



UNIVERSITI PUTRA MALAYSIA

***CHARACTERIZATION OF RECENTLY ISOLATED NEWCASTLE
DISEASE VIRUSES AND DEVELOPMENT OF INACTIVATED
VACCINE USING GENOTYPE VII NEWCASTLE DISEASE VIRUS***

ODAY ABDUL RAZZAQ ABDUL WAHHAB ALJUMAILI

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By

ODAY ABDUL RAZZAQ ABDUL WAHHAB ALJUMAILI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the
Degree of Doctor of Philosophy**

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DEDICATION

The sake of Allah, my Creator and my Master,

My great teacher and messenger, Mohammed (May Allah bless and grant him), who taught us the purpose of life.

Am dedicating this thesis to two beloved people who have meant and continue to mean so much to me. Although they are no longer of this world, their memories continue to regulate my life.

First and foremost, to my Father whose love for me knew no bounds and, who taught me the value of hard work. Thank you so much "Haj Abdulrazzaq", I will never forget you.

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Amen.

To my mother, who continues praying to me, supporting and developing and who has been a source of encouragement and inspiration to me throughout my life, very special thanks for her.

My dearest wife, who leads me through the valley of darkness with light of hope and support,

My beloved sisters;

My beloved kids: Mostafa, Dhuha, and Ghena whom I can't force myself to stop loving. To all my family, the symbol of love and giving,

My friends who encourage and support me,

All the people in my life who touch my heart,

I dedicate this research to you .

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

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October 2017

Chairman : Professor Aini bt Ideris, PhD
Faculty : Veterinary Medicine

Newcastle Disease (ND) is a highly contagious and economically devastating disease of poultry in many parts of the world. At present, limited molecular epidemiological data are available regarding the causes of ND outbreaks in vaccinated chickens in commercial poultry farms. Knowing the genomic characteristics of Newcastle disease virus (NDV) infecting commercial poultry operations despite the chickens were vaccinated may give important insights on the infection dynamics of these viruses. In addition, molecular analyses at the subgenotype level and studies on the relationship of Malaysian NDVs with other isolates from around the world are lacking. Although many countries, including Malaysia maintain a stringent vaccination policy against ND, there are indications that ND outbreaks can still occur despite intensive vaccination. Virulent genotype VII NDV from China, Indonesia, Korea and Malaysia share only 82 to 87% similarity in amino acid residues of F and HN antigens respectively with B1 and LaSota vaccine strains (genotype II). While these genotype II-based vaccines prevent disease, they cannot stop viral shedding in the environment. Hence, the need for the so-called genotype-matched vaccines, which have been shown to reduce virus shedding, compared to genotype-mismatched vaccines is highly anticipated. Therefore, in the present study, a molecular epidemiological investigation is conducted to characterize six NDVs isolated from vaccinated commercial poultry flocks. To better understand the epidemiology of Newcastle disease outbreak, a partial F gene and HN gene were amplified from UPM-IBS 046/2014, UPM-IBS 060/2014, UPM-IBS 061/2014, UPM-IBS 074/2014, UPM-IBS 160/2015, and UPM-IBS 162A/2015 isolates by using conventional one step reverse

transcription-polymerase chain reaction (RT-PCR) and then conducted sequence and phylogenetic analysis. Furthermore, inactivation of NDV IBS/025/2013 strain (naturally recombinant strain) by two different method i.e. Ultraviolet type C (UVC) and Binary ethylinimine (BEI), and tested the inactivation by inoculation of the inactivated virus in SPF eggs for two passages was successfully executed.

Six NDV isolates which were recovered from ND outbreaks in chicken flocks in Malaysia were genotypically characterized. All the isolates had close phylogenetic relationship with previously characterized isolates from Malaysia as well as different countries within genotype VIIa, and genetically on the basis of the fusion (F) protein cleavage site. Among these, six NDV isolates showed an F protein cleavage site motif ¹¹²RRQKRF¹¹⁷, ¹¹²KRRKRF¹¹⁷, ¹¹²KRRKRF¹¹⁷, ¹¹²KRRKRF¹¹⁷, ¹¹²KRRKRF¹¹⁷, and ¹¹²KRRKRF¹¹⁷.

In the present study, NDV IBS/025/2013 strain (a naturally recombinant virus strain) was successfully inactivated by UVC light+Riboflavin and chemical (BEI), and then passaged consecutively two times in SPF chicken embryonated eggs. Results from the virus inactivation study revealed that exposure to UVC light for 14 hours or treatment of the virus with BEI for 21 hours at 37°C successfully inactivated the virus as evidenced by its inability to kill SPF chicken embryonated eggs 6 days post-inoculation in two consecutive passages. Both inactivated vaccines were emulsified in two different adjuvants, *Nigella sativa* oil adjuvant, and Freund's incomplete adjuvant to produce vaccines in the form of water in oil (W/O). A stable W/O vaccine with two different adjuvants was successfully formulated. To test the efficacy of each vaccine formulation, ten days old SPF chickens were randomly divided into 11 groups, with 11 chickens in each group. The first 5 groups were vaccinated with different formulations of the vaccine, but the remaining groups were vaccinated with LaSota live vaccine (+L) in different formulations. One group was kept as control. The result for in vivo experiment indicated that the BEI-black seed oil (BEI-BSO), BEI- Incomplete Freund's adjuvant (BEI-IFA) as well as commercial vaccine, were fully protective against a virulent NDV challenge. Although all vaccinated groups had significantly lower mortality rate than unvaccinated-challenged group, full protection from death was observed in birds of group BEI-BSO, BEI-IFA, Commercial, UVC-BSO+L, UVC-IFA+L, BEI-BSO+L, BEI-IFA+L, and Commercial +L followed by group UVC-IFA with 33% mortality. Among vaccinated groups, the best results with regards to clinical signs and gross lesions were obtained in groups that were vaccinated with killed and live vaccine together, and groups of killed vaccine especially commercial and BEI-IFA; followed by group BEI-BSO with 10% morbidity. Vaccination provided high HI antibody titers in most of the vaccinated groups excluding UVC-BSO

and UVC-IFA. The chickens in all vaccinated-challenged groups shed the challenge virus from day 3 days post challenge. Insignificant differences were observed in the frequencies of virus detection among vaccinated groups (UVC-BSO and UVC-IFA) and positive control with different incidence. Duration of virulent virus shedding in infected birds of vaccinated groups were different. Vaccination programs used in groups of killed and live vaccine have shortened the duration of virus shedding (only at 3 days post challenge), while control group and UVC-BSO group started from day 3 post challenge continued to shed the virulent virus in the feces until the end of the experiment. Meanwhile the UVC-IFA started to shed the virus from day 5 post challenge until the end of the experiment. Finally, the BEI-BSO, BEI-IFA and commercial groups started to shed the virus from day 7 post challenge until the end of the experiment with reduced titer at 10days post challenge. The measurement of potency for the inactivated vaccine BEI-BSO and BEI-IFA by the use of the mortality as the metric, resulted in $10^{-7.612}$ of the full dose for BEI-BSO while $10^{-7.532}$ was for BEI-IFA.

In conclusion, the etiologic agents of the ND outbreaks recently reported in vaccinated chickens in Malaysia were found belonging to the velogenic genotype VIIa strain. The exposure of the NDV to UVC light + riboflavin for 14 hours or the exposure of the virus to BEI treatment for 21 hours at 37°C was found to be adequate for the complete inactivation of the virus as demonstrated by its failure to induce mortality in the SPF chicken embryonated eggs, 6 days post inoculation in two successive passages. The preparation of stable water in oil emulsion from both black seed oil and incomplete Freund's adjuvant was successfully achieved in this study. An inactivated ND oil-emulsified vaccine from NDV IBS/025/13 high pathogenic viruses provides protection in young chickens against NDV IBS 002/11 genotype VII virus isolate. The BEI-black seed oil and BEI- Freund's adjuvant as well as commercial vaccine were demonstrated in this study to be capable of offering full protection against virulent NDV challenge.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENCIRIAN VIRUS PENYAKIT NEWCASTLE YANG DIPENCILKAN
BARU-BARU INI DAN PEMBANGUNAN VAKSIN TIDAK AKTIF
MENGUNAKAN VIRUS PENYAKIT NEWCASTLE GENOTIP VII**

Oleh

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Penyakit Newcastle (ND) ialah sejenis penyakit yang amat mudah berjangkit dan mengakibatkan kemusnahan dari segi ekonomi di kebanyakan bahagian di dunia ini. Pada masa kini, data epidemiologi molekular berkenaan penyebab wabak ND di ladang ternakan ayam komersil yang telah divaksinasi adalah terhad. Pengetahuan berkenaan ciri-ciri genomik virus penyakit Newcastle (NDV) yang menjangkiti operasi ternakan ayam komersil meskipun selepas vaksinasi mampu memberikan tanggapan yang penting berkenaan dinamik jangkitan virus ini. Tambahan pula, analisis molekular pada tahap subgenotip dan kajian berkenaan hubungan NDV di Malaysia dengan penciran lain di seluruh dunia adalah terhad. Walaupun kebanyakan negara-negara, termasuk Malaysia mengekalkan polisi vaksinasi yang ketat terhadap ND, terdapat tanda-tanda bahawa wabak ND masih boleh berlaku sungguhpun vaksinasi intensif telah dijalankan. NDV genotip VII virulen dari Cina, Indonesia, Korea, dan Malaysia berkongsi hanya 82–87% persamaan dalam residu asid amino pada antigen-antigen F dan HN masing-masing dengan strain vaksin B1 dan LaSota (genotip II). Walaupun vaksin berasaskan genotip II ini mampu menghalang penyakit, ia tidak berupaya menghalang peluruhan virus ke persekitaran. Oleh yang demikian, keperluan bagi vaksin berpadanan-genotip, yang telah berjaya menunjukkan pengurangan peluruhan virus, berbanding vaksin tak berpadanan-genotip adalah sangat diharapkan. Oleh itu, di dalam kajian ini, satu penyelidikan epidemiologi molekular dijalankan bagi mencirikan enam NDV yang telah dipencilkan dari kumpulan ternakan ayam komersil yang divaksinasi. Bagi lebih memahami epidemiologi wabak penyakit Newcastle, gen separa F dan gen HN telah

diampifikasi daripada pencilan-pencilan UPM-IBS 046/2014, UPM-IBS 060/2014, UPM-IBS 061/2014, UPM-IBS 074/2014, UPM-IBS 160/2015, dan UPM-IBS 162A/2015 dengan menggunakan kaedah lazim satu langkah tindakbalas rantaian polimerase-transkripsi berbalik (RT-PCR), dan kemudian pengendalian analisis jujukan dan filogenetik. Selanjutnya, penyahaktifan strain IBS 025 (strain rekombinan semulajadi) melalui dua kaedah berbeza iaitu UVC dan BEI, dan pengujian penyahaktifan dengan menginokulasi virus tidak aktif itu ke dalam telur SPF sebanyak dua laluan telah berjaya dilakukan.

Enam pencilan NDV yang telah didapati dari wabak ND pada kumpulan ayam di Malaysia telah dicirikan secara genotip. Kesemua pencilan tersebut mempunyai hubungan filogenetik yang rapat dengan pencilan yang telah dicirikan terdahulu di samping negara-negara berbeza dalam genotip VIIa, dan secara genetik pada asas tapak belahan protein lakuran (F). Di antara kesemuanya, enam pencilan-pencilan NDV menunjukkan motif tapak belahan protein F ¹¹²RRQKRF¹¹⁷, ¹¹²KRRKRF¹¹⁷, ¹¹²KRRKRF¹¹⁷, ¹¹²KRRKRF¹¹⁷, ¹¹²KRRKRF¹¹⁷, dan ¹¹²KRRKRF¹¹⁷ masing-masing.

Selanjutnya, dalam kajian ini, strain IBS 025/13 (strain rekombinan semulajadi) telah berjaya dinyahaktifkan dengan lampu UVC+Riboflavin dan bahan kimia (BEI), dan telah dilakukan berturutan sebanyak dua kali dalam telur ayam berembrio SPF. Keputusan dari kajian ketidakaktifan virus menunjukkan bahawa pendedahan kepada lampu UVC selama 14 jam atau rawatan virus dengan BEI selama 21 jam pada 37°C telah berjaya menyahaktifkan virus tersebut sebagaimana dibuktikan dengan ketidakupayaannya untuk mematikan telur ayam berembrio SPF pada 6 hari pos-inokulasi dalam dua laluan berturutan. Kedua-dua vaksin tidak aktif tersebut telah diemulsifikasikan dalam dua adjuvan berbeza, adjuvan minyak *Nigella sativa*, dan adjuvan tak lengkap Freund's bagi menghasilkan vaksin dalam bentuk air dalam minyak (W/O). Tambahan lagi, satu vaksin W/O yang stabil dengan dua adjuvan berbeza telah berjaya diformulasi. Bagi menguji keberkesanan setiap formulasi vaksin, ayam SPF berumur sepuluh hari telah dibahagikan secara rawak kepada 11 kumpulan, dengan 11 ekor ayam dalam setiap kumpulan. Lima kumpulan pertama telah divaksinasi dengan formulasi vaksin yang berbeza, tetapi kumpulan selanjutnya telah divaksinasi dengan vaksin hidup LaSota (+L) dalam formulasi berbeza. Satu kumpulan dikekalkan sebagai kawalan. Keputusan bagi eksperimen in vivo menunjukkan bahawa minyak BEI-bijian hitam (BEI-BSO), BEI-adjuvan tak lengkap Freund's (BEI-IFA) di samping vaksin komersil, adalah melindungi sepenuhnya daripada cabaran NDV virulen. Walaupun kesemua kumpulan divaksinasi mempunyai kadar kematian lebih rendah yang ketara berbanding kumpulan cabaran-tak divaksinasi, perlindungan penuh daripada kematian

telah diperhatikan pada ayam dalam kumpulan BEI-BSO, BEI-IFA, komersil, UVC-BSO+L, UVC-IFA+L, BEI-BSO+L, BEI-IFA+L, dan komersil +L diikuti dengan kumpulan UVC-IFA dengan 33% kematian. Di kalangan kumpulan divaksinasi, keputusan paling baik berkenaan tanda-tanda klinikal dan lesi kasar didapati dari kumpulan yang divaksinasi dengan vaksin terbunuh dan hidup bersama-sama, dan kumpulan vaksin terbunuh terutamanya komersil dan BEI-IFA; diikuti dengan kumpulan BEI-BSO dengan 10% morbiditi. Vaksinasi menyebabkan titer antibodi HI yang tinggi pada kebanyakan kumpulan divaksinasi melainkan UVC-BSO dan UVC-IFA. Ayam di dalam kesemua kumpulan cabaran-vaksinasi meluruhkan virus cabaran dari hari ketiga pos cabaran. Perbezaan yang tidak ketara diperhatikan dalam kekerapan pengesanan virus di kalangan kumpulan divaksinasi (UVC-BSO dan UVC-IFA), dan kawalan positif dengan kejadian berbeza. Tempoh peluruhan virus virulen pada ayam dijangkiti dalam kumpulan divaksinasi adalah berbeza. Program vaksinasi yang digunakan pada kumpulan vaksin terbunuh dan hidup telah memendekkan tempoh peluruhan virus (hanya pada hari ketiga pos cabaran), manakala kumpulan kawalan dan UVC-BSO bermula dari hari ketiga pos cabaran berterusan meluruhkan virus virulen di dalam najis hingga ke akhir eksperimen. Sementara itu, kumpulan UVC-IFA mula meluruhkan virus dari hari kelima pos cabaran hingga ke akhir eksperimen. Akhirnya, kumpulan-kumpulan BEI-BSO, BEI-IFA dan komersil mula meluruhkan virus dari hari ketujuh pos cabaran hingga ke akhir eksperimen dengan penurunan titer pada hari kesepuluh pos cabaran. Pengukuran potensi vaksin dinyahaktif BEI-BSO dan BEI-IFA dengan menggunakan kematian sebagai metrik, memberikan keputusan $10^{-7.612}$ untuk dos lengkap BEI-BSO manakala $10^{-7.532}$ untuk BEI-IFA.

Sebagai kesimpulan, agen etiologi wabak ND yang dilaporkan baru-baru ini pada ayam divaksinasi di Malaysia tergolong dalam strain velogenik genotip VIIa. Pendedahan NDV tersebut pada lampu UVC+riboflavin selama 14 jam atau perawatan virus tersebut dengan rawatan BEI selama 21 jam pada 37°C ditemui adalah mencukupi bagi penyahaktifan lengkap virus sebagaimana ditunjukkan dengan kegagalannya untuk menyebabkan kematian pada telur ayam berembrio SPF, 6 hari pos inokulasi dalam dua laluan berturutan. Penyediaan emulsi stabil air dalam minyak daripada kedua-dua minyak bijan hitam dan adjuvant Freund's tak lengkap telah berjaya dicapai dalam kajian ini. Satu vaksin ND tidak aktif minyak-teremulsi daripada virus sangat patogenik IBS/025 berupaya melindungi ayam muda terhadap pencilan virus IBS/002 genotip VII. Minyak BEI-bijan hitam dan BEI-adjuvan Freund's di samping vaksin komersil telah dibuktikan dalam kajian ini berupaya menawarkan perlindungan sepenuhnya terhadap cabaran NDV virulen.

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I certify that a Thesis Examination Committee has met on 26 October 2017 to conduct the final examination of Oday Abdul Razzaq Abdul Wahhab Aljumaili on his thesis entitled "Characterization of Recently Isolated Newcastle Disease Viruses and Development of Inactivated Vaccine using Genotype VII Newcastle Disease Virus" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

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TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iv
ACKNOWLEDGEMENTS	vii
APPROVAL	viii
DECLARATION	x
LIST OF TABLES	xv
LIST OF FIGURES	xvii
LIST OF APPENDICES	xix
LIST OF ABBREVIATIONS	xx
 CHAPTER	
1 INTRODUCTION	1
1.1 Introduction	1
1.2 Hypothesis	4
 2 LITERATURE REVIEW	5
2.1 Virus classification	5
2.2 Structure and properties	7
2.3 Viral Glycoproteins	8
2.4 Replication of the virus	8
2.4.1 Virus adsorption and entry	8
2.4.2 Transcription	9
2.4.3 Replication	10
2.4.4 Virus assembly and release	10
2.5 Clinical signs of Newcastle disease	11
2.6 Isolation of NDV from field samples	12
2.7 Molecular basis of pathogenicity of ND	13
2.8 Immunity to Newcastle disease virus	14
2.8.1 Innate immune response to NDV infection in poultry	14
2.8.2 Antibody response to infection and vaccination with NDV	15
2.8.3 Cellular immunity induced by NDV	16
2.9 Conventional veterinary vaccines	17
2.9.1 Conventional live vaccines	17
2.9.2 Conventional lentogenic vaccines	18
2.9.3 Conventional inactivated vaccines	18
2.10 Recombinant vector vaccines	19
2.11 Type of emulsion used for veterinary vaccine	20
2.11.1 Water in oil emulsions (W/O)	20
2.11.2 Water in oil in water emulsion (W/O/W)	21
2.11.3 Oil in water emulsions (O/W)	21

2.11.4	Virus inactivation	21
2.11.5	Chemicals inactivating materials	22
2.11.6	Physicals inactivating materials	23
2.12	Adjuvants	24
2.13	Chicken Vaccine Adjuvants	24
2.14	Antigen delivery strategies	26
2.15	Economic and Public Health Significance	26
2.16	Prevention and control	26
3	ISOLATION AND MOLECULAR CHARACTERIZATION OF GENOTYPE VII NEWCASTLE DISEASE VIRUSES FROM ND VACCINATED FARMS	28
3.1	Introduction	28
3.2	Materials and Methods	29
3.2.1	Sample Specimens	29
3.2.2	Viral RNA Extraction	30
3.2.3	F and HN gene RT-PCR amplification	31
3.2.4	Agarose Gel Electrophoresis of RT-PCR Product	32
3.2.5	Sequencing of RT-PCR Product	32
3.2.6	Virus Propagation	34
3.2.7	Hemagglutination Test	35
3.3	Results	35
3.3.1	Phylogenetic Analysis	37
3.3.2	Detection of NDV and virus isolation	41
3.4	Discussion	43
4	PREPARATION OF INACTIVATED GENOTYPE VII NDV VACCINES USING DIFFERENT INACTIVATION METHODS AND DIFFERENT OIL BASED ADJUVANTS	45
4.1	Introduction	45
4.2	Materials and Methods	46
4.2.1	Virus preparations	46
4.2.2	Extraction of black seed oil	46
4.2.3	Gas Chromatography Analysis on Black Seed Oil	46
4.2.4	Inactivation of the virus	46
4.2.5	Testing the infectivity	48
4.2.6	Hemagglutination Assay	48
4.2.7	TEM negative stain	48
4.2.8	Preparation of inactivated NDV oil adjuvant vaccines	48
4.2.9	Stability tests	49
4.3	Results	49
4.3.1	Gas Chromatography and mass spectroscopy Analysis on Black Seed Oil	49
4.3.2	Binary ethylenimine inactivation	51
4.3.3	UVC light inactivation:	53

4.3.4	Effect of the UVC and BEI Virus Inactivation on Hemagglutination Activity	55
4.3.5	Transmission Electron Microscopy	56
4.3.6	Stability tests	58
4.4	Discussion	58
5	EFFICACY AND POTENCY OF INACTIVATED NDV VACCINE	61
5.1	Introduction	61
5.2	Materials and Methods	62
5.2.1	Chickens and Husbandry	62
5.2.2	Vaccine and Challenge Strains	63
5.2.3	Efficacy Trial	63
5.2.4	Potency test	64
5.2.5	Samplings	64
5.2.6	Hemagglutination Test	66
5.2.7	Hemagglutination Inhibition Test	66
5.2.8	Virus Shedding Measurement	66
5.2.9	Pathogenicity Scoring System	68
5.2.10	Statistics	68
5.3	Results	68
5.3.1	Hemagglutination Inhibition Test	68
5.3.2	Potency result	71
5.3.3	Scoring of Mortality, Morbidity and Pathogenicity	73
5.3.4	Virus Shedding	75
5.3.5	Cloacal Virus Shedding	75
5.4	Discussion	78
5.5	Conclusion	80
6	GENERAL DISCUSSION, CONCLUSION AND RECOMMENDATIONS	81
	REFERENCES	88
	APPENDICES	112
	BIODATA OF STUDENT	128
	LIST OF PUBLICATIONS	129

LIST OF TABLES

Table	Page
2.1 Genotypes of class II NDV in a summarized form (Kiarash, 2014)	6
3.1 Clinical descriptions and vaccination history of NDV samples specimen used in this study.	30
3.2 Primers used for RT-PCR amplification for detection of NDV.	32
3.3 Referential Newcastle disease virus (NDV) strains used in this work for phylogenetic analysis of the F gene and HN gene.	33
3.4 The cleavage site of F gene.	41
3.5 C terminus extension length of NDV HN gene.	42
3.6 Comparison of group mean distance of six Malaysian isolates of NDV showed lowest distance to Genotype VII and highest distance to Genotype II.	42
3.7 Comparison of group mean distance of six Malaysian isolates of NDV showed lowest distance to NDV sub genotype VIIa and highest distance to NDV sub genotype VIIh.	43
4.1 Composition of essential oil from seeds of methanolic extraction of Nigella Sativa and the pharmacological action.	51
4.2 First passage SPF eggs inoculation with 4mM BEI inactivated NDV groups.	52
4.3 First passage SPF eggs inoculation with 10mM BEI inactivated NDV groups.	52
4.4 Second passage SPF eggs inoculation with 4mM BEI inactivated NDV groups.	53
4.5 Second passage SPF eggs inoculation with 10mM BEI inactivated groups.	53

4.6	First passage SPF eggs inoculation with UVC only inactivated NDV groups.	54
4.7	First passage SPF eggs inoculation with UVC + riboflavin inactivated NDV groups.	54
4.8	Second passage SPF eggs inoculation with UVC + riboflavin inactivated NDV groups.	55
4.9	HA titer before and after the inactivation of NDV by different inactivation methods.	56
5.1	The Newcastle disease virus vaccines utilize in vaccine efficacy trial.	63
5.2	Primers and probes used in one-step real-time RT-PCR for evaluation of viral load for detection of NDV.	67
5.3	HI antibody level (in Log ₂) using homologous NDV antigens. Values with significance difference are marked with (*) with P-value < 0.05 as statistically significant relationship.	70
5.4	Reed and Muench method for potency test calculation of BEI -BSO NDV inactivated vaccine.	72
5.5	Reed and Muench method for potency test calculation of BEI- IFA NDV inactivated vaccine.	72
5.6	Morbidity and mortality rates and scores of NDV vaccinated groups.	74
5.7	Cloacal ND virus shedding from vaccinated challenged chickens. Values with significance difference are marked with (*) with P-value < 0.05 as statistically significant relationship.	77

LIST OF FIGURES

Figure		Page
2.1	Newcastle disease virus structure and genomic organization (Yusoff and Tan, 2001).	7
2.2	Schematic diagram of Paramyxovirus transcription and replication (Yan, 2008).	10
2.3	Schematic diagram of the life cycle of Newcastle disease virus; NC refers to the nucleocapsid (Takimoto and Portner, 2004).	11
3.1	Agarose gel electrophoresis analysis of RT-PCR for amplification of partial F gene of NDV. Band of the expected size of 535 bp were detected from all the tested samples. Lane M: 100 bp DNA ladder (Fermentas, USA), Lane 1: UPM-IBS 046/2014, Lane 2: UPM-IBS 060/2014, Lane 3: UPM-IBS 061/2014, Lane 4: UPM-IBS 074/2014, Lane 5: UPM-IBS 160/2015, Lane 6: UPM-IBS 162A/2015, Lane 7: negative control, and Lane 8: Positive control.	36
3.2	Agarose gel electrophoresis analysis of RT-PCR for amplification of partial HN gene of NDV. Band of the expected size of 386bp were detected from all the tested samples. Lane M: 100 bp DNA ladder (Fermentas, USA), Lane 1: UPM-IBS 046/2014, Lane 2: UPM-IBS 060/2014, Lane 3: UPM-IBS 061/2014, Lane 4: UPM-IBS 074/2014, Lane 5: UPM-IBS 160/2015, Lane 6: UPM-IBS 162A/2015, Lane 7: negative control, and Lane 8: Positive control.	36
3.3	Phylogenetic analysis of Malaysian NDV isolates based on partial sequence of F protein gene. Viruses highlighted with the coloured circle (●) were characterised in this study. The phylogenetic tree was constructed by maximum likelihood method as implemented in MEGA 6. Numbers at the nodes indicate a confidence level of bootstrap analysis with 1000 replications as a percentage value. Grouping of the isolates as genotype VII.	38
3.4	Phylogenetic analysis of Malaysian NDV isolates based on partial sequence of F protein gene. Viruses highlighted with the coloured circle (●) were characterised in this study. The phylogenetic tree was constructed by	39

maximum likelihood method as implemented in MEGA 6. Numbers at the nodes indicate a confidence level of bootstrap analysis with 1000 replications as a percentage value. Grouping of the isolates as subgenotype VIIa.

3.5	Phylogenetic analysis of Malaysian NDV isolates based on partial sequence of HN protein gene of NDV isolates. Viruses highlighted with the coloured circle (●) were characterized in this study. The phylogenetic tree was constructed by maximum likelihood method as implemented in MEGA 6. Numbers at the nodes indicate a confidence level of bootstrap analysis with 1000 replications as a percentage value.	40
4.1	GC-MS chromatogram of oil extract of <i>Nigella sativa</i>	50
4.2	TEM analysis of NDV virion following UVC and BEI inactivation variations in the ultrastructure of the live and inactivated viruses, A)live IBS025 complete structure (nucleic acid) and external glycoprotein (HN and F) (white arrow) , B)12 hr UVC destruction of the external glycoprotein, C) 14hr, and D) 16hr UVC completely destruction of external glycoprotein , E) 21hr BEI and F) 48hr BEI no variance on the external glycoprotein comparable to live NDV IBS 025 (100 nm).	57
4.3	Dilution test for the inactivated vaccines indicated that the vaccine preparation is formulated as water in oil emulsion (arrows), (A)UVC-BSO, (B) BEI-BSO, (C) UVC-IFA, (D) BEI-IFA, (E) Commercial vaccine.	58
5.1	The flowchart of the trial of NDV vaccine efficacy.	65
5.2	Antibody titers of NDV following vaccination with inactivated vaccine (A) groups vaccinated with inactivated vaccine only, (B) groups vaccinated with inactivated and live vaccine.	69
5.3	A linear relationship between quantification cycle (Cq) and 10-fold serial dilution of RNA. Standard curve was generated using serially diluted RNA of NDV IBS 002/11	75

LIST OF APPENDICES

Appendix	Page
A Buffer, Chemicals and Reagents	112
B Cleavage site of F gene, all Malaysian NDV isolated categorized as velogenic NDV due to the multiple amino acids and have phenylalanine at position 117 observed in cleavage site of F protein.	113
C Analysis of c terminus extension length of HN gene revealed that all the 6 isolates have no amino acid extension length and ended with KDDRV* with predicted total length of 571 amino acids for HN protein.	116
D EID50 Recorded Data	119
E Cold pressed extraction for Black seed by using the manual pressed machine	120
F GC-MS Analysis Conditions.	121
G UVC light inactivation method.	122
H Formulation of water in oil emulsion.	123
I HLB scale of typical emulsifiers.	124
J Reed and Muench method for calculation of PD60 for BEI-BSO vaccine.	125
K Morbidity and mortality rate graph for the groups vaccinated with inactivated vaccine only	126
L Morbidity and mortality rate graph for the groups vaccinated with inactivated vaccine with live vaccine together	127

LIST OF ABBREVIATIONS

aa	Amino Acid
APMV	Avian Paramyxovirus
BLAST	Basic Local Alignment Search Tool
BSO	Black seed oil
DNA	Deoxyribonucleic Acid
dpc	Day post challenge
dsRNA	Double strand Deoxyribonucleic Acid
EID ₅₀	Embryo infective dose 50
ELISA	Enzyme-linked immunosorbent assays
F	Fusion Protein
F0	Fusion Protein 0
F1	Fusion Protein 1
F2	Fusion Protein 2
GC-MS	Gas chromatography-mass spectrometry
GIT	Gastrointestinal Tract
HI	Hemagglutination Inhibition
HN	Hemagglutinin-Neuraminidase
IBD	Infectious Bursal Disease
ICPI	Intra Cerebral Pathogenicity Index
IFA	Incomplete Freund's adjuvant
IVPI	Intra Venous Pathogenicity Index
L	Large Polymerase Protein
M	Matrix Protein
MEGA	Molecular Evolutionary Genetics Analysis
mM	Millimolar
NCBI	National Centre for Biotechnology Information
ND	Newcastle Disease
NDV	Newcastle Disease Virus
nm	Nano meter
NP	Nucleocapsid Protein
NV-ND	Neurotropic Velogenic Newcastle Disease
O/W	Oil in water
OIE	Office International des Epizooties
P	Phosphoprotein Protein
PBMC	Peripheral Blood Mononuclear Cells
PBS	Phosphate Buffered Saline
PCR	Polymerase Chain Reaction
PD60	Protective dose 60
RNA	Ribonucleic Acid
rpm	Revolutions Per Minute
RT	Reverse Transcription
RT-PCR	Reverse Transcription-Polymerase Chain Reaction

SPF	Specific-Pathogen-Free
TAE	Tris-Acetate-EDTA
UVC	Ultraviolet light type C
VG/GA	Villegas-Glisson/University of Georgia
VV-ND	Viscerotropic Velogenic Newcastle Disease
W/O	Water in oil
W/O/W	Water in oil in water



CHAPTER 1

INTRODUCTION

1.1 Introduction

Since over 80 years of the Newcastle disease (ND) discovery in Java and England (Doyle, 1927; Kraneveld, 1926) and vaccine against NDV introduction in 1950s in controlling NDV disease (Alexander and Senne, 2008), Newcastle disease is yet the one of the most significant avian infections which lingers and causes high loss economically in the poultry industry. Due to its global epidemics and geographical spread, the Organization for Animal Health (OIE) of the world enlisted Newcastle disease to be a notifiable disease (OIE 2013). Newcastle disease is as a result of infection with Newcastle disease virus (NDV). This virus is an extremely infectious agent which is able to cause high dead rate in unvaccinated flocks, beside the subclinical forms of Newcastle disease in flocks that are vaccinated and/or Newcastle disease virus exposed flocks which might possess synergist consequence with other viral and or bacterial infections which can lead to additional severe disease and higher losses economically (Swayne and King, 2003).

Newcastle disease virus (NDV) is an avian paramyxovirus serotype-1 (APMV-1) which is a member of sub-family Paramyxovirinae from the family Paramyxoviridae and in the Mononegavirales virus order (Fauquet and Fargette, 2005; Mayo, 2002). NDV possess a negative sense, single stranded RNA genome that encodes for 6 genes (Lyles et al., 2013). Even though the entire APMV-1 viruses comes from a single serotype, they possess different genomic structures and are separated into different genotypes (Diel et al., 2012). Newcastle disease viruses are classified due to their F gene to 2 basic classes. Class one (I) Newcastle disease viruses are typically isolated from water fowls (Anatidae) and shore birds whereas, Class two (II) Newcastle disease viruses induces disease in poultry, and are alienated into ten (10) different genotypes (Miller et al., 2010). Certain infectious Newcastle disease virus which are isolates from several nations within Europe during 1990s were notice not to fit into whichever of recognized genotypes of those time, therefore, were categorized as genotype VII NDVs (Lomniczi et al., 1998). These Newcastle disease viruses are thought to be from East-Asia (Lomniczi et al., 1998), and subsequently extent to Middle-East, Europe and Africa (Bogoyavlenskiy et al., 2009; Wang et al., 2006). Recently research revealed that currently genotype VII is the main circulating NDV in South-East Asia which causes main epidemics comprising vaccinated flocks against Newcastle disease virus (Umali et al., 2013; Yi et al., 2011; Tan et al., 2010; Cho et al., 2008) .

From the time when NDV was discovered, countless determinations have been executed in controlling Newcastle disease in poultry industry. Apart from biosecurity and practice of good farm husbandry, the control of Newcastle disease is by vaccinating the flocks. Presently, numerous diverse vaccines are accessible in the market for Newcastle disease control both in large layer/ broiler chicken farms and also chickens of backyard village. Majority of the vaccines is from genotype II of class II of Newcastle disease viruses (Chong et al., 2010). Malaysia is like many other nations, poultry farmers utilized low pathogenic Newcastle disease viruses viz. lentogenic NDV such as Hitchner B1 and LaSota, as live vaccines for the protection against Newcastle disease. Apart from the two live vaccines, other Newcastle disease virus vaccine strains for example S, Ulster 2C, NDV-6/10 and enteric vaccine strain VG-GA is also been utilized (Aini, 2006). Numerous different types of genetically produced ND vaccines have been produced and tried experimentally. Nevertheless, very limited recombinant Newcastle disease virus vaccines are commercially obtainable, viz., herpesvirus turkey virus (HVT) based Newcastle disease virus vaccine (Palya et al., 2012). These vaccines have displayed encouraging outcomes in rendering defense against experimental trial with Newcastle disease virus velogenic strain.

Even though it is likely to evaluate vaccine efficiency by laboratory scale experimentations, it is very challenging to estimate the vaccine efficiency in the field (Chulan et al., 1982). Therefore, failure of vaccination after Newcastle disease virus vaccination has been stated. Amongst the primary issues that help to poor Newcastle disease virus vaccine induced immunity are incorrect vaccination dosage and timing, existence of simultaneous infection specifically immunosuppressive agents for example chicken infectious anemia (CAV), infectious bursal disease (IBD) or Marek's disease, nutritive insufficiencies in addition to mycotoxins in feed being the likely cause (s) for the breakdown in the vaccine inducing protection (Habibian et al., 2014; Zhang et al., 2012; Saif, 1991). Nonetheless, current research revealed that, commercially obtained Newcastle disease virus vaccines offer diverse level of protection against challenged with diverse genotypes of Newcastle disease virus (Hu et al., 2009; Miller et al., 2009) rising the status of affinity between vaccine and field strains of NDV. Additionally, it has been established that, LaSota vaccine, a genotype II NDV is not efficient in decreasing shedding of virus and clinical symptom after experimental trial with genotype VII isolates as equated to the reverse genetic designed genotype VII vaccine (Hu et al. 2009; Cho et al. 2008b). Nevertheless, the significance of virus shedding and transmission to vulnerable flocks is yet to be clear. Several research revealed that Newcastle disease virus vaccine was capable of protecting against death (disease immunity) nonetheless incapable in providing sterilizing immunity through prevention of infection after challenging with velogenic NDV (Cornax et al., 2012; Ezema et al., 2009).

Since the year 2000, genotype VII NDV has been reported in unvaccinated and vaccinated flocks of chicken in Malaysia (Berhanu et al., 2010; Tan et al., 2010). Recently investigation revealed that genotype VII NDV yet circulate amongst the chicken flocks in the country in spite of the widespread usage of LaSota based vaccines (Roohani et al., 2015). Nevertheless, inadequate investigation have being done in gaining access to the capability of genotype II (LaSota, B1, VG/GA, Avinew) (genotype mismatched vaccine) and genotype VII (genotype matched vaccine) in rendering defense against experimental trial with velogenic genotype VII in specific-pathogen-free (SPF) and large flocks of chicken.

The Malaysian strain (IBS025/13) has been characterized as natural recombinant strain between genotype II and genotype VII (Satharasinghe et al., 2016). It has nucleocapsid protein and phosphoprotein genes of genotype II whereas surface glycoproteins (fusion, hemagglutinin-neuraminidase) and large polymerase of genotype VII this strain has been used in this study to prepare the inactivated vaccines.

Typically, two leading types of vaccines are utilized in protecting animals and man clinically and virologically on experimental trials: improved live virus (attenuated virus) vaccines and killed virus (KV; inactivated virus) vaccines. KV vaccines are regularly chosen as a result of it safety reason, nevertheless certain problems could occur when using them. Initially, inactivation of virus could later reverse to be imperfect with epidemics post vaccination which could be a consequence (Patil et al., 2002; Beck and Strohmaier, 1987). Additionally, the viral-neutralizing epitopes could be damaged at the course of inactivation, resulting to reduced neutralizing antibody reaction and a reduced defense on experimental trial (Cham et al., 2006; FAO, 1995). Hence, effective excellence control of the inactivated antigen is essential in evaluating virus inactivation and the consequence of the inactivation process on neutralizing epitopes. However, currently existing viral vaccines are unable to cope with many types of prevailing viruses in the field. Therefore, new vaccines have to be created from the strains responsible for new outbreaks (Lee et al., 2012).

These days, commercial available killed vaccines are commonly primed from mineral oil as an adjuvant. The mineral oil can lead to opposing outcome plus tissue reaction; remains of vaccine in tissues of the poultry, the mineral could result to carcinogenic agent on the poultry and poultry products consumers. Due to the opposing properties of the mineral oil, considerable investigation had been introduced to overcome the difficulties through discovering other types of oil which could be replaced for the mineral oil (Stone, 1997; Gupta et al., 1993; Yamanaka et al., 1993). In the year 1996, inactivated vaccine in form

of water in oil in water (WOW) was developed through the use of subunit virus, acquired from whole or incomplete interruption by Tween 80, as an antigen (Cajavec et al., 1996). The vaccine delivered a less viscidness, high steadiness with lowered percentage of mineral oil in the vaccine to sustain the useful vaccination with stress-free washing of vaccination apparatus (Kaleta and Baldauf, 1988). Preceding report show that the WOW vaccine appeared to result in less reactions of tissue as compare to water in oil (WO) vaccine due to less liquid paraffin concentration (Fukanoki et al., 2001).

Vaccines produced from natural oil delivered lesser efficacy of stimulating HI antibody and higher viscosity as equated to vaccines produced from mineral oil (Stone, 1993).

1.2 Hypothesis

The hypothesis of this investigation is that

- 1- The genotype of Newcastle disease virus isolated from ND epidemics from vaccinated chicken farms belongs to velogenic genotype VII.
- 2- The inactivated NDV vaccine using naturally occurring recombinant genotype VII is able to induce better immune responses and protection against challenge with virulent genotype VII NDV.
- 3- The inactivated NDV vaccine formulated in black seed oil (*Nigella Sativa* oil) as adjuvant is able to induce better protection compared to inactivated NDV vaccine formulated in Freund's incomplete adjuvants against genotype VII NDV challenge.

To address the entire hypotheses, the specific objectives of this investigation are as follows:

- 1- To isolate and perform molecular characterization of Newcastle disease virus (NDV) from Newcastle disease outbreaks originated from NDV vaccinated farms.
- 2- To evaluate the inactivation of ND virus through different methods (UVC only, UVC+ Riboflavin and conventional BEI inactivation).
- 3- To study the adjuvant properties of black seed oil with incomplete Freund's adjuvant of inactivated ND vaccine in the form of W/O.
- 4- To determine the efficacy and potency of the developed NDV vaccines against velogenic genotype VII challenge in specific-pathogen-free flocks of chickens.

REFERENCES

- Aaskov, J., Williams, L., and Yu, S. (1997). A candidate Ross River virus vaccine: preclinical evaluation. *Vaccine*, 15(12-13), 1396-404.
- Abolnik, C., Horner, R. F., Bisschop, S. P. R., Parker, M. E., Romito, M., and Viljoen, G. J. (2004). A phylogenetic study of South African Newcastle disease virus strains isolated between 1990 and 2002 suggests epidemiological origins in the Far East. *Archives of Virology*, 149(3), 603-19.
- Adair, B. (2000). Immunopathogenesis of chicken anemia virus infection. *Developmental & Comparative Immunology*, 24(2-3), 247-255.
- Ahmed, K. A., Saxena, V. K., Ara, A., Singh, K. B., Sundaresan, N. R., Saxena, M., and Rasool, T. J. (2007). Immune response to Newcastle disease virus in chicken lines divergently selected for cutaneous hypersensitivity. *International Journal of Immunogenetics*, 34(6), 445-455.
- Aini, I. (2006). Newcastle disease. In M. Zamri-Saad (Ed.), *Disease of Poultry in Southeast Asia* (pp. 225-233). Serdang, Malaysia, Malindo Printers Sdn. Bhd.
- Al-Garib, S. O., Gielkens, A. L. J., Gruys, E., and Koch, G. (2003). Immunoglobulin class distribution of systemic and mucosal antibody responses to Newcastle disease in chickens. *Avian Diseases*, 47(1), 32-40.
- Al-Garib, S. O., Gielkens, A. L. J., Gruys, E., and Kochi, G. (2003). Review of Newcastle disease virus with particular references to immunity and vaccination. *World's Poultry Science Journal*, 59(2), 185-200.
- Alders, R. (1999). Controlling Newcastle Disease in village chickens in Mozambique. *African Studies Review and Newsletter*, 21, 43-44.
- Alderson, T. (1964). The mechanism of formaldehyde-induced mutagenesis. The monohydroxymethylation reaction of formaldehyde with adenylic acid as the necessary and sufficient condition for the mediation of the mutagenic activity of formaldehyde. *Mutation Research*, 77-85.
- Aldous, E. W., and Alexander, D. J. (2001). Detection and differentiation of Newcastle disease virus (avian paramyxovirus type 1). *Avian Pathology*, 30, 117-128.

- Aldous, E. W., Mynn, J. K., Banks, J., and Alexander, D. J. (2003). A molecular epidemiological study of avian paramyxovirus type 1 (Newcastle disease virus) isolates by phylogenetic analysis of a partial nucleotide sequence of the fusion protein gene. *Avian Pathology*, 32(3), 239–256.
- Alexander, D. J. (2000). Newcastle disease and other avian paramyxoviruses. In *Revue scientifique et technique (International Office of Epizootics)* (12th ed., pp. 443–462).
- Alexander, D. J. (2008). Newcastle Disease, Other Avian Paramyxoviruses, and Pneumovirus Infections. In Y. M. Saif (Ed.), *Diseases of Poultry* (pp. 63–87). Wiley-Blackwell.
- Alexander, D. J., and Senne, D. A. (2008). Newcastle Disease, Other Avian Paramyxoviruses, and Pneumovirus Infections. In *Diseases of Poultry* (pp. 75–100). Wiley-Blackwell.
- Allan, W. H. (1978). A review of Newcastle disease and its control. In *Proceedings of the APHCA Poultry Diseases Workshop, Kuala Lumpur, Malaysia*, (pp. 94–101).
- Allan, W. H., and Borland, L. J. (1979). The stress index: A method for indicating the pathogenicity of vaccinal Newcastle disease virus when administered by aerosol. *Avian Pathology*, 8(4), 401–409.
- Asif, M., Jenkins, K. A., Hilton, L. S., Kimpton, W. G., Bean, A. G. D., and Lowenthal, J. W. (2004). Cytokines as adjuvants for avian vaccines. *Immunology and Cell Biology*, 82(6), 638–643.
- Aucouturier, J., Dupuis, L., and Ganne, V. (2001). Adjuvants designed for veterinary and human vaccines. *Vaccine*, 19(17–19), 2666–2672.
- Bahnemann, G., and Mesquita, A. (1987). Oil adjuvant vaccine against foot-and-mouth disease. In *Boletín del centro panamericano de fiebre aftosa* (pp. 25–30).
- Bahnemann, H. G. (1976). Inactivation of viruses in serum with binary ethyleneimine. *Journal of Clinical Microbiology*, 3(2), 209–210.
- Ballagi-Pordány, a, Wehmann, E., Herczeg, J., Belák, S., and Lomniczi, B. (1996). Identification and grouping of Newcastle disease virus strains by restriction site analysis of a region from the F gene. *Archives of Virology*, 141(2), 243–61.
- Barnett, P. V., Pullen, L., Williams, L., and Doel, T. R. (1996). International bank for foot-and-mouth disease vaccine: Assessment of Montanide

- ISA 25 and ISA 206, two commercially available oil adjuvants. *Vaccine*, 14(13), 1187–1198.
- Bazin, H. (2003). A brief history of the prevention of infectious diseases by immunisations. *Comparative Immunology, Microbiology and Infectious Diseases*, 26(5–6), 293–308.
- Beck, E., and Strohmaier, K. (1987). Subtyping of European foot-and-mouth disease virus strains by nucleotide sequence determination. *Journal of Virology*, 61(5), 1621–1629.
- Bell, J. G. (2001). A Comparison of the Different Vaccines Available for the Control of Newcastle Disease in Village Chickens. In *ACIAR proceedings* (pp. 56–60).
- Bell, J. G., Belarbi, D. A., and Amara, A. (1990). A controlled vaccination trial for Newcastle disease under village conditions. *Preventive Veterinary Medicine*, 9, 295–300.
- Berhanu, A., Ideris, A., Omar, A. R., and Bejo, M. (2010). Molecular characterization of partial fusion gene and C-terminus extension length of haemagglutinin-neuraminidase gene of recently isolated Newcastle disease virus isolates in Malaysia. *Virology Journal*, 7(1), 183.
- Bey, R., Simonson, R., and Garcon, N. (1998). Formulation of Vaccines. In *Development and Formulation of Veterinary Dosage Forms* (pp. 283–304).
- Biancifiori, F., and Fioroni, A. (1983). An occurrence of Newcastle disease in pigeons: Virological and serological studies on the isolates. *Comparative Immunology, Microbiology and Infectious Diseases*, 6(3), 247–252.
- Blumberg, B. M., and Kolakofsky, D. (1981). Intracellular vesicular stomatitis virus leader RNAs are found in nucleocapsid structures. *Journal of Virology*, 40(2), 568–576.
- Blumberg, B. M., Leppert, M., and Kolakofsky, D. (1981). Interaction of VSV leader RNA and nucleocapsid protein may control VSV genome replication. *Cell*, 23(3), 837–845.
- Bogoyavlenskiy, A., Berezin, V., Prilipov, A., Usachev, E., Lyapina, O., Korotetskiy, I., Zaitceva, I., Asanova, S., Kydyrmanov, A., Daulbaeva, K., Shakhvorostova, L., Sayatov, M., and King, D. (2009). Newcastle disease outbreaks in Kazakhstan and Kyrgyzstan during 1998, 2000, 2001, 2003, 2004, and 2005 were caused by viruses of the genotypes VIIb and VIIId. *Virus Genes*, 39(1), 94–101.

- Boven, M., Bouma, A., Fabri, T. H. F., Katsma, E., Hartog, L., and Koch, G. (2008). Herd immunity to Newcastle disease virus in poultry by vaccination. *Avian Pathology*, 37(1), 1-5.
- Bowersock, T. L., and Martin, S. (1999). Vaccine delivery to animals. *Advanced Drug Delivery Reviews*, 38(2), 167-194.
- Broo, K., Wei, J., Marshall, D., Brown, F., Smith, T. J., Johnson, J. E., Schneemann, A., and Siuzdak, G. (2001). Viral capsid mobility: A dynamic conduit for inactivation. *Proceedings of the National Academy of Sciences*, 98(5), 2274-2277.
- Burleson, F. G., Chambers, T. M., and Wiedbrauk, D. L. (1992). *Virology A Laboratory Manual*.
- Cajavec, S., Bidin, Z., Sladic, D., and Pokric, B. (1996). Tween 80-Solubilized Newcastle Disease Virus Prepared as a Water-in-Oil-in-Water Vaccine. *Avian Diseases*, 40(1), 193.
- Calnek, B. W., Harris, R. W., Buscaglia, C., Schat, K. A., and Lucio, B. (1998). Relationship between the immunosuppressive potential and the pathotype of Marek's disease virus isolates. *Avian Diseases*, 42(1), 124-132.
- Cannon, M. J., and Russell, P. H. (1986). Secondary in vitro stimulation of specific cytotoxic cells to newcastle disease virus in chickens. *Avian Pathology*, 15(4), 731-740.
- Capua, I., Dalla, P. M., Mutinelli, F., Marangon, S., and Terregino, C. (2002). Newcastle disease outbreaks in Italy during 2000. *The Veterinary Record*, 150(18), 565-8.
- Cattoli, G., Fusaro, A., Monne, I., Molia, S., Le Menach, A., Maregeya, B., Nchare, A., Bangana, I., Maina, a. G., N'Goran Koffi, J. N., Thiam, H., Bezeid, O. E. M. a, Salviato, A., Nisi, R., Terregino, C., and Capua, I. (2011). Emergence of a new genetic lineage of Newcastle disease virus in West and Central Africa-Implications for diagnosis and control. *Veterinary Microbiology*, 142(3-4), 168-176.
- Cham, B. E., Vickery, K., Tohidi-Esfahani, R., and Cossart, Y. (2006). Delipidation of a hepadnavirus: Viral inactivation and vaccine development. *Journal of Virological Methods*, 137(1), 160-163.
- Chimeno Zoth, S., Gómez, E., Carrillo, E., and Berinstein, A. (2008). Locally produced mucosal IgG in chickens immunized with conventional

- vaccines for Newcastle disease virus. *Brazilian Journal of Medical and Biological Research*, 41(4), 318–323.
- Cho, S. H., Kwon, H. J., Kim, T. E., Kim, J. H., Yoo, H. S., and Kim, S. J. (2008). Variation of a Newcastle Disease Virus Hemagglutinin-Neuraminidase Linear Epitope. *Journal of Clinical Microbiology*, 46(4), 1541–1544.
- Cho S.H, Kwon, H. J., Kim, T. E., Kim, J. H., Yoo, H. S., Park, M. H., Park, Y. H., and Kim, S. J. (2008). Characterization of a recombinant Newcastle disease virus vaccine strain. *Clinical and Vaccine Immunology : CVI*, 15(10), 1572–9.
- Choi, K. S., Kye, S. J., Kim, J. Y., Damasco, V. R., Sorn, S., Lee, Y. J., Choi, J. G., Kang, H. M., Kim, K. Il, Song, B. M., and Lee, H. S. (2013). Molecular epidemiological investigation of velogenic Newcastle disease viruses from village chickens in Cambodia. *Virus Genes*, 47(2), 244–249.
- Choi, K. S., Kye, S. J., Kim, J. Y., and Lee, H. S. (2013). Genetic and antigenic variation of shedding viruses from vaccinated chickens after challenge with virulent Newcastle disease virus. *Avian Diseases*, 57(2), 303–306.
- Chong, Y. L., Padhi, A., Hudson, P. J., and Poss, M. (2010). The effect of vaccination on the evolution and population dynamics of avian paramyxovirus-1. *PLoS Pathogens*, 6(4), 1–11.
- Chulan, U., Ibrahim, A. L., Mustaffa Babjee, A., and Sheikh-Omar, A. R. (1982). Vaccination against newcastle disease. *Tropical Animal Health and Production*, 14(3), 177–184.
- Collins, M. S., Bashiruddin, J. B., and Alexander, D. J. (1993). Deduced amino acid sequences at the fusion protein cleavage site of Newcastle disease viruses showing variation in antigenicity and pathogenicity. *Archives of Virology*, 128(3–4), 363–370.
- Collins, M. S., Strong, I., and Alexander, D. J. (1994). Evaluation of the molecular basis of pathogenicity of the variant Newcastle disease viruses termed "pigeon "MV-1 viruses." *Archives of Virology*, 134, 403–411.
- Cornax, I., Miller, P. J., and Afonso, C. L. (2012). Characterization of Live LaSota Vaccine Strain-Induced Protection in Chickens upon Early Challenge with a Virulent Newcastle Disease Virus of Heterologous Genotype. *Avian Diseases Digest*, 7(3), 464–470.
- Czeglédi, A., Ujvári, D., Somogyi, E., Wehmann, E., Werner, O., and Lomniczi, B. (2006). Third genome size category of avian paramyxovirus serotype

- 1 (Newcastle disease virus) and evolutionary implications. *Virus Research*, 120(1-2), 36-48.
- Czerkinsky, C., Anjuere, F., McGhee, J. R., George-Chandy, A., Holmgren, J., Kieny, M. P., Fujiyashi, K., Mestecky, J. F., Pierrefite-Carle, V., Rask, C., and Sun, J. Bin. (1999). Mucosal immunity and tolerance: Relevance to vaccine development. *Immunological Reviews*, 170, 197-222.
- Dalsgaard, K., Hilgers, L., and Trouvé, G. (1990). Classical and New Approaches to Adjuvant Use in Domestic Food Animals. In *Immunomodulation in Domestic Food Animals* (pp. 121-160).
- Darnell, M. E. R., Subbarao, K., Feinstone, S. M., and Taylor, D. R. (2004). Inactivation of the coronavirus that induces severe acute respiratory syndrome, SARS-CoV. *Journal of Virological Methods*, 121(1), 85-91.
- Degefa, T., Dadi, L., Yami, A., K, G. M., and Nassir, M. (2004). Technical and economic evaluation of different methods of Newcastle disease vaccine administration. *Journal of Veterinary Medicine*, 51, 365-369.
- Delrue, I., Delputte, P. L., and Nauwynck, H. J. (2009). Assessing the functionality of viral entry-associated domains of porcine reproductive and respiratory syndrome virus during inactivation procedures, a potential tool to optimize inactivated vaccines. *Veterinary Research*, 40(6), 62.
- Delrue, I., Verzele, D., Madder, A., and Nauwynck, H. J. (2012). Inactivated virus vaccines from chemistry to prophylaxis: merits, risks and challenges. *Expert Review of Vaccines*, 11(6), 695-719.
- Desbat, B., Lancelot, E., Krell, T., Nicolai, M. C., Vogel, F., Chevalier, M., and Ronzon, F. (2011). Effect of the β -propiolactone treatment on the adsorption and fusion of influenza A/Brisbane/59/2007 and A/New Caledonia/20/1999 virus H1N1 on a dimyristoylphosphatidylcholine/ganglioside GM3 mixed phospholipids monolayer at the air-water interface. *Langmuir*, 27(22), 13675-13683.
- Diel, D. G., da Silva, L. H. A., Liu, H., Wang, Z., Miller, P. J., and Afonso, C. L. (2012). Genetic diversity of avian paramyxovirus type 1: Proposal for a unified nomenclature and classification system of Newcastle disease virus genotypes. *Infection, Genetics and Evolution*, 12(8), 1770-1779.
- Dimmock, N. J. (1967). Differences between the thermal inactivation of picornaviruses at "high" and "low" temperatures. *Virology*, 31(1953), 338-353.

- Doel, T. R. (1997). Potency assessment of inactivated viral vaccines. In *Vaccine manual* (pp. 395–409).
- Dolganiuc, V., McGinnes, L., Luna, E. J., and Morrison, T. G. (2003). Role of the cytoplasmic domain of the Newcastle disease virus fusion protein in association with lipid rafts. *Journal of Virology*, 77(24), 12968–12979.
- Doms, R. W., Lamb, R. A., Rose, J. K., and Helenius, A. (1993). Folding and assembly of viral membrane proteins. *Virology*, 193, 545–562.
- Dortmans, J. C. F. M., Peeters, B. P. H., and Koch, G. (2012). Newcastle disease virus outbreaks: Vaccine mismatch or inadequate application. *Veterinary Microbiology*, 160(1–2), 17–22.
- Doyle, T. M. (1927). A hitherto unrecorded disease of fowls due to a filter-passing virus. *Journal of Comparative Pathology and Therapeutics*, 40, 144–169.
- Dupuis, M., Murphy, T. J., Higgins, D., Ugozzoli, M., van Nest, G., Ott, G., and McDonald, D. M. (1998). Dendritic cells internalize vaccine adjuvant after intramuscular injection. *Cellular Immunology*, 186(1), 18–27.
- El-Bagoury, G. F., Nasr, M. H. M., El-Habbaa, A. S., and Hala, M. E. M. (2015). A trial to improve stability and immunogenicity of inactivated NDV vaccine with paraffin oil adjuvant using aluminum stearate. *Benha Veterinary Medical Journal*, 28, 199–209.
- Ezema, W. S., Okoye, J. O. A., and Nwanta, J. A. (2009). LaSota vaccination may not protect against the lesions of velogenic newcastle disease in chickens. *Tropical Animal Health and Production*, 41(4), 477–484.
- FAO. (1995). Overview: the present state of veterinary vaccine development. In *FAO Manual for the Production and Quality Control of Veterinary Vaccines for Use in Developing Countries* (pp. 127–142).
- Fauquet, C. M., and Fargette, D. (2005). International Committee on Taxonomy of Viruses and the 3,142 unassigned species. *Virology Journal*, 2, 64.
- Fentie, T., Dadi, K., Kassa, T., Sahle, M., and Cattoli, G. (2014). Effect of vaccination on transmission characteristics of highly virulent Newcastle disease virus in experimentally infected chickens. *Avian Pathology*, 43(August), 1–25.
- Ferreira, L., Muñoz-Barroso, I., Marcos, F., Shnyrov, V. L., and Villar, E. (2004). Sialidase, receptor-binding and fusion-promotion activities of

Newcastle disease virus haemagglutinin-neuraminidase glycoprotein: A mutational and kinetic study. *Journal of General Virology*, 85(7), 1981–1988.

Freund J. (1951). The effect of paraffin oil and mycobacteria. *American Journal of Clinical Pathology*, 7, 645–656.

Fujii, Y., Sakaguchi, T., Kiyotani, K., and Yoshida, T. (1999). Comparison of substrate specificities against the fusion glycoprotein of virulent Newcastle disease virus between a chick embryo fibroblast processing protease and mammalian subtilisin-like proteases. *Microbiology and Immunology*, 43(2), 133–140.

Fukanoki, S., Iwakura, T., Iwaki, S., Matsumoto, K., Takeda, R., Ikeda, K., Shi, Z., and Mori, H. (2001). Safety and efficacy of water-in-oil-in-water emulsion vaccines containing Newcastle disease virus haemagglutinin-neuraminidase glycoprotein. *Avian Pathology*, 30(5), 509–16.

Gali-Muhtasib, H., El-Najjar, N., and Schneider-Stock, R. (2006). The medicinal potential of black seed (*Nigella sativa*) and its components. *Advances in Phytomedicine*, 2(C), 133–153.

Ganne, V., Eloit, M., Laval, A., Adam, M., and Trouve, G. (1994). vaccine by the addition of oil adjuvants. *Vaccine*, 12(13), 1190–1196.

Garten, W., Berk, W., Nagai, Y., Rott, R., and Klenk, H. D. (1980). Mutational changes of the protease susceptibility of glycoprotein F of Newcastle disease virus: Effects on pathogenicity. *Journal of General Virology*, 50(1), 135–147.

Gates, K. S., Noonan, T., and Dutta, S. (2004). Biologically Relevant Chemical Reactions of N7-Alkylguanine Residues in DNA. *Chemical Research in Toxicology*, 17(7), 839–856.

Gavin, A. L., Hoebe, K., Duong, B., Ota, T., Martin, C., Beutler, B., and Nemazee, D. (2006). Adjuvant-Enhanced Antibody Receptor Signaling. *Science*, 314, 1936–1938.

Ghosheh, O. A., Houdi, A. A., and Crooks, P. A. (1999). High performance liquid chromatographic analysis of the pharmacologically active quinones and related compounds in the oil of the black seed (*Nigella sativa* L.). *Journal of Pharmaceutical and Biomedical Analysis*, 19(5), 757–762.

- Ghumman, J. S., Wiggins, A. D., and Bankowski, R. A. (1976). Antibody Response and Resistance of Turkeys to Newcastle Disease Vaccine Strain LaSota. *Avian Diseases*, 20(1), 1-8.
- Giambrone, J. J., and Closser, J. (1990). Effect of breeder vaccination on immunization of progeny against Newcastle disease. *Avian Diseases*, 34(1), 114-119.
- Glickman, R. L., Syddall, R. J., Iorio, R. M., Sheehan, J. P., and Bratt, M. A. (1988). Quantitative basic residue requirements in the cleavage-activation site of the fusion glycoprotein as a determinant of virulence for Newcastle disease virus. *Journal of Virology*, 62(1), 354-356.
- Gohm, D. S., Thur, B., and Hofmann, M. a. (2000). Detection of Newcastle disease virus in organs and faeces of experimentally infected chickens using RT-PCR. *Avian Pathology*, 29(2), 143-152.
- Gupta, R. K., Relyveld, E. H., Lindblad, E. B., Bizzini, B., Ben-Efraim, S., and Gupta, C. K. (1993). Adjuvants – a balance between toxicity and adjuvanticity. *Vaccine*, 11(3), 293-306.
- Habibian, M., Ghazi, S., Moeini, M. M., and Abdolmohammadi, A. (2014). Effects of dietary selenium and vitamin E on immune response and biological blood parameters of broilers reared under thermoneutral or heat stress conditions. *International Journal of Biometeorology*, 58(5), 741-752.
- Hamal, K. R., Burgess, S. C., Pevzner, I. Y., and Erf, G. F. (2006). Maternal antibody transfer from dams to their egg yolks, egg whites, and chicks in meat lines of chickens. *Poultry Science*, 85(8), 1364-72.
- Hanson, R. p., and Brandly, C. A. (1958). Newcastle Disease. *Annals of the New York Academy of Sciences*, 70, 585-597.
- Hanson, R. P., and Brandly, C. A. (1955). Identification of vaccine strains of Newcastle disease virus. *Science*, 122, 156-157.
- Haque, M., Hossain, M., Islam, M., Zinnah, M., Khan, M., and Islam, M. (2010). Isolation and Detection of Newcastle Disease Virus From Field Outbreaks in Broiler and Layer Chickens By Reverse Transcription-Polymerase Chain Reaction. *Bangladesh Journal of Veterinary Medicine*, 8, 87-92.
- Hasson, S. S. A. A., Al-Busaidi, J. K. Z., and Sallam, T. A. (2015). The past, current and future trends in DNA vaccine immunisations. *Asian Pacific Journal of Tropical Biomedicine*, 5(5), 344-353.

- Henning, J., Meers, J., and Davies, P. R. (2005). Exposure of rabbits to ultraviolet light-inactivated rabbit haemorrhagic disease virus (RHDV) and subsequent challenge with virulent virus. *Epidemiology and Infection*, 133(4), 731–5.
- Herbert, W. J. (1965). Multiple emulsions: a new form of mineral-oil antigen adjuvant. *The Lancet*, 286(7416), 771.
- Herczeg, J., Pascucci, S., Massi, P., Luini, M., Selli, L., Capua, I., and Lomniczi, B. (2001). A longitudinal study of velogenic Newcastle disease virus genotypes isolated in Italy between 1960 and 2000 A longitudinal study of velogenic Newcastle disease virus genotypes isolated in Italy between 1960 and 2000. *Avian Pathology*, 30, 163–168.
- Hitchner, S. B., and Johnson, E. P. (1948). A virus of low virulence for immunizing fowls against Newcastle disease; avian pneumoencephalitis. *Veterinary Medicine*, 43(12), 525–530.
- Horne MT. (1997). Technical aspects of the administration of vaccines. *Developments in Biological Standarization*, 90, 79–89.
- Hoss A., EC, Z., and R Zawatzky. (1989). Differential Expression of Interferon Alpha and Beta Induced with Newcastle Disease Virus in Mouse Macrophage Cultures. *Journal of General Virology*, 70, 575–589.
- Hu, S., Ma, H., Wu, Y., Liu, W., Wang, X., Liu, Y., and Liu, X. (2009). A vaccine candidate of attenuated genotype VII Newcastle disease virus generated by reverse genetics. *Vaccine*, 27(6), 904–910.
- Huovilainen, A., Ek-Kommonen, C., Manvell, R., and Kinnunen, L. (2001). Phylogenetic analysis of avian paramyxovirus 1 strains isolated in Finland. *Archives of Virology*, 146(9), 1775–1785.
- Ibrahim, A. L., Chulan, U., and Babjee, A. M. (1980). The immune response of chickens vaccinated against newcastle disease with live newcastle disease v4 vaccine. *Australian Vererinary Journal*, 56, 29–33.
- Ideris, A., Ibrahim, A. L., and Spradbrow, P. B. (1990). Vaccination of chickens against Newcastle disease with a food pellet vaccine. *Avian Pathology*, 19(2), 371–84.
- Jang, S. I., Lillehoj, H. S., Lee, S. H., Lee, K. W., Lillehoj, E. P., Bertrand, F., Dupuis, L., and Deville, S. (2011). Montanide™ ISA 71 VG adjuvant enhances antibody and cell-mediated immune responses to profilin subunit antigen vaccination and promotes protection against *Eimeria*

acervulina and *Eimeria tenella*. *Experimental Parasitology*, 127(1), 178–183.

Jansen, T., Hofmans, M. P. M., Theelen, M. J. G., Manders, F. G. A., and Schijns, V. E. J. C. (2007). Dose and timing requirements for immunogenicity of viral poultry vaccine antigen: investigations of emulsion-based depot function. *Avian Pathology*, 36(5), 361–365.

Jansen, T., Hofmans, M. P. M., Theelen, M. J. G., Manders, F., and Schijns, V. E. J. C. (2006). Structure- and oil type-based efficacy of emulsion adjuvants. *Vaccine*, 24(26), 5400–5.

Jansen, T., Hofmans, M. P. M., Theelen, M. J. G., and Schijns, V. E. J. C. (2005). Structure-activity relations of water-in-oil vaccine formulations and induced antigen-specific antibody responses. *Vaccine*, 23(8), 1053–1060.

Jeurissen, S. H., Boonstra-Blom, a G., Al-Garib, S. O., Hartog, L., and Koch, G. (2000). Defence mechanisms against viral infection in poultry: a review. *The Veterinary Quarterly*, 22(4), 204–8.

Johnson, M., Zaretskaya, I., Raytselis, Y., Merezuk, Y., McGinnis, S., and Madden, T. L. (2008). NCBI BLAST: a better web interface. *Nucleic Acids Research*, 36(Web Server issue), W5–9. <https://doi.org/10.1093/nar/gkn201>

Joshi, P. C., and Keane, T. C. (2010). Investigation of riboflavin sensitized degradation of purine and pyrimidine derivatives of DNA and RNA under UVA and UVB. *Biochemical and Biophysical Research Communications*, 400(4), 729–733.

Kaleta, E. F., and Baldauf, C. (1988). Newcastle disease in free-living and pet birds. In D. J. Alexander (Ed.), *Newcastle Disease* (Vol. 8, pp. 197–246). Boston, MA: Springer US.

Kapczynski, D. R., Afonso, C. L., and Miller, P. J. (2013). Immune responses of poultry to Newcastle disease virus. *Developmental and Comparative Immunology*, 41(3), 447–453.

Kapczynski, D. R., and King, D. J. (2005). Protection of chickens against overt clinical disease and determination of viral shedding following vaccination with commercially available Newcastle disease virus vaccines upon challenge with highly virulent virus from the California 2002 exotic Newcastl. *Vaccine*, 23(26), 3424–3433.

Khan, S. A., Khan, A. M., Karim, S., Kamal, M. A., Damanhour, G. A., and Mirza, Z. (2016). Panacea seed “*Nigella*”: A review focusing on

regenerative effects for gastric ailments. *Saudi Journal of Biological Sciences*, 23(4), 542–553.

- Kiarash, R. S. (2014). Characterization of Newcastle disease virus (NDV) isolated from NDV vaccinated broiler farms and investigation of vaccine efficacy against challenge with velogenic genotype VII NDV. Universiti Putra Malaysia.
- Kim, L. M., King, D. ., Suarez, D. ., Wong, C. ., and Afonso, C. . (2007). Characterization of class I newcastle disease virus isolates from Hong Kong live bird markets and detection using real-time reverse transcription-PCR. *Journal of Clinical Microbiology*, 45(4), 1310–1314.
- Kim, L. M., King, D. J., Curry, P. E., Suarez, D. L., Swayne, D. E., Stallknecht, D. E., Slemons, R. D., Pedersen, J. C., Senne, D. A., Winker, K., and Afonso, C. L. (2007). Phylogenetic Diversity among Low-Virulence Newcastle Disease Viruses from Waterfowl and Shorebirds and Comparison of Genotype Distributions to Those of Poultry-Origin Isolates. *Journal of Virology*, 81(22), 12641–12653.
- King, D. J. (1991). Evaluation of Different Methods of Inactivation of Newcastle Disease Virus and Avian Influenza Virus in Egg Fluids and Serum. *Avian Diseases*, 35(3), 505.
- Kou, Y. T., Chueh, L. L., and Wang, C. H. (1999). Restriction fragment length polymorphism analysis of the F gene of Newcastle disease viruses isolated from chickens and an owl in Taiwan. *The Journal of Veterinary Medical Science / the Japanese Society of Veterinary Science*, 61(11), 1191–5.
- Kraneveld, F. C. (1926). A poultry disease in the Dutch East Indies. *Nederlands-Indische Bladen Voor Diergeneeskunde*, 38, 448–450.
- Kwon, H. J., Cho, S. H., Ahn, Y. J., Seo, S. H., Choi, K. S., and Kim, S. J. (2003). Molecular epidemiology of Newcastle disease in Republic of Korea. *Veterinary Microbiology*, 95(1–2), 39–48.
- Lai, M. C., and Ibrahim, A. L. (1983). Scanning electron microscopy of tracheal epithelium of chickens infected with velogenic viscerotropic Newcastle disease virus. *Avian Diseases*, 27(2), 393–404.
- Laliberte, J. P., McGinnes, L. W., Peeples, M. E., and Morrison, T. G. (2006). Integrity of membrane lipid rafts is necessary for the ordered assembly and release of infectious Newcastle disease virus particles. *Journal of Virology*, 80(21), 10652–62.

- Lam, K. M., Kabbur, M. B., and Eiserich, J. P. (1996). Newcastle disease virus-induced functional impairments and biochemical changes in chicken heterophils. *Veterinary Immunology and Immunopathology*, 53(3-4), 313-327.
- Lamb, R. A., and Kolakofsky, D. (2001). Paramyxoviridae: The Viruses and Their Replication. In *Fields Virology* (Vol. 41, pp. 1449-1491). B. N. Field. USA,: Lippincott Williams & Wilkins.
- Lamb, R. A., and Parks, G. D. (2006). Paramyxoviridae: The viruses and their replication. In *Fields virology* (pp. 1449-1491). B. N. Field. USA, Lippincott Williams & Wilkins.
- Lascelles, A. K., Eagleson, G., Beh, K. J., and Watson, D. L. (1989). Significance of Freund's adjuvant/antigen injection granuloma in the maintenance of serum antibody response. *Veterinary Immunology and Immunopathology*, 22(1), 15-27.
- Le, L., Brasseur, R., Wemers, C., Meulemans, G., and Burny, A. (1988). Fusion (F) protein gene of newcastle disease virus: Sequence and hydrophobicity comparative analysis between virulent and avirulent strains. *Virus Genes*, 1(4), 333-350.
- Lee, N. H., Lee, J. A., Park, S. Y., Song, C. S., Choi, I. S., and Lee, J. B. (2012). A review of vaccine development and research for industry animals in Korea. *Clinical and Experimental Vaccine Research*, 1(1), 18-34.
- de Leeuw, O., and Peeters, B. (1999). Complete nucleotide sequence of Newcastle disease virus: evidence for the existence of a new genus within the subfamily Paramyxovirinae. *Journal of General Virology*, 80(1), 131-136.
- Liu, X. F., Wan, H. Q., Ni, X. X., Wu, Y. T., and Liu, W. B. (2003). Pathotypical and genotypical characterization of strains of Newcastle disease virus isolated from outbreaks in chicken and goose flocks in some regions of China during 1985-2001. *Archives of Virology*, 148, 1387-1403.
- Lomniczi, B., Wehmann, E., Herczeg, J., Ballagi-Pordány, A., Kaleta, E. F., Werner, O., Meulemans, G., Jorgensen, P. H., Manté, A. P., Gielkens, A. L. J., Capua, I., and Damoser, J. (1998). Newcastle disease outbreaks in recent years in Western Europe were caused by an old (VI) and a novel genotype (VII). *Archives of Virology*, 143(1), 49-64.
- Lyles, D. S., Kuzmin, I. V., and Rupprecht, C. E. (2013). Paramyxoviridae. In *Fields Virology* (6th ed., pp. 957-995). USA ,Lippincott williams & wilkins, a wolters kluwer business two commerce square.

- Mady, W. H., Arafa, A., Hussein, A. S., Aly, M. M., and Madbouly, H. M. (2013). Nigella Sativa Oil as an Immunostimulant Adjuvant in H5 Based DNA Vaccine of H5N1 Avian Influenza Virus.
- Mahan, S. M., Kumbula, D., Burrridge, M. J., and Barbett, A. F. (1998). The inactivated Cowdria ruminantium vaccine for heartwater protects against heterologous strains and against laboratory and field tick challenge. *Vaccine*, 16(11/12), 1203–1211.
- Majdalawieh, A. F., Hmaidan, R., and Carr, R. I. (2010). Nigella sativa modulates splenocyte proliferation, Th1/Th2 cytokine profile, macrophage function and NK anti-tumor activity. *Journal of Ethnopharmacology*, 131(2), 268–275.
- Manin, T. B., Shcherbakova, L. O., Bochkov, I. A., El'nikov, V. V., Pchelkina, I. P., Starov, S. K., and Drygin, V. V. (2002). Characteristics of Field Isolates of Newcastle Disease Virus Isolated in the Course of Outbreaks in the Poultry Plant in the Leningrad Region in 2000. *Vopr Virusol*, 47, 41–43.
- Marangon, S., and Busani, L. (2006). The use of vaccination in poultry production Herd immunity. *Revue Scientifique et Technique (International Office of Epizootics)*, 26(1), 265–274.
- Marino, O. C., and Hanson, R. P. (1987). Cellular and Humoral Response of in Ovo-Bursectomized Chickens to Experimental Challenge with Velogenic Newcastle Disease Virus. *Avian Diseases*, 31(2), 293.
- Martin, L. a, Smith, T. J., Obermoeller, D., Bruner, B., Kracklauer, M., and Dharmaraj, S. (2001). RNA Purification. *Molecular Biology Problem Solver: A Laboratory Guide*, 7, 197–224.
- Mase, M., Imai, K., Sanada, Y., Sanada, N., Yuasa, N., Imada, T., Tsukamoto, K., and Yamaguchi, S. (2002). Phylogenetic Analysis of Newcastle Disease Virus Genotypes Isolated in Japan. *Journal of Clinical Microbiology*, 40(10), 3826–3830.
- Mayo, M. A. (2002). A summary of taxonomic changes recently approved by ICTV. *Archives of Virology*, 147(8), 1655–1656.
- Mestecky, J., E.Lamm, M., Strober, W., Bienenstock, J., R.McGhee, J., and Mayer, L. (2007). *Mucosal Immunology* (third). Elsevier academic press.
- Miller, P. J., Afonso, C. L., El Attrache, J., Dorsey, K. M., Courtney, S. C., Guo, Z., and Kapczynski, D. R. (2013). Effects of Newcastle disease virus

vaccine antibodies on the shedding and transmission of challenge viruses. *Developmental and Comparative Immunology*, 41(4), 505–513.

Miller, P. J., Decanini, E. L., and Afonso, C. L. (2010). Newcastle disease: Evolution of genotypes and the related diagnostic challenges. *Infection, Genetics and Evolution*, 10(1), 26–35.

Miller, P. J., Estevez, C., Yu, Q., Suarez, D. L., and King, D. J. (2009). Comparison of viral shedding following vaccination with inactivated and live Newcastle disease vaccines formulated with wild-type and recombinant viruses. *Avian Diseases*, 53(1), 39–49.

Miller, R. L., and Plagemann, P. G. (1974). Effect of ultraviolet light on mengovirus: formation of uracil dimers, instability and degradation of capsid, and covalent linkage of protein to viral RNA. *Journal of Virology*, 13(3), 729–39.

Mirza, A. M., Sheehan, J. P., Hardy, L. W., Glickman, R. L., and Iorio, R. M. (1993). Structure and function of a membrane anchor-less form of the hemagglutinin-neuraminidase glycoprotein of newcastle disease virus. *Journal of Biological Chemistry*, 268(28), 21425–21431.

Mittal, S. K., HogenEsch, H., and Park, K. (2006). Vaccines and Other Immunological Products. In *Encyclopedia of pharmaceutical technology* (pp. 3908–3927).

Le Moignic, P. (1916). Les vaccines en emulsion dans les corps gras ou “lipo-vaccins”. *Comptes Rendus de La Societe de Biologie*, 79, 201–203.

Munir, M., Cortey, M., Abbas, M., Qureshi, Z. ul A., Afzal, F., Shabbir, M. Z., Khan, M. T., Ahmed, S., Ahmad, S., Baule, C., Ståhl, K., Zohari, S., and Berg, M. (2012). Biological characterization and phylogenetic analysis of a novel genetic group of Newcastle disease virus isolated from outbreaks in commercial poultry and from backyard poultry flocks in Pakistan. *Infection, Genetics and Evolution*, 12(5), 1010–1019.

Nagai, Y., and Klenk, H. D. (1977). Activation of precursors to both glycoproteins of Newcastle disease virus by proteolytic cleavage. *Virology*, 77, 125–134.

Nagai, Y., Klenk, H. D., and Rott, R. (1976). Proteolytic cleavage of the viral glycoproteins and its significance for the virulence of Newcastle disease virus. *Virology*, 72(2), 494–508.

Nagai, Y., Ogura, H., and Klenk, H. (1976). Studies on the assembly of the envelope of Newcastle disease virus. *Virology*, 69(2), 523–538.

- El Najjar, F., Schmitt, A. P., and Dutch, R. E. (2014). Paramyxovirus glycoprotein incorporation, assembly and budding: A three way dance for infectious particle production. *Viruses*, 6(8), 3019–3054.
- O'Hagan, D. T. (2000). Vaccine Adjuvants: Preparation Methods and Research Protocols. Humana Press, Totowa, New Jersey.
- OIE. (2008). Newcastle disease. In Manual of Diagnostic Tests and Vaccines for Terrestrial Animals (Mammals, Birds and Bees). (Vol. 1, pp. 576–589).
- OIE. (2012). Biotechnology in the diagnosis of infectious diseases. In *manual of diagnostic tests and vaccines for terrestrial animals (mammals, birds and bees)* (Seventh Ed, Vol. 2, pp. 1322–1337).
- OIE. (2012). Newcastle disease. In Manual of diagnostic tests and vaccines for terrestrial animals: mammals, birds and bees. In Biological Standards Commission (Vol. 1, pp. 555–574).
- OIE. (2013). Manual of Diagnostic Tests and Vaccines for Terrestrial Animals. *World Organisation for Animal Health*, 1185–1191.
- OIE. (2013). Protecting animals , preserving our future terrestrial animal health code (Twenty-sec, Vol. 2).
- Palmer, R. M., Ferrige, A. G., and Moncada, S. (1987). Nitric oxide release accounts for the biological activity of endothelium-derived relaxing factor. *Nature*, 327(6122), 524–526.
- Palya, V., Kiss, I., Tatár-Kis, T., Mató, T., Felföldi, B., and Gardin, Y. (2012). Advancement in Vaccination Against Newcastle Disease: Recombinant HVT NDV Provides High Clinical Protection and Reduces Challenge Virus Shedding with the Absence of Vaccine Reactions. *Avian Diseases*, 56(2), 282–287.
- Pantua, H. D., McGinnes, L. W., Peeples, M. E., and Morrison, T. G. (2006). Requirements for the assembly and release of Newcastle disease virus-like particles. *Journal of Virology*, 80(22), 11062–11073.
- Patil, P. K., Suryanarayana, V., Bist, P., Bayry, J., and Natarajan, C. (2002). Integrity of GH-loop of foot-and-mouth disease virus during virus inactivation: Detection by epitope specific antibodies. *Vaccine*, 20(7–8), 1163–1168.
- Perdiz, D., Gróf, P., Mezzina, M., Nikaido, O., Moustacchi, E., and Sage, E. (2000). Distribution and repair of bipyrimidine photoproducts in solar

UV-irradiated mammalian cells: Possible role of dewar photoproducts in solar mutagenesis. *Journal of Biological Chemistry*, 275(35), 26732–26742.

Peroulis-Kourtis, I., O'Riley, K., Grix, D., Condron, R. J., and Ainsworth, C. (2002). Molecular characterisation of Victorian Newcastle disease virus isolates from 1976 to 1999. *Australian Veterinary Journal*, 80(7), 422–424.

Perozo, F., Marcano, R., and Afonso, C. (2012). Biological and phylogenetic characterization of a genotype VII Newcastle disease virus from Venezuela: Efficacy of field vaccination. *Journal of Clinical Microbiology*, 50(4), 1204–1208.

Perozo, F., Villegas, P., Dolz, R., Afonso, C. L., and Purvis, L. B. (2008). The VG/GA strain of Newcastle disease virus: mucosal immunity, protection against lethal challenge and molecular analysis. *Avian Pathology*, 37(3), 237–245.

Pharmacopoeia, E. (2008). Newcastle disease vaccine (inactivated). In *European Pharmacopoeia* (6th ed., pp. 937–939). Boston: Academic Publishers.

Rasoli, M., Yeap, S. K., Tan, S. W., Moeini, H., Ideris, A., Bejo, M. H., Alitheen, N. B. M., Kaiser, P., and Omar, A. R. (2014). Alteration in lymphocyte responses, cytokine and chemokine profiles in chickens infected with genotype VII and VIII velogenic Newcastle disease virus. *Comparative Immunology, Microbiology and Infectious Diseases*, 37(1), 11–21.

Rauw, F., Gardin, Y., Palya, V., Anbari, S., Lemaire, S., Boschmans, M., van den Berg, T., and Lambrecht, B. (2010). Improved vaccination against Newcastle disease by an in ovo recombinant HVT-ND combined with an adjuvanted live vaccine at day-old. *Vaccine*, 28(3), 823–833.

Reynolds, D. L., and Maraqa, A. D. (2000). Protective immunity against Newcastle disease: the role of cell-mediated immunity. *Avian Diseases*, 44(1), 138–44.

Rifkin, D. B., and Quigley, J. P. (1974). Virus-induced modification of cellular membranes related to viral structure. *Annual Review of Microbiology*, 325–351.

Roohani, K., Tan, S. W., Yeap, S. K., Ideris, A., Bejo, M. H., and Omar, A. R. (2015). Characterisation of genotype VII Newcastle disease virus (NDV) isolated from NDV vaccinated chickens, and the efficacy of LaSota and recombinant genotype VII vaccines against challenge with velogenic NDV. *Journal of Veterinary Science*, 16(4), 447–457.

- Roy, P., Venugopalan, A. T., and Manvell, R. (2000). Characterization of Newcastle Disease Viruses Isolated from Chickens and Ducks in Tamilnadu, India. *Veterinary Research Communications*, 24(2), 135–142.
- Russell, P. H., Dwivedi, P. N., and Davison, T. F. (1997). The effects of cyclosporin A and cyclophosphamide on the populations of B and T cells and virus in the Harderian gland of chickens vaccinated with the Hitchner B1 strain of Newcastle disease virus. *Veterinary Immunology and Immunopathology*, 60(1–2), 171–185.
- Saif, Y. M. (1991). Immunosuppression induced by infectious bursal disease virus. *Veterinary Immunology and Immunopathology*, 30(1), 45–50.
- Sakaguchi, T., Toyoda, T., Gotoh, B., Inocencio, N. M., Kuma, K., Miyata, T., and Nagai, Y. (1989). Newcastle disease virus evolution. *Virology*, 169(2), 260–272.
- Salem, M. L. (2005). Immunomodulatory and therapeutic properties of the *Nigella sativa* L. seed. *International Immunopharmacology*, 5(13–14), 1749–1770.
- Samal, S. K., and Collins, P. L. (1996). RNA replication by a respiratory syncytial virus RNA analog does not obey the rule of six and retains a nonviral trinucleotide extension at the leader end. *Journal of Virology*, 70(8), 5075–82.
- Satharasinghe, D. A., Murulitharan, K., Tan, S. W., Yeap, S. K., Munir, M., Ideris, A., and Omar, A. R. (2016). Detection of inter-lineage natural recombination in avian paramyxovirus serotype 1 using simplified deep sequencing platform. *Frontiers in Microbiology*, 7, 1–14.
- Scalzo, A. A., Elliott, S. L., Cox, J., Gardner, J. O. Y., Moss, D. J., and Suhrbier, A. (1995). Induction of Protective Cytotoxic T Cells to Murine Cytomegalovirus by Using a Nonapeptide and a Human-Compatible Adjuvant (Montanide ISA 720), 69(2), 1306–1309.
- Schat, K. A., Kaspers, B., and Kaiser, P. (2014). Autoimmune Diseases of Poultry. In *Avian Immunology* (Second edi, pp. 315–332). Elsevier Ltd, USA.
- Scheid, A., and Choppin, P. W. (1973). Isolation and Purification of the Envelope Proteins of Newcastle Disease Virus. *Journal of Virology*, 11(2), 263–271.
- Schijns, V. E. (2000). Immunological concepts of vaccine adjuvant activity. *Current Opinion in Immunology*, 12(4), 456–463.

- Schlegel, A., Immelmann, A., and Kempf, C. (2001). in the Presence of Guanidine Hydrochloride. *Transfusion*, 41, 382–389.
- Schuy, W., Garten, W., Linder, D., and Klenk, H. D. (1984). The carboxyterminus of the hemagglutinin-neuraminidase of Newcastle disease virus is exposed at the surface of the viral envelope. *Virus Research*, 1(5), 415–426.
- Seal, B. S., King, D. J., and Sellers, H. S. (2000). The avian response to Newcastle disease virus. *Developmental and Comparative Immunology*, 24(2–3), 257–268.
- Senne, D. A., King, D. J., and Kapczynski, D. R. (2004). Control of Newcastle disease by vaccination (Conference Paper). *Developments in Biologicals*, 119, 165–170.
- Shams, H. (2005). Recent developments in veterinary vaccinology. *The Veterinary Journal*, 170, 289–299.
- Sharma, J. M. (1999). Introduction to poultry vaccines and immunity. *Advances in Veterinary Medicine*, 41, 481–494.
- Sharma, N. K., Ahirwar, D., Jhade, D., and Gupta, S. (2009). Medicinal and Phamacological Potential of Nigella sativa : A Review. *Ethnobotanical Review*, 13, 1–8.
- Sick, C., Schneider, K., Staeheli, P., and Weining, K. C. (2000). Novel Chicken CXC and CC Chemokines. *Cytokine*, 12(3), 181–186.
- Sick, C., Schultz, U., Münster, U., Meier, J., Kaspers, B., and Staeheli, P. (1998). Promoter structures and differential responses to viral and nonviral inducers of chicken type I interferon genes. *Journal of Biological Chemistry*, 273(16), 9749–9754.
- Singh, M., and O'Hagan, D. T. (2002). Recent advances in vaccine adjuvants. *Pharmaceutical Research*, 19(6), 715–28.
- Sinha, R. P., and Häder, D. P. (2002). UV-induced DNA damage and repair: a review. *Photochemical & Photobiological Sciences*, 1(4), 225–236.
- Spilki, F. R., Almeida, R. S. De, and Arns, C. W. (2010). Antibody responses in mice after immunization with inactivated bovine respiratory syncytial virus using different adjuvants. *Ciência Rural, Santa Maria*, 40, 2332–2337.

- Spradbrow, A. P. B., Ibrahim, A. L., and Kim, S. J. (1978). Use of an Avirulent Australian Strain of Newcastle Disease Virus as a Vaccine. *Avian Diseases*, 22(2), 329-335.
- Spradbrow, P. B., MacKenzie, M., and Grimes, S. E. (1995). Recent isolates of Newcastle disease virus in Australia. *Veterinary Microbiology*, 46(1-3), 21-28.
- Steward, M., Vipond, I. B., Millar, N. S., and Emmerson, P. T. (1993). RNA editing in Newcastle disease virus. *Journal of General Virology*, 74(12), 2539-2547.
- Stone, H. D. (1993). Efficacy of experimental animal and vegetable oil-emulsion vaccines for Newcastle disease and avian influenza. *Avian Diseases*, 37(2), 399-405.
- Stone, H. D. (1997). Newcastle disease oil emulsion vaccines prepared with animal, vegetable, and synthetic oils. *Avian Diseases*, 41(3), 591-7.
- Stone, H. D., Brugh, M., and Beard, C. W. (1981). Comparison of Three Experimental Inactivated Oil-Emulsion Newcastle Disease Vaccines. *Avian Diseases*, 25(4), 1070-1076.
- Stone, H. D., Brugh, M., Hopkins, S. R., Yoder, H. W., and Beard, C. W. (1978). Preparation of inactivated oil-emulsion vaccines with avian viral or Mycoplasma antigens. *Avian Diseases*, 22(4), 666-674.
- Sun, A. H. ling, Wang, Y. feng, Tong, G. zhi, Zhang, P. jun, Miao, D. yuan, Zhi, D., Wang, M., Wang, M., Sun, H. ling, Wang, A. B. C. Y. feng, Tong, B. G. zhi, Zhang, A. B. D. P. jun, Miao, C. D. yuan, Zhi, C. H. dong, Wang, M., and B, M. W. (2008). Protection of Chickens from Newcastle Disease and Infectious Laryngotracheitis with a Recombinant Fowlpox Virus Co-Expressing the F , HN Genes of Newcastle Disease Virus and gB Gene of Infectious Laryngotracheitis Virus. *Avian Diseases*, 52, 111-117.
- Susta, L., Miller, P. J., Afonso, C. L., and Brown, C. C. (2011). Clinicopathological characterization in poultry of three strains of Newcastle disease virus isolated from recent outbreaks. *Veterinary Pathology*, 48(2), 349-60.
- Swayne, D. E. (2006). Principles for vaccine protection in chickens and domestic waterfowl against avian influenza: Emphasis on Asian H5N1 high pathogenicity avian influenza. *Annals of the New York Academy of Sciences*, 1081, 174-181.

- Swayne, D. E., Glisson, J. R., McDougald, L. R., Nolan, L. K., Suarez, D. L., and Nair, V. (2013). *Diseases of Poultry* (13th ed.). John Wiley & Sons, Inc.
- Swayne, D. E., and King, D. J. (2003). Avian influenza and Newcastle disease. *Journal of the American Veterinary Medical Association*, 222(11), 1534–1540.
- Swayne, D. E., and King, D. J. (2003). Zoonosis Update Avian Influenza and Newcastle disease. *Journal of the American Veterinary Medical Association*, Vol. 222,(1534–1540).
- Szretter, K. J., Balish, A. L., and Katz, J. M. (2006). Influenza: propagation, quantification, and storage. In *Current Protocols in Microbiology* (pp. 1–22).
- Takada, A., and Kida, H. (1996). Protective immune response of chickens against Newcastle disease, induced by the intranasal vaccination with inactivated virus. *Veterinary Microbiology*, 50(1–2), 17–25.
- Takimoto, T., and Portner, A. (2004). Molecular mechanism of paramyxovirus budding. *Virus Research*, 106, 133–145.
- Tamam, S. M., Hussein, A. S., Arafa, A. M., and Madbouly, H. M. (2015). Preparation and evaluation of inactivated avian metapneumovirus vaccine from recently isolated Egyptian strain. *Journal of Applied Poultry Research*, 24(2), 168–176.
- Tamura, K., Peterson, D., Peterson, N., Stecher, G., Nei, M., and Kumar, S. (2011). MEGA5: Molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Molecular Biology and Evolution*, 28(10), 2731–2739.
- Tan, S. W., Ideris, A., Omar, A. R., Yusoff, K., and Hair-Bejo, M. (2010). Sequence and phylogenetic analysis of Newcastle disease virus genotypes isolated in Malaysia between 2004 and 2005. *Archives of Virology*, 155(1), 63–70.
- Tano, Y., Shimizu, H., Martin, J., Nishimura, Y., Simizu, B., and Miyamura, T. (2007). Antigenic characterization of a formalin-inactivated poliovirus vaccine derived from live-attenuated Sabin strains. *Vaccine*, 25(41), 7041–7046.
- Terregino, C., Cattoli, G., Grossele, B., Bertoli, E., Tisato, E., and Capua, I. (2003). Characterization of Newcastle disease virus isolates obtained from Eurasian collared doves (*Streptopelia decaocto*) in Italy. *Avian Pathology*, 32(1), 63–8.

- Tirumurugaan, K. G., Vinupriya, M. K., Vijayarani, K., and Kumanan, K. (2011). Analysis of the fusion protein cleavage site of newcastle disease virus isolates from India reveals preliminary evidence for the existence of II, VI and VII genotypes. *Indian Journal of Virology*, 22(2), 131-137.
- Totté, P., Esteves, I., Gunter, N., Martinez, D., and Bensaida, A. (2002). Evaluation of several flow cytometric assays for the analysis of T-cell responses in goats. *Cytometry*, 49(2), 49-55.
- Tsai, H. J., Chang, K. H., Tseng, C. H., Frost, K. M., Manvell, R. J., and Alexander, D. J. (2004). Antigenic and genotypical characterization of Newcastle disease viruses isolated in Taiwan between 1969 and 1996. *Veterinary Microbiology*, 104(1-2), 19-30.
- Uittenbogaard, J. P., Zomer, B., Hoogerhout, P., and Metz, B. (2011). Reactions of β -propiolactone with nucleobase analogues, nucleosides, and peptides: Implications for the inactivation of viruses. *Journal of Biological Chemistry*, 286(42), 36198-36214.
- Umali, D. V., Ito, H., Suzuki, T., Shiota, K., Katoh, H., and Ito, T. (2013). Molecular epidemiology of Newcastle disease virus isolates from vaccinated commercial poultry farms in non-epidemic areas of Japan. *Virology Journal*, 10, 330.
- Umar, S., Rehman, A., Younus, M., Qamar-Un-Nisa, Ali, A., Shahzad, M., Shah, M. A. A., Munir, M. T., Aslam, H. Bin, and Yaqoob, M. (2015). Effects of *Nigella sativa* on immune responses and pathogenesis of avian influenza (H9N2) virus in turkeys. *Journal of Applied Poultry Research*, 25(1), 95-103.
- Villar, E., and Barroso, I. M. (2006). Role of sialic acid-containing molecules in paramyxovirus entry into the host cell: A minireview. *Glycoconjugate Journal*, 23(1-2), 5-17.
- Wanasawaeng, W., Tawatsin, A., Sasipreeyajan, J., Poomvises, P., and Chansiripornchai, N. (2009). Development of Inactivated Newcastle Disease Vaccine using Palm Oil as an Adjuvant. *Thai Journal of Veterinary Medicine*, 39(1), 9-16.
- Wang, J. yu, Liu, W. hua, Ren, J. juan, Tang, P., Wu, N., and Liu, H. jen. (2013). Complete genome sequence of a newly emerging Newcastle disease virus. *Genome Announcements*, 1(3), 3-4.
- Wang, Z., Liu, H., Xu, J., Bao, J., Zheng, D., Sun, C., Wei, R., Song, C., and Chen, J. (2006). Genotyping of Newcastle disease viruses isolated from

2002 to 2004 in China. *Annals of the New York Academy of Sciences*, 1081, 228–239.

Wehmann, E., Herczeg, J., Ballagi-pordny, A., and Lomniczi, B. (1997). Rapid identification of Newcastle disease virus vaccine strains LaSota and B-1 by restriction site analysis of their matrix gene. *Vaccine*, 15(12/13), 1430–1433.

Wehmann, E., Herczeg, J., Tanyi, J., Wehmann, O., Nagy, E., and Lomniczi, B. (1999). Lentogenic field isolates of Newcastle disease virus isolated in Canada and Hungary are identical with the vaccine type used in the region. *Avian Pathology*, 28(1), 6–12.

Westbury, H. A., Parsons, G., and Allan, W. H. (1984). Comparison of the immunogenicity of Newcastle disease virus strains V4, Hitchner B1 and La Sota in chickens. 2. Tests in chickens with maternal antibody to the virus. *Australian Veterinary Journal*, 61(1), 10–3.

Wilden, H., Schirmacher, V., and Fournier, P. (2011). Important role of interferon regulatory factor (IRF) -3 in the interferon response of mouse macrophages upon infection by Newcastle disease virus. *International Journal of Oncology*, 39, 493–504.

Willey, J., Sherwood, L., and Woolverton, C. (2008). Recombinant DNA Technology. In *Prescott, Harley, and Klein's Microbiology* (7th ed., pp. 357–382). New York: McGraw-Hill.

Xiao, S., Paldurai, A., Nayak, B., Samuel, A., Bharoto, E. E., Prajitno, T. Y., Collins, P. L., and Samal, S. K. (2012). Complete Genome Sequences of Newcastle Disease Virus Strains Circulating in Chicken Populations of Indonesia. *Journal of Virology*, 86(10), 5969–5970.

Xie, Z., Xie, L., Chen, A., Liu, J., Pang, Y., Deng, X., Xie, Z., and Fan, Q. (2012). Complete genome sequence analysis of a Newcastle disease virus isolated from a wild egret. *Journal of Virology*, 86(24), 13854–13855.

Yamanaka, M., Okabe, T., Nakai, M., and Goto, N. (1993). Local pathological reactions and immune response of chickens to ISA-70 and other adjuvants containing Newcastle disease virus antigen. *Avian Diseases*, 37(2), 459–466.

Yan, Y. (2008). Role of Noncoding Regions in Newcastle Disease Virus Replication and pathogenesis. University of Maryland.

Yang, C. yao, Shieh, H. K., Lin, Y. L., and Chang, P. chun. (1999). Newcastle Disease Virus Isolated from Recent Outbreaks in Taiwan

Phylogenetically Related to Viruses (Genotype VII) from Recent Outbreaks in Western Europe. *Avian Diseases*, 43(1), 125.

Yi, J., Liu, C., Chen, B., and Wu, S. (2011). Molecular Characterization of a Virulent Genotype VII_d Strain of Newcastle Disease Virus from Farmed Chickens in Shanghai Molecular Characterization of a Virulent Genotype VII_d Strain of Newcastle Disease Virus from Farmed Chickens. *Avian Diseases*, 55(2), 279–284.

Yohannes, T., Sharma, A. K., Singh, S. D., and Goswami, T. K. (2012). Immunopathological effects of experimental T-2 mycotoxocosis in broiler chicken co-infected with infectious bronchitis virus (IBV). *Veterinary Immunology and Immunopathology*, 146(3–4), 245–253.

Yu, L., Wang, Z., Jiang, Y., Chang, L., and Kwang, J. (2001). Characterization of Newly Emerging Newcastle Disease Virus Isolates from the People's Republic of China and Taiwan. *Journal of Clinical Microbiology*, 39(10), 3512–3519.

Yuan, P., Paterson, R. G., Leser, G. P., Lamb, R. A., and Jardetzky, T. S. (2012). Structure of the Ulster Strain Newcastle Disease Virus Hemagglutinin-Neuraminidase Reveals Auto-Inhibitory Interactions Associated with Low Virulence. *PLoS Pathogens*, 8(8), 1–15.

Yusoff, K., and Tan, W. S. (2001). Newcastle disease virus: macromolecules and opportunities. *Avian Pathology*, 30(5), 439–455.

Zawatzky, R., Wurmbaek, H., Falk, W., and Homfeld, A. (1991). Endogenous Interferon Specifically Regulates Newcastle Disease Virus-Induced Cytokine Gene Expression in Mouse Macrophages. *Journal of Virology*, 65(9), 4839–4846.

Zhang, W., and Sun, Z. (2008). Random local neighbor joining: A new method for reconstructing phylogenetic trees. *Molecular Phylogenetics and Evolution*, 47(1), 117–128.

Zhang, Z. W., Wang, Q. H., Zhang, J. L., Li, S., Wang, X. L., and Xu, S. W. (2012). Effects of oxidative stress on immunosuppression induced by selenium deficiency in chickens. *Biological Trace Element Research*, 149(3), 352–361.