



UNIVERSITI PUTRA MALAYSIA

***ENZYMATIC ESTERIFICATION OF KOJIC ACID AND PALMITIC ACID
BY IMMOBILIZED LIPASE FOR SYNTHESIS OF KOJIC ACID
PALMITATE***

NURAZWA BINTI ISHAK

FBSB 2015 175



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By

NURAZWA BINTI ISHAK

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirement for the Degree of Master of Science**

April 2015

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

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April 2015

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Kojic acid (5-hydroxy-2-hydroxymethyl-4-pyrone) is an organic acid produced from various carbon sources in an aerobic fermentation by many species of *Aspergillus*, *Penicillium* and *Acetobacter*. The importance of kojic acid (KA) is recently focused on its role as whitening agent in cosmetic formulation. Kojic acid is water soluble and has low stability towards light exposure. KA has also been criticized for weak depigmenting effect and unstable for long storage. The hydrophilic property of KA has restricted its application in cosmetic, oily food and pharmaceutical products. In order to improve the chemical and biological activities of KA, its derivatives with new and improved chemical properties and biological activities needs to be developed. Various KA derivatives such as KA esters have been synthesized at industrial scale. KA esters are normally produced via chemical process where strong acid or alkali is used. This chemical process is not environmentally friendly and also produces complex mixtures that make the product purification difficult and high cost.

The possibility of using lipase, lipozyme RMIM, in the esterification of KA with palmitic acid (PA) in acetone to synthesize KA palmitate (KAP) was investigated in this study. Preliminary, the effects of organic solvent, substrate ratio, enzyme loading, temperature and reaction time on the yield of KAP were evaluated. The appropriate ranges for each variable were subsequently used for optimization using response surface methodology (RSM). The optimal reaction condition for ester production was then applied in 500 mL stirred tank reactor (STR) using two types of impeller [Rushton turbine (RT) and Pitch blade disc turbine (PBDT)] to investigate the effect of agitation speed on the esterification performance. Among the organic solvent tested for esterification to synthesize KAP, acetone was the preferred solvent. Optimal conditions for esterification as suggested by RSM were as follows: PA to KA ratio, 6.74; enzyme loading, 0.59 g; reaction temperature, 45.9°C and reaction time, 20 h, which gave the percentage of esterification of 64.47%. For the esterification in STR, the percentage yield of KAP was significantly higher for RT than PBDT at all agitation speeds tested (150 to 450 rpm). The highest yield of KAP (82.14%) was obtained in STR with RT agitated at 250 rpm. Results from this study have demonstrated that substantially high yield of KA esters could be produced by enzymatic esterification using lipase. This alternative

method has potential to be used industrially. Since the use of hazardous chemical can be minimized, enzymatic synthesis of KA esters is more natural and appears to be more appealing to the customers than the chemical process.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia Sebagai memenuhi keperluan untuk Ijazah Sarjana Sains

**PENGESTERAN BERENZIM ASID KOJIK DAN ASID PALMITIK
MENGUNAKAN LIPASE TERSEKAT GERAK UNTUK SINTESIS ASID
KOJIK PALMITIK**

Oleh

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Asid kojik (5-hydroxy-2-hydroxymethyl-4-pyrone) adalah asid organik yang dihasilkan daripada pelbagai sumber karbon menggunakan fermentasi aerobik oleh spesies *Aspergillus*, *Penicillium* dan *Acetobacter*. Kepentingan asid kojik (KA) kini tertumpu kepada fungsinya sebagai agen pencerah dalam formulasi kosmetik. KA adalah larut air dan tidak stabil terhadap pendedahan kepada cahaya matahari. KA mempunyai kesan depigmentasi yang lemah dan tidak stabil untuk simpanan jangka lama. Sifat kesukaannya kepada air menjadikan kegunaannya dalam kosmetik, makanan berasaskan minyak dan farmaseutikal terhad. Bagi meningkatkan aktiviti biokimia KA, terbitan KA dengan sifat biokimia yang lebih baik harus dibangunkan. Pelbagai terbitan KA seperti ester KA dihasilkan pada skala industri. Ester KA biasanya dihasilkan melalui proses kimia dalam kehadiran asid atau alkali kuat. Proses kimia ini tidak mesra alam dan menghasilkan campuran kompleks yang sukar ditulenkan dengan kos yang tinggi.

Kajian ini mengkaji kebolehan enzim lipase, Lipozyme RMIM dalam pengesteran KA dan asid palmitik (PA) dalam aseton untuk menghasilkan asid kojik palmitik (KAP). Eksperimen pengesteran pada permulaannya dilakukan untuk menilai kesan pelarut yang sesuai, nisbah substrat, jumlah enzim, suhu dan masa tindak balas pada hasil KAP. Julat daripada setiap pembolehubah kemudiannya digunakan untuk pengoptimuman menggunakan kaedah tindak balas permukaan (RSM). Keadaan optimum tindak balas penghasilan ester ini digunakan dalam pengesteran menggunakan 500 mL reaktor tangki teraduk (STR) untuk mengkaji kesan kelajuan pengadukan bagi dua jenis pengaduk [*Rushton turbine* (RT) dan *Pitch blade disc turbine* (PBDT)] terhadap prestasi pengesteran. Aseton didapati merupakan pelarut pilihan berbanding pelarut lain. Keadaan optimum pengesteran yang dicadangkan oleh RSM adalah seperti berikut: nisbah PA kepada KA, 6:74; jumlah enzim, 0.59 g; suhu, 45.9°C; dan masa tindak balas, 20 jam, yang memberikan peratusan pengesteran sebanyak 64.47%. Peratusan hasil asid kojik palmitik adalah lebih tinggi bagi RT berbanding PBDT pada semua kelajuan pengadukan (150-450 rpm) yang diuji bagi pengesteran dalam STR. Hasil tertinggi KAP (82.14%) diperolehi dalam STR menggunakan RT apabila diaduk pada kelajuan 250 rpm. Hasil kajian ini menunjukkan jumlah KAP yang tinggi boleh dihasilkan oleh pengesteran berenzim menggunakan lipase. Kaedah alternatif ini berpotensi digunakan dalam

perindustrian. Oleh kerana penggunaan bahan kimia berbahaya dapat dikurangkan, sintesis enzim ester KA adalah lebih asli dan kelihatan lebih menarik kepada pelanggan.



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This thesis submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirements for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

ANN	Artificial Neural Network
ANOVA	Analysis of variance
CCRD	Central composite rotatable design
Di	Impeller diameter
Dt	Tank diameter
FBR	Fluidized Bed Reactor
FT-IR	Fourier Transform-Infrared Spectroscopy
GC	Gas Chromatography
GC-MS	Gas Chromatography-Mass Spectrometry
HL	Fluid depth
KA	Kojic acid
KAP	Kojic acid palmitate
L	Tank height
MPOB	Malaysian Palm Oil Board
NMR	Nuclear Magnetic Resonance
PBDT	Pitch Blade Disc Turbine
PBR	Packed Bed Reactor
R2	Coefficient of determination
Rf	Retention factor
Rt	Retention time
rpm	Rotation per minute
RSM	Response Surface Methodology
RT	Rushton Turbine
STR	Stirred Tank Reactor
TAG	Triacylglycerol
TLC	Thin Layer Chromatography
UV	Ultraviolet
v/v	Volume per volume
W	Ribbon width



CHAPTER 1

INTRODUCTION

Kojic acid (2-hydroxymethyl-5-hydroxy- γ -pyrone), an organic acid, is normally produced in an aerobic fermentation by fungi such as *Aspergillus* and *Penicillium* or bacteria such as *Acetobacter*. Kojic acid has diverse application in medical, food, agricultural, chemistry and cosmetic industry. In medicine, it is used as an anti-inflammatory drug and painkiller (Kayahara *et al.*, 1990). Kojic acid is widely used as a precursor for flavor enhancer and as an anti-browning agent to prevent agricultural products such as crustacean, meat and fresh vegetables from blackening (Le Blanch and Akers, 1989). Kojic acid is also served as a preservative and an antioxidant for oils and fats. Moreover, kojic acid is recognized as important intermediates in the production of the chemicals and pharmaceuticals.

Among of the aforesaid applications, a desire to maintain a youthful appearance has propelled the recent surge of kojic acid in cosmetic industry. The cosmetic formulation with the presence of kojic acid is intended not only to improve visual appearance of the skin but also to offer long lasting effect. In this case, kojic acid acts as a whitening agent and a protective against ultraviolet (Noh *et al.*, 2009; Masse *et al.*, 2001). Kojic acid not only effective to treat melasma, but it also has the ability to suppress hyperpigmentation in human skin by restraining the formation of melanin through inhibition of tyrosinase activity. Convincing evidence which shows that kojic acid is effective in inhibiting melanin synthesis as tested either *in vitro* and *in vivo* have been reported (Nohynek *et al.*, 2004).

Despite of its myriad application, kojic acid is water-soluble and unstable at high temperature for long term storage and also sensitive towards exposure to light. These characteristics restricted its application to be directly incorporated in oil based cosmetic product (Masse *et al.*, 2001). Research on the conversion of kojic acid into stable derivatives aims at improving its lipophilic characteristic is of interest of many researchers. This is usually focussed on the modification of the C-5 or C-7 hydroxyl through esterification process which can be achieved via chemical or enzymatic approach (Lajis *et al.*, 2013).

At present, many esters are industrially manufactured by chemical methods because it is claimed as economical process. However, chemical esterification involves high temperature and extreme pressure which limits this method for the esterification of unstable compound. Furthermore, chemical esterification has low reaction rates with simultaneous production of unwanted side products, which make product purification more complicated with increasing cost. Enzymatic esterification appears to be more appealing to the customer since there is a rapidly growing demand for natural, cleaner, alternative technologies that produces less waste and also to avoid the use of toxic reagent. Liu and Shaw (1998) had successfully performed the enzymatic synthesis of kojic acid esters from various sources of acyl donor. Various parameters such as type of solvent, substrate ratio, enzyme loading, reaction time and temperature may affect the enzymatic esterification of kojic acid esters.

Optimization of the process reaction is essential to maximize the production and to reduce the cost. Process optimization via the classical one-variable-at-a-time approach is simple to plan and execute. However, this method is inefficient and fails to detect any interaction amongst the reaction variables. Response surface methodology (RSM), a statistical tool, is commonly used to overcome such drawbacks. RSM has been successfully applied to study and optimize the enzymatic synthesis of kojic acid monolaurate (Chen *et al.*, 2002). For industrial production, large scale reactor shall be used to perform the esterification at optimal conditions to maximize the yield and productivity. Stirred tank reactor (STR), which is the conventional mixing vessel, has advantage of low capital cost and low operating costs. STR has been used in many lipase-catalyzed esterification process for production of various esters (Mat Radzi *et al.*, 2010; Keng *et al.*, 2008). A wide variety of impeller designs are available to produce different flow pattern inside the stirred tank vessel to accommodate the specific requirement of the enzymatic process. Appropriate degree of mixing and flow pattern is required in enzymatic process employing immobilized enzyme to ensure the homogenous particle in suspension is achieved. Excessive shear effect created by the agitated impeller may cause damage to immobilized enzyme particle, which in turn, reduce the activity and reusability of the enzyme. The yield and productivity of the enzymatic esterification process may also be improved using suitable mode of reactor operation. For example, production of lauroyl kojic acid through lipase-catalyzed esterification has been improved in continuous STR (Kobayashi *et al.*, 2001).

Therefore, this particular work was carried out with the following objectives:

1. To investigate the possibility of using commercial immobilized lipase, which are known to have specificity towards ester bonds, for the synthesis of kojic acid palmitate by esterification in solvent system.
2. To optimize the reaction parameters for the synthesis of kojic acid palmitate in shake flask reaction using response surface methodology (RSM).
3. To investigate the effect of agitation speed on the synthesis of kojic acid palmitate in stirred tank reactor employing two different designs of impeller.

REFERENCES

- Abd Rahman., N.F., Basri, M., Abdul Rahman, M.B., Raja Abdul Rahman, R.N.Z. and Salleh, A.B. (2011). High yield lipase-catalyzed synthesis of engkabang fat esters for the cosmetic industry. *Bioresource Technology*, 102:2168-2176.
- Abdul Wahab, R., Basri, M., Raja Abdul Rahman, R.N.Z., Salleh, A.B., Abdul Rahman, A.B., Chaibakhsh, N. and Leow T.C. (2014). Enzymatic production of a solvent-free menthyl butyrate via response surface methodology catalyzed by a novel thermostable lipase from *Geobacillus zalihae*. *Biotechnology & Biotechnology Equipment*, 28(6):1065-1072.
- Adams, E.M. and Allen, H.C. (2013). Palmitic acid on salt subphases and in mixed monolayers of cerebrosides: application to atmospheric aerosol chemistry. *Atmosphere*, 4: 315-336.
- Adamopoulos, L. (2006). *Understanding the Formation of Sugar Fatty Acid Esters*. Master's Thesis, North Carolina State University, North Carolina, United States.
- Adnani, A., Basri, M., Chaibakhsh, N., Abdul Rahman, M.B. and Salleh, A.B. (2011). Artificial neural network analysis of lipase-catalyzed synthesis of sugar alcohol ester. *Industrial Crops and Products*, 33:42-28.
- Al-Edresi, S. and Baie, S. (2010). In-vitro and in-vivo evaluation of a photo-protective kojic dipalmitate loaded into nano-creams. *Asian Journal of Pharmaceutical Sciences*, 5(6):251-265.
- Andualema, B. and Gessesse, A. (2012). Microbial lipases and their industrial applications: Review. *Biotechnology*, 11(3):100-118.
- Aracil, J., Garcia, T. and Martinez, M. (1993). Enzymatic synthesis of an analogue of jojoba oil: Optimization by statistical analysis. *Enzyme and Microbial Technology*, 15:607-611.
- Arcos, J.A., Bernabe, M. and Otero, C. (1998). Quantitative enzymatic production of 1,6-diacyl fructofuranoses. *Enzyme and Microbial Technology*, 22:27-35.
- Ariff, A.B., Salleh, M.S., Ghani, B., Hassan, M.A., Rusul, G. and Karim, M.I.A. (1996). Aeration and yeast extract requirements for kojic acid production by *Aspergillus flavus* link. *Enzyme and Microbial Technology*, 19:545-550.
- Ashari, S.E., Mohamad, R., Ariff, A., Basri, M. and Salleh, A.B. (2009). Optimization of enzymatic synthesis of palm-based kojic acid ester using response surface methodology. *Journal of Oleo Science*, 58(10):503-510.
- Basiron, Y. (2007). Palm oil production through sustainable plantations. *European Journal of Lipid Science and Technology*, 109:289-295.
- Basri, M., Ampon, K., Wan Yunus, W.M.Z., Razak, C.N.A. and Salleh, A.B. (1995). Enzymatic synthesis of fatty esters by hydrophobic lipase derivatives immobilized on organic polymer beads. *Journal of American Oil Chemist' Society*, 72(4):407-411.

- Basri, M., Raja Abdul Rahman, R.N.Z., Ebrahimpour, A., Salleh, A.B., Gunawan, E.R. and Abdul Rahman, M.B. (2007). Comparison of estimation capabilities of response surface methodology (RSM) with artificial neural network (ANN) in lipase-catalyzed synthesis of palm-based wax ester. *BMC Biotechnology*, 7:53-63.
- Basri, M., Raja Abdul Rahman, R.N.Z. and Salleh, A.B. (2013). Specialty oleochemicals from palm oil via enzymatic synthesis. *Journal of Oil Palm Research*, 25(1):22-35.
- Bartal, N., Serrati, G., Szewczyk, D. and Waterman, J. (2009). *Modelling of a Catalytic Packed Bed Reactor and Gas Chromatograph Using COMSOL Multiphysics*. Degree's Thesis, Worcester Polytechnic Institute, United States.
- Bentley, R. (2006). From miso, saké and shoyu to cosmetics: A century of science for kojic acid. *Natural Products Report*, 23:1046-1062.
- Betiku, E. and Ajala, S. O. (2014). Modelling and optimization of *Thevetia peruviana* (yellow oleander) oil biodiesel synthesis via *Musa paradisiacal* (plantain) peels as heterogenous base catalyst: A case of artificial neural network vs response surface methodology. *Industrial Crops and Products*, 53:314-322.
- Bezerra, M.A., Santelli, R.E., Oliveira, E.P., Villar, L.S. and Escaleira, L.A. (2008). Response surface methodology (RSM) as a tool for optimization in analytical chemistry. *Talanta*, 77:965-977.
- Bidin, H., Basri, M., Radzi, S.M., Ariff, A., Rahman, R.N.Z.A. and Salleh, A.B. (2009). Optimization of lipase-catalyzed synthesis of palm amino acid surfactant using response surface methodology (RSM). *Industrial Crops and Products*, 30:206-211.
- Bloomer, S., Adlercreutz, P. and Mattiasson, B. (1992). Facile synthesis of fatty acid esters in high yields. *Enzyme and Microbial Technology*, 14(7):546-552.
- Box, G.E.P. and Wilson, K.B. (1951). On the experimental attainment of optimum conditions. *Journal of the Royal Statistical Society: Series B Methodological*, 13:1-45.
- Bradoo, S., Saxena, R.K. and Gupta, R. (1999). High yields of ascorbyl palmitate by thermostable lipase-mediated esterification. *Journal of the American Oil Chemists' Society*, 76:1291-1295.
- Bransova, J., Brtko, J., Uher, M. and Novotny, L. (1995). Antileukemic activity of 4-pyranone derivatives. *International Journal of Biochemistry and Cell Biology*, 27:701-706.
- Bransova, J., Uher, M. and Brtko, J. (1998). Regulation of selected biological processes in neoplastic cell lines by halogen derivatives of 5-hydroxy-2-hydroxymethyl-4-pyranone. *Anticancer Research*, 18:4423-4428.
- Bransova, J., Uher, M., Novotny, L. and Brtko, J. (1997). 5-Benzoyloxy-2-thiocyanatoethyl-4-pyranone, a novel heterocyclic compound: Synthesis,

- structure determination and effects on neoplastic cell growth. *Anticancer Research*, 17:1175-1178.
- Brtko, J., Rondahl, L., Fickova, M., Hudecova, D., Eybl, V. and Uher, M. (2004). Kojic acid and its derivatives: History and present state of art. *Central European Journal of Public Health*, 12:16-18.
- Burkert, J.M., Maugeri, F. and Rodrigues, M.I. (2004). Optimization of cellular lipase production by *Geotrichum sp.* using factorial design. *Bioresource Technology*, 91:77-84.
- Cabanes, J., Chazarra, S. and Garcia, F. (1994). Kojic acid, a cosmetics skin whitening agent, is a slow-binding inhibitor of catecholase activity of tyrosinase. *Journal of Pharmaceutical and Pharmacology*, 46:982-985.
- Carrea, G. and Riva, S. (2000). Properties and synthetic applications of enzymes in organic solvents. *Angewandte Chemie International Edition*, 39:2226-2254.
- Castillo, E., Pezzotti, F., Navarro, A. and López-Munguía, A. (2003). Lipase catalyzed synthesis of xylitol monoesters: Solvent engineering approach. *Journal Biotechnology*, 102:251-259.
- Chaibakhsh, N., Basri, M., Abdul Rahman, M.B., Adnani, A. and Salleh, A.B. (2012). Lipase-catalyzed synthesis of ergosterol ester. *Biocatalysis and Agricultural Biotechnology*, 1:51-56.
- Chaudhary, J., Pathak, A.N. and Lakhawat, S. (2014). Production technology and applications of kojic acid. *Annual Research & Review in Biology*, 4(21):3165-3196.
- Chen, C.S., Liu, K.J., Lou, Y.H. and Shieh, C.J. (2002). Optimisation of kojic acid monolaurate synthesis with lipase PS from *Pseudomonas cepacia*. *Journal of the Science of Food and Agriculture*, 82:601-605.
- Cho, J.C., Rho, H.S., Baek, H.S., Ahn, S.M., Woo, B.Y., Hong, Y.D., Cheon, J.W., Heo, J.M., Shin, S.S., Park, Y.H. and Suh, K.D. (2012). Depigmenting activity of new kojic acid derivative obtained as a side product in the synthesis of cinnamate of kojic acid. *Bioorganic & Medicinal Chemistry Letters*, 22:2004-2007.
- Chowdary, G.V. and Prapulla, S.G. (2002). The influence of water activity on the lipase-catalyzed synthesis of butyl butyrate by transesterification. *Process Biochemistry*, 38:393-397.
- Ciftci, O.N., Fadliloglu, S., Gogus, F. and Guven, A. (2008). Prediction of a model enzymatic acidolysis system using neural networks. *Grasas Y. Aceites*, 59:375-382.
- Colla, L.M., Rizzardi, J., Pinto, M.H., Reinehr, C.O., Bertolin, T.E. and Vieira Costa, J.A. (2010). A simultaneous production of lipases and biosurfactants by submerged and solid-state bioprocess. *Bioresource Technology*, 101:8308-8314.

- Dahlan, I., Harun, A. and Najafpour, G.D. (2009). Free *Candida rugosa* lipase-catalyzed synthesis of citronellyl butyrate in n-hexane by direct esterification: effect of reaction parameters. *Jurnal Teknologi*, 50:29-40.
- Daniel, R.M., Dines, M. and Petach, H. (1996). The denaturation and degradation of stable enzyme at high temperature. *Biochemical Journal*, 317:1-11.
- Datta, S., Christena, L.R., Rajaram, Y.R.S. (2013). Enzyme immobilization: an overview on techniques and support materials. *Biotechnology*, 3:1-9.
- Degn, P. and Zimmermann, W. (2001). Optimization of carbohydrate fatty acid ester synthesis in organic media by a lipase from *Candida antarctica*. *Biotechnology and Bioengineering*, 74(6):483-491.
- Dholakiya, B.Z. (2012). Super phosphoric acid catalyzed biodiesel from low cost feed stock. *Archives of Applied Science Research*, 4(1):551-561.
- Doran, M.P. (2003). *Bioprocess Engineering Principles*. London: Academic Press.
- El-Aasar, S.A. (2006). Cultural conditions studies on kojic acid production by *Aspergillus paraciticus*. *International Journal of Agriculture and Biology*, 8:468-473.
- El-Boulifi, N, Ashari, S.E., Serrano, M., Aracil, J. and Martinez, M. (2014). Solvent-free lipase-catalyzed synthesis of a novel hydroxyl-fatty acid derivatives of kojic acid. *Enzyme and Microbial Technology*, 55:128-132.
- Elibol, M. and Ozer, D. (2000). Influence of oxygen transfer on lipase production by *Rhizopus arrhizus*. *Process Biochemistry*, 36:329-352.
- Facioli, N.L. and Barrera, A.D. (2001). Optimization of enzymatic esterification of soybean oil deodoriser distillate. *Journal of the Science and Agriculture*, 12:1193-1198.
- Federation of Asian Chemical Societies (FACS) (2000). *Newsletter*. Chemical Industry in Malaysia with Special Reference to Palm-Based Industry.
- Ferreira, B.S., Fernandes, P. and Cabral, J.M.S. (2001). Design and modelling of immobilized biocatalytic reactors. In *Multiphase Bioreactor Design*, ed. J.M.S. Cabral, M. Mota, and J. Tramper, pp. 85-114. London: Taylor & Francis.
- Food and Drug Administration (FDA) (2012). *Reports*. Guidance for Industry Q3C Impurities: Residual Solvents.
- Gandhi, N.N., Patil, N.S., Sawant, S.B. and Joshi, J.B. (2000). Lipase-catalyzed esterification. *Catalysis Reviews Science and Engineering*, 42(4):439-480.
- Ghorbani, F., Younesi, H., Ghasempouri, S.M., Zinatizadeh, A.A., Amini, M. and Daneshi, A. (2008). Application of response surface methodology for optimization of cadmium biosorption in an aqueous solution by *Saccharomyces cerevisiae*. *Chemical Engineering Journal*, 145:267-275.

- Giovanni, M. (1983). Response surface methodology and product optimization. *Food Technology*, November Issue: 41-45.
- Gogoi, S. and Dutta, N.N. (2009). Kinetics and mechanism of esterification of isoamyl alcohol with acetic acid by immobilized lipase. *Indian Journal of Chemical Technology*, 16:209-215.
- Goswami, D., De, S. and Basu, K. (2012). Effects of process variables and additives on mustard oil hydrolysis by porcine pancrease lipase. *Brazilian Journal of Chemical Engineering*, 29(3):449-460.
- Gumel, A.M., Annuar, M.S.M., Heidelberg, T. and Chisti, Y. (2011). Lipase mediated synthesis of sugar fatty esters. *Process Biochemistry*, 46:2079-2090.
- Gunasekaran, V. and Das, D. (2005). Lipase fermentation: progress and prospects. *Indian Journal of Biotechnology*, 4:437-445.
- Gunstone, F.D. (2001). Basic oleochemicals, oleochemical products and new industrial oils. In *Oleochemical Manufacture and Applications*, ed. F.D. Gunstone, and R.J. Hamilton, pp 1-22. Sheffield: Sheffield Academic Press.
- Gunawan, E.R., Basri, M., Abdul Rahman, M.B., Salleh, A.B. and Abdul Rahman, R.N.Z. (2005). Study on response surface methodology (RSM) of lipase-catalyzed synthesis of palm-based wax ester. *Enzyme and Microbial Technology*, 37:739-744.
- Gunawan, E.R. and Suhendra, D. (2008). Wax ester production by alcoholysis of palm oil fractions. *Indonesian Journal of Chemistry*, 8(3):356-362.
- Haaland, P.D. (1989). *Experimental Design in Biotechnology*. New York: Marcel Dekker, Inc.
- Halling, P.J. (1992). Salts hydrates for water activity control with biocatalyst in organic media. *Biotechnology and Bioengineering*, 6(3): 271-276.
- Hari Krishna, S., Divakar, S., Prapulla, S.G. and Karanth, N.G. (2001). Enzymatic synthesis of isoamyl acetate using immobilized lipase from *Rhizomucor miehei*. *Journal of Biotechnology*, 87:193-201.
- Hassan, F., Shah, A.A. and Hameed, A. (2006). Industrial applications of microbial lipases. *Enzyme and Microbial Technology*, 39:235-251.
- Hudecova, D., Jantova, S., Melnik, M. and Uher, M. (1996). New azidometalkojates and their biological activity. *Folia Microbiologica*, 41(6):473-476.
- Husaini, N.A. (2012). *The Jordan-Pi Sigma Neural Network for Temperature Prediction*, Master's Thesis, Universiti Tun Hussein Onn Malaysia, Johor, Malaysia.
- Ikeda, T. and Tsutsumi, T. (1990). Function and skin depigmental activity of crude drugs. *Fragrance Journal*, 6:59-61.

- Jaeger, K.E., Dijkstra, B.W. and Reetz, M.T. (1999). Bacterial biocatalysis: molecular biology, three-dimensional structures, and biotechnological applications of lipases. *Annual Review of Microbiology*, 53:315-351.
- Jegannathan, K.R. and Nielson, P.H. (2013). Environmental assessment of enzyme use in industrial production: A literature review. *Journal of Cleaner Production*, 42:228-240.
- Kaatz, H., Streffer, K., Wollenberger, U. and Peret, M.G. (1999). Inhibition of mushroom tyrosinase by kojic acid octanoates. *Zeitschrift für Naturforschung*, 54c:70-74.
- Kadic, A., Palmqvist, B. and Liden, G. (2014). Effects of agitation on particle size distribution and enzymatic hydrolysis of pretreated spruce and giant reed. *Biotechnology for Biofuels*, 7:77-86.
- Kahn, V., Linder, P. and Zakin, V. (1994). Effect of kojic acid on the oxidation of o-dihydroxyphenols by mushroom tyrosinase. *Journal of Food Chemistry*, 18:253-271.
- Kamini, N.R., Fujii, T., Kurosu, T. and Iefuji, H. (2000). Production and purification and characterization of an extracellular lipase from yeast, *Cryptococcus* sp. S-2. *Process Biochemistry*, 36:317-324.
- Kapoor, M. and Gupta, M.N. (2012). Lipase promiscuity and its biochemical applications. *Process Biochemistry*, 47:555-569.
- Kang, I.J., Pfomm, P.H. and Rezac, M.E. (2005). Real time measurement and control of thermodynamic water activities for enzymatic catalysis in hexane. *Journal of Biotechnology*, 119(2):147-154.
- Kang, S.S., Kim, H.J., Jin, C. And Lee, Y.S. (2009). Synthesis of tyrosinase inhibitory (4-oxo-4H-pyran-2-yl)acrylic acid ester derivatives. *Bioorganic and Medicinal Chemistry Letters*, 19:188-191.
- Kayahara, H., Shibata, N., Tadasa, K., Maedu, H., Kotani, T. and Ichimoto, I. (1990). Amino acids and peptide derivatives of kojic acid and their antifungal properties. *Agriculture and Biological Chemistry*, 54:2441-2442.
- Keng, P.S., Basri, M., Rahman, M.B.A., Salleh, A.B., Rahman, R.N.Z.A. and Ariff, A. (2005). Optimization of palm-based wax ester production using statistical experimental designs. *Journal of Oleo Science*, 54:519-528.
- Keng, P.S., Basri, M., Ariff, A.B., Abdul Rahman, M.B., Abdul Rahman, R.N.Z. and Salleh, A.B. (2008). Scale-up synthesis of lipase-catalyzed palm esters in stirred-tank reactor. *Bioresource Technology*, 99(14):6097-6104.
- Khamaruddin, N.H., Basri, M., Lian, G.E.C., Salleh, A.B., Raja Abdul Rahman, R.A. Z., Ariff, A., Mohamad, R. and Awang, R. (2008). Enzymatic synthesis and characterization of palm-based kojic acid ester. *Journal of Oil Palm Research*, 20:461-469.

- Khan, A.A. and Alzohairy, M.A. (2010). Recent advances and applications of immobilized enzyme technologies: A review. *Research Journal of Biological Sciences*, 5(8):565-575.
- Kichner, G., Scollar, M.P. and Klibanov, A.M. (1985). Resolution of racemic mixtures via lipase catalysis in organic solvents. *Journal of American Oil Chemistry Society*, 107: 7072-7076.
- Kim, D.H., Hwang, J.S., Baek, H.S., Kim, K.J., Lee, B.G., Chang, I., Kang, H.H. and Lee, O.S. (2003). Development of 5-[(3-Aminopropyl)phosphinooxyl]-2-(hydroxymethyl)-4H-pyran-4-one as a novel whitening agent. *Chemical and Pharmaceutical Bulletin*, 51(2):113-116.
- Kobayashi, Y., Kayahara, H., Tadasa, K. and Tanaka, H. (1996). Synthesis of n-kojic-amino acid and n-kojic-amino acid-kojiate and their tyrosinase inhibitory activity. *Bioorganic and Medicinal Chemistry Letters*, 6(12):1303-1308.
- Kobayashi, T., Adachi, S., Nakanishi, K. and Matsuno, R. (2001). Semi-continuous production of lauroyl kojic acid through lipase-catalyzed condensation in acetonitrile. *Biochemical Engineering Journal*, 9:85-89.
- Kongkkrapan, I. (2011). Transforming oleochemical landscape-The way forward for south east asia, Emery Oleochemical Group. *International Palm Oil Congress-Malaysia*.
- Kotulova, D., Uher, M. and Brtko, J. Anti-inflammatory preparation. SK Patent No. 277821, 1994.
- Kuan, J.L., Alhindra, N. and Shaw, J.F. (2001). Lipase-catalyzed synthesis of fatty acid diethanolamides. *Journal of Agriculture and Food Chemistry*, 49(12):5761-5764.
- Kumar, A., Sharma, P. and Kanwar, S.S. (2012). Lipase catalyzed esters syntheses in organic media: A review. *International Journal of Institutional Pharmacy and Life Sciences*, 2(2): 90-119.
- Laane, C., Boeren, S., Vos, K. and Veeger, C. (1987). Rules for optimization of biocatalysis in organic solvents. *Biotechnology and Bioengineering*, 30:81-87.
- Lajis, A.F., Hamid, M., Ariff, A. (2012). Depigmenting effect of kojic acid esters in hyperpigmented B16F1 melanoma cells. *Journal of Biomedicine and Biotechnology*. Article ID 952452, 9 pages.
- Lajis, A.F., Basri, M., Mohamad, R., Hamid, M., Ashari, S.E., Ishak, N., Zoolkiflie, A. and Ariff, A. (2013). Enzymatic synthesis of kojic acid esters and their potential industrial applications. *Chemical Papers*, 67(6):573-585.
- Le Blanch, D.T. and Akers, H.A. (1989). Maltol and ethyl maltol from larch tree to successful food additives. *Food Technology*, 26:78-87.

- Li, C., Sun, J., Li, T., Liu, S.Q. and Huang, D. (2014). Chemical and enzymatic synthesis of a library of 2-phenetyl esters and their sensory attributes. *Food Chemistry*, 154:205-210.
- Ling, H.H. (1999). *Production of Kojic Acid by Aspergillus flavus and Aspergillus oryzae and Its Purification Procedures*. Degree's Thesis, Universiti Putra Malaysia, Selangor, Malaysia.
- Liu, K.J. and Shaw, J.F. (1998). Lipase-catalyzed synthesis of kojic acid esters in organic solvents. *Journal of the American Oil Chemists' Society*, 75:1507-1511.
- Liu, X., Gong, I., Xin, M., Liu, J. (1999). The synthesis of sucrose ester and selection of its catalyst. *Journal of Molecular Catalysis A: Chemical*, 147:37-40.
- Loudon, G.M. (1995) Enzymes: Biological catalyst. In *Organic Chemistry* (3rd Ed.). Cummings Publishing Company, pp. 1308-1309.
- Malaysian Investment Development Authority (MIDA) (2014). Seminar on oleochemical industry in Malaysia, Kuala Lumpur, Malaysia.
- Malaysian Palm Oil Board (MPOB) (2014). *Reports*. Overview of the Malaysian Oil Palm Industry 2013.
- Manohar, B. and Divakar, S. (2005). An artificial neural network analysis of porcine pancreas lipase catalyzed esterification of anthranilic acid with methanol. *Process Biochemistry*, 40:3372-3376.
- Manosroi, A., Wongtrakul, P., Manosroi, J., Midorikawa, U., Hanyu, Y., Yuasa, M., Sugawara, F., Sakai, H. and Abe, M. (2005). The entrapment of kojic oleate in bilayer vesicles. *International Journal of Pharmaceutics*, 298(1):13-25.
- Marchitan, N., Cojucaru, C., Mereuta, A., Duca, G., Cretescu, I. and Gonta, M. (2010). Modelling and optimization of tartaric acid reactive extraction from aqueous solutions: A comparison between response surface methodology and artificial neural network. *Separation and Purification Technology*, 75:273-285.
- Masse, M.O., Duvallet, V., Borremans, M. and Goeyens, L. (2001). Identification and quantitative analysis of kojic acid and arbutine in skin-whitening cosmetics. *International Journal of Cosmetic Science*, 23:219-232.
- Mat Radzi, S., Basri, M., Salleh, A.B., Ariff, A., Mohamad, R., Abdul Rahman, M.B. and Raja Abdul Rahman, R.N.Z. (2005a). High performance enzymatic synthesis of oleyl oleate using immobilised lipase from *Candida rugosa*. *Electronic Journal of Biotechnology*, 8(3):291-298.
- Mat Radzi, S., Basri, M., Salleh, A. B., Ariff, A., Mohammad, R., Abdul Rahman, M. B. and Raja Abdul Rahman, R.N.Z. (2005b). Large scale production of liquid wax ester by immobilized lipase. *Journal of Oleo Science*, 54(4):203-209.
- Mat Radzi, S., Basri, M., Salleh, A.B., Mohamad, R., Abdul Rahman, M.B. and Abdul Rahman, R.N.Z. (2006). Optimization study of large-scale enzymatic synthesis

- of oleyl oleate, a liquid wax ester, by response surface methodology. *Journal of Chemical Technology and Biotechnology*, 81(3):374-380.
- Mat Radzi, S., Mohamad, R., Basri, M., Salleh, A.B., Ariff, A., Abdul Rahman, M.B. and Raja Abdul Rahman, R.N.Z. (2010). Kinetics of enzymatic synthesis of liquid wax ester from oleic acid and oleyl alcohol. *Journal of Oleo Science*, 59(3): 127-34.
- McDonough, R.J. (1992). *Mixing for the Process Industries*. New York: Van Norstrand Reinhold.
- McCulloch, W.S. and Pitts, W.H. (1943). A logical calculus of the ideas immanent in nervous activity. *Bulletin of Mathematical Biophysics*, 5:115-133.
- Mohamad, R., Madihah, S. and Ariff, A.B. (1998). Isolation of a kojic acid producing fungus capable of using starches as a carbon sources. *Letter of Applied Microbiology*, 26:27-30.
- Mohamad, R., Mohamed, M.S., Suhaili, N., Salleh, M.M. and Ariff, A. (2010). Kojic acid: Applications and development of fermentation process for production. *Biotechnology and Molecular Biology Reviews*, 5(2):24-37.
- Montgomery, D.C. and Runger, G.C. (2003). *Applied Statistics and Probability for Engineers: Design and Analysis of Experiments*. John Wiley & Sons, Inc.
- Myers, R.H. and Montgomery, D.C. (2002). *Response surface methodology: Process and product optimization using experiments* (2nd Edition). USA: John Wiley & Sons, Inc.
- Nagai, S. and Izumi, T. (1981). U. S. Patent No 4278656 A.
- Nandan, R. and Polasa, H. (1985). Inhibition of growth of kojic acid biosynthesis in *Aspergillus* by some chlorinated hydrocarbons. *Indian Journal of Microbiology*, 25:21-25.
- Neta, N.S., Peres, A.M, Teixeira, J.A. and Rodrigues, L.R. (2011). Maximization of fructose esters synthesis by response surface methodology. *New Biotechnology*, 28:349-355.
- Noh, J.I., Kwak, S.Y., Seo, H.S., Seo, J.H., Kim, B.G. and Lee, Y.S. (2009). Kojic acid-amino acid conjugates as tyrosinase inhibitors. *Bioorganic and Medicinal Chemistry Letters*, 19:5586-5589.
- Nohynek, G.J., Kirkland, D., Marzin, D., Toutain, H., Leclerc-Ribaud, C. and Jinnai, H. (2004). An assessment of the genotoxicity and human health risk of topical use of kojic acid [5-hydroxy-2-(hydroxymethyl)-4h-pyran-4-one]. *Food and Chemical Toxicology*, 42:93-105.
- Noor, I.M., Hasan, M. and Ramachandran, K.B. (2003). Effect of operating variables on the hydrolysis rate of palm oil by lipase. *Process Biochemistry*, 39:13-20.

- Novozyme A/S Bagsvaerd, Denmark. (2002). Product Sheet Lipozyme RMIM 2000-10503-04.
- Obiri, O.O. (2006). *Synthesis Of Lipase-Catalyzed Saccharide-Fatty Acid Esters Using A Packed Bed Bioreactor System With Continuous Re-Circulation Of Reaction Medium: A Continuation Of Batch-Mode-Related Research*. Master's Thesis, University of Tennessee, Knoxville, United States.
- O'Brien, R.D. (2004). *Fats and Oils: Formulating and Processing for Applications*. Boca Raton: CRC Press.
- Ohyama, Y. and Mishima, Y. (1990). Melanosis-inhibitory effect of kojic acid and its action mechanism. *Fragrance Journal*, 6:53-58.
- Ozturk, B. (2001). *Immobilization of Lipase from Candida rugosa on Hydrophobic and Hydrophilic Supports*. Master's Thesis, Izmir Institute of Technology, Turkey.
- Ramamurthi, S. and McCurdy, A.R. (1994). Lipase-catalyzed esterification of oleic acid and methanol in hexane - A kinetic study. *Journal of American Oil Chemists' Society*, 71(9):927-930.
- Ramero, M.D., Calvo, L., Alba, C., Daneshfar, A.D. and Ghaziaskar, H.S. (2005). Enzymatic synthesis of isoamyl acetate with immobilized *Candida antarctica* lipase in n-hexane. *Enzyme and Microbial Technology*, 37:42-48.
- Ray, A. (2012). Application of lipase in industry. *Asian Journal of Pharmacy and Technology*, 2(2):33-37.
- Reetz, M.T. (2002). Lipases as practical biocatalysts. *Current Opinion in Chemical Biology*, 6:145-150.
- Rho, H.S., Baek, H.S., You, J.W., Kim, S., Lee, J.Y., Kim, D.H. and Chang, I.S. (2007). New 5-hydroxy-2-(hydroxymethyl)-4H-pyran-4-one derivatives has both inhibitory and antioxidant properties. *Bulletin of the Korean Chemical Society*, 28(3):471-473.
- Rho, H.S., Ahn, S.M., Yoo, D.S., Kim, M.K., Cho, D.H. and Cho, J.Y. (2010a). Kojyl thioether derivatives having both tyrosinase inhibitory and anti-inflammatory properties. *Bioorganic & Medicinal Chemistry Letters*, 20:6569-6571.
- Rho, H.S., Yoo, D.S., Ahn, S.M., Kim, M.K., Cho, D.H. and Cho, J.Y. (2010b). Inhibitory activities of kojyl thioether derivatives against nitric oxide production induced by lipopolysaccharide. *Bulletin of the Korean Chemical Society*, 31(11):3463-3466.
- Rho, H.S., Goh, M., Lee, J., Ahn, S.M., Yeon, J., Yoo, D.S., Kim, D.H., Kim, H.G. and Cho, J.Y. (2011). Ester derivatives of kojic acid and polyphenols containing adamantane moiety with tyrosinase inhibitory and anti-inflammatory properties. *Bulletin of the Korean Chemical Society*, 32(4):1411-1414.
- Roig, M.G., Bello, J.F., Velasco, F.V., De Celis, C.D. and Cachaza, J.M. (1987). Applications of immobilized lipase. *Biochemical Education*, 15(4):198-208.

- Rondahl, L., Uher, M. and Brtko, J. (2003). Syntheses and structure determinations of some selenocyanato- and thiocyanato-kojic acid derivatives. *Heterocyclic Communications*, 9: 257-258.
- Sahoo, S. (2012). *Fluidized Bed Reactor: Design and Application for Abatement of Fluoride*. Degree's Thesis, National Institute of Technology, Roukela, India.
- Sambanthamurthi, R., Sundram, K. and Tan, Y.A. (2000). Chemistry and biochemistry of palm oil. *Journal of Progress in Lipid Research*, 39: 507-558.
- Schmidt, R.D. and Veger, R. (1998). Lipases: Interfacial enzymes with attractive applications. *Angewandte Chemie International Edition*, 37:608-1633.
- Scinderm. (2014). Ingredient Glossary for Scinderm Products. http://www.scinderm.co.za/SCINDERM_GLOSSARY_OF_SKIN_CARE_INGREDIENTS.pdf, Retrieved 19th August 2014.
- Scragg, A.H. (1991). *Bioreactors in biotechnology*. London: Ellis Horwood.
- Servat, F., Montet, D., Pina, M., Galzy, P., Arnaud, A., Ledon, H., Marcou, L. and Graillie, J. (1990). Synthesis of fatty hydroxamic acid catalyzed by the lipase of *Mucor miehei*. *Journal of Oil Chemistry Society*, 67(10): 646-649.
- Sani, W. (2014). *Multistage Methanolysis of Crude Palm Oil for Biodiesel Production in a Pilot Plant*. Phd Thesis, Universiti Tun Hussein Onn, Malaysia.
- Sharma, R., Chisti, Y. and Banerjee, U.C. (2001). Production, purification, characterization and application of lipases. *Biotechnology Advances*, 19:627-662.
- Sharma, R., Thakur, V., Sharma, M. and Birkeland, N.K. (2013). *Thermophilic microbes in environmental and industrial biotechnology: Biotechnology of Thermophiles*. Springer, London.
- Sharma, S. and Kanwar, S.S. (2014). Organic solvent tolerant lipases and applications. *The Scientific World Journal, Review Article*, Article ID 625258, 15 pages.
- Sheldon, R. (2001). Catalytic reactions in ionic liquids. *Chemical Communication*. 2399–2407.
- Shieh, C.J., Liao, H.F. and Lee, C.C. (2003). Optimization of lipase-catalyzed biodiesel by response surface methodology. *Bioresource Technology*, 88:103-106.
- Shihani, N., Kumbhar, B.K. and Kulshreshtha, M. (2006). Modeling of extrusion process using response surface methodology and artificial neural networks. *Journal of Engineering Science and Technology*, 1(1):31-40.
- Silva, G.F., Camargo, F.L. and Ferreira, A.L.O. (2011). Application of response surface methodology for optimization of biodiesel production by transesterification of soybean oil with ethanol. *Food Processing Technology*, 92:407-413.

- Soo, E.L., Salleh, A.B., Basri, M., Rahman, R.N.Z.A. and Kamaruddin, K. (2004). Response surface methodological study on lipase-catalyzed synthesis of amino acid surfactants. *Process Biochemistry*, 39:1511-1518.
- Stergiou, P.Y., Foukis, A., Filippou, M., Koukouritaki, M., Parapouli, M., Theodorou, L.G., Hatziloukas, E., Afendra, A., Pandey, A. and Papamichael, E.M. (2013). Advances in lipase-catalyzed esterification reactions. *Journal of Biotechnology Advances*, 31:1846-1859.
- Stojavonic, M., Velickovic, D., Dimitrijevic, A., Milosavic, N., Jugovic, Z.K. and Bezbradica, D. (2013). Lipase-catalyzed synthesis of ascorbyl oleate in acetone: Optimization of reaction conditions and lipase reuseability. *Journal of Oleo Science*, 62(8):591-603.
- Sun, P., Yang, H., Wang, Y., Liu, K. and Xu, Y. (2013). Lipase-catalyzed synthesis and characterization of stearic acid dextrin ester. *Research in Health and Nutrition*, 1(1):7-11.
- Sun, W.J., Zhao, H.X., Cui, F.J., Li, Y.H., Yu, S.L., Zhou, Q., Qian, J.Y. and Dong, Y. (2013). D-isoascorbyl palmitate: lipase-catalyzed synthesis, structural characterization and process optimization using response surface methodology. *Chemistry Central Journal*, 7:114-126.
- Syamsul, K.M.W., Salina, M.R., Siti, S.O. and Hanina, M.N. (2010). Green synthesis of lauryl palmitate via lipase-catalyzed reaction. *World Applied Sciences Journal*, 11(4):401-407.
- Tewari, Y.B., Schantz, M.M. and Vanderah, D.J. (1999). Thermodynamics of the lipase-catalyzed esterification of 1-dodecanoic acid with menthol in organic solvents. *Journal of Chemical Engineering Data*, 44: 641-647.
- Turon, F., Caro, Y., Villeneuve, P. and Graille, J. (2003). Effect of water content and temperature on *Carica papaya* lipase catalyzed esterification and transesterification reactions. *John Libbey Eurotext*, 10:400-4004.
- Uher, M., Kyselicova, L., Rajniakova, O., Hudecova, D., Bransova, J. and Brtko, J. (1997). Derivatives of 4h-pyran-4-one with the atom of sulfur in the side chain. *Chemistry Papers*, 51: 421-426.
- Vachalkova, A., Bransova, J., Brtko, J., Uher, M. and Novotny, L. (1996). Polarographic behaviour of kojic acid and its derivatives, determination of potential carcinogenicity and correlation of these properties with their other attributes. *Neoplasma*, 43: 265-269.
- Villeneuve, P., Muderhwa, J.M., Graile, J. and Hass, M.J. (2000). Customizing lipases for biocatalysis: A survey of chemical physical and molecular biological approaches. *Journal of Molecular Catalysis B: Enzymatic*, 9:113-148.
- Wang, X., Miao, S., Wang, P. and Zhang, S. (2012). Highly efficient synthesis of sucrose monolaurate by alkaline protease Protex 6L. *Bioresource Technology*, 109:7-12.

- Watanabe, Y., Miyawaki, Y., Adachi, S., Nakanishi, K. and Matsuno, R. (2001b). Equilibrium constant for lipase-catalyzed condensation of mannose and lauric acid in water-miscible organic solvents. *Enzyme Microbiology Technology*, 29: 494–498.
- Watanabe-Akanuma, M., Inaba, Y. and Ohta, T. (2007). Mutagenicity of uv-irradiated maltol in *Salmonella typhimurium*. *Mutagenesis*, 22:43–47.
- Witek-Krowiak, A., Chojnacka, K., Podstawczyk, D., Dawiec, A. and Pokomeda, K. (2014). Applications of response surface methodology and artificial neural network methods in modelling and optimization of biosorption process. *Bioresource Technology*, 160:150–160.
- Xu, Y. (2012). *Process Technology for Immobilized Lipase-Catalyzed Reaction*. Phd Thesis, Technical University of Denmark, Kongens Lyngby, Denmark.
- Yabuta, T. (1924). The constitution of kojic acid: A d-pyrone derivative formed by *Aspergillus flavus* from carbohydrates. *Journal of Chemical Society Transaction*. 125:575–587.
- Yee, L.N., Akoh, C.C. and Phillips, R.S. (1997). Lipase PS-catalyzed transesterification of citronellyl butyrate and geranyl caproate: effect of reaction parameters. *Journal of the American Oil Chemists' Society*, 74:255–260.
- Youcefi, S., Bouzit, M., Ameer, H., Kamla, Y. and Youcefi A. (2013). Effect of some design parameters on the flow fields and power consumption in a vessel stirred by a rushton turbine. *Chemical and Process Engineering*, 34(2):293–307.
- Yuan, X., Liu, J., Zeng, G., Shi, J., Tong, J. and Huang, G. (2008). Optimization of conversion of waste rapeseed oil with high FFA to biodiesel using response surface methodology. *Renewable Energy*, 33:1678–1684.
- Zhang, X., Kobayashi, T., Adachi, S. and Matsuno, R. (2002). Lipase-catalyzed synthesis of 6-O-vinylacetyl glucose in acetonitrile. *Biotechnology Letters*. 24: 1097–1100.
- Zhao, H., Zhang, Y., Lu, F., Bie, X., Lu, Z. and Ning, H. (2011). Optimized enzymatic synthesis of ascorbyl esters from lard using Novozyme 435 in co-solvent mixture. *Journal of Molecular Catalysis B: Enzymatic*, 69:107–111.
- Zhen, H. and Xiang, X.Y. (2004). Regression-based artificial neural network methodology in response surface methodology. *Transactions of Tianjin University*, 10(2):153–157.
- Zirak, M. and Eftekhari-sis, B. (2015). Kojic acid in organic synthesis. *Turkish Journal of Chemistry*, 1:1–58.