



UNIVERSITI PUTRA MALAYSIA

***EFFECTS OF PRENATAL AND POSTNATAL TOCOTRIENOL- RICH
FRACTION SUPPLEMENTATION ON COGNITIVE FUNCTION
DEVELOPMENT IN F1 MALE RATS***

GOWRI A/P NAGAPAN

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By

GOWRI A/P NAGAPAN

**Thesis submitted to the School of Graduate Studies,
Universiti Putra Malaysia, in fulfillment of the Requirement for the Degree of
Master of Science**

August 2014

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

EFFECTS OF PRENATAL AND POSTNATAL TOCOTRIENOL- RICH FRACTION SUPPLEMENTATION ON COGNITIVE FUNCTION DEVELOPMENT IN F1 MALE RATS

By

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August 2014

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The intake of specific nutrients during the critical period in early life influence cognitive and behavioural development profoundly. Antioxidants such as vitamin E have been postulated to be pivotal in this process, as vitamin E is able to protect the growing brain from oxidative stress. Currently, tocotrienols are gaining much attention due to their potent antioxidant and neuroprotective properties. It is thus compelling to look at the effects of prenatal and early postnatal tocotrienols supplementation, on cognition and behavioural development among offspring of individual supplemented with tocotrienols. Therefore, this study was aimed to investigate potential prenatal and early postnatal influence of Tocotrienol-Rich Fraction (TRF) supplementation on vitamin E content in plasma and tissues; fatty acid profiles in brain and cognitive function developments in male F1 rats.

Eight-week-old adult female Sprague Dawley (SD) rats (F0 generation) were randomly assigned into five groups of two animals each. The animals were fed either with the base diet as control (CTRL), base diet plus vehicle (VHCL), base diet plus docosahexanoic acid (DHA), base diet plus Tocotrienol-Rich fraction (TRF), and base diet plus both docosahexanoic acid, and tocotrienol rich fraction (DTRF) diets for two weeks prior to mating. The F0 females were maintained on their respective treatment diets throughout the gestation and lactation periods. Pups (F1 generation) derived from these dams were raised with their dams from birth till four weeks post natal. The male pups were weaned at eight weeks postnatal, after which they were grouped into five groups of ten animals each, and fed with the same diets as their dams for another eight weeks. Learning and behavioural experiments were conducted only in F1 male rats using the Morris water maze (MWM).

The vitamin E content of the diets, plasma and tissues of the male F1 rats were determined using standard extraction and High Performance Liquid Chromatography procedures. Results showed that vitamin E content, α -tocopherol and α -tocotrienol were found increased in the plasma, brain and liver of the TRF and DTRF groups than CTRL group ($P<0.05$). δ -Tocotrienol was found increased in the adipose of TRF and DTRF groups than CTRL group ($P<0.05$). Results also showed that the DHA content and total n-3 PUFA in the brain of the DHA and DTRF groups were significantly higher and their n-3: n-6 ratio significantly lower than CTRL group ($P<0.05$). TRF supplementation had no detectable effects on the PUFA profiles of the brain tissues of male F1 rats. Animals supplemented with either DHA and/or TRF such as DHA, TRF and DTRF groups demonstrated better spatial learning and re-learning ability compared to the CTRL animals ($P<0.05$). The cognitive parameters of TRF supplemented animals from the current study were comparable to that of the DHA group which was a positive control for this experiment.

In conclusion, the current study demonstrated that prenatal and postnatal intake of TRF increases the α -tocotrienol level in the progeny's brain. This results in better behavioural performance and cognitive function development in the F1 progeny.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Master Sains

**KESAN SUPLEMENTASI PECAHAN KAYA-TOKOTRIENOL (TRF)
SEMASA PRANATAL DAN PASCANATAL KE ATAS PERKEMBANGAN
FUNGSI KOGNITIF PADA TIKUS JANTAN F1**

Oleh

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Pengambilan nutrien khusus mempengaruhi perkembangan kognitif dan tingkah laku dalam kehidupan awal. Antioksidan seperti vitamin E memainkan peranan penting dalam proses ini. Ini kerana vitamin E dapat melindungi otak daripada tegasan oksidatif. Tokotrienol semakin banyak mendapat perhatian sebagai komponen penting antioksidan dan pelindung saraf pada masa ini. Sehubungan itu, adalah penting untuk mengkaji kesan suplementasi tokotrienol semasa pranatal dan pascanatal pada perkembangan fungsi kognitif. Kajian ini bertujuan untuk menyelidik kesan pengambilan pecahan kaya-tokotrienol (*Tocotrienol-rich fraction, TRF*) semasa pranatal dan pascanatal pada tahap vitamin E, profil asid lemak dalam tisu dan plasma, serta kesannya ke atas perkembangan fungsi kognitif pada progeni tikus jantan F1.

Tikus betina Sprague Dawley (SD) dewasa berumur lapan minggu telah dibahagikan secara rawak kepada lima kumpulan yang terdiri daripada dua haiwan setiap kumpulan. Tikus ini diberi makan diet rawatan yakni, diet asas (CTRL), diet ditambah dengan pengantara (VHCL), diet ditambah dengan asid dokosaheksanoik (DHA), diet dengan pecahan kaya-tokotrienol (TRF), dan diet ditambah dengan asid dokosaheksanoik serta pecahan kaya-tokotrienol (DTRF) selama dua minggu sebelum mengawan dengan tikus dewasa jantan. Tikus betina generasi F0 diberi makan diet rawatan masing-masing sepanjang tempoh bunting dan laktasi. Progeni tikus jantan dicerai susu pada masa lapan minggu. Selepas itu tikus jantan F1 telah dikumpulkan ke dalam lima kumpulan dengan sepuluh haiwan setiap kumpulan, dan dirawat dengan diet yang sama seperti induk tikus masing-masing selama lapan minggu lagi. Penilaian

fungsi kognitif hanya dijalankan pada progeni jantan F1 menggunakan Instrumen Pagar Sesat Air Morris (*Morris Water Maze*, MWM).

Tahap kandungan vitamin E dalam diet, plasma dan tisu tikus jantan F1 ditentukan menggunakan kaedah Kromatografi Cecair Berprestasi Tinggi (*High Performance Liquid Chromatography*, HPLC). Keputusan menunjukkan bahawa tahap vitamin E, α - tokoferol dan α - Tokotrienol didapati meningkat dalam plasma, otak dan hati pada kumpulan TRF dan DTRF berbanding kumpulan CTRL ($P < 0.05$). δ -tokotrienol didapati meningkat dalam tisu adipos daripada kumpulan TRF dan DTRF berbanding CTRL ($P < 0.05$). Keputusan juga menunjukkan bahawa kandungan DHA dan asid lemak poli tak tepu n-3 (PUFA n-3) telah meningkat dalam otak haiwan dari kumpulan DHA dan DTRF dengan nisbah PUFA n-3 : PUFA n-6 yang lebih rendah berbanding kumpulan CTRL ($P < 0.05$). Suplementasi TRF juga tidak memberi kesan signifikan kepada profil asid lemak politaktepu otak tikus jantan F1. Hasil kajian menunjukkan bahawa haiwan yang diberi makan sama ada DHA dan/atau TRF menunjukkan prestasi fungsi kognitif yang lebih baik berbanding dengan haiwan CTRL ($P < 0.05$). Tahap fungsi kognitif kumpulan TRF daripada kajian ini juga setanding dengan kumpulan DHA yang merupakan haiwan kawalan positif dalam eksperimen ini.

Kesimpulannya, kajian ini menunjukkan bahawa suplementasi TRF semasa pranatal dan pascanatal meningkatkan tahap α -tocotrienol dalam tisu otak. Ini seterusnya berkait dengan peningkatan fungsi kognitif di kalangan progeni jantan F1.

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I certify that a Thesis Examination Committee has met on 26 August 2014 to conduct the final examination of **Gowri Nagapan** on her thesis entitled "**Effects of Prenatal and Postnatal Tocotrienol- Rich Fraction Supplementation on Cognitive Function Development in F1 Male Rats**" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

ABC	ATP binding cassette
AD	Alzheimer's disease
APO B	apolipoprotein B
BDNF	Brain derived neurotropic factor
CA	Cornu ammonis
CEHC	Carboxyethyl hydroxychroman
CNS	Central nervous system (CNS)
EDTA	Ethylene-diamine-tetra-acetate
FAC	Fatty acid composition
FAME	Fatty Acid Methyl Esters
GC	Gas-liquid chromatography
GRAS	Generally regarded has safe
HDL	High-density lipoproteins
HMG-CoA	3-hydroxy-3-methylglutaryl coenzyme A
HO-3	heme oxygenase 3
HPLC	High Performance Liquid Chromatography
L1	LINE-1
LDL	Low-density lipoprotein
LPL	Lipoprotein lipase
LTD	Long-term depression
LTM	Long- term memory
LTP	Long-term potentiation
MDR2	multidrug resistance 2 GeneTransporter glycoprotein
MWM	Morris water maze
NaCl	Sodium chloride
PMC	2, 2, 5, 7, 8-pentamethyl-6-hydroxychroman
ppm	Parts per million

PUFA	Polyunsaturated fatty acid
ROS	Reactive oxygen species
SREBPs	Sterol regulatory element binding protein
STM	Short- term memory
VLDL	Very-low-density lipoprotein
α -CEHC	2,5,7,8-tetramethyl-2(2'carboxyyethyl)-6-hydroxychroman
α -TTP	α -tocopherol-transfer protein
γ -CEHC	2,7,8-trimethyl-2(2'carboxyyethyl)-6-hydroxychroman
α -	Alpha
β -	Beta
γ -	Gamma
δ -	Delta



CHAPTER 1

INTRODUCTION

1.1 Overview

The brain requires adequate nutrition for optimum growth, development and maturation. Several micronutrients and polyunsaturated fatty acids (PUFAs) are known to be essential for proper structure of brain tissue, healthy neurochemistry and maturation of the brain (Bourre, 2006a; Bourre, 2006b). Adequate intake of these nutrients results in proper cognitive development. As the brain development occurs during foetal and early postnatal life, inadequate maternal intake of nutrition not only alters the brain development, but also causes impact on cognitive abilities that can have long lasting and irreversible effect (Lucas *et al.*, 2001a). In the developing brain, macronutrients such as proteins and PUFAs are critically important for the synthesis of nucleic acids, neurotransmitters, growth factors and formation of cell membranes, synapse and myelin (Bourre, 2006b). Micronutrient such as vitamin E is important in developing brain as an antioxidant to protect the vulnerable brain cells from lipid peroxidation and also for normal neurological function (Ramakrishna, 1999).

The brain develops very rapidly during the last trimester of foetal life (prenatal) and the first two years (postnatal) of childhood (Rassin *et al.*, 2003). Research has shown that exposure to certain conditions such as diseases (gestational diabetes, hypertension), nutrient deficiency, exposure to alcohol and smoking can have long lasting effects on cognition (Roseboom *et al.*, 2012). Specifically, n-3 fatty acids, iodine, iron and choline have been shown to influence brain development and impact cognitive ability and behaviour in human and animals (Lucas *et al.*, 2001b). Animal studies have shown that low n-3 PUFAs intake leads to decrease levels of n-3 PUFAs in the brain and in turn resulted in memory impairment and behavioural depression (Su H.M., 2010; Watanabe *et al.*, 2004). Other nutrient such as vitamin E also had demonstrated significant neuroprotective properties (Khanna *et al.*, 2003, 2005; Sen *et al.*, 2000, 2003). The brain is highly susceptible to oxidative damage as it rich in PUFAs that form plasma membrane of neural cells. Vitamin E has been regarded as an important component of biological membrane as it may act as chain-breaking antioxidant to protect lipid membrane from oxidative damage (Suzuki *et al.*, 1993). Tocotrienols in particular have been reported to exert antioxidant activity in many *in vitro* and *in vivo* systems (Ghafoorunissa *et al.*, 2004; Nesaretnam *et al.*, 1993; Noguchi *et al.*, 2003; Yoshida *et al.*, 2003; Yoshida *et al.*, 2007; Qureshi *et al.*, 2000). Tocotrienols may quench lipid radicals and in the process become radicals themselves, albeit less reactive ones. These radicals then reduced back to native form by vitamin C or indirectly by thiol antioxidants (glutathione and lipoic acid) (Rimbach *et al.*, 2002).

Vitamin E is a lipid soluble natural antioxidant, which comprised of two major families of tocopherols and tocotrienols. The tocotrienols are the major form of vitamin E found in palm oil with the ratio of 30 % tocopherols and 70 % tocotrienols (Aggarwal *et al.*, 2012). Accumulation of tocotrienol in tissue has very tangible health benefits. Studies have shown that tocotrienols have potent antioxidant (Azlina *et al.*, 2005), cardioprotective (Das *et al.*, 2008), hypocholesterolemic (Qureshi *et al.*, 1986, 1991, 2000, 2001) anti-cancer (Sundram *et al.*, 1989; Komityama *et al.*, 1989; Nesaretnam *et al.*, 2004, 2008;) and neuroprotective (Khanna *et al.*, 2003, 2005; Sen *et al.*, 2000, 2003) properties that are not exhibited by tocopherols. Tocotrienol also is a nutrient certified as GRAS (generally regarded as safe) by the U.S. Food and Drug Administration (US FDA) GRAS notice number (GRN) 307 in April 2010 (Park *et al.*, 2011).

The integrity of the neural membrane is crucial to the neural functions. For living beings, the nervous system acts as a central coordinating centre for the various physiological, locomotor, and cognitive functions central to an animal's survival. Cognition refers to the mental processes involve in memory and learning (Wainwright & Colombo, 2006). Various factors such as nutrition, environment and genetic have strong influences on memory and learning. Nutrition affects cognition and mental health as brain structure and function are dependent on nutritional inputs (Dauncey & Bicknell, 1999). Various dietary factors such as n-3 fatty acids, antioxidants vitamins, minerals, curcumins and flavonoids have been identified as having beneficial effects on cognition (Gomez, 2008). This is because these dietary factors can affect multiple brain processes by regulating neurotransmitter pathways, synaptic transmission, membrane fluidity and signal transduction pathways associated with synaptic plasticity (Gomez, 2008). In contrast, diets rich in saturated and trans fats are known to affect cognition adversely. This is typically attributed to the role of these fats in reducing the synaptic plasticity mediated by the hippocampal brain derived neurotropic factor (BDNF) mediated synaptic plasticity (Molteni, 2002).

Accumulation of tocotrienols in the brain is required to protect vulnerable neurons from oxidative stress, enhance existing neuronal function and stimulate neuronal regeneration. Long-term oral supplementation was reported to be effective in delivering tocotrienols to vital organs without dependence on tocopherol-transfer protein (TTP) (Khanna *et al.*, 2005). The delivery was more pronounced in the fetal brain when pregnant rats were fed with tocotrienols (Roy *et al.*, 2002). Previous animal studies on cognition have focused on role of tocotrienols on improvement and prevention of cognitive impairments caused by diabetes (Kuhad *et al.*, 2009), alcoholism (Tiwari *et al.*, 2009), oxidative stress (Tiwari *et al.*, 2009) and aging (Taridi *et al.*, 2011, 2014). The roles of tocotrienols in neuroprotection have been reported in a number of studies. However, their exact contribution to neuroprotection afforded by the dam to her developing foetus and subsequent generations remained to be investigated. Aptly so, there are limited numbers of scientific publications in the trans-generational influence of tocotrienols supplementation on cognition and behaviour development. Thus, this study attempted to investigate the potential trans-generational

effects of maternal TRF supplementation on cognitive function development in the F1 males at maturity.

1.2 Hypothesis

It was hypothesized that continuous maternal TRF supplementation during gestation, lactation and post-weaning will improve cognitive function development in the F1 rats.

1.3 Objectives

The objectives of the present study were:

- a) To investigate the prenatal and postnatal effects of TRF supplementation on the brain fatty acids profile and vitamin E content in plasma and tissues of the F1 male rats.
- b) To investigate the prenatal and postnatal effects of TRF supplementation on cognitive function development in F1 male rats.

REFERENCES

- Adachi, H., & Ishii, N. (2000). Effects of tocotrienols on life span and protein carbonylation in *Caenorhabditis elegans*. *The Journals of Gerontology Series A: Biological Sciences and Medical Sciences*, 55(6), B280-B285. .
- Adachi, K., Miki, M., Tamai, H., Tokuda, M., & Mino, M. (1990). Adipose tissues and vitamin E. *Journal of Nutritional Science and Vitaminology*, 36(4), 327-337.
- Aggarwal, B., & Nesaretnam, K. (2012). Vitamin E tocotrienols: life beyond tocopherols. *Genes & nutrition*, 7(1), 1-1.
- Ambrogini, P., Ciuffoli, S., Lattanzi, D., Minelli, A., Bucherelli, C., Baldi, E., & Cuppini, R. (2011). Maternal dietary loads of α -tocopherol differentially influence fear conditioning and spatial learning in adult offspring. *Physiology & Behavior*, 104(5), 809-815.
- Ang, S. Y., & Lee, K. (2010). Exploring developmental differences in visual short-term memory and working memory. *Developmental psychology*, 46(1), 279.
- Arnsten, A. F. (1993). Catecholamine mechanisms in age-related cognitive decline. *Neurobiology of Aging*, 14(6), 639-641.
- Azlin, M. N., Nafeera, M. I., & Khalid, B. A. K. (2005). Effects of tocotrienol on lipid peroxidation in experimental gastritis induced by restraint stress. *Pakistan J Nutr*, 4(2), 69-72.
- Azzi, A., & Stocker, A. (2000). Vitamin E: non-antioxidant roles. *Progress in Lipid Research*, 39(3), 231-255.
- Begum, N., & Terao, J. (2002). Protective effect of α -tocotrienol against free radical-induced impairment of erythrocyte deformability. *Bioscience, Biotechnology, and Biochemistry*, 66(2), 398-403.
- Berger-Sweeney, J., Arnold, A., Gabeau, D., & Mills, J. (1995). Sex differences in learning and memory in mice: effects of sequence of testing and cholinergic blockade. *Behavioral Neuroscience*, 109(5), 859.
- Berlin, E., Bhathena, S. J., Judd, J. T., Nair, P. P., Peters, R. C., Bhagavan, H. N., & Taylor, P. R. (1992). Effects of omega-3 fatty acid and vitamin E supplementation on erythrocyte membrane fluidity, tocopherols, insulin binding, and lipid composition in adult men. *The Journal of Nutritional Biochemistry*, 3(8), 392-400.
- Betti, M., Ambrogini, P., Minelli, A., Floridi, A., Lattanzi, D., Ciuffoli, S., & Cuppini, R. (2011). Maternal dietary loads of α -tocopherol depress protein kinase C signaling and synaptic plasticity in rat postnatal developing hippocampus and promote permanent deficits in adult offspring. *The Journal of Nutritional Biochemistry*, 22(1), 60-70.

- Salucci, S., Ambrogini, P., Lattanzi, D., Betti, M., Gobbi, P., Galati, C., & Minelli, A. (2014). Maternal dietary loads of alpha-tocopherol increase synapse density and glial synaptic coverage in the hippocampus of adult offspring. *European Journal of Histochemistry*, 58(2).
- Bhathena, S. J., & Chow, C. K. (2000). Dietary fatty acids and fatty acid metabolism in diabetes. *Fatty Acids in Foods and Their Health Implications*. (Ed. 2), 915-961.
- Birringer, M., Pfluger, P., Kluth, D., Landes, N., & Brigelius-Flohé, R. (2002). Identities and differences in the metabolism of tocotrienols and tocopherols in HepG2 cells. *The Journal of Nutrition*, 132(10), 3113-3118.
- Bliss, T., & Collingridge, G. A. (1993). Synaptic model of memory: long-term potentiation in the hippocampus. *Nature*, 361, 31-39.
- Bourre, J. M. (2006). Effects of nutrients (in food) on the structure and function of the nervous system: update on dietary requirements for brain. Part 1: micronutrients. *Journal of Nutrition Health and Aging*, 10(5), 377.
- Bourre, J. M. (2006). Effects of nutrients (in food) on the structure and function of the nervous system: update on dietary requirements for brain. Part 2: macronutrients. *Journal of Nutrition Health and Aging*, 10(5), 386.
- Brown, J. E., & Wahle, K. W. (1990). Effect of fish-oil and vitamin E supplementation on lipid peroxidation and whole-blood aggregation in man. *Clinica Chimica Acta*, 193(3), 147-156.
- Budin, S. B., Taib, I. S., Jayusman, P. A., Chiang, H., Ramalingam, A., Ghazali, A. R., & Mohamed, J. (2014). Ameliorative Effect of Palm Oil Tocotrienol Rich Fraction on Brain Oxidative Stress in Fenitrothion-administered Rats. *Sains Malaysiana*, 43(7), 1031-1036.
- Chautan, M., Calaf, R., Leonardi, J., Charbonnier, M., Andre, M., Portugal, H., & Nalbone, G. (1990). Inverse modifications of heart and liver alpha-tocopherol status by various dietary n-6/n-3 polyunsaturated fatty acid ratios. *Journal of Lipid Research*, 31(12), 2201-2208.
- Chong, Z. Z., Li, F., & Maiese, K. (2005). Oxidative stress in the brain: novel cellular targets that govern survival during neurodegenerative disease. *Progress in Neurobiology*, 75(3), 207-246.
- Chopra, K., & Tiwari, V. (2013). Tocotrienol and Cognitive Dysfunction Induced by Alcohol. In *Alcohol, Nutrition, and Health Consequences* (pp. 181-202). Humana Press.
- Cockburn, F. (1994). Neonatal brain and dietary lipids. *Arch. Dis. Child. Fetal Neonatal Ed.*, 70, 1-2.
- Crawford, M. A., Williams, G., Hassam, A. G., & Whitehouse, W. L. (1976). Essential fatty acids and fetal brain growth. *The Lancet*, 307(7957), 452-453.

- Dalla, C., Edgecomb, C., Whetstone, A. S., & Shors, T. J. (2007). Females do not express learned helplessness like males do. *Neuropsychopharmacology*, 33(7), 1559-1569.
- Das, S., Lekli, I., Das, M., Szabo, G., Varadi, J., Juhasz, B., & Das, D. K. (2008). Cardioprotection with palm oil tocotrienols: comparison of different isomers. *American Journal of Physiology-Heart and Circulatory Physiology*, 294(2), H970-H978.
- Dauncey, M. J., & Bicknell, R. J. (1999). Nutrition and neurodevelopment: mechanisms of developmental dysfunction and disease in later life. *Nutrition Research Reviews*, 12(02), 231-253.
- Dauncey, M. J. (2009). New insights into nutrition and cognitive neuroscience. *Proceedings of the Nutrition Society*, 68(04), 408-415.
- Davis, H. P., Indowu, A., & Gibbson, G. E. (1983). Improvement of 8-arm maze performance in aged Fisher 344 rats with 3, 4,-diaminopyridine. *Experimental Aging Research*, 9, 211-214.
- Davis, S., Butcher, S. P., & Morris, R. G. (1992). The NMDA receptor antagonist D-2-amino-5-phosphonopentanoate (D-AP5) impairs spatial learning and LTP in vivo at intracerebral concentrations comparable to those that block LTP in vitro. *The Journal of Neuroscience*, 12(1), 21-34.
- Decker, M. W. (1987). The effects of aging on hippocampal and cortical projections of the forebrain cholinergic system. *Brain Research Reviews*, 12(4), 423-438.
- Dröge, W. (2003). Oxidative stress and aging. In *Hypoxia* (pp. 191-200). Springer US.
- Dullemeijer, C., Durga, J., Brouwer, I. A., van de Rest, O., Kok, F. J., Brummer, R. J. M., & Verhoef, P. (2007). n-3 Fatty acid proportions in plasma and cognitive performance in older adults. *The American Journal of Clinical Nutrition*, 86(5), 1479-1485.
- Mazlan, M., & Sue Mian, T., Mat Top G, Zurinah Wan Ngah, W. (2006) Comparative effects of alpha-tocopherol and gamma-tocotrienol against hydrogen peroxide induced apoptosis on primary-cultured astrocytes. *J Neurol Sci*, 243, 5-12.
- Eichenbaum, H., & Cohen, N. J. (2001). *From conditioning to conscious recollection: Memory Systems of The Brain*. Oxford University Press.
- Ebrahimi, M., Rajion, M. A., Goh, Y. M., & Sazili, A. Q. (2012). Impact of different inclusion levels of oil palm (*Elaeis guineensis* Jacq.) fronds on fatty acid profiles of goat muscles. *Journal of Animal Physiology and Animal Nutrition*, 96(6), 962-969.
- Erhard, H. W., Boissy, A., Rae, M. T., & Rhind, S. M. (2004). Effects of prenatal under nutrition on emotional reactivity and cognitive flexibility in adult sheep. *Behavioural Brain Research*, 151(1), 25-35.

- Eriksson, P., Perfilieva, E., Bjork-Eriksson, T., Alborn, A., Nordborg, C., Peterson, D., & Gage, F. (1998). Neurogenesis in the adult human hippocampus. *Nature Medicine*, 4, 1313-1317.
- Farquharson, J., Jamieson, E. C., Abbasi, K. A., Patrick, W. J., Logan, R. W., & Cockburn, F. (1995). Effect of diet on the fatty acid composition of the major phospholipids of infant cerebral cortex. *Archives of Disease in Childhood*, 72(3), 198-203.
- Floyd, R. A., & Carney, J. M. (1992). Free radical damage to protein and DNA: mechanisms involved and relevant observations on brain undergoing oxidative stress. *Annals of Neurology*, 32(S1), S22-S27.
- Frank, J., Chin, X. W. D., Schrader, C., Eckert, G. P., & Rimbach, G. (2012). Do tocotrienols have potential as neuroprotective dietary factors. *Ageing Research Reviews*, 11(1), 163-180.
- Gallagher, M., & Rapp, P. (1997). The use of animal models to study the effects of aging on cognition. *Annu Rev Psychol*, 48, 339-370.
- Hemalatha, S., & Rao, M. V. V. (2004). Sesame lignans enhance antioxidant activity of vitamin E in lipid peroxidation systems. *Molecular and Cellular Biochemistry*, 262(1-2), 195-202.
- Goh, S. H., Hew, N. F., Norhanom, A. W., & Yadav, M. (1994). Inhibition of tumour promotion by various palm-oil tocotrienols. *International Journal of Cancer*, 57(4), 529-531.
- Gómez-Pinilla, F. (2008). Brain foods: the effects of nutrients on brain function. *Nature Reviews Neuroscience*, 9(7), 568-578.
- Gould, E., Beylin, A., Tanapat, P., Reeves, A., & Shors, T. J. (1999). Learning enhances adult neurogenesis in the hippocampal formation. *Nature Neuroscience*, 2(3), 260-265.
- Grachev, I. D., & Apkarian, A. V. (2000). Chemical heterogeneity of the living human brain: A proton MR spectroscopy study on the effects of sex, age, and brain region. *Neuro Image*, 11, 554-563.
- Hajjar, T., Meng, G. Y., Rajion, M. A., Vidyadaran, S., Othman, F., Farjam, A. S., & Ebrahimi, M. (2012). Omega 3 polyunsaturated fatty acid improves spatial learning and hippocampal Peroxisome Proliferator Activated Receptors (PPAR α and PPAR γ) gene expression in rats. *BMC Neuroscience*, 13(1), 109.
- Halliwel, B. (2006). Oxidative stress and neurodegeneration: where are we now? *Journal of Neurochemistry*, 97(6), 1634-1658.
- Hebb, 1949 D.O. Hebb the Organization of Behavior John Wiley & Sons, Inc, New York (1949)

- Hiura, Y., Tachibana, H., Arakawa, R., Aoyama, N., Okabe, M., & Sakai, M. (2009). Specific accumulation of gamma- and delta-tocotrienols in tumor and their antitumor effect in vivo. *Journal of Nutritional Biochemistry*, 20:607–613.
- Hodges, H. (1995). Maze procedures: the radial-arm and water maze compared. *Cognitive Brain Research*, 3(3), 167-181.
- Hornstra, G. (2000). Essential fatty acids in mothers and their neonates. *The American Journal of Clinical Nutrition*, 71(5), 1262s-1269s.
- Hosomi, A., Arita, M., Sato, Y., Kiyose, C., Ueda, T., Igarashi, O., & Inoue, K. (1997). Affinity for [alpha]-tocopherol transfer protein as a determinant of the biological activities of vitamin E analogs. *Febs Letters*, 409(1), 105-108.
- Igarashi, M., DeMar, J. C., Ma, K., Chang, L., Bell, J. M., & Rapoport, S. I. (2007). Docosahexaenoic acid synthesis from α -linolenic acid by rat brain is unaffected by dietary n-3 PUFA deprivation. *Journal of Lipid Research*, 48(5), 1150-1158.
- Innis, S. M. (2007). Dietary (n-3) fatty acids and brain development. *The Journal of Nutrition*, 137(4), 855-859.
- Innis, S. M. (2008). Dietary omega 3 fatty acids and the developing brain. *Brain Research*, 1237, 35-43.
- Jensen, M., Skarsfeldt, T., & Hey, C.-E. (1996). Correlation between level of (n - 3) polyunsaturated fatty acids in brain phospholipids and learning ability in rats. A multiple generation study. *Biochimica Et Biophysica Acta*, 1300, 203-209.
- Jiang, Q., Christen, S., Shigenaga, M. K., & Ames, B. N. (2001). γ -Tocopherol, the major form of vitamin E in the US diet, deserves more attention. *The American Journal of Clinical Nutrition*, 74(6), 714-722.
- Johnson, E. A. (2010). A study of the effects of immersion on short-term spatial memory.
- Johnson, S. A., Lampert-Etchells, M., Pasinetti, G. M., Rozovsky, I., & Finch, C. E. (1992). Complement mRNA in the mammalian brain: responses to Alzheimer's disease and experimental brain lesioning. *Neurobiology of Aging*, 13(6), 641-648.
- Jordan, K., Wustenberg, T., Heinze, H.-J., Peters, M., & Jancke, L. (2002). Women and men exhibit different cortical activation patterns during mental rotation tasks. *Neuropsychologia*, 40, 2397–2408.
- Kaempf-Rotzoll, D. E., Traber, M. G., & Arai, H. (2003). Vitamin E and transfer proteins. *Current Opinion in Lipidology*, 14(3), 249-254.
- Kamal-Eldin, A., & Appelqvist, L. Å. (1996). The chemistry and antioxidant properties of tocopherols and tocotrienols. *Lipids*, 31(7), 671-701.

- Kamat, J. P., & Devasagayam, T. P. A. (1995). Tocotrienols from palm oil as potent inhibitors of lipid peroxidation and protein oxidation in rat brain mitochondria. *Neuroscience Letters*, 195(3), 179-182.
- Kamat, J. P., Sarma, H. D., Devasagayam, T. P. A., Nesaretnam, K., & Basiron, Y. (1997). Tocotrienols from palm oil as effective inhibitors of protein oxidation and lipid peroxidation in rat liver microsomes. *Molecular and Cellular Biochemistry*, 170(1-2), 131-138.
- Kamat, J. P., & Devasagayam, T. P. A. (1995). Tocotrienols from palm oil as potent inhibitors of lipid peroxidation and protein oxidation in rat brain mitochondria. *Neuroscience Letters*, 195(3), 179-182.
- Kanaya, Y., Doi, T., Sasaki, H., Fujita, A., Matsuno, S., Okamoto, K. & Nanjo, K. (2004). Rice bran extract prevents the elevation of plasma peroxy lipid in KKAY diabetic mice. *Diabetes Research and Clinical Practice*, 66, S157-S160.
- Kawakami, Y., Tsuzuki, T., Nakagawa, K., & Miyazawa, T. (2007). Distribution of tocotrienols in rats fed a rice bran tocotrienol concentrate. *Bioscience, Biotechnology, and Biochemistry*, 71(2), 464-471.
- Khanna, S., Patel, V., Rink, C., Roy, S., & Sen, C. K. (2005). Delivery of orally supplemented α -tocotrienol to vital organs of rats and tocopherol-transport protein deficient mice. *Free Radical Biology and Medicine*, 39(10), 1310-1319.
- Khanna S, Roy S, Slivka A, Craft K S T, Chaki S, Rink C, Notestine A M, Devries C, Parinandi N L, Chandan K Sen (2005). Neuroprotective Properties of the Natural Vitamin E α -Tocotrienol. *Stroke*, 36: e144-e152.
- Khanna, S., Roy, S., Parinandi, N. L., Maurer, M., & Sen, C. K. (2006). Characterization of the potent neuroprotective properties of the natural vitamin E α -tocotrienol. *Journal of Neurochemistry*, 98(5), 1474-1486.
- Khanna, S., Roy, S., Ryu, H., Bahadduri, P., Swaan, P. W., Ratan, R. R., & Sen, C. K. (2003). Molecular Basis of Vitamin E Action tocotrienol modulates 12-lipoxygenase, a key mediator of glutamate-induced neurodegeneration. *Journal of Biological Chemistry*, 278(44), 43508-43515
- Komiyama, K., Iizuka, K., Yamaoka, M., Watanabe, H., Tsuchiya, N., & Umezawa, I. (1989). Studies on the biological activity of tocotrienols. *Chemical & Pharmaceutical Bulletin*, 37(5), 1369-1371.
- Kuhad A, Bishnoi M, Tiwari V, Bishnoi M, Chopra K (2009). Suppression of NF- κ B signalling pathway by tocotrienol can prevent diabetes associated cognitive deficits. *Pharmacology Biochemistry Behaviour*, 92(2):251-259
- Kumar, K. S., Raghavan, M., Hieber, K., Ege, C., Mog, S., Parra, N., & Papas, A. (2006). Preferential radiation sensitization of prostate cancer in nude mice by nutraceutical antioxidant γ -tocotrienol. *Life Sciences*, 78(18), 2099-2104.

- Lacreuse, A., Herndon, J. G., Killiany, R., Rosene, D. L., & Moss, M. B. (1999). Spatial cognition in rhesus monkeys: Male superiority declines with age. *Hormones and Behavior*, *36*, 70-76.
- Lamberty, Y., & Gower, A. J. (1988). Investigation into sex-related differences in locomotor activity, place learning and passive avoidance responding in NMRI mice. *Physiology & Behavior*, *44*(6), 787-790. 790.
- Landon, J., Davison, M., Krägeloh, C. U., Thompson, N. M., Miles, J. L., Vickers, M. H. & Breier, B. H. (2007). Global undernutrition during gestation influences learning during adult life. *Learning & Behaviour*, *35*(2), 79-86.
- Lehtinen, M. K., & Bonni, A. (2006). Modeling oxidative stress in the central nervous system. *Current molecular Medicine*, *6*(8), 871-881.
- Leibovitz, B. E., Hu, M. L., & Tappel, A. L. (1990). Lipid peroxidation in rat tissue slices: effect of dietary vitamin E, corn oil-lard and menhaden oil. *Lipids*, *25*(3), 125-129.
- Lodge, J. K., Ridlington, J., Leonard, S., Vaule, H., & Traber, M. G. (2001). α - and γ -Tocotrienols are metabolized to carboxyethyl-hydroxychroman derivatives and excreted in human urine. *Lipids*, *36*(1), 43-48.
- Loganathan, R., Selvaduray, K. R., Nesaretnam, K., & Radhakrishnan, A. K. (2013). Tocotrienols promote apoptosis in human breast cancer cells by inducing poly (ADP-ribose) polymerase cleavage and inhibiting nuclear factor kappa-B activity. *Cell Proliferation*, *46*(2), 203-213.
- Loo, G., Berlin, E., Peters, R. C., Kliman, P. G., & Wong, H. Y. (1991). Effect of dietary corn, coconut, and menhaden oils on lipoprotein, liver, and heart membrane composition in the hypercholesterolemic rabbit. *The Journal of Nutritional Biochemistry*, *2*(11), 594-603.
- Lucas, A. (1998). Programming by early nutrition: an experimental approach. *The Journal of Nutrition*, *128*(2), 401S-406S.
- Lucas A (2001). Role of nutritional programming in determining adult morbidity. *Arch Dis Child*, *71*(4):288–290. Lucas, A., Morley, R., & Isaacs, E. (2001). Nutrition and mental development. *Nutrition Reviews*, *59*(8), S24-S33.
- Lucas, A., Morley, R., & Isaacs, E. (2001). Nutrition and mental development. *Nutrition Reviews*, *59*(8), S24-S33.
- Luine, V., Bowling, D., & Hearn, M. (1990). Spatial memory deficits in aged rats: contributions of monoaminergic systems. *Brain Research*, *537*(1), 271-278.
- Makrides, M., Gibson, R. A., McPhee, A. J., Yelland, L., Quinlivan, J., & Ryan, P. (2010). Effect of DHA supplementation during pregnancy on maternal depression and neurodevelopment of young children: a randomized controlled trial. *Jama*, *304*(15), 1675-1683.

- Maren, S., & Baudry, M. (1995). Properties and mechanisms of long-term synaptic plasticity in the mammalian brain: relationships to learning and memory. *Neurobiology of Learning and Memory*, 63(1), 1-18.
- Martin, J. H. (2003). Lymbic system and cerebral circuits for emotions, learning, and memory. *Neuroanatomy: text and atlas (third Ed.)*. McGraw-Hill Companies. (p. p. 382).
- Martinez, M. (1992). Tissue levels of polyunsaturated fatty acids in early human development. *J. Pediatr*, 120, 129–138.
- Martínez, M., & Mougán, I. (1998). Fatty acid composition of human brain phospholipids during normal development. *Journal of Neurochemistry*, 71(6), 2528-2533.
- Martínez-Serrano, A., & Björklund, A. (1998). Ex vivo nerve growth factor gene transfer to the basal forebrain in presymptomatic middle-aged rats prevents the development of cholinergic neuron atrophy and cognitive impairment during aging. *Proceedings of the National Academy of Sciences*, 95(4), 1858-1863.
- Mazlan, M., Then, S. M., Mat Top, G., & Zurinah Wan Ngah, W. (2006). Comparative effects of α -tocopherol and γ -tocotrienol against hydrogen peroxide induced apoptosis on primary-cultured astrocytes. *Journal of The Neurological Sciences*, 243(1), 5-12.
- Mccann, J. C., & Ames, B. N. (2005). Is docosahexaenoic acid, an n \times 3 long-chain polyunsaturated fatty acid, required for development of normal brain function? An overview of evidence from cognitive and behavioral tests in humans and animals. *American Journal of Clinical Nutrition*, 82(1), 281-295.
- McQuade, J. M. S. 2002. The Involvement of DFF45 and c-fos in Hippocampal Plasticity and Function. PhD Thesis, University of Cincinnati, USA.
- Meydani, M., Evans, W. J., Handelman, G., Biddle, L., Fielding, R. A., Meydani, S. N., & Cannon, J. G. (1993). Protective effect of vitamin E on exercise-induced oxidative damage in young and older adults. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, 264(5), R992-R998.
- Molteni, R., Barnard, R. J., Ying, Z., Roberts, C. K., & Gomez-Pinilla, F. (2002). A high-fat, refined sugar diet reduces hippocampal brain-derived neurotrophic factor, neuronal plasticity, and learning. *Neuroscience*, 112(4), 803-814.
- Morris, R. G. (1981). Spatial Localization Does Not Require the Presence of Local Cues. *Learning and Motivation*, 12(2), 239-260.
- Morris, R. G., Garrud, P., Rawlins, J. N. P., & O'Keefe, J. (1982). Place navigation impaired in rats with hippocampal lesions. *Nature*, 297, 681-683.
- Moser, E. I., Kropff, E., & Moser, M. B. (2008). Place cells, grid cells, and the brain's spatial representation system. *Neuroscience*, 31(1), 69.

- Nakagawa, K., Shibata, A., Yamashita, S., Tsuzuki, T., Kariya, J., Oikawa, S., & Miyazawa, T. (2007). In vivo angiogenesis is suppressed by unsaturated vitamin E, tocotrienol. *The Journal of Nutrition*, 137(8), 1938-1943.
- Nesaretnam K, Ambra R, Selvaduray KR, Radhakrishnan A, Reimann K, Razak G, Virgili F (2004). Tocotrienol-Rich Fraction from palm oil affects gene expression in tumors resulting from MCF-7 cell inoculation in athymic mice. *Lipids*, 39(5): 459-467.
- Nesaretnam, K., Guthrie, N., Chambers, A. F., & Carroll, K. K. (1995). Effect of tocotrienols on the growth of a human breast cancer cell line in culture. *Lipids*, 30(12), 1139-1143.
- Nesaretnam, K., Mahalingam, D., Radhakrishnan, A. K., & Premier, R. (2010). Supplementation of tocotrienol-rich fraction increases interferon-gamma production in ovalbumin-immunized mice. *European Journal of Lipid Science and Technology*, 112(5), 531-536.
- Nesaretnam, K., Stephen, R., Dils, R., & Darbre, P. (1998). Tocotrienols inhibit the growth of human breast cancer cells irrespective of estrogen receptor status. *Lipids*, 33(5), 461-469.
- Nesaretnam, K., Ambra, R., Selvaduray, K. R., Radhakrishnan, A., Canali, R., & Virgili, F. (2004). Tocotrienol-rich fraction from palm oil and gene expression in human breast cancer cells. *Annals of the New York Academy of Sciences*, 1031(1), 143-157.
- Nesaretnam, K., Devasagayam, T. P. A., Singh, B. B., & Basiron, Y. (1993). Influence of palm oil or its tocotrienol-rich fraction on the lipid peroxidation potential of rat liver mitochondria and microsomes. *Biochemistry and molecular biology international*, 30, 159-159.
- Nesaretnam, K., Koon, T. H., Selvaduray, K. R., Bruno, R. S., & Ho, E. (2008). Modulation of cell growth and apoptosis response in human prostate cancer cells supplemented with tocotrienols. *European Journal of Lipid Science and Technology*, 110(1), 23-31.
- Niculescu, M. D., & Lupu, D. S. (2009). High fat diet-induced maternal obesity alters fetal hippocampal development. *International Journal of Developmental Neuroscience*, 27(7), 627-633.
- Noguchi, N., Hanyu, R., Nonaka, A., Okimoto, Y., & Kodama, T. (2003). Inhibition of THP-1 cell adhesion to endothelial cells by α -tocopherol and α -tocotrienol is dependent on intracellular concentration of the antioxidants. *Free Radical Biology and Medicine*, 34(12), 1614-1620.
- Nurk, E., Drevon, C. A., Refsum, H., Solvoll, K., Vollset, S. E., Nygård, O., & Smith, A. D. (2007). Cognitive performance among the elderly and dietary fish intake: the Hordaland Health Study. *The American Journal of Clinical Nutrition*, 86(5), 1470-1478.
- Olton, D. S. (1976). Remembrance on placed passed: spatial memory in rats. *Journal Experimental Physiology Behavior*, 2, 97-116.

- Ong, A. S. H., Natural sources of tocotrienol, in: Packer, L., Fuchs, J. (Eds.), *From Natural Sources of Tocotrienols in Vitamin E in Health and Disease*. Marcel Dekker, New York 1993, pp. 3–8.
- Osakada, F., Hashino, A., Kume, T., Katsuki, H., Kaneko, S., & Akaike, A. (2004). α -Tocotrienol provides the most potent neuroprotection among vitamin E analogs on cultured striatal neurons. *Neuropharmacology*, 47(6), 904-915.
- Packer, L., Weber, S. U., & Rimbach, G. (2001). Molecular aspects of α -tocotrienol antioxidant action and cell signalling. *The Journal of Nutrition*, 131(2), 369S-373S.
- Park, H. A., Kubicki, N., Gnyawali, S., Chan, Y. C., Roy, S., Khanna, S., Sen, C. K. (2011). Natural vitamin E α -tocotrienol protects against ischemic stroke by induction of multidrug resistance-associated protein 1. *Stroke*, 42(8):2308-14.
- Parker, R. A., Pearce, B. C., Clark, R. W., Gordon, D. A., & Wright, J. J. (1993). Tocotrienols regulate cholesterol production in mammalian cells by post-transcriptional suppression of 3-hydroxy-3-methylglutaryl-coenzyme A reductase. *Journal of Biological Chemistry*, 268(15), 11230-11238.
- Patel, V., Khanna, S., Roy, S., Ezziddin, O., & Sen, C. K. (2006). Natural vitamin E α -tocotrienol: Retention in vital organs in response to long-term oral supplementation and withdrawal. *Free Radical Research*, 40(7), 763-771.
- Perrot-Sinal, T. S., Kostenuik, M. A., Ossenkopp, K. P., & Kavaliers, M. (1996). Sex differences in performance in the Morris water maze and the effects of initial nonstationary hidden platform training. *Behavioral Neuroscience*, 110(6), 1309.
- Pomeroy, S., & Kim, JY. (2000). Biology and pathology of neuronal development. *Ment Ret and Dev Disabil Res Rev*, 6, 41-46.
- Qureshi, A. A., Qureshi, N., Wright, J. J., Shen, Z., Kramer, G., Gapor, A., ... & Peterson, D. M. (1991). Lowering of serum cholesterol in hypercholesterolemic humans by tocotrienols (palmvitee). *The American Journal of Clinical Nutrition*, 53(4), 1021S-1026S.
- Qureshi, A. A., Burger, W. C., Peterson, D. M., & Elson, C. E. (1986). The structure of an inhibitor of cholesterol biosynthesis isolated from barley. *Journal of Biological Chemistry*, 261(23), 10544-10550.
- Qureshi, A. A., Mo, H., Packer, L., & Peterson, D. M. (2000). Isolation and identification of novel tocotrienols from rice bran with hypocholesterolemic, antioxidant, and antitumor properties. *Journal of Agricultural and Food Chemistry*, 48(8), 3130-3140.
- Rajion, M.A. (1985). *Essential fatty acid metabolism in the fetal and newborn lamb*. Ph.D. Thesis, University of Melbourne, Australia.
- Ramakrishna, T. (1999). Vitamins and brain development. *Physiological Research*, 48, 175-188.

- Rassin DK, Smith KE (2003). Nutritional approaches to improve cognitive development during infancy: antioxidant compounds. *Acta Paediatrica*, 92: 34-41.
- Rimbach, G., Minihane, A. M., Majewicz, J., Fischer, A., Pallauf, J., Virgli, F., & Weinberg, P. D. (2002). Regulation of cell signalling by vitamin E. *Proceedings of The Nutrition Society*, 61(04), 415-425.
- Roseboom, T. J., & Watson, E. D. (2012). The next generation of disease risk: are the effects of prenatal nutrition transmitted across generations? Evidence from animal and human studies. *Placenta*, 33, e40-e44.
- Roy, S., Lado, B. H., Khanna, S., Sen, C. K. (2002). Vitamin E sensitive genes in the developing rat fetal brain: a high-density oligonucleotide microarray analysis. *FEBS Letters*, 530(1-3): 17-23.
- Schultz, M., Leist, M., Petrzika, M., Gassmann, B., & Brigelius-Flohe, R. (1995). Novel urinary metabolite of alpha-tocopherol, 2, 5, 7, 8-tetramethyl-2 (2'-carboxyethyl)-6-hydroxychroman, as an indicator of an adequate vitamin E supply. *The American Journal of Clinical Nutrition*, 62(6), 1527S-1534S
- Sen, C. K., Khanna, S., & Roy, S. (2004). Tocotrienol: the natural vitamin E to defend the nervous system. *Annals of the New York Academy of Sciences*, 1031(1), 127-142.
- Sen, C. K., Khanna, S., & Roy, S. (2006). Tocotrienols: vitamin E beyond tocopherols. *Life Sciences*, 78(18), 2088-2098.
- Sen, C. K., Khanna, S., Roy, S., & Packer, L. (2000). Molecular Basis of Vitamin E Action Tocotrienol potently inhibits glutamate-induced pp60c-Src Kinase activation and death of ht4 neuronal cells. *Journal of Biological Chemistry*, 275(17), 13049-13055.
- Sen, C. K., Rink, C., & Khanna, S. (2010). Palm oil-derived natural vitamin E α -tocotrienol in brain health and disease. *Journal of the American College of Nutrition*, 29(sup3), 314S-323S.
- Serbinova, E., Kagan, V., Han, D., & Packer, L. (1991). Free radical recycling and intramembrane mobility in the antioxidant properties of alpha-tocopherol and alpha-tocotrienol. *Free Radical Biology and Medicine*, 10(5), 263-275.
- Shah, S., Gapor, A., & Sylvester, P. W. (2003). Role of caspase-8 activation in mediating vitamin E-induced apoptosis in murine mammary cancer cells. *Nutrition and Cancer*, 45(2), 236-246.
- Shen, J., Barnes, C. A., McNaughton, B. L., Skaggs, W. E., & Weaver, K. L. (1997). The effect of aging on experience-dependent plasticity of hippocampal place cells. *The Journal of Neuroscience*, 17(17), 6769-6782.
- Sherry, D. F., Jacobs, L. F., & Gaulin, S. J. C. (1992). Spatial memory and adaptive specialization of the hippocampus. *Trends in Neuroscience*, 15, 298-303.
- Shettleworth, S. J. (2001). Animal cognition and animal behaviour. *Animal Behaviour*, 61(2), 277-286.

- Song, B. L., & DeBose-Boyd, R. A. (2006). Insight-dependent ubiquitination and degradation of 3-hydroxy-3-methylglutaryl coenzyme A reductase stimulated by δ - and γ -tocotrienols. *Journal of Biological Chemistry*, 281(35), 25054-25061
- Spencer, J. P. (2008). Food for thought: the role of dietary flavonoids in enhancing human memory, learning and neuro-cognitive performance. *Proceedings of the Nutrition Society*, 67(02), 238-252.
- Squire, L. R. (1992). Memory and the hippocampus: a synthesis from findings with rats, monkeys, and humans. *Psychological Review*, 99(2), 195.
- Squire, L. R. (2004). Memory systems of the brain: a brief history and current perspective. *Neurobiology of Learning and Memory*, 82(3), 171-177.
- Squire, L. R., & Alvarez, P. (1995). Retrograde amnesia and memory consolidation: a neurobiological perspective. *Current Opinion in Neurobiology*, 5(2), 169-177.
- Squire, L. R., & Zola-Morgan, S. (1991). The medial temporal lobe memory system. *Science*, 253(5026), 1380-1386.
- Stone, W. L., Papas, A. M., LeClair, I. O., Qui, M., & Ponder, T. (2002). The influence of dietary iron and tocopherols on oxidative stress and ras-p21 levels in the colon. *Cancer Detection and Prevention*, 26(1), 78-84.
- Su, H. M. (2010). Mechanisms of n-3 fatty acid-mediated development and maintenance of learning memory performance. *The Journal of Nutritional Biochemistry*, 21(5), 364-373.
- Suarna, C., Hood, R. L., Dean, R. T., & Stocker, R. (1993). Comparative antioxidant activity of tocotrienols and other natural lipid-soluble antioxidants in a homogeneous system, and in rat and human lipoproteins. *Biochimica et Biophysica Acta (BBA)-Lipids and Lipid Metabolism*, 1166(2), 163-170.
- Sundram, K., Ismail, A., Hayes, K. C., Jeyamalar, R., & Pathmanathan, R. (1997). Trans (elaidic) fatty acids adversely affect the lipoprotein profile relative to specific saturated fatty acids in humans. *The Journal of Nutrition*, 127(3), 514S-520S.
- Sundram, K., Khor, H. T., Ong, A. S., & Pathmanathan, R. (1989). Effect of dietary palm oils on mammary carcinogenesis in female rats induced by 7, 12-dimethylbenz (a) anthracene. *Cancer Research*, 49(6), 1447-1451.
- Sutcliffe, J. S., Marshall, K. M., & Neill, J. C. (2007). Influence of gender on working and spatial memory in the novel object recognition task in the rat. *Behavioural Brain Research*, 177(1), 117-125.
- Suzuki, Y. J., Tsuchiya, M., Wassall, S. R., Choo, Y. M., Govil, G., Kagan, V. E., & Packer, L. (1993). Structural and dynamic membrane properties of. Alpha.-tocopherol and. Alpha.-tocotrienol: implication to the molecular mechanism of their antioxidant potency. *Biochemistry*, 32(40), 10692-10699.

- Swanson, J. E., Ben, R. N., Burton, G. W., & Parker, R. S. (1999). Urinary excretion of 2, 7, 8-trimethyl-2-(β -carboxyethyl)-6-hydroxychroman is a major route of elimination of γ -tocopherol in humans. *Journal of Lipid Research*, 40(4), 665-671.
- Sylvester, P. W., Nachnani, A., Shah, S., & Briski, K. P. (2002). Role of GTP-binding proteins in reversing the antiproliferative effects of tocotrienols in preneoplastic mammary epithelial cells. *Asia Pacific Journal of Clinical Nutrition*, 11(s7), S452-S459.
- Taridi, N. M., Yahaya, M. F., Teoh, S. L., Latiff, A. A., Ngah, W. Z., Das, S., & Mazlan, M. (2010). Tocotrienol rich fraction (TRF) supplementation protects against oxidative DNA damage and improves cognitive functions in Wistar rats. *La Clinica Terapeutica*, 162(2), 93-98.
- Taridi, N. M., Rani, N. A., Latiff, A. A., Ngah, W. Z. W., & Mazlan, M. (2014). Tocotrienol Rich Fraction Reverses Age-Related Deficits in Spatial Learning and Memory in Aged Rats. *Lipids*, 49(9), 855-869.
- Tiwari, V., Kuhad, A., Bishnoi, M., & Chopra, K. (2009). Chronic treatment with tocotrienol, an isoform of vitamin E, prevents intracerebroventricular streptozotocin-induced cognitive impairment and oxidative–nitrosative stress in rats. *Pharmacology Biochemistry and Behaviour*, 93(2), 183-189.
- Tiwari, V., Kuhad, A., & Chopra, K. (2009). Suppression of neuro-inflammatory signaling cascade by tocotrienol can prevent chronic alcohol-induced cognitive dysfunction in rats. *Behavioural Brain Research*, 203(2), 296-303.
- Tonkiss, J., Shultz, P. L., Shumsky, J. S., & Galler, J. R. (1997). Development of spatial navigation following prenatal cocaine and malnutrition in rats: lack of additive effects. *Neurotoxicology and Teratology*, 19(5), 363-372.
- Tozuka, Y., Kumon, M., Wada, E., Onodera, M., Mochizuki, H., & Wada, K. (2010). Maternal obesity impairs hippocampal BDNF production and spatial learning performance in young mouse offspring. *Neurochemistry international*, 57(3), 235-247.
- Traber, M. G., & Kayden, H. J. (1989). Preferential incorporation of alpha-tocopherol vs gamma-tocopherol in human lipoproteins. *The American Journal of Clinical Nutrition*, 49(3), 517-526.
- Traber, M. G., Burton, G. W., & Hamilton, R. L. (2004). Vitamin E trafficking. *Annals of the New York Academy of Sciences*, 1031(1), 1-12.
- Traber, M. G., Lane, J. C., Lagmay, N. R., & Kayden, H. J. (1992). Studies on the transfer of tocopherol between lipoproteins. *Lipids*, 27(9), 657-663.
- Traber, M. G., Olivecrona, T., & Kayden, H. J. (1985). Bovine milk lipoprotein lipase transfers tocopherol to human fibroblasts during triglyceride hydrolysis in vitro. *Journal of Clinical Investigation*, 75(5), 1729.

- Traber, Maret G (2004). Vitamin E nuclear receptors and xenobiotic metabolism. *Arch Biochem Biophys*, 423(1):6-11.
- Tuzcu, M., & Baydas, G. (2006). Effect of melatonin and vitamin E on diabetes-induced learning and memory impairment in rats. *European Journal of Pharmacology*, 537(1), 106-110.
- Uauy, R., & Dangour, A. D. (2006). Nutrition in brain development and aging: role of essential fatty acids. *Nutrition Reviews*, 64(s2), S24-S33.
- Uauy, R., Mena, P., & Rojas, C. (2000). Essential fatty acids in early life: structural and functional role. *Proceedings of the Nutrition Society*, 59(01), 3-15.
- Venga, L. M., Granholmb, A. C., & Rose, G. M. (2003). Age-related sex differences in spatial learning and basal forebrain cholinergic neurons in F344 rats. *Physiology & Behavior*, 80, 27– 36.
- Vorhees, C. V., & Williams, M. T. (2006). Morris water maze: procedures for assessing spatial and related forms of learning and memory. *Nature Protocols*, 1(2), 848-858.
- Wainwright, P. E. (2002). Dietary essential fatty acids and brain function : a developmental perspective on mechanisms. *Journal of Nutrition*, 61, 61-69. Doi: 10.1079/PNS2001130
- Wainwright, P. E., & Colombo, J. (2006). Nutrition and the development of cognitive functions: interpretation of behavioral studies in animals and human infants. *The American Journal of Clinical Nutrition*, 84(5), 961-970.
- Wainwright, P. E., Huang, Y. S., Bulman-Fleming, B., Mills, D. E., Redden, P., & McCutcheon, D. (1991). The role of n- 3 essential fatty acids in brain and behavioural development: a cross-fostering study in the mouse. *Lipids*, 26(1), 37-45.
- Wang, H., Hu, Y., & Tsien, J. Z. (2006). Molecular and systems mechanisms of memory consolidation and storage. *Progress in Neurobiology*, 79(3), 123-135.
- Watanabe, S., Kanada, S., Takenaka, M., & Hamazaki, T. (2004). Dietary n-3 fatty acids selectively attenuate LPS-induced behavioral depression in mice. *Physiology & Behavior*, 81(4), 605-613.
- Wauben, I. P., & Wainwright, P. E. (1999). The influence of neonatal nutrition on behavioral development: a critical appraisal. *Nutrition Reviews*, 57(2), 35-44.
- White, C. L., Pistell, P. J., Purpera, M. N., Gupta, S., Fernandez-Kim, S. O., Hise, T. L & Bruce-Keller, A. J. (2009). Effects of high fat diet on Morris maze performance, oxidative stress, and inflammation in rats: contributions of maternal diet. *Neurobiology of Disease*, 35(1), 3-13.

- Williams, C. L., Barnett, A. M., & Meck, W. H. (1990). Organizational effects of early gonadal secretions on sexual differentiation in spatial memory. *Behavioral Neuroscience*, 104(1), 84.
- Yamashita, K., Ikeda, S., Iizuka, Y., & Ikeda, I. (2002). Effect of sesaminol on plasma and tissue α -tocopherol and α -tocotrienol concentrations in rats fed a vitamin E concentrate rich in tocotrienols. *Lipids*, 37(4), 351-358.
- Yoshida, Y., Niki, E., & Noguchi, N. (2003). Comparative study on the action of tocopherols and tocotrienols as antioxidant: chemical and physical effects. *Chemistry and Physics of Lipids*, 123(1), 63-75.
- Yoshida, Y., Saito, Y., Jones, L. S., & Shigeri, Y. (2007). Chemical reactivities and physical effects in comparison between tocopherols and tocotrienols: physiological significance and prospects as antioxidants. *Journal of Bioscience and Bioengineering*, 104(6), 439-445.
- Yurko-Mauro, K., McCarthy, D., Rom, D., Nelson, E. B., Ryan, A. S., Blackwell, A., ... & Stedman, M. (2010). Beneficial effects of docosahexaenoic acid on cognition in age-related cognitive decline. *Alzheimer's & Dementia*, 6(6), 456-464.
- Zarkovic, K. (2003). 4-hydroxynonenal and neurodegenerative diseases. *Molecular Aspects of Medicine*, 24(4), 293-303.
- Zhang, Y., Li, N., & Yang, Z. (2010). Perinatal food restriction impaired spatial learning and memory behavior and decreased the density of nitric oxide synthase neurons in the hippocampus of adult male rat offspring. *Toxicology Letters*, 193(2), 167-172.