



UNIVERSITI PUTRA MALAYSIA

***CLINICOPATHOLOGICAL CHANGES OF THE NERVOUS SYSTEM
FOLLOWING INOCULATION OF PASTEURELLA MULTOCIDA B:2 AND
ITS IMMUNOGENS IN BUFFALO CALVES***

ALI DHIAA MARZA

FPV 2017 2



**CLINICOPATHOLOGICAL CHANGES OF THE NERVOUS SYSTEM
FOLLOWING INOCULATION OF *PASTEURELLA MULTOCIDA* B:2 AND
ITS IMMUNOGENS IN BUFFALO CALVES**

By

ALI DHIAA MARZA

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

January 2017



© COPYRIGHT UPM

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

To my family who made my professional progress possible

To my instructors, teachers, mentors who made it a reality

To the fellow researchers who may be able to use the results of this research

I dedicate my work to all with love and gratitude.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Doctor of Philosophy

**CLINICOPATHOLOGICAL CHANGES OF THE NERVOUS SYSTEM
FOLLOWING INOCULATION OF *PASTEURELLA MULTOCIDA* B:2 AND
ITS IMMUNOGENS IN BUFFALO CALVES**

By

ALI DHIAA MARZA

January 2017

Chairman : Associate Professor Faez Firdaus Jesse Abdullah, PhD
Faculty : Veterinary Medicine

Haemorrhagic septicaemia (HS) is an acute, highly fatal septicaemic disease affecting cattle and buffaloes with high morbidity and mortality rates. *Pasteurella multocida* E:2 (African serotype) and B:2 (Asian serotype) are responsible of the disease in Africa and Asia, respectively. Despite continuing researches on the pathogenesis of *P. multocida*, the mechanisms by which these bacteria and its lipopolysaccharide (LPS) and outer membrane proteins (OMPs) immunogens develop the diseases are poorly understood. It is well known that HS affects mainly the respiratory and digestive tracts of cattle and buffaloes. However, involvement of other sites such as the nervous system in the pathogenesis of HS has been reported in previous studies, including outbreaks and pathogenesis studies of HS, without details. Therefore, this study was conducted to investigate in details the involvement of the nervous system towards *P. multocida* B:2 and its lipopolysaccharides (LPS) and outer membrane proteins (OMPs) immunogens in buffalo calves.

Twenty one clinically healthy, non-pregnant and non-lactating swamp buffalo calves were used in this study and divided into 7 groups of 3 buffaloes in each one. Buffaloes calves of group 1 (Ctrl) was inoculated orally with 10 ml of sterile phosphate buffered (PBS) as a negative control. Group 2 (Pmor) and group 3 (Pmsc) were inoculated with 10 ml of 1×10^{12} cfu/ml of *P. multocida* B:2 whole bacteria orally and subcutaneously, respectively. Group 4 (LPSor) and group 5 (LPSiv) were inoculated with 10 ml of LPS broth extracted from 1×10^{12} cfu/ml of *P. multocida* B:2 orally and intravenously, respectively. Group 6 (OMPor) and group 7 (OMPsc) were inoculated with 10 ml of OMPs broth extracted from 1×10^{12} cfu/ml of *P. multocida* B:2 orally and subcutaneously, respectively. All calves in Pmor, LPSor, LPSiv and OMPor and Ctrl groups survived during the experiment and euthanized at the end of the experiment at 504 h (21 days) post inoculation, while calves in Pmsc and OMPsc groups had to be euthanized after 12 h and 72 h post infection,

respectively as they develop severe clinical signs of HS according to the guideline of the Institutional Animal Care and Use Committee (IACUC). In order to monitor both general clinical and neurological responses during the experiment, detailed clinical examination were performed based on modified scoring system including; total general clinical score (TGCS) and total nervous clinical score (TNCS). The TGCS includes scores of mean rectal temperature (MRT), mean respiratory rate (MRR) and mean heart rate (MHR), in addition to scores of demeanour, appetite, nasal discharge and nature of respiration. While TNCS includes scores of mentation, gait abnormalities, fixation reflex, menace response, pupillary light reflex, perineal reflex, photomotor reflex, tongue tone test, gag reflex, swallowing reflex, panniculus reflex, pedal reflex, patellar reflex and palpebral reflex. In addition, pathophysiological changes in the cerebrospinal fluid (CSF) were also evaluated which include selected proinflammatory cytokines and acute phase proteins. After euthanization of the calves, complete post mortem, microbiological and histopathological examinations were conducted on the calves in the studies groups.

Clinically, significant differences ($P<0.05$) in both TGCS and TNCS were found among the studies groups compare with Ctrl group. However, Pmsc and OMPsc groups developed severe clinical signs and recorded sharp increase in both TGCS and TNCS that lead to euthanization of the calves in these groups at 12 and 72 h post inoculation, respectively, and this indicates direct associated of increased nervous clinical signs with disease severity. While less severe TGCS and TNCS were recorded in Pmor, LPSor, LPSiv and OMP groups and calves in these groups were able to survive until the end of experiment at 504 h post inoculation, and this indicates that calves in these groups were able to overcome the infection with *P. multocida* B:2 or inoculation of its LPS and OMPs immunogens.

At necropsy, different levels of significance ($P<0.05$) in mean gross lesion score (MGLS) were recorded in buffalo calves in the studied groups including the cerebrum, cerebellum, brainstem and spinal cord when compared with calves of Ctrl group. However, the cerebrum and brainstem being the most affected sites of the nervous system. On the other hand, examination of the cranial and spinal nerves did not reveal any significant changes. The pattern of the pathological lesions reflect the severity of the clinical signs where Pmsc and OMPsc groups represented the per acute stage of the disease and congestion and hemorrhage in the brain and spinal cord were the most prominent lesions. However, oedema was noted in the brain of the buffalo calves in OMPsc rather than Pmsc group. While the pathological changes were different in LPSiv group in which oedema was more prominent than congestion and hemorrhage. Less MGLS were observed in the Pmor, LPSor and OMPor groups indicating the less effect of the oral route on the nervous system. Additionally, positive bacterial re-isolation and polymerase chain reaction (PCR) confirmation of *P. multocida* B:2 from different parts of the nervous system in calves of Pmsc group confirm the ability of these bacteria to cross blood brain barrier (BBB) especially at the per acute stage of the disease and support the involvement of the nervous system in the pathogenesis of HS.

The histopathological examination of the nervous system of the inoculated groups reflect the previously observed gross pathological findings, and significant increase ($P<0.05$) has been found in the total mean histopathological lesion score (MHLS) of the treated groups: Pmor, Pmsc, LPSor, LPSiv, OMPor and OMPsc when compared with calves of Ctrl group. The histopathological changes recorded in this study included; inflammatory cells infiltration, oedema and spongiosis, gliosis, congestion and hemorrhage, in addition to necrosis and degeneration and confirm the ability of *P. multocida* B:2 and its LPS and OMPs immunogens to cross the BBB and induce their pathological effects. While the most affected sites of the nervous system were the cerebrum and brainstem. Although various histopathological lesion were reported in different parts of the nervous tissues, the route of inoculation was observed to play a significant role in determination of the severity of the lesion as subcutaneous and intravenous routes in Pmsc, OMPsc and LPSiv groups showed the most pronounced and higher lesion distribution than respected orally inoculated groups.

Pathophysiological changes including; classical parameters, proinflammatory cytokines and acute phase proteins (APPs) also showed various levels of difference in the studied groups. Classical parameters showed significant differences ($P<0.05$) with high values recorded in the CSF total leukocytic count (CSF-TLC) and CSF total protein (CSF-TP) especially Pmsc and OMPsc groups as indication of acute body response to the disease produced. Significant increase also observed in the levels of CSF cytokines including; tumor necrosis factor- α (CSF-TNF- α), interleukins (CSF-IL-1) and (CSF-IL-6) in addition to CSF APPs including; haptoglobin (CSF-Hp), serum amyloid A (CSF-SAA) and C-reactive protein (CSF-CRP) concentrations which indicate that the subcutaneous route of inoculation would induce more severe reactions rather than oral route.

According to presented evidences, it can be concluded that *P. multocida* B:2 and its LPS and OMPs immunogens were able to cross the BBB and induce pathological changes in different sites of the nervous system. The produced neuropathological changes could make alteration in animal behavior with the evidence of absence of some of the neurological responses especially in Pmsc and OMPsc groups, and this was supported by the increased concentrations of proinflammatory cytokines and APPs in the CSF in these groups. Accordingly, these results provide for the first time strong evidence of the involvement of the nervous system of buffalo calves experimentally inoculated of *P. multocida* and its immunogens in the pathogenesis of HS, and shows that the type of inoculum and route of inoculation strongly affect the induced pathological changes of the nervous system and also the severity of disease produced.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

**PERUBAHAN KLINIKOPATHOLOGI SISTEM SARAF BERIKUTAN
INOKULASI DENGAN *PASTEURELLA MULTOCIDA* B:2 DAN IMUNOGEN
DALAM ANAK KERBAU**

Oleh

ALI DHIAA MARZA

Januari 2017

Pengerusi : Profesor Madya Faez Firdaus Jesse Abdullah, PhD
Fakulti : Perubatan Veterinar

Hawar berdarah (HS) adalah akut, penyakit septisemia yang membawa maut yang melibatkan lembu dan kerbau dengan morbiditi dan mortaliti kadar yang tinggi. *Pasteurella multocida* E: 2 (serotype Afrika) dan B: 2 (serotype Asia) adalah bertanggungjawab ke atas penyakit masing-masing di Afrika dan Asia. Walaupun kajian berterusan ke atas patogenesis *P. multocida*, mekanisme bakteria ini dan lipopolisakarida (LPS) dan protein membran luar (OMPs) immunogens yang menyebabkan penyakit ini kurang difahami. Adalah diketahui umum bahawa HS memberi kesan terutamanya kepada salur pernafasan dan pencernaan lembu dan kerbau. Walau bagaimanapun, penglibatan bahagian lain seperti sistem saraf dalam patogenesis HS telah dilaporkan dalam kajian sebelum ini, termasuk wabak dan kajian patogenesis HS tetapi tanpa butiran yang terperinci. Oleh itu, kajian ini telah dijalankan untuk mengkaji secara terperinci penglibatan sistem saraf terhadap *P. multocida* B: 2 dan lipopolisakarida (LPS) dan protein membran luar (OMPs) immunegen bagi anak kerbau.

Dua puluh satu ekor kerbau yang sihat secara klinikal, tidak hamil dan bukan menyusu telah digunakan dalam kajian ini dan dibahagikan kepada 7 kumpulan dimana 3 anak krbau dalam setiap satu kumpulan. Anak kerbau dari kumpulan 1 (Ctrl) telah disuntik secara oral dengan 10 ml bufer fosfat steril (PBS) sebagai kawalan negatif. Kumpulan 2 (Pmor) dan kumpulan 3 (Pmsc) telah disuntik keseluruhan secara oral masing-masing dengan 10 ml larutan 1×10^{12} cfu/ml bakteria *P. multocida* B:2. Kumpulan 3 (LPSor) dan kumpulan 4 (LPSiv) telah disuntik keseluruhan secara oral masing-masing dengan larutan ekstrak daripada 1×10^{12} cfu/ml *P. multocida* B: 2. Kumpulan 6 (OMPor) dan kumpulan 7 (OMPsc) telah disuntik keseluruhan secara oral masing-masing dengan larutan ekstrak daripada 1×10^{12} cfu/ml *P. multocida* B: 2. Semua anak kerbau dalam kumpulan Pmor, LPSor, LPSiv dan OMPor dan kawalan terselamat semasa eksperimen dan

dikorbankan pada akhir eksperimen pada 504 jam (21 hari) selepas inokulasi, manakala anak-anak kerbau dalam kumpulan Pmsc dan OMPsc terpaksa dimatikan masing-masing selepas 12 jam dan 72 jam selepas inokulasi, kerana mereka menunjukkan tanda-tanda klinikal HS yang teruk dan mengikut garis panduan Jawatankuasa Penjagaan dan Penggunaan Haiwan Makmal (IACUC). Dalam usaha untuk memantau kedua-dua tindak balas klinikal dan neurologi umum semasa eksperimen, pemeriksaan klinikal secara terperinci telah dijalankan berdasarkan sistem penskoran yang diubahsuai termasuk; jumlah skor umum klinikal (TGCS) dan jumlah skor klinikal saraf (TNCS). TGCS termasuk purata skor suhu rektum (MRT), purata kadar pernafasan (MRR) dan purata kadar jantung (KSM), sebagai tambahan kepada skor sikap, selera makan, pengeluaran dari hidung dan jenis pernafasan. TNCS termasuk puluhan status mentaliti, keabnormalan gaya berjalan, penetapan refleks, tindak balas ancaman, tindakbalas cahaya, tindakbalas palpebral, tindakbalas photomotor, ujian nada lidah, tindakbalas gag, tindakbalas menelan, tindakbalas refleks, tindakbalas panniculus, tindakbalas pedal, tindakbalas patellar dan tindakbalas perineal. Di samping itu, perubahan patofisiologikal dalam cecair serebrospina (CSF) juga telah dinilai termasuk sitokine proinflamator yang dipilih dan protein fasa akut. Selepas anak kerbau dikorbankan, bedah siasat lengkap, mikrobiologi dan peperiksaan histopatologi dijalankan ke atas anak kerbau dalam kumpulan kajian ini.

Secara klinikal, perbezaan ketara ($P < 0.05$) dalam TGCS dan TNCS telah dijumpai dalam kumpulan kajian berbanding kumpulan kawalan. Walaubagaimanapun, kumpulan Pmsc and OMPsc telah menunjukkan tanda-tanda klinikal yang teruk dan direkodkan meningkat dalam TGCS dan TNCS dimana menyebabkan banyak anak dalam kumpulan ini masing-masing dikorbankan pada 12 dan 72 jam selepas inokulasi dan ia menunjukkan berkait rapat dengan peningkatan tanda saraf klinikal dengan penyakit berjangkit. Manakala pengurangan dalam kumpulan TGCS dan TNCS telah direkodkan dalam kumpulan Pmor, LPSor, LPSiv dan OMP dan anak kerbau dalam kumpulan ini mampu untuk bertahan sehingga akhir eksperimen iaitu pada 504 jam selepas inokulasi dan ini menunjukkan bahawa anak kerbau dalam kumpulan ini mampu untuk mengatasi kesan dengan *P. multocida* B:2 atau inokulasi LPS dan OMPs immunogens.

Pada ujian nekropsi, kadar perbezaan ketara ($P < 0.05$) dalam purata skor lesi kasar (MGLS) telah direkodkan dalam kajian kumpulan anak kerbau termasuk serebrum, serebellum, saraf otak dan serat tunjang apabila dibandingkan dengan kumpulan kawalan. Walaubagaimanapun, serebrum dan saraf otak adalah paling banyak terkesan dalam sistem saraf. Disamping itu, pemeriksaan pada saraf kranial dan saraf spinal tidak menunjukkan sebarang perubahan. Paten kajian patalogikal menunjukkan, kadar jangkitan pada tahap penyakit yang merebak secara pesat dan gangguan dalam otak dan sarat tunjang adalah paling banyak dalam kajian. Walaubagaimanapun, oedema telah dinyatakan dalam otak anak kerbau adalah tinggi dalam OMPsc berbanding kumpulan Pmsc. Manakala perubahan secara patalogikal adalah berbeza dalam kumpulan LPSiv dimana oedema adalah lebih banyak berbanding kongest and pendarahan. MGLS telah didapati kurang dalam kumpulan Pmor, LPSor dan OMPor dimana menunjukkan kurang kesan secara oral pada sistem saraf. Tambahan, bakteria positif yang diasingkan dan sistem penjujukan DNA

(PCR) telah mengesahkan bahawa *P. multocida* B:2 daripada bahagian lain pada sistem saraf dalam anak kerbau bagi kumpulan Pmsc menunjukkan kemampuan bakteria ini untuk mengatasi banteng darah otak terutamanya pada peringkat penyakit berjangkit dan menyokong penglibatan sistem saraf dalam patogenesis HS.

Pemeriksaan histopatologikal pada sistem saraf bagi kumpulan inokulasi telah menunjukkan perkaitan pada penemuan pathologi kasar, dan secara perbezaan ketara telah meningkat ($P < 0.05$) telah ditemui dalam purata skor histopatologikal (MHLS) bagi kumpulan rawatan: Pmor, Pmsc, LPSor, LPSiv, OMPor dan OMPsc berbanding kumpulan kawalan anak kerbau. Perubahan histopatologikal telah direkodkan dalam kajian ini termasuklah kehadiran sel radang, oedema dan spongiosis, gliosis, kongesi dan perdarahan, tambahan nekrosis dan penglibatan telah mengesahkan kemampuan *P. multocida* B:2 serta LPS dan OMPs immunogens untuk melawan BBB dan menyebabkan kesan pathologikal. Manakala yang paling terkesan ialah pada serebrum dan sistem saraf. Walaubagaimanapun, kepelbagaian histopathologikal telah dilaporkan dalam pelbagai laporan pada sistem saraf, kadar inokulasi telah memainkan peranan dalam penentuan tahap penyakit subkutaneous and intravenous bagi kumpulan Pmsc, OMPsc and LPSiv telah menunjukkan peningkatan dalam ketaburan lesi penyakit berbanding kumpulan inokulasi yang lain.

Perubahan pathofisiologikal termasuklah: parameter klasikal, sitokine dan tahap akut proteins (APPs) telah menunjukkan pelbagai tahap perbezaan dalam kumpulan kajian. Parameter klasikal menunjukkan perbezaan ketara ($P < 0.05$) dengan nilai tinggi direkodkan dalam CSF jumlah leukosit (CSF-TLC) dan CSF jumlah protein (CSF-TP) terutamanya kumpulan Pmsc dan OMPsc dimana menunjukkan tindakbalas badan kepada penyakit telah dihasilkan. Perbezaan ketara juga telah dijumpai pada CSF sitokine, termasuklah nekrosis otak, factor $-\alpha$ (CSF-TNF- α), interleukine (CSF-IL-1) dan (CSF-IL-6) tambahan kepada konsentrasi CSF APPs termasuklah ; haptoglobin (CSF-Hp), serum amyloid A (CSF-SAA) dan C-reactive protein (CSF-CRP) dimana menunjukkan inokulasi subkutis boleh menyebabkan banyak tindakbalas berbanding secara oral.

Berdasarkan kepada penemuan, ia boleh disimpulkan bahawa *P. multocida* B:2 dan LPS serta OMPs immunogens mampu untuk melawan BBB and membawa kepada kesan perubahan pathologikal pada bahagian berbeza dalam sistem saraf. Perubahan neuropathologikal juga mampu untuk mengubah tindakbalas terutamanya dalam kumpulan Pmsc dan OMPsc, dan ia telah disokong oleh peningkatan konsentrasi proinflamatori sitokien dan kumpulan APPs dalam CSF. Tambahan, keputusan yang dihasilkan telah menyokong secara kuat bahawa penglibatan sistem saraf dalam anak kerbau yang telah diinokulasi dengan *P. multocida* dan immunogens dalam pathogenesis HS telah menunjukkan bahawa inokulasi memberi kesan kepada perubahan pathologikal bagi sistem saraf dan menyebabkan penghasilan penyakit.

ACKNOWLEDGEMENTS

I would give the praise to Allah almighty, the most beneficent the most merciful who keep inspiring me, guiding me and always looking after me and directing me toward the utmost goodness.

I also would like to express my sincere gratitude and appreciations to my supervisor Associate Professor Dr.Faez Firdaus Jesse Abdullah for his professional insight, proficient guidance, encouragement, suggestion, time and patience throughout my study period. Thanks a lot for his kind and helpful supervision.

Many thanks and gratitude also goes to the supervisory committee for their guidance, advice and supervision starting with Professor Dr.Abdul Rahman Omar, Professor Dr.Zuki Abu Bakar and Professor Dr.Mohd Zamri Saad. Many thanks also go to Mr. Jefri Norsidin, Mr.Yap Keng Chee, Mr.Ganesanmurthi Perumal and Mr.Saiquzaman Ali for their help during the experiment work in laboratory. I am heartily grateful to my best friends Dr.Eric Lim Teik Chung and Dr. Hayder Hamzah Ibrahim for helpful and friendliness.

I also would like to express my utmost appreciation and gratitude to Universiti Putra Malaysia, School of Graduate Studies and Ministry of Higher Education for giving me the opportunity to pursue this study.

My thanks and appreciations also go to Taman Pertanian Universiti, UPM, Faculty of Veterinary Medicine, UPM, Department of Veterinary Clinical Studies and Department of Microbiology and Pathology, Faculty of Veterinary Medicine for providing me with all the facilities pertaining to my research. This study was funded by the Research University Grant Scheme (RUGS), Universiti Putra Malaysia.

I certify that a Thesis Examination Committee has met on 20 January 2017 to conduct the final examination of Ali Dhiaa Marza on his thesis entitled "Clinicopathological Changes of the Nervous System Following Inoculation of *Pasteurella multocida* B:2 and its Immunogens in Buffalo Calves" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy:

Members of the Thesis Examination Committee were as follows:

Noordin bin Mohamed Mustapha, PhD

Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Chairman)

Hazilawati binti Hamzah, PhD

Associate Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Internal Examiner)

Siti Khairani binti Bejo, PhD

Associate Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Internal Examiner)

Peter David Eckersall, PhD

Professor
University of Glasgow
United Kingdom
(External Examiner)



NOR AINI AB. SHUKOR, PhD
Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 22 March 2017

This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Faez Firdaus Jesse Abdullah, PhD

Associate Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Chairman)

Abdul Rahman Omar, PhD

Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

Zuki Abu Bakar, PhD

Professor
Faculty of Veterinary Medicine
Universiti Malaysia Kelantan
(Member)

Mohd Zamri-Saad, PhD

Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: _____ Date: _____

Name and Matric No: Ali Dhiaa Marza Alhashimi, GS40040

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: _____

Name of Chairman
of Supervisory
Committee:

Associate Professor Dr. Faez Firdaus Jesse Abdullah

Signature: _____

Name of Member
of Supervisory
Committee:

Professor Dr. Abdul Rahman Omar

Signature: _____

Name of Member
of Supervisory
Committee:

Professor Dr. Zuki Abu Bakar

Signature: _____

Name of Member
of Supervisory
Committee:

Professor Dr. Mohd Zamri-Saad

TABLE OF CONTENTS

		Page
ABSTRACT		i
ABSTRAK		iv
ACKNOWLEDGEMENTS		vii
APPROVAL		viii
DECLARATION		x
LIST OF TABLES		xvi
LIST OF FIGURES		xviii
LIST OF ABBREVIATIONS		xxvii
CHAPTER		
1	INTRODUCTION	1
	1.1 General introduction	1
	1.2 Problem statement	3
	1.3 Hypothesis of the study	3
	1.4 Objectives of the study	4
 2	LITERATURE REVIEW	 5
	2.1 <i>Pasteurella multocida</i>	5
	2.1.1 Cell morphology and growth characteristics	6
	2.1.2 Serological typing	7
	2.1.3 Virulence factors	7
	2.1.3.1 Capsule	7
	2.1.3.2 Outer membrane proteins	8
	2.1.3.3 Lipid polysaccharides	8
	2.1.3.4 Other factors	9
	2.2 Haemorrhagic septicaemia	9
	2.2.1 Source and routes of infection	10
	2.2.2 Pathogenesis	11
	2.2.3 Clinical signs	12
	2.2.4 Gross pathology	13
	2.2.5 Histopathology	14
	2.2.6 Animal models for study of haemorrhagic septicaemia	16
	2.3 Body responses due to <i>P. multocida</i> infection	16
	2.3.1 Routine biomarkers	16
	2.3.2 Proinflammatory cytokines	18
	2.3.3 Acute phase protein	19
	2.4 Diagnosis of haemorrhagic septicaemia	21
	2.4.1 Clinical diagnosis	21
	2.4.2 Laboratory diagnosis	22
	2.4.2.1 Bacteriological methods	22
	2.4.2.2 Serological tests	23
	2.4.2.3 Molecular biology methods	24
	2.5 Nervous system involvement caused by bacterial infection	24
	2.5.1 Normal structure and function of the nervous system	24
	2.5.2 Cerebrospinal fluid	27

2.5.3	Immunity of the nervous system	27
2.5.4	Bacterial infections of the nervous system	28
2.5.5	Clinical manifestations in the nervous system	28
2.5.5.1	Fever	29
2.5.5.2	Abnormal Limb movement	29
2.5.5.3	Abnormal vision	31
2.5.5.4	Abnormal mouth movement	32
2.5.5.5	Abnormal mentation	32
2.5.5.6	Depression	33
2.5.6	Pathologic findings in the nervous system	33
2.5.6.1	Response to injury	33
2.5.6.2	Oedema and vascular disturbances	34
2.5.7	Diagnosis of the nervous system	35
3	GENERAL MATERIALS AND METHODS	36
3.1	Animals	36
3.2	Bacterial strain	36
3.3	Preparation of inocula	36
3.3.1	<i>Pasteurella multocida</i> B:2 whole bacteria	36
3.3.2	<i>Pasteurella multocida</i> B:2 lipopolysaccharide	37
3.3.3	<i>Pasteurella multocida</i> B:2 outer membrane proteins	37
3.3.4	Sodium dodecyl sulphate-polyacrylamide gel electrophoresis	37
3.4	Experimental design	38
3.5	Clinical monitoring	40
3.6	Collection and examination of the CSF and blood	43
3.6.1	Collection of the CSF and blood	43
3.6.2	Classical examination of the CSF, blood and serum	44
3.6.3	Determination of acute phase proteins and cytokines levels in CSF and serum using ELISA	45
3.7	Necropsy	45
3.8	Bacterial isolation and polymerase chain reaction	46
3.8.1	Bacterial isolation	46
3.8.2	Polymerase chain reaction	46
3.8.2.1	DNA extraction	46
3.8.2.2	Primers	46
3.8.2.3	PCR procedure	47
3.8.2.4	Gel electrophoresis	47
3.9	Histopathological examination	47
3.10	Statistical analysis	48
4	CLINICAL SIGNS AND NEUROLOGICAL RESPONSES FOLLOWING INOCULATION OF <i>PASTEURELLA MULTOCIDA</i> B:2 AND ITS IMMUNOGENS IN BUFFALO CALVES	49
4.1	Introduction	49
4.2	Material and methods	50
4.3	Results	50
4.3.1	Rectal temperature	51
4.3.2	Respiratory rate	53

4.3.3	Heart rate	55
4.3.4	Total general clinical score	57
4.3.5	Total nervous clinical score	60
4.4	Discussion and conclusion	63
5	GROSS PATHOLOGICAL CHANGES AND BACTERIOLOGICAL EXAMINATION OF THE NERVOUS SYSTEM FOLLOWING INOCULATION OF <i>PASTEURELLA MULTOCIDA</i> B:2 AND ITS IMMUNOGENS IN BUFFALO CALVES	70
5.1	Introduction	70
5.2	Material and methods	71
5.3	Results	71
5.3.1	Gross pathology findings	71
5.3.1.1	Cerebrum	74
5.3.1.2	Cerebellum	78
5.3.1.3	Brainstem	82
5.3.1.4	Spinal cord	86
5.3.1.5	Cranial and spinal nerves	89
5.3.2	Bacterial isolation and polymerase chain reaction	90
5.4	Discussion and conclusion	92
6	HISTOPATHOLOGICAL CHANGES OF THE NERVOUS SYSTEM FOLLOWING INOCULATION OF <i>PASTEURELLA MULTOCIDA</i> B:2 AND ITS IMMUNOGENS IN BUFFALO CALVES	96
6.1	Introduction	96
6.2	Material and methods	97
6.3	Results	97
6.3.1	Cerebrum	100
6.3.2	Cerebellum	105
6.3.3	Brainstem	110
6.3.4	Spinal cord	115
6.3.5	Cranial nerves	120
6.3.6	Spinal nerves	125
6.4	Discussion and conclusion	130
7	CYTOLOGICAL ANALYSIS AND DETERMINATION OF PROINFLAMMATORY CYTOKINES AND ACUTE PHASE PROTEINS LEVELS IN THE CEREBROSPINAL FLUID AND BLOOD FOLLOWING INOCULATION OF <i>PASTEURELLA MULTOCIDA</i> B:2 AND ITS IMMUNOGENS IN BUFFALO CALVES	135
7.1	Introduction	135
7.2	Material and methods	136
7.3	Results	136
7.3.1	Classical parameters in CSF and blood	137
7.3.2	Proinflammatory cytokines	139
7.3.2.1	Tumor necrosis factor- α (TNF- α)	139
7.3.2.2	Interleukin-1(IL-1)	141

7.3.2.3	Interleukin-6 (IL-6)	142
7.3.3	Acute phase proteins	144
7.3.3.1	Haptoglobin (Hp)	144
7.3.3.2	Serum amyloid A (SAA)	145
7.3.3.3	C-reactive protein (CRP)	147
7.4	Discussion and conclusion	148
8	GENERAL DISCUSSION, CONCLUSIONS AND RECOMMENDATIONS	155
8.1	General discussion	155
8.2	Conclusions	165
8.3	Recommendations	165
	REFERENCES	166
	APPENDICES	198
	BIODATA OF STUDENT	214
	LIST OF PUBLICATIONS	215

LIST OF TABLES

Table		Page
2.1	Involvement of the nervous system due to <i>P. multocida</i> in different animals	15
3.1	The total general and total nervous clinical scores evaluated in the buffaloes are as shown in the table below	41
3.2	Relation between nervous reflex or test and their respective organ (s) with response	42
5.1	Mean gross lesion score in the groups of buffalo calves following inoculation of <i>P. multocida</i> B:2 and its LPS and OMPs extracts via different routes	72
5.2	Bacterial isolation and PCR of <i>P. multocida</i> B:2 from the nervous system of buffalo calves in Pmor, Pmsc and Ctrl groups	88
5.3	Biochemical tests results of the re-isolated bacteria from different site of the nervous system of buffalo calves in Pmsc group	89
6.1	Total mean histopathological lesion score in the groups of buffalo calves following inoculation of <i>P. multocida</i> B:2 and its LPS and OMPs extracts via different routes	96
6.2	Mean histopathological lesion score in the cerebrum of all groups of buffalo calves following inoculation of <i>P. multocida</i> B:2 and its LPS and OMPs extracts via different routes	99
6.3	Mean histopathological lesion score in the cerebellum of all groups of buffalo calves following inoculation of <i>P. multocida</i> B:2 and its LPS and OMPs extracts via different routes	104
6.4	Mean histopathological lesion score in the brainstem of all groups of buffalo calves following inoculation of <i>P. multocida</i> B:2 and its LPS and OMPs extracts via different routes	109
6.5	Mean histopathological lesion score in the spinal cord of all groups of buffalo calves following inoculation of <i>P. multocida</i> B:2 and its LPS and OMPs extracts via different routes	114
6.6	Mean histopathological lesion score in the cranial nerves of all groups of buffalo calves following inoculation of <i>P. multocida</i> B:2 and its LPS and OMPs extracts via different routes	119
6.7	Mean histopathological lesion score in the spinal nerves of all	124

groups of buffalo calves following inoculation of *P. multocida* B:2 and its LPS and OMPs extracts via different routes

7.1	Total leukocytes count, total protein and glucose concentrations in the CSF of the groups of buffalo calves following inoculation of <i>P. multocida</i> B:2 and its LPS and OMPs extracts via different routes	136
7.2	Total leukocytes count and serum total proteins (TP) and serum glucose (Glu) concentrations of the groups of buffalo calves following inoculation of <i>P. multocida</i> B:2 and its LPS and OMPs extracts via different routes	137
7.3	Tumor necrosis factor - α concentration in CSF and serum of the buffalo calves following inoculation of <i>P. multocida</i> B:2 and its LPS and OMPs extracts via different routes	138
7.4	Interleukin-1 concentration in CSF and serum of the buffalo calves following inoculation of <i>P. multocida</i> B:2 and its LPS and OMPs extracts via different routes	140
7.5	Interleukin-6 concentration in CSF and serum of the buffalo calves following inoculation of <i>P. multocida</i> B:2 and its LPS and OMPs extracts via different routes	141
7.6	Haptoglobin concentration in CSF and serum of the buffalo calves following inoculation of <i>P. multocida</i> B:2 and its LPS and OMPs extracts via different routes	143
7.7	Serum amyloid A concentration in CSF and serum of the buffalo calves following inoculation of <i>P. multocida</i> B:2 and its LPS and OMPs extracts via different routes	144
7.8	C-reactive protein concentration in CSF and serum of the buffalo calves following inoculation of <i>P. multocida</i> B:2 and its LPS and OMPs extracts via different routes	146

LIST OF FIGURES

Figure		Page
2.1	Bovine brain: (A) ventral section; (B) dorsal section	25
2.2	Blood brain barrier: (A) blood cerebrospinal fluid barrier; (B) interstitial fluid (ISF)	28
3.1	Flowchart of experimental design of inoculation of <i>P. multocida</i> B:2 and its immunogens, the LPS and OMPs, via different routes using buffalo calves model	39
3.2	Lumbosacral CSF tap: (A) anatomical reference points to locate the puncture site; (B) spinal needle	43
3.3	Cerebrospinal fluid was obtained from the lumbosacral region of the spine of a buffalo calf	44
4.1	Mean rectal temperature (MRT) expressed as (\pm SEM) over time in the groups of buffalo calves following inoculation of <i>P. multocida</i> B:2 and its LPS and OMPs extracts via different routes	52
4.2	Mean respiratory rate (MRR) expressed as (\pm SEM) over time in the groups of buffalo calves following inoculation of <i>P. multocida</i> B:2 and its LPS and OMPs extracts via different routes	54
4.3	Mean heart rate (MHR) expressed as (\pm SEM) over time in the groups of buffalo calves following inoculation of <i>P. multocida</i> B:2 and its LPS and OMPs extracts via different routes	56
4.4	Mean total general clinical score (TGCS) expressed as (\pm SEM) over time in the groups of buffalo calves following inoculation of <i>P. multocida</i> B:2 and its LPS and OMPs extracts via different routes	59
4.5	Mean total nervous clinical score (TNCS) expressed as (\pm SEM) over time in the groups of buffalo calves following inoculation of <i>P. multocida</i> B:2 and its LPS and OMPs extracts via different routes	62
5.1	Mean gross lesion score in the groups of buffalo calves following inoculation of <i>P. multocida</i> B:2 and its LPS and OMPs extracts via different routes	73

5.2	Cerebrum of a buffalo calf from <i>P. multocida</i> oral group showing mild congestion in leptomeninges of the cerebrum	74
5.3	Cerebrum of a buffalo calf from <i>P. multocida</i> subcutaneous group showing severe congestion of the cerebrum. Note the prominent cerebral vessels (bridging veins) that extend across to the superior aspect of the cerebral hemispheres, which are congested in the context of swelling	75
5.4	Cerebrum of a buffalo calf from lipopolysaccharide oral group showing mild congestion in leptomeninges of the cerebrum	75
5.5	Cerebrum of a buffalo calf from lipopolysaccharide intravenous group showing porridge-like structure of the brain eventually reduced the consistency of putty. The surface of the brain with cerebral oedema demonstrates widened gyri with a flattened surface. The sulci are narrowed in addition to mild congestion	76
5.6	Cerebrum of a buffalo calf from outer membrane proteins oral group showing oedema and mild congestion in leptomeninges of cerebrum	76
5.7	Cerebrum of a buffalo calf from outer membrane proteins subcutaneous group showing the prominent cerebral hemorrhage and swelling of the brain inside the skull vault bones has resulted in flattening of the gyri and narrowing of the sulci of the cerebral convexities	77
5.8	Cerebellum of a buffalo calf of <i>P. multocida</i> oral group showing mild congestion in leptomeninges of cerebellum	78
5.9	Cerebellum of a buffalo calf of <i>P. multocida</i> subcutaneous group showing moderate congestion of cerebellum. Also, swelling of the brain inside the skull vault bones has resulted in flattening of the gyri and narrowing of the sulci of the cerebral convexities, Note the prominent cerebral vessels, which are congested in the context of swelling	78
5.10	Cerebellum of a buffalo calf of lipopolysaccharide oral group showing mild congestion in leptomeninges of cerebellum	79
5.11	Cerebellum of a buffalo calf of lipopolysaccharide intravenous group showing mild to moderate hemorrhage of cerebellum	79
5.12	Cerebellum of a buffalo calf of outer membrane proteins oral group showing no clear lesion in cerebellum	80

5.13	Cerebellum of a buffalo calf of outer membrane proteins subcutaneous group showing moderate congestion of cerebellum with oedema. In addition, swelling of the brain inside the skull vault bones has resulted in flattening of the gyri and narrowing of the sulci of the cerebral convexities	80
5.14	Brainstem of a buffalo calf of <i>P. multocida</i> oral group showing normal to mild congestion of the surface of brainstem	81
5.15	Brainstem of a buffalo calf of <i>P. multocida</i> subcutaneous group showing moderate congestion of brainstem	82
5.16	Brainstem of a buffalo calf of lipopolysaccharide oral group showing normal to mild congestion of brainstem	82
5.17	Brainstem of a buffalo calf of lipopolysaccharide intravenous group showing mild to moderate congestion of the surface of brainstem	83
5.18	Brainstem of a buffalo calf of outer membrane proteins oral group showing normal to mild congestion of brainstem	83
5.19	Brainstem of a buffalo calf of outer membrane proteins subcutaneous group showing congestion and hemorrhage of brainstem	84
5.20	Spinal cord of a buffalo calf of <i>P. multocida</i> oral group showing no clear lesion of leptomeninges of spinal cord in: (A) thoracic region. (B) lumbosacral region	85
5.21	Spinal cord of a buffalo calf of <i>P. multocida</i> subcutaneous group showing moderate to severe congestion of leptomeninges of spinal cord in: (A) thoracic region. (B) lumbosacral region. Note the dark-black discoloration under the dura mater of the spinal cord. This is a subdural hemorrhage	85
5.22	Spinal cord of a buffalo calf of lipopolysaccharide oral group showing no clear lesion of leptomeninges of spinal cord in: (A) thoracic region. (B) lumbosacral region	86
5.23	Spinal cord of a buffalo calf of lipopolysaccharide intravenous group showing mild congestion of pia mater of spinal cord in: (A) thoracic region. (B) lumbosacral region. Note the mild dark-black discoloration due to subdural hemorrhage under the dura mater	86
5.24	Spinal cord of a buffalo calf of outer membrane proteins oral group showing no clear lesion of leptomeninges of spinal cord	87

	in: (A) thoracic region. (B) lumbosacral region	
5.25	Spinal cord of a buffalo calf of outer membrane proteins subcutaneous group showing subdural hemorrhage of leptomeninges of spinal cord in: (A) thoracic region. (B) lumbosacral region	87
5.26	Positive bacterial isolation of <i>P. multocida</i> B:2 from a buffalo calf of Pmsc group: (A) four cerebral lobes. (B) spinal cord	89
5.27	Polymerase chain reaction (PCR) confirmation of <i>P. multocida</i> B:2 isolates from different parts of the nervous system of calves of Pmsc grouped. Lane M, Marker; lanes 1 and 2, positive and negative controls; lanes 3-7 represent cerebrum, cerebellum, midbrain, medulla oblongata and spinal cord; lane 8 represent CSF	90
6.1	Total mean histopathological lesion score (\pm SEM) in the groups of buffalo calves following inoculation of <i>P. multocida</i> B:2 and its LPS and OMPs extracts via different routes	97
6.2	Photomicrograph of the cerebrum (temporal lobe) of a buffalo calf from <i>P. multocida</i> oral group showing (a) congested blood vessels; (b) perineuronal satellitosis typified by the presence of oligodendroglial cells around necrotic neurons and (c) microgliosis in an area of chromatolysis, (H&E, \times 200).	100
6.3	Photomicrograph of the cerebrum (temporal lobe) of a buffalo calf from <i>P. multocida</i> subcutaneous group showing (a) congested blood vessels; (b) diffuse aggregation of microglial cells with proliferation of oligodendroglial and astrocytes adjacent to the congested blood vessel (encephalitis) and (c) neuronal degeneration.	100
6.4	Photomicrograph of the cerebrum (frontal lobe) of a buffalo calf from lipopolysaccharide oral group showing (a) marked gliosis in the pia mater of the meninges; (b) prominent congested blood vessels in the pia mater and cerebral molecular layer with evidence of spongiosis (arrows); (c) perivascular lymphocytic cells aggregation in the cerebral cortex and (d) gliosis scattered in molecular layer.	101
6.5	Photomicrograph of the cerebrum (parietal lobe) of a buffalo calf from lipopolysaccharide intravenous group showing (a) marked microgliosis in the meninges (meningitis); (b) multiple microglial nodules scattered in the brain parenchyma consisting of aggregations of microglial cells, astrocytes and neutrophils (encephalitis); (c) congested blood vessels with	101

vacuolations due to spongiosis (arrow) in the molecular layer of the cerebral cortex.

- 6.6 Photomicrograph of the cerebrum (frontal lobe) of a buffalo calf from outer membrane proteins oral group showing (a) congested blood vessels in the cerebral tissue (b) satellitosis typified by proliferation of oligodendroglial cells around a degenerative neuron, (H&E, $\times 100$) 102
- 6.7 Photomicrograph of the cerebrum (frontal lobe) of a buffalo calf from outer membrane proteins subcutaneous group showing (a) nodular lesion characterized by aggregation of microglia around degenerating and necrotic neurons forming gitter cells; (b) satelliosis around cavitation formed by chromatolysis of a neuron and (c) congested capillaries with vacuolations due to spongiosis (arrows) in the cerebral tissue, (H&E, $\times 200$) 102
- 6.8 Photomicrograph of the cerebellum of a buffalo calf from *P. multocida* oral group showing (a) spongiosis characterized by vacuolations in the granular and Purkinje layers and (b) mild gliosis in the molecular layer. 105
- 6.9 Photomicrograph of the cerebellum of a buffalo calf from *P. multocida* subcutaneous group showing (a) hemorrhage in the cortex of the gray matter (molecular, Purkinje and granular layers); (b) red Purkinje cells characterized by pyknotic nuclei and eosinophilic cytoplasm of Purkinje cells; (c) spongiosis in the Purkinje and molecular layers and (d) Gliosis in the granular layer. 105
- 6.10 Photomicrograph of the cerebellum of a buffalo calf from lipopolysaccharide oral group showing (a) congestion of blood vessels in the molecular and granular layers; (b) spongiosis typified by vacuolations in the Purkinje layer and (c) marked gliosis in molecular layer, (H&E, $\times 200$) 106
- 6.11 Photomicrograph of the cerebellum of a buffalo calf from lipopolysaccharide intravenous group showing (a) congested blood vessels in the granular, molecular and Purkinje layers; (b) central chromatolysis in Purkinje cells and (c) gliosis in the molecular layer. 106
- 6.12 Photomicrograph of the cerebellum of a buffalo calf from outer membrane proteins oral group showing (a) congestion of blood vessels and capillaries, and spongiosis in the granular and Purkinje layers. 107
- 6.13 Photomicrograph of the cerebellum of a buffalo calf from outer membrane proteins subcutaneous group showing (a) 107

satellitosis around a necrotic neuron; (b) perivascular spongiosis around congested blood vessels in molecular layer and (c) complete disappearance of granular and Purkinje cells, (H&E, ×200).

- 6.14 Photomicrograph of the brainstem of a buffalo calf from *P. multocida* oral group showing (a) an area of focal hemorrhage close to a degenerative neuron and (b) proliferation of oligodendroglial and microglial cells around a necrotic neuronal cells, (H&E, ×200) 110
- 6.15 Photomicrograph of the brainstem of a buffalo calf from *P. multocida* subcutaneous group showing (a) a large congested blood vessel with perivascular oedema; (b) necrotic neurons typified by eosinophilic cytoplasm (c) mild areas of spongiosis characterized by vacuolations, (H&E, ×100) 110
- 6.16 Photomicrograph of the brainstem of a buffalo calf from lipopolysaccharide oral group showing (a) neuronal necrosis with satellitosis and (b) spongiosis around congested blood vessels, (H&E, ×100). 111
- 6.17 Photomicrograph of the brainstem of a buffalo calf from lipopolysaccharide intravenous group showing (a) a necrotic neuron with satellitosis; (b) vascular congestion with evidence of perivascular spongiosis; (c) oligodendroglial cells in an area of neuronal chromatolysis; (d) presence of degenerative neurons typified by vacuolar cytoplasm. 111
- 6.18 Photomicrograph of the brainstem of a buffalo calf from outer membrane proteins oral group showing (a) vascular congestion; (b) a degenerative neuron with chromatolysis and loss of Nissl substances (c) extensive spongiosis typified by vacuolations, (H&E, ×100). 112
- 6.19 Photomicrograph of the brainstem of a buffalo calf from outer membrane proteins subcutaneous group showing (a) vascular congestion with perivascular leucocytic cuffing (b) neuronal degeneration typified by absence of nuclear detail; (c) proliferation of oligodendroglial cells around degenerative neuronal cell and (d) gliosis scattered in the parenchyma, (H&E, ×100) 112
- 6.20 Photomicrograph of the spinal cord (thoracic region) of a buffalo calf from *P. multocida* oral group showing (a) congested blood vessels ; (b) necrosis with central chromatolysis of neuron cell body and proliferation of oligodendroglial cells, (H&E, ×200) 115
- 6.21 Photomicrograph of the spinal cord (cervical region) of a 115

- buffalo calf from *P. multocida* subcutaneous group showing (a) congested blood vessels with evidence of perivascular hemorrhages and (b) proliferation of oligodendroglial cells around a necrotic neuron characterized by cytoplasmic eosinophilia and nuclear karyorrhexis, (H&E, ×200).
- 6.22 Photomicrograph of the spinal cord of a buffalo calf from lipopolysaccharide oral group showing (a) perivascular spongiosis around congested blood vessels ; (b) necrosis of a neuron characterized by homogeneous, eosinophilic cytoplasm and nuclear pyknosis (c) gliosis in the parenchyma, (H&E, ×200) 116
- 6.23 Photomicrograph of the spinal cord of a buffalo calf from lipopolysaccharide intravenous group showing (a) a degenerative neuron with loss of nuclear detail ; (b) perivascular spongiosis around congested blood vessels in gray and white matter; (c) central chromatolysis of neuronal cell body characterized by karyorrhexis of nuclei with disappear of Nissl granules and proliferation of oligodendroglial cells , (H&E, ×200) 116
- 6.24 Photomicrograph of the spinal cord of a buffalo calf from outer membrane proteins oral group showing (a) perineural oedema around a necrotic neuron and (b) spongiosis around blood vessel, (H&E, ×200) 117
- 6.25 Photomicrograph of the spinal cord of a buffalo calf from outer membrane proteins subcutaneous group showing (a) extensive hemorrhage in the neuropil of the gray matter with (b) spongiosis around necrotic neurons typified by shrunken cytoplasm and nuclear pyknosis, (H&E, ×200). 117
- 6.26 Photomicrograph of the cranial nerve of a buffalo calf of *P. multocida* oral group showing (a) mononuclear cells infiltration between nerve fiber and around blood vessels, (H&E, ×200) 120
- 6.27 Photomicrograph of the cranial nerve of a buffalo calf of *P. multocida* subcutaneous group showing (a) congested blood vessels and inflammatory cells infiltration and (b) demyelination of axonal sheath, (H&E, ×200) 120
- 6.28 Photomicrograph of the cranial nerve of a buffalo calf of lipopolysaccharide oral group showing (a) wallerian degeneration of axon characterized by elongated irregular cleft with proliferation of oligodendroglial cells arranged linearly fascicular, (H&E, ×200) 121

- 6.29 Photomicrograph of the cranial nerve of a buffalo calf of lipopolysaccharide intravenous group showing (a) congested blood capillaries with inflammatory cells infiltration in the endoneurium; (b) mild gliosis and (c) vacuolation of fibers, (H&E, ×100) 121
- 6.30 Photomicrograph of the cranial nerve of a buffalo calf of outer membrane proteins oral group showing (a) congested blood capillaries with inflammatory cells infiltration in the endoneurium; (b) mild gliosis and (c) vacuolation of fibers, (H&E, ×200) 122
- 6.31 Photomicrograph of the cranial nerve of a buffalo calf of outer membrane proteins subcutaneous group showing (a) mild congestion of blood capillaries; (b) inflammatory cells infiltration between nerve fibers and (c) early axon degeneration in which the myelin sheath has collapsed forming balls and ovoids, (H&E, ×200) 122
- 6.32 Photomicrograph of a longitudinal section of a spinal nerve of a buffalo calf of *P. multocida* oral group showing (a) a few vacuolar fibers and (b) presence of eosinophilic cytoplasm (b), (H&E, ×200) 125
- 6.33 Photomicrograph of a longitudinal section of a spinal nerve of a buffalo calf of *P. multocida* subcutaneous group showing (a) demyelination of axon in addition, Schwann cells distal to the transaction also proliferate and make new myelin; (b) congested blood vessels in perineurium of spinal nerve and (c) degeneration of Schwann cells characterized of eosinophilic cytoplasm, (H&E, 400×) 125
- 6.34 Photomicrograph of a longitudinal section of a spinal nerve of a buffalo calf of lipopolysaccharide oral group showing (a) few vacuolar fibers, (H&E, ×200) 126
- 6.35 Photomicrograph of a longitudinal section of a spinal nerve of a buffalo calf of lipopolysaccharide intravenous group showing (a) few vacuolar fibers and (b) spindle shaped nuclei that appeared eosinophilic, (H&E, ×400) 126
- 6.36 Photomicrograph of a longitudinal section of a spinal nerve of a buffalo calf of outer membrane proteins oral group showing (a) few vacuolar fibers and (b) spindle shaped nuclei that appeared eosinophilic, (H&E, ×400) 127
- 6.37 Photomicrograph of a longitudinal section of a spinal nerve of a buffalo calf of outer membrane proteins subcutaneous group showing (a) few vacuolar fibers and (b) presence of vascular congestion, (H&E, ×400) 127

- 7.1 TNF- α concentration (\pm SEM) in CSF and serum of the groups of buffalo calves following inoculation of *P. multocida* B:2 and its LPS and OMPs extracts via different routes. All values were expressed as (\pm SEM), a, b, c, d, e and f values superscript within columns are significantly different at ($P < 0.05$) 138
- 7.2 IL-1 concentration (\pm SEM) in CSF and serum of the groups of buffalo calves following inoculation of *P. multocida* B:2 and its LPS and OMPs extracts via different routes. All values were expressed as (\pm SEM), a, b, c, d, e and f values superscript within columns are significantly different at ($P < 0.05$) 139
- 7.3 IL-6 concentration (\pm SEM) in CSF and serum of the groups of buffalo calves following inoculation of *P. multocida* B:2 and its LPS and OMPs extracts via different routes. All values were expressed as (\pm SEM), a, b, c, d, e and f values superscript within columns are significantly different at ($P < 0.05$) 141
- 7.4 Hp concentration (\pm SEM) in CSF and serum of the groups of buffalo calves following inoculation of *P. multocida* B:2 and its LPS and OMPs extracts via different routes. All values were expressed as (\pm SEM), a, b, c, d, e and f values superscript within columns are significantly different at ($P < 0.05$) 142
- 7.5 SAA concentration (\pm SEM) in CSF and serum of the groups of buffalo calves following inoculation of *P. multocida* B:2 and its LPS and OMPs extracts via different routes. All values were expressed as (\pm SEM), a, b, c, d, e and f values superscript within columns are significantly different at ($P < 0.05$) 144
- 7.6 CRP concentration (\pm SEM) in CSF and serum of the groups of buffalo calves following inoculation of *P. multocida* B:2 and its LPS and OMPs extracts via different routes. All values were expressed as (\pm SEM), a, b, c, d, e and f values superscript within columns are significantly different at ($P < 0.05$) 145

LIST OF ABBREVIATIONS

APP	Acute phase proteins
APTT	Activated partial thromboplastin time
BHI	Brain heart infusion
cfu	colony forming units
CSF	Cerebrospinal fluid
Ctrl	Control
ELISA	Enzyme linked immunosorbent assay
h	hour
Hp	Haptoglobin
HR	Heart rate
HRP-avidin	Horse radish peroxidase avidin
HS	Haemorrhagic septicaemia
IACUC	Institutional Animal Care and Use Committee
ICR	Institute of Cancer Research
IL-1	Interleukins-1
IL-6	Interleukins-6
LPS	Lipopolysaccharide
LPSiv	LPS of <i>P. multocida</i> B:2 in oculted via intravenous route
LPSor	LPS of <i>P. multocida</i> B:2 inoculated via oral route
MHLS	Mean histopathological lesion score
ml	millilitre
MSLS	Mean surface lesion score
min	minute
ng	nanograms

nm	nanometers
OD	Optical density
OMP _{or}	OMP of <i>P. multocida</i> B:2 inoculated via oral route
OMPs	Outer membrane proteins
OMP _{sc}	OMP of <i>P. multocida</i> B:2 inoculated subcutaneous route
PBS	Phosphate buffered saline
PCR	Polymerase chain reaction
pg	picogram
P _{mor}	<i>P. multocida</i> B:2 inoculated via oral route
P _{msc}	<i>P. multocida</i> B:2 inoculated via subcutaneous route
rpm	revolutions per minute
RR	Respiratory rate
RT	Rectal temperature
SAA	Serum amyloid A
Sec	second
SDS-PAGE	Sodium dodecyl sulphate-polyacrylamide gel electrophoresis
SIM	Sulphur indole motility
TGCS	Total general clinical score
TMB-Substrate	Tetramethylbenzidine liquid substrate
TNCS	Total nervous clinical score
TSI	Triple sugar iron
VRI	Veterinary Research Institute

CHAPTER 1

INTRODUCTION

1.1 General Introduction

Pasteurella multocida is the causative agent of a wide range of diseases in many species of animals, including haemorrhagic septicaemia (HS) in cattle and buffalo, atrophic rhinitis in swine, fowl cholera in poultry and respiratory disease in ungulates and rabbits (De Alwis, 1999; Harper *et al.*, 2006; Wilkie *et al.*, 2012). It also affect human and causes wound abscesses and meningitis especially after cat or dog bites (Wade *et al.*, 1999; Green *et al.*, 2002). Haemorrhagic septicaemia (HS) is an acute septicemic disease principally affecting cattle and buffaloes caused by specific serotypes B:2 and E:2 of *P. multocida* in Asia and Africa, respectively (De Alwis, 1999; World Organization for Animal Health, 2012; Jesse *et al.*, 2013d). The disease has economic impact due to high mortality rates among the affected animals especially buffaloes (De Alwis, 1999; Annas *et al.*, 2015a). It is a well-known fact that HS occurs most commonly in cattle and buffaloes. However, buffaloes are thought to be more susceptible than cattle, and young animals have been observed to be more susceptible than adults (Horadagoda *et al.*, 2002; Annas *et al.*, 2015a). Moreover, predisposing factors such as movement of animals, poor husbandry conditions and weather changes could vastly affect the immunostatus of the animals and finally result in explosive outbreaks (De Alwis, 1992b; Benkirane and De Alwis, 2002; McFadden *et al.*, 2011).

Complex interactions between specific host factors such as species, age and immune status, and virulence factors of *P. multocida* B:2, mainly lipopolysaccharides (LPS), outer membrane proteins (OMPs), capsule and adhesions are thought to play a key role in causing HS (Harper *et al.*, 2012; Othman *et al.*, 2012; Jesse *et al.*, 2013a; Jesse *et al.*, 2013d; Kharb and Charan, 2013). The LPS of *P. multocida* constitutes the major components of the bacterial cell surface and it has a major role in the disease pathogenesis (Horadagoda *et al.*, 2001; Harper *et al.*, 2012; Jesse *et al.*, 2013a). Additionally, outer membrane proteins of *P. multocida* are thought to have a role in the disease pathogenesis (Hatfaludi *et al.*, 2010), and recent studies showed the ability of *P. multocida* B:2 OMPs to induce the disease in both murine model and real hosts (Abdullah *et al.*, 2013a; Jesse *et al.*, 2013f). Furthermore, in experimental studies related to pathogenesis of HS, different routes of inoculation of *P. multocida* B:2 or its immunogens LPS and OMPs have been found to induce different pathogenesis pathways and finally might affect the produced clinical and pathological pictures of the disease (Dowling *et al.*, 2002; Horadagoda *et al.*, 2002; Jesse *et al.*, 2013d; Annas *et al.*, 2014b). Respiratory route (intranasal infection by aerosols or intratracheal) and oral drenching results in a longer course of disease and more profound lesions, while subcutaneous inoculation results in rapid onset of disease, a shorter course and less marked pathological lesions mainly describes as peracute form (Dowling *et al.*, 2002; Abubakar *et al.*, 2013; Jesse *et al.*, 2013d; Annas *et al.*, 2015b).

Buffaloes affected by HS outbreaks usually show short clinical course lasting a few hours with signs of severe depression, dyspnea, elevated temperature, salivation, nasal discharge followed by swelling of the ventral cervical region and brisket, respiratory distress and finally death (Carter and De Alwis, 1992; Khan *et al.*, 2011).

On the other hand, in experimental infection studies using *P. multocida* B:2, three main phases usually could be observed following short incubation period, one of temperature elevation, a phase of clinical signs and a terminal phase of recumbency (De Alwis, 1999; Dowling *et al.*, 2002; Jesse *et al.*, 2013d). Nevertheless, in many cases, overlap between the clinical phases are occur with varying degrees with less define phases when the disease course is short (Odugbo *et al.*, 2005; Jesse *et al.*, 2013d). Examination of the nervous system is one of the most important factors that can precisely identify the neuroanatomic location or locations of any abnormalities (Jackson *et al.*, 2002b). Accordingly, any defect in a specific anatomic region of the nervous system could lead to affect the associated organ and finally produced the related clinical signs (Constable, 2004). Signs such as abnormalities in gait, decreased rumen motility, defect in vision and recumbency at the final stage of the disease might be related to the defect in the corresponding part of the nervous system. Rumen hypomotility 0/min after 4 h post infection has been reported by Jesse *et al.* (2013d) in a study involved buffalo calves inoculated intramuscularly by 1×10^{12} cfu/mL of *P. multocida* B:2. In the same manner, recumbency and inability to rise were the most prominent signs of abnormal gait especially at the end stage of the endotoxic shock (De Alwis, 1999; Abubakar and Zamri-Saad, 2011). Although manifestation of the nervous clinical signs of animals affected by *P. multocida* B:2 is rarely reported, lack of special interest in investigating the possible clinical and neurological responses following infection with *P. multocida* B:2 could be a major contributor to the rarity of such responses (Marza *et al.*, 2015).

During necropsy examination, the main focus of the examiner is usually paid on the pathological changes of the respiratory and gastrointestinal tracts as main affected sites of animals affected by HS, and this render the examination of other parts of the body seldom investigated (Dowling *et al.*, 2002; Odugbo *et al.*, 2005; Jesse *et al.*, 2013d). However, recent studies reported the involvement of urinary tract as new localization site of *P. multocida* B:2 in buffalo surviving experimental HS, suggesting their role in the pathogenesis of HS. (Annas *et al.*, 2014a; Annas *et al.*, 2014b). While the involvement of the nervous system due to *P. multocida* has been reported mainly in human (Zaramella *et al.*, 1999; Green *et al.*, 2002), few studies reported the presence of meningitis or encephalitis in outbreaks of HS as central nervous tissues are not often routinely examined (Lane *et al.*, 1992). Nevertheless, recent experimental studies on pathogenesis of HS re-emphasized the involvement of nervous system with successful bacterial isolation of *P. multocida* B:2 from the brain (Abubakar and Zamri-Saad, 2011; Jesse *et al.*, 2013c; Khaleel *et al.*, 2014). These findings rise a question about the ability of this enigmatic pathogen to involve other sites like the nervous system (Marza *et al.*, 2015).

Besides clinical and pathological changes, pathophysiological responses have been investigated in the pathogenesis studies of HS using murine and buffalo models (Abdullah *et al.*, 2013b; Abdullah *et al.*, 2014b; Ali *et al.*, 2014). Cytokines including interleukine-1 β (IL-1 β), interleukine-6 (IL-6) and tumor necrosis factor- α (TNF- α) have been shown able to reproduce toxic effects induced by LPS of *P. multocida* (Horadagoda *et al.*, 2001; Harper *et al.*, 2011; Abdullah *et al.*, 2013a; Abubakar *et al.*, 2013; Ali *et al.*, 2014). Moreover, administration of LPS has been reported to induce neuroinflammation and increases proinflammatory cytokines, such as IL-1 β , IL-6 and TNF- α (Song and Wang, 2011). Acute phase proteins (APPs) also have been investigated extensively as potential biomarkers for detection of HS (Horadagoda *et al.*, 2001; Abdullah *et al.*, 2013b; Khaleel *et al.*, 2013). Jesse *et al.* (2013a) reported an increased in serum amyloid A (SAA), haptoglobin (Hp) after experimental infection of buffalo calves with *P. multocida* B:2 and inoculation of its immunogens, LPS and OPMs. Although extensive studies have been done on cytokines and APPs using different animal models (Abdullah *et al.*, 2013b; Jesse *et al.*, 2013a; Khaleel *et al.*, 2013; Abdullah *et al.*, 2014b; Ali *et al.*, 2014), cytokine and AAPPs responses and their effect on the nervous system, especially the cerebrospinal fluid (CSF), are new area need to be studied (Marza *et al.*, 2015).

According to the previous facts, it is clear that there are clinical, pathological and pathophysiological evidences about the involvement of the nervous system in the pathogenesis of *P. multocida* B:2 infection. Such evidences have been generated from different studies. However, these studies lack detailed information about the effect of *P. multocida* B:2 on different parts of the nervous system and further studies are needed for better understanding of the pathogenesis of such devastating disease.

1.2 Problem statement

Despite continuing researches on pathogenesis of *P. multocida* B:2 for several decades, the involvement of the nervous system as a newly investigated localization site during the disease course of HS has not been studied in details yet in buffaloes as a natural host

1.3 Hypothesis of the study

Experimental infection of buffaloes with *P. multocida* B:2 and its LPS and OPMs immunogens via different routes could induce clinical, pathological and pathophysiological changes in the nervous system.

1.4 Objectives of the study

The objectives of the study were:

1. To evaluate clinical signs and neurological responses of buffaloes inoculated with *P. multocida* B:2 whole bacteria and its LPS and OMPs immunogens extracts using oral, subcutaneous and intravenous routes.
2. To detect the presence of *P. multocida* B:2 in nervous tissues and cerebrospinal fluid (CSF) of buffaloes inoculated with *P. multocida* B:2 via various route using standard bacterial culture and polymerase chain reaction (PCR).
3. To determine pathological and histopathological changes of different anatomic regions of the nervous system of buffaloes inoculated with *P. multocida* B:2 whole bacteria and its LPS and OMPs immunogens extracts using oral, subcutaneous and intravenous routes.
4. To evaluate classical parameters, cytokine and acute phase proteins (APPs) responses in the CSF of buffaloes inoculated with *P. multocida* B:2 whole bacteria and its LPS and OMPs immunogens extracts using oral, subcutaneous and intravenous routes.

REFERENCES

- Aalbæk, B., Eriksen, L., Rimler, R. B., Leifsson, P. S., Basse, A., Christiansen, T. and Eriksen, E. (1999). Typing of *Pasteurella multocida* from haemorrhagic septicaemia in Danish fallow deer (*Dama dama*). *Acta Pathologica, Microbiologica et Immunologica Scandinavica*, 107(7-12), 913-920.
- Abbott, N. J., Patabendige, A. A., Dolman, D. E., Yusof, S. R. and Begley, D. J. (2010). Structure and function of the blood–brain barrier. *Neurobiology of Disease*, 37(1), 13-25.
- Abdelbaset, A., Abd Ellah, M. R., Abd ElGhaffar, S. and Sadiék, A. (2014). Acute-phase proteins in different pathological conditions at the lungs of buffaloes. *Comparative Clinical Pathology*, 23(4), 823-828.
- Abdullah, F. F. J., Adamu, L., Osman, A. Y., Haron, A. W., Saharee, A. A., Abdullah, R., Saad, M. Z. and Zakaria, Z. (2013a). Biochemical and hematological alterations in mice inoculated with outer membrane protein, lipopolysaccharides and whole cells of *Pasteurella multocida* type B: 2. *American Journal of Animal and Veterinary Sciences*, 8(3), 152-158.
- Abdullah, F. F. J., Adamu, L., Osman, A. Y., Saad, M. Z., Zakaria, Z., Abdullah, R. and Saharee, A. A. (2013b). Acute phase protein profile and clinicopathological changes in mice associated with the infection of *Pasteurella multocida* type B and the bacterial lipopolysaccharide and outer membrane protein immunogens. *Journal of Animal and Veterinary Advances*, 12(2), 186-193.
- Abdullah, F. F. J., Adamu, L., Abba, Y., Tijjani, A., Mohammed, K., Omar, A. and Saharee, A. A. (2014a). Effect of dose dependent oral inoculation of *Pasteurella multocida* type B: 2 in mice: molecular detection and histopathological evaluation. *Research Opinions in Animal and Veterinary Sciences*, 4(10), 535-539.
- Abdullah, F. F. J., Ali, O. S., Adamu, L., Abba, Y., Hamzah, H. B., Mohd-Azmi, M. L., Haron, A. W. and Zamri-saad, M. (2014b). Modifications in serum amyloid A and haptoglobin in mice following oral inoculation of graded doses of *P. multocida* type B: 2 and its lipopolysaccharide. *Research Opinions in Animal and Veterinary Sciences*, 4(11), 587-592.
- Abubakar, M., Zamri-Saad, M. and Jasni, S. (2013). Ultrastructural changes and bacterial localization in buffalo calves following oral exposure to *Pasteurella multocida* B: 2. *Pakistan Veterinary Journal*, 33(1), 101-106.
- Abubakar, M., Zamri-Saad, M., Jasni, S. and Zuki, A. (2016). Immunohistochemical evaluation of lesions in the gastrointestinal tract of buffalo (*Bubalus bubalis*) calves orally exposed to *Pasteurella multocida* B: 2. *Sokoto Journal of Veterinary Sciences*, 14(1), 21-27.

- Abubakar, M. S. and Zamri-Saad, M. (2011). Clinico-pathological changes in buffalo calves following oral exposure to *Pasteurella multocida* B:2. *Basic and Applied Pathology*, 4(4), 130-135.
- Adamczyk, K., Pokorska, J., Makulska, J., Earley, B. and Mazurek, M. (2013). Genetic analysis and evaluation of behavioural traits in cattle. *Livestock Science*, 154(1), 1-12.
- Aguirre, G. K., Komáromy, A. M., Cideciyan, A. V., Brainard, D. H., Aleman, T. S., Roman, A. J., Avants, B. B., Gee, J. C., Korczykowski, M. and Hauswirth, W. W. (2007). Canine and human visual cortex intact and responsive despite early retinal blindness from RPE65 mutation. *PLoS Medicine*, 4(6), 1117-1128.
- Ainsworth, D. M. (1999). Rhodococcal infections in foals. *Equine Veterinary Education*, 11(4), 191-198.
- Al-Hasani, K., Boyce, J., McCarl, V. P., Bottomley, S., Wilkie, I. and Adler, B. (2007). Identification of novel immunogens in *Pasteurella multocida*. *Microbial Cell Factories*, 6(1), 1-5
- Al Laham, S. A. and Al Fadel, F. M. (2013). The anti-bacterial effect of punica granatum extracts against antibiotic resistant *Pasteurella haemolytica*. *Jundishapur Journal of Microbiology*, 6(9), 1-5
- Allam, N.G. and Elsliek, S.E., 2011. Evaluation of lipopolysaccharide, exopolysaccharide and mutant strain as subunit vaccines against *Serratia marcescens* W225. *African Journal of Microbiology Research*, 5(12), pp.1389-1397.
- Alarcón, M., Gulla, S., Røsaeg, M., Rønneseth, A., Wergeland, H., Poppe, T., Nilsen, H. and Colquhoun, D. (2015). Pasteurellosis in lump sucker *Cyclopterus lumpus*, farmed in Norway. *Journal of Fish Diseases*, 39(4), 489-495.
- Alçığır, M., Vural, S. A., Müştak, K. and Tunç, A. S. (2014). Naturally infection with *Pasteurella multocida* in a rabbit: the pathological and microbiological findings. *Ankara Universitesi Veteriner Fakultesi Dergisi*, 61(2), 147-150.
- Ahn, J.H., Hong, I.P., Bok, J.I., Kim, B.Y., Song, J. and Weon, H.Y., (2012). Pyrosequencing analysis of the bacterial communities in the guts of honey bees *Apis cerana* and *Apis mellifera* in Korea. *Journal of Microbiology*, 50(5), 735-745.
- Ali, O. S., Adamu, L., Jesse, F. F. A., Ilyasu, Y., Abba, Y., Hamzah, H., Mohd-Azmi, M. L., Haron, A. W. and Saad, M. Z. (2014). Alteration in interleukin-1 β and interleukin-6 in mice inoculated through the oral route using graded doses of *Pasteurella multocida* type B:2 and its lipopolysaccharide. *American Journal of Animal and Veterinary Sciences*, 9(4), 177-184.

- Ali, O. S., Adamu, L., Abdullah, F. F. J., Abba, Y., Hamzah, H. B., Mohd-Azmi, M., Haron, A. W. and Zamri-Saad, M. (2015). Haematological and histopathological vicissitudes following oral inoculation of graded doses of *Pasteurella multocida* type B:2 and its lipopolysaccharide in mice. *Veterinary Science and Technology*, 6(2), 1-7
- Ametaj, B. N., Hosseini, A., Odhiambo, J. F., Iqbal, S., Sharma, S., Deng, Q. Dunn, S. M. (2011). Application of acute phase proteins for monitoring inflammatory states in cattle. In F. Veas (Ed.), *Acute Phase Proteins as early Non-Specific Biomarkers of Human and Veterinary Diseases: China*, InTech Open Access Publisher. 299-354
- Annas, S., Zamri-Saad, M., Abubakar, M., Jesse, F. and Zunita, Z. (2014a). Distribution of *Pasteurella multocida* B: 2 in the respiratory, gastrointestinal and urinary tracts of buffaloes following experimental subcutaneous inoculation. *Journal of Veterinary Science and Technology*, 5(3), 1-6.
- Annas, S., Zamri-Saad, M., Jesse, F. F. and Zunita, Z. (2014b). New sites of localisation of *Pasteurella multocida* B: 2 in buffalo surviving experimental haemorrhagic septicaemia. *BMC Veterinary Research*, 10(88), 1-7.
- Annas, S., Zamri-Saad, M., Jesse, F. F. A. and Zunita, Z. (2015a). Comparative clinicopathological changes in buffalo and cattle following infection by *Pasteurella multocida* B:2. *Microbial Pathogenesis*, 88, 94-102.
- Annas, S., Abubakar, M., Zamri-Saad, M., Jesse, F. and Zunita, Z. (2015b). Pathological changes in the respiratory, gastrointestinal and urinary tracts of buffalo calves following experimental hemorrhagic septicaemia. *Pakistan Veterinary Journal*, 35(4), 430-435.
- Anoop, A., Singh, P. K., Jacob, R. S. and Maji, S. K. (2010). CSF biomarkers for Alzheimer's disease diagnosis. *International Journal of Alzheimer's Disease*, 2010, 1-7
- Aquino, R. S., Lee, E. S. and Park, P. W. (2010). Diverse functions of glycosaminoglycans in infectious diseases. *Progress in Molecular Biology and Translational Science*, 93, 373-394.
- Armstrong, G., Sen, R. and Wilkinson, J. (2000). *Pasteurella multocida* meningitis in an adult: case report. *Journal of Clinical Pathology*, 53(3), 234-235.
- Aronoff, D.M. and Neilson, E.G., (2001). Antipyretics: mechanisms of action and clinical use in fever suppression. *The American Journal of Medicine*, 111(4), 304-315.
- Aubert, A. and Renault, J. (2008). Cytokines and immune-related behaviors. *Neuroimmune Biology*, 6, 527-547.

- Audoy-Rémus, J., Richard, J.-F., Soulet, D., Zhou, H., Kubes, P. and Vallières, L. (2008). Rod-shaped monocytes patrol the brain vasculature and give rise to perivascular macrophages under the influence of proinflammatory cytokines and angiopoietin-2. *The Journal of Neuroscience*, 28(41), 10187-10199.
- Avakian, A. P., Kleven, S. and Glisson, J. (1988). Evaluation of the specificity and sensitivity of two commercial enzyme-linked immunosorbent assay kits, the serum plate agglutination test, and the hemagglutination-inhibition test for antibodies formed in response to *Mycoplasma gallisepticum*. *Avian Diseases*, 1988, 262-272.
- Aydogdu, I., Ertekin, C., Tarlaci, S., Turman, B., Kiylioglu, N. and Secil, Y. (2001). Dysphagia in Lateral Medullary Infarction (Wallenberg's Syndrome) An acute disconnection syndrome in premotor neurons related to swallowing activity. *Stroke*, 32(9), 2081-2087.
- Banani, M., Hablolvarid, M., Momayez, R., Nouri, A., Ghodsian, N., Ashtari, A. and Mirzaei, S. (2015). Isolation of *Ornithobacterium rhinotracheale* from the brains of commercial broiler breeder chickens with meningitis and encephalitis. *Archives of Razi*, 70(3), 203-209.
- Basagoudanavar, S. H., Singh, D. K. and Varshney, B. C. (2006). Immunization with outer membrane proteins of *Pasteurella multocida* (6:B) provides protection in mice. *Journal of Veterinary Medicine*, 53(10), 524-530.
- Bellino, C., Miniscalco, B., Bertone, I., Cagnasso, A., Occhiena, E., Gianella, P. and D'Angelo, A. (2015). Analysis of cerebrospinal fluid from cattle with central nervous system disorders after storage for 24 hours with autologous serum. *BMC Veterinary Research*, 11(1), 1-5.
- Benkirane, A. and De Alwis, M. (2002). Haemorrhagic septicaemia, its significance, prevention and control in Asia. *Veterinary Medicine-Czech*, 47(8), 234-240.
- Bischoff, K. and Mukai, M. (2012). Toxicity of over-the-counter drugs. *Veterinary Toxicology: Basic and Clinical Principles*, 2012, 363-390
- Biswas, A., Shivachandra, S., Saxena, M., Kumar, A., Singh, V. and Srivastava, S. (2004). Molecular variability among strains of *Pasteurella multocida* isolated from an outbreak of haemorrhagic septicaemia in India. *Veterinary Research Communications*, 28(4), 287-298.
- Bogdanski, R., Blobner, M., Becker, I., Hänel, F., Fink, H., and Kochs, E. (2000). Cerebral histopathology following portal venous infusion of bacteria in a chronic porcine model. *The Journal of the American Society of Anesthesiologists*, 93(3), 793-804.
- Borghese, A. and Mazzi, M. (2005). Buffalo population and strategies in the world. *Buffalo Production and Research*, 67, 1-39.

- Boulant, J. A. (2000). Role of the preoptic-anterior hypothalamus in thermoregulation and fever. *Clinical Infectious Diseases*, 31(5), 157-161.
- Boveja, B.R. and Widhany, A., Boveja Birinder R, 2006. Method and system for modulating the vagus nerve (10th cranial nerve) with electrical pulses using implanted and external components, to provide therapy neurological and neuropsychiatric disorders. U.S. Patent 7,076,307.
- Boyce, J., Lo, R., Wilkie, I. and Adler, B. (2004). *Pasteurella* and *Mannheimia*. *Pathogenesis of Bacterial Infections in Animals*. 3rd Edition. Oxford, UK: Blackwell Publishing, 2004, 273-294.
- Boyce, J. D. and Adler, B. (2000). The capsule is a virulence determinant in the pathogenesis of *Pasteurella multocida* M1404 (B:2). *Infection and Immunity*, 68(6), 3463-3468.
- Boyce, J. D. and Adler, B. (2006). How does *Pasteurella multocida* respond to the host environment. *Current Opinion in Microbiology*, 9(1), 117-122.
- Brandt, T., Dieterich, M. and Danek, A. (1994). Vestibular cortex lesions affect the perception of verticality. *Annals of Neurology*, 35(4), 403-412.
- Braun, U., Feige, K., Schweizer, G. and Pospischill, A. (2005). Clinical findings and treatment of 30 cattle with botulism. *Veterinary Record: Journal of the British Veterinary Association*, 156(14), 438-414.
- Brenner, D. J., Fanning, G. R., Knutson, J. K. L., Steigerwalt, A. G. and Krichevsky, M. I. (1984). Attempts to classify *Herbicola* group-*Enterobacter* agglomerans strains by deoxyribonucleic acid hybridization and phenotypic tests. *International Journal of Systematic and Evolutionary Microbiology*, 34(1), 45-55.
- Brickell, S., Thomas, L., Long, K., Panaccio, M. and Widders, P. (1998). Development of a PCR test based on a gene region associated with the pathogenicity of *Pasteurella multocida* serotype B: 2, the causal agent of haemorrhagic septicaemia in Asia. *Veterinary Microbiology*, 59(4), 295-307.
- Briggs, D. and Skeeles, J. (1984). An enzyme-linked immunosorbent assay for detecting antibodies to *Pasteurella multocida* in chickens. *Avian Diseases*, 1984, 208-215.
- Brough, D., Tyrrell, P. J. and Allan, S. M. (2011). Regulation of interleukin-1 in acute brain injury. *Trends in Pharmacological Sciences*, 32(10), 617-622.
- Budras, K.-D., Habel, R. E. Mulling, C. K., Greenhough, P. K. (2011). *Bovine Anatomy*. 2nd Edition.: Schlütersche, UK. 34-35.

- Carlson, N. G., Wieggel, W. A., Chen, J., Bacchi, A., Rogers, S. W. and Gahring, L. C. (1999). Inflammatory cytokines IL-1 α , IL-1 β , IL-6, and TNF- α impart neuroprotection to an excitotoxin through distinct pathways. *The Journal of Immunology*, 163(7), 3963-3968.
- Carter, G. and De Alwis, M. (1989). Haemorrhagic septicaemia. *Pasteurella In Pasteurella and Pasteurellosis*, Adlam C. and Rutter JM., London, UK, Academic Press, 131-160.
- Catry, B., Haesebrouck, F., Vlieghe, S. D., Feyen, B., Vanrobaeys, M., Opsomer, G., Schwarz, S. and Kruif, A. D. (2005). Variability in acquired resistance of *Pasteurella* and *Mannheimia* isolates from the nasopharynx of calves, with particular reference to different herd types. *Microbial Drug Resistance*, 11(4), 387-394.
- Cebra, C., Anderson, D. E., Tibary, A., Van Saun, R. J. and Johnson, L. W. (2014). *Llama and alpaca care: Medicine, surgery, reproduction, nutrition, and herd health*. 1st Edition. Elsevier Health Sciences. 450-455.
- Cece, J. A., Lawson, W., Biller, H. F., Eden, A. R. and Parisier, S. C. (1987). Complications in the management of large glomus jugulae tumors. *The Laryngoscope*, 97(2), 152-157.
- Ceciliani, F., Ceron, J., Eckersall, P. and Sauerwein, H. (2012). Acute phase proteins in ruminants. *Journal of Proteomics*, 75(14), 4207-4231.
- Chandrasekaran, A., Sengupta, S., Berry, D.A., Holley, K., Zhao, G. and Sasisekharan, R., (2013). Methods and compositions related to the modulation of intercellular junctions. Massachusetts Institute of Technology. U.S. Patent 8,529,889.
- Cheng, N., He, R., Tian, J., Patrick, P. Y. and Richard, D. Y. (2008). Cutting edge: TLR2 is a functional receptor for acute-phase serum amyloid A. *The Journal of Immunology*, 181(1), 22-26.
- Chowdhury, M., Mitra, J., Sarkar, S., Samanta, T. and Roy, B. (2014). PCR and electron microscopy based diagnosis of an outbreak of haemorrhagic septicaemia in buffalo and its control in a farm of West Bengal, India. *Exploratory Animal and Medical Research*, 4(1), 86-94.
- Christensen, H., Bisgaard, M., Bojesen, A. M., Mutters, R. and Olsen, J. E. (2003). Genetic relationships among avian isolates classified as *Pasteurella haemolytica*, 'Actinobacillus salpingitidis' or *Pasteurella anatis* with proposal of *Gallibacterium anatis* gen. nov., comb. nov. and description of additional genomospecies within *Gallibacterium* gen. nov. *International Journal of Systematic and Evolutionary Microbiology*, 53(1), 275-287.

- Chung, E. L., Jesse, F. F., Ibrahim, H. H., Marza, A. D., Zamri-Saad, M., Haron, A. W., Lila, M. A. M. and Norsidin, M. J. (2016). Clinico-pathology, hematology and biochemistry responses in buffaloes towards *Pasteurella multocida* type B: 2 immunogen lypopolysaccharide via oral and intravenous routes of infection. *Microbial Pathogenesis*, 91, 141-154.
- Chung, J. Y., Wilkie, I., Boyce, J. D., Townsend, K. M., Frost, A. J., Ghoddusi, M. and Adler, B. (2001). Role of capsule in the pathogenesis of fowl cholera caused by *Pasteurella multocida* serogroup A. *Infection and Immunity*, 69(4), 2487-2492.
- Chung, E. L., Jesse, F. F. A., Adamu, L., Marza, A. D., Ibrahim, H. H., Zamri-Saad, M., Haron, A. W., Saharee, A. A., Mohd Azmi, M. and Omar, A. R. (2015). Clinico-pathology, hematology, and biochemistry responses toward *Pasteurella multocida* Type B: 2 via oral and subcutaneous route of infections. *Veterinary World*, 8(6), 783-792.
- Clyne, B. and Olshaker, J.S., 1999. The C-reactive protein. *The Journal of Emergency Medicine*, 17(6), 1019-1025.
- Confer, A. W. and Ayalew, S. (2013). The OmpA family of proteins: roles in bacterial pathogenesis and immunity. *Veterinary Microbiology*, 163(3), 207-222.
- Constable, P. D. (2004). Clinical examination of the ruminant nervous system. *Veterinary Clinics of North America: Food Animal Practice*, 20(2), 185-214.
- Cray, C., Zaias, J. and Altman, N. H. (2009). Acute phase response in animals: A review. *Comparative Medicine*, 59(6), 517-526.
- Crilly, J.P., Rzechorzek, N. and Scott, P., (2015). Diagnosing limb paresis and paralysis in sheep. *In practice*, 37(10), 490-507.
- Cunningham, E. T. and Sawchenko, P. E. (2000). Dorsal medullary pathways subserving oromotor reflexes in the rat: implications for the central neural control of swallowing. *Journal of Comparative Neurology*, 417(4), 448-466.
- Dabo, S., Taylor, J. and Confer, A. (2007). *Pasteurella multocida* and bovine respiratory disease. *Animal Health Research Reviews*, 8(02), 129-150.
- Dabo, S. M., Confer, A. W. and Murphy, G. L. (1997). Outer membrane proteins of bovine *Pasteurella multocida* serogroup A isolates. *Veterinary Microbiology*, 54(2), 167-183.
- Dabo, S. M., Confer, A. W., Montelongo, M. and Lu, Y.-S. (1999). Characterization of rabbit *Pasteurella multocida* isolates by use of whole-cell, outer-membrane, and polymerase chain reaction typing. *Comparative Medicine*, 49(5), 551-559.

- Dagleish, M. P., Finlayson, J., Bayne, C., MacDonald, S., Sales, J. and Hodgson, J. C. (2010). Characterization and time course of pulmonary lesions in calves after intratracheal infection with *Pasteurella multocida* A:3. *Journal of Comparative Pathology*, 142(2-3), 157-169.
- Davies, A., Stone, S., Kidd, D. and MacMahon, J. (1995). Pharyngeal sensation and gag reflex in healthy subjects. *The Lancet*, 345(8948), 487-488.
- Davies, R. L., MacCorquodale, R. and Reilly, S. (2004). Characterisation of bovine strains of *Pasteurella multocida* and comparison with isolates of avian, ovine and porcine origin. *Veterinary Microbiology*, 99(2), 145-158.
- Davis, J. L., Gilger, B. C., Spaulding, K., Robertson, I. D. and Jones, S. L. (2002). Nasal adenocarcinoma with diffuse metastases involving the orbit, cerebrum, and multiple cranial nerves in a horse. *Journal of the American Veterinary Medical Association*, 221(10), 1460-1463.
- Dawkins, H., Johnson, R., Spencer, T. and Adler, B. (1990). *Pasteurella multocida* infections in mice with reference to haemorrhagic septicaemia in cattle and buffalo. *Immunology and Cell Biology*, 68, 57-61.
- De Alwis, M. and Carter, G. (1980). Preliminary field trials with a streptomycin-dependent vaccine against haemorrhagic septicaemia. *The Veterinary Record*, 106(21), 435-437.
- De Alwis, M.C.L. (1992b). Haemorrhagic septicaemia- A general review. *British Veterinary Journal*, 148, 99-112.
- De Alwis, M. C. L. (1992). Pasteurellosis in production animals: a review. In Australian Centre for International Agricultural Research (ACIAR) Proceedings, Vol. 43, 11-22.
- DeAlwis, M.C.L. (1981). Mortality among cattle and buffaloes in Sri Lanka due to Haemorrhagic septicaemia. *Tropical Animal Health and Production*, 13: 195-202.
- De Alwis, M. (1995). Haemorrhagic septicaemia (*Pasteurella multocida* serotype B: 2 and E: 2 infection) in cattle and buffaloes *Haemophilus, Actinobacillus, and Pasteurella*. London: Plenum Press, 9-24.
- De Alwis, M. C. (1999). Haemorrhagic septicaemia. Canberra, Australia: In Australian Centre for International Agricultural Research (ACIAR) proceedings, Vol. 43, 26-51.
- De Alwis, M. C. L., Wijewardana, T., Gomis, A. U. and Vipulasiri, A. A. (1990). Persistence of the carrier status in haemorrhagic septicaemia (*Pasteurella multocida* serotype 6:B infection) in buffaloes. *Tropical Animal Health and Production*, 22(3), 185-194.

- De Alwis, M. C. L. (1992b). Haemorrhagic septicaemia-A general review. *British Veterinary Journal*, 148(2), 99-112.
- de Leon-Casasola, O. A. (2007). Pain pathways and mechanisms of neuropathic pain. *Acta Neurológica Colombiana*, 30(1), 133-138
- de Oliveira Filho, J. X., Morés, M. A., Rebelatto, R., Agnol, A. M., Plieski, C. L., Klein, C. S., Barcellos, D. E. and Morés, N. (2015). *Pasteurella multocida* type A as the primary agent of pneumonia and septicaemia in pigs1. *Pesquisa Veterinária Brasileira*, 35(8), 716-724.
- de Vries, H. E., Kuiper, J., de Boer, A. G., Van Berkel, T. J. and Breimer, D. D. (1997). The blood-brain barrier in neuroinflammatory diseases. *Pharmacological Reviews*, 49(2), 143-156.
- DeAngelis, P. L., Gunay, N. S., Toida, T., Mao, W.-j. and Linhardt, R. J. (2002). Identification of the capsular polysaccharides of type D and F *Pasteurella multocida* as unmodified heparin and chondroitin, respectively. *Carbohydrate Research*, 337(17), 1547-1552.
- DeAngelis, P. L. and White, C. L. (2004). Identification of a distinct, cryptic heparosan synthase from *Pasteurella multocida* types A, D, and F. *Journal of Bacteriology*, 186(24), 8529-8532.
- Dehkordi, F. S., Haghghi Borujeni, M. R., Rahimi, E. and Abdizadeh, R. (2013). Detection of *Toxoplasma gondii* in raw caprine, ovine, buffalo, bovine, and camel milk using cell cultivation, cat bioassay, capture ELISA, and PCR methods in Iran. *Foodborne Pathogens and Disease*, 10(2), 120-125.
- Deisenhammer, F., Bartos, A., Egg, R., Gilhus, N., Giovannoni, G., Rauer, S. and Sellebjerg, F. (2006). Guidelines on routine cerebrospinal fluid analysis. Report from an EFNS task force. *European Journal of Neurology*, 13(9), 913-922.
- Del Río, M., Gutiérrez, C. and Ferri, E. R. (2003). Value of indirect hemagglutination and coagglutination tests for serotyping *Haemophilus parasuis*. *Journal of Clinical Microbiology*, 41(2), 880-882.
- Di Napoli, M., Godoy, D. A., Campi, V., Masotti, L., Smith, C. J., Jones, A. R. P., Hopkins, S. J., Slevin, M., Papa, F. and Mogoanta, L. (2012). C-reactive protein in intracerebral hemorrhage Time course, tissue localization, and prognosis. *Neurology*, 79(7), 690-699.
- Dinarello, C. A. (1996). Thermoregulation and the pathogenesis of fever. *Infectious Disease Clinics of North America*, 10(2), 433-449.
- Dinarello, C. A. (2007). Historical insights into cytokines. *European Journal of Immunology*, 37(1), 34-45.

- Divers, T. J. (2004). Acquired spinal cord and peripheral nerve disease. *Veterinary Clinics of North America: Food Animal Practice*, 20(2), 231-242.
- Donkin, J. J. and Vink, R. (2010). Mechanisms of cerebral oedema in traumatic brain injury: therapeutic developments. *Current Opinion in Neurology*, 23(3), 293-299.
- Dow, S. W., Lecouteur, R. A., Henik, R. A., Jones, R. L. and Poss, M. L. (1988). Central nervous system infection associated with anaerobic bacteria in two dogs and two cats. *Journal of Veterinary Internal Medicine*, 2(4), 171-176.
- Dowling, A., Hodgson, J. C., Schock, A., Donachie, W., Eckersall, P. D. and McKendrick, I. J. (2002). Experimental induction of pneumonic pasteurellosis in calves by intratracheal infection with *Pasteurella multocida* biotype A:3. *Research in Veterinary Science*, 73(1), 37-44.
- Dowling, A., Hodgson, J. C., Dagleish, M. P., Eckersall, P. D. and Sales, J. (2004). Pathophysiological and immune cell responses in calves prior to and following lung challenge with formalin-killed *Pasteurella multocida* biotype A:3 and protection studies involving subsequent homologous live challenge. *Veterinary Immunology and Immunopathology*, 100(3-4), 197-207.
- Drevets, D. A. and Leenen, P. J. (2000). Leukocyte-facilitated entry of intracellular pathogens into the central nervous system. *Microbes and Infection*, 2(13), 1609-1618.
- Driver, L., Ayyangar, R. and Van Tubbergen, M. (2015). Language Development in Disorders of Communication and Oral Motor Function. *Pediatric Rehabilitation*, 2015, 53-77
- Dunn, A. J., Swiergiel, A. H. and Beaurepaire, R. d. (2005). Cytokines as mediators of depression: What can we learn from animal studies? *Neuroscience and Biobehavioral Reviews*, 29(4-5), 891-909.
- Dziva, F., Muhairwa, A. P., Bisgaard, M. and Christensen, H. (2008). Diagnostic and typing options for investigating diseases associated with *Pasteurella multocida*. *Veterinary Microbiology*, 128(1-2), 1-22.
- Ebersole, J. L. and Cappelli, D. (2000). Acute-phase reactants in infections and inflammatory diseases. *Periodontology 2000*, 23(1), 19-49.
- Eckersall, P. (2000). Recent advances and future prospects for the use of acute phase proteins as markers of disease in animals. *Revue De Medecine Veterinaire*, 151(7), 577-584.
- Eckersall, P., Young, F., McComb, C., Hogarth, C., Safi, S., Weber, A., McDonald, T., Nolan, A. and Fitzpatrick, J. (2001). Acute phase proteins in serum and milk from dairy cows with clinical mastitis. *The Veterinary Record*, 148(2), 35-41.

- Eckersall, P. and Bell, R. (2010). Acute phase proteins: Biomarkers of infection and inflammation in veterinary medicine. *The Veterinary Journal*, 185(1), 23-27.
- Eckersall, P.D., 2006, December. Measurement of acute phase proteins as biomarkers of disease in production animals. In Proceedings of the 57th Annual Meeting of the American College of Veterinary Pathologists and the 41st Annual Meeting of the *American Society for Veterinary Clinical Pathology 2006*, 2-6.
- Edelman, D. B. and Seth, A. K. (2009). Animal consciousness: a synthetic approach. *Trends in Neurosciences*, 32(9), 476-484.
- Ellis, R. (1967). *Pasteurella septica* infection in respiratory disease. *Thorax*, 22(1), 79-81.
- Eriksen, L., Aalbæk, B., Leifsson, P. S., Basse, A., Christiansen, T., Eriksen, E. and Rimler, R. (1999). Hemorrhagic septicaemia in fallow deer (*Dama dama*) caused by *Pasteurella multocida*. *Journal of Zoo and Wildlife Medicine*, 285-292.
- Fagiolo, A., Roncoroni, C., Lai, O. and Borghese, A. (2005). Chapter XIII buffalo pathologies. *Buffalo Production and Research, 2005*, 249-296.
- Fecteau, G. and George, L. W. (2004). Bacterial meningitis and encephalitis in ruminants. *Veterinary Clinics of North America: Food Animal Practice*, 20(2), 363-377.
- Finnie, J. (2001). Animal models of traumatic brain injury: a review. *Australian Veterinary Journal*, 79(9), 628-633.
- Flores-Tena, F.J., Guerrero-Barrera, A.L., Avelar-González, F.J., Ramírez-López, E.M. and Martínez-Saldaña, M.C., (2007). Pathogenic and opportunistic Gram-negative bacteria in soil, leachate and air in San Nicolás landfill at Aguascalientes, Mexico. *Revista Latinoamericana de Microbiología*, 49(1-2), 25-30.
- Foged, N., Nielsen, J. and Pedersen, K. (1988). Differentiation of toxigenic from nontoxigenic isolates of *Pasteurella multocida* by enzyme-linked immunosorbent assay. *Journal of Clinical Microbiology*, 26(7), 1419-1420.
- Frandsen, R. D., Wilke, W. L. and Fails, A. D. (2009). *Anatomy and physiology of farm animals*. 7th Edition, John Wiley and Sons.
- Fraser, J. A., Biousse, V. and Newman, N. J. (2010). The neuro-ophthalmology of mitochondrial disease. *Survey of Ophthalmology*, 55(4), 299-334.
- Fuller, T. E., Kennedy, M. J. and Lowery, D. E. (2000). Identification of *Pasteurella multocida* virulence genes in a septicemic mouse model using signature-tagged mutagenesis. *Microbial Pathogenesis*, 29(1), 25-38.

- Galdiero, M., A., Cantisani, M., Tarallo, R. Falanga and Galdiero, S. (2012). Septic shock by Gram-negative infections: Role of outer membrane proteins. In R. Fernandez (Ed.), *Severe Sepsis and Septic Shock -Understanding a Serious Killer*: China, InTech Publisher, 1-22.
- Ganheim, C., Alenius, S. and Persson Waller, K. (2007). Acute phase proteins as indicators of calf herd health. *The Veterinary Journal*, 173(3), 645-651.
- Garman, R. H. (2011). Histology of the central nervous system. *Toxicologic Pathology*, 39(1), 22-35.
- Gilbert, T. M. and Blatteis, C. M. (1977). Hypothalamic thermoregulatory pathways in the rat. *Journal of Applied Physiology*, 43(5), 770-777.
- Gillig, P. M. and Sanders, R. D. (2010). Cranial nerves IX, X, XI, and XII. *Psychiatry*, 7(5), (1550-5952).
- Giuliano, F., Rampin, O. and Allard, J. (2002). Neurophysiology and pharmacology of female genital sexual response. *Journal of Sex and Marital Therapy*, 28(1), 101-121.
- Giuliano, F. and Rampin, O. (2004). Neural control of erection. *Physiology and Behavior*, 83(2), 189-201.
- Glorioso, J., Jones, G., Rush, H., Pentler, L., Darif, C. and Coward, J. (1982). Adhesion of type A *Pasteurella multocida* to rabbit pharyngeal cells and its possible role in rabbit respiratory tract infections. *Infection and Immunity*, 35(3), 1103-1109.
- Glushakova, O. Y., Glushakov, A. V., Miller, E. R., Valadka, A. B. and Hayes, R. L. (2016). Biomarkers for acute diagnosis and management of stroke in neurointensive care units. *Brain Circulation*, 2(1), 28-47
- Goldman, E. and Green, L. H. (2008). *Practical Handbook of Microbiology* 2nd Edition, Boca Raton: Taylor and Francis Group.
- Gray, L. D. and Fedorko, D. P. (1992). Laboratory diagnosis of bacterial meningitis. *Clinical Microbiology Reviews*, 5(2), 130-145.
- Graydon, R., Patten, B. and Hamid, H. (1992). The pathology of experimental haemorrhagic septicaemia in cattle and buffalo *Pasteurellosis in production animals*. Canberra, Australia: In Australian Centre for International Agricultural Research (ACIAR) proceedings, Vol. 43, 105-107
- Green, B. T., Ramsay, K. M. and Nolan, P. E. (2002). *Pasteurella multocida* meningitis: case report and review. *Scandinavian Journal of Infectious Diseases*, 34(3), 213-217.

- Gruys, E., Toussaint, M. J. M., Niewold, T. A. and Koopmans, S. J. (2005). Acute phase reaction and acute phase proteins. *Journal of Zhejiang University Science. B*, 6(11), 1045-1056.
- Habek, M., Brinar, M., Brinar, V. V. and Poser, C. M. (2006). Psychiatric manifestations of multiple sclerosis and acute disseminated encephalomyelitis. *Clinical Neurology and Neurosurgery*, 108(3), 290-294.
- Harper, M., Boyce, J. D. and Adler, B. (2006). *Pasteurella multocida* pathogenesis: 125 years after Pasteur. *FEMS Microbiology Letters*, 265(1), 1-10.
- Harper, M., Cox, A. D., Adler, B. and Boyce, J. D. (2011). *Pasteurella multocida* lipopolysaccharide: The long and the short of it. *Veterinary Microbiology*, 153(1-2), 109-115.
- Harper, M., Boyce, J. and Adler, B. (2012). The Key Surface Components of *Pasteurella multocida*: Capsule and Lipopolysaccharide. In K. Aktories, J. H. C. Orth and B. Adler (Eds.), *Pasteurella multocida*. Springer Berlin Heidelberg, 361, 39-51.
- Hashimoto, T., Yonetani, M. and Nakamura, H. (2004). Selective brain hypothermia protects against hypoxic-ischemic injury in newborn rats by reducing hydroxyl radical production. *Kobe Journal of Medical Sciences*, 49(3/4), 83-92.
- Hatfaludi, T., Al-Hasani, K., Boyce, J. D. and Adler, B. (2010). Outer membrane proteins of *Pasteurella multocida*. *Veterinary Microbiology*, 144(1-2), 1-17.
- Heddleston, K. L., Gallagher, J. E. and Rebers, P. A. (1972). Fowl cholera: Gel diffusion precipitin test for serotyping *Pasteurella multocida* from avian species. *Avian Diseases*, 16, 925-936.
- Heegaard, P. M., Klausen, J., Nielsen, J. P., González-Ramón, N., Piñeiro, M., Lampreave, F. and Alava, M. A. (1998). The porcine acute phase response to infection with *Actinobacillus pleuropneumoniae*. Haptoglobin, C-reactive protein, major acute phase protein and serum amyloid A protein are sensitive indicators of infection. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*, 119(2), 365-373.
- Herpers, B. L., Endeman, H., De Jong, B. A., De Jongh, B. M., Grutters, J. C., Biesma, D. H. and Velzen-Blad, V. (2009). Acute-phase responsiveness of mannose-binding lectin in community-acquired pneumonia is highly dependent upon MBL2 genotypes. *Clinical and Experimental Immunology*, 156(3), 488-494.
- Hinchcliff, K. W. (1995). Cerebrospinal fluid analysis in the diagnosis of neurological disease in large animals. *British Veterinary Journal*, 151(6), 599-602.

- Hirsh, D. C., Jessup, D. A., Snipes, K. P., Carpenter, T. E., Hird, D. W. and McCapes, R. H. (1990). Characteristics of *Pasteurella multocida* isolated from waterfowl and associated avian species in California. *Journal of Wildlife Diseases*, 26(2), 204-209.
- Hodgson, J. C., Finucane, A., Dagleish, M. P., Ataei, S., Parton, R. and Coote, J. G. (2005). Efficacy of vaccination of calves against hemorrhagic septicaemia with a live *aroA* derivative of *Pasteurella multocida* B:2 by two different routes of administration. *Infection and Immunity*, 73(3), 1475-1481.
- Hodgson, J. C. (2006). Endotoxin and Mammalian Host Responses During Experimental Disease. *Journal of Comparative Pathology*, 135(4), 157-175.
- Hodgson, J. C., Dagleish, M. P., Gibbard, L., Bayne, C. W., Finlayson, J., Moon, G. M. and Nath, M. (2013). Seven strains of mice as potential models of bovine pasteurellosis following intranasal challenge with a bovine pneumonic strain of *Pasteurella multocida* A:3; comparisons of disease and pathological outcomes. *Research in Veterinary Science*, 94(3), 634-640.
- Horadagoda, N., Knox, K., Gibbs, H., Reid, S., Horadagoda, A., Edwards, S. and Eckersall, P. (1999). Acute phase proteins in cattle: discrimination between acute and chronic inflammation. *The Veterinary Record*, 144(16), 437-441.
- Horadagoda, N., Hodgson, J., Moon, G., Wijewardana, T. and Eckersall, P. (2001). Role of endotoxin in the pathogenesis of haemorrhagic septicaemia in the buffalo. *Microbial Pathogenesis*, 30(3), 171-178.
- Horadagoda, N., Hodgson, J., Moon, G., Wijewardana, T. and Eckersall, P. (2002). Development of a clinical syndrome resembling haemorrhagic septicaemia in the buffalo following intravenous inoculation of *Pasteurella multocida* serotype B: 2 endotoxin and the role of tumour necrosis factor- α . *Research in Veterinary Science*, 72(3), 194-200.
- Howard, R., Brammer, M., David, A., Woodruff, P. and Williams, S. (1998). The anatomy of conscious vision: an fMRI study of visual hallucinations. *Nature Neuroscience*, 1(8), 738-742.
- Huang, W.-T., Tsai, S.-M. and Lin, M.-T. (2001). Involvement of brain glutamate release in pyrogenic fever. *Neuropharmacology*, 41(7), 811-818.
- Hunt, M. L., Adler, B. and Townsend, K. M. (2000). The molecular biology of *Pasteurella multocida*. *Veterinary Microbiology*, 72(1-2), 3-25.
- Hunt, M. L., Boucher, D. J., Boyce, J. D. and Adler, B. (2001). In vivo-expressed genes of *Pasteurella multocida*. *Infection and Immunity*, 69(5), 3004-3012.
- Hussaini, J. and Jumahat, N.M., (2014). Characterization of recombinant protein of *Pasteurella multocida* serotype B. *Life Science Journal*, 11(12) .13-19.

- Iatrou, C., Domaigne, C., Thomas, R. and Nye, D. (2002). The effect of selective brain cooling on intracerebral temperature during craniotomy. *Anaesthesia and Intensive Care*, 30(2), 167.
- Idoate, I., Vander Ley, B., Schultz, L. and Heller, M. (2015). Acute phase proteins in naturally occurring respiratory disease of feedlot cattle. *Veterinary Immunology and Immunopathology*, 163(3-4), 221-226.
- Iovane, G., Pagnini, P., Galdiero, M., Cipollaro de l'Ero, G., Vitiello, M., D'Isanto, M. and Marcatili, A. (1998). Role of *Pasteurella multocida* porin on cytokine expression and release by murine splenocytes. *Veterinary Immunology and Immunopathology*, 66(3), 391-404.
- Isenberg, J. S., Roberts, D. D. and Frazier, W. A. (2012). Prevention of tissue ischemia, related methods and compositions, U.S. Google Patents: 8,236,313.
- Jackson, P. G., Cockcroft, P. D. and Elmhurst, S. (2002). *Clinical examination of farm animals*: Oxford, UK: Blackwell Science Ltd, 141-124
- Jacobsen, S., Andersen, P., Toelboell, T. and Heegaard, P. M. (2004). Dose dependency and individual variability of the lipopolysaccharide-induced bovine acute phase protein response. *Journal of Dairy Science*, 87(10), 3330-3339.
- Jankovic, J. and Patel, S. C. (1983). Blepharospasm associated with brainstem lesions. *Neurology*, 33(9), 1237-1237.
- Jawetz, E. (1950). A pneumotropic *Pasteurella* of laboratory animals. Bacteriological and serological characteristics of the organism. *Journal of Infectious Diseases*, 86(2), 172-183.
- Jennifer, C. D. M., Baba. (2000). Evaluation and treatment of swallowing impairments. *American Family Physician*, 61(8), 2453-2462.
- Jergens, A. E., Schreiner, C. A., Frank, D. E., Niyo, Y., Ahrens, F. E., Eckersall, P. D., Benson, T. J., and Evans, R. (2003). A scoring index for disease activity in canine inflammatory bowel disease. *Journal of Veterinary Internal Medicine*, 17, 291-297.
- Jesse, F., Adamu, L., Abdinasir, Y., Zakaria, Z. and Abdullah, R. (2013a). Acute phase protein profile in calves following infection with whole cell, lipopolysaccharide and outer membrane protein extracted from *Pasteurella multocida* type B: 2. *Asian Journal of Animal and Veterinary Advances*, 8(4), 655-662.
- Jesse, F. F., Affandi, S. A., Osman, A. Y., Adamu, L., Saad, M. Z., Haron, A. W., Omar, A. R., Sabri, J. and Saharee, A. A. (2013b). Clinico-pathological features in mice following oral exposure to *Pasteurella multocida* B: 2. *IOSR Journal of Agriculture and Veterinary Science*, 3(4), 35-39.

- Jesse, F. F. A., Khaleel, M. M., Adamu, L., Osman, A. Y., Haron, A. W., Saad, M. Z. and Omar, A. R. (2013c). Polymerase chain reaction detection of *Pasteurella multocida* type B:2 in mice infected with contaminated river water. *American Journal of Animal and Veterinary Sciences*, 8(3), 146-151.
- Jesse, F. F. A., Lawan, A., Abdinasir, Y. O., Zakaria, Z., Abdullah, R., Zamri, M. S. and Saharee, A. A. (2013d). Clinico-pathological responses of calves associated with infection of *Pasteurella multocida* type B and the bacterial lipopolysaccharide and outer membrane protein immunogens. *International Journal of Animal and Veterinary Advances*, 5(5), 190-198.
- Jesse, F. F. A., Adamu, L., Hazirah, N., Osman, A. Y., Mansor, R., Haron, A. W., Saad, M. Z., Omar, A. R. and Saharee, A. A. (2013e). Clinical and reproductive pathological changes associated with *Brucella melitensis* and its lipopolysaccharides in female mice via oral inoculation. *American Journal of Animal and Veterinary Sciences*, 8(3), 104-111.
- Jesse, F. F. A., Abdinasir, Y. O., Lawan, A., Zakaria, Z., Abdullah, R., Zamri, M. S. and Saharee, A. A. (2013f). Haematological and biochemical alterations in calves following infection with *Pasteurella multocida* type B: 2, bacterial lipopolysaccharide and outer membrane protein (OMP) immunogens. *Asian Journal of Animal and Veterinary Advances*, 8(6), 806-813.
- Jesse, F. F. A., Osman, A. Y., Adamu, L., Yusof, M., Syamil, M., Omar, A. R., Saharee, A. A., Haron, A. W. and Abdullah, R. (2013g). Polymerase chain reaction detection of *Pasteurella multocida* type B: 2 in mice following oral inoculation. *Asian Journal of Animal and Veterinary Advances*, 8(3), 493-501.
- Jesse, F. F. A., Adamu, L., Tijjani, A., Mohammed, K., Abba, Y., Sadiq, M. A., Zainal, R., Sabu, M. J., Saharee, A. A. and Haron, A. W. (2014). Hormonal and histopathological alterations in pituitary glands and reproductive organs of male and female mice orally inoculated with *Pasteurella multocida* type B: 2 and its lipopolysaccharides. *American Journal of Animal and Veterinary Sciences*, 9, 200-212.
- Johnston, L., Simmons, G. and McGavin, M. (1962). A viral meningo-encephalitis in calves. *Australian Veterinary Journal*, 38(4), 207-215.
- Jones, M. L. and Allison, R. W. (2007). Evaluation of the ruminant complete blood cell count. *Veterinary Clinics of North America: Food Animal Practice*, 23(3), 377-402.
- Joseph, P. (1979). Haemorrhagic septicaemia in Peninsular Malaysia. *Kajian Veterinar*, 11, 65-79.
- Juma, W. M., Lira, A., Marzuk, A., Marzuk, Z., Hakim, A. M. and Thompson, C. S. (2011). C-reactive protein expression in a rodent model of chronic cerebral hyperperfusion. *Brain Research*, 1414, 85-93.

- Katoch, S., Rana, V., Shukla, P. and Wann, A. (2015). *Pasteurella multocida* and its virulence determinants. *North-East Veterinarian*, 14(4), 35-36.
- Kaushik, S., Tan, A. G., Mitchell, P. and Wang, J. J. (2007). Prevalence and associations of enhanced retinal arteriolar light reflex: a new look at an old sign. *Ophthalmology*, 114(1), 113-120.
- Kaya, S., Kacar, C., ÖĞÜN, M., and Kuru, M. (2016). Evaluation of serum c-reactive protein and natural antibodies in cows with endometritis. *Kafkas Üniversitesi Veteriner Fakültesi Dergisi*, 22(5), 709-715.
- Kennedy, P. C., Biberstein, E. L., Howarth, J., Frazier, L. M. and Dungworth, D. (1960). Infectious meningo-encephalitis in cattle, caused by a haemophilus-like organism. *American Journal of Veterinary Research*, 21, 403-409.
- Kessell, A., Finnie, J. and Windsor, P. (2011). Neurological diseases of ruminant livestock in Australia. III: bacterial and protozoal infections. *Australian Veterinary Journal*, 89(8), 289-296.
- Khaleel, M. M., Abdullah, F. F. J., Adamu, L., Osman, A. Y., Haron, A. W., Saad, M. Z. and Omar, A. R. (2013). Acute phase protein responses in mice infected with river water contaminated by *Pasteurella multocida* type B: 2. *American Journal of Animal and Veterinary Sciences*, 8(3), 159- 164.
- Khaleel, M. M., Abdullah, F. F. J., Adamu, L., Abba, Y., Haron, A. W., Saad, M. Z. and Omar, A. R. (2014). Histopathological changes in mice infected with river water contaminated by *Pasteurella multocida* type B:2. *American Journal of Animal and Veterinary Sciences*, 9(2), 71-76.
- Khan, A., Saleemi, M. K., Khan, M. Z., Gul, S. T., Irfan, M. and Qamar, M. S. (2011). Hemorrhagic septicaemia in buffalo (*Bubalus bubalis*) calves under sub-tropical conditions in Pakistan. *Pakistan Journal of Zoology*, 43, 295-302.
- Kharb, S. and Charan, S. (2011). Mucosal immunization provides better protection than subcutaneous immunization against *Pasteurella multocida* (B:2) in mice preimmunized with the outer membrane proteins. *Veterinary Research Communications*, 35(7), 457- 461.
- Kharb, S. and Charan, S. (2013). Mouse model of haemorrhagic septicaemia: dissemination and multiplication of *Pasteurella multocida* B:2 in vital organs after intranasal and subcutaneous challenge in mice. *Veterinary Research Communications*, 37(1), 59-63.
- Khattari, S., Kinjavdekar, P., Amarpal, H. P. A., Pawde, A. M., Kumar, R. and Singh, J. (2013). Dexmedetomidine with butorphanol and propofol for total intravenous anesthesia in uraemic buffalo calves. *Advances in Animal and Veterinary Sciences*, 1, 15-23.

- Khin, M. N., Zamri-Saad, M. and Noordin, M. (2010). Pathological changes in the lungs of calves following intratracheal exposure to *Pasteurella multocida* B: 2. *Pertanika Journal of Tropical Agricultural Science*, 33(1), 113-117.
- Kim, K. S. (2008). Mechanisms of microbial traversal of the blood–brain barrier. *Nature Reviews Microbiology*, 6(8), 625-634.
- Kiš, I., Foršek, J., Matijatko, V., Kučer, N. and Šimonji, K. (2008). Steroid-responsive meningitis-arteritis in a dog—a case report. *Veterinarski Arhiv*, 78(6), 529-538.
- Kitamura, N., Yamada, J. and Yamashita, T. (1986). Immunohistochemical study on the distribution of neuron-specific enolase-and peptide-containing nerves in the reticulorumen and the reticular groove of cattle. *Journal of Comparative Neurology*, 248, 223-234.
- Konno, S., Moriwaki, M. and Nakagawa, M. (1982). Akabane disease in cattle: congenital abnormalities caused by viral infection. Spontaneous disease. *Veterinary Pathology Online*, 19(3), 246-266.
- Konold, T., Bone, G., Ryder, S., Hawkins, S., Courtin, F. and Berthelin-Baker, C. (2004). Clinical findings in 78 suspected cases of bovine spongiform encephalopathy in Great Britain. *Veterinary Record: Journal of the British Veterinary Association*, 155(21), 659-666
- Konuk, N., Tekin, I., Ozturk, U., Atik, L., Atasoy, N., Bektas, S. and Erdogan, A. (2007). Plasma levels of tumor necrosis factor-alpha and interleukin-6 in obsessive compulsive disorder. *Mediators of Inflammation*, 2007.
- Kpodékon, M. (1982). Pathology and pathogenesis of ear and brain complications of pasteurellosis in rabbits bred for food. *Annales de Recherches Veterinaires. Annals of Veterinary Research*, 14(3), 225-232.
- Kubatzky, K. (2012). *Pasteurella multocida* and Immune Cells. In K. Aktories, J. H. C. Orth and B. Adler (Eds.), *Pasteurella multocida*. Springer Berlin Heidelberg. 361, 53-72.
- Kumar, V., Singh, S., Kumar, A. and Peshin, P. (2014). Evaluation of propofol-halothane anaesthesia in buffalo calves (*Bubalus bubalis*). *Indian Journal of Veterinary Surgery*, 35(1), 53-54.
- Kupfer, D. J. (1991). Hypersomnia in bipolar depression: a comparison with narcolepsy using the multiple sleep latency test. *The American Journal of Psychiatry*, 148(9), 1177-1181
- Kurćubić, V. S., Đoković, R. D., Ilić, Z. Ž., Stojković, J. S., Petrović, M. P. and Caro-Petrović, V. (2014). Modern approach to the enigma of bovine respiratory disease complex: a review. *Pakistan Veterinary Journal*, 34, 11-17.

- Laemmli, U. K. (1970). Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature*, 227(5259), 680-685.
- Lane, E. P., Kock, N. D., Hill, F. W. G. and Mohan, K. (1992). An outbreak of haemorrhagic septicaemia (septicaemic pasteurellosis) in cattle in Zimbabwe. *Tropical Animal Health and Production*, 24(2), 97-102.
- Launay, F., Young, P., Sterk, S., Blokland, M. and Kennedy, D. (2004). Confirmatory assay for zeranol, taleranol and the *Fusarium* spp. toxins in bovine urine using liquid chromatography-tandem mass spectrometry. *Food Additives and Contaminants*, 21(1), 52-62.
- Leake, J. A., Albani, S., Kao, A. S., Senac, M. O., Billman, G. F., Nespeca, M. P., Paulino, A. D., Quintela, E. R., Sawyer, M. H. and Bradley, J. S. (2004). Acute disseminated encephalomyelitis in childhood: epidemiologic, clinical and laboratory features. *The Pediatric Infectious Disease Journal*, 23(8), 756-764.
- Lee, L., Harkness, K. L., Sabbagh, M. A. and Jacobson, J. A. (2005). Mental state decoding abilities in clinical depression. *Journal of Affective Disorders*, 86(2), 247-258.
- Lee, M.-Y., Kim, S. Y., Choi, J.-S., Lee, I.-H., Choi, Y.-S., Jin, J. Y., Park, S.-J., Sung, K.-W., Chun, M.-H. and Kim, I.-S. (2002). Upregulation of haptoglobin in reactive astrocytes after transient forebrain ischemia in rats. *Journal of Cerebral Blood Flow and Metabolism*, 22(10), 1176-1180.
- Lewis, R., Behnke, J. M., Cassidy, J., Stafford, P., Murray, N. and Holland, C. V. (2007). The migration of *Ascaris suum* larvae, and the associated pulmonary inflammatory response in susceptible C57BL/6j and resistant CBA/Ca mice. *Parasitology*, 134(09), 1301-1314.
- Liao, C.-M., Huang, C., Hsuan, S.-L., Chen, Z.-W., Lee, W.-C., Liu, C.-I., Winton, J. R. and Chien, M.-S. (2006). Immunogenicity and efficacy of three recombinant subunit *Pasteurella multocida* toxin vaccines against progressive atrophic rhinitis in pigs. *Vaccine*, 24(1), 27-35.
- Licinio, J. and Wong, M. (1999). The role of inflammatory mediators in the biology of major depression: central nervous system cytokines modulate the biological substrate of depressive symptoms, regulate stress-responsive systems, and contribute to neurotoxicity and neuroprotection. *Molecular Psychiatry*, 4(4), 317-327.
- Lillie, L. (1974). The bovine respiratory disease complex. *The Canadian Veterinary Journal*, 15(9), 233.
- LimTeik, C., Faez Firdaus, J., Adamu, L., Marza, A. D., Ibrahim, H. H., Zamri-Saad, M., Haron, A. W., Saharee, A. A., Mohd Azmi, M. and Omar, A. R. (2015). Clinico-pathology, hematology, and biochemistry responses toward *Pasteurella multocida* Type B: 2 via oral and subcutaneous route of infections. *Veterinary World*, 8(6), 783-792.

- Lin, H., Bhatia, R. and Lal, R. (2001). Amyloid β protein forms ion channels: implications for Alzheimer's disease pathophysiology. *The Federation of American Societies for Experimental Biology Journal*, 15(13), 2433-2444.
- Lisanby, S. H., Schlaepfer, T. E., Fisch, H.-U. and Sackeim, H. A. (2001). Magnetic seizure therapy of major depression. *Archives of General Psychiatry*, 58(3), 303-305.
- Lorenz, M. D., Coates, J. and Kent, M. (2010). *Handbook of veterinary neurology*: 5th Edition; Elsevier/Saunders: St. Louis, MI, USA, 2011; 28.
- Loretti, A. P., Colodel, E. M., Driemeier, D., Corrêa, A. M., Bangel, J. J. and Ferreiro, L. (2003). Neurological disorder in dairy cattle associated with consumption of beer residues contaminated with *Aspergillus clavatus*. *Journal of Veterinary Diagnostic Investigation*, 15(2), 123-132.
- Luzina, I. G., Todd, N. W., Sundararajan, S. and Atamas, S. P. (2015). The cytokines of pulmonary fibrosis: Much learned, much more to learn. *Cytokine*, 74(1), 88-100.
- Mackintosh, C., Haigh, J. and Griffin, F. (2002). Bacterial diseases of farmed deer and bison. *Revue Scientifique et Technique-Office International Des Epizooties*, 21(1), 249-264.
- Malik, V., Kinjavdekar, P., Aithal, H. and Pawde, A. (2011). Comparative evaluation of halothane anaesthesia in medetomidine-butorphanol and midazolam-butorphanol premedicated water buffaloes (*Bubalus bubalis*). *Journal of the South African Veterinary Association*, 82(1), 8-17.
- Marandi, M. V., Dubreuil, J. and Mittal, K. (1996). The 32 kDa major outer-membrane protein of *Pasteurella multocida* capsular serotype D. *Microbiology*, 142(1), 199-206.
- Marandi, M. V. and Mittal, K. (1997). Role of outer membrane protein H (OmpH)- and OmpA-specific monoclonal antibodies from hybridoma tumors in protection of mice against *Pasteurella multocida*. *Infection and Immunity*, 65(11), 4502-4508.
- Marques, F., Palha, J. A., Sousa, J. C., Correia-Neves, M. and Sousa, N. (2011). Brain Barriers and the Acute-Phase Response, *China, InTech Publisher*, 137-152.
- Marshall, M., Robison, R. and Jensen, M. (1981). Use of an enzyme-linked immunosorbent assay to measure antibody responses in turkeys against *Pasteurella multocida*. *Avian Diseases*, 25(4), 964-971.
- Martin, R. E. and Sessle, B. J. (1993). The role of the cerebral cortex in swallowing. *Dysphagia*, 8(3), 195-202.
- Marza, A. D., Jesse, F. F. A., Ahmed, I. M., Chung, E. L., Ibrahim, H. H., Zamri Saad, M., Omar, A. R., Abu Bakar, M. Z., Saharee, A. A., Haron, A. W. and

- Mohd Lila, M. A. (2015). Involvement of nervous system in cattle and buffaloes due to *Pasteurella multocida* B:2 infection: A review of clinicopathological and pathophysiological changes. *Journal of Advanced Veterinary and Animal Research*, 2(3), 252-262.
- Marza, A. D., Jesse, F. F. A., Ahmed, I. M., Teik Chung, E. L., Ibrahim, H. H., Zamri-Saad, M., Omar, A. R., Abu Bakar, M. Z., Saharee, A. A., Haron, A. W., Alwan, M. J. and Mohd Lila, M. A. (2016). Involvement of the nervous system following experimental infection with *Pasteurella multocida* B:2 in buffalo (*Bubalus bubalis*): A clinicopathological study. *Microbial Pathogenesis*, 93, 111-119.
- Masujin, K., Matthews, D., Wells, G. A., Mohri, S. and Yokoyama, T. (2007). Prions in the peripheral nerves of bovine spongiform encephalopathy-affected cattle. *Journal of General Virology*, 88(6), 1850-1858.
- Matre, R. and Vedeler, C. A. (1987). Demonstration of human erythrocyte C3b receptors (CR1) by haemadsorption and indirect haemagglutination techniques. *Journal of Immunological Methods*, 96(1), 139-144.
- McEwen, S. A. and Fedorka-Cray, P. J. (2002). Antimicrobial use and resistance in animals. *Clinical Infectious Diseases*, 34(3), 93-106.
- McFadden, A., Christensen, H., Fairley, R., Hill, F., Gill, J., Keeling, S. and Spence, R. (2011). Outbreaks of pleuritis and peritonitis in calves associated with *Pasteurella multocida* capsular type B strain. *New Zealand Veterinary Journal*, 59(1), 40-45.
- McGrotty, Y. L., Arteaga, A., Knottenbelt, C. M., Ramsey, I. K., and Eckersall, P. D. (2005). Haptoglobin concentrations in dogs undergoing trilostane treatment for hyperadrenocorticism. *Veterinary Clinical Pathology*, 34, 255-258.
- Melnikow, E., Schoenfeld, C., Spehr, V., Warrass, R., Gunkel, N., Duszenko, M., Selzer, P. and Ullrich, H. (2008). A compendium of antibiotic-induced transcription profiles reveals broad regulation of *Pasteurella multocida* virulence genes. *Veterinary Microbiology*, 131(3), 277-292.
- Merritt, S. L., Cohen, F. L. and Smith, K. M. (1992). Depressive symptomatology in narcolepsy. *Loss, Grief and Care*, 5(3-4), 53-59.
- Mitra, J., Chowdhury, M. and Bhattacharya, C. (2013). Outbreak of hemorrhagic septicaemia in free range buffalo and cattle grazing at riverside grassland in Murshidabad District, West Bengal, India. *Exploratory Animal and Medical Research*, 3(2), 178-182.
- Mittal, R., Krishnan, S., Gonzalez-Gomez, I. and Prasadarao, N. V. (2011). Deciphering the roles of outer membrane protein A extracellular loops in the pathogenesis of *Escherichia coli* K1 meningitis. *Journal of Biological Chemistry*, 286(3), 2183-2193.

- Mobasheri, A. and Cassidy, J. P. (2010). Biomarkers in veterinary medicine: towards targeted, individualised therapies for companion animals. *The Veterinary Journal*, 185(1), 1-3.
- Moolchand, M., Kachiwal, A., Soomro, S. and Bhutto, Z. (2014). Comparison of sedative and analgesic effects of xylazine, detomidine, and medetomidine in sheep. *Egyptian Journal of Sheep and Goat Sciences*, 9(2), 43-48.
- Morrish, R. B. (2001). Method of suppression and prevention of the gag reflex: Google Patents: 6,192,889.
- Murata, H. (2007). Stress and acute phase protein response: an inconspicuous but essential linkage. *Veterinary Journal*, 173(3), 473-474.
- Mustafa, M. M., Lebel, M. H., Ramilo, O., Olsen, K. D., Reisch, J. S., Beutler, B., and McCracken, G. H. (1989). Correlation of interleukin-1 β and cachectin concentrations in cerebrospinal fluid and outcome from bacterial meningitis. *The Journal of Pediatrics*, 115(2), 208-213.
- Nakagawa-Tosa, N., Morimatsu, M., Kawasaki, M., Nakatsuji, H., Syuto, B. and Saito, M. (1995). Stimulation of haptoglobin synthesis by interleukin-6 and tumor necrosis factor, but not by interleukin-1, in bovine primary cultured hepatocytes. *The Journal of Veterinary Medical Science/the Japanese Society of Veterinary Science*, 57(2), 219-223.
- Namioka, M. and Murata, S. (1961). Serological studies on *Pasteurella multocida*. II: Characteristics of the somatic 'O' antigen of the organism. *Cornell Veterinarian*, 51(507-521).
- Nemzek, J. A., Hugunin, K. and Opp, M. R. (2008). Modeling sepsis in the laboratory: merging sound science with animal well-being. *Comparative Medicine*, 58(2), 120-128.
- Nudelman, Y. and Tunkel, A. R. (2009). Bacterial meningitis. *Drugs*, 69(18), 2577-2596.
- Odugbo, M., Turaki, U., Itodo, A., Okwori, A. and Yakubu, R. (2005). Experimental hemorrhagic septicaemia of calves with *Pasteurella multocida* Serotype E: 2: clinical, pathologic and microbiologic studies. *Revue d'Elevage et de Medecine Veterinaire des Pays Tropicaux*, 58(3), 133-137
- Ohga, S., Aoki, T., Okada, K., Akeda, H., Fujioka, K., Ohshima, A., ... and Ueda, K. (1994). Cerebrospinal fluid concentrations of interleukin-1 beta, tumour necrosis factor-alpha, and interferon gamma in bacterial meningitis. *Archives of Disease in Childhood*, 70(2), 123-125.
- Okamoto, K., Tashiro, A., Chang, Z. and Bereiter, D. A. (2010). Bright light activates a trigeminal nociceptive pathway. *Pain*, 149(2), 235-242.

- Okay, S., Özcengiz, E., Gürsel, İ. and Özcengiz, G. (2012). Immunogenicity and protective efficacy of the recombinant *Pasteurella* lipoprotein E and outer membrane protein H from *Pasteurella multocida* A:3 in mice. *Research in Veterinary Science*, 93(3), 1261-1265.
- Othman, S., Parton, R. and Coote, J. (2012). Interaction between mammalian cells and *Pasteurella multocida* B:2. Adherence, invasion and intracellular survival. *Microbial Pathogenesis*, 52(6), 353-358.
- Otomaru, K., Kubota, S. and Tokimori, M. (2015). Maternally and naturally acquired antibodies to *Pasteurella multocida* in Japanese Black calves. *Pakistan Veterinary Journal*, 35(1), 108-110.
- Pal, P., Calne, D., Calne, S. and Tsui, J. (2000). Botulinum toxin A as treatment for drooling saliva in PD. *Neurology*, 54(1), 244-244.
- Pan, W., Zadina, J. E., Harlan, R. E., Tweber, J., Banks, W. A. and Kastin, A. J. (1997). Tumor necrosis factor- α : a neuromodulator in the CNS. *Neuroscience and Biobehavioral Reviews*, 21(5), 603-613.
- Parker, G., Hadzi-Pavlovic, D., Boyce, P., Wilhelm, K., Brodaty, H., Mitchell, P., Hickie, I. and Eyers, K. (1990). Classifying depression by mental state signs. *The British Journal of Psychiatry*, 157(1), 55-65.
- Patel, S., Joshi, D., Raval, S., Patel, B., Patel, J., Chauhan, H., Shah, N. (2016). Clinicopathological studies of *Pasteurella multocida* B: 2 experimental infection in rabbits. *The Indian Journal of Animal Sciences*, 86(4), 380-386
- Pavan, B., Dalpiaz, A., Ciliberti, N., Biondi, C., Manfredini, S. and Vertuani, S. (2008). Progress in drug delivery to the central nervous system by the prodrug approach. *Molecules*, 13(5), 1035-1065.
- Pawde, A., Kinjavdekar, P., Aithal, H., Pratap, K. and Bisht, G. (2000). Detomidine-Diazepam-Ketamine Anaesthesia in Buffalo (*Bubalus bubalis*) Calves. *Journal of Veterinary Medicine Series A*, 47(3), 175-179.
- Petzold, A., Brettschneider, J., Jin, K., Keir, G., Murray, N., Hirsch, N., Itoyama, Y., Reilly, M., Takeda, A. and Tumani, H. (2009). CSF protein biomarkers for proximal axonal damage improve prognostic accuracy in the acute phase of Guillain-Barré syndrome. *Muscle and Nerve*, 40(1), 42-49.
- Pfannkuche, H., Reiche, D., Hoppe, S. and Schemann, M. (2002). Cholinergic and noncholinergic innervation of the smooth muscle layers in the bovine abomasum. *Anatomy Record*, 267, 70-77.
- Plessers, E., Wyns, H., Watteyn, A., Pardon, B., De Backer, P. and Croubels, S. (2015). Characterization of an intravenous lipopolysaccharide inflammation model in calves with respect to the acute-phase response. *Veterinary Immunology and Immunopathology*, 163(1-2), 46-56.

- Podnar, S. (2004). Bilateral vs. unilateral electromyographic examination of the external anal sphincter muscle. *Clinical Neurophysiology*, 34(3), 153-157.
- Polizopoulou, Z. (2010). Haematological tests in sheep health management. *Small Ruminant Research*, 92(1), 88-91.
- Popoff, M. (2011). Multifaceted interactions of bacterial toxins with the gastrointestinal mucosa. *Future Microbiology*, 6(7), 763-797.
- Post, A. and Hoshizaki, T. B. (2012). Mechanisms of brain impact injuries and their prediction: a review. *Trauma*, 14(4), 327-349.
- Poulton, P. (2013). Musculo-skeletal examination and diagnosis of the downer cow. *Cattle Practice*, 21, 174-180.
- Praveena, P. E., Periasamy, S., Kumar, A. A. and Singh, N. (2010). Cytokine profiles, apoptosis and pathology of experimental *Pasteurella multocida* serotype A1 infection in mice. *Research in Veterinary Science*, 89(3), 332-339.
- Prince, G. H. and Smith, J. (1966). Antigenic studies on *Pasteurella multocida* using immunodiffusion techniques: I. Identification and nomenclature of the soluble antigens of a bovine haemorrhagic septicaemia strain. *Journal of Comparative Pathology*, 76(3), 321-314.
- Pulzova, L., Bhide, M. R. and Andrej, K. (2009). Pathogen translocation across the blood-brain barrier. *FEMS Immunology and Medical Microbiology*, 57(3), 203-213.
- Radaelli, S. T. and Platt, S. R. (2002). Bacterial Meningoencephalomyelitis in Dogs: A Retrospective Study of 23 Cases (1990-1999). *Journal of Veterinary Internal Medicine*, 16(2), 159-163.
- Radostits, O. M., Gay, C. C., Hinchcliff, K. W. and Constable, P. D. (2006). *Veterinary medicine: a textbook of the diseases of cattle, horses, sheep, pigs and goats*: 10th Edition. Saunders Ltd, Philadelphia, USA
- Raetz, C. R. and Whitfield, C. (2002). Lipopolysaccharide endotoxins. *Annual Review of Biochemistry*, 71(1), 635- 700.
- Rafidah, O., and Zamri-Saad, M. (2013). Effect of dexamethasone on protective efficacy of live gdhA derivative *Pasteurella multocida* B: 2 vaccine. *Asian Journal of Animal and Veterinary Advances*, 8(3), 1-7
- Raison, C. L., Borisov, A. S., Majer, M., Drake, D. F., Pagnoni, G., Woolwine, B. J., Vogt, G. J., Massung, B. and Miller, A. H. (2009). Activation of central nervous system inflammatory pathways by interferon-alpha: relationship to monoamines and depression. *Biological Psychiatry*, 65(4), 296-303.

- Raivich, G., Bohatschek, M., Kloss, C. U., Werner, A., Jones, L. L. and Kreutzberg, G. W. (1999). Neuroglial activation repertoire in the injured brain: graded response, molecular mechanisms and cues to physiological function. *Brain Research Reviews*, 30(1), 77-105.
- Ranjan, R., Panda, S., Acharya, A., Singh, A. and Gupta, M. (2011). Molecular diagnosis of haemorrhagic septicaemia-A review. *Veterinary World*, 4(4), 189-192.
- Rhoades, K. and Rimler, R. (1987). Capsular groups of *Pasteurella multocida* isolated from avian hosts. *Avian Diseases*, 895-898.
- Rhoades, K. R., Heddleston, K. L. and Rebers, P. A. (1967). Experimental hemorrhagic septicaemia: gross and microscopic lesions resulting from acute infections and from endotoxin administration. *Canadian Journal of Comparative Medicine and Veterinary Science*, 31(9), 226.
- Rimler, R. (1992). Serology and virulence of haemorrhagic septicaemia *Pasteurella multocida* isolated from domestic and feral ruminants. *Pasteurellosis in Production Animals*, In Australian Centre for International Agricultural Research (ACIAR) proceedings, Vol. 43, 44-46.
- Rimler, R. B. (2000). Restriction endonuclease analysis using HhaI and HpaII to discriminate among group B *Pasteurella multocida* associated with haemorrhagic septicaemia. *Journal of Medical Microbiology*, 49(1), 81-87.
- Ritchie, R. F., Palomaki, G. E., Neveux, L. M., Navolotskaia, O., Ledue, T. B. and Craig, W. Y. (1999). Reference distributions for the negative acute-phase serum proteins, albumin, transferrin and transthyretin: a practical, simple and clinically relevant approach in a large cohort. *Journal of Clinical Laboratory Analysis*, 13(6), 273-279.
- Rivera, L. R., Thacker, M. and Furness, J. B. (2009). High-and medium-molecular-weight neurofilament proteins define specific neuron types in the guinea-pig enteric nervous system. *Cell and Tissue Research*, 335(3), 529-538.
- Roier, S., Fenninger, J. C., Leitner, D. R., Rechberger, G. N., Reidl, J. and Schild, S. (2013). Immunogenicity of *Pasteurella multocida* and *Mannheimia haemolytica* outer membrane vesicles. *International Journal of Medical Microbiology*, 303(5), 247-256.
- Rollin, B. E. (2005). On understanding animal mentation. *Mental Health and Well-Being in Animals*, 2005, 3-14.
- Rommel, O., Malin, J.-P., Zenz, M. and Jänig, W. (2001). Quantitative sensory testing, neurophysiological and psychological examination in patients with complex regional pain syndrome and hemisensory deficits. *Pain*, 93(3), 279-293.

- Rose, W. and Rac, R. (1957). Encephalitis in cattle due to *Pasteurella*. *Australian Veterinary Journal*, 33(5), 124-124.
- Ross, R. F. (2006). *Pasteurella multocida* and its role in porcine pneumonia. *Animal Health Research Reviews*, 7(1-2), 13-29.
- Ruffolo, C. G., Tennent, J. M., Michalski, W. P. and Adler, B. (1997). Identification, purification, and characterization of the type 4 fimbriae of *Pasteurella multocida*. *Infection and Immunity*, 65(1), 339-343.
- Ryckman, L. R., Krahwinkel, D. J., Sims, M. H., Donnell, R. L., Moore, P. F. and Shelton, G. D. (2005). Dysphagia as the primary clinical abnormality in two dogs with inflammatory myopathy. *Journal of the American Veterinary Medical Association*, 226(9), 1519-1523.
- Saharee, A. A., 2006. Haemorrhagic septicaemia in cattle and buffaloes: Are we ready for freedom. Inaugural Lecture, Universiti Putra Malaysia, 1-29.
- Saharee, A., Salin, N., Rasedee, A. and Jainudee, M. (1993). *Haemorrhagic septicaemia carriers among cattle and buffalo in Malaysia Pasteurellosis in production animals*. In Australian Centre for International Agricultural Research (ACIAR) proceedings, Vol. 43, 89-91.
- Saleh, M. A., Rateb, H. and Misk, N. (2008). Comparison of blood serum proteins in water buffaloes with traumatic reticuloperitonitis and sequellae. *Research in Veterinary Science*, 85(2), 208-213.
- Salgado, F. J., Arias, P., Canda-Sánchez, A., Nogueira, M. and Veas, F. (2011). Acute phase proteins as biomarkers of disease: from bench to clinical practice. *Acute Phase Proteins as Early non-Specific Biomarkers of Human and Veterinary Diseases*. Rijeka, Croatia: InTech., 127-174
- Sawada, T., Rimler, R. B. and Rhoades, K. R. (1982). Indirect hemagglutination test that uses glutaraldehyde-fixed sheep erythrocytes sensitized with extract antigens for detection of *Pasteurella* antibody. *Journal of Clinical Microbiology*, 15(5), 752-756.
- Schild, A. L., Fiss, L., Damé, M. C., Uzal, F. A., Soares, M. P., Schuch, L. F., Flores, E. F. and Riet-Correa, F. (2011). Congenital hydranencephaly and cerebellar hypoplasia in water buffalo in southern Brazil. *Journal of Veterinary Diagnostic Investigation*, 23(3), 603-609.
- Scott, P. (1993). A field study of ovine listerial meningo-encephalitis with particular reference to cerebrospinal fluid analysis as an aid to diagnosis and prognosis. *British Veterinary Journal*, 149(2), 165-170.
- Scott, P. (2012). Clinical, ultrasonographic and pathological description of bladder distension with consequent hydroureters, severe hydronephrosis and perirenal fluid accumulation in two rams putatively ascribed to pelvic nerve dysfunction. *Small Ruminant Research*, 107(1), 45-48.

- Scott, P. R. (1995). The collection and analysis of cerebrospinal fluid as an aid to diagnosis in ruminant neurological disease. *British Veterinary Journal*, 151(6), 603-614.
- Scott, P. R. (2004). Diagnostic techniques and clinicopathologic findings in ruminant neurologic disease. *Veterinary Clinics of North America: Food Animal Practice*, 20(2), 215-230.
- Shafarin, M., Zamri-Saad, M., Khairani, B. S. and Saharee, A. A. (2009). Pathological changes in the respiratory tract of goats infected by *Pasteurella multocida* B: 2. *Journal of Comparative Pathology*, 140(2), 194-197.
- Shah, N. H., Biewenga, J., Shah, N. H. and de Graaf, F. K. (1996). Vacuolating cytotoxic activity of *Pasteurella multocida* causing haemorrhagic septicaemia in buffalo and cattle. *FEMS Microbiology Letters*, 143(1), 97-101.
- Shah, N. n. H. and Shah, N. H. (1998). Identification and immunogenicity of polysaccharide antigens of *Pasteurella multocida* strains involved in haemorrhagic septicaemia. *Pakistan Veterinary Journal*, 18, 192-196.
- Shimetani, N., Ichikawa, K., Shibuya, M., Mashiko, T., Matsuyama, N., Kanoh, Y. and Ohtani, H. (1998). Quantitative levels of serum amyloid A protein and other proteins in cerebrospinal fluid and serum of patients with meningitis. *Rinsho byori. The Japanese Journal of Clinical Pathology*, 46(9), 930-935.
- Shirzad Aski, H. and Tabatabaei, M. (2016). Occurrence of virulence-associated genes in *Pasteurella multocida* isolates obtained from different hosts. *Microbial Pathogenesis*, 96, 52-57.
- Shivachandra, S., Viswas, K. and Kumar, A. (2011a). A review of hemorrhagic septicaemia in cattle and buffalo. *Animal Health Research Reviews*, 12(01), 67-82.
- Shivachandra, S. B., Viswas, K. N. and Kumar, A. A. (2011b). A review of hemorrhagic septicaemia in cattle and buffalo. *Animal Health Research Reviews*, 12(01), 67-82.
- Shoshani, J., Kupsky, W. J. and Marchant, G. H. (2006). Elephant brain: Part I: Gross morphology, functions, comparative anatomy, and evolution. *Brain Research Bulletin*, 70(2), 124-157.
- Skinner, J. G. (2001). International standardization of acute phase proteins. *Veterinary Clinical Pathology*, 30(1), 2-7.
- Smith, R. S. (1991). The macrophage theory of depression. *Medical Hypotheses*, 35, 298-306.
- Smith, W. (1993). Responses of laboratory animals to some injectable anaesthetics. *Laboratory Animals*, 27(1), 30-39.

- Son, Y.-S., Park, H.-J., Kwon, O.-B., Jung, S.-C., Shin, H.-C. and Lim, S. (2002). Antipyretic effects of acupuncture on the lipopolysaccharide-induced fever and expression of interleukin-6 and interleukin-1 β mRNAs in the hypothalamus of rats. *Neuroscience Letters*, 319(1), 45-48.
- Song, C. and Wang, H. (2011). Cytokines mediated inflammation and decreased neurogenesis in animal models of depression. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*, 35(3), 760-768.
- Stenken, J. A. and Poschenrieder, A. J. (2015). Bioanalytical chemistry of cytokines- A review. *Analytica Chimica Acta*, 853(0), 95-115.
- Stokol, T., Divers, T. J., Arrigan, J. W. and McDonough, S. P. (2009). Cerebrospinal fluid findings in cattle with central nervous system disorders: a retrospective study of 102 cases (1990–2008). *Veterinary Clinical Pathology*, 38(1), 103-112.
- Stumhofer, J. S., Laurence, A., Wilson, E. H., Huang, E., Tato, C. M., Johnson, L. M., Villarino, A. V., Huang, Q., Yoshimura, A. and Sehy, D. (2006). Interleukin 27 negatively regulates the development of interleukin 17–producing T helper cells during chronic inflammation of the central nervous system. *Nature Immunology*, 7(9), 937-945.
- Srivastava, S. K, 1998. Outer membrane protein of *Pasteurella multocida* serotype B:2 is immunogenic and antiphagocytic. *Indian Journal of Experimental Biology*, 36(5), 530-532.
- Sundaram, C., Shankar, S., Thong, W. K. and Pardo-Villamizar, C. A. (2011). Pathology and diagnosis of central nervous system infections. *Pathology Research International*, 2011, 878263, 1-4
- Surguy, S. M., Duricki, D. A., Reilly, J. M., Lax, A. J. and Robbins, J. (2014). The actions of *Pasteurella multocida* toxin on neuronal cells. *Neuropharmacology*, 77, 9-18.
- Swartz, M. N. and Kunz, L. J. (1959). *Pasteurella multocida* infections in man: report of two cases meningitis and infected cat bite. *New England Journal of Medicine*, 261(18), 889-893.
- Tabatabaei, M., Liu, Z., Finucane, A., Parton, R. and Coote, J. (2002). Protective immunity conferred by attenuated aroA derivatives of *Pasteurella multocida* B: 2 strains in a mouse model of hemorrhagic septicaemia. *Infection and Immunity*, 70(7), 3355-3362.
- Tatro, J. B. (2000). Endogenous antipyretics. *Clinical Infectious Diseases: An Official Publication of the Infectious Diseases Society of America*, 3(5), 190-201.

- Teeling, J. and Perry, V. (2009). Systemic infection and inflammation in acute CNS injury and chronic neurodegeneration: underlying mechanisms. *Neuroscience*, 158(3), 1062-1073.
- Tomer, P., Chaturvedi, G. C., Minakshi, Malik, P. and Monga, D. P. (2002). Comparative analysis of the outer membrane protein profiles of isolates of the *Pasteurella multocida* (B:2) associated with haemorrhagic septicaemia. *Veterinary Research Communications*, 26(7), 513-522.
- Tóthová, C., Nagy, O., Seidel, H., and Kováč, G. (2011). Acute phase proteins as markers of diseases in farm animals. In F. Veas (Ed.), *Acute Phase Proteins as Early Non-Specific Biomarkers of Human and Veterinary Diseases*, 231-252
- Tothova, C., Nagy, O. and Kovac, G. (2013). The use of acute phase proteins as biomarkers of diseases in cattle and swine. *Acute Phase Proteins*, 2013, 103-138.
- Townsend, K. M., Frost, A. J., Lee, C. W., Papadimitriou, J. M. and Dawkins, H. J. (1998). Development of PCR assays for species- and type-specific identification of *Pasteurella multocida* isolates. *Journal of Clinical Microbiology*, 36(4), 1096-1100.
- Townsend, K. M., Boyce, J. D., Chung, J. Y., Frost, A. J. and Adler, B. (2001). Genetic organization of *Pasteurella multocida* cap loci and development of a multiplex capsular PCR typing system. *Journal of Clinical Microbiology*, 39(3), 924-929.
- Uhlar, C. M. and Whitehead, A. S. (1999). Serum amyloid A, the major vertebrate acute-phase reactant. *European Journal of Biochemistry*, 265(2), 501-523.
- Ungar, A., Mussi, C., Del Rosso, A., Noro, G., Abete, P., Ghirelli, L., Cellai, T., Landi, A., Salvioli, G. and Rengo, F. (2006). Diagnosis and characteristics of syncope in older patients referred to geriatric departments. *Journal of the American Geriatrics Society*, 54(10), 1531-1536.
- Urieli-Shoval, S., Cohen, P., Eisenberg, S. and Matzner, Y. (1998). Widespread expression of serum amyloid A in histologically normal human tissues: predominant localization to the epithelium. *Journal of Histochemistry and Cytochemistry*, 46(12), 1377-1384.
- van Furth, A. M., Seijmonsbergen, E. M., Langermans, J. A., Groeneveld, P. H., de Bel, C. E., and van Furth, R. (1995). High levels of interleukin 10 and tumor necrosis factor α in cerebrospinal fluid during the onset of bacterial meningitis. *Clinical Infectious Diseases*, 21(1), 220-222.
- Van Nuffel, A., Zwertvaegher, I., Pluym, L., Van Weyenberg, S., Thorup, V. M., Pastell, M., Sonck, B. and Saeys, W. (2015). Lameness detection in dairy cows: Part 1. How to distinguish between non-lame and lame cows based on differences in locomotion or behavior. *Animals*, 5(3), 838-860.

- Verma, R. and Jaiswal, T. N. (1998). Haemorrhagic septicaemia vaccines. *Vaccine*, 16(11–12), 1184-1192.
- Villard, L., Gauthier, D., Lacheretz, A., Abadie, G., Game, Y., Maurin, F., Richard, Y., Borges, E. and Kodjo, A. (2006). Serological and molecular comparison of *Mannheimia haemolytica* and *Pasteurella trehalosi* strains isolated from wild and domestic ruminants in the French Alps. *The Veterinary Journal*, 171(3), 545-550.
- Visser, M., Bouter, L. M., McQuillan, G. M., Wener, M. H. and Harris, T. B. (1999). Elevated C-reactive protein levels in overweight and obese adults. *Journal of the American Medical Association*, 282(22), 2131-2135.
- Volanakis, J. E. (1982). Complement activation by c-reactive protein complexes. *Annals of the New York Academy of Sciences*, 389(1), 235-250.
- Wade, T., Booy, R., Teare, E. and Kroll, S. (1999). *Pasteurella multocida* meningitis in infancy—(a lick may be as bad as a bite). *European Journal of Pediatrics*, 158(11), 875-878.
- Washburn, K. E. and Streeter, R. N. (2004). Congenital defects of the ruminant nervous system. *Veterinary Clinics of North America: Food Animal Practice*, 20(2), 413-434.
- Welles, E., Tyler, J., Sorjonen, D. and Whatley, E. (1992). Composition and analysis of cerebrospinal fluid in clinically normal adult cattle. *American Journal of Veterinary Research*, 53(11), 2050-2057.
- Whay, H., Main, D., Green, L. and Webster, A. (2003). observations and investigation of farm. *The Veterinary Record*, 153, 197-202.
- Wickham, S., Lu, B., Ash, J. and Carr, D. J. (2005). Chemokine receptor deficiency is associated with increased chemokine expression in the peripheral and central nervous systems and increased resistance to herpetic encephalitis. *Journal of Neuroimmunology*, 162(1), 51-59.
- Wijewardana, T. (1992). Haemorrhagic septicaemia. *Reviews in Medical Microbiology*, 3, 59-63.
- Wilcockson, D. C., Campbell, S. J., Anthony, D. C. and Perry, V. H. (2002). The systemic and local acute phase response following acute brain injury. *Journal of Cerebral Blood Flow and Metabolism*, 22(3), 318-326.
- Wilkie, I. W., Harper, M., Boyce, J. D. and Adler, B. (2012). *Pasteurella multocida*: Diseases and pathogenesis. In K. Aktories, J. H. C. Orth and B. Adler (Eds.), *Pasteurella multocida*. Springer Berlin Heidelberg, 361, 1-22.
- Willoughby Jr, R. E., Tieves, K. S., Hoffman, G. M., Ghanayem, N. S., Amlie-Lefond, C. M., Schwabe, M. J., Chusid, M. J. and Rupprecht, C. E. (2005). Survival after treatment of rabies with induction of coma. *New England Journal of Medicine*, 352(24), 2508-2514.

- Wilson, B. A. and Ho, M. (2013). *Pasteurella multocida*: from Zoonosis to Cellular Microbiology. *Clinical Microbiology Reviews*, 26(3), 631-655.
- Windsor, P., Kessell, A. and Finnie, J. (2011). Neurological diseases of ruminant livestock in Australia. V: congenital neurogenetic disorders of cattle. *Australian Veterinary Journal*, 89(10), 394-401.
- Wolfger, B., Timsit, E., White, B. J. and Orsel, K. (2015). A systematic review of bovine respiratory disease diagnosis focused on diagnostic confirmation, early detection, and prediction of unfavorable outcomes in feedlot cattle. *Veterinary Clinics of North America: Food Animal Practice*, 31(3), 351-365.
- World Organization for Animal Health. (2012). Haemorrhagic septicaemia chapter 2.4.12 *Terrestrial animal health code*. OIE: Paris.
- Xiao, C.-G., Du, M.-X., Li, B., Liu, Z., Chen, M., Chen, Z.-H., Cheng, P., Xue, X.-N., Shapiro, E. and Lepor, H. (2005). An artificial somatic-autonomic reflex pathway procedure for bladder control in children with spina bifida. *The Journal of Urology*, 173(6), 2112-2116.
- Xu-Cai, Y. O., Brotman, D. J., Phillips, C. O., Michota, F. A., Tang, W. W., Whinney, C. M., Panneerselvam, A., Hixson, E. D., Garcia, M. and Francis, G. S. (2008). Outcomes of patients with stable heart failure undergoing elective noncardiac surgery. *Paper presented at the Mayo Clinic Proceedings*.83(3), 280-288
- Yasojima, K., Schwab, C., McGeer, E. G. and McGeer, P. L. (2000). Human neurons generate C-reactive protein and amyloid P: upregulation in Alzheimer's disease. *Brain Research*, 887(1), 80-89.
- Yi, S.-m., Zhu, J.-l., Fu, L.-l. and Li, J.-r. (2010). Tea polyphenols inhibit *Pseudomonas aeruginosa* through damage to the cell membrane. *International Journal of Food Microbiology*, 144(1), 111-117.
- Young, P. J. and Saxena, M. (2014). Fever management in intensive care patients with infections. *Critical Care*, 18(2), 2-8.
- Zald, D. H. and Pardo, J. V. (1999). The functional neuroanatomy of voluntary swallowing. *Annals of Neurology*, 46(3), 281-286.
- Zamri-Saad, M., Effendy, A., Maswati, M., Salim, N. and Sheikh-Omar, A. (1996). The goat as a model for studies of pneumonic pasteurellosis caused by *Pasteurella multocida*. *British Veterinary Journal*, 152(4), 453-458.
- Zamri-Saad, M., Ernie, Z. and Sabri, M. (2006). Protective effect following intranasal exposure of goats to live *Pasteurella multocida* B: 2. *Tropical Animal Health and Production*, 38(7-8), 541-546.
- Zaramella, P., Zamorani, E., Freato, F., Cattai, M. and Meloni, G. A. (1999). Neonatal meningitis due to a vertical transmission of *Pasteurella multocida*. *Pediatrics International*, 41(3), 307-310.

Zhao, X., Song, S., Sun, G., Strong, R., Zhang, J., Grotta, J. C. and Aronowski, J. (2009). Neuroprotective role of haptoglobin after intracerebral hemorrhage. *The Journal of Neuroscience*, 29(50), 15819-15827.

Zweigner, J., Schumann, R. R. and Weber, J. R. (2006). The role of lipopolysaccharide-binding protein in modulating the innate immune response. *Microbes and Infection*, 8(3), 946-952.

