

# **UNIVERSITI PUTRA MALAYSIA**

ISOLATION, CHARACTERIZATION AND PATHOGENICITY OF EPIZOOTIC ULCERATIVE SYNDROME-RELATED Aphanomyces TOWARD AN IMPROVED DIAGNOSTIC TECHNIQUE

SEYEDEH FATEMEH AFZALI

FPV 2014 7



## ISOLATION, CHARACTERIZATION AND PATHOGENICITY OF EPIZOOTIC ULCERATIVE SYNDROME-RELATED Aphanomyces TOWARD AN IMPROVED DIAGNOSTIC TECHNIQUE



By

SEYEDEH FATEMEH AFZALI

Thesis Submitted to the School of Graduate Study, Universiti Putra Malaysia, in Fulfillment of the Requirement for the Degree of Doctor of Philosophy

August 2014

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



### **DEDICATION**

This dissertation is lovingly dedicated to my kind family.

A special feeling of gratitude to my great parents who inspired my life through their gritty strength, enduring faith, and boundless love for family. My nice sisters and brother have never left my side and have supported me throughout the process. I also dedicate this work and give special thanks to my best friend "Hasti" for being there for me throughout the entire doctorate program.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

## ISOLATION, CHARACTERIZATION AND PATHOGENICITY OF EPIZOOTIC ULCERATIVE SYNDROME-RELATED Aphanomyces TOWARD AN IMPROVED DIAGNOSTIC TECHNIQUE

By

## SEYEDEH FATEMEH AFZALI

#### August 2014

#### Chair: Associate Professor Hassan Hj Mohd Daud, PhD Faculty: Veterinary Medicine

Epizootic ulcerative syndrome (EUS) is a seasonal and severely damaging disease in wild and farmed freshwater and estuarine fishes. The disease has been spread through countries of the Asia-Pacific region with dire consequences to the fish resources and livelihood of fishermen. It has been a major concern almost all over the world since 1972. Epizootic ulcerative syndrome is a disease which manifested with severe skin and muscle ulceration and caused heavy mortalities in freshwater fishes. The aquatic fungus, Aphanomyces invadans, which belongs to the family Saprolegniacea, has been identified as the causative agent of EUS. Up to date no effective prophylactic measures and no protective vaccines are available against this disease. If scientific development could not solve this microbiological problem, it is likely to impact a noticeable negative income in the future especially for fish farmers who rely on wild-caught fish for income. Thus this study aimed to (i) isolate and identify Aphanomyces spp. from Malaysian water bodies and fish farms, (ii) determine the pathogenicity of A. invadans on the Malaysian local fish, and (iii) improve a molecular technique (PCR) for a rapid and reliable detection of EUS infection.

Four hundred sixty one water and 235 fish were sampled from different water bodies and fish farms in Selangor state of Malaysia from February 2011 until February 2013. Oomycete fungi were isolated by applying bait methods using hempseed and corn, and identified according to their hyphae, sporangium and oogonium morphological characteristics.

Through experimentally infection studies, Snakehead fish (*Channa striata*) (positive control), Moonlight gourami (*Trichopodus microlepis*), Snakeskin gourami (*Trichopodus pectoralis*), Koi carp (*Cyprinus carpio carpio*), Broadhead catfish (*Clarias macrocephalus*), Goldfish (*Carasius auratus auratus*), Climbing perch (*Anabas testudineus*) and Tilapia (*Oreochromis niloticus*) (negative control) were challenged by intramuscular injection and cohabitation using zoospores of a reference *A. invadans* NJM9701 (isolated from naturally infected Ayu by Dr. Hatai in Japan, 1997). *Aphanomyces invadans* was able to be re-isolated from experimentally infected Moonlight gourami and Koch's postulates were fulfilled to confirm the exact source of infection in this study. *Aphanomyces invadans* DNA were extracted from experimentally infected fish skin and muscle at different days of post

i

inoculation and were detected by the PCR method by using the primer set 1APM 1 F, 1APM 6R which were commercially available in the market.

From 73 water samples which were positive for fungi, 31 isolates were identified as Saprolegnia spp., 27 isolates as Achlya spp., 12 isolates as Aphanomyces spp., and three isolates as Allomyces spp. Among of 235 naturally infected fish, 62 samples were positive for fungi infection which identified as Saprolegnia (34 samples) and Achlya (28 samples). Snakeheads experimentally infected with local isolates of Aphanomyces did not show any EUS typical clinical signs and no mortalities were observed in any group during observation period, which indicated that the local isolates of Aphanomyces spp., were of saprophytic strains. Snakehead, Gouramy, Koi carp, Broadhead catfish, Goldfish and Climbing perch injected with zoospores from reference strain developed lesions that were grossly and histopathologically identical to those observed in naturally infected fish and 100% mortalities were observed. Histopathological studies showed severe cellular inflammatory infiltration, granulomatous formations and presence of invasive fungal hyphae in zoospores injected fish skins and muscles. The DNA extraction protocol used in this study was successful in isolating A. invadans genomic DNA from fish muscles and pure cultured fungus, and the improved PCR assay also was able to detect the presence of A. invadans DNA in experimentally infected fish skin and muscle from day one post inoculation.

This study was the first research conducted on freshwater aquatic fungi in Malaysia and successfully showed the presence of *Aphanomyces* spp., and other oomycete fungi in Malaysian water bodies. It is found that Malaysian Moonlight gourami, Snakeskin gourami, Koi carp and Broadhead catfish are highly susceptible while Goldfish and Climbing perch are moderately susceptible to infection by *A. invadans* via intramuscular injection. The infection is also capable of being transferred to healthy susceptible fish through the water column. It is concluded that by applying PCR assay *A. invadans* could be detected in clinical samples in very early stages of disease. Because of the presence of *Aphanomyces* spp., and EUS-susceptible fish in freshwater resources of Malaysia, there is potential risk of EUS outbreak in the region which thus must be avoided by good prophylactic measures and rigid farm biosecurity. Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

## PEMENCILAN, PENCIRIAN DAN PATOGENISITI Aphanomyces BERKAITAN SINDROM EPIZOOTIK ULSERATIF, KE ARAH PENAMBAIKAN TEKNIK DIAGNOSTIK

Oleh

## SEYEDEH FATEMEH AFZALI

#### **Ogos 2014**

#### Pengerusi: Professor Madya Hassan Hj Mohd Daud, PhD Fakulti: Perubatan Veterinar

Sindrom epizootik ulseratif (EUS) adalah penyakit bermusim dan membawa kerosakan yang serius dalam ikan air tawar dan muara yang diternak atau liar. Penyakit ini telah merebak ke serata negara di kawasan Asia-Pasifik dengan kesan yang menakutkan kepada sumber perikanan dan kehidupan nelayan. Ianya telah menjadi perhatian utama di seluruh dunia semenjak tahun 1972. Sindrom epizootik ulseratif adalah penyakit yang menunjukkan ulser teruk di kulit dan otot dan menyebabkan kematian yang tinggi dalam ikan air tawar. Fungus Aphanomyces invadans, yang tergolong dalam keluarga Saprolegniacea telah dikenalpasti sebagai agen penyebab EUS. Setakat ini tidak ada langkah profilaktik yang efektif dan vaksin perlindung terhadap EUS. Jika pembangunan saintifik tidak berjaya menyelesaikan masalah mikrobiologi ini, ianya akan memberi impak negatif yang jelas pada masa hadapan terutama kepada yang bergantung terhadap tangkapan ikan liar sebagai sumber pendapatan. Oleh itu kajian ini bertujuan untuk (i) memencil dan mengenalpasti spesis Aphanomyces daripada perairan dan ladang ikan, (ii) menentukan patogenisiti A. invadans dalam ikan tempatan Malaysia, dan (iii) peningkatkan teknik molekular (PCR) untuk pengesanan jangkitan EUS secara cepat dan kebolehpercayaan.

Sampel air dan ikan telah diperoleh di perairan dan ladang ikan yang berlainan dari Februari 2011 sehingga Februari 2013 dalam Selangor, Malaysia. Kulat Oomycete dipencilkan melalui kaedah umpan menggunakan biji hemp dan jagung dan ia dikenalpasti mengikut pencirian hifa, sporangium dan oogonium.

Melalui kajian infeksi artifisial, ikan Haruan (*Channa striata*), Gourami bulan (*Trichopodus microlepis*), Gourami kulit ular (*Trichopodus pectoralis*), ikan Koi (*Cyprinus carpio var. carpio*), ikan Keli bunga (*Clarias macrocephalus*), ikan Mas (*Carasius auratus var. auratus*), ikan Puyu (*Anabas testudineus*) dan Tilapia (*Oreochromis niloticus*) telah disuntik dengan zoospora rujukan *A. invadans* (NJM9701) secara intraotot dan jangkitan secara kohabitasi. Untuk memenuhi postulat Koch, *A. invadans* telah berjaya diisolasi semula daripada ikan Gourami bulan yang telah dijangkiti secara artifisial. Asid deoksiribonukleik (DNA) *A. invadans* telah diekstrak daripada tisu ikan tersebut pada peringkat infeksi penyakit yang berbeza dan dikenalpasti melalui kaedah reaksi berantai polimerase (PCR) menggunakan pasangan primer "1APM 1 F, 1APM 6R" yang ada dalam pasaran.

Daripada 73 sampel air positif kulat, 31 isolat telah dikenalpasti sebagai Saprolegnia spp., 27 isolat Achlya spp., 12 isolat Aphanomyces spp., dan tiga isolat Allomyces spp. Dalam kalangan 235 ikan yang terinfeksi, 62 sampel adalah positif kulat Saprolegnia (34 sampel) dan selebihnya adalah Achlya (28 sampel). Ikan haruan yang telah dijangkiti dengan isolat Aphanomyces tempatan tidak menunjukkan sebarang tanda klinikal lazim EUS dan tiada sebarang kematian dilihat sepanjang tempoh pemerhatian dalam mana-mana kumpulan ikan. Ini menunjukkan bahawa isolat Aphanomyces spp. tempatan adalah strain saprofitik. Ikan Haruan, Gourami, Koi dan Keli bunga, ikan Mas, dan Puyu yang telah disuntik dengan zoospora rujukan dilihat mengalami lesi yang serupa dengan simptom yang ada pada ikan dijangkiti EUS secara semulajadi dan 100% kematian telah dicatatkan. Pemerhatian histopatologi menunjukkan terdapat inflamasi susupan sel yang teruk, pembentukan granuloma dan kehadiran hifa kulat invasif pada kulit dan otot ikan yang disuntik zoospora. Kaedah pengekstrakan DNA yang digunakan telah berjaya memencilkan DNA genomik A. invadans dari otot ikan dan kulat yang dikultur, dan asai PCR juga berjaya mengenalpasti kehadiran DNA A. invadans daripada tisu ikan yang dijangkiti secara artifisial mulai hari pertama selepas suntikan.

Kajian ini merupakan kajian ulung yang dilakukan terhadap kulat akuatik air tawar di Malaysia dan telah berjaya membuktikan kehadiran *Aphanomyces* spp. dan kulat oomysit yang lain dalam perairan di Malaysia. Selain itu, didapati ikan Gourami bulan, Gourami kulit ular, Koi dan Keli bunga sangat peka terhadap infeksi *A. invadans* secara intarotot manakala ikan Mas dan Puyu adalah sederhana peka. Infeksi ini juga didapati mudah untuk menjangkiti ikan peka yang sihat melalui kolum air. Asai PCR yang dilakukan sangat sensitif terhadap kehadiran *A. invadans* dalam sampel klinikal dan dapat dikenalpasti pada peringkat awal jangkitan lagi. Disebabkan kulat spesis *Aphanomyces* dan ikan peka-EUS dijumpai di dalam sumber air tawar di Malaysia, terdapat kemungkinan berlakunya ledakan penyebaran EUS di kawasan ini. Justeru, langkah profilatik yang baik dan kaedah penternakan yang mementingkan biokeselamatan perlulah diberi perhatian.

### ACKNOWLEDGEMENTS

All praise and gratitude will be to God the almighty for his mercy and support during course of our life and moments of truth.

First and foremost, I would like to acknowledge my deep gratitude and appreciation to my dear supervisors Associate Professors Dr. Hassan, Dr. Rahim and Dr. Sharifpour for their continual support and endless encouragement and patience, without all nothing would have been accomplished.

My special thanks go to Dr. Birgit Oidtmann (England) for sending us *A. invadans* strain (NJM9701) which without it my research could not be done, I could never forgot her kindness. I also would like to thank Dr. Hatai from Japan who helped me in identification of aquatic fungi, and Dr. Lilley from England for sharing his vast knowledge of fungi isolation with me.

I hereby would like to thank all people who somehow helped me to fulfill my PhD research program.



I certify that a Thesis Examination Committee has met on 13 August 2014 to conduct the final examination of Seyedeh Fatemeh Afzali on her thesis entitled "Isolation, Characterization and Pathogenicity of Epizootic Ulcerative Syndrome-Related *Aphanomyces* Toward an Improved Diagnostic Technique" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

#### Mohamed Ali bin Rajion, PhD

- Professor Faculty of Ve

Faculty of Veterinary Medicine Universiti Putra Malaysia (Chairman)

## Mohd Zamri bin Saad, PhD Professor

Faculty of Veterinary Medicine Universiti Putra Malaysia (Internal Examiner)

#### Siti Khairani binti Bejo, PhD

Associate Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Internal Examiner)

#### K.M Shankar, PhD

Professor Animal and Fisheries Sciences University India (External Examiner)

NORITAH OMAR, PhD Associate Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 19 September 2014

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

#### Hassan Hj Mohd Daud, PhD

Associate Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Chairman)

## Abdul Rahim Mutalib, PhD

Associate Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Member)

## Issa Sharifpour, PhD

Associate Professor Department of International and Scientific Relations and Information Iranian Fisheries Research Organization (Member)

## Jasni Bin Sabri, PhD

Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Member)

## **BUJANG BIN KIM HUAT, PhD**

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

## DECLARATION

#### **Declaration by graduate student**

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:

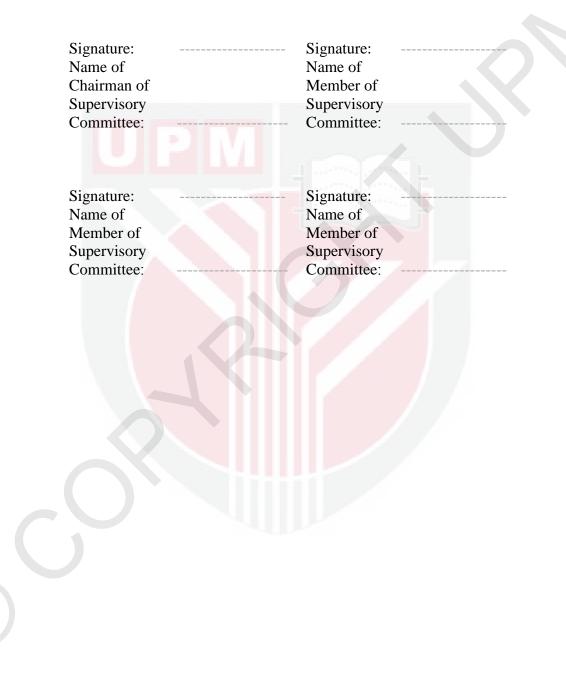
\_ Date:

Name and Matric No.: Seyedeh Fatemeh Afzali, GS29933

## **Declaration by Members of Supervisory Committee**

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.



# TABLE OF CONTENTS

		Page
ABSTRACT		i
ABSTRAK		iii
ACKNOWLED	GEMENTS	v
APPROVAL		vi
DECLARATION	N	viii
LIST OF TABL	ES	xii
LIST OF FIGUE	RES	xiii
LIST OF APPE	NDICS	xxii
LIST OF ABBR	EVIATIONS	xxiii
CHAPTER		
	TRODUCTION	1
	Objectives	3
	Hypothesis	3
	TERATURE REVIEW	4
2.1	Epizootic Ulcerative Syndrome (EUS)	4
	2.1.1 EUS Outbreak	4
	2.1.2 Etiology Agent	5
	2.1.3 Epidemiology of EUS	6
	2.1.4 Environmental EUS Risk Factors	9
	2.1.5 Pathogenesis	9
	2.1.6 EUS Clinical Signs and Gross Pathology	10
2.2	Aphanomyces invadans	11
	2.2.1 Life Cycle of A. invadans	11
	2.2.2 Transmission Mechanisms	13
	2.2.3 Ecology of A. invadans	13
2.3		13
	2.3.1 Taxonomy of Aphanomyces spp.	15
	Isolation and Identification of Saprolegniaceae	16
2.5	Characterization and Diagnosis of EUS	18
	2.5.1 Histopathology and Experimental Infection Study	18
	2.5.2 Polymerase Chain Reaction (PCR)	21
	Economic and Social Impacts of EUS	23
2.7	Importance of EUS as OIE-Listed Disease	24
3 ISC	DLATION AND IDENTIFICATION OF Aphanomyces	25
	ECIES FROM NATURAL WATER BODIES AND FISH	
	RMS IN SELANGOR, MALAYSIA	
	Introduction	25
3.2	Materials and Methods	28
	3.2.1 Sampling	28
	3.2.2 Pilot Study	30
	3.2.3 Fungi Isolation and Identification	30
	3.2.4 Histopathology Examination	32
3.3	Results	33
	3.3.1 Isolation and Identification	33
	3.3.2 Histopathology	43

Х

	3.4 Discussion and Conclusion	47
4	EXPERIMENTAL INFECTION OF EPIZOOTIC ULCERATIVE SYNDROME (EUS) AGENT, A. invadans IN MALAYSIAN LOCAL FISH USING ZOOSPORES	50
	4.1 Introduction	50
	4.2 Materials and Methods	51
	4.2.1 Aphanomyces invadans strain	51
	4.2.2 Maintenance of <i>A. invadans</i> Cultures	51
	4.2.3 Inducing Sporulation in <i>A. invadans</i> Cultures	52
	4.2.4 Experimental Challenge	52
	4.2.5 Re-isolation of <i>A. invadans</i> and Fulfilling Koch's Postulate	53
	4.2.6 Histopathology	54
	4.2.7 Statistical Analyzing	54
	4.3 Results	56
	4.3.1 Infection by Intramuscular Injection	56
	4.3.2 Statistical Analysing	94
	4.3.3 Infection by Cohabitation	96
	4.3.4 Re-isolation of <i>A. invadans</i> and Fulfilling Koch's	97
	postulate	00
	4.4 Discussion and Conclusion	98
5	IMPROVEMENT OF POLYMERASE CHAIN REACTION	103
	(PCR) METHOD FOR DETECTION A. invadans	
	5.1 Introduction	103
	5.2 Methodology	103
	5.2.1 Fungi and Fish Tissues Tested	103
	5.2.2 DNA Preparation	105
	5.2.3 Aphanomyces invadans-specific Primers	106
	5.2.4 Single PCR Assay	106
	5.2.5 Agarose Gel Electrophoresis 5.3 Results	107 108
	5.3.1 Detection of <i>A. invadans</i> in Fish Muscle	108
	5.3.2 Detection of <i>Aphanomyces</i> spp. and other	115
	Oomycete Fungi	115
	5.4 Discussion and Conclusion	116
6	SUMMARY, CONCLUSION AND RECOMMENDATION FOR FUTURE RESEARCH	120
	6.1 Summary	120
	6.2 Conclusion and Recommendation	123
		10-
	RENCES	125
	NDICES	141
	ATA OF STUDENT DF PUBLICATIONS	150 151
		1.51

# LIST OF TABLES

Table		Page
2.1	Fish species susceptible to infection with A. invadans	8
3.1	Sampling stations and the number of water samples which was taken from each station	29
3.2	Aquatic fungi found in 14 stations of Selangor state water bodies	33
3.3	Aquatic fungi isolated from naturally infected fish in Selangor state	34
3.4	Aquatic fungi isolated from naturally infected fish in Selangor state	35
4.1	Characteristic histopathological findings compared among eight fish species infected with <i>A. invadans</i>	93
4.2	Skin lesion score and significancy of disease in <i>A. invadans</i> -experimentally challenged fish	94
5.1	Oomycete isolates used for PCR	104
5.2	Clinical specimens were tested in the PCR assays. Amplification and absence of amplification are shown as + and – respectively	109

# LIST OF FIGURES

Figure		Page
2.1	Map showing the geographical spread of EUS in the last three decades	5
2.2	Smear preparation of <i>A. invadans</i> . Typical <i>A. invadans</i> non-septate hyphae are shown with cluster of encysted primary zoospores (arrow) (X100)	12
2.3	The asexual life cycle of <i>A. invadans</i>	12
2.4	Zoosporangia formation and dehiscence in <i>Saprolegnia</i> , <i>Achlya</i> and <i>Aphanomyces</i>	14
3.1	Map of peninsular Malaysia showing the location of Selangor state (arrow)	26
3.2	Isolation of fungi by baiting methods. (a) Showing fungal hyphae growing on maize bait (circle), and (b) hemp seeds bait (circle)	30
3.3	Fungal infected fish collected from natural water bodies and fish farms. (a) Snakehead fish (arrow) and (b) Climbing perch (arrow) with red ulcers, (c) River catfish with cotton like whitish colonies on the head and body surface	31
3.4	Identical asexual reproduction <i>Saprolegnia</i> spp. isolated from water and fish. Wet mount preparation of <i>Saprolegnia</i> showing (a) aseptate hyphae and sporangium (arrow) with immature zoospores inside (SAWP01 isolated from recreational pond) (Bar = 65 $\mu$ m), (b) Saprolegnoid mode of zoospore release in <i>Saprolegnia</i> sp. (SACF02 isolated from Snakehead fish) (Bar 40 $\mu$ m)	36
3.5	Fruiting bodies of asexual and sexual reproductions of <i>Achlya</i> isolated from natural pond (ACWP02) and Shark catfish (ACCF01). (a) Asexual reproduction: mature sporangium (large arrow) and new sporangium (small arrow). (b) Sexual reproduction: Oogonium (female) (circle) with oospores inside (arrow) touched by many antheridial branches (male). (c) High magnification of Oogonium with Oospores inside. (d) High magnification of Oogonium with antheridium (arrow) attached	37
3.6	Morphological characteristics of <i>Allomyces</i> sp., isolate ALWP02. (a) Vacuolate vegetative hyphae showing dichotomous branch (arrow), septate hyphae with rhizoid structure (R) (Bar 100µm). (b) Zoosporangia in chains (circle) and various sites of exit pores (arrow) (Bar 20µm)	38

- 3.7 Wet mount preparation of saprophytic *Aphanomyces* spp. isolated 39 from different water bodies. Showing aseptate vegetative hyphae (arrow) and zoospores with Achlyoid type clusters (circle) (Bar 50µm). (a) ASFF01 and (b) ASFF02 isolated from fish farm. (c) ASP07 isolated from natural pond. (d) ASE06 isolated from estuary. (e) ASP08 isolated from recreational pond. (f) ASL010 isolated from lake
- 3.8 Asexual reproduction of *Aphanomyces* spp. (a-b) Cluster of encysted 40 primary zoospores. (C) Secondary zoospore
- 3.9 Cultural characteristics of isolated fungi cultured on Glucose–Yeast 41 (GY) media. (a) Cotton like and whitish colonies of *Saprolegna* sp. isolate SAWP05. (b) Puffy and whitish colonies of *Achlya* sp. isolate ACCF03. Colonies of *Aphanomyces* sp. isolate ASFT6 (c) and *Allomyces* sp. isolate ALWP01 (d) growing on hemp seeds
- 3.10 Snakehead *Channa Striata* experimentally injected with saprophytic 42 *Aphanomyces* isolate ASFT02 showed some reddening in injection area which was healed after 3 dpi
- 3.11 Mild organizing macrophage responses to the injection of the 43 saprophytic *Aphanomyces* isolate ASFT02. No fungus and only very limited myonecrosis were detected in this section at 7 dpi (H&E, X200)
- 3.12 Gross and microscopic pathological changes of skin of naturally 44 infected Snakehead *Channa Striata* by *Saprolegnia* sp. (a) Grey whitish cotton like growth (arrow) on *Saprolegnia* sp. infected Snakehead. (b) Normal muscle of uninfected Snakehead. (c) Degeneration, severe necrotizing, distribution of melanin pigments (arrow) with mild cellular infiltration (CI) in skin (H&E, X200)
- 3.13 Gross and microscopic pathological changes of skin of infected Silver
  barb (*Puntius* sp.) by *Saprolegnia dicilina*. (a) Whitish discoloured patch (arrow) on *Saprolegnia dicilina* dorsal muscle in infected Silver barb. (b) Normal skin and muscle of uninfected River barb. (c) Degeneration and severe necrotizing (arrow). (d) Severe necrosis (N) and distribution of melanin pigments (arrow) in skin (H&E, X200)
- 3.14 Gross and microscopic pathological changes of skin of infected Shark 46 catfish (*Pangasius* sp.) by *Achlya* sp. (a) Cotton like whitish colony on infected fish body (arrow). (b) Normal muscle of uninfected Shark catfish. (c) Muscle necrosis (N), macrophages engulfing muscle debris (arrow) and (d) Muscle necrosis (N) with melanin pigments diposition (arrow) of infected Shark catfish (H&E, X200)
- 4.1 The injection site (star) in intramuscularly infected fish where placed 53 at the left side of the body below the dorsal fin.

xiv

- 4.2 Visual description of skin lesion scoring system of examined fish is 55 showed using EUS-affected snakehead lesions. (a) Score 1: Skin blanching, lost of scale and epithelial cells. (b) Score 2: Red spot and marked swelling. (c) Score 3: Ulcerative lesion. (d) Score 4: Deep ulcers involving underlying muscles
- 4.3 Smear preparation of EUS fungus "*Aphanomyces invadans*". (a) Typical *A. invadans* non-septate hyphae showing cluster of encysted primary zoospores (X200). (b) Achyloid clusters (arrow), hyphae and lateral evacuation tube of *A. invadans*, (X400)

56

- 4.4 Snakehead experimentally injected by *A. invadans* zoospores isolate 57 JM9701. Showed (a) some red hemorrhagic lesions on the injected site (6 dpi), and (b) dermal ulcer penetrating musculature (9 dpi)
- 4.5 Histopathological characteristic of Snakehead intramuscularly 58 infected by *A. invadans* NJM9701 zoospores. The site of injection showed: (a) Disorganization of muscle fibers, mono-nucleated inflammatory cells and edema at 1 dpi. (b) Thickening of blood vessels and hemorrhage (H) at 2 dpi (H & E, 200X, Bar = 80µm). (c) Myonecrosis at 2 dpi (H & E, 100X, Bar = 160µm). (d) Myophagia (M) and severe muscle degeneration (MD) in the lesion area at 4 dpi (H & E, 400X, Bar = 40µm)
- 4.6 Histopathological characteristic of Snakehead intramuscularly 60 infected by *A. invadans* NJM9701 zoospores. The site of injection showed: (a) Free fungal hyphae (arrow) with melanin deposit around the area at 6 dpi (PAS, 200X, Bar =  $20\mu$ m). (b) Initiation of granulomatous formation (arrow) and severe cellular infiltration (CI) at 8 dpi (H & E, 200X, Bar =  $80\mu$ m). (c) High magnification of a granuloma with necrotic center which is surrounded by fibroblast layers (F) at 12 dpi (H & E, 400X, Bar =  $40\mu$ m). (d) Fusion of granulomata leading to the formation of a giant granulomata (G) with necrotic deposition at centre at 14 dpi (H & E, 100X, Bar =  $160\mu$ m)
- 4.7 Moonlight gourami experimentally infected by *A. invadans* zoospores isolate NJM9701. Showed (a) whitish fungal colonies with red ulcer on injection site (5 dpi), and (b) deep penetrating focal red ulcer exposed the underlying musculature (8 dpi)
- 4.8 Snakeskin gouramis experimentally infected by *A. invadans* 62 zoospores isolate NJM9701. Showed whitish fungal colonies and red ulcer on injection site and died at 9 dpi
- 4.9 Histopathological characteristic of Moonlight gourami 64 intramuscularly infected by *A. invadans* NJM9701 zoospores. The site of injection showed: (a) severe cellular infiltration (CI) at 24 h after inoculation. (b) Severe Myophagia (M) with moth-eaten muscle fibers at 2 dpi. (c) Empty blood vessel (arrows) and hemorrhages (H) at 4 dpi (H & E, 200X, Bar = 20µm). (d) Free fungal hyphae in the

infected area at 6 dpi (arrows) (PAS, 100X, Bar =  $100\mu m$ )

- 4.10 Histopathological characteristic of Snakeskin gourami 65 intramuscularly infected by *A. invadans* NJM9701 zoospores. The site of injection showed: (a) Mononuclear inflammatory response and cellular infiltration (CI) with initiation of Myophagia (M) at 1 dpi. (b) Myophagia (M), muscle degeneration (MD), shrinkage and necrosis with free hyphae inside (arrow) at 2 dpi. (c) Severe Myophagia with empty blood vessles (arrows) at 4 dpi (H & E, 200X, Bar = 20µm). (d) Free fungal hyphae in the infected area at 6 dpi (arrows) (PAS, 100X, Bar = 100µm)
- 4.11 Histopathological characteristic of Moonlight gourami 66 intramuscularly infected by *A. invadans* NJM9701 zoospores. The site of injection showed: (a) Initiation of granulomatous formation (arrow) at 8 dpi (Bar =  $20\mu$ m). (b) Severe myonecrosis with increase of the number of granulomata (arrows) at 10 dpi (Bar =  $20\mu$ m). (c) Fusion of granulomata leading to formation giant granulomata (G) at 12 dpi (Bar =  $20\mu$ m). (d) Vaculization and Rupture of the muscles, macrophages activities, and free fungal hyphae (arrows) at 14 dpi (Bar =  $80\mu$ m) (H & E, 200X)
- 4.12 Histopathological characteristic of Snakeskin gourami 67 intramuscularly infected by *A. invadans* NJM9701 zoospores. The site of injection showed: (a) Vacuolization (V) and initiation of fibroblast activity and granulomatous reaction (circles) at 8 dpi. (b) Increase of the number of granulomata surrounded by fibroblas layers (arrows) at 10 dpi. (c) Fusion of granulomata resulting in formation giant granulomata (G) with fungal hyphae at the center at 12 dpi. (d) High magnification of giant granulomata with fungal hyphae (arrows) inside (H & E, 200X, Bar = 20µm).
- 4.13 Koi carp fish experimentally infected by *A. invadans* zoospores 68 isolate NJM9701. Showed (a) reddening with scale loss (4 dpi), and (b) deep red ulcer on skin (8 dpi)
- 4.14 Histopathological characteristic of Koi carp intramuscularly infected by *A. invadans* NJM9701 zoospores. The site of injection showed: (a) Muscle degeneration (MD) with severe hemorrhages at 1 dpi (H & E, 200X, Bar =  $80\mu$ m). (b) Higher magnification of muscle with hemorrhages spots inside the infected area at 2 dpi (H & E, 400X, Bar =  $40\mu$ m). (c) Presence of non-capsulated hyphae (arrows) at 4 dpi (PAS, 200X, Bar =  $50\mu$ m). (d) Severe inflammatory response with replacement of muscle by fibrous tissue (F), and melanin deposition in infection areas (arrows) at 8 dpi (H & E, 100X, Bar =  $160\mu$ m)
- 4.15 Histopathological characteristic of Koi carp intramuscularly infected 72 by *A. invadans* NJM9701 zoospores. The site of injection showed: (a) A granuloma (circle) surrounded by epithelioid cells, and blood

vessels containing inflammatory cells (arrow) at 10 dpi (H & E, 200X, Bar =  $80\mu$ m). (b) Severe inflammatory response and initiation of fibroblast activities with hemorrhages in muscle fibers (arrows) at 12 dpi (H & E, 200X, Bar =  $40\mu$ m). (c) Severe muscle degeneration (MD) with a limited numbers of small granulomata (arrows) at 14 dpi (H & E, 400X, Bar =  $40\mu$ m). (d) Non-capsulated hyphae (arrows) in necrotic areas at 18 dpi (PAS, 200X, Bar =  $20\mu$ m)

73

75

- 4.16 Broadhead catfish experimentally infected by *A. invadans* zoospores isolate NJM9701. Showed (a) red ulcer (6 dpi), and (b) deep red ulcer with losing skin color (10 dpi) on injected site
- 4.17 Histopathological characteristic of Broadhead catfish intramuscularly infected by *A. invadans* NJM9701 zoospores. The site of injection sh owed: (a) Blood vessel containing inflammatory cells (arrow) and Myophagia (M) at 1 dpi (H & E, 400X, Bar = 40 $\mu$ m). (b) Severe muscle degeneration, edema (E), hemorrhages (H) and severe Myophagia (M) with formation of Langhan type giant cells (LG) at 2 dpi (H & E, 400X, Bar = 40 $\mu$ m). (c) Presence of non-capsulated hyphae (arrows) at 4 dpi (PAS, 200X, Bar = 50 $\mu$ m). (d) Langhan (LG) and Foreign body (FG) type giant cells with hyphae in center at 8 dpi (H & E, 400X, Bar = 40 $\mu$ m)
- 4.18 Histopathological characteristic of Broadhead catfish intramuscularly 77 infected by *A. invadans* NJM9701 zoospores. The site of injection showed: (a) Complete degeneration of muscle fibers, increase of cellular infiltration (CI) with formation of multinucleated giant cells (arrow) at 10 dpi (H & E, 200X, Bar = 80µm). (b) A granuloma (circle) comprising two layers connective tissue with giant cells inside, and Langhan type giant cells (LG) with hyphae at center (arrow) at 12 dpi. (c) Foreign body type giant cells with connective tissues around (FG) at 14 dpi. (d) Severe spread of free fungal hyphae (arrow) in infection area at 21 dpi (H & E, 400X, Bar = 40µm)
- 4.19 Goldfish experimentally infected by *A. invadans* zoospores isolate 78 NJM9701. Showed (a) whitish fungal colony (4 dpi), and (b) deep red ulcer on injection site (8 dpi)
- 4.20 Histopathological characteristic of Goldfish intramuscularly infected by *A. invadans* NJM9701 zoospores. The site of injection showed: (a) Hemorrhages inside the muscles with mononuclear inflammatory cells at 2 dpi (H & E, 200X, Bar =  $80\mu$ m). (b) Muscles degeneration with severe Myophagia (M) at 4 dpi. (c) Presence of both types of multinucleated giant cells (arrows), Langhans (LG) and Foreign body (FG) giant cells at 6 dpi. (d) A granuloma (circle) with necrotic center surrounded by lacunae-like cells (arrows) at 8 dpi, (H & E, 400X, Bar =  $40\mu$ m)
- 4.21 Histopathological characteristic of Goldfish intramuscularly infected 82 by *A. invadans* NJM9701 zoospores. The site of injection showed: (a)

Increasing the number of granulomata surrounded by fibroblast layers (arrows) at 10 dpi (H & E, 200X, Bar =  $80\mu$ m). (b) Free fungal hyphae in necrotic area (arrows) at 12 dpi (PAS, 200X, Bar =  $80\mu$ m). (c) Severe muscle necrosis and degeneration of muscle bundle at 12 dpi. (d) Well developed granulomata (G) with necrotic area and degenerated hyphae at the center at 21 dpi (H & E, 100X, Bar =  $80\mu$ m)

83

85

- 4.22 Climbing perch experimentally infected by *A. invadans* zoospores isolate NJM9701. Showed (a) red ulcer and scale loss (6 dpi), and (b) open red ulcer (10 dpi) on injected site
- 4.23 Histopathological characteristic of Climbing perch intramuscularly infected by *A. invadans* NJM9701 zoospores. The site of injection showed: (a) Mononuclear inflammatory response, severe Myophagia (M) and hemorrhages (H) at 2 dpi (H & E, 400X, Bar = 40µm). (b) Severe muscle degeneration (MD) with initiation of granulomatus reaction (circle) at 4 dpi (H & E, 200X, Bar = 80µm). (c) Increase of cellular infiltration (CI), granulomatus response (circle) and formation of fibroblast layers (F) around granulomata at 6 dpi (H & E, 100X, Bar = 160µm). (d) Presence of encapsulated hyphae at the center of granulomata and non-capsulated hyphae (arrows) in infection area at 8 dpi (PAS, 200X, Bar = 50µm)
- 4.24 Histopathological characteristic of Climbing perch intramuscularly infected by *A. invadans* NJM9701 zoospores. The site of injection showed: (a) Increasing of the number of granulomata surrounded by fibroblast layers (arrows) at 10 dpi. (b) A typical granuloma (circle) comprising connective tissues and epithelioid cells encapsulated fungus hyphae in the mycotic lesion at 12 dpi (H & E, 400X, Bar =  $40\mu$ m). (c) Fusion of granulomata resulting in formation giant granulomata (G) between muscle fibers at day 14 pi (H & E, 100X, Bar =  $160\mu$ m). (d) Formation of hematoma with blood clot (BC) inside at 28 dpi (H & E, 200X, Bar =  $80\mu$ m)
- 4.25 Tilapia experimentally infected by *A. invadans* zoospores isolate 88 NJM9701. Showed (a) reddening on injection site at 2 dpi, and (b) recovered after a couple of days (14 dpi)

4.26 Histopathological characteristic of Tilapia intramuscularly infected by *A. invadans* NJM9701 zoospores. The site of injection showed: (a) Severe Hemorrhages (H) and edema at 1 dpi (H & E, 200X, Bar = 80µm). (b) Severe Myophagia (M), and initiation of encapsulating response by giant cells around the hyphae in the mycotic lesion (arrows) with lacunae-like cells (diamond) around at 2 dpi (H & E, 400X, Bar = 40µm). (c) Foreign body type giant cell (arrow) and well developed granulomata surrounded by fibroblast layers and degenerated hyphae inside (circle) at 4 dpi (H & E, 200X, Bar = 80µm). (d) Increasing of granulomatous tissues which filled the entire defect areas and initiation of healing process at 8 dpi (H & E, 100X,

xviii

#### $Bar = 160 \mu m$ )

- 4.27 Histopathological characteristic of Tilapia intramuscularly infected by *A. invadans* NJM9701 zoospores. The site of injection showed: (a) Encapsulation fungi by Foreign body type giant cells (FG) and a granuloma (arrow) surrounding by thick fibroblast layers at 14 dpi (H & E, 400X, Bar = 40µm). (b) A big granuloma encapsulated many fungi in the mycotic lesion at 21 dpi (PAS, 200X, Bar = 20µm). (c) Fusion of granulomata and formation giant granulomata (G) with degenerated fungus hyphae and necrotic material (N) inside at center at 28 dpi (H & E, 200X, Bar = 80µm). (d) Regenerated muscle in healed area at 35 dpi (H & E, 200X, Bar = 80µm)
- 4.28 Final score of susceptibility among examined fish species. SP1: 95 Snakehead; SP2: Moonlight gourami; SP3: Snakeskin gourami; SP4: Koi carp; SP5: Broadhead catfish; SP6: Goldfish; SP7: Climbing perch; SP8:Tilapia
- 4.29 Gross pathology and histopathological characteristics of Snakehead 96 (*Channa Striata*) infected by cohabitation at 14 dpi. (a) Snakehead showing red ulcers on the body surface. (b) Degeneration of muscle fiber (MD) and formation of granolumata (arrows) with fibroblast layers around (H&E, X400, Bar =  $40\mu$ )
- 4.30 Gross pathology and histopathological characteristics of Moonlight 97 gourami infected by *A. invadans* zoospores isolate MG001. (a) Moonligh gourami showed EUS-like lesion on injection site at 6 dpi. (b) EUS-effected moonlight gourami showed secondary infection of *Saprolegnia* sp. on injection site at 3 dpi. (c) Achlyoid cluster (arrows) of primary zoospores of *A. invadans* isolate MG001. (d) Degeneration of muscle fiber (MD), severe cellular infiltration (CI), hemorrhages and formation of granolumata (arrows) with fibroblast layers around in infection area (H&E, X200, Bar = 20μ)
- 5.1 Agarose gel showing the detection of PCR products. The left margin 108 in figure (M) indicates the position of size markers in base pairs (100-1000 bp). Lane N: negative control with no DNA template. Lanes P: positive control with genomic DNA of *A. invadans* NJM 9701. Lanes 1: genomic DNA of experimentally EUS-infected Snakehead muscle with *A. invadans* NJM 9701 zoospores. Lane 2: genomic DNA of *A. invadans* MG001 cultured hyphae
- 5.2

Agarose gel showing the PCR products, from Snakehead fish tissue 110 DNA obtained by amplification of genomic DNA of Snakehead lesion infected with *A. invadans* NJM9701. The left margin in figure (M) indicates the position of size markers in base pairs (100-1000 bp). Lane N: negative control with no DNA template. Lanes P: positive control with genomic DNA of pure cultured *A. invadans* NJM 9701. Lanes 1-9: genomic DNA of intact Snakehead from day 1, 2, 4, 6, 8, 10, 12, 14 and 21 post-injection. Lane 10: genomic DNA from non infected Snakehead injected with APW

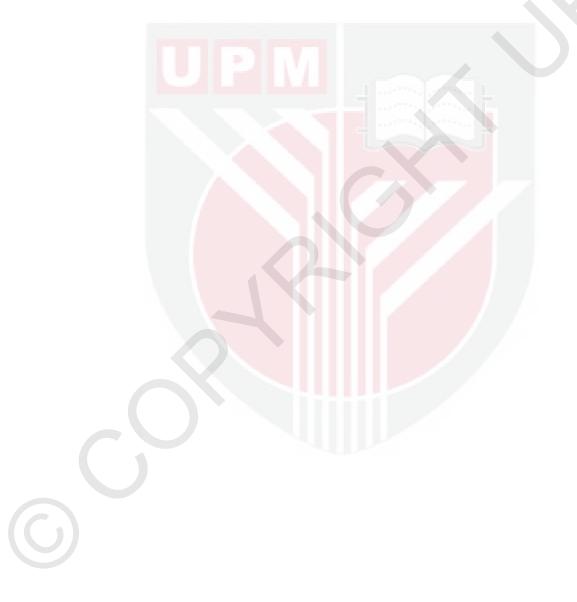
- 5.3 Agarose gel showing the PCR products, from Moonlight gourami (A) 111 and Snakeskin gourami (B) muscle DNA obtained by amplification of genomic DNA of Gouramies lesion infected with *A. invadans* NJM9701. The left margin in figure (M) indicates the position of size markers in base pairs (100-1000 bp). Lane N: negative control with no DNA template. Lanes P: positive control with genomic DNA of pure cultured *A. invadans* NJM 9701. Lanes 1-8: genomic DNA of intact Gouramies from day 1, 2, 4, 6, 8, 10, 12 and 14 post-injection. Lane 10: genomic DNA from non infected Gouramies injected with APW
- 5.4 Agarose Gel Showing the PCR Products, from Koi Carp Fish Muscle 112 DNA Obtained by Amplification of Genomic DNA of Koi Carp Infected with *A. invadans* NJM9701. The left margin in figure (M) indicates the position of size markers in base pairs (100-1000 bp). Lane N: negative control with no DNA template. Lanes P: positive control with genomic DNA of pure cultured *A. invadans* NJM 9701. Lanes 1-9: genomic DNA of intact koi carp from day 1, 2, 4, 6, 8, 10, 12, 14 and 18 post-injection. Lane 10: genomic DNA from non lesioned koi carp injected with APW
- 5.5 Agarose gel showing the PCR products, from Broadhead catfish 112 muscle DNA obtained by amplification of genomic DNA of fish infected with *A. invadans* NJM9701. The left margin in figure (M) indicates the position of size markers in base pairs (100-1000 bp). Lane N: negative control with no DNA template. Lanes P: positive control with genomic DNA of pure cultured *A. invadans* NJM 9701. Lanes 1-9: genomic DNA of intact Broadhead catfish from day 1, 2, 4, 6, 8, 10, 12, 14 and 20 post-injection. Lane 10: genomic DNA from non infected Broadhead catfish injected with APW
- 5.6 Agarose gel showing the PCR products, from Goldfish muscle tissue
  5.6 DNA obtained by amplification of genomic DNA of Goldfish lesion infected with *A. invadans* NJM9701. The left and right margins in figure (M) indicate the position of size markers in base pairs (100-1000 bp). Lane N: negative control with no DNA template. Lane P: positive control with genomic DNA of pure cultured *A. invadans* NJM 9701. Lanes 1-10: genomic DNA of intact Goldfish from day 1, 2, 4, 6, 8, 10, 12, 14, 21 and 22 post-injection. Lane 11: genomic DNA from non infected Goldfish injected with APW
- 5.7 Agarose gel showing the PCR products, from Climbing perch muscle 113 tissue DNA obtained by amplification of genomic DNA of fish lesion infected with *A. invadans* NJM9701. The left margin in figure (M) indicates the position of size markers in base pairs (100-1000 bp). Lane N: negative control with no DNA template. Lane P: positive control with genomic DNA of pure cultured *A. invadans* NJM 9701. Lanes 1-10: genomic DNA of intact Climbing perch from day 1, 2, 4,

6, 8, 10, 12, 14, 21 and 28 post-injection. Lane 11: genomic DNA from non infected Climbing perch injected with APW

- 5.8 Agarose gel showing the PCR products, from Tilapia muscle tissue 114 DNA Obtained by amplification of genomic DNA of fish lesion infected with *A. invadans* NJM9701. The left margin in figure (M) indicates the position of size markers in base pairs (100-1000 bp). Lane N: negative control with no DNA template. Lane P: positive control with genomic DNA of pure cultured *A. invadans* NJM 9701. Lanes 1-11: genomic DNA of intact Tilapia from day 1, 2, 4, 6, 8, 10, 12, 14, 21, 28 and 35 post-injection. Lane 12: genomic DNA from non infected Tilapia injected with APW
- 5.9 Agarose gel showing the detection of PCR products from oomycete 115 fungi DNA obtained by amplification of genomic DNA of fungi. The left margin in figure (M) indicates the position of size markers in base pairs (100-1000 bp). Lane N: negative control with no DNA template. Lanes P: positive control with genomic DNA of pure cultured A. *invadans* NJM 9701. Lanes 1-12: genomic DNA of 12 isolates of *Aphanomyces* spp. Lanes 13-14: genomic DNA of 2 isolates of *Saprolegnia* spp. Lanes 14-16: genomic DNA of 2 isolates of *Achlya* spp. Lane 17: genomic DNA of *Allomyces* sp

# LIST OF APPENDICS

Appendix		Page
А	Formulae for Media	142
В	Histopathology Staining Procedures	144
С	OIE-Listed diseases, infections and infestations in force in 2013	146
D	Control fish	148



# LIST OF ABBREVIATIONS

AAHRI	Aquatic Animal Health Research Institute, Thailand
ACIAR	Australian Centre for International Agriculture
	C C
APW	Autoclaved pond water
CsCasp10	Channa striata Caspase 10 (amino acid)
EDTA	Ethilen diamina tetraacetic acid
EFSA	European Food Safety Authority
EUS	Epizootic ulcerative syndrome
FAO	Food and Agriculture Organisation of the United Nations
GPY	Glucose peptone yeast
GP	Glucose peptone
GY	Glucose yeast
H&E	Haematoxylin and Eosin
IFAT	Immunofluorescence antibody technique
MAb	Monoclonal antibody
MG	Mycotic granulomatosis
MGC	Multinucleate giant cell
MW	Molecular weight
OIE	Office Internationale des Epizooties
PAb	Polyclonal antibody
PBS	Phosphate buffered saline
PG-1	Peptone glucose media one
PDA	Potato Dextrose Agar
RAPD	Random amplification of polymorphic DNA
RSD	Red spot disease
SDS	Sodium dodecyl sulphate
SDS-PAGE	Sodium dodecyl sulphate polyacrylamide gel electrophorsis
TBS	Trizma buffered saline
UM	Ulcerative mycosis
UV	Ultra-violet

6

#### **CHAPTER 1**

#### **INTRODUCTION**

Epizootic Ulcerative Syndrome (EUS) is a dangerous fish disease of wide range of fresh and brackish water wild and farmed fish throughout the world. It causes serious economic losses in many countries during the last four decades (Baldock *et al.*, 2005). The first EUS onset was reported in Japan in 1971, and later in 24 countries within four continents, viz. Northern America, Southern Africa, Asia and Australia (OIE, 2013; Oidtmann, 2011). *Aphanomyces invadans* is a causative agent of EUS (Saylor, 2010; Baldock *et al.*, 2005; Ahmed and Hoque 1999; Lilley *et al.*, 1997) by producing a proteolytic enzyme that helps it to penetrate the fish tissue causing shallow to deep ulcers (Chinabut and Roberts, 1999), leading to high mortality in fish population (Kamilya and Baruah, 2013).

The actual amount of economic losses in the aquaculture industry worldwide due to EUS is estimated to be just over USD 9 billion (Harikrishnan *et al.*, 2010) per year, which is about 15% of the value of the world's farmed fish and shellfish production. Furthermore, decreasing fish biomass causing unchangeable damage to the aquatic biodiversity is some indirect impacts of this destructive disease.

Diagnosis of EUS is difficult, as this fungus does not produce sexual structure which is essential for morphological identification. Thus, diagnosis done by observation granulomatous response in histopathology sections and must be confirmed by polymerase chain reaction (PCR) amplification. For rapid detection of uncultivable or fastidious microorganisms and characterization of the pathogen, PCR-based systems which detect the etiologic agents of disease directly from clinical samples, without the need for culture, have been useful (Tang *et al.*, 1997). It is also very specific due to the nature and orientation of the oligo-nucleotide primers that are required to allow amplification to proceed (Shariff *et al.*, 2000). Polymerase chain reaction techniques may solve the problems associated with the identification of pathogenic *A. invadans* which is so difficult and time consuming (Kuan *et al.*, 2013; Phadee *et al.*, 2004).

Epizootic ulcerative syndrome is a worldwide disease and has high mortality in farmed and wild fish. The control of disease in wild fish populations in open water bodies is most likely impossible (Fairweather, 1999), however, it is based on water treatment and management strategies (Lilley *et al.*, 1998). On the other hand, there is no effective prophylactic measure for *A. invadans*-infected fish in the wild and in aquaculture ponds. Attempts at using green water, ash, lime, salt (Noga, 2010) and neem (*Azadirachta indica*) seeds or branches for prophylactic treatments of the EUS-infected fish in fish ponds gave variable results (Clifton and Alderman, 2006), and accumulation of these residues cause pollution and made consumers reluctant during the last few years. There is no protective vaccine available (OIE, 2013), however, Snakehead fish that had been immunized with an extract of *A. invadans* elicited humoral immune response (Arockiaraja *et al.*, 2012; Thompson *et al.*, 1997). Since vaccinations are also complicated and expensive method, at present it could not be practical way for prevention EUS (Newman *et al.*, 2003).



So, if scientific development could not solve this ecological problem, it is likely to impose a noticeable cost in the future to the next generation especially for farmers who rely on fishing for income and fisher's livelihood and so on people's health. It can be expected to culture EUS resistant fish species in fish farms in the coming future to decrease fish losses arising from EUS outbreak.

Epizootic ulcerative syndrome was reported for the first time in Southern Peninsular Malaysia in 1979 and later, in rice-field fishes in Northern Malaysia and affected some Malaysian important fish like Snakehead, Snakeskin gourami, Catfish and Anabas (Lilley et al., 1998), but no scientific work was done on EUS until present. In addition the other reason which led us to conduct research on EUS in Malaysia is that EUS is listed in OIE aquatic animal diseases list (OIE, 2013) and all OIE member countries (including Malaysia) are obliged to conduct research on OIE listed diseases to make an official report for any occurrence of disease. So far, there has been conducted no studies on the aquatic pathogenic oomycetes specially EUS Aphanomyces in Malaysia which has high production and international trade of fish in the world (Ng and Tan, 1997). International trade in aquaculture animals still causes spread of major infectious diseases. Further un-restricted trade in aquatic animals without the knowledge of whether the animals from one country to another serve as a vector for a particular disease are already having a major negative impact on aquaculture (Eli, 2008). This research was conducted in three main chapters; the first chapter investigated EUS related Aphanomyces infection (ERA) in a selected area of Malaysia (Selangor state) where is economically important in terms of aquaculture and fish industry. The second chapter aimed to fulfill experimental infection studies on Malaysian local fish to investigate the susceptibility of selected fish to EUS. Finally, the third chapter aimed to establish and improve molecular method for detection of EUS in fish.

However, a number of studies have done on EUS in the world to isolate and characterize the etiological agent of EUS, but Malaysian local fish have not previously been experimentally challenged and the potential impact of an introduction of the pathogen into Malaysia on wild and farmed fish populations is unclear. Hence, the general aim of this study is characterization and isolation of *Aphanomyces* spp., and establishing diagnostic technique for detection *A. invadans* to gain insights into the EUS and Malaysian local fish susceptibility to this world wild disease, in order to create a technical pathway for future study on EUS and decrease economical impacts associated with EUS likely onset in Malaysia.

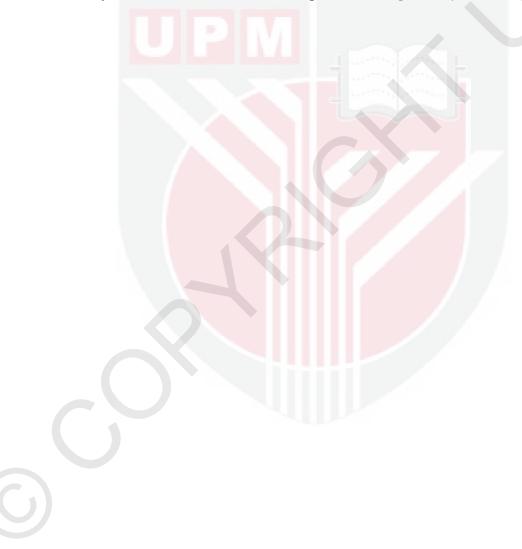
# 1.1 Objectives

Current study was conducted to:

- 1. Isolate, identify and characterize *Aphanomyces* spp. from fish and water in Selangor state, Malaysia.
- 2. Determine pathogenicity of isolated *Aphanomyces* spp. to the most susceptible fish to the EUS (Snakehead, *Channa striata*).
- 3. Assess the virulence of *A. invadans* strain in the most important local fishes of Malaysia (Snakehead, Moonlight gourami, Snakeskin gourami, Koi Carp, Goldfish, Broadhead catfish, Climbing perch and Tilapia).
- 4. Establish a PCR method for rapid and reliable diagnosis of A. invadans.

## **1.2 Hypothesis**

Aquatic fungi infections are common in Malaysian water bodies, and local Malaysian freshwater fishes are susceptible to the *Aphanomyces invadans* infection.



#### REFERENCES

- Abdul Salam, M.N., and Gopinath, N. (2006). Riverine fish and fisheries in Malaysia: An ignored resource. Aquatic Ecosystem Health & Management. 9(2): 159-164.
- Abking, N., Fuangsawat, W., and Lawhavinit, O.A. In Achlya spp. isolated from eggs of the Mekong Giant Catfish (Pangasianodon gigas, Chevey). Proceedings of the 47th Kasetsart University Annual Conference on Veterinary Medicine, Kasetsart, March. 17-20, 2009. Kasetsart University: Kasetsart, 2009.
- Adil, B., Shankar, K.M., Naveen Kumar, B.T., Patil, R., Ballyaya, A., Ramesh, K.S., Byadgi, V., and Siriyappagouder, P. (2013). Development and standardization of a monoclonal antibody-based rapid flow-through immunoassay for the detection of Aphanomyces invadans in the field. Journal of Veterinary Science. 14(4): 413-419.
- Ahilan, B., Felix N., and Jamesson, J.D. (2009). Goldfish. Delhi. India: Daya Publishing House. pp87.
- Ahmed, G.U., and Hoque, M.A. (1999). Mycotic involvement in epizootic ulcerative syndrome of freshwater fishes of Bangladesh: a histopathological study. Asian Fishery Society. 12: 381–390.
- Ahmed, M., and Rab, M.A. (1995). Factors affecting outbreaks of epizootic ulcerative syndrome in farmed and wild fish in Bangladesh. Journal of Fish Diseases. 18: 263-271.
- Alderman, D.J. (1982). In vitro testing of fisheries chemotherapeutants. Journal of Fish Diseases. 5: 112–123.
- Alderman, D.J., and Polglase, J.L. (1986). Aphanomyces astaci: isolation and culture. Journal of Fish Biology Diseases. 9: 367-379.
- Ali, A.B. (1993). Aspects of the fecundity of the feral catfish, Clarias macrocephalus (Gunther), population obtained from the rice fields used for rice-fish farming, in Malaysia. Hydrobiologia. 254(2): 81-89.
- Alias, S.A., and Jones, E.B.G. (2000). Colonization of mangrove wood by marine fungi at Kuala Selangor mangrove stand, Malaysia. Fungal Diversity. 5: 9-21.
- Andrew, T. G., Huchzermeyer, K. D. A., Mbeha, B. C., and Nengu, S. M. (2008). Epizootic ulcerative syndrome affecting fish in the Zambezi river system in southern Africa. Veterinary Record. 163(21): 629-631.
- Arockiaraj, J., Avin, F. A., Vanaraja, P., Easwvaran, S., Singh, A., Othman, R.Y., and Bhassu, S. (2012). Immune role of MrNFκBI-α, an IκB family member characterized in prawn "M. rosenbergii". Fish & Shellfish Immunology. 33(3): 619–625.
- Balasuriya, K.S.W., Kulathilake, M., and Subasinghe, R. In Preliminary investigations into the experimental transmission syndrome in fish in Sri Lanka 662. Proceeding of the 2nd Asian Fisheries Forum, Manila, Philippine, 1990. Hirano, R., and Hanyu, I. (Ed.); Asian Fish Society: Manila, 1990.
- Baldock, F.C., Blazer, V., Callinan, R., Hatai, K., Karunasagar, I., Mohan, C.V., and Bondad-Reantaso, M.G. (2005). In Outcomes of a short expert consultation on epizootic ulcerative syndrome (EUS): Re-examination of causal factors, case definition and nomenclature. Proceeding of the Diseases in Asian Aquaculture V. Walker P., Laster R. & Bondad-Reantaso MG. (Ed.); Manila, Asian Fisheries Society: 555–585.

- Ballesteros, I., Martín, M.P., and Diéguez-Uribeondo, J. (2006). First isolation of Aphanomyces frigidophilus (Saprolegniales) in Europe. Mycotaxon. 95: 335–340.
- Barron, G.L. (2003). Predatory fungi, wood decay, and the carbon cycle. Biodiversity. 4: 3–9.
- Barua, G. (1994). In The status of epizootic ulcerative syndrome of fish of Bangladesh. Proceedings of the ODA Regional Seminar on Epizootic Ulcerative Syndrome, Roberts, Bangkok, Thailand, Jan.1994. R.J., Campbell, B. and Macrae, I.H. (Ed.); Aquatic Animal Health Research Institute: Bangkok, 1994.
- Baruah, A., Saha, R.K., and Kamilya, D. (2012). Inter-Species Transmission of the Epizootic Ulcerative Syndrome (EUS) Pathogen, Aphanomyces invadans, and Associated Physiological Responses. The Israeli Journal of Aquaculture.696: 9.
- Cole, B.E., Tamaru, C.S., Bailey, R., and Brown, C. (1999). A manual for commercial production of the gourami, Trichogaster trichopterus, a temporary paired spawner. Hawaii: University of Hawaii Sea Grant College Program.
- Blazer, V.S., Vogelbein, W.K., Densmore, C.L., May, E.B., Lilley, J.H., and Zwerner, D.E. (1999). Aphanomyces as a cause of ulcerative skin lesions of menhaden from Chesapeake Bay tributaries. Journal of Aquatic Animal Health. 11(4): 340-349.
- Blazer, V.S., Lilley, J.H., Schill, W.B., Kiryu, Y., Densmore, C.L., Panyawachira, V., and Chinabut, S. (2002). Aphanomyces invadans in Atlantic Menhaden along the East Coast of the United States. Journal of Aquatic Animal Health. 14 (1): 1-10
- Bly, J.E., Lawson, L.A, Dale, D.J., Szalai, A.J., Durborow, R.M., and Clem, L.W. (1992). Winter Saprolegniosis in channel catfish. Diseases of Aquatic Organisms. 13: 155-164.
- Bondad-Reantaso, M.G. (2004). Trans-boundary aquatic animal diseases/pathogens; Capacity and Awareness Building on Import Risk Analysis (Ira) for Aquatic Animals: (FWG/01/2002), 9.
- Bondad-Reantaso, M.G., Hatai, K., and Kurata, O. In Aphanomyces from EUSinfected fish in the Philippines and Bangladessh and MG-infected fish from Japan: II. Pathogenicity studies. Proceeding of the Fourth Symposium on Diseases in Asian Aquaculture, Cebu, Philippines, Nov. 22-26, 1999. Fish Health Section, Asian Fisheries Society: Cebu, 1999.
- Boys, C.A., Rowland, S.J., Gabor, M., Gabor, L., Marsh, I.B., Hum, S., and Callinan, R.B. (2012). Emergence of epizootic ulcerative syndrome in native fish of the Murray–Darling river system, Australia: hosts, distribution and possible vectors. PLoS One 7:e35568.
- Bruno, D.W. and Stamps, D.J. (1987). Saprolegniasis of Atlantic salmon, Salmo salar L., fry. Journal of Fish Diseases. 10: 513–517.
- Bruno, D.W., and Wood, B.P. (1999). Saprolegnia and other oomycetes. Mycotaxon. 95: 335–340.
- Bullis, R.A., Noga, E.J., and Levy, M.G. (1990). Immunological relationship of the fish-pathogenic oomycete Saprolegnia parasitica to other oomycetes and unrelated fungi. Journal of Aquatic Animal Health. 2(3): 223-227.
- Buttrell, E.S. (1974). Parasitism of fungi on vascular plants. Mycologia. 66: 1-15.

- Callinan, R.B., and Keep, J.A. (1989). Bacteriology and parasitology of red spot disease in sea mullet, Mugil cephalus L., from eastern Australia. Journal of Fish Diseases. 12(4): 349–356.
- Callinan, R.B., Paclibare, J.O., Bondad-Reantaso, M.G., Chin, J.C., and Gogolewski, R.P. (1995). Aphanomyces species associated with epizootic ulcerative syndrome (EUS) in the Philippines and red spot disease (RSD) in Australia: preliminary comparative studies. Diseases of Aquatic Organisms. 21: 233.
- Cao, H., Zheng, W., Xu, J., Ou, R., He, S., and Yang, X. (2012). Identification of an isolate of Saprolegnia ferax as the causal agent of saprolegniosis of Yellow catfish (Pelteobagrus fulvidraco) eggs. Veterinary research communications. 36(4): 239-244.
- Catap, E. S., and Munday, B. L. (1998). In Effects of variations of water temperature and dietary lipids on the expression of experimental epizootic ulcerative syndrome (EUS) in sand whiting, Sillago ciliata. Proceedings of the International Symposium on Diseases in Marine Aquaculture, Hiroshima, Japan, October. 3-6, 1997. Vol. 33, No. 4, pp. 327-335.
- Catap, E. S., and Munday, B. L. (2002). Development of a method for reproducing epizootic ulcerative syndrome using controlled doses of Aphanomyces invadans in species with different salinity requirements. Aquaculture. 209(1-4): 35–47.
- Chaturvedi, B. (2009). Studies on periodicity and effects of certain physico-chemical Factors on life cycle of an aquatic fungus: Achlya diffusa Harvey ex Johnson. Master of Science Dissertation, DDU Gorakhpur University, Gorakhpur.
- Chinabut, S. (1998). Epizootic ulcerative syndrome: Information up to 1997. Fish Pathology. 33: 321-326.
- Chinabut, S. (1989). Studies on the inflammatory response of the striped snakehead Channa striatus (Fowler). Doctoral Dissertation, University of Stirling, Stirling, UK.
- Chinabut, S., Roberts, R.J., Willoughby, G.R., and Pearson, M.D. (1995). Histopathology of snakehead, Channa striatus (Bloch), experimentally infected with the specific Aphanomyces fungus associated with epizootic ulcerative syndrome (EUS) at different temperatures. Journal of Fish Diseases. 18(1): 41-47.
- Chinabut, S., Roberts, R.J., Willoughby, G.R., and Pearson, M.D. (2006). Histopathology of snakehead, Channa striatus (Bloch), experimentally infected with the specific Aphanomyces fungus associated with epizootic ulcerative syndrome (EUS) at different temperatures. Journal of Fish Diseases. 18(1): 41–47.
- Chong, V.C., Lee, P.K. Y., and Lau, C.M. (2010). Diversity, extinction risk and conservation of Malaysian fishes. Journal of fish biology. 76(9): 2009-2066.
- Choogo K., Hangomba B., Samui K.L., Syachaba M., Phuri H., Maguswi C., Muyangaali k., Bwalya G., and Mataa, L. (2009). Environmental and climatic factors associated with epizootic ulcerative syndrome in fish from the Zambezi floodplains, Zambia. Bulletin of Environmental Contamination and Toxicology. 4: 474-478.

Chukanhom, K., and Hatai, K. (2004). Freshwater fungi isolated from eggs of the common carp (Cyprinus carpio) in Thailand. Mycoscience. 45(1): 42–48.

Clifton- Hadley, R.S., and Alderman, D.J. (1987). The effects of malachite green upon proliferative kidney disease. Journal of Fish Diseases. 10(2): 101-107.

- Coker, W.C., and Matthews, V.D. (1937). Blastocladiales, Monoblepharidales, Saprolegniales. N. Amer. North American Flora. 2(1): 1–76.
- Cruz-Lacierda, E.R., and Shariff, M. In The haematological changes in snakehead (Ophiocephalus striatus) affected by epizootic ulcerative syndrome. Proceeding of the 3rd Asian Fisheries Forum, Manila, Philippine, 1994. Chou, L.M., Munro, A.D., Lam, T.J., and Chen, T.W. (Ed.); Asian Fisheries Society : Manila, 1994.
- Czeczuga, B., and Mazalska, B. (2000). Zoosporic aquatic fungi growing on avian excrements in various types of water bodies. Limnologica-Ecology and Management of Inland Waters. 30(4): 323–330.
- Czeczuga, B., and Muszynska, E. (2004). Aquatic zoosporic fungi from baited spores of cryptogams. Fungal Diversity. 16: 11–22.
- Czeczuga, B., Godlewska, A., Mazalska, B., and MuszyA ska, E. (2010). Straminipilous organisms growing on herbivorous pirapitinga (Piaractus brachypomus) and carnivorous piranha (Pygocentrus nattereri) from Poland. Brazilian Journal of Biology. 70(2): 335-339.
- Czeczuga, B., Kiziewicz B., and Danilkiewicz, Z. (2002). Zoosporic fungi growing on specimens of certain fish species recently introduced to Polish waters. ACTA Ichthyologica Et Piscatoria. 32(2): 117-125.
- Czeczuga, B., Kiziewicz, B., and Godlewska, A. (2003). Zoosporic fungi growing on eggs of Coregonus lavaretus holsatus Thienemann, 1916 from Lake Wdzydze in Kaszuby. Polish Journal of Environmental Studies. 13(4): 355–359.
- Czeczuga, B., Kiziewicz, B., and Godlewska, A. (2004). Zoosporic fungi growing on eggs of Coregonus lavaretus holsatus Thienemann, 1916 from Lake Wdzydze in Kaszuby. Polish Journal of Environmental Studies. 13(4): 355-359.
- Dahail, S.P., Shrestha, M.K., Pradhan S.K., and Jha, D.K. (2008). In Occurrence of epizootic ulcerative syndrome in pond fish of Kapilvastu District of Nepal. Proceeding of Diseases in Asian Aquaculture VI. Manilla, Philippines, 2005. Bondad-Reantaso, M.G., Mohan, C.V., Crumlish, M. and Subasinghe, R.P. (Ed.); Asian Fisheries Society: Manila, 2005. 505 pp.
- Dahal, S.P. (2003). Country report (prepared and submitted in the capacity of 'Focal Point 'for fish disease reporting-Nepal) to Quarterly Aquatic Animal Disease Report (Asia and Pacific region). QAAD Report 2003/1.
- Deng, G., Li, S., Xie, J., Bai, J., Chen, K., Ma, D., Jiang, X., Lao, H., and Yu, L. (2011). Characterization of a ranavirus isolated from cultured largemouth bass (Micropterus salmoides) in China. Aquaculture. 312: 198–204.
- Dick, M.W. (1973). Saprolegniales: The Fungi, an advanced treatise. London: Academic Press.
- Dick, M.W. (1995). Sexual reproduction in the Peronosporomycetes (chromista fungi). Canadian Journal of Botany. 73: 712–724.
- Dick, M.W. (2001). Straminipilous Fungi: systematics of the Peronosporomycetes including accounts of the marine straminipilous protists, the plasmodiophorids and similar organisms. Dordrecht: Kluwer Academic Publishers.
- Diéguez-Uribeondo, J., García, M.A., Cerenius, L., Kozubíková, E., Ballesteros, I., Windels, C., and Weiland, J. (2009). Phylogenetic relationships among plant and animal parasites, and saprotrophs in Aphanomyces (Oomycetes). Fungal Genetics and Biology. 46(5): 365–376.

- Dykstra M.J., Noga EJ., Levine J.F., Moye D.W., and Hawkins J.H. (1986). Characterization of the Aphanomyces species involved with ulcerative mycosis (UM) in menhaden. Mycologia. 78: 664-672.
- Dykstra, M.J., and Kane, A.S. (2000). Pfiesteria piscicida and ulcerative mycosis of Atlantic menhaden—current status of understanding. Journal of Aquatic Animal Health. 12(1): 18-25.
- Dykstra, M. J., Noga, E. J., Levine, J. F., Moye, D. W., and Hawkins, J. H. (1986). Characterization of the Aphanomyces species involved with ulcerative mycosis (UM) in menhaden. Mycologia. 664–672.
- Dykstra, M.J., Levine, J.F., Noga, E.J., Hawkins, J.H., Gerdes, P., Grier, H., and Strake, D. (1989). Ulcerative mycosis: a serious menhaden disease of the southeastern coastal fisheries of the United States. Journal of Fish Diseases. 12(2): 175-178.
- Ebrahimzadeh, M.H, Hoseinifard, S., Khosravi, A., Soltani M., and Yoosefian, M. (2007). Isolation and Identification of Saprophytic fungi from Rainbow trout Infected eggs in farms of Mazandaran Province. Iranian Journal of Veterinary Research. 62(3): 163-168.
- Egusa, S., and Masuda, N. (1971). A new fungal disease of Plecoglossus altivelis. Fish Pathology. 6: 41–46.
- Eli, K.R. (2008). Overview of the OIE's involvement in terms of aquatic animal diseases. Report of a seminar of World Organization for Animal Health (OIE).
- El-Sharouny H.M., Badran R.A.M. (1995). Experimental transmission and pathogenicity of some zoosporic fungi to Tilapia fish. Mycopathologia. 132(2): 95.
- European Food Safety Authority (EFSA). (2011). Scientific Opinion on Epizootic Ulcerative Syndrome. EFSA Journal. 9(10): 2387, 1-58.
- Fadaeifard, F., Raissy, M., Bahrami, H., Rahimi, E., and Najafipoor, A. (2011). Freshwater fungi isolated from eggs and broodstocks with an emphasis on Saprolegnia in rainbow trout farms in west Iran. African Journal of Microbiology Research. 4(22): 3647-3651.
- Fairweather, D.J. (2000). Development of a Bath Challenge System to Study Component Causes, and Preventative Treatments, of Epizootic Ulcerative Syndrome (EUS) in Snakehead Fish (Channa Striata). Doctoral dissertation, University of Plymouth, UK.
- FAO Report of the international emergency disease investigation task force on a serious finfish disease in Southern Africa; Food and Agriculture Organization of the United Nations: Rome, 2009.
- Ferguson, H.W. (1989). Systemic pathology of fish: A text and atlas of comparative tissue responses in diseases of teleosts. Iowa State University Press.
- Firoozbakhsh, F., Ebrahimzade Moosavi, H., and Khosravi, A. (2005). Isolation of Pathogenic and saprophytic fungi from Cyprinid gill lesions. Iranian Journal of Veterinary Research. 60(1): 15-19.
- Fitzsimmons, K., Martinez-Garcia, R., and Gonzalez-Alanis, P. (2011). Why tilapia is becoming the most important food fish on the planet. Better Science, Better Fish, Better Life. 9.
- Fraser, G.C., Callinan, R.B., and Calder, L.M. (1992). Aphanomyces species associated with red spot disease: an ulcerative disease of estuarine fish from eastern Australia. Journal of Fish Diseases. 15(2): 173–181.

- Fregenedaa Grandes, J.M., RodrAgueza Cadenas, F., and Allera Gancedo, J.M. (2007). Fungi isolated from cultured eggs, alevins and broodfish of brown trout in a hatchery affected by saprolegniosis. Journal of Fish Biology. 71(2): 510–518.
- Frerichs, G.N. (1995). Viruses associated with the epizootic ulcerative syndrome (EUS) of fish in South-East Asia. Veterinary Research. 26(5-6): 449–454.
- Gam, L.H., Leow, C.Y., and Baie, S. (2006). Proteomic analysis of snakehead fish (Channa striata) muscle tissue. Malaysian Journal of Biochemistry and Molecular Biology. 14(1): 25-32.
- Gayathri D., Shankar, K.M., and Mohan, C.V. (2004). Monoclonal antibody based immunodot test for epizootic ulcerative syndrome pathogen, Aphanomyces invadans. Current Science. 87: 289-291.
- Gupta, M.V., and Acosta, B.O. (2004). A review of global tilapia farming practices. Aquaculture Asia. 9: 7-12.
- Hanjavanit, C., Hiroki, S., and Hatai, K. (1997). Mycotic granulomatosis found in two species of ornamental fishes imported from Singapore. Mycoscience. 38: 433-436.
- Harikrishnan, R., Balasundaram, C., and Heo, M. (2010). Supplementation Diet Containing Probiotics, Herbal and Azadirachtin on Hematological and Biochemical Changes in Cirrhina mrigala Against "Aphanomyces invadans". Journal of Fisheries and Aquaculture. 4: 1-11.
- Hatai K., Nakamura K., Rha S.A., Yuasa K. and Wada S. (1994). Aphanomyces infection in dwarf gourami (Colisa lalia). Fish Pathology. 29: 95–99.
- Hatai, K. (1980). Studies on the pathogenic agents of Saprolegniasis in fresh water fishes. Special Report of Nagasaki Prefectural Institute of Fisheries. 8: 95 pp.
- Hatai, K., Egusa, S., Takahashi, S., and Ooe, K. (1977). Study on the pathogenic fungus of mycotic granulomatosis: Isolation and pathogenicity of the fungus from cultured Ayu infected with the disease. Fish Pathology. 12:129-133.
- Hawke, J.P., Grooters, A.M., and Camus, A.C. (2003). Ulcerative mycosis caused by Aphanomyces invadans in channel catfish, black bullhead, and bluegill from southeastern Louisiana. Journal of Aquatic Animal Health. 15(2): 120-127.
- Hossain, M.B., Amin, S.M.N., Shamsuddin, M., and Minar, M.H. (2013). Use of Aqua-chemicals in the Hatcheries and Fish Farms of Greater Noakhali, Bangladesh. Asian Journal of Animal and Veterinary Advances. 8(2).
- Hossain, M.F., Rahman, M.M., and Sayed, M.A. (2013). Experimental Infection of Indigenous Climbing Perch Anabas testudineus with Aeromonas hydrophila Bacteria. Progressive Agriculture. 22(1-2): 105-114.
- Hudspeth, D.S.S., Nadler, S.A., Hudspeth, M.E.S. (2000). A COX2 molecular mhylogeny of the Peronosporomycetes. Mycologia. 92: 674–684.
- Hulvey, J.P., Padgett, D.E., and Bailey, J.C. (2007). Species boundaries within Saprolegnia (Saprolegniales, Oomycota) based on morphological and DNA sequence data. Mycologia. 99(3): 421–429.
- Hussein, M.M., Hatai, K., and Nomura, T. (2001). Saprolegniosis in salmonids and their eggs in Japan. Journal of Wildlife Diseases. 37(1): 204–207.
- Iqbal, Z., and Mumtaz, R. (2013). Some fungal pathogens of an ornamental fish, black moor (Carassius auratus L.). European Journal of Veterinary Medicine. 2(1).
- John, K.R., and George, M.R. (2012). Viruses Associated with Epizootic Ulcerative Syndrome: An Update. Indian Journal of Virology. 23(2): 106-113.

- Johnson, R.A., Zabrecky, J., Kiryu, Y., and Shields, J.D. (2004). Infection experiments with Aphanomyces invadans in four species of estuarine fish. Journal of Fish Diseases. 27(5): 287–295.
- Johnson, T.W., Seymour, R.L., Padgett, D.E. (2002). Biology and systematics of the Saprolegniaceae. USA: University of North Carolina at Wilmington, Department of Biological Sciences.
- Johnson, T.W. (1956). The Genus Achlya, Morphology and Taxonomy. USA: University of Michigan Press, Ann Arbor.
- Jones, E.G., and Pang, K.L. (2012). Tropical aquatic fungi. Biodiversity and Conservation. 21(9): 2403-2423.
- Kamilya, D., and Baruah, A. (2013). Epizootic ulcerative syndrome (EUS) in fish: history and current status of understanding. Reviews in Fish Biology and Fisheries. 1-12.
- Kanchanakhan, S. (1996). Epizootic ulcerative syndrome (EUS): a new look at the old story. AAHRI News letter. 5: 2–3.
- Kanchanakhan, S., Chinabut, S., Tonguthai, K. and Richards, R.H. In Epizootic ulcerative syndrome of fishes: rhabdovirus infection and EUS induction experiments in snakehead fish. Proceeding of the Diseases in Asian Aquaculture IV. Fish Health Section, Asian Fisheries Society, Manila, Philippine. Lavilla-Pitogo, C. R. and Cruz-Lacierda, E. R. (Ed.); Asian Fisheries Society: Manila, 2002.
- Khan, M.H., Lilley, J.H., Subasinghe, R. P., Reantaso, M.B., and MacRae, I. H. (2002). Risk factors and socio-economic impacts associated with epizootic ulcerative syndrome (EUS) in Bangladesh. Fao Fisheries Technical Paper. 27–39.
- Khan, M. H., Marshall, L., Thompson, K. D., Campbell, R. E., and Lilley, J. H. (1998). Susceptibility of five fish species (Nile tilapia, rosy barb, rainbow trout, stickleback and roach) to intramuscular injection with the oomycete fish pathogen, Aphanomyces invadans. Bulletin-European Association of Fish Pathologists. 18: 192–197.
- Khan, M.H., Lilley, J.H., Subasinghe, R.P., Reantaso, M.B., and MacRae, I.H. (2002). Risk factors and socio-economic impacts associated with epizootic ulcerative syndrome (EUS) in Bangladesh. FAO Fisheries Technical Paper. 27–39.
- Khan, M.R., Rahman, M.M., Shamsuddin, M., Islam, M.R, and Rahman, M. (2011). Present status of aqua drugs and chemicals in Mymensingh District. Journal of Bangladesh Society Agricultural Sciences Technology. 8: 169-174.
- Kiryu, Y., Shields, J.D., Vogelbein, W.K., Kator, H., and Blazer, V.S. (2003). Infectivity and pathogenicity of the oomycete Aphanomyces invadans in Atlantic menhaden Brevoortia tyrannus. Diseases of Aquatic Organisms. 54(2): 135–146.
- Kiryu, Y., Shields, J.D., Vogelbein, W.K., Zwerner, D.E., Kator, H., and Blazer, V. S. (2002). Induction of skin ulcers in Atlantic menhaden by injection and aqueous exposure to the zoospores of Aphanomyces invadans. Journal of Aquatic Animal Health. 14(1): 11–24.
- Kitancharoen, N., and Hatai, K. (1997). Aphanomyces frigidophilus sp. nov. from eggs of Japanese char, Salvelinus leucomaenis. Mycoscience. 38(2): 135–140.

- Kitancharoen, N., Hatai, K., Ogihara R., and Aye, D.N.N. (1995). A new record of Achlya klebsiana from snakehead, Channa striatus, with fungal infection in Myanmar. Mycoscience. 36: 235-238.
- Kiziewicz, B. (2005). Aquatic fungi growing on seeds of plants in various types of water bodies of Podlasie Province. Polish Journal of Environmental Studies. 14(1): 49–55.
- Kiziewicz, B., and Nalepa, T. F. (2008). Some fungi and water molds in waters of Lake Michigan with emphasis on those associated with the benthic amphipod Diporeia spp. Journal of Great Lakes Research. 34(4): 774-780.
- Kiziewicz, B., and Nalepa, T.F. (2008). Some fungi and water molds in waters of Lake Michigan with emphasis on those associated with the benthic amphipod Diporeia spp. Journal of Great Lakes Research. 34(4): 774-780.
- Kohinoor, A.H.M., Akhteruzzaman, M., Hussain, M.G., Shah, M.S. (1991). Observation on the induced breeding of koi fish (Anabas testudineus (Bloch) in Bangladesh. Bangladesh Journal of Fisheries. 14(1-2): 73-77.
- Kottelat, M., Whitten, A.J., Kartikasari, S.N., and Wirjoatmodjo, S. (1993). Freshwater Fishes of Western (Indonesia and Sulawesi). Hong Kong: Periplus Editions. 221 pp.
- Krishna, L., Gupta, V.K., Katoch, R.C., and Singh, D. (1990). Saprolegniasis in Indian major carps-an investigation. Indian Veterinary Journal. 67: 554–555.
- Kuan, G.C., Sheng, L.P., Rijiravanich, P., Marimuthu, K., Ravichandran, M., Yin, L.S., and Surareungchai, W. (2013). Gold-nanoparticle based electrochemical DNA sensor for the detection of fish pathogen "Aphanomyces invadans". Talanta. 117: 312-317.
- Kumamaru, A. (1973). A fungal disease of fishes in lake Kasumigiura and lake Kitaura; Report Stan: Ibaraki Prefecture, Japan, 11, 129-142.
- Kumaresan, V., Bhatt, P., Palanisamy, R., Gnanam, A.J., Pasupuleti, M., and Arockiaraj, J. (2014). A murrel cysteine protease, cathepsin L: bioinformatics characterization, gene expression and proteolytic activity. Biologia. 69(3): 395-406.
- Kurata, O., Kanai, H., and Hatai, K. (2000). Hemagglutinating and Hemolytic Capacities of Aphanomyces piscicida. Fish Pathology. 35(1): 29–33.
- Kurata, O., Sanpei, K., Hikiji, K., and Hatai, K. (2002). A Galactose-Binding Protein Revealed as a Hemagglutinin in Aphanomyces piscicida. Fish Pathology. 37(1): 1–6.
- Kwanprasert, P., Hanjavanit, C., and Kitancharoen, N. (2007). Characteristics of "Achlya bisexualis" Isolated from Eggs of NileTilapia (Oreochromis niloticus Linn.). Warasan Wichai Mokho Journal. 12(3): 195 202.
- Lawhavinit, O., Chukanhom, K., and Hatai, K. (2002). Effect of Tetrahymena on the occurrence of achlyosis in the guppy Poecilia reticulata. Mycoscience. 43: 27–31.
- Leclerc, M.C., Guillot, J., and Deville, M. (2000). Taxonomic and phylogenetic analysis of Saprolegniaceae (Oomycetes) inferred from LSU rDNA and ITS sequence comparisons. Antonie van Leeuwenhoek. 77: 369–377.
- Lilley, Beakes, G.W., and Hetherington, C.S. (2001). Characterization of Aphanomyces invadans isolates using pyrolysis mass spectrometry (PyMS) Charakterisierung von Aphanomyces invadans und verwandten Arten mittels Pyrolyse massen spektrometrie (PyMS). Mycoses: 44(9-10): 383–389.

- Lilley, J.H., and Roberts, R. J. (1997). Pathogenicity and culture studies comparing the Aphanomyces involved in epizootic ulcerative syndrome (EUS) with other similar fungi. Journal of Fish Diseases. 20(2): 135–144.
- Lilley, J. H., Thompson, K.D., and Adams, A. (1997). Characterization of Aphanomyces invadans by electrophoretic and Western blot analysis. Diseases of Aquatic Organisms. 30: 187–197.
- Lilley, J., Callinan, R.B., Chinabut, S., and Kanchanakhan, S. (1998). Epizootic Ulcerative Syndrome (EUS) technical handbook (p. 88 pp). The Aquatic Animal Health Research Institute, Bangkok.
- Lilley, J., Hart, D., Panyawachira, V., Kanchanakhan, S., Chinabut, S., Soderhall, K., and Cerenius, L. (2003). Molecular characterization of the fish pathogenic fungus "Aphanomyces invadans". Journal of Fish Diseases. 26(5): 263–275.
- Lilley, J.H., Phillips, M.J. and Tonguthai, K. (1992). A Review of Epizootic Ulcerative Syndrome (EUS) in Asia. Aquatic Animal Health Research Institute and Network of Aquaculture Centres in Asia-Pacific, Bangkok, Thailand. 73pp.
- Lilley, JH. In Assaying pond water for spores of saprolegniaceous fungi. Proceedings of the Seminar on Fisheries, Sep. 16-18, 1992. National Inland Fisheries Insistute: Bangkhen, 1992. pp 79-82.
- Lio-Po, G.D., Albright, A., Michel, C., and Leano, EM. (1998). Experimental induction of lesions in snakeheads (Ophicephalus striatus) and catfish (Clarias batrachus) with Aeromonas hydrophila, Aquaspirillum sp., Pseudomonas sp. and Streptococcus sp. Journal of Applied Ichthyology. 14:75–9.
- Lio-Po, G.D., Albright, A., Traxler, G., Leano, E.M. (2001). Pathogenicity of the epizootic ulcerative syndrome (EUS) associated rhabdovirus to snakehead Ophicephalus striatus. Fish Pathology. 36: 57–66.
- Lio-Po, G.D., Albright, L.J., Traxler, G.S., Leano, E.M. (2003). Horizontal transmission of epizootic ulcerative syndrome (EUS)-associated virus in the snakehead Ophicephalus striatus under simulated natural conditions. Diseases of Aquatic Organisms. 57: 213–20.
- Lio-Po, G.D., Traxler, G.S., Albright, L.J., Leano, E.M. (2000). Characterization of a virus obtained from snakeheads Ophicephalus striatus with epizootic ulcerative syndrome (EUS) in the Philippines. Diseases of Aquatic Organisms. 43:191–8.
- Lumanlan-Mayo, SC., Callinan, RB., Paclibare, JO., Catup, ES., Fraser, GC. (1997). Epizootic Ulcerative Syndrome (EUS) in Rice-Fish Culture Systems: an Overview of Field Experiments 1993-1995. Diseases in Asian Aquaculture. 129-138.
- Luna, L.G. (1968). Manual of histologic staining methods of the Armed Forces Institute of Pathology (third.). New York: McGraw-Hill.
- Marcos-Lopez, M., Gale, P., Oidtmann, B.C., and Peeler, E.J. (2010). Assessing the impact of climate change on disease emergence in freshwater fish in the United Kingdom. Transboundary Emerging Diseases. 57: 293–304.
- Marimuthu, K., Arumugam, J., Sandragasan, D., and Jegathambigai, R. (2009). Studies on the fecundity of native fish climbing perch (Anabas testudineus, Bloch) in Malaysia. American-European Journal of Sustainable Agriculture. 3(3): 266-274.
- Miles, D. (2001). Studies on Host Responses to Aphanomyces invadans. Doctoral Dissertation, University of Stirling, UK.

- Miyazaki T., and Egusa, S. (1972). Studies on mycotic granulomatosis in freshwater fishes: Mycotic granulomatosisp revailed in goldfish. Fish Pathology. 7: 15-25.
- Miyazaki, T., and Egusa, S. (1973). Studies on mycotic granulomatosis in freshwater fishes: Mycotic granulomatosisin some wild fishes. Fish Pathology. 8: 44-47.
- Mohamed, A., and Mahmoud, A.M. In Seasonal study, histopathological and treatment trial on saprolegniosis in some fish farms. Proceeding of the 2004 First Scientific Conference of Veterinary Medicine. Moshtohor, Banha, Egypt, 2004. pp. 1-4.
- Mohan, C.V. (2002). Inflammatory response of Indian Major Carps to Aphanomyces invadans, fungal pathogen of EUS. IFS Report. 36.
- MOSTE (1997). Assessment of Biological Diversity in Malaysia; Kuala Lumpur: Ministry of Science, Technology and the Environment, Malaysia.
- Murray, N. (2002). Import Risk Analysis Animal and Animal Products. Ministry of Agriculture and Forestry, Wellington, New Zealand.
- Murray, N., MacDiarmid, S.C., Wooldridge, M., Gummow, B., Morley, R.S., Weber, S.E. Giovannini, A., and Wilson, D. (2004). Handbook on Import Risk Analysis for Animals and Animal Products – Introduction and Qualitative Risk Analysis. Paris: OIE (World Organisation for Animal Health).
- Nabi, N., Jabeen, M., and Hasnain, A. (2000). Recovery of multiple drug resistant pseudomonads associated with an ulcerative condition in an airbreathing murrel, Channa gachua Bl. Asian Fisheries Science. 13: 105–115.
- Naik, M.G., Rajesh, K.M., and Shankar, K.M. (2012). Monoclonal antibody (MAb) based immunodot for early detection of Aphanomyces invadans in fish. International Journal of Science. 1(1): 47-55.
- Nejadsattari, T. (2000). Occurrence and distribution of aquatic Saprolegniaceae in the northwest and south of Tehran. Iranian International Journal of Science. 2: 91-98.
- Newman, D.J., Cragg, G.M., and Snader, K.M. (2003). Natural products as sources of new drugs over the period 1981-2002. Journal of natural products. 66(7): 1022-1037.
- Ng, P.K.L., and Tan, H.H. (1997). Freshwater Fishes of Southeast Asia: potential for the aquarium fish trade and conservation issues. Aquarium Sciences and Conservation. 1(2): 79–90.Noga, E. J. (2010). Fish disease: diagnosis and treatment. John Wiley & Sons.
- Noga, E.J., Khoo, L., Stevens, J.B., Fan, Z., and Burkholder, J.M. (1996). Novel toxic dinoflagellate causes epidemic disease in estuarine fish. Marine Pollution Bulletin. 32: 219–224.
- Oidtmann, B. (2011). Review of Biological Factors Relevant to Import Risk Assessments for Epizootic Ulcerative Syndrome (Aphanomyces invadans). Transboundary and Emerging Diseases. 59(1): 26–39.
- Oidtmann, B., Bausewein, S., Holzle, L., Hoffmann, R., and Wittenbrink, M. (2002). Identification of the crayfish plague fungus Aphanomyces astaci by polymerase chain reaction and restriction enzyme analysis. Veterinary Microbiology. 85: 183–194.
- Oidtmann, B., Schaefers, N., Cerenius, L., Soderhall, K., and Hoffmann, R. W. (2004). Detection of genomic DNA of the crayfish plague fungus Aphanomyces astaci (Oomycete) in clinical samples by PCR. Veterinary microbiology. 100(3): 269–282.

- Oidtmann, B., Steinbauer, P., Geiger, S., and Hoffmann, R. W. (2008). Experimental infection and detection of Aphanomyces invadans in European catfish, rainbow trout and European eel. Diseases of Aquatic Organisms. 82(3): 195–207.
- OIE. (2013). Infection with Aphanomyces invadans (Epizootic Ulcerative Syndrome). OIE Aquatic Animal Health Standards Commission. www.oie.int/international-standard-setting/aquatic-manual/access-online.
- OIE. (2010). Chapter 1 .2. Criteria for Listing Aquatic Animal Diseases Article 1.2.1,http://www.oie.int/fileadmin/Home/eng/Health\_standards/aahc/2010/ch apitre\_1.1.2.pdf.
- Otaye, D.O. (2005). Repeated Emergence, Motility, and Autonomous Dispersal by Sporangial and Cyst Derived Zoospores of Phytophthora. Doctoral dissertation, Oklahoma State University, USA.
- Othman, S.H., Hong, Y.B., and Salam, M.N. A. (2002). A Rapid Assessment of the Economic Benefits and Stakeholder Analysis of Fisheries Resources in the Selangor River Basin; Kuala Selangor: Forests for Water, Water for Life (FWWL) Project 409/98 (4), Malaysia.
- Pal, J., Pradhan, K. (1990). Bacterial involvement in ulcerative condition of airbreathing fish from India. Journal of Fish Biology. 36: 833–9.
- Panyawachira, V., Lilley, J.H., Hart, D. and Kanchanakhan, S. (2000). A PCR-based technique for the identification of "Aphanomyces invadans". AAHRI news letter. 22-26.
- Panyawachira, V., Lilley, J.H., Hart, D., and Kanchana Khan S. In A PCR-based technique for the identification of Aphanomyces invadans. Proceeding of the Fourth Symposium on Diseases in Asian Aquaculture, Cebu, Phillipines, Nov. 22-26, 1999. Asian Fisheries Society: Cebu, 1999.
- Peduzzi, P. and Bizzozero, S. (1977). Immunological investigation of four Saprolegnia species with parasitic activity in fish: serological and kinetic characterization of a chymotrypsin-like activity. Microbe Ecology. 3: 107-118.
- Petersen, A.B., Olson, L.W., and Rosendahl, S. (1996). Use of polyclonal antibodies to detect oospores of "Aphanomyces". Mycological Research. 100(4): 495-499.
- Petersen, A.B., Rosendhal, S. (2000). Phylogeny of the Peronosporomycetes (Oomycota) based on partial sequences of the large ribosomal subunit (LSU rDNA). Mycol. Res. 104: 1295–1303.
- Phadee, P., Kurata, O., Hatai, K., Hirono, I., and Aoki, T. (2004). Detection and identification of fish-pathogenic Aphanomyces piscicida using polymerase chain reaction (PCR) with species-specific primers. Journal of Aquatic Animal Health. 16(4): 220–230.
- Post, G. (1983). Textbook of Fish Health. USA: TFH Publications, 288 pp.
- Prabhuji, S. K. (2010). Sexual reproduction in water moulds-1: General aspects related to family saprolegniaceae. International Journal of Plant Reproductive Biology. 2: 17–30.
- Prabhuji, S.K. (2005). Occurrence and Phenology of the Oomycetes with special reference to Saprolegniaceae. Frontiers in Plant Sciences. 129–142.
- Pradhan, P. K., Mohan, C. V, Shankar, K. M., and Kumar, B. M. (2007). Sequential inflammatory response of fingerlings of Indian major carps to Aphanomyces invadans. Indian Journal of Fisheries. 54(4): 389–396.

- Pradhan, P.K., Mohan, C.V., Shankar, K.M. and Mohana Kumar, B. In Infection experiments with Aphanomyces invadans in advanced fingerlings of four different carp species. Proceeding of the Diseases in Asian Aquaculture VI, Fish Health Section, Asian Fisheries Society, Manila, Philippines, 2008. Bondad-Reantaso, M.G., Mohan, C.V., Crumlish, M. and Subasinghe, R.P. (Ed.); Asian Fisheries Society: Manila, 2008.
- Ramaiah, N. (2006). A review on fungal diseases of algae, marine fishes, shrimps and corals. Indian Journal of Marine Sciences. 35(4): 380-387.
- Report of The Meeting of The OIE Ad Hoc Group on the OIE List of Aquatic Animal Diseases (Finfish); OIE ad hoc Group on the OIE List of Aquatic Animal Diseases, September 2012, (p. 20).
- Riethmüller, A., Voglmayr, H., Göker, M., Weib, M., Oberwinkler, F. (2002). Phylogenetic relationships of the downy mildews (Peronosporales) and related groups based on nuclear large subunit ribosomal DNA sequences. Mycologia. 94: 834–849.
- Riethmüller, A., Weib, M., Oberwinkler, F. (1999). Phylogenetic studies of Saprolegniomycetidae and related groups based on nuclear large subunit of ribosomal DNA sequences. Canadian Journal Botany. 77: 1790–1800.
- Roberts R.J. and Bullock A. M. (1981). Recent observations on the pathological effect of ultraviolet light on fish skin. Fish Pathology. 15: 237-239.
- Roberts, R., Willoughby, L., and Chinabut, S. (2006). Mycotic aspects of epizootic ulcerative syndrome (EUS) of Asian fishes. Journal of Fish Diseases. 16(3): 169-183.
- Roberts, R.J. (2012). Fish pathology. Wiley-Blackwell.
- Roberts, R.J., Frerichs, G.N., Tonguthai, K., and Chinabut, S. (1994). Epizootic ulcerative syndrome of farmed and wild fishes. Recent advances in aquaculture. 5.
- Roberts, R.J., Willoughby, L.G., Chiabut, S. (1993). Mycotic aspects of epizootic ulcerative syndrome (EUS) of Asian fishes. Journal of Fish Diseases. 16:169–183
- Roberts, T.R. (1989). The Freshwater Fishes of Western Borneo (Kalimantan Barat, Indonesia). San Francisco, California Academy of Sciences. 210 pp.
- Royo, F., Andersson, G., Bangyeekhun, E., Muzquiz, J. L., Soderhall, K., and Cerenius, L. (2004). Physiological and genetic characterisation of some new Aphanomyces strains isolated from freshwater crayfish. Veterinary microbiology. 104(1-2): 103–112.
- Sa, T.T., and Boon, Y.H. (2010). 19.2 Malaysia–Introduction (Encyclopedia of the World's Coastal Landforms). USA: Springer.
- Safar, H.M., binti Sarji, A., and Gunaratne, S.A. (2000). 1. NATIONAL PROFILE 1.1 Geography Malaysia is situated in Southeast Asia, bordering Thailand, Singapore, Indonesia, and the Philippines. It is a federation of 13 states, straddling the South-China Sea. Eleven of the states are in the peninsula, with Thailand to the north and Singapore. Handbook of the media in Asia. 317.
- Saha, D., and Pal, J. (2002). In vitro antibiotic susceptibility of bacteria isolated from EUS- affected fishes in India. Letters in applied microbiology. 34(5): 311-316.
- Salam, A., and Gopinath, N. (2006). Riverine fish and fisheries in Malaysia: An ignored resource. Aquatic ecosystem Health & Management. 9(2): 159-164.

- Sammut, J., White, I., Melville, D. (1996). Acidification of an estuarine tributary in eastern Australia due to drainage of acid sulfate soils. Marine and Freshwater Research. 47 (5): 669-684.
- Sarkar, U.K., Deepak, P.K., Kapoor, D., Negi, R.S., Paul, S.K., and Singh, S. (2005). Captive breeding of climbing perch Anabas testudineus (Bloch, 1792) with Wova- FH for conservation and aquaculture. Aquaculture Research. 36(10): 941-945.
- Saylor, R. K., Miller, D. L., Vandersea, M. W., Bevelhimer, M. S., Schofield, P. J., and Bennett, W. A. (2010). Epizootic ulcerative syndrome caused by Aphanomyces invadans in captive bullseye snakehead Channa marulius collected from south Florida, USA. Diseases of Aquatic Organisms. 88(2): 169–175.
- Schneider, C.L., and Yoder, D.L. (1973). Development of a methodology for the production of Aphanomyces cochlioides oöspores in vitro. Journal of American Society. Sugar Beet Technologists. 17: 230-239.
- Scott, W.W. (1961). A monograph of the genus Aphanomyces. Technical Bulletin. Virginia Agricultural Experiment Station. 151.
- Seymour, R.L. (1970). The genus Saprolegnia. Nova Hedwigia. 19: 1–14.
- Shafer, T.H., Padgett, D.E. and Celio, D.A. (1990). Evidence for enhanced salinity tolerance of a suspected fungal pathogen of Atlantic menhaden (Brevoortia tyrannus Latrobe). Journal of Fish Diseases. 13: 335-344.
- Shanor, L., and Saslow, B. (1944). Aphanomyces a fish parasite. Mycologia. 36: 413-415.
- Shariff, M., Soon, S., Lee, K.L., and Tan, L.T. (2000). Practical problems with PCR detection in Asia: the importance of standardization. DNA-Based Molecular Diagnostic Techniques: Research Needs for Standardisation and Validation of the Detection of Aquatic Animal Pathogens and Diseases. 45-51.
- Sharifpour, I. (1997). Histology of the inflammatory response of carp (Cyprinus carpio L.) to various stimuli. Doctoral dissertation, University of Stirling, UK.
- Sharma, P., and Sihag R.C. (2013).Pathogenicity Test of Baterial and Fungal Fish Pathogens in Cirrihinus mrigala Infected with EUS disease. Pakistanian Journal of Biological Sciences. 16 (20): 1204-1207.
- Shrestha, G.B. In Status of epizootic ulcerative syndrome (EUS) and its effects on aquaculture in Nepal. Proceedings of the ODA Regional Seminar on Epizootic Ulcerative Syndrome, Jan. 25-27, 1994. R.J. Roberts, B. Campbell and I.H. MacRae (Eds.); Aquatic Animal Health Research Institute: Bangkok, 1994.
- Singh, A. (2009). Studies on periodicity and effects of certain physico-chemical Factors on life cycle of an aquatic fungus: Allomyces recurvus Prabhuji and Sinha. Master of Science Dissertation, DDU Gorakhpur University, Gorakhpur.
- Singhal, R.N., Jeet, S. and Davies, R.W. (1987). Experimental transmission of Saprolegnia and Achlya to fish. Aquaculture. 64: 1–7.
- Smith, D., and Onions, A.H.S. (1994). The Preservation and Maintenance of Living Fungi. International Mycological Institute. 122pp.
- Smith, D., and Ryan, M.J. (2008). The impact of OECD best practice on the validation of cryopreservation techniques for microorganisms. Cryoletters. 29: 63–72.

- Songe, M.M., Hang'ombe, M.B., Phiri, H., Mwase, M., Choongo, K., Vander Waal, B., and Kanchanakhan, S. (2012). Field observations of fish species susceptible to epizootic ulcerative syndrome in the Zambezi River basin in Sesheke District of Zambia. Tropical Animal Health and Production. 1–5.
- Sosa, E.R., Landsberg, J.H., Stephenson, C.M., Forstchen, A.B., Vandersea, M.W., and Litaker, R.W. (2007). Aphanomyces invadans and ulcerative mycosis in estuarine and freshwater fish in Florida. Journal of Aquatic Animal Health. 19(1): 14–26.
- Sparrow, F.D. (1960). Aquatic Phycomycetes. University of Michigan Press: Ann Arbor.
- Srivastava, R.C. (1980). Fungal parasites of certain freshwater fish of India. Aquaculture. 21:387–392.
- Steciow, M.M. (2001). Achlya fuegiana, a new species from Tierra Del Fuego province (Argentina). Mycologia. 93(6): 1195-1199.
- Stevens, R.B. (1974). Mycology Guidebook. Stirling: University of Washington Press.
- Stueland, S., Hatai, K., and Skaar, I. (2005). Morphological and physiological characteristics of Saprolegnia spp. strains pathogenic to Atlantic salmon, Salmo salar L. Journal of Fish Diseases. 28(8): 445–453.
- Takuma, D., Sano, A., and Hatai, K. (2013). Two new species, Aphanomyces izumoensis sp. nov. and Aphanomyces shimanensis sp. nov. isolated from Ice Fish Salangichthys microdon. International Journal of Research in Pure and Applied Microbiology. 3(3): 67-76.
- Takuma, D., Sano, A., Wada, S., Kurata, O., and Hatai, K. (2010). A new species, Aphanomyces salsuginosus sp. nov., isolated from ice fish Salangichthys microdon. Mycoscience. 51(6): 432–442.
- Tang, Y.W., Procop, G.W., and Persing, D.H. (1997). Molecular diagnostics of infectious diseases. Clinical chemistry. 43(11): 2021–2038.
- Tay, Y.L., Loong, A.M., Hiong, K.C., Lee, S.J.J., Tng, Y.Y.M., Wee, N.L.J., Lee, S.M.L., Wang, W.P., Chew, S.F., Wilson, J.M., Ip, Y.K. (2006). Active ammonia transport and excretory nitrogen metabolism in the climbing perch, Anabas testudineus, during 4 days of emersion or 10 minutes of forced exercise on land. Journal of Experimental Biology. 209: 4475-4489.
- Teoh, C.Y., Turchini, G.M., and Ng, W.K. (2011). Erratum to "Genetically improved farmed Nile tilapia and red hybrid tilapia showed differences in fatty acid metabolism when fed diets with added fish oil or a vegetable oil blend". Aquaculture. 316(1): 144-154.
- Teugels, G.G., Diego, R.C., Pouyaud L., and Legendre, M. (1999). Redescription of Clarias macrocephalus (Siluriformes: Clariidae) from Southeast Asia. Cybium. 23(3): 285-295.
- Thompson, K.D., Lilley, J.H., Chen, S.C., Adams, A., and Richards, R.H. (1999). The immune response of rainbow trout (Oncorhynchus mykiss) against Aphanomyces invadans. Fish and Shellfish Immunology. 9:195-201.
- Thompson, K.D., Lilley, J.H., Chinabut, S., and Adams, A., (1997). The antibody response of snakehead Channa striata Bloch to Aphanomyces invaderis. Fish Shellfish Immunol. 7: 349–353.
- Tonguthai, K. (1985). A preliminary account of ulcerative fish diseases in the-Indo-Pacific region (a comprehensive study based on Thai experiences). National Inland Fisheries Institute, Bangkok, Thailand. 39 p.

- Torres, J.L., Tajima, K., and Shariff, M. (1993). Numerical taxonomy and virulence screening of Aeromonas spp. isolated from healthy and ulcerative disease syndrome-positive fishes. Asian Fisheries Science. 6: 11–12.
- Vandersea, M.W., Litaker, R.W., Yonnish, B., Sosa, E., Landsberg, J.H., Pullinger, C., Moon-Butzin, P. (2006). Molecular assays for detecting Aphanomyces invadans in ulcerative mycotic fish lesions. Applied and environmental microbiology. 72(2): 1551.
- Vogelbein, W. K., Shields, J. D., Haas, L. W., Reece, K. S., and Zwerner, D. E. (2001). Skin ulcers in estuarine fishes: a comparative pathological evaluation of wild and laboratory-exposed fish. Environmental Health Perspectives. 109(5): 687.
- Wada, S., Rha, S., Kondo, T., Suda, H., Hatai, K., and Ishi, H. (1996). Histopathology of Ayu and Carp artificially infected with Aphanomyces piscicida. Fish Pathology. 31.2: 71–80.
- Wada, S., Yuasa, K., Rha, S., Nakamura, K., and Hatai, K. (1994). Histopathology of Aphanomyces infection in dwarf gourami "Colisa lalia". Fish Pathology. 29: 229–237.
- Whittington, R.J., and Chong, R. (2007). Global trade in ornamental fish from an Australian perspective: The case for revised import risk analysis and management strategies. Preventive Veterinary Medicine. 81: 92–116.
- Willoughby, L. G. (1994). Fungi and fish diseases. Pisces Press.
- Willoughby, L.G. (1986). An ecological study of water as the medium for growth and reproduction of the Saprolegnia from salmonid fish. Transactions of the British Mycological Society: 87: 493–502.
- Willoughby, L.G., Pickering, A.D., and Johnson, H.G. (1984). Polycell-gel assay of water for spores of Saprolegniaceae (fungi), especially those of the Saprolegnia pathogen of fish. Hydrobiologia. 114: 237–248.
- Willoughby, L.G., Roberts, R.J., and Chinabut, S. (1995). Aphanomyces invaderis sp. nov., the fungal pathogen of freshwater tropical fish affected by epizootic ulcerative syndrome. Journal of Fish Diseases. 18: 273-275.
- Wolf, F.T. (1944). The aquatic oomycetes of Wisconsin. The University of Wisconsin press.
- Wong, S.L. (2004). Matang Mangroves: A Century of Sustainable Management; Petaling Jaya: Sasyaz Holdings Private Ltd, Malaysia.
- Wood, S.E., and Willoughby, L.G. (1986). Ecological observation on the fungal colonization of fish by Saprolegniaceae in Windermere. Journal of Applied Ecology. 23: 737–749.
- Yahya, M.A., and Singh, H.R. In An assessment of the distribution of the freshwater fishes of the Taman Negara Pahang, Malaysia. Proceeding of the Symposium on Business, Engineering and Industrial Applications (ISBEIA), Bandung, Indonesia, Sept. 23-26, 2012. Institute of Electrical and Electronics Engineers (IEEE): Bandung, 2012.
- Yu Abit, L., Kamaruddin, I.S., Mohd-Rozhan, Z., Ina-Salwany, M.Y., and Mustafa-Kamal, A.S. (2012). Fish Biodiversity Survey (2009) of Streams in the Ayer Hitam Forest Reserve, Puchong, and Selangor. Pertanika Journal of Tropical Agricultural Sciences. 35(1): 15-19.
- Yusoff, F.M. and Gopinath, N. (1995). The status of inland fisheries in Malaysia. Indo-Pacific Fishery Commission. FAO Fisheries Report. 512.
- Zgozi, S.W. (2007). The distribution and abundance of fish and macro invertebrate communities in relation to environmental factors in Klang Strait (Malaysia).

Doctoral dissertation, Institute of Postgraduate Studies and Research, University of Malaya, Kuala Lumpur, Malaysia.

