



**UNIVERSITI PUTRA MALAYSIA**

***HEPATOPROTECTIVE ACTIVITY OF *Dicranopteris linearis* (Burm. f.)  
Underw. LEAF METHANOL EXTRACT AND ITS ETHYL ACETATE  
FRACTION***

**FARAH HIDAYAH BINTI KAMISAN**

**FPSK(P) 2017 41**



**HEPATOPROTECTIVE ACTIVITY OF *Dicranopteris linearis* (Burm. f.)  
Underw. LEAF METHANOL EXTRACT AND ITS ETHYL ACETATE  
FRACTION**

**By**

**FARAH HIDAYAH BINTI KAMISAN**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfillment of the Requirements for the Degree of Doctor of  
Philosophy**

**September 2017**

## **COPYRIGHT**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

**HEPATOPROTECTIVE ACTIVITY OF *Dicranopteris linearis* (Burm. f.)  
Underw. LEAF METHANOL EXTRACT AND ITS ETHYL ACETATE  
FRACTION**

By

**FARAH HIDAYAH BINTI KAMISAN**

**September 2017**

**Chairman : Associate Professor Zainul Amiruddin Zakaria, PhD**  
**Faculty : Medicine and Health Sciences**

Liver disease is a major global concern with extremely poor prognosis and high mortality rate due to the lack of effective preventive or treatment options. Besides, a reliable liver protective drug with fewer side effects is still lacking in modern medicines. Therefore, attempts are perpetually being made to investigate several alternative therapies that use natural plants. In the first part of the *in vivo* study, methanol extract (MEDL) of *Dicranopteris linearis* leaves was investigated for hepatoprotective activity against carbon tetrachloride (CCl<sub>4</sub>)-induced and paracetamol (PCM)-induced liver damage. Rats (n=6) used in the study were divided into several groups and given a daily administration of 10% dimethyl sulfoxide (negative control group), 200 mg/kg Silymarin (positive control group) or MEDL (50, 250 or 500 mg/kg) for 7 days, followed by the induction of hepatotoxicity either CCl<sub>4</sub> or PCM. Crude methanolic extracts also were further partitioned using solvents of increasing polarity: petroleum ether <ethyl acetate <water. Semi-purified partitions were called petroleum ether (PEDL), ethyl acetate (EADL) and aqueous (AQDL) extracts. In the second part of *in vivo* study, these partitions were assayed on PCM-induced hepatotoxicity study. All the semi-purified partitions with intermediate doses (250 mg/kg) were tested on the PCM-induced hepatotoxicity model. The best partition proceeded with another two doses of 50 mg/kg and 500 mg/kg. Blood samples and livers were collected and subjected to biochemical and microscopic analyses. The crude and all the semi-purified partitions were also subjected to antioxidant assays, 2, 2-diphenyl-1-picrylhydrazyl radical scavenging assay (DPPH), superoxide dismutase scavenging activity assay (SOD), and oxygen radical absorbance capacity assay (ORAC), and anti-inflammatory assays using lipoxigenase (LOX) and xanthine oxidase (XO) assay. Besides that, total phenolic content (TPC), phytochemical screening and high-performance liquid chromatography (HPLC) analysis and gas chromatography-mass spectrometer (GCMS) were also carried out. MEDL exhibited significant (p<0.05) hepatoprotective activity against both inducers. Pre-

treatment of MEDL at a high dose markedly attenuated the increase in serum alanine aminotransferase (ALT), aspartate aminotransferase (AST) and alkaline phosphatase (ALP) and prevented a marked decrease in superoxide dismutase (SOD) and catalase (CAT) levels in CCl<sub>4</sub>-and PCM-induced hepatotoxic rats. These observations were supported by the histologic findings and scoring, where the liver tissues in groups pre-treated with MEDL and Silymarin showed mild necrosis and inflammation of the hepatocytes compared to the DMSO pre-treated group (negative control group). For the partition extracts, EADL partition exhibited the best activity in protecting the liver against PCM-induced toxicity. Interestingly, EADL, at a low dose (50 mg/kg) significantly ( $p < 0.05$ ) reduced ALP, ALT and AST to  $198.5 \pm 19.78$  U/L,  $108.7 \pm 29.00$  U/L and  $313.0 \pm 65.99$  U/L, respectively and the total bilirubin levels ( $1.133 \pm 0.1687$   $\mu$ mol/L) in PCM-induced hepatotoxic rats. EADL also increased the activity of SOD ( $17.98 \pm 0.09$  U/g tissue) and CAT ( $116.9 \pm 2.71$  U/g tissue) while significantly attenuated the malondialdehyde (MDA) levels of the liver to  $2.61 \pm 0.70$   $\mu$ M in the liver homogenates. EADL partition also ameliorated histopathological changes to liver tissues by the PCM intoxication. In the *in-vitro* antioxidant assay, MEDL showed the highest DPPH- and superoxide anion-radical scavenging activity ( $98.94 \pm 1.14\%$  and  $93.2 \pm 1.18\%$ ) as well as high TPC ( $1757.25 \pm 29.39$  mg/100g GAE) and ORAC ( $24\ 272.50 \pm 2056.53$   $\mu$ M TE/ 100g) values, indicating a high antioxidant activity. In addition, EADL with low TPC ( $352.18 \pm 48.40$  mg/100g GAE) exhibited a high scavenging activity against DPPH- and superoxide anion with  $93.68 \pm 3.0\%$  and  $92.6 \pm 2.17\%$ , respectively. EADL was also the highest in ORAC value with  $555\ 000 \pm 12\ 700$   $\mu$ M TE/ 100g. The phytochemical screening of MEDL and EADL showed the present of saponins, flavonoids, tannins and polyphenolic compounds. HPLC analysis of MEDL also identified the presence of flavonoids, Rutin and Quercetin in the extracts. Moreover, GCMS analysis revealed the presence of 48 volatiles compounds in the MEDL extracts with some of them reported to exhibit antioxidant and anti-inflammatory activity. MEDL and EADL also exerted hepatoprotective activity that could be partly contributed by its antioxidant activity and high phenolic content, and the presence of various bioactive compounds that might act synergistically to enhance the hepatoprotective effect.

Abstrak thesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**KESAN HEPATOPROTEKTIF OLEH EKSTRAK METANOL DAN  
PECAHAN ETIL ESITAT EKSTRAK DARI DAUN *Dicranopteris linearis*  
(Burm. f.) Underw**

Oleh

**FARAH HIDAYAH BINTI KAMISAN**

**September 2017**

**Pengerusi : Profesor Madya Zainul Amiruddin Zakaria, PhD**

**Fakulti : Perubatan dan Sains Kesihatan**

Penyakit hati adalah antara perkara yang membimbangkan secara global dan penyakit ini mempunyai prognosis yang lemah dan kadar kematian yang tinggi disebabkan oleh kekurangan kaedah pencegahan yang efektif atau pilihan rawatan. Selain itu, dalam sistem perubatan moden, terdapat juga kekurangan dadah yang merawat hati dengan kurang kesan sampingan. Oleh itu, usaha dilakukan untuk mengkaji terapi alternatif dengan menggunakan tumbuhan semulajadi. Untuk bahagian pertama kajian in vivo, ekstrak metanol daun *Dicranopteris linearis* (MEDL) telah dikaji untuk aktiviti hepatoprotektif melawan kerosakan hati oleh karbon tetraklorida (CCl<sub>4</sub>) dan parasetamol (PCM). Tikus-tikus (n=6) dibahagikan kepada beberapa kumpulan dengan diberi sekali sehari 10% dimetil sulfoksida (kumpulan kawalan negatif), 200 mg/kg Silymarin (kumpulan kawalan positif), atau MEDL (50, 250 or 500 mg/kg) masing-masing selama 7 hari diikuti dengan induksi kerosakan hati oleh CCl<sub>4</sub> dan PCM. Ekstrak methanol kemudian melalui kaedah partisi dengan menggunakan larutan berbeza polarity secara menaik: petroleum eter < etil asetat < air. Partisi-partisi semi tulen tersebut dipanggil petroleum eter (PEDL), Etil asetat (EADL) dan akua (AQDL). Untuk bahagian kedua kajian in vivo, partisi-partisi tersebut diuji untuk mencegah kerosakan hati oleh PCM. Pada dos sederhana (250 mg/kg), semua partisi-partisi diuji untuk mencegah kerosakan hati oleh PCM. Partisi yang terbaik dilanjutkan untuk ujian pada dos 50 mg/kg dan 500 mg/kg. Sampel darah dan hati dikumpul untuk analisa biokimia dan mikroskopi. Ekstrak mentah MEDL dan semua partisi-partisinya juga diuji untuk antioksidan asai iaitu asai skaveng radikal 2, 2-difenil-1-picirilhidrazil (DPPH), asai skaveng radikal superoksida dismutase (SOD), dan asai kapasiti penyerapan oksigen radikal (ORAC), dan asai anti-inflamasi iaitu asai lipoksigenas (LOX) and xanthine oxidase (XO). Selain itu, jumlah kandungan fenol (TPC), saringan fitokimia, analisa kromatografi cecair berprestasi tinggi (HPLC) dan analisa kromatografi gas- spectrometer jisim (GCMS) juga dijalankan. MEDL menunjukkan aktiviti hepatoprotektif yang ketara (p < 0.05) mencegah

kerusakan hati. Pra-rawatan dengan MEDL pada dos tinggi menurunkan peningkatan paras alanin aminotransferas (ALT), aspartate aminotransferas (AST), dan alkalin fosfatas (ALP) dalam serum secara ketara dan juga mencegah penurunan paras enzim superoksida dismutase (SOD) dan katalas (CAT) dalam hati tikus yang telah dirangsang kerosakan hati oleh CCl<sub>4</sub> dan PCM. Hasil pemerhatian ini juga disokong oleh hasil dari analisa dan indeks histology, dimana hati yang telah pra-rawat oleh MEDL dan Silymarin menunjukkan kurang nekrosis dan inflamasi apabila dibandingkan dengan kumpulan kawalan negative. Untuk ekstrak partisi, partisi EADL menunjukkan aktiviti yang terbaik dalam melindungi hati dari kerosakan oleh PCM. Menariknya, EADL pada dos rendah 50 mg/kg, menurunkan paras ALP, ALT dan AST yang ketara kepada  $198.5 \pm 19.78$  U/L,  $108.7 \pm 29.00$  U/L, dan  $313.0 \pm 65.99$  U/L dan juga paras jumlah bilirubin dalam serum kepada  $1.133 \pm 0.1687$   $\mu\text{mol/L}$ . EADL juga telah meningkatkan aktiviti SOD ( $17.98 \pm 0.09$  U/g tisu) dan CAT ( $116.9 \pm 2.71$  U/g tisu) manakala menurunkan secara ketara paras malondialdihida (MDA) kepada  $2.61 \pm 0.70$   $\mu\text{M}$  dalam hati. Partisi EADL juga mengurangkan perubahan histopatologi kepada tisu hati dari toksik oleh PCM. Dalam asai in vitro antioksidan, MEDL menunjukkan aktiviti yang tertinggi dalam skaveng radikal DPPH dan superoksida ( $98.94 \pm 1.14\%$  dan  $93.2 \pm 1.18\%$ ) dan tinggi TPC ( $1757.25 \pm 29.39$  mg/100g GAE) dan nilai ORAC ( $24\ 272.50 \pm 2056.53$   $\mu\text{M TE/ 100g}$ ) menunjukkan tinggi aktiviti antioksidan. Tambahan pula, EADL dengan jumlah TPC yang rendah ( $352.18 \pm 48.40$  mg/100g GAE) menunjukkan aktiviti yang tinggi dalam skaveng radikal DPPH dan superoksida, masing-masing dengan  $93.68 \pm 3.0\%$  dan  $92.6 \pm 2.17\%$ . EADL juga tinggi nilai ORAC sebanyak  $555\ 000 \pm 12\ 700$   $\mu\text{M TE/ 100g}$ . Dalam saringan fitokimia MEDL dan EADL, menunjukkan adanya sebatian saponin, flavonoid, tannin dan juga polifenolik. Analisa HPLC MEDL juga telah mengenal pasti adanya Rutin dan Quercetin di dalam ekstrak. Selain itu, analisa GCMS juga merungkai terdapat 48 sebatian yang meruap terdapat dalam ekstrak, dimana sebahagian dari sebatian tersebut mempunyai aktiviti antioksidan dan antiinflamasi. MEDL dan EADL mempunyai aktiviti heptoprotektif mungkin disumbang oleh aktiviti antioksidan dan jumlah kandungan fenol di dalamnya, serta disebabkan kehadiran beberapa sebatian bioaktif yang bertindak secara bersinergisma antara satu sama lain untuk mempertingkatkan efek hepatoprotektif.

## ACKNOWLEDGEMENTS

Alhamdulillah, All Praise and Thanks to Allah Subhanahu wa ta'ala for helping me in completing my journey in pursuing the degree of Doctor of Philosophy.

I would like to express my greatest appreciation to my supervisor, Associate Professor Dr Zainul Amiruddin Zakaria who has never stopped encouraging and helping me throughout the course of my study. I thank him for giving me the opportunity to be a postgraduate student under his supervision.

My appreciation also goes to my co-supervisor, Associate Professor Dr Norhafizah Mohtarruddin for her compassion in guiding me in the histological study that is undeniably vital in completing this project. I would also like thank my co-supervisor, Dr Siti Farah Binti Md Tohid and Professor Dr. Mohd Zaki Salleh for the opportunity to conduct a research under their guidance and for their continuous encouragement and willingness to share their knowledge with me.

My sincere appreciation also goes to the staff members of the Pharmacology and Toxicology Laboratory, Histopathology and Chemical Pathology Laboratory and also those from the Animal House, Faculty of Medicine and Health Science, UPM for their cooperation in allowing me to perform my research in the laboratory unit and the excellent facilities that have helped the analyses to run smoothly.

I would also like to acknowledge and express my gratitude to all my fellow friends, Farhana, Roihana, Tavamani, Siti Syariah, Nur Diyana, Nur Liana, Erin, Wahida, Khusairi, Mohd Hijaz, Muhammad Hafiz, Salahuddin and Fauzi Fahmi for their support, assistance and kindness towards me in completing this research project successfully.

Last but not least, my heartfelt gratitude goes to my beloved parents, Encik Kamisan bin Misran and Puan Nor'Aishah Zakaria, my beloved husband, Mohammad Sya'rani Najmuddin and my little daughter Nur Syifa Humaira for their love, patience, moral support and encouragement that have truly motivated me to complete my journey as a postgraduate student successfully. To my family, including members of my family in-law and everyone who has been involved in this project whether directly or indirectly, thank you so much. I can never repay your supports and love. May Allah bless all of them.

Thank you so much.



I certify that a Thesis Examination Committee has met on 7 September 2017 to conduct the final examination of Farah Hidayah binti Kamisan on her thesis entitled "Hepatoprotective Activity of *Dicranopteris linearis* (Burm.f.) Underw. Leaf Methanol Extract and its Ethyl Acetate Fraction" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

**Muhammad Nazrul Hakim bin Abdullah, PhD**

Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Chairman)

**Mohamad Taufik Hidayat bin Baharuldin, PhD**

Associate Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Internal Examiner)

**Patimah binti Ismail, PhD**

Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Internal Examiner)

**Lyndy Joy Mcgaw, PhD**

Professor  
University of Pretoria  
South Africa  
(External Examiner)



**NOR AINI AB. SHUKOR, PhD**

Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 30 November 2017

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

**Zainul Amiruddin Zakaria, PhD**

Associate Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Chairman)

**Norhafizah Mohtarrudin, PhD**

Associate Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Member)

**Siti Farah Md Tohid, PhD**

Senior Lecturer  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Member)

**Mohd. Zaki Salleh, PhD**

Professor  
Faculty of Pharmacy  
Universiti Teknologi MARA  
(Member)

---

**ROBIAH BINTI YUNUS, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_

Date: \_\_\_\_\_

Name and Matric No.: Farah Hidayah binti Kamisan, GS 32383

## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: \_\_\_\_\_

Name of  
Chairman of  
Supervisory  
Committee:

Associate Professor Dr. Zainul Amiruddin Zakaria

Signature: \_\_\_\_\_

Name of  
Member of  
Supervisory  
Committee:

Associate Professor Dr. Norhafizah Mohtarrudin

Signature: \_\_\_\_\_

Name of  
Member of  
Supervisory  
Committee:

Dr. Siti Farah Md Tohid

Signature: \_\_\_\_\_

Name of  
Member of  
Supervisory  
Committee:

Professor Dr. Mohd. Zaki Salleh

## TABLE OF CONTENTS

	<b>Page</b>
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENTS</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	viii
<b>LIST OF TABLES</b>	xv
<b>LIST OF FIGURES</b>	xvii
<b>LIST OF ABBREVIATIONS</b>	xix
<b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	<b>1</b>
1.1 Background of the study	1
1.2 Problem statement	2
1.3 Justification of the study	2
1.4 Hypothesis	3
1.5 Objective of the study	3
1.5.1 General objective	3
1.5.2 Specific objectives	3
<b>2 LITERATURE REVIEW</b>	<b>5</b>
2.1 Liver	5
2.1.1 Anatomy of the liver	5
2.1.2 Histology of the liver	6
2.1.3 Blood supply of the liver	7
2.1.4 Function of the liver	7
2.2 Liver disease	7
2.3 Pathohistological changes of hepatic injury	8
2.3.1 Steatosis	8
2.3.2 Necrosis	8
2.3.3 Inflammation	9
2.4 Hepatotoxicant	9
2.4.1 Carbon tetrachloride	9
2.4.2 Paracetamol	9
2.5 Biomarkers of liver injury	10
2.5.1 Serum liver enzymes	10
2.5.2 Liver oxidative stress marker	11
2.6 Agents used in treatment of liver disease	11
2.6.1 Conventional medicines available for liver disease	11
2.6.2 Side effects of available liver disease medicines	12
2.7 Antioxidant and anti-inflammatory as a mechanism of liver protection	12
2.8 Natural products	13

2.8.1	Natural products with liver protection activity	13
2.8.2	Silymarin	13
2.9	<i>Dicranopteris linearis</i>	14
2.9.1	Origin and geographical distribution	14
2.9.2	Botany description	14
2.9.3	Traditional practices	14
2.9.4	Research background for <i>D.linearis</i>	15
<b>3</b>	<b>GENERAL METHODOLOGY</b>	16
3.1	Plant material and collection	16
3.2	Methanol crude extracts preparation	16
3.3	Animals and chemicals	16
3.4	Hepatoprotective assays	17
3.5	Measurement of serum liver enzymes and bilirubin	17
3.6	Estimation of antioxidant enzymes activity in liver homogenate	17
3.7	Histological analysis	18
3.8	Statistical analysis	18
<b>4</b>	<b>ACUTE TOXICITY STUDY OF METHANOL EXTRACT OF <i>Dicranopteris linearis</i></b>	19
4.1	Introduction	19
4.2	Methodology	19
4.2.1	Experimental animals	19
4.2.2	Acute toxicity study	20
4.2.3	Hematological and biochemical analyses	20
4.3	Results	21
4.3.1	Effects of acute oral administration of MEDL on general signs, body weight and relative organ weight	21
4.3.2	Effects of acute oral administration of MEDL on the haematological and biochemical parameters in mice	23
4.3.3	Histopathological study on various organs following the acute oral administration of MEDL in mice	26
4.4	Discussion	33
<b>5</b>	<b>EVALUATION OF HEPATOPROTECTIVE ACTIVITY OF METHANOLIC EXTRACT OF <i>Dicranopteris linearis</i> LEAVES</b>	34
5.1	Introduction	34
5.2	Methodology	34
5.2.1	Experimental Design for Hepatoprotective Activity	34
5.2.2	Assessment of Hepatoprotective Activity	35
5.3	Results	36
5.3.1	CCl <sub>4</sub> -induced hepatotoxicity assay	36
5.3.1.1	Effects of MEDL on the body weight and liver weight after induction with CCl <sub>4</sub>	36
5.3.1.2	Effects of extracts on serum liver enzymes	36
5.3.1.3	Effects of MEDL extracts on SOD and CAT activities	38

5.3.1.4	Histopathological study of the CCl <sub>4</sub> -induced hepatotoxic liver with and without pre-treatment with MEDL	38
5.3.2	PCM- induced hepatotoxicity assay	44
5.3.2.1	Effects of MEDL on the body weight and liver weight after induction with PCM	44
5.3.2.2	Effects of extracts on serum biochemical parameters	44
5.3.2.3	Effects of extracts on SOD and CAT activities	44
5.3.2.4	Histopathological study of the PCM-induced hepatotoxic liver with and without pre-treatment with MEDL	46
5.4	Discussion	50
<b>6</b>	<b>EVALUATION OF HEPATOPROTECTIVE ACTIVITY OF PETROLEUM ETHER, ETHYL ACETATE AND AQUEOUS PARTITIONS</b>	<b>54</b>
6.1	Introduction	54
6.2	Methodology	54
6.2.1	Preparation of various partitions	54
6.2.2	Hepatoprotective assays	55
6.2.3	Estimation of lipid peroxidation level	56
6.3	Results	56
6.3.1	Extraction yield	56
6.3.2	Effects of partitions on body and liver weights after induction with PCM	56
6.3.3	Effects of PEDL, EADL and AQDL on serum biochemical parameters	56
6.3.4	Effects of EADL and AQDL on MDA, SOD and CAT levels	60
6.3.5	Histopathological study of the PCM-induced hepatotoxic liver with and without pre-treatment of partitions	61
6.4	Discussion	67
<b>7</b>	<b>ANTIOXIDANT ACTIVITY AND TOTAL PHENOLIC CONTENT OF THE METHANOLIC EXTRACT OF <i>Dicranopteris linearis</i> LEAVES AND ITS PARTITIONS</b>	<b>70</b>
7.1	Introduction	70
7.2	Methodology	70
7.2.1	Total phenolic content	70
7.2.2	2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging activity	71
7.2.3	Superoxide anion radical scavenging activity	71
7.2.4	Oxygen radical absorbance capacity (ORAC)	72

7.3	Result	72
7.3.1	Total phenolic content	72
7.3.2	2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging activity	72
7.3.3	Superoxide anion radical scavenging	73
7.3.4	Oxygen radical absorbance capacity (ORAC)	73
7.4	Discussion	75
<b>8</b>	<b>ASSESSMENT OF THE ANTI-INFLAMMATORY ACTIVITY OF THE METHANOLIC EXTRACT OF <i>Dicranopteris linearis</i> LEAVES AND ITS PARTITIONS</b>	<b>79</b>
8.1	Introduction	79
8.2	Methodology	80
8.2.1	Lipoxygenase assay	80
8.2.2	Xanthine oxidase assay	80
8.3	Results	80
8.4	Discussion	81
<b>9</b>	<b>PHYTOCHEMICAL SCREENING, HPLC AND GC-MS ANALYSIS OF METHANOLIC EXTRACT OF <i>Dicranopteris linearis</i> LEAVES AND ITS PARTITIONS</b>	<b>83</b>
9.1	Introduction	83
9.2	Methodology	83
9.2.1	Phytochemical screening	83
9.2.1.1	Test for alkaloids	84
9.2.1.2	Test for saponins	84
9.2.1.3	Test for flavonoids	84
9.2.1.4	Test for tannins and polyphenolic compounds	84
9.2.1.5	Test for triterpenes/ steroids	84
9.2.2	HPLC analysis of MEDL crude extract, and EADL and AQDL partitions	85
9.2.3	Identification of flavonoids in MEDL via HPLC analysis	85
9.2.4	GC-MS analysis of MEDL crude extract	86
9.3	Results	86
9.3.1	Phytochemical screening	86
9.3.2	HPLC profiling of MEDL crude extract	86
9.3.3	GC-MS profile of crude MEDL	91
9.3.4	HPLC profiling of EADL and AQDL partitions	95
9.4	Discussion	100
<b>10</b>	<b>GENERAL DISCUSSION, SUMMARY, CONCLUSION AND RECOMMENDATIONS</b>	<b>103</b>
10.1	General discussion	103
10.2	Summary	106
10.3	Limitation of the study	106
10.4	Conclusion and Recommendations	106



<b>REFERENCES</b>	108
<b>APPENDICES</b>	126
<b>BIODATA OF STUDENT</b>	128
<b>LIST OF PUBLICATIONS</b>	129



## LIST OF TABLES

Table		Page
4.1	Haematology value of mice treated with single dose of <i>Dicranopteris linearis</i> extract	24
4.2	Biochemistry value of mice treated with single dose of <i>Dicranopteris linearis</i> methanol extract	25
5.1	CCl <sub>4</sub> -induced hepatotoxicity treatment groups	35
5.2	PCM-induced hepatotoxicity treatment groups	35
5.3	Effects of MEDL on percentage change of body and liver weight in CCl <sub>4</sub> - treated rats	37
5.4	Effects of MEDL on serum liver biomarkers of CCl <sub>4</sub> - treated rats	37
5.5	Effects of MEDL extracts on SOD and catalase activities	38
5.6	Histopathological scoring of the liver section of CCl <sub>4</sub> - induced hepatic injury rats pre-treated with MEDL	43
5.7	Effects of MEDL on percentage change of body and liver weight in PCM- treated rats	45
5.8	Effects of MEDL on serum liver biomarkers of PCM- treated rats	45
5.9	Antioxidant enzyme activities in liver tissue of PCM- treated rats, SOD (U/g tissue) and CAT (U/g tissue)	46
5.10	Histopathological scoring of the liver section of PCM- induced hepatic injury rats pre-treated with MEDL	50
6.1	PCM-induced hepatotoxicity treatment groups of partition extracts	55
6.2	Effects of PEDL, EADL and AQDL on percentage change of liver weight in PCM- treated rats	58
6.3	Effects of PEDL, EADL and AQDL on ALT, AST, ALP (U/L), and total bilirubin (umol/L)	59
6.4	Effects of EADL and AQDL on SOD, CAT and MDA levels	60

6.5	Histopathological scoring of the liver section of PCM- induced hepatic injury rats pre-treated with PEDL, EADL and AQDL	67
7.1	Total phenolic content of MEDL extract and its partitions	73
7.2	The DPPH radical scavenging activity of MEDL and its partitions	74
7.3	Superoxide scavenging activity of MEDL and its partition	74
7.4	The total oxygen radical absorbance capacity (ORAC) value of MEDL and its partitions	75
8.1	Effects of extracts on the anti-inflammatory mediators using the in vitro lipoxygenase and xantine oxidase assays	81
9.1	The solvent system used for HPLC profiling	85
9.2	Comparison on the presence of phytochemical constituents of the leaves in different extracts of <i>D. linearis</i> leaves	90
9.3	GCMS profile shows the volatile phytoconstituents of MEDL	93
9.4	Volatile compounds with anti-inflammatory and antioxidant activities	95

## LIST OF FIGURES

Figure		Page
2.1	Anatomy of liver	5
2.2	Schematic representation of hepatic acinus	6
2.3	<i>Dicranopteris linearis</i>	15
4.1	Effects of methanol extract of <i>D. linearis</i> on body weight of male (A) and female (B) ICR mice in acute oral toxicity	22
4.2	Liver photomicrographs of male mice of normal control, male and female mice treated with 5000 mg/kg MEDL	27
4.3	Kidney photomicrographs of male mice of normal control, male and female mice treated with 5000 mg/kg MEDL	28
4.4	Spleen photomicrographs of male mice of normal control, male and female mice treated with 5000 mg/kg MEDL	29
4.5	Lung photomicrographs from Male mice of normal control, male and female mice treated with 5000 mg/kg MEDL	30
4.6	Stomach photomicrographs of Male mice of normal control, male and female mice treated with 5000 mg/kg MEDL	31
4.7	Heart photomicrographs of Male mice of normal control, male and female mice treated with 5000 mg/kg MEDL	32
5.1(a),(b)	Liver photomicrographs of normal control and CCl <sub>4</sub> -intoxicated rats	40
5.1(c),(d)	Liver photomicrographs of male rats pre-treated with 200 mg/kg Silymarin and 50 mg/kg MEDL followed by CCl <sub>4</sub> intoxication	41
5.1(e),(f)	Liver photomicrographs of male rats pre-treated with 250 mg/kg MEDL and 500 mg/kg MEDL followed by CCl <sub>4</sub> intoxication	42
5.2(a),(b)	Liver photomicrographs of normal control and PCM-intoxicated rats.	47
5.2(c),(d)	Liver photomicrographs of male rats pre-treated with 200 mg/kg silymarin and pre-treated with 50 mg/kg MEDL followed by PCM	48

5.2(e),(f)	Liver photomicrographs of male rats pre-treated with 250 mg/kg and 500 mg/kg MEDL followed by PCM	49
6.1(a),(b)	Liver photomicrographs of normal male rats and male rats treated with 3g/kg PCM	62
6.1(c),(d)	Liver photomicrographs of male rats pre-treated with 200 mg/kg silymarin and 50 mg/kg EADL followed by PCM intoxication	63
6.1(e),(f)	Liver photomicrographs of male rats pre-treated with 250 mg/kg EADL and 500 mg/kg EADL followed by PCM intoxication	64
6.1(g),(h)	Liver photomicrographs of male rats pre-treated with 50 mg/kg AQDL and 250 mg/kg AQDL followed by PCM intoxication	65
6.1(i),(j)	Liver photomicrographs of male rats pre-treated with 500 mg/kg AQDL and 250 mg/kg PEDL followed by PCM intoxication	66
9.1	HPLC profile of MEDL	88
9.2	The UV spectra analysis of MEDL	89
9.3	GCMS chromatogram of MEDL	92
9.4	The HPLC profile of EADL	96
9.5	The UV spectra analysis of EADL	97
9.6	The HPLC profile of AQDL	98
9.7	The UV spectra analysis of AQDL.	99

## LIST OF ABBREVIATIONS

ALP	Alkaline phosphatase
ALT	Alanine aminotransferase
ANOVA	Analysis of variance
APAP	Acetyl-para-aminophenol
AQDL	Aqueous extract of <i>Dicranopteris linearis</i>
AST	Aspartate aminotransferase
ATP	Adenosine triphosphate
CAT	Catalase
CAM	Complementary and alternative medicine
CCl <sub>3</sub> ·	Trichloromethyl free radical
CCl <sub>4</sub>	Carbon tetrachloride
Cl <sub>3</sub> COO·	Trichloromethyl peroxy
CMC	Sodium carbomethyl cellulose
COX	Cyclooxygenase
CYP450	Cytochrome P450
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
DPPH	2,2-diphenyl-1-picrylhydrazyl
EADL	Ethyl acetate extract of <i>Dicranopteris linearis</i>
EDTA	Ethylenediaminetetra acetic acid
FDA	Food and Drug Administration
eNOS	Endothelial nitric oxide synthase
GSH	Glutathione
Hb	Haemoglobin
HPLC	High performance liquid chromatography
i.p	Intraperitoneally
IC <sub>50</sub>	Median inhibitory concentration
IBS	Institute of Bioscience
ICR	Institute of Cancer Research
iNOS	Inducible nitric oxide synthase
LD <sub>50</sub>	Lethal dose of 50% population
LOX	Lipoxygenase
MCHC	Mean corpuscular haemoglobin concentration
MCV	Mean corpuscular volume
MDA	Malondialdehyde
MEDL	Methanol extract of <i>Dicranopteris linearis</i>
MeOH	Methanol
NAC	N-acetyl cysteine
NAPQI	N-acetyl-p-benzoquinoneimine
NO	Nitric oxide
OECD	Organisation for Economic Cooperation and Development
o.p	Orally
ORAC	Oxygen radical absorbance capacity
PCM	Paracetamol
PCV	Packed cell volume

PEDL	Petroleum ether extract of <i>Dicranopteris linearis</i>
RBC	Red blood cell
ROW	Relatives organ weight
ROS	Reactive oxygen species
SEM	Standard error mean
SOD	Superoxide dismutase
TB	Total bilirubin
TPC	Total phenolic content
WBC	White blood cell
WHO	World Health Organization
w/v	Weight/ volume
XO	Xanthine oxidase



# CHAPTER 1

## INTRODUCTION

### 1.1 Background of the study

Liver is one of the most vital organs involved with almost all the biochemical pathways to growth, nutrient supply, fight against disease, energy provision and reproduction. Nevertheless, factors such as toxic chemicals, alcohol abuse and viral infections can lead to different extent of liver injury. Liver disease can be classified as acute or chronic hepatitis (inflammatory liver disease), hepatosis (non-inflammatory diseases) and cirrhosis (degenerative disorder leading to fibrosis of the liver) (Kumar et al., 2012). Nowadays, liver disease has become one of the main concerns threatening human health at high prevalence (Hou et al., 2013). For this reason, medical experts and the general population are increasingly paying attention to ways to maintain a healthy liver. Scientific research on natural plants or their compounds with potential hepatoprotective activity for functional food and nutraceutical development has also been an important topic in many fields, including nutrition and pharmaceutical.

Moreover, the World Health Organization (WHO) recorded that 80% of the world's population depends on either, wholly or partly on plant-derived pharmaceuticals (MacDonald et al., 2016). The world population have shifted from synthetic drugs to natural products because of the very expensive cost and associated intolerable side effects of most synthetic drugs used for hepatic disorders (Khan et al., 2017; MacDonald et al., 2016). Herbal medicines have gained significance and popularity in recent years due to safety, efficacy and cost effectiveness associated with them. Extracts of a wide range of natural plants have been used as raw drugs, and they possess varied medicinal properties. Besides, these plants also contain a wide range of phyto-constituents that can be used to treat diseases. The different parts that are used include the leaves, stems, flowers, fruits and twigs exudates (MacDonald et al., 2016).

One group of plants in biodiversity that has been neglected as far as its economic value is concerned is the pteridophytes. Used as folk medicine, the pteridophytes, which constitute fern and ferns allies, have been known to man for more than 2000 years (Talukdar et al., 2011). *Dicranopteris linearis* is one of the neglected plants that is widely distributed in Malaysia, and it has been used in folk medicine in some countries including Malaysia, Papua New Guinea and India (Zakaria et al., 2008). The Malays call it "Resam" and have been using its leaves to control fever as it helps in reducing body temperature. *D.linearis* leaves are squeezed in water before the water is drunk as tonic, or the wet leaves are applied to one's body as poultice (Zakaria et al., 2008). Several other biological attributes of *D. linearis* leaves have also been reported; nevertheless, none on them were on the



hepatoprotective properties. Therefore, this study is to assess the potential of *D. linearis* leaves as a hepatoprotective agent.

## 1.2 Problem statement

Liver disease is a health problem worldwide. The causes that may contribute to liver disease are excessive alcohol consumption, drug overdose (eg. Paracetamol), viral infection and obesity. According to the Malaysian Liver Registry, Association of Clinical Registries Malaysia (2005), digestive disorder was responsible for 3.2% of the total burden disease in Malaysia in 2000, and cirrhosis of the liver was the main and important cause of burden of disease among digestive disorders causal as it was responsible for up to 60% of the total burden in this category. In Malaysia, liver cirrhosis is mostly caused by the viral hepatitis B and C infections and also alcohol consumption (Qua and Goh, 2011). However, recently there is a rising prevalence of non-alcoholic fatty liver disease (NAFLD) due to diabetes and obesity which are common among Malaysians. The fatty accumulation in liver could result in liver inflammation, liver tissue scarring (fibrosis), cirrhosis and even liver cancer (Ho, 2013). In addition, overdose of paracetamol (PCM) is recognised to result in a range of hepatic damage, leading to acute liver failure (ALF) and death. This problem is infrequent in Asia; however, it is a common means of self-poisoning worldwide because of its wide availability and accessibility (Marzilawati et al., 2012). Acute liver failure due to PCM overdose has been extensively reported in the United Kingdom (UK), the United States (US), France, Canada and Australia. 60 to 75% of acute liver failure in the UK was due to paracetamol overdoses (Marzilawati et al., 2012).

## 1.3 Justification of the study

Despite remarkable advances in modern medicines, liver disease remains a health problem worldwide. Hence, the search for new treatments is still ongoing. The currently used conventional or synthetic drugs in the treatment of liver disease are insufficient and have severe side effects. Besides, some of the available treatments also have low bioavailability and poor solubility (Khan et al., 2017). Due to the serious side effects of available treatments, there is an urgent need to investigate and develop more efficient hepatoprotective agents with high bioavailability, better solubility, but at the same time, inexpensive and safe. In recent times, focus on plant research has increased all over the world and interestingly, utilisation of herbal medicines to prevent various chronic diseases has been a common clinical practice for long in Asian countries (Ghosh et al., 2011). Thus, investigation of hepatoprotective activity of methanol extract of *D. linearis* (MEDL) leaves and its partitions, may contribute to the identification of another natural plant product that may be effective as a hepatoprotective agent. To date, there is no complete study on the hepatoprotective activity of methanol extract of *D. linearis* leaves and its partitions. Recently, investigations of *D. linearis* leaves have demonstrated that they exert anti-oxidant and anti-inflammatory activities (Zakaria et al., 2011; Zakaria et al., 2008). Both activities are crucial as a mechanism of liver protection against toxic substances. Literature review shows that some plants are useful in

protecting the liver by improving the antioxidant and anti-inflammatory statuses (Makni et al., 2011; Makni et al., 2010; Makni et al., 2008). In addition, phytochemical studies on *D.linearis* leaves have reported to show the presence of various types of flavonoids, particularly of flavonol 3-o-glycosides types and triterpenes, saponins and steroids (Hussaini et al., 2012). Flavonoids possess a wide range of bioactive capacities, especially their antioxidant activity as antioxidants play a significant role in liver protection (Pareek et al., 2013). Supported by these facts, *D. linearis* leaves may also have the potential to exert hepatoprotective activities thus, justifying the investigation of hepatoprotective activity of *D. linearis*. This study may contribute in the search for other plant products that can be used in the treatment of liver disease.

#### **1.4 Hypothesis**

Methanol extract of *D. linearis* leaves (MEDL) possesses hepatoprotective activity against both carbon tetrachloride (CCl<sub>4</sub>)- and Paracetamol (PCM)-induced liver injury, and one of its partitions either petroleum ether partition (PEDL), ethyl acetate partition (EADL) or aqueous partition (AQDL) possesses the best hepatoprotective agent against PCM-induced liver injury.

#### **1.5 Objective of the study**

##### **1.5.1 General objective**

This study aimed to investigate the *in vivo* hepatoprotective properties of the methanol extract of *Dicranopteris linearis* leaves (MEDL) and its petroleum ether (PEDL), ethyl acetate (EADL) and aqueous (AQDL) partitions using rat models.

##### **1.5.2 Specific objectives**

1. To determine the acute toxicity effects of *D. linearis* methanol extract *in vivo*.
2. To investigate the potential of the methanol extract of *D. linearis* (MEDL) as a hepatoprotective agent against CCl<sub>4</sub> and PCM toxicity.
3. To determine whether (PEDL, EADL or AQDL) is the best to possess the hepatoprotective activity against PCM- induced liver injury.
4. To assess the antioxidant activities of MEDL and its partitions via determination of 2, 2-diphenyl-1-picrylhydrazyl radical scavenging (DPPH), superoxide dismutase scavenging activity (SOD), and oxygen radical absorbance capacity (ORAC) assays that may be involved to act as a pathway in hepatoprotective properties of *Dicranopteris linearis* against liver toxicants.
5. To assess the *in vitro* anti-inflammatory activity of MEDL and its partitions using lipoxygenase and xanthine oxidase assays.

6. To screen the bioactive compounds present in MEDL crude extract and its partitions using high performance liquid chromatography (HPLC) and gas chromatography-mass spectrometer (GCMS).



## REFERENCES

- Acamovic, T., and Brooker, J. D. (2005). Biochemistry of plant secondary metabolites and their effects in animals. *Proceedings of the Nutrition Society*, 64(03), 403-412.
- Ackermann, J. A., Hofheinz, K., Zaiss, M. M., and Krönke, G. (2016). The double-edged role of 12/15-lipoxygenase during inflammation and immunity. *Biochimica et Biophysica Acta (BBA)-Molecular and Cell Biology of Lipids*.
- Ahmed, M.B., and Khater, M.R., (2001). Evaluation of the protective potential of *Ambrosia maritima* extract on acetaminophen- induced liver damage. *Journal of Ethnopharmacology*. 75, 169- 174.
- Ahsan, M. R., Islam, K. M., Bulbul, I. J., Musaddik, M. A., and Haque, E. (2009). Hepatoprotective activity of methanol extract of some medicinal plants against carbon tetrachloride-induced hepatotoxicity in rats. *European Journal of Scientific Research*, 37(2), 302-310.
- Akamatsu, K., Yamasaki, Y., Nishikawa, M., Takakura, Y., and Hashida, M. (2001). Synthesis and pharmacological activity of a novel water-soluble hepatocyte-specific polymeric prodrug of prostaglandin E 1 using lactosylated poly (L-glutamic hydrazide) as a carrier. *Biochemical Pharmacology*, 62(11), 1531-1536.
- Alinezhad, H., Baharfar, R., Zare, M., Azimi, R., Nabavi, S. F., and Nabavi, S. M. (2012). Biological activities of ethyl acetate extract of different parts of *Hyssopus angustifolius*. *Pharmaceutical Biology*, 50(8), 1062-1066.
- Ashoush, I. S., El-Batawy, O. I., and El-Shourbagy, G. A. (2013). Antioxidant activity and hepatoprotective effect of pomegranate peel and whey powders in rats. *Annals of Agricultural Sciences*, 58(1), 27-32.
- Association of Clinical Registry Malaysia. *Malaysia Liver Registry*, [http:// www.Acrm.org.my/affiliatedDB.php](http://www.Acrm.org.my/affiliatedDB.php) (accessed 22 September 2017).
- Atiba, A. S., Abbiyesuku, F. M., Oparinde, D. P., Temitope, A., and Akindele, R. A. (2016). Plasma Malondialdehyde (MDA): an indication of liver damage in women with pre-eclampsia. *Ethiopian Journal of Health Sciences*, 26(5), 479-486.
- Azlim Almey, A. A., Ahmed Jalal Khan, C., Syed Zahir, I., Mustapha Suleiman, K., Aisyah, M. R., and Kamarul Rahim, K. (2010). Total phenolic content and primary antioxidant activity of methanolic and ethanolic extracts of aromatic plants' leaves. *International Food Research Journal*, 17(4).

- Babu, B. H., Shylesh, B. S., and Padikkala, J. (2001). Antioxidant and hepatoprotective effect of *Acanthus ilicifolius*. *Fitoterapia*, 72(3), 272-277.
- Bagban, I. M., Roy, S. P., Chaudhary, A., Das, S. K., Gohil, K. J., and Bhandari, K. K. (2012). Hepatoprotective activity of the methanolic extract of *Fagonia indica* Burm in carbon tetra chloride induced hepatotoxicity in albino rats. *Asian Pacific Journal of Tropical Biomedicine*, 2(3), S1457-S1460.
- Bharathi, T. R., Nadafi, R., and Prakash, H. S. (2014). In vitro antioxidant and anti-inflammatory properties of different Solvent Extracts of *Memecylon talbotianum* Brandis. *International Journal of Phytopharmacy*, 4(6), 148-152.
- Bhawna, S., and Kumar, S. U. (2009). Hepatoprotective activity of some indigenous plants. *International Journal of Pharm Tech Research*, 4, 1330-1334.
- Bhoopat, L., Srichairatanakool, S., Kanjanapothi, D., Taesotikul, T., Thananchai, H., and Bhoopat, T. (2011). Hepatoprotective effects of lychee (*Litchi chinensis* Sonn.): a combination of antioxidant and anti-apoptotic activities. *Journal of Ethnopharmacology*, 136(1), 55-66.
- Blois, M. S. (1958). Antioxidant determinations by the use of a stable free radical. *Nature*, 181(4617), 1199-1200.
- Boyle, S. P., Dobson, V. L., Duthie, S. J., Hinselwood, D. C., Kyle, J. A., and Collins, A. R. (2000). Bioavailability and efficiency of rutin as an antioxidant: a human supplementation study. *European Journal of Clinical Nutrition*, 54(10), 774-782.
- Brink, M., and Achigan-Dako, E. G. (2012). Plant Resources of Tropical Africa 16 Fibres. *Economic Botany*, 66(3), 123-125.
- Brufau, G., Canela, M. A., and Rafecas, M. (2008). Phytosterols: physiologic and metabolic aspects related to cholesterol-lowering properties. *Nutrition Research*, 28(4), 217-225.
- Calixto, J. B., Otuki, M. F., and Santos, A. R. (2003). Anti-inflammatory compounds of plant origin. Part I. Action on arachidonic acid pathway, nitric oxide and nuclear factor  $\kappa$  B (NF- $\kappa$ B). *Planta Medica*, 69(11), 973-983.
- Chin, W.Y. (1992). *A Guide to Medicinal Plants*. Singapore Sci. Centre, p. 24.
- Conforti, F., Sosa, S., Marrelli, M., Menichini, F., Statti, G. A., Uzunov, D., and Menichini, F. (2009). The protective ability of Mediterranean dietary plants against the oxidative damage: the role of radical oxygen species in

inflammation and the polyphenol, flavonoid and sterol contents. *Food Chemistry*, 112(3), 587-594

Coruh, N., Celep, A. S., and Özgökçe, F. (2007). Antioxidant properties of *Prangos ferulacea* (L.) Lindl., *Chaerophyllum macropodium* Boiss. and *Heracleum persicum* Desf. from Apiaceae family used as food in Eastern Anatolia and their inhibitory effects on glutathione-S-transferase. *Food Chemistry*, 100(3), 1237-1242.

Cotoi, C. G., and Quaglia, A. (2016). Normal Liver Anatomy and Introduction to Liver Histology. In *Textbook of Pediatric Gastroenterology, Hepatology and Nutrition* (pp. 609-612). Springer International Publishing.

Crankshaw, D. L., Berkeley, L. I., Cohen, J. F., Shirota, F. N., and Nagasawa, H. T. (2002). Double-prodrugs of L-cysteine: Differential protection against acetaminophen-induced hepatotoxicity in mice. *Journal of Biochemical and Molecular Toxicology*, 16(5), 235-244.

Croteau, R., Kutchan, T. M., and Lewis, N. G. (2000). Natural products (secondary metabolites). *Biochemistry and Molecular Biology of Plants*, 24, 1250-1319.

Dai, J., and Mumper, R. J. (2010). Plant phenolics: extraction, analysis and their antioxidant and anticancer properties. *Molecules*, 15(10), 7313-7352.

Darus, F. M., Buyong, F., and Abdullah, S. (2004). Fern Tree (*Gleichenia Linearis*) As Metal Sorbent For Lead Ions Removal.

Das, P., and Holt, S. (2011). Liver disease and renal dysfunction. *Medicine*, 39(8), 492-496.

Davis, R. H., and Maro, N. P. (1989). Aloe vera and gibberellin. Anti-inflammatory activity in diabetes. *Journal of the American Podiatric Medical Association*, 79(1), 24-26.

Dehmlow, C., Murawski, N., and de Groot, H. (1996). Scavenging of reactive oxygen species and inhibition of arachidonic acid metabolism by silibinin in human cells. *Life Sciences*, 58(18), 1591-1600.

Dehshiri, M. M., Aghamollaei, H., Zarini, M., Nabavi, S. M., Mirzaei, M., Loizzo, M. R., and Nabavi, S. F. (2013). Antioxidant activity of different parts of *Tetrataenium lasiopetalum*. *Pharmaceutical Biology*, 51(8), 1081-1085.

Denecker, G., Vercammen, D., Declercq, W. and Vandenaabeele, P. (2001). Review Apoptotic and necrotic cell death induced by death domain receptors. *Cellular and Molecular Life Science*, 58: 356-370.

Derrickson, B., and Tortora, G. J. (2006). *Principles of Anatomy and Physiology*. Wiley.

- Desai, F., Ramanathan, M., Fink, C. S., Wilding, G. E., Weinstock-Guttman, B., and Awad, A. B. (2009). Comparison of the immunomodulatory effects of the plant sterol  $\beta$ -sitosterol to simvastatin in peripheral blood cells from multiple sclerosis patients. *International Immunopharmacology*, 9(1), 153-157.
- Devkar, S. T., Kandhare, A. D., Zanwar, A. A., Jagtap, S. D., Katyare, S. S., Bodhankar, S. L., and Hegde, M. V. (2016). Hepatoprotective effect of withanolide-rich fraction in acetaminophen-intoxicated rat: decisive role of TNF- $\alpha$ , IL-1 $\beta$ , COX-II and iNOS. *Pharmaceutical Biology*, 54(11), 2394-2403.
- Dixit, N., Baboota, S., Kohli, K., Ahmad, S., and Ali, J. (2007). Silymarin: A review of pharmacological aspects and bioavailability enhancement approaches. *Indian Journal of Pharmacology*, 39(4), 172.
- Domitrović, R., Jakovac, H., and Blagojević, G. (2011). Hepatoprotective activity of berberine is mediated by inhibition of TNF- $\alpha$ , COX-2, and iNOS expression in CCl 4-intoxicated mice. *Toxicology*, 280(1), 33-43.
- Domitrović, R., Jakovac, H., Marchesi, V. V., Vladimir-Knežević, S., Cvijanović, O., Tadić, Ž., and Rahelić, D. (2012). Differential hepatoprotective mechanisms of rutin and quercetin in CCl4-intoxicated BALB/cN mice. *Acta Pharmacologica Sinica*, 33(10), 1260-1270.
- Dorman, R. B., Wunder, C., Saba, H., Shoemaker, J. L., Macmillan-Crow, L. A., and Brock, R. W. (2005). NAD (P) H oxidase contributes to the progression of remote hepatic parenchymal injury and endothelial dysfunction, but not microvascular perfusion deficits. *American Journal of Physiology-Gastrointestinal and Liver Physiology*. 290: G1025–G1032.
- El-Beshbishy, H. A., Mohamadin, A. M., Nagy, A. A., and Abdel-Naim, A. B. (2010). Amelioration of tamoxifen-induced liver injury in rats by grape seed extract, black seed extract and curcumin. *Indian Journal of Experimental Biology*. 48:280–288.
- Fakurazi, S., Hairuszah, I., and Nanthini, U. (2008). Moringa oleifera Lam prevents acetaminophen induced liver injury through restoration of glutathione level. *Food and Chemical Toxicology*, 46(8), 2611-2615.
- Fakurazi, S., Sharifudin, S. A., and Arulselvan, P. (2012). Moringa oleifera hydroethanolic extracts effectively alleviate acetaminophen-induced hepatotoxicity in experimental rats through their antioxidant nature. *Molecules*, 17(7), 8334-8350.
- Feldman, B.V., Zinkl, J.G., Jain, N.C., (2000). *Schalm's Veterinary Hematology*, 5th ed. Lea Febiger, Philadelphia, 1210–1218.

- Fiebrich, F., and Koch, H. (1979). Silymarin, an inhibitor of lipoxygenase. *Cellular and Molecular Life Sciences*, 35(12), 1548-1550.
- Fontana, R. J. (2009). Side effects of long-term oral antiviral therapy for hepatitis B. *Hepatology*, 49(S5).
- Fujisawa, K., Yabuuchi, C., Izawa, T., Kuwamura, M., Takasu, N., Torii, M., and Yamate, J. (2013). Expression patterns of heat shock protein 25 in carbon tetrachloride-induced rat liver injury. *Experimental and Toxicologic Pathology*, 65(5), 469-476.
- Gao, J., Sun, C. R., Yang, J. H., Shi, J. M., Du, Y. G., Zhang, Y. Y., and Wan, H. T. (2011). Evaluation of the hepatoprotective and antioxidant activities of *Rubus parvifolius* L. *Journal of Zhejiang University SCIENCE B*, 12(2), 135-142.
- Gardner, C., and Laskin, D., (2007). Sinusoidal cells in liver injury and repair. In: Sahu, S. (Ed.), *Hepatotoxicity: From Genomics to in Vitro and in Vivo Models*. John Wiley & Sons, Ltd, West Sussex, 341-370.
- Gebhardt, R. (2002). Oxidative stress, plant-derived antioxidants and liver fibrosis. *Planta Medica*, 68(04), 289-296.
- Geller, S. A., and Petrovic, L. M. (2004). *Biopsy Interpretation of The Liver*. Lippincott Williams & Wilkins.
- Ghosh, N., Ghosh, R., Mandal, V., Mandal, S.C. (2011). Recent advances in herbal medicine for treatment of liver diseases. *Pharmaceutical Biology*, 49, 970-988.
- Gregory, M., Vithalrao, K. P., Gregory, F., and Kalaichelvan, V. K. (2009). Anti-ulcer (ulcer-preventive) activity of *Ficus arnottiana* Miq. (Moraceae) leaf methanolic extract. *American Journal of Pharmacology and Toxicology*, 4(3), 89-93.
- Guardia, T., Rotelli, A. E., Juarez, A. O., and Pelzer, L. E. (2001). Anti-inflammatory properties of plant flavonoids. Effects of rutin, quercetin and hesperidin on adjuvant arthritis in rat. *Il Farmaco*, 56(9), 683-687.
- Guay, J., Bateman, K., Gordon, R., Mancini, J., and Riendeau, D. (2004). Carrageenan-induced paw edema in rat elicits a predominant prostaglandin E2 (PGE2) response in the central nervous system associated with the induction of microsomal PGE2 synthase-1. *Journal of Biological Chemistry*, 279(23), 24866-24872.
- Gutiérrez, R. M., & Solís, R. V. (2009). Hepatoprotective and inhibition of oxidative stress in liver of *Prostechea michuacana*. *Records of Natural Products*, 3(1), 46.



- Gutowska, I., Jakubczyk, K., Dec, K., Baranowska-Bosiacka, I., Drozd, A., Janda, K., and Chlubek, D. (2014). Effect of the extract from nettle (*Urtica dioica* L.) fruit cluster on the synthesis of pro-inflammatory agents in hepatocytes treated with fluoride. *Res Rep Fluoride*, 47(2), 109-18.
- Harizal, S. N., Mansor, S. M., Hasnan, J., Tharakan, J. K. J., and Abdullah, J. (2010). Acute toxicity study of the standardized methanolic extract of *Mitragyna speciosa* Korth in rodent. *Journal of Ethnopharmacology*, 131(2), 404-409.
- Hinson, J.A., Reid, A.B., McCullough, S.S., James, L.P., (2004). Acetaminophen – induced hepatotoxicity: role of metabolite activation, reactive oxygen/nitrogen species, and mitochondrial permeability transition. *Drug Metabolism Review*. 36, 805- 822.
- Ho, F. (2013). Eliminating fatty liver disease. *TheStar Online* (Malaysia), August 4, 2013, <http://www.thestar.com.my/lifestyle/health/2013/08/04/eliminating-fatty-liver-disease> (accessed 22 September 2017).
- Hollman, P. C. (2004). Absorption, bioavailability, and metabolism of flavonoids. *Pharmaceutical Biology*, 42(sup1), 74-83.
- Hou, F., Zhang, R., Zhang, M., Su, D., Wei, Z., Deng, Y., and Tang, X. (2013). Hepatoprotective and antioxidant activity of anthocyanins in black rice bran on carbon tetrachloride-induced liver injury in mice. *Journal of Functional Foods*, 5(4), 1705-1713.
- Hsiao, G., Shen, M. Y., Lin, K. H., Lan, M. H., Wu, L. Y., Chou, D. S., and Sheu, J. R. (2003). Antioxidative and hepatoprotective effects of *Antrodia camphorata* extract. *Journal of Agricultural and Food Chemistry*, 51(11), 3302-3308.
- Huang, B., Ban, X., He, J., Zeng, H., Zhang, P., and Wang, Y. (2010). Hepatoprotective and antioxidant effects of the methanolic extract from *Halenia elliptica*. *Journal of Ethnopharmacology*, 131(2), 276-281.
- Huang, D., Ou, B., Hampsch-Woodill, M., Flanagan, J. A., and Prior, R. L. (2002). High-throughput assay of oxygen radical absorbance capacity (ORAC) using a multichannel liquid handling system coupled with a microplate fluorescence reader in 96-well format. *Journal of Agricultural and Food Chemistry*, 50(16), 4437-4444.
- Hubert, D. J., Dawe, A., Florence, N. T., Gilbert, K. D., Angele, T. N., Buonocore, D., and Paul, M. F. (2011). In vitro hepatoprotective and antioxidant activities of crude extract and isolated compounds from *Ficus gnaphalocarpa*. *Inflammopharmacology*, 19(1), 35-43.

- Hung, O.L., and Nelson, L., (2004). *Tintinalli's Emergency Medicine: A Comprehensive Study Guide*, 6 ed. McGraw-Hill, New York.
- Husain, K., and Hazelrigg, S. R. (2002). Oxidative injury due to chronic nitric oxide synthase inhibition in rat: effect of regular exercise on the heart. *Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease*, 1587(1), 75-82.
- Hussaini, J., Othman, N. A., Abdulla, M. A., Majid, N. A., Farooq, H. M., and Ismail, S. (2012). Gastroprotective effects of *Dicranopteris linearis* leaf extract against ethanol-induced gastric mucosal injury in rats. *Scientific Research and Essays*, 7(18), 1761-1767.
- Ikhiri, K., Boureima, D., Dicko, D., and Koulodo, D. (1992). Chemical screening of medicinal plants used in the traditional pharmacopoeia of Niger. *International Journal of Pharmacognosy*, 30(4), 251-262.
- Ilahi, I., Khan, J., Ghaffar, R., Hussain, A., Rahman, K., Wahab, S., and Zeb, L. (2016). In vitro antioxidant and hepatoprotective activities of *Paeonia emodi* (Wall.) rhizome methanol extract and its phenolic compounds rich fractions. *Pakistan Journal of Pharmaceutical Science*, 29(5), 1787-1794.
- Irondi, E. A., Oboh, G., Agboola, S. O., Boligon, A. A., and Athayde, M. L. (2016). Phenolics extract of *Tetrapleura tetraptera* fruit inhibits xanthine oxidase and Fe<sup>2+</sup>-induced lipid peroxidation in the kidney, liver, and lungs tissues of rats in vitro. *Food Science and Human Wellness*, 5(1), 17-23.
- Jaeschke, H., Knight, T. R., and Bajt, M. L. (2003). The role of oxidant stress and reactive nitrogen species in acetaminophen hepatotoxicity. *Toxicology Letters*, 144(3), 279-288.
- Jaeschke, H., McGill, M. R., Williams, C. D., and Ramachandran, A. (2011). Current issues with acetaminophen hepatotoxicity—a clinically relevant model to test the efficacy of natural products. *Life Sciences*, 88(17), 737-745.
- Jain, A., Soni, M., Deb, L., Jain, A., Rout, S. P., Gupta, V. B., and Krishna, K. L. (2008). Antioxidant and hepatoprotective activity of ethanolic and aqueous extracts of *Momordica dioica* Roxb. leaves. *Journal of Ethnopharmacology*, 115(1), 61-66.
- James, L. P., Letzig, L., Simpson, P. M., Capparelli, E., Roberts, D. W., Hinson, J. A., and Lee, W. M. (2009). Pharmacokinetics of acetaminophen-protein adducts in adults with acetaminophen overdose and acute liver failure. *Drug Metabolism and Disposition*, 37(8), 1779-1784.
- James, L.P., McCullough, S.S., Knight, T.R., Jaeschke, H., Hinson, J.A., (2003). Acetaminophen toxicity in mice lacking NADPH oxidase activity: role of

peroxynitrite formation and mitochondrial oxidant stress. *Free Radical Research*. 37, 1289-1297.

- Kabera, J. N., Semana, E., Mussa, A. R., and He, X. (2014). Plant secondary metabolites: biosynthesis, classification, function and pharmacological properties. *Journal of Pharmacy and Pharmacology*, 2, 377-392.
- Kakarla, L., Mathi, P., Allu, P. R., Rama, C., and Botlagunta, M. (2014). Identification of human cyclooxygenase-2 inhibitors from *Cyperus scariosus* (R. Br) rhizomes. *Bioinformation*, 10(10), 637.
- Kalaisezhiyen, P., and Sasikumar, V. (2012). GC-MS evaluation of chemical constituents from methanolic leaf extract of *Kedrostis foetidissima* (Jacq.) Cogn. *Asian Journal of Pharmaceutical and Clinical Research*, 5(4), 77-81.
- Kalemci, S., Zeybek, A., Intepe, Y. S., Uner, A. G., Acar, T., Yaylali, A., and Sütçü, R. (2012). Methyl palmitate attenuates lipopolysaccharide-induced acute lung injury in mice. *La Clinica Terapeutica*, 164(6), e453-9.
- Kamiyama, T., Sato, C., Liu, J., Tajiri, K., Miyakawa, H., and Marumo, F. (1993). Role of lipid peroxidation in acetaminophen-induced hepatotoxicity: comparison with carbon tetrachloride. *Toxicology Letters*, 66(1), 7-12.
- Kanel, G.C, and Korula, .(2005). *Atlas of Liver Pathology*. 2<sup>nd</sup> ed. 1-6 USA: Elsevier Inc.
- Katsube, T., Tsurunaga, Y., Sugiyama, M., Furuno, T., and Yamasaki, Y. (2009). Effect of air-drying temperature on antioxidant capacity and stability of polyphenolic compounds in mulberry (*Morus alba* L.) leaves. *Food Chemistry*, 113(4), 964-969.
- Keays, R., Harrison, P. M., Wendon, J. A., Forbes, A., Gove, C., and Alexander, G. J. (1991). A prospective controlled trial of intravenous N-acetylcysteine in paracetamol-induced fulminant hepatic failure. *BMJ*, 303, 1024-1029.
- Kelly, G. S. (1998). Clinical applications of N-acetylcysteine. *Alternative medicine review: a journal of clinical therapeutic*, 3(2), 114-127.
- Khan, M. A., Ahmad, W., Ahmad, M., & Nisar, M. (2017). Hepatoprotective effect of the solvent extracts of *Viola canescens* Wall. ex. Roxb. against CCl<sub>4</sub> induced toxicity through antioxidant and membrane stabilizing activity. *BMC Complementary and Alternative Medicine*, 17(1), 10.
- Kim, Y., You, Y., Yoon, H. G., Lee, Y. H., Kim, K., Lee, J., and Jun, W. (2014). Hepatoprotective effects of fermented *Curcuma longa* L. on carbon tetrachloride-induced oxidative stress in rats. *Food Chemistry*, 151, 148-153.

- Klein, A. S., Hart, J., Brems, J. J., Goldstein, L., Lewin, K., and Busuttill, R. W. (1989). Amanita poisoning: treatment and the role of liver transplantation. *The American journal of Medicine*, 86(2), 187-193.
- Krishnaiah, D., Sarbatly, R., and Nithyanandam, R. (2011). A review of the antioxidant potential of medicinal plant species. *Food and Bioproducts Processing*, 89(3), 217-233.
- Kumar, S. V., Sanjeev, T., Ajay, S., Kumar, S. P., and Anil, S. (2012). A review on hepatoprotective activity of medicinal plants. *International Journal of Advanced Research in Pharmaceutical & Bio Sciences*, 1, 31-38.
- Kumar, V., Abbas, A. K., Fausto, N., and Mitchell, R. N. (2007). Robbins basic pathology. 8<sup>th</sup> Edition. *Saunders Elsevier*, 1- 32.
- Lai, H. Y., Lim, Y. Y., and Tan, S. P. (2009). Antioxidative, tyrosinase inhibiting and antibacterial activities of leaf extracts from medicinal ferns. *Bioscience, Biotechnology, and Biochemistry*, 73(6), 1362-1366.
- Laskin, D.L., (2009). Macrophages and inflammatory mediators in chemical toxicity: a battle offorces. *Chemical Research in Toxicology* .22,1376–1385.
- Li, X., Wang, Z. G., Chen, H. H., and Liu, S. G. (2014). The antioxidant methyl 3-(3, 5-di-tert-butyl-4-hydroxyphenyl) propionate. *Acta Crystallographica Section C: Structural Chemistry*, 70(11), 1050-1053.
- Lin, C. C., Huang, P. C., and Lin, J. M. (2000). Antioxidant and hepatoprotective effects of Anoectochilus formosanus and Gynostemma pentaphyllum. *The American Journal of Chinese Medicine*, 28(01), 87-96.
- Liu, F., Ooi, V. E. C., and Chang, S. T. (1997). Free radical scavenging activities of mushroom polysaccharide extracts. *Life Sciences*, 60(10), 763-771.
- Liu, Y. J., Du, J. L., Cao, L. P., Jia, R., Shen, Y. J., Zhao, C. Y., and Yin, G. J. (2015). Anti-inflammatory and hepatoprotective effects of Ganoderma lucidum polysaccharides on carbon tetrachloride-induced hepatocyte damage in common carp (Cyprinus carpio L.). *International Immunopharmacology*, 25(1), 112-120.
- Liu, Y. T., Lu, B. N., and Peng, J. Y. (2011). Hepatoprotective activity of the total flavonoids from Rosa laevigata Michx fruit in mice treated by paracetamol. *Food Chemistry*, 125(2), 719-725.
- Long, L. H., and Halliwell, B. (2001). Antioxidant and prooxidant abilities of foods and beverages. *Methods in Enzymology*, 335, 181-190.

- Lowry, O. H., Rosebrough, N. J., Farr, A. L., and Randall, R. J. (1951). Protein measurement with the Folin phenol reagent. *The Journal of Biological Chemistry*, 193(1), 265-275.
- Lu, B., Xu, Y., Xu, L., Cong, X., Yin, L., Li, H., and Peng, J. (2012). Mechanism investigation of dioscin against CCl<sub>4</sub>-induced acute liver damage in mice. *Environmental Toxicology and Pharmacology*, 34(2), 127-135.
- Luster, M. I., Simeonova, P. P., Gallucci, R. M., Matheson, J. M., and Yucelsoy, B. (2000). Immunotoxicology: role of inflammation in chemical-induced hepatotoxicity. *International Journal of Immunopharmacology*, 22(12), 1143-1147.
- MacDonald, I., Oghale, O. U., Ikechi, E. G., & Orji, O. A. (2016). Hepatoprotective potentials of *Picralima nitida* against in vivo carbon tetrachloride-mediated hepatotoxicity. *The Journal of Phytopharmacology*, 5(1): 6-9.
- Makni, M., Chtourou, Y., Fetoui, H., Garoui, E. M., Boudawara, T., & Zeghal, N. (2011). Evaluation of the antioxidant, anti-inflammatory and hepatoprotective properties of vanillin in carbon tetrachloride-treated rats. *European Journal of Pharmacology*, 668(1), 133-139.
- Makni, M., Fetoui, H., Gargouri, N.K., Garoui, M., Jaber, H., Makni, J., Boudawara, T., Zeghal, N., 2010. Hypolipidemic and hepatoprotective seeds mixture diet rich in  $\omega$ -3 and  $\omega$ -6 fatty acids. *Food and Chemical Toxicology*. 48, 2239–2246.
- Malik, A., Anis, I., Khan, S. B., Ahmed, E., Ahmed, Z., Nawaz, S. A., and Choudhary, M. I. (2004). Enzymes inhibiting lignans from *Vitex negundo*. *Chemical and Pharmaceutical Bulletin*, 52(11), 1269-1272.
- Mamat, S. S., Kamisan, F. H., Zainulddin, W. W., Ismail, N. A., Yahya, F., Din, S. S., and Zakaria, Z. A. (2013). Effect of methanol extract of *Dicranopteris linearis* leaves against paracetamol-and carbon tetrachloride (CCl<sub>4</sub>)-induced liver toxicity in rats. *Journal of Medicinal Plants Research*, 7(19), 1305-1309.
- Mantawy, E. M., Tadros, M. G., Awad, A. S., Hassan, D. A., and El-Demerdash, E. (2012). Insights antifibrotic mechanism of methyl palmitate: impact on nuclear factor kappa B and proinflammatory cytokines. *Toxicology and Applied Pharmacology*, 258(1), 134-144.
- Martínez-Clemente, M., Ferré, N., Titos, E., Horrillo, R., González-Pérez, A., Morán-Salvador, E., and Clària, J. (2010). Disruption of the 12/15-lipoxygenase gene (Alox15) protects hyperlipidemic mice from nonalcoholic fatty liver disease. *Hepatology*, 52(6), 1980-1991.

- Marzilawati, A. R., Ngau, Y. Y., and Mahadeva, S. (2012). Low rates of hepatotoxicity among Asian patients with paracetamol overdose: a review of 1024 cases. *BMC Pharmacology and Toxicology*, 13(1), 8.
- McComb, R. B., Bowers Jr, G. N., and Posen, S. (2013). *Alkaline phosphatase*. Springer Science & Business Media.
- McGill, M. R., Williams, C. D., Xie, Y., Ramachandran, A., and Jaeschke, H. (2012). Acetaminophen-induced liver injury in rats and mice: comparison of protein adducts, mitochondrial dysfunction, and oxidative stress in the mechanism of toxicity. *Toxicology and Applied Pharmacology*, 264(3), 387-394.
- Mihailović, V., Mihailović, M., Uskoković, A., Arambašić, J., Mišić, D., Stanković, V., and Matić, S. (2013). Hepatoprotective effects of *Gentiana asclepiadea* L. extracts against carbon tetrachloride induced liver injury in rats. *Food and Chemical Toxicology*, 52, 83-90.
- Mohamed, E. A. H., Lim, C. P., Ebrika, O. S., Asmawi, M. Z., Sadikun, A., and Yam, M. F. (2011). Toxicity evaluation of a standardised 50% ethanol extract of *Orthosiphon stamineus*. *Journal of Ethnopharmacology*, 133(2), 358-363.
- Mohammadian, A., Moradkhani, S., and Ataie, S. (2016). Antioxidative and hepatoprotective effects of hydroalcoholic extract of *Artemisia absinthium* L. in rat. *Journal of HerbMed Pharmacology*, 5(1), 29-32.
- Mukherjee, P. K. (2003). Plant products with hypocholesterolemic potentials. *Advances in Food and Nutrition Research*, 47, 277-338.
- Mukinda, J. T., & Syce, J. A. (2007). Acute and chronic toxicity of the aqueous extract of *Artemisia afra* in rodents. *Journal of Ethnopharmacology*, 112(1), 138-144.
- Murugan, R., and Parimelazhagan, T. (2014). Comparative evaluation of different extraction methods for antioxidant and anti-inflammatory properties from *Osbeckia parvifolia* Arn.—An in vitro approach. *Journal of King Saud University-Science*, 26(4), 267-275.
- Naik, S. R., and Panda, V. S. (2007). Antioxidant and hepatoprotective effects of *Ginkgo biloba* phytosomes in carbon tetrachloride-induced liver injury in rodents. *Liver International*, 27(3), 393-399.
- Negishi, J. N., Sidle, R. C., Noguchi, S., Nik, A. R., and Stanforth, R. (2006). Ecological roles of roadside fern (*Dicranopteris curranii*) on logging road recovery in Peninsular Malaysia: Preliminary results. *Forest Ecology and Management*, 224(1), 176-186.

- New South Wales Flora Online. <http://plantnet.rbgsyd.nsw.gov.au/cgi-bin/NSWfl.pl?page=nswfl&lvl=sp&name=Dicranopteris~linearis> accessed on 25<sup>th</sup> September 2017.
- Nithianantham, K., Shyamala, M., Chen, Y., Latha, L. Y., Jothy, S. L., and Sasidharan, S. (2011). Hepatoprotective potential of *Clitoria ternatea* leaf extract against paracetamol induced damage in mice. *Molecules*, *16*(12), 10134-10145.
- Oh, H., Kim, D. H., Cho, J. H., and Kim, Y. C. (2004). Hepatoprotective and free radical scavenging activities of phenolic petrosins and flavonoids isolated from *Equisetum arvense*. *Journal of Ethnopharmacology*, *95*(2), 421-424.
- Organization of Economic Co-Operation and Development (OECD), (2001). *Guideline for testing of chemicals, acute oral toxicity—acute toxicity class method. Tech. Rep. 423*, OECD,
- Orhan, I. E., Şener, B., and Musharraf, S. G. (2012). Antioxidant and hepatoprotective activity appraisal of four selected *Fumaria* species and their total phenol and flavonoid quantities. *Experimental and Toxicologic Pathology*, *64*(3), 205-209.
- Otsuka, H. (2005). Purification by Solvent Extraction Using Partition Coefficient. *Natural Products Isolation*, *20*, 269.
- Panteghini, M. (1990). Aspartate aminotransferase isoenzymes. *Clinical Biochemistry*, *23*(4), 311-319.
- Pareek, A., Godavarthi, A., Issarani, R., and Nagori, B. P. (2013). Antioxidant and hepatoprotective activity of *Fagonia schweinfurthii* (Hadidi) Hadidi extract in carbon tetrachloride induced hepatotoxicity in HepG2 cell line and rats. *Journal of Ethnopharmacology*, *150*(3), 973-981.
- Parmar, S. R., Vashrambhai, P. H., and Kalia, K. (2010). Hepatoprotective activity of some plants extract against paracetamol induced hepatotoxicity in rats. *Journal of Herbal Medicine and Toxicology*, *4*(2), 101-106.
- Patel, S. J., Milwid, J. M., King, K. R., Bohr, S., Iracheta-Vellve, A., Li, M., and Yarmush, M. L. (2012). Gap junction inhibition prevents drug-induced liver toxicity and fulminant hepatic failure. *Nature Biotechnology*, *30*(2), 179-183.
- Pithayanukul, P., Nithitanakool, S., and Bavovada, R. (2009). Hepatoprotective potential of extracts from seeds of *Areca catechu* and nutgalls of *Quercus infectoria*. *Molecules*, *14*(12), 4987-5000.
- Polson, J., and Lee, W. M. (2005). AASLD position paper: the management of acute liver failure. *Hepatology*, *41*(5), 1179-1197.

- Popović, M., Kaurinović, B., Trivić, S., Mimica-Dukić, N., and Bursać, M. (2006). Effect of celery (*Apium graveolens*) extracts on some biochemical parameters of oxidative stress in mice treated with carbon tetrachloride. *Phytotherapy Research*, 20(7), 531-537.
- Porchezian, E. and Ansari, S. H. (2005). Hepatoprotective activity of *Abutilon indicum* on experimental liver damage in rats. *Phytomedicine* 12: 62-64
- Post-White, J., Ladas, E. J., and Kelly, K. M. (2007). Advances in the use of milk thistle (*Silybum marianum*). *Integrative Cancer Therapies*, 6(2), 104-109.
- Prabakar, K., and Wembonyama, J. P. (2016). Phytochemical profiling of the aqueous leaf extracts of *Blepharis maderaspatensis* (L.) Heyne ex Roth its HPLC, GC-MS, and column chromatographic analysis. *Imperial Journal of Interdisciplinary Research*, 2(7).
- Pradhan, S. C., and Girish, C. (2006). Hepatoprotective herbal drug, silymarin from experimental pharmacology to clinical medicine. *Indian Journal of Medical Research*, 124(5), 491.
- Qua, C. S., and Goh, K. L. (2011). Liver cirrhosis in Malaysia: peculiar epidemiology in a multiracial Asian country. *Journal of Gastroenterology and Hepatology*, 26(8), 1333-1337.
- Rabelo, T. K., Guimarães, A. G., Oliveira, M. A., Gasparotto, J., Serafini, M. R., de Souza Araújo, A. A., and Gelain, D. P. (2016). Shikimic acid inhibits LPS-induced cellular pro-inflammatory cytokines and attenuates mechanical hyperalgesia in mice. *International Immunopharmacology*, 39, 97-105.
- Rabelo, T. K., Zeidán-Chuliá, F., Caregnato, F. F., Schnorr, C. E., Gasparotto, J., Serafini, M. R., and Gelain, D. P. (2015). In vitro neuroprotective effect of shikimic acid against hydrogen peroxide-induced oxidative stress. *Journal of Molecular Neuroscience*, 56(4), 956-965.
- Rackova, L., Oblozinsky, M., Kostalova, D., Kettmann, V., and Bezakova, L. (2007). Free radical scavenging activity and lipoxygenase inhibition of *Mahonia aquifolium* extract and isoquinoline alkaloids. *Journal of Inflammation*, 4(1), 15.
- Raja, D. P., Manickam, V. S., De Britto, A. J., Gopalakrishnan, S., Ushioda, T., Satoh, M., and Tanaka, N. (1995). Chemical and chemotaxonomical studies on *Dicranopteris* species. *Chemical and Pharmaceutical Bulletin*, 43(10), 1800-1803.
- Rajeh, M. A. B., Kwan, Y. P., Zakaria, Z., Latha, L. Y., Jothy, S. L., and Sasidharan, S. (2012). Acute toxicity impacts of *Euphorbia hirta* L extract on behavior, organs body weight index and histopathology of organs of the mice and *Artemia salina*. *Pharmacognosy Research*, 4(3), 170.



- Ramaiah, S. K. (2007). A toxicologist guide to the diagnostic interpretation of hepatic biochemical parameters. *Food and Chemical Toxicology*, 45(9), 1551-1557.
- Ranawat, L., Bhatt, J. and Patel, J. (2010). Hepatoprotective activity of ethanolic extracts of bark of *Zanthoxylum armatum* DC in CCl<sub>4</sub> induced hepatic damage in rats. *Journal of Ethnopharmacology* 127: 777-780.
- Reihani, S. F. S., and Azhar, M. E. (2012). Antioxidant activity and total phenolic content in aqueous extracts of selected traditional Malay salads (Ulam). *International Food Research Journal*, 19(4), 1439-1444.
- Rice-Evans, C., Halliwell, B., and Lunt, G. G. (1995). *Free Radicals and Oxidative Stress: Environment, Drugs and Food Additives*. Portland Press Ltd..
- Robbers, J. E., and Tyler, V. E. (1999). *Tyler's Herbs of Choice. The Therapeutic Use of Phytomedicinals*. Haworth Press Inc..
- Rodzi, R., Cheah, Y. L., Ooi, K. K., Othman, F., Mohtarrudin, N., Tohid, S. F., and Zakaria, Z. A. (2013). Chemopreventive potential of methanol extract of *Dicranopteris linearis* leaf on DMBA/croton oil-induced mouse skin carcinogenesis. *African Journal of Pharmacy and Pharmacology*, 7(35), 2484-2498.
- Sabir, S. M., and Rocha, J. B. T. (2008). Water-extractable phytochemicals from *Phyllanthus niruri* exhibit distinct in vitro antioxidant and in vivo hepatoprotective activity against paracetamol-induced liver damage in mice. *Food Chemistry*, 111(4), 845-851.
- Saliou, C., Rihn, B., Cillard, J., Okamoto, T., and Packer, L. (1998). Selective inhibition of NF- $\kappa$ B activation by the flavonoid hepatoprotector silymarin in HepG2. *FEBS Letters*, 440(1-2), 8-12.
- Saller, R., Meier, R., and Brignoli, R. (2001). The use of silymarin in the treatment of liver diseases. *Drugs*, 61(14), 2035-2063.
- Seeff, L. B., Lindsay, K. L., Bacon, B. R., Kresina, T. F., and Hoofnagle, J. H. (2001). Complementary and alternative medicine in chronic liver disease. *Hepatology*, 34(3), 595-603.
- Sharawy, M. H., El-Agamy, D. S., Shalaby, A. A., and Ammar, E. S. M. (2013). Protective effects of methyl palmitate against silica-induced pulmonary fibrosis in rats. *International Immunopharmacology*, 16(2), 191-198.
- Shyamal, S., Latha, P. G., Shine, V. J., Suja, S. R., Rajasekharan, S., and Devi, T. G. (2006). Hepatoprotective effects of *Pittosporum neelgherrense* Wight & Arn., a popular Indian ethnomedicine. *Journal of Ethnopharmacology*, 107(1), 151-155.

- Sies, H. (1993). Strategies of antioxidant defense. *The FEBS Journal*, 215(2), 213-219.
- Singh, D., Arya, P. V., Aggarwal, V. P., and Gupta, R. S. (2014). Evaluation of antioxidant and hepatoprotective activities of *Moringa oleifera* Lam. leaves in carbon tetrachloride-intoxicated rats. *Antioxidants*, 3(3), 569-591.
- Singh, R., Kumar, S., Rana, A. C., and Sharma, N. (2012). Different models of hepatotoxicity and related liver diseases: a review. *International Research Journal of Pharmacy*, 3(7), 86-95.
- Singleton, V. L., and Rossi, J. A. (1965). Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. *American Journal of Enology and Viticulture*, 16(3), 144-158.
- Singleton, V. L., Orthofer, R., and Lamuela-Raventós, R. M. (1999). [14] Analysis of total phenols and other oxidation substrates and antioxidants by means of folin-ciocalteu reagent. *Methods in Enzymology*, 299, 152-178.
- Somchit, M. N., Zuraini, A., Bustamam, A. A., Somchit, N., Sulaiman, M. R., and Noratunlina, R. (2005). Protective activity of turmeric (*Curcuma longa*) in paracetamol-induced hepatotoxicity in rats. *International Journal of Pharmacology*, 1, 252-256.
- Sowndhararajan, K. and Kang, S. C., (2013). Protective effect of ethyl acetate fraction of *Acacia ferruginea* DC. Against ethanol-induced gastric ulcer in rats. *Journal of Ethnopharmacology*, 148: 175-181.
- Stankovic, M. S. (2011). Total phenolic content, flavonoid concentration and antioxidant activity of *Marrubium peregrinum* L. extracts. *Kragujevac Journal of Sciences*, 33(2011), 63-72.
- StuartXchange Organisation. <http://www.stuartxchange.org/Kilob.html> accessed on 27th September 2017.
- Sundari, K., Karthik, D., Ilavenil, S., Kaleeswaran, B., Srigopalram, S., and Ravikumar, S. (2013). Hepatoprotective and proteomic mechanism of *Sphaeranthus indicus* in paracetamol induced hepatotoxicity in wistar rats. *Food Bioscience*, 1, 57-65.
- Talukdar, A. D., Tarafdar, R. G., Choudhury, M. D., Nath, D., and Choudhury, S. (2011). A review on pteridophyte antioxidants and their potential role in discovery of new drugs. *Assam University Journal of Science and Technology*, 7(1), 151-155.
- Thabrew, M. and Joice, P. (1987). *Planta Medica.*, 53: 239–241

- Thanabhorn, S., Jaijoy, K., Thamaree, S., Ingkaninan, K., and Panthong, A. (2006). Acute and subacute toxicity study of the ethanol extract from *Lonicera japonica* Thunb. *Journal of Ethnopharmacology*, 107(3), 370-373.
- Tosun, M., Ercisli, S., Sengul, M., Ozer, H., Polat, T., and Ozturk, E. (2009). Antioxidant properties and total phenolic content of eight *Salvia* species from Turkey. *Biological Research*, 42(2), 175-181.
- Tsimogiannis, D., Samiotaki, M., Panayotou, G., and Oreopoulou, V. (2007). Characterization of flavonoid subgroups and hydroxy substitution by HPLC-MS/MS. *Molecules*, 12(3), 593-606.
- Vasudeva, S. M. (1999). Economic importance of pteridophytes. *Indian Fern Journal*, 16(1-2), 130-152.
- Vaziri, N. D., Dicus, M., Ho, N. D., Boroujerdi-Rad, L., and Sindhu, R. K. (2003). Oxidative stress and dysregulation of superoxide dismutase and NADPH oxidase in renal insufficiency. *Kidney international*, 63(1), 179-185.
- Venkatakishore, T., Rao, M. P., Thulasi, B., Durga, P. K., Harikrishna, B., and Prasanth, S. S. (2016). Hepatoprotective activity of ethanolic extract of aerial part of *gymnema sylvestre* against ccl4 and paracetamol induced hepatotoxicity in rats. *World Journal of Pharmacy and Pharmaceutical Sciences*. 5(12): 1007-1016.
- Vilegas, W., Sanommiya, M., Rastrelli, L., and Pizza, C. (1999). Isolation and structure elucidation of two new flavonoid glycosides from the infusion of *Maytenus aquifolium* leaves. Evaluation of the antiulcer activity of the infusion. *Journal of Agricultural and Food Chemistry*, 47(2), 403-406.
- Vimala, S., Rohana, S., Rashih, A. A., and Juliza, M. (2012). Antioxidant evaluation in Malaysian medicinal plant: *Persicaria minor* (Huds.) leaf. *Science Journal of Medicine and Clinical Trials*, 2012.
- Wallace, J.L., (2004). Acetaminophen hepatotoxicity: NO to the rescue. *British Journal of Pharmacology*. 143, 1-2.
- White, C. P., Hirsch, G., Patel, S., Adams, F., and Peltekian, K. M. (2007). Complementary and alternative medicine use by patients chronically infected with hepatitis C virus. *Canadian Journal of Gastroenterology and Hepatology*, 21(9), 589-595.
- Wild Life of Hawaii. <https://wildlifeofhawaii.com/flowers/454/dicranopteris-linearis-old-world-forkedfern/> accessed on 25<sup>th</sup> September 2017.
- Wong, W. L., Abdulla, M. A., Chua, K. H., Kuppusamy, U. R., Tan, Y. S., and Sabaratnam, V. (2012). Hepatoprotective effects of *Panus giganteus* (Berk.)

Corner against thioacetamide-(TAA-) induced liver injury in rats. *Evidence-Based Complementary and Alternative Medicine*, 2012.

- Wu, Y., Yang, L., Wang, F., Wu, X., Zhou, C., Shi, S., and Zhao, Y. (2007). Hepatoprotective and antioxidative effects of total phenolics from *Laggetera pterodonta* on chemical-induced injury in primary cultured neonatal rat hepatocytes. *Food and Chemical toxicology*, 45(8), 1349-1355.
- Yadav, S., and Bhadoria, B. K. (2005). Two dimeric flavonoids from *Bauhinia purpurea*. *Indian Journal of Chemistry*. 44B:2604–2607.
- Yahya, F., Mamat, S. S., Kamarolzaman, M. F. F., Seyedan, A. A., Jakius, K. F., Mahmood, N. D., and Somchit, M. N. (2013). Hepatoprotective activity of methanolic extract of *Bauhinia purpurea* leaves against paracetamol-induced hepatic damage in rats. *Evidence-Based Complementary and Alternative Medicine*, 2013.
- Yang, C. C., Fang, J. Y., Hong, T. L., Wang, T. C., Zhou, Y. E., and Lin, T. C. (2013). Potential antioxidant properties and hepatoprotective effects of an aqueous extract formula derived from three Chinese medicinal herbs against CCl<sub>4</sub>-induced liver injury in rats. *International Immunopharmacology*, 15(1), 106-113.
- Yang, J. I. A. N., Gadi, R. A. M. A., and Thomson, T. A. L. E. N. E. (2011). Antioxidant capacity, total phenols, and ascorbic acid content of noni (*Morinda citrifolia*) fruits and leaves at various stages of maturity. *Micronesica*, 41(2), 167-176.
- Yapar, K., Kart, A., Karapehliyan, M., Atakisi, O., Tunca, R., Erginsoy, S., and Cital, M. (2007). Hepatoprotective effect of L-carnitine against acute acetaminophen toxicity in mice. *Experimental and Toxicologic Pathology*, 59(2), 121-128.
- Yayla, M., Halici, Z., Unal, B., Bayir, Y., Akpinar, E., and Gocer, F. (2014). Protective effect of Et-1 receptor antagonist bosentan on paracetamol induced acute liver toxicity in rats. *European Journal of Pharmacology*, 726, 87-95.
- Yen, F. L., Wu, T. H., Lin, L. T., and Lin, C. C. (2007). Hepatoprotective and antioxidant effects of *Cuscuta chinensis* against acetaminophen-induced hepatotoxicity in rats. *Journal of Ethnopharmacology*, 111(1), 123-128.
- Yeşilada, E., and Takaishi, Y. (1999). A saponin with anti-ulcerogenic effect from the flowers of *Spartium junceum*. *Phytochemistry*, 51(7), 903-908.
- Zafrani, E. S. (2004). Non-alcoholic fatty liver disease: an emerging pathological spectrum. *Virchows Archiv*, 444(1), 3-12.

- Zakaria, Z. A. (2007). Free radical scavenging activity of some plants available in Malaysia. *Iranian Journal of Pharmacology & Therapeutics*, 6(1), 87-91.
- Zakaria, Z. A., Ghani, Z. D. F. A., Nor, R. N. S. R. M., Gopalan, H. K., Sulaiman, M. R., Jais, A. M. M., ...and Ripin, J. (2008). Antinociceptive, anti-inflammatory, and antipyretic properties of an aqueous extract of *Dicranopteris linearis* leaves in experimental animal models. *Journal of Natural Medicines*, 62(2), 179-187.
- Zakaria, Z. A., Ghani, Z. D. F. A., Nor, R. N. S. R. M., Gopalan, H. K., Sulaiman, M. R., and Abdullah, F. C. (2006). Antinociceptive and anti-inflammatory activities of *Dicranopteris linearis* leaves chloroform extract in experimental animals. *Yakugaku Zasshi*, 126(11), 1197-1203.
- Zakaria, Z. A., Mohamed, A. M., Jamil, N. M., Rofiee, M. S., Somchit, M. N., Zuraini, A., ... and Sulaiman, M. R. (2011). In vitro cytotoxic and antioxidant properties of the aqueous, chloroform and methanol extracts of *Dicranopteris linearis* leaves. *African journal of Biotechnology*, 10(2), 273-282.
- Zakaria, Z. A., Rahim, A., Hafiz, M., Mohtarrudin, N., Kadir, A. A., Cheema, M. S., and Tohid, S. F. M. (2016a). Acute and sub-chronic oral toxicity studies of methanol extract of *clinacanthus nutans* in mice. *African Journal of Traditional, Complementary & Alternative Medicines*, 13(2).
- Zakaria, Z. A., Yahya, F., Mamat, S. S., Mahmood, N. D., Mohtarrudin, N., Taher, M., and Salleh, M. Z. (2016b). Hepatoprotective action of various partitions of methanol extract of *Bauhinia purpurea* leaves against paracetamol-induced liver toxicity: involvement of the antioxidant mechanisms. *BMC Complementary and Alternative Medicine*, 16(1), 175.
- Zhu, R., Wang, Y., Zhang, L., and Guo, Q. (2012). Oxidative stress and liver disease. *Hepatology Research*, 42(8), 741-749.