



UNIVERSITI PUTRA MALAYSIA

***PHYTOCHEMICAL CONSTITUENTS FROM *Calophyllum buxifolium*
Vesque AND *Calophyllum hosei* Ridl AND THEIR BIOLOGICAL
ACTIVITIES***

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FS 2017 72



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AND *Calophyllum hosei* Ridl AND THEIR BIOLOGICAL ACTIVITIES**

By

SHAARI BIN DAUD

**Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfillment of the Requirements for the degree of Doctor of Philosophy**

May 2017

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

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May 2017

Chairman: Professor Gwendoline Ee Cheng Lian, PhD
Faculty: Science

Extensive phytochemical and biological studies to search for bioactive secondary metabolites from our Malaysian flora stem bark of two *Calophyllum* spp., *C. buxifolium* and *C. hosei* were carried out. These two plants were subjected to detail isolation work which involved extraction and purification of chemical components using various chromatographic techniques. All the pure compounds were identified using numerous spectroscopic analysis, such as 1D and 2D NMR, MS, IR, UV and also by comparison with literature data. These techniques have led to the isolation and elucidation of several chemical constituents of different type of classes which are the xanthenes, coumarins, flavonoid, terpenoid and sterols. The crude extracts and several pure compounds from both plants species were also screened for their cytotoxic, anti-inflammatory and antimicrobial activities using MTT (Microculture Tetrazolium Salt), nitric oxide (NO) and disc diffusion assay. Human promyelocytic leukemia (HL60) and human breast adenocarcinoma (MCF7) cell lines were used in the cytotoxic assay. The anti-inflammatory activity was tested against RAW 264.7 murine macrophage cells in vitro that were treated with lipopolysaccharide (LPS) which induce inflammatory response. The antimicrobial activity was tested against three Gram positive bacteria, *Bacillus subtilis* B145, *Staphylococcus aureus* S276 and *Staphylococcus epidermis* S273 and two Gram negative bacteria, *Escherichia coli* E266 and *Serratia marcescens* S381.

Detail work on the stem bark of *C. buxifolium* a plant never reported before for its phytochemistry has led to the isolation of two new chemical components, buxixanthone (215) and benjaminin (223) together with another 14 known compounds that consists of xanthenes, flavonoids and terpenoids. Meanwhile from the study on three different extracts from the stem bark of *C. hosei* also an unreported plant, has successfully result in one new coumarin, hoseimarin (222) along with other 12 known xanthenes and one common triterpenoid. There has been no previous report on the phytochemical work on the *C. hosei*.

The crude extracts for both plants gave a weak cytotoxic activity through MTT assay against two human cancer cell lines, HL60 (human promyelocytic leukemia) and MCF7 (human breast adenocarcinoma) cell line. Only four compounds, benjaminin (**223**), mangostingone (**220**), rubraxanthone (**221**) and β -mangostin (**178**) gave some activities towards the HL60 cell line with IC₅₀ values of 96.13, 46.92, 10.53 and 7.16 μ g / mL, respectively.

Anti-inflammatory assay was carried out for the crude extracts and 14 pure compounds using the nitric oxide (NO) assay. Most of the crude extracts for both plants gave a weak inhibition of NO production in RAW 264.7 cells. Eight xanthones, rubraxanthone (**221**), buxixanthone (**215**), ananixanthone (**183**), dombakinoxanthone (**95**), macluraxanthone (**87**), β -mangostin (**178**), 1,3,7-trihydroxy-2-(3-methylbut-2-enyl)-xanthone (**218**) and mangostingone (**220**) gave IC₅₀ values of 6.45, 6.84, 7.14, 7.57, 9.07, 11.68, 12.05 and 12.34 μ g / ml indicating their strong efficacy in reducing the NO production in RAW 264.7 cells. On the other hand, six more pure compounds gave mild inhibition on NO production in RAW 264.7 cells.

All the crude extracts for *C. buxifolium* and *C. hosei* gave no inhibition towards three Gram positive bacteria, *Bacillus subtilis* B145, *Staphylococcus aureus* S276 and *Staphylococcus epidermis* S273 and two Gram negative bacteria, *Escherichia coli* E266 and *Serratia marcescens* S381.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**SEBATIAN FITOKIMIA DARI *Calophyllum buxifolium* Vesque DAN
Calophyllum hosei Ridl DAN AKTIVITI BIOLOGIKALNYA**

Oleh

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Kajian intensif fitokimia dan biologi dalam pencarian metabolit sekunder dari flora Malaysia telah dijalankan terhadap dua spesies *Calophyllum*, iaitu *C. buxifolium* dan *C. hosei*. Dua pokok ini akan dikaji secara terperinci di mana ia melibatkan pengekstrakan dan penulenan komponen kimia dengan menggunakan pelbagai kaedah kromatografi. Semua sebatian tulen dikenalpasti menggunakan pelbagai analisis spektroskopi seperti 1D dan 2D Resonan Magnetik Nuklear (RMN), spektroskopi jisim (MS), Infra merah (IR), ultra violet (UV) dan juga perbandingan dengan data terdahulu.

Melalui teknik-teknik ini ia telah membolehkan pengasingan dan pengenalpastian untuk beberapa jenis kelas sebatian kimia iaitu xanthon, kumarin, flavonoid, terpenoid dan sterol. Ekstrak mentah dan beberapa sebatian tulen daripada kedua-dua spesies pokok ini juga diuji untuk aktiviti sitotoksik, anti keradangan dan aktiviti mikrobial dengan menggunakan asai MTT (Garam tetrazolium mikrokultur), nitrik oksida (NO) dan cerakin penyebaran cakera. Dua talian sel karsinoma manusia iaitu sel karsinoma promyelocytic leukemia (HL60) dan sel karsinoma payu dara (MCF7) telah digunakan di dalam ujian sitotoksik ini. Anti keradangan pula telah diuji secara *in vitro* dengan menggunakan sel RAW 264.7 murin makrofaj yang telah dirawat dengan lipopolisakarida (LPS) di mana ia bertindak sebagai agen keradangan. Tiga bakteria gram positif iaitu *Bacillus subtilis* B145, *Staphylococcus aureus* S276 dan *Staphylococcus epidermis* S273 dan dua bakteria gram negatif, *Escherichia coli* E266 dan *Serratia marcescens* S381 telah digunakan dalam menguji aktiviti antimikrobial.

Kajian fitokimia yang terperinci terhadap kulit batang pokok *C. buxifolium* telah membolehkan pengasingan dua sebatian kimia baru iaitu buksixanthon (**215**) dan benjaminin (**223**) bersama dengan 13 lagi xanthon sebatian biasa yang terdiri daripada xanthan, flavonoid, triterpenoid dan sterol. Sementara itu kajian terhadap tiga ekstrak mentah daripada satu lagi pokok yang belum pernah dikaji iaitu *C. hosei* telah berjaya mengasingkan satu kumarin yang baru iaitu hoseimarin (**222**) bersama dengan 12 lagi xanthon dan satu triterpenoid biasa. Tiada laporan tentang kajian fitokimia terhadap *C. hosei* dilaporkan sebelum ini.

Ekstrak mentah dari kedua-dua pokok memberi aktiviti sitotoksik yang lemah terhadap dua talian sel barah iaitu HL60 (human promyelocytic leukemia) dan MCF7 (human breast adenocarcinoma). Hanya empat sebatian kimia iaitu, benjaminin (**223**), mangostingon (**220**), rubraxanthon (**221**) and β -mangostin (**178**) memberi aktiviti terhadap sel HL60 masing-masing dengan nilai IC_{50} 96.13, 46.92, 10.53 dan 7.16 $\mu\text{g} / \text{mL}$.

Kebanyakan ekstrak mentah daripada kedua-dua tumbuhan ini memberi nilai perencatan penghasilan NO pada sel RAW 264.7. Lapan xanthon, rubraxanthon (**221**), buksixanthon (**215**), ananixanthon (**183**), dombakinaxanthon (**95**), makluraxanthon (**87**), β -mangostin (**178**), 1,3,7-trihidroksi-2-(3-metilbut-2-enil)-xanthon (**218**) dan mangostingon (**220**) memberi nilai IC_{50} 6.45, 6.84, 7.14, 7.57, 9.07, 11.68, 12.05 dan 12.34 $\mu\text{g} / \text{ml}$ menunjukkan keberkesanan yang kuat dalam menurunkan penghasilan NO di dalam sel RAW 264.7. Manakala, enam lagi sebatian kimia tulen memberi perencatan sederhana pada penghasilan NO dalam sel RAW 264.7.

Semua ekstrak mentah untuk *C. buxifolium* dan *C. hosei* tidak memberikan sebarang perencatan terhadap tiga bakteria Gram positif, *Bacillus subtilis* B145, *Staphylococcus aureus* S276 dan *Staphylococcus epidermis* S273 dan dua bakteria Gram negatif, *Escherichia coli* E266 dan *Serratia marcescens* S381.

ACKNOWLEDGEMENTS

Bismillahirrahmanirahim

First of all, I am grateful to my supervisor, Prof. Dr. Gwendoline Ee Cheng Lian, who always understood and supported my research project. Besides that, she also gave precious advice and continuous guidance throughout my research project from which I was able to gain a lot of experience and knowledge. It was a pleasure working with her. I also want to thank my co-supervisors, Dr Emilia Abdul Malek and the Associate Prof Dr. Zuraini Ahmad who gave their precious time, support and motivated me in order to finish my research project. Special thanks to Dr. Felix Yong, Dr. Teh Soek Sin, Nurnabila, Nurfarahdilla for their guidance in the anti-inflammatory work.

I also would like to express my gratitude to all my labmates, Syarik, Azri, Sham, Irene, Kar Wei, Thiru, Ain, Mr. Muniandy and Ka Wong for their willingness to help and share their knowledge as well as be a good listener for me. On the other hand, all of them had made an awesome working environment which all of us unite as one team. I am really happy to have worked with all of them. Special thanks goes to Nazil, Kamilah, Ms. Shareena Safiai and Mr. Johadi Iskandar in their guidance in the NMR instrument. I am also grateful to Mr. Zainal Abidin Kassim and Mrs. Rusnani Amirudin for running all my compounds for GCMS and IR analysis. I am highly indebted to Prof. Dr. Jegak Uli for collection of plant samples.

This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfillment of requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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- the research conducted and the writing of this thesis was under our supervision
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LIST OF ABBREVIATIONS

δ	Chemical shift in ppm
λ_{max}	Wavelength maxima in nm
$^{\circ}\text{C}$	Degree celcius
^1H	Proton
^{13}C	Carbon-13
IC_{50}	Initial Concentration to kill 50% of cells
cm	centimeter
d	Doublet
D	Deuterated proton
DEPT	Distortionless Enhancement by Polarization Transfer
EIMS	Electron ionization mass spectrometry
EtOH	Ethanol
FTIR	Fourier Transform Infra-Red
g	Gram
GC-MS	Gas chromatography-mass spectroscopy
HIV	Human immunodeficiency virus
HPLC	High performance liquid chromatography
Hz	Hertz
IR	Infra-Red
J	Coupling constant
Lit.	Literature review
m	Multiplet
m	meter
$[\text{M}]^+$	Molecular ion

mg	Milligram
MHz	Megahertz
m.p.	Melting point
MS	Mass Spectrometry
m/z	Mass per charge
NMR	Nuclear Magnetic Resonance
nm	nanometer
R _f	Retention Factor
s	Singlet
t	Triplet
TLC	Thin layer chromatography
TMS	Tetramethylsilane
μg	microgram
μM	Micro molar
UATR	Universal Attenuated Total Reflection
UV	Ultraviolet
ν _{max}	Wavenumber maxima in cm ⁻¹

CHAPTER 1

INTRODUCTION

1.1 General introduction

Phytochemistry are usually referred or deals to the analysis of plant chemical that found in plants, which gave health benefits for humans further than those attributed to macronutrients and micronutrients (Hasler *et al.*, 1999). Meanwhile, phytochemicals can be described as plant chemicals that protect plant cells from environmental hazards such as pollution, stress, UV exposure, drought and pathogenic attack. Phytochemicals can be classified to primary or secondary compounds. The common sugars, amino acids, proteins, purines and chlorophyll's are consider as primary constituents. On the other hand, alkaloids, flavonoids, saponins, terpenes, lignans, phenolics and glucosides are the remaining plant chemicals whereas classified as secondary constituents (Hana Ni, 1998). Phenolics are the most numerous and structurally diverse plant phytoconstituents based on literature survey. Phytochemical constituents also can be referred as natural products since it source come from plants.

Natural products can be defined as secondary metabolites produced by organisms in response to external stimuli such as nutritional changes, infections and competition. Natural products are usually isolated as biologically active pharmacophores from plants, fungi, bacteria, protozoans, insects and animal. These valuable natural products can be highlighted by their usage today in medical and animal health industries such as lovastatin (anticholesterolemic agent), paclitaxel and doxorubicin (antitumor agents), erythromycin (antibiotic), and amphotericin B (fungicidal agent).

Natural products have played an important role in the drug discovery process. Most of the old society and agrarian societies use plant-derived natural products as therapies for quite a number of diseases ranging from infections to emphysema. About one third of the top selling drugs in the world are natural products or their derivatives. Natural products in extract form consist of various secondary metabolites. Some of these metabolites can react as active components and give positive effects to some diseases. In order to obtain these active pure compounds, extraction and isolation processes need to be carried out. However these processes are tedious and time consuming. The inavailability of sufficient plant materials, selection and implementation of appropriate screening bioassay and to obtain these bioactive compounds in large amounts are also the constraints for the search for new lead compounds in drug discovery.

Even though the process of finding new active compounds from natural product research is difficult compared to the synthetic conventional drug research most users still prefer drugs that are derived from natural products. This is probably due to the fact that the natural product secondary metabolites have fewer side effects as compared to synthetic drugs and they are also safe therapeutic agents (Deltito *et al.*, 1998).

Moreover, the increasing public concern for a healthy lifestyle has made the natural extracts to be more reliable as nutraceuticals and agrochemicals for them.

Since secondary metabolites are structurally complex molecules, characterization need to be carried out using modern spectroscopic instruments such as gas chromatography-mass spectrometry (GCMS), nuclear magnetic resonance (NMR), infrared (IR) and ultra-violet spectroscopy (UV). Identification of naturally occurring products can be solved by natural product chemists using those instruments. Alkaloids, steroids, coumarins, xanthenes, acids and flavonoids are the commonly isolated secondary metabolites that have been reported to possess medicinal properties.

1.2 Botany of Plants Studied

1.2.1 The Family Clusiaceae

The Clusiaceae family, formerly known as the Guttiferae, belongs to the order of Malpighiales. They consists of about 40 genera and over 1600 species of trees and shrubs (Goh *et al.*, 1992). Basically there are 4 important genera of the Clusiaceae of importance namely *Garcinia*, *Calophyllum*, *Mesua* and *Mammea*. *Calophyllum* species are important for their timbers, while some species of *Garcinia* produce edible fruits and important resin in pigment manufacturing.

Members of the Clusiaceae are usually found as trees or shrubs and can be recognized by the latex in their stem, leaves, fruit and bark and fruits of capsules for seeds. The flowers are bee-pollinated, actinomorphic with numerous stamens and separate petals. The leaves are variable in size with opposite leaves often with fine parallel nerves and without true stipulates.

1.2.2 The Genus *Calophyllum*

The *Calophyllum* plants are usually referred to as “beautiful leaf”. This is because the name “*Calophyllum*” comes from the Greek language, in which “kalos” means beautiful, while “phullon” means leaf. This genus has around 180-200 species of tropical evergreen trees in the family Clusiaceae. It is widely distributed in Australasia, Madagascar, Eastern Africa, South and Southeast Asia, the Pacific islands, the West Indies and Latin America (Morel *et al.*, 2000). There are a few common names for *Calophyllum*, whereas based on the geographical areas such as Malaysia it is known as Bintagor tree, India as Poon tree, and Latin America as Guanandi, Jacareuba or Santa Maria.

Calophyllum plants are mainly widespread from the sea-shore to the highest mountain tops, mixed peat swamps, freshwater swamp and mixed dipterocarp forests. Trees from

this genus are usually found as small to medium-sized, without buttresses. Meanwhile, the inner barks of this genus are pink to red, laminated, exuding sticky and yellow or white varnish. On the other hand, leaves of this genus are generally narrow and less channeled above while the flowers and fruits are favoured by bees, birds and small mammals. Wood from this genus are usually used for local purposes while the large round nuts have contributed to the main source for seed oil production (Dweck *et al.*, 2002).

1.2.3 The Species *Calophyllum buxifolium*

Calophyllum buxifolium is a small gnarled tree with low buttresses. This tree can reach 80 ft. in height. This tree has ruggedly thick flaky bark surface and is chocolate-brown colour. Furthermore, the inner bark is orange-brown and exudate is yellow in colour while the leaves are broadly elliptic and variable in shape but always thickly coriaceous. The flower is 7 mm wide and the fruit 2.5 cm long, 1.8 cm wide, ellipsoid and apiculate. This fruit will be chocolate-brown in colour when it dries.

This species is usually distributed at Philippines, Sabah, Sarawak and Brunei. Their habitat is common on skeletal soils or on sandy plateaux. There are no phytochemical works yet for this species.



Figure 1.1: The Leaves and Twigs of *Calophyllum buxifolium*

1.2.4 The Species *Calophyllum hosei*

Calophyllum hosei is widely distributed in Borneo, Sarawak and Brunei. *Calophyllum hosei* is a small to medium sized tree which occasionally can grow up to 80 ft tall and 5 ft wide. This species also has a glabrous pubescent bud and shiny leaf blade which then changes to copper-brown when the leaf dries up. *Calopyllum hosei* commonly grows in ground-water podsols and shallow peat.

There are no reports on their uses and phytochemistry.



Figure 1.2: The Leaves and Twigs of *Calophyllum hosei*

1.3 Problem Statement

Cancer disease is one of the major threats to human beings in this world. The number of people suffering from this disease keeps on increasing year by year. As we know inflammation is one of the causes that contribute to cancer disease. Inflammation is a normal and main process created by nature by our body. The process of inflammation helps to get rid of unwanted bacteria, and other invaders. Inflammation also helps our body to clean up dead cells from infections. Chronic inflammation can drive to cancer disease. Thus, research on the finding of active crude extracts or pure compounds from natural resources for future anticancer and anti-inflammatory drugs need to be carried out.

The use of natural products as therapeutics and their medicinal properties have been known since a long time ago. This can be clearly seen by the ancient society using plant crude extracts in their routine life in therapies for treating diseases. Synthetic drugs that we have been using to treat disease are now believed to give long term side effects. Hence, work on the discovery of drugs from natural active compounds that have less side effects from plants is still in high demand.

The discovery of new chemical constituents that have bioactive properties through screening of natural products from *Calophyllum* which are not accessible by other technologies will be of great importance to the pharmaceutical industry and public health. The isolation techniques and structural elucidation of new compounds as well as bioassay results will also be useful for other natural product chemists for future research work. There are no reports on the phytochemistry of *Calophyllum buxifolium* and *Calophyllum hosei*. The significant discovery of lead compounds from this research

can contribute to the search for alternative drugs to the presently available drugs in the market.

1.4 Objectives of Study

The main objectives of this study are:

1. To extract and isolate the chemical components from the stem bark of *Calophyllum buxifolium* and *Calophyllum hosei*
2. To identify and elucidate the structures of the pure components using various spectroscopic methods.
3. To screen and evaluate the anti-inflammatory, anti-cancer and anti-microbial biological activities of each crude extract and pure isolated constituents.

BIBLIOGRAPHY

- Ali, M.S., Mahmud, S., Perveen, S., Ahmad, V.U. and Rizwani, G.H. (1999). Epimers from the leaves of *Calophyllum inophyllum*. *Phytochemistry* 50:1385-1389.
- Ampofo, S. A. and Waterman, P. G. (1986). Xanthones and neoflavonoids from two Asian species of *Calophyllum*. *Phytochemistry* 25(11):2617-2620.
- Babu, V., Arya, R., Ilyas, M. and Nasim, K. T. (1994). 9-hydroxy-2,2,6,7-tetramethyl-2H-[1]-benzopyran-(1-phenylethylene-10-yl)[3,2-b]-dihdropyran-4-one from *Calophyllum tomentosum*. *Phytochemistry* 35(2):507-510.
- Bandara, B. M. R., Dharmaratne, H. R. W., Sotheeswaran, S., and Balasubramaniam, S. (1986). Two chemically distinct groups of *Calophyllum* species from Sri Lanka. *Phytochemistry* 25 (2):425-428.
- Bhanu. S. and Scheinmann, F. (1975). Xanthones from the heartwood of *Calophyllum ramiflorum*. *Phytochemistry* 14:298-299.
- Cao, S.G., Sim, K.Y., Pereira, J. and Goh, S.H. (1998). Coumarins from *Calophyllum teysmannii*. *Phytochemistry* 47(5):773-777.
- Chaturvedula, V.S.P. and Prakash, I. (2012). Isolation of Stigmasterol and β -Sitosterol from the dichloromethane extract of *Rubus suavissimus* *International Current Pharmaceutical Journal* 1(9):239-242.
- Chen, G.-Y., Zhu, G.-Y., Han, C.-R., Zhao, J., Song, X.-P. and Fong, W.-F. (2008). A new pyranoxanthone from the stems of *Calophyllum membranaceum*. *General Papers* xiii:249-254.
- Cortez, D.A.G., Young, M.C.M, Marston, A., Wolfender, J.-L. and Hostettmann, K. (1998). Xanthones, Triterpenes and A Biphenyl from *Kielmeyera coriacea*. *Phytochemistry*. 7:1367-1374.
- Dachriyanus, Wahyuni, F.S., Shaari, K., Stanslas, J. and Lajis, N. H. (2015). Cytotoxic xanthones from the stem bark of *Garcinia cowa*. *Journal of Chemical and Pharmaceutical Research*, 2015, 7(1):227-236.
- Deltito, J. and Beyer, D. (1998). The scientific, quasi-scientific and popular literature on the use of St. John's Wort in the treatment of depression. *Journal of Affective Disorders* 51(3):345-351.
- Dharmaratne, H.R. and Marasinghe, G.P. (2009a). New methylethers of cordatolides from *Calophyllum cordato-oblongum* and their synthesis. *Natural Product Research* 23(10):883-890.
- Dharmaratne, H.R., Napagoda, M.T. and Tennakoon, S.B. (2009b). Xanthones from roots of *Calophyllum thwaitesii* and their bioactivity. *Natural Product Research* 23(6):539-545.

- Dharmaratne, H. R. W., Sotheeswaran, S., and Balasubramaniam, S. (1984). Triterpenes and neoflavanoids of *Calophyllum lankaensis* and *Calophyllum thwaitesii*. *Phytochemistry* 23(11):2601-2603.
- Dharmaratne, H.R., Sotheeswaran, S., Balasubramaniam, S. and Waight, E.S. (1985). Triterpenoids and coumarins from the leaves of *Calophyllum cordatoblongum*. *Phytochemistry* 24(7):1553-1556.
- Dharmaratne, H. R. W., Sotheeswaran, S., Balasubramaniam, S. and Reisch, J. (1984). Xanthenes from roots of three *Calophyllum* species. *Phytochemistry* 25(8):1957-1959.
- Dharmaratne, H.R., Wanigasekera, W.M., Mata-Greenwood, E. and Pezzuto, J.M. (1998b). Inhibition of human immunodeficiency virus type 1 reverse transcriptase activity by cordatolides isolated from *Calophyllum cordatoblongum*. *Planta Medica* 64(5):460-461.
- Dharmaratne, H.R., Wijesinghe, W.M. and Thevanasem, V. (1999). Antimicrobial activity of xanthenes from *Calophyllum* species, against methicillin-resistant *Staphylococcus aureus* (MRSA). *Journal of Ethnopharmacology* 66(3):339-342.
- Dharmaratne, H. R. W. and Wuesinghe, W. M. N. M. (1997). A trioxxygenated diprenylated chromenxanthone from *Calophyllum moonii*. *Phytochemistry* 46(7):1293-1295.
- Dweck, A.C. and Meadows, T. (2002). Tamanu (*Calophyllum inophyllum*) - the African, Asian, Polynesian and Pacific Panacea. *Int J Cosmet Sci.* 24(6):341-348.
- Ee, G.C., Kua, A.S., Lim, C.K., Jong, V. and Lee, H.L. (2006). Inophyllin A, a new pyranoxanthone from *Calophyllum inophyllum* (Guttiferae). *Natural Product Research* 20(5):485-491.
- Ee, G.C.L., Kua, A.S.M., Cheow, Y.L., Lim, C.K., Jong, V. and Rahmani, M. (2004a). A new pyranoxanthone inophyllin B from *Calophyllum inophyllum*. *Natural Product Sciences* 10(5):220-222.
- Ee, G.C.L., Mah, S.H., Rahmani, M., Taufiq-Yap, Y.H., Teh, S.S. and Lim, Y.M. (2011a). A new furanoxanthone from the stem bark of *Calophyllum inophyllum*. *Journal of Asian Natural Products Research* 13(10):956-960.
- Ee, G.C.L., Mah, S.H., Teh, S.S., Rahmani, M., Go, R. and Taufiq-Yap, Y.H. (2011b). Soulamarin, a new coumarin from stem bark of *Calophyllum soulattri*. *Molecules* 16(11):9721-9727.
- Ee, G.C.L., Ng, K.N., Taufiq-Yap, Y.H., Rahmani, M., Ali, A.M. and Muse, R. (2004b). Mucigerin, a new coumarin from *Calophyllum mucigerum* (Guttiferae). *Natural Product Research* 18(2):123-128.

- Ferchichi, L., Derbré, S., Mahmood, K., Touré, K., Guilet, D., Litaudon, M., Awang, K., Hadi, A. H. A., Ray, A. M. L. and Richomme, P. (2012). Bioguided fractionation and isolation of natural inhibitors of advanced glycation end-products (AGEs) from *Calophyllum flavoramulum*. *Phytochemistry* 78:98-106.
- Goh, S.H. and Jantan, I. (1991). A xanthone from *Calophyllum inophyllum*. *Phytochemistry* 30(1):366-367.
- Gautier, J., Kunesch, G. and Polonsky, J. (1972). Structure of calophynic acid, a novel constituent of *Calophyllum inophyllum*. *Tetrahedron Letters* 27:2715-2718.
- Goh, S.H., Jantan, I. and Waterman, P.G. (1992). Neoflavonoid and biflavonoid constituents of *Calophyllum inophylloide*. *Journal of Natural Products* 55(10):1415-1420.
- Govindachari, T.R., Viswanathan, N., Pai, B.R., Rao, R. and Srinivasan, M. (1967). Triterpenes of *Calophyllum inophyllum* Linn. *Tetrahedron* 23(4):1901-1910.
- Guerreiro, E., Kunesch, G. & Poloivsky, J. (1971). Les constituants des grains de *Calophyllum chapelierii* (Guttiferae). *Phytochemistry* 10:2139-2145
- Gunatilaka, A.A.L., Jasmin De Silve, A.M.Y., Subramanian, S.S., Balasubramaniam, S. and Wazeer, M.I.M. (1984). Terpenoid and biflavanoid constituent of *Calophyllum calaba* and *Garcinia spicata* from Sri Lanka. *Phytochemistry* 23(2):323-328.
- Hahn Ni (1998). Is Phytoestrogens Nature's Cure for What Ails Us? A Look at the Research. *Journal of the American Dietetic Association* 98: 974-976.
- Hasler C.M. and Blumberg J.B. (1999) Symposium on Phytochemicals: Biochemistry and Physiology. *Journal of Nutrition* 129: 756S-757S.
- Jackson, B., Locksley, H.D. and Scheinmann, F. (1968). The isolation of 6-(3,3-dimethylallyl)-1,5-dihydroxyxanthone and two related metabolites from *Calophyllum scriblitifolium* Henderson and Wyatt. *Tetrahedron* 24:3059-3068.
- Jackson, B., Locksley, H.D. and Scheinmann, F. (1969). The isolation of 6-desoxyjacareubin, 2-(3,3-dimethylallyl)-1,3,5,6-tetrahydroxyxanthone and jacareubin from *Calophyllum inophyllum*. *Phytochemistry* 8:927-929.
- Ha, L.D., Hansen, P.E., Duus, F., Pham, H.D. and Nguyen L-H.D. (2012). A new chromanone acid from the bark of *Calophyllum dryobalanoides*. *Phytochemistry Letters* 5:287-291.
- Hay, A-E., He'lesbeuxa, J-J., Duvala, O., LabaRedb, M., Grellierb, P. and Richommea, P. (2004). Antimalarial xanthenes from *Calophyllum caledonicum* and *Garcinia vieillardii*. *Life Sciences* 75:3077-3085.

- Honda, P.A., Ferreira, I.C., Cortez, D.A., Amado, C.A., Silveira, T.G., Brenzan, M.A. and Lonardoni, M.V. (2010). Efficacy of components from leaves of *Calophyllum brasiliense* against *Leishmania (Leishmania) amazonensis*. *Phytomedicine* 17(5):333-338.
- Hostettmann K. Plant-derived molluscicides of current importance. Economic and Medicinal Plant Research. Vol. 3. London: Academic Press; 1989.
- Iinuma, M., Ito, T., Tosa, H., Tanaka, T., Miyake, R. and Chelladura, V. (1997). Prenylated xanthonoids from *Calophyllum apetalum*. *Phytochemistry* 46(8):1423-1429.
- Iinuma, M., Tosa, H., Tanaka, T. and Yonemori, S. (1994a). Two new xanthenes in the underground part of *Calophyllum inophyllum*. *Heterocycles* 37(2):833-838.
- Iinuma, M., Tosa, H., Tanaka, T. and Yonemori, S. (1994b). Two xanthenes from root bark of *Calophyllum inophyllum*. *Phytochemistry* 35(2):527-532.
- Iinuma, M., Tosa, H., Tanaka, T. and Yonemori, S. (1995). Two xanthenes from roots of *Calophyllum inophyllum*. *Phytochemistry* 38(3):725-728.
- Iinuma, M., Tosa, H., Toriyama, N., Tanaka, T., Ito, T. and Celladurai, V. (1996). Six xanthenes from *Calophyllum austroindicum*. *Phytochemistry* 43(3):681-685.
- Ismail, A.A.F, Ee G₂C₂L, Daud, S., Teh, S.S., Hashim, H.M. and Awang, K., (2015). Venuloxanthone, a new pyranoxanthone from the stem bark of *Calophyllum venulosum*. *Journal of Asian Natural Products Research* 17:1104-1108.
- Ito, C., Itoigawa, M., Mishina, Y., Filho, V.C., Enjo, F., Tokuda, H., Nishino, H. and Furukawa, H. (2003). Chemical constituents of *Calophyllum brasiliense*. 2. Structure of three new coumarins and cancer chemopreventive activity of 4-substituted coumarins. *Journal of Natural Products* 66(3):368-371.
- Ito, C., Itoigawa, M., Mishina, Y., Filho, V.C., Mukainaka, T., Tokuda, H., Nishino, H. and Furukawa, H. (2002). Chemical constituents of *Calophyllum brasiliensis*: structure elucidation of seven new xanthenes and their cancer chemopreventive activity. *Journal of Natural Products* 65:267-272.
- Itoigawa, M., Ito, C., Tan, H.T., Kuchide, M., Tokuda, H., Nishino, H. and Furukawa, H. (2001). Cancer chemopreventive agents, 4-phenylcoumarins from *Calophyllum inophyllum*. *Cancer Letters* 169(1):15-19.
- Jun, C.J., Min, X, De, L.S., Ying, W.H. and Chu, X.J. (2001). Chemical Constituents of *Calophyllum polyanthum*. *Acta Botanica Yunnanica* 23(4):521-526.
- Jung, H-A., Su, B-N., Keller, W.J., Mehta, R. G. and Kinghorn, D. (2006). Antioxidant Xanthenes from the Pericarp of *Garcinia mangostana* (Mangosteen) *J. Agric. Food Chem.* 54:2077-2082.

- Joaquim, C. B., Mara, S. P. A., & Marcelino, S. N. (1998). A prenylated xanthone from the bark of *Symphonia globulifera*. *Phytochemistry* 49(4):1159-1160.
- Karunanayake, S., Sotheeswaran, S. and Sultanbawa, M.U.S. (1979). Zeyloxanthone, a new tetrahydroxyxanthone from *Calophyllum zeylanicum* (Guttiferae). *Tetrahedron Letters* 16:1449-1450.
- Karunanayake, S., Sotheeswaran, S., Uvais, M., Sultanbawa, M.U.S. and Balasubramaniam, S. (1981). Xanthones and triterpenes of *Calophyllum tomentosum*. *Phytochemistry* 20(6):1303-1304.
- Kashman, Y., Gustafson, K.R., Fuller, R.W., Cardellina, J.H., McMahon, J.B., Currens, M.J., Buckheit, R.W., Jr., Hughes, S.H., Cragg, G.M. and Boyd, M.R. (1992). The calanolides, a novel HIV-inhibitory class of coumarin derivatives from the tropical rainforest tree, *Calophyllum lanigerum*. *Journal of Medicinal Chemistry* 35(15):2735-2743.
- Kawazu, K., Ohigashi, H., Takahashi, N. and Mitsui, T. (1972). Piscicidal constituents of *Calophyllum inophyllum*. *Bulletin of the Institute for Chemical Research, Kyoto University* 50(3):160-167.
- Kijjoo, A., Gonzalez, M.J., Pinto, M.M., Silva, A.M., Anantchoke, C. and Herz, W. (2000). Xanthones from *Calophyllum teysmannii* var. *inophylloide*. *Phytochemistry* 55(7):833-836.
- Kumar, V., Ramachandran, S. and Sultanbawa, M. U. S. (1976). Xanthones and triterpenoids from timber of *Calophyllum inophyllum*. *Phytochemistry* 15:2016-2017.
- Kumar, V., Sotheeswaran, S., Surendrakumar, S. and Balasubramaniam, S. (1982). Calocalabaxanthone, the putative isoprenyl precursor of calabaxanthone from *Calophyllum calaba*. *Phytochemistry* 21(3):807-809.
- Lamari, F.N. and Cordopatis, P. (2008). Exploring the potential of Natural Products in Cancer treatment. *Anticancer Therapeutics* 1.
- Li, Z.-L., Liu, D., Li, D.-Y. and Hua, H.-M. (2011). A novel prenylated xanthone from the stems and leaves of *Calophyllum inophyllum*. *Natural Product Research* 25(9):905-908.
- Lopes, J.L.C., Lopes, J.N.C., Gilbert, B., Bonini, S.E. (1977). Osajaxanthone from *Kielmeyera coriacea*. *Phytochemistry* 16:1101-1101.
- Mah, S.H., Ee, G.C.L., Rahmani, M., Taufiq-Yap, Y.H., Go, R. and Teh, S.S. (2011a). A new pyranoxanthone from the stem bark of *Calophyllum inophyllum*. *Letters in Organic Chemistry* 8(6):447-449.
- Mah, S.H., Ee, G.C.L., Rahmani, M., Taufiq-Yap, Y.H., Sukari, M.A. and Teh, S.S. (2011b). A new pyranoxanthone from *Calophyllum soulattri*. *Molecules* 16 (5):3999-4004.

- Mah, S.H., Ee, G.C.L., Teh, S.S., Rahmani, M., Lim, Y.M. and Go, R. (2012). Phylatrin, a new cytotoxic xanthone from *Calophyllum soulattri*. *Molecules* 17(7):8303-8311.
- Mah, S. H., Ee, G. C. L., Teh, S. S. and Sukari, M. A. (2015). Antiproliferative xanthone derivatives from *Calophyllum inophyllum* and *Calophyllum soulattri*. *Pak. J. Pharm. Sci.* 28(2):425-429.
- McKee, T.C., Fuller, R.W., Covington, C.D., Cardellina, J.H., 2nd, Gulakowski, R.J., Krepps, B.L., McMahon, J.B. and Boyd, M.R. (1996). New pyranocoumarins isolated from *Calophyllum lanigerum* and *Calophyllum teysmannii*. *Journal of Natural Products* 59(8):754-758.
- Mesquita A.A.L, Oliveira W.G.D., Neiva R.M.T. (1975). Xanthenes from *Tovomita pyrifolium*. *Phytochemistry* 14:803-80.
- Ming, M., Zhang, X., Hai-Feng Chen, H-F., Zhu, L-J., Zeng, D-Q., Yang, J., Wu, G-X., Wu, Y-Z., and Yao, X-S. (2016). RXR α transcriptional inhibitors from the stems of *Calophyllum membranaceum*. *Fitoterapia* 108:66-72.
- Morel, C., Seraphin, D., Oger, J.M., Litaudon, M., Sevenet, T., Richomme, P. and Bruneton, J. (2000). New xanthenes from *Calophyllum caledonicum*. *Journal of Natural Products* 63(11):1471-1474.
- Morel, C., Seraphin, D., Teyrouz, A., Larcher, G., Bouchara, J.P., Litaudon, M., Richomme, P. and Bruneton, J. (2002). New and antifungal xanthenes from *Calophyllum caledonicum*. *Planta Medica* 68(1):41-44.
- Mosmann, T. (1983). Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *Journal of Immunological Methods* 65:55-63.
- Nasir, N.D., Rahmani, M., Shaari, K., Kassim, N.K., Go, R., Stanslas, J. and Jeyaraj, E.J. (2013). Xanthenes from *Calophyllum gracilipes* and Their Cytotoxic Activity (Zanton daripada *Calofilum gracilipes* dan Aktiviti Ketoksikannya). *Sains Malaysiana* 42(9):1261-1266.
- Nguyen, L-T. T., Nguyen, H. T., Barbic, M., Brunner, G., Heilmann, J., Pham, H. D., Nguyen, D. M. and Nguyen. L-H. D. (2012). Polyisoprenylated acylphloroglucinols and a polyisoprenylated tetracyclicxanthone from the bark of *Calophyllum thorelii*. *Tetrahedron Letters* 53:4487–4493.
- Nigam, S.K. and Mitra, C.R. (1968). Constituents of *Calophyllum tomentosum* and *Calophyllum apetalum* nuts: structure of a new 4-alkyl- and of two new 4-phenyl-coumarins. *Planta Medica* 16(4):450-457.
- Ogunnusi, T.A., Oso, B.A. and Dosumu, O.O. (2010). Isolation and antibacterial activity of triterpenes from *Euphorbia kamerunica* Pax. *Int. J. Biol. Chem. Sci.* 4(1):158-167.

- Patil, A.D., Freyer, A.J., Eggleston, D.S., Haltiwanger, R.C., Bean, M.F., Taylor, P.B., Caranfa, M.J., Breen, A.L., Bartus, H.R. and Johnson, R.K. (1993). The inophyllums, novel inhibitors of HIV-1 reverse transcriptase isolated from the Malaysian tree, *Calophyllum inophyllum* Linn. *Journal of Medicinal Chemistry* 36(26):4131-4138.
- Prasad, J., Shrivastava, A., Khanna, A. K., Bhatia, G., Awasthi, S.K. and Narender, T. (2012). Antidyslipidemic and antioxidant activity of the constituents isolated from the leaves of *Calophyllum inophyllum*. *Phytomedicine* 19:1245– 1249.
- Pengsuparp, T., Serit, M., Hughes, S.H., Soejarto, D.D. and Pezzuto, J.M. (1996). Specific inhibition of human immunodeficiency virus type 1 reverse transcriptase mediated by soulattrolide, a coumarin isolated from the latex of *Calophyllum teysmannii*. *Journal of Natural Products* 59(9):839-842.
- Ravelonjato, B., Kunesch, N. and Poisson, J.E. (1987). Neoflavanoids from the stem bark of *Calophyllum verticillatum*. *Phytochemistry* 26(11):2973-2976.
- Ravelonjato, B., Libot, F., Ramiandrasoa, F., Kunesch, N., Gayral, P. and Poisson, J. (1992). Molluscicidal constituents of *Calophyllum* from Madagascar: activity of some natural and synthetic neoflavonoids and khellactones. *Planta Medica* 58(1):51-55.
- Reyes-Chilpa, R., Estrada-Muniz, E., Apan, T. R., Amekraz, B., Aumelas, A., Jankowski, C. K. and Vazquez-Torres, M. (2004). Cytotoxic effects of mammea type coumarins from *Calophyllum brasiliense*. *Life Sciences* 75:1635-1647.
- Sahimi, M. S. M., Ee, G. C. L., Mahaiyiddin, A. G., Daud, S., Teh, S. S., See, I. and Sukari, M. A. (2015). A New Natural Product Compound Benjaminin from *Calophyllum benjaminum*. *Pertanika J. Trop. Agric. Sci.* 38(1): 1-6
- Samaraweera, U., Sotheeswaran, S. and Sultanbawa, M.U.S. (1981). Calozeylanic acid, a new bark acid from three *Calophyllum* species (Guttiferae). *Tetrahedron Letters* 22(50):5083-5086.
- Sen, A. K., Sarkar, K. K., Majumder, P. C., and Banerji, N. 1981. Minor Xanthones of *Garcinia mangostana*. *Phytochemistry* 20(1): 183-185.
- Shen, Y.C., Hung, M.C., Wang, L.T. and Chen, C.Y. (2003). Inocalophyllins A, B and their methyl esters from the seeds of *Calophyllum inophyllum*. *Chemical & Pharmaceutical Bulletin* 51(7):802-806.
- Somanathan, R., Sultanbawa, S. and Uvais, M. (1974). Chemical investigation of Ceylonese plants. Part VIII. Trapezifolixanthone, a new di-isoprenylated xanthone from the bark of *Calophyllum trapezifolium* Thw. (Guttiferae). *Journal of the Chemical Society, Perkin Transactions* 1:2515-2517.

- Souza, M., D, O, C., Beserra, A.M., Martins, D.C., Real, V.V., Santos, R.A., Rao, V.S., Silva, R.M. and Martins, D.T. (2009). *In vitro and in vivo anti-Helicobacter pylori activity of Calophyllum brasiliense* Camb. *Journal of Ethnopharmacology* 123(3):452-428.
- Spino, C., Dodier, M. and Sotheeswaran, S. (1998). Anti-HIV coumarins from *Calophyllum* seed oil. *Bioorganic & Medicinal Chemistry Letters* 8(24):3475-3478.
- Waterman, P. G. and Crichton, E. G. (1980). Xanthenes and biflavonoids from *Garcinia densivenia* stem bark. *Phytochemistry* 19: 2733.
- Wei, D.-J., Mei, W.-L., Zhong, H.-M., Zeng, Y.-B., Wu, X.-D. and Dai, H.-F. (2011). A new prenylated xanthone from the branches of *Calophyllum inophyllum*. *Journal of Asian Natural Products Research* 13(3):265-269.
- Xiao, Q., Zeng, Y.B., Mei, W.L., Zhao, Y.X., Deng, Y.Y. and Dai, H.F. (2008). Cytotoxic prenylated xanthenes from *Calophyllum inophyllum*. *Journal of Asian Natural Products Research* 10(9-10):993-997.
- Yates P., and Bhat H.B. (1968). Structure of β -mangostin. *Canadian Journal of Chemistry* 46:3770-3772.
- Yimdjo, M.C., Azebaze, A.G., Nkengfack, A.E., Meyer, A.M., Bodo, B. and Fomum, Z.T. (2004). Antimicrobial and cytotoxic agents from *Calophyllum inophyllum*. *Phytochemistry* 65(20):2789-2795.
- Zakaria, M. B., Vijayasekaran, Ilham, Z. and Muhamad, N. A. (2014). Anti-Inflammatory Activity of *Calophyllum inophyllum* Fruits Extracts. *Procedia Chemistry* 13:218-220.
- Zou, J., Wu, J., Liu, S.-Z. and Zhao, W.-M. (2010). New coumarins and triterpenes from *Calophyllum inophyllum*. *Helvetica Chimica Acta* 93:1812-1821.
- Zhang, H., Tang, B. and Row, K. (2014). Extraction of catechin compounds from green tea with a new green solvent. *Chem. Res. Chin. Univ.* 30:37-41.