



UNIVERSITI PUTRA MALAYSIA

***OPTIMIZATION OF ANTIOXIDANTS IN Curcuma zedoaria
(CHRISTM.)ROSCOE LEAVES AND THEIR FUNCTIONAL COSMETIC
PROPERTIES***

NUR FAUWIZAH BINTI AZAHAR

FS 2017 66



**OPTIMIZATION OF ANTIOXIDANTS IN *Curcuma zedoaria* (CHRSTM.)
ROSCOE LEAVES AND THEIR FUNCTIONAL COSMETIC PROPERTIES**

NUR FAUWIZAH BINTI AZAHAR

**Thesis Submitted to the School of Graduated Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for Degree of Master of Science**

June 2017

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of the thesis presented to the Senate of Universiti Putra Malaysia in
fulfillment of requirement for the degree of Master of Science

**OPTIMIZATION OF ANTIOXIDANTS IN *Curcuma zedoaria* (CHRSTM.)
ROSCOE LEAVES AND THEIR FUNCTIONAL COSMETIC PROPERTIES**

By

NUR FAUWIZAH BINTI AZAHAR

June 2017

Chairman : Associate Professor Siti Salwa Abd Gani, PhD
Faculty : Science

Medicinal plants especially leaves are an ample source of numerous pharmacologically active molecules. Therefore, continuous exploration from the leaves of *Curcuma zedoaria* has been extensively probe for its therapeutic value. In this study, the fourier transform infrared (FTIR) spectroscopy analysis was employed and it is found that the compounds present in the zedoary leaves samples are carboxylic acid, carbonyl, benzene and phenol. The levels of mineral content in zedoaria leaves were divided into two groups which are essential heavy metals; Mn (328.30 ppm), Fe (128.35 ppm), Cu (4.21 ppm), Ni (1.16 ppm), Cr (0.90 ppm) and non-essential heavy metals; (Pb) and (Cd) recorded to have 0.17 ppm and 0.03 ppm respectively which considered under permissible range for consumable medicinal plants. The extraction of antioxidants compounds from zedoary leaves were obtained by using response surface methodology (RSM). This method is used to optimize and simultaneously investigate the influence of condition variables such as temperature, time and solvent concentration towards highest total antioxidants assay. Result shows that at the optimized condition for high total antioxidants activity was found in zedoary leaves at 75°C, 92 mins and 90:10 v/v% ethanol concentrations exhibited (DPPH-radical scavenging (85.76%), β-carotene bleaching test (81.35%), ferric reducing antioxidant power assay (FRAP) (5.08 mM Fe²⁺ g/DW), total phenolic (125.75 mg/g GAE) and total flavonoid content (6.12 mg QE/g of extract). The optimized zedoary leaves were further explored for its functional cosmetics towards sun protection factor (SPF), anti-microbial test, anti-tyrosinase inhibition test, total carotenoids content, total ascorbic acid and total anthocyanin content. The results revealed that SPF value is 29.94 and fall into region UVC and UVB, the inhibition zone activity for bacteria *Bacillus Sublis* is 7.0 mm at concentration 100 mg mL⁻¹, tyrosinase inhibition recorded to have 31% and total carotenoids content, ascorbic acid including anthocyanins content is 8.36 g BET/100 g, 1.2577 mg/100 g and 4.25 mg/100 g respectively. To conclude, there could be clear potential for the utilization of *Curcuma zedoaria* leaves as a good source of antioxidants for diverse cosmeceutical, medicinal and food additives applications.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PENGOPTIMUMAN ANTIOKSIDA DALAM DAUN KUNYIT PUTIH *Curcuma zedoaria* (CHRISTM.) ROSCOE DAN FUNGSI SIFAT-SIFAT KOSMETIK

Oleh

NUR FAUWIZAH BINTI AZAHAR

Jun 2017

Pengerusi : Professor Madya Siti Salwa Abd Gani, PhD
Fakulti : Sains

Tumbuhan perubatan terutamanya dari daun adalah kaya dengan sumber molekul aktif farmakologi. Oleh itu, penerokaan berterusan dari daun kunyit putih telah disiasat secara meluas untuk nilai terapeutiknya. Dalam kajian ini, analisa FTIR menunjukkan sebatian yang hadir dalam daun kunyit putih ini adalah asid karbolik, karbonil, benzena dan fenol. Tahap kandungan mineral dalam daun zedoaria ini dibahagikan kepada dua kumpulan iaitu logam berat penting; Mn (328.30 ppm), Fe (128.35 ppm), Cu (4.21 ppm), Ni (1.16 ppm), Cr (0.90 ppm) dan logam berat tidak penting (Pb) dan (Cd) direkodkan masing-masing mempunyai berat 0.17 ppm dan 0.03 ppm iaitu di bawah julat yang dibenarkan untuk pengambilan tumbuhan perubatan. Kaedah gerak balas permukaan (RSM) telah digunakan untuk pengesektrakan antioksidan dari daun kunyit putih. Kaedah ini digunakan untuk pengoptimaan dan penyiasatan secara serentak oleh pengaruh tiga boleh ubah iaitu suhu, masa dan kepekatan pelarut ke arah jumlah antiokida yang tinggi. Data menunjukkan bahawa keadaan pengoptimuman untuk jumlah antioksidan yang tinggi dari daun kunyit putih adalah 75°C, 92 min dan 90:10 v/v% kepekatan etanol mempaparkan (DPPH-perangkap radikal (85.76%), ujian pelunturan β- karotena (81.35%), pengurangan kuasa antiokida ferik (FRAP) (5.08 mM Fe²⁺ g/DW), jumlah fenolik (125.75 mg/g GAE) dan jumlah kandungan flavonoid (6.12 mg QE/g). Penerokaan daun kunyit putih yang telah dioptimumkan diteruskan untuk fungsi kosmetik iaitu faktor perlindungan matahari (SPF), ujian anti-mikrob, ujian perencutan anti-tyrosinase, kandungan karotenoid, asid askorbik dan kandungan antosianin. Hasil kajian menunjukkan bahawa nilai SPF adalah 29.94 dan berada di kawasan UVC dan UVB, aktiviti zon perencutan untuk bakteria *Bacillus Sublis* adalah 7.0 mm pada kepekatan 100 mg mL⁻¹, data peratusan perencutan anti-tyrosinase sebanyak 31% dan jumlah kandungan karotenoid, asid askorbik termasuk antosianin masing-masing mengandungi 8.36 g BET/100 g, 1.2577 mg/100 g dan 4.25 mg/100 g secara turut. Secara kesimpulan, daun kunyit putih mempunyai potensi yang jelas sebagai sumber antioksidan yang baik untuk diaplakasikan dalam pelbagai kosmeseutikal, ubat-ubatan dan makanan.

ACKNOWLEDGEMENTS

In The Name of ALLAH, The Most Merciful and Most Beneficent

Allhamdulilah, all praises, glory and thanks to Allah, the almighty lord of the universe. Peace and blessing to Nabi Muhammad S.A.W., all the prophets, his families and all muslims. Only by His grace and mercy, this thesis can be completed.

Foremost, I would like to express my sincere gratitude to Assoc. Prof. Dr. Siti Salwa Abd. Gani, and Dr. Nor Fadzillah Mohd Mokhtar for patiently guiding me through the course of this thesis to its eventual end enlightening me scientifically and resolving my technical crises. Their patience, critics, motivation and immense knowledge throughout this study is greatly appreciated.

Secondly, I am very thankful to my parents Azahar bin Othman and Aishah binti Salleh, my brother and sister for their encouragement, love and kindness in my life. The holistic life during conducting this research is unable to achieve without the companion from all of you,

Finally, special thanks to my beloved friends Ramya, Ain, Farah, Kak Haila and colleagues who in many ways contributed to the success of my thesis. Without support from them, this thesis would not be the same as presented here. Thank you.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Siti Salwa Abd Gani, PhD
Associate Professor
Faculty of Science
Universiti Putra Malaysia
(Chairman)

Nor Fadzillah Mohd Mokhtar, PhD
Senior Lecturer
Faculty of Science
Universiti Putra Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD
Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:

Name of Chairman of
Supervisory Committee:

Associate Professor Dr. Siti Salwa binti Abd Gani

Signature:

Name of Member of
Supervisory Committee:

Dr Nor Fadzillah Mohd Mokhtar

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	ii
ACKNOWLEDGEMENTS	iii
APPROVAL	iv
DECLARATION	vi
LIST OF TABLES	x
LIST OF FIGURES	xi
LIST OF ABBREVIATIONS	xii
CHAPTER	
1. INTRODUCTION	1
1.1. Background	1
1.2. Objectives	2
2. LITERATURE REVIEW	3
2.1. Skin Aging and Antioxidants	3
2.2. Cosmetic Antioxidants-Natural Versus Synthetic	4
2.3. Medicinal Plant	5
2.4. Potential Leaves as a Biomarker Antioxidants	6
2.5. <i>Curcuma</i> , Zingiberaceae Family	6
2.5.1. <i>Curcuma zedoaria</i>	7
2.5.2. Traditional Uses	8
2.5.3. Phytochemistry and Chemical Constituents	9
2.6. Response Surface Methodology (RSM)	9
2.7. Pharmacological Identified from <i>Curcuma zedoaria</i> Extract	10
2.7.1. Anti- Oxidant Activity	10
2.8. Cosmetics and Functional Cosmetics	11
2.8.1. Sun Protection Factor (SPF)	12
2.8.2. Anti-Microbial Activity	13
2.8.3. Anti-Tyrosinase Activity	14
2.8.4. Carotenoids, Ascorbic Acids and Anthocyanins Content	14
3. MATERIALS AND METHODS	16
3.1. Materials and Reagents	16
3.2. Phytochemical in <i>Curcuma zedoaria</i> Leaves	16
3.2.1. Fourier Transform Infrared (FTIR) Spectroscopy Analysis	16
3.2.2. Mineral Content	16
3.3. Response Surface Methodology (RSM)	17
3.3.1. Experimental Design	17

3.3.2.	Statistical Analysis and Optimization	18
3.4.	Plant Extraction	19
3.5.	Antioxidant Test	19
3.5.1.	DPPH- Radical Scavenging Activity	19
3.5.2.	β - carotene Bleaching (BCB) Assay	20
3.5.3.	Ferric Reducing Antioxidant Power Assay (FRAP)	20
3.5.4.	Total Phenolic Content	21
3.5.5.	Total Flavonoid Content	21
3.6.	Functional Cosmetic Properties of <i>Curcuma zedoaria</i> Leaves-Based Optimum Conditions	21
3.6.1.	Sun Protection Factor (SPF)	21
3.6.2.	Anti-Microbial Test	22
3.6.3.	Anti-Tyrosinase Inhibition Test	23
3.6.4.	Total Carotenoids Content	23
3.6.5.	Total Ascorbic Acid Content	23
3.6.6.	Total Anthocyanins Content	24
4.	RESULTS AND DISCUSSION	25
4.1.	Phytochemical in <i>Curcuma zedoaria</i> Leaves	25
4.1.1.	FTIR	25
4.1.2.	Mineral Content	26
4.2.	Response Surface Methodology	28
4.2.1.	Fitting The Response Surface Models (RSM)	28
4.3.	Antioxidants (RSM) Analysis	32
4.3.1.	DPPH-Free Radical Scavenging	32
4.3.2.	B-carotene Bleaching (BCB) Assay	35
4.3.3.	Ferric Reducing Antioxidant Power Assay (FRAP)	38
4.3.4.	Total Phenolic Content	40
4.3.5.	Total Flavonoid Content	43
4.4.	Optimization of <i>Curcuma zedoaria</i> Leaves	46
4.4.1.	Optimal Conditions	46
4.5.	Functional Cosmetic Properties of <i>Curcuma zedoaria</i> Leaves-Based Optimized Conditions	47
4.5.1.	Sun Protection Factor (SPF) Value	47
4.5.2.	Anti-Microbial Test	49
4.5.3.	Anti-Tyrosinase Inhibition Test	51
4.5.4.	Total Carotenoids Content	53
4.5.5.	Total Ascorbic Content	54
4.5.6.	Total Anthocyanins Content	55
5.	CONCLUSION	57
REFERENCES		58
APPENDICES		69
BIODATA OF STUDENT		87
LIST OF PUBLICATIONS		88

LIST OF TABLES

Table		Page
1	Taxonomic classification of <i>Curcuma zedoaria</i>	7
2	Traditional uses of different parts of <i>Curcuma zedoaria</i>	8
3	Chemical constituents in <i>Curcuma zedoaria</i>	9
4	Types of bacteria and its diameter of zone of inhibition (mm)	13
5	Independent test variables and their coded and uncoded value used for CCD matrix	18
6	20 series of experimental runs	18
7	Normalized extract function used in the calculation of SPF	22
8	List of bacteria tested	22
9	FTIR peak values of <i>Curcuma zedoaria</i> leaves	26
10	Concentrations (mg/kg) heavy metals in <i>Curcuma zedoaria</i> leaves	28
11	The experimental data obtained for the three responses based on the CCD matrix	30
12	Analysis of variance (ANOVA) for the model	31
13	Comparison between the predicted and experimental values for antioxidants from extracts of <i>Curcuma zedoaria</i> leaves	47
14	SPF value of ethanolic extract of <i>Curcuma zedoaria</i> leaves	49
15	Zone inhibition of bacteria and sample tested	51

LIST OF FIGURES

Figure		Page
1	Illustration of aging process	4
2	Reaction of Vitamin C with reactive radical	4
3	<i>Curcuma zedoaria</i> leaves	8
4	FTIR spectrum of <i>Curcuma zedoaria</i> leaves	26
5	Response surface plots for the effects of temperature, time and ethanol concentration on DPPH scavenging activity of <i>Curcuma zedoaria</i> leaves extracts.	35
6	Response surface plots for the effects of temperature, time and ethanol concentration on β -carotene of <i>Curcuma zedoaria</i> leaves extracts.	37
7	Response surface plots for the effects of temperature, time and ethanol concentration on FRAP reducing assay of <i>Curcuma zedoaria</i> leaves extracts.	40
8	Response surface plots for the effects of temperature, time and ethanol concentration on phenolic content of <i>Curcuma zedoaria</i> leaves extracts.	43
9	Response surface plots for the effects of temperature, time and ethanol concentration on flavonoid content of <i>Curcuma zedoaria</i> leaves extracts.	46
10	SPF spectra for <i>Curcuma zedoaria</i> leaves	48
11	Anti-tyrosinase activity against mushroom tyrosinase	52
12	Percentage inhibition of <i>Curcuma zedoaria</i> leaves, kojic and ascorbic acid	52
13	Standard calibration curve of β -carotene	54
14	Calibration curve of ascorbic acid	55

LIST OF ABBREVIATIONS

AA	Ascorbic acid
Anova	Analysis of variance
AOAC	Association of Analytical Communities
Ar	Arsenic
BCB	B-carotene bleaching
CCD	Central composite design
Cd	Cadmium
Cr	Chromium
Cu	Copper
cm	Centimetre
DPPH	1,1-diphenyl-2-picrylhdrazyl
Fe	Iron
FRAP	Ferric ion Reducing Antioxidant Power Assay
FTIR	Fourier Transform Infrared
GCMS	Gas Chromatography –Mass Spectroscopy
mg/L	Milligram per litre
Mn	Manganese
min	Minute
MPAES	Microwave plasma- atomic emission spectroscopy
mL	Mililiter
Ni	Nickel
nm	Nanometer
Pb	Lead
RSM	Response Surface Methodology
SPF	Sun Protection Factor
TFC	Total flavonoid content
TPC	Total phenolic content
µg/L	Microgram per litre
UV-Vis	Ultra Violet –Visible Infrared Spectroscopy
Zn	Zinc

CHAPTER I

INTRODUCTION

1.1 Background

Plants are substantial source of natural antioxidants. Active constituents present in natural antioxidants such as phenolic, carotenoids, flavonoids, folic acid, benzoic acid, and tocopherol are secondary metabolites of the plants which can provide various potential treatment and prevention of cancer, cardiovascular diseases, and neurodegenerative diseases (Ghasemzadeh et al., 2010; Karabegović et al., 2014). Antioxidant is a highly reactive compound help to delay or inhibit the initiation and propagation of oxidizing chain reactions involving uncontrollable metabolic processes from reactive oxygen species (ROS) and free radical species (Chanda and Nagani, 2010). Practically, radical damage can be treated by both synthetic and natural antioxidants. However, the use of artificial antioxidants such as butylated hydroxytoluene (BHT) and butylated hydroxyanisole (BHA) are suspected to increase the risk of cancer and liver damage in humans (Hue et al., 2012).

Recently, it has been investigated that countless by-products from the processing plants like leaves, barks, fruit pods, shells and even pulp contain numerous biologically active compounds. It was reported to contain high level of phytochemicals sources such as antioxidants, antimutagens and anticarcinogenic. For instance, the leaves from tea, lemon and papaya are explored and found to contain high level of phytochemicals constituents which resulted in the development of natural antioxidant formulations for cosmetic and medicinal applications.

Fortunately, Malaysia is one of world's richest tropical rainforest that can provide useful primary medicinal plants and herbs. In recent times, Zingiberaceae from the largest family of Zingiberales is getting recognition from scientists all over the world for their potential benefit to living organisms (Das and Rahman, 2012). One of the genus of Zingiberaceae families that has higher potential is *Curcuma zedoaria*, which is popularly known as white turmeric, zedoaria or gajutsu (Lobo et al., 2009). Despite numerous studies on *Curcuma zedoaria* such as rhizomes and roots which cater enormous functional components, however very limited scientific information on antioxidant properties conducted on *Curcuma zedoaria* leaves for its therapeutic potential. In this study, the antioxidant activity and functional cosmetic properties of *Curcuma zedoaria* leaves were investigated and explored.

1.2 Objectives

The general objective of this study was to investigate the phytochemical of *Curcuma zedoaria* leaves as potential for antioxidant activity and its cosmetic functional properties.

The specific objectives are as follow;

- i. To identify the phytochemicals constituents present in *Curcuma zedoaria* leaves via FTIR and mineral content.
- ii. To investigate the five different antioxidants assay (DPPH-radical scavenging, β -carotene, Ferric reducing antioxidant power assay (FRAP), total phenolic and total flavonoid content) present in *Curcuma zedoaria* leaves and to optimize an antioxidants activity using Response Surface Methodology (RSM)
- iii. To determine the functional cosmetic properties *Curcuma zedoaria* leaves extract with respect to the sun protection factor (SPF), anti-microbial test, anti-tyrosinase inhibition test, total carotenoids, ascorbic acid and anthocyanin content.

REFERENCES

- Abdul Karim, A.; Azlan, A.; Ismail, A.; Hashim, P.; Abd Gani, S.S.; Zainudin, B.H.; Abdullah, N.A. Phenolic composition, antioxidant, anti-wrinkles, and tyrosinase inhibitory activities of cocoa pod extract. *BMC Complement. Altern. Med.* 2014. 14, 381
- Abdul Rohman. Mini review analysis of curcuminoids in food and pharmaceutical products. *Int. Food Res. J.* 2012. 19(1), 19-27.
- Ahirrao, R.A.; Patel, M.R.; Pokal, D.M.; Patil, J.K.; Suryawanshi, H.P. Phytochemical screening of leaves of *Jatropha curcas* plant. *Int. J. Res. Ayurveda Pharm.* 2011. 2(4), 1324-1327.
- Ajmal, K.; Shahid, J.; Atif, M.; Tahir, M.; Abid, N.; Abdul, M. Heavy metals status of soil and vegetables grown on peri-urban area of Lahore district. *Soil Environ.* 2013. 32, 49-54.
- Akhila, S.; Vijayalakshmi, N.G. Phytochemical studies on carica papaya leaf juice. *Int. J. Pharma. Sci. Res.* 2015. 6(2), 880-883.
- Amado, I.R.; Franco, D.; Sánchez, M.; Zapata, C.; Vázquez, J.A. Optimisation of antioxidant extraction from Solanum tuberosum potato peel waste by surface response methodology. *Food Chem.* 2014. 165, 290–299.
- Amit, G.; Ashawat, M.S.; Shailendra, S.; Swarnlata, S. Phytosome: A novel approach towards functional cosmetics. *J. Plant Sci.* 2007, 2(6), 644-649.
- Angel, G.R.; Vimala, B.; Bala, N. Phenolic content and antioxidant activity in five underutilized starch curcuma species. *Int.J. Pharmacognasy Phytochem. Res.* 2012. 4(2), 69-73.
- Archana, A.; Bharathi, V.D.; Saraboji, S.; Thirunavukkarassu,; Nithya, R. Optimization and extraction of phenolic compounds from Capsicum annuum using response surface methodology. *Int. J. Emerg. Res. Manag. Technol.* 2015. 4(7), 204-211.
- Ashawat, M.S.; Banchlor, M.; Saraf, S.; Saraf, S. Herbal cosmetics: Trends in skin care formulation. *Pharmacogn.J.* 2009. 3(5), 72-79.
- Baby, T.; Saraswathi, U.; Revathi, S.; Malathi, M. Screening of antioxidant and antityrosinase activities of a herbal formulations. *Int. J. Innov. Drug Discv.* 2014. 4(2), 93-98.
- Bachir Bey, M.; Meziant, L.; Benchikh, Y.; Louaileche, H. Deployment of response surface methodology to optimize recovery of dark fresh fig (*Ficus carica* L., var. Azenjar) total phenolic compounds and antioxidant activity. *Food Chem.* 2014. 162, 277–282.

- Banisalam, B.; Sani, W.; Philip, K.; Imdadul, H.; Khorasani, A. Comparison Between in vitro and in vivo Antibacterial Activity of Curcuma zedoaria from Malaysia. *African J. Biotechnol.* 2011.10, 11676–11681.
- Barbes, L.; Barbulescu, A.; Radulescu,C.; Sithi,C.; Chelarescu, E.D. Determination of heavy metals in leaves and bark of *populus nigra L* by atomic absorption spectrometry. *Rom reports Phy.* 2014. 66,877-886.
- Bazykina, N.I.; Nikolaevskii,A.N.; Filippenko,T.A.; Kolerva,V.G. Optimization of conditions for the extraction of natural antioxidants from raw plant materials. *Pharm.Chem.J.* 2002. 36, 46-49.
- Benzie,I.E.F.,Strain,J. The ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power": the FRAP assay. *Anal.Biochem.* 1996. 239, 70-76.
- Blois,M.S. Antioxidant determinations by the use of a stable free radical. *Nature.* 1958. 181, 1199-1200.
- Bonn mee.A.; Srisomsap.C.; Karchanat A.; Sangvanich. P. An antioxidant protein in Curcuma comosa Roxb.Rhizomes. *Food Chem.* 2011. 124, 476-480.
- Brophy, J.J. Essential oils from Three Curcuma Species Collected in Thailand. *Perspec. Nat. Prod.Chem.* 2005. 3, 37-41.
- Chanda, S. V.; Nagani, K.V. Antioxidant Capacity of Manilkara zapota L . Leaves Extracts Evaluated by Four in vitro Methods. *Nat.Sci.* 2010. 8, 260–266.
- Chang, S.K.; Yim, H.S. Influence of extraction conditions on antioxidant properties of passion fruit (*passiflora edulis*) peel. *Acta Sci.Pol.,Technol.Aliment.* 2014. 13, 257-265.
- Chaveerach, A.; Sudmoon, R.; Tanee, T.; Mokkamul, P.; Sattayasai, N.; Sattayasai,J. Two New Species of (Zingiberaceae) used as cobra bite. *J. Syst. Evol.* 2008. 46(1), 80-88.
- Chitta Ranjan Sarkar.; Lima Das.; Bagmitta Bhagawati.; Bhabesh, C.H.G. A comparative study of carotenoids extraction from algae in different solvent systems. *Asian J. Plant Sci. Res.* 2012. 2(4), 546-549.
- Cho, W.Y.; Kim, S.J. Antioxidative actions of Curcuma zedoaria extract with inhibition of inducible nitric oxide synthase (iNOS) induction and lipid peroxidation. *J.Med. Plants Res.* 2012. 6(22), 3837-3844.
- Chowdury, J.A.; Islam, M.S.; Asifuzzamman, S.K.; Islam. M.K. Antibacterial and cytotoxic activity screening leaf extract of vitex negundo (*Fam:Verbenaceae*). *J. Pharma. Sci. Res.* 2009. 4, 103-108.
- Curcuma zedoaria leaves. Zedoary (Curcuma zedoaria) Overview, Health Benefits, Side effects. Retrieved 11 June 2017 from www.tipdisease.com/UsefulHerbal/
- Das, K.; Rahman, M.A. Analgesic and antimicrobial activities of Curcuma zedoaria. *Int. J. Pharm. Pharm. Sci.* 2012. 4, 322–328.

- De Oliveira Junior, R.G.; Araújo, C. de S.; Souza, G.R.; Guimarães, A.L.; De Oliveira, A.P.; de Lima-Saraiva, S.R.G.; Morais, A.C.S.; dos Santos, J.S.R.; Almeida, J.R.G. da S. In vitro antioxidant and photoprotective activities of dried extracts from *Neoglaziovia variegata* (Bromeliaceae). *J. Appl. Pharm. Sci.* 2013. 3, 122–127.
- Dipali Gupta. UV absorbing properties of some plant derived extracts. *Res. J.Chem. Environ.Sci.* 2013. 34-36.
- Ebrahimzadeh, M.A.; Enayatifard, R.; Khalili, M.; Ghafarrloo, M.; Saeedi, M.; Charati.Y.Corellation between sun protection factor and antioxidant activity, phenol and flavonoid contents of some medicinal plants. *Iranian J. Pharma. Res.* 2014. 13(3), 1041-1047.
- Erna, W.I.H.; Ahmad Ziad, S.; Mimi Sakinah, A.M.. Assessment of Heavy metals tolerance in leaves, stems and flowers of stevia rebaudiana plant. *Procedia Environ.Sci.* 2014. 20, 386-393.
- FDA. Sunscreen drugs products for over the counter human use. Department of health, education and welfare, USA. 2001. *Fed. Regist.* 64/27687, 276.
- Fidrianny, I.; Rahmiyani, I.; Wirasutisna, K.R. Antioxidant capacities from various leaves extracts of four varieties mangoes using DPPH, ABTS assay and correlation with total phenolic , flavonoid, carotenoid. *Int.J. Pharma. Pharma.Sci.* 2013.5, 189-194.
- Garcia-Mendoza, M.D.P.; Espinosa-Pardo, F.A.; Baseggio, A.M.; Barbero, G.F.; Junior, M.R.M.; Rostagno, M.A.; Martinez, J. Extraction of phenolic compounds and anthocyanins from jucara (*Euterpe edulis* Mart.) residues using pressurized liquids and supercritical fluids. *J. Supercrit. Fluids.* 2016. 119, 9-16.
- Garg, S.N.; Naquvi, A.A.; Bansal, R.P.; Bahl, J.R.; Kumar, S. Chemical Composition of the essential oil from the leaves of *Curcuma zedoaria* rosc. of Indian Origin. *J. Essential Oil Res.* 2005. 17, 29-31.
- Gediya, S.K.; Mistry, R.B.; Patel,U.K.; Blessy, M.; Jain, H.N. Herbal plant : used as cosmetics. *Sch. Res. Libr.* 2011. 24-32.
- Ghasemzadeh, A.; Jaafar, H.Z.E.; Rahmat, A. Antioxidant activities, total phenolics and flavonoids content in two varieties of malaysian young ginger (*Zingiber officinale* Roscoe). *Molecules.* 2010. 15, 4324–4333.
- Gomes, T.; Delgado, T.; Ferreira, A.; Pereira, J.A.; Baptista, P.; Casal, S.; Ramalhosa, E. Application of response surface methodology for obtaining lettuce (*Lactuca sativa* L.) by-products extracts with high antioxidative properties. *Ind.Crops Prod.* 2013. 44, 622–629.

- Gulsen, O.; Roose, M.L. Lemons: diversity and relationships with selected Citrus genotypes as measured with nuclear genome markers. *J Amer Soc Hort. Sci.* 2001. 126, 309-317.
- Hamdi,O.A.A.; Ye,L.J.; Kamarudin, M.N.A.; Hazni,H.; Paydar,M.; Looi, C.Y.; Shilpi, J.A.; Kadir,H.A.; Awang,K. Neuroprotective and antioxidants constituents from curcuma zedoaria rhizomes. *Rec. Nat. Prod.* 2015. 9(3), 349-355.
- Herrling, T.; Jung, K.; Chatelain, E.; Langenauer.M. Radical skin/ sun protection factor RSF-protection against UV-induced free radicals in skin. *SOFW. J.* 2016. 132, 24-30.
- Himaja,M.; Anand, R.; Ramana,M.V.; Anand,M.; Karigar,A. Phytochemical screening and antioxidant activity of rhizome part of Curcuma Zedoaria. *Int. J. Res. Ayurveda Pharm.* 2010. 2, 414-417.
- Hue, S.; Boyce, A.N.; Somasundram, C. Antioxidant Activity , Phenolic and Flavonoid Contents in the Leaves of Different Varieties of Sweet Potato (*Ipomoea batatas*). *Aust. J. Crop Sci.* 2012, 6, 375–380.
- Hussain, I.; Khan, L.; Khan, M.A.; Khan, F.U.; Ayaz, S.; Khan, F.U. UV spectrophotometric analysis profile of ascorbic acid in medicinal plants of Pakistan. *World App. Sci. J.* 2010. 9(7), 800-803.
- Ifesan, B.O.T.; Fashakin, J.F., Ebosele, F.; Oyerinde, A.S. Antioxidant and Antimicrobial properties of selected plant leaves. 2013.3(3), 465-473.
- Illustration of aging process. Retrieved 10 June 2017 from <http://shiraesthetics.com/how-skin-ages/>.
- Isla, M.; Cordero, A.; Diaz, L.; Perez-Perez, E.M.; Vit, P. Cosmetic properties of honey. 1. Antioxidant activity. In Vit P & Roubik DW, eds. Stingless bees process honey and pollen in cerumen pots. Facultad de Farmacia y Bioanalisis, Universidad de Los Andes, Merida,Venezuela, 1-8. Retrieved from <http://www.saber.ula.ve/handle/123456789/35292>
- Janghel, E.K.; Santosh, S.; Pervez, Y. A new method for determination of ascorbic acid in fruit juices, pharmaceuticals and biological sample. *J. Sci. Ind. Res.* 2012.71, 549-555.
- Jarup, L. Hazards of heavy metal contamination. *Br. Med. Bull.* 2013. 68(1), 167-182.
- Karnjanawipagul, P.; Nittayanuntawech, W.; Rojsanga, P.; Suntornsuk, L. Analysis of b-carotene in carrot by spectrometry. *Mahidol Univ. J. Pharm. Sci.* 2010.37, 8-16.
- Kapoor,V.P. Herbal Cosmetics for Skin and Hair Care. *Nat. Prod. Rad.* 2005, 4(4), 306-314.
- Karabegović, I.T.; Stojičević, S.S.; Veličković, D.T.; Todorović, Z.B.; Nikolić, N.Č.; Lazić, wM.L. The effect of different extraction techniques on the composition

- and antioxidant activity of cherry laurel (*Prunus laurocerasus*) leaf and fruit extracts. *Ind. Crops Prod.* 2014. 54, 142–148.
- Kavanaugh, E.E. Tentative final monograph for OTC Sunscreen. *Cosmet, Toilet. and Fragr. Assoc.* Retrieved on 20170142 at www.fda.gov/ohrms/dockets/daily
- Khalaf, N A.; Shakya, A.K.; Al-othman, A.; El-agbar, Z.; Farah, H. Antioxidant activity of some common plants. *Turk J Biol.* 2007. 32, 51-55.
- Kumar, D.; Jain, V.K.; Shanker, G.; Srivastava,A. 2003. Citric Acid Production by Solid State Fermentation Using Sugarcane Bagasse. *J Process Biochem,* 38,1731-1738.
- Khoo, H.E.; Azlan,A.; Ismail,A.; Abas.F. Response surface methodology optimization for extraction of phenolics and antioxidant capacity in defatted dabai parts. *Sains malaysiana,* 2013. 42, 949-954.
- Kozubek, A.; Zarnowski, R.; Stasiuk, M.; Gubernator, J. Natural amphiphilic phenols as bioactive compounds. *Cell. Mol. Biol. Let.* 2001. 6, 351-355.
- Lai, H.; Lim, Y. Evaluation of Antioxidant Activities of the Methanolic Extracts of Selected Ferns in Malaysia. *Int. J. Environ. Sci. Dev.* 2011. 2, 442–447.
- Lakshmi, S.; Padmaja, G.; Remani, P.; Antitumour effects of isocurcumenol isolated from curcuma zedoaria rhizomes on human and murine cancer cells. *Int. J. Med. Chem.* 2011, 13.
- Lawal, A.O.; Batagarawa, S.M.; Oyeyinka,O.D.; Lawal,M.O. Estimation of heavy metals in Neem Tree leaves along katsina-Dutsinma-Funtua-Highway in Katsina State of Nigeria. *J.Appl.Sci.Environ.Manage.* 2011. 15, 327-330.
- Lee, J.H.; Cho, S.; Paik, H.D.; Choi, C.W.; Nam, K.T.; Hwang, S.G.; Kim, S.K. Investigation on antibacterial and antioxidant activities , phenolic and flavonoid contents of some thai edible plants asn an alternative for antibiotics. *Asian Australas. J Anim. Sci.* 2014. 27(10), 1461-1467.
- Lemos, L.; Morelli, L.; Prado, M.A. Ultrasonics Sonochemistry Extraction optimization for antioxidant phenolic compounds in red grape jam using ultrasound with a response surface methodology.*Ultrason.sonochemistry.* 2012. 19, 1144–1149.
- Lobo, R.; Prabhu, K.S.; Shirwaikar, A.; Shirwaikar, A.Curcuma zedoaria Rosc. (white turmeric): a review of its chemical, pharmacological and ethnomedicinal properties. *J. Pharm. Pharmacol.* 2009. 61, 13–21
- Lu, J.; Zhou, C.; Rong, O.; Xu, Y. Optimization of microwave-assisted extraction of flavonoids from *Cryptotaenia japonica* hassk using response surface methodology. *Adv. J. food Sci. Technol.* 2013. 5, 310-317.
- Lu, J.M.; Lin, P.H.; Yao, Q.; Chen, C. Chemical and molecular mechanisms of antioxidants : experimental approaches and model systems. *J.Cell. Mol. Med.* 2010. 14(4), 840-860.

- Maizura, M.; Aminah, M.; Wan Aida, M. Total phenolic and antioxidant activity of Kesum (*Polygonum minus*), ginger (*Zingiber officinale*) and turmeric (*Curcuma Longa*) extract. *Int. Food Res. J.* 2011. 18,529-534.
- Majeed, M.; Hussain, AB.; Chatha, S.A.S.; Khosa, M.K.K.; Kamal, G.M.; Kamal, M.A.; Zhang, X.; Liu, M. Optimization protocol for the extraction of antioxidant components from *Origanum vulgare* leaves using response surface methodology. *Saudi J. Biol. Sci.* 2015. Article in press.
- Malek, S.N.A; Lee, G.S.; Hong, S.L.; Yaacoh, H.; Abdul Wahad, N.; Faizal Weber, J.F.; Ali shah, S.A. Phytochemical and cytotoxic investigations of Curcuma manga rhizomes. *Molecules*. 2011. 16, 4540-4548.
- Mansur, J.S.; Breder, M.V.R.; Mansur, M.C.A.; Azulay, R.D. Determination of sun protection factor for spectrophotometry. *An Bras Dermatol.* 1986. 61(4), 121-124.
- Maobe, M.A.G.; Gatabe,E.; Gitu,L.; Rotish,H. Profile of Heavy metals in selected medicinal plants used for the treatment of diabetes, malaria and pneumonia in Kisii Region, southwest kenya. *Glob. J. Pharmacol.* 2012. 6,245-251.
- Marija Radojkovića.; Zoran Zekovića.; Stela Jokićb, S.V. Determination of optimal extraction parameters of mulberry leaves using Response Surface Methodology (RSM) Materials and Methods. *Rom. Biotechnol. Lett.* 2012, 17, 7295–7308.
- Masnec, I.S.; Situm, M. Skin Aging. *Orig. Sci. Pap.* 2010.49, 515-519.
- Mbanga,L.; Mulenga, M.; Mpiana, P.T.; Bokolo, K.; Mumbwa,M.; Mvingu.K. Determination of sun protection factor (SPF) os some body creams and lotions marketed in Kinshasa by ultraviolet spectrophotometry. *Int. J. Advance Res Chem Sci.* 2014. 1, 7-13
- Mechanism of electron transfer of antioxidant. Retrieved 2016 from <http://evolutionaryhealthperspective.wordpress.com>
- Mensor, L.L.; Menezes, F.S.; Leitao, G.G.; Reis, A.S.; Dos Santos, T.C.; Coube, C.S.; Leitao, G. Screening of Brazilian plant extracts for Antioxidant activity by the Use of DPPH Free radical method. *Phytother. Res.* 2001. 15, 127-130.
- Mitra,K.; Uddin,N. Total phenolics, flavonoids, proanthocyanidins, ascorbic acid contents and in-vitro antioxidant activities of newly developed isolated soya protein. *Discourse J. Agric. Food Sci.* 2014. 2,160-168.
- Mohd Taib, S.H.; Abd Gani, S.S.; Ab Rahman, M.Z.; Basri, M.; Ismail, A.; Shamsudin, R. Formulation and process optimizations of nano-cosmeceuticals containing purified swiftlet nest. 2015. 5, 42322–42328.
- Mohammad, N.; Huyop, F.; Aboul-Enein, H.Y.; Mahat, N.A.; Abdul Wahab, R. Response surface methodological approach for optimizing production of

- geranyl propionate catalysed by carbon nanotubes nanobioconjugates. *Biotechnol. Biotechnol. Equip.* 2015. 29(4), 732-739.
- More, B.H.; Sakharwarde, S.N.; Tembhurne, S.V.; Sakarkar, D.M. Evaluation of sunscreen activity of cream containing leaves extract of *Butea monosperma* for topical applications. *Int. J. of Res. Cos. Sci.* 2013. 3(1), 1-6.
- Muthu Kumar, T.; Mary Violet Christy,A.; Anusha Mangadu.; Malaisamy, M.; Sivaraj,C.; Arjun, P.; Rahman, N.; Balasubramanian, K. Anticancer and antioxidant activity of curcuma zedoaria and curcuma amada rhizome extracts. *J. Acad. Ind. Res.* 2012. 12, 91-96.
- Narayanaswamy, N.; Duraisamy, A.; Balakrishnan, K.P. Screening of some medicinal plants for their antityrosinase and antioxidant activities. *Int. J. of Pharmatech. Res.* 2011. 3(2), 1107-1112.
- Nworie F.S.; Jedidiah, J. Determination of ascorbic acid levels in Hibiscus Sabdariffa (Zobo rod) and other tropical leaves. *Int. J.Sci.Res.* 2012. 3(6), 1505-1508.
- Ogbonda, G.E.; Kabari,L.G. Heavy metal content in bitter leaf (*Vernonia amygdalina*) Grown Along Heavy traffic routes in port Harcourt. *Intech.* 2013. 202-210.
- Olives Barba, A.I.; Camara Hurtado, M.; Sanchez Mata.; Fernandez Ruiz,V.; de Tejada, M.L.S.. Application of UV-Vis detection-HPLC method for a rapid determination of lycopene and β-carotene in vegetables. *Food Chem.* 2006. 95,328-336.
- Omaruyi, B.E.; Afolayan, A.J.; Bradley, G. The inhibitory effect of *Masembryanthemum edule* (L.) bolus essential oil on some pathogenic fungal isolates. *BMC complement. Altern. Med.* 2014. 14, 168.
- Paramapojn,S.; Gritsanapan,W. Free radical scavenging activity determination and quantitative analysis of curcuminoids in curcuma zedoaria rhizome extract by HPLC method. *Curr. Sci.* 2009. 97, 1069-1073.
- Phan, M.G.; Van, N.G.; Phan, T.S. Antimicrobial activity of sesquiterpene constituents from Curcuma species of Vietnam. *Tap Chi Hoa Hoc*, 2000. 38, 91-94.
- Philip Koshy.; Abd malek, S.N.; Sani, W.; Shin, S.K.; Kumar, S.; Iai, H.S.; Serm, L.G.; Rahman, S.N.S.A. Antimicrobial activity of some medicinal plants from Malaysia. *American J App Sci.* 2009. 6 (8), 1613-1617.
- Pisoschi, A.M.; Pop, A.; Negulescu, G.P.; Pisoschi, A. Determination of Ascorbic acid content of some fruit juices and wine by voltammetry performed at Pt and carbon paste electrodes. *Molecules.* 2011.16, 1349-1365.
- Radwan, M.A.; Salama, A.K. Market basket survey for some heavy metals in Egyptian fruits and vegetables. *Food chem. Toxicol.* 2006. 44, 1273-1278.
- Rahiman, R.; Mohd Ali, M.A.; Ab-Rahman, M.S. Carotenoids concentration detection investigation : A review of current status and future trend. *Int. J. Biosci. Biochem Bioinforma.* 2013. 3(5), 466-472.

- Rahman, A.; Afroz, M.; Islam. R.; Islam, K.D.; Hossain, A.; Na, M. In vitro antioxidant potential of the essential oil and leaf extracts of curcuma zedoaria rosc. *J. App. Pharma.Sci.* 2014. 4(2), 107-111.
- Rahman Khan, M.M.; Rahman, M.M.; Islam, M.S., Begum, S.A. A simple UV-spectrophotometric method for the determination of vitamin c content in various fruits and vegetables at Syllent area in Bangladesh. *J. Biol. Sci.* 2006. 6(2), 388-392.
- Raimundo, G.O.J.; Camila S.J.; Grasielly, R.S.; Amanda, L.G.; Ana P.O.; Sarah, R.G.L.; Amanda, C.S.M.; Jessica, S.R.S.; Jackson, R.G.S.A. In vitro antioxidant and photoprotective activities of dried extracts from Neoglaziovia variegata (Bromeliaceae). *J. App. Pharm. Sci.* 2013. 3(1), 122-127.
- Rajha,H.N.; Darra,N.E.; Hobaika,Z.; Boussetta,N.; Vorobiev,E.; Maraoun,R.G.; Louka, N. Extraction of total phenolic compound, flavonoids, anthocyanins, and tannins from grape byproducts by response surface methodology. Influence of Solid-Liquid ratio, particle size, time, temperature and solvent mixtures on the optimization process. *Food Nutr.Sci.* 2014. 5, 397-409.
- Rasanu, N.; Magearu, V.; Matei, N.; Soceanu, A. *analele Universitatii din Bucuresti.* 2005. I (2), 167-172.
- Riaz, T.; Abbasi, M.A.; Rehman, A.U.; Shahzadi,T.; Qureshi, M.Z.; Khan, K.M. Antioxidant activity and radical scavenging effects of various fractions from curcuma zedoaria. *Asian J. Pharma. Biol. Res.* 2011. 4, 525-533.
- Roseiro, L.B.; Tavares, C.S.; Roseiro, J.C.; Rauter, A.P. Antioxidants from aqueous decoction of carob pods biomass (*Ceretonia siliqua* L.): Optimisation using response surface methodology and phenolic profile by capillary electrophoresis. *Ind. Crops Prod.* 2013. 44, 119–126.
- Roslan, J.; Mustapa, K.S.M.; Yunos, M.D.; Abdullah N. Optimization of Enzymatic Hydrolysis of Tilapia Muscle (*Oreochromis niloticus*) using Response surface methodology (RSM). *Sains Malaysiana.* 2014. 43(11), 1715-1723.
- Rutkowski, M.; Grzegorczyk, K. Modifications of spectrophotometric methods for antioxidative vitamins determination convenient in analytic practice. *Acta Sci. Pol. Techno. Aliment.* 2007. 6 (3), 17-28.
- Saeed, N.; Kham, M.R.; Shabbir, M.. Antioxidant activity, total phenolic and total flavonoid content of whole plant extracts *Torilis leptophylla* L. *BMC complement. Altern. Med.* 2012. 12, 221.
- Sahu, R.; Saxena,J. Phytochemical Analysis of Curcuma Amada by FTIR and UV-VIS Spectroscopic Analysis. *Int. J. Pharm. Res. Sch.* 2015.4, 458-461.
- Sai-Ut, S.; Benjakul, S.; Kraithong, S.; Rawdkuen, S. Optimization of antioxidants and tyrosinase inhibitory activity in mango peels using response surface methodology. *LWT - Food Sci. Technol.* 2015. 64, 742–749.

- Samuagam, L.; Cm, S.; Ga, A.; Pn, O.; Hs, Y. The Effect of Extraction Conditions on Total Phenolic Content and Free Radical Scavenging Capacity of Selected Tropical Fruits' Peel. *Heal. Environ. J.* 2013. 4, 80–102.
- Sasidhran,S.; Chen,Y.; Saravanan,D.; Sundram,K.M.; Latha,Y.L. Extraction, isolation, and characterization of bioactive compounds from plants extracts. *Afr J. Tradit. Complement. Altern. Med.* 2011. 8(1), 1-10.
- Sekhar, T.C.; Anju.G. Antioxidant activity by DPPH Radical Scavenging Method of Ageratum conyzoides Linn.Leaves. *Am. J. Etnomedicine.* 2014. 1(4), 244-249.
- Shahriar,M. Antimicrobial activity of the rhizomes of curcuma zedoaria. *J. Bangladesh Acad. Sci.* 2010. 34, 201-203.
- Sheng, Z.L.; Wan, P.F.; Dong, C.L.; Li, Y.H. Optimization of total flavonoids content extracted from Flos Populi using response surface methodology. *Ind. Crops Prod.* 2013. 43, 778–786.
- Singh, B.S.; Priya, K.R.; Rojita, M.; Amrita, P.; Praveen, B. Antimicrobial properties of few plants used in traditional system of medicine. *Int.J.Res. Ayurveda Pharm.* 2012. 3(4), 563-564.
- Singleton,V.L.; Rossi,J.A. Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagent. *Am.J.Enol.Vitic.* 1965. 16, 144-153.
- Solomon-Wisdo, G.O.; Ugoh, S.C.; Mohammed, B. Phytochemical screening and antimicrobial activities of Annona muricata (L) leaf extract. *Am. J. Biol. Chem. Pharma.Sci.* 2014. 2(1), 1-7.
- Soni Himesh.; Patel, S.S.; Mishra K.; Nayak, G.; Singhai,A.K. Qualitative and quantitative profile of curcumin from ethanolic extract of curcuma longa. *Int.Res. J. Pharm.* 2011. 2, 180-184.
- Soo, E.; Salleh, A.; Basri, M.; Rahman, R.; Kamaruddin, K. Response surface methodological study on Lipase-catalyzed synthesis of Amino acid surfactants. *J. process Biochem.* 2004. 39,1511-1518.
- Srivastava,S.; Mehrotra,S.; Rawat, A.K.S. Pharmacognostic evaluation of the rhizomes of curcuma zedoaria Rosc. *Pharmacog. J.* 2011. 3(21), 18-24.
- Stankovic, M. Total phenolic content, flavonoid concentration and antioxidant activity of Marrubium peregrinum L. extracts. *Kragujevac J. Sci.* 2011. 33, 63-72.
- Suva, M.A. Evaluation of Sun Protection Factor of ZIngiber Officinale Roscoe Extract by Ultraviolet Spectroscopy Method. *J. Pharma. Sci. Tech.* 2014. 3(2), 95-97.
- Taheri, S.; Abdullah, T.L.; Karimi, E.; Oskoueian, E.; Ebrahimi, A.M. Antioxidant capacities and total phenolic contents enhancement with acute Gamma irradiation in Curcuma alismatifolia (Zingiberaceae) leaves. *Int. J. Mol. .Sci.* 2014. 15, 13078-13090.

- Tan, M.C.; Tan, C.P.; Ho, C.W. Effects of extraction solvent system, time and temperature on total phenolic content of henna (*Lawsonia inermis*) stems. *Int. Food Res. J.* 2013, 20, 3117–3123.
- Tang, Y.; Cai, W.; Xu, B. Profiles of phenolics, carotenoids and antioxidant capacities of thermal processed white, yellow, orange and purple sweet potatoes grown in Guilin, China. *Food Sci. Hum. Wellness.* 2015, 4, 123-132.
- Taofiq, O.; Gonzalez-Paramas, A.M.; Martins, A.; Barreiro, M.F.; Ferreira, I.C.F.R. Mushrooms extracts and compounds in cosmetics, cosmeceuticals and nutricosmetics-A review. *Ind. Crops. Prod.* 2016, 90, 38-48.
- Taxonomic classification of *Curcuma zedoaria*. USDA, NRCS. 2017. The plants database Retrieved 30 January 2017 from <http://plants.usda.gov/java/>
- Tepsorn Racha. Antimicrobial activity of Thai Traditional Medicinal plant extract incorporated Alginate-tapioca Starch based edible Films against food related bacteria including Foodborne pathogens. *PhD Thesis.* University of Hohenheim, Thailand. 2009.
- Tholkappiyavathi, K.; Selvan, K.M.; Neyanila, S.K.; Yoganandam, G.P.; Gopal, V. A Concise Review on Curcuma Zedoaria. *Int. J. Phyther.* 2013. 2, 1–4.
- Vaibhaz, S.; Lakshman. K. Tyrosinase enzyme inhibitory activity of selected Indian Herbs. *Int. J. Res. Pharma. Biomed. Sci.* 2012. 3(3), 977-982.
- Vajić, U.J.; Grujić-Milanović, J.; Živković, J.; Šavikin, K.; Gođevac, D.; Miloradović, Z.; Bugarski, B.; Mihailović-Stanojević, N. Optimization of extraction of stinging nettle leaf phenolic compounds using response surface methodology. *Ind. Crops Prod.* 2015.74, 912–917.
- Vardhan, A.; Khan, S.; Pandey, B. Screening of plant parts for anti-tyrosinase activity by tyrosinase assay using mushroom tyrosinase. *Indian J. Sci. Res.* 2014. 4(1), 134-139.
- Varzakas, T.; Kiokias,S.; HPLC analysis and determination carotenoid pigment in commercially available plant extracts. *Curr. Res. Nutr. Food Sci.* 2016. 4(1), 1-14.
- Vázquez,G.; Fernández-Agulló,A.;Gómez-Castro,C.;Freire,M.S.;Antoreanna,G.;González-Álvarez,J. Response surface optimization of antioxidants extraction from chesnut (*Castanea sativa*) bur. *Ind. Crops Prod.* 2012. 35, 126-134.
- Wang, L.; Liu, Y. Optimization of solvent extraction conditions for total carotenoids in rapeseed using response surface methodology. *Nat. Sci.* 2009. 1, 23–29.
- Wani, S.M.; Jan, N.; Wani, T.A.; Ahmad, M.; Masoodi, F.A.; Gani,A. Optimization of antioxidant activity and total polyphenols of dried apricot fruit extracts (*Prunus armeniaca L.*) using response surface methodology. *J. Saudi Soc. Agric. Sci.* 2015. Article in press

- Wilson, B.; Abraham, G.; Manju, V.S.; Mathew, M.; Vimala, B.; Sundaresan, S.; Nambisan, B. Antimicrobial activity of Curcuma zedoaria and Curcuma malabarica tubers. *J. Ethnopharm.* 2005. 99, 147-151.
- Wojdylo A.; Oszmianski J.; Czemerys, R. Antioxidant activity and phenolic compounds in 32 selected herbs. *Food Chem.* 2007. 105, 940-949.
- Xu, P.; Bao, J.; Gao, J.; Zhou, T.; Wang, Y. Optimization of extraction of phenolic antioxidants from tea (*camellia sinensis* L.) fruit peel biomass using response surface methodology. *Bioresources*. 2012. 2, 2431-2443.
- Yaar,M.,Eller,M.S.,and Gilchrest,B.A.,(2002).Fifty Years of Skin Aging. The Society for Investigative Dermatology,Inc,7,51-58.
- Yadaz, N.; Yadaz, R.; Goyal, A. Chemistry of terpenoids. *Int. J. Pharm. Sci. Rev. Res.* 2014. 27(2), 272-278.
- Yilmaz, Y.; Toledo, R.T. Oxygen radical absorbance capacities of grape/wine industry byproducts and effect of solvent type on extraction of grape seed polyphenols. *J. Food Compos. Anal.* 2006. 19, 41-48.
- Yoswathana, N. Optimization of ScCO₂ Extraction of Rambutan Seed Oil Using Response Surface Methodology. *Int. J. Chem. Eng. Appl.* 2013. 4, 187-190.
- You, D.H.; Park, J.W.; Yuk,H.G.; Lee, S.C. Antioxidant and tyrosinase inhibitory activities of different parts of guava (*psidium guavaja* L.). *Food Sci. Biotechnol.* 2011. 20(4), 1095-1100.
- Zaveska, E.; Fer, T.; Sida, O.; Krak, K.; Marhold, K.; Skornickova, J.L. Phylogeny of Curcuma (Zingiberaceae) based on plastid and nuclear sequences: proposal of the new subgenus. *Ecomata. Taxon.* 2012. 61, 747-763.
- Zhang, L.; Ravipati, A.S.; Kooyalamudi, S.R.; Jeong, S.C.; Reddy, N.; Bartlett, J.; Smith, P.T.; de la Cruz, M.; Monteiro, M.C.; Melguizo, A.; Jimenez, E.; Vicente, F. Anti-fungal and anti-bacterial activities of ethanol extracts of selected traditional Chinese medicinal herbs. *Asian Pac.J. Trop.Med.* 2013. 673-681.
- Zhao, L.C.; Liang, J.; Li, W.; Cheng, K.M.; Xia, X.; Deng, X.; Yang, G.L. The use of response surface methodology to optimize the ultrasound-assisted extraction of five anthraquinones from rheum palmatum L. *Molecules*. 2011. 16, 5928-5937.