

UNIVERSITI PUTRA MALAYSIA

EFFECTS OF TOPICAL APPLICATION OF MIXTURES OF MORINDA CITRIFOLIA L., MELASTOMA MALABATHRICUM L., AND LAWSONIA INERMIS L. ETHANOLIC EXTRACTS ON EXCISION WOUND IN SPRAGUE DAWLEY RATS

ALI KHAIRULLAH ZAHI ALSAEED

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By

ALI KHAIRULLAH ZAHI ALSAEED

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirement for the Degree of Master of Science

January 2015

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Abstract of thesis presented to the senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

EFFECTS OF TOPICAL APPLICATION OF MIXTURES OF Morinda citrifolia L., Melastoma malabathricum L., and Lawsonia inermis L. ETHANOLIC EXTRACTS ON EXCISION WOUND IN SPRAGUE DAWLEY RATS

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January 2015

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Faculty: Veterinary Medicine

The eligibility of attempting to repair excision wounds has long-term been appreciated since the last century, at least in the sensation of providing a spotless wound, avoiding formalization of purulent and pulverization as well as to boost a granulation and reepithelisation. It has been a particular vision in medical practice as excision wound denatures cellular protein, inhibits cellular metabolism hence secondary interference of local vascular supply. Therefore, the present study was designed to investigate the effects of the mixture of ethanolic extracts of *Morinda citrifolia* fruits, *Melastoma malabathricum* leaves and *Lawsonia inermis* leaves on the excision wound healing as these natural herbs have been traditionally used in excision wound treatment. Prior to wound healing experiment, the dermal toxicity levels of each herbal extract were determined via dermal toxicity experiments using the Organization for Economic Cooperation and Development (OECD) standard guidelines.

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Results showed that the lethal dose 50 (LD₅₀) of each herb was more than 5000 mg/kg body weight, while the no observed adverse effect level (NOAEL) for each extract was more than 2000 mg/kg body weight. Based on the dermal toxicity results, mixtures of the herbal extracts at 0.5% and 1% were selected for treatment in the wound healing study. An area of uniform wound 2 cm in diameter was inflicted on 120 adult male Sprague Dawley weighing between 250-350 g. The animals were divided into five groups with six animals in each group, representing a control and experimental groups. Mixture of the herbs at 0.5%, mixture of the herbs at 1%, silver sulphadiazine as a standard treatment and paraffin were applied once daily, except in the control group where wounds were left without any topical treatment.

The rats were closely monitored to assess any changes. The rats were euthanized at 4, 8, 12 and 21 days post wounding. The macroscopic appearances of excision wounds were evaluated and recorded. The percentages of wound contraction, wound size and wound epithelisation were measured and analyzed. In addition, histopathological examination of the skin was also performed qualitatively and quantitatively using haematoxylin and eosin (H&E), van Gieson and immunohistochemistry staining methods.

Results obtained from this study revealed that mixture of three herbs at 0.5% showed advanced effects to decrease the period of wound healing process, wound contraction and wound size as compared to the other experimental groups. Quantitative evaluation of the number of inflammatory cells (polymorphonuclear cells and macrophages) from day 4 to day 21 in wounds treated with mixture of herbs at 0.5% demonstrated significant (p < 0.05) decrease in number of inflammatory cells (fibroblasts) increased from day 4 to day 21, while the number of proliferative cells (fibroblasts) increased from day 8 to day 21. Mixture of the herbs at 0.5% also showed potential to preserve viable dermal tissues and induce a well-formed of angiogenesis with better organisation as compared to the other experimental groups.

On the other hand, the re-epithelisation and level of collagen formation was increased gradually in group that received mixture of herbs at 0.5% compared to the other groups. Qualitative and quantitative evaluation of the expression of vascular endothelial growth factor (VEGF), transforming growth factor alpha (TGF α) and transforming growth factor beta (TGF β) proteins in wounds treated with mixture of herbs at 0.5% and 1% recorded gradual rise in the expression of these three growth factors in wound area. In conclusion, the present study showed that mixture of the three herbs at 0.5% had the most superior treatment agent for excision wounds fallowed by mixture of the herbs at 1% in relation to the macroscopic and histopathological evaluation.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

KESAN PENGGUNAAN TOPIKAL CAMPURAN EKSTRAK ETANOL Morinda citrifolia L., Melastoma malabathricum L., DAN Lawsonia inermis L. KE ATAS LUKA EKSISI PADA TIKUS SPRAGUE DAWLEY

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Kesesuaian dalam cubaan untuk merawat luka eksisi telah lama diberi penghargaan semenjak seabad yang lalu, sekurang-kurangnya dari sudut memastikan luka tiada parut, mengelakkan penghasilan dan pengumpulan nanah, dan juga untuk merangsang pembetukan granul dan epitelium. Adalah menjadi satu prinsip dalam amalan perubatan yang mana luka eksisi menyebabkan penguraian protin sel dan merencat metabolisma sel dan menyebabkan gangguan kedua kepada bekalan darah setempat. Oleh itu, kajian ini telah dibuat untuk menyiasat kesan campuran ekstrak etanol buah *Morinda citrifolia*, *Melastoma malabathricum* dan *Lawsonia inermis* ke atas penyumbuhan luka eksisi berikutan herba asli ini telah digunakan secara tradisional dalam rawatan luka eksisi. Sebelum ekperimen penyumbuhan luka dilaksanakan, aras toksisiti kulit bagi setiap ekstrak herba ditentukan dengan mejalankan eksperimen toksisiti kulit berpandukan garis panduan Pertubuhan Kerjasama dan Pembangunan Ekonomi (OECD).



Keputusan menunjukkan bahawa Dos Maut 50 (LD_{50}) bagi setiap herba adalah melebihi 5000 mg/kg berat badan, sementara aras kesan teruk yang tidak nampak (NOAEL) bagi setiap herba ialah melebihi 2000 mg/kg berat badan. Berdasarkan ujian tosisiti kulit ini, campuran tiga jenis herba pada 0.5% dan 1% telah dipilih dalam kajian merawat penyumbuhan luka. Satu kawasan luka samarata bergaris pusat 2 cm telah dibuat pada 120 tikus jantan Sprague Dawley dengan berat badan antara 250-350g. Tikus tersebut dibahagikan kepada lima kumpulan dengan enam ekar tikus dalam setiap kumpulan, mewakili kumpulam kawalan dan rawatan. Campuran herba pada 0.5%. campuran pada 1%, sulfadiazin sulfur sebagai rawatan standard dan paraffin disapukan sekali sehari, kecuali kumpulan rawatan dimana luka dibiarkan tanpa sebarang rawatan setempat.

Tikus tersebut diperhatikan secara dekat untuk menilai sebarang perubahan. Tikus tersebut dimatikan pada hari ke 4, 8, 12 dan 21 selepas dilukakan. Penampilan luka eksisi dinilaikan dan dicatatkan. Peratus kontraksi luka, saiz luka dan luka epithelisation

pembentukan diukur dan dianalisa. Sebagai tambahan, pemeriksaan histopatologi ke atas kulit dibuat secara kualitatif dan kuantitatif menggunakan kaedah perwarnaan hematosilin dan eosin (H&E), van Gieson dan imunohistokima.

Keputusan yang diperolehi daripada kajian ini menunujukkan campuran ketiga-tiga herba pada 0.5% memberi kesan bagus bagi mengurangkan tempoh masa pemyumbuhan luka., kontraksi luka dan saiz luka berbanding dengan kumpulan eksperimen yang lain. Penilaian kuantitatif ke atas jumlah sel inflamasi (sel polimorfonuklear dan makrofaj) daripada hari ke 4 hingga hari ke 21 dalam rawatan luka menggunakan campuran herba pada 0.5% menunjukkan penurunan seerti (p < 0.05) sel inflamasi daripada hari ke 4 hingga hari ke 21, sementara jumlah sel proliferative (fibroblast) meningkat daripada hari ke 8 hingga hari ke 21. Campuran herba pada 0.5% menunjukkan potensi untuk mengutuhkan tisu kulit hidup dan merangsang pembentukkan salur darah baharu yang sempurna dengan orgnisasi yang lebih baik berbanding kumpulan eksperimen yang lain.

Dalam pada itu, pembentukkan semula epitelium dan aras pembentukkan kolagen adalah meningkat secara gradual pada kumpulan yang menerima campuran herba pada 0.5% berbanding kumpulan lain. Penilaian kualitatif dan kuantitatif ke atas ekspresi protin factor tumbesaran endotelial vascular (VEGF), faktor tumbesaran perubahan alpha (TGF α) dan faktor tumbesaran perubahan beta (TGF β) pada luka yang dirawat dengan herba pada 0.5% dan 1% menunjukkan peningkatan secara gradual ekspresi protin bagi ketiga-tiga faktor tumbesaran tersebut pada kawasan luka. Kesimpulannya, kajian ini menunjukkan bahawa campuran ketiga-tiga herba pada 0.5% memberikan agen rawatan yang paling bagus untuk luka eksisi diikuti dengan campuran herba pada 1% berdasarkan penilaian makroskopik dan histopatologi.

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"In the name of Allah, the most beneficent and the most merciful"

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I certify that a Thesis Examination Committee has met on 16 Januari 2015 to conduct the final examination of Ali Khairullah Zahi on his thesis entitled "Effects of Topical Application of Mixtures of *Morinda citrifolia* L., *Melastoma malabathricum* L., and *Lawsonia inermis* L. Ethanolic Extracts on Excision Wound in Sprague Dawley Rats" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U. (A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

mm	Micrometer
LDH	Lactate dehydrogenase
ALT	Alanine transaminase
AST	Aspartate transaminase
H&E	Hematoxylin and eosin
SSD	Silver sulphadiazine
G	Gram
°C	Celsius degree
EDTA	Ethylene diamine tetra acetic acid
PCV	Packed cell volume
RBC	Red blood cell
SD	Standard deviation
Cm	Centimetre
WBC	White blood cell
Kg	Kilogram
mg	Milligram

CHAPTER 1

INTRODUCTION

Wound is a physical injury which leads to breakage of the skin. Wound healing is a natural response to tissue injury which consists of complex and sophisticated cascade of events involving various cellular, molecular and biochemical processes and resulting in the healing of the wound and the restoration of the intact functional barrier (Kondo, 2007). Generally, wound healing process is categorised into three integrating and interfering phases: 1) the inflammatory phase, which is the first phase that consists of the establishment of homeostasis and inflammation, 2) the proliferative phase, which is the second phase that consists of granulation, contraction, and epithelialisation, and 3) the remodelling or maturation phase, which is the final phase of wound healing process that eventually determines the strength and appearance of the healed tissue (Mantle *et al.*, 2001).

Plant extracts have been used as a wound healing agents for long time (Wang *et al.*, 2002). The use of traditional medicinal remedies and plants in the treatment of burns and wounds is viewed as an essential mode to improve healing processes and also to reduce the financial burden especially in the economically deprived societies of the developing world. Several plants and herbs have been used experimentally to treat skin disorders such as wound injuries in traditional medicine (Mahmood *et al.*, 2009). There are a lot of studies have been done on natural herbs and plants for wound treatment. Most of the natural herbs and plants that have been studied contain active compounds like triterpenes, alkaloids, flavonoids and other biomolecules, that reported assisted in boosting and promoting the process of wound healing via influencing one or more of the phases of the healing process (Phillipson, 2001; Soma *et al.*, 2012). It is reported that extracts of the medical plants and herbal as well as natural compounds involve in different and a few stages of wound healing which include coagulation, inflammation, fibroplasia, collagenation, epithelisation and wound contraction (Hemmati and Mohammadian, 2000).

In this study, three herbal extracts namely *Morinda citrifolia* fruits, *Melastoma malabathricum* and *Lawsonia inermis* leaves were investigated for their effectiveness in accelerating wound healing process. *M. citrifolia* is one of the most significant herbs that used in wound healing researches. All findings of the studies reported that *M. citrifolia* reduced the period of wound healing via reducing the period of inflammation phases, increasing the rate of cells proliferation and increase the level of epithelisation (Shivananda Nayak *et al.*, 2007; Vijaykumar *et al.*, 2008; Afa Palu *et al.*, 2010). *M. malabathricum* is another important plant used in wound healing researches, especially in Malaysia. There are various studies done on *M. malabathricum* on the wound healing activities of this herb. A few studies recorded that *M. malabathricum* reduced the time of bleeding and repair of scar formation (Sulaiman *et al.*, 2004; Sunilson *et al.*, 2008; Manicam *et al.*, 2010; Choudhury *et al.*, 2011; Nurdiana and Marziana, 2013).



L. inermis is also one of the significant herbs employed in traditional medicine for wound treatment. A study conducted on the effects of *L. inermis* on wound healing activities indicated that *L. inermis* reduced time of inflammatory phase of wound healing, increase epithelisation and scar formation (Sakarkar *et al.*, 2004).

Problem statement

Recently, the necessity for the improvement of treatment of wound healing using natural herbs instead of using chemicals has been increased. This is attributed to the side effects of using chemical treatment, difficulties of preparing chemicals and the expenses. Although numerous studies have been conducted on wound healing using natural herbs including *M. citrifolia*, *M. malabathricum* and *L. inermis*, none of the studies investigated the effects of mixtures of these three herbs in accelerating wound healing process.

Hypothesis

It is hypothesised that mixtures of the three herbs will increase the activity of wound healing in rats.

Objectives

The aim of this study was to evaluate the effects of topical application of mixtures of *M*, *citrifolia*, *M*. *malabathricum*, and *L*. *inermis* on excision wound healing process in male Sprague Dawley rats.

The specific objectives of this study were:

- 1. to investigate the acute dermal toxicity of ethanolic extracts of *M. citrifolia* fruits, *M. malabathricum* leaves, and *L. inermis* leaves at doses of 2000 mg/kg and 5000 mg/kg body weight in female rats,
- 2. to investigate the sub-acute dermal toxicity of ethanolic extracts of *M. citrifolia* fruits, *M. malabathricum* leaves, and *L. inermis* leaves at doses of 500 mg/kg, 1000 mg/kg, and 2000 mg/kg body weight in male rats, and
- 3. to evaluate the effectiveness of mixtures of the herbal extracts at different concentrations for topical treatment of excision wounds via macroscopic, microscopic examination.

REFERENCES

- Adeneye, A.A., Ajagbonna, O.P., Adeleke, T.I., Bello, S.O. (2006) Preliminary toxicity and phytochemical studies of the stem bark aqueous extract of Musanga cecropioides in rats. J. Ethnopharmacol 105, 374-379.
- Afa Palu, Chen Su, Bing-Nan Zhou, Brett West, and Jarakae Jensen. (2010) Wound Healing Effects of Noni (Morinda citrifolia L.) Leaves: A Mechanism Involving its PDGF/A2A Receptor Ligand Binding and Promotion of Wound Closure. Phytother. Res. 24: p.1437-1441.
- Aghel N, Ameri A, Ebrahimi P. (2005) Essential oil of Lawsonia inermis growing in iran: chemical composition and antifungal activity. First Seminar of Medicinal & Natural Products Chemistry Shiraz, Iran. May 10-11.
- Ajagbonna, O.P., Onifade, K.I., Suleiman, U. (1999) Haematological and biochemical changes in rats given extract of C. pocera. Sokoto. J. Vet. Sci. 1, 3-12.
- Akanmu, M.A., Iwalewa, E.O., Elujoba, A.A., Adelusola, K.A. (2004) Toxicity potentials of Cassiafistula fruits as laxative with reference to senna. Afr. J. Biomed. Res. 7, 23-26.
- Alade, G.O., Akanmu, M.A., Obuotor, E.M., Osasan, S.A., Omobuwajo, O.R. (2009) Acute and oral sub-acute toxicity of methanolic extract of Bauhinia monandra leaf in rats. Afr. J. Pharm. Pharmacol3, 354-358.
- Alen, Y., Nakajima, S., Nitoda, T., Baba, N., Kanzaki, H., andKawazu, K. (2000) "Antinematodal activity of some tropical rainforest plants against the pinewood nematode Bursaphelenchus xylophilus," Zeitschrift fur Naturforschung C, vol. 55, no. 3-4, pp. 295-299.
- Aljady, A.M., Kamaruddin, M.Y., Jamal, A.M., and Mohd.yassim, M.Y. (2000) Biochemical study on the efficacy of Malaysian honey on inflicted wounds: an animal model. Medical Journal of Islam Academy of Sciences13(3), 125-132.
- Aljady, A.M.A. (2003) Biochemical study on the wound healing property of honey. PHD thesis. University of Malaya, Kuala Lumpur.
- Ali, N.A.A., Julich, W.D., Kusnick, C., Lindequist, U. (2001) Screening of Yemeni medicinal plants for antibacterial and cytotoxic activities. J. Ethnopharmacol74(2):173-179.
- Alia, B.H., Bashir, A.K., Tanira, M.O.M. (1995) Anti-inflammatory, antipyretic and analgesic effects of Lawsonia inermis L. (henna) in rats. Pharmacol 51:356-363.
- Anwar, M.N., Begum, J., Yusuf, M., Chowdhury, J.U., Khan, S., Nural, M. (2007) Antifungal activity of forty higher plants against phytopathogenic fungi. Bangladesh J Microbiol 24(1):76-78.
- Arayne, M.S., Sultana, N., Mirza, A.Z., Zuberi, M.H., Siddiqui, F.A. (2007) In vitro hypoglycemic activity of methanolic extract of some indigenous plants. Pak J Pharm Sci 20(4):268-273.

- Asahina, A.Y., Ebesu, J.S.M., Ichinotsubo, D., Tongson, J., Hokama, Y. (1994) Effect of okadaic acid (OA) and Noni fruit extraction in the synthesis of tumor necrosis factor-a (TNF-a) by peripheral blood mononuclear (PBN) cells in vitro. The Proceedings of the International Symposium of Ciguatera and Marine Natural Products. p 197-205.
- Asante-Duah, K. (2002) Public Health Risk Assessment for Human Exposure to Chemicals (illustrated.); Kluwer Academic Publishers: Dordrecht, The Netherlands. Volume 6.
- Atkinson, N. (1956) Antibacterial substances from flowering plants. 3. Antibacterial activity of dried Australian plants by rapid direct plate test. Australian Journal of Experimental Biology 34, 17–26.
- Attinger, C.E., Janis, J.E., Steinberg, J. et al., (2006) Clinical approach to wounds: debridement and wound bed preparation including the use of dressings and wound-healing adjuvants. Plast Reconstr. Surg 117(7 suppl): 72S-109S.
- Baba-Moussa, F., Nacoulma, O., Ouattara, A., Nguyen, H.P., Akpagana, K., Bouchet, P. (1997) Antibacterial activity of total aqueous extracts of Combretum micranthum, Lawsonia inermis and Waltheria indica, plants from west African pharmacopoeia. Revue de Medecines et Pharmacopees Africaines11-12:197-203.
- Bagi, M.K., Kakrani, H.K., Kalyani, G.A., Dennis, T.J., Jagdale, M.H. (1988) Experimental evaluation of pharmacological activity of Lawsonia alba seed oil. Fitoterapia 59(1):39-42.
- Baie, S.H. and Sheikh, K.A. (2000) The wound healing properties of Channa striatus cetrimide cream tensile strength measurement. Journal of Ethnopharmacology 71(1), 93-100.
- Ballard, K. and Bazter, H. (2000) Development in wound care for difficult manage wounds. Br. J. Nursing 9(7) 405-412.
- Bischoff, M., Kinzl, L., Schmelz, A. (1999) The complicated wound. Unfallchirurg 102: 797-804 [in German].
- Brannon, H.L. (2007) Skin anatomy. Retrieved on 30 October 2010 from http://dermatology.about.com/cs/skinanatomy.html.
- Brown, L.F., Yeo, K.T., Berse, B., Yeo, T.K., Senger, D.R., Dvorak, H.F. andvande water, L. (1992) Expression of vascular permeability factor (vascular endothelial growth factor) by epidermal keratinocytes during wound healing. J Exp Med 176: 1375-1379.
- Cardon, D. (2003). Le Monde des Teintures Naturelles. Belin, Paris. *Chemistry*,49, 4478-4481.
- Carol, S.A. (1995) Acute, Sub-chronic and Chronic Toxicology. In CRC Handbook of Toxicology; Michael, J.D., Mannfred, A.H., Eds. CRC Press Inc. Boca Raton, FL, USA 51-104.
- Chan-Blanco, Y., Vaillant, F., Perez, A.M., Reynes, M., Brillouet, J.M. and Brat, P. (2006) The noni fruit (Morinda citrifolia L.): A review of Agricultural research,

nutritional and therapeutic properties, Journal of Food Composition Analysis, Vol.19, pp. 645-654, ISSN 0889-1575.

- Chauhan, M.G., Pillai, A.P.G. (2007) Microscopic profile of powdered drug used in Indian system of medicine, Edn 1, Vol. 2, Gujarat Ayurved University, Jamnagar, Gujarat pp. 84-85.
- Chetty, K.M. (2008) Flowering plants of Chittoor, Edn 1, Andhra Pradesh pp. 132.
- Chopra, R.N., Nayer, S.L., Chopra, I.C. (1956) Glossary of India Medicinal Plants, CSIR Publications, New Delhi pp. 151.
- Choudhury, M.D., Nath, D., Talukdar, A.D. (2011) Antimicrobial activity of Melastoma malabathricum L. Assam University Journal of Science & Technology, 7, 76-78.
- Chunhieng, M.T. (2003) De'veloppement de nouveaux aliments sante tropicale: application a` la noix du Bre'sil Bertholettia excelsa et au fruit de Cambodge Morinda citrifolia. Ph.D. thesis, INPL, France.
- Clark, R.A.F. (1985) Cutaneous tissue repair: basic biologic concedarations. I.J.Am Acad Dermal 13: 701-725.
- Clark, R.A.F (1996) Wound repair: overview and general conciderations. In: Clark, R.A.F, (editor) The molecular and cellular biology of wound repair (2nd ed). London: Plemum Press. 3-50.
- Coffey RJ, J.R., Bascom, C.C., Sipes, N.J., Graves-deal, R., Weissman, B.E. and Moses, H.L. (1988) Selective inhibition of growth-related gene expression in murine keratinocytes by transforming growth factor. Mol Cell Biol 8: 3088-3093.
- Dama, L.B., Poul, B.N., Jadhav, B.V. (1999) Antimicrobial activity of Napthoquinonic compounds. Journal of Ecotoxicology and Environmental Monitoring 8:213-215.
- Davidson, J.M. (1998). Animal models for wound repair. Arch Dermatol Res. 290 (Suppl): S1-S11.
- Davis, M., Dunkley, P., Hareden, R.M., Harding, K., Laidlaw, J.M., Moris, R.m. and wood, R.A.B. (1992) The wound programme. Center for Medical Education. Dundee.
- Davis, S.C. and Perez, R. (2009) Cosmeceuticals and natural products: wound healing. Clinics in dermatology 27, 502-506.
- Desmouliere, A., Geinoz, A., Gabbiani, F. and Gabbiani, G. (1993) Transforming growth factor-beta 1 induces alpha-smooth muscle actin expression in granulation tissue myofibroblasts and in quiescent and growing cultured fibroblasts. J Cell Biol 122: 103-111.
- Dikshit, V., Dikshit, J., Saraf, M., Thakur, V., Sainis, K. (2000) Immunomodulatory activity of naphthoquinone fraction of Lawsonia inermis Linn. Phytomedicine (Jena) 7:102 103.

- Dittmar, A. (1993) *Morinda citrifolia* L, Use in indigenous Samoan medicine. Journal of Herbs, Spices and Medicine Plants 1, 77-92.
- Diwan, P.V., Tillo, L.D., Kulkarni, D.R. (1982) Influence of Tridax procumbens on wound healing. Indian J Med Res 75: 460-464.
- Dixit, S.N., Srivastava, H.S., Tripathi, R.D. (1980) Lawsone, the antifungal antibiotic from the leaves of *Lawsonia inermis* and some aspects of its mode of action. Indian Phytopathol 31:131-133.
- Dixon, A.R., McMillen, H., Etkin, N.L. (1999) Ferment this: the transformation of Noni, a traditional Polynesian medicine (Morinda citrifolia, Rubiaceae). Ecological Botony 53, 51-68.
- Dorsett-Martin, W.A. (2004) Rat models in skin wound healing: a review. Wound Rep Reg 12: 591-599.
- Dybing, E., Doe, J., Groten, J., Kleiner, J., O'Brien, J. (2002) Hazard characterization of chemicals in food and diet: dose response, mechanism and extrapolation issues. Food Chem. Toxicol 42, 237-282.
- Dyson, M., Young, S., Pendle, C.L., Webster, D. F. and Lang, S.M. (1988). Comparison of the effects of moist and dry conditions on dermal repair. Journal of investigative dermatology, 91(5), 434-439.
- Eaglastein, W. (1989) Wound healing and aging. Clin. Geriat. Med 5(1).183-188.
- Eaton, D.L., Klaassen, C.D. (1996) Principles of toxicology. In Casarett and Doull's Toxicology: The Basic Science of Poisons, 5th ed; Klaassen, C.D. Ed.; McGraw-Hill: New York, NY, USA p. 13.
- Elkins, R. (1998) Hawaiian Noni (Morinda citrifolia) Prize Herb of Hawaii and the South Pacific. Woodland Publishing, Utah.
- Enoch, S. and Leaper, D.J. (2007) Basic science of wound healing. Surgery, 2-7.
- Frank, S., Hubner, G., Breier, G., Longaker, M.T., Greenhalgh, D.G. and Werner, S. (1995) Regulation of vascular endothelial growth factor expression in cultured keratinocytes: implications for normal and impaired wound healing. J Biol Chem 270: 12607-12613.
- Friedrich, L. (2010) Nutrition and wound healing: from kitchen to the pharmacy. Retrieve from http://www.anfponline.org/Events/11Annual-handouts/lfriedrichnutritionWoundHealing.pdf.
- Gabrie, A., Mussman, J., Rosenberg, L.Z. and Torre, J.I.d.I. (2009) Wound Healing, Growth Factors (Publication Retrived 25 February 2009 from eMedicine from web MD:http:/emedicine.medscape. Com/article/1298196-overview.
- Gal, P., Kilik, R., Mokry, M., Vidinsky, B., Valsilenko, T., Mozes, S., Bobrov, N., Tomori, Z., Bober, J., Lenhardt, L. (2008) Simple method of open skin wound healing model in corticosteriod treated and diabetic rats: standardization of semi quantitative and quantitative histological assessments. Veterinary Medicina: 53 (12): 652-59.

- Gailit, J., Welch, M.P., and Clark, R.A. (1994) TGF-beta 1 stimulates expression of keratinocyte integrins during re-epithelialization of cutaneous wounds. J. Invest Dermatol 103: 221–227.
- Ghosh, S., Samanta, A., Mandal, N.B., Bannerjee, S. and Chattopadhyay, D. (2012) Evaluation of the wound healing activity of methanol extract of Pedilanthus tithymaloides L. Poit leaf and its isolated active constituents in topical formulation. Journal of ethnopharmacology 142(3), 714-722.
- Glerup, P., TAHITIAN, T.N.J. (2001) A 13-week oral (gavage) toxicity study in rats. Cantox Biologisk Laboratorium A/S, DK-426. Lille Skensved, Denmark.
- Gottrup, F. (2001) Experimental wound healing research: the use of models. EWMA Journal, 2:5-8.
- Grosvenor, P.W., Gothard, P.K., McWilliam, N.C., Supriono, A. and Gray, D.O. (1995) "Medicinal plants from Riau Province, Sumatra, Indonesia. Part 1: uses," Journal of Ethnopharmacology, vol. 45, no. 2, pp. 75-95.
- Gupta, A., Saifi, A.Q., Modi, N.T., Mishra, N. (1986) Anti-inflammatory activity of some active principles of Lawsonia inermis leaves. Indian Journal of Pharmacology 18(6):113-114.
- Gupta, A.K. (2003) Quality standards of Indian medicinal plants. Indian council of medicinal research 1:123-129.
- Habbal, O.A., Al-Jabri, A.A., El-Hag, A.H., Al-Mahrooqi, Z.H., Al Hashmi, N.A. (2005) In vitro antimicrobial activity of Lawsonia inermis Linn (henna)-A pilot study on the Omani henna. Saudi Medical Journal 26:69-72.
- Halloran and Slavin. (2002) Pathophysiology of Wound Healing, Surgury (Oxford) 20 (5).
- Hamdi, Y.P., Benazzouz, M., Belkhiri, H., Chari, Z., Serakta, M., Bensgni, L. (1997) Healing effect of Lawsonia inermis L. (henna) as exemplified by the third degree burns. Revue de Medecines et Pharmacopees Africaines 11-12:151-156.
- Harizal, S.N., Mansor, S.M., Hasnan, J., Tharakan, J.K.J., Abdullah, J. (2010) Acute toxicity study of the standardized methanolic extract of Mitragyna speciosa Korth in Rodent. J. Ethnopharmacol 131, 404-409.
- Hebda, P.A. (1988) Stimulatory effects of transforming growth factor-beta and epidermal growth factor on epidermal cell outgrowth from porcine skin explant cultures. J Invest Dermatol 91: 440-445.
- Hemmati, A.A. and Mohammadian, F. (2000) An investigation into the effects of mucilage of quince seeds on wound healing in rabbit. Journal of Herbs, Spices and Medicinal Plants, vol. 7, no. 4, pp. 41-46.
- Hiramatsu, T., Imoto, M., Koyano, T., Umezawa, K. (1993) Induction of normal phenotypes in rats-transformed cells by damnacanthal from Morinda citrifolia. Cancer Lett 73: 161-6.

- Hirazumi, A., Furusawa, E., Chou, S.C., Hokama, Y. (1994) Anticancer activity of Morinda citrifolia (noni) on intraperitoneally im planted Lewis lung carcinoma in syngeneic mice. Proc West Pharmacol Soc. 37: 145-6.
- Hiwasa, T., Arase, Y., Chen, Z., Kita, K., Umezawa, K., Ito, H. et al., (1999) Stimulation of ultraviolet induced apoptosis of human fibroblast UVr-1 cells by tyrosine kinase inhibitors. FEBS Lett 444: 173-6.
- Iocono, J.A., Colleran, K.R., Remick, D.G., Gillespie, B.W., Ehrlich, H.P. and Garner, W.L. (2000) Interleukin-8 levels and activity in delayed-healing human thermal wounds. Wound Repair and Regeneration, 8(3), 216-225.
- Iversen, P.O., Nicolaysen, G. (2003) Water for life. J. Norw. Med. Assoc 123, 3402-3405.
- Kampfer, H., Pfeilschifter, J. and Frank, S. (2001) Expressional regulation of angiopoietin-1 and -2 and the Tie-1 and -2 receptor tyrosine kinases during cutaneous wound healing: a comparative study of normal and impaired repair. Lab Invest 81: 361–373.
- Khan, M.M., Ali, A., Jain, D.C., Bhakuni, R.S., Zaim, M., Thakur, R.S. (1991) Occurrence of some antiviral sterols in Artemisia annua. Plant Sci. 75(2):161-165.
- Khanna, S., Venojarvi, M., Roy, S., Sharma, N., Trikha, P., Bagchi, D. and Sen, C.K. (2002) Dermal wound healing properties of redox-active grape seed proanthocyanidins. Free Radical Biology and Medicine, 33(8), 1089-1096.
- Khoo, Y.T., Halim, A.S., Singh, K.K. B. and Mohamad, N.A. (2010) Wound contraction effects and antibacterial properties of Tualang honey on full-thickness burn wounds in rats in comparison to hydrofibre. BMC complementary and alternative medicine, 10 (1), 48.
- Kirkland, D., Marzin, D. (2003) An assessment of the genotoxicity of 2-hydroxy-1, 4-naphthoquinone, the natural dye ingredient of Henna. Mutat Res. 537(2):183-199.
- Kirtikar, K.R., Basu, B.D. (1975) Indian Medicinal Plants; International Book Distributors: Dehradun, India, Volume 2, p. 858.
- Klaassen, C.D. (2001) Principles of Toxicology. In Casarett and Doull's Toxicology: The Basic Science of Poisons, 5th ed.; McGraw-Hill: New York, NY, USA, p. 13.
- Koay, S.S. (2008) Establishment of cell suspension culture of Melastoma malabathricum L. for the production of anthocyanin, Ph.D. thesis, Universiti Sains Malaysia, Pulau Pinang, Malaysia.
- Kondo, T. (2007) Timing of skin wounds. Legal Medicine, 9 (2), 109-114.
- Korani, M., Rezayat, S.M., Gilani, K., Arbabi-Bidgoli, S., Adeli, S. (2011) Acute and sub-chronic dermal toxicity of nanosilver in guinea pig. International Journal of Nanomedicine6: 855–862.

- Kumar, R.A., Kokate, C.K., Rambhau, D., Rao, Y.M. (1985) Studies in Lawsonia inermis lawsone and its oxazine derivatives as potential anticoagulant agents. Indian Journal of Pharmaceutical Sciences. 47.
- Labler, L., Mica, L., Härter, L. et al., (2006) Influence of V.A.C. therapy on cytokines and growth factors in traumatic wounds. Zentralbl Chir. 131(suppl 1): S62-S67 [in German].
- Lauer, G., Sollberg, S., Cole, M., Flamme, I., Sturzebecher, J., Mann, K., Krieg, T. and Eming, S.A. (2000) Expression and proteolysis of vascular endothelial growth factor is increased in chronic wounds. J Invest Dermatol 115: 12-18.
- Lazurus, G.S., Cooper, D.M., Knighton, D.R. et al., (1994) Definitions and guidelines for assessment of wounds and evaluation of healing. Arch Dermatol 130: 489-493.
- Li, J., Chen, J. and Kirsner, R. (2007) Pathophysiology of acute wound healing. Clinics in Dermatology. 25, 9-18.
- Ling, K.H., Kian, C.T. and Hoon, T.C. (2009) A Guide to Medicinal Plants. An Illustrated, Scientific and Medicinal Approach, World Scientific, Singapore.
- Lipnick, R.L., Cotruvo, J.A., Hill, R.N., Bruce, R.D., Stitzel, K.A., Walker, A.P., Chu, I., Goddard, M., Segal, L., Springer, J.A. and Myers, R.C. (1995) Comparison of the up-and down, conventional LD50, and fixed-dose acute toxicity procedures. Fd. Chem. Toxicol. 33: 223-231.
- Locher, C.P., Burch, M.T., Mower, H.F., Berestecky, H., Davis, H., Van Polel, B., Lasure, A., Vander-Berghe, D.A., Vlieti-Nick, A.J. (1995) Anti-microbial activity and anti-complement activity of extracts obtained from selected Hawaiian medicinal plants. Journal of Ethnopharmacology 49, 23-32.
- Lorena, D., Uchio, K. and Costa, A.M. (2002) Normal scarring: importance of Myofibroblasts. Wound Repair and Regeneration 10(2): 86-93.
- Mackay, D. and Miller, A.L. (2003) Nutritional support for wound healing. Alternative medicine review 8(4) 359-374.
- Mahmood, A.A., Khaled, A.Ahmed., Hapipah, M.Ali., suzita, M.Noor., Salmah. and Ismail. (2009) Wound Healing Activities of Rafflesia Hasseltii Extract in Rats. J. Clin. Biochem. Nutr.45:p. 304-308.
- Manicam, C., Abdullah, J.O., Tohit, E.M., Seman, Z., Chin, S.C. and Hamid, M. (2010) In vitro anticoagulant activities of Melastoma malabathricum Linn. aqueous leaf extract: A preliminary novel finding. Journal of Medicinal Plants Research, 4(14), 1464-1472.
- Malekzadeh, F. (1968) Antimicrobial activity of Lawsonia inermis L. Appl. Microbiol. 16:663-664.
- Mantle, D., Gok, M.A. and Lennard, T.W.J. (2001) Adverse and beneficial effects of plant extracts on skin and skin disorders. Adverse Drug Reactions and Toxicological Reviews, vol. 20, no.2, pp. 89-103.

- Martini, F.H. (2006) Fundamentals of anatomy and physiology (7th ed.). San Francisco: Benjamin Cummings.
- Meyer, K. (2001) "Revision of the Southeast Asian genus Melastoma (Melastomataceae)," Blumea: Journal of Plant Taxonomy and Plant Geography, vol. 46, no. 2, pp. 351-398.
- McClatchey, W. (2002) From Polynesian healers to health food stores: changing perspectives of Morinda citrifolia (Rubiaceae). Integral Cancer Therapy 1, 110–120.
- McKoy, M.L.G., Thomas, E.A., Simon, O.R. (2002) Preliminary investigation of the anti-inflammatory properties of an aqueous extract from Morinda citrifolia (Noni). Pharmacological Society 45, 76–78.
- Melisa, C. (2000) Oestrogens and wound healing. Maturitas 34, 195-210.
- Merchandetti, M. and Cohen, A.J. (2008) Wound healing, healing and repair. Retrieved on 11 November 2010, from emedicine from webMD. MacLellan, D.G.(2000).Chronic wound management. aust Prescr, 23,6-9.
- Middelkoop, E., Van-Den Bogaerdt, A.J., Lamme, E.N., Hoekstra, M.J., Brandsma, K. and Ulrich, M.M.W. (2004) Porcine wound models for skin substitution and burn treatment. Biomaterials, 25(9), 1559-1567.
- Mikhaeil, B.R., Badria, F.A., Maatooq, G.T., Amer, M.M.A. (2004) Antioxidant and immunomodulatory constituents of henna leaves. Zeitschrift fuer Naturforschung Section C Journal of Biosciences. 59:468-476.
- Mohd, Z., Abdul-Hamid, A., Osman, A. (2001) Antioxidative activity extracts from Mengkudu (Morinda citrifolia L.) root, fruit and leaf. Food Chemistry 78, 227–231.
- Mohsin, A., Shah, A.H., Al-Yahya, M.A., Tariq, M., Tanira, M.O.M., Ageel, A.A. (1989) Analgesic, antipyretic activity and phytochemical screening of some plants used in traditional Arab system of medicine. Fitoterapia 60(2):174-177.
- Moon, C.H. and Crabtree, T.G. (2003) New wound dressing techniques to accelerate wound healing. Current Treatment Options in Infectious Diseases 5:251-260.
- Mortimer, D. (2007) Moist wound dressings and pressure relieving surfaces mechanisms, materials and a review of some cost-effectivness findings. Working paper 104. Centre for Health Program Evaluation. West Heidelberg, Australia.
- Morton, J.J., Malone, M.H. (1972) Evaluation of vulnerary activity by an open wound procedure in rats. Arch. Int. Pharmacodyn 196:117–26.
- Morton, J.F. (1992) The ocean-going Noni, or Indian mulberry (Morinda citrifolia, Rubiaceae) and some of its "colourful" relatives. Ecological Botony 46, 241–256.
- Moshi, M.J. (2007) Brine shrimp toxicity evaluation of some Tanzanian plants used traditionally for the treatment of fungal infections. Afr. J. Tradit. Complement. Altern. Med 4, 219-225.

- Muhammad, H.S., Muhammad, S. (2005) The use of Lawsonia inermis Linn. (henna) in the management of burn wound infections. African Journal of Biotechnology 4:934-937.
- Munshi, S.R., Shetye, T.A., Nair, R.K. (1977) Anti-fertility activity of three indigenous plant preparations. Planta Med 31:73-75.
- Murrell, G.A.C., Pilowsky, E. and Murrell, T.G.C. (1987) A hypothesis on the resolution of dupuytrens contracture with allopurinol. Speculations in Science and Technology, 10(2), 107-112.
- Natarajan, M.R., Lalithakumar, D. (1987) Leaf extracts of Lawsonia inermis as antifungal agent. Curr Sci 56:1021-1022.
- Nayak, B.S., Isitor, G., Davis, E.M., Pillai, G.K. (2007). The evidence based wound healing activity of Lawsonia inermis Linn. Phytotherapy Research 21(9):827-831.
- Nurdiana, S. and Marziana, N. (2013) Wound Healing Activities of Melastoma malabathricum Leaves Extract in Sprague Dawley Rats. Int. J. Pharm. Sci. Rev. Res., 20(2),04, 20-23.
- OECD. (1995) OECD Guideline for Testing of Chemicals. Repeated Dose 28-Day Oral Toxicity Study in Rodents; Organisation for Economic Co-operation and Development: Paris, France.
- Owen, D.A.A. (1987) Inflammation-histamine and 5 hydroxytryptamine. British Medical Bulletin. 43:256.
- Peacock, E.E., Van Winkle, V. (1984) Inflammation and the cellular response to injury. In Wound Healing. Edited by E.E. Peacock, E.E. and Van Winkle, V. Philadelphia, W.B. Saunders.
- Percival, M. (1997) Nutritional support for connective tissue repair and wound healing NUT026/98.
- Perez, B., Davis, S.C. (2008) Relevance of Animal Models for Wound Healing. Retrieved on 9 Dec 2009 from http://www.woundsresearch.com/article/8200.
- Peters, K.G., Devries, C. and Williams, L.T. (1993) Vascular endothelial growth factor receptor expression during embryogenesis and tissue repair suggests a role in endothelial differentiation and blood vessel growth. Proc Natl Acad Sci USA 90: 8915-8919.
- Phillipson, J.D. (2001) Phytochemistry and medicinal plants. Phytochemistry 56(3),237-243.
- Rajenderan, M.T. (2010) "Ethno medicinal uses and antimicrobial properties of Melastoma malabathricum," SEGi Review, vol. 3, pp. 34-44.
- Rasal, V.P., Sinnathambi, A., Ashok, P. and Yeshmaina, S. (2008) Wound healing and antioxidant activities of Morinda citrifolia leaf extract in rats. Iranian Journal of Pharmacology & Therapeutics (IJPT), 7(1), 49-52.

- Raveesha, K.A., Satish, S., Mohana, D.C., Raghavendra, M.P. (2007) Antifungal activity of some plant extracts against important seed borne pathogens of Aspergillus sp. J. Agr Technol 3(1):109-119.
- Raza, M., Al-Shabanah, O.A., El-Hadiyah, T.M., Al-Majed, A.A. (2002) Effect of prolonged vigabatrin treatment on haematological and biochemical parameters in plasma, liver and kidney of Swiss albino mice. Sci. Pharm 70, 135-145.
- Reddy, K.R. (1988) Folk medicine from Chittoor district Andhra Pradesh, India used in the treatment of jaundice. International Journal of Crude Drug Research 26:137-140.
- Rhiouania, H., El-Hilalya, J., Israili, Z.H., Lyoussia, B. (2008) Acute and subchronic toxicity of an aqueous extract of the leaves of Herniaria glabra in rodents. J. Ethnopharmacol 118,378-386.
- Richard, D., Forrest, M.B. (1982) Early history of wound treatment. Journal of the Royal Society of Medicine Vol. 75. p:198-205.
- Rivera, A.E., Spencer, J.M. (2007) Clinical aspects of fullthickness wound healing. Clin Dermatol 25: 39-48.
- Roberts, A.B., Sporn, M.B., Assoian, R.K., Smith, J.M., Roche, N.S., Wakefield, L.M., Heine, U.I., Liotta, L.A., Falanga, V., Kehrl, J.H. and Fauci, A.S. (1986) Transforming growth factor type beta: rapid induction of fibrosis and angiogenesis in vivo and stimulation of collagen formation in vitro. Proc Natl Acad Sci USA 83: 4167-4171.
- Roberts, A.B., Sporn, M.B. (1996) Transforming growth factor. In: The Molecular and Cellular Biology of Wound Repair (2nd ed.), edited by Clark, R.A.F. New York: Plenum. p. 275-308.
- Robson, M.C., Steed, D.L., Franz, M.G. (2001) Wound healing: biologic features and approaches to maximize healing trajectories. Curr Probl Surg 38: 72-140.
- Rosly, S.M., Shanmugavelu, S., Murugaiyah, M., Hadijah, H., Ahmad-Tarmizi, S., Noridayusni, Y. and Subramaniam, K. (2011) Sub-chronic Oral Toxicity Study of Morinda citrifolia in Spraque Dawley Rats. Pertanika J. Trop. Agric. Sci. 34 (2): 341-349.

Ross, I.A. (2001) Medical Plants of the World. Chemical Constituents, Traditional and Modern Medical Uses. Humana Press, New Jersey.

Rowett, H.G.Q. (1968) In: The rat as a small mammal (2edn.), John Murray Ltd:3.

- Rozaini, M.Z., Zuki, A.B.Z., Noordin, M.M., Norimah, Y. and Nazrul Hakim, A. (2005) Macroscopic evaluation of burn wounds healing progress treated with diffrent types of honey. Pakistan Journal of Biological Scinces 8(5):672-8. ISSN 1028-8880.
- Saadabi, M.A.A. (2007) Evaluation of Lawsonia inermis L. (Sudanese Henna) Leaf extracts as an antimicrobial agent. Res J Bio Sci. 2(4):419-423.

- Sakarkar, D.M., Sakarkar, U.M., Shrikhande, V.N., Vyas, J.V., Mandavgade, S., Jaismal, S.B. and Purohit, R.N. (2004) Wound healing properties of Henna Leaves. Natural Products Radiance. vol. 3(6). p 406-412.
- Salawu, O.A., Chindo, B.A., Tijani, A.Y., Obidike, I.C., Salawu, T.A., James Akingbasote, A. (2009) Acute and sub-acute toxicological evaluation of the methanolic stem bark extract of Crossopteryx febrifuga in rats. Afr. J. Pharm. Pharmacol 3, 621-626.
- salmon, J.A., Higgs, G.A. (1987) Prostaglandin and leukotrienes as inflammatory mediators. British Medical Bulletin. 43: 285.
- Santos Heredero, F.X., Hamann, C., Obispo Martin, J.M., Rodriguez Arias, C. and Coca Menchero, S. (1996) Experimental burn models. Ann Burns Fire Disaster 9, 96.
- Sasidharan, S., Darah, I., Jain, K. (2008) In vivo and in vitro toxicity study of Gracilaria changii. Pharm.Biol 46, 413-417.
- Sastri, B.N. (1962) The Wealth of India: Raw Materials. Edn 6, Vol. (L-M), CSIR, New Delhi, pp. 47-50.
- Scortichini, M., Pia Rossi, M. (1991) Preliminary in vitro evaluation of the antimicrobial activity of triterpenes and terpenoids towards Erwinia amylovora (Burrill). J. Bacteriol 71:109–12.
- Sellheyer, K., Bickenbach, J.R., Rothnagel, J.A., Bundman, D., Longley, M.A., Krieg, T., Roche, N.S., Roberts, A.B. and Roop, D.R. (1993) Inhibition of skin development by over expression of transforming growth factor-1 in the epidermis of transgenic mice. Proc Natl Acad Sci USA 90: 5237–5241.
- Shaw, T.J., Martin, P. (2009) Wound repair at a glance. Journal of cell science. 122: 3209-3213.
- Shivananda Nayak, B., Steve Sandiford. and Anderson Maxwell. (2007) Evaluation of the Wound-healing Activity of Ethanolic Extract of Morinda citrifolia L. Leaf. eCAM, 6(3) p 351–356.
- Singh, S., Shrivastava, N.M., Modi, N.T., Saifi, A.Q. (1982) Anti-inflammatory activity of Lawsonia inermis. Current Science (Bangalore) 51:470-471.
- Singh, Y., Ikahihifo, T., Panuve, M., Slatter, C. (1984) Folk medicine in Tonga. A study on the use of herbal medicines for obstetric and gynacological conditions and disorders. J. Ethnopharm 12: 305-25.
- Singh, V.K., Pandey, D.K. (1989) Fungitoxic studies on bark extract of Lawsonia inermis against ringworm fungi. Hindusthan Antibiot Bull 31(1-2):32-35.
- Solomon, N. (1999) The Noni phenomenon. Discover the powerful tropical healer that fights cancer, lowers high blood pressure and relieves chronic pain. Direct Source Publishing.
- Solomon, N. (1999) The tropical fruit with 101 medicinal uses, NONI juice. 2nd ed. Woodland Publishing.



Solomon, N. (1999) The Noni Phenomenon. Direct Source Publishing, Utah.

- Steven, K.R., Mylecrdfaine, L. (1994) Issues in Chronic Toxicology. In Principles and Methods of Toxicology, 3rd ed.; Hayes, A.W., Ed.; Ravan Press: New York, NY, USA, p. 673.
- Su, C., Wang, M., Nowicki, D., Jensen, J., Anderson, G. (2001) Selective COX-2 inhibition of Morinda citrifolia (Noni) in vitro. In: The Proceedings of the Eicos anoids and other Bioactive Lipids in Cancer, Inflammation and Related Disease. The 7th Annual Conference, 2001 October 14-17. Loews Vanderbilt Plaza, Nashville, Tennessee, USA.
- Su, W.H., Cheng, M.H., Lee, W.L., Tsou, T.S., Chang, W.H., Chen, C.S. and Wang, P.H. (2010) Nonsteroildal Anti-inflammatory Drugs for wounds: pain relief or excessive scar formation. Mediator of inflammation 2010. DOIL 10.115/2010/4132238.
- Sulaiman, M.R., Somchit, M.N., Israf, D.A., Ahmad, Z., Moin, S. (2004) Antinociceptive effect of Melastoma malabathricum ethanolic extract in mice, Fitoterapia, 75,667-672.
- Sunilson, A.J., James, J., Thomas, J. (2008) Antibacterial and wound healing activities of Melastoma malabathricum Linn. African Journal of infectious Diseases, 2,68-73.
- Sunilson, J.A.J., James, J.J., Thomas, J., Jayaraj, P., Varatharajan, R. and Muthappan, M. (2008) "Antibacterial and wound healing activities of Melastoma malabathricum Linn.," African Journal of Infectious Disease, vol. 2, pp. 68-73.
- Sunilson, J.A.J., Anandarajagopal, K., Kumari, A.V.A.G. and Mohan, S. (2009) "Antidiarrhoeal activity of leaves of Melastomamalabathricum linn," Indian Journal of Pharmaceutical Sciences, vol. 71, no. 6, pp. 691–695.
- Sussman, C., Bates Jensen, B.M. (2001) Wound healing physiology and chronic wound healing in wound care. A collaborative Practice Manual for Physical Therapists and Nurses. (Sussman, C. and Bates Jensen, B.M.eds), p. 26-51. Maryland: Aspen Publisher, Inc.
- Swift, M.E., Kleinman, H.K. and Dipietro, L.A. (1999) Impaired wound repair and delayed angiogenesis in aged mice. Lab Invest 79: 1479-1487.
- Syahmi, A.R.M., Vijayarathna, S., Sasidharan, S., Yoga Latha, L., Kwan, Y.P., Lau, Y.L., Shin, L.N., Chen, Y. (2010) Acute oral toxicity and brine shrimp lethality of Elaeis guineensis Jacq., (Oil Palm leaf) methanol extract. Molecules 15, 8111-8121.
- Syamsudin, I., Winarno, H. (2008) The effects of Inai (Lawsonia inermis) leave extract on blood sugar level: An Experimental Study. Res .J. Pharmacol 2(2):20-23.
- Szycher, M., Lee, S.J. (1992) Modern wound dressings: a systematic approach to wound healing. J. Biomater Appl 7: 142 213.

- Teo, S.D., Stirling, S., Thomas, A., Kiorpes, A., Vikram, K. (2002) A 90-day oral gavage toxicity study of D-methylphenidate and D, L methylphenidate in Sprague-dawley rats. Toxicology 179,183-196.
- Thomas, S. (1990) Wound management and dressing. London: the Pharmaceutical press.
- Thomas, R.III. (2003) Wound healing. eMEdicine World Medical Library (www.emedic.com).
- Thomas Romo, III., Pearson, J.M., Yalamanchili, H. and Zoumalan, R.A. (2008) Wound Healing, Skin (Publication. Retrieved 25 February 2010, from eMedicine from Web MD: http://emedicine.medscape.com/article/884594-overview.
- Toree, J. I. D. L., and Chambers, J.A. (2008). Wound Healing, chronic Wounds (Publication. Retrieved 25 February 2010, from EMedicine from Web MD: http://emedicine.medscape.com/article/1298452-overview.
- Tomlinson, A., Ferguson, M.W.J. (2003) Chapter 26: wound healing. In: Winyard PG, willoughby D A. Methods in molecular biology, vol. 225: Inflammation protocols. Totowa, NJ: Hummana Press Inc; p249-60.
- Tripathi, R.D., Srivastava, H.S., Dixit, S.N. (1978) A fungi toxic principle from the leaves of Lawsonia inermis. Experientia (Basel) 34:51-52.
- Truong, N., Vern, A.K., Latenser, B.A., wiley, D.E. and Walter, R.J. (2005) Comparison of dermal substitutes in wound healing utilizing a nude mouse model. J. Burn Wounds., 14(4);(4ed.) 72-78.
- Tsuchiya, H., Sato, M., Miyazaki, T., Fujiwara, S., Tanigaki, S., Ohyama, M. et al., (1996) Comparative study on the antibacterial activity of phytochemical flavanones against methicillin-resistant Staphylococcus aureus. J Ethnopharmacol 50:27-34.
- Ukong, S., Ampawong, A. and Kengkoom, K. (2008) Collagen measurement and staining pattern of wound healing comparison with fixation and stains.
- Umezawa, K. (1992) Isolation of 1-methoxy-2-foremyl-3-hydroxy anthraquinone from M. citrifolia and neoplasm inhibitors con taining the same. Japan Kokai Tokyo Koho J. P 06 87, 736 (94-87, 736) Appl 92/264, 311 07.
- Vaghasiya, Y.K., Shukla, V.J., Chanda, S.V. (2011) Acute oral toxicity study of Pluchea arguta boiss extract in mice. J. Pharmacol. Toxicol 6, 113-123.
- Van Valkenberg, J.L.C.H., and Bunyapraphatsara, N. (2001) "Melastoma malabathricum L.," in Plant Resources of South-East Asia No. 12(2): Medicinal and Poisonous Plants 2, J. L. C. H. van Valkenburg and N. Bunyapraphatsara, Eds., pp. 365–366, Backhuys, Leiden, The Netherlands.
- Vanwijck, R. (2001) Surgical biology of wound healing. Bull Mem Acad R Med Belg 115: 175-184.
- Vasudevan, T.N., Laddha, K.S. (2003) Herbal drug microscopy, Edn 1, Yucca publishing house, Dombivli. pp. 68-69.

- Watson, T. (2006) Soft tissue wound healing review. Retrieved on 11 Nov 2010 from http://www.electrotherapy.org/downlowds/Modalities/tissue%20repair.pdf.
- Wang, M., Kikuzaki, H., Jin, Y., Nakatani, N., Zhu, N., Csiszar, K. et al., (2000) Novel glycosides from noni (Morinda citrifolia). J. Nat Prod 63: 1182-3.
- Wang, M.Y., Su, C. (2001) Cancer preventive effect of Morinda citrifolia (Noni). Ann NY Acad Sci 952: 161-8.
- Wang, M.Y., West, B.J., Jensen, C.J., Nowicki, D., SU, C., Palu, A.K. and Anderson, G. (2002) Morinda citrifolia (Noni): A literature review and recent advances in Noni research. Acta Pharmacol Sin, 23(12):p. 1127-41.
- Wheater, P.R., Young, B., Lowe, J.S., Stevens, A. and Heath, J.W. (2006) Wheatear's functional histology: a text and colour atlas (5th ed.). London: Churchill Livingstone.
- Winter, G. (1962) Formation of scab and the rate of epithelization of superficial wounds. Nature, 193:293-4.
- Wong, W. (2008) Melastoma malabathricum: Too Beautiful to Be Called a Weed, Green Culture, Singapore.
- Younos, C., Rolland, A., Fleurentin, J., Lanhers, M.C., Misslin, R., Mortier, F. (1990) Analgesic and behavioral effects of Morinda citrifolia. Planta Medicine 56, 430–434.
- Zakaria, M. and Mohd, M.A. (1994) Traditional Malay Medicinal Plants, Fajar Bakti Sdn. Bhd., Kuala Lumpur, Malaysia.
- Zakaria, Z.A., Raden, M. N. R. N. S., Hanan Kumar, H. et al., (2006) "Antinociceptive, anti-inflammatory and antipyretic properties of Melastoma malabathricum leaves aqueous extract in experimental animals," Canadian Journal of Physiology and Pharmacology, vol. 84, no. 12, pp. 1291-1299.
- Zambruno, G., Marchisio, P.C., Marconi, A., Vaschieri, C., Melchiori, A., Giannetti, A. and Deluca, M. (1995) Transforming growth factor-beta 1 modulates beta 1 and beta 5 integrin receptors and induces the de novo expression of the alpha v beta 6 heterodimer in normal human keratinocytes: implications for wound healing. J. Cell Biol 129: 853-865.
- Zhengyi, W., Raven, P.H. and Hong, D.Y. (2007) Flora of China (Vol. 13) (Clusiaceae through Araliaceae), Missouri Botanical Garden Press, St. Louis, Mo, USA.