



**UNIVERSITI PUTRA MALAYSIA**

***SYNTHESIS AND CHARACTERIZATION OF SILVER /KAPPA-CARRAGEENAN NANOPARTICLES USING GREEN METHODS AND EVALUATION OF THEIR ANTIBACTERIAL ACTIVITIES***

**RANDA FAWZI ELSUPIKHE**

**FS 2017 71**



**SYNTHESIS AND CHARACTERIZATION OF SILVER /KAPPA-CARRAGEENAN NANOPARTICLES USING GREEN METHODS AND EVALUATION OF THEIR ANTIBACTERIAL ACTIVITIES**

By

**RANDA FAWZI ELSUPIKHE**

**Thesis Submitted to the School of Graduates Studies, Universiti Putra Malaysia,  
in Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

**March 2017**

## **COPYRIGHT**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



## DEDICATION

I dedicate this work to Allah for his tender unlimited and to my precious parents, my dear mum **Fatma Elgandoz** and my darling dad **Fawzi Elsupikhe**, which without their giving and their education, I could not make any success. I dedicate it with special thanks to my dear's Husband **Taha Husin** for his encouragements, understandings and helping during my study, which without him I would never able to finish my PhD and to my sweetheart daughters **Touka and Jana** for giving me the hope. I owe my loving thanks to my dear sister **Hanin**.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in Fulfillment  
of the requirement for the Degree of Doctor of Philosophy

**SYNTHESIS AND CHARACTERIZATION OF SILVER /KAPPA-  
CARRAGEENAN NANOPARTICLES USING GREEN METHODS AND  
EVALUATION OF THEIR ANTIBACTERIAL ACTIVITIES**

By

**RANDA FAWZI ELSUPIKHE**

**March 2017**

**Chairman : Professor Mansor Bin Ahmad, PhD**  
**Faculty : Science**

Nanoscale materials have received extensive attention because their unusual properties that differ significantly from bulk sample of the same material. Nanoparticles are particles with size less than 100 nm which small in diameter, but larger in surface area. Silver nanoparticles (Ag-NPs) are being increasingly used in consumer products such as water purification, household cleaning agents and huge in current many exclusive medical applications such as biological engineering. Synthesis of Ag-NPs has attracted the scientists' attention in recent years due to the huge advantages and applications of Ag-NPs especially as antimicrobial agent. Chemical methods have been used for the synthesis of Ag-NPs, but these methods have a lot of disadvantages because most of the chemical that have been used for synthesis the nanoparticles are too expensive and toxic, which are responsible for various biological risks. Also, most of the chemical methods for synthesis Ag-NPs are not able to control the size of the NPs. Furthermore, the agglomeration between the nanoparticles lead to bad results in antibacterial application. In this work, the green methods for synthesis Ag-NPs have been used for solving these problems and  $\kappa$ -carrageenan polymer has been used as a stabilizer to prevent this agglomeration. Ag-NPs in  $\kappa$ -carrageenan synthesized by different green methods (stirring method, UV- irradiation ultrasonic-irradiation) at room temperature were developed to prepare and control the size of Ag-NPs. Parameters such as the time of stirring, time of irradiation, ultrasonic amplitude, concentration of  $\text{AgNO}_3$  and concentration of  $\kappa$ -carrageenan have been optimized.  $\kappa$ -carrageenan was used as an eco-friendly stabilizer and  $\text{AgNO}_3$  as producer. Formation of Ag/ $\kappa$ -carrageenan was determined by the UV-visible spectra, which improved the formation of Ag-NPs by surface plasmon resonance in range 300-450 nm. The FT-IR spectra indicated the presence of  $\kappa$ -carrageenan in capping with Ag-NPs. The XRD analysis showed that the Ag-NPs were of face-centred cubic structure. TEM images illustrated the well dispersed of Ag-NPs with similar particle size. SEM images displayed the change on the surface morphology of the  $\kappa$ -carrageenan and illustrated the shape of the Ag-NPs.

EDXRF spectra of Ag-NPs in  $\kappa$ -carrageenan confirmed the presence of elemental compounds without any impurity peak. The antibacterial properties of the synthesized nanoparticles were evaluated using agar diffusion methods. Four species of bacteria were used in this study, including two Gram-positive and two Gram-negative bacteria: Methicillin Resistant *Staphylococcus aureus* (MRSA), *Bacillus subtilis*, *Pseudomonas aeruginosa* and *Escherichia coli* (*E-coli*). Optimized parameters in the stirring method for synthesis Ag-NPs were: 48 h of stirring times, 0.2 M of  $\text{AgNO}_3$  and 0.3%  $\kappa$ -carrageenan, which produced, the size and the concentration of Ag-NPs of 32 nm and 0.065 M, respectively. The good condition of UV-irradiation method for synthesis Ag-NPs were 60 min irradiation time, 0.2 M  $\text{AgNO}_3$  and 0.3%  $\kappa$ -carrageenan, which produced, the size of and the concentration Ag-NPs of 14 nm and 0.12 M, respectively. The conditions of the ultrasonic-irradiation method for synthesis Ag-NPs that give the best results were 90 min irradiation time, 0.15 M  $\text{AgNO}_3$ , 0.3 %  $\kappa$ -carrageenan and 60 amplitude, which produced the size and the concentration of Ag-NPs of 1.21 nm and 0.22 M, respectively. All Ag-NPs from the above methods were in spherical shape. The different methods demonstrated different results on anti-bacterial activity, which depended on the size and concentration of Ag-NPs. The stability test by using zeta potential analysis proved the Ag-NPs that synthesized by stirring method, UV-irradiation and ultrasonic irradiation were stable. The comparison between the three methods according to the size and concentration of Ag-NPs and the effect on the bacterial activity showed that, the ultrasonic irradiation synthesis was the best method for synthesis of Ag-NPs because the high yield and with a small size of Ag-NPs which lead to a high effect on the bacterial activity.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

**SINTESIS DAN PENCIRIAN NANOPARTIKEL PERAK/KAPPA-KARAGEENAN MENGGUNAKAN KAEDAH HIJAU DAN PENILAIAN AKTIVITI ANTIBAKTERIA**

Oleh

**RANDA FAWZI ELSUPIKHE**

Mac 2017

**Pengerusi : Profesor Mansor Bin Ahmad, PhD**  
**Fakulti : Sains**

Bahan nano telah mendapat perhatian meluas kerana sifat yang luar biasa mereka yang berbeza dengan ketara daripada sampel pukal daripada bahan yang sama. Nanopartikel adalah zarah dengan saiz kurang daripada 100 nm, iaitu diameter yang kecil tetapi kawasan permukaan yang lebih besar. Nanopartikel perak (Ag-NPs) semakin banyak digunakan dalam produk pengguna seperti ejen pembersihan air, ejen pembersihan rumah, dan besar dalam aplikasi perubatan eksklusif seperti kejuruteraan biologi. Sintesis Ag-NPs telah menarik perhatian para saintis dalam tahun-tahun kebelakangan ini disebabkan oleh kelebihan yang besar dan aplikasi Ag-NPs terutamanya sebagai ejen anti-mikrob. Kaedah kimia telah digunakan untuk sintesis Ag-NPs, tetapi kaedah ini mempunyai banyak kelemahan kerana kebanyakan bahan kimia yang digunakan untuk sintesis nanopartikel terlalu mahal dan toksik, yang bertanggungjawab untuk pelbagai risiko biologi. Juga sebahagian besar daripada kaedah kimia untuk sintesis Ag-NPs tidak dapat mengawal saiz NPs. Tambahan pula, pengumpulan zarah nano membawa kepada keputusan yang buruk dalam kegunaan sebagai anti-bakteria. Dalam kajian ini, kaedah hijau untuk sintesis Ag-NPs telah digunakan bagi menyelesaikan masalah ini dan polimer  $\kappa$ -karageenan telah digunakan sebagai penstabil untuk mengelakkan pengumpulan. Ag-NPs dalam  $\kappa$ -karageenan disintesis oleh kaedah hijau yang berbeza (kaedah kacau, penyinaran-UV dan penyinaran-ultrasonik) pada suhu bilik dibangunkan untuk menyediakan dan mengawal saiz nanopartikel Ag-NPs. Parameter seperti waktu kacau, masa penyinaran, amplitud ultrasonik, kepekatan AgNO<sub>3</sub> dan kepekatan  $\kappa$ -karageenan telah dioptimumkan.  $\kappa$ -karageenan digunakan sebagai penstabil mesra alam dan AgNO<sub>3</sub> sebagai pengeluar. Bentuk Ag/ $\kappa$ -karrageenan ditentukan oleh spektra UV-boleh dilihat yang dipertingkatkan pembentukan Ag-NPs oleh resonans plasmon permukaan dalam julat 300-450 nm. Spektra FT-IR menunjukkan kehadiran  $\kappa$ -karageenan dalam liputan dengan Ag-NPs. Analisis XRD menunjukkan Ag-NPs adalah berstruktur kiub berpusatkan muka. Imej TEM menggambarkan juga Ag-NPs tersebar dengan saiz zarah yang sekata. Imej SEM

memaparkan perubahan pada morfologi permukaan  $\kappa$ -karageenan dan menggambarkan bentuk Ag-NPs. Spektra EDXRF Ag-NPs dalam  $\kappa$ -karageenan mengesahkan kehadiran sebatian unsur tanpa puncak junub. Sifat antibakteria nanopartikel yang disintesis dinilai menggunakan kaedah penyebaran agar. Empat spesies bakteria telah digunakan dalam kajian ini, termasuk dua bakteria Gram-positif dan dua bakteria Gram-negatif: *Staphylococcus aureus* tahan-Methicillin (MRSA), *subtilis Bacillus*, *Pseudomonas aeruginosa* dan *Escherichia coli* (*E-coli*). Parameter optimum bagi kaedah kacau untuk sintesis Ag-NPs adalah: 48 j masa kacau, 0.2 M AgNO<sub>3</sub> dan 0.3%  $\kappa$ -carrageenan, yang menghasilkan saiz dan kepekatan Ag-NPs masing-masing 32 nm dan 0.065 M. Keadaan terbaik bagi kaedah penyinaran-UV untuk sintesis Ag-NPs adalah 60 min masa penyinaran, 0.2 M AgNO<sub>3</sub> dan 0.3%  $\kappa$ -carrageenan, yang menghasilkan saiz dan kepekatan Ag-NPs masing-masing 14 nm dan 0.12 M. Keadaan bagi kaedah penyinaran-ultrasonik untuk sintesis Ag-NPs yang memberikan hasil yang terbaik adalah 90 min masa penyinaran, 0.15 M AgNO<sub>3</sub>, 0.3%  $\kappa$ -carrageenan dan 60 amplitud yang menghasilkan saiz dan kepekatan Ag-NPs masing-masing 1.21 nm dan 0.22 M. Semua Ag-NPs dari kaedah di atas adalah dalam bentuk sfera. Kaedah yang berbeza menunjukkan keputusan yang berbeza pada aktiviti antibakteria bergantung kepada saiz dan kepekatan Ag-NPs. Ujian kestabilan dengan menggunakan analisis potensi zeta membuktikan Ag-NPs yang disintesis oleh kaedah kacau, UV-sinaran dan penyinaran-ultrasonik adalah stabil. Perbandingan antara ketiga-tiga kaedah mengikut saiz dan kepekatan Ag-NPs dan kesan ke atas aktiviti bakteria menunjukkan, sintesis penyinaran-ultrasonik adalah kaedah terbaik untuk sintesis Ag-NPS kerana hasil yang tinggi dengan saiz Ag-NPs kecil yang membawa kepada kesan yang tinggi kepada aktiviti bakteria



## ACKNOWLEDGEMENTS

First of all, Praise and gratitude be to ALLAH, almighty which it would have been impossible to accomplish this work without his gracious.

I owe thanks and gratitude to Libyan Higher Education for sponsorship and support. I would like also to thank all Libyan staff in the embassy for cooperation, encouragement and support during my study in Malaysia. I owe a great debt to my supervisor Prof. Dr. Mansor Bin Ahmad, for unlimited advice; suggestions and encouragement throughout my study. As such, I want to express gratitude to members of the supervisory committee, Dr. Norhazlin Zainuddin and Dr. Nor Azowa Ibrahim for their guidance and constant support through the research. I admire their devotion to science. My sincere thanks to Dr. Kamyar Shameli for advice and help during the first year of my study.

Appreciation is also given to my best friends for their help and encouragement which keep me going and I wish for them all the best in their life.

In gratitude, finally I want to express to all the staff and lecturer of Department of Chemistry, Faculty of Science and Universiti Putra Malaysia that gave me the opportunity to study at UPM. I will fondly remember your support, knowledge, assistance, advice, and teaching. I thank the administrators, the Dean and staffs of the Chemistry Department for the assistance provided throughout the duration of my study at UPM.

I certify that a Thesis Examination Committee has met on 21 March 2017 to conduct the final examination of Randa Fawzi El Supikhe on her thesis entitled "Synthesis and Characterization of Silver/Kappa-Carrageenan Nanoparticles using Green Methods and Evaluation of their Antibacterial Activities" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

**Irmawati binti Ramli, PhD**

Associate Professor  
Faculty of Science  
Universiti Putra Malaysia  
(Chairman)

**Azmi bin Zakaria, PhD**

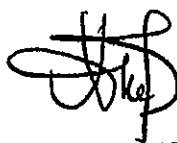
Professor  
Faculty of Science  
Universiti Putra Malaysia  
(Internal Examiner)

**Mohd Zobir bin Hussein, PhD**

Professor  
Institute of Advance Technology  
Universiti Putra Malaysia  
(Internal Examiner)

**Sabu Thomas, PhD**

Professor  
Mahatma Gandhi University  
India  
(External Examiner)



---

**NOR AINI AB. SHUKOR, PhD**

Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 6 July 2017

This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

**Mansor Bin Ahmad, PhD**

Professor  
Faculty of Science  
Universiti Putra Malaysia  
(Chairman)

**Nor Azowa Ibrahim, PhD**

Associate Professor  
Faculty of Science  
Universiti Putra Malaysia  
(Member)

**Norhazlin Zainuddin, PhD**

Senior Lecturer  
Faculty of Science  
Universiti Putra Malaysia  
(Member)

---

**ROBIAH BINTI YUNUS, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No.: Randa Fawzi Elsupikhe, GS36871

## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: \_\_\_\_\_

Name of Chairman  
of Supervisory  
Committee:

Professor Dr. Mansor Bin Ahmad  
\_\_\_\_\_

Signature: \_\_\_\_\_

Name of Member  
of Supervisory  
Committee:

Associate Professor Dr. Nor Azowa Ibrahim  
\_\_\_\_\_

Signature: \_\_\_\_\_

Name of Member  
of Supervisory  
Committee:

Dr. Norhazlin Zainuddin  
\_\_\_\_\_

## TABLE OF CONTENTS

	<b>Page</b>
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENTS</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	viii
<b>LIST OF TABLES</b>	xii
<b>LIST OF FIGURES</b>	xiv
<b>LIST OF ABBREVIATIONS</b>	xxii
<b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	<b>1</b>
1.1 Background study	1
1.2 Problem statement	3
1.3 Objectives	4
1.4 Research hypothesis	4
1.5 Research questions	4
<b>2 LITERATURE REVIEW</b>	<b>5</b>
2.1 Silver Nanoparticles	5
2.2 Synthesis of Ag-NPs	6
2.2.1 Synthesis of Ag-NPs by using chemical methods	6
2.2.2 Synthesis Ag-NPs by using green methods	6
2.3 Stabilizers	14
2.4 Carrageenan polymer	15
2.5 Characterization of Silver Nanoparticles	18
2.6 Application of Silver Nanoparticles	19
<b>3 MATERIALS AND METHODS</b>	<b>22</b>
3.1 Materials	22
3.2 Methods	22
3.2.1 Preparation of Ag-NPs in $\kappa$ -carrageenan by using stirring times at room temperature	22
3.2.2 Preparation of Ag-NPs in $\kappa$ -carrageenan by using UV- Irradiation	24
3.2.3 Preparation of AgNPs in $\kappa$ -carrageenan by using ultrasonic irradiation	26
3.3 Characterization	29
3.3.1 Ultraviolet-Visible Spectroscopy	29
3.3.2 X-Ray Diffraction	29
3.3.3 Scanning Electron Microscopy	29
3.3.4 Transmission Electron Microscopy	29
3.3.5 Energy-Dispersive X-ray Spectroscopy	30
3.3.6 Fourier Transformed-Infrared Spectroscopy	30
3.4 Antibacterial test	30

3.5	Stability test of Ag-NPs	30
<b>4</b>	<b>RESULTS AND DISCUSSION</b>	<b>31</b>
4.1	Preparation of Ag-NPs in $\kappa$ -carrageenan by using stirring time at room temperature.	31
4.1.1	Effect of reaction's time on synthesis of AgNPs.	32
4.1.2	The effect of the concentration of AgNO <sub>3</sub> on synthesis of AgNPs at stirring time 48h.	41
4.1.3	The effect of the concentrations of $\kappa$ -carrageenan on synthesis of AgNPs at stirring time 48h.	49
4.2	Preparation of Ag-NPs in $\kappa$ -carrageenan by using UV-irradiation at room temperature	58
4.2.1	The effect of UV-irradiation's time on synthesis of Ag-NPs	58
4.2.2	The effect of the concentration of AgNO <sub>3</sub> on synthesis of AgNPs for 60 min UV-irradiation	68
4.2.3	Effect of the concentration of $\kappa$ -carrageenan on synthesis of Ag-NPs for 60 min UV-irradiation	77
4.3	Preparation of Ag-NPs in $\kappa$ -carrageenan by using ultrasonic irradiation at room temperature.	86
4.3.1	Effect of ultra-sonic irradiation times on synthesis of AgNPs	86
4.3.2	Effect of the concentration of AgNO <sub>3</sub> on synthesis of Ag-NPs at ultrasonic irradiation 90 min.	94
4.3.3	Effect of the concentration of $\kappa$ -carrageenan at ultrasonic irradiation 90 min.	103
4.3.4	Effect of the amplitude of ultrasonic irradiation on synthesis of AgNPs at ultrasonic irradiation 90 min.	113
4.4	Zeta potential analysis	122
4.5	Comparison Study on the Methods that have been Used for Synthesis Ag-NPs	123
4.5.1	Concentration of Ag-NPs	123
4.5.2	Comparative study	124
<b>5</b>	<b>CONCLUSIONS AND RECOMMENDATION</b>	<b>126</b>
5.1	Conclusion	126
5.2	Recommendations for future research	127
	<b>REFERENCES</b>	<b>128</b>
	<b>BIODATA OF STUDENT</b>	<b>142</b>
	<b>LIST OF PUBLICATIONS</b>	<b>143</b>

## LIST OF TABLES

<b>Table</b>	<b>Page</b>
1.1 Typical nanostructure categories	2
2.1 Advantages of synthesis Ag-NPs without using a reducing agent	8
2.2 Advantages of synthesis Ag-NPs using physical irradiation as a reducing agent	14
4.1 Antibacterial activities effect of $\kappa$ -carrageenan, AgNO <sub>3</sub> / $\kappa$ -carrageenan (0 h), Ag-NPs at different stirring times 3, 24 and 48 h and	40
4.2 Antibacterial activities effect of $\kappa$ -carrageenan, AgNO <sub>3</sub> / $\kappa$ -carrageena, Ag-NPs at different concentration of AgNO <sub>3</sub> 0.10, 0.20 and 0.25 M for 48 h stirring time.	49
4.3 Antibacterial activities effect of $\kappa$ -carrageenan, AgNO <sub>3</sub> / $\kappa$ -carrageenan, Ag-NPs at different concentrations of $\kappa$ -carrageenan 0.10, 0.20 and 0.30 % for stirring time at 48h and $\kappa$ -carrageenan	58
4.4 Antibacterial activities effect of $\kappa$ -carrageenan, AgNO <sub>3</sub> / $\kappa$ -carrageenan (0 min), Ag-NPs at different UV-irradiation times 10, 30 and 60 min	67
4.5 Antibacterial activities effect of $\kappa$ -carrageenan, AgNO <sub>3</sub> / $\kappa$ -carrageenan, Ag-NPs at different concentration of AgNO <sub>3</sub> 0.1, 0.2 and 0.25 M for 60 minutes of UV- irradiation.	76
4.6 Antibacterial activities effect of carrageenan, AgNO <sub>3</sub> / $\kappa$ -carrageenan Ag-NPs at different concentration of $\kappa$ -carrageenan 0.1, 0.2 and 0.3 % and $\kappa$ - at 60 minutes of UV- irradiation.	85
4.7 Antibacterial activities effect of $\kappa$ -carrageenan, AgNO <sub>3</sub> / $\kappa$ -carrageenan (0 min), Ag-NPs at different times of ultrasonic irradiation at 30,50 and 90 min	94
4.8 Antibacterial activities effect of $\kappa$ -carrageenan, AgNO <sub>3</sub> / $\kappa$ -carrageenan, Ag-NPs at different concentrations of AgNO <sub>3</sub> (0.05, 0.15 and 0.25 M) for ultrasonic irradiation 90 min	103
4.9 Antibacterial activities effect of $\kappa$ -carrageenan, AgNO <sub>3</sub> / $\kappa$ -carrageenan (A0), Ag-NPs at different concentration of $\kappa$ -carrageenan at 0.1, 0.2 and 0.3 % respectively for ultrasonic irradiation 90	112



- 4.10 Antibacterial activities effect of  $\kappa$ -carrageenan,  $\text{AgNO}_3$ /  $\kappa$ -carrageenan, Ag-NPs at different ultrasonic irradiation amplitudes 30, 60 and 80 122
- 4.11 Illustrates the different between the three methods according to the size of Ag-NPs and the yield of the reaction. 125



## LIST OF FIGURES

Figure		Page
1.1	Top-down and Bottom-up Strategies	1
2.1	Scheme illustrates the methods of preparation Ag-NPs	6
2.2	Different types of carrageenan	16
2.3	Application of carrageenan	17
2.4	Applications of Ag-NPs	19
3.1	Flowchart of synthesis of Ag-NPs uses different stirring times	23
3.2	Flowchart of synthesis of Ag-NPs uses different concentrations of AgNO <sub>3</sub> at stirring time 48 h.	23
3.3	Flowchart of synthesis of Ag-NPs uses different concentrations of $\kappa$ -carrageenan at stirring time 48 h.	24
3.4	Flowchart of synthesis of Ag-NPs uses different UV- irradiation time	24
3.5	Flowchart of synthesis of Ag-NPs uses different concentrations of AgNO <sub>3</sub> at UV-irradiation 60 min.	25
3.6	Flowchart of synthesis of Ag-NPs uses different concentrations of $\kappa$ -carrageenan at UV-irradiation 60 min.	25
3.7	Flowchart of synthesis of Ag-NPs uses different ultrasonic - irradiation time	26
3.8	Flowchart of synthesis of Ag-NPs uses different concentrations of AgNO <sub>3</sub> at ultrasonic -irradiation 90 min.	27
3.9	Flowchart of synthesis of Ag-NPs uses different concentrations of $\kappa$ -carrageenan at ultrasonic -irradiation 90 min.	27
3.10	Flowchart of synthesis of Ag-NPs uses different ultrasonic amplitude at ultrasonic -irradiation 90 min.	28
3.11	Schematic illustration the synthesized AgNPs in $\kappa$ -carrageenan using different green methods	28
4.1	Scheme illustrates the formation of Ag-NPs in $\kappa$ -carrageenan by stirring time at room temperature	31

4.2	Photograph of AgNO <sub>3</sub> /κ-carrageenan (0 h) and Ag-NPs in κ-carrageenan suspensions at different stirring time (3-52 h).	32
4.3	(A) The UV-visible absorption spectra for AgNO <sub>3</sub> /κ-carrageenan (a) and Ag-NPs in κ-carrageenan suspensions b-g at different stirring times, (B) The relation between stirring time and wavelength and (C) The relation between stirring time and the absorbance	34
4.4	X-ray diffraction patterns of κ-carrageenan (a) and Ag-NPs in κ-carrageenan, (b) after stirring time 48 h	35
4.5	TEM images and corresponding size distributions of Ag-NPs in κ-carrageenan at stirring times 3 h (A), 24 h (B) and 48 h (C)	35
4.6	SEM images of κ-carrageenan and Ag-NPs in κ-carrageenan at stirring times 3 h (a), 24 h (b) and 48 h (c)	36
4.7	EDXRF spectra of κ-carrageenan and AgNPs in κ-carrageenan carrageenan at stirring times 3h (a), 24h (b) and 48h (c)	38
4.8	FT-IR spectra of κ-carrageenan (a) and Ag-NPs in κ-carrageenan at 48 h stirring time (b)	39
4.9	Antibacterial property of κ-carrageenan, AgNO <sub>3</sub> / κ-carrageenan (0 h) and Ag NPs in κ-carrageenan at different stirring times 3, 24 and 48 h against different types of bacteria	40
4.10	(A) UV-Visible absorption spectra of Ag-NPs in κ-carrageenan suspensions, at different concentrations of AgNO <sub>3</sub> 0.05, 0.1, 0.15, 0.2 and 0.25 M a-e respectively, for stirring time 48h. B) The relation between the concentration of AgNO <sub>3</sub> and the wavelength and (C) The relation between the concentration of AgNO <sub>3</sub> and the absorbance	43
4.11	X-ray diffraction patterns of Ag-NPs in κ-carrageenan at different concentrations of AgNO <sub>3</sub> (0.05, 0.10, 0.15, 0.20 and 0.25 M), (a-e) respectively for stirring time 48h	44
4.12	TEM images and corresponding size distributions of Ag-NPs in κ-carrageenan at different concentrations of AgNO <sub>3</sub> 0.1 M (A), 0.2 M (B) and 0.25 M (C) for stirring time 48 h.	45
4.13	SEM images of Ag-NPs in κ-carrageenan at different concentrations of AgNO <sub>3</sub> 0.1 M (a), 0.2 M (b) and 0.25 M (c) for stirring time 48 h.	45

4.14	EDXRF spectra of Ag in $\kappa$ -carrageenan at different concentrations of $\text{AgNO}_3$ , (0.1, 0.2, and 0.25 M, (a-c)) respectively for stirring time 48 h	46
4.15	FT-IR spectra of $\kappa$ -carrageenan (a) and Ag/ $\kappa$ -carrageenan at different concentrations of $\text{AgNO}_3$ 0.1, 0.15, 0.2, 0.25 M (b-e) for stirring time 48 h.	47
4.16	Antibacterial effect of $\kappa$ -carrageenan, $\text{AgNO}_3/\kappa$ -carrageenan and Ag- NPs in $\kappa$ -carrageenan at different concentrations of $\text{AgNO}_3$ 0.10 M, 0.20 M and 0.25 M for 48 h stirring time against different types of bacteria.	48
4.17	(A) UV-Visible absorption spectra of Ag-NPs in $\kappa$ -carrageenan synthesised, at different concentrations of $\kappa$ -carrageenan (0.1, 0.15, 0.2, 0.25 and 0.3 % (a-e) for stirring time 48 h. (B) The relation between the concentrations of $\kappa$ -carrageenan and wavelength and (C) The relation between the concentrations of $\kappa$ -carrageenan and the absorbance	51
4.18	X-ray diffraction patterns of Ag-NPs in $\kappa$ -carrageenan at different concentrations of $\kappa$ -carrageenan (0.1, 0.15, 0.2, 0.25 and 0.3 % for a, b, c, d, and e) respectively for stirring time 48 h	52
4.19	TEM images and corresponding size distributions of Ag-NPs in $\kappa$ -carrageenan synthesised at different concentrations of $\kappa$ -carrageenan 0.1 % (A), 0.2 % (B) and 0.3 % (C), for stirring time 48 h.	53
4.20	SEM images of Ag-NPs in $\kappa$ -carrageenan synthesised at different concentrations of $\kappa$ -carrageenan 0.1 % (a), 0.2 % (b) and 0.3 % (c), at stirring time 48h.	54
4.21	EDXRF spectra of Ag in $\kappa$ -carrageenan synthesised at different concentrations of $\kappa$ -carrageenan 0.1% (a), 0.2% (b) and 0.3 (c), for stirring time 48h.	55
4.22	FT-IR spectra of $\kappa$ -carrageenan (a) and Ag/ $\kappa$ -carrageenan that synthesised at different concentrations of $\kappa$ -carrageenan 0.1, 0.15, 0.2, 0.25 and 0.3% (b-f), for stirring time 48h	56
4.23	Antibacterial effect of $\kappa$ -carrageenan, $\text{AgNO}_3/\kappa$ -carrageenan and Ag-NPs in $\kappa$ -carrageenan at different concentrations of $\kappa$ -carrageenan 0.1, 0.2 and 0.3% stirring time at 48h	57
4.24	Picture of $\text{AgNO}_3/\kappa$ -carrageenan (0 min) and Ag/ $\kappa$ -carrageenan (10- 70 min) suspensions at different UV irradiation times	59

4.25	(A) UV-visible absorption spectra of Ag+/κ-carrageenan (a) and Ag- NPs in κ-carrageenan at different UV- irradiation times 10, 20, 30, 40, 50, 60 and 70 min (b–h), (B) The relation between the UV- irradiation and the wavelength and (C) The relation between the UV- irradiation and the absorbance	61
4.26	x-ray diffraction data of κ-carrageenan (a) and Ag-NPs in κ-carrageenan synthesized at 60 min UV- irradiation times (b).	62
4.27	TEM images and corresponding size distributions of Ag-NPs in κ-carrageenan at 10, 30 and 60 min of UV- irradiation respectively (a–c).	63
4.28	SEM images and corresponding size distributions of AgNPs in carrageenan after 10, 30 and 60 min of UV- irradiation respectively (a–c)	63
4.29	EDXRF spectra of Ag in κ-carrageenan at different uv- irradiation's time (10, 30 and 60 min) (a–c) respectively.	65
4.30	FT–IR spectra of κ-carrageenan (a) and Ag-NPs in κ-carrageenan synthesized at 60 minutes of UV- irradiation (b)	66
4.31	Antibacterial effect of κ-carrageenan, AgNO <sub>3</sub> / κ-carrageenan (0 min) and Ag- NPs and κ-carrageenan at different UV-irradiation times 10, 30 and 60 min against different types of Bacteria	67
4.32	(A) UV-visible absorption spectra of Ag-NPs in κ-carrageenan suspensions, at different concentration of AgNO <sub>3</sub> 0.05, 0.1, 0.15, 0.2 and 0.25 M (a–e) for 60 minutes of UV- irradiation, (B) The relation between the AgNO <sub>3</sub> concentration and the wavelength and (C) The relation between the AgNO <sub>3</sub> concentration and the absorbance.	70
4.33	X-ray diffraction patterns of Ag-NPs in κ-carrageenan at different concentrations of AgNO <sub>3</sub> (0.05, 0.10, 0.15, 0.20 and 0.25 M) (a–e) at UV-irradiation for 60 min.	71
4.34	TEM images and corresponding size distributions of Ag-NPs in κ-carrageenan at different concentrations of AgNO <sub>3</sub> (0.1 M (a), 0.2 M (b) and 0.25 M, (c) at UV-irradiation for 60 min.	72
4.35	SEM images for Ag-NPs in κ-carrageenan at different concentrations of AgNO <sub>3</sub> 0.1 M (a), 0.2 M (b) and 0.25 M (c) at UV-irradiation 60 min	72
4.36	EDXRF spectra of Ag in κ-carrageenan at different concentrations of AgNO <sub>3</sub> , 0.1 M (a), 0.2 M (b) and 0.25 M, (c) at UV-irradiation for 60 min.	74

4.37	FT-IR spectra of $\kappa$ -carrageenan (a) and Ag/ $\kappa$ -carrageenan at different concentration of AgNO <sub>3</sub> (0.05 , 0.1, 0.15, 0.2, 0.25 M (b-f) respectively under UV-irradiation for 60 min	75
4.38	Antibacterial effect of $\kappa$ -carrageenan, AgNO <sub>3</sub> / $\kappa$ -carrageenan (A <sub>0</sub> ) and Ag-NPs in $\kappa$ -carrageenan at different concentration of AgNO <sub>3</sub> (0.1, 0.2 and 0.25 M for 60 minutes of UV-irradiation against different types of bacteria	76
4.39	(A) UV-visible absorption spectra of Ag-NPs in $\kappa$ -carrageenan at different concentration $\kappa$ -carrageenan 0.1, 0.15, 0.2, 0.25 and 0.3 (a-e), for 60 minutes of UV- irradiation. B) The relation between the $\kappa$ -carrageenan concentration and the wavelength and (C) The relation between the $\kappa$ -carrageenan concentration and the absorbance.	79
4.40	X-ray diffraction patterns for the Ag-NPs in $\kappa$ -carrageenan at different concentrations of $\kappa$ -carrageenan 0.1, 0.15, 0.2, 0.25 and 0.3 at UV-irradiation for 60 min.	80
4.41	TEM images and corresponding size distributions of Ag-NPs in $\kappa$ -carrageenan at different concentrations of $\kappa$ -carrageenan 0.1 % (a), 0.2 % (b) and 0.3 % (c) respectively, for UV-irradiation 60 min.	81
4.42	SEM images of AgNPs in $\kappa$ -carrageenan at different concentrations of $\kappa$ -carrageenan, (0.1 % (a), 0.2 % (b) and 0.3 % (c) respectively, for UV-irradiation 60 min	81
4.43	EDXRF spectra of Ag in $\kappa$ -carrageenan at different concentration of $\kappa$ -carrageenan 0.1 % (a), 0.2 % (b) and 0.3 % (c) respectively, for UV-irradiation 60 min	83
4.44	FT-IR spectra of $\kappa$ -carrageenan (a) and Ag/ $\kappa$ -carrageenan at different concentrations of $\kappa$ -carrageenan (0.1,0.15,0.2,0.25 and 0.3 % (b-f) respectively under uv-irradiation for 60 min	84
4.45	Antibacterial effect of $\kappa$ -carrageenan, AgNO <sub>3</sub> / $\kappa$ -carrageenan and Ag-NPs in $\kappa$ -carrageenan at different concentration of $\kappa$ -carrageenan (0.1, 0.2 and 0.3 % respectively for 60 minutes of UV- irradiation against different types of bacteria.	85
4.46	Photograph of AgNO <sub>3</sub> / $\kappa$ -carrageenan (0 min) and Ag-NPs in $\kappa$ carrageenan (10-100 min) suspensions at different ultrasonic irradiation time.	86

4.47	(A) UV-visible absorption spectra of AgNO <sub>3</sub> /κ-carrageenan Ag-NPs in κ-carrageenan at different ultrasonic irradiation time (10, 30, 50, 70, 90 and 100 min) (b-g). (B) The relation between the ultrasonic-irradiation- and the wavelength and (C) The relation between the ultrasonic- irradiation and the absorbance	88
4.48	X-ray diffraction patterns (a) of the κ-carrageenan and (b) of Ag-NPs in κ-carrageenan at 90 min ultrasonic irradiation.	89
4.49	TEM images and corresponding size distributions of Ag-NPs in κ-carrageenan after (30, 50 and 90 min) ultrasonic irradiation times respectively (a –c).	90
4.50	SEM images of Ag-NPs in κ-carrageenan after 30, 50 and 90 min of ultrasonic irradiation respectively (a–c).	90
4.51	EDXRF spectroscopy of Ag-NPs in κ-carrageenan at different irradiation times (30, 50 and 90 minutes (a-c).	91
4.52	FT–IR spectra for (a) κ-carrageenan and (b) Ag/κ-carrageenan at ultrasonic irradiation 90 min	92
4.53	Antibacterial effect of κ-carrageenan ,Ag+/κ-carrageenan (0 min) and Ag-NPs in κ-carrageenan a different times of ultrasonic irradiation 30,50 and 90 min against different type of bacteria.	93
4.54	A) UV-visible absorption spectra of Ag-NPs in κ-carrageenan at different concentrations of AgNO <sub>3</sub> (0.05, 0.10, 0.15, 0.20 and 0.25 M) respectively (a–e) at ultrasonic irradiation 90 min. (B) The relation between the AgNO <sub>3</sub> concentration and the wavelength and (C) The relation between the AgNO <sub>3</sub> concentration and the absorbance	96
4.55	X-ray diffraction patterns of the Ag-NPs in κ-carrageenan at different concentrations of AgNO <sub>3</sub> (0.05, 0.10, 0.15, 0.20 and 0.25 M) at ultrasonic irradiation 90 min.	97
4.56	TEM images and corresponding size distributions for Ag-NPs in κ-carrageenan at different concentrations of AgNO <sub>3</sub> (0.05 M (a), 0.15 M (b) and 0.25 M (c) at ultrasonic irradiation 90 min.	98
4.57	SEM images for Ag-NPs in κ-carrageenan at different concentrations of AgNO <sub>3</sub> 0.05 M (a), 0.15 M (b) and 0.25 M (c) at ultrasonic irradiation 90 min.	99
4.58	EDXRF spectroscopy of Ag-NPs in κ-carrageenan at different concentrations of AgNO <sub>3</sub> (0.05 M (a), 0.15 M (b) and 0.25 M (c) at ultrasonic irradiation 90 min.	100

4.59	FT-IR spectra of $\kappa$ -carrageenan (a) and Ag/ $\kappa$ -carrageenan at different concentrations of AgNO <sub>3</sub> , (0.05, 0.10, 0.15, 0.20 and 0.25 M) (b-f respectively at ultrasonic irradiation 90 min.	101
4.60	Antibacterial effect of $\kappa$ -carrageenan, Ag+/ $\kappa$ -carrageenan and Ag-NPs in $\kappa$ -carrageenan at different concentrations of AgNO <sub>3</sub> (0.05 0.15 and 0.25 M) against different type of bacteria for ultrasonic irradiation 90 min.	102
4.61	(A) UV-visible absorption spectra of Ag-NPs in $\kappa$ -carrageenan at different concentration $\kappa$ -carrageenan (0.1, 0.15, 0.2, 0.25 and 0.3 % (a-e)) respectively for ultrasonic irradiation 90 min (B) The relation between the $\kappa$ -carrageenan concentration and the wavelength and The relation between the $\kappa$ -carrageenan concentration and the absorbance	105
4.62	X-ray diffraction patterns of Ag-NPs in $\kappa$ -carrageenan at different concentrations of $\kappa$ -carrageenan (0.1, 0.15, 0.2, 0.25 and 0.3 %).	106
4.63	TEM images and corresponding size distributions for Ag-NPs in $\kappa$ -carrageenan at different concentration of $\kappa$ -carrageenan 0.1 % (a), 0.2 % (b) and 0.3 % (c) at ultrasonic irradiation 90 min.	108
4.64	SEM images of Ag-NPs in $\kappa$ -carrageenan at different concentration of $\kappa$ -carrageenan 0.1 % (a), 0.2 % (b) and 0.3 % (c) at ultrasonic irradiation 90 min.	109
4.65	EDXRF spectra of Ag-NPs in $\kappa$ -carrageenan synthesised at different concentration of $\kappa$ -carrageenan 0.1 % (a), 0.2 % (b) and 0.3 % (c) at ultrasonic irradiation 90 min	110
4.66	FT-IR spectra of $\kappa$ -carrageenan (a) and Ag/ $\kappa$ -carrageenan at different concentrations of $\kappa$ -carrageenan 0.1, 0.15, 0.2, 0.25 and 0.3 % (b-f)) respectively for ultrasonic irradiation 90 min.	111
4.67	Antibacterial effect of $\kappa$ -carrageenan, Ag+/ $\kappa$ -carrageenan and Ag-NPs in $\kappa$ -carrageenan at different concentration of $\kappa$ -carrageenan at (0.1, 0.2, and 0.3 %) for ultrasonic irradiation 90 min against different type of bacteria	112
4.68	Photograph of AgNO <sub>3</sub> / $\kappa$ -carrageenan (0) and Ag-NP $\kappa$ -carrageenan suspensions for different ultrasonic irradiation amplitudes 10, 30, 60, 80 and 100, respectively at ultrasonic irradiation 90 min.	113



4.69	(A) UV-visible absorption spectra of Ag/ $\kappa$ -carrageenan at different ultrasonic amplitudes (10, 30, 60, 80 and 100) (a–e) respectively, for ultrasonic irradiation 90 min (B) The relation between the ultrasonic amplitudes and the wavelength and (C) The relation between the ultrasonic amplitudes and the absorbance	116
4.70	X-ray diffraction patterns of the Ag-NPs in $\kappa$ -carrageenan at different ultrasonic amplitudes 10, 30, 60, 80 and 100 (a–e) respectively for ultrasonic irradiation 90 min.	117
4.71	TEM images and corresponding size distributions of Ag-NPs in $\kappa$ -carrageenan at different ultrasonic amplitudes at (30 (a), 60 (b) and 80 (c) respectively, for ultrasonic irradiation 90 min.	118
4.72	SEM images of Ag-NPs in $\kappa$ -carrageenan at different ultrasonic irradiation amplitudes at (30 (a), 60 (b) and 80 (c) for ultrasonic irradiation 90 min.	118
4.73	EDXRF results of Ag-NPs in $\kappa$ -carrageenan at different ultrasonic irradiation amplitudes at (30 (a), 60 (b) and 80 (c) for ultrasonic irradiation 90 min.	119
4.74	FT–IR spectra of $\kappa$ -carrageenan (a) and Ag/ $\kappa$ -carrageenan at different ultrasonic amplitudes (10, 30, 60, 80 and 100) (b–f) respectively for ultrasonic irradiation 90 min.	120
4.75	Effect of $\kappa$ -carrageenan, Ag+/ $\kappa$ -carrageenan (0) and Ag-NPs in $\kappa$ -carrageenan at different ultrasonic irradiation amplitudes 30, 60 and 80 against different type of bacteria.	121
4.76	Zeta potential of AgNPs that synthesized using stirring time (A), UV-irradiation (B) and ultrasonic irradiation (C)	123
4.77	The standard curve between the concentration of Ag-NPs and their absorbance for measuring the concentration of Ag-NPs that were synthesised from stirring time at room temperature, u-v irradiation and ultrasonic irradiation, (a, b and c), respectively.	124

## LIST OF ABBREVIATIONS

Ag -NPs	Silver nanoparticles
fcc	face-centred cubic
SPR	Surface Plasmon Resonance
MHA	Mueller-Hinton Agar
MRSA	methicillin-resistant <i>Staphylococcus aureus</i>
NPs	Nanoparticles
<i>E. coli</i>	<i>Escherichia coli</i>
PNP	Polymer nanoparticles
AgNO <sub>3</sub>	Silver nitrate
TEM	Transmission electron microscopy
UV-Vis	UV-Visible spectroscopy
SEM	Scanning electron microscopy
FT-IR	Fourier transform infrared
PXRD	Powder X-ray diffraction
EDXRF	Energy dispersive X-ray fluorescence spectrometer

# CHAPTER 1

## INTRODUCTION

### 1.1 Background study

Publications in the field of nanotechnology have grown dramatically in the last two decades. Nanotechnology is the process of engineering of functional systems at the nanoscale level (1-100 nm ) through structural modifications of their shapes and sizes (Silva, *et al.*, 2004). The conceptual underpinnings of nanotechnologies are first laid out in 1959 by a physicist Richard Feynman in his lecture entitled “There’s plenty of room at the bottom”. Feynman explored the possibility of manipulating material at the scale of individual atoms and molecules (Sahoo, *et al.*, 2007). Nanofabrication methods are divided into two major categories. “Top-down” and “Bottom-up” methods, according to the processes involved in creating the nanoscale structures. A top-down approach corresponds to using nanofabrication tools that are controlled by external experimental parameters to create nanoscaled structures/ functional devices with the desired shape and characteristics starting from larger dimensions and reducing them to the required values. On the other hand, bottom-up approaches seek to have molecular or atomic components built up into more complex nanoscale assemblies or self directed assemblies based on complex mechanisms and technologies (Biswas *et al.*, 2012).

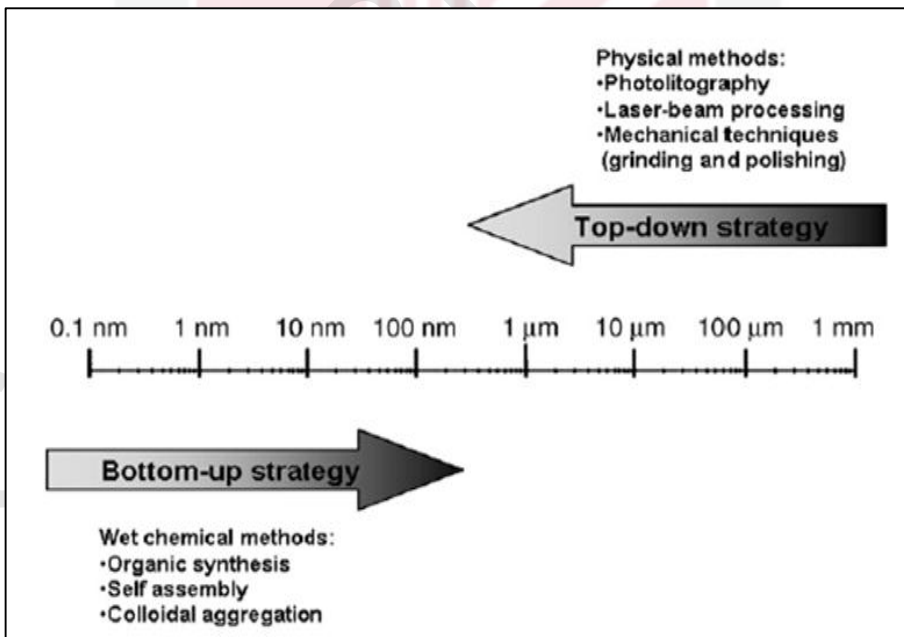


Figure 1.1 : Top-down and Bottom-up Strategies (Source: Nam and Lead, 2008)

The properties of materials with nanometer dimensions are completely different from those of atomic and bulky materials and this is mainly due to the nanometer size of the materials which cause to have unique properties, for example, a large fraction of surface atoms, high surface energy and reduced imperfections which do not exist in the corresponding bulk materials. Table 1.1 lists some typical dimensions of nanostructures (Rao and Cheetham, 2010).

A nanoparticle is the most fundamental component in the fabrication of a nanostructure, and is far smaller than the average everyday object, that is described by Newton's laws of motion, but are bigger than an atom or a simple molecule that are governed by quantum mechanics (Horikoshi *et al.*, 2013).

**Table 1.1 : Typical nanostructure categories (Rao & Cheetham, 2010).**

Structure	Size Diameter (nm)	Materials
Nanocrystals and clusters (quantum dots)	Radius. 1-10 nm	Insulators, metals, semiconductors, magnetic materials
Other Nanoparticles	Radius. 1-100 nm	Ceramic oxides
Nanowires	Radius. 1-100 nm	Metals, semiconductors, oxide, sulphides, nitrites
Nanotubes	Radius. 1-100 nm	Carbon layered chalcogenides
Nanoporous solids (pore)	Radius. 0.5-30 nm	Zeolites, phosphates etc.
2-Dimensional arrays of Nanoparticles	Area. Several nm <sup>2</sup> -µm <sup>2</sup>	Metals, semiconductors, magnetic materials
Surfaces and thin films	Thickness 1–1000 nm	Insulators, metal, DNA
3-Dimensional structures	Several nm	Semiconductors, magnetic material

Today, metal nanoparticles are important in a variety of scientific fields. Metal nanoparticles especially those containing gold (Au), silver (Ag), platinum (Pt) etc. have been of particular interest in recent years because of their unique and attractive optical and electronic properties., which are significantly different from those of bulk materials (Mohan *et al.*, 2014).

Metal nanoparticles are important due to their interesting and unusual properties such as. large optical fields, strong and well pronounced Raman scattering and light absorption effects (Jain *et al.*, 2008). Size, shap and surface morphology play vital roles in controlling the physical, chemical, optical, and electronic properties of the metal nanoparticles (Raveendran *et al.*, 2003). Metallic nanoparticles find excellent potential in biomedical sciences and engineering fields because of their huge potential in nanotechnology, hence opening a wide range of potential applications in biotechnology (Mohan *et al.*, 2010).

Silver nanoparticles (Ag NPs) have been used as antimicrobial agents, usually in the form of polymer nanocomposites, this bactericidal effect of Ag-NPs has resulted in their global application in various consumer products, e.g., deodorants, toys, humidifiers, filters and also the food and feed industry as instance, packaging materials and nursing bottles (Morones *et al.*, 2005). The broad range of targets within the bacteria makes metal nanoparticles a novel substitute for traditional antibacterial drugs, due to the significantly lower skin absorption and internal organ deposition and their relatively lower toxicity compared to silver sulfadiazine (Galdiero *et al.*, 2011).

The most common chemical approaches, including chemical reduction using a variety of inorganic and organic reducing agents, physicochemical reduction, and radiolysis are broadly used for the synthesis of silver nanoparticles. Recently, nanoparticle synthesis has been among the most interesting scientific areas of inquiry and there is growing attention to produce nanoparticles using environmentally friendly methods (green chemistry). Green synthesis approaches include polysaccharides, biological, and irradiation method which has advantages over conventional methods involving chemical agents related to environmental toxicity (Korbekandi & Irvani, 2012).

The field of polymer nanoparticles (PNP) is quickly expanding and playing a pivotal role in a wide spectrum of areas ranging from electronics to photonics, conducting materials to sensors, medicine to biotechnology, pollution control to environmental technology, and so forth, during the past decade (Rao *et al.*, 2011). Alternative synthetic strategies based on using polymers as both the reducing and stabilizing agents for the generation of stable metal nanoparticles without the use of an additional stabilizing agent have been developed recently (Sardar *et al.*, 2007).

## **1.2 Problem statement**

Nanotechnology is able to create new materials and devices with a huge range of applications, such as in electronics, medicine, biomaterials, and energy production. Furthermore, nanotechnology raises many of the same issues as any new technology, including concerns about the toxicity and the environmental impact of nanomaterials. Most of the chemical methods that have been used for the synthesis of nanoparticles are too expensive and involve the uses of toxic and hazardous chemicals which are responsible for various biological risks. Furthermore, the environment is undergoing great damage because a large amount of hazardous and unwanted chemical, gases or substances are released by man-made processes. Furthermore, most of chemical methods for the synthesis of silver nanoparticles cannot control the size and the distribution of nanoparticles. On the other hand, Ag- nanoparticles have regained importance due to increasing bacterial resistance to antibiotics. Pathogenic bacteria are becoming much resistant to antibiotics, which are produced on a continuous basis for combating infections caused by microorganisms. At present, antibiotics that are resisted by every single pathogenic organism, makes the fight much more challenging and is a problem that needs to be addressed (Raffi *et al.*, 2010).

### 1.3 Objectives

The specific objectives of this study are

- 1- To study the synthesis of Ag-NPs in  $\kappa$ -carrageenan by stirring time and characterize their physical, chemical and morphological properties.
- 2- To study the synthesis of Ag-NPs in  $\kappa$ -carrageenan by using physical methods as a reducing agent (UV- irradiation and ultrasonic irradiation) and characterize their physical, chemical and morphological properties.
- 3- To evaluate the antibacterial activity of Ag-NPs by using Mueller-Hinton Agar diffusion (MHA) test.

### 1.4 Research hypothesis

If the preparation of Ag-Nps has done by green methods, it will be safer to the environment. However, when the polymer has used as a stabilizer in the synthesis of Ag-NPs, the Ag-NPs will not agglomerate with each other. Furthe more, if the time of reactions, concentrations of the stabilizer and the concentrations of metal producer have optimized, the size of Ag-NPs will be controlled. Also, when the size of Ag-NPs decreased, it will give good results as an antibiotic.

### 1.5 Research questions

- 1- Are the green methods can synthesize Ag-NPs?
- 2- Is the polymer ( $\kappa$ -carrageenan) able to coat the Ag-NPs?
- 3- Are optimized the experimental parameters (time of reaction, concentrations of the  $\kappa$ -carrageenan and the  $\text{AgNO}_3$ ) can control the size of Ag-NPs?
- 4- Does the size of Ag-NPs influence their antibacterial properties?
- 5- Are the Ag-NPs that synthesize using green methods stable?

## REFERENCES

- Abad, L. V., Kudo, H., Saiki, S., Nagasawa, N., Tamada, M., Fu, H., DeLaRosa, M. (2010). Radiolysis studies of aqueous  $\kappa$ -carrageenan. *Nuclear Instruments and Methods in Physics Research, Section B. Beam Interactions with Materials and Atoms*, 268 (10), 1607–1612.
- Abad, L. V., Kudo, H., Saiki, S., Nagasawa, N., Tamada, M., Katsumura, Y. De La Rosa, M. (2009). Radiation degradation studies of carrageenans. *Carbohydrate Polymers*, 78 (1), 100–106.
- Abou El-Nour, K. M. M., Eftaiha, A., Al-Warthan, A., Ammar, R. (2010). Synthesis and applications of silver nanoparticles. *Arabian Journal of Chemistry*, 3 (3), 135–140.
- Ahamed, M., AlSalhi, M. S., Siddiqui, M. K. J. (2010a). Silver nanoparticle applications and human health. *Clinica Chimica Acta*, 411 (23-24), 1841–1848.
- Ahmad, M. B., Darroudi, M., Shameli, K., Abdullah, A. H., Ibrahim, N. A., Hamid, A. A., Zargar, M. (2009a). Antibacterial effect of synthesized silver/montmorillonite nanocomposites by UV-irradiation method. *Research Journal of Biological Sciences*.
- Ahmad, M. Bin, Tay, M. Y., Shameli, K., Hussein, M. Z., Lim, J. J. (2011). Green synthesis and characterization of silver/chitosan/polyethylene glycol nanocomposites without any reducing agent. *International Journal of Molecular Sciences*, 12 (8), 4872–4884.
- Ahmad, M., Shameli, K., Darroudi, M., Wan Md. Zin Wan, Y., Ibrahim, N. A., Rustaiyan, A., Abdollahi, Y. (2009b). Synthesis and Characterization of Silver / Clay / Chitosan Bionanocomposites by UV-Irradiation Method. *International Journal of Nanomedicine*, 6 (12), 2030–2035.
- Ahmad, N., Sharma, S., Alam, M. K., Singh, V. N., Shamsi, S. F., Mehta, B. R., Fatma, A. (2010b). Rapid synthesis of AgNPs using dried medicinal plant of basil. *Colloids and Surfaces B. Biointerfaces*, 81 (1), 81–86.
- Aliste, A. J., Vieira, F. F., Mastro, N. L. Del. (2000). Radiation effects on agar, alginates and carrageenan to be used as food additives. *Radiation Physics and Chemistry*, 57, 305–308.
- Akbal, A., Turkdemir, M. H., Cicek, A., Ulug, B. (2016). Relation between Silver Nanoparticle Formation Rate and Antioxidant Capacity of Aqueous Plant Leaf Extracts. *Journal of Spectroscopy*, (2016) 1-6.

- Barrena, R., Casals, E., Colón, J., Font, X., Sánchez, A., Puntès, V. (2009). Evaluation of the ecotoxicity of model nanoparticles. *Chemosphere*, 75 (7), 850-857.
- Balavandy, S. K., Shameli, K., Abidin, Z. Z. (2015). Rapid and Green Synthesis of AgNPs via Sodium Alginate Media, *Electrochem. Sci*, 10, 486–497.
- Balavandy, S. K., Shameli, K., Biak, D. R. B. A., Abidin, Z. Z. (2014). Stirring time effect of AgNPs prepared in glutathione mediated by green method. *Chemistry Central Journal*, 8 (1), 11.
- Baldevraj, R. S. M., Jagadish, R. S. (2011). Multifunctional and Nanoreinforced Polymers for Food Packaging. *Multifunctional and Nanoreinforced Polymers for Food Packaging*, 368–420.
- Bhui, D. K., Bar, H., Sarkar, P., Sahoo, G. P., De, S. P., Misra, A. (2009). Synthesis and UV–vis spectroscopic study of silver nanoparticles in aqueous SDS solution. *Journal of Molecular Liquids*, 145 (1), 33–37.
- Bindhu, M. R., Umadevi, M. (2015). Antibacterial and catalytic activities of green synthesized silver nanoparticles. *Spectrochimica Acta Part A. Molecular and Biomolecular Spectroscopy*, 135, 373–378.
- Biswas, A., Bayer, I. S., Biris, A. S., Wang, T., Dervishi, E., Faupel, F. (2012). Advances in top-down and bottom-up surface nanofabrication. Techniques, applications & future prospects. *Advances in Colloid and Interface Science*, 170 (1-2), 2–27.
- Bogle, K. A., Dhole, S. D., Bhoraskar, V. N. (2006). Silver nanoparticles. synthesis and size control by electron irradiation. *Nanotechnology*, 17 (13), 3204.
- Byeon, J. H., Kim, Y. W. (2012). A novel polyol method to synthesize colloidal silver nanoparticles by ultrasonic irradiation. *Ultrasonics sonochemistry*, 19 (1), 209-215.
- Campo, V. L., Kawano, D. F., Silva, D., Carvalho, I. (2009). Carrageenans. Biological properties, chemical modifications and structural analysis - A review. *Carbohydrate Polymers*, 77 (2), 167–180.
- Clermont, O., Bonacorsi, S., Bingen, E. (2000). Rapid and simple determination of the *Escherichia coli* phylogenetic group. *Applied and environmental microbiology*, 66 (10), 4555-4558.
- Chen, D.-H., Hsieh, C.-H. (2002). Synthesis of nickel nanoparticles in aqueous cationic surfactant solutions. *Journal of Materials Chemistry*. 12(8), 2412-2415.
- Chen, J., Zheng, X., Dong, J., Chen, Y., Tian, J. (2015). Optimization of effective high hydrostatic pressure treatment of *Bacillus subtilis* in Hami melon juice. *LWT - Food Science and Technology*, 60 (2), 1168–1173.



- Cheviron, P., Gouanvé, F., Espuche, E. (2014). Green synthesis of colloid silver nanoparticles and resulting biodegradable starch/silver nanocomposites. *Carbohydrate Polymers*, 108, 291–298.
- Dadgostar, N., Ferdous, S., Henneke, D. (2010). Colloidal synthesis of copper nanoparticles in a two-phase liquid-liquid system. *Materials Letters*, 64 (1), 45–48.
- Daniel-Da-Silva, A. L., Lopes, A. B., Gil, A. M., Correia, R. N. (2007). Synthesis and characterization of porous  $\kappa$ -carrageenan/calcium phosphate nanocomposite scaffolds. *Journal of Materials Science*, 42 (20), 8581–8591.
- Darroudi, M., Ahmad, M. B., Zak, A. K., Zamiri, R., Hakimi, M. (2011). Fabrication and characterization of gelatin stabilized AgNPs under UV-Light. *International Journal of Molecular Sciences*, 12 (9), 6346–6356.
- Darroudi, M., Ahmad, M. Bin, Abdullah, A. H., Ibrahim, N. A., Shameli, K. (2010). Effect of accelerator in green synthesis of silver nanoparticles. *International Journal of Molecular Sciences*, 11 (10), 3898–3905.
- Darroudi, M., Ahmad, M. Bin, Zamiri, R., Zak, a. K., Abdullah, A. H., Ibrahim, N. A. (2011a). Time-dependent effect in green synthesis of silver nanoparticles. *International Journal of Nanomedicine*, 6 (1), 677–681.
- Darroudi, M., Khorsand Zak, A., Muhamad, M. R., Huang, N. M., Hakimi, M. (2012). Green synthesis of colloidal AgNPs by sonochemical method. *Materials Letters*, 66 (1), 117–120.
- Das, M. R., Sarma, R. K., Saikia, R., Kale, V. S., Shelke, M. V., Sengupta, P. (2011). Synthesis of silver nanoparticles in an aqueous suspension of graphene oxide sheets and its antimicrobial activity. *Colloids and Surfaces B. Biointerfaces*, 83 (1), 16–22.
- Datta, S., Mody, K., Gopalsamy, G., Jha, B. (2011). Novel application of  $\kappa$ -carrageenan. As a gelling agent in microbiological media to study biodiversity of extreme alkaliphiles. *Carbohydrate Polymers*, 85 (2), 465–468.
- Dubey, S. P., Lahtinen, M., Sillanpää, M. (2010). Tansy fruit mediated greener synthesis of silver and gold nanoparticles. *Process Biochemistry*. 45(7), 1065–1071.
- Elechiguerra, J. L., Burt, J. L., Morones, J. R., Camacho-Bragado, A., Gao, X., Lara, H. H., Yacaman, M. J. (2005). Interaction of silver nanoparticles with HIV-1. *Journal of Nanobiotechnology*, 3 (1), 1–10.
- Enright, M. C., Enright, M. C., Robinson, D. A., Robinson, D. A., Randle, G., Randle, G Spratt, B. G. (2002). The evolutionary history of methicillin-resistant *Staphylococcus aureus* (MRSA). *Proceedings of the National Academy of Sciences of the United States of America*, 99 (11), 7687–92.

- Fabrega, J., Luoma, S. N., Tyler, C. R., Galloway, T. S., Lead, J. R. (2011). Silver nanoparticles. *Behaviour and effects in the aquatic environment. Environment International*, 37 (2), 517–531.
- Fernandez Rivas, D., Stricker, L., Zijlstra, A. G., Gardeniers, H. J. G. E., Lohse, D., Prosperetti, A. (2013). Ultrasound artificially nucleated bubbles and their sonochemical radical production. *Ultrasonics Sonochemistry*, 20 (1), 510–524.
- Gashti, M. P., Almasian, A. (2012). Synthesizing tertiary silver/silica/kaolinite nanocomposite using photo-reduction method. Characterization of morphology and electromagnetic properties. *Composites Part B. Engineering*, 43 (8), 3374–3383.
- Ge, L., Li, Q., Wang, M., Ouyang, J., Li, X., Xing, M. M. Q. (2014). Nanosilver particles in medical applications. Synthesis, performance, and toxicity. *International Journal of Nanomedicine*, 9 (1), 2399–2407.
- Gedanken, A. (2003). Sonochemistry and its application to nanochemistry. *Current Science*, 85 (12), 1720–1722.
- Ghorbani, H. R., Safekordi, A. A., Attar, H., Sorkhabadi, S. M. (2011). Biological and non-biological methods for silver nanoparticles synthesis. *Chemical and Biochemical Engineering Quarterly*, 25 (3), 317-326.
- Gómez-Acosta, a., Manzano-Ramírez, a., López-Naranjo, E. J., Apatiga, L. M., Herrera-Basurto, R., Rivera-Muñoz, E. M. (2015). Silver nanostructure dependence on the stirring-time in a high-yield polyol synthesis using a short-chain PVP. *Materials Letters*, 138 (2000), 167–170.
- Galdiero, S., Falanga, A., Vitiello, M., Cantisani, M., Marra, V., Galdiero, M. (2011). Silver nanoparticles as potential antiviral agents. *Molecules*, 16(10), 8894-8918.
- Guzmán, M. G., Dille, J., Godet, S. (2009). Synthesis of silver nanoparticles by chemical reduction method and their antibacterial activity. *Int J Chem Biomol Eng*, 2 (3), 104-111.
- Guzman, M., Dille, J., Godet, S. (2012). Synthesis and antibacterial activity of AgNPs against gram-positive and gram-negative bacteria. *Nanomedicine. Nanotechnology, Biology, and Medicine*, 8 (1), 37–45.
- Hassan Korbekandi and Siavash Irvani. (2010). Silver Nanoparticles. In Abbass A. Hashim (Ed.), *The Delivery of Nanoparticles*. Iran. <http://doi.org/10.5772/186>
- He, B., Tan, J. J., Liew, K. Y., Liu, H. (2004). Synthesis of size controlled Ag nanoparticles. *Journal of Molecular Catalysis A. Chemical*, 221 (1-2), 121–126.

- He, C., Liu, L., Fang, Z., Li, J., Guo, J., Wei, J. (2014). Formation and characterization of silver nanoparticles in aqueous solution via ultrasonic irradiation. *Ultrasonics Sonochemistry*, 21 (2), 542–548.
- Hebeish, A., Hashem, M., El-Hady, M. M. A., Sharaf, S. (2013). Development of CMC hydrogels loaded with silver nano-particles for medical applications. *Carbohydrate Polymers*, 92 (1), 407–13.
- Horikoshi, S., Serpone, N. (Eds.). (2013). *Microwaves in nanoparticle synthesis. fundamentals and applications*. John Wiley & Sons.
- Huang, H. H., Ni, X. P., Loy, G. L., Chew, C. H., Tan, K. L., Loh, F. C., Xu, G. Q. (1996). Photochemical Formation of Silver Nanoparticles in Poly (N –vinyl pyrrolidone). *Langmuir*, 12 (12), 909–912.
- Huang, H., Yang, X. (2004). Synthesis of polysaccharide-stabilized gold and silver nanoparticles. A green method. *Carbohydrate Research*, 339(15), 2627–2631.
- Huang, X., El-Sayed, M. a. (2010). Gold nanoparticles. Optical properties and implementations in cancer diagnosis and photothermal therapy. *Journal of Advanced Research*, 1 (1), 13–28.
- Iravani, S. (2011). Green synthesis of metal nanoparticles using plants. *Green Chemistry*, 13 (10), 2638.
- Jain, P. K., Huang, X., El-Sayed, I. H., El-Sayed, M. a. (2008). Noble metals on the nanoscale. Optical and photothermal properties and some applications in imaging, sensing, biology, and medicine. *Accounts of Chemical Research*, 41 (12), 1578–1586.
- Jacobs, C., Müller, R. H. (2002). Production and characterization of a budesonide nanosuspension for pulmonary administration. *Pharmaceutical research*, 19 (2), 189-194.
- Joseph, S., Mathew, B. (2014). Synthesis of AgNPs by Microwave irradiation and investigation of their Catalytic activity, 3, 185–191.
- Jurasekova, Z., Sanchez-Cortes, S., Tamba, M., Torreggiani, a. (2011). AgNPs active as surface-enhanced Raman scattering substrates prepared by high energy irradiation. *Vibrational Spectroscopy*, 57 (1), 42–48.
- Jyoti, K., Baunthiyal, M., Singh, A. (2015). Characterization of silver nanoparticles synthesized using *Urtica dioica* Linn. leaves and their synergistic effects with antibiotics. *Journal of Radiation Research and Applied Sciences*. 9 ( 2 0 1 6 ) 217-227.

- Kamat, P. V., Flumiani, M., Hartland, G. V. (1998). Picosecond Dynamics of Silver Nanoclusters. Photoejection of Electrons and Fragmentation. *The Journal of Physical Chemistry B*, 102 (17), 3123–3128.
- Kannan, R. R. R., Stirk, W. A., Van Staden, J. (2013). Synthesis of AgNPs using the seaweed *Codium capitatum* P.C. Silva (Chlorophyceae). *South African Journal of Botany*, 86, 1–4.
- Kasaai, M. R. (2013). Input power-mechanism relationship for ultrasonic Irradiation. Food and polymer applications. *Natural Science*, 05 (08), 14–22.
- Kelly, K. L., Kelly, K. L., Coronado, E., Zhao, L., Coronado, E., Schatz, G. C., Schatz, G. C. (2003). The Optical Properties of Metal Nanoparticles. The Influence of Size, Shape, and Dielectric Environment. *Journal of Physical Chemistry B*, 107 (3), 668–677.
- Khan, Z., Al-Thabaiti, S. A., Obaid, A. Y., Al-Youbi, a. O. (2011). Preparation and characterization of AgNPs by chemical reduction method. *Colloids and Surfaces B. Biointerfaces*, 82 (2), 513–517.
- Khanna, P. K., Singh, N., Charan, S., Subbarao, V. V. V. S., Gokhale, R., Mulik, U. P. (2005). Synthesis and characterization of Ag/PVA nanocomposite by chemical reduction method. *Materials Chemistry and Physics*, 93 (1), 117–121.
- Kharissova, O. V., Dias, H. V. R., Kharisov, B. I., Pérez, B. O., Pérez, V. M. J. (2013). The greener synthesis of nanoparticles. *Trends in Biotechnology*, 31 (4), 240–248.
- Korbekandi, H., Iravani, S. (2012). Silver nanoparticles. In *The delivery of nanoparticles*. InTech.
- Kouvaris, P., Delimitis, A., Zaspalis, V., Papadopoulos, D., Tsipas, S. a., Michailidis, N. (2012). Green synthesis and characterization of silver nanoparticles produced using *Arbutus Unedo* leaf extract. *Materials Letters*, 76, 18–20.
- Kreyling, W. G., Semmler-Behnke, M., Chaudhry, Q. (2010). A complementary definition of nanomaterial. *Nano Today*, 5 (3), 165–168.
- Krstić, J., Spasojević, J., Radosavljević, A., Šiljegović, M., Kačarević-Popović, Z. (2014). Optical and structural properties of radiolytically in situ synthesized AgNP sstabilized by chitosan/poly (vinyl alcohol) blends. *Radiation Physics and Chemistry*, 96, 158–166.
- Kumar, B., Smita, K., Cumbal, L., Debut, A., Pathak, R. N. (2014). Sonochemical synthesis of silver nanoparticles using starch. a comparison. *Bioinorganic chemistry and applications*, 2014.

- Lee, W., Kim, K.-J., Lee, D. G. (2014). A novel mechanism for the antibacterial effect of AgNPs on Escherichia coli. *Biometals. An International Journal on the Role of Metal Ions in Biology, Biochemistry, and Medicine*, 1191–1201.
- Li, L., Ni, R., Shao, Y., Mao, S. (2014). Carrageenan and its applications in drug delivery. *Carbohydrate Polymers*, 103 (1), 1–11.
- Li, W. R., Xie, X. B., Shi, Q. S., Zeng, H. Y., Ou-Yang, Y. S., Chen, Y. Ben. (2010). Antibacterial activity and mechanism of AgNPson Escherichia coli. *Applied Microbiology and Biotechnology*, 85 (4), 1115–1122.
- Li, X., Liu, B., Ye, W., Wang, X., Sun, R. (2015). Effect of rectorite on the synthesis of Ag NP and its catalytic activity. *Materials Chemistry and Physics*, 151, 301–307.
- Liu, J., Li, X., Zeng, X. (2010). AgNPs prepared by chemical reduction-protection method, and their application in electrically conductive silver nanopaste. *Journal of Alloys and Compounds*, 494 (1-2), 84–87.
- Liu, F. K., Hsu, Y. C., Tsai, M. H., & Chu, T. C. (2007). Using  $\gamma$ -irradiation to synthesize Ag nanoparticles. *Materials Letters*, 61(11), 2402-2405.
- Liu, L., Yang, Y., Liu, P., Tan, W. (2014). The influence of air content in water on ultrasonic cavitation field. *Ultrasonics Sonochemistry*, 21 (2), 566–571.
- Liu, Z., Wang, H., Li, H., & Wang, X. (1998). Red shift of plasmon resonance frequency due to the interacting Ag nanoparticles embedded in single crystal SiO<sub>2</sub> by implantation. *Applied physics letters*, 72 (15), 1823-1825.
- Livermore, D. M. (2002). Multiple mechanisms of antimicrobial resistance in Pseudomonas aeruginosa. our worst nightmare? *Clinical Infectious Diseases. An Official Publication of the Infectious Diseases Society of America*, 34 (5), 634–640.
- Longano, D., Ditaranto, N., Sabbatini, L., Torsi, L., Cioffi, N. (2012). Synthesis and antimicrobial activity of copper nanomaterials *Nano-Antimicrobials* (pp. 85-117). Springer.
- Lu, W., Senapati, D., Wang, S., Tovmachenko, O., Singh, A. K., Yu, H., Ray, P. C. (2010). Effect of surface coating on the toxicity of silver nanomaterials on human skin keratinocytes. *Chemical Physics Letters*, 487 (1-3), 92–96.
- Mankad, V., Kumar, R. K., Jha, P. K. (2013). Investigation of Blue-Shifted Plasmon Resonance. An Optical Properties Study of Silver Nanoparticles. *Nanoscience and Nanotechnology Letters*, 5 (8), 889-894.
- Marimuthu, S., Rahuman, A. A., Rajakumar, G., Santhoshkumar, T., Kirthi, A. V., Jayaseelan, C., Kamaraj, C. (2011). Evaluation of green synthesized silver nanoparticles against parasites. *Parasitology Research*, 108 (6), 1541-1549.

- Martins, J. T., Cerqueira, M. a., Bourbon, A. I., Pinheiro, A. C., Souza, B. W. S., Vicente, A. a. (2012). Synergistic effects between  $\kappa$ -carrageenan and locust bean gum on physicochemical properties of edible films made thereof. *Food Hydrocolloids*, 29 (2), 280–289.
- Misra, N., Biswal, J., Gupta, A., Sainis, J. K., Sabharwal, S. (2012). Gamma radiation induced synthesis of gold nanoparticles in aqueous polyvinyl pyrrolidone solution and its application for hydrogen peroxide estimation. *Radiation Physics and Chemistry*, 81 (2), 195–200.
- Mohan, S., Oluwafemi, O. S., George, S. C., Jayachandran, V. P., Lewu, F. B., Songca, S. P., Thomas, S. (2014). Completely green synthesis of dextrose reduced silver nanoparticles, its antimicrobial and sensing properties. *Carbohydrate Polymers*, 106, 469–474.
- Morones, J. R., Elechiguerra, J. L., Camacho, A., Holt, K., Kouri, J. B., Ramírez, J. T., Yacaman, M. J. (2005). The bactericidal effect of silver nanoparticles. *Nanotechnology*, 16 (10), 2346–2353.
- Mukherjee, P., Ahmad, A., Mandal, D., Senapati, S., Sainkar, S. R., Khan, M. I., Sastry, M. (2001). Fungus-Mediated Synthesis of Silver Nanoparticles and Their Immobilization in the Mycelial Matrix. A Novel Biological Approach to Nanoparticle Synthesis. *Nano Letters*, 1 (10), 515–519.
- Musa, A., Ahmad, M. B., Hussein, M. Z., Mohd Izham, S., Shameli, K., Abubakar Sani, H. (2016). Synthesis of nanocrystalline cellulose stabilized copper nanoparticles. *Journal of Nanomaterials*, 2016, 8.
- Nagata, Y., Watanabe, Y., Fujita, S., Dohmaru, T., Taniguchi, S. (1992). Formation of colloidal silver in water by ultrasonic irradiation. *Journal of the Chemical Society, Chemical Communications*, (21), 1620.
- Nam, Y.J., Lead, J.R. (2008). Manufactured nanoparticles: An overview of their chemistry, interactions and potential environmental implications. *Science of the Total Environment*, (400) 396-414.
- Naraginti, S., Sivakumar, a. (2014). Eco-friendly synthesis of silver and gold nanoparticles with enhanced bactericidal activity and study of silver catalyzed reduction of 4-nitrophenol. *Spectrochimica Acta - Part A. Molecular and Biomolecular Spectroscopy*, 128, 357–362.
- Narayanan, K. B., Sakthivel, N. (2010). Biological synthesis of metal nanoparticles by microbes. *Advances in Colloid and Interface Science*, 156 (1-2), 1–13.
- Omrani, a. A., Taghavinia, N. (2012). Photo-induced growth of AgNPs using UV sensitivity of cellulose fibers. *Applied Surface Science*, 258 (7), 2373–2377.

- Prasad, V., D'Souza, C., Yadav, D., Shaikh, A. J., Vigneshwaran, N. (2006). Spectroscopic characterization of zinc oxide nanorods synthesized by solid-state reaction. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 65(1), 173-178.
- Pastoriza-Santos, I., Liz-Marzán, L. (1999). Formation and stabilization of silver nanoparticles through reduction by N, N-dimethylformamide. *Langmuir*, 15 (4), 948–951.
- Pavia, D., Lampman, G., Kriz, G., Vyvyan, J. (2008). *Introduction to spectroscopy*. Cengage Learning.
- Pereira, L., Amado, A. M., Critchley, A. T., van de Velde, F., Ribeiro-Claro, P. J. a. (2009). Identification of selected seaweed polysaccharides (phycocolloids) by vibrational spectroscopy (FTIR-ATR and FT-Raman). *Food Hydrocolloids*, 23 (7), 1903–1909.
- Pereira, L., Sousa, A., Coelho, H., Amado, A. M., Ribeiro-Claro, P. J. a. (2003). Use of FTIR, FT-Raman and <sup>13</sup>C-NMR spectroscopy for identification of some seaweed phycocolloids. *Biomolecular Engineering*, 20 (4-6), 223–228.
- Pokropivny, V. V., Skorokhod, V. V. (2007). Classification of nanostructures by dimensionality and concept of surface forms engineering in nanomaterial science. *Materials Science and Engineering. C*, 27 (5), 990-993.
- Prajapati, V. D., Maheriya, P. M., Jani, G. K., Solanki, H. K. (2014). Carrageenan. A natural seaweed polysaccharide and its applications. *Carbohydrate Polymers*, 105 (1), 97–112.
- Radzig, M. A., Nadtochenko, V. A., Koksharova, O. A., Kiwi, J., Lipasova, V. A., Khmel, I. A. (2013). Antibacterial effects of silver nanoparticles on gram-negative bacteria. influence on the growth and biofilms formation, mechanisms of action. *Colloids and Surfaces B. Biointerfaces*, 102, 300-306.
- Raffi, M., Mehrwan, S., Bhatti, T. M., Akhter, J. I., Hameed, A., Yawar, W., ul Hasan, M. M. (2010). Investigations into the antibacterial behavior of copper nanoparticles against Escherichia coli. *Annals of microbiology*, 60 (1). 75-80.
- Rai, M., Yadav, A., Gade, A. (2009). AgNPs as a new generation of antimicrobials. *Biotechnology Advances*, 27 (1), 76–83.
- Rao, C. N. R., Cheetham, a K. (2010). Science and technology of nanomaterials. current status and future prospects. *Society*, 11 (12), 2887–2894.
- Rao, J. P., Geckeler, K. E. (2011). Polymer nanoparticles. Preparation techniques and size-control parameters. *Progress in Polymer Science (Oxford)*, 36 (7), 887–913.

- Raveendran, P., Fu, J., Wallen, S. L. (2003). Completely “Green” Synthesis and Stabilization of Metal Nanoparticles. *Journal of the American Chemical Society*,
- Relleve, L., Nagasawa, N., Luan, L. Q., Yagi, T., Aranilla, C., Abad, L., Dela Rosa, a. (2005). Degradation of carrageenan by radiation. *Polymer Degradation and Stability*, 87 (3), 403–410.
- Remita, S., Fontaine, P., Lacaze, E., Borensztein, Y., Sellame, H., Farha, R., Goldmann, M. (2007). X-ray radiolysis induced formation of silver nanoparticles. A SAXS and UV-visible absorption spectroscopy study. *Nuclear Instruments and Methods in Physics Research, Section B. Beam Interactions with Materials and Atoms*, 263 (2), 436–440.
- Sadeghi, M., Soleimani, F. (2011). Synthesis of Novel Polysaccharide-Based Superabsorbent Hydro Gels Via Graft Copolymerization of Vinyllic Monomers onto Kappa-Carrageenan. *International Journal of Chemical Engineering and Applications*, 2 (5), 304–306.
- Saeb, A. T. M., Alshammari, A. S., Al-Brahim, H., Al-Rubeaan, K. a. (2014). Production of Silver Nanoparticles with Strong and S Antimicrobial Activity against Highly Pathogenic and Multidrug Resistant Bacteria. *The Scientific World Journal*, 2014, 704708.
- Sahoo, S. K., Parveen, S., Panda, J. J. (2007). The present and future of nanotechnology in human health care. *Nanomedicine. Nanotechnology, Biology, and Medicine*, 3 (1), 20–31.
- Saifuddin, N., Wong, C. W., Yasumira, a. a. N. (2009). Rapid Biosynthesis of AgNPs Using Culture Supernatant of Bacteria with Microwave Irradiation. *E-Journal of Chemistry*, 6 (1), 61–70.
- Sakamoto, M., Fujistuka, M., Majima, T. (2009). Light as a construction tool of metal nanoparticles. Synthesis and mechanism. *Journal of Photochemistry and Photobiology C. Photochemistry Reviews*, 10 (1), 33–56.
- Salkar, R. a., Jeevanandam, P., Aruna, S. T., Kolytyn, Y., Gedanken, a. (1999). The sonochemical preparation of amorphous silver nanoparticles. *Journal of Materials Chemistry*, 9 (6), 1333–1335.
- Sardar, R., Park, J.-W., Shumaker-Parry, J. S. (2007). Polymer-induced synthesis of gold and silver nanoparticles and subsequent ligand exchange in water. *Langmuir. The ACS Journal of Surfaces and Colloids*, 23(23), 11883–11889.
- Schodek, D. L., Ferreira, P., Ashby, M. F. (2009). *Nanomaterials, nanotechnologies and design. an introduction for engineers and architects*. Butterworth-Heinemann.



- Seney, C. S., Gutzman, B. M., Goddard, R. H. (2009). Correlation of Size and Surface-Enhanced Raman Scattering Activity of Optical and Spectroscopic Properties for Silver Nanoparticles. *The Journal of Physical Chemistry C*, 113 (1), 74–80.
- Shahverdi, A. R., Fakhimi, A., Shahverdi, H. R., Minaian, S. (2007). Synthesis and effect of AgNPs on the antibacterial activity of different antibiotics against *Staphylococcus aureus* and *Escherichia coli*. *Nanomedicine. Nanotechnology, Biology, and Medicine*, 3 (2), 168–171.
- Shameli, K., Ahmad, M. Bin, Al-Mulla, E. J., Shabanzadeh, P., Bagheri, S. (2013). Antibacterial effect of silver nanoparticles on talc composites. *Research on Chemical Intermediates*, 1–13.
- Shameli, K., Ahmad, M. Bin, Jazayeri, S. D., Sedaghat, S., Shabanzadeh, P., Jahangirian, H., Abdollahi, Y. (2012a). Synthesis and characterization of polyethylene glycol mediated AgNPs by the green method. *International Journal of Molecular Sciences*, 13 (6), 6639–6650.
- Shameli, K., Ahmad, M. Bin, Shabanzadeh, P., Al-Mulla, E. J., Zamanian, A., Abdollahi, Y., Haroun, R. Z. (2014). Effect of *Curcuma longa* tuber powder extract on size of silver nanoparticles prepared by green method. *Research on Chemical Intermediates*, 40 (3), 1313–1325.
- Shameli, K., Ahmad, M. Bin, Yunus, W. M. Z. W., Ibrahim, N. A., Gharayebi, Y., Sedaghat, S. (2010). Synthesis of silver/montmorillonite nanocomposites using  $\gamma$ -irradiation. *International Journal of Nanomedicine*, 5 (1), 1067–1077.
- Shameli, K., Ahmad, M. Bin, Yunus, W. M. Z. W., Rustaiyan, A., Ibrahim, N. A., Zargar, M., Abdollahi, Y. (2010). Green synthesis of silver/montmorillonite/chitosan bionanocomposites using the UV irradiation method and evaluation of antibacterial activity. *International Journal of Nanomedicine*, 5 (1), 875–887.
- Shameli, K., Ahmad, M. Bin, Zamanian, A., Sangpour, P., Shabanzadeh, P., Abdollahi, Y., Zargar, M. (2012b). Green biosynthesis of silver nanoparticles using *Curcuma longa* tuber powder. *International Journal of Nanomedicine*, 7, 5603–5610.
- Shameli, K., Ahmad, M. Bin, Zargar, M., Yunus, W. M. Z. W., Ibrahim, N. A. (2011). Fabrication of silver nanoparticles doped in the zeolite framework and antibacterial activity. *International Journal of Nanomedicine*, 6, 331–341.
- Shameli, K., Bin Ahmad, M., Jazayeri, S. D., Shabanzadeh, P., Sangpour, P., Jahangirian, H., Gharayebi, Y. (2012c). Investigation of antibacterial properties silver nanoparticles prepared via green method. *Chemistry Central Journal*.

- Shameli, K., Bin Ahmad, M., Zargar, M., Yunus, W. M. Z. W., Ibrahim, N. A., Shabanzadeh, P., Moghaddam, M. G. (2011a). Synthesis and characterization of silver/montmorillonite/chitosan bionanocomposites by chemical reduction method and their antibacterial activity. *International Journal of Nanomedicine*, 6, 271–284.
- Shameli, K., Mansor Bin Ahmad, M., Mohsen, Z., Yunus, W. Z., Ibrahim, N. A., Rustaiyan, A. (2011b). Synthesis of silver nanoparticles in montmorillonite and their antibacterial behavior. *International Journal of Nanomedicine*, 581.
- Sharma, V. K., Yngard, R. a., Lin, Y. (2009). Silver nanoparticles. Green synthesis and their antimicrobial activities. *Advances in Colloid and Interface Science*, 145 (1-2), 83–96.
- Silva, G. a. (2004). Introduction to nanotechnology and its applications to medicine. *Surgical Neurology*, 61 (3), 216–220.
- Sondi, I., Salopek-Sondi, B. (2004). AgNPs as antimicrobial agent. A case study on *E. coli* as a model for Gram-negative bacteria. *Journal of Colloid and Interface Science*, 275 (1), 177–182.
- Sondi, I., Goia, D. V., Matijević, E. (2003). Preparation of highly concentrated s dispersions of uniform silver nanoparticles. *Journal of Colloid and Interface Science*, 260 (1), 75–81.
- Song, J. Y., Kim, B. S. (2009). Rapid biological synthesis of silver nanoparticles using plant leaf extracts. *Bioprocess and Biosystems Engineering*, 32 (1), 79–84.
- Spadaro, D., Barletta, E., Barreca, F., Currò, G., Neri, F. (2010). Synthesis of PMA stabilized AgNPs by chemical reduction process under a two-step UV irradiation. *Applied Surface Science*, 256 (12), 3812–3816.
- Stamplecoskie, K. G., Scaiano, J. C. (2010). Light emitting diode irradiation can control the morphology and optical properties of silver nanoparticles. *Journal of the American Chemical Society*, 132 (6), 1825-1827
- Sun, C., Qu, R., Chen, H., Ji, C., Wang, C., Sun, Y., Wang, B. (2008). Degradation behavior of chitosan chains in the “green” synthesis of gold nanoparticles. *Carbohydrate Research*, 343 (15), 2595–2599.
- Tamboli, D. P., Lee, D. S. (2013). Mechanistic antimicrobial approach of extracellularly synthesized AgNPs against gram positive and gram negative bacteria. *Journal of Hazardous Materials*, 260, 878–884.
- Tan, Y., Dai, X., Li, Y., Zhu, D. (2003). Preparation of gold, platinum, palladium and AgNPs by the reduction of their salts with a weak reductant–potassium bitartrate. *Journal of Materials Chemistry*, 13 (5), 1069–1075.

- Temgire, M. K., & Joshi, S. S. (2004). Optical and structural studies of silver nanoparticles. *Radiation Physics and Chemistry*, 71(5), 1039-1044.
- Thakkar, K. N., Mhatre, S. S., Parikh, R. Y. (2010). Biological synthesis of metallic nanoparticles. *Nanomedicine. Nanotechnology, Biology, and Medicine*, 6 (2), 257–262.
- Torreggiani, a., Jurasekova, Z., D'Angelantonio, M., Tamba, M., Garcia-Ramos, J. V., Sanchez-Cortes, S. (2009). Fabrication of Ag nanoparticles by  $\gamma$ -irradiation. Application to surface-enhanced Raman spectroscopy of fungicides. *Colloids and Surfaces A. Physicochemical and Engineering Aspects*, 339 (1-3), 60–67.
- Tran, Q. H., Nguyen, V. Q., Le, A. T. (2013). Silver nanoparticles. synthesis, properties, toxicology, applications and perspectives. *Advances in Natural Sciences. Nanoscience and Nanotechnology*, 4 (3), 1-20.
- Venkatapurwar, V., Pokharkar, V. (2011). Green synthesis of AgNPs using marine polysaccharide. Study of in-vitro antibacterial activity. *Materials Letters*, 65 (6),
- Vigneshwaran, N., Nachane, R. P., Balasubramanya, R. H., Varadarajan, P. V. (2006). A novel one-pot “green” synthesis of silver nanoparticles using soluble starch. *Carbohydrate Research*, 341 (12), 2012–2018.
- Wang, H., Qiao, X., Chen, J., Wang, X., Ding, S. (2005). Mechanisms of PVP in the preparation of silver nanoparticles. *Materials Chemistry and Physics*, 94 (2-3), 449–453.
- Wang, S. M., Huang, Q. Z., Wang, Q. S. (2005a). Study on the synergetic degradation of chitosan with ultraviolet light and hydrogen peroxide. *Carbohydrate Research*, 340 (6), 1143–1147.
- Wani, I., Ganguly, A., Ahmed, J., Ahmad, T. (2011). Silver nanoparticles. Ultrasonic wave assisted synthesis, optical characterization and surface area studies. *Materials Letters*, 65 (3), 520–522.
- Wani, I., Khatoon, S., Ganguly, A., Ahmed, J., Ganguli, A. K., Ahmad, T. (2010). Silver nanoparticles. Large scale solvothermal synthesis and optical properties. *Materials Research Bulletin*, 45 (8), 1033–1038.
- Xu, G. N., Qiao, X. L., Qiu, X. L., Chen, J. G. (2008). Preparation and characterization of stable monodisperse AgNPs via photoreduction. *Colloids and Surfaces A. Physicochemical and Engineering Aspects*, 320 (1-3), 222–226.
- Yin, H., Yamamoto, T., Wada, Y., Yanagida, S. (2004). Large-scale and size-controlled synthesis of AgNPs under microwave irradiation. *Materials Chemistry and Physics*, 83 (1), 66–70.

- Zargar, M., Shameli, K., Najafi, G. R., Farahani, F. (2014). Plant mediated green biosynthesis of silver nanoparticles using *Vitex negundo* L. extract. *Journal of Industrial and Engineering Chemistry*, 1–7.
- Zargar, M., Hamid, A. A., Bakar, F. A., Shamsudin, M. N., Shameli, K., Jahanshahi, F., Farahani, F. (2011). Green synthesis and antibacterial effect of silver nanoparticles using *Vitex negundo* L. *Molecules*, 16(8), 6667-6676.
- Zhang, Q., Yang, Z., Ding, B., Lan, X., Guo, Y. (2010). Preparation of copper nanoparticles by chemical reduction method using potassium borohydride. *Transactions of Nonferrous Metals Society of China*. 6326 (10) 60047-7
- Zhang, W., Qiao, X., Chen, J. (2007). Synthesis of silver nanoparticles-Effects of concerned parameters in water/oil microemulsion. *Materials Science and Engineering B. Solid-State Materials for Advanced Technology*, 142 (1), 1–15.
- Zhang, X. F., Liu, Z. G., Shen, W., Gurunathan, S. (2016). Silver Nanoparticles: Synthesis, Characterization, Properties, Applications, and Therapeutic Approaches. *International Journal of Molecular Sciences*, 17(9), 1534.
- Zhu, J., Liu, S., Palchik, O., Koltypin, Y., Gedanken, A. (2000). Shape-controlled synthesis of silver nanoparticles by pulse sonoelectrochemical methods. *Langmuir*, 16 (16), 6396-6399.
- Zielińska, A., Skwarek, E., Zaleska, A., Gazda, M., Hupka, J. (2009). Preparation of AgNPs with controlled particle size. *Procedia Chemistry*, 1 (2), 1560–1566.