



UNIVERSITI PUTRA MALAYSIA

***ANTI-OBESITY AND ANTI-OXIDATIVE EFFECTS OF METHANOLIC
Albizia myriophylla (L. BENTH) BARK EXTRACT IN OBESE MICE***

AZMAH SA'AT

FPV 2017 12



**ANTI-OBESITY AND ANTI-OXIDATIVE EFFECTS OF METHANOLIC
Albizia myriophylla (L. BENTH) BARK EXTRACT IN OBESE MICE**

By

AZMAH SA'AT

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

July 2017

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

**ANTI-OBESITY AND ANTI-OXIDATIVE EFFECTS OF METHANOLIC
Albizia myriophylla (L. BENTH) BARK EXTRACT IN OBESE MICE**

By

AZMAH SA'AT

July 2017

Chairman : Goh Yong Meng, DVM, PhD
Faculty : Veterinary Medicine

Globally and in Malaysia, obesity causes early morbidity and mortality and is costly to treat. The antioxidant activities in herbal plants have been suggested as one of the mechanisms working against obesity. Hence, this study examined the antioxidant activity of *Albizia myriophylla* bark or known locally as tebu gajah. Its effect on anti-obesity via *in vivo* test in high-fat diet-induced obesity in mice consisting of the normal control group (NC), the high-fat diet control group (HFDC), the high fat diet group treated with *Albizia myriophylla* methanol extract at 10 mg/kg, 20 mg/kg and 30 mg/kg for 5 weeks. Antioxidant activity of the methanol extract and its derived fractions namely hexane, chloroform, ethyl acetate, butanol and a residual aqueous fraction of the bark of ABZ was assessed. In (1,1-diphenyl-2-picrylhydrazyl) radical scavenging test, (2,2'-azinobis 3-ethyl-6-sulfonic acid) radical scavenging test and reducing activity on ferrous iron test, the total antioxidant capacity was found to be varied in different fractions. The IC₅₀ calculated value of the three assays showed that the methanolic extract of ABZ bark had the lowest IC₅₀ value for each assay, compared to the other extracts signifying highest antioxidant activity and was selected for use in the anti-obesity study. The body weight of mice, adipose cellularity study, and levels of cholesterol, triglyceride, low-density lipoprotein (LDL), the malondialdehyde levels in the muscles of the obese mice, the serum aspartate transaminase (AST) and serum alanine transaminase (ALT) were analysed. Results showed significant weight loss with significant reduction in the number and size of adipose cells in the mice treated with 20 mg/kg and 30 mg/kg of methanolic extract of ABZ bark. In addition, there was a significant reduction of blood cholesterol, triglyceride and LDL following treatment with 20 mg/kg and 30 mg/kg of methanolic extract of ABZ bark, compared to HFDC. Furthermore, the significant reduction of AST, ALT and malondialdehyde in the 20 mg/kg and 30 mg/kg treated group, compared with HFDC, suggest the safety of methanolic extract of ABZ bark in a mice model. The regulatory effects of methanolic extract of ABZ bark on genes including hormone sensitive lipase (HSL), peroxisome proliferator-activated receptors α and γ (PPAR α and PPAR γ), sterol regulatory element-binding protein-2 (SREBP2) and Stearoyl-Coenzyme A-Desaturase (SCD)

gene expression and adipocytokines including tumor necrosis factor- α (TNF α), interleukin-6 (IL-6), adiponectin and leptin involved in lipid metabolism were also studied. The obese mice treated with the 30 mg/kg of methanolic extract of ABZ showed significant up-regulated of PPAR α and significant down-regulated activity of PPAR γ , SREBP2, and SCD gene expression when compared to the HFDC group. In addition, the mice treated with 20mg/kg and 30 mg/kg of ABZ methanolic extract showed a significant reduction of TNF α and IL-6 and significant increase of adiponectin and leptin compared to the HFDC. In conclusion, the antioxidant properties of 20mg/kg to 30 mg/kg methanolic extract of ABZ bark is able to avert obesity by regulation of the genes and adipocytokines involved in lipid metabolism.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**AKTIVITI ANTI-OBESITI DAN ANTIOKSIDAN EKSTRAK
METANOL KULIT *Albizia myriophylla* (L. BENTH) PADA TIKUS OBES**

Oleh

AZMAH SA'AT

Julai 2017

Pengerusi : Goh Yong Meng, DVM, PhD
Fakulti : Perubatan Veterinar

Di peringkat global dan di Malaysia, obesiti merupakan antara penyebab morbiditi dan kematian di peringkat awal berserta kos rawatan yang mahal. Aktiviti antioksidan dalam tumbuhan herba telah dicadangkan sebagai salah satu daripada mekanisme untuk mengatasi masalah obesiti. Oleh itu, kajian ini mengkaji aktiviti antioksidan *Albizia myriophylla* (ABZ) yang lebih dikenali sebagai tebu gajah di Malaysia. Kesannya kepada anti-kegemukan melalui ujian *in vivo* dalam obesiti yang dirangsang melalui diet lemak tinggi di kalangan tikus yang terdiri dari kumpulan kawalan normal (NC), kumpulan diet lemak tinggi (HFDC), kumpulan diet lemak tinggi yang di rawat dengan ekstrak metanol kulit ABZ pada aras 10 mg/kg, 20 mg/kg dan 30 mg/kg turut dikaji selama 5 minggu. Aktiviti antioksidan ekstrak metanol dan pecahan ekstrak yang diperolehi daripadanya seperti hexana, klorofom, etil asetat, butanol dan sisa pecahan akueus daripada kulit ABZ telah dilakukan dengan menggunakan analisis kimia secara *in vitro*. Dalam ujian memerangkap radikal DPPH (1,1-diphenyl-2-picrylhydrazyl), ujian memerangkap radikal ABTS (2,2'-azinobis 3-ethyl-benzothiazoline-6-sulfonic acid) dan ujian pengurangan aktiviti terhadap ion ferus (FRAP), jumlah keupayaan antioksidan didapati berbeza-beza dalam fraksi yang berlainan. Nilai IC₅₀ daripada tiga kajian ini menunjukkan bahawa ekstrak metanolik kulit ABZ menunjukkan nilai IC₅₀ yang terendah untuk ketiga-tiga ujian, dibandingkan dengan ekstrak yang lain, serta menunjukkan aktiviti antioksidan yang tertinggi. Dapatan-dapatan di atas menunjukkan bahawa ekstrak metanolik kulit ABZ adalah sumber antioksidan semulajadi yang berpotensi dan telah dipilih untuk digunakan dalam kajian anti-obesiti. Penilaian berat badan tikus, kajian sellulariti adipos, dan kadar ketinggian kolesterol, trigliserida, lipoprotein berketumpatan rendah (LDL), aras malondialdehyde dalam otot tikus gemuk, serum aspartate transaminase (AST) dan serum alanine transaminase (ALT) turut dikaji. Hasilnya telah menunjukkan kehilangan berat yang signifikan berserta penurunan ketara dalam bilangan dan saiz sel-sel adipos pada tikus-tikus yang telah diberikan 20 mg/kg dan 30 mg/kg ekstrak metanolik kulit ABZ. Tambahan pula, berikutan rawatan 20 mg/kg dan 30 mg/kg ekstrak metanol dahan ABZ, terdapat pengurangan kolesterol, TG dan LDL yang

ketara dilihat dalam kumpulan-kumpulan ini. Di samping itu, pengurangan AST, ALT dan malondialdehyde yang ketara dalam kumpulan 20 mg/kg dan 30 mg/kg dibandingkan dengan kumpulan HFDC, menggambarkan ekstrak metanolik kulit ABZ adalah selamat digunakan dalam model tikus. Kesan pengawalan ekstrak metanol ABZ pada gen yang melibatkan lipase hormon sensitif (HSL), peroksisom reseptor proliferasi-diaktifkan α dan γ (α PPAR dan PPAR γ), sterol peraturan unsur mengikat protein-2 (SREBP2) dan stearoyl-Koenzim A-desaturase (SCD) dengan adipositokin termasuk tumor nekrosis faktor- α (TNF α), interleukin-6 (IL-6), adiponektin dan leptin yang terlibat dalam metabolisme lemak juga telah dikaji. Tikus gemuk yang dirawat dengan 30 mg/kg ekstrak metanol ABZ menunjukkan aktiviti yang ketara kepada kenaikan kadar aktiviti PPAR α dan penurunan kadar aktiviti PPAR γ , SREBP2, dan SCD ekspresi gen apabila dibandingkan dengan kumpulan HFDC. Di samping itu, kumpulan yang dirawat dengan 20 mg/kg dan 30 mg/kg menunjukkan pengurangan ketara TNF α dan IL-6 serta menunjukkan peningkatan ketara adiponektin dan leptin berbanding dengan kumpulan HFDC. Kesimpulannya, sifat-sifat antioksidan 20 mg/kg kepada 30 mg/kg ekstrak metanol kulit ABZ mampu mengelakkan kegemukan dengan pengawalan gen dan adipositokin yang terlibat dalam metabolisme lemak.

ACKNOWLEDGEMENTS

In the name of Allah, the Most Benevolent and the Most Merciful. Alhamdulillah, praise to Almighty of Allah for giving me the strength to complete my research and this thesis. First of all, I wish to express my deepest thanks and appreciation to my supervisor Assoc. Prof. Dr. Goh Yong Meng and co-supervisor Dr. Mahdi Ebrahimi for their excellent supervision, guidance and patience. I am also grateful to my co-supervisors Prof. Dr. Noordin Mohamed Mustapha and Dr. Siti Khadijah Adam for their sound advice during the research.

I would like to thank Dr. Nurawati Syakroni for providing assistance and advice in running the research and Prof. Dr. Rozita Rosli for permission on using the bark of *Albizia myriophylla* for my current study.

Heartful thanks are extended to Dr. Mohammad Fasakh Jahroumi for his technical assistance and support. My gratitude extends to my colleagues, Tarlan and Samiah for their friendship, encouragement, useful discussion and much appreciated help.

Finally, I am deeply grateful to my understanding husband Shah Nahar Ismail, my supportive parents, Sa'at Mohd. Yasin and Misah Haron and last but not least, my loving kids which include Amirah, Rais, Nisa, Idris and Ishak for their cooperation and patience in making this thesis a reality for us, especially so, for my son Rais, who has Dandy Walker Variant Syndrome. He has been my inspiration in completing this thesis.

I certify that a Thesis Examination Committee has met on 20 July 2017 to conduct the final examination of Azmah binti Sa'at on her thesis entitled "Anti-Obesity and Anti-Oxidative Effects of Methanolic *Albizia myriophylla* (L.Benth) Bark Extract in Obese Mice" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Arifah binti Abdul Kadir, PhD

Associate Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Chairman)

Lokman Hakim bin Idris, PhD

Senior Lecturer
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Internal Examiner)

Hazilawati binti Hamzah, PhD

Associate Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Internal Examiner)

Penbe Sema Temizer Ozan, PhD

Professor
Firat University
Turkey
(External Examiner)



NOR AINI AB. SHUKOR, PhD

Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 28 September 2017

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Goh Yong Meng, DVM, PhD

Associate Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Chairman)

Mahdi Ebrahimi, PhD

Lecturer
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

Noordin Mohamed Mustapha, DVM, PhD

Professor
Faculty of Veterinary Medicine,
Universiti Putra Malaysia
(Member)

Siti Khadijah Adam, PhD

Senior Lecturer
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Azmah binti Sa'at (GS36085)

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:

Name of

Chairman of

Supervisory Committee:



Assoc. Prof. Dr. Goh Yong Meng

Signature:

Name of

Member of

Supervisory Committee:




Dr. Mahdi Ebrahimi

Signature:

Name of

Member of

Supervisory Committee:



Prof. Dr. Noordin Mohamed Mustapha

Signature:

Name of

Member of

Supervisory Committee:



Dr. Siti Khadijah Adam

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xv
LIST OF FIGURES	xvi
LIST OF ABBREVIATIONS	xx
 CHAPTER	
 1 INTRODUCTION	 1
1.1 Significance of the Study	2
1.2 General Objective	3
1.3 Specific Aims	3
 2 LITERATURE REVIEW	 4
2.1 Naturopathy and Obesity	4
2.2 Obesity Definition and Related Diseases	6
2.3 Epidemiology of Obesity in Malaysia	6
2.4 Predisposing Factors of Obesity	7
2.4.1 Age	7
2.4.2 Genetic	8
2.4.3 Medications Induce Obesity	8
2.4.4 Environmental Factors	8
2.4.5 Endocrinological Causes	9
2.4.6 Psychological Causes	10
2.5 Anti-obesity Drugs	10
2.6 Antioxidant Effects of Natural Products	10
2.7 Natural Products for Obesity Treatment	11
2.8 Mechanism of Anti-obesity Effects of Polyphenols and Catechins	12
2.8.1 Polyphenols	12
2.8.2 Catechin	12
2.9 <i>Albizia myriophylla</i>	13
2.10 Toxicity Studies of Albizia Species	14
2.11 Anti-obesity Bioactive Compounds from ABZ	14
2.11.1 Saponins	14
2.11.2 Spermine-type alkaloids	16
2.11.3 Flavonol	16
2.11.4 Phenolic Acid	17
2.12 Adipose Tissue as an Endocrine Organ	19
2.13 Adipocytokines	19
2.13.1 Adiponectin	20
2.13.2 Tumor Necrosis Factor Alpha (TNF α)	20

2.13.3	Interleukin-6 (IL-6)	20
2.13.4	Leptin	21
2.14	Lipogenic Gene Expression	21
2.15	House Keeping Gene (HKG)	22
2.16	β Actin	22
2.17	Peroxisome Proliferator-activated Receptors	22
2.18	Peroxisome Proliferator-activated Receptors Gamma (PPAR γ)	23
2.19	Peroxisome Proliferator-activated Receptors Alpha (PPAR α)	24
2.20	Hormone-sensitive Lipase (HSL)	24
2.21	Sterol Regulatory Element-Binding Protein 2 (SREBP2)	25
2.22	Stearoyl-Coenzyme A-Desaturase (SCD)	25
2.23	Oxidative Stress in the Development of Obesity	26
2.24	Relationship Between Adipose Tissue and Fatty Acids	27
2.25	Lipid Peroxidation and Obesity (Malondialdehyde)	27
2.26	Obesity Induced Inflammatory Changes in Adipose Tissue	28
2.27	Obesity and Adipose Tissue Cellularity	28
2.28	Mice Obesity Model	28
2.29	Summary	29
3.	PREPARATION OF VARIOUS FRACTIONS FROM <i>ALBIZIA MYRIOPHYLLA</i> (L. Benth) BARK AND DETERMINATION OF ITS PHENOLIC COMPOUND AND ITS ANTIOXIDANT ACTIVITIES	30
3.1	Introduction	30
3.2	Materials and Methods	30
3.2.1	Sample Preparation	30
3.2.1.1	Collection of Plant Material	30
3.2.2	The <i>Albizia myriophylla</i> Bark Extracts	31
3.2.2.1	Preparation of Dried Barks of ABZ	31
3.2.2.3	Preparation of Aqueous ABZ Bark Extract	31
3.2.3	Preparation of Methanol ABZ Bark Extract	31
3.2.4	Preparation of Hexane, Chloroform, Ethyl Acetate and Butanol ABZ Bark Extract	31
3.2.5	Determination of Total Phenolic Contents of ABZ Bark Extracts	33
3.2.6	Determination of Total Flavonoid Contents of ABZ Bark Extracts	33
3.2.7	Determination of Total Saponin Contents of ABZ Bark Extracts	33
3.2.8	Antioxidants Activity Determinations	34
3.2.8.1	Total Antioxidants Assay	34
3.2.8.2	DPPH Scavenging Activity	34
3.2.8.3	FRAP Assay	34
3.2.9	Statistical Analysis	35
3.3	Results	35
3.3.1	The Total Quantity of Aqueous, Methanol, Hexane, Ethyl Acetate, Chloroform and Butanol Bark Extracts of ABZ	35
3.3.2	The Total Phenolic Acid Content of ABZ Bark	35

	Extracts	
3.3.3	The Total Flavonoid Content of ABZ Bark Extracts	36
3.3.4	The Total Saponin Content of ABZ Bark Extracts	37
3.3.5	ABTS Assay if ABZ Bark Extracts ABTS Assay of ABZ Bark Extracts	38
3.3.6	DPPH Radical Scavenging Activity of ABZ Bark Extracts	39
3.3.7	FRAP Assay of ABZ Bark Extracts	40
3.3.8	The IC ₅₀ Values of ABZ Bark Extracts Based on ABTS, DPPH and FRAP Assay of ABZ Bark Extracts	41
3.4	Discussion	42
3.4.1	The Phenolics, Saponins, and Flavonoids Found in Various Extracts of ABZ Bark	42
3.4.2	The Antioxidant Activity and IC ₅₀ of Various Extracts of ABZ Bark	45
3.5	Conclusion	46
4.	ANTI-OBESITY EFFECTS OF <i>ALBIZIA MYRIOPHYLLA</i> (L. Benth) METHANOLIC BARK EXTRACT IN OBESE ICR MICE	47
4.1	Introduction	47
4.2	Materials and Methods	48
4.2.1	Animal Model	48
4.2.1.1	Preparation of Animal Model	48
4.2.2	Creation of Obese Mice	48
4.2.3	Study Design for the Treatment Group	48
4.2.4	Biochemical Analysis	50
4.2.4.1	Serum AST, ALT, Triglyceride, HDL and LDL Level	50
4.2.5	Cholesterol Level in Muscle	50
4.2.6	Malondialdehyde Antioxidant Assay	50
4.2.6.1	Reagent Preparation	50
4.2.6.2	Malondialdehyde Assay Procedure	51
4.2.7	Fatty Acid Profile Determination from Muscle	51
4.2.7.1	Materials and Chemicals	51
4.2.7.2	Total Lipid Extraction	51
4.2.7.3	Gas Liquid Chromatography	52
4.2.8	Histology of Adipose Cells and Liver	53
4.2.8.1	Haematoxylin and Eosin (H & E) Stain of Adipose Tissue and Liver	53
4.2.8.2	Quantitative Histology of the Adipose Tissue Including the Number, Diameter and Area	54
4.2.8.3	Hepatic Lesion Scoring Based on the Haematoxylin and Eosin staining of the Liver Cells	54
4.2.9	Statistical Analysis	54
4.3	Results	54
4.3.1	Effect of Methanolic Extract of ABZ Bark on Body	54

	Weight of High-fat Diet-Induced Obesity in Mice	
4.3.2	Effect of Methanolic Extract of ABZ Bark on Weight of Adipose Tissue	56
4.3.3	Effect of Methanolic Extract of ABZ Bark on Cholesterol, Triglyceride, HDL and LDL Level in Mice	57
4.3.4	Effect of Methanolic Extract of ABZ Bark on Malondialdehyde Level	59
4.3.5	Effect of Methanolic Extract of ABZ Bark on the Fatty Acid Composition	60
4.3.6	Effect of Methanolic Extract of ABZ Bark on the Adipose Cellularity	62
4.3.7	Effect of Methanolic Extract of ABZ Bark on Liver Cells or Hepatocytes	66
4.3.8	Effect of Methanolic Extract of ABZ Bark on Relative Weight of Liver and Kidney, AST and ALT	67
4.4	Discussion	70
4.4.1	Fatty Acid Composition in Obese Mice Treated With Methanolic Extract of ABZ Bark	70
4.4.2	Adipose Cellularity in Obese Mice Treated With Methanolic Extract of ABZ Bark	70
4.4.3	Effects of Methanolic Extract of ABZ Bark on Hyperlipidemia	72
4.4.4	Effects of High-Fat Diet and Methanolic Extract of ABZ Bark on Liver Cells	73
4.5	Conclusion	74
5.	REGULATION OF GENES AND ADIPOCYTOKINES INVOLVED IN LIPID METABOLISM IN HIGH-FAT DIET-INDUCED OBESITY IN ICR MICE FED WITH METHANOLIC EXTRACT OF <i>ALBIZIA MYRIOPHYLLA</i> (L. BENTH) BARK	75
5.1	Introduction	75
5.2	Materials and Methods	76
5.2.1	Liver Gene Transcription	76
5.2.2	Tissue Collection for RNA Extraction and Purification	76
5.2.3	RNeasy Lipid Tissue Mini Kit for RNA Extraction from Liver	77
5.2.4	Quantitect® Reverse Transcription Kit for Real-time PCR Purposes	77
5.2.5	ELISA Assay Procedure for Adipocytokines which includes Adiponectin, Leptin, TNF α and IL-6	79
5.2.6	Serum Adiponectin Assay	79
	5.2.6.1 Reagents	79
	5.2.6.2 Reagents Preparation	79
5.2.7	Serum Leptin Assay	80
	5.2.7.1 Reagents	80
	5.2.7.2 Reagents Preparation	80
5.2.8	Serum TNF α Assay	81

5.2.8.1	Reagents	81
5.2.8.2	Reagents Preparation	81
5.2.9	Serum IL-6 Assay	81
5.2.9.1	Reagents	81
5.2.9.2	Reagents Preparation	82
5.2.10	Statistical Analysis	82
5.3	Results	82
5.3.1	Effects of Methanolic Extract of ABZ on Liver Gene Transcription including HSL, PPAR α , PPAR γ , SREBP2 and SCD gene	82
5.3.2	Effect of Methanolic Extract of ABZ on Adiponectin, Leptin, TNF α and IL-6 Level	85
5.4	Discussion	88
5.4.1	Regulation of Gene Transcription in Obesity	88
5.4.2	Regulation of Adipocytokines in Obesity	91
5.5	Conclusion	92
6.	GENERAL DISCUSSION, CONCLUSION AND FURTHER WORK	94
6.1	General Discussion	94
6.2	Conclusion	96
6.3	Further Work	97
	REFERENCES	98
	APPENDICES	124
	BIODATA OF STUDENT	133
	PUBLICATION	134

LIST OF TABLES

Table		Page
3.1	The IC ₅₀ Values of ABZ Bark Extracts Based on ABTS, DPPH and FRAP Assay of ABZ Bark Extracts	42
4.1	Treatment groups on methanol ABZ bark extract	49
4.2a	Morphological characteristics of adipose cells	53
4.2b	Morphological characteristics of focal inflammation of the hepatic cells	54
4.2c	Grading of focal inflammation of the hepatic cells	54
4.3	Effect of ABZ bark on body weight in mice	55
4.4	Fatty acid composition in normal and obese mice treated with methanolic extract of ABZ bark	61
4.5	The number of adipose cells with diameter of (<10, 10 to <20, 20 to <30, 30 to <40 and ≥ 40) µm within normal control (NC) mice, high-fat diet control (HFDC) mice, high fat diet mice treated with methanolic extract of ABZ at 10 mg/kg (HFDABZ1), 20 mg/kg (HFDABZ2) and 30 mg/kg (HFDABZ3)	64
4.6	The number of adipose cells with area of <200, 200 to <400, 400 to <600, 600 to <800, 800 to < 1000 and ≥1000 µm ² within normal control (NC) mice, high-fat diet control (HFDC) mice, high fat diet mice treated with methanolic extract of ABZ at 10 mg/kg (HFDABZ1), 20 mg/kg (HFDABZ2) and 30 mg/kg (HFDABZ3)	65
5.1	Names and sequences of the primers used in this study	78

LIST OF FIGURES

Figure	Page
2.1 <i>A. myriophylla</i> tree with leaves and bark	5
2.2 Hypothesis of the possible mechanism involved in anti-obesity and hypoglycemic action of <i>A. myriophylla</i> phytochemical properties	14
2.3 Albizia saponin A,B,C,D,E found in <i>A. myriophylla</i>	15
2.4 The phytochemical properties of genus Albizia	18
2.5 Actions of agent onto PPAR γ causing a cascade which regulates the adipogenesis pathway and glucose homeostasis, thus releasing the adipocytokines and genes involved	23
3.1 Extraction procedure of bark of <i>A. myriophylla</i>	32
3.2 The phenolic contents in 250 μ g of aqueous, methanol, hexane, ethyl acetate, chloroform and butanol <i>A. myriophylla</i> bark extract	36
3.3 The flavonoid levels in 250 μ g of aqueous, methanol, hexane, ethyl acetate, chloroform and butanol <i>A. myriophylla</i> bark extract	37
3.4 The saponin levels in 250 μ g of aqueous, methanol, hexane, ethyl acetate, chloroform and butanol <i>A. myriophylla</i> bark extract	38
3.5 The percentage of inhibition in ABTS antioxidant test in 20 μ g/ml, 40 μ g/ml, 60 μ g/ml, 80 μ g/ml and 100 μ g/ml of aqueous, methanol, hexane, ethyl acetate, butanol and chloroform <i>A. myriophylla</i> bark extracts	39
3.6 The percentage of inhibition in DPPH antioxidant test in 20 μ g/ml, 40 μ g/ml, 60 μ g/ml, 80 μ g/ml and 100 μ g/ml of aqueous, methanol, hexane, ethyl acetate, butanol and chloroform <i>A. myriophylla</i> bark extracts	40
3.7 The percentage of inhibition in FRAP antioxidant test in 20 μ g/ml, 40 μ g/ml, 60 μ g/ml, 80 μ g/ml and 100 μ g/ml of aqueous, methanol, hexane, ethyl acetate, butanol and chloroform <i>A. myriophylla</i> bark extracts	41
4.1 Effect of methanol ABZ extract on weight of adipose tissue in mice. Normal control mice (NC), high-fat diet control mice (HFDC), high fat diet mice treated with methanolic extract of ABZ at 10 mg/kg (HFDABZ1), 20 mg/kg (HFDABZ2) and 30 mg/kg (HFDABZ3).	56

4.2	Cholesterol levels in muscles in mice. Normal control mice (NC), high-fat diet control mice (HFDC), high fat diet mice treated with methanolic extract of ABZ at 10 mg/kg (HFDABZ1), 20 mg/kg (HFDABZ2) and 30 mg/kg (HFDABZ3).	57
4.3	Serum triglyceride levels in mice. Normal control mice (NC), high-fat diet control mice (HFDC), high fat diet mice treated with methanolic extract of ABZ at 10 mg/kg (HFDABZ1), 20 mg/kg (HFDABZ2) and 30 mg/kg (HFDABZ3).	58
4.4	Serum low-density lipoprotein (LDL) levels in mice. Normal control mice (NC), high-fat diet control mice (HFDC), high fat diet mice treated with methanolic extract of ABZ at 10 mg/kg (HFDABZ1), 20 mg/kg (HFDABZ2) and 30 mg/kg (HFDABZ3).	58
4.5	Serum high-density lipoprotein in mice. Normal control mice (NC), high-fat diet control mice (HFDC), high fat diet mice treated with methanolic extract of ABZ at 10 mg/kg (HFDABZ1), 20 mg/kg (HFDABZ2) and 30 mg/kg (HFDABZ3).	59
4.6	Level of malondialdehyde (MDA) in muscles of mice. Normal control mice (NC), high-fat diet control mice (HFDC), high fat diet mice treated with methanolic extract of ABZ at 10 mg/kg (HFDABZ1), 20 mg/kg (HFDABZ2) and 30 mg/kg (HFDABZ3).	60
4.7	Histology of adipose cells marked by the white arrow, in NC and HFDC mice treated with <i>Albizia myriophylla</i> for 5 weeks	62
4.8	The number of adipose cells in mice (cells/ μm^2). Normal control mice (NC), high-fat diet control mice (HFDC), high fat diet mice treated with methanolic extract of ABZ at 10 mg/kg (HFDABZ1), 20 mg/kg (HFDABZ2) and 30 mg/kg (HFDABZ3).	63
4.9	Histology of liver in NC mice and HFDC mice, depicting the hepatocytes or liver cells, marked by the white arrow	66
4.10	Histology of liver cells in NC and HFDC treated with methanol ABZ extract for 5 weeks	66
4.11	Hepatic lesion scoring involving inflammatory foci in hepatic cells of mice. Normal control mice (NC), high-fat diet control mice (HFDC), high fat diet mice treated with methanolic extract of ABZ at 10 mg/kg (HFDABZ1), 20 mg/kg (HFDABZ2) and 30 mg/kg (HFDABZ3)	67
4.12	Effect of methanol ABZ extract on the relative weight of liver tissue collected from mice. Normal control mice (NC), high-fat diet control mice (HFDC), high fat diet mice treated with methanolic extract of ABZ at 10 mg/kg (HFDABZ1), 20 mg/kg (HFDABZ2) and 30 mg/kg (HFDABZ3)	68

4.13	Effect of methanol ABZ extract on the relative weight of kidney collected after sacrificing the mice. Normal control mice (NC), high-fat diet control mice (HFDC), high fat diet mice treated with methanolic extract of ABZ at 10 mg/kg (HFDABZ1), 20 mg/kg (HFDABZ2) and 30 mg/kg (HFDABZ3)	68
4.14	Serum aspartate transaminase levels in mice. Normal control mice (NC), high-fat diet control mice (HFDC), high fat diet mice treated with methanolic extract of ABZ at 10 mg/kg (HFDABZ1), 20 mg/kg (HFDABZ2) and 30 mg/kg (HFDABZ3).	69
4.15	Serum alanine transaminase levels in mice. Normal control mice (NC), high-fat diet control mice (HFDC), high fat diet mice treated with methanolic extract of ABZ at 10 mg/kg (HFDABZ1), 20 mg/kg (HFDABZ2) and 30 mg/kg (HFDABZ3).	69
5.1	Comparison of HSL relative gene transcription in the liver tissue of mice. Normal control mice (NC), high-fat diet control mice (HFDC), high fat diet mice treated with methanolic extract of ABZ at 10 mg/kg (HFDABZ1), 20 mg/kg (HFDABZ2) and 30 mg/kg (HFDABZ3).	83
5.2	Comparison of PPAR α relative gene transcription in the liver tissue of mice. Normal control mice (NC), high-fat diet control mice (HFDC), high fat diet mice treated with methanolic extract of ABZ at 10 mg/kg (HFDABZ1), 20 mg/kg (HFDABZ2) and 30 mg/kg (HFDABZ3).	83
5.3	Comparison of PPAR γ relative gene transcription in the liver tissue of mice. Normal control mice (NC), high-fat diet control mice (HFDC), high fat diet mice treated with methanolic extract of ABZ at 10 mg/kg (HFDABZ1), 20 mg/kg (HFDABZ2) and 30 mg/kg (HFDABZ3).	84
5.4	Comparison of SREBP2 relative gene transcription in the liver tissue of mice. Normal control mice (NC), high-fat diet control mice (HFDC), high fat diet mice treated with methanolic extract of ABZ at 10 mg/kg (HFDABZ1), 20 mg/kg (HFDABZ2) and 30 mg/kg (HFDABZ3).	84
5.5	Comparison of SCD relative gene transcription in the liver tissue of mice. Normal control mice (NC), high-fat diet control mice (HFDC), high fat diet mice treated with methanolic extract of ABZ at 10 mg/kg (HFDABZ1), 20 mg/kg (HFDABZ2) and 30 mg/kg (HFDABZ3).	85
5.6	Serum Adiponectin levels in mice. Normal control mice (NC), high-fat diet control mice (HFDC), high fat diet mice treated with methanolic extract of ABZ at 10 mg/kg (HFDABZ1), 20 mg/kg (HFDABZ2) and 30 mg/kg (HFDABZ3).	86

5.7	Serum leptin levels in mice. Normal control mice (NC), high-fat diet control mice (HFDC), high fat diet mice treated with methanolic extract of ABZ at 10 mg/kg (HFDABZ1), 20 mg/kg (HFDABZ2) and 30 mg/kg (HFDABZ3).	87
5.8	Serum tumor necrosis factor α (TNF α) levels in mice. Normal control mice (NC), high-fat diet control mice (HFDC), high fat diet mice treated with methanolic extract of ABZ at 10 mg/kg (HFDABZ1), 20 mg/kg (HFDABZ2) and 30 mg/kg (HFDABZ3).	87
5.9	Serum interleukin-6 (IL-6) levels in mice. Normal control mice (NC), high-fat diet control mice (HFDC), high fat diet mice treated with methanolic extract of ABZ at 10 mg/kg (HFDABZ1), 20 mg/kg (HFDABZ2) and 30 mg/kg (HFDABZ3).	88
5.10	The mechanism involved in anti-obesity and hypoglycemic action of 30 mg/kg methanolic extract of ABZ bark	93

LIST OF ABBREVIATIONS

ABTS	2-2'-azinobis (3-ethyl-benzothiazoline-6-sulfonic acid)
ABZ	<i>Albizia myriophylla</i>
ADP	Adenosine diphosphate kinase
AKT	Protein kinase B
ALT	Alanine Transaminase
AMPK	Adenosine monophosphate kinase
AST	Aspartate transaminase
BAT	Brown adipose tissue
BMI	Body mass index
CAT	Catalase
CT	Cycle Threshold
CVD	Cardiovascular Disease
DNA	Deoxyribonucleic acid
DPPH	2,2-diphenyl-1-picrylhydrazyl
ELISA	Enzyme-linked immunosorbent assay
FRAP	ferric reducing antioxidant power
GP	Glutathione peroxidase
H&E	Haematoxylin and eosin
HDL	High-Density Lipoprotein
HFD	High-fat diet
HFDABZ1	High-fat diet mice treated with ABZ 10 mg/kg
HFDABZ2	High-fat diet mice treated with ABZ 20 mg/kg
HFDABZ3	High-fat diet mice treated with ABZ 30 mg/kg
HFDC	High-fat diet control

HKG	House keeping gene
HMG-CoA-R	Hydroxy-methylglutaryl-Coenzyme A reductase
HRP	Horseradish peroxidase enzyme
HSL	Hormone-sensitive lipase
IC	Inhibition concentration
IL-6	Interleukin-6
LDL	Low-Density Lipoprotein
MDA	Malondialdehyde
mRNA	Messenger ribonucleic acid
mTOR	Mechanism target of Rapamycin
MUFA	Monounsaturated fatty acid
NAFLD	Non-alcoholic fatty liver disease
NC	Normal control
NF-kB	Nuclear factor kappa-light-chain-enhancer of activated B cells
pAMPK	Phosphorylation of Adenosine monophosphate-activated protein kinase
PCR	Polymerase chain reaction
PPAR α	Peroxisome proliferator-activated receptor alpha
PPAR γ	Peroxisome proliferator-activated receptor gamma
PUFA	Polyunsaturated fatty acid
qPCR	Quantitative reverse transcription PCR
ROS	Reactive oxygen species
RT-PCR	Reverse transcription polymerase chain reaction
SCD	Stearoyl-Coenzyme A-Desaturase
SFA	Saturated fatty acid
SOD	Sodium dismutase

SREBP	Sterol Regulatory Element-Binding Protein
T2DM	Type 2 diabetes mellitus
TG	Triglyceride
TMB	3,3,5,5'-tetramethylbenzidine
TNF- α	Tumor necrosis factor alpha
TSFA	Total saturated fatty acid
VLDL	Very low-density lipoprotein
WAT	White adipose tissue



CHAPTER 1

INTRODUCTION

Obesity is fast becoming a major health concern throughout the world (Hurt *et al.*, 2010). The predisposing factors of obesity have multi-factorial facets involving diet, sedentary lifestyle, dysregulation of the genes and adipocytokines involved in the lipid metabolism (Valavanis *et al.*, 2010). The health burden from obesity is largely driven by an increased risk of type 2 diabetes, cardiovascular diseases, and several forms of cancer such as colon and breast cancer (Renehan *et al.*, 2008). The medical costs of obesity represent the monetary value of health-care resources devoted to managing obesity-related disorders, including the costs incurred by excess use of ambulatory care, hospitalisation, drugs, radiological or laboratory tests, and long term care (Guh *et al.*, 2009). Therefore, obesity proves to be a challenge in the prevention and treatment of the foreseeable conditions stated above.

Obesity is closely related to increasing inflammatory environment and enhanced oxidative stress conditions in the cells (Savini *et al.*, 2013). Obesity consisting of adipose cells accumulation is generally known as a crucial energy supplier and storage and was recently acknowledged as an important endocrine tissue which is actively involved in lipid metabolism (Wang *et al.*, 2014). Undeniably, the endocrine function of the adipose tissue plays a major role in the energy balance with the involvement of the adipocyte derivatives involving the pro-inflammatory and anti-inflammatory adipocytokines (Wang and Huang, 2015). When the generation and release of pro-inflammatory adipocytokines such as TNF α and IL-6 exceed the production of anti-inflammatory adipocytokines such as adiponectin and leptin, systemic inflammation and obesity associated metabolic disorders occurs (Johnson *et al.*, 2012). Hence, the development of obesity is related to the chronic low-grade inflammatory condition (Wang *et al.*, 2014).

Dietary consumption of nutrients rich with anti-inflammatory bioactive components from plants which are high with antioxidant activities such as omega-3 fatty acids and polyphenols, has demonstrated anti-inflammatory activity with reduced inflammation in breast cancer tissue and obesity (Kalupahana *et al.* 2011; Siriwardhana *et al.*, 2013; Savini *et al.*, 2013; Khan and Mukhtar, 2013; Jeong *et al.*, 2016). A review article by Esfahani *et al.* (2011), stated that consumption of mixed fruits and herbal supplements, considerably reduces the oxidative stress activity and elevates the antioxidant levels in the systemic circulation. This evidence supports the dietary guidelines of Malaysians which emphasize the importance of a diet rich in high fiber diet consisting of fruits and vegetables, to prevent the development of obesity (Tee, 2011).

Several researches stated that polyphenols consumption prevents development of obesity via numerous probable mechanism which includes reducing adipogenesis via suppressing the growth and differentiation of adipocytes by inhibiting the PPAR γ and SREBP2 gene expression (Mayoral *et al.*, 2015; Moseti *et al.*, 2016), reducing the

inflammatory activity and attenuating the oxidative stress condition via stimulation of the PPAR α gene expression which reduces the anti-inflammatory precursors namely the TNF α and IL-6, and increases the anti-inflammatory adipocytokines which are the adiponectin and leptin (Jung and Choi, 2014; Jia *et al.*, 2016) and also by reduction of the cholesterol synthesis via the suppression of SREBP2 gene (Taylor *et al.*, 2011). In addition, the down regulation of SCD gene will help in stimulating the insulin sensitivity action, thus producing effective energy utilization (Karahashi *et al.*, 2013).

Among the most common pharmacological drugs concurrently used as an anti-obesity agent in the clinical settings are orlistat, which inhibits the gastric and pancreatic lipases thus, reducing the lipid absorption from the gut and sibutramine, which acts as an oral anorexiant (Kang and Park, 2012). The high rate of side effects such as myocardial ischemia specifically associated with sibutramine, has been the reason for the drug to be withdrawn from the market in several countries (Kang and Park, 2012). This side effect seen in the current synthetic anti-obesity drug stresses the necessity of finding a suitable replacement of alternative with a natural or herbal product.

1.1 Significance of Study

Natural or herbal products have shown great promise as novel drug leads (Cragg and Newman, 2013). Numerous herbal products have been used as an anti-obesity agent. For example, astaxanthin, a dietary carotenoid reduces lipid accumulation in mice through the inhibition of PPAR γ and activation of PPAR α (Jia *et al.*, 2016). Consequently, for a more efficient treatment and control of obesity or weight gain and to satisfy unmet medical needs, there is a necessity to develop new anti-obesity agents that specifically up-regulate the PPAR α expression and down-regulate the PPAR γ and SREBP2 expression. To explore this possibility, in this study, the researcher has used the ABZ bark and its various fractions extracted through maceration process, and tested its antioxidant activities and gene transcription studies.

In Peninsular Malaysia, ABZ or tebu gajah aqueous bark extracts was used as a tonic drink which was mixed with other herbal supplementation. It was taken as an alternative medication for the treatment of diabetes. Following this, a study by Saat *et al.* (2012) has proven that the ABZ aqueous bark extract had significant hypoglycemic activity in streptozotocin nicotinamide induced diabetic rat. As diabetes is closely associated with obesity, with similar pathogenesis, ABZ bark extract may have a potential anti-obesity effect, as well, via indirectly from the antioxidant or free radical scavenging activity. A patent in the United States had used ABZ bark to prevent obesity but not as an anti-obesity agent, as the bitter after taste of saponin thwarts over eating (Patent Application 20040146468).

In the present study, the effect of high fat diet on the liver of the obese mice was also evaluated. The levels of liver enzymes which are AST and ALT, along with the hematoxylin and eosin staining of the hepatocytes of the mice were done. The safety of consumption of the selected doses of ABZ bark in mice was investigated through

evaluation of the obese mice liver. The histology of the kidney was not done due to financial constraint.

Thus, the significance of this study is in the hope that the anti-obesity action of *Albizia myriophylla* bark extract will provide an alternative and play a significant role in body weight management.

1.2 Hypothesis

The antioxidant activity of *A. myriophylla* will regulate the adipogenesis pathway by controlling the adipocytokines and genes involved in the lipid metabolism, thus leading to a reduction in the number and size of adipocyte cells, eventually controlling obesity and is safe in mice model.

1.3 General Objective

The main objective of this study was to investigate the potential anti-obesity activity of *A. myriophylla* bark extract in lipid metabolism and gene regulation of obesity-induced mice.

1.4 Specific Aims

1. To determine the polyphenol composition and content in ABZ bark extract and its antioxidant activity.
2. To analyse the body composition, adipocyte cellularity and blood serum parameters in obesity-induced mice given ABZ bark extract.
3. To evaluate the mRNA expression patterns of different genes in the liver.
4. To evaluate the effect of ABZ bark extract in obesity-induced mice liver.

REFERENCES

- Aali, M., Shahrababak, H.M., Shahrababak, M.M., Sadeghi, M. and Kohram, H. (2016). Polymorphism in the SCD gene is associated with meat quality and fatty acid composition in Iranian fat-and thin-tailed sheep breeds. *Livestock Science*, 188, 81–90.
- Abdollahi, M., Zuki, A.B.Z., Goh, Y.M., Rezaeizadeh, A. and Noordin, M.M. (2010). The effects of *Momordica charantia* on the liver in streptozotocin-induced diabetes in neonatal rats. *African Journal of Biotechnology*, 9(31), 5004–5012.
- Abidov, M., Ramazanov, Z., Seifulla, R. and Grachev, S. (2010). The effects of *Xanthigen* in the weight management of obese premenopausal women with nonalcoholic fatty liver disease and normal liver fat. *Diabetes, Obesity and Metabolism*, 12, 72–81.
- Agassandian, M. and Mallampalli, R.K. (2013). Surfactant phospholipid metabolism. *Biochimica Biophysica Acta*, 1831(3), 612–625.
- Aggarwal, B.B., Prasad, S., Reuter, S., Kannappan, R., Yadav, V.R., Park, B., Kim, J.H., Gupta, S.C., Phromnoi, K., Sundaram, C., Prasad, S., Chaturvedi, M.M. and ung, B. (2011). Identification of novel anti-inflammatory agents from ayurvedic medicine for prevention of chronic diseases: “Reverse pharmacology” and “Bedside to bench” approach. *Current Drug Targets*, 12(11), 1595–1653.
- Ahmad, Q.I., Ahmad, C.B. and Ahmad, S.M. (2010). Childhood obesity. *Indian Journal of Endocrinology and Metabolism*, 14(1): 19–25.
- Ahmadian, M., Suh, J.M., Hah, N., Liddle, J., Atkins, A.R., Downes, M. and Evans, R.M. (2013). PPAR γ signaling and metabolism: The good, the bad and the future. *Nature Medicine*, 19, 557–566.
- Ahmed, H.H., Metwally, F.M., Asmaa, H.R., Zaazaa, M., Ezzat, S.M. and Salama, M.M. (2014). *Moringa oleifera* offers a multi-mechanistic approach for management of obesity in rats. *International Journal of Pharmaceutical Sciences Review and Research*, 29(2), 98–106.
- Ahn, J., Lee, H., Kima, S., Parka, J. and Taeyoul, H. (2008). The anti-obesity effect of quercetin is mediated by the AMPK and MAPK signaling pathways. *Biochemical and Biophysical Research Communications*, 373(4), 545–549.
- Ahn, T.G., Tang, G., Lee, H.M., Kim, M.D., Choi, H.Y. and Park, K.S. (2013). Molecular mechanisms underlying the anti-obesity potential of prunetin, an O-methylated isoflavone. *Biochemical Pharmacology*, 85(10), 1525–1533.
- Akinpelu, B.A., Igbeneghu, O.A., Awotunde, A.I., Iwalewa, E.O. and Oyedapo, O.O. (2014). Antioxidant and antibacterial activities of saponin fractions of *Erythrophyleum suaveolens* (Guill and Perri.) stem bark extract. *Scientific Research and Essays*, 9(18), 826–833.
- Akpan, E.J., Okokon, J.E. and Offong, E. (2012). Antidiabetic and hypolipidemic activities of ethanolic leaf extract and fractions of *Melanthera scandens*. *Asian Pacific Journal of Tropical Biomedicine*, 2(7), 523–527.

- Alfaia, C.M.M., Ribeiro, V.S.S., Lourenc, M.R.A., Auaresma, M.A.G., Martins, S.I.V., Portugal, A.P.V., Fontes, C.M.G., Bessa, R.J.B., Castro, M.L.F. and Prater, J.A.M. (2006). Fatty acid composition, conjugated linoleic acid isomers and cholesterol in beef from crossbred bullocks intensively produced and from Alentejana purebred bullocks reared according to Carnalentejana-PDO specifications. *Meat Science* 72, 425-436.
- Ali, A.T., Hochfeld, W.E., Myburgh, R. and Pepper, M.S. (2013). Adipocyte and adipogenesis. *European Journal of Cell Biology*, 92, 229– 236.
- Amirkhizi, F., Siassi, F., Djalali, M. and RahimiForoushani, A. (2010). Evaluation of oxidative stress and total antioxidant capacity in women with general and abdominal adiposity. *Obesity Research of Clinical Practice*, 4, 209-216.
- Amirkhizi, F., Siassi, F., Djalali, M. and Shahraki, S.H. (2014). Impaired enzymatic antioxidant defense in erythrocytes of women with general and abdominal obesity. *Obesity Research and Clinical Practice*, 8, 26-34.
- Amornchat, C., Kraivaphan, P., Dhanabumi, C., Tandhachoon, K., Trirattana, T. and Choonharcongdj, S. (2006). Effect of cha-em Thai mouthwash on salivary levels of mutans streptococci and total Ig A. *Southeast Asian Journal of Tropical Medicine and Public Health*, 37, 528-531.
- Anandacoomarasamy, A., Caterson, I.D. and Leibman, S (2009). Influence of BMI on health-related quality of life: comparison between an obese adult cohort and age-matched population norms. *Obesity*, 17, 2114–2118.
- Andriantsitohaina, R., Auger, C., Chataigneau, T., Etienne-Selloum, N., Li, H., Martinez, M.C., Schini-Kerth, V.B. and Laher, I. (2012). Molecular mechanisms of the cardiovascular protective effects of polyphenols. *British Journal of Nutrition*, 108, 1–18.
- Arenas, P.M., Molaes, S., Contreras, A.A., Doumecq, B. and Gabrielli, F. (2013). Ethnobotanical, micrographic and pharmacological features of plant-based weight-loss products sold in naturist stores in Mexico City: the need for better quality control. *Acta Botanica Brasilica*, 27(3), 560-579.
- Arner, E., Westermark, P.O., Spalding, K.L., Britton, T., Rydén, M., Frisén, J., Bernard, S. and Arner, P. (2010). Adipocyte Turnover: Relevance to Human Adipose Tissue Morphology. *Diabetes*, 59, 105-109.
- Asilmaz, E., Cohen, P., Miyazaki, M., Dobrzyn, P., Ueki, K. and Fayzikhodjaeva, G. (2004). Site and mechanism of leptin action in a rodent form of congenital lipodystrophy. *Journal of Clinical Investigation*, 113, 414–424.
- Atay, Z. and Bereket, A. (2016). Current status on obesity in childhood and adolescence: Prevalence, etiology, co-morbidities and management. *Obesity medicine*, 3, 1-9.
- Autieri, M.V. (2012). Pro and anti-inflammatory cytokine networks in atherosclerosis. *International Scholarly Research Network Vascular Medicine*, 987629, 1-17.
- Awad, A.B. and Chattopadhyay, J.P. (1986). Effect of dietary saturated fatty acids on hormone-sensitive lipolysis in rat adipocytes. *Journal of Nutrition*, 116, 1088-1094.

- Bais, S., Singh, G.S. and Sharma, R. (2014). Anti-obesity and hypolipidemic activity of *Moringa oleifera* leaves against high fat diet-induced obesity in rats. *Advances in Biology*, 2014, 1-9.
- Bak, M.J., Kim, K.B., Jun, M. and Jeong, W.S. (2014). Safety of red ginseng oil for single oral administration in Sprague-Dawley rats. *Journal of Ginseng Research*, 38, 78-81.
- Berry, D.C., Stenesen, D., Zeve, D. and Graff, J.M. (2013). The developmental origins of adipose tissue. *Development*, 140(19), 3939–3949.
- Boeing, J.S., Barizão, E.O., Silva, B.C., Montanher, P.F., Almeida, V.C. and Visentainer, J.V. (2014). Evaluation of solvent effect on the extraction of phenolic compounds and antioxidant capacities from the berries: application of principal component analysis. *Chemistry Central Journal*, 8(48), 1-9.
- Bonet, M.L., Ribot, J. and Palou, A. (2012). Lipid metabolism in mammalian tissues and its control by retinoic acid. *Biochimica Biophysica Acta*, 1821, 177-189.
- Buysschaerta, B., Aydinb, S., Morellea, J., Hermansc, M.P., Jadoula, M. and Demoulina N. (2016). Weight loss at a high cost: Orlistat-induced late-onset severe kidney disease. *Diabetes and Metabolism*, 42, 62–64.
- Cao, H., Gerhold, K., Mayers, J.R., Wiest, M.M., Watkins, S.M. and Hotamisligil, G.S. (2008). Identification of a lipokine, a lipid hormone linking adipose tissue to systemic metabolism. *Cell*, 134, 933–944.
- Cardellini, M., Andreozzi, F., Laratta, E., Marini, M.A., Laurom R., Hribal, M.L., Perticone, F. and Sesti, G. (2007). Plasma interleukin-6 levels are increased in subjects with impaired glucose tolerance but not in those with impaired fasting glucose in a cohort of Italian Caucasians. *Diabetes Metabolism Research and Reviews*, 23(2), 141-145.
- Cawthorn, W.P., Scheller, E.L. and MacDougald, O.A. (2012). Adipose tissue stem cells meet preadipocyte commitment: going back to the future. *Journal of Lipid Research*, 53, 227-246.
- Chan, R.S.M. and Woo, J. (2010). Prevention of overweight and obesity: How effective is the current public health approach. *International Journal of Environmental Research and Public Health*, 7, 765-783.
- Chara, K.J. and Oltmanns, K.M. (2014). Obesity – A neuropsychological disease? Systematic review and neuropsychological model. *Progress in Neurobiology*, 114, 84–101.
- Cheng, N., Ren, N., Gao, H., Lei, X., Zheng, J. and Cao, W. (2013). Antioxidant and hepatoprotective effects of Schisandra chinensis pollen extract on CCl₄-induced acute liver damage in mice. *Food and Chemical Toxicology*, 55, 234–240.
- Chevassus, H., Gaillard, J.B., Farret, A., Costa, F., Gabillaud, I., Mas, E., Dupuy, A.M., Michel, F., Centiê, C., Renard, E., Galtier, F. and Petit, P. (2010). A fenugreek seed extract selectively reduces spontaneous fat intake in overweight subjects. *European Journal of Clinical Pharmacology*, 66, 449-455.

- Choi, B.K., Park, S.B., Lee, D.R., Lee, H.J., Jin, Y.Y., Yang, S.H. and Suh, J.W. (2016). Green coffee bean extract improves obesity by decreasing body fat in high-fat diet-induced obese mice. *Asian Pacific Journal of Tropical Medicine*, 9(7), 635–643.
- Choi, H., Won, E., Jang, Y.P. and Choung, S. (2013). Antiobesity Effect of *Codonopsis lanceolata* in High-Calorie/High-Fat-Diet-Induced Obese Rats. *Evidence-Based Complementary and Alternative Medicine*, 210297, 1-9.
- Choudharya, I., Chimanpurea, V., Patil, A., Mukhopadhyayab, R., Paranjapea, R. and Bhattacharyaa, J. (2013). Single step detection of HIV-1 proviral DNA and housekeeping β actin gene from dried blood spots by a multiplex polymerase chain reaction. *Journal of Virological Methods*, 187, 203– 206.
- Clemens, R., Kranz, S., Mobley, A.R., Nicklas, T.A., Raimondi, M.P. Rodriguez, J.C., Slavin, J.L. and Warshaw, H. (2012). Filling America's fiber intake gap: Summary of a roundtable to probe realistic solutions with a focus on grain-based foods. *The Journal of Nutrition*, 142, 1390–1401.
- Coelho, I., Pequeto, D.C.T., Borghetti, G., Aikawa, J., Yamaguchi, A.A., Brito, G.A.P., Yamazaki, R.K., Scorsato, A.P., Fernandes, L.C., Coimbra, T.M. and Fernandez, R. (2016). Chronic fish oil supplementation partially reverses renal alterations in mice fed with a high-fat diet. *Journal of Functional Foods*, 26, 196–207.
- Coelho, M., Oliveira, T. and Fernandes, R. (2013). Biochemistry of adipose tissue: an endocrine organ. *Archives of Medical Science*, 9(2), 191-200.
- Cohen, P., Miyazaki, M., Succi, N.D., Hagge-Greenberg, A., Liedtke, W. and Soukas, A.A. (2002). Role for stearoyl-CoA desaturase-1 in leptin mediated weight loss. *Science*, 297, 240–243.
- Contreras-Zentella, M.L. and Hernández-Muñoz, R. (2015). Is liver enzyme release really associated with cell necrosis induced by oxidant stress? *Oxidative Medicine and Cellular Longevity*, 2016 (2016), 3529149, 1-12.
- Cook, K.S., Min, H.Y., Johnson, D., Chaplinsky, R.J., Flier, J.S., Hunt, C.R. and Spiegelman, B.M. (1987). Adipsin: a circulating serine protease homolog secreted by adipose tissue and sciatic nerve. *Science*, 237(4813), 402-405.
- Cook, N.C. and Samman, S. (1996). Review: flavonoids-chemistry, metabolism, cardioprotective effects and dietary sources. *Journal of Nutritional Biochemistry*, 7(2), 66–76.
- Copes, L.E., Schutz, H., Dlugosz, E.M., Acosta, W., Chappell, M.A. and Junior, Garland, T.J. (2015). Effects of voluntary exercise on spontaneous physical activity and food consumption in mice: Results from an artificial selection experiment. *Physiology and Behavior*, 149, 86–94.
- Cragg, G.M. and Newman, D.J. (2013). Natural products: A continuing source of novel drug leads. *Biochimica Biophysica Acta*, 1830(6), 3670-3695.
- Das, N., Islam, M.E., Jahan, N., Islam, M.S., Khan, A., Islam, M.R. and Parvin, S. (2014). Antioxidant activities of ethanol extracts and fractions of *Crescentia cujete* leaves and stem bark and the involvement of phenolic

- compounds. *BioMedCentral Complementary and Alternative Medicine*, 14(45), 1-9.
- Dashty, M., (2014). A quick look at biochemistry: Lipid metabolism. *Journal of Diabetes and Metabolism*, 5(1), 1-17.
- Datau, E., Surachmanto, E.E., Pandelaki, K. and Langi, J. (2010). Efficacy of *Nigella sativa* on serum free testosterone and metabolic disturbances in central obese male. *Acta Medical Indonesia*, 42, 130-134.
- Denroche, H.C., Huynh, F.K. and Kieffer, T.J. (2012). The role of leptin in glucose homeostasis. *Journal of Diabetes Investigation*, 3(2), 115-129.
- Després, J.P., Poirier, P., Bergeron, J., Tremblay, A., Lemieux, I. and Alméras, N. (2008). From individual risk factors to the metabolic syndrome to global cardiometabolic risk. *European Heart Journal*, 10, 24-33.
- Deurenberg, P., Weststrate, J.A. and Seidell, J.C. (1991). Body mass index as a measure of body fatness: age and sex specific prediction formulas. *British Journal of Nutrition*, 65, 105.
- Deurenberg-Yap, M., S.K., Lin, V.F.P., Tan, B.Y., van Staveren, W.A. and Deurenberg, P. (2001). Relationships between indices of obesity and its comorbidities in multi-ethnic Singapore. *International Journal of Obesity*, 25, 1554-1562.
- Dhurandhar, E.J. and Keith, S.W. (2014). The aetiology of obesity beyond eating more and exercising less. *Best Practice and Research Clinical Gastroenterology*, doi: 10.1016/j.bpg.2014.07.001.
- Ding, L., Li, J., Song, B., Xiao, X., Huang, W. and Zhang, B. (2014). Andrographolide prevents high-fat diet-induced obesity in C57BL/6 mice by suppressing the sterol regulatory element-binding protein pathway. *Journal of Pharmacology and Experimental Therapeutics*, 351(2), 474-483.
- Ding, L., Li, J., Song, B., Xiao, X., Zhang, B., Qi, M., Huang, W., Yang, Li. and Wang, Z. (2016). Curcumin rescues high fat diet-induced obesity and insulin sensitivity in mice through regulating SREBP pathway. *Toxicology and Applied Pharmacology*, 304, 99-109.
- Dinstel, R.R., Cascio, J. and Koukel, S. (2013). The antioxidant level of Alaska's wild berries: high, higher and highest. *International Journal of Circumpolar Health*, 72, 796-802.
- Dixon, J.L., Furukawa, S. and Ginsberg, H.N. (1991). Oleate stimulates secretion of apolipoprotein B-containing lipoproteins from Hep G2 cells by inhibiting early intracellular degradation of apolipoprotein B. *Journal of Biological Chemistry*, 266, 5080-5086.
- Dzialis, M., Mierziak, J., Korzun, U., Preisner, M., Szopa, J. and Kulma, A. (2016). The Potential of plant phenolics in prevention and therapy of skin disorders. *International Journal of Molecular Sciences*, 17(160), 1-41.
- Ebrahimi, M., Rajion, M.A., Goh, Y.M. and Farjam, A.S. (2014). Omega-3 Fatty Acid Enriched Chevron (Goat Meat) Lowers Plasma Cholesterol Levels and Alters Gene Expressions in Rats. *BioMed Research International*, 749341, 1-8.

- Ebrahimi, M., Rajion, M.A., Goh, Y.M. and Sazili, A.Q. (2012). Impact of different inclusion levels of oil palm (*Elaeis guineensis* Jacq.) fronds on fatty acid profiles of goat muscles. *Journal of Animal Physiology and Animal Nutrition*, 96(6), 962-969.
- Ebrahimi, M., Rajion, M.A., Goh, Y.M., Farjam, A.S., Oskoueian, E. and Jafari, S. (2015). Diet high in α -linolenic acid up-regulate PPAR- α gene expression in the liver of goats. *Electronic Journal of Biotechnology*, 18, 210-214.
- Ebrahimi, M., Rajion, M.A., Goh, Y.M., Sazili, A.Q. and Schonewille, J.T. (2013). Effect of Linseed Oil Dietary Supplementation on Fatty Acid Composition and Gene Expression in Adipose Tissue of Growing Goats. *BioMed Research International*, 194625, 1-11.
- Eisenberg, T., Knauer, H., Schauer, A., Büttner, S., Ruckstuhl, C. and Carmona-Gutierrez, D. (2009). Induction of autophagy by spermidine promotes longevity. *Natural Cell Biology*, 11(11), 1305-1314.
- Eldin, E.E.M.N., Almarzouki, A., Assiri, A.M., Elsheikh, O.M., B.E.A., Mohamed, and Babakr, A.T. (2014). Oxidized low density lipoprotein and total antioxidant capacity in type-2 diabetic and impaired glucose tolerance Saudi men. *Diabetology and Metabolic Syndrome*, 6(94), 1-10.
- El-Haleim, E.A.A., Bahgat, A.K. and Saleh, S. (2016). Effects of combined PPAR- γ and PPAR- α agonist therapy on fructose induce NASH in rats: Modulation of gene expression. *European Journal of Pharmacology*, 773, 59-70.
- Elhanati, S., Kanfi, Y., Varvak, A., Roichman, A., Carme-Gross, I., Barth, S., Gibor, G. and Cohen, H.Y. (2013). Multiple regulatory layers of SREBP1/2 by SIRT6. *Cell Reports*, 4, 905-912.
- El-Kadre, L.J. and Tinoco, A.C.A. (2013). Interleukin-6 and obesity: The crosstalk between intestine, pancreas and liver. *Current Opinion in Clinical Nutrition & Metabolic Care*, 16, 564-568.
- Engelmann, G.S., Richardson, A., Katz, A. and Fierer, J.A. (1981). Age-related changes in isolated rat hepatocytes, comparison of size, morphology, binucleation and protein content. *Mechanism of Ageing and Development*, 16, 385-395.
- Eriksson, J.W. (2007). Metabolic stress in insulin's target cells leads to ROS accumulation – A hypothetical common pathway causing insulin resistance. *Federation of European Biochemical Societies Letters*, 581, 3734-3742.
- Erlund, I., Koli, R., Alfthan, G., Marniemi, J., Puukka, P., Mustonen, P., Mattila, P. and Jula, A. (2010). Favorable effects of berry consumption on platelet function, blood pressure, and HDL cholesterol. *American Journal of Clinical Nutrition*, 87, 323-331.
- Esfahani, A., Wong, J.M., Truan, J., Villa, C.R., Mirrahimi, A. and Srichaikul, K. (2011). Health effects of mixed fruit and vegetable concentrates: a systematic review of the clinical interventions. *The Journal of American College of Nutrition*, 30, 285-94.

- Esteghamati, A., Mazaheri, T., Rad, M.V. and Noshad, S. (2015). Complementary and alternative medicine for the treatment of obesity: A critical review. *International Journal of Endocrinology Metabolism*, 13(2), 1-9.
- Evans, C.A.R., Miller, N.J. Bolwell, P.G., Broamley, P.M. and Pridham.J.B. (1995). The relative antioxidant activities of plant derived polyphenolic flavonoids,.*Free Radical Research*, 22(4), 375–383.
- Facchini, F.S., Hua, N.W., Reaven, G.M. and Stoohs, R.A. (2000). Hyperinsulinemia: the missing link among oxidative stress and age-related diseases? *Free Radical of Biology Medicine*, 29, 1302-1306.
- Fidrianny, I., Permatasari, L. and Wirasutisna, K.R. (2013). Antioxidant activities frm various bulbs extracts of three kinds allium using DPPH, ABTS assays and correlation with total phenolic, flavonoid, carotenoid content. *International Journal of Research and Pharmaceutical Sciences*, 4(3), 438-444.
- Flowers, J.B., Rabaglia, M.E., Schueler, K.L., Flowers, M.T., Lan, H. and Keller, M.P. (2007). Loss of stearyl-CoA desaturase-1 improves insulin sensitivity in lean mice but worsens diabetes in leptin-deficient obese mice. *Diabetes*, 56, 1228–1239.
- Folch, J., Lees, M. and Stanley, G.H.S. (1957).A simple method for the isolation and purification of total lipids from animal tissues.*The Journal of Biological Chemistry*, 226, 497-509.
- Fuhrman, B., Buch, S., Vaya, J., Coleman, R., Hayek, T. and Aviram, M. (1997). Licorice extract and its major polyphenol glabridin protect low density lipoprotein against lipid peroxidation: in vitro and ex vivo studies in humans and in arteriosclerotic apolipoprotein E-deficient mice. *American Journal of Clinical Nutrition*, 66,267–275
- Fuhrman, B., Rosenblat, M., Hayek, T., Coleman, R. and Aviram, M. (2000). Ginger extract consumption reduces plasma cholesterol, inhibits LDL oxidation and attenuates development of atherosclerosis in atherosclerotic, Apolipoprotein E-deficient mice. *Journal of Nutrition*, 130, 1124–1131.
- Funtikova, A.N., Navaro, E., Bawaked, R.A., Fito, M. and Schröder, H. (2015).Impact of diet on cardiometabolic health in children and adolescents. *Nutrition Journal*, 14, 118-129.
- Gao, C.L., Zhu, C. and Zhao, Y.P. (2010). Mitochondrial dysfunction is induced by high levels of glucose and free fatty acids in 3T3L1 adipocytes. *Molecular and Cellular Endocrinology*, 320(2), 25-33.
- Ge, F., Zhou, S., Hu, C., Lobdell, H. and Berk, P.D. (2010). Insulin- and leptin-regulated fatty acid uptake plays a key causal role in hepatic steatosis in mice with intact leptin signaling but not in ob/ob or db/db mice. *American Journal of Physiology Gastrointestinal Liver Physiology*, 299, G855–G866.
- Ghee, L.K. (2016).Areview of adult obesity research in Malaysia.*Medical Journal of Malaysia*, 71(1), 1-19.
- González Jiménez E. (2010). Evaluación de la eficacia de una intervención educativa sobre nutrición y actividad física en niños y adolescentes escolares con

sobrepeso y obesidad de Granada y provincia. *Tesis Doctoral. Universidad de Granada*.

- González-Castejón, M. and Rodríguez-Casado, A. (2011). Dietary phytochemicals and their potential effects on obesity: A review. *Pharmacological Research*, 64, 438-455.
- Gowda, S., Desai, P.B., Hull, V.V., Math, A.A.K., Vernekar, S.N. and Kulkarni, S.S. (2009). A review on laboratory liver function tests. *Pan African Medical Journal*, 3(17), 1-11.
- Grygiel-Górniak, B. (2014). Peroxisome proliferator-activated receptors and their ligands: nutritional and clinical implications: A review. *BioMedCentral Nutrition Journal*, 13(17), 1-10.
- Guh, D.P., Zhang, W., Bansback, N., Amarsi, Z., Birmingham, C.L. and Anis, A.H. (2009). The incidence of co-morbidities related to obesity and overweight: a systematic review and meta-analysis. *BioMedCentral Public Health*, 9, 88.
- Gujral, J.S., Knight, T.R., Farhood, A., Bajt, M.L. and Jaeschke, H. (2002). Mode of cell death after acetaminophen overdose in mice: Apoptosis or oncotic necrosis? *Toxicological Sciences*, 67, 322-328.
- Gujral, U.P., Pradeepa, R., Weber, M.B., Narayan, K.M.V. and Mohan, V. (2013). Type 2 diabetes in South Asians: similarities and differences with white Caucasian and other populations. *Annals of the New York Academy of Sciences*, 1281, 51-63.
- Gutiérrez-Juárez, R., Pocai, A., Mulas, C., Ono, H., Bhanot, S., Monia, B.P. and Rossetti, L. (2006) Critical role of stearoyl-CoA desaturase-1(SCD1) in the onset of diet-induced hepatic insulin resistance. *Journal of Clinical Investigation*, 116, 1686-1695.
- Han, J.M., Lee, J.S., Kim, H.G., Seol, I.C., Im, H.J., Cho, J.H. and Son, C.G. (2015). Synergistic effects of *Artemisia iwayomogi* and *Curcuma longa* radix on high-fat diet-induced hyperlipidemia in a mouse model. *Journal of Ethnopharmacology* 173 (2015) 217-224.
- Hara, A. and Radin, N.S. (1978). Lipid extraction of tissues with a low-toxicity solvent. *Analytical Biochemistry*, 90, 420-426.
- Hariri, N. and Thibault, L. (2010). High-fat diet-induced obesity in animal models. *Nutrition Research Reviews*, 23, 270-299.
- Harris, R.B.S. (2014). Direct and indirect effects of leptin on adipocyte metabolism. *Biochimica Biophysica Acta*, 1842, 414-423.
- Hasani-Ranjbar, S., Nayeibi, M., Larijani, B. and Abdollahi, M. (2009). . A systemic review of the efficacy and safety of herbal medicines used in the treatment of obesity. *World Journal of Gastroenterology*, 15, 3070-3085.
- Hasani-Ranjbar, S., Nayeibi, N., Moradi, Mehri, A., Larijani, B. and Abdollahi, M. (2010). The efficacy and safety of herbal medicines used in the treatment of hyperlipidemia: a systemic review. *Current Pharmaceutical Design*, 16, 2935-2947.

- He, R., Chen, L., Lin, B., Matsui, Y., Yao, X. and Kurihara, H. (2009). Beneficial effects of oolong tea consumption on diet-induced overweight and obese subjects. *Journal of Chinese Integrated Medicine*, 15, 34-41.
- Hill, J.O., Wyatt, H.R. and Peters, J.C. (2012). Energy, balance and obesity. *Circulation*, 126(1), 126-132.
- Hofman, R. and Gerber, M. (2015). Food processing and the mediterranean diet. *Nutrients*, 7, 7925-7964
- Hossain, M.A., Al-Raqmi, K.A.S., Al-Mijizy, Z.H., Weli, A.M. and Al-Riyami, Q. (2013). Study of total phenol, flavonoids contents and phytochemical screening of various leaves crude extracts of locally grown *Thymus vulgaris*. *Asian Pacific Journal of Tropical Biomedicine*, 3(9), 705-710.
- Hotamisligil, G.S., Shargill, N.S. and Spiegelman, B.M. (1993). Adipose expression of tumor necrosis factor- α direct role in obesity-linked insulin resistance. *Science*, 259, 87-89.
- Hu, C.C., Qing, K. and Chen, Y. (2004). Diet-induced changes in stearoyl-CoA desaturase 1 expression in obesity-prone and -resistant mice. *Obesity Research*, 12, 1264-1270.
- Huang, D., Ou, B., Hampsch-Woodill, M., Flanagan, J.A. and Prior, R.L. (2002). High throughput assay of oxygen radical absorbance capacity (ORAC) using a multichannel liquid handling system coupled with a microplate fluorescence reader in 96-well format. *Journal of Agriculture and Food Chemistry*, 50, 4437-4444.
- Huang, J., Wang, Y., Xie, Z., Zhou, Y., Zhang, Y. and Wan, X. (2014). The anti-obesity effects of green tea in human intervention and basic molecular studies. *European Journal of Clinical Nutrition*, 68(10), 1075-1087.
- Huerta-Leidenz N. O., Cross H. R., Savell J. W., Lunt D. K., Baker J. F., and Smith S. B. (1996). Fatty acid composition of subcutaneous adipose tissue from male calves at different stages of growth. *Journal of Animal Sciences*, 74, 1256-1264.
- Hurt, R.T., Kulisek, C., Buchanan, L.A. and McClave, S.A. The obesity epidemic: challenges, health initiatives, and implications for gastroenterologists. *Gastroenterology and Hepatology*, 6(12), 780-792.
- Hwang, I.C., Park, J.Y., Ahn, H.Y., Kim, K.K., Suh, H.S., Ko, K.D. and Kimb, K. (2014). Effects of CYP3A5, CYP2C19, and CYP2B6 on the clinical efficacy and adverse outcomes of sibutramine therapy: A crucial role for the CYP2B6*6 allele. *Clinica Chimica Acta*, 428, 77-81.
- Ikarashi, N., Toda, T., Okaniwa, T., Ito, K., Ochiai, W. and Sugiyama. (2011). Anti-obesity and anti-diabetic effects of acacia polyphenol in obese diabetic KKAY mice fed high-fat diet. *Evidence-Based Complementary Medicine*. 952031, 1-10.
- Iloki-Assanga, S.B., Lewis-Luján, L.M., Lara-Espinoza, C.L., Gil-Salido, A.A., Fernandez-Angulo, D., Rubio-Pino, J.L. and Haines, D.D. (2015). Solvent effects on phytochemical constituent profiles and antioxidant activities, using four different extraction formulations for analysis of *Bucida buceras* L.

- and *Phoradendron californicum*. *BioMedCentral Research Notes*, 8(396), 1-14.
- Inadera, H. (2008). The usefulness of circulating adipokine levels for the assessment of obesity-related health problems. *International Journal of Medical Sciences*, 5(5), 248-262
- Jang, W.S. and Choung, S.Y. (2013). Antiobesity Effects of the Ethanol Extract of *Laminaria japonica* Areshoung in High-Fat-Diet-Induced Obese Rat. *Evidence-Based Complementary and Alternative Medicine*, 492807, 1-17.
- Jeong, Y.J., Sohm, E.H., Jung, Y.H., Yoon, W.J., Cho, Y.M., Kim, I., Lee, S.R. and Kang, S.C. (2016). Anti-obesity effect of *Crinum asiaticum* var. *japonicum* Baker extract in high-fat diet-induced and monogenic obese mice. *Biomedicine and Pharmacotherapy*, 82, 35-43.
- Jernås, M., Palming, J., Sjöholm, K., Jennische, E., Svensson, P.A., Gabrielsson, B.G., Levin, M., Sjögren, A., Rudemo, M., Lystig, T.C., Carlsson, B., Carlsson, L.M. and Lönn, M. (2006). Separation of human adipocytes by size: hypertrophic fat cells display distinct gene expression. *Federation of American Societies for Experimental Biology Journal*, 20, 1540-1542.
- Jiang, G., Li, Z., Liu, F., Ellsworth, K., Dallas-Yang, Q. and Wu, M. (2005). Prevention of obesity in mice by antisense oligonucleotide inhibitors of stearoyl-CoA desaturase-1. *Journal of Clinical Investigation*, 115, 1030-1038.
- Jie, Z.Y., Yue-qin, L., Shan-yan, C. and Jie, S. (2012). Effects of dietary energy level on the expression of the HSL gene in different tissues of sheep. *Journal of Integrative Agriculture*, 11(7), 1167-1172.
- Jiménez, E.G. (2013). Obesity: Etiologic and pathophysiological analysis. *Endocrinological Nutrition*, 60(1), 17---24.
- Jo, J., Gavrilova, O., Pack, S., Jou, W. Mullen, S., Sumner, A., Cushman, S.W. and Periwai, V. (2009). Hypertrophy and/or Hyperplasia: Dynamics of Adipose Tissue Growth. *PLoS Computational Biology*, 5(3), 1-5.
- Joycharat, N., Boonma, C., Thammavong, S., Yingyongnarongkul B.E., Limsuwan, S. and Voravuthikunchai, S.P. (2016). Chemical constituents and biological activities of *Albizia myriophylla* wood. *Pharmaceutical Biology*, 54(1), 62-73.
- Jung, U.J. and Choi, M. (2014). Obesity and its metabolic complications: The role of adipokines and the relationship between obesity, inflammation, insulin resistance, dyslipidemia and nonalcoholic fatty liver disease. *International Journal of Molecular Sciences*, 15, 6184- 6223.
- Kadir, N.A.A.A., Rahmat, A. and Jaafat, H.Z.E. (2015). Protective effects of tamarillo (*Cyphomandra betacea*) extract against high fat diet Induced obesity in Sprague-Dawley rats. *Journal of Obesity*, 846041, 1-8.
- Kalupahana, N.S. and Moustaid-Moussa, N. (2012). The adipose tissue renin-angiotensin system and metabolic disorders: a review of molecular mechanisms. *Critical Review of Biochemistry and Molecular Biology*, 47, 379-390.

- Kalupahana, N.S., Claycombe, K.J. and Moustaid-Moussa, N. (2011). (n-3) Fatty acids alleviate adipose tissue inflammation and insulin resistance: mechanistic insights. *Advance Nutrition*, 2, 304–316.
- Kalupahana, N.S., Moustaid-Moussa, N. and Claycombe, K.J. (2012). Immunity as a link between obesity and insulin resistance. *Molecular Aspects Medicine*, 33, 26–34.
- Kang, J.G. and Park, C.Y. (2012). Anti-obesity drugs: A review about their effects and safety. *Diabetes and Metabolism Journal*, 36, 13–25.
- Karahashi, M., Ishii, F., Yamazaki, T., Imai, K., Mitsumoto, A., Kawashima, Y. and Kudo, N. Up-regulation of stearyl-CoA desaturase 1 increases liver MUFA content in obese Zucker but not Goto-Kakizaki rats. *Lipids*, 48, 457–467.
- Kaur, J. (2014). A comprehensive review on metabolic syndrome. *Cardiology Research and Practice*, 943162, 1–21.
- Kaur, R., Arora, S. and Singh, B. (2008). Antioxidant activity of the phenol rich fractions of leaves of *Chukrasia tabularis* A. Juss. *Bioresource Technology*, 99, 7692–7698.
- Kawashima, Y. and Kozuka, H. (1982). Increased activity of stearyl-CoA desaturation in liver from rat fed clofibrate. *Biochimica Biophysica Acta*, 713, 622–628.
- Kedare, S.B. and Singh, R.P. (2011). Genesis and development of DPPH method of antioxidant assay. *Food Science Technology*, 48(4), 412–422.
- Kêkê, L.M., Samouda, H., Jacobs, J., di Pompeo, C., Lemdani, M., Hubert, H., Zitouni D. and Guinhouya, B.C. (2015). Body mass index and childhood obesity classification systems: A comparison of the French, International Obesity Task Force (IOTF) and World Health Organization (WHO) references. *Epidemiology and Public Health*, 63, 173–182.
- Kesh, S.B., Sarkar, D. and Manna, K. (2016). High-fat diet-induced oxidative stress and its impact on metabolic syndrome: A review. *Asian Journal of Pharmaceutical and Clinical Research*, 9(1), 47–52.
- Khan, N. and Mukhtar, H. (2013). Tea and health: Studies in humans. *Current Pharmaceutical Design*, 19(34), 6141–6147.
- Khatoon, M., Islam, E., Islam, R., Rahman, A.A., Alam, A.H.M.K., Khondkar, P., Rashid M. and Parvin, S. (2013). Estimation of total phenol and in vitro antioxidant activity of *Albizia procera* leaves. *BioMed Central Research Notes*, 6(121), 1–7.
- Kim, H., Hiraishi, A., Tsuchiya, K. and Sakamoto, K. (2010). Epigallocatechingallate suppresses the differentiation of 3T3-L1 preadipocytes through transcription factors FoxO1 and SREBP1c. *Cytotech*, 62(3), 245–255.
- Kishimoto, T. (2010). IL-6: From its discovery to clinical applications. *International Immunology*, 22(5), 347–352.
- Klaus, S., Pultz, S., Thone-Reineke, C. and Wolfram, S. (2005). Epigallocatechingallate attenuates diet-induced obesity in mice by decreasing

- energy absorption and increasing fat oxidation. *International Journal of Obesity*, 29, 615-623.
- Klover, P.J., Zimmers, T.A., Koniaris, L.G. and Mooney, R.A. (2003). Chronic exposure to interleukin-6 causes hepatic insulin resistance in mice. *Diabetes*, 52, 2784-2789.
- Knowler, W.C., Pettitt, D.J., Saad, M.F. and Bennett, P.H. (1990). Diabetes mellitus in the Pima Indians: incidence, risk factors and pathogenesis. *Diabetes Metabolic*
- Kokila, K., Priyadarshini, S.D. and Sujatha, V. (2013). Phytopharmacological properties of Albizia species: A review. *International Journal of Pharmacy and Pharmaceutical Sciences*, 5(3), 70-73.
- Kopaei, M.R., Setorki, M., Douadi, M., Baradaran, A. and Nasri, H. (2014). Atherosclerosis: Process, indicators, risk factors and new hopes. *International Journal of Preventive Medicine*, 5(8), 927-946.
- Kostapanos, M.S., Kei, A. and Elisaf, M.S. (2013). Current role of fenofibrate in the prevention and management of non-alcoholic fatty liver disease. *World Journal of Hepatology*, 5 (9), 470-478.
- Kovacova, Z., Tencerova, M., Roussel, B., Wedellova, Z., Rossmeislova, L., Langin, D., Polak, J. and Stich, V. (2012). The impact of obesity on secretion of adiponectin multimeric isoforms differs in visceral and subcutaneous adipose tissue. *International Journal of Obesity*, 36, 1360-1365.
- Kranendonk, M.E.G., Herwaarden, J.A.V., Stupkova, T., Jager, W.D., Vink, A., Moll, F.L., Kalkhoven, E. and Visseren F.L.J. (2015). Inflammatory characteristics of distinct abdominal adipose tissue depots relate differently to metabolic risk factors for cardiovascular disease. Distinct fat depots and vascular risk factors. *Atherosclerosis*, 239, 419-427.
- Kubota, K., Sumi, S., Tojo, H. and Sumi-Inoue, Y. (2011). Improvements of mean body mass index and body weight in pre-obese and overweight Japanese adults with black Chinese tea (Pu-Erh) water extract. *Nutritional Research*, 31, 421-428.
- Kudo, N., Toyama, T., Mitsumoto, A. and Kawashima, Y. (2003). Regulation by carbohydrate and clofibrate of palmitoyl-CoA chain elongation in the liver of rats. *Lipids*, 38, 531-537.
- Kumar, R.S., Raj Kapoor, B. and Perumal, P. (2012). Antioxidant activities of *Indigofera cassioides* Rottl. Ex. DC. Using various in vitro assay models. *Asian Pacific Journal of Tropical Biomedicine*, 2(4), 256-261.
- Khennouf, S., Amira, S., Arrar, L. and Baghiani, A. (2010). Effect of some phenolic compounds and quercus tannins on lipid peroxidation. *World Applied Sciences*, 8, 1144-1149.
- Kumar, S. and Pandey, A.K. (2013). Chemistry and biological activities of flavonoids: An overview. *The Scientific World Journal*, 162750, 1-16.
- Kumar, S., Gupta, A. and Pandey, A.K. (2013). *Calotropis procera* root extract has capability to combat free radical mediated damage. *Pharmacology*, 691372, 1-8.

- Kumar, S., Gupta, A. and Pandey, A.K. (2013). Lai, N., Ims, J.K., Jeon, N.L. and Lee, K. (2012). Adipocyte induction of preadipocyte differentiation. *Tissue Engineering*, 18(12), 958-967.
- Lampidonis, A.D., Rogdakis, E., Voutsinas, G.E. and Stravopodis D.J. (2011). The resurgence of Hormone-Sensitive Lipase (HSL) in mammalian lipolysis. *Gene*, 477, 1-11.
- Lee, D.R., Lee, Y.S., Choi, B.K., Lee, H.J., Park, S.B., Kim, T.M., Oh, H.J., Yang, S.H. and Suh, J.W. (2015). Roots extracts of *Adenophora triphylla* var. *japonica* improve obesity in 3T3-L1 adipocytes and high-fat diet-induced obese mice. *Asian Pacific Journal of Tropical Medicine*, 8(11), 898-906.
- Lee, S.M., Cho, Y.H., Lee, S.Y., Jeong, D.W., Cho, A., Jeon, J.S., Park, E., Kim, Y.J., Lee, J.G., Yi, Y.H., Tak, Y.J., Hwang, H.R., Lee, S. and Hand, H. (2015). Urinary malondialdehyde is associated with visceral abdominal obesity in middle-aged men. *Mediators of Inflammation*, 524291, 1-6.
- Legrand, P., Catheline, D., Fichot, M.C. and Lemarchal, P. (1997). Inhibiting D9-desaturase activity impairs triacylglycerol secretion in cultured chicken hepatocytes. *Journal of Nutrition*, 127, 249-256.
- Leite, A.V., Malta, L.G., Riccio, M.F., Eberlin, M.N., Pastore, G.M. and Marostica, M.R. (2011). Antioxidant potential of rat plasma by administration of freeze-dried jaboticaba peel (*Myrciaria jaboticaba* Vell Berg). *Journal of Agricultural and Food Chemistry*, 59(6), 2277-2283.
- Lenon, G.B., Li, K.X., Chang, Y.H., Yang, A.W., De Costa, C., Li, C.G., Cohen, M., Mann, N. and Xue, C.C.L. (2012). Efficacy and safety of a Chinese herbal medicine formula (RCM-104) in the management of simple obesity: a randomized, placebo controlled clinical trial. *Evidence Based Complementary and Alternative Medicine*, 2012, 1-11.
- Lenquiste, A., Batista, A.G., Marineli, R.D.S., Dragano, N.R.V. and Marostica, M.R. (2012). Freeze-dried jaboticaba peel added to high-fat diet increases HDL-cholesterol and improves insulin resistance in obese rats. *Food Research International*, 49(1), 153-160.
- Li, J., He, X., Li, M., Zhao, W., Liu, L. and Kong, X. (2015). Chemical fingerprint and quantitative analysis for quality control of polyphenols extracted from pomegranate peel by HPLC. *Food Chemistry*, 176, 7-11.
- Li, X.C., Wang X.Z. and Chen, D.F. (2011). Antioxidant activity and mechanism of protochatechuic acid in vitro. *Journal of Functional Food in Health and Disease*, 1, 232-244.
- Li, Z., Yang, L., Wang, J., Shi, W., Pawar, R.A., Liu, Y., Xu, C., Cong, W., Hua, Q., Lu, T., Xia, F., Guo, W., Zhao, M. and Zhang, Y. (2010). β -Actin is a useful internal control for tissue-specific gene expression studies using quantitative real-time PCR in the half-smooth tongue sole *Cynoglossus semilaevis* challenged with LPS or *Vibrio anguillarum*. *Fish and Shellfish Immunology*, 29, 89-93.
- Liu, H., Liu, J.Y., Wu, X. and Zhang, J. (2010). Biochemistry, molecular biology, and pharmacology of fatty acid synthase, an emerging therapeutic target and

- diagnosis/prognosis marker. *International Journal of Biochemistry and Molecular Biology*, 1(1), 69-89.
- Lumeng, C.N. and Saltiel, A.R. (2011). Inflammatory links between obesity and metabolic disease. *Journal of Clinical Investigations*, 121(6), 2111-2117.
- Lundgren, M., Svensson, M., Lindmark, S., Renstrom, F., Ruge, T. and Eriksson, J.W. (2007). Fat cell enlargement is an independent marker of insulin resistance and hyperleptinaemia. *Diabetologica*, 50, 625-633.
- Lutz, T.A. and Woods, S.C. (2012). Overview of animal models of obesity. *Current Protocol in Pharmacology*, 5, 1-22.
- Ma, R.C.W. and Chan, J.C.N. (2013). Type 2 diabetes in East Asians: similarities and differences with populations in Europe and the United States. *Annals of the New York Academy of Sciences*, 1281, 64-91.
- Madeo, F., Tavernarakis, N. and Kroemer, G. (2010). Can autophagy promote longevity? *Natural Cell Biology*, 12(9), 842-846.
- Madhikarmi, N.L., Murthyl, K.R.S., Rajagopal, G. and Singh, P.P. (2013). Lipid peroxidation and antioxidant status in patients with type 2 diabetes in relation to obesity in Pokhara – Nepal. *Journal of Diabetology*, 1(3), 1-8.
- Matough, F.A., Budin, S.B., Hamid, Z.A., Alwahaibi, N. and Mohamed, J. (2012). The role of oxidative stress and antioxidants in diabetic complications. *Sultan Qaboos University Medical Journal*, 12(1), 5-18.
- Makki, K., Froguel, P. and Wolowczuk, I. (2013). Adipose tissue in obesity-related inflammation and insulin resistance: cells, cytokines, and chemokines. *International Scholarly Research Network Inflammation*, 139239, 1-12.
- Mandal, S., Yadav, S., Yadav, S. and Nema, R.N. (2009). Antioxidants: A Review. *Journal of Chemical and Pharmaceutical Research*, 1(1), 102-104.
- Manickam, M., Ramanathan, M., Farboodiney Jahromi, M.A., Chansouria, J.P.N. and Ray, A.B. (1997). Antihyperglycemic activity of phenolics from *Pterocarpus marsupium*. *Journal of Natural Product*, 60, 609-610.
- Massaro, M., Scoditti, E., Pellegrino, M., Carluccio, M.A., Calabriso, N., Wabitsch, M., Storelli, C., Wright, M. and Caterina R.D. (2016). Therapeutic potential of the dual peroxisome proliferator activated receptor (PPAR) α/γ agonist aleglitazar in attenuating TNF α mediated inflammation and insulin resistance in human adipocytes. *Pharmacological Research*, 107, 125-136.
- Mastrocola, R., Guglielmotto, M., Medana, C., Catalano, Cutrupi, S., Borghi, R., Tamagno, E., Boccuzzi, G. and Aragno, M. (2011). Dysregulation of SREBP2 induces BACE1 expression. *Neurobiology of Disease*, 44, 116-124.
- Mayne, S.T., Playdon, M.C. and Rock, C.L. (2016). Diet, nutrition, and cancer: past, present and future. *Nature Reviews Clinical Oncology*, 13, 504-515.
- Mayoral, R., Osborn, O., McNelis, J., Johnson, A.M., Oh, D.Y., Izqueirido, C.I., Chung, H., Li, P., Traves, P.G., Bandyopadhyay, G., Pessentheiner, A.R., Ofrecio, J.M., Cook, J.R., Qiang, L., Accili, D. and Olefsky, J.M. (2015). Adipocyte SIRT1 knockout promotes PPAR γ activity, adipogenesis and

- insulin sensitivity in chronic-HFD and obesity. *Molecular Metabolism*, 4, 378-391.
- McArdle, M.A., Finucane, O.M., Connaughton, R.M., McMorrow, A.M. and Roche, H.M. (2013). Mechanisms of obesity-induced inflammation and insulin resistance: insights into the emerging role of nutritional strategies. *Frontiers in Endocrinology*, 5, 1-23.
- McKeage, K. and Keating, G.M. (2011). Fenofibrate: A review of its use in dyslipidaemia. *Drugs*, 71 (14), 1917–1946.
- Meda, A., Lamien, C.E., Romito, M., Millogo, J. and Nacoulma, O.G. (2005). Determination of the total phenolic, flavonoid and proline contents in Burkina Fasan honey, as well as their radical scavenging activity. *Food Chemistry*, 91 (3), 571–577.
- Mehenni, C., Atmani-Kilani, D., Dumarc, S., Perrin, D., Gerardin, P. and Atmani, D. (2016). Hepatoprotective and antidiabetic effects of *Pistacia lentiscus* leaf and fruit extracts. *Journal of Food and Drug Analysis*, 24, 653-669.
- Miyazaki, M., Kim, Y.C., Gray-Keller, M.P., Attie, A.D. and Ntambi, J.M. (2000). The Biosynthesis of hepatic cholesterol esters and triglycerides is impaired in mice with a disruption of the gene for stearoyl-CoA desaturase 1. *Journal of Biological Chemistry*, 275, 30132–30138.
- Miyazaki, Y., Pipek, R., Mandarino, L.J. and DeFronzo, R.A. (2003). Tumor necrosis factor α and insulin resistance in obese type 2 diabetic patients. *International Journal of Obesity and Related Metabolic Disorder*, 27, 88-94.
- Mohamed, G.A., Ibrahim, S.R.M., Elkhayat, E.S. and El Dine, R.S. (2014). Natural anti-obesity agents. *Bulletin of Faculty of Pharmacy, Cairo University*, 52, 269-284.
- Mohammadzadeh, G. and Ghaffari, M (2014). Additional effect of diabetes mellitus type 2 on the risk of coronary artery disease: Role of serum adiponectin. *Iranian Red Crescent Medical Journal*, 16,(1), 1-7.
- Mohamud, W.N.W., Musa, K.I., Khir, A.S.M., Ismail, A.A., Ismail, I.S., Kadir, K.A., Kamaruddin, N.A., Yaacob, N.A., Mustafa, N, Ali, O, Isa, S.H.M. and Bebakar, W.M.W. (2011). Prevalence of overweight and obesity among adult Malaysians: an update. *Asian Pacific Journal of Clinical Nutrition*, 20(1), 35-41.
- Mohd-Radzman, N.B., Ismail, W.I.W., Adam, Z., Jaapar, S.S. and Adam A. (2013). Potential roles of *Stevia rebaudiana bertonii* in abrogating insulin resistance and diabetes: A review. *Evidence-Based Complementary Medicine*, 718049, 1-10.
- Moller, D.E. (2000). Potential role of TNF- α in the pathogenesis of insulin resistance and type 2 diabetes. *Trends in Endocrinology and Metabolism*, 11(6), 212-217.
- Morsella, E., Criollo, A., Navas, C.R. and Clegg D.J (2014). Chronic high-fat diet consumption impairs metabolic health of male mice. *Inflammatory Cell Signal*, 1(6), 561-572.

- Morselli, E., Mariño, G., Bennetzen, M.V., Eisenberg, T., Megalou, E. and Schroeder, S. (2011). Spermidine and resveratrol induce autophagy by distinct pathways converging on the acetylproteome. *Journal of Cell Biology*, 192(4), 615-629.
- Moseti, D., Regassa, A. and Kin, W.K. (2016). Molecular regulation of adipogenesis and potential anti-adipogenic bioactive molecules. *International Journal of Molecular Science*, 17(124), 1-24.
- Mulvihill, E.E. and Huff, M.W. (2010). Antiatherogenic properties of flavonoids: implications for cardiovascular health. *Canadian Journal of Cardiology*, 26(A), 17-21.
- Musman, M. (2010). Toxicity of *Barringtonia racemosa* (L.) kernel extract on *Pomacea canaliculata* (Ampullariidae). *Tropical Life and Sciences Research*, 21, 41-50.
- Ngondi, J.L., Etoundi, B.C., Nyamongo, C.B., Mbofung, C.M.F. and Oben, J.E. (2009). IGOB131, a novel seed extract of the West African plant *irvingia gabonensis* significantly reduces body weight and improves metabolic parameters in overweight humans in a randomized double-blind placebo controlled investigation. *Lipids Health and Disease*, 8, 1-7.
- Nichols, M.S., Silva-Sanigorski, A.D., Clearly, J.E., Goldfeld, S.R., Colahan, A. and Swinburn, B.A. (2011). Decreasing trends in overweight and obesity among an Australian population of preschool children. *International Journal of Obesity*, 35, 916-924.
- Nicholson, S.K., Tucker, G.A. and Brameld, J.M. (2008). Effects of dietary polyphenols on gene expression in human vascular endothelial cells. *Proceedings of the Nutrition Society*, 67, 42-47.
- Nikolic, D., Katsiki, N., Montalto, G., Isenovic, E.R., Mikhailidis, D.P. and Rizzo, M. (2013). Lipoprotein subfractions in metabolic syndrome and obesity: Clinical significance and therapeutic approaches. *Nutrients*, 5, 928-948.
- Nilsson, C., Raun, K., Yan, F.F., Larsen, M.O. and Christensen, M.T. (2012). Laboratory animals as surrogate models of human obesity. *Acta Pharmacologica Sinica*, 33, 173-181.
- Nita, M. and Grzybowski, A. (2016). The role of the reactive oxygen species and oxidative stress in the pathomechanism of the age-related ocular diseases and other pathologies of the anterior and posterior eye segments in adults. *Oxidative Medicine and Cellular Longevity*, 3164734, 1-23.
- Nobili, S., Lippi, D., Witort, E., Donnini, M. and Bausi, L. (2009). Natural compounds for cancer treatment and prevention. *Pharmacological Research*, 59(6), 365-378.
- Ntambi, J.M. and Miyazaki, M. (2004). Regulation of stearoyl-CoA desaturases and role in metabolism. *Progress in Lipid Research*, 43, 91-104.
- Ntambi, J.M., Miyazaki, M., Stoehr, J.P., Lan, H., Kendzierski, C.M. and Yandell, B.S. (2002). Loss of stearoyl-CoA desaturase-1 function protects mice against adiposity. *Proceedings of the National Academy of Sciences of the United States of America*, 99, 11482-11486.

- Nunez, I.N. Galdeano, C.M., de LeBlanc, A.M. and Perdíg, G. (2014). Evaluation of immune response, microbiota, and blood markers after probiotic bacteria administration in obese mice induced by a high-fat diet. *Nutrition*, 30, 1423–1432.
- Nworgu, Z.A.M., Onwukaeme, D.N., Afolayan A.J., Ameachine, F.C. and Ayinde, B.A. (2008). Preliminary studies of blood pressure lowering effect of *Nauclea latifolia* in rats. *African Journal of Pharmacy and Pharmacology*, 2(2), 37-41.
- Oberly, D.W. (1998). Free radicals and diabetes. *Free Radical of Biology Medicine*, 5, 113-124.
- Oliveira L.L., Carvalho M.V. and Melo, L. (2014). Health promoting and sensory properties of phenolic compounds in food. *Revista Ceres*, 61, 764-779.
- Ondrejovičová, I., Muchová, J., Mišlanová, C., Nagyová, Z. and Ďuračková, Z. (2010). Hypercholesterolemia, oxidative stress and gender dependence in children. *Prague Medical Report*, 111(4), 300–312.
- Ouchi, N., Parker, J.L., Lugus, J.J. and Walsh, K. (2011). Adipokine in inflammation and metabolic disease. *Nature Reviews Immunology*, 11(2), 85-97.
- Ouchii, N., Ohashii, K., Shibata, R. and Murohara, T. (2012). Adipocytokines and obesity-linked disorders. *Nagoya Journal of Medicine*, 74, 19-30.
- Padro, G., Badimon, T. and Vilahur, L. (2010). Nutraceuticals and atherosclerosis human trials. *Cardiovascular Therapy*, 28(4), 202-215.
- Padwal, R.S. and Majumdar, S.R. (2007). Drug treatments for obesity: orlistat, sibutramine, and rimonabant. *Lancet*, 369, 71–77.
- Pandey, A.K. (2007). Anti-staphylococcal activity of a pan-tropical aggressive and obnoxious weed *Pariheniumhisterophorus*: an *invitro* study. *National Academy Science Letters*, 30(11-12), 383–386.
- Panmei, C., Singh, P.K., Gautam, S., Variyar, P.S., Shantibala Devi, G.A. and Sharma, A. (2007). Phenolic acids in *Albizia* bark used as a starter for rice fermentation in Zou preparation. *Journal of Food, Agricultural and Environment*, 5, 147-150.
- Pannangpatch, P., Kongyingyoes, B., Kukongviriyapan, V. and Kukongviriyapan, U. (2006). Hypoglycemic activity of three Thai traditional medicine regimens in streptozotocin-induced diabetic rats. *Khon kaen University Research Journal*, 11(2), 159-168.
- Park, A., Kim, W.K. and Bae, K. (2014). Distinction of white, beige and brown adipocytes derived from mesenchymal stem cells. *World Journal of Stem Cells*, 6(1), 33-42.
- Parola, M. and Marra, F. (2011). Adipokines and redox signaling: Impact on fatty liver disease. *Antioxidants and Redox Signalling*, 15 (2), 461-483.
- Parthasarathy, S., Raghavamenon, A., Garelnabi, M.O. and Santanam N. (2010). Oxidized low-density lipoprotein. *Methods in Molecular Biology*, 610, 403–417.
- Pasaribu, T., Astuti, D.A. Wina, E, Sumiati. and Setiyono, A. (2014). Saponin content of *Sapindus rarak* pericarp affected by particle size and type of solvent, its

- biological activity on *Eimeria tenella* oocysts. *International Journal of Poultry Science*, 13 (6), 347-352.
- Paz-Filho, G., Mastronardi, C., Wong, Ma. and Licinio, J. (2012). Leptin therapy, insulin sensitivity, and glucose homeostasis. *Indian Journal of Endocrinology and Metabolism*, 16(3), 549–555.
- Perez-Vizcaino, F. and Duarte, J. (2010). Phytochemicals and cardiovascular protection: flavonols and cardiovascular disease. *Molecular Aspects of Medicine*, 31 (6), 478-494.
- Pham-Huy, L.A., He, H. and Phan-Huy, C. (2008). Free, radicals, antioxidants in disease and health. *International Journal of Biomedical Sciences*, 4, 89-86.
- Picchi, M.G., Mattos, A.M., Barbosa, M.R., Duarte, C.P. Gandini, M.A., Portari, G.V. and Jordão A.A. (2011). A high-fat diet as a model of fatty liver disease in rats. *Acta Cirúrgica Brasileira*, 26 (2), 25-31.
- Pichiah, P.B.T., Suriyakalaa, U., Kamalakkannan, S., Kokilavani, P., Kalaiselvi, S., Sankar Ganesh, D., Gowri, J., Archunan, G., Cha, Y.S. and Achiraman, S. (2011). Spermidine may decrease ER stress in pancreatic beta cells and may reduce apoptosis via activating AMPK dependent autophagy pathway. *Medical hypotheses*, 77 (4), 677–679.
- Pienaar, A.E. (2015). Prevalence of overweight and obesity among primary school children in a developing country: NW-CHILD longitudinal data of 6–9-yr-old children in South Africa. *BioMed Central Obesity*, 2(2), 1-10.
- Pincu, Y., Huntsman, H.D., Zou, K., Lisio, M.D., Mahmassani, Z.S., Munroe, M.R., Garg, K., Jensen, T. and Boppart, M.D. (2016). Diet-induced obesity regulates adipose-resident stromal cell quantity and extracellular matrix gene expression. *Stem Cell Research*, 17, 181–190.
- Pi-Sunyer, X (2009). The medical risks of obesity. *Postgraduate Medicine*, 121(6), 21-33.
- Popeijus, H.E., Saris, W.H.M. and Mensink, R.P. (2008). Role of stearoyl-CoA desaturases in obesity and the metabolic syndrome. *International Journal of Obesity*, 32, 1076–1082.
- Popkin, B.M., Adair, L.S. and Ng, S.W. (2012). Now and then: The global nutrition transition: The pandemic of obesity in developing countries. *Nutritional Review*, 70(1), 3–21.
- Pou, K.M., Massaro, J.M., Hofman, U., Vasan, R.S., Maurovich-Horvat, P. and Larson, M.G. (2007). Visceral and subcutaneous adipose tissue volumes are cross-sectionally related to markers of inflammation and oxidative stress. *Circulation*, 116, 1234-1241.
- Powell, D.A., Black, W.C.B., Bleasby, K., Chan, C., Deschenes, D., Gagnon, M., Gordon, R., Guay, J., Guiral, S., Hafey, M.J., Huang, Z., Isabel, E., Leblanc, Y., Styhler, A., Xu, L.J., Zhang, L. and Oballa, R.M. (2011). Nicotinic acids: Liver-targeted SCD inhibitors with preclinical anti-diabetic efficacy. *Bioorganic and Medicinal Chemistry Letters*, 21, 7281–7286.
- Powell, L.M. and Nguyen, B.T. (2013). Fast-food and full-service restaurant consumption among children and adolescents: Impact on energy, beverage

- and nutrient intake. *Journal of Americal Medical Association Paediatrics*, 167(1), 14-20.
- Pumthong, G., Nathason, A., Tuseewan, M., Pinthong, P., Klangrapun, S., Thepsuriyanon, D. and Kotta, P. (2015). Complementary and alternative medicines for diabetes mellitus management in ASEAN countries. *Complementary Therapies in Medicine*, 2015, 1-9.
- Qatanani, M. and Lazar, M.A. (2007). Mechanisms of obesity-associated insulin resistance: many choices on the menu. *Genes and Development*, 21, 1443-1455.
- Qin, N., Chen, Y., Jin, M.N., Zhang, C., Qiao, W., Yue, X.L., Duan, H.Q. and Niu, W.Y. (2016). Anti-obesity and anti-diabetic effects of flavonoid derivative (Fla-CN) via microRNA in high fat diet induced obesity mice. *European Journal of Pharmaceutical Sciences*, 82, 52–63.
- Quan, H.Y., Kim Do, Y. and Vhyng, S.H. (2013). Caffeine attenuates lipid accumulation via activation of AMP-activated protein kinase signaling pathway in HepG2 cells. *Biochemistry and Molecular Biology Reports*, 46, 207-212.
- Quettier-Deleu, C., Gressier, B., Vasseur, J., Dine, T., Brunet, C., Luyckx, M., Cazin, M., Cazin, J.C., Bailleul, F. and Trotin, F. (2000). Phenolic compounds and antioxidant activities of buckwheat (*Fagopyrum esculentum* Moench) hulls and flour. *Journal of Ethnopharmacology*, 72(1-2), 35-42.
- Rahal, A., Kumar, A., Singh, V., Yadav, B., Tiwari, R., Chakraborty, S. and Dhama, K. (2014). Oxidative stress, prooxidants, and antioxidants: The interplay. *BioMed Research International*. 761264, 1-19.
- Rajendra, P., Nandakumar, N., Rengarajan, T., Palaniswami, R., Gnanadhas, E.N., Lakshminarasiah, U., Gopas, J. and Nishigaki, I. (2014). Antioxidants and human diseases. *Clinica Chimica Acta*, 436, 332-347.
- Rajion, M.A., McLean, J.G. and Cahill, R.N. (1985). Essential fatty acids in the fetal and newborn lamb. *Aust J Biol Sci*. 1985;38(1):33-40. *Review*, 6 (1), 1-27.
- Renehan, A.G., Tyson, M., Egger, M., Heller, R.F. and Zwahlen, M. (2008). Body-mass index and incidence of cancer: a systematic review and meta-analysis of prospective observational studies. *Lancet*, 371, 569–578.
- Rezende, L.F.M., Rey-López, J.P., Matsudo, V.K.R. and Luiz, O.C. (2014). *BioMed Central Public Health*, 14(333), 1-9.
- Richard, D., Kefi, K., Barbe, U., Poli, A., Bausero, P. and Visioli, F. (2009). Weight and plasma lipid control by decaffeinated green tea. *Pharmacological Research*, 59(5), 351-354.
- Rivera, H.M., Christiansen, K.J. and Sullivan, E.L. (2015). The role of maternal obesity in the risk of neuropsychiatric disorders. *Frontiers of Neuroscience*. 9, 194-210.
- Romero-Corral, A., Caples, D.M., Lopez-Jimenez, F. and Somers, V.K. (2010). Interactions between obesity and obstructive sleep apnea. *Chest*, 137(3), 711-719.

- Rosini, T.C., Silva A.S.R. and Moraes, C. (2012). Diet-induced obesity: rodent model for the study of obesity-related disorders. *Revista da Associação Médica Brasileira*, 58(3), 383-387.
- Rucker, D., Padwal, R., Li, S.K., Curioni, C. and Lau, D.C. (2007). Long term pharmacotherapy for obesity and overweight: updated meta-analysis. *British Medical Journal*, 335, 1194-1199.
- Rydén, M., Jocken, J., van Harmelen, V., Dicker, A., Hoffstedt, J., Wirén, M., Blomqvist, L., Mairal, A., Langin, D., Blaak, E. and Arner, P. (2007). Comparative studies of the role of hormone-sensitive lipase and adipose triglyceride lipase in human fat cell lipolysis. *American Journal of Physiology, Endocrinology and Metabolism*, 292, 1847-1855.
- Saat, A., Syakroni, N. and Rosli, R. (2012). Potential hypoglycemic property of *Albizia myriophylla* in streptozotocin-nicotinamide induced diabetic rats. *International Journal of Pharmacy and Pharmaceutical Sciences*, 4(3), 427-431.
- Saat, A., Syakroni, N. and Rosli, R. (2013). Potential hypoglycemic activity of *Albizia myriophylla* and virgin coconut oil in streptozotocin induced diabetic rats. *International Journal of Pharmacy and Pharmaceutical Sciences*, 5(4), 199-202.
- Sahoo, K., Sahoo, B., Choudhury, A.K., Sofi, N.Y., Kumar, R. and Bhadoria, A.S. (2015). Childhood obesity: causes and consequences. *Journal of Family Medicine and Primary Care*, 4(2), 187-192.
- Samoylenko, V., Jacob, M.R., Khan, S.I., Zhao, J., Tekwani, B.L., Midiwo, J.O., Walker, L.A. and Muhammad, I. (2009). Antimicrobial, antiparasitic and cytotoxic spermine alkaloids from *Albizia schimperiana*. *Natural Products Communications*, 4(6), 791-796.
- Santos, N.H.A., Fiaccone, R.L., Barreto, M.L., Silva, L.A. and Silva, R.C.R. Association between eating patterns and body mass index in a sample of children and adolescents in Northeastern Brazil. *Cadernos de Saúde Pública*, Rio de Janeiro, 30(10), 2235-2245.
- Savage, D.B., Petersen, K.F. and Shulman, G. (2007). Disordered lipid metabolism and the pathogenesis of insulin resistance. *Physiological Reviews*, 87(2), 507-520.
- Savini, I., Catani, M.V., Evangelista, D., Gasperi, V. and Avigliano, L (2013). Obesity-associated oxidative stress: Strategies finalized to improve redox state. *International Journal of Molecular Sciences*, 14, 10497-10538.
- Sebastiano, K.M.D. and Mourtzakis, M. (2014). The role of dietary fat throughout the prostate cancer trajectory. *Nutrients*, 6, 6095-6109.
- Selassie, M. and Sinha, A.C. (2011). The epidemiology and aetiology of obesity: A global challenge. *Best Practice and Research Clinical Anaesthesiology*, 25, 1-9.
- Shao, W. and Espenshade, P.J. (2012). Expanding roles for SREBP in metabolism. *Cell Metabolism*, 16(4), 414-419.
- Sharma, J. (2010). Etiology of obesity- What a family practitioner needs to know. *BioMedCentral Obesity*, 513, 1-9.

- Shen, B., Chen, H., Shen, C., Xu, P., Li, J., Shen, G., Yuan, H. and Han, Jin. (2015). Hepatoprotective effects of lignans extract from *Herpetospermum caudigerum* against CCl₄-induced acute liver injury in mice. *Journal of Ethnopharmacology*, 164, 46-52.
- Shields, M. and Tremblay, M.S. (2010). Canadian childhood obesity estimates based on WHO, IOTF and CDC cut-points. *International Journal of Paediatrics Obesity*, 5(3), 265-273.
- Shukri, R., Mohamed, S. and Mustapha, M.N. (2010). Cloves protect the heart, liver and lens of diabetic rats. *Food Chemistry*, 122, 1116-1121.
- Siahpoosh, A. and Mehrpeyma, M. (2014). Antioxidant effects of *Albizia lebbek* and *Prosopis juliflora* barks. *International Journal of Biosciences*, 5(9), 273-284.
- Sies, H (2010). Polyphenols and health: update and perspectives. *Archives Biochemistry Biophysics*, 501, 2-5.
- Singh, D., Cho, W.C. and Upadhyay G. (2015). Drug induced liver toxicity and prevention by herbal antioxidants: An overview. *Frontiers in Physiology*, 6(363), 1-18.
- Singh, M. (2014). Mood, food and obesity. *Frontiers in Psychology*, 5, 1-20.
- Siriwardhana, N., Kalupahana, N.S., Cekanova, M., Lemieux, M., Greer, B. and Moustaid-Moussa, N. (2013). Modulation of adipose tissue inflammation by bioactive food compounds. *Journal of Nutritional Biochemistry*, 24, 613-623.
- Sivakrishnan, S., Kavitha, J. and Muthu, A.K. (2012). Antioxidant potential, total phenolic and flavonoids content of aerial parts of ethanolic extract of *Albizia procera* (Family: Mimosoideae). *Asian Journal of Pharmaceutical and Clinical Research*, 6, 108-110.
- Skinner, A.C. and Skelton, J.A. (2014). Prevalence and trends in obesity and severe obesity among children in the United States, 1999-2012. *The Journal of American Medical Association Pediatrics*, 168 (6), 561-566.
- Skinner, J.D., Carruth, B.R. and Moran, J. (1999). Fruit juice is not related to children's growth. *Pediatrics*, 103, 58-64.
- Skurk, T., Albert-Huber, C., Herder, C. and Hauner, H. (2007). Relationship between adipocyte size and adipokine expression and secretion. *Journal of Clinical Endocrinology and Metabolism*, 92, 1023-1033.
- Smith, A.Y.R. and Adanlawo, I.G. (2014). *In vitro* and *in vivo* antioxidant activity of saponin extractd from the root of *Garcinia kola* (bitter kola) on alloxan-induced diabetic rats. *World Journal of Pharmacy and Pharmaceutical Sciences*, 3(7), 8-26.
- Son, M.J., Rico, C.W., Nam, S.H. and Kang, M.Y. (2010). Ferulic Acid on the lipid metabolism and antioxidative status in high-fed mice. *Journal of Clinical Biochemistry and Nutrition*, 46, 150-156.
- Soran, H., Schofield, J.D. and Durrington, P.N. (2015). Antioxidant properties of HDL. *Frontiers in Pharmacology*. 6(222), 1-6.
- Sparg, S.G., Light, M.E. and van Staden, J. (2004). Biological activities and distribution of plant saponins. *Journal of Ethnopharmacology*, 94, 219-243.

- Srinivasan, M., Sudheer, A.R. and Menon, V.P. (2007). Ferulic acid therapeutic potential through its antioxidant property. *Journal of Clinical Biochemistry and Nutrition*, 40, 92-100.
- Stich, V., Harant, I., Gliszinski, I., Crampes, F., Berlan, M., Kunesova, M., Hainer, V., Dauzats, M., Rivi re, D. and Garrigues, M. (1997). Adipose tissue lipolysis and hormone sensitive lipase expression during very-low-calorie diet in obese female identical twins. *Journal of Clinical Endocrinology and Metabolism*, 82, 739-744.
- Stothard, K.J., Tennant, P.W.G., Bell, R. and Rankin, J. (2009). Maternal overweight and obesity and the risk of congenital anomalies: a systematic review and meta-analysis. *Journal of American Medical Association*, 301, 636-650.
- Styskal, J., Van Remmen, H., Richardson, A. and Salmon, A.B. (2012). Oxidative stress and diabetes: what can we learn about insulin resistance from antioxidant mutant mouse models? *Free Radical Biology and Medicine*, 52(1), 46-58.
- Sudheer, A., Muthukumaran, S., Kalpana, C., Srinivasan, M. and Menon, V.P. (2007). Protective effect of ferulic acid in nicotine-induced DNA damage and cellular changes in cultured rat peripheral blood lymphocytes. A comparison with N-acetylcysteine. *Toxicological In Vitro*, 21, 576-585.
- Sun, H., He, S. and Shi, M. (2014). Adjuvant-active fraction from *Albizia julibrissin* saponins improves immune responses by inducing cytokine and chemokine at the site of injection. *International Immunopharmacology*, 22, 346-355.
- Sun, K., Kusminski, C.M. and Scherer, P.E. (2011). Adipose tissue remodeling and obesity. *Journal of Clinical Investigation*, 121, 2094-2101.
- Suzuki, K., Jayasena, C.N. and Bloom, S.R. (2012). Obesity and Appetite Control. *Experimental Diabetes Research*, 2012, 1-19.
- Taheri, E., Djalali, M., Saedisomeolia, A., Moghadam, A.M., Djazayeri, A. and Qorbani, M. (2012). The relationship between the activities of antioxidant enzymes in red blood cells and body mass index in Iranian type 2 diabetes and healthy subjects. *Journal of Diabetes and Metabolic Disorders*, 11(3), 1-5.
- Tanaka, S., Yagi, Y., Yamazaki, T., Mitsumoto, A., Kobayashi, D., Kudo, N. and Kawashima, Y. (2012). Characterization of fatty acid profile in the liver of SHR/NDmcr-cp (cp/cp) rats, a model of the metabolic syndrome. *Biological and Pharmaceutical Bulletin*, 35, 184-191.
- Tang, J.J., Li, J.G., Qi, W.W., Li, P.S. and Li, B.L. (2011). Inhibition of SREBP by a small molecule, botulin, improves hyperlipidemia and insulin resistance and reduces atherosclerotic plaques. *Cell Metabolism*, 13(1), 44-56.
- Tao, R., Xiong, X., DePinho, R.A., Deng, C.X. and Dong, X.C. (2013). Hepatic SREBP-2 and cholesterol biosynthesis are regulated by FoxO3 and Sirt6. *Journal of Lipid Research*, 54(10), 2745-2753.
- Taylor, H.E., Linde, M.E., Khatua, A.K., Popik, W. and Hildreth, J.E.K. (2011). Sterol regulatory element-binding protein 2 couples HIV-1 transcription to cholesterol homeostasis and T Cell Activation. *Journal of Virology*, 85, 7699-7709.

- Tchernof, A. and Després, J. (2013). Pathophysiology of human visceral obesity: An update. *Physiological Review*, 93, 359-404.
- Tee, E.E. (2011). Development and promotion of Malaysian Dietary Guidelines. *Asian Pacific Journal of Clinical Nutrition*, 20(3), 455-461.
- Teodoro, J.S., Varela, A.T. and Rolo, A.P (2014). High-fat and obesogenic diets: current and future strategies to fight obesity and diabetes. *Genes and Nutrition*, 9, 406-421.
- Thaipong, K., Boonprakob, U., Crosby, K., Zevallos, L.C. and Byrne, D.H. (2006). Comparison of ABTS, DPPH, FRAP, and ORAC assays for estimating antioxidant activity from guava fruit extracts. *Journal of Food Composition and Analysis*, 19, 669-675.
- Theis, N.D. 2013. Histopathology of alcoholic liver disease. *Clinical Liver Disease A Multimedia Review Journal*, 2(2), 64-67.
- Thomas, T.R., Pellechia, J., Rector, R.S., Sun, G.Y., Sturek, M.S. and Laughlin, M.H. (2002). Exercise training does not reduce hyperlipidemia in pigs fed a high-fat diet. *Metabolism*, 51(12), 1587-1595
- Tiwari, P., Kumar, B., Kaur, M., Kaur, G. and Kaur, H. (2011). Phytochemical screening and extraction: A review. *Internationale Pharmaceutica Scientia*, 1(1), 98-106.
- Touyz, R.M. (2004). Reactive oxygen species, vascular oxidative stress, and redox signaling in hypertension. What is the clinical significance? *Hypertension*, 44, 248-252.
- Toyama, T., Kudo, N., Hibino, Y., Mitsumoto, A., Nishikawa, M. and Kawashima, Y. (2007). Effects of pioglitazone on stearyl-CoA desaturase in obese Zucker fa/fa rats. *Journal of Pharmacological Sciences*, 104, 137-145.
- Tresserra-Rimbau, A., Rimm, E.B., Medina-Remón, A., Martínez-González, M.A., López-Sabater, M.C., Covas, M.I., Corella, D., Salas-Salvadó, J., Gómez-Gracia, E., Lapetra, J., Arós, F., Fiol, M., Ros, E., Serra-Majem, L., Pintó, X., Muñoz, M.A., Gea, A., Ruiz-Gutiérrez, V., Estruch, R. and Lamuela-Raventós, R.M. (2014). Polyphenol intake and mortality risk: a re-analysis of the PREDIMED trial. *BioMed Central Medicine*, 12(77), 1-11.
- Tschöp, M. and Heiman, M.L. (2001). Rodent obesity models: an overview. *Experimental and Clinical Endocrinology and Diabetes*, 109(6), 307-319.
- Tunsaringkarn, T., Rungsiyothin, A. and Ruangrunsi, N. (2008). α -Glucosidase inhibitory activity of Thai Mimosaceae plant extracts. *Journal of Health Research*, 22(1), 29-33.
- Tyagi, S., Gupta, P., Saini, A.S., Kaushal, C. and Sharma, S. (2011). The peroxisome proliferator-activated receptor: A family of nuclear receptors role in various diseases. *Journal of Advance Pharmaceutical and Technological Research*, 2(4), 236-240.
- Ueno, H., Saitoh, Y., Mizuta, M., Shiiya, T., Noma, K. and Mashiba, S. (2011). Fenofibrate ameliorates insulin resistance, hypertension and novel oxidative stress markers in patients with metabolic syndrome. *Obesity Research and Clinical Practise*, 5(4), 267-360.

- Valavanis, I.K., Mougiakakou, S.G., Grimaldi, K.A. and Nikita, K.S. (2010). A multifactorial analysis of obesity as CVD risk factor: Use of neural network based methods in a nutrigenetics context. *BioMedCentral Bioinformatics*, 11(453), 1-10.
- Valko, M., Leibfritz, D., Moncol, J., Cronin, M.T.D., Mazur, M. and Telser, J. (2007). Free radicals and antioxidants in normal physiological functions and human disease. *International Journal of Biochemical Cell Biology*, 39, 44-84.
- Vanzo, A., Vrhovsek, U., Tramer, F., Mattivi, F. and Passamonti, S. (2011). Exceptionally fast uptake and metabolism of cyanidin 3-glucoside by rat kidneys and liver. *Journal of Natural Products*, 74(5), 1049-1054.
- Vasselli, J.R., Scarpace, P.J., Harris, R.B.S. and Banks, W.A. (2013). Dietary components in the development of leptin resistance. *Advances in Nutrition*, 4, 164-175.
- Vecina, J.F., Oliveira, A.G., Araujo, T.G., Baggio, S.R., Torello, C.O., Saad, M.J.A. and de Souza Queiroz, M.L. (2014). Chlorella modulates insulin signaling pathway and prevents high-fat diet-induced insulin resistance in mice. *Life Sciences*, 95, 45-52.
- Verma, R.K. and Paraidathathu, T. (2014). Herbal medicines used in the traditional indian medicinal system as a therapeutic treatment option for overweight and obesity management: A review. *International Journal of Pharmacy and Pharmaceutical Sciences*, 6(2), 40-47.
- Vladimir-Knežević, S., Blažeković, B., Štefan, M.B. and Babac, M. (2012). Plant polyphenols as antioxidants influencing the human health. *Phytochemicals as Nutraceuticals – Global Approaches to Their Role in Nutrition and Health*, 155-180.
- Wang, C.Y. and Liao, J.K. (2012). A mouse model of diet-induced obesity and insulin resistance. *Methods in Molecular Biology*, 821, 421-433.
- Wang, Q., Zhang, M., Xu, M., Gu, W., Xi, Y., Qi, L., Li, B. and Wang, W. (2015). Brown adipose tissue activation is inversely related to central obesity and metabolic parameters in adult human. *PLoS ONE*, 10(4), 1-13.
- Wang, S., Moustaid-Moussa, N., Chen, L., Mo, H., Shastri, A., Su, R., Bapat, P., Kwun, I. and Shen, C.L. (2014). Novel insights of dietary polyphenols and obesity. *Journal of Nutritional Biochemistry*, 25, 1-18.
- Wang, X., Xu, J., Yang, M. and Zhou, H. (2011). Chloroform extract of Tibetan herbal medicine *Dracocephalum tanguticum* Maxim. inhibits proliferation of T98G glioblastomas cells by modulating Caspase-3 cleavage and expression of Bax and p21. *Journal of Medicinal Plants Research*, 5(25), 6024-6031.
- Wang, Y., Beydoun, M.A., Liang, L., Caballero, B. and Kumanyika, S.K. (2008). Will all Americans become overweight or obese? Estimating the progression and cost of the US obesity epidemic. *Obesity*, 16, 2323-2330.
- Wang, Y. and Huang, F. (2015). N-3 polyunsaturated fatty acids and inflammation in obesity: Local effect and systemic benefit. *BioMedCentral Research International*, 581469, 1-16.

- Wang, Y.C., McPherson, K., Matsh, T., Gortmaker, S.L. and Brown, M. (2011). Health and economic burden of the projected obesity trends in the USA and the UK. *Lancet*, 378, 815-825.
- Watanabe, H., Nakano, T., Saito, R., Akasaka, D., Saito, K., Ogasawara, H., Minashima, T., Miyazawa, K., Kanaya, T., Takakura, I., Inoue, N., Ikeda, I., Chen, X., Miyake, M., Kitazawa, H., Shirakawa, H., Sato, K., Tahara, K., Nagasawa, Y., Rose, M.T., Ohwada, S., Watanabe, K. and Aso, H. (2016). Serotonin improves high fat diet induced obesity in mice. *PLoS ONE*, 11(1), 1-14.
- Weaver, J.U. (2008). Classical endocrine disease causing obesity. *Frontiers of Hormone Research*, 36, 212-228.
- Wei, Y., Chen, K., Adam, I., Whaley-Connell, T., Stump, C.S., Ibdah, J.A. and Sowers, J.R. (2007). Skeletal muscle insulin resistance: role of inflammatory cytokines and reactive oxygen species. *The American Journal of Physiology, Regulatory, Integrative and Comparative Physiology*, 294, 673-680.
- Weyer, C., Foley, J.E., Bogardus, C., Tataranni, P.A. and Pratley, R.E. (2000). Enlarged subcutaneous abdominal adipocyte size, but not obesity itself, predicts type II diabetes independent of insulin resistance. *Diabetologica*, 43, 1298-1506.
- Wilson, T.A., Nicolosi, R.J., Woolfrey, B. and Kritchevsky, D. (2007). Rice bran oil and oryzanol reduce plasma lipid and lipoprotein cholesterol concentrations and aortic cholesterol ester accumulation to a greater extent than ferulic acid in hypercholesteremic hamsters. *Journal of Nutritional Biochemistry*, 18, 105-112.
- Wiss, B.E. (2004). The inflammatory syndrome: the role of adipose tissue cytokines in metabolic disorders linked to obesity. *Journal of American Society of Nephrology*, 15, 2792-2800.
- Withrow, D. and Alter, D.A. (2011). The economic burden of obesity worldwide: a systematic review of the direct costs of obesity. *Obesity Review*, 12, 131-141.
- Wizturn, J.L. (1994). The oxidation hypothesis of atherosclerosis. *Lancet*, 344, 793-795.
- Woo, W.S. and Kang, S.S. (1984). Isolation of a new monoterpene conjugated triterpenoid from the stem bark of *Albizia julibrissin*. *Journal of Natural Products*, 47(3), 547-549.
- Xia, Q. and Grant, S.F.A. (2013). The genetics of human obesity. *Annals of the New York Academy of Sciences*, 1281, 178-190.
- Xiao, X., Li, M., Wang, K., Qin, Q. and Chen, X. (2011). Characterization of large yellow croaker (*Pseudosciaena crocea*) β -actin promoter supports β -actin gene as an internal control for gene expression modulation and its potential application in transgenic studies in fish. *Fish and Shellfish Immunology*, 30, 1072-1079.
- Xu, Z.J., Fan, J.G., Ding, X.D., Qiao, L. and Wang, G.L. (2010). Characterization of high-fat, diet-induced, non-alcoholic steatohepatitis with fibrosis in rats. *Digestive Disease and Sciences*, 55, 931-940.

- Yadav, S.S., Galib, Prajapati, P.K. and Harisha, C.R. (2011). Pharmacognostical screening and phytochemical evaluation of *Albizia lebbek* Benth.Heartwood.*Asian Journal of Biomedical and Pharmaceutical Sciences*.1(5), 1-6.
- Yaturu, S. (2011).Obesity and type 2 diabetes.*Journal of Diabetes Mellitus*, 1(4), 79-95.
- Yoshikawa, M., Morikawa, T., Nakano, K., Pongpiriyadacha, Y., Murakami, T. and Matsuda, H. (2002).Characterization of new sweet triterpene saponins from *Albizia myriophylla*. *Journal of Natural Products*, 65(11), 1638-1642.
- Zamora, R. and Hidalgo, F.J. (2016).Lipoproteins.*Reference Module in Food Science, from Encyclopedia of Food and Health*, 544-549.
- Zhang, G.F., Yang, Z.B., Wang, Y., Yang, W.R., Jiang, S.Z. and Gai, G.S. (2009). Effects of ginger root (*Zingiber officinale*) processed to different particle sizes on growth performance, antioxidant status and serum metabolites of broiler chickens. *Poultry Science*, 88, 2159-2166.
- Zhang, H.H., Halbleib, M., Ahmad, F., Manganiello, V.C. and Greenberg, A.S. (2002). Tumour necrosis factor- α stimulates lipolysis in differentiated human adipocytes through activation of extracellular signal-related kinase and elevation of intracellular cAMP. *Diabetes*, 51, 2929-2935.
- Zheng, C.D., Duan, Y.Q., Gao, J.M. and Ruan, Z.G. (2010). Screening for anti-lipase properties of 37 traditional Chinese medicinal herbs.*Journal of Chinese Medical Association*, 73(6).319-324.
- Zhou, Y. and and Rui, L. (2013).Leptin signaling and leptin resistance.*Frontier Medicine*, 7(2), 207–222.
- Zhu, X.X., Ma, S.T., Eirin, A., Woolard, J.R., Hickson, L.J., Sun, D., Lerman, A. and Lerman, L.O. (2016).Functional plasticity of adipose-derived stromal cells during development of obesity.*Stem Cells Translational Medicine*, 5, 893-900.
- Zinman, B., Hanley, A.J., Harris, S.B., Kwan, J. and Fantus, I.G. (1999).Circulating tumor necrosis factor α concentrations in a native Canadian population with high rates of type 2 diabetes mellitus.*Journal of Clinical Endocrinology and Metabolism*, 84, 272–278.
- Zubairi, S.I., Sarmidi, M.R. and Aziz, R.A. (2014). The effects of raw material particles size, types of solvents and solvent-to-solid ratio on the yield of rotenone extracted from *Derris elliptica* roots. *Sains Malaysiana*, 43, 707-71