8(2), 247-252.
- Shang, K., Bai, Y.-P., Wang, C., Wang, Z., Gu, H.-Y., Du, X., Mukaida, N. (2012). Crucial involvement of tumor-associated neutrophils in the regulation of chronic colitis-associated carcinogenesis in mice. *PloS One*, 7(12), e51848.
- Shang, X., Yao, G., Ge, J., Sun, Y., Teng, W., & Huang, Y. (2009). Procyanidin Induces Apoptosis and Necrosis of Prostate Cancer Cell Line PC-3 in a Mitochondrion-Dependent Manner. *Journal of Andrology*, 30(2), 122-126.
- Sheng, H., Shao, J., Morrow, J. D., Beauchamp, R. D., & DuBois, R. N. (1998). Modulation of apoptosis and Bcl-2 expression by prostaglandin E2 in human colon cancer cells. *Cancer Research*, 58(2), 362-366.
- Shiraki, M., Aihara, H., Kinouchi, Y., Takahashi, S., Oki, M., Noguchi, M., Shimosegawa, T. (2004). IL-12 p40 prevents the development of chronic enterocolitis in IL-10-deficient mice. *Laboratory Investigation*, 84(11), 1491-1500.
- Sica, A., Schioppa, T., Mantovani, A., & Allavena, P. (2006). Tumour-associated macrophages are a distinct M2 polarised population promoting tumour progression: potential targets of anti-cancer therapy. *European Journal of Cancer*, 42(6), 717-727.
- Siegel, R., Ma, J., Zou, Z., & Jemal, A. (2014). Cancer statistics, 2014. *CA: A Cancer Journal for Clinicians*, 64(1), 9-29.
- Signorelli, P., & Ghidoni, R. (2005). Resveratrol as an anticancer nutrient: molecular basis, open questions and promises. *The Journal of Nutritional Biochemistry*, 16(8), 449-466.
- Singleton, V. L., Orthofer, R., & Lamuela-Raventos, R. M. (1999). Analysis of total phenols and other oxidation substrates and antioxidants by means of Folin-Ciocalteu reagent. *Methods in Enzymology*, 299, 152-178.

- Smith, J. A. (1994). Neutrophils, host defense, and inflammation: a double-edged sword. *Journal of Leukocyte Biology*, 56(6), 672-686.
- Society, A. C. (2009). *Cancer Facts & Figures for Hispanics*: American Cancer Society.
- Sohn, O. S., Ishizaki, H., Yang, C. S., & Fiala, E. S. (1991). Metabolism of azoxymethane, methylazoxymethanol and N-nitrosodimethylamine by cytochrome P450IIE1. *Carcinogenesis*, 12(1), 127-131.
- Spencer, J. P., Schroeter, H., Crossthwaite, A. J., Kuhnle, G., Williams, R. J., & Rice-Evans, C. (2001). Contrasting influences of glucuronidation and O-methylation of epicatechin on hydrogen peroxide-induced cell death in neurons and fibroblasts. *Free Radical Biology and Medicine*, 31(9), 1139-1146.
- Stanković, M. S. (2011). Total phenolic content, flavonoid concentration and antioxidant activity of *Marrubium peregrinum* L. extracts. *Kragujevac Journal Science*, 33(2011), 63-72.
- Steinberg, F. M., Bearden, M. M., & Keen, C. L. (2003). Cocoa and chocolate flavonoids: implications for cardiovascular health. *Journal of the American Dietetic Association*, 103(2), 215-223.
- Stolfi, C., Rizzo, A., Franzè, E., Rotondi, A., Fantini, M. C., Sarra, M., Franceschilli, L. (2011). Involvement of interleukin-21 in the regulation of colitis-associated colon cancer. *The Journal of Experimental Medicine*, 208(11), 2279-2290.
- Strober, W., Fuss, I., & Mannon, P. (2007). The fundamental basis of inflammatory bowel disease. *The Journal of Clinical Investigation*, 117(3), 514-521.
- Subbaramaiah, K., Telang, N., Ramonetti, J. T., Araki, R., DeVito, B., Weksler, B. B., & Dannenberg, A. J. (1996). Transcription of cyclooxygenase-2 is enhanced in transformed mammary epithelial cells. *Cancer Research*, 56(19), 4424-4429.
- Suzuki, A., Hanada, T., Mitsuyama, K., Yoshida, T., Kamizono, S., Hoshino, T., Takeda, K. (2001). CIS3/SOCS3/SSI3 plays a negative regulatory role in STAT3 activation and intestinal inflammation. *The Journal of Experimental Medicine*, 193(4), 471-482.
- Szlosarek, P. W., Grimshaw, M. J., Kulbe, H., Wilson, J. L., Wilbanks, G. D., Burke, F., & Balkwill, F. R. (2006). Expression and regulation of tumor necrosis factor α in normal and malignant ovarian epithelium. *Molecular Cancer Therapeutics*, 5(2), 382-390.

- Takahashi, M., Fukuda, K., Ohata, T., Sugimura, T., & Wakabayashi, K. (1997). Increased expression of inducible and endothelial constitutive nitric oxide synthases in rat colon tumors induced by azoxymethane. *Cancer Research*, 57(7), 1233-1237.
- Talalay, P. (1992). Chemical protection against cancer by induction of electrophile detoxication (phase II) enzymes. *Cellular and Molecular Targets for Chemoprevention*, 193-205.
- Tang, W., Wang, W., Zhang, Y., Liu, S., Liu, Y., & Zheng, D. (2009). Tumour necrosis factor-related apoptosis-inducing ligand (TRAIL)-induced chemokine release in both TRAIL-resistant and TRAIL-sensitive cells via nuclear factor kappa B. *Federation European Biochemical Societies journal*, 276(2), 581-593.
- Tarka Jr, S., Morrissey, R., Apgar, J., Hostetler, K., & Shively, C. (1991). Chronic toxicity/carcinogenicity studies of cocoa powder in rats. *Food and Chemical Toxicology*, 29(1), 7-19.
- Teo, S., Stirling, D., Thomas, S., Hoberman, A., Kiorpis, A., & Khetani, V. (2002). A 90-day oral gavage toxicity study of d-methylphenidate and d, l-methylphenidate in Sprague-Dawley rats. *Toxicology*, 179(3), 183-196.
- Terzić, J., Grivennikov, S., Karin, E., & Karin, M. (2010). Inflammation and colon cancer. *Gastroenterology*, 138(6), 2101-2114. e2105.
- Thaker, A. I., Shaker, A., Rao, M. S., & Ciorba, M. A. (2012). Modeling colitis-associated cancer with azoxymethane (AOM) and dextran sulfate sodium (DSS). *Journal of Visualized Experiments: JoVE*(67).
- Tian, Y., Wang, K., Wang, Z., Li, N., & Ji, G. (2013). Chemopreventive effect of dietary glutamine on colitis-associated colon tumorigenesis in mice. *Carcinogenesis*, 00 (00) 1-8.
- Tiffany, R. (1978). *Oncology for Nurses and Health Care Professionals: Pathology, diagnosis, and treatment* (Vol. 1): Taylor & Francis.
- Tomas-Barberán, F. A., Cienfuegos-Jovellanos, E., Marín, A., Muguerza, B., Gil-Izquierdo, A., Cerdá, B., . . . Ibarra, A. (2007). A new process to develop a cocoa powder with higher flavonoid monomer content and enhanced bioavailability in healthy humans. *Journal of Agricultural and Food Chemistry*, 55(10), 3926-3935.
- Tosun, M., Ercisli, S., Sengul, M., Ozer, H., Polat, T., & Ozturk, E. (2009). Antioxidant properties and total phenolic content of eight Salvia species from Turkey. *Biological Research*, 42(2), 175-181.

- Triantafillidis, J. K., Nasioulas, G., & Kosmidis, P. A. (2009). Colorectal cancer and inflammatory bowel disease: epidemiology, risk factors, mechanisms of carcinogenesis and prevention strategies. *Anticancer Research*, 29(7), 2727-2737.
- Tsuda, T., Ueno, Y., Yoshikawa, T., Kojo, H., & Osawa, T. (2006). Microarray profiling of gene expression in human adipocytes in response to anthocyanins. *Biochemical Pharmacology*, 71(8), 1184-1197.
- Ullman, T., Croog, V., Harpaz, N., Sachar, D., & Itzkowitz, S. (2003). Progression of flat low-grade dysplasia to advanced neoplasia in patients with ulcerative colitis. *Gastroenterology*, 125(5), 1311-1319.
- Ullman, T. A., & Itzkowitz, S. H. (2011). Intestinal inflammation and cancer. *Gastroenterology*, 140(6), 1807-1816. e1801.
- Van Dyke, T., & Serhan, C. (2003). Resolution of inflammation: a new paradigm for the pathogenesis of periodontal diseases. *Journal of Dental Research*, 82(2), 82-90.
- van Erk, M. J., Roepman, P., van der Lende, T. R., Stierum, R. H., Aarts, J., van Bladeren, P. J., & van Ommen, B. (2005). Integrated assessment by multiple gene expression analysis of quercetin bioactivity on anticancer-related mechanisms in colon cancer cells in vitro. *European Journal of Nutrition*, 44(3), 143-156.
- Van Kemseke, C., Belaiche, J., & Louis, E. (2000). Frequently relapsing Crohn's disease is characterized by persistent elevation in interleukin-6 and soluble interleukin-2 receptor serum levels during remission. *International Journal of Colorectal Disease*, 15(4), 206-210.
- Vanamala, J., Leonardi, T., Patil, B. S., Taddeo, S. S., Murphy, M. E., Pike, L. M., Turner, N. D. (2006). Suppression of colon carcinogenesis by bioactive compounds in grapefruit. *Carcinogenesis*, 27(6), 1257-1265.
- Vauzour, D., Rodriguez-Mateos, A., Corona, G., Oruna-Concha, M. J., & Spencer, J. P. (2010). Polyphenols and human health: prevention of disease and mechanisms of action. *Nutrients*, 2(11), 1106-1131.
- Velioglu, Y., Mazza, G., Gao, L., & Oomah, B. (1998). Antioxidant activity and total phenolics in selected fruits, vegetables, and grain products. *Journal of Agricultural and Food Chemistry*, 46(10), 4113-4117.
- Velmurugan, B., Singh, R. P., Tyagi, A., & Agarwal, R. (2008). Inhibition of azoxymethane-induced colonic aberrant crypt foci formation by silibinin in male Fisher 344 rats. *Cancer Prevention Research*, 1(5), 376-384.

- Vinson, J. A., Proch, J., Bose, P., Muchler, S., Taffera, P., Shuta, D., Agbor, G. A. (2006). Chocolate is a powerful ex vivo and in vivo antioxidant, an antiatherosclerotic agent in an animal model, and a significant contributor to antioxidants in the European and American Diets. *Journal of Agricultural and Food Chemistry*, 54(21), 8071-8076.
- Vinson, J. A., Proch, J., & Zubik, L. (1999). Phenol antioxidant quantity and quality in foods: cocoa, dark chocolate, and milk chocolate. *Journal of Agricultural and Food Chemistry*, 47(12), 4821-4824.
- Visioli, F., Bernaert, H., Corti, R., Ferri, C., Heptinstall, S., Molinari, E., Vinson, J. A. (2009). Chocolate, lifestyle, and health. *Critical Reviews in Food Science and Nutrition*, 49(4), 299-312.
- Volate, S. R., Davenport, D. M., Muga, S. J., & Wargovich, M. J. (2005). Modulation of aberrant crypt foci and apoptosis by dietary herbal supplements (quercetin, curcumin, silymarin, ginseng and rutin). *Carcinogenesis*, 26(8), 1450-1456.
- Waldner, M. J., & Neurath, M. F. (2008). Cytokines in colitis-associated cancer: potential drug targets? *Inflammation & Allergy-Drug Targets (Formerly Current Drug Targets-Inflammation & Allergy)*, 7(3), 187-194.
- Wang, J., & Mazza, G. (2002). Effects of anthocyanins and other phenolic compounds on the production of tumor necrosis factor α in LPS/IFN- γ -activated RAW 264.7 macrophages. *Journal of Agricultural and Food Chemistry*, 50(15), 4183-4189.
- Wang, S., Liu, Z., Wang, L., & Zhang, X. (2009). NF-kappaB signaling pathway, inflammation and colorectal cancer. *Cell Molecular Immunology*, 6(5), 327-334.
- Wang, S., Meckling, K. A., Marcone, M. F., Kakuda, Y., & Tsao, R. (2011). Can phytochemical antioxidant rich foods act as anti-cancer agents? *Food Research International*, 44(9), 2545-2554.
- Wang, Z., Jin, H., Xu, R., Mei, Q., & Fan, D. (2009). Triptolide downregulates Rac1 and the JAK/STAT3 pathway and inhibits colitis-related colon cancer progression. *Experimental and Molecular Medicine*, 41(10), 717-727.
- Ward, J. (1974). Morphogenesis of chemically induced neoplasms of the colon and small intestine in rats. *Laboratory Investigation; a Journal of Technical Methods and Pathology*, 30(4), 505-513.
- Weisburger, J. H. (2001). Chemopreventive effects of cocoa polyphenols on chronic diseases. *Experimental Biology and Medicine*, 226(10), 891-897.
- Wertz, I. E., & Dixit, V. M. (2010). Signaling to NF- κ B: regulation by ubiquitination. *Cold Spring Harbor Perspectives in Biology*, 2(3), a003350.

- Williams, T., & Peck, M. (1977). Role of prostaglandin-mediated vasodilatation in inflammation. *Nature*, 270, 530-532.
- Wirtz, S., Neufert, C., Weigmann, B., & Neurath, M. F. (2007). Chemically induced mouse models of intestinal inflammation. *Nature Protocols*, 2(3), 541-546.
- Wirtz, S., & Neurath, M. F. (2007). Mouse models of inflammatory bowel disease. *Advanced Drug Delivery Reviews*, 59(11), 1073-1083.
- Wiseman, H., & Halliwell, B. (1996). Damage to DNA by reactive oxygen and nitrogen species: role in inflammatory disease and progression to cancer. *Biochemical Journal*, 313, 17-29.
- Wollgast, J., & Anklam, E. (2000). Polyphenols in chocolate: is there a contribution to human health? *Food Research International*, 33(6), 449-459.
- Wong, R. (2011). Apoptosis in cancer: from pathogenesis to treatment. *Journal of Experimental and Clinical Cancer Research*, 30(1), 87.
- Wurochekke, A., Anthony, A., & Obidah, W. (2008). Biochemical effects on the liver and kidney of rats administered aqueous stem bark extract of Xemenia Americana. *African Journal of Biotechnology*, 7(16).
- Xie, J., & Itzkowitz, S. H. (2008). Cancer in inflammatory bowel disease. *World Journal of Gastroenterology: WJG*, 14(3), 378.
- Yamamoto, T. (2000). Molecular mechanism of monocyte predominant infiltration in chronic inflammation: mediation by a novel monocyte chemotactic factor, S19 ribosomal protein dimer. *Pathology International*, 50(11), 863-871.
- Yamamoto, Y., & Gaynor, R. B. (2001). Therapeutic potential of inhibition of the NF- κ B pathway in the treatment of inflammation and cancer. *Journal of Clinical Investigation*, 107(2), 135.
- Yan, Y., Kolachala, V., Dalmasso, G., Nguyen, H., Laroui, H., Sitaraman, S. V., & Merlin, D. (2009). Temporal and spatial analysis of clinical and molecular parameters in dextran sodium sulfate induced colitis. *PloS One*, 4(6), e6073.
- Yarbro, C., Wujcik, D., & Gobel, B. H. (2010). *Cancer nursing: principles and practice*: Jones and Bartlett Learning.
- Yin, C., Knudson, C. M., Korsmeyer, S. J., & Van Dyke, T. (1997). Bax suppresses tumorigenesis and stimulates apoptosis in vivo. *Nature*, 385(6617), 637-640.
- Zeller, J. L., Lynn, C., & Glass, R. M. (2008). Colon Cancer. *Journal of American Medical Association*, 300(23), 2816-2816.
- Zheng, P., Niu, F.-L., Liu, W.-Z., Shi, Y., & Lu, L.-G. (2005). Anti-inflammatory mechanism of oxymatrine in dextran sulfate sodium-induced colitis of rats. *World Journal of Gastroenterology*, 11(31), 4912.

Zhu, Y., Zhu, M., & Lance, P. (2012). iNOS signaling interacts with COX-2 pathway in colonic fibroblasts. *Experimental Cell Research*, 318(16), 2116-2127.

