



UNIVERSITI PUTRA MALAYSIA

***GERMINATION, SEEDLING GROWTH AND ANATOMICAL RESPONSES
OF *Cucumis sativus* cv. MTi2 IN DIFFERENT SALTS AND
DEVELOPMENT OF GERMINATION ENHANCER***

SAMAR JASIM MOHAMMED

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SAMAR JASIM MOHAMMED

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfillment of the Requirements for the Degree of
Master of Science**

April 2017

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Master of Science

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April 2017

Chairman : Associate Professor Rosimah Nulit, PhD
Faculty : Science

Salinity continues to be one of the most serious environmental problems. One of the strategies in dealing with salinity is producing salt tolerant plants and understanding the effects of salinity on crops. Seed germination is an important stage in the life cycle of plants and understanding of tolerance to salinity during the germination stage is crucial for the establishment and management of plant in saline soils. Many studies had been conducted using the effect of NaCl on seed germination, and although there are other types of salt found in soil, until to date, studies about these salts are still limited. Therefore, this study aimed to compare the effects of NaCl, KCl, MgCl₂, MgSO₄ and CaCl₂ on the germination of MTi2 seed, early seedling growth and anatomical of seedling cotyledon leaves. This study also aimed to develop the liquid enhancer in order to improve the germination of salt stressed MTi2 seed. Five types of salts (NaCl, KCl, MgCl₂, MgSO₄ and CaCl₂) at different concentration (50,100,150, and 200 mM) and deionized water as a control were used with nine replicates for each treatment. A 10 sterilized MTi2 seeds were placed in petri dishes containing 5 ml of deionized water or each salinity solution and placed in a completely randomized design in the growth room at 25 ± 1°C. The number of germinated seeds was recorded daily until day 8. On day 8, the length of the hypocotyl, radicle and the biomass of seedlings was measured and the cotyledon leaves were fixed for anatomical study. Germination percentage, germination rate, seed vigor, relative salt injury rate, salt tolerance was calculated. To develop liquid enhancer, sterilized MTi2 seeds were primed with 300mM NaCl for 72 hours and then treated with Salicylic acid alone (SA) (0.25, 0.5, 0.75, 1mM) and KCl alone (10, 20, 30, 40, 50 mM) and deionized water as a control. Germination parameters were calculated as mentioned before. Following this, the ideal concentration of the SA and KCl was mixed together and was used to study its effectiveness as a germination enhancer on the salt-stressed MTi2 seeds. Data were analyzed using SPSS windows version 22. Data are subjected two way ANOVA at

confidence level, $p \leq 0.05$ to determine the significant difference between treatment and followed by Tukey at $p \leq 0.05$ for means comparison. The Study found that the germination response of MTi2 seed on NaCl, KCl, MgCl₂, MgSO₄ and CaCl₂ is significantly different. MTi2 seeds are able to germinate in KCl and NaCl until the high concentration (200 mM) which is % GP more than 50%. In addition, MTi2 seeds able to germinate in medium and low concentration of CaCl₂ (50-100 mM) and MgSO₄ (50 mM) but unable to germinate in any concentration of MgCl₂. The germination rates and the vigor of seed found higher in KCl and NaCl but lower in CaCl₂ and MgSO₄. Results also found Relatives Salts Injury Rate of MTi2 germinating seed are very low in KCl and NaCl but higher in CaCl₂ and MgSO₄. The tolerance level of MTi2 seed on different types of salts can be concluded as follows, KCl > NaCl > CaCl₂ > MgSO₄ > MgCl₂. This study also found that 50 mM KCl increased the germination percentage, germination rate and the vigor of MTi2 seeds. Histological studies revealed that changes of MTi2 cotyledon leaves as a response to NaCl, KCl, MgCl₂, MgSO₄ and CaCl₂ are different. The degree histological changes of MTi2 cotyledon leaves increased as the concentration of salts increased. Another type of salts significantly changes the structure and arrangement of the upper epidermis cells, mesophyll tissue and lower epidermis cells, also reduce of intracellular space among the mesophyll cells. Results also found that no anatomical changes of MTi2 cotyledon leaves in any concentration of KCl. Results showed that 0.5-0.75 mM SA and 20-30 mM KCl are the ideal concentration that increased significantly the germination and the early growth of MTi2 seedling compared with control treatment. Moreover, the combination of the ideal concentration of SA (0.5 -0.75 mM) and KCl (20-30 mM) increased the germination and the early growth of MTi2 seedlings more than 1x higher than SA and KCl individually. Salt-stressed MTi2 seeds treated with salicylic acid (SA) and low level of (KCl) can contribute in mitigation of the deleterious effects of salinity stress and improve seed germination percentage, germination rate, seed vigor, seedling length and seedling biomass. It can be concluded that the tolerance level of MTi2 seed on different types of salts can be summarized as follows, KCl > NaCl > CaCl₂ > MgSO₄ > MgCl₂. Low concentration of a mixture of KCl and SA can be used as an enhancer to increase germination of salt stressed MTi2 seeds.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Master Sains

**PERCAMBAHAN, PERTUMBUHAN ANAK BENIH, GERAKBALAS
ANATOMI *Cucumis sativus* cv. MTi2 TERHADAP BERLAINAN
GARAM-GARAM DAN PEMBANGUNAN PENGGALAK
PERCAMBAHAN BAGI BIJI BENIH TIMUN YG DIBERIKAN TEGASAN
GARAM**

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Kemasinan terus menjadi salah satu masalah alam sekitar yang serius. Salah satu strategi dalam menanganinya dengan menghasilkan tumbuhan yang toleran dengan kemasinan dan memahami kesannya kepada tanaman. Percambahan benih adalah satu peringkat yang penting dalam kitaran hidup tumbuhan dan kefahaman toleransi kepada kemasinan pada peringkat ini adalah penting bagi pertumbuhan pokok. Banyak kajian telah dijalankan menggunakan kesan NaCl pada percambahan benih, dan walau kandungan tanah terdapat garam yang lain, kajian mengenai garam-garam ini masih terhad. Oleh itu, kajian ini dijalankan bertujuan untuk membandingkan kesan NaCl, KCl, MgCl₂, MgSO₄ dan CaCl₂ pada percambahan biji benih timun MTi2, pertumbuhan awal anak benih dan anatomi daun kotiledon anak benih. Kajian ini juga bertujuan untuk membuat satu larutan formulasi yang dapat meningkatkan percambahan garam benih MTi2 yang diberikan tegasan garam. Lima jenis garam (NaCl, KCl, MgCl₂, MgSO₄ dan CaCl₂) pada kepekatan yang berbeza (50, 100, 150, dan 200 mM) dan air ternyahion sebagai kawalan digunakan dengan sembilan replikat pada setiap rawatan. Sebanyak 10 biji benih MTi2 disterilkan diletakkan di dalam piring petri yang diberi 5 ml air ternyahion atau larutan garam dan diatur secara rawak di ruang pertumbuhan pada suhu 25±1°C. Bilangan benih bercambah dicatatkan setiap hari sehingga hari ke 8. Pada hari ke 8, panjang hipokotil, radikel dan biomas benih diukur. Daun kotiledon diambil dan diawetkan untuk kajian anatomi. Peratusan percambahan, kadar percambahan, vigor benih, kadar kecederaan relatif, toleransi terhadap garam dikira. Untuk menyediakan formulasi, biji benih MTi2 telah direndam dengan 300mM NaCl selama 72 jam untuk memberikan tegasan garam kepada biji benih MTi2 dan kemudian dirawat secara berasingan dengan asid salisilik sahaja (SA) (0.25, 0.5, 0.75, 1mM) dan KCl (10, 20, 30, 40, 50 mM) dan air ternyahion sebagai kawalan.

Ini bertujuan untuk mendapatkan kepekatan yang ideal. Parameter percambahan dikira seperti yang dinyatakan sebelum ini. Kepekatan SA dan KCl yang telah dicampurkan bersama-sama dan digunakan untuk mengkaji keberkesannya sebagai penggalak untuk percambahan bijibenh MTi2 yang telah diberikan tegasan garam. Data dianalisis dengan menggunakan SPSS versi window 22. Analisis data menggunakan "two way " ANOVA pada tahap aras keertian, $p=0.05$ untuk menentukan perbezaan signifikan di antara rawatan-rawatan dan diikuti oleh DMRT pada $p=0.05$ untuk perbandingan antara purata rawatan. Kajian mendapati bahawa tindakbalas percambahan benih MTi2 pada NaCl, KCl, MgCl₂, MgSO₄ dan CaCl₂ adalah sangat berbeza. Benih MTi2 didapati bercambah dalam KCl dan NaCl sehingga kepekatan yang tinggi (200 mM) dimana peratus percambahan (% GP) lebih daripada 50%. Bijibenh MTi2 hanya boleh bercambah dalam kepekatan CaCl₂ yang sederhana (50-100 mM) dan pada kepekatan yang rendah (50 mM) dalam MgSO₄ tetapi tidak bercambah dalam mana-mana kepekatan MgCl₂. Kadar percambahan dan vigor benih didapati lebih tinggi dalam rawatan KCl dan NaCl berbanding dalam rawatan CaCl₂ dan MgSO₄. Keputusan juga menunjukkan kadar kecederaan relatif benih MTi2 sangat rendah dalam KCl dan NaCl tetapi lebih tinggi dalam CaCl₂ dan MgSO₄. Oleh itu, tahap toleransi benih MTi2 terhadap garam dapat disimpulkan seperti berikut, KCl > NaCl > CaCl₂ > MgSO₄ > MgCl₂. Kajian ini juga mendapati bahawa 50 mM KCl meningkat peratusan percambahan, indeks percambahan dan vigor benih MTi2. Kajian histologi menunjukkan bahawa perubahan histologi MTi2 daun kotiledon sebagai tindak balas kepada garam-garam adalah berbeza. Tahap perubahan histologi MTi2 daun kotiledon meningkat seiring dengan kepekatan garam. Rawatan benih MTi2 dengan garam didapati mengubah struktur dan susunan sel-sel epidermis atas, tisu-tisu mesofil dan sel-sel epidermis. Ruang intraselular antara sel-sel mesophyll juga semakin mengecil disebabkan oleh kesan tegasan larutan garam. Keputusan juga mendapati bahawa tiada perubahan anatomi MTi2 daun kotiledon dalam mana-mana kepekatan KCl. Hasil kajian menunjukkan bahawa 0.5-0.75 mM SA dan 20-30 mM KCl adalah kepekatan yang ideal yang dapat meningkat dengan kadar percambahan dan pertumbuhan awal MTi2 anak benih berbanding dengan rawatan kawalan. Selain itu, kombinasi campuran SA (0.5 -0.75 mM) dan KCl (20-30 mM) meningkatkan percambahan dan pertumbuhan awal anak benih MTi2 1x lebih tinggi daripada rawatan SA dan KCl secara berasingan. Sebagai kesimpulan, tahap toleransi benih MTi2 terhadap pelbagai jenis garam boleh diringkaskan seperti berikut, KCl > NaCl > CaCl₂ > MgSO₄ > MgCl₂. Kombinasi campuran SA dan KCl boleh digunakan sebagai penggalak untuk meningkatkan percambahan benih MTi2 yang diberikan tegasan garam.

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I certify that a Thesis Examination Committee has met on 21 April 2017 to conduct the final examination of Samar Jasim Mohammed on her thesis entitled "Germination, Seedling Growth, and Anatomical Responses of *Cucumis sativus* cv. MTi2 in Different Salts and Development of Germination Enhancer" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

pH	Potential of hydrogen
Na ⁺	Sodium ion
Ca ²⁺	Calcium ion
Mg ²⁺	Magnesium ion
Cl ⁻¹	Chloride ion
SO ₄ ⁻²	Sulfate ion
HCO ₃ ⁻¹	Bicarbonate ion
NO ₃ ⁻²	Nitrate ion
CO ₃ ⁻²	Carbonate ion
K ⁺	Potassium ion
EC	Electrical conductivity
mm/l	Millimole/liter
mM	Millimole
SA	Salicylic acid
cm	Centimeter
D _{Sm} ⁻¹	DeciSemens per meter
NaCl	Sodium chloride
KCl	Potassium chloride
MgSO ₄	Magnesium sulfate
MgCl ₂	Magnesium chloride
CaCl ₂	Calcium chloride
GP	Germination percentage
GR	Germination rate

(%)	Percentage
(g)	Gram
HSD	(Honestly significant difference)
FAO	Food and Agriculture Organization



CHAPTER 1

INTRODUCTION

1.1 Background of the study

Plant exposes to abiotic and biotic stresses since decades ago that cause severe effects on metabolism, growth, development, and productivity (Vorasoot *et al.*, 2003; Kaur *et al.*, 2008; Thakur *et al.*, 2010; Doupis *et al.*, 2011). One of worldwide problem stress is salinity and caused about 323 million hectares worldwide land salinized (Brinkman, 1980) is estimated to exceed 400 million hectares by 2025 (Hakim *et al.*, 2014; Flowers & Muscolo, 2015).

Salinity changes soil properties, both physically and chemically, this eventually change the osmotic pressure (Rowell, 1988; Hossain *et al.*, 2015). According to Zhao *et al.*, (2007), soil salinity causes substantial reduction in crop yields, particularly in arid and semi-arid regions with naturally high soil salt and low rainfall that inhibits leaching. The pH and availability of nutrients of soil such as phosphorus, iron and manganese affected by salinity (Hassan *et al.*, 1970).

Adaptation of plants to salinity during germination and early seedling stages is critical for the plant stand to be established. The common cations in soil are Na^+ , Ca^{2+} , Mg^{2+} , meanwhile Cl^- , SO_4^{2-} , and HCO_3^- are the common anions (Flower *et al.*, 1997; Hasegawa *et al.*, 2000; Ali, 2010). Current studies found that different salts cause different effects on the germination and the growth of plants (Ghanad, 2016), this is due to the osmotic and ionic stress in each type of salts had different degrees of influence on germination and development (Panuccio, 2014). Studies demonstrate that the germination of plant is strongly affected by the nature of the ions in the salt solutions and their interactions (Sosa *et al.*, 2005). Seeds have different response to levels of salt during germination (Hussain *et al.*, 2013). Variety or cultivar of plant species vary in various ontogenetic stages to salt tolerance, which give scope for choice of genotype for salt tolerance, indicating the differential response of genotypes in plant species to salinity (Ashagre *et al.*, 2013).

1.2 Problem Statement, Justification and Objectives of the Study

Germination stage in the life cycle of the plant is important and understanding the tolerance level of seed at this stage is needed. Many studies have been conducted extensively on the effect of NaCl on seed germination, but until to date, the study of the different type of salt on the germination of seed still limited.

The impact of salinity depends on its type, level and the genotype of plant species (Adolf, 2012; Ghanad, 2016). This study was conducted on the Malaysia cucumber cv. MTi2 which is the best known and most popular cucumber cultivars among the locals. Cucumber (*Cucumis sativus*) belongs to the gourd, family Cucurbitaceae. It is a widely cultivated, creeping vine that bears cylindrical, fruits that are used as culinary vegetables (Grubben *et al.*, 2004). The cucumber originated from South Asia, but is currently found on most continents. This plant is an important greenhouse crop in semi- arid areas with saline ground water. Therefore, it is important more study to be done on the impact of salinity on germination of this plant (Sato *et al.*, 2006). Cucumber plant was classified as glycophytes and moderately sensitive to salinity. In addition, previous studies focused on other cucumber cultivars to reveal the tolerance to salinity, however, tolerance study of Malaysian cucumber cv. MTi2 is still not done yet.

Therefore, the objectives of the present study are:

- i. To study the effects of different concentration of NaCl, KCl, MgCl₂, MgSO₄, and CaCl₂ on the germination, early seedling growth and the anatomical changes of *C. sativus* cv. MTi2.
- ii. To develop the liquid enhancer in order to improve the germination of salt stressed *C. sativus* cv. MTi2 seed.

REFERENCES

- Abari, A. K., Nasr, M. H., Hojjati, M., & Bayat, D. (2011). Salt effects on seed germination and seedling emergence of two *Acacia* species. *African Journal of Plant Science*, 5(1), 52-56.
- Abdul-Baki, A. A., & Anderson, J. D. (1973). Vigor determination in soybean seed by multiple criteria. *Crop science*, 13(6), 630-633.
- Achakzai, A. K. K. (2009). Effect of water stress on imbibition, germination and seedling growth of maize cultivars. *Sarhad Journal of agriculture*, 25(2), 165-172.
- Adolf, V. I., Shabala, S., Andersen, M. N., Razzaghi, F., & Jacobsen, S. E. (2012). Varietal differences of quinoa's tolerance to saline conditions. *Plant and Soil*, 357(1-2), 117-129.
- Afzal, I., Rauf, S., Basra, S. M. A., & Murtaza, G. (2008). Halopriming improves vigor, metabolism of reserves and ionic contents in wheat seedlings under salt stress. *Plant Soil Environ*, 54(9), 382-388.
- Ahmadi, A., Emam, Y., & Pessaraki, M. (2009). Response of various cultivars of wheat and maize to salinity stress. *Journal of Food Agriculture Environ*, 7(1), 123-128.
- Aishah, H. S., Saberi, A. R., Halim, R. A., & Zaharah, A. R. (2010). Salinity effects on germination of *forage sorghum*. *Journal of Agronomy*, 9 (4), 169-174.
- Al Sahil, A. A. (2016). Effect of Gibberellic and Salicylic Acids Pre-Soaking on Seed Germination Attributes of Cucumber (*Cucumis sativus L.*) under Induced Salt Stress. *Cercetari Agronomice in Moldova*, 49(1), 99-109.
- Ali, A. (2010). Exploring the possibility of transforming food crops for salinity tolerance using the TMT gene encoding thiol methyltransferase enzyme. (Master dissertation, University of Waterloo).
- Ali, A., Hyder, S. I., Arshadullah, M., & Bhatti, S. U. (2012). Potassium chloride as a nutrient seed primer to enhance salt-tolerance in maize. *Pesquisa Agropecuária Brasileira*, 47(8), 1181-1184.
- Almansouri, M., Kinet, J. M., & Lutts, S. (2001). Effect of salt and osmotic stresses on germination in durum wheat (*Triticum durum Desf.*). *Plant and Soil*, 231(2), 243-254.
- Amirjani, M. R. (2011). Pigments and Enzyme Activity of Rice. *International Journal of Botany*, 7(1), 73-81.

- Anaya, F., Fghire, R., Wahbi, S., & Loutfi, K. (2015). Influence of salicylic acid on seed germination of *Vicia faba L.* under salt stress. *Journal of the Saudi Society of Agricultural Sciences*.
- Anonymous (2012). Nutritional recommendations for cucumber (*Cucumis sativus L.*) in the open fields, tunnel and greenhouse. Haifa, Pioneering the Future pp 45. <http://www.haifagroup.com/files/Guides/Cucumber.pdf>
- Asaadi, A. M. (2009). Investigation of salinity stress on seed germination of *Trigonella foenum-graecum*. *Research Journal of Biological Sciences*, 4(11), 1152-1155.
- Ashagre, H., Hamza, I. A., Fasika, E., & Temesgen, F. (2013). Effect of salinity stress on germination and seedling vigour of chickpea (*Cicer arietinum L.*) cultivars. *Academia Journal of Agricultural Research*, 1(9), 161-166.
- Ashraf, M. (2002). Salt tolerance of cotton: some new advances. *Critical Reviews in Plant Sciences*, 21(1), 1-30.
- Ashraf, M. (2009). Biotechnological approach of improving plant salt tolerance using antioxidants as markers. *Biotechnology advances*, 27(1), 84-93.
- Ashraf, M. (2010). Registration of 'S-24'spring wheat with improved salt tolerance. *Journal of plant registrations*, 4(1), 34-37.
- Ashraf, M., & McNeilly, T. (2004). Salinity tolerance in *Brassica oilseeds*. *Critical Reviews in Plant Sciences*, 23(2), 157-174.
- Aydinsakir, K., Ulukapi, K., Kurum, R., & Buyuktas, D. (2013). Effects of different salt sources and concentrations on germination and seedling growth of some pumpkin seeds varieties used as rootstock. *African Journal of Agricultural*, 8(19), 2254-2252.
- Aziz, I., & Khan, M. A. (2001). Effect of Seawater on the Growth, Ion Content and Water Potential of *Rhizophora mucronata Lam.* *Journal of Plant Research*, 114(3), 369-373.
- Baghbani, A., Forghani, A. H., & Kadkhodaie, A. (2013). Study of salinity stress on germination and seedling growth in greenhouse cucumber cultivars. *Journal of Basic and Applied Scientific Research*, 3, 1137-1140.
- Bahrani, A., & Pourreza, J. (2012). Gibberlic acid and salicylic acid effects on seed germination and seedlings growth of wheat (*Triticum aestivum L.*) under salt stress condition. *World App Sci J*, 18(5), 633-641.

- Baninasab, B., & Baghbanha, M. R. (2013). Influence of salicylic acid pre-treatment on emergence and early seedling growth of cucumber (*Cucumis sativus*) under salt stress. *International Journal of Plant Production*, 7(2).
- Barmon, P. K. (2013). Effect Of Different Salt Concentrations On Germination And Growth Of Eight Varieties Of Cucumber (Doctoral Dissertation).
- Bijanazadeh, E., & Kazemeini, S. A. (2014). Tissue architecture changes of expanding barley (*Hordeum vulgare* L.) leaf under salt stress. *Australian Journal of Crop Science*, 8(10), 1373.
- Bot, A., Nachtergaele, F., & Young, A. (2000). Land resource potential and constraints at regional and country levels(pp.62) Rom: food and agriculture organization.
- Bradford, K. J. (1990). A water relations analysis of seed germination rates. *Plant Physiology*, 94(2), 840-849
- Brinkman R (1980) Saline and sodic soils. In: Land reclamation and water management International Institute for Land Reclamation and Improvement (ILRI), Wageningen, The Netherlands. pp62-68.
- Cakmak, I. (2005). The role of potassium in alleviating detrimental effects of abiotic stresses in plants. *Journal of Plant Nutrition and Soil Science*, 168(4), 521-530.
- Çavuşoğlu, K., Kiliç, S., & Kabar, K. (2008). Effects of some plant growth regulators on leaf anatomy of radish seedlings grown under saline conditions. *Journal of Applied Biological Sciences*, 2(2), 47-50.
- Chehregani, A., Mohsenzade, F., & Ghanad, M. O. N. A. (2011). Male and female gametophyte development in *Cichorium intybus*. *International Journal Of Agriculture & Biology*, 13, 603-606.
- Clarkson, D. T., & Hanson, J. B. (1980). The mineral nutrition of higher plants. *Annual review of plant physiology*, 31(1), 239-298.
- Cronquist, A. (1981). *An integrated system of classification of flowering plants*. Columbia University Press. Dekker, New York, pp. 199-244.
- Deef, H. E. (2007). Influence of salicylic acid on stress tolerance during seed germination of *Triticum aestivum* and *Hordeum vulgare*. *Advances in biological research*, 1 (1-2), 40-48.
- Deivanai, S., Xavier, R., Vinod, V., Timalata, K., & Lim, O. F. (2011). Role of exogenous proline in ameliorating salt stress at early stage in two rice cultivars. *Journal of Stress Physiology & Biochemistry*, 7(4).337-341

- Dezfuli, P. M., Sharif-Zadeh, F., & Janmohammadi, M. (2008). Influence of priming techniques on seed germination behavior of maize inbred lines (*Zea mays* L.). *Journal of Agricultural and Biological Science*, 3(3), 22-25.
- Dolatabadian, A., Sanavy, S. A. M. M., & Ghanati, F. (2011). Effect Of Salinity On Growth, Xylem Structure And Anatomical Characteristics Of Soybean. *Notulae Scientia Biologicae*, 3(1), 41.
- Dolatabadian, A., Sanavy, S. A. M. M., & Ghanati, F. (2011). Effect of salinity on growth, xylem structure and anatomical characteristics of soybean. *Notulae Scientia Biologicae*, 3(1), 41.
- Domroes, M. (1991). The tropical forest ecosystem: Reviewing the effects of deforestation on climate and environment. In *The Global Environment* (pp. 70-80). Springer Berlin Heidelberg.
- Dotaniya, M. L., Meena, V. D., Basak, B. B., & Meena, R. S. (2016). Potassium uptake by crops as well as microorganisms. In *Potassium Solubilizing Microorganisms for Sustainable Agriculture* (pp. 267-280). Springer India.
- Doupis, G., Chartzoulakis, K., Beis, A., & Patakas, A. (2011). Allometric and biochemical responses of grapevines subjected to drought and enhanced ultraviolet-B radiation. *Australian Journal of Grape and Wine Research*, 17(1), 36-42.
- Egli, D. B., & Rucker, M. (2012). Seed vigor and the uniformity of emergence of corn seedlings. *Crop Science*, 52(6), 2774-2782.
- Elouaer, M. A., & Cherif, H. (2012). Effect of NaCl priming duration and concentration on germination behavior of Tunisian safflower. *Journal of Stress Physiology & Biochemistry*, 8(3).
- El-Tayeb, M. A. (2005). Response of barley grains to the interactive effect of salinity and salicylic acid. *Plant Growth Regulation*, 45(3), 215-224.
- Erdal, S., Aydın, M., Genisel, M., Taspınar, M. S., Dumlupınar, R., Kaya, O., & Gorcek, Z. (2011). Effects of salicylic acid on wheat salt sensitivity. *African Journal of Biotechnology*, 10(30), 5713-5718.
- Fageria, N. K. (1985). Salt tolerance of rice cultivars. *Plant and soil*, 88(2), 237-243.
- FAO. (2015) status of world soil resources .retrieved 2nd september , 2016 from <https://www.fao.org/3/a-bc595e.pdf>

- Farahbakhsh, H. (2012). Germination and seedling growth in un-primed and primed seeds of fenel as affected by reduced water potential induced by NaCl. *International Research Journal of Applied and Basic Sciences.*, 3(4), 737-744.
- Fawzy, Z. F., El-Nemr, M. A., & Saleh, S. A. (2007). Influence of levels and methods of potassium fertilizer application on growth and yield of eggplant. *Journal of Applied Sciences Research*, 3(1), 42-49.
- Fisarakis, I., Chartzoulakis, K., & Stavrakas, D. (2001). Response of Sultana vines (*V. vinifera* L.) on six rootstocks to NaCl salinity exposure and recovery. *Agricultural water management*, 51(1), 13-27.
- Flowers TJ, Muscolo A (2015) Short Communication Special Issue: Physiology and Ecology of Halophytes — Plants Living in Salt-Rich Environments Introduction to the Special Issue: *Halophytes in a changing world*. 7, 1–5
- Flowers, T. J., & Flowers, S. A. (2005). Why does salinity pose such a difficult problem for plant breeders. *Agricultural water management*, 78(1), 15-24.
- Flowers, T. J., Garcia, A., Koyama, M., & Yeo, A. R. (1997). Breeding for salt tolerance in crop plants—the role of molecular biology. *Acta Physiologiae Plantarum*, 19(4), 427-433.
- Foolad, M. R. (2004). Recent advances in genetics of salt tolerance in tomato. *Plant Cell, Tissue and Organ Culture*, 76(2), 101-119.
- Gairola, K. C., Nautiyal, A. R., & Dwivedi, A. K. (2011). Effect of temperatures and germination media on seed germination of *Jatropha curcas* Linn. *Advances In Bioresearch*, 2(2), 66-71.
- Ghanad, M., Nulit, R. B., Go, R., & Yien, C. Y. S. (2016). The effects of NaCl, KCl and MgCl₂ on the germination of *Brassica rapa* var. parachinensis seed. *International Journal of Biological Research*, 4(1), 52-55.
- Gharib, F. A., & Hegazi, A. Z. (2010). Salicylic acid ameliorates germination, seedling growth, phytohormone and enzymes activity in bean (*Phaseolus vulgaris* L.) under cold stress. *Journal of American Science*, 6(10), 675-683.
- Ghoulam, C., Foursy, A., & Fares, K. (2002). Effects of salt stress on growth, inorganic ions and proline accumulation in relation to osmotic adjustment in five sugar beet cultivars. *Environmental and experimental Botany*, 47(1), 39-50.
- Gregorio, G. B. (1997). Tagging salinity tolerance genes in rice using amplified fragment length polymorphism (AFLP). Ph.D. Thesis, University of the Philippines, Los Banose, 118p.

- Grime, J. P., & Campbell, B. D. (1991). *Growth rate, habitat productivity and plant strategy as predictors of stress response*. In H. A. Mooney, W. E. Winner, & E. J. Pell (Eds.). *Response of plants to multiple stresses*. Academic Press, Orlando. pp.143-159.
- Grubben, G. J. H., & Denton, O. A. (2004). *Plant Resources of Tropical Africa 2. Vegetables*. PROTA Foundation, Wageningen, Netherlands. *backhuys Publishers, Leiden, Netherlands/CTA, Wgeningen Netherlands*. [Http://www.hort.purdue.edu/newcrop.duke_energy/moringa, htm](http://www.hort.purdue.edu/newcrop.duke_energy/moringa.htm). Accessed on, 4(05), 2008.
- Habibi, A., & Abdoli, M. (2013). Influence of salicylic acid pre-treatment on germination, vigor and growth parameters of garden cress (*Lepidium sativum*) seedlings under water potential loss at salinity stress. *International Research Journal of Applied and Basic Sciences*, 4(6), 1393-1399.
- Haifa (2014) Nutritional recommendation for Cucumber in open fields, tunnels and greenhouse. Haifa Pioneering the future.pp5.<http://www.haifagroup.com/files/Guides/Cucumber.pdf>
- Hakim, M. A., Juraimi, A. S., Hanafi, M. M., Ali, E., Ismail, M. R., Selamat, A., & Karim, S. R. (2014). Effect of salt stress on morpho-physiology, vegetative growth and yield of rice. *Journal of Environmental Biology*, 35(2), 317.
- Hara, M., Furukawa, J., Sato, A., Mizoguchi, T., & Miura, K. (2012). Abiotic stress and role of salicylic acid in plants. *In Abiotic Stress Responses in Plants*. Springer, New York. pp. 235-251.
- Hasanuzzaman, M., Nahar, K., & Fujita, M. (2013). Plant response to salt stress and role of exogenous protectants to mitigate salt-induced damages. *In Ecophysiology and responses of plants under salt stress*. Springer, New York. pp. 25-87.
- Hasegawa, P.M., Bressan, R.A., Zhu, J.K. and Bohnert, H.J. 2000. Plant cellular and molecular responses to high salinity. *Annual Review of Plant Physiology and Plant Molecular Biology*, 51,463-499.
- Hashim, G. M. (2003). Salt-affected Soils of Malaysia. In *Proceedings of the Workshop on Soil Science in Malaysia towards 2020*.
- Hassan, N. A., Drew, J. V., Knudsen, D., & Olson, R. A. (1970). Influence of soil salinity on production of dry matter and uptake and distribution of nutrients in barley and corn: I. Barley (*Hordeum vulgare* L.). *Agronomy Journal*, 62(1), 43-45.
- Hayat, S., Ali, B., & Ahmad, A. (2007). Salicylic acid: biosynthesis, metabolism and physiological role in plants. In *Salicylic acid: A plant hormone*. Springer, Netherlands. pp. 1-14.

- Herrero, J., & Pérez-Coveta, O. (2005). Soil salinity changes over 24 years in a Mediterranean irrigated district. *Geoderma*, 125(3), 287-308.
- Hester, M. W., Mendelssohn, I. A., & McKee, K. L. (2001). Species and population variation to salinity stress in *Panicum hemitomon*, *Spartina patens*, and *Spartina alterniflora*: morphological and physiological constraints. *Environmental and Experimental Botany*, 46(3), 277-297.
- Hokmalipour, S. (2015). Effect of Salinity and Temperature on Seed Germination and Seed Vigor Index of Chicory (*chichoriumintynus* L.), Cumin (*CuminumCyminium* L.) and Fennel (*Foeniculum Vulgare*). *Indian Journal of Science and Technology*, 8(35).
- Hossain, N., Muhibbullah, M., Ali, K. M. B., & Molla, M. H. (2015). Relationship between Soil Salinity and Physico-chemical Properties of Paddy Field Soils of Jhilwanja Union, Cox's Bazar, Bangladesh. *Journal of Agricultural Science*, 7(10), 166.
- Hu, Y., Fromm, J., & Schmidhalter, U. (2005). Effect of salinity on tissue architecture in expanding wheat leaves. *Planta*, 220(6), 838-848.
- Hussain, I., Uddin, M. B., & Aziz, M. G. (2011). Optimization of antinutritional factors from germinated wheat and mungbean by Response Surface Methodology. *International Food Research Journal*, 18(3).
- Hussain, S., Khaliq, A., Matloob, A., Wahid, M. A., & Afzal, I. (2013). Germination and growth response of three wheat cultivars to NaCl salinity. *Soil Environ*, 32(1), 36-43. Germination and growth response of three wheat cultivars to NaCl salinity. *Soil Environ*, 32(1), 36-43.
- Hussein, M. M., & Abou-Baker, N. H. (2014). Growth and mineral status of moringa plants as affected by silicate and salicylic acid under salt stress. *International Journal of Plant & Soil Science, Tarakeswar, India*, 3, 163-177.
- Hussein, M. M., Balbaa, L. K., & Gaballah, M. S. (2007). Salicylic acid and salinity effects on growth of maize plants. *Research Journal of Agriculture and biological Sciences*, 3(4), 321-328.
- Isla, R., Aragüés, R., & Royo, A. (1998). Validity of various physiological traits as screening criteria for salt tolerance in barley. *Field Crops Research*, 58(2), 97-107.
- Jadhav, S. H., & Bhamburdekar, S. B. (2011). Effect of salicylic acid on germination performance in groundnut. *International Journal of Applied Biology and Pharmaceutical Technology*, 2(4):224-227.
- Jamil, M., & Rha, E. S. (2004). The effect of salinity (NaCl) on the germination and seedling of sugar beet (*Beta vulgaris* L.) and cabbage (*Brassica oleracea* L.). *Plant resources*, 7(3), 226-232.

- Jamil, M., Charnikhova, T., Houshyani, B., van Ast, A., & Bouwmeester, H. J. (2012). Genetic variation in strigolactone production and tillering in rice and its effect on *Striga hermonthica* infection. *Planta*, 235(3), 473-484.
- Kandil, A. A., Sharief, A. E., Abido, W. A. E., & Ibrahim, M. M. (2012). Effect of salinity on seed germination and seedling characters of some *forage sorghum* cultivars. *International Journal of Agriculture Sciences*, 4(7), 306.
- Kang, H. M., & Saltveit, M. E. (2002). Chilling tolerance of maize, cucumber and rice seedling leaves and roots are differentially affected by salicylic acid. *Physiologia Plantarum*, 115(4), 571-576.
- Karimi, E., Abdolzadeh, A., & Sadeghipour, H. R. (2012). Increasing salt tolerance in Olive, *Olea europaea* L. plants by supplemental potassium nutrition involves changes in ion accumulation and anatomical attributes. *International Journal of plant production*, 3(4), 49-60.
- Kaur, G., Kumar, S., Nayyar, H., & Upadhyaya, H. D. (2008). Cold Stress Injury during the Pod-Filling Phase in Chickpea (*Cicer arietinum* L.): Effects on Quantitative and Qualitative Components of Seeds. *Journal of Agronomy and Crop Science*, 194(6), 457-464.
- Kaymakanova, M. (2009). Effect of salinity on germination and seed physiology in bean (*Phaseolus vulgaris* L.). *Biotechnology & Biotechnological Equipment*, 23(sup1), 326-329.
- Keshavarzi, M. H. B. (2012). The effect of drought stress on germination and early growth of *Sesamum indicum* seedling's varieties under laboratory conditions. *International Journal of Agricultural Management and Development*, 2(4), 271-275.
- Khajeh-Hosseini, M., Powell, A. A., & Bingham, I. J. (2003). The interaction between salinity stress and seed vigour during germination of soyabean seeds. *Seed Science and technology*, 31(3), 715-725.
- Khan, M. A., Gul, B., & Weber, D. J. (2001). Effect of salinity on the growth and ion content of *Salicornia rubra*. *Communications in soil science and plant analysis*, 32(17-18), 2965-2977.
- Khayatnezhad, M., Gholamin, R., Jamaati-e-Somarin, S. H., & Zabihi-
emahmoodabad, R. (2010). Study of NaCl salinity effect on wheat (*Triticum aestivum* L.) cultivars at germination stage. *American-Eurasian Journal Agriculture Environ Science*, 9, 128-132.
- Kim, S. G., & Park, C. M. (2008). Gibberellic acid-mediated salt signaling in seed germination. *Plant Signaling & Behavior*, 3(10), 877-879.

- Kishor, P. K., Sangam, S., Amrutha, R. N., Laxmi, P. S., Naidu, K. R., Rao, K. R. S. S., ... & Sreenivasulu, N. (2005). Regulation of proline biosynthesis, degradation, uptake and transport in higher plants: its implications in plant growth and abiotic stress tolerance. *Current Science*, 88(3), 424-438.
- Kobayashi, H., Masaoka, Y., & Sato, S. (2005). Effects of excess magnesium on the growth and mineral content of rice and *Echinochloa*. *Plant production science*, 8(1), 38-43.
- Koyro, H. W. (2006). Effect of salinity on growth, photosynthesis, water relations and solute composition of the potential cash crop halophyte *Plantago coronopus* (L.). *Environmental and Experimental Botany*, 56(2), 136-146.
- Kurban, H., Saneoka, H., Nehira, K., Adilla, R., Premachandra, G. S., & Fujita, K. (1999). Effect of salinity on growth, photosynthesis and mineral composition in leguminous plant *Alhagi pseudoalhagi* (Bieb.). *Soil science and plant nutrition*, 45(4), 851-862.
- Kurum, R., Ulukapi, K., Aydinsakir, K., & Onus, A. N. (2013). The influence of salinity on seedling growth of some pumpkin varieties used as rootstock. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 41(1), 219.
- Lee, S., Kim, S. G., & Park, C. M. (2010). Salicylic acid promotes seed germination under high salinity by modulating antioxidant activity in *Arabidopsis*. *New Phytologist*, 188(2), 626-637.
- Leigh, R. A., & Wyn Jones, R. G. (1984). A hypothesis relating critical potassium concentrations for growth to the distribution and functions of this ion in the plant cell. *New Phytologist*, 97(1), 1-13.
- Li, J. Z., & Hong, Z. F. (2005). Effect of NaCl with CaCl stress on seed germination and seedling of cucumber. *China Journal of Jilin Agricultural University*, 27(2), 175-178.
- Li, Y. (2008). Effect of salt stress on seed germination and seedling growth of three salinity plants. *Pakistan journal of biological sciences: PJBS*, 11(9), 1268-1272.
- Llanes, A., Bertazza, G., Palacio, G., & Luna, V. (2013). Different sodium salts cause different solute accumulation in the halophyte *Prosopis strombulifera*. *Plant Biology*, 15(s1), 118-125.
- Llanes, A., Reinoso, H., & Luna, V. (2005). Germination and early growth of *Prosopis strombulifera* seedlings in different saline solutions. *World Journal of Agricultural Sciences*, 1(2):120-128.

- Lovato, M. B., & Lemos, P. M. F. J. (1994). Germination in *Stylosanthes humilis* populations in the presence of NaCl. *Australian Journal of Botany*, 42(6), 717-723.
- Maas, E. V. (1984). Salt tolerance of plants. *Handbook of plant science in agriculture*, 2, 57-75.
- Maas, E. V., & Grieve, C. M. (1990). Spike and Leaf Development of Sal-Stressed Wheat. *Crop Science*, 30(6), 1309-1313.
- Mahajan, S., & Tuteja, N. (2005). Cold, salinity and drought stresses: an overview. *Archives of biochemistry and biophysics*, 444(2), 139-158.
- Mane, A. V., Deshpande, T. V., Wagh, V. B., Karadge, B. A., & Samant, J. S. (2011). A critical review on physiological changes associated with reference to salinity. *International Journal of Environmental Sciences*, 1(6), 1192.
- Mansour, M. M. F. (1997). Cell permeability under salt stress. In :JaiwalP.K.,Gulati A.ed.s.):Strategies For Improving Salt Olerance In Higher Plants.Oxford and IBH Publishing CO., New Delhi.p 87-110
- Matsuoka, M. (2003). Gibberellin signaling: how do plant cells respond to GA signals. *Journal of Plant Growth Regulation*, 22(2), 123-125.
- Meloni, D. A., Oliva, M. A., Ruiz, H. A., & Martinez, C. A. (2001). Contribution of proline and inorganic solutes to osmotic adjustment in cotton under salt stress. *Journal of Plant Nutrition*, 24(3), 599-612.
- Mian, A. A., Senadheera, P., & Maathuis, F. J. (2011). Improving crop salt tolerance: anion and cation transporters as genetic engineering targets. *Plant Stress*, 5, 64-72.
- Mitra, G. N. (2015). Chloride (Cl⁻) Uptake. *In Regulation of Nutrient Uptake by Plants* (pp. 167-173). Springer India.
- Mittler, R. (2002). Oxidative stress, antioxidants and stress tolerance. *Trends in plant science*, 7(9), 405-410.
- MOA (Ministry of agriculture (2014). Perangkaan agromakanan 2014/ Agrofood statistics (2014) information, PJ:PP 60-63
- Mondo, V. H. V., Cicero, S. M., Dourado-Neto, D., Pupim, T. L., & Dias, M. A. N. (2013). Seed vigor and initial growth of corn crop. *Journal of Seed Science*, 35(1), 64-69.
- Moradi, F., Ismail, A. M., Egdane, J., & Gregorio, G. B. (2003). Salinity tolerance of rice during reproductive development and association with tolerance at the seedling stage. *Indian Journal Of Plant Physiology*, 8(1), 105-116.

- Morais, M. C., Panuccio, M. R., Muscolo, A., & Freitas, H. (2012). Salt tolerance traits increase the invasive success of *Acacia longifolia* in Portuguese coastal dunes. *Plant Physiology and Biochemistry*, 55, 60-65.
- Morais, M. C., Panuccio, M. R., Muscolo, A., & Freitas, H. (2012a). Does salt stress increase the ability of the exotic legume *Acacia longifolia* to compete with native legumes in sand dune ecosystems. *Environmental and Experimental Botany*, 82, 74-79.
- Mostafa, D. M. (2004). Metabolic imbalance and salinity tolerance of two maize cultivars (Doctoral dissertation, M. Sc. Thesis. El-Minia University. Elminia, Egypt 1-195).
- Moud, A. M., & Maghsoudi, K. (2008). Salt stress effects on respiration and growth of germinated seeds of different wheat (*Triticum aestivum* L.) cultivars. *World Journal of Agricultural Sciences*, 4(3), 351-358.
- Munns, R. (2002). Comparative physiology of salt and water stress. *Plant, cell & environment*, 25(2), 239-250.
- Munns, R. (2005). Genes and salt tolerance: bringing them together. *New phytologist*, 167(3), 645-663
- Munns, R., & Tester, M. (2008). Mechanisms of salinity tolerance. *Annual Review of Plant Biology*, 59, 651-681.
- Munns, R., James, R. A., & Läuchli, A. (2006). Approaches to increasing the salt tolerance of wheat and other cereals. *Journal of Experimental Botany*, 57(5), 1025-1043
- Mutlu, S., Karadağoğlu, Ö., Atici, Ö., & Nalbantoğlu, B. (2013). Protective role of salicylic acid applied before cold stress on antioxidative system and protein patterns in barley apoplast. *Biologia Plantarum*, 57(3), 507-513.
- Naseri, R., Mirzaei, A., Emami, T., & Vafa, P. (2012). Effect of salinity on germination stage of rapeseed cultivars (*Brassica napus* L.). *International Journal of Crop Science*, 4(13), 918-922.
- Nawaz, K., Hussain, K., Majeed, A., Khan, F., Afghan, S., & Ali, K. (2010). Fatality of salt stress to plants: Morphological, physiological and biochemical aspects. *African Journal of Biotechnology*, 9(34).
- Naz, F., Gul, H., & Hamayun, M. (2014). Effect of NaCl Stress on *Pisum sativum* Germination and Seedling Growth with the Influence of Seed Priming with Potassium (KCl and KOH), *American-Eurasian Journal Agriculture & Environ Science*, 14 (11), 1304-1311.

- Noreen, S. I. B. G. H. A., & Ashraf, M. U. H. A. M. M. A. D. (2008). Alleviation of adverse effects of salt stress on sunflower (*Helianthus annuus* L.) by exogenous application of salicylic acid: growth and photosynthesis. *Pakistan Jurnal Botany*, 40(4), 1657-1663.
- Nozulaidi, M., Khairi, M., & Jahan, M. S. (2015). Effects of different salinity levels on rice production. *Austuralia Journal of Basic and Applied Science*, 9, 524-530.
- Okçu, G., Kaya, M. D., & Atak, M. (2005). Effects of salt and drought stresses on germination and seedling growth of pea (*Pisum sativum* L.). *Turkish journal of agriculture and forestry*, 29(4), 237-242.
- Ola, H., Reham, E. F., Eisa, S. S., & Habib, S. A. (2012). Morpho-anatomical changes in salt stressed kallar grass (*Leptochloa fusca* L. Kunth). *Research Journal of Agriculture and Biological Sciences*, 8(2), 158-166.
- Omami, E. N. (2005). Response of *Amaranth* to salinity stress (Doctoral dissertation, University of Pretoria).
- Owino, D. O., & Ouma, G. (2011). Effect of potassium priming on papaya (*Carica papaya* var. kamiya). *Journal of Animal & Plant Sciences*, 11(2), 1418-1423.
- Pal, P., Yadav, K., Kumar, K., & Singh, N. (2016) Cumulative effect of potassium and gibberellic acid on growth, biochemical attributes and productivity of F1 hybrid cucumber. *Environmental and Experimental Biology* 14: 57–61
- Panuccio, M. R., Jacobsen, S. E., Akhtar, S. S., & Muscolo, A. (2014). Effect of saline water on seed germination and early seedling growth of the halophyte *quinoa*. *Annals of Botany* , 6, plu047.
- Pareek, A., Singla, S. L., & Grover, A. (1997). Salt Responsive Proteins/Genes In Crop Plants. In *Strategies For Improving Salt Tolerance In Higher Plants* (eds P.K.)Jaiwal,R.P.singh, and A.Gulati Oxford and IBH Puplication Co.,New Delhi, pp. 365-391
- Parida, A. K., & Das, A. B. (2005). Salt tolerance and salinity effects on plants: a review. *Ecotoxicology and environmental safety*, 60(3), 324-349.
- Parida, A. K., Das, A. B., & Mitra, B. (2004). Effects of salt on growth, ion accumulation, photosynthesis and leaf anatomy of the *mangrove*, *Bruguiera parviflora*. *Trees*, 18(2), 167-174.
- Pessarakli, M. (2001). Handbook of Plant and Crop Physiology. Second Edition. Marcel Dekker, Inc, NewYork. pp. 139, 569.

- Pirasteh-Anosheh, H., & Emam, Y. (2012). Manipulation of morpho-physiological traits in bread and durum wheat by using PGRs at different water regimes. *Journal of Crop production and processing*, 5, 29-45.
- Pirasteh-Anosheh, H., Emam, Y., Ashraf, M., & Foolad, M. R. (2012). Exogenous application of salicylic acid and chlormequat chloride alleviates negative effects of drought stress in wheat. *Advanced Studies in Biology*, 11(4), 501-520.
- Prajapati, K., & Modi, H. A. (2012). The importance of potassium in plant growth—A review. *Indian Journal of Plant Sciences*, 1, 177-186.
- Rabie, G. H. (2005). Influence of arbuscular mycorrhizal fungi and kinetin on the response of mungbean plants to irrigation with seawater. *Mycorrhiza*, 15(3), 225-230.
- Rahman, S., Miyake, H., & Takeoka, Y. (2002). Effects of exogenous glycinebetaine on growth and ultrastructure of salt-stressed rice seedlings (*Oryza sativa* L.). *Plant Production Science*, 5(1), 33-44.
- Rajabi, A., Khayamim, S., Abbasi, Z., & Ober, E. (2014). Salt Stress and Sugar Beet Improvement: *Challenges and Opportunities*. In *Improvement of Crops in the Era of Climatic Changes* (pp. 121-150). Springer, New York.
- Rajjou, L., Duval, M., Gallardo, K., Catusse, J., Bally, J., Job, C., & Job, D. (2012). Seed germination and vigor. *Annual Review of Plant Biology*, 63, 507-533.
- Ramoliya, P. J., & Pandey, A. N. (2003). Soil salinity and water status affect growth of Phoenix dactylifera seedlings. *New Zealand Journal of Crop and Horticultural Science*, 31(4), 345-353.
- Rao, K., Raghavendra, A., & Reddy, K. (2006). Physiology and molecular biology of stress tolerance in plants. *Springer Science & Business Dordrecht*: Springer. pp 43, 68, 75.
- Ratnakar, A., & Rai, A. (2013). Effect of sodium chloride salinity on seed germination and early seedling growth of *Trigonella foenum-graecum* L. var. Peb. *Octa Journal of Environmental Research*, 1(4).
- Reddy, M. A., Francies, R. M., Rasool, S. N., & Reddy, V. R. P. (2014). Breeding for tolerance to stress triggered by salinity in rice. *International Journal of Applied Biology and Pharmaceutical Technology*, 5(1), 167-176.

- Renner, S. S., Schaefer, H., & Kocyan, A. (2007). Phylogenetics of Cucumis (Cucurbitaceae): Cucumber (*C. sativus*) belongs in an Asian/Australian clade far from melon (*C. melo*). *Bio Med Central Evolutionary Biology*, 7(1), 1.
- Rodríguez, A. A., Stella, A. M., Storni, M. M., Zulpa, G., & Zaccaro, M. C. (2006). Effects of cyanobacterial extracellular products and gibberellic acid on salinity tolerance in *Oryza sativa* L.. *Saline Systems*, 2(1), 1.
- Rodríguez, P., Torrecillas, A., Morales, M. A., Ortuno, M. F., & Sánchez-Blanco, M. J. (2005). Effects of NaCl salinity and water stress on growth and leaf water relations of *Asteriscus maritimus* plants. *Environmental and Experimental Botany*, 53(2), 113-123.
- Romero-Aranda, R., Soria, T., & Cuartero, J. (2001). Tomato plant-water uptake and plant-water relationships under saline growth conditions. *Plant Science*, 160 (2), 265-272.
- Rowell, D. L., & Wild, A. (1988). Soil acidity and alkalinity. Russell's soil conditions and plant growth. *Eleventh edition*, 844-898.
- Salama, K. H., Mansour, M. M., & Hassan, N. S. (2011). Choline priming improves salt tolerance in wheat (*Triticum aestivum* L.). *Journal of Basic and Applied Sciences*, 5, 126-132.
- Sangha, M. K. (2015). Screening and biochemical analyses for salinity tolerance in cotton (Doctoral dissertation, PAU).
- Santelia, D., & Lawson, T. (2016). Rethinking Guard Cell Metabolism. *Plant Physiology*, 172(3), 1371-1392.
- Sato, S., Sakaguchi, S., Furukawa, H., & Ikeda, H. (2006). Effects of NaCl application to hydroponic nutrient solution on fruit characteristics of tomato (*Lycopersicon esculentum* Mill.). *Scientia horticultrae*, 109(3), 248-253.
- Shaaban, S. A. S. (2016). Botanical Studies on Wheat Plants (*Triticum aestivum* L.) Grown Under Saline Conditions And Its Response To Foliar Application By Some Organic Substances. Cu Theses.
- Shakirova, F. M., Sakhabutdinova, A. R., Bezrukova, M. V., Fatkhutdinova, R. A., & Fatkhutdinova, D. R. (2003). Changes in the hormonal status of wheat seedlings induced by salicylic acid and salinity. *Plant Science*, 164(3), 317-322.
- Shannon, M. C., Grieve, C. M., & Francois, L. E. (1994). Whole-plant response to salinity. *Plant-environment interactions*, 199-244

- Shereen, A., Mumtaz, S., Raza, S., Khan, M. A., & Solangi, S. (2005). Salinity effects on seedling growth and yield components of different inbred rice lines. *Pakistan Journal Botany* 37(1), 131-139
- Simon-Sarkadi, L., Kocsy, G., & Sebestyén, Z. (2002). Effect of salt stress on free amino acid and polyamine content in cereals. *Acta Biol Szegediensis*, 46, 73-75.
- Sohan, D., Jasoni, R., & Zajicek, J. (1999). Plant–water relations of NaCl and calcium-treated sunflower plants. *Environmental and Experimental Botany*, 42(2), 105-111.
- Sohrabikertabad, S., Ghanbari, A., Mohassel, M., Mahalati, M. N., & Gherekhloo, J. (2013). Effect of desiccation and salinity stress on seed germination and initial plant growth of *Cucumis melo*. *Planta Daninha*, 31(4), 833-841.
- Soo, S. W. (1972). Semi-detailed Soil Survey of the Kedah-Perlis Coastal Plain. Kuala Lumpur: Department of Agriculture.
- Soo, S.W. 1984. Semi-detailed Soil Survey of North-West Selangor. *Soils & Analytical Services Branch, Ministry of Agric.*, Kuala Lumpur.
- Sosa, L., Llanes, A., Reinoso, H., Reginato, M., & Luna, V. (2005). Osmotic and specific ion effects on the germination of *Prosopis strombulifera*. *Annals of Botany*, 96(2), 261-267.
- Sparks, D. L., & Huang, P. M. (1985). Physical chemistry of soil potassium. *Potassium in agriculture*, (potassiuminagri), 201-276.
- Srivastava, N. K., & Srivastava, A. K. (2007). Influence of gibberellic acid on ¹⁴CO₂ metabolism, growth, and production of alkaloids in *Catharanthus roseus*. *Photosynthetica*, 45(1), 156-160.
- Steduto, P., Albrizio, R., Giorio, P., & Sorrentino, G. (2000). Gas-exchange response and stomatal and non-stomatal limitations to carbon assimilation of sunflower under salinity. *Environmental and Experimental Botany*, 44(3), 243-255.
- Sudharani, M., Reddy, P. R., & Jayalakshmi, V. (2012). A Comprehensive Review On “Genetic Components Of Salinity Tolerance In Rice (*Oryza Sativa* L.)”. *International Journal of Applied Biology and Pharmaceutical Technol*, , ((3, ,312-322.
- Szepesi, Á., Csiszár, J., Gémes, K., Horváth, E., Horváth, F., Simon, M. L., & Tari, I. (2009). Salicylic acid improves acclimation to salt stress by stimulating abscisic aldehyde oxidase activity and abscisic acid accumulation, and increases Na⁺ content in leaves without toxicity symptoms in *Solanum lycopersicum* L. *Journal of plant physiology*, 166(9), 914-925.

- Tabaei-Aghdaei, S., & Harrison, P. (2000). Expression of dehydration-stress related genes in crown of wheat grass species having contrasting acclimation to salt, cold and drought. *Plant Cell Environ*, 23, 561-571.
- Taiz, L & Zeiger, E. (2006). *Plant Physiology*. Fourth Edition. *Sinauer, Sunderland, Mass, USA*. Pp 33, 612.
- Talbott, L. D., & Zeiger, E. (1996). Central roles for potassium and sucrose in guard-cell osmoregulation. *Plant Physiology*, 111(4), 1051-1057.
- Teh, C. Y., Mahmood, M., Shaharuddin, N. A., & Ho, C. L. (2015). In vitro rice shoot apices as simple model to study the effect of NaCl and the potential of exogenous proline and glutathione in mitigating salinity stress. *Plant Growth Regulation*, 75(3), 771-781
- Thakur, P., Kumar, S., Malik, J. A., Berger, J. D., & Nayyar, H. (2010). Cold stress effects on reproductive development in grain crops: an overview. *Environmental and Experimental Botany*, 67(3), 429-443.
- Thomson, M. J., Ocampo, D. M., Egdane, J., Katimbang, M., Singh, R. K., Gregorio, G., & Ismail, M. (2007, January). QTL mapping and marker assisted backcrossing for improved salinity tolerance in rice. *In Plant and animal genomes XV conference, San Diego, CA* (pp. 13-17).
- Tobe, K., Li, X., & Omasa, K. (2004). Effects of five different salts on seed germination and seedling growth of *Haloxylon ammodendron* (Chenopodiaceae). *Seed Science Research*, 14(04), 345-353.
- Trichopoulou, A., Lagiou, P., Kuper, H., & Trichopoulos, D. (2000). Cancer and Mediterranean dietary traditions. *Cancer Epidemiology Biomarkers & Prevention*, 9(9), 869-873.
- Tsegay, B. A., & Gebreslassie, B. (2014). The effect of salinity (NaCl) on germination and early seedling growth of *Lathyrus sativus* and *Pisum sativum* var. *abyssinicum*. *African Journal of Plant Science*, 8(5), 225-231.
- Turhan, A., Kuşçu, H., & Şeniz, V. (2011). Effects of different salt concentrations (NaCl) on germination of some spinach cultivars. *Uludağ Üniversitesi Ziraat Fakültesi Dergisi*, 25(1).
- USDA (1954) Reclamation and improvement of saline and sodic soils. USDA Handbook 60. Riverside, California.
- Vicente, O., Boscaiu, M., Naranjo, M. Á., Estrelles, E., Bellés, J. M., & Soriano, P. (2004). Responses to salt stress in the halophyte *Plantago crassifolia* (Plantaginaceae). *Journal of Arid Environments*, 58(4), 463-481.

- Vinod, K. K., Krishnan, S. G., Babu, N. N., Nagarajan, M., & Singh, A. K. (2013). Improving salt tolerance in rice: Looking beyond the conventional. *In Salt Stress in Plants* (pp. 219-260). Springer New York.
- Vorasoot, N., Songsri, P., Akkasaeng, C., Jogloy, S., & Patanothai, A. (2003). Effect of water stress on yield and agronomic characters of peanut (*Arachis hypogaea* L.). *Songklanakar J. Sci. Technol*, 25(3), 283-288.
- Wallender, W. W., & Tanji, K. K. (2011). Agricultural salinity assessment and management (No. Ed. 2). *American Society of Civil Engineers* (ASCE) No. 71. chapter 11 pp:343-370.
- Wang, Y., & Wu, W. H. (2013). Potassium transport and signaling in higher plants. *Annual Review of Plant Biology*, 64, 451-476.
- Wang, Z. L., Li, P. H., Fredricksen, M., Gong, Z. Z., Kim, C. S., Zhang, C., ... & Zhao, Y. X. (2004). Expressed sequence tags from *Thellungiella halophila*, a new model to study plant salt-tolerance. *Plant Science*, 166(3), 609-616.
- Wankhade, S. D., Bahaji, A., Mateu-Andrés, I., & Cornejo, M. J. (2010). Phenotypic indicators of NaCl tolerance levels in rice seedlings: variations in development and leaf anatomy. *Acta physiologiae plantarum*, 32(6), 1161-1169.
- War, A. R., Paulraj, M. G., War, M. Y., & Ignacimuthu, S. (2011). Role of salicylic acid in induction of plant defense system in chickpea (*Cicer arietinum* L.). *Plant signaling & behavior*, 6(11), 1787-1792.
- Wehner, T. C., & Guner, N. (2002, August). Growth stage, flowering pattern, yield, and harvest date prediction of four types of cucumber tested at 10 planting dates. In XXVI International Horticultural Congress: *Advances in Vegetable Breeding* 637 (pp. 223-230).
- Wicke, B., Smeets, E., Dornburg, V., Vashev, B., Gaiser, T., Turkenburg, W., & Faaij, A. (2011). The global technical and economic potential of bioenergy from salt-affected soils. *Energy & Environmental Science*, 4(8), 2669-2681.
- Wong, N. C. & Jaafar, A. M.: 1993, Soil Chemical Characteristics of Vegetable Plots at Cameron Highlands, MARDI Report No. 170, pp 9.
- Yadav, S., Irfan, M., Ahmad, A., & Hayat, S. (2011). Causes of salinity and plant manifestations to salt stress: A review. *Journal of Environmental Biology*, 32(5), 667
- Yalpani, N., Enyedi, A. J., León, J., & Raskin, I. (1994). Ultraviolet light and ozone stimulate accumulation of salicylic acid, pathogenesis-related proteins and virus resistance in tobacco. *Planta*, 193(3), 372-376.

- ZHANG, Z. G., & SHANG, Q. M. (2010). Promoting effect of salicylic acid and chitosan on germination of cucumber seeds under NaCl stress. *China Vegetables*, 8, 008.
- Zhani, K., Ben, F. M., Mani, F., & Hannachi, C. (2012). Impact of salt stress (NaCl) on growth, chlorophyll content and fluorescence of Tunisian cultivars of chili pepper (*Capsicum frutescens* L.). *Journal of Stress Physiology & Biochemistry*, 8(4).
- Zhao, J., Ren, W., Zhi, D., Wang, L., & Xia, G. (2007). Arabidopsis DREB1A/CBF3 bestowed transgenic tall fescue increased tolerance to drought stress. *Plant Cell Reports*, 26(9), 1521-1528.
- Zhu, J. K. (2000). Genetic analysis of plant salt tolerance using Arabidopsis. *Plant Physiology*, 124(3), 941-948.
- Zhu, J. K. (2001). Plant salt tolerance. *Trends in Plant Science*, 6(2), 66-71.
- Zhu, J. K. (2002). Salt and drought stress signal transduction in plants. *Annual review of plant biology*, 53, 247.