



**UNIVERSITI PUTRA MALAYSIA**

***DEVELOPMENT OF A SINGLE-STEP PLASMA IRON DETECTION  
METHOD***

**LIM WAI FENG**

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## **DEVELOPMENT OF A SINGLE-STEP PLASMA IRON DETECTION METHOD**

**By**

**LIM WAI FENG**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfillment of the Requirement for the Degree of Doctor of Philosophy**

**August 2017**

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

## **DEVELOPMENT OF A SINGLE-STEP PLASMA IRON DETECTION METHOD**

By

**LIM WAI FENG**

**August 2017**

**Chairman : Lai Mei I, PhD**  
**Faculty : Medicine and Health Sciences**

Iron deficiency anaemia (IDA) is the most common cause of anaemia worldwide that affects almost two billion people in many developing countries including Malaysia. The gold standard for identifying iron deficiency is a direct test of bone marrow iron. Bone marrow aspiration is too invasive in nature for routine use. Therefore, peripheral whole blood and serum/plasma are used to assess iron status through haematological and biochemistry tests, respectively. Iron status quantification can be limited in some areas due to high cost and lack of access to these analysers. Therefore, a low-cost and efficient technique was designed to detect iron status using human plasma. Currently, no single definitive diagnosis can assess iron status effectively except bone marrow iron. Although ferritin is a common practice, it can be confounded by inflammation and required haemoglobin level to detect iron deficiency anaemia. To devise a technique for field studies, plasma iron (PI) was chosen due to its simplicity without involving multiple steps like ferritin. Firstly, a recipe to rapidly induce iron release from human plasma was identified, comprising 720 mM citric acid, 20 mM ascorbic acid, 100 mM thiourea and 3 mM ferrozine. Next, a total of 190 samples were collected, i.e. 10 inflammation (Infla), 31 iron deficiency with and without anaemia (IDwwA), 114 normal iron (NI) and 35 iron overload (IOL), respectively. These samples were subjected to PI screening using the freeze-dried version of the concocted recipe. By comparing the current technique (termed Prototype\_PI) to autoanalyser (termed Cobas\_PI), Prototype\_PI and Cobas\_PI across all samples were ranged from 148.3-2744.4  $\mu\text{g/L}$  and 184.0-2918.0  $\mu\text{g/L}$ , respectively with 72.1-157.4% of recoveries. Only nine samples were found to be beyond 80-120% of the acceptance range. Both methods correlated well with a Spearman rho coefficient of 0.967. In Passing-Bablok analysis, both methods did not differ by any constant or proportional error but has random error, with residual standard deviation (RSD) of 61.5  $\mu\text{g/L}$  across all samples. The Bland-Altman's limit of agreement (LoA) was -239.7 to 104.8  $\mu\text{g/L}$  with a mean difference of -67.5  $\mu\text{g/L}$ . Concordance (CCC) and intraclass correlation coefficient (ICC) of 0.980 and 0.994, respectively, indicating a good agreement between two methods. Across iron status, each group indicated good agreement with values more than 0.9 for Spearman rho coefficient, CCC and ICC. LoAs were -156.3 to 65.6  $\mu\text{g/L}$  (IDwwA), -225.8 to 86.5  $\mu\text{g/L}$  (NI) and -336.0 to 129.3  $\mu\text{g/L}$  (IOL) with a mean difference of -45.35  $\mu\text{g/L}$ , -69.65  $\mu\text{g/L}$  and -103.32  $\mu\text{g/L}$ , respectively.

Similarly, neither constant nor proportional error found across iron status, indicating random error contributed to the difference between both methods. As compared to Cobas\_PI, Prototype\_PI has a sensitivity of 87.5% (91.7%) and a specificity of 97.1% (96.8%) in diagnosis of IDwwoA in male (female), respectively. However, the ability of Prototype\_PI to diagnose IOL in male (female) was reported to have lower sensitivity, i.e. 71.4% (male) and 80.8% (female) but 100% specificity, respectively. By comparing to ferritin level, both Prototype\_PI and Cobas\_PI found to have moderate sensitivity and specificity. This project concluded that Prototype\_PI could screen PI successfully as comparable to Cobas\_PI for diagnosis of IDwwoA but less accurate in IOL screening. Further justification has to be done by performing double-blind study.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

## **PEMBANGUNAN KAEDAH PENGESANAN ZAT BESI PLASMA DENGAN SATU LANGKAH**

Oleh

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**Ogos 2017**

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Anemia kekurangan zat besi merupakan anemia yang paling biasa di seluruh dunia dan melibatkan hampir dua bilion orang dan kebanyakan negara-negara membangun termasuk Malaysia. 'Gold standard' untuk mengesan kekurangan zat besi adalah ujian terus atas zat besi di sumsum tulang. Namun begitu, Aspirasi sumsum tulang adalah terlalu invasif untuk penggunaan rutin. Oleh itu, darah periferi dan plasma telah digunakan untuk mengukur tahap zat besi, iaitu melalui ujian hematologi dan biokimia masing-masing. Kuantifikasi tahap zat besi terhad di beberapa daerah kerana kos yang tinggi dan kekurangan akses terhadap autoanalyser. Oleh itu, satu kaedah yang berkos rendah dan teknik yang berkesan telah dicipta untuk mengukur tahap zat besi dalam plasma manusia. Masa kini, tiada satupun diagnosis muktamad yang dapat mengukur tahap zat besi secara berkesan kecuali pengukuran zat besi di sumsum tulang. Walaupun feritin merupakan amalan biasa, dan tahapnya akan meningkat disebabkan oleh mana-mana gejala meradang dan memerlukan tahap hemoglobin untuk mengesan anemia kekurangan zat besi. Untuk merangka teknik untuk bidang kajian, besi plasma (PI) telah dipilih kerana ia lebih mudah tanpa melibatkan pelbagai langkah seperti feritin. Satu resipi untuk mendorong pembebasan zat besi dari plasma manusia dengan cepat telah dikenalpasti, termasuk 720 mM asid sitrik, 20 mM asid askorbik, 100 mM thiourea dan 3 mM ferrozine. Seterusnya, sejumlah 190 sampel telah dikumpul iaitu 10 individu yang beradang (Infla), 31 individu yang kekurangan zat besi dengan dan tanpa anemia (IDwwoA), 114 individu yang mempunyai zat besi tahap biasa (NI) dan 35 individu yang mempunyai bebanan zat besi (IOL) masing-masing. Sampel-sampel ini telah disaringkan dengan menggunakan resipi yang telah dibeku dan kering. Dengan membandingkan teknik semasa (Prototip\_PI) dengan autoanalyser (Cobas\_PI), Prototip\_PI dan Cobas\_PI berjulat dari 148.3-2744.4 µg/L dan 184.0-2918.0 µg/L ke atas semua sampel masing-masing dan mempunyai pemulihan sebanyak 72.1-157.4%. Hanya terdapat 9 sampel yang terkeluar daripada julat yang dibenarkan, iaitu 80-120%. Kedua-dua kaedah ini berkorelasi baik dengan koefisien spearman rho iaitu 0.967. Dalam analisis Passing-Bablok, tiada sebarang ralat konstant atau berkadar didapati dalam kedua-dua kaedah tetapi mempunyai ralat rawak dengan sisa sisihan piawai sebanyak 61.5 µg/L ke atas semua sampel. Had persetujuan dengan analisis Bland-Altman adalah -239.7 ke 104.8

$\mu\text{g/L}$  dengan perbezaan purata sebanyak  $-67.5 \mu\text{g/L}$ . Konkordans (CCC) dan pekali korelasi intraclass (ICC) adalah sebanyak 0.980 dan 0.994 masing-masing, menunjukkan persetujuan yang baik antara kedua-dua kaedah tersebut. Setiap kumpulan tahap zat besi menunjukkan persetujuan yang baik dengan nilai melebihi 0.9 untuk koefisien spearman rho, CCC dan ICC. Had persetujuan dengan analisis Bland-Altman adalah  $-156.3$  ke  $65.6 \mu\text{g/L}$  (IDwwoA),  $-225.8$  ke  $86.5 \mu\text{g/L}$  (NI) dan  $-336.0$  ke  $129.3 \mu\text{g/L}$  (IOL) dengan perbezaan purata sebanyak  $-45.35 \mu\text{g/L}$ ,  $-69.65 \mu\text{g/L}$  dan  $-103.32 \mu\text{g/L}$  masing-masing. Serupanya, tiada ralat konstant ataupun berkadar didapati tetapi ralat rawak yang menyumbang kepada perbezaan antara kedua-dua kaedah. Berbanding dengan Cobas\_PI, Prototip\_PI mempunyai tahap sensitiviti sebanyak 87.5% (91.7%) dan spesifisiti sebanyak 97.1% (96.8%) dalam diagnosis IDwwoA untuk lelaki (perempuan) masing-masing. Namun begitu, kemampuan Prototip\_PI untuk diagnosis IOL dalam lelaki (perempuan) dilaporkan mempunyai sensitiviti yang rendah iaitu 71.4% (lelaki) dan 80.8% (perempuan) tetapi mempunyai spesifisiti 100% masing-masing. Dengan berbanding dengan tahap feritin, kedua-dua Prototip\_PI dan Cobas\_PI didapati mempunyai tahap sensitiviti dan spesifisiti yang sederhana. Projek ini menyimpulkan bahawa Prototip\_PI mampu membuat penyaringan PI dengan berjaya untuk diagnosis IDwwoA tetapi kurang memuaskan dalam diagnosis IOL berbanding dengan Cobas\_PI. Justifikasi yang selanjutnya seharusnya dilakukan dengan melaksanakan kajian dalam sampel secara rawak.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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## LIST OF ABBREVIATIONS

3D	Three-dimensional
AAS	Atomic absorption spectrophotometer
AsC	Ascorbic acid
AUC	Area under curve
BHT	2,6-bis[hydroxy(methyl)amino]-1,3,5-triazine
BPY	2,2'-bipyridine
BSA	Bovine serum albumin
c-SBF	Conventional-SBF
CA	Citric acid
CAD	Computer-aided-design
CCC	Lin's concordance correlation coefficient
cHP	Commercially available human plasma
CI	Confidence interval
Cobas_PI	Plasma iron measured from Cobas
CRP	C-reactive protein
CRM	Certified reference material
CV	Coefficient variation
DFO	Desferoxamine
DI	Deionised water
EDTA	Ethylenediaminetetraacetic acid
ELISA	Enzyme-linked immunosorbent assay
EPO	Erythropoietin
EqaSF	Equine spleen apoferritin
EqSF	Equine spleen ferritin
FdWC	Freeze-dry with prior concentrating
FdwoC	Freeze-dry without prior concentrating
FER	Ferene
FMNH <sub>2</sub>	Reduced flavin mononucleotide
FMN	Flavin mononucleotide
FRC	Fragmented red cells
FRN	Ferritin
FRZ	Ferrozine
Hb	Haemoglobin
HCT	Haematocrit
HEPES	4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid)
HuaTF	Human apotransferrin
HuLF	Human liver ferritin
HuTF	Human transferrin
HyHCl	Hydroxylamine hydrochloride
i-SBF	Ionised-SBF
I <sub>0</sub>	Incident radiant energy
ICC	Intraclass correlation coefficient
ICP-MS	Inductively coupled plasma mass spectrometry
ID	Iron deficiency
IDA	Iron deficiency anaemia
IDE	Iron deficiency erythropoiesis
IDwoA	iron deficiency with and without anaemia
Infla	Inflammation

IOL	Iron overload
IRF	Immature reticulocyte fraction
LDR	Light-dependent resistor
LED	Light-emitting diode
LG	Logarithm-transformed
LoA	Limit of agreement
LOD	Limit of detection
LOQ	Limit of quantitation
LR-	Likelihood ratio negative
LR+	Likelihood ratio positive
m-SBF	Modified-SBF
MCH	Mean cell haemoglobin
MCHC	Mean cell haemoglobin concentration
MCV	Mean cell volume
MCV <sub>r</sub>	mean reticulocyte volume
MES	2-(N-Morpholino)ethanesulfonic acid, 4-Morpholineethanesulfonic acid
MFR	Middle-fluorescent reticulocytes
Mops	3-Morpholinopropanesulfonic acid
NA	Not available
NI	Normal iron
noPC	Without pathlength correction
NPV	Negative predictive value
NRBCs	Nucleated red blood cells
OFR	Out of range
op-amp	Operational amplifier
PEG	Polyethylene glycol
PCDI	Pathlength correction with deionised water
PCr-SBF	Pathlength correction with revised-simulated body fluid
PCV	Packed cell volume
PI	Plasma iron
POC	Point-of-care
PPV	Positive predictive value
Prototype_PI	Plasma iron measured from Prototype
r-SBF	Revised-SBF
RBC	Red blood cell
RCF	Red cell flag
RDW	Red cell distribution width
RDWI	Red cell distribution width index
ReSD	Relative standard deviation
Ret-Hb/CHr/Ret-Y	Reticulocyte haemoglobin content
Ret-Hbe	reticulocyte haemoglobin equivalent
RMI	Reticulocyte maturity index
ROC	Receiver operative curve
RSD	Residual standard deviation
SBF	Simulated body fluid
SD	Standard deviation
SDT	Sodium dithionite
SI	Serum iron
sTfR	Soluble transferrin receptor
sTfR-F index	Ratio of transferrin receptor to ferritin



Tf	Serum transferrin
TfR	Transferrin receptors
TIBC	Total iron binding capacity
TRIS	Tris(hydroxymethyl)aminomethane
TU	Thiourea
WBC	White blood cells
WHO	World Health Organization
ZPP	Zinc protoporphyrin
%HYPO	Percentage of hypochromic red cells
%HYPOr	Percentage of hypochromic reticulocytes
%T	Percentage of transmittance
%TSAT	Transferrin saturation





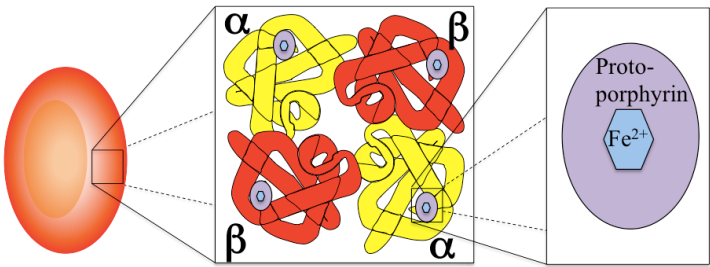
# CHAPTER 1

## INTRODUCTION

### 1.1 Anaemia

A single red blood cell (RBC) consists of millions of haemoglobins, which function as an important oxygen-carrier from the lungs to the tissues in our bodies. Haemoglobin comprises of two  $\alpha$ - and two  $\beta$ -globin chains; each binding to a haem group. The haem group consists of a cyclic protoporphyrin ring and an iron core (**Figure 1.1**) (Hoffbrand et al., 2006).

Anaemia is described as a condition when RBCs are not supplying sufficient oxygen throughout the body for the body's normal physiological need (Hoffbrand et al., 2006; WHO, 2011). The three main causes of anaemia are contributed by (1) key micronutrient deficiencies for RBC synthesis, such as iron, folate, vitamin B<sub>12</sub> and vitamin A deficiencies; (2) inherited conditions that affect haemoglobin structure or function, such as  $\alpha$ - or  $\beta$ -thalassaemia and sickle cell anaemia; and (3) infectious diseases that cause intravascular haemolysis, such as malaria, hookworm and schistosomiasis (WHO, 2007; Miller et al., 2013; Pasricha, 2014). Anaemia is a public health problem, mainly attributed by iron deficiency anaemia (Hoffbrand et al., 2006; WHO, 2001; WHO, 2015).

Structure	Red blood cell	Haemoglobin	Heme
			
Content	<ul style="list-style-type: none"> <li>• millions of haemoglobin</li> </ul>	<ul style="list-style-type: none"> <li>• two <math>\alpha</math>-globin chains</li> <li>• two <math>\beta</math>-globin chains</li> </ul>	<ul style="list-style-type: none"> <li>• iron</li> <li>• protoporphyrin</li> </ul>
Deficiency /faulty	<ul style="list-style-type: none"> <li>• anaemia</li> </ul>	<ul style="list-style-type: none"> <li>• <math>\alpha</math>-thalassaemia</li> <li>• <math>\beta</math>-thalassaemia</li> </ul>	<ul style="list-style-type: none"> <li>• iron deficiency anaemia</li> <li>• sideroblastic anaemia</li> </ul>

**Figure 1.1: Red blood cell structure.** Adult red blood cells (RBCs) consist of millions of haemoglobins: two  $\alpha$ -globin chains (shown in orange colour) and two  $\beta$ -globin chains (shown in red colour); each globin has a heme group made of a protoporphyrin ring (shown in purple colour) and an iron core (shown in blue colour). Reduced production of globin chains and iron leads to a low haemoglobin content in the RBCs resulting in anaemia.

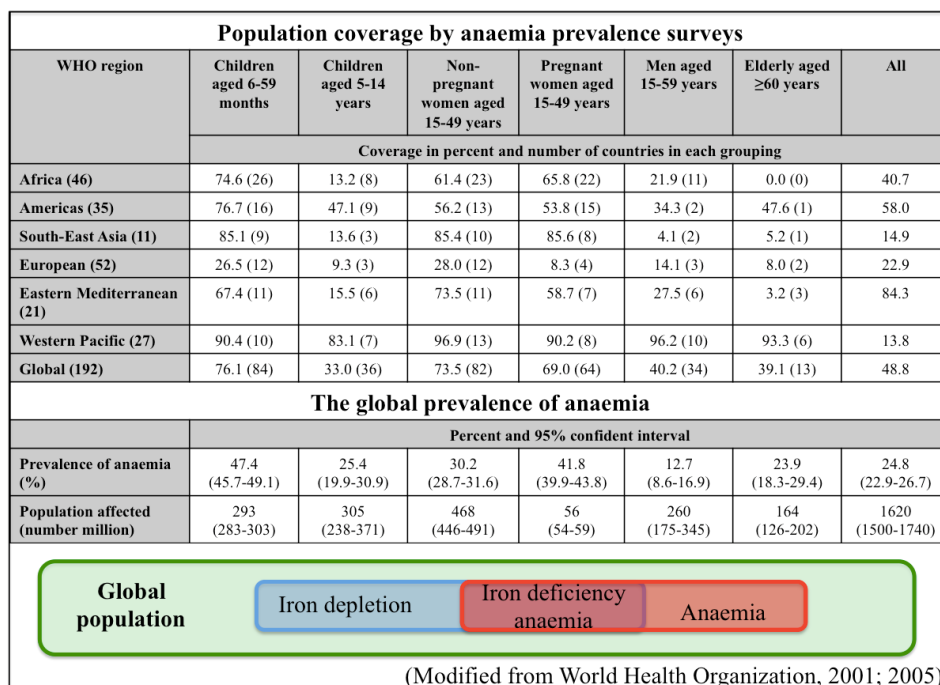
## **1.2 Iron deficiency anaemia**

Iron deficiency anaemia (IDA) is the most common cause of anaemia worldwide due to a deficient iron supply to produce functional haemoglobin (Hoffbrand et al., 2006; WHO, 2001). Causes of IDA are dietary iron deficiency, iron malabsorption, chronic blood loss from gastrointestinal bleeding, the maternal and perinatal period of iron deficiency anaemia and certain infectious diseases like malaria and hookworm infestations (Miller et al., 2013).

High-risk groups that are most vulnerable to iron deficiency are infants, adolescents, women of reproductive age, pregnant and breastfeeding women, postmenopausal women, elderly people in terms of physiological demand (age and gender-related factors); vegetarians in terms of dietary habit; patients with chronic renal failure undergoing haemodialysis and receiving erythropoietin in terms of pathological demand, individuals in resource-poor area in terms of socioeconomic influences and others (Hoffbrand et al., 2006; Provan, 2013). Impairment of oxygen delivery in iron deficiency anaemia may lead to weakness, lethargy, dyspnoea, unusual headaches, taste disturbances, difficulty in concentration, poor work capacity and productivity as well as decreased cognitive performance and physical development, are of major concerns (Kassebaum et al., 2014; Provan, 2013).

## **1.3 Prevalence of iron deficiency anaemia**

To date, there are no direct global estimates for iron deficiency, instead a comprehensive global estimate of anaemia based on haemoglobin level, has often been used as a proxy indicator of iron deficiency (WHO, 2001; WHO, 2015; Pasricha, 2010). Anaemia is a public health problem that affects almost two billion people globally, in both non-industrialised and industrialised countries, with 50% of the anaemic causes can be attributed by iron deficiency anaemia (IDA) (**Figure 1.2**) (WHO, 2001; WHO 2011). The prevalence of global anaemia was 32.9% (2010) and 29.4% (2011); respectively (Kassebaum et al., 2014; Pasricha, 2014; WHO, 2015). In Malaysia, almost two million women of reproductive age are anaemic (McLean et al., 2009).



**Figure 1.2: The surveys of global prevalence of anaemia in WHO region with respect to the number of countries and percentage of population involved.** Anaemia is the most common nutritional disorder in the world, mainly contributed by iron deficiency anaemia. Affected groups are children, pregnant women, all women, men and elderly (Modified from WHO, 2001; WHO, 2015).

In 2011, it is estimated that almost 800 million children and women suffered from anaemia globally, including 43% of children, 38% of pregnant women, 29% of non-pregnant women and 29% of all women of reproductive age (**Figure 1.3-1.6**) (WHO, 2015). Additional meta-analysis on the effect of iron supplementation is an approach to estimate the attribution of iron deficiency to the prevalence of anaemia, based on haemoglobin level (WHO, 2015). These analyses indicated that iron supplementation did improve around 42% of anaemic children and 50% anaemic women; respectively (WHO, 2015).

It is a serious public health problem whereby essential control strategies have to be implemented (WHO, 2001). In 2012, World Health Organization (WHO) has endorsed the second global nutrition target, in which by the year 2025, 50% reduction of anaemia in women of reproductive age (pregnant and non-pregnant women), i.e. from prevalence of anaemia 29.4% (2011) to 14.7% (2025) (WHO, 2014; WHO, 2015). **Figure 1.7-1.8** showed the latest prevalence and number cases of anaemia globally using WHO Nutrition Tracking Tool (WHO, 2016).

## Infants and children aged 6-59 months

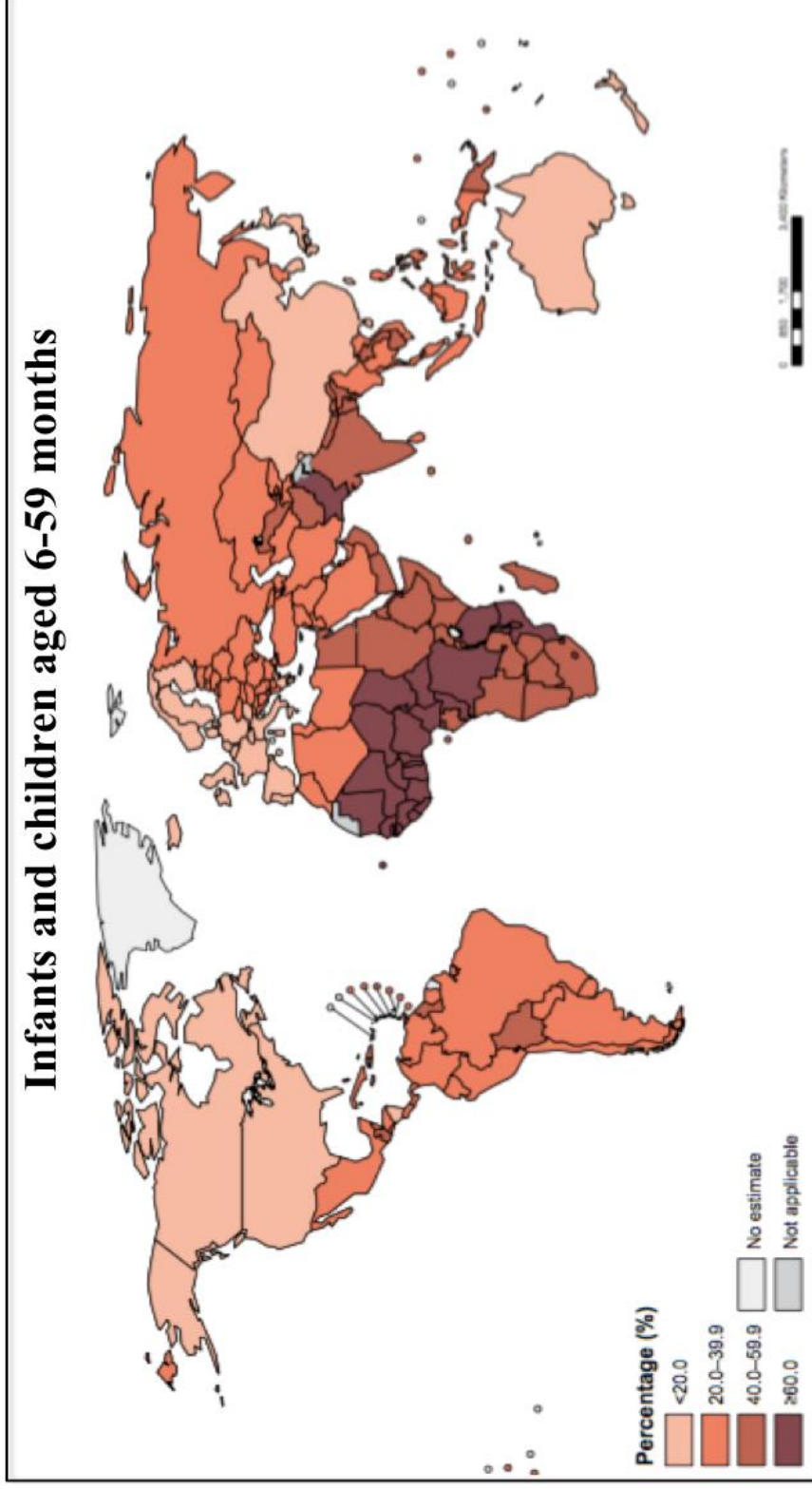


Figure 1.3: The global prevalence of anaemia in high-risk group of infants and children aged 6-59 months (Adapted from WHO, 2015).

## All women aged 15-49 years

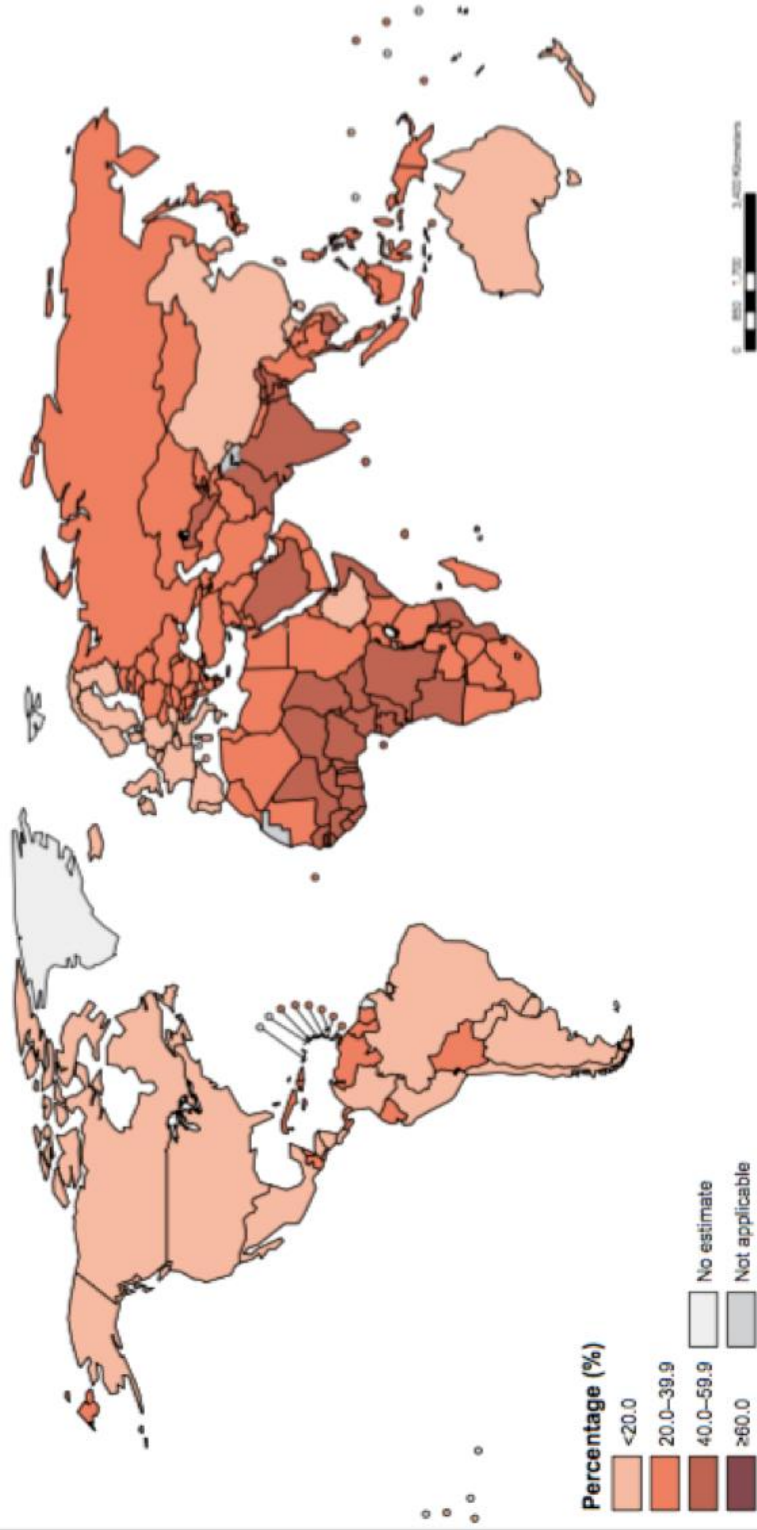


Figure 1.4: The global prevalence of anaemia in high-risk group of all women aged 15-49 years (Adapted from WHO, 2015).



## Pregnant women aged 15-49 years

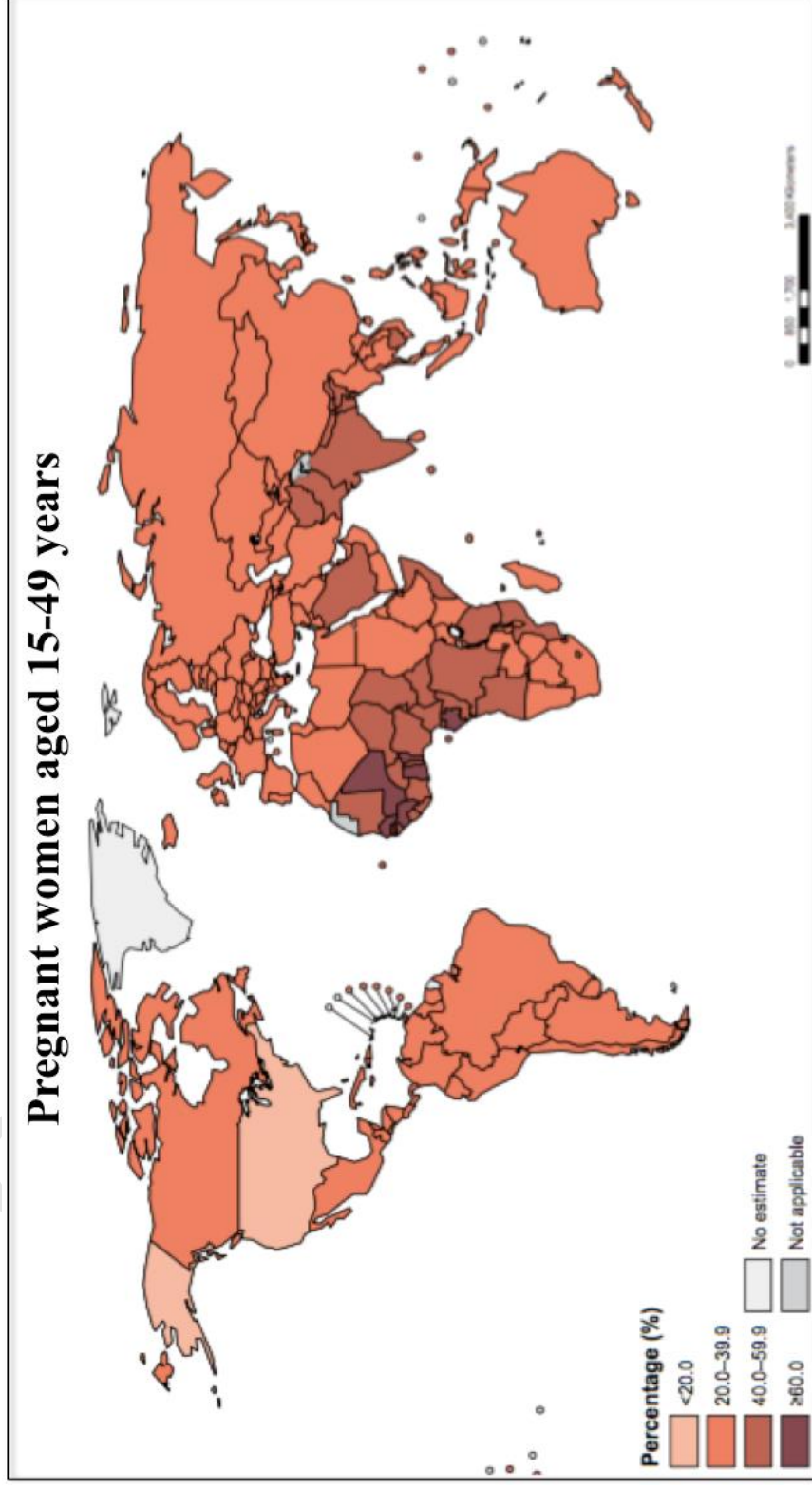


Figure 1.5: The global prevalence of anaemia in high-risk group of pregnant women aged 15-49 years (Adapted from WHO, 2015).

## Non-pregnant women aged 15-49 years

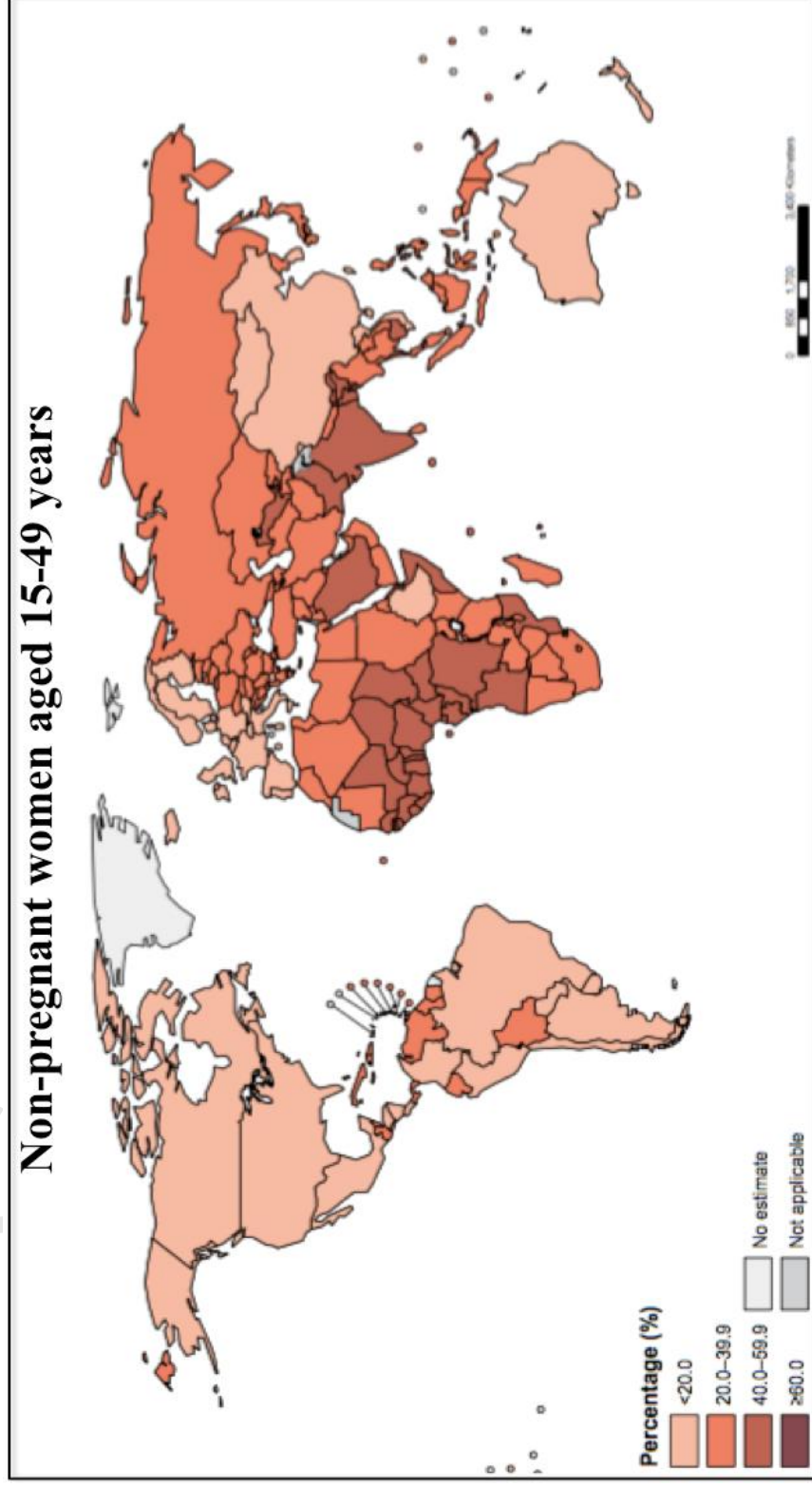
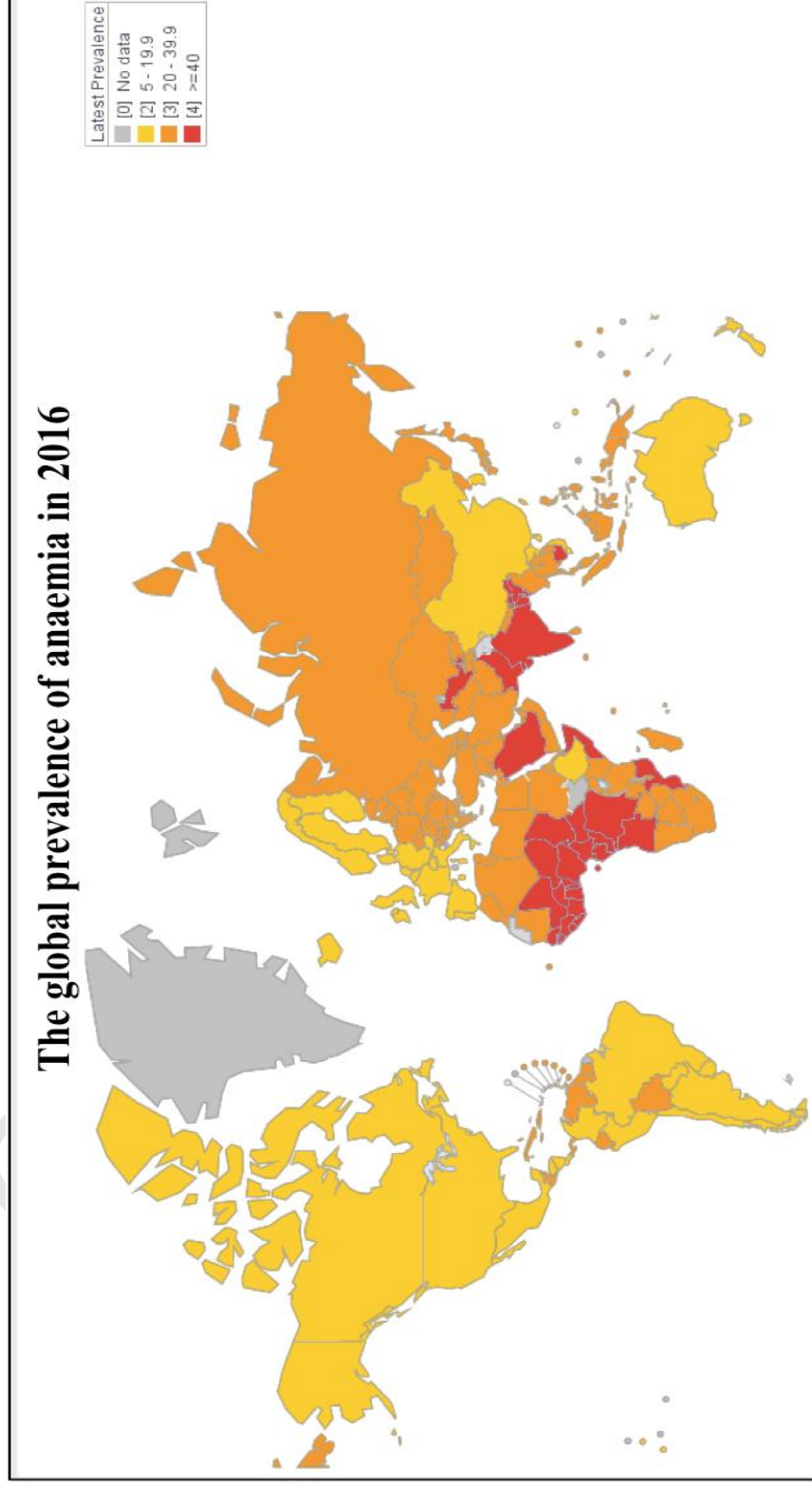


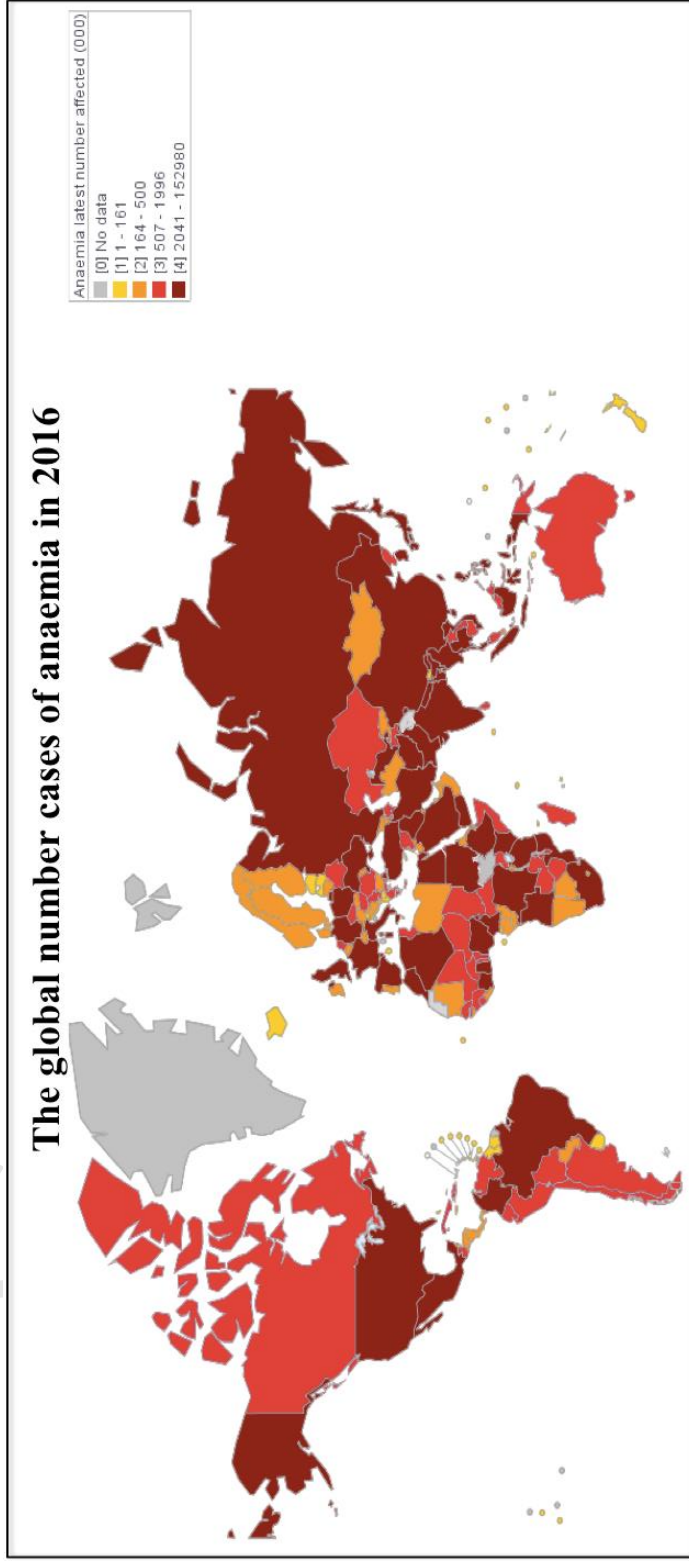
Figure 1.6: The global prevalence of anaemia in high-risk group of non-pregnant women aged 15-49 years (Adapted from WHO, 2015).

## The global prevalence of anaemia in 2016



**Figure 1.7: The global prevalence of anaemia in 2016** (Adapted from <http://www.who.int/nutrition/trackingtool/en/>, accessed on 2<sup>nd</sup> March 2016).





**Figure 1.8: The global number cases of anaemia in 2016** (Adapted from <http://www.who.int/nutrition/trackingtool/en/>, accessed on 2<sup>nd</sup> March 2016).

#### 1.4 Problem statement

Although there have been many efforts to improve nutritional well-being to eradicate iron deficiency anaemia, the condition is still common globally (WHO, 2001; WHO, 2005). Since the prevalence of iron deficiency is referred to the prevalence of anaemia based on haemoglobin levels, other causes for non-anaemic iron deficiency could have been easily underestimated (WHO, 2001; WHO, 2005).

Iron deficiency anaemia is characterised by smaller RBCs (microcytic) and reduced haemoglobin content (hypochromic) with pencil cells and target cells in the blood film and reduced mean cell volume (MCV) and mean cell haemoglobin (MCH) in red cell indices (Hoffbrand et al., 2006; Lewis et al., 2007; Provan, 2013). However, other diseases such as thalassaemia, chronic anaemia and sideroblastic anaemia have similar clinical symptoms; i.e. microcytic hypochromic anaemia in terms of blood film and red cell indices. Therefore, misdiagnosis is easy unless iron status is assessed.

To effectively fight iron deficiency anaemia, there is an urgent need to have better information in assessing iron status of populations, especially in rural areas (WHO, 2001; WHO, 2005). However, among currently available iron parameters, not one parameter alone can be used to confirm iron deficiency. Instead a combination of several indicators is needed for a definite conclusion (WHO, 2005).

Serum/plasma ferritin is commonly used to assess the body's iron status (Haskin et al., 1952). However, the serum/plasma ferritin test might only be available in some areas and have to be quantified using a biochemistry autoanalyser or enzyme-linked immunosorbent assay (ELISA). This assay requires trained staff for blood collection and assay runs. Not only that, the sample quality may be compromised during transportation and the results take a while to be released. Thus far, there is no low-cost and efficient iron tool for iron deficiency anaemia screening in market yet, especially in field studies. Furthermore, to target rural area, a simple iron tool might be useful, accompanying with simple blood collection (WHO, 2005).

Serum/plasma ferritin is an acute phase protein, rising with any inflammatory state. However, studies have shown that iron available in the ferritin could clearly distinguish those with iron overload from those with elevated ferritin due to inflammation, as the iron level is not affected by inflammation (Herbert et al., 1997).

Serum/plasma iron is abundant in blood plasma, which includes all iron, especially from transferrin-bound iron apart from ferritin-bound iron. Transferrin-bound iron is readily released in acidic condition while iron release from ferritin is not clear (Iron Panel of the International Committee for Standardization in Haematology, 1990). It was reported that there was only 25% of iron release from ferritin of  $>1000 \mu\text{g/L}$  using serum iron method according to the modifications to the Iron Panel of the International Committee for Standardization in Haematology (ICSH) reference method (Iron Panel of the International Committee for Standardization in Haematology, 1990).

## **1.5 Hypothesis**

The hypothesis of this project is that the concocted recipe is able to detect plasma ferritin iron and plasma iron spontaneously in individuals with iron deficiency, normal iron status and  $\beta$ -thalassaemia individuals with iron overload as a result from blood transfusion and increased iron absorption from the intestine.

## **1.6 Objectives**

The general objective is to develop a single-step plasma iron detection method.

The specific objectives are:

1. to identify a suitable method to rapidly detect iron from human plasma.
2. to quantify the iron status from a selected cohort of subjects.
3. to calculate the sensitivity and specificity of the screening test.

## REFERENCES

- Adams and Overman, E., 1909. The Reduction of Copper Sulphate with Hydroxylamin. *Journal of the American Chemical Society*, 31(6):637-640.
- Aisen, P., Enns, C., and Wessling-resnick, M., 2001. Chemistry and biology of eukaryotic iron metabolism. *The International Journal of Biochemistry & Cell Biology*, 33:940–959.
- Akahane, W., Ijiri, H., and Hanada, T., 2010. Method of Determining Iron Concentration. Patnet Number: US 7,785,892 B2. Retrieved 12 February 2017 from <https://docs.google.com/viewer?url=patentimages.storage.googleapis.com/pdfs/US7785892.pdf>
- Akobeng, A. K., 2006. Understanding diagnostic tests 1: sensitivity, specificity and predictive values. *Acta Paediatrica*, 96:338–341.
- Al-samarrai, A. H., Adaay, M. H., Al-tikriti, K. A., and Al-anzy, M. M., 2007. Evaluation of some essential element levels in thalassemia major patients in Mosul district, Iraq. *Saudi Medical Journal*, 29(1):94–97.
- Anderson, 1964. The Copper-catalysed Oxidation of Hydroxylamine. *Analyst*, 89:357-362.
- Andres, R. T., and Ill, F. S., 1993. Using the Electrician's Multimeter in the Chemistry Teaching Laboratory. *Journal of Chemical Education*, 70(9):514–517.
- AOAC, 2016. Appendix F: Guidelines for Standard Method Performance Requirements. Retrieved 12 February 2017 from [http://www.eoma.aoac.org/app\\_f.pdf](http://www.eoma.aoac.org/app_f.pdf)
- Archararit N, Chuncharunee S, Pornvoranunt A, Atamasirikul K, Rachakom B, Atichartakarn V., 2000. Serum C-reactive protein level in postsplenectomized thalassemic patients. *Journal of Medical Association of Thailand*, 83(Suppl1):S63-69.
- Arosio, P., Ingrassia, R., and Cavadini, P., 2009. Ferritins: A family of molecules for iron storage, antioxidation and more. *Biochimica et Biophysica Acta*, 1790:589–599.
- Aslan, D., Gümrük, F., Gürgey, A., and Altay, Ç, 2002. Importance of RDW value in differential diagnosis of hypochrome anemias. *American Journal of Hematology*, 69:31-33.
- ASTM. Standard Test Method for Iro in Trace Quantities Using the FerroZine Method. ASTM International. Retrieved 12 February 2017 from <ftp://185.72.26.245/Astm/2/01/Section%2015/ASTM1505/PDF/E1615.pdf>

- Aulakh, R., Sohi, I., Singh, T., and Kakkar, N., 2009. Red cell distribution width (RDW) in the diagnosis of iron deficiency with microcytic hypochromic anemia. *Indian Journal of Pediatrics*, 76:265-267.
- Australasian Association of Clinical Biochemists., 2013. Guidelines for the Evaluation of PoCT Instruments that Provide Quantitative Results.
- Azma, R. Z., Ainoon, O., Azlin, I., Hamenuddin, H., Hadi, N. A., Tatt, W. K., Syazana, I. N., Asmaliza, A. M., Das, S., and Hamidah, N. H., 2012. Prevalence of iron deficiency anaemia and thalassaemia trait among undergraduate medical students. *La Clinica Terapeutica*, 163(4):287-291.
- Åsberg, A. E., Mikkelsen, G., Aune, M. W., and Åsberg, A., 2013. Empty iron stores in children and young adults – the diagnostic accuracy of MCV, MCH, and MCHC. *International Journal of Laboratory Hematology*.
- Bableswhar, R.S., Roy, M., Bali, A., Patil, P.V., and Inumella, S., 2013. Intensive method of assessment and classification of the bone marrow iron status: A study of 80 patients. *Indian Journal Pathology Microbiology*, 56(1):16- 9.
- Bablok, W., and Passing, H., 1985. Application of Statistical Procedures in Analytical Instrument Testing. *Journal of Automatic Chemistry/Journal of Clinical Laboratory Automation*, 7(2):74-79.
- Backe, P. H., Ytre-Arne, M., Røhr, Å. K., Brodtkorb, E., Fowler, B., Rootwelt, H., Bjørås, M., and Mørkrid, L., 2013. Novel Deletion Mutation Identified in a Patient with Late-Onset Combined Methylmalonic Acidemia and Homocystinuria, cblC Type. *Journal of Inherited Metabolic Disease Reports*, 11:79–85.
- Battin, E. E., Lawhon, A., Brumaghim, J. L., and Hamilton, D. H., 2009. Using Proteins in a Bioinorganic Laboratory Experiment: Iron Loading and Removal from Transferrin. *Journal of Chemical Education*, 86(8):969–972.
- Bejjani, S., Pullakhandam, R., Punjal, R., and Nair, K. M., 2007. Gastric digestion of pea ferritin and modulation of its iron bioavailability by ascorbic and phytic acids in caco-2 cells. *World Journal of Gastroenterology*, 13(14):2083–2088.
- Bilić-Zulle, L., 2011. Comparison of methods: Passing and Bablok regression. *Biochemia Medica*, 21:49-52.
- Bland, J.M. and Altman, D., 1986. Statistical methods for assessing agreement between two methods of clinical measurement. *The lancet*, 327:307-310.
- Bou-Abdallah, F., McNally, J., Liu, X. X., and Melman, A., 2011. Oxygen Catalyzed Mobilization of Iron from Ferritin by Iron (III) Chelate Ligands. *Chemical Communications*, 47:731–733.
- Bovy, C., Tsobo, C., Crapanzano, L., Rorive, G., Beguin, Y., Albert, A., and Paulus, J., M., 1999. Factors determining the percentage of hypochromic red blood cells in hemodialysis patients. *Kidney International*, 56(3):113-119.

- Boyer, R. F., Clark, H. M., and LaRoche, A. P., 1988. Reduction and Release of Ferritin Iron By Plant Phenolics. *Journal of Inorganic Biochemistry*, 32:171–181.
- Braun, J., Lindner, K., Schreiber, M., Heidler, R. A., and Hörl, W. J., 1997. Percentage of hypochromic red blood cells as predictor of erythropoietic and iron response after i.v. iron supplementation in maintenance haemodialysis patients. *Nephrology Dialysis Transplantation*, 12:1173-1181.
- Briggs, C., Rogers, R., Thompson, B., and Machin, S. J., 2001. New red cell parameters on the Sysmex XE-2100 as potential markers of functional iron deficiency. *Sysmex Journal International*, 11(2):63-68.
- Bruce, G.R. and Gill, P.S., 1999. Estimates of precision in a standard additions analysis. *Journal of Chemical Education*, 76:805-807.
- Brugnara, C., Laufer, M. R., Friedman, A. J., Bridges, K., and Platt, O., 1994. Reticulocyte hemoglobin content (CHr): early indicator of iron deficiency and response to therapy [letter]. *Blood*, 83:3100-3101.
- Brugnara, C., Schiller, B., and Moran, J., 2006. Reticulocyte hemoglobin equivalent (Ret-He) and assessment of iron-deficient states. *Clinical Laboratory of Haematology*, 28:303-308.
- Brugnara, C., Zurakowski, D., DiCanzio, J., Boyd, T and Platt, O., 1999. Reticulocyte hemoglobin content to diagnose iron deficiency in children. *The Journal of American Medical Association*, 281(23):2225-2230.
- Bush, 2003. Why Doesn't My Heparinized Plasma Specimen Remain Anticoagulated? A Discussion on Latent Fibrin Formation in Heparinized Plasma, *Prenanalytical Solutions*, 13(2):9-10
- Buttarelli, M., and Pleani, M., 2008. Automated blood cell counts. *American Society for Clinical Pathology*, 130:104-116.
- Buttarelli, M., Temporin, V., Ceravolo, R., Farina, G., and Bulian, P., 2004. The new reticulocyte parameter (Ret-Y) of the Sysmex XE 2100. *American Society for Clinical Pathology*, 121-489-495.
- Cagle, F. W. M., and Smith, G. F., 1947. 2,2'-Bipyridine Ferrous Complex Ion as Indicator in the Determination of Iron. *Analytical Chemistry*, 19(6):384–385.
- Camacho, F., Páez, M. P., Jiménez, M. C., and Fernández, M., 1997. Application of the sodium dithionite oxidation to measure oxygen transfer parameters. *Chemical Engineering Science*, 52(8):1387–1391.
- Camaschella, C., 2015. Iron deficiency: new insights into diagnosis and treatment. *American Society of Hematology Education Program*, 8–13.
- Cassanelli, S., and Moulis, J., 2001. Sulfide is an efficient iron releasing agent for mammalian ferritins. *Biochimica et Biophysica Acta*, 1547:174–182.



- Cerioti, F. and Cerioti, G., 1980. Improved direct specific determination of serum iron and total iron-binding capacity. *Clinical chemistry*, 26:327-331.
- Cheong, S. K., Lim, Y. C., and Mok, K. L., 1991. A Freeze-dried Method for Preparation of G6PD Reagent Tubes. *Malaysian Journal of Pathology*, 13(1):51-52.
- Chesher, D., 2008. Evaluating Assay Precision. *The Clinical Biochemist Reviews*, 29:23-26.
- Chin, C. D., Linder, V., and Sia, S. K., 2012. Lab on a Chip Commercialization of microfluidic point-of-care diagnostic devices. *Lab on a Chip*, 12:2118-2134.
- Choi and Pai, 2001. Reticulocyte subpopulations and reticulocyte maturity index (RMI) rise as body iron status falls. *American Journal of Hematology*, 67:130-135.
- Choi and Pai, 2003. Association between serum transferrin receptor concentrations and erythropoietic activities according to body iron status. *Annals of Clinical and Laboratory Science*, 33(3):279-284.
- Choi, H. S., Song, S. H., Lee, J. H., Kim, H., and Yang, H. R., 2012. Serum hepcidin levels and iron parameters in children with iron deficiency. *The Korean Journal of Hematology*, 47(4):286-92.
- Cobas, 2015. Insert Fe Iron from the Roche/Hitachi analyser, 2015-03, Version 18.0. Code: 11965239001V18.
- Cook, J. D., Flowers, C. H., and Skikne, B. S., 2003. The quantitative assessment of body iron. *Blood*, 101:3359-3363.
- Cornbleet, P. J., and Gochman, N., 1979. Incorrect Least-Squares Regression Coefficients in Method- Comparison Analysis. *Clinical Chemistry*, 25(3):432-438.
- Cullen, O., Söffker, J., Höpfel, M., Bremer, C., Schlaghecken, R., Mehrens, T., Assmann, G, and Schaefer, R., M., 1999. Hypochromic red cells and reticulocyte haemoglobin content as markers of iron-deficient erythropoiesis in patients undergoing chronic haemodialysis. *Nephrology Dialysis Transplantation*, 14:659-665.
- Dale, J. C., Burritt, M. F., and Zinsmeister, A. R., 2002. Diurnal variation of serum iron, iron-binding capacity, transferrin saturation, and ferritin levels. *Clinical Chemistry*, 117:802-808.
- Delgado, J., Quintero-ortega, I. A., and Vega-gonzalez, A., 2014. From Voltage to Absorbance and Chemical Kinetics Using a Homemade Colorimeter. *Journal of Chemical Education*, 91(12):2158-2162.
- DeLong, E. R., DeLong, D. M., Clarke-Pearson, D. L., 1988. Comparing the areas under two or more correlated receiver operating characteristic curves: a nonparametric approach. *Biometrics*, 44:837-845.

- Dewitte, K., Fierens, C., Stöckl, D., and Thienpont, L. M., 2002. Application of the Bland–Altman Plot for Interpretation of Method- Comparison Studies: A Critical Investigation of Its Practice. *Clinical Chemistry*, 48(5):799–801.
- Dixit, M., and Kulkarni, P. K., 2011. Enhancing solubility and dissolution of Mefenamic acid by freeze drying. *Elixir International Journal*, 39:5026–5029.
- Dixit, M., Kini, A. G., Kulkarni, P. K., and Shivakumar, H. G., 2012. A Novel Technique To Enhancing The Solubility And Dissolution of Flutamide using Freeze Drying. *Turkish Journal of Pharmaceutical Sciences*, 9(2):139–149.
- Domenico, I. D., Vaughn, M. B., Li, L., Bagley, D., Musci, G., Ward, D. M., and Kaplan, J., 2005. Ferroportin-mediated mobilization of ferritin iron precedes ferritin degradation by the proteasome. *European Molecular Biology Organization*, 25:5396-5404.
- Donlin, M. J., Frey, R. F., Putnam, C., Proctor, J., and Bashkin, J. K., 1998. Analysis of Iron in Ferritin, the Iron-Storage Protein: A General Chemistry Experiment. *Journal of Chemical Education*, 75(4):437.
- Drees, J.C. and Wu, A.H., 2010. Analytical techniques. Clinical chemistry: techniques, principles, correlations. 6th ed. Lybrary of Congresss Cataloging in Publications Data. Baltimore 130-165.
- Duffy, J. R., and Gaudin, J., 1977. Copper interference in the determinatio of iron in serum using ferrozine. *Clinical Biochemistry*, 10(3):122-3.
- Earley, J. J., Kuivaniemi, H., Prockop, D. J., and Tromp, G., 1993. Efficient DNA sequencing on microtiter plates using dried reagents and Bst DNA polymerase. *The Journal of Sequencing and Mapping*, 4(2):79–85.
- Eivazi-Ziaei, J., Dastgiri, S., Pourebrahim, S., and Soltanpour, R., 2008. Usefulness of red blood cell flags in diagnosing and differentiating thalassemia trait from iron-deficiency anemia. *Hematology*, 13(4):253-256.
- Ellison, S.L. and Thompson, M., 2008. Standard additions: myth and reality. *Analyst* 133, 992-997.
- Elmagirbi, A., Sulistyarti, H., and Atikah, 2012. Study of ascorbic acid as iron(III) reducing agent for spectrophotometric iron speciation. *The Journal of Pure and Applied Chemistry Research*, 1(1):11-17.
- Empson, M. B., 2001. Statistics in the Pathology Laboratory; Characteristics of Diagnostic Tests. *Pathology*, 33:93-95.
- Feeney, G. P., Carter, K., Masters, G. S., Jackson, H. A., Cavil, I and Worwood, M., 2005. Changes in erythropoiesis in hereditary hemochromatosis are not mediated by HFE expression in nucleated red cells. *Haematologica*, 90(2):180-187.



- Fersht, A. R., and Petrovich, M., 2013. Reply to Campos and Munoz: Why phosphate is a bad buffer for guanidinium chloride titrations. *Proceedings of the National Academy of Sciences*, 110(14):1244–1245.
- Fomovska, G., Lo, R., and Perreault, S., 2008a. Assymetric Polysulfone Membranes for Plasma Separation From Whole Blood: Plasma Protein Binding/Recovery. Poster of Pall Life Sciences.
- Fomovska, G., Lo, R., and Smith, M. A., 2008b. Generation of High Quality Plasma for the Detection of Protein Biomarkers from the Whole Blood Using Vivid™Plasma Separation Membrane. Poster of Pall Life Sciences.
- Fomovska, G., Stanchfield, I., and Dubitsky, A., 2009. Vivid™Plasma Separation Membrane: Inter- and Intra-Lot Consistency for Plasma Separation for Lateral Flow Assays. Poster of Pall Life Sciences.
- Fomovsky, M., Fomovska, G., and Bormann, T., 2012. Centrifuge-free Isolation of Liquid Plasma from Clinical Samples of Whole Blood. Poster of Pall Life Sciences.
- Funk, F., Lenders, J-P., Crichton, R. R., and Schneider, W., 1985. Reductive mobilisation of ferritin iron. *European Journal of Biochemistry*, 152:167-172.
- Galanello, R., and Origa, R., 2010. Beta-thalassemia. *Orphanet Journal of Rare Diseases*, 5(11).
- Ganz, T., and Nemeth, E., 2012. Hepcidin and iron homeostasis. *Biochimica et Biophysica Acta*, 1823:1434–1443.
- Garcia-Granero, M., 2005. Lin's Concordance Correlation Coefficient. Retrieved 12 February, 2017 from <http://gjyp.nl/marta/>.
- Garcia-Mira, M. M., and Sanchez-ruiz, J. M. 2001. pH Corrections and Protein Ionization in Water/Guanidinium Chloride. *Biophysical Journal*, 81:3489–3502.
- George, E., Ng, M. L., and Tan, J. A. M. A., 2008. Erythrocyte Zinc Protoporphyrin in Beta-Thalassaemia Carriers. *Malaysian Journal of Medicine and Health Sciences*, 4(1):51-55.
- Ghasemi, A. and Zahediasl, S., 2012. Normality tests for statistical analysis: a guide for non-statisticians. *International journal of endocrinology and metabolism*, 10:486-489.
- Giavarina, D., 2015. Understanding bland altman analysis. *Biochemia Medica*, 25:141-151.
- Giraud, B., Frasca, D., Debaene, B. and Mimoz, O., 2013. Comparison of haemoglobin measurement methods in the operating theatre. *British journal of Anaesthesia*, 111:946-954.

- Good, N. E., Winget, G. D., Winter, W., Connolly, T. N., Izawa, S., and Singh, R. M. M., 1966. Hydrogen Ion Buffers for Biological Research. *Biochemistry*, 5(2):467–477.
- Gubala, V., Harris, L. F., Ricco, A. J., Tan, M. X., and Williams, D. E., 2012. Point of Care Diagnostics: Status and Future. *Analytical Chemistry*, 84(2):487–515.
- Gunasekaran, S., Natarajan, R. K., and Renganayaki, V., 2008. UV Visible Spectrophotometric Approach and Absorption Model for the Discrimination of Diseased Blood. *Asian Journal of Chemistry*, 20(1):48–54.
- Gunasekaran, S., and Sankari, G., 2004. FTIR and UV-Visible Spectral Study on Normal and Diseased Blood Samples. *Asian Journal of Chemistry*, 16(3-4):1779–1786.
- Gunasekaran, S., and Uthra, D., 2008. FTIR and UV-Visible Spectral Study on Normal and Jaundice Blood Samples. *Asian Journal of Chemistry*, 20(7):5695–5703.
- Gálvez, N., Ruiz, B., Cuesta, R., Colacio, E. and Domínguez-Vera, J.M., 2005. Release of iron from ferritin by aceto- and benzohydroxamic acids. *Inorganic chemistry*, 44: 2706–2709.
- Hajian-Tilaki, K., 2013. Receiver Operating Characteristic (ROC) Curve Analysis for Medical Diagnostic Test Evaluation. *Caspian Journal of Internal Medicine*, 4(2):627–635.
- Hall, T. G., Smukste, I., Bresciano, K. R., Wang, Y., McKearn, D., and Savage, R. E., 2012. Identifying and Overcoming Matrix Effects in Drug Discovery and Development, Tandem Mass Spectrometry - Applications and Principles, Dr Jeevan Prasain (Ed.), ISBN: 978-953-51-0141-3, InTech, Retrieved 12 February 2017 from <http://cdn.intechopen.com/pdfs/29012.pdf>
- Hamilton, L. D., Gubler, C. J., Cartwright, G. E., and Wintrobe, M. M., 1950. Diurnal variation in plasma iron level of man. *Proceedings of the Society of Experimental Biology and Medicine*, 75(1):65–68.
- Han and Kishimoto, 1997. Reticulocyte maturity index reflects erythropoietin effects in hemodialysis patients. *Osaka City Medical Journal*, 43(1):69–76.
- Hanneman, S.K., 2008. Design, analysis and interpretation of method-comparison studies. *AACN Advanced Critical Care*, 19:223.
- Harrington, J.M., Young, D.J., Essader, A.S., Sumner, S.J. and Levine, K.E., 2014. Analysis of human serum and whole blood for mineral content by ICP-MS and ICP-OES: development of a mineralomics method. *Biological trace element research*, 160:132–142.
- Harrison, P. M., 1986. The Structure and Function of Ferritin. *Biochemical Education*, 14(4):154–162.

- Harrison, P. M., and Arosio, P., 1996. The ferritins: molecular properties, iron storage function and cellular regulation. *Biochimica et Biophysica Acta*, 1275:161–203.
- Harrison, R.O. and Hammock, B.D., 1988. Location dependent biases in automatic 96-well microplate readers. *Journal Association of Official Analytical Chemists*, 71, 981-987.
- Harthoorn-Lasthuizen, E. J., Lindemans, J., and Langenhuijsen, M. M. A. C., 2009. Combined use of erythrocyte zinc protoporphyrin and mean corpuscular volume in differentiation of thalassemia from iron deficiency anemia. *European Journal of Haematology*, 60(4):245-251.
- Harvey, D., a Chapter 3 The Vocabulary of Analytical Chemistry. Analytical Chemistry 2.0, pp.41-62. Retrieved 12 February 2017 from [http://dpwadweb.depauw.edu/harvey\\_web/eTextProject/pdfFiles/Chapter3.pdf](http://dpwadweb.depauw.edu/harvey_web/eTextProject/pdfFiles/Chapter3.pdf)
- Harvey, D., b. Chapter 5 Standardizing Analytical Methods. Analytical Chemistry 2.0, pp.153- 208. Retrieved 12 February 2017 from <https://www.saylor.org/site/wp-content/uploads/2012/07/Chapter511.pdf>
- Haskins, D., Stevens, A. R., Finch, S., and Finch, C. A., 1952. Iron metabolism, iron stores in man as measured by phlebotomy. *The Journal of Clinical Investigation*, 543-547.
- Hem, 1960. Some Chemical Relationship among Sulfur Species and Dissolved Ferrous Iron. *Water Supply Paper*, 1459C:57-73.
- Hemocue, 2015. Hemocue® Hb 301 System. Retrieved 12 February 2017 from <http://www.hemocue.com/~media/hemocue-images/hemocuedotcom-images/product-images/hb/pdf-folders-etc/web-update-01092015.pdf?la=en>
- Hemocue. Hemocue® Urine Albumin Microcuvettes and the Hemocue® Urine Albumin Analyzer. Retrieved 12 February 2017 from [http://www.cliawaived.com/web/items/pdf/HMC-AL201ONE\\_Hemocue\\_Albumin\\_Product\\_Insert~2012file3.pdf](http://www.cliawaived.com/web/items/pdf/HMC-AL201ONE_Hemocue_Albumin_Product_Insert~2012file3.pdf)
- Hendriks, G., Uges, D.R.A. and Franke, J.P., 2008. pH adjustment of human blood plasma prior to bioanalytical sample preparation. *Journal of pharmaceutical and biomedical analysis*, 47:126-133.
- Hennessy, D. J., Reid, G. R., Smith, F. E., and Thompson, S. L., 1984. Ferene-a new spectrophotometric reagent for iron. *Canadian Journal of Chemistry*, 62:721–724.
- Herbert, V. D., Shaw, S., and Jayatilleke, E., 1996. Method for measuring total body tissue iron stores. United States Patent. Patent Number: 5,552,268. Retrieved 12 February 2017 from <https://docs.google.com/viewer?url=patentimages.storage.googleapis.com/pdfs/US5552268.pdf>

- Herbert, V., Jayatilleke, E., Shaw, S., Rosman, A. S., Giardina, P., Grady, R. W., Bowman, B., and Gunter, E. W., 1997. Serum ferritin iron, a new test, measures human body iron stores unconfounded by inflammation. *Stem cells*, 15:291-296.
- Herbert, V., Shaw, S., Jayatilleke, E., and Stopler-kasdan, T., 1994. Most Free-Radical Injury Is Iron-Related: It Is Promoted by Iron, Hemin, Holoferritin and Vitamin C, and Inhibited by Desferoxamine and Apoferritin. *Stem Cells*, 12:289–303.
- Hess, A. S., Shardell, M., Johnson, J. K., Thom, K. A., Strassle, P., Netzer, G., and Harris, A. D., 2012. Methods and recommendations for evaluating and reporting a new diagnostic test. *European Journal of Clinical Microbiology & Infectious Diseases*, 31(9):2111–2116.
- Hoffbrand, A. V., Moss, P. A. H., and Pettit, J. E., 2006. Essential Haematology, Fifth Edition. Blackwell Publishing.
- Hollis, S., 1996. Analysis of method comparison studies. *Annals of Clinical Biochemistry*, 33, 1–4.
- Hoppler, M., Schönbächler, A., Meile, L., Hurrell, R. F., and Walczyk, T., 2008. Ferritin-Iron Is Released during Boiling and In Vitro Gastric Digestion. *The Journal of Nutrition*, 138(5):878–84.
- Hoyer, K., 1944. Physiologic variations in the iron content of human blood serum. *Acta Medica Scandinavica*, 119:562-576.
- Huang, F., Johnson, C. M., Petrovich, M., and Fersht, A. R., 2013. Don' t waste good methods on bad buffers and ambiguous data. *Proceedings of the National Academy of Sciences*, 110(5):331–332.
- Hynes, M. J., and Coinceanainn, M. O., 2002. Investigation of the release of iron from ferritin by naturally occurring antioxidants. *Journal of Inorganic Biochemistry*, 90:18–21.
- Hinzmann., 2003. Iron Metabolism, Iron Deficiency and Anaemia. *Sysmex Journal International*, 13(2):65–74.
- Iron Panel of the International Committee for Standardization in Haematology, 1990. Revised recommendations for the measurements of the serum iron in human blood. *British Journal of Haematology*, 75:615-616.
- Jafarian-Dehkordi, A., Saghiaie, L., and Movahedi, N., 2008. A new spectrophotometric method for direct determination of iron (III) in serum. *Daru-Journal of Faculty of Pharmacy*, 16(2):76–82.
- Jokhio, R., Khan, Y., Chughtai, L. A., and Mughal, Z., 2009. C-reactive (CRP) Protein in Transfusion Dependent Thalassaemic Patients. *Pakistan Journal of Physiology*, 5(2):20–23.

- Jones, T., Spencer, R., and Walsh, C., 1978. Mechanism and Kinetics of Iron Release from Ferritin by Dihydroflavins and Dihydroflavin Analogues, 17(19):4011–4017.
- Kalra, K., 2011. Method Development and Validation of Analytical Procedures, Quality Control of Herbal Medicines and Related Areas, Prof. Yukihiro Shoyama (Ed.), ISBN: 978-953-307-682-9, InTech, Retrieved 22 January 2017 from <https://www.researchgate.net/file.PostFileLoader.html?id=57516dd1cbd5c2368e71535c&assetKey=AS:368859312869376@1464954321455>
- Shabir, G. A. Step-by-Step Analytical Methods Validation and Protocol in the Quality System Compliance Industry. *Institute of Validation Technology*, 4–14.
- Karagulle, M., Gündüz, E., Mutlu, F. Ş., and Akay, M., O., 2013. Clinical significance of reticulocyte hemoglobin content in the diagnosis of iron deficiency anemia. *Turkish Journal of Hematology*, 30:153-156.
- Karimi, S. A., McGarraugh, G., and Yu, Y. S., 1996. Control Solution for a Blood Glucose Monitor. United States Patent. Patent Number: 5,605,837. Retrieved 12 February 2017 from <https://docs.google.com/viewer?url=patentimages.storage.googleapis.com/pdfs/US5605837.pdf>
- Kassebaum, N. J., Jasrasaria, R., Naghavi, M., Wulf, S. K., Johns, N., Lozano, R., Regan, M., Weatherall, D., Chou, D. P., Eisele, T. P., Flaxman, S. R., Pullan, R. L., Brooker, S. J., and Murray, C. J. L., 2014. A systematic analysis of global anemia burden from 1990 to 2010. *Blood*, 123(5):615-624.
- Kemna, E. H. J. M., Tjalsma, H., Willems, H. L., and Swinkels, D. W., 2008. Hepcidin: from discovery to differential diagnosis. *Haematologica*, 93(1):90–97.
- Kidane, T. Z., Sauble, E., and Linder, M. C., 2005. Release of iron from ferritin requires lysosomal activity. *American Journal of Physiology Cell Physiology*, 291:C445-C455.
- Kim, M., Rho, Y., Jin, K. S., Ahn, B., Jung, S., Kim, H., and Ree, M., 2011. pH-Dependent Structures of Ferritin and Apoferritin in Solution: Disassembly and Reassembly. *Biomacromolecules*, 12:1629–1640.
- Konz, T., Añón-alvarez, E., Montes-bayon, M., and Sanz-medel, A., 2013. Antibody Labeling and Elemental Mass Spectrometry (Inductively Coupled Plasma-Mass Spectrometry) Using Isotope Dilution for Highly Sensitive Ferritin Determination and Iron-Ferritin Ratio Measurements. *Analytical Chemistry*, 85(17):8334–8340.
- Kotisaari, S., Romppanen, J., Penttilä, I., and Punnonen, K., 2002. The Advia 120 red blood cell and reticulocyte indices are useful in diagnosis of iron-deficiency anemia. *European Journal of Haematology*, 68:150-156.



- Kotz, L., Kaiser, G., Tschopel, P. and Tolg, G. 1972. Theory of sample preparation using acid digestion, pressure digestion and microwave digestion (microwave decomposition). *Analytical Chemistry*, 260:207-209.
- Kumar, R. and Indrayan, A., 2011. Receiver operating characteristic (ROC) curve for medical researchers. *Indian pediatrics*, 48:277-287.
- Kurz, G., Lopez-Calle, E., 2015. Multi-application Approach for Photometric Determination of An Analyte in a Fluid Sample on an Automated Analyzer. United States Patent Application Publication. Pub. No.: US 2015/0044780 A1. Retrieved 12 February 2017 from <http://patentimages.storage.googleapis.com/pdfs/US20150044780.pdf>
- Lalkhen, A. G., and McCluskey, A., 2008. Clinical tests: sensitivity and specificity. *Continuing Education in Anaesthesia, Critical Care & Pain*, 8(6):221–223
- Lamhaut, L., Apriotesei, R., Combes, X., Lejay, M., Carli, P. and Vivien, B., 2011. Comparison of the accuracy of noninvasive hemoglobin monitoring by spectrophotometry (SpHb) and HemoCue® with automated laboratory hemoglobin measurement. *The Journal of the American Society of Anesthesiologists*, 115:548-554.
- Lampinen, J., Raitio, M., Perälä, A., Oranen, H., and Harinen, R., 2012. Microplate Based Pathlength Correction Method for Photometric DNA Quantification Assay. Application Note of Thermo Scientific.
- Lenk, G., Hansson, J., Wijngaart, van der, Stemme, G., and Roxhed, N., 2015. Capillary Driven and Volume-metred Blood-plasma Separation. *Transducers*, 335–338.
- Lewis, S. M., Bain, B. J., and Bates, I. Dacie and Lewis PRACTICAL HAEMATOLOGY, Tenth edition, Churchill Livingstone.
- Li, M., Jia, X., Yang, J., Deng, J., and Zhao, G., 2012. Effect of tannic acid on properties of soybean (Glycine max) seed ferritin: A model for interaction between naturally-occurring components in foodstuffs. *Food Chemistry*, 133:410–415.
- LifeScan. SureStep® Technology. LifeScan makes getting accurate glucose results perfectly easy.
- Lim, W.F., Muniandi, L., George, E., Sathar, J., and The, L.K., Gan, G.G., and Lai, M.I., 2012.  $\alpha$ -haemoglobin stabilising protein expression is influenced by mean cell haemoglobin and HbF levels in HbE/ $\beta$ -thalassaemia individuals. *Blood Cells, Molecules, and Diseases*, 48(1):17-21.
- Linnet, K., 1993. Evaluation of Regression Procedures for Methods Comparison Studies. *Clinical Chemistry*, 39(3):424–432.

- Listowsky, I., Blauer, G., England, S., and Bethel, J. J., 1972. Denaturation of Horse Spleen Ferritin in Aqueous Guanidinium Chloride Solutions. *Biochemistry*, 11(11):2176–2182.
- Liu, J., Campos, L. A., Cerminara, M., Wang, X., Ramanathan, R., English, D. S., and Muñoz, V., 2012. Exploring one-state downhill protein folding in single molecules. *Proceedings of the National Academy of Sciences*, 109(1):179–184.
- Liu, X., 2012. Classification accuracy and cut point selection. *Statistics in Medicine*, 31:2676–2686.
- Liu, X., Jin, W., & Theil, E. C., 2003. Opening protein pores with chaotropes enhances Fe reduction and chelation of Fe from the ferritin biomineral. *Proceedings of the National Academy of Sciences*, 100(7):3653–3658.
- Lott, J.A. and Khabbaza, E., 1985. Haemoglobin analysis on whole blood by reflectance photometry. *Journal of Analytical Methods in Chemistry*, 7:197–200.
- Ludbrook, J., 2010. Linear regression analysis for comparing two measurers or methods of measurement: But which regression? *Clinical and Experimental Pharmacology and Physiology*, 37:692–699.
- Macdougall, 1998. What is the most appropriate strategy to monitor functional iron deficiency in the dialyzed patient on rhEPO therapy? Merits of percentage hypochromic red cells as a marker of functional iron deficiency. *Nephrology Dialysis Transplantation*, 13:847–849.
- Macdougall, I. C., Cavill, I., Hulme, B., Bain, B., McGregor, E., McKay, P., Sanders, E., Coles, G. A., and Williams, J. D., 1992. Detection of functional iron deficiency during erythropoietin treatment: a new approach. *British Medical Journal*, 304:225–226.
- Magari, R.T., 2002. Statistics for laboratory method comparison studies. *BioPharm*, 15:28–34.
- Magge, H., Sprinz, P., Adams, W. G., Drainoni, M-L., and Meyers, A., 2013. Zinc Protoporphyrin and Iron Deficiency Screening. *JAMA Pediatrics*, 167(4):361–367.
- Malakar, R., Kour, M., Ahmed, A., Malviya, S. N., and Dangi, C. B. S., 2014. Trace Elements Ratio in Patients of Haemoglobinopathy. *International Journal of Current Microbiology and Applied Sciences*, 3(6):81–92.
- Mannheim, B., 1985. Application of statistical procedures in analytical instrument testing. *Journal of Automatic Chemistry/Journal of Clinical Laboratory Automation*, 7(2):74–79.
- Marinova, M., and Vladimirova, L., 2009. Atomic Absorption Assessment of Mineral Iron Quantity in Ferritin. *Bulgarian Journal of Physics*, 43:192–195.

- Martinez, A. W., Phillips, S. T., and Whitesides, G. M., 2010. Diagnostics for the Developing World: Microfluidic Paper-Based Analytical Devices. *Analytical Chemistry*, 82:3–10.
- May, M. E., and Fish, W. W., 1978. The UV and Visible Spectral Properties of Ferritin. *Archives of Biochemistry and Biophysics*. 190(2):720-725.
- McGown, 2000. UV absorbance measurements of DNA in microplates. *BioTechniques*, 28:60-64.
- McKenna, C. E., Gutheil, W. G., and Song, W., 1991. A method for preparing analytically pure sodium dithionite. Dithionite quality and observed nitrogenase-specific activities. *Biochimica et Biophysica Acta*, 1075:109–117.
- McLean E, Cogswell M, Egli I, Wojdyla D and de Benoist B., 2009. Worldwide prevalence of anaemia, WHO Vitamin and Mineral Nutrition Information System, 1993-2005. *Public Health Nutrition*, 12: 444-454.
- MedCalc., 2017. Passing-Bablok regression. [https://www.medcalc.org/manual/passing-bablok\\_regression.php](https://www.medcalc.org/manual/passing-bablok_regression.php)
- Melman, G., Bou-abdallah, F., Vane, E., Maura, P., Arosio, P., and Melman, A., 2013. Iron release from ferritin by flavin nucleotides. *Biochimica et Biophysica Acta*, 1830:4669–4674.
- Metzgeroth, G., Adelberger, V., Dorn-Beineke, A., Kuhn, C., Schatz, M., Maywald, O., Bertsch, T., Wisser, H., Hehlmann, R., and Hastka, J., 2005. Soluble transferrin receptor and zinc protoporphyrin – competitors or efficient partners? *European Journal of Haematology*, 75(4):309-317.
- Miller, 2013. Iron deficiency anemia: a common and curable disease. *Cold Spring Harbor Perspectives in Medicine*, 3:a011866.
- Miwa, N., Akiba, T., Kimata, N., Hamaguchi, Y., Arakawa, Y., Tamura, T., Nitta, K., and Tsuchiya, K., 2009. Usefulness of measuring reticulocyte hemoglobin equivalent in the management of haemodialysis patients with iron deficiency. *International Journal of Laboratory Hematology*, 32:248-255.
- Mohsen, U. A., Elaish, K. I. A., and Issa, M., 2013. Determination of Iron in Blood Serum by Spectrophotometric and Atomic Absorption Methods as a Comparative Study. *Cukurova Medical Journal*, 38(3):358–364.
- Monadi, M., Firouzjahi, A., Hosseini, A., Javadian, Y., Sharbatdaran, M., and Heidari, B., 2016. Serum C-reactive protein in asthma and its ability in predicting asthma control, a case-control study. *Caspian Journal of Internal Medicine*, 7(1):37–42.
- Morey, T. E., Gravenstein, N., and Rice, M. J., 2011. Let's Think Clinically Instead of Mathematically About. *Anesthesia & Analgesia*, 113(1)89–91.
- Mwangi, M. N., Maskey, S., Andang'o P. EA., Shinali, N. K., Roth, J. M., Trijsburg, L., Mwangi, A. M., Zuilhof, H., van Lagen, B., Savelkoul, H FJ., Demir, A.



- Y., Verhoef, H., 2014. Diagnostic utility of zinc protoporphyrin to detect iron deficiency in Kenyan pregnant women. *BioMed Central Medicine*, 12(229).
- Naing, L., Winn, T., and Rusli, B. N., 2006. Practical issues in calculating the sample size for prevalence studies. *Medical Statistics*, 1:9-14.
- Nandagopalan. How to Construct and Use Error Grids for Evaluating Quantitative Diagnostic Assays. Clinical and Laboratory Standards Institute. Retrieved 12 February 2017  
<http://eo2.commpartners.com/users/clsi/downloads/121025CLSIHANDOUT.pdf>
- Nandagopalan, S., Carey, R. N., Levine, J. B., Miller, W. G., and Pennello, G., 2012. How to Construct and Use Error Grids for Evaluating Quantitative Diagnostic Assays; Approved Guideline. CLSI document EP27-A. Retrieved 12 February 2017 from [http://shop.clsi.org/site/Sample\\_pdf/EP27AE\\_sample.pdf](http://shop.clsi.org/site/Sample_pdf/EP27AE_sample.pdf)
- Naqvi. H., Naqvi, Faizan-ul-Hassan, Naqvi, I. H. ,Farhan, M., Abbas, T., Yang, L., and Gul, A., 2014. Serum Hcpidin: Its Correlation with Serum Ferritin, Serum Iron and Hemoglobin in Patients of Iron Deficiency Anemia. *Immunology, Endocrine & Metabolic Agents in Medicinal Chemistry*, 14(20):105-113.
- Nesa, A., Tayab, M. A., Sultana, T., Khondker, L., Rahman, Q., Karim, A., and Ahmed, A. N. N., 2009. RDWI is Better Discriminant than RDW in Differentiation of Iron Deficiency Anaemia and Beta Thalassaemia Trait, *Bangladesh Journal of Child Health*, 33(3):100-103.
- Neufeld, L., Garcia-Guerra, A., Sacher-Francia, D., Newton-Sachez, O., Ramiez-Villalobos, M. D., Rivera-Dommarco, J., 2002. Hemoglobin measured by Hemocue and a reference method in venous and capillary blood: a validation study. *Salud Pública de México*, 44(3):219-227.
- Nickerson, C. A. E., 2012. A Note on “A Concordance Correlation Coefficient to Evaluate Reproducibility.” *Biometrics*, 53(4):1503–1507.
- Nkrumah, B., Nguah, S. B., Sarpong, N., Dekker, D., Idriss, A., May, J., and Adu-Sarkodie, Y., 2011. Hemoglobin estimation by the HemoCue® portable hemoglobin photometer in a resource poor setting. *BMC Clinical Pathology*, 11(5).
- Nóbrega, J. A., Pirola, C., Fialho, L. L., Rota, G. de Campos Jordão, C. E., and Pollo, P., 2012. Microwave-assisted digestion of organic samples: How simple can it become? *Talanta*, 98:272–276.
- Olatunya, O. S., Olu-taiwo, A., Ogundare, E. O., Oluwayemi, I. O., Olaleye, A. O., Fadare, J. O., Adekoya-Benson, T., Fatunla, O., Agaja, O. T., Omoniyi, E., and Oluwadiya, K. S., 2016. Evaluation of a Portable Haemoglobin Metre Performance in Children with Sick Cell Disease and Implications for Healthcare in Resource-poor Settings. *Journal of Tropical Pediatrics*, 0:1–8.

- Ong, K.H., Tan, H.L., Lai, H.C. and Kuperan, P., 2005. Accuracy of various iron parameters in the prediction of iron deficiency in an acute care hospital. *Annals-Academy of Medicine Singapore*, 34:437.
- Oskam, I. C., Ropstad, E., Andersen, B. K., Fredriksen, B., Larsen, S., Dahl, E., Andresen, O., 2008. Testicular germ cell development in relation to 5 $\alpha$ -androstenedione levels in pubertal entire male pigs. *Theriogenology*, 69:967–76.
- Oyane, A., Kim, H., Furuya, T., Kokubo, T., Miyazaki, T., and Nakamura, T., 2003a. Preparation and assessment of revised simulated body fluids. *Journal of Biomedical Materials Research Part A*, 65(2):188–95.
- Oyane, A., Onuma, K., Ito, A., Kim, H., Kokubo, T., and Nakamura, T., 2003b. Formation and growth of clusters in conventional and new kinds of simulated body fluids. *Journal of Biomedical Materials Research Part A*, 64(2):339–48.
- O'Shaughnessy, D. F., Atterbury, C., Bolton Maggs, P., Murphy, M., Thomas, D., Yates, S., Williamson, L. M., British Committee for Standards in Haematology and Blood Transfusion Task Force, 2004. Guidelines for the use of fresh-frozen plasma, cryoprecipitate and cryosupernatant. *British Journal of Haematology*, 126(1):11–28.
- Pall, 2009. Optimised, highly efficient membrane for one-step plasma separation from whole blood without the use of centrifugation. Product Data of Pall Life Sciences.
- Palmer, K. F., and Williams, D., 1974. Optical properties of water in the near infrared. *Journal of the Optical Society of America*, 64(8):1107–1110.
- Parkes, J. L., Slatin, S. L., Pardo, S., and Gindberg, B. H., 2000. A New Consensus Error Grid to Evaluate the Clinical Significance of Inaccuracies in the Measurement of Blood Glucose. *Diabetes Care*, 23(8):1143–1148.
- Pasricha, S., 2014. Anemia: a comprehensive global estimate. *Red Cells, Iron & Erythropoiesis*, 123(5):611–612.
- Pasricha, S., Mcquillen, Z., Westerman, M., Keller, A., Nemeth, E., Ganz, T and Wood, E., 2011. Serum hepcidin as a diagnostic test of iron deficiency in premenopausal female blood donors. *Haematologica*, 96(8):1099–1105.
- Parischa, S-R. S., Flecknoe-Brown, S. C., Allen, K. J., Gibson, P. R., McMahon, L. P., Olynyk, J. K., Roger, S. D., Savoia, H. F., Tampi, R., Thomson, A. R., Wood, E. M., and Robinson, K. L., 2010. Diagnosis and management of iron deficiency anaemia: a clinical update. *Medical Journal of Australia*, 193(9):525-532.
- Passing, H. and Bablok, W., 1983. A new biometrical procedure for testing the equality of measurements from two different analytical methods. Application of linear regression procedures for method comparison studies in clinical chemistry, Part I. *Clinical Chemistry and Laboratory Medicine*, 21:709-720.

- Phiri, K. S., Calis, J. C. J., Kachala, D., Borgstein, E., Waluza, J., Bates, I., Brabin B., and van Hensbroek, M. B., 2009. Improved method for assessing iron stores in the bone marrow. *Journal of Clinical Pathology*, 62, 685–689.
- Pilon, V. A., Howanitz, P. J., Howanitz, J. H., and Domres, N., 1981. Day-to-Day Variation in Serum Ferritin Concentration in Healthy Subjects. *Clinical Chemistry*, 27(1):78–82.
- Pisaruka, J., and Dymond, M. K., 2016. A low volume 3D-printed temperature-controllable cuvette for UV visible spectroscopy. *Analytical Biochemistry*, 510:52–55.
- Plath, L. D., Ozdemir, A., Aksenov, A. A., and Bier, M. E., 2015. Determination of Iron Content and Dispersity of Intact Ferritin by Superconducting Tunnel Junction Cryodetection Mass Spectrometry. *Analytical Chemistry*, 87:8985–8993.
- Pujara, K. M., Bhalara, R. V., and Dhruva, G. A., 2014. A study of bone marrow iron storage in hematological disorder. *International Journal of Health & Allied Sciences*, 3(4), 221–224.
- Punnonen, K., Irjala, K., and Rajamäki, A., 1997. Serum transferrin receptor and its ratio to serum ferritin in the diagnosis of iron deficiency. *Blood*, 89(3):1052-7.
- Puretec Industrial Water. The Relationship between pH and Deionized Water. Retrieved 12 February 2017 from <http://puretecwater.com/downloads/relationship-between-ph-and-deionized-water.pdf>
- Provan, D. Chapter 1 Iron deficiency anaemia. Retrieved 22 January 2017 from [http://www.blackwellpublishing.com/content/BPL/Images/Content\\_store/Sample\\_Chapter/9781405153539/9781405153539\\_4\\_001.pdf](http://www.blackwellpublishing.com/content/BPL/Images/Content_store/Sample_Chapter/9781405153539/9781405153539_4_001.pdf)
- Raja, S.O., Shaw, J., Chattopadhyay, A., Chatterjee, S., Bhattacharya, M. and Dasgupta, A.K., 2014. Synchronous fluorescence based one step optical method for assessing oxidative stress and its dependence on serum ferritin. *Analytical Methods*, 6:6228-6231.
- Ramsay, W. N., 1952. The Determination of Iron in Blood Plasma or Serum. *Biochemical Journal*, 53(2):227–231.
- Ren, Y., and Walczyk, T., 2014. Quantification of ferritin bound iron in human serum using species-specific isotope dilution mass spectrometry. *Metallomics*, 6:1709–1717.
- Rich, D., 2005. The Removal of Total Phosphorus from Natural Waters by Precipitation, PhD Thesis, Northwestern International University.
- Salvi, G., Rios, P. D. L., and Vendruscolo, M., 2005. Effective Interactions Between Chaotropic Agents and Proteins. *Proteins*, 61:492–499.

- Schmid, F-X., 2001. Biological Macromolecules: Spectrophotometry Concentrations. *Encyclopedia of Life Sciences*.
- Schobert, B., and Tschesche, H., 1978. Unusual solution properties of proline and its interaction with proteins. *Biochimica et Biophysica Acta*, 541(2):270-7.
- Schuepbach, R.A., Bestmann, L., Béchir, M., Fehr, J. and Bachli, E.B., 2011. High prevalence of iron deficiency among educated hospital employees in Switzerland. *International Journal of Biomedical Science*, 7:150.
- Schweitzer, L., and Cornett, C., 2008. Determination of Heavy Metals in Whole Blood Using Inductively-Coupled Plasma-Mass Spectrometry: A Comparison of Microwave and Dilution Techniques. *The Big M*, 4:75-83.
- Seise, B., Pollok, S., Seyboldt, C., and Weber, K., 2013. Dry-reagent-based PCR as a novel tool for the rapid detection of *Clostridium* spp. *Journal of Medical Microbiology*, 62:1588–1591.
- Shah, P.P., Desai, S.A., Modi, D.K. and Shah, S.P., 2014. Assessing diagnostic accuracy of Haemoglobin Colour Scale in real-life setting. *Journal of health, population, and nutrition*, 32:51.
- Sheftel, A. D., Mason, A. B., and Ponka, P., 2012. The long history of iron in the Universe and in health and disease. *Biochimica et Biophysica Acta*, 1820:161–187.
- Sundberg, R. D., and Broman, H., 1955. The Application of the Prussian Blue Stain to Previously Stained Films of Blood and Bone Marrow. *Blood*, 10:160–166.
- Shrivastava, A., and Gupta, V. B., 2011. Methods for the determination of limit of detection and limit of quantitation of the analytical methods. *Chronicles of Young Scientists*, 2(1):21–25.
- Shrout, P. E., and Fleiss, J. L., 1979. Intraclass Correlations : Uses in Assessing Rater Reliability. *Psychological Bulletin*, 86(2):420–428.
- Sirivech, B. S., Frieden, E., and Osaki, S., 1974. The Release of Iron from Horse Spleen Ferritin by Reduced Flavins. *Biochemical Journal*, 143:311–315.
- Skikne, B. S., Flower, C. H., and Cook, J. D., 1990. Serum transferrin receptor: a quantitative measure of tissue iron deficiency. *Blood*, 75:1870–1876.
- Statland, B. E., Winkel, P., and Bokelund, H., 1976. Variation of serum iron concentration in young healthy men: Within-day and day-to-day changes. *Clinical Biochemistry*, 9(1):26–29.
- Stojanović, M., Apostolović, M., Stojanović, D., Milošević, Z., Toplaović, A., Lakušić, V. M., and Golubović, M., 2014. Understanding sensitivity, specificity and predictive values. *Practical Advices for Physicians*, 71(11)1062–1065.

- Stookey, L. L., 1970. Ferrozine-A New Spectrophotometric Reagent for Iron. *Analytical Chemistry*, 42(7):779–781.
- Stumm, W., and Lee, G. F., 1961. Oxygenation of Ferrous Iron. *Industrial & Engineering Chemistry*, 53(2):143–146.
- Sultana, G. S., Haque, S. A., Sultana, T., Rahman, Q., and Ahmed, A. N. N., 2011. Role of red cell distribution width (RDW) in the detection of iron deficiency anaemia in pregnancy within the first 20 weeks of gestation. *Bangladesh Medical Research Council Bulletin*, 37:102–105.
- Swinehart, D. F., 1962. The Beer-Lambert. *Journal of Chemical Education*, 39(7):333–335.
- Sánchez, P., Gálvez, N., Colacio, E., Miñones, E., and Domínguez-Vera, J., 2005. Catechol releases iron(III) from ferritin by direct chelation without iron(II) production. *Dalton Transactions*, 21(4):811–813.
- Theil, E. C., 1987. Ferritin: Structure, Gene Regulation, and Cellular Function in Animals, Plants, and Microorganisms. *Annual Review of Biochemistry*, 56:289–315.
- Theil, E. C., 2013. Ferritin: The Protein Nanocage and Iron Biomineral in Health and in Disease. *Inorganic Chemistry*, 52(21):12223–33.
- Thermo Scientific, 2010. Thermo Scientific SkanIt Software 3.2 for Multiskan® GO. User Manual of Thermo Scientific.
- Thomas, C., Kirschbaum, A., Boehm, D., and Thomas, L., 2006. The Diagnostic Plot. *Medical Oncology*, 23(1):23–36.
- Thomas, C., and Thomas, L., 2002. Biochemical Markers and Hematologic Indices in the Diagnosis of Functional Iron Deficiency. *Clinical Chemistry*, 48(7):1066–1076.
- Tietz, N. W., Rinker, A. D., and Morrison, S. R., 1996. When is a serum iron really a serum iron? A follow-up study on the status of iron measurements in serum. *Clinical Chemistry*, 42(1):109–111.
- Tietz, W., Rinker, A. D., and Morrison, R., 1994. When Is a Serum Iron Really a Serum Iron? The Status of Serum Iron Measurements. *Clinical Chemistry*, 40(4):546–551.
- Topham, R., Goger, M., Pearce, K., and Schultz, P., 1989. The mobilization of ferritin iron by liver cytosol A comparison of xanthine and NADH as reducing substrates. *Biochemical Journal*, 261:137–143.
- Trnka, H., Rantanen, J., and Grohgan, H., 2015. Well-plate freeze-drying: a high throughput platform for screening of physical properties of freeze-dried formulations. *Pharmaceutical Development and Technology*, 20(1):65–73.



- Turgut, S., Hacio, S., Emmungil, G., Turgut, G., and Keskin, A., 2009. Relations between Iron Deficiency Anemia and Serum Levels of Copper, Zinc, Cadmium and Lead. *Polish Journal of Environmental Studies*, 18(2):273–277.
- Twomey, P.J. and Kroll, M.H., 2008. How to use linear regression and correlation in quantitative method comparison studies. *International Journal of Clinical Practice*, 62:529-538.
- Ullrich, C., Wu, A., Armsby, C., Rieber, S., Wingerter, S., Brugnara, C., Shapiro, D., and Bernstein, H., 2005. Screening healthy infants for iron deficiency using reticulocyte hemoglobin content. *The Journal of American Medical Association*, 294(8):924-930.
- UNODC, 2009. Guidance for the Validation of Analytical Methodology and Calibration of Equipment Used for Testing of Illicit Drugs in Seized Materials and Biological Specimens. United Nations, New York.
- Uppal, V. and Uppal, N., 2015. An Unrecorded Pre-Pre-Analytical Error in Serum Iron Analysis. *Journal of clinical and diagnostic research*, 9(11):BL01.
- Urrechaga, E., Borque, L., and Escanero, J. F., 2009. Potential utility of the new Sysmex XE 5000 red blood cell extended parameters in the study of disorders of iron metabolism. *Clinical Chemistry and Laboratory Medicine*, 47(11):1411-1416.
- Urrechaga, E., Borque, L., and Escanero, J. F., 2012. Percentage of hypochromic erythrocytes as a potential marker of iron availability. *Clinical Chemistry and Laboratory Medicine*, 50(4):685-687.
- Urrechaga, E., Borque, L., and Escanero, J. F., 2012. Erythrocyte and reticulocyte indices in the assessment of erythropoiesis activity and iron availability. *International Journal of Laboratory Hematology*, 35:144-149.
- Urrechaga, E., Borque, L., and Escanero, J. F., 2013. Biomarkers of hypochromia: the contemporary assessment of iron status and erythropoiesis. *BioMed Research International*, Article ID 603786.
- Ventola, C. L., 2014. Medical Applications for 3D Printing: Current and Projected Uses. *Pharmacy and Therapeutics*, 39(10):704–711.
- Verschoor and Molot, 2013. A comparison of three colorimetric methods of ferrous and total retractive iron measurement in freshwaters. *Limnology and Oceanography: Methods*, 11:113-125.
- Vidyashankar, P., Alan, F. A., Niwruiti, K. H., Arun, H., Harinakshi, R., and Bhusri, S., 2013. Diagnosis of iron deficiency of chronic kidney disease: validity of iron parameters, reticulocytes hemoglobin content (CHr) and hypochromic red cells in inflammatory state. *International Journal of Current Research and Review*, 05(21):83-91.

- Viollier, E., Inglett, P. W., Hunter, K., Roychoudhury, A. N., and Cappellen, P. Van., 2000. The ferrozine method revisited : Fe(II)/ Fe(III) determination in natural waters. *Applied Geochemistry*, 15:785–790.
- Walter, P.B., Macklin, E. A., Porter, J., Evans, P., Kwiatkowski, J. L., Neufeld, E. J., Coates, T., Giardina, P. J., Vichinsky, E., Olivieri, N., Alberti, D., Holland, J., and Harmatz, P., 2008. Inflammation and oxidant-stress in  $\beta$ -thalassemia patients treated with iron chelators deferasirox (ICL670) or deferoxamine: an ancillary study of the Novartis CICL670A0107 trial. *Haematologica*, 93(6):817-825.
- Wang, A., Zhou, K., Qi, X., and Zhao, G., 2014. Phytoferritin Association Induced by EGCG Inhibits Protein Degradation by Proteases. *Plant Foods for Human Nutrition*, 69:386–391.
- Watson, P. F., and Petrie, A., 2010. Method agreement analysis: A review of correct methodology. *Theriogenology*, 73:1167–1179.
- Wentholt, I.M., Hoekstra, J.B. and DeVries, J.H., 2006. A critical appraisal of the continuous glucose–error grid analysis. *Diabetes Care*, 29:1805-1811.
- WHO, 2001. Iron Deficiency Anaemia: Assessment, Prevention, and Control. Retrieved 22 January 2017 from [http://apps.who.int/iris/bitstream/10665/66914/1/WHO\\_NHD\\_01.3.pdf?ua=1](http://apps.who.int/iris/bitstream/10665/66914/1/WHO_NHD_01.3.pdf?ua=1)
- WHO, 2007. Assessing the iron status of populations. Retrieved 22 January 2017 from [http://www.who.int/nutrition/publications/micronutrients/anaemia\\_iron\\_deficiency/9789241596107.pdf](http://www.who.int/nutrition/publications/micronutrients/anaemia_iron_deficiency/9789241596107.pdf)
- WHO, 2011. Haemoglobin Concentrations for the Diagnosis of Anaemia and Assessment of Severity. Retrieved 22 January 2017 from [http://apps.who.int/iris/bitstream/10665/85839/3/WHO\\_NMH\\_NHD\\_MNM\\_11.1\\_eng.pdf?ua=1](http://apps.who.int/iris/bitstream/10665/85839/3/WHO_NMH_NHD_MNM_11.1_eng.pdf?ua=1)
- WHO, 2014. Global nutrition targets 2025: anaemia policy brief. Retrieved 22 January 2017 from [http://apps.who.int/iris/bitstream/10665/148556/1/WHO\\_NMH\\_NHD\\_14.4\\_eng.pdf?ua=1](http://apps.who.int/iris/bitstream/10665/148556/1/WHO_NMH_NHD_14.4_eng.pdf?ua=1)
- WHO, 2015. The global prevalence of anaemia in 2011. Retrieved 22 January 2017 from [http://apps.who.int/iris/bitstream/10665/177094/1/9789241564960\\_eng.pdf](http://apps.who.int/iris/bitstream/10665/177094/1/9789241564960_eng.pdf)
- WHO, 2016. Global targets tracking tool. Retrieved 22 January 2017 from <http://www.who.int/nutrition/trackingtool/en/>
- Williamson, 1989. Reduction of Indigo: Sodium Hydrosulfite as a Reducing Agent. *Letters*, 66(4):359.



- Williamsson, A., Wahlqvist, S., Nilsson, S-E. Lilja, J., Jannsson, L., and Nilsson Bertil., 1997. Capillary microcuvette. United States Patent. Patent Number: 5,674,457. Retrieved 12 February 2017 from <https://docs.google.com/viewer?url=patentimages.storage.googleapis.com/pdf/s/US5674457.pdf>
- Williamson, M. A, Snyder, L. M., Wallach, J. B. *Wallach's interpretation of diagnostic tests*. 9th ed. Wolters Kluwer/Lippincott Williams & Wilkins Health: Philadelphia; 2011.
- Wiltink, W. F., Kruithof, J., Mol, C., GréBos, M., and Eijk, H. G. V., 1973. Diurnal and Nocturnal Variations of the Serum Iron in Normal Subjects. *Clinica Chimica Acta*, 49(1): 99–104.
- Yamanishi, H., Iyama, S., Fushimi, R., Amino, N., 1996. Interference of ferritin in measurement of serum iron concentrations: comparison by five methods. *Clinical Chemistry*, 42(2):331-332.
- Yap, B.K., Lai, M.I., Lim, W.F., Talik, N.A., Sankar, P. and Nasser, A. A., 2016. 'Iron lady' Handheld Body Iron Store Reader. Pertandingan Rekacipta dan Inovasi IPTS at Universiti Tenaga Nasional, Malaysia, 15<sup>th</sup> – 16<sup>th</sup> November 2016.
- Yasmin, S., Andrews, S. C., Moore, G. R., and Le Brun, N. E., 2011. A new role for heme, facilitating release of iron from the bacterioferritin iron biomineral. *Journal of Biological Chemistry*, 286:3473–3483.
- Yee, H. V, and Goodwin, J. F., 1974. Simultaneous Determination of Copper and Iron in a Single Aliquot of Serum. *Clinical Chemistry*, 20(2):188–191.
- Youden, W. J., 1950. An index for rating diagnostic tests. *Cancer*, 3:32-35.
- Yu, K. H., 2011. Effectiveness of zinc protoporphyrin/heme ratio for screening iron deficiency in. *Nutrition Research and Practice*, 5(1):40–45.
- Zaki, R., Bulgiba, A., Ismail, R. and Ismail, N.A., 2012. Statistical methods used to test for agreement of medical instruments measuring continuous variables in method comparison studies: a systematic review. *PloS One*, 7:e37908.
- Zhou, X., Yan, H., Xing, Y., Dang, S., Shuoma, B., and Wang, D., 2009. Evaluation of a portable hemoglobin photometer in pregnant women in a high altitude area: a pilot study. *BioMed Central Public Health*, 9(228).
- Zou, K. H., O'Malley, A. J., Mauri, L., 2007. Receiver operating characteristic analysis for evaluation diagnostic tests and predictive models. *Circulation*, 115: 654-57.