



UNIVERSITI PUTRA MALAYSIA

**CHARACTERIZATION AND CYTOTOXICITY OF CLAUSENIDIN
FROM *Clausena excavata* Burm f. AND ITS EFFECTS ON CELL CYCLE
REGULATION AND APOPTOSIS OF LIVER (HEPG2) AND COLON
(HT29) CANCER CELL LINES.**

PETER MAITALATA WAZIRI

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**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

February 2017

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DEDICATION

This work is dedicated first and foremost to God Almighty Whose grace and providence made this project a successful voyage. It is also dedicated to everyone who loves knowledge.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment
of the requirement for the degree of Doctor of Philosophy.

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By

PETER MAITALATA WAZIRI

February 2017

Chairman : Professor Rasedee Abdullah, PhD
Faculty : Veterinary Medicine

Clausena excavata Burm. f. is a wild shrub from the *Rutaceae* family predominantly found in tropical Asia. The plant is traditionally used in the treatment of cancers; however, its mechanism of anticancer action is still unknown. Among phytochemicals present in *C. excavata* are alkaloids, coumarins and limonoid. Clausenidin, a pyranocoumarin isolated from *C. excavata* is postulated to have anticancer effects. Thus, the objective of this study is to determine the *in vitro* anticancer effect of clausenidin on the liver (HepG2) and colon (HT-29) cancer cell lines.

The cytotoxicity and effect of clausenidin on the HepG2 and HT-29 cell cycles were determined via acridine orange/propidium iodide, reactive oxygen species (ROS), annexin V and cell cycle assays. DNA fragmentation and ultrastructural analyses, caspase-3 and -9 as well as MMP assays of the clausenidin-treated HepG2 and HT-29 cells were also performed to determine the mode of cell death. In addition, apoptosis-related genes and proteins were also analyzed using qPCR and Western blot respectively to further verify the effects of clausenidin on HepG2 and HT-29 cells.

The IC₅₀ of clausenidin in HepG2 and HT-29 cells was 7.7 and 13.8 µg/mL at 72 hours of treatment, respectively. The results reveal that clausenidin induced G0/G1 and G2/M cell cycle arrest of HepG2 cells in a dose- and time-dependent manner. Clausenidin also caused depolarization of the mitochondrial membrane that resulted in the release of cytochrome C and significant ($p<0.05$) upregulation of Bax, Apaf-1, Smac, Caspase-3 and -9 proteins, suggesting involvement of the mitochondria pathway of apoptosis. It was also observed that the caspase-8, TNFR1, TRAIL, FADD and Fas proteins, which are key regulators of the extrinsic pathway of apoptosis were upregulated. Gene studies showed significant ($p<0.05$) increases in caspase-8 and -9 expression that further corroborates the involvement of the extrinsic and intrinsic

pathways in the apoptosis of clausenidin-treated HepG2 cells. The JNK, Bax, Apaf 1, cytochrome C, p53 and p21 genes were also found to be significantly ($p<0.05$) upregulated while Bcl-2, Bcl-x and HSP70 were downregulated. The findings suggest that clausenidin also suppressed the inducers of angiogenesis in HepG2 cells. Clausenidin-mediated apoptosis was evident by the increase in DNA fragmentation, typical microscopic and ultrastructural features of apoptosis.

Clausenidin induced responses in the HT-29 cells like that of the HepG2 cells, except that it did not cause significant ($p>0.05$) change in caspase-8, JNK and VEGF protein expressions. The HT-29 cells treated with clausenidin entered a G0/G1 cell cycle arrest. These cells also underwent depolarization of mitochondrial membrane resulting in cytochrome C release and subsequent increase in caspase-9 and Bax protein expressions that resulted in the activation of the intrinsic pathway of apoptosis. Clausenidin induced activation of caspase-3 that caused fragmentation of DNA and nuclei of the HT-29 cells, which are hallmarks of apoptosis. Upon treatment with clausenidin, HT-29 cells also showed increased production of ROS that is postulated to contribute to their death. The clausenidin-treated HT-29 cells like the HepG2 cells showed typical features of apoptosis although some cells underwent necrosis.

In conclusion, the study showed clausenidin isolated from *C. excavata* induced death of the HepG2 and HT-29 cells especially via apoptosis. Thus, clausenidin has the potential to be developed as a therapeutic compound for the treatment of liver and colon cancers.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENCIRIAN DAN KESITOKSIKAN KLAUSENIDIN YANG DIEKSTRAK
DARIPADA *Clausena excavata* Burm f. DAN KESANNYA TERHADAP
PENGAWALATURAN KITARAN SEL DAN APOPTOSIS SEL KANSER
HATI (HEPG2) DAN KOLON (HT29)**

Oleh

PETER MAITALATA WAZIRI

Februari 2017

**Pengerusi : Profesor Rasedee Abdullah, PhD
Fakulti : Perubatan Veterinar**

Clausena excavata Burm. f. adalah pokok renek liar daripada keluarga *Rutaceae* yang kebanyakannya ditemui di Asia tropika. Tumbuhan ini digunakan secara tradisional untuk merawat kanser; walau bagaimanapun, mekanisme tindakan anti-kansernya masih belum diketahui. Antara fitokimia di dalam *C. excavata* adalah alkaloid, koumarin dan limonoid. Klausenidin, suatu piranokoumarin yang diasing daripada *C. excavata* dipostulat mempunyai kesan antikanser. Oleh itu, objektif kajian ini ialah untuk menentukan kesan antikanser klausenidin secara *in vitro*.

Kesitotoksikan dan kesan klausenidin terhadap kitaran sel HepG2 dan HT-29 ditentukan melalui assai aridina jingga/propidium iodida, spesies oksigen reaktif (ROS), aneksin V, dan kitaran sel. Analisis penyepihan DNA dan ultrastruktur, dan assai kaspase 3 dan 9, dan MMP terhadap HepG2 dan HT-29 terperlaku klausenidin dilakukan juga untuk menentutkan cara kematian sel. Di samping itu, gen dan protein berkaitan apoptosis masing-masing dianalisis menggunakan qPCR dan sap Western untuk mengesahkan kesan klausenidin terhadap sel HepG2 dan HT-29.

Nilai IC₅₀ klausenidin terhadap sel HepG2 dan HT-29 masing-masing adalah dengan 7.7 dan 13.8 µg/mL, selepas 72 jam perlakuan. Hasil kajian menunjukkan klausenidin mengaruh hentian kitaran sel pada fasa G0/G1 dan G2/M secara sandaran dos dan masa. Klausenidin juga menyebabkan penyahkutuhan membran mitokondrion yang membawa kepada pembebasan sitokrom C dan peningkatan secara tererti ($p<0.05$) pengaturan naik protein Bax, Apaf -1, Smac, kaspase-3 dan -9 dan ini menyarankan penglibatan apoptosis arah laluan mitokondria. Peningkatan pengaturan naik protein kaspase 8, TNFR1, TRAIL, Fadd, dan Fas, yang merupakan pengatur utama apoptosis laluan ekstrinsik, telah juga dicerapkan. Kajian gen menunjukkan peningkatan tererti ($p<0.05$) dalam pernyataan kaspase-8 dan -9 and ini lagi mengesahkan penglibatan

apoptosis laluan ekstrinsik dan intrinsik sel HepG2 terpelaku klausenidin. Pengaturan naik Gen JNK, Bax, Apaf 1, sitokrom C, p53, dan p21 didapati berlaku secara tererti ($p<0.05$) manakala Bcl-2, Bcl-x, dan HSP70 mengalami pengaturan turun. Hasil kajian menyarankan yang klausenidin juga menindas pengaruh angiogenesis pada sel HepG2. Apoptosis teraruh klausenidin jelas berlaku dengan peningkatan penyerpihan DNA, ciri mikroskopi and ultrastruktur apoptosis yang tipikal.

Klausenidin mengaruh gerak balas sel HT-29 sama seperti yang berlaku terhadap sel HepG2, kecuali ianya tidak menyebabkan perubahan tererti ($p>0.05$) dalam penyataan protein kaspase-8, JNK, dan VEGF. Sel HT-29 terperlaku klausenidin mengalami hentian fasa G0/G1 dalam kitaran selnya. Sel ini juga mengalami penyahkutuhan membran mitokondrion yang menyebabkan pembebasan sitokrom C dan seterusnya peningkatan dalam penyataan protein kaspase-9 dan Bax yang membawa kepada pengaktifan arah lalaun intrinsik apoptosis. Klausenidin mengaruhkan pengaktifan kaspase-3 yang menyebabkan penyerpihan DNA dan nukleus sel HT-29, iaitu petanda pasti apoptosis. Apabila terperlaku klausenidin, sel HT-29 juga menunjukkan peningkatan pengeluaran ROS yang dipostulatkan sebagai penyumbang kepada kematianya. Sel HT-29 terperlaku klausenidin seperti sel HepG2 menunjukkan ciri apoptosis yang tipikal walaupun sesetengahnya mengalami nekrosis.

Kesimpulannya, kajian ini menunjukkan klausenidin yang diasingkan daripada *C. excavata* mengaruh kematian sel HepG2 dan HT-29 khususnya melalui apoptosis. Oleh itu, klausenidin berpotensi untuk dibangunkan sebagai sebatian terapeutik untuk rawatan kanser hati dan kolon.

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I certify that a Thesis Examination Committee has met on 17 February 2017 to conduct the final examination of Peter Maitalata Waziri on his thesis entitled "Characterization and Cytotoxicity of Clausenidin from *Clausena excavate* Burm.f. and its Effects on Cell Cycle Regulation and Apoptosis of Liver (HepG2) and Colon (HT29) Cancer Cell Lines" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

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LIST OF ABBREVIATIONS

Bax	Bcl-2 associated protein
Bcl-2	B-cell lymphoma 2
CAD	Caspase activated DNase
CDK	Cyclin dependent kinase
DIMS	Direct Infusion Mass spectra
DMSO	Dimethyl sulphoxide
DNA	Deoxyribonucleic acid
EDTA	Ethylene diamine tetraacetic acid
FITC	Fluorescene isothiocyanate
HepG2	Human hepatocellular carcinoma
HT-29	Human adenocarcinoma cells
HSP	Heat shock proteins
IC ₅₀	Inhibitory concentration (50%)
MMP	Mitochondrial Membrane Potential
mRNA	Messenger Ribonucleic acid
MTT	3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide
NMR	Nuclear Magnetic Resonance
OD	Optical density
PBS	Phosphate Buffered Saline
ROS	Reactive oxygen species
TAE	Tris acetate EDTA
Tris-HCl	Tris hydrochloric acid
TLC	Thin Layer Chromatography
VEGF	Vascular endothelial growth factor

CHAPTER ONE

INTRODUCTION

1.1 Overview of *Clausena excavata* Burm. f.

Clausena excavata Burm.f. is a wild medicinal shrub from the *Rutaceae* family. It is locally known as *Chemen* or *Cherek Hitam* in Malaysia and *San Soak* in Thailand (Descola and Pálsson, 1996; Khare, 2008). *Clausena* is a genus of 14 species predominantly found in Tropical Asia (Shier, 1983; Arbab *et al.*, 2012). In Thailand, the extracts of *Clausena excavata* is used for the treatment of cancers. The use of this plant in traditional medicine is based on hearsay and previous experience and with little or no scientific evidence supporting its therapeutic uses. The phytochemistry of *C. excavata* are currently being extensively studied and many secondary metabolites that are beneficial to health have been identified through various extraction and identification techniques (Zhi, 2006; Kongkathip *et al.*, 2010; Arbab *et al.*, 2013). Coumarins, alkaloids and limonoids are among the most abundant natural products in *C. excavata*. Recently, the effect of the extracts of *C. excavata* and some of its compounds were determined *in vitro* on a panel of cancer cell lines (Wu *et al.*, 1994; Ali *et al.*, 2000; Su *et al.*, 2009). Clausenidin is one of the compounds identified as an anti-cancer agent that contributes to the therapeutic effect of *C. excavata*. However, the mechanism of action of clausenidin is still unknown.

1.2 Common cancers affecting man

Cancer is a group of diseases characterized by uncontrolled cell growth and division and formation of tumors that invade and destroy tissues. The disease is a global problem and affects quality of life (Connolly and Rose, 1998). Based on WHO reports, the number of cancer deaths worldwide exceeds that from coronary heart disease and stroke combined together (Society, 2013).

The rate of increase in cancer cases is alarming and by the GLOBOCAN's estimates, in 2012 over 14 million cancer cases have been diagnosed, of which 8 million deaths were mainly the result of lung, breast, liver and colon cancers (Ferlay *et al.*, 2013). The cost of management of cancer amounts to approximately USD125 billion globally in 2010 and it is projected to increase to USD158 billion by year 2020 (Jemal *et al.*, 2004). Cancer management is costly, as such, cancer therapy remains an exclusive privilege of the well-developed and wealthy nations of the world. Ministry of Health of Malaysia reported that malignant neoplasms constitute the third major leading cause of death in Peninsular Malaysia (Omar *et al.*, 2006). The situation is even worse in Africa where cancer burden is expected to increase by 85% by the year 2030 (Morhason-Bello *et al.*, 2013).

Liver and colon cancers are peculiar to Africa and Malaysia, respectively. Liver cancer usually appear in form of hepatocellular carcinoma (HCC), hepatoblastoma, or

cholangiocarcinoma (Jemal *et al.*, 2009). The cancer more often affects men than women (Society, 2013). Among the causes of liver cancers is chronic viral hepatitis. Liver cancers, like other cancer are treated by surgery, radiotherapy and chemotherapeutic drugs.

Colon cancer cases are also on the increase worldwide. The cancer is also known as colorectal cancer because carcinomas of the colon and rectum have similar pathology. Over 95% of colon cancers are adenocarcinomas (Society, 2013). Among major risk factors to colon cancer are diet, age and unhealthy lifestyle. Survivability from colon cancers is poor because there is no single efficient method that could be used to permanently remove the cancer tissues.

1.3 Anticancer compounds of plant sources

According WHO reports, 80% of the world's population depend largely on plant-based traditional medicine (Newman *et al.*, 2003; Koehn and Carter, 2005). These medicinal plants contain phytochemicals and nutrients and while curing diseases are also nutritious (Taylor *et al.*, 2001; Krief *et al.*, 2005). Historically, plants are the oldest and most sought sources of chemotherapeutic compounds. Currently, there are an estimated 200,000 to 500,000 plant species; some of which are potential sources of therapeutic compounds (Raven, 1988; Borris, 1996). Less than 10% of these plants are consumed as food and approximately 20% are being used for therapeutic purposes (Moerman *et al.*, 1996). Some herbs and plants were shown to be effective in the treatment of early stage cancers (Cragg and Newman, 2005). About 115,000 plant extracts from over 30,000 plant species globally have been screened for their anticancer properties by the United States National Cancer Institute (Shoeb, 2006; Tan and Zhou, 2006).

In tropical Asia, *Clausena excavata* Burm f. is commonly used for the treatment of different ailments. The use of this plant is an ancient folklore that is still practiced to date. About four pyranocoumarins were isolated from *C. excavata* and their effect on cancers were determined *in vitro* on a panel of cancer cell lines (Su *et al.*, 2009). Among these pyranocoumarins is clausenidin that was postulated to have anti-cancer properties. In our study, clausenidin was extracted from fresh roots of *C. excavata* and purified to its crystalline form. To determine the potential of clausenidin as an anticancer agent, its effects were investigated *in vitro* using the liver (HepG2) and colon (HT-29) cancer cell lines.

1.4 Research problem

Current chemotherapeutic drugs are plagued with undue side-effects and eventual development of drug-resistance. The anticancer drug currently in use are expensive and not affordable by people of low income groups of which the majority are in underdeveloped and developing countries.

1.5 Hypothesis

1.5.1 Null Hypothesis

Clausenidin is not cytotoxic to cancer cells.

1.5.2 Alternate Hypothesis

Clausenidin from *C. excavata* is a potent inhibitor of tumor cell growth and cancer spread through the production proteins of the apoptosis pathways.

1.6 Objective

The general objective of the study is to evaluate the cytotoxic and anti-proliferative effects of clausenidin on liver and colon cancer cell lines. The specific objectives are to:

1. characterize clausenidin isolated from *Clausena excavata*.
2. determine the cytotoxic effects of clausenidin on the HepG2 and HT-29 cell lines *in vitro*.
3. determine the effects of clausenidin on genes and proteins involved in the regulation of cell cycle and apoptosis of the HepG2 and HT-29 cells.
4. determine the pathways involved in the death of HepG2 and HT-29 cells induced by clausenidin.

1.7 Justification

At present, cancer is synonymous with death sentence because of the devastating consequences of the disease. It is projected that by the year 2030, 17 million people would have died of cancers. The significant majority of cancer cases and deaths are expected to be in Asia and Africa, where good and structured health care and services are not in place. The cost of treatment and management of the disease is enormous, especially with the use of costly modern drugs. In addition, modern drugs have deleterious effects that can lower quality of life of cancer patients.

Therapeutic compounds from natural sources are not only in abundance but also cheap to produce. Cheap drugs and compounds will benefit poor societies either by extending life of cancer patients or improving quality of life. Thus, the discovery of natural anticancer compounds may alleviate the sufferings from this debilitating disease.

REFERENCES

- Akerele, O. (1993). Natures medicinal bounty: dont throw it away. *World Health Forum*, 14. 390-395.
- Alwan, A. (2011). Global status report on noncommunicable diseases *World Health Organization*, 100-108, http://www.who.int/nmh/publications/ncd_report2010/en/. (accessed on 31 October, 2016).
- Ali, A. M., Ismail, N. H., Mackeen, M., Yazan, L., Mohamed, S., Ho, A. and Lajis, N. (2000). Antiviral, cytotoxic and antimicrobial activities of anthraquinones isolated from the roots of *Morinda elliptica*. *Pharmaceutical Biology*, 38, 298-301.
- Amin, F., Bowen, I. D., Szegedi, Z., Mihalik, R. and Szende, B. (2000). Apoptotic and non-apoptotic modes of programmed cell death in MCF-7 human breast carcinoma cells. *Cell Biology International*, 24, 253-260.
- Arbab, I. A., Abdul, A. B., Aspollah, M., Abdullah, R., Abdelwahab, S. I., Ibrahim, M. Y. and Ali, L. Z. (2012). A review of traditional uses, phytochemical and pharmacological aspects of selected members of *Clausena* genus (*Rutaceae*). *Journal of Medicinal Plants Research*, 6(38), 5107-5118.
- Arbab, I. A., Abdul, A. B., Sukari, M. A., Abdullah, R., Syam, S., Kamalidehghan, B., Ibrahim, M. Y., Taha, M. M. E., Abdelwahab, S. I. and Ali, H. M. (2013). Dentatin isolated from *Clausena excavata* induces apoptosis in MCF-7 cells through the intrinsic pathway with involvement of NF-κB signalling and G0/G1 cell cycle arrest: a bioassay-guided approach. *Journal of Ethnopharmacology*, 145, 343-354.
- Ashhab, Y., Alian, A., Polliack, A., Panet, A. and Yehuda, D. B. (2001). Two splicing variants of a new inhibitor of apoptosis gene with different biological properties and tissue distribution pattern. *Federation of European Biochemical Societies Letters*, 495(1-2), 56-60.
- Au-Yeung, K. K. W., Liu, P. L., Chan, C., Wu, W. Y., Lee, S. S. T. and Ko, J. K. S. (2008). Herbal isoprenols induce apoptosis in human colon cancer cells through transcriptional activation of PPAR γ . *Cancer Investigation*, 26(7), 708-717.
- Auyeung, K. K. W. and Ko, J. K. S. (2010). Novel herbal flavonoids promote apoptosis but differentially induce cell cycle arrest in human colon cancer cell. *Investigational New Drugs*, 28(1), 1-13.
- Avendaño, C. and Menendez, J. C. (2015). Medicinal chemistry of anticancer drugs. Hungary: Elsevier; 2015. Pp95-100.

- Bai, L. and Zhu, W. G. (2006). P53: structure, function and therapeutic applications. *Journal of Cancer Molecules*, 2(4), 141-153.
- Balandrin, M. F., Klocke, J., Wurtele, E. S. and Bollinger, W. H. (1985). Natural plant chemicals: sources of industrial and medicinal materials. *Journal of Science(Washington)*, 228(4704), 1154-1159.
- Bamias, A. and Dimopoulos, M. A. (2003). Angiogenesis in human cancer: implications in cancer therapy. *European Journal of Internal Medicine*, 14(8), 459-469.
- Beckner, M. E. and Liotta, L. A. (1997). Growth factors and cytokines in tumor invasion and metastasis. *Growth Factors and Cytokines in Health and Disease*, 3, 381-437.
- Beekman, K. W. and Hussain, M. (2008). Hormonal approaches in prostate cancer: application in the contemporary prostate cancer patient. *Urologic Oncology: Seminars and Original Investigations*, 26, 415-419.
- Beere, H. M., Wolf, B. B., Cain, K., Mosser, D. D., Mahboubi, A., Kuwana, T., Tailor, P., Morimoto, R. I., Cohen, G. M. and Green, D. R. (2000). Heat Shock Protein 70 inhibits apoptosis by preventing recruitment of procaspase-9 to the Apaf-1 apoptosome. *Nature Cell Biology*, 2, 469-475.
- Beesoo, R., Neergheen-Bhujun, V., Bhagooli, R. and Bahorun, T. (2014). Apoptosis inducing lead compounds isolated from marine organisms of potential relevance in cancer treatment. *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, 768, 84-97.
- Behzadian, M. A., Windsor, L. J., Ghaly, N., Liou, G., Tsai, N. T. and Caldwell, R. B. (2003). VEGF-induced paracellular permeability in cultured endothelial cells involves urokinase and its receptor. *The FASEB Journal*, 17(6), 752-754.
- Benchimol, S. and Minden, M. D. (1998). Viruses, Oncogenes and tumor suppressor genes. The basic science of oncology. McGraw Hill, Pp79-105.
- Bisacchi, D., Benelli, R., Vanzetto, C., Ferrari, N., Tosetti, F. and Albini, A. (2003). Anti-angiogenesis and angioprevention: mechanisms, problems and perspectives. *Cancer Detection and Prevention*, 27(3), 229-238.
- Bishop, J. M. and Weinberg, R. A. (1996). Scientific American molecular oncology: Scientific American, Mosby.
- Borris, R. P. (1996). Natural products research: perspectives from a major pharmaceutical company. *Journal of Ethnopharmacology*, 51(1), 29-38.
- Bowen, I., Bowen, S. and Jones, A. (1998). The cell cycle. Mitosis and apoptosis: Matters of life and death.(Eds. Bowen Id, Bowen Sm). London, New York: Chapman and Hall, Pp28-59.

- Bowen, I. D. (1990). Programmed Cell Death In Tumours And Tissues, Springer Science and Business Media.
- Bowen, I. D., Mullarkey, K. and Morgan, S. (1996). Programmed cell death during metamorphosis in the blow-fly *Calliphora vomitoria*. *Microscopy Research and Technique*, 34, 202-217.
- Bryant, J. A., Garland, J. M. and Hughes, S. G. (2000). *Programmed cell death in animals and plants*, Bios Scientific.
- Burri, P. H. and Djonov, V. (2002). Intussusceptive angiogenesis - the alternative to capillary sprouting. *Molecular Aspects of Medicine*, 23(6), 1-27.
- Carrasco, R. A., Stamm, N. B., Marcusson, E., Sandusky, G., Iversen, P. and Patel, B. K. (2011). Antisense inhibition of survivin expression as a cancer therapeutic. *Molecular Cancer Therapeutics*, 10, 221-232.
- Chen, Z., Naito, M., Hori, S., Mashima, T., Yamori, T. and Tsuruo, T. (1999). A human IAP-family gene, apollon, expressed in human brain cancer cells. *Biochemical and Biophysical Research Communications*, 264(3), 847-854.
- Clarke, A., Purdie, C., Harrison, D., Morris, R., Bird, C., Hooper, M. and Wyllie, A. (1993). Thymocyte apoptosis induced by p53-dependent and independent pathways. *Nature*, 362, 849.
- Connolly, J. M. and Rose, D. P. (1998). Angiogenesis in two human prostate cancer cell lines with differing metastatic potential when growing as solid tumors in nude mice. *The Journal of Urology*, 160(3), 932-936.
- Cordell, G. A., Beecher, C. W. and Pezzuto, J. M. (1991). Can ethnopharmacology contribute to the development of new anticancer drugs? *Journal of Ethnopharmacology*, 32(1-3), 117-133.
- Cragg, G. M. and Newman, D. J. (2005). Biodiversity: A continuing source of novel drug leads. *Pure and Applied Chemistry*, 77(1), 7-24.
- Crutchley, A., Thompson, A., Chan, H. and Reed, M. (2013). Current controversies in breast cancer surgery. *Clinical Oncology*, 25(2), 101-108.
- Danial, N. N. and Korsmeyer, S. J. (2004). Cell death: critical control points. *Cell*, 116(2), 205-219.
- De Leon, M. P., Di Gregorio, C., Roncucci, L., Benatti, P., Fante, R. and Rossi, G. (1998). Epidemiology of tumors of the colon and rectum. Incidence, mortality, familiarity and survival in the Health Care District of Modena, 1984–1995. In University did Modeme ARTI.
- Deorukhkar, A., Krishnan, S., Sethi, G. and Aggarwal, B. B. (2007). Back to basics: how natural products can provide the basis for new therapeutics. *Expert Opinion on Investigational Drugs*, 16(11), 1753-1773.

- Descola, P. and Pálsson, G. (1996). Nature and society: anthropological perspectives: In London: Routledge, Taylor and Francis. Pp529-544.
- Dewson, G. and Kluck, R. (2010). Bcl-2 family-regulated apoptosis in health and disease. *Cell Health and Cytoskeleton*, 2, 9-22.
- Disis, M. L. (2006). Immunotherapy of cancer. Springer, Humana press Inc., Pp 487-602.
- Dunn, S. E., Hardman, R. A., Kari, F. W. and Barrett, J. C. (1997). Insulin-like growth factor 1 (IGF-1) alters drug sensitivity of Hbl100 human breast cancer cells by inhibition of apoptosis induced by diverse anticancer drugs. *Cancer Research*, 57, 2687-2693.
- Earnshaw, W. C. (1995). Nuclear changes in apoptosis. *Current Opinion in Cell Biology*, 7, 337-343.
- El-deiry, W. S., Harper, J. W., O'connor, P. M., Velculescu, V. E., Canman, C. E., Jackman, J., Pietsch, J. A., Burrell, M., Hill, D. E. and Wang, Y. (1994). WAF1/CIP1 is induced in p53-mediated G1 arrest and apoptosis. *Cancer Research*, 54, 1169-1174.
- El-Serag, H. B. and Mason, A. C. (2000). Risk factors for the rising rates of primary liver cancer in the United States. *Archives of Internal Medicine*, 160(21), 3227-3230.
- Etkin, N. L. and Elisabetsky, E. (2005). Seeking a transdisciplinary and culturally germane science: The future of ethnopharmacology. *Journal of Ethnopharmacology*, 100(1), 23-26.
- Evan, G. I. and Vousden, K. H. (2001). Proliferation, cell cycle and apoptosis in cancer. *Nature*, 411, 342-348.
- Farber, E. (1956). Similarities in the sequence of early histological changes induced in the liver of the rat by ethionine, 2-acetylamino-fluorene and 3'-methyl-4 dimethylaminoazobenzene. *Cancer Research*, 16(2), 142-155.
- Farber, E. and Cameron, R. (1980). The sequential analysis of cancer development. *Advances in Cancer Research*, 31, 125-226.
- Ferlay, J., Shin, H. R., Bray, F., Forman, D., Mathers, C. and Parkin, D. M. (2010). Estimates of worldwide burden of cancer in 2008: Globocan 2008. *International Journal of Cancer*, 127, 2893-2917.
- Ferlay, J., Soerjomataram, I., Ervik, M., Dikshit, R., Eser, S., Mathers, C., Rebelo, M., Parkin, D., Forman, D. and Bray, F. (2013). GLOBOCAN 2012 v1. 0. *Cancer incidence and mortality worldwide: IARC CancerBase*, 127(12), 2893-2917.

- Festjens, N., Berghe, T. V., Cornelis, S. and Vandenabeele, P. (2007). RIP1, a kinase on the crossroads of a cell's decision to live or die. *Cell Death and Differentiation*, 14(3), 400-410.
- Feuerstein, G., Ruffolo, R. R. and Yue, T. L. (1997). Apoptosis and congestive heart failure. *Trends in Cardiovascular Medicine*, 7(7), 249-255.
- Fidler, I. J. (2003). The pathogenesis of cancer metastasis: the 'seed and soil' hypothesis revisited. *Nature Reviews Cancer*, 3(6), 453-458.
- Fink, S. L. and Cookson, B. T. (2005). Apoptosis, pyroptosis and necrosis: mechanistic description of dead and dying eukaryotic cells. *Infection and Immunity*, 73(4), 1907-1916.
- Folkman, J. (2002). Role of angiogenesis in tumor growth and metastasis. *Seminars in Oncology* 29(6), 15-18.
- Franceschi, S. and Wild, C. P. (2013). Meeting the global demands of epidemiologic transition—The indispensable role of cancer prevention. *Molecular Oncology*, 7(1), 1-13.
- Fridman, J. S. and Lowe, S. W. (2003). Control of apoptosis by p53. *Oncogene*, 22, 9030-9040.
- Friesen, C., Fulda, S. and Debatin, K. (1997). Deficient activation of the cd95 (apo-1/fas) system in drug-resistant cells. *Leukemia*, 11, 1833-1841.
- Fulda, S., Los, M. J., Friesen, C. and Debatin, K. M. (1998). Chemosensitivity of solid tumor cells in vitro is related to activation of the CD95 system. *International Journal of Cancer*, 76(1), 105-114.
- Fulda, S., Meyer, E. and Debatin, K. M. (2002). inhibition of TRAIL-induced apoptosis by bcl-2 overexpression. *Oncogene*, 21(15), 2283-2294.
- Fulda, S. (2010). Evasion of apoptosis as a cellular stress response in cancer. *International Journal of Cell Biology*, 2010, 1-6.
- Gangwar, R., Mandhani, A. and Mittal, R. D. (2009). Caspase-9 and caspase-8 gene polymorphisms and susceptibility to bladder cancer in north Indian population. *Annals of Surgical Oncology*, 16(7), 2028-2034.
- Gels, M. P. C. P. (2001). Molecupage precast protein gels. Technical Manual. Molecutools Ireland, Pp7-11.
- Gething, M. J. and Sambrook, J. (1992). Protein folding in the cell. *Nature*, 355, 33.
- Ghobrial, I. M., Witzig, T. E. and Adjei, A. A. (2005). Targeting apoptosis pathways in cancer therapy. *A Cancer Journal for Clinicians*, 55(3), 178-194.

- Gimlette, J. D. and Thomson, H. W. (1939). A Dictionary of Malayan Medicine. Humphrey Milford, Oxford University Press. Pp1287-1186.
- Giovannucci, E. (2002). Modifiable risk factors for colon cancer. *Gastroenterology Clinics of North America*, 31(4), 925-943.
- González-Sarriás, A., Li, L. and Seeram, N. P. (2012). Anticancer effects of maple syrup phenolics and extracts on proliferation, apoptosis and cell cycle arrest of human colon cells. *Journal of Functional Foods*, 4(1), 185-196.
- Grieve, C. M. and Scora, R. W. (1980). Flavonoid distribution in the Aurantioideae (*Rutaceae*). *Systematic Botany*, 5, 39-53.
- Gross, A., McDonnell, J. M. and Korsmeyer, S. J. (1999). Bcl-2 family members and the mitochondria in apoptosis. *Genes and Development*, 13(15), 1899-1911.
- Häcker, G. (2000). The morphology of apoptosis. *Cell and tissue research*, 301(1), 5-17.
- Hafidh, R. R., Abdulamir, A. S., Bakar, F. A., Jalilian, F. A., Jahanshiri, F., Abas, F. and Sekawi, Z. (2013). Novel anticancer activity and anticancer mechanisms of *Brassica oleracea* L. Var. Capitata F. Rubra. *European Journal of Integrative Medicine*, 5, 450-464.
- Hanahan, D. and Weinberg, R. A. (2000). The hallmarks of cancer. *Cell*, 100(1), 57-70.
- Harris, A. (2003). Angiogenesis as a new target for cancer control. *European Journal of Cancer Supplements*, 1(2), 1-12.
- Hartwell, L. H. and Kastan, M. B. (1994). Cell cycle control and cancer. *Science*, 266(5192), 1821-1828.
- He, H. P., Shen, Y. M., Zuo, G. Y., Zhu, W. M., Yang, X. S. and Hao, X. J. (2000). Two new O-terpenoidal coumarins, excavacoumarin A and B from *Clausena excavata*. *Journal of Heterocycles*, 53, 1807-1810.
- Hecht, J., Trarbach, T., Jaeger, E., Hainsworth, J., Wolff, R., Lloyd, K., Bodoky, G., Borner, M., Laurent, D. and Jacques, C. (2005). A randomized, double-blind, placebo-controlled, phase III study in patients (Pts) with metastatic adenocarcinoma of the colon or rectum receiving first-line chemotherapy with oxaliplatin/5-fluorouracil/leucovorin and PTK787/ZK 222584 or placebo (CONFIRM-1). Paper presented at the ASCO Annual Meeting Proceedings 23 (16), LB-A3. <http://www.oncologink.org/print/pdf/12776> (Accessed on 31 October 2016)
- Hengartner, M. O. (2001). Apoptosis: corralling the corpses. *Cell*, 104(3), 325-328.
- Hewitson, P., Glasziou, P., Watson, E., Towler, B. and Irwig, L. (2008). Cochrane systematic review of colorectal cancer screening using the fecal occult blood

- test (hemoccult): an update. *The American Journal of Gastroenterology*, 103(6), 1541-1549.
- Holmgren, L., Szeles, A., Rajnavölgyi, E., Folkman, J., Klein, G., Ernberg, I. and Falk, K. I. (1999). Horizontal transfer of DNA by the uptake of apoptotic bodies. *Blood*, 93(11), 3956-3963.
- Huang, S. C., Wu, P. L. and Wu, T. S. (1997). Two coumarins from the root bark of *Clausena excavata*. *Phytochemistry*, 44(1), 179-181.
- Hurley, K. E. and Chapman, P. B. (2005). Helping melanoma patients decide whether to choose adjuvant high-dose interferon- α 2b. *The Oncologist*, 10(9), 739-742.
- Hussain, S. P. and Harris, C. C. (1998). Molecular epidemiology of human cancer: contribution of mutation spectra studies of tumor suppressor genes. *Cancer Research*, 58, 4023-4037.
- Ickes, K., Dewalt, S. J. and Thomas, S. C. (2003). Resprouting of woody saplings following stem snap by wild pigs in a Malaysian rain forest. *Journal of Ecology*, 91(2), 222-233.
- Isa, N. M., Abdul, A. B., Abdelwahab, S. I., Abdullah, R., Sukari, M. A., Kamalidehghan, B., Hadi, A. H. A. and Mohan, S. (2013). Boesenbergin A, a chalcone from *Boesenbergia rotunda* induces apoptosis via mitochondrial dysregulation and cytochrome c release in A549 cells *in vitro*: involvement of hsp70 and bcl2/bax signalling pathways. *Journal of Functional Foods*, 5, 87-97.
- Ito, C., Itoigawa, M., Katsuno, S., Omura, M., Tokuda, H., Nishino, H. and Furukawa, H. (2000). Chemical constituents of *Clausena excavata*: isolation and structure elucidation of novel furanone-coumarins with inhibitory effects for tumor-promotion 1. *Journal of Natural Products*, 63, 1218-1224.
- Jatoi, A., Ellison, N., Burch, P. A., Sloan, J. A., Dakhil, S. R., Novotny, P., Tan, W., Fitch, T. R., Rowland, K. M. and Young, C. Y. (2003). A phase II trial of green tea in the treatment of patients with androgen independent metastatic prostate carcinoma. *Cancer*, 97, 1442-1446.
- Jemal, A., Clegg, L. X., Ward, E., Ries, L. A., Wu, X., Jamison, P. M., Wingo, P. A., Howe, H. L., Anderson, R. N. and Edwards, B. K. (2004). Annual report to the nation on the status of cancer, 1975–2001, with a special feature regarding survival. *Cancer*, 101, 3-27.
- Jemal, A., Siegel, R., Ward, E., Hao, Y., Xu, J. and Thun, M. J. (2009). Cancer statistics, 2009. *A Cancer Journal for Clinicians*, 59(4), 225-249.
- Johnstone, R. W., Ruefli, A. A. and Lowe, S. W. (2002). Apoptosis: A link between cancer genetics and chemotherapy. *Cell*, 108, 153-164.

- Joshi, B. and Kamat, V. (1966). Structures of clausenin, clausenidin and a synthesis of clausenin and xanthoxyletin. *Tetrahedron Letters*, 7(46), 5767-5773.
- Karp, G. (2008). Cell and Molecular Biology: Concepts and Experiments 5th Edition with Study. John Wiley and Sons, Pp653-657.
- Kaufmann, S. H. and Earnshaw, W. C. (2000). Induction of apoptosis by cancer chemotherapy. *Experimental Cell Research*, 256(1), 42-49.
- Khare, C. P. (2008). Indian medicinal plants: an illustrated dictionary. Springer-verlag, Pp294-299.
- Ko, L. J. and Prives, C. (1996). P53: puzzle and paradigm. *Genes and Development*, 10, 1054-1072.
- Koehn, F. E. and Carter, G. T. (2005). The evolving role of natural products in drug discovery. *Nature Reviews Drug Discovery*, 4(3), 206-220.
- Kongkathip, B., Kongkathip, N., Sunthitikawinsakul, A., Napaswat, C. and Yoosook, C. (2005). Anti-Hiv-1 constituents from *Clausena excavata*: carbazoles and a pyranocoumarin. *Phytotherapy Research*, 19, 728-731.
- Kongkathip, B., Suttiprabha, S., Yoosook, C., Mongkolsook, Y. and Kongkathip, N. (2010). Determination of a pyranocoumarin and three carbazole compounds in *Clausena excavata* by RP-HPLC. *Journal of Chromatographic Science*, 48(6), 445-449.
- Koopman, G., Reutelingsperger, C., Kuijten, G., Keehnen, R., Pals, S. and Van Oers, M. (1994). Annexin V for flow cytometric detection of phosphatidylserine expression on B cells undergoing apoptosis. *Blood*, 84, 1415-1420.
- Korsmeyer, S. J., Shutter, J. R., Veis, D. J., Merry, D. E. and Oltvai, Z. N. (1993). Bcl-2/Bax: a rheostat that regulates an anti-oxidant pathway and cell death. *Seminars in Cancer Biology*, 4(6), 327-332.
- Krepela, E., Dankova, P., Moravcikova, E., Krepelova, A., Prochazka, J., Cermak, J., Schutzner, J., Zatloukal, P. and Benkova, K. (2009). Increased expression of inhibitor of apoptosis proteins, survivin and XIAP, in non-small cell lung carcinoma. *International Journal of Oncology*, 35, 1449-1462.
- Krief, S., Hladik, C. M. and Haxaire, C. (2005). Ethnomedicinal and bioactive properties of plants ingested by wild chimpanzees in Uganda. *Journal of Ethnopharmacology*, 101(1), 1-15.
- Kroemer, G., El-Deiry, W., Golstein, P., Peter, M., Vaux, D., Vandenebeele, P., Zhivotovsky, B., Blagosklonny, M., Malorni, W. and Knight, R. (2005). Classification of cell death: recommendations of the nomenclature committee on cell death. *Cell Death and Differentiation*, 12, 1463-1467.

- Kroemer, G., Galluzzi, L. and Brenner, C. (2007). Mitochondrial membrane permeabilization in cell death. *Physiological reviews*, 87(1), 99-163.
- Kroemer, G., Galluzzi, L., Vandenabeele, P., Abrams, J., Alnemri, E., Baehrecke, E., Blagosklonny, M. V., El-Deiry, W. S., Goldstein, P. and Green, D. (2009). Classification of cell death: recommendations of the nomenclature committee on cell death 2009. *Cell Death and Differentiation*, 16(1), 3-11.
- Kuerbitz, S. J., Plunkett, B. S., Walsh, W. V. and Kastan, M. B. (1992). Wild-type p53 is a cell cycle checkpoint determinant following irradiation. *Proceedings of the National Academy of Sciences*, 89, 7491-7495.
- Kumar, V., Reisch, J. and Wickramasinghe, A. (2008). Glycomaurin and glycomaurrol, new carbazole alkaloids from *Glycosmis mauritiana* (Rutaceae) bark. *Australian Journal of Chemistry*, 42(8), 1375-1379.
- Kummalue, T., Pornchai, O., Jiratchariyakul, W., Chanchai, M., Pattanapanyasat, K., Sukapirom, K. and Iemsri, S. (2007). Antiproliferative effect of *Erycibe elliptilimba* on human breast cancer cell lines. *Journal of Ethnopharmacology*, 110(3), 439-443.
- LaCasse, E., Mahoney, D., Cheung, H., Plenchette, S., Baird, S. and Korneluk, R. (2008). IAP-targeted therapies for cancer. *Oncogene*, 27(48), 6252-6275.
- Lane, D. P. (1992). Cancer. p53, guardian of the genome. *Nature*, 358, 15-16.
- Lanza, R. (2013). Essentials of stem cell biology 3rd edition. Academic Press, Pp95-106.
- Lavrik, I., Golks, A. and Krammer, P. H. (2005). Death receptor signaling. *Journal of Cell Science*, 118(2), 265-267.
- Lavrik, I. N., Golks, A. and Krammer, P. H. (2005). Caspases: pharmacological manipulation of cell death. *The Journal of Clinical Investigation*, 115(10), 2665-2672.
- Lengauer, C., Kinzler, K. W. and Vogelstein, B. (1998). Genetic instabilities in human cancers. *Nature*, 396(6712), 643-649.
- Levine, A. J. (1997). P53, the cellular gatekeeper for growth and division. *Cell*, 88(3), 323-331.
- Levine, A. J., Momand, J. and Finlay, C. A. (1991). The p53 tumour suppressor gene. *Nature*, 351(6326), 453-456.
- Li, S., Zhao, Y., He, X., Kim, T. H., Kuharsky, D. K., Rabinowich, H., Chen, J., Du, C. and Yin, X. M. (2002). Relief of extrinsic pathway inhibition by the Bid-dependent mitochondrial release of Smac in Fas-mediated hepatocyte apoptosis. *Journal of Biological Chemistry*, 277, 26912-26920.

- Liamarkopoulos, E., Gazouli, M., Aravantinos, G., Tzanakis, N., Theodoropoulos, G., Rizos, S. and Nikiteas, N. (2011). Caspase-8 and caspase-9 gene polymorphisms and susceptibility to gastric cancer. *Gastric Cancer*, 14(4), 317-321.
- Liekens, S., De Clercq, E. and Neyts, J. (2001). Angiogenesis: regulators and clinical applications. *Biochemical Pharmacology*, 61(3), 253-270.
- Lin, L., Zhou, D., Liu, K., Wang, F., Lan, S. and Ye, X. (2005). [Analysis on the prognostic factors in patients with large hepatocarcinoma treated by shentao ruangan pill and hydroxycamptothecine]. *Zhongguo Zhong xi yi jie he za zhi Zhongguo Zhongxiyi jiehe zazhi= Chinese journal of integrated traditional and Western medicine/Zhongguo Zhong xi yi jie he xue hui, Zhongguo Zhong yi yan jiu yuan zhu ban*, 25(1), 8-11.
- Loeb, L. A. (1991). Mutator phenotype may be required for multistage carcinogenesis. *Cancer Research*, 51(12).
- Look, A. T., Hayes, F. A., Shuster, J. J., Douglass, E. C., Castleberry, R. P., Bowman, L., Smith, E. and Brodeur, G. (1991). Clinical relevance of tumor cell ploidy and N-myc gene amplification in childhood neuroblastoma: a pediatric oncology group study. *Journal of Clinical Oncology*, 9, 581-591.
- Lopes, R. B., Gangeswaran, R., McNeish, I. A., Wang, Y. and Lemoine, N. R. (2007). Expression of the IAP protein family is dysregulated in pancreatic cancer cells and is important for resistance to chemotherapy. *International Journal of Cancer*, 120(11), 2344-2352.
- Lynch, H. T., Schuelke, G. S., Kimberling, W. J., Albano, W. A., Lynch, J. F., Biscone, K. A., Lipkin, M. L., Deschner, E. E., Mikol, Y. B. and Sandberg, A. A. (1985). Hereditary nonpolyposis colorectal cancer (Lynch syndromes I and II). II. Biomarker studies. *Cancer*, 56(4), 939-951.
- Ma, J. and Waxman, D. J. (2008). Combination of antiangiogenesis with chemotherapy for more effective cancer treatment. *Molecular Cancer Therapeutics*, 7(12), 3670-3684.
- Maecker, H. L., Koumenis, C. and Giaccia, A. J. (2000). P53 promotes selection for Fas-mediated apoptotic resistance. *Cancer Research*, 60, 4638-4644.
- Majno, G. and Joris, I. (1995). Apoptosis, oncosis and necrosis. An overview of cell death. *The American Journal of Pathology*, 146(1), 3-15.
- Manosroi, A., Saraphanchotiwitthaya, A. and Manosroi, J. (2005). *In vitro* immunomodulatory effect of *Pouteria cambodiana* (Pierre Ex Dubard) baehni extract. *Journal of Ethnopharmacology*, 101, 90-94.
- Manser, C. N., Bachmann, L. M., Brunner, J., Hunold, F., Bauerfeind, P. and Marbet, U. A. (2012). Colonoscopy screening markedly reduces the occurrence of

- colon carcinomas and carcinoma-related death: a closed cohort study. *Gastrointestinal Endoscopy*, 76(1), 110-117.
- Marmot, M., Atinmo, T., Byers, T., Chen, T., Hirohata, T., Jackson, A., James, W., Kolonel, L., Kumanyika, S., Leitzmann, C. and Mann, J. (2007). Food, nutrition, physical activity and the prevention of cancer: a global perspective. *American Institute for Cancer Research*, 1, 277-280.
- Marsden, V. S., O'Connor, L., O'Reilly, L. A., Silke, J., Metcalf, D., Ekert, P. G., Huang, D. C., Cecconi, F., Kuida, K. and Tomaselli, K. J. (2002). Apoptosis initiated by bcl-2-regulated caspase activation independently of the cytochrome c/apaf-1/caspase-9 apoptosome. *Nature*, 419, 634-637.
- Matsuyama, R., Reddy, S. and Smith, T. J. (2006). Why do patients choose chemotherapy near the end of life? A review of the perspective of those facing death from cancer. *Journal of Clinical Oncology*, 24(21), 3490-3496.
- McCarthy, N. J. and Evan, G. I. (1997). 15 methods for detecting and quantifying apoptosis. *Current Topics in Developmental Biology*, 36, 259-278.
- Mendel, D. B., Laird, A. D., Smolich, B. D., Blake, R. A., Liang, C., Hannah, A. L., Shaheen, R. M., Ellis, L. M., Weitman, S. and Shawver, L. K. (2000). Development of SU5416, a selective small molecule inhibitor of VEGF receptor tyrosine kinase activity, as an anti-angiogenesis agent. *Anti-cancer Drug Design*, 15, 29-41.
- Meyerowitz, B. E. (1980). Psychosocial correlates of breast cancer and its treatments. *Psychological Bulletin*, 87(1), 108-131.
- Milliken, W. and Albert, B. (1996). The use of medicinal plants by the Yanomami Indians of Brazil. *Economic Botany*, 50(1), 10-25.
- Minn, A., Rudin, C. M., Boise, L. H. and Thompson, C. B. (1995). Expression of bcl-xL can confer a multidrug resistance phenotype. *Blood*, 86(5), 1903-1910.
- Minn, A. J., Vélez, P., Schendel, S. L., Liang, H., Muchmore, S. W., Fesik, S. W., Fill, M. and Thompson, C. B. (1997). Bcl-xL forms an ion channel in synthetic lipid membranes. *Nature*, 385(6614), 353-357.
- Miquel, C., Borrini, F., Grandjouan, S., Aupérin, A., Viguer, J., Velasco, V., Duvillard, P., Praz, F. and Sabourin, J. C. (2005). Role of Bax mutations in apoptosis in colorectal cancers with microsatellite instability. *American Journal of Clinical Pathology*, 123, 562-570.
- Moerman, N., van Dam, F. S., Muller, M. J. and Oosting, H. (1996). The Amsterdam preoperative anxiety and information scale (APAIS). *Anesthesia and Analgesia*, 82(3), 445-451.
- Mohan, S., Abdul, A. B., Abdelwahab, S. I., Al-Zubairi, A. S., Sukari, M. A., Abdullah, R., Taha, M. M. E., Ibrahim, M. Y. and Syam, S. (2010). Typhonium

- flagelliforme induces apoptosis in CEMss cells via activation of caspase-9, PARP cleavage and cytochrome C release: Its activation coupled with G0/G1 phase cell cycle arrest. *Journal of Ethnopharmacology*, 131, 592-600.
- Mohd, W. N. I. Z. W., Zain, A. R., Othman, F. and Yap, T. Y. H. (2009). Antiproliferative properties of clausine-B against cancer cell lines. *The Malaysian Journal of Medical Sciences*, 16(3), 29-34.
- Morhason-Bello, I. O., Odedina, F., Rebbeck, T. R., Harford, J., Dangou, J. M., Denny, L. and Adewole, I. F. (2013). Challenges and opportunities in cancer control in Africa: a perspective from the African organisation for research and training in cancer. *The Lancet Oncology*, 14(4), 142-151.
- Müller, M., Wilder, S., Bannasch, D., Israeli, D., Lehlbach, K., Li-Weber, M., Friedman, S. L., Galle, P. R., Stremmel, W. and Oren, M. (1998). P53 activates the CD95 (Apo-1/Fas) gene in response to DNA damage by anticancer drugs. *Journal of Experimental Medicine*, 188, 2033-2045.
- Nagata, S. (2000). Apoptotic DNA fragmentation. *Experimental Cell Research*, 256, 12-18.
- Nakamura, K., Takemura, Y., Ju-ichi, M., Ito, C. and Furukawa, H. (1998). Three new coumarins from *Clausena excavata*. *Heterocycles*, 3(48), 549-553.
- Ncube, N., Afolayan, A. and Okoh, A. (2008). Assessment techniques of antimicrobial properties of natural compounds of plant origin: current methods and future trends. *African Journal of Biotechnology*, 7(12), 1797-1806.
- Newman, D. J., Cragg, G. M. and Snader, K. M. (2003). Natural products as sources of new drugs over the period 1981-2002. *Journal of Natural Products*, 66(7), 1022-1037.
- Noor Rain, A., Khozirah, S., Mohd Ridzuan, M., Ong, B., Rohaya, C., Rosilawati, M., Hamdino, I., Badrul, A. and Zakiah, I. (2007). Antiplasmodial properties of some Malaysian medicinal plants. *Tropical Biomedicine*, 24, 29-35.
- Nordenstedt, H., White, D. L. and El-Serag, H. B. (2010). The changing pattern of epidemiology in hepatocellular carcinoma. *Digestive and Liver Disease*, 42, S206-S214.
- O'Brien, M. A. and Kirby, R. (2008). Apoptosis: A review of pro-apoptotic and anti-apoptotic pathways and dysregulation in disease. *Journal of Veterinary Emergency and Critical Care*, 18(6), 572-585.
- O'Byrne, K. J. and Steward, W. P. (2001). Tumour angiogenesis: a novel therapeutic target in patients with malignant disease. *Emerging Drugs*, 6(1), 155-174.
- Omar, Z. A., Ali, Z. M. and Tamin, N. S. I. (2006). Malaysian cancer statistics - data and figure, Peninsular Malaysia 2006. *National cancer registry, ministry of health Malaysia*, 11-12. <https://www.scribd.com/doc/45028125/Malaysia-Cancer-Statistics> (Accessed on 31 October, 2016).

- Oren, M. and Rotter, V. (1999). Introduction: p53—the first twenty years. *Cellular and Molecular Life Sciences*, 55(1), 9-11.
- Owen-Schaub, L. B., Zhang, W., Cusack, J. C., Angelo, L. S., Santee, S. M., Fujiwara, T., Roth, J. A., Deisseroth, A. B., Zhang, W. W. and Kruzel, E. (1995). Wild-type human p53 and A Temperature-Sensitive Mutant Induce Fas/Apo-1 Expression. *Molecular and Cellular Biology*, 15, 3032-3040.
- Pan, M. H., Chen, W. J., Lin-Shiau, S. Y., Ho, C. T. and Lin, J. K. (2002). Tangeretin induces cell-cycle G1 arrest through inhibiting cyclin-dependent kinases 2 and 4 activities as well as elevating cdk inhibitors p21 and p27 in human colorectal carcinoma cells. *Carcinogenesis*, 23, 1677-1684.
- Park, J. Y., Park, J. M., Jang, J. S., Choi, J. E., Kim, K. M., Cha, S. I., Kim, C. H., Kang, Y. M., Lee, W. K. and Kam, S. (2006). Caspase-9 promoter polymorphisms and risk of primary lung cancer. *Human Molecular Genetics*, 15, 1963-1971.
- Parsell, D. and Lindquist, S. (1993). The function of Heat-Shock Proteins in stress tolerance: Degradation and reactivation of damaged proteins. *Annual Review of Genetics*, 27, 437-496.
- Parsell, D. A., Taulien, J. and Lindquist, S. (1993). The role of Heat-Shock Proteins in thermotolerance. *Molecular Chaperones*. Springer.
- Peltomäki, P., Aaltonen, L. A., Sistonen, P., Pylkkänen, L., Mecklin, J. P., Järvinen, H., Green, J. S., Jass, J. R., Weber, J. L. and Leach, F. S. (1993). Genetic mapping of a locus predisposing to human colorectal cancer. *Science*, 260, 810-810.
- Pfister, H. (1984). Biology and biochemistry of papillomaviruses. *Reviews of Physiology, Biochemistry and Pharmacology*, 99, 111-181.
- Potterat, O., Puder, C., Bolek, W., Wagner, K., Ke, C., Ye, Y. and Gillardon, F. (2005). Clausine Z, a new carbazole alkaloid from *Clausena excavata* with inhibitory activity on CDK5. *Die Pharmazie-An International Journal of Pharmaceutical Sciences*, 60(8), 637-639.
- Preston, T. J., Abadi, A., Wilson, L. and Singh, G. (2001). Mitochondrial contributions to cancer cell physiology: potential for drug development. *Advanced Drug Delivery Reviews*, 49(1), 45-61.
- Prior, B. M., Yang, H. and Terjung, R. L. (2004). What makes vessels grow with exercise training? *Journal of Applied Physiology*, 97(3), 1119-1128.
- Qi, F., Li, A., Inagaki, Y., Gao, J., Li, J., Kokudo, N., Li, X. K. and Tang, W. (2010). Chinese herbal medicines as adjuvant treatment during chemo- or radiotherapy for cancer. *Bioscience Trends*, 4, 297-307.

- Quirk, D. M., Rattner, D. W., Fernandez-del Castillo, C., Warshaw, A. L. and Brugge, W. R. (1997). The use of endoscopic ultrasonography to reduce the cost of treating ampullary tumors. *Gastrointestinal Endoscopy*, 46(4), 334-337.
- Qutub, A. A. and Popel, A. S. (2009). Elongation, proliferation and migration differentiate endothelial cell phenotypes and determine capillary sprouting. *BMC Systems Biology*, 3(1), 13-22.
- Raffo, A. J., Perlman, H., Chen, M. W., Day, M. L., Streitman, J. S. and Buttyan, R. (1995). Overexpression of bcl-2 protects prostate cancer cells from apoptosis *in vitro* and confers resistance to androgen depletion *in vivo*. *Cancer Research*, 55(19), 4438-4445.
- Raica, M., Cimpean, A. M. and Ribatti, D. (2009). Angiogenesis in pre-malignant conditions. *European Journal of Cancer*, 45(11), 1924-1934.
- Rao, C. V. (2005). Immunology: a textbook. Alpha Science International Limited, Pp326.
- Raven, P. H. (1988). Our diminishing tropical forests: in Biodiversity. National Academy Press, Pp119-122.
- Ridley, H. N. (1925). The Flora Of The Malay Peninsula. London, L. Reeve and Cooperative Limited, Pp498-511.
- Roncucci, L. and Mariani, F. (2015). Prevention of colorectal cancer: How many tools do we have in our basket? *European Journal of Internal Medicine*, 26(10), 752-756.
- Santisuk, T. (1988). An account of the vegetation of northern Thailand. *Journal of Geological Research* 5, Pp101.
- Saraste, A. and Pulkki, K. (2000). Morphologic and biochemical hallmarks of apoptosis. *Cardiovascular Research*, 45(3), 528-537.
- Schmitt, C. A., Fridman, J. S., Yang, M., Lee, S., Baranov, E., Hoffman, R. M. and Lowe, S. W. (2002). A senescence program controlled by p53 and p16 INK4a contributes to the outcome of cancer therapy. *Cell*, 109, 335-346.
- Schneider, P. and Tschopp, J. (2000). Apoptosis induced by death receptors. *Pharmaceutica Acta Helvetiae*, 74(2), 281-286.
- Schuster, M., Nechansky, A. and Kircheis, R. (2006). Cancer immunotherapy. *Biotechnology Journal*, 1(2), 138-147.
- Selby, J. V., Friedman, G. D., Quesenberry Jr, C. P. and Weiss, N. S. (1992). A case-control study of screening sigmoidoscopy and mortality from colorectal cancer. *New England Journal of Medicine*, 326(10), 653-657.

- Sharif, M. and Waznah, N. (2009). *Chemical constituents and bioactivities of Clausena excavata*. Masters Thesis, UPM, Pp19-24.
- Sharma, H., Parihar, L. and Parihar, P. (2011). Review on cancer and anticancerous properties of some medicinal plants. *Journal of Medicinal Plants Research*, 5(10), 1818-1835.
- Sherr, C. J. (1996). Cancer cell cycles. *Science*, 274(5293), 1672-1677.
- Shier, W. (1983). An undergraduate experiment to demonstrate the use of cyto-toxic drugs in cancer-chemotherapy. *American Association of College of Pharmacy*, 47, Pp216-220.
- Shoeb, M. (2006). Anticancer agents from medicinal plants. *Bangladesh Journal of Pharmacology*, 1(2), 35-41.
- Shu, Y. Z. (1998). Recent natural products based drug development: a pharmaceutical industry perspective. *Journal of Natural Products*, 61(8), 1053-1071.
- Simon, H. U., Haj-Yehia, A. and Levi-Schaffer, F. (2000). Role of reactive oxygen species (ROS) in apoptosis induction. *Apoptosis*, 5(5), 415-418.
- Skulachev, V. P. (1996). Why are mitochondria involved in apoptosis? Permeability transition pores and apoptosis as selective mechanisms to eliminate superoxide-producing mitochondria and cell. *Federation of European Biochemical Societies letters*, 397(1), 7-10.
- Slatter, T. L., Hung, N., Campbell, H., Rubio, C., Mehta, R., Renshaw, P., Williams, G., Wilson, M., Engelmann, A. and Jeffs, A. (2011). Hyperproliferation, cancer and inflammation in mice expressing a Δ133p53-like isoform. *Blood*, 117, 5166-5177.
- Small, S., Keerthivasan, G., Huang, Z., Gurbuxani, S. and Crispino, J. D. (2010). Overexpression of survivin initiates hematologic malignancies *in vivo*. *Leukemia*, 24, 1920-1926.
- Society, A. C. (2013). Cancer facts and figures 2013: American Cancer Society Atlanta, Pp1-25.
- Sripisut, T. and Laphookhieo, S. (2010). Carbazole alkaloids from the stems of *Clausena excavata*. *Journal of Asian Natural Products Research*, 12(7), 614-617.
- Steller, H. (1995). Mechanisms and genes of cellular suicide. *Science*, 267(5203), 1445-1449.
- Stroh, C. and Schulze-Osthoff, K. (1998). Death by a thousand cuts: An ever increasing list of caspase substrates. *Cell Death and Differentiation*, 5, 997-1000.

- Su, C. R., Yeh, S. F., Liu, C. M., Damu, A. G., Kuo, T. H., Chiang, P. C., Bastow, K. F., Lee, K. H. and Wu, T. S. (2009). Anti-HBV and cytotoxic activities of pyranocoumarin derivatives. *Bioorganic and Medicinal Chemistry*, 17, 6137-6143.
- Sunthitikawinsakul, A., Kongkathip, N., Kongkathip, B., Phonnakhu, S., Daly, J. W., Spande, T. F., Nimit, Y. and Rochanaruangrai, S. (2003). Coumarins and carbazoles from *Clausena excavata* exhibited antimycobacterial and antifungal activities. *Planta Medica*, 69, 155-157.
- Syama, S., Bustamama, A., Abdullaib, R., Sukaric, M. A., Hashimd, N. M., Ghaderiand, M., Rahmanic, M., Mohane, S., Abdelwahabe, S. I. and Alif, H. M. (2013). β -Mangostin induces p53-dependent G2/M cell cycle arrest and apoptosis through ROS mediated mitochondrial pathway and NfkB suppression in mcf-7 cells. *Journal of Functional Foods*, 6, 290-304.
- Szegezdi, E., Fitzgerald, U. and Samali, A. (2003). Caspase-12 and ER-stress-mediated apoptosis. *Annals of the New York Academy of Sciences*, 1010(1), 186-194.
- Takahashi, Y., Kitadai, Y., Bucana, C. D., Cleary, K. R. and Ellis, L. M. (1995). Expression of vascular endothelial growth factor and its receptor, KDR, correlates with vascularity, metastasis and proliferation of human colon cancer. *Cancer Research*, 55(18), 3964-3968.
- Takemura, Y., Nakamura, K., Hirusawa, T., Ju-Ichi, M., Ito, C. and Furukawa, H. (2000). Four new furanone-coumarins from *Clausena excavata*. *Chemical and Pharmaceutical Bulletin*, 48(4), 582-584.
- Tan, N. H. and Zhou, J. (2006). Plant cyclopeptides. *Chemical Reviews*, 106(3), 840-895.
- Tan, M., Muhammad, T. T., Najimudin, N. and Sulaiman, S. (2005). Growth arrest and non-apoptotic programmed cell death associated with the up-regulation of C-MYC mRNA expression in T-47D breast tumor cells following exposure to *Epipremnum pinnatum* (L.) Engl. hexane extract. *Journal of Ethnopharmacology*, 96, 375-383.
- Taufiq-Yap, Y., Peh, T., Ee, G., Rahmani, M., Sukari, M., Ali, A. and Muse, R. (2007). A new cytotoxic carbazole alkaloid from *Clausena excavata*. *Natural Product Research*, 21(9), 810-813.
- Taylor, J., Rabe, T., McGaw, L., Jäger, A. and Van Staden, J. (2001). Towards the scientific validation of traditional medicinal plants. *Plant Growth Regulation*, 34(1), 23-37.
- Theodoropoulos, G. E., Gazouli, M., Vaiopoulou, A., Leandrou, M., Nikouli, S., Vassou, E., Kouraklis, G. and Nikiteas, N. (2011). Polymorphisms of caspase-8 and caspase-9 gene and colorectal cancer susceptibility and prognosis. *International Journal of Colorectal Disease*, 26, 1113-1118.

- Thompson, C. B. (1995). Apoptosis in the pathogenesis and treatment of disease. *Science*, 267(5203), 1456-1462.
- Thorgeirsson, S. S. and Grisham, J. W. (2002). Molecular pathogenesis of human hepatocellular carcinoma. *Nature Genetics*, 31(4), 339-346.
- Trevor, A. J., Katzung, B. G., Masters, S. B. and Kruidering-Hall, M. (2010). *Pharmacology Examination and Board Review*: McGraw-Hill Medical New York, Pp716-722.
- United Nations, D. O. E. and Social Affairs, P. D. (2012). Changing levels and trends in mortality: the role of patterns of death by cause. United Nations publication, ST/ESA/SER.A/318,110.<http://www.un.org/en/development/desa/population/publications/mortality/changingLevelsandtrends.shtml> (Accessed on 31 October, 2016).
- Van Schil, P. E., Balduyck, B., De Waele, M., Hendriks, J. M., Hertoghs, M. and Lauwers, P. (2013). Surgical treatment of early-stage non-small-cell lung cancer. *European Journal of Cancer Supplements*, 11(2), 110-122.
- Vermes, I., Haanen, C., Steffens-Nakken, H. and Reutellingsperger, C. (1995). A novel assay for apoptosis flow cytometric detection of phosphatidylserine expression on early apoptotic cells using fluorescein labelled annexin V. *Journal of Immunological Methods*, 184, 39-51.
- Vikhanskaya, F., Lee, M. K., Mazzoletti, M., Broggini, M. and Sabapathy, K. (2007). Cancer-derived p53 mutants suppress p53-target gene expression - potential mechanism for gain of function of mutant p53. *Nucleic Acids Research*, 35(6), 2093-2104.
- Vucic, D. and Fairbrother, W. J. (2007). The inhibitor of apoptosis proteins as therapeutic targets in cancer. *Clinical Cancer Research*, 13(20), 5995-6000.
- Vucic, D., Stennicke, H. R., Pisabarro, M. T., Salvesen, G. S. and Dixit, V. M. (2000). ML-IAP, a novel inhibitor of apoptosis that is preferentially expressed in human melanomas. *Current Biology*, 10(21), 1359-1366.
- Waljee, J. F. and Newman, L. A. (2007). Neoadjuvant systemic therapy and the surgical management of breast cancer. *Surgical Clinics of North America*, 87(2), 399-415.
- Wallace-Brodeur, R. and Lowe, S. (1999). Clinical implications of p53 mutations. *Cellular and Molecular Life Sciences*, 55, 64-75.
- Wang, S., Zheng, Z., Weng, Y., Yu, Y., Zhang, D., Fan, W., Dai, R. and Hu, Z. (2004). Angiogenesis and anti-angiogenesis activity of Chinese medicinal herbal extracts. *Life Sciences*, 74, 2467-2478.

- Waziri, P. M., Abdullah, R., Yeap, S. K., Omar, A. R., Kassim, N. K., Malami, I., How, C. W., Etti, I. C. and Abu, M. L. (2016a). Clausenidin induces caspase-dependent apoptosis in colon cancer. *BMC Complementary and Alternative Medicine*, 16, 256.
- Waziri, P. M., Abdullah, R., Yeap, S. K., Omar, A. R., Abdul, A. B., Kassim, N. K., Malami, I., Karunakaran, T. and Imam, M. U. (2016b). Clausenidin from *Clausena excavata* induces apoptosis in HepG2 cells via the mitochondrial pathway. *Journal of Ethnopharmacology*, 194, 549-558.
- Wei, Y., Fan, T. and Yu, M. (2008). Inhibitor of apoptosis proteins and apoptosis. *Acta Biochimica et Biophysica Sinica*, 40(4), 278-288.
- Wiart, C., Mogana, S., Khalifah, S., Mahan, M., Ismail, S., Buckle, M., Narayana, A. and Sulaiman, M. (2004). Antimicrobial screening of plants used for traditional medicine in the state of Perak, Peninsular Malaysia. *Fitoterapia*, 75, 68-73.
- Williams, G. T. and Smith, C. A. (1993). Molecular regulation of apoptosis: genetic controls on cell death. *Cell*, 74(5), 777-779.
- Wong, R. S. (2011). Apoptosis in cancer: from pathogenesis to treatment. *Journal of Experimental and Clinical Cancer Research*, 30(1), 87.
- Wu, T. S. and Furukawa, H. (1982). Biological and phytochemical investigation of *Clausena excavata*. *Journal of Natural Products*, 45(6), 718-720.
- Wu, C. C., Ko, F. N., Wu, T. S. and Teng, C. M. (1994). Antiplatelet effects of clausine-D isolated from *Clausena excavata*. *Biochimica et Biophysica Acta (Bba)-General Subjects*, 1201, 1-6.
- www.everythingmaths.co.za/science/lifesciences/grade-10/03-cell-division/03-cell-division-02.cnxmlplus
- Xin, Z. Q., Lu, J. J., Ke, C. Q., Hu, C. X., Lin, L. P. and Ye, Y. (2008). Constituents from *Clausena excavata*. *Chemical and Pharmaceutical Bulletin*, 56(6), 827-830.
- Xu, L., Li, H., Xu, Z., Wang, Z., Liu, L., Tian, J., Sun, J., Zhou, L., Yao, Y. and Jiao, L. (2012). Multi-center randomized double-blind controlled clinical study of chemotherapy combined with or without traditional Chinese medicine on quality of life of postoperative non-small cell lung cancer patients. *BMC Complementary and Alternative Medicine*, 12(1), 112.
- Yamamoto, S., Sobue, T., Kobayashi, M., Sasaki, S. and Tsugane, S. (2003). Soy, isoflavones and breast cancer risk in Japan. *Journal of the National Cancer Institute*, 95(12), 906-913.

- Yamamoto, Y. and Gaynor, R. B. (2001). Therapeutic potential of inhibition of the NF-κB pathway in the treatment of inflammation and cancer. *The Journal of Clinical Investigation*, 107(2), 135-142.
- Yan, X., Zhou, T., Tao, Y., Wang, Q., Liu, P. and Liu, C. (2010). Salvianolic acid B attenuates hepatocyte apoptosis by regulating mediators in death receptor and mitochondrial pathways. *Experimental Biology and Medicine*, 235(5), 623-632.
- Yang, J. C., Sherry, R. M., Steinberg, S. M., Topalian, S. L., Schwartzenruber, D. J., Hwu, P., Seipp, C. A., Rogers-Freezer, L., Morton, K. E. and White, D. E. (2003). Randomized study of high-dose and low-dose interleukin-2 in patients with metastatic renal cancer. *Journal of Clinical Oncology*, 21, 3127-3132.
- Yano, S., Shinohara, H., Herbst, R. S., Kuniyasu, H., Bucana, C. D., Ellis, L. M., Davis, D. W., McConkey, D. J. and Fidler, I. J. (2000). Expression of vascular endothelial growth factor is necessary but not sufficient for production and growth of brain metastasis. *Cancer Research*, 60, 4959-4967.
- Yoshida, T. (1996). Graft compatibility of citrus with plants in the *Aurantioideae* and their susceptibility to *Citrus tristeza* virus. *Plant Disease*, 80(4), 414-417.
- Zain, W. N., Rahmat, A., Othman, F. and Yap, T. Y. H. (2009). Antiproliferative properties of clausine-B against cancer cell lines. *Malaysian Journal of Medical Sciences*, 16(3), 31-36.
- Zamzami, N. and Kroemer, G. (2003). Apoptosis: mitochondrial membrane permeabilization—the whole story? *Current Biology*, 13(2), R71-R73.
- Zhang, A. and Lin, G. (2000). The first synthesis of clausenamine-A and cytotoxic activities of three biscarbazole analogues against cancer cells. *Bioorganic and Medicinal Chemistry Letters*, 10(10), 1021-1023.
- Zhang, J., Wu, Y., Zhao, X., Luo, F., Li, X., Zhu, H., Sun, C. and Chen, K. (2014). Chemopreventive effect of flavonoids from ougan (*Citrus reticulata* Cv. *Suavissima*) fruit against cancer cell proliferation and migration. *Journal of Functional Foods*, 10, 511-519.
- Zhi, N. (2006). Chemical constituents of essential oil from leaves of three species of *Clausena*. *J. Biomass Chemical Engineering*, 13, 244-254.
- Zhong, C., Guo, R., Shi, M., Wei, W., Yu, W. and Li, J. (2006). Expression and clinical significance of VEGF and MMP-9 in hepatocellular carcinoma. *Ai zheng=Aizheng= Chinese Journal of Cancer*, 25(5), 599-603.
- Ziegler, U. and Groscurth, P. (2004). Morphological features of cell death. *Physiology*, 19(3), 124-128.
- Zilli, M., Grassadonia, A., Tinari, N., Di Giacobbe, A., Gildetti, S., Giampietro, J., Natoli, C. and Iacobelli, S. (2009). Molecular mechanisms of endocrine

resistance and their implication in the therapy of breast cancer. *Biochimica et Biophysica Acta (BBA)-Reviews on Cancer*, 1795(1), 62-81.

