



**UNIVERSITI PUTRA MALAYSIA**

***ANTI-ATHEROTHROMBOTIC EFFECTS OF *Berberis vulgaris* L.,  
*Teucrium polium* L. AND *Orthosiphon stamineus* BENTH EXTRACTS In  
Vitro AND *Berberis vulgaris* L. EXTRACTS In Vivo***

**NURUL HUDA MOHD NOR**

**FPSK(P) 2017 31**



**ANTI-ATHEROTHROMBOTIC EFFECTS OF *Berberis vulgaris* L.,  
*Teucrium polium* L. AND *Orthosiphon stamineus* BENTH EXTRACTS *In*  
*Vitro* AND *Berberis vulgaris* L. EXTRACTS *In Vivo***

By

**NURUL HUDA MOHD NOR**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfilment of Requirements for the Degree of Doctor of Philosophy**

**July 2017**

## **COPYRIGHT**

All material contained within the thesis, including without text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the Degree of Doctor of Philosophy

***ANTI-ATHEROTHROMBOTIC EFFECTS OF *Berberis vulgaris* L., *Teucrium polium* L. AND *Orthosiphon stamineus* BENTH EXTRACTS In Vitro AND *Berberis vulgaris* L. EXTRACTS In Vivo***

By

**NURUL HUDA MOHD NOR**

**July 2017**

**Chairman : Prof. Fauziah Othman, PhD**  
**Faculty : Medicine and Health Sciences**

Coronary artery disease is a group of diseases that includes stable angina, unstable angina, myocardial infarction and sudden cardiac death. Coronary artery disease is a leading cause of mortality and morbidity worldwide. The pathogenesis mainly due to atherosclerosis, plaque rupture and platelet thrombus formation. The main risk factors for coronary artery diseases are obesity, hypercholesterolemia, smoking, diabetes and high blood pressure. As part of disease management, anticoagulant and antiplatelet drugs are the options of treatment together with lipid-lowering medication.

In combating this disease, medicinal plants comprised of anti-atherothrombotic effects can be options other than drug therapies that may be considered to have lesser adverse effects. Nevertheless, the effectiveness of medicinal plants in treating coronary atherothrombotic disease has yet to be fully explored. Therefore, the haematological, biochemical, gross and histological effect of *Berberis vulgaris* L, *Teucrium polium* Land *Orthosiphon stamineus* Benth extracts in preventing and treating coronary atherothrombotic disease were studied at the *in vitro* (phase I) and *in vivo* (phase II) level.

In phase I, three types of extraction including aqueous, methanol and polysaccharide of the *B. vulgaris*, *T. polium* and *O. stamineus* were studied for antiplatelet and anticoagulant effect using human whole blood. All extracts were subjected to the prothrombin time (PT) and activated partial thromboplastin time (APTT) test for anticoagulant activity and then investigated using an electrical impedance method for antiplatelet activity. *B. vulgaris* aqueous extract (BVAE), *B. vulgaris* polysaccharide extract (BVPE) *T. polium* aqueous extract (TPAE) and *T. polium* polysaccharide extract (TPPE) were found to significantly prolong the coagulation time in a concentration dependent manner ( $p < 0.05$ ). In addition, phytochemical screening revealed that aqueous extract of all three medicinal plants contained polysaccharides. Hence, it was

concluded that the present of polysaccharides in the medicinal plants, especially in *B. vulgaris* and *T. polium* but not in *O. stamineus* played a role in prolonging blood clotting in coagulation activity. However, this current study on also proved that not all polysaccharides gave equal effects on coagulation test. Though, for antiplatelet activity, the BVAE was the most effective sample against platelet aggregation caused by arachidonic acid (AA) and collagen. These effects are probably due to the present of berberine content in *B. vulgaris* and higher total phenolic compound thus inhibit platelet aggregation activity.

In continuation of result in phase I, BVAE was proceeded for phase II (*in vivo*) due to its optimum effect of anticoagulation and antiplatelet activity. Phase II was divided into two parts, preventive and treatment studies. Atherosclerotic-induced male New Zealand white rabbits were divided into nine groups and antihyperlipidemic effect, anti-inflammatory effect, antiplatelet properties, anticoagulation properties, liver function test and renal profile were studied. After the treatment period for preventive study (10 weeks) and treatment study (12 weeks) were completed, the rabbits were sacrificed for gross and histological analyses (H&E, Masson trichrome and Modified Verhoeff stains), immunohistochemistry analyses (RAM 11) and biochemistry analyses were performed on extracts and control groups in both studies. All data were analysed using one-way ANOVA followed by LSD's post-hoc test. The values were considered significant when  $p$  value is less than 0.05.

The current study provided biochemical and histological evidences that BVAE possesses antihyperlipidemic effect through significant reduction on total cholesterol, triglyceride and low-density lipoproteins levels, and histologically for intima-media ratio and collagen score compared to control ( $p < 0.05$ ). For the anti-inflammatory effect, BVAE was shown to attenuate inflammatory cells biochemically including TNF-alpha and interleukin-6 levels, and significant reduction on macrophages cells number in concentration dependent manner ( $p < 0.05$ ).

Toxicity analysis showed both concentrations of the extracts had no effect on the liver and renal function biochemically and histologically These finding provide an important evidence that BVAE is safe to use, and possible to proceed for further testing in a clinical setting. In conclusion, *B. vulgaris* had been proven to have anti-atherothrombotic properties and can be considered as a safe option for anti-atherothrombotic agent.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**KESAN-KESAN ANTI-ATHEROTHROMBOSIS DALAM EKSTRAK *Berberis vulgaris* L., *Teucrium polium* L. DAN *Orthosiphon stamineus* BENTH KE ATAS MODEL *In Vitro* DAN EKSTRAK *Berberis vulgaris* L. KE ATAS MODEL *In Vivo***

Oleh

**NURUL HUDA MOHD NOR**

**Julai 2017**

**Pengerusi : Prof. Fauziah Othman, PhD**  
**Fakulti : Perubatan dan Sains Kesihatan**

Penyakit koronari arteri adalah penyakit jantung termasuklah angina yang stabil, angina tidak stabil dan infarksi miokardium. Penyakit koronari arteri adalah penyebab utama kematian dan morbiditi di seluruh dunia. Patogenesis utamanya adalah disebabkan oleh aterosklerosis plak dan pembentukan thrombus platelet. Faktor-faktor utama bagi penyakit atherothrombotik koronari adalah obesiti, hyperkolesterolemia, merokok, diabetik dan tekanan darah tinggi. Antikoagulan dan antiplatelet adalah pilihan rawatan untuk penyakit ini bersama-sama dengan ubat kolesterol.

Dalam memerangi penyakit ini, tumbuh-tumbuhan perubatan yang mempunyai kesan anti-atherothrombotic boleh menjadi pilihan selain daripada rawatan yang mempunyai kesan sampingan yang lebih rendah. Walau bagaimanapun, keberkesanan tumbuhan dalam merawat penyakit atherotrombotik koronari masih belum diterokai sepenuhnya. Oleh itu, kesan hematologi, biokimia, makroskopik dan histologi ke atas *Berberis vulgaris* L, *Teucrium polium* L dan *Orthosiphon stamineus* Benth ekstrak dalam mencegah dan merawat penyakit atherotrombotik koronari dikaji di peringkat *in vitro* (fasa I) dan *in vivo* (fasa II).

Dalam fasa I, tiga jenis ekstrak termasuk air, metanol dan polisakarida dari *B. vulgaris*, *T. polium* dan *O. stamineus* telah dikaji untuk kesan antiplatelet dan antikoagulan menggunakan sampel darah manusia. Semua ekstrak telah dikaji untuk aktiviti antikoagulan melalui ujian masa prothrombin (PT) dan masa tromboplastin (APTT) dan juga menggunakan kaedah impedans elektrik untuk aktiviti antiplatelet. Ekstrak air *B. vulgaris* (BVAE), ekstrak polisakarida *B. vulgaris* (BVPE) ekstrak air *T. polium* (TPAE) dan ekstrak polisakarida *T. polium* (TPPE) didapati memanjangkan masa pembekuan mengikut kepekatan ( $p < 0.05$ ). Di samping itu, pemeriksaan phytokimia juga mendedahkan bahawa ekstrak air dari ketiga-tiga tumbuhan ubatan ini

mengandung polisakarida. Oleh itu, disimpulkan bahawa polisakarida dalam tumbuhan perubatan, terutama di *B. vulgaris* dan *T. polium* tetapi tidak pada *O. stamineus*, memainkan peranan dalam memanjangkan pembekuan darah dalam aktiviti pembekuan. Walau bagaimanapun, kajian semasa ini juga membuktikan bahawa tidak semua polisakarida memberi kesan yang sama pada ujian koagulasi. Untuk aktiviti antiplatelet pula, BVAE adalah sampel yang paling berkesan terhadap agregasi platelet yang disebabkan oleh asid arakidonik (AA) dan kolagen. Kesan-kesan ini mungkin disebabkan oleh kandungan berberine dalam *B. vulgaris* dan sebatian fenolik yang lebih tinggi sekali gus menghalang aktiviti agregasi platelet.

Kesinambungan dengan keputusan dalam fasa I, BVAE diteruskan untuk fasa II (*in vivo*) disebabkan oleh kesan optimum aktiviti antikoagulasi dan antiplatelet. Fasa II dibahagikan kepada dua bahagian, kajian pencegahan dan kajian rawatan. Arnab jantan jenis New Zealand yang bewarna putih diaruhkan kepada arterosklerosis dan kesan antihyperlipidemic, kesan anti-radang, antiplatelet, antikoagulasi, ujian fungsi hati dan profil renal dikaji. Selepas tempoh rawatan untuk kajian pencegahan (10 minggu) dan kajian rawatan (12 minggu) telah selesai, arnab dikorbankan untuk analisis makroskopik dan histologi (H&E, Masson trichrome dan Modified Verhoeff noda), analisis imunohistokimia (RAM 11) dan analisis biokimia dilakukan pada ekstrak dan kumpulan kawalan dalam kedua-dua kajian. Semua data dianalisis dengan menggunakan satu arah ANOVA diikuti oleh ujian post-hoc LSD. Nilai dianggap bererti apabila nilai  $p$  kurang dari 0.05.

Kajian semasa menyediakan bukti biokimia bahawa BVAE mempunyai kesan antihyperlipidemic melalui pengurangan jumlah kolesterol, trigliserida dan tahap lipoprotein berketumpatan rendah, dan secara histologi untuk nisbah intima-media dan skor kolagen berbanding dengan kawalan ( $p < 0.05$ ). Untuk kesan anti-radang, BVAE ditunjukkan untuk menurunkan sel-sel radang secara biokimia termasuk tahap TNF-alpha dan interleukin-6, dan secara histologi melalui pengurangan ketara pada jumlah sel makrofaj mengikut kepekatan ( $p < 0.05$ ).

Analisis toksik menunjukkan kedua-dua kepekatan ekstrak tidak memberi kesan ke atas fungsi hati dan buah pinggang secara biokimia dan histologikal. Penemuan ini memberikan bukti penting bahawa BVAE adalah selamat untuk digunakan dan dilanjutkan ke ujian secara klinikal. Kesimpulannya, *B. vulgaris* telah terbukti mempunyai sifat anti-atherotrombotik dan dapat dianggap sebagai pilihan yang selamat untuk agen anti-atherotrombotik.

## ACKNOWLEDGEMENTS

In the name of Allah, the Most Gracious, the Most Merciful. Alhamdulillah, thanks to Allah for giving me this opportunity to complete my PhD study. There are many people to whom I am most grateful for their thoughtfulness in helping and supporting me throughout the journey.

Firstly, I would like to express my sincere gratitude to all my supervisors, for your guidance, motivation and encouragement. I am so thankful to my main supervisor; Professor Fauziah Othman for enlightening me with hope and knowledge to make my journey toward the end of my PhD achievable. Special thanks to my co-supervisors; Dr Sabariah Md Noor for her motherly help and professional guidance and Dr Eusni Mohd Tohit, for wise advice and always believed in me.

Secondly, I would like to thank all my lab mates and staffs in Anatomy and Histology laboratory, Haematology laboratory and Chemistry Pathology laboratory for limitless intellectual and emotional support. Also thanks to my wonderful officemates for cheering and making my life in UPM remarkable. It was a real pleasure to get to know and spend time with you during my training and lab works.

An acknowledgement to Dr Halijah from MARDI, Dr Khor Kuan Hua from Faculty of Veterinary Medicine UPM, Pn Rosniza from Malaysian Institute of Nuclear Technology for allowing and guiding me on using facilities for sample analysis and test.

Thank you to my beloved husband, Muhammad Azlan Muad for his countless help, encouragement and support throughout my PhD journey. He has always been there for me, especially in taking care of our children, accompanying me during conference and helping hand during animal study. Special thanks go to my family, especially my parents and parents-in-law for the love, care and prayers. To Haniza, Rosfayati, Melati, Azuin, Liyana, Rozaini, Raihanah, Salma, Husna, Zalinda, Zalina, Baizura, Noraini, Fatin and Ely, I'm really blessed to be part of your life.

Last but not least, I thank to everyone, although not individually named here, who had contributed directly or indirectly to my project and thesis.



I certify that a Thesis Examination Committee has met on 24 July 2017 to conduct the final examination of Nurul Huda binti Mohd Nor on her thesis entitled "Anti-Atherothrombotic Effects of *Berberis vulgaris* L., *Teucrium polium* L. and *Orthosiphon stamineus* Benth Extracts *In Vitro* and *Berberis vulgaris* L. Extracts *In Vivo*" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

**Rozi binti Mahmud, PhD**

Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Chairman)

**Patimah binti Ismail, PhD**

Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Internal Examiner)

**Zainul Amiruddin bin Zakaria, PhD**

Associate Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Internal Examiner)

**Mary Jane Black, PhD**

Professor  
Monash University  
Australia  
(External Examiner)



---

**NOR AINI AB. SHUKOR, PhD**  
Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 28 September 2017

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

**Fauziah Othman, PhD**

Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Chairman)

**Sabariah Md Noor, MPath**

Doctor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Member)

**Eusni Rahayu Mohd Tohit, MPath**

Doctor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Member)

---

**ROBIAH BINTI YUNUS, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rule 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice- Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No.: Nurul Huda Binti Mohd Nor / GS 36563

## Declaration by Members of Supervisory Committee

This is to confirm that:

- The research conducted and the writing of this thesis was under our supervision;
- Supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: \_\_\_\_\_

Name of Chairman of

Supervisory Committee: Prof. Dr. Fauziah Othman

Signature: \_\_\_\_\_

Name of Member of

Supervisory Committee: Dr. Sabariah Md Noor

Signature: \_\_\_\_\_

Name of Member of

Supervisory Committee: Dr. Eusni Rahayu Mohd Tohit

## TABLE OF CONTENTS

	<b>Page</b>
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENTS</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	viii
<b>LIST OF TABLES</b>	xiv
<b>LIST OF FIGURES</b>	xv
<b>LIST OF ABBREVIATIONS</b>	xix
<b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	<b>1</b>
1.1 Introduction	1
1.2 Research Hypothesis	2
1.3 Research Objectives	3
1.3.1 General Objective	3
1.3.2 Specific Objectives	3
<b>2 LITERATURE REVIEW</b>	<b>4</b>
2.1 Coronary Artery Disease	4
2.2 History and Incidence of Atherothrombosis	4
2.3 Atherothrombotic Complication	5
2.3.1 Chronic Ischaemia	6
2.3.2 Acute Ischaemic Condition	6
2.4 Risk Factor for Atherothrombosis	7
2.4.1 Major Risk Factors of Atherothrombosis	7
2.4.2 Minor Risk Factors of Atherosclerosis	9
2.5 Pathogenesis of Atherothrombosis and Coronary Artery Diseases	10
2.5.1 Endothelial Dysfunction	10
2.5.2 Inflammation	11
2.5.3 Foam Cell Formation	11
2.5.4 Vascular Smooth Muscle Cell Migration and Proliferation	12
2.5.5 Platelet: A Necessary Mediator in Atherosclerosis	12
2.5.6 The Atheromatous Plaque Rupture and Thrombus Formation	12
2.6 Morphology Classification of Atherothrombotic Lesion	13
2.7 Interaction Between Platelets and The Coagulation System	14
2.7.1 Primary Haemostasis	15
2.7.2 Vasoconstriction, Vasodilatation and Platelet Activating Agents	17
2.7.3 Secondary Haemostasis	18
2.8 Treatment for Atherothrombotic Complications	19
2.8.1 Antiplatelet Drugs	19
2.8.2 Anticoagulation Drugs	21
2.9 Medicinal Herbs in Anti-atherothrombotic Study	22
2.10 Berberis vulgaris L	23
2.11 Teucrium polium L	25

2.12	Orthosiphon stamineus Benth	26
2.13	Animal Model in Atherothrombosis Study	28
2.14	Histological Examination	28
	2.14.1 Masson's Trichrome Staining	28
	2.14.2 Modified Verhoeff Staining	29
	2.14.3 Immunohistochemistry	29
2.15	Haematological Investigations	30
	2.15.1 Platelet Function Test	30
	2.15.2 Basic Coagulation Test	30
2.16	Enzyme Linked Immunosorbent Assay	31

<b>3</b>	<b>MATERIALS AND METHODS</b>	<b>32</b>
3.1	Plant Material	32
	3.1.1 Preparation of Plants Crude Methanol Extract (BVME, TPME and OSME)	32
	3.1.2 Preparation of Plants Crude Polysaccharide Extracts (BVPE, TPPE and OSPE)	32
	3.1.3 Preparation of Plants Crude Aqueous Extract (BVAE, TPAE and OSAE)	33
3.2	Anti-Atherothrombosis effect of BV, OS and TP in <i>in vitro</i> Study (Phase I)	33
	3.2.1 The Subjects Studied	34
	3.2.2 Preparation of Blood Sample	35
	3.2.3 Coagulation Test	35
	3.2.4 Platelet Aggregation Test	36
3.3	Experimental Design for Phase II ( <i>in vivo study</i> )	37
	3.3.1 Chemical and Apparatus	39
	3.3.2 Phytochemical Screening for Carbohydrate	39
	3.3.3 Monosaccharide Content Analysis	39
	3.3.4 Animal Studies	40
	3.3.5 Induction of Atherosclerosis	40
3.4	Anti-Atherothrombosis Properties of BVAE ( <i>in vivo</i> )	41
	3.4.1 Approach 1: Preventive Study	41
	3.4.2 Approach 2: Treatment Study	41
3.5	Measurement of Biochemical Analysis	42
	3.5.1 Measurement of Fasting Serum Lipid	42
	3.5.2 Liver Function Test (LFTs)	43
	3.5.3 Renal Profile (RP)	43
	3.5.4 ELISA	43
3.6	Measurement of Haematology Parameters	43
	3.6.1 Full Blood Count	44
	3.6.2 Platelet Function Test	44
	3.6.3 Coagulation Function Test	44
3.7	Collection of Organ Samples	45
	3.7.1 Macroscopic Study	45
	3.7.2 Histological Staining	45
	3.7.3 Immunohistochemistry Analysis	46
3.8	Scoring	47
	3.8.1 Intima-Media Ratio	47
	3.8.2 Numerical Scoring for Histological Analysis	47
	3.8.3 Numerical Scoring for Assessing Toxicity	47

3.8.4	Scoring for Immunohistochemistry Staining of RAM	48
	11	
3.9	Statistical Analysis	49
<b>4</b>	<b>RESULTS</b>	<b>50</b>
4.1	<i>In-vitro</i> Anti-Atherothrombosis Study (Phase I)	50
4.1.1	Baseline FBC in Human Blood Sample	50
4.1.2	Anticoagulation Effect (PT and APTT) of BV, TP and OS Using Aqueous, Methanol and Polysaccharide Crude Extracts	50
4.1.3	Antiplatelet Effects of BVAE, BVPE, TPAE and TPPE <i>In Vitro</i>	57
4.2	Phytochemical Screening for Carbohydrate	58
4.2.1	Monosaccharide Content Analysis	58
4.3	Anti-Atherothrombosis Effects of BVAE in Preventive Study (Phase II)	62
4.3.1	Body Weight Profile of Rabbits	62
4.4	Antihyperlipidemic Effects of BVAE in Preventive Study	63
4.4.1	Effects of BVAE on Cholesterol (TC) and Triglyceride (TG) Level	63
4.5	Histomorphometric Analysis and Macroscopic Observation of Aorta in Preventive Study	65
4.6	Microscopic Analysis of Aorta in Preventive Study	67
4.6.1	Haematoxylin and Eosin Staining of Aorta	67
4.6.2	Masson's Trichrome Staining of Aorta	70
4.6.3	Modified Verhoeff Staining of Aorta	74
4.7	Anti-Inflammatory Effect of BVAE in Preventive Study	77
4.7.1	Serum Tumour Necrosis Factor Alpha	77
4.7.2	Serum Interleukin-6	77
4.7.3	Immunohistochemistry	78
4.8	Full Blood Count in Preventive Study	82
4.9	Antiplatelet Properties	82
4.10	Anticoagulant Properties	83
4.11	Toxicity Analysis of BVAE in Preventive Study	85
4.11.1	Liver Function Tests	85
4.11.2	Renal Profile	85
4.11.3	Mean Lesion Scoring for Kidney	87
4.11.4	Mean Lesion Scoring for Liver	89
4.12	<i>In vivo</i> Anti Atherothrombosis Effects in Treatment Study (Phase II)	91
4.12.1	Body Weight Profile of Rabbits	91
4.13	Antihyperlipidemic Effects of BVAE in Treatment Groups	92
4.13.1	Fasting Lipid Profile	92
4.14	Histomorphometric Analysis and Macroscopic Observation of Aorta in Treatment Study	92
4.15	Microscopic analysis of Aorta in Treatment Groups	95
4.15.1	Haematoxylin and Eosin Staining of Aorta	95
4.15.2	Masson's Trichrome Staining of Aorta	98
4.15.3	Modified Verhoeff Staining of Aorta	102
4.16	Anti-Inflammatory Effects of BVAE in Treatment Study	105
4.16.1	Serum Tumour Necrosis Factor Alpha	105

4.16.2	Serum Interleukin-6	106
4.16.3	Immunohistochemistry Finding in Treatment Groups	107
4.17	Haematological Parameter Effects of BVAE in Treatment Groups	111
4.17.1	Full Blood Count	111
4.18	Antiplatelet Properties	112
4.19	Anticoagulant Properties	113
<b>5</b>	<b>DISCUSSION</b>	<b>115</b>
5.1	Different Methods of Plants Extraction	115
5.2	In-vitro Study	116
5.2.1	Anticoagulation Effect of BV, TP and OS Plants Crude Extracts	116
5.2.2	Antiplatelet Effects of BV and TP Crude Extracts	117
5.3	<i>In vivo</i> Study	118
5.3.1	Polysaccharide Compound of BVAE	118
5.3.2	Antihyperlipidemic Effects of BVAE in Preventive and Treatment Study	119
5.3.3	Anti-inflammatory Properties in Preventive and Treatment Studies	120
5.3.4	Antiplatelet Effect in Preventive and Treatment Studies	121
5.3.5	Anticoagulation Properties in Preventive and Treatment Studies	123
5.4	Histology Evaluation of Preventive and Treatment Studies	124
5.5	Immunohistochemical Evaluation in Preventive and Treatment Studies	126
5.6	Toxicity Analysis in Preventive and Treatment Studies	126
<b>6</b>	<b>CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH</b>	<b>128</b>
6.1	Conclusion	128
6.2	Recommendation for Future Research	129
	<b>REFERENCES</b>	<b>130</b>
	<b>APPENDICES</b>	<b>150</b>
	<b>BIODATA OF STUDENT</b>	<b>174</b>
	<b>LIST OF PUBLICATIONS</b>	<b>175</b>



## LIST OF TABLES

Table		Page
3.1	The Preventive Study	41
3.2	The Treatment Study	42
3.3	Lesion Scoring for Liver and Kidney	48
3.4	Score for Marker RAM 11 Using Immunohistochemistry Techniques	49
4.1	Mean Values of Main Parameter of Full Blood Count of Human Blood For In Vitro Study	50
4.2	Inhibition of Platelet Aggregation (%) in Human Whole Blood with BVPE, BVAE, TPPE and TPPE using Platelet Agonists of Collagen (2 µg/ml), ADP (10 µM) and AA (0.5 µM).	57
4.3	Effect of BVAE on Cholesterol and Triglyceride Levels	64
4.4	Effects of <i>B. vulgaris</i> on Haemoglobin, White Cell Count and Platelet of Rabbits as Compared to Control and Cholesterol Treated Groups.	82
4.5	The Effects of BVAE on Liver Function Enzymes of Preventive Study.	85
4.6	Effect of BVAE 50 and Simvastatin on Lipid Profile.	92
4.7	Effects of <i>B. vulgaris</i> on Haemoglobin, White Cell Count and Platelet of Rabbits as Compared to Control and Cholesterol Treated Groups.	112

## LIST OF FIGURES

Figure		Page
2.1	Major Clinical Manifestation of Atherothrombosis Disease.	5
2.2	Pathogenesis of Atherothrombosis.	10
2.3	A Modified AHA Lesion Morphology Classification of Coronary Atherosclerosis According to Gross Pathological and Clinical Finding.	13
2.4	The Platelet Activation Pathway and Coagulation Cascade.	15
2.5	Primary Haemostasis; The Platelet Response.	17
2.6	Secondary Haemostasis; The Coagulation Cascade.	18
2.7	Flower, Fruits and Tree of <i>B. vulgaris</i> .	24
2.8	Tree and Dried Flower of <i>T. polium</i> .	25
2.9	Tree and Dried Leaves of <i>O. stamineus</i> .	27
3.1	Experimental Design of Study for Phase I (in vitro)	34
3.2	Experimental Design of Study for Phase II (in vivo).	38
4.1	Effects of BVPE, TPPE and OSPE on Prothrombin Time (seconds) of Human Plasma.	51
4.2	Effects of BVME, TPME and OSME on Prothrombin Time (seconds) of Human Plasma.	52
4.3	Effects of BVAE, TPAE and OSAE on Prothrombin Time (seconds) of Human Plasma.	53
4.4	Effects of BVPE, TPPE and OSPE on Activated Partial Thromboplastin Time in seconds of Human Plasma.	54
4.5	Effects of BVME, TPME and OSME on Activated Partial Thromboplastin Time (seconds) of Human Plasma.	55
4.6	Effect of BVAE, TPAE and OSAE on Activated Partial Thromboplastin Time in seconds of Human Plasma.	56
4.7	LC-MS/MS Chromatograms Showing Mannose in BVAE.	59
4.8	LC-MS/MS Chromatograms Showing Glucose in BVAE	60
4.9	LC-MS/MS Chromatograms Showing Galactose in BVAE	61

4.10	Body Weight Profiles in Atherosclerosis induced Rabbits as Compared to Control Group in Preventive Study.	62
4.11	Histomorphometric Analysis of Intimal Lesion Area.	65
4.12	The Macroscopic Observation of The Atheromatous Plaque Formation on The Luminal Surface of The Descending of Aorta Stained with Sudan IV.	66
4.13	Intima-media Ratio of Aorta in Preventive Study.	67
4.14	Photomicrograph of Intimal Hyperplasia of Arch of Aorta.	70
4.15	The Mean Value of Collagen Score in Aorta Stained with Masson's Trichrome in Area (%).	71
4.16	Photomicrograph of Aorta from Normal Rabbit Group.	72
4.17	Photomicrograph of Aorta from Control Rabbit Group.	73
4.18	Photomicrograph of Aorta from BVAE 25 mg/kg Rabbit Group.	73
4.19	Photomicrograph of Aorta from BVAE 50 mg/kg Rabbit Group.	74
4.20	Photomicrograph of Aorta from Normal Rabbit Group.	75
4.21	Photomicrograph of Aorta from Control Rabbit Group.	75
4.22	Photomicrograph of Aorta from BVAE 25 mg/kg Rabbit Group.	76
4.23	Photomicrograph of Aorta from BVAE 50 mg/kg Rabbit Group.	76
4.24	Effect of BVAE on Serum TNF- $\alpha$ of Rabbits.	77
4.25	Effect of BVAE on Serum IL-6 of Rabbits.	78
4.26	Effect of BVAE on Mean RAM II Scoring of Aorta in Preventive Study.	79
4.27	Photomicrograph of Aorta in Normal Rabbit Group.	80
4.28	Photomicrograph of Macrophages in Fibrous Plaque in Aorta of Control Rabbit Group.	80
4.29	Photomicrograph of Macrophages in An Early Fibrous Plaque in Aorta of BVAE 25 mg/kg Rabbit Group.	81
4.30	Photomicrograph of Macrophages in An Early Fibrous Plaque in Aorta of BVAE 50 mg/kg Rabbit Group.	81

4.31	Inhibition of Platelet Aggregation (%) on Rabbit Whole Blood by BVAE Using Platelet Agonists of Collagen (2µg/mL).	83
4.32	Effects of BVAE on Activated Partial Thromboplastin Time (seconds).	84
4.33	The Effects of BVAE on Urea of Atherosclerotic-induced Rabbits in Preventive Study.	86
4.34	The Effects of BVAE on Creatinine Level of Atherosclerotic-induced Rabbits in Preventive Study.	86
4.35	Photomicrographs of Renal Corpuscle in Kidney in Preventive Study.	88
4.36	Photomicrograph of Portal Triad in Liver in Preventive Study.	90
4.37	The Body Weight Profile of Atherosclerotic-induced Rabbits as Compared to Control Group in Treatment Study.	91
4.38	Histomorphometric Analysis of Intimal Lesion Area in Treatment Study.	93
4.39	Macroscopic Observation of The Atheromatous Plaque Formation on The Luminal Surface of the Descending Aorta Stained with Sudan IV in Treatment Study.	94
4.40	Intima-media Ratios of Rabbits in Treatment Groups.	95
4.41	Photomicrographs of Intimal Hyperplasia of Aorta in Treatment Study.	98
4.42	Mean Scoring of Collagen in Aorta Stained with Masson Trichrome in Treatment Study.	99
4.43	Photomicrograph of Aorta from Control Treatment Group.	100
4.44	Photomicrograph of Aorta from BVAE 50 Treatment Group.	100
4.45	Photomicrograph of Aorta from Aspirin Treatment Group.	101
4.46	Photomicrograph of Aorta from Warfarin Rabbit Group.	101
4.47	Photomicrograph of Aorta from Simvastatin Treatment Group.	102
4.48	Photomicrograph of Aorta from Control Treatment Group.	103
4.49	Photomicrograph of Aorta from BVAE 50 mg/kg Treatment Group.	103
4.50	Photomicrograph of Aorta from Aspirin Treatment Group.	104
4.51	Photomicrograph of Aorta from Warfarin Treatment Group.	104

4.52	Photomicrograph of Aorta from Simvastatin Treatment Group.	105
4.53	Effect of BVAE 50 on Serum TNF- $\alpha$ of Rabbit in Treatment Study.	106
4.54	Effect of BVAE 50 on Serum IL-6 of Rabbit in Treatment Study.	107
4.55	Effect of BVAE 50 on RAM II Scoring of Aorta in Treatment Study.	108
4.56	Photomicrograph of Macrophage Localisation in Aorta of Control Group in Treatment Study.	109
4.57	Photomicrograph of Macrophage Localisation in Aorta of BVAE 50 Group in Treatment Study.	109
4.58	Photomicrograph of Macrophage Localisation in Aorta of Aspirin Group in Treatment Study.	110
4.59	Photomicrograph of Macrophage Localisation in Aorta of Warfarin Group in Treatment Study.	110
4.60	Photomicrograph of Macrophage Localisation in Aorta of Simvastatin Group Treatment Study.	111
4.61	Inhibition of Platelet Aggregation (%) on Rabbit Whole Blood by BVAE and Aspirin Using Platelet Agonists of Collagen (2 $\mu$ g/mL).	113
4.62	The effect of BVAE 50 and warfarin-treated group on activated partial thromboplastin time (seconds).	114

## LIST OF ABBREVIATIONS

15-R-HETE	15- R- hydroxyicosatetraenoic acid
5-HT	5- hydroxytryptamine
AA	Arachidonic Acid
ACS	Acute Coronary Syndrome
ADP	Adenosine Diphosphate
AGE	Advance Glycation End- product
ALP	Alkaline Phosphatase
ALT	Alanine Aminotransferase
APTT	Activated Partial Thromboplastin Time
AST	Aspartate Aminotransferase
BMI	Body Mass Index
BV	<i>Berberis vulgaris</i>
BVAE	<i>Berberis vulgaris</i> L Aqueous Extract
BVME	<i>Berberis vulgaris</i> L Methanol Extract
BVPE	<i>Berberis vulgaris</i> L Polysaccharide Extract
CAD	Coronary Artery Disease
cAMP	Cyclic adenosine monophosphate
CHD	Coronary Heart Disease
COX	Cyclooxygenase
CRP	C-reactive Protein
CVD	Coronary Vascular Disease
ELISA	Enzyme Linked Immunosorbent Assay
FBC	Full blood count
FGF- 2	Fibroblast growth factor 2
GBD	Global Burden of Disease Study
GPIa	Glycoprotein Ia
GPIIa/IIIb	Glycoprotein IIb /IIIa
H&E	Haematoxylin and Eosin
HDL	High Density Lipoprotein
HK	High Molecular Weight Kininogen
ICAM	Intercellular Cell Adhesion Molecule
IFN- $\gamma$	Interferon-gamma
IHC	Immunohistochemistry
IHD	Ischemic Heart Disease
IL	Interleukin
INR	International Normalized Ratio
IU/L	International Unit per Liter
LDL	Low Density Lipoprotein
LOX	Lipoxygenase
MI	Myocardial Infarction
MOH	Ministry of Health
NCD	Non-communicable Disease
NO	Nitric Oxide
OS	<i>Orthosiphon stamineus</i> Benth
OSAE	<i>Orthosiphon stamineus</i> Benth Aqueous Extract
OSME	<i>Orthosiphon stamineus</i> Benth Methanol Extract
OSPE	<i>Orthosiphon stamineus</i> Benth Polysaccharide Extract
oxLDL	Oxidize Low Density Lipoprotein

PAF	Platelet Activating Factor
PDGF	Platelet-derived Growth Factor
PG	Prostaglandin
PK	Prekallikrein
PT	Prothrombin Time
ROS	Reactive Oxygen Species
SGOT	Serum Glutamic Oxaloacetic Transaminase
SMC	Smooth Muscle Cell
TF	Tissue Factor
TIA	Transient Ischemic Attack
TNF- $\alpha$	Tumour Necrosis Factor-alpha
TP	Teucrium polium
TPAE	Teucrium polium L Aqueous Extract
TPME	<i>Teucrium polium</i> L Methanol Extract
TPPE	<i>Teucrium polium</i> L Polysaccharide Extract
TXA <sub>2</sub>	Thromboxane A <sub>2</sub>
VCAM	Vascular Cell Adhesion Molecule
VSMC	Vascular Smooth Muscle Cell
vWF	Von Willebrand
WBC	White Blood Cell
WHI	Woman Health Initiative
WHO	World Health Organization

# CHAPTER 1

## INTRODUCTION

### 1.1 Introduction

The World Health Organization (World Health Organization, 2011) reported that coronary artery diseases (CADs) are the leading causes of death and disability in the world. Globally, it is estimated about 17.3 million people died from CADs in 2008 and by 2030, approximately 23.6 million of the world population will die from this disease (Lee, 2006). The risk factor for CADs is mainly due to atherosclerosis which leads to arterial thrombosis.

The pathogenesis of atherothrombosis started with endothelial disturbance by many noxious stimuli, including oxidised LDL cholesterol, glycation end-products, smoking, hypertension and others leading to inflammation, oxidation of more lipoproteins, smooth muscle cell proliferation, platelet activation, hypercoagulation and thrombosis formation. Hypercholesterolemia with high LDL causes release of more platelet-activating factor that, in turn, increased the production of inflammatory factors including activation of macrophages and released of cytokines. The process encompasses chronic progressive atherosclerosis punctuated by acute processes such as plaque rupture and platelet thrombus formation in areas of progressive stenosis. The final pathway to arterial thrombosis is atherosclerosis plaque rupture, thus the effective way to treat or to prevent this event is limiting or eradicating platelet-dependent thrombus formation.

Anticoagulant and antiplatelet drugs together with lipid lowering medication are the cornerstone in the management of coronary artery diseases. An ideal anticoagulant should be effective, safe, lack of serious toxicity, and have a wide therapeutic window with less monitoring. It should be also available orally (for long-term use), safe during pregnancy, at low cost and a short half-life for drugs used in the acute setting of thrombosis or has long half-life for prophylaxis (Moll and Roberts, 2002). Aspirin is the most commonly used antiplatelet drug worldwide in both primary and secondary prevention of CVDs. However, the antiplatelet effects of aspirin may vary between individuals. A proportion of patients prescribed aspirin suffer recurrent thromboembolic vascular events ('aspirin resistance') or had increased risk of bleeding (Hankey and Eikelboom, 2006).

The pharmaceutical industry uses animal polysaccharides on a large scale, especially for treatment of CADs. Heparin is an example of an animal based, widely used as anticoagulant for the treatment and prevention of thrombotic diseases and for maintaining blood fluidity in extracorporeal devices (Johnell et al., 2002). Unfortunately, the main complication with heparin as an anticoagulant includes occasionally life-threatening bleeding and heparin-induced thrombocytopenia (Greinacher and Warkentin, 2006).



Medicinal plants serve as a greater resource for new medication and their potential is now becoming a topic of interest for researchers all over the world. WHO has recommended medicinal plants to be used more effectively in the healthcare system (Lee, 2006).

Three plants named *Berberis vulgaris* L, *Teucrium polium* L and *Orthosiphon stamineus* Benth are herbs that had been used widely in Asia in daily diets or medicinal purposes. These plants have well-known benefits toward human health (Esmaeili, Zohari and Sadeghi, 2009; Fatehi-Hassanabad et al., 2005; Yuliana et al., 2009). The promising properties from *B. vulgaris*, *T. polium* and *O. stamineus* lead this study to determine their potential anti-atherothrombotic properties. Polysaccharides isolated from higher plants do not contain sulphate groups and their anticoagulant activity is due to the presence of hexuronic acids residues, like GlcA or GalA, and its derivatives (Yoon et al., 2002).

All three plants have been studied for various cardiovascular effects that play important roles in CAD management. These include antihyperlipidemia, cardiogenic and antiarrhythmia effects for *B. vulgaris* (Imanshahidi and Hosseinzadeh, 2008), anti-inflammatory effect for *T. polium* (Passos et al., 2007) and antihyperlipidemia effects for *O. stamineus* (Sriplang et al., 2007). All these cardiovascular effects suggest possible potentials of the plant to be investigated for anti-atherothrombotic properties.

This research resulted to determine the medicinal plants effects of *B. vulgaris*, *T. polium* and *O. stamineus* on atherothrombotic diseases as an herbal treatment. As compared to current synthetic medicine, herbal therapy has shown lower toxicity, easier availability and better acceptability, thus patients with CAD favour to use complementary and alternative medicine (Kristoffersen et al., 2017). Therefore, this study aims to determine the use of *B. vulgaris*, *T. polium* and *O. stamineus* in phase I and *B. vulgaris* aqueous extract as a treatment in atherothrombotic diseases.

## **1.2 Research Hypothesis**

It is hypothesized that the three medicinal plants, *B. vulgaris* fruits, *T. polium* and *O. stamineus* leaves possess potential anti-atherothrombotic effects *in vitro* and *in vivo* experiments. These plants seem as complementary agent with no side effect, to the current medicines which are known with adverse side effects.

### 1.3 Research Objectives

#### 1.3.1 General Objective

To determine anti-atherothrombotic effects of *B. vulgaris*, *T. polium*, and *O. stamineus* extracts *in vitro* and *in vivo* animal models.

#### 1.3.2 Specific Objectives

1. To determine the effect of aqueous extract, methanol extract and polysaccharides crude extract of *B. vulgaris* fruits, *T. polium*, and *O. stamineus* leaves on the basic blood coagulation (Prothrombin Time and Activated Partial Thrombiplastin Time) in human plasma and platelet function test in human whole blood.
2. To identify specific monosaccharides from aqueous extracts of *B. vulgaris*
3. To investigate the anti hyperlipidemia effect of *B. vulgaris* aqueous extract (BVAE) on lipid profile (HDL, LDL, TG and Cholesterol) in atherosclerotic-induced rabbits.
4. To determine the antiplatelet and anti-coagulation effects of BVAE on haematological parameters (full blood count, basic coagulation test and platelet function test) using *in vivo* model.
5. To determine the anti-inflammatory effect of BVAE on interleukin-6, tumor necrosis factor alpha (TNF-  $\alpha$ ) and macrophages in the atherosclerotic plaque in atherosclerotic-induced rabbits.
6. To investigate the effects caused by BVAE on gross anatomical and histological changes of aorta and vital organs of *in vivo* model.
7. To determine toxicity effect of BVAE on liver function test (ALT, AST and ALP), kidney function (RP and creatinine).

## REFERENCES

- Abdelwahab, S. I., Mohan, S., Mohamed Elhassan, M., Al-Mekhlafi, N., Mariod, A. A., Abdul, A. B., Abdulla, M. A., and Alkharfy, K. M. (2010). Antiapoptotic and antioxidant properties of orthosiphon stamineus benth (cat's whiskers): Intervention in the bcl-2-mediated apoptotic pathway. *Evidence-Based Complementary and Alternative Medicine*, 12(2).100-111.
- Abdullah, N. R., Ismail, Z., and Ismail, Z. (2009). Acute toxicity of orthosiphon stamineus benth standardized extract in sprague dawley rats. *Phytomedicine*, 16(2), 222-226.
- Abulude, F. O. (2007). Phytochemical screening and mineral contents of leaves of some nigerian woody plants. *Research Journal of Phytochemistry*, 1(1), 33-39.
- Acheson, J., Danta, G., and Hutchinson, E. (1969). Controlled trial of dipyrindamole in cerebral vascular disease. *British Medical Journal*, 1(5644), 614-615.
- Adams, R. L., and Bird, R. J. (2009). Review article: Coagulation cascade and therapeutics update: Relevance to nephrology. Part 1: Overview of coagulation, thrombophilias and history of anticoagulants. *Nephrology*, 14(5), 462-470.
- Adcock, D. M., Kressin, D. C., and Marlar, R. A. (1997). Effect of 3.2% vs 3.8% sodium citrate concentration on routine coagulation testing. *American Journal of Clinical Pathology*, 107(1), 105-110.
- Aghayan, S. S., Mogadam, H. K., Fazli, M., Darban-Sarokhalil, D., Khoramrooz, S. S., Jabalameli, F., Yaslianifard, S., and Mirzaii, M. (2017). The effects of berberine and palmatine on efflux pumps inhibition with different gene patterns in pseudomonas aeruginosa isolated from burn infections. *Avicenna Journal of Medical Biotechnology*, 9(1), 2.
- Akowuah, G., Ismail, Z., Norhayati, I., and Sadikun, A. (2005). The effects of different extraction solvents of varying polarities on polyphenols of orthosiphon stamineus and evaluation of the free radical-scavenging activity. *Food Chemistry*, 93(2), 311-317.
- Al-Zubairi, A. S., Abdul, A. B., Abdelwahab, S. I., Peng, C. Y., Mohan, S., and Elhassan, M. M. (2011). Eleusine indica possesses antioxidant, antibacterial and cytotoxic properties. *Evidence-Based Complementary and Alternative Medicine*, 2011.
- Almatar, M., Ekal, H., and Rahmat, Z. (2014). A glance on medical applications of orthosiphon stamineus and some of its oxidative compounds. *International Journal of Pharmaceutical Science*, 24, 83-88.

- Alpert, A. J. (1990). Hydrophilic-interaction chromatography for the separation of peptides, nucleic acids and other polar compounds. *Journal of Chromatography A*, 499, 177-196.
- Angiolillo, D. J., and Ferreiro, J. L. (2013). Antiplatelet and anticoagulant therapy for atherothrombotic disease: The role of current and emerging agents. *American Journal of Cardiovascular Drugs*, 13(4), 233-250.
- Bahramikia, S., and Yazdanparast, R. (2012). Phytochemistry and medicinal properties of *Teucrium polium* l.(lamiaceae). *Phytotherapy Research*, 26(11), 1581-1593.
- Bancroft, J. D., and Cook, H. C. (1994). *Manual of histological techniques and their diagnostic application*. London:Churchill Livingstone.
- Basila, D., and Yuan, C.-S. (2005). Effects of dietary supplements on coagulation and platelet function. *Thrombosis Research*, 117(1), 49-53.
- Bennett, J. S. (2001). Novel platelet inhibitors. *Annual Review of Medicine*, 52(1), 161-184.
- Berillis, P. (2013). The role of collagen in the aorta's structure. *The Open Circulation and Vascular Journal*, 6, 1-8.
- Bilia, A., Giomi, M., Innocenti, M., Gallori, S., and Vincieri, F. (2008). Hplc–dad–esi–ms analysis of the constituents of aqueous preparations of verbena and lemon verbena and evaluation of the antioxidant activity. *Journal of Pharmaceutical and Biomedical Analysis*, 46(3), 463-470.
- Bithell, T. (1993). *Blood coagulation* (9th Ed). Wintrobe's clinical hematology. Philadelphia 566-615: Lea and Febiger.
- Bocan, T., Bak Mueller, S., Mazur, M. J., Uhlendorf, P. D., Quenby Brown, E., and Kieft, K. A. (1993). The relationship between the degree of dietary-induced hypercholesterolemia in the rabbit and atherosclerotic lesion formation. *Atherosclerosis*, 102(1), 9-22.
- Bots, M. L., Hoes, A. W., Koudstaal, P. J., Hofman, A., and Grobbee, D. E. (1997). Common carotid intima-media thickness and risk of stroke and myocardial infarction: The rotterdam study. *Circulation*, 96(5), 1432-1437.
- Briffa, T. G., Nedkoff, L., Knuiman, M., Norman, P. E., Hung, J., Hankey, G. J., Thompson, P. L., Geelhoed, E., Hickling, S., Sanfilippo, F., Bremner, A., and Hobbs, M. (2013). Downward trend in the prevalence of hospitalisation for atherothrombotic disease. *International Journal of Cardiology*, 164(2), 185-192. doi: <http://dx.doi.org/10.1016/j.ijcard.2011.06.122>
- Brown, B. A. (1988). *Haematology: Principles and procedures*: Philadelphia: Lea and Febiger.

- Burtis, C., and Ashwood, E. (1999). *Tietz textbook of clinical chemistry and molecular Diagnostic*. Amsterdam: Elsevier Health Sciences.
- Cai, W., Xie, L., Chen, Y., and Zhang, H. (2013). Purification, characterization and anticoagulant activity of the polysaccharides from green tea. *Carbohydrate Polymers*, 92(2), 1086-1090.
- Cardinal, D. C., and Flower, R. J. (1980). The electronic aggregometer: A novel device for assessing platelet behavior in blood. *Journal of Pharmacological Methods*, 3(2), 135-158.
- Cataldo, G., Heiman, F., Lavezzari, M., and Marubini, E. (1998). Indobufen compared with aspirin and dipyridamole on graft patency after coronary artery bypass surgery: Results of a combined analysis. *Coronary Artery Disease*, 9(4), 217-222.
- Changizi Ashtiyani, S., Zarei, A., Taheri, S., Rezaei, A., Golshan, M., and Ghafarzadegan, R. (2013). A comparative study of hypolipidemic activities of the extracts of melissa officinalis and Berberis vulgaris Lin rats. *Journal of Medicinal Plants*, 3(47), 38-47.
- Charo, I. F., and Taub, R. (2011). Anti-inflammatory therapeutics for the treatment of atherosclerosis. *Nature Reviews Drug Discovery*, 10(5), 365-376.
- Chi, L., Peng, L., Pan, N., Hu, X., and Zhang, Y. (2014). The anti-atherogenic effects of berberine on foam cell formation are mediated through the upregulation of sirtuin 1. *International Journal of Molecular Medicine*, 34(4), 1087-1093.
- Chou, T.-C., Fu, E., Wu, C.-J., and Yeh, J.-H. (2003). Chitosan enhances platelet adhesion and aggregation. *Biochemical and Biophysical Research Communications*, 302(3), 480-483.
- Clinic, M. (2011). Activated partial thromboplastin time (aptt), plasma. Retrieved from <http://www.mayomedicallaboratories.com/>
- Cohen, M., and Pollett, J. (1976). Prostaglandin e2 prevents aspirin and indomethacin damage to human gastric mucosa. *Surgical* 1(2), 100-102.
- Coiffic, A., Cazes, E., Janvier, G., Forestier, F., Lanza, F., Nurden, A., and Nurden, P. (1999). Inhibition of platelet aggregation by abciximab but not by aspirin can be detected by a new point-of-care test, the hemostatus. *Thrombosis Research*, 95(2), 83-91.
- Cordier, W., and Steenkamp, V. (2012). Herbal remedies affecting coagulation: A review. *Pharmaceutical Biology*, 50(4), 443-452. doi: 10.3109/13880209.2011.611145
- Corti, R., Fuster, V., and Badimon, J. J. (2003). Pathogenetic concepts of acute coronary syndromes. *Journal of the American College of Cardiology*, 41(4s1), S7-S14.

- Corti, R., Hutter, R., Badimon, J. J., and Fuster, V. (2004). Evolving concepts in the triad of atherosclerosis, inflammation and thrombosis. *Journal of Thrombosis and Thrombolysis*, 17(1), 35-44.
- Dang, X., Miao, J.-j., Chen, A.-q., Li, P., Chen, L., Liang, J.-r., Xie, R.-m., and Zhao, Y. (2015). The antithrombotic effect of rsnk in blood-stasis model rats. *Journal of Ethnopharmacology*, 173, 266-272.
- Danihelka, J., Chrtek Jr, J., and Kaplan, Z. (2012). Checklist of vascular plants of the czech republic. *Preslia*, 84(3), 647-811.
- De Caterina, R., Husted, S., Wallentin, L., Andreotti, F., Arnesen, H., Bachmann, F., Baigent, C., Huber, K., Jespersen, J., and Kristensen, S. D. (2013). General mechanisms of coagulation and targets of anticoagulants (section i). *Thrombosis and Haemostasis*, 109(2013), 569-579.
- De Caterina, R., Libby, P., Peng, H.-B., Thannickal, V. J., Rajavashisth, T., Gimbrone Jr, M., Shin, W. S., and Liao, J. K. (1995). Nitric oxide decreases cytokine-induced endothelial activation. Nitric oxide selectively reduces endothelial expression of adhesion molecules and proinflammatory cytokines. *Journal of Clinical Investigation*, 96(1), 60.
- de Groot, P. G., Urbanus, R. T., and Roest, M. (2012). Platelet interaction with the vessel wall (Ed). *Antiplatelet agents* (pp. 87-110). New York: Springer.
- De La Cruz, J. P., Villalobos, M. a. A., Carmona, J. A., Martín-Romero, M., Smith-Agreda, J. M. a., and de la Cuesta, F. S. (2000). Antithrombotic potential of olive oil administration in rabbits with elevated cholesterol. *Thrombosis Research*, 100(4), 305-315.
- de Sauvage Nolting, P. R., de Groot, E., Zwinderman, A. H., Buirma, R. J., Trip, M. D., and Kastelein, J. J. (2003). Regression of carotid and femoral artery intima-media thickness in familial hypercholesterolemia: Treatment with simvastatin. *Archives of Internal Medicine*, 163(15), 1837-1841.
- De Schryver, E. L., Algra, A., and van Gijn, J. (2007). Dipyridamole for preventing stroke and other vascular events in patients with vascular disease. *The Cochrane Library*, 2007(1), 1-101.
- Depta, J. P., and Bhatt, D. L. (2015). New approaches to inhibiting platelets and coagulation. *Annual Review of Pharmacology and Toxicology*, 55, 373-397.
- Derosa, G., D'Angelo, A., Bonaventura, A., Bianchi, L., Romano, D., and Maffioli, P. (2013). Effects of berberine on lipid profile in subjects with low cardiovascular risk. *Expert Opinion on Biological Therapy*, 13(4), 475-482.
- Devaraj, S., Xu, D. Y., and Jialal, I. (2003). C-reactive protein increases plasminogen activator inhibitor-1 expression and activity in human aortic endothelial cells implications for the metabolic syndrome and atherothrombosis. *Circulation*, 107(3), 398-404.

- Drozd, N., Kuznetsova, S., Lapikova, E., Davydova, A., Makarov, V., Kuznetsov, B., Butylkina, A., Vasil'eva, N., and Skvortsova, G. (2007). Anticoagulant activity of arabinogalactane sulfate and cedar bark extract studied in vitro. *Eksperimental'naiia i Klinicheskaia Farmakologiya*, 71(4), 30-34.
- El Sayed, M., Ghareeb, D., Sarhan, E., and Khalil, A. (2011). Therapeutic bio-screening of the bioactive ingredients of berberis vulgaris. *Functional Plant Science and Biotechnology*, 5(1), 63-68.
- El-Sayed, M. M., Ghareeb, D. A., Talat, H. A., and Sarhan, E. M. (2013). High fat diet induced insulin resistance and elevated retinol binding protein 4 in female rats; treatment and protection with Berberis vulgaris L extract and vitamin a. *Pakistan Journal Pharmacological Science*, 26, 1189-1195.
- El-Wahab, A. E. A., Ghareeb, D. A., Sarhan, E. E., Abu-Serie, M. M., and El Demellawy, M. A. (2013). In vitro biological assessment of Berberis vulgaris L and its active constituent, berberine: Antioxidants, anti-acetylcholinesterase, anti-diabetic and anticancer effects. *BMC Complementary and Alternative Medicine*, 13(1), 218.
- Esmaili, M. A., Zohari, F., and Sadeghi, H. (2009). Antioxidant and protective effects of major flavonoids from Teucrium polium L on  $\beta$ -cell destruction in a model of streptozotocin-induced diabetes. *Planta Medica*, 75(13), 1418-1420.
- Fang, M. C. (2011). *Inpatient anticoagulation (Vol. 4)*: New Jersey: John Wiley and Sons.
- Fang, P., Zhang, D., Cheng, Z., Yan, C., Jiang, X., Kruger, W. D., Meng, S., Arning, E., Bottiglieri, T., and Choi, E. T. (2014). Hyperhomocysteinemia potentiates hyperglycemia-induced inflammatory monocyte differentiation and atherosclerosis. *Diabetes*, 14(8), 10-15.
- Fatehi-Hassanabad, Z., Jafarzadeh, M., Tarhini, A., and Fatehi, M. (2005). The antihypertensive and vasodilator effects of aqueous extract from Berberis vulgaris L fruit on hypertensive rats. *Phytotherapy Research*, 19(3), 222-225.
- Feghali, C. A., and Wright, T. M. (1997). Cytokines in acute and chronic inflammation. *Front Bioscience*, 2(1), 12-26.
- Fitzgerald, M. A. (2000). What are the reasons for an elevated alkaline phosphatase? Retrieved from <http://www.medscape.com/viewarticle/413420>
- Force, U. P. S. T. (2002). Aspirin for the primary prevention of cardiovascular events: Recommendation and rationale. *Annals of Internal Medicine*, 136(2), 157.
- Fuster, V., Moreno, P. R., Fayad, Z. A., Corti, R., and Badimon, J. J. (2005). Atherothrombosis and high-risk plaque: Part i: Evolving concepts. *Journal of the American College of Cardiology*, 46(6), 937-954.

- Gale, A. J. (2011). Continuing education course# 2: Current understanding of hemostasis. *Toxicologic Pathology*, 39(1), 273-280.
- Garcia, D. A., Baglin, T. P., Weitz, J. I., and Samama, M. M. (2012). Parenteral anticoagulants: Antithrombotic therapy and prevention of thrombosis: American college of chest physicians evidence-based clinical practice guidelines. *Chest Journal*, 141(2\_suppl), e24S-e43S.
- Garg, U. C., and Hassid, A. (1989). Nitric oxide-generating vasodilators and 8-bromo-cyclic guanosine monophosphate inhibit mitogenesis and proliferation of cultured rat vascular smooth muscle cells. *Journal of Clinical Investigation*, 83(5), 1774.
- Gawaz, M. (2004). Role of platelets in coronary thrombosis and reperfusion of ischemic myocardium. *Cardiovascular Research*, 61(3), 498-511.
- George, S. J., and Lyon, C. (2010). Pathogenesis of atherosclerosis. In S. J. George & J. Johnson (Ed.), *Atherosclerosis* (pp. 3-19). Weinheim: Wiley.
- Gharaibeh, M. N., Elayan, H. H., and Salhab, A. S. (1988). Hypoglycemic effects of teucrium polium. *Journal of Ethnopharmacology*, 24(1), 93-99.
- Ghooi, R., Thatte, S., and Joshi, P. (1995). The mechanism of action of aspirin—is there anything beyond cyclo-oxygenase? *Medical Hypotheses*, 44(2), 77-80.
- Glauser, B. F., Rezende, R. M., Melo, F. R., Pereira, M. S., Francischetti, I. M., Monteiro, R. Q., Rezaie, A. R., and Mourão, P. A. (2009). Anticoagulant activity of a sulfated galactan: Serpin-independent effect and specific interaction with factor xa. *Thrombosis and Haemostasis*, 102(6), 1183.
- Greinacher, A., and Warkentin, T. E. (2006). Recognition, treatment, and prevention of heparin-induced thrombocytopenia: Review and update. *Thrombosis Research*, 118(2), 165-176.
- Guh, J. H., Ko, F. N., Jong, T. T., and Teng, C. M. (1995). Antiplatelet effect of gingerol isolated from zingiber officinale. *Journal of Pharmacy and Pharmacology*, 47(4), 329-332.
- Gurbel, P. A., Bliden, K. P., DiChiara, J., Newcomer, J., Weng, W., Neerchal, N. K., Gesheff, T., Chaganti, S. K., Etherington, A., and Tantry, U. S. (2007). Evaluation of dose-related effects of aspirin on platelet function. *Circulation*, 115(25), 3156-3164.
- Hadaruga, D. I., Hadaruga, N. G., Bandur, G. N., Ravis, A., Costescu, C., Ordodi, V. L., and Ardelean, A. (2010). Berberis vulgaris L extract/ $\beta$  cyclodextrin nanoparticles synthesis and characterization. *Revista de Chimie(Bucharest)*, 61, 669-675.
- Hall, J. E. (2011). *Guyton and hall textbook of medical physiology* (12th Ed.). Amsterdam: Elsevier Health Sciences.



- Hankey, G. J., and Eikelboom, J. W. (2006). Aspirin resistance. *Lancet*, 367(9510), 606-617.
- Hirano, R., Sasamoto, W., Matsumoto, A., Itakura, H., Igarashi, O., and Kondo, K. (2001). Antioxidant ability of various flavonoids against dpph radicals and ldl oxidation. *Journal of Nutritional Science and Vitaminology*, 47(5), 357-362.
- Hirsh, J., Dalen, J. E., Anderson, D. R., Poller, L., Bussey, H., Ansell, J., and Deykin, D. (2001). Oral anticoagulants: Mechanism of action, clinical effectiveness, and optimal therapeutic range. *Chest Journal*, 119(1\_suppl), 8S-21S.
- Hou, M., Xia, M., Zhu, H., Wang, Q., Li, Y., Xiao, Y., Zhao, T., Tang, Z., Ma, J., and Ling, W. (2007). Lysophosphatidylcholine promotes cholesterol efflux from mouse macrophage foam cells via ppar $\gamma$ -lxr $\alpha$ -abca1-dependent pathway associated with apoe. *Cell Biochemistry and Function*, 25(1), 33-44.
- Hsi, E. D. (2011). A volume in the series: Foundations in diagnostic pathology (2nd Ed.) Hematopathology (pp. 20-30). Amsterdam: Elsevier Health Sciences.
- Huang, C. G., Chu, Z. L., Wei, S. J., Jiang, H., and Jiao, B. H. (2002). Effect of berberine on arachidonic acid metabolism in rabbit platelets and endothelial cells. *Thrombosis Research*, 106(4), 223-227.
- Hughes, S., and Hayman, L. L. (2004). Improving cardiovascular health in women: An opportunity for nursing. *Journal of Cardiovascular Nursing*, 19(2), 145-147.
- Hutter, R., Valdiviezo, C., Sauter, B. V., Savontaus, M., Chereshev, I., Carrick, F. E., Bauriedel, G., Lüderitz, B., Fallon, J. T., and Fuster, V. (2004). Caspase-3 and tissue factor expression in lipid-rich plaque macrophages evidence for apoptosis as link between inflammation and atherothrombosis. *Circulation*, 109(16), 2001-2008.
- Huxley, R. R., Barzi, F., Woo, J., Giles, G., Lam, T. H., Rahimi, K., Konety, S., Ohkubo, T., Jee, S. H., Fang, X., and Woodward, M. (2014). A comparison of risk factors for mortality from heart failure in asian and non-asian populations: An overview of individual participant data from 32 prospective cohorts from the asia-pacific region. *BMC Cardiovascular Disorders*, 14, 61-61. doi: 10.1186/1471-2261-14-61
- Hyafil, F., Cornily, J.-C., Rudd, J. H., Machac, J., Feldman, L. J., and Fayad, Z. A. (2009). Quantification of inflammation within rabbit atherosclerotic plaques using the macrophage-specific ct contrast agent n1177: A comparison with 18f-fdg pet/ct and histology. *Journal of Nuclear Medicine*, 50(6), 959-965.
- Ignatavicius, D. (2002). *Medical -surgical nursing; critical thinking for collaborative care* (4 ed.). Philadelphia: Saunders.
- Imanshahidi, M., and Hosseinzadeh, H. (2008). Pharmacological and therapeutic effects of *Berberis vulgaris* L and its active constituent, berberine. *Phytotherapy Research*, 22(8), 999-1012.

- Imenshahidi, M., and Hosseinzadeh, H. (2016). *Berberis vulgaris* and berberine: An update review. *Phytotherapy Research*, 16(1), 1-5.
- Ingerman-Wojenski, C. M., and Silver, M. J. (1984). A quick method for screening platelet dysfunctions using the whole blood lumi-aggregometer. *Thrombosis and Haemostasis*, 51(2), 154-156.
- Investigators, (2002). Risks and benefits of estrogen plus progestin in healthy postmenopausal women: Principal results from the women's health initiative randomized controlled trial. *Journal of American Medical Association*, 288(3), 321-333.
- Ismail, O., and Maskon, O. (2012). Annual report of the national cardiovascular disease database(ncvd) - percutaneous coronary intervention (pci) registry 2010 - 2012 (pp. 105- 127). Malaysia.
- Ivanovska, N., and Philipov, S. (1996). Study on the anti-inflammatory action of *Berberis vulgaris* Lroot extract, alkaloid fractions and pure alkaloids. *International Journal of Immunopharmacology*, 18(10), 553-561.
- Jain, M. K., and Ridker, P. M. (2005). Anti-inflammatory effects of statins: Clinical evidence and basic mechanisms. *Nature Reviews Drug Discovery*, 4(12), 977-987.
- Jang, H.-D., Chang, K.-S., Huang, Y.-S., Hsu, C.-L., Lee, S.-H., and Su, M.-S. (2007). Principal phenolic phytochemicals and antioxidant activities of three chinese medicinal plants. *Food Chemistry*, 103(3), 749-756.
- Jantan, I., Jumuddin, F. A., Saputri, F. C., and Rahman, K. (2011). Inhibitory effects of the extracts of *Garcinia* species on human low-density lipoprotein peroxidation and platelet aggregation in relation to their total phenolic contents. *Journal of Medical Plants Research*, 5(13), 2699-2709.
- Javadzadeh, S. M., and Fallah, S. R. (2012). Therapeutic application of different parts *berberis vulgaris*. *International Journal of Agriculture and Crop Science*, 4, 404-408.
- Jeong, H. W., Hsu, K. C., Lee, J.-W., Ham, M., Huh, J. Y., Shin, H. J., Kim, W. S., and Kim, J. B. (2009). Berberine suppresses proinflammatory responses through ampk activation in macrophages. *American Journal of Physiology-Endocrinology and Metabolism*, 296(4), E955-E964.
- Johnell, M., Elgue, G., Larsson, R., Larsson, A., Thelin, S., and Siegbahn, A. (2002). Coagulation, fibrinolysis, and cell activation in patients and shed mediastinal blood during coronary artery bypass grafting with a new heparin-coated surface. *The Journal of Thoracic and Cardiovascular Surgery*, 124(2), 321-332.
- Jones, C. R., Hatley, O. J., Ungell, A.-L., Hilgendorf, C., Peters, S. A., and Rostami-Hodjegan, A. (2016). Gut wall metabolism. Application of pre-clinical models

for the prediction of human drug absorption and first-pass elimination. *The American Association of Pharmaceutical Science Journal*, 18(3), 589-604.

- Kahn, M. L., Nakanishi-Matsui, M., Shapiro, M. J., Ishihara, H., and Coughlin, S. R. (1999). Protease-activated receptors 1 and 4 mediate activation of human platelets by thrombin. *The Journal of Clinical Investigation*, 103(6), 879-887.
- Kang, D. G., Sohn, E. J., Kwon, E. K., Han, J. H., Oh, H., and Lee, H. S. (2002). Effects of berberine on angiotensin-converting enzyme and no/cgmp system in vessels. *Vascular Pharmacology*, 39(6), 281-286.
- Kearon, C., Johnston, M., Moffat, K., McGinnis, J., and Ginsberg, J. S. (1998). Effect of warfarin on activated partial thromboplastin time in patients receiving heparin. *Internal Medicine*, 158(10), 1140-1143.
- Keerthisingam, C. B., Jenkins, R. G., Harrison, N. K., Hernandez-Rodriguez, N. A., Booth, H., Laurent, G. J., Hart, S. L., Foster, M. L., and McAnulty, R. J. (2001). Cyclooxygenase-2 deficiency results in a loss of the anti-proliferative response to transforming growth factor- $\beta$  in human fibrotic lung fibroblasts and promotes bleomycin-induced pulmonary fibrosis in mice. *The American Journal of Pathology*, 158(4), 1411-1422.
- Khoo, L. T., Abas, F., Abdullah, J. O., Mohd Tohit, E. R., and Hamid, M. (2014). Anticoagulant activity of polyphenolic-polysaccharides isolated from *melastoma malabathricum* l. *Evidence-Based Complementary and Alternative Medicine*, 2014.
- Kim, J.-A., Kim, J.-E., Song, S. H., and Kim, H. K. (2015). Influence of blood lipids on global coagulation test results. *Annals of Laboratory Medicine*, 35(1), 15-21.
- Kim, M.-S., and Lee, K.-A. (2006). Antithrombotic activity of methanolic extract of *umbilicaria esculenta*. *Journal of Ethnopharmacology*, 105(3), 342-345.
- Knodell, R. G., Ishak, K. G., Black, W. C., Chen, T. S., Craig, R., Kaplowitz, N., Kiernan, T. W., and Wollman, J. (1981). Formulation and application of a numerical scoring system for assessing histological activity in asymptomatic chronic active hepatitis. *Hepatology*, 1(5), 431-435.
- Koga, J.-i., and Aikawa, M. (2012). Crosstalk between macrophages and smooth muscle cells in atherosclerotic vascular diseases. *Vascular Pharmacology*, 57(1), 24-28.
- Končić, M. Z., Kremer, D., Karlović, K., and Kosalec, I. (2010). Evaluation of antioxidant activities and phenolic content of *Berberis vulgaris* L. and *berberis croatica* horvat. *Food and Chemical Toxicology*, 48(8), 2176-2180.
- Kong, L., Luo, C., Li, X., Zhou, Y., and He, H. (2013). The anti-inflammatory effect of kaempferol on early atherosclerosis in high cholesterol fed rabbits. *Lipids In Health And Disease*, 12, 115-115. doi: 10.1186/1476-511X-12-115

- Koniari, I., Mavrilas, D., Papadaki, H., Karanikolas, M., Mandellou, M., Papalois, A., Koletsis, E., Dougenis, D., and Apostolakis, E. (2011). Structural and biomechanical alterations in rabbit thoracic aortas are associated with the progression of atherosclerosis. *Lipids in Health and Disease*, 10(1), 125.
- Koo, K. L., Ammit, A. J., Tran, V. H., Duke, C. C., and Roufogalis, B. D. (2001). Gingerols and related analogues inhibit arachidonic acid-induced human platelet serotonin release and aggregation. *Thrombosis Research*, 103(5), 387-397.
- Krishnaiah, D., Sarbatly, R., and Nithyanandam, R. (2011). A review of the antioxidant potential of medicinal plant species. *Food and Bioproducts Processing*, 89(3), 217-233.
- Ku, S.-K., and Bae, J.-S. (2014). Antiplatelet and antithrombotic activities of purpurogallin in vitro and in vivo. *BMB Reports*, 47(7), 376.
- Lee, H. (2006). Antiplatelet property of curcuma longa l. Rhizome-derived ar-turmerone. *Bioresource Technology*, 97(12), 1372-1376.
- Léon, C., Ravanat, C., Freund, M., Cazenave, J.-P., and Gachet, C. (2003). Differential involvement of the p2y1 and p2y12 receptors in platelet procoagulant activity. *Arteriosclerosis, Thrombosis, and Vascular Biology*, 23(10), 1941-1947.
- Lequin, R. M. (2005). Enzyme immunoassay (eia)/enzyme-linked immunosorbent assay (elisa). *Clinical Chemistry*, 51(12), 2415-2418.
- Levey, A. S., Coresh, J., Balk, E., Kausz, A. T., Levin, A., Steffes, M. W., Hogg, R. J., Perrone, R. D., Lau, J., and Eknoyan, G. (2003). National kidney foundation practice guidelines for chronic kidney disease: Evaluation, classification, and stratification. *Annals of Internal Medicine*, 139(2), 137-147.
- Levi, M., van der Poll, T., and Büller, H. R. (2004). Bidirectional relation between inflammation and coagulation. *Circulation*, 109(22), 2698-2704.
- Li, H., Dong, B., Park, S. W., Lee, H.-S., Chen, W., and Liu, J. (2009). Hnf1 $\alpha$  plays a critical role in pcsk9 gene transcription and regulation by a natural hypocholesterolemic compound berberine. *Journal of Biological Chemistry*, jbc. M109. 052407.
- Li, X., Fries, S., Li, R., Lawson, J. A., Propert, K. J., Diamond, S. L., Blair, I. A., FitzGerald, G. A., and Grosser, T. (2014). Differential impairment of aspirin-dependent platelet cyclooxygenase acetylation by nonsteroidal antiinflammatory drugs. *Proceedings of the National Academy of Sciences*, 111(47), 16830-16835.
- Libby, P., Ridker, P. M., and Hansson, G. K. (2009). Inflammation in atherosclerosis: From pathophysiology to practice. *Journal of the American College of Cardiology*, 54(23), 2129-2138. doi: <http://dx.doi.org/10.1016/j.jacc.2009.09.009>

- Lillie, R. (1940). Further experiments with the masson trichrome modification of mallory's connective tissue stain. *Stain Technology*, 15(1), 17-22.
- Lin, C.-N., Lu, C.-M., Lin, H.-C., Fang, S.-C., Shieh, B.-J., Hsu, M.-F., Wang, J.-P., Ko, F.-N., and Teng, C.-M. (1996). Novel antiplatelet constituents from formosan moraceous plants. *Journal of Natural Products*, 59(9), 834-838.
- Liu, Z. C., and Uetrecht, J. P. (2000). Metabolism of ticlopidine by activated neutrophils: Implications for ticlopidine-induced agranulocytosis. *Drug Metabolism and Disposition*, 28(7), 726-730.
- Ljubuncic, P., Dakwar, S., Portnaya, I., Cogan, U., Azaizeh, H., and Bomzon, A. (2006). Aqueous extracts of *Teucrium polium* L possess remarkable antioxidant activity in vitro. *Evidence-Based Complementary and Alternative Medicine*, 3(3), 329-338.
- López, J. A., and Dong, J.-F. (1997). Structure and function of the glycoprotein ib-ix-v complex. *Current Opinion in Hematology*, 4(5), 323-329.
- Lu, H. T., and Nordin, R. B. (2013). Ethnic differences in the occurrence of acute coronary syndrome: Results of the malaysian national cardiovascular disease (ncvd) database registry (march 2006-february 2010). *BMC Cardiovascular Disorders*, 13(1), 1.
- Maedeker, J. A., Stoka, K. V., Bhayani, S. A., Gardner, W. S., Bennett, L., Procknow, J. D., Staiculescu, M. C., Walji, T. A., Craft, C. S., and Wagenseil, J. E. (2016). Hypertension and decreased aortic compliance due to reduced elastin amounts do not increase atherosclerotic plaque accumulation in *ldlr*<sup>-/-</sup> mice. *Atherosclerosis*, 249, 22-29.
- Mahmoudvand, H., Sharififar, F., Sharifi, I., Ezatpour, B., Harandi, M. F., Makki, M. S., Zia-Ali, N., and Jahanbakhsh, S. (2014). In vitro inhibitory effect of *Berberis vulgaris* L(berberidaceae) and its main component, berberine against different leishmania species. *Iranian Journal of Parasitology*, 9(1), 28.
- Marcovina, S. M., and Koschinsky, M. L. (1998). Lipoprotein (a) as a risk factor for coronary artery disease. *The American Journal of Cardiology*, 82(12), 57U-66U.
- Massberg, S., Brand, K., Grüner, S., Page, S., Müller, E., Müller, I., Bergmeier, W., Richter, T., Lorenz, M., and Konrad, I. (2002). A critical role of platelet adhesion in the initiation of atherosclerotic lesion formation. *The Journal of Experimental Medicine*, 196(7), 887-896.
- Mentré, V., Bulliot, C., Linsart, A., and Ronot, P. (2014). Reference intervals for coagulation times using two point-of-care analysers in healthy pet rabbits (*oryctolagus cuniculus*). *Veterinary Record: Journal of the British Veterinary Association*, 174(27).

- Michelson, A. (2013). Platelet function testing in cardiovascular diseases. *Hematology*,12(2),100-103.
- Mohamed, E. A. H., Lim, C. P., Ebrika, O. S., Asmawi, M. Z., Sadikun, A., and Yam, M. F. (2011). Toxicity evaluation of a standardised 50% ethanol extract of *orthosiphon stamineus*. *Journal of Ethnopharmacology*, 133(2), 358-363.
- Mohammadi, A., Sahebkar, A., Kermani, T., Zhilae, M., Tavallaie, S., and Mobarhan, M. G. (2014). Barberry administration and pro-oxidant–antioxidant balance in patients with metabolic syndrome. *Iranian Red Crescent Medical Journal*, 16(12).
- Moll, S., and Roberts, H. R. (2002, 2002). Overview of anticoagulant drugs for the future. *Hematology*, 12(2),112-120.
- Motalleb, G., Hanachi, P., Fauziah, O., and Asmah, R. (2012). Effect of *Berberis vulgaris* Lfruit extract on alpha-fetoprotein gene expression and chemical carcinogen metabolizing enzymes activities in hepatocarcinogenesis rats. *Iranian Journal of Cancer Prevention*, 1(1), 33-42.
- Movahedi, A. (2014). Comparative and synergistic effects of *orthosiphon stamineus*, *teucrium polium*, *Berberis vulgaris* Lof serum blood cancer markers, glucocorticoid and histology of hepatocarcinogenic rats. (Unpublished doctoral dissertation), Universiti Putra Malaysia, Malaysia.
- Movahedi, A., Basir, R., Rahmat, A., Charaffedine, M., and Othman, F. (2014). Remarkable anticancer activity of *Teucrium polium* L on hepatocellular carcinogenic rats. *Evidence-Based Complementary and Alternative Medicine*, 2014.
- Muzaffar, S., Shukla, N., and Jeremy, J. Y. (2005). Nicotinamide adenine dinucleotide phosphate oxidase: A promiscuous therapeutic target for cardiovascular drugs? *Trends in Cardiovascular Medicine*, 15(8), 278-282.
- Nakashima, Y., Plump, A. S., Raines, E. W., Breslow, J. L., and Ross, R. (1994). Apoe-deficient mice develop lesions of all phases of atherosclerosis throughout the arterial tree. *Arteriosclerosis, Thrombosis, and Vascular Biology*, 14(1), 133-140.
- Naqvi, T. Z., Shah, P. K., Ivey, P. A., Molloy, M. D., Thomas, A. M., Panicker, S., Ahmed, A., Cercek, B., and Kaul, S. (1999). Evidence that high-density lipoprotein cholesterol is an independent predictor of acute platelet-dependent thrombus formation. *The American Journal of Cardiology*, 84(9), 1011-1017.
- Nassar, T., Sachais, B. S., Akkawi, S. e., Kowalska, M. A., Bdeir, K., Leitersdorf, E., Hiss, E., Ziporen, L., Aviram, M., and Cines, D. (2003). Platelet factor 4 enhances the binding of oxidized low-density lipoprotein to vascular wall cells. *Journal of Biological Chemistry*, 278(8), 6187-6193.

- Neville, B., Fareed, J., Florian-Kujawski, M., Cera, L., Duff, R., Valero, A., Beusing, R., Hoppensteadt, D., and Kennedy, R. (2006). Coagulation profiling of human, non-human primate, pig, dog, rabbit, and rat plasma: Pharmacologic implications. *The FASEB Journal*, 20(4), A655-A656.
- Niazmand, S., Esparham, M., Hassannia, T., and Derakhshan, M. (2011). Cardiovascular effects of *Teucrium polium* l. Extract in rabbit. *Pharmacognosy Magazine*, 7(27), 260-264. doi: 10.4103/0973-1296.84244
- Nissen, S. E., Nicholls, S. J., Wolski, K., Rodés-Cabau, J., Cannon, C. P., Deanfield, J. E., Després, J.-P., Kastelein, J. J., Steinhubl, S. R., and Kapadia, S. (2008). Effect of rimonabant on progression of atherosclerosis in patients with abdominal obesity and coronary artery disease: The stradivarius randomized controlled trial. *Journal of American Medical Association*, 299(13), 1547-1560.
- Nurtjahja-Tjendraputra, E., Ammit, A. J., Roufogalis, B. D., Tran, V. H., and Duke, C. C. (2003). Effective anti-platelet and cox-1 enzyme inhibitors from pungent constituents of ginger. *Thrombosis Research*, 111(4-5), 259-265. doi: <http://dx.doi.org/10.1016/j.thromres.2003.09.009>
- Ohashi, K., Bohgaki, T., and Shibuya, H. (2000). Antihypertensive substance in the leaves of kumis kucing (*orthosiphon aristatus*) in java island. *Yakugaku zasshi: Journal of the Pharmaceutical Society of Japan*, 120(5), 474-482.
- Olas, B., Wachowicz, B., Tomczak, A., Erler, J., Stochmal, A., and Oleszek, W. (2008). Comparative anti-platelet and antioxidant properties of polyphenol-rich extracts from: Berries of *aronia melanocarpa*, seeds of grape and bark of *yucca schidigera* in vitro. *Platelets*, 19(1), 70-77.
- Olmez, E., and Ilhan, M. (1992). Evaluation of the alpha-adrenoceptor antagonistic action of berberine in isolated organs. *Arzneimittel-Forschung*, 42(9), 1095-1097.
- Panchal, S. K., Poudyal, H., and Brown, L. (2012). Quercetin ameliorates cardiovascular, hepatic, and metabolic changes in diet-induced metabolic syndrome in rats. *The Journal of Nutrition*, 142(6), 1026-1032.
- Paniccia, R., Priora, R., Liotta, A. A., and Abbate, R. (2014). Platelet function tests: A comparative review. *Vascular Health And Risk Management*, 11, 133-148.
- Park, Y., Franchi, F., Rollini, F., and Angiolillo, D. J. (2015). Update on oral antithrombotic therapy for secondary prevention following non-st segment elevation myocardial infarction. *Trends in Cardiovascular Medicine*, 15(1), 100-105.
- Passos, G. F., Fernandes, E. S., da Cunha, F. M., Ferreira, J., Pianowski, L. F., Campos, M. M., and Calixto, J. B. (2007). Anti-inflammatory and anti-allergic properties of the essential oil and active compounds from *cordia verbenacea*. *Journal of Ethnopharmacology*, 110(2), 323-333.

- Patrono, C. (1994). Aspirin as an antiplatelet drug. *New England Journal of Medicine*, 330(18), 1287-1294.
- Pawlaczyk, I., Capek, P., Czerchawski, L., Bijak, J., Lewik-Tsirigotis, M., Pliszczak-Król, A., and Gancarz, R. (2011). An anticoagulant effect and chemical characterization of *lythrum salicaria* l. Glycoconjugates. *Carbohydrate Polymers*, 86(1), 277-284.
- Pawlaczyk, I., Czerchawski, L., Pilecki, W., Lamer-Zarawska, E., and Gancarz, R. (2009). Polyphenolic-polysaccharide compounds from selected medicinal plants of asteraceae and rosaceae families: Chemical characterization and blood anticoagulant activity. *Carbohydrate Polymers*, 77(3), 568-575.
- Perk, J., De Backer, G., Gohlke, H., Graham, I., Reiner, Ž., Verschuren, M., Albus, C., Benlian, P., Boysen, G., and Cifkova, R. (2012). European guidelines on cardiovascular disease prevention in clinical practice (version 2012). *European Heart Journal*, 33(13), 1635-1701.
- Petersen, M., and Simmonds, M. S. (2003). Rosmarinic acid. *Phytochemistry*, 62(2), 121-125.
- Pettersen, A., Arnesen, H., and Seljeflot, I. (2015). A brief review on high on-aspirin residual platelet reactivity. *Vascular Pharmacology*, 67, 6-9.
- Pliquett, R. U., Cornish, K. G., Peuler, J. D., and Zucker, I. H. (2003). Simvastatin normalizes autonomic neural control in experimental heart failure. *Circulation*, 107(19), 2493-2498.
- Plutzky, J., and Libby, P. (2003). Pathophysiology of atherosclerosis heart disease. In A. Tonkin (Ed.), *Atherosclerosis And Heart Disease* (pp. 1-12). United Kingdom: Martin Dunitz.
- Prakash, P., Misra, A., Surin, W. R., Jain, M., Bhatta, R. S., Pal, R., Raj, K., Barthwal, M. K., and Dikshit, M. (2011). Anti-platelet effects of curcuma oil in experimental models of myocardial ischemia-reperfusion and thrombosis. *Thrombosis Research*, 127(2), 111-118.
- Prasad, K. (1997). Dietary flax seed in prevention of hypercholesterolemic atherosclerosis. *Atherosclerosis*, 132(1), 69-76.
- Prociuk, M., Edel, A., Richard, M., Gavel, N., Ander, B., Dupasquier, C., and Pierce, G. (2008). Cholesterol-induced stimulation of platelet aggregation is prevented by a hempseed-enriched diet this article is one of a selection of papers published in the special issue bridging the gap: Where progress in cardiovascular and neurophysiologic research meet. *Canadian Journal Of Physiology And Pharmacology*, 86(4), 153-159.
- Radomski, M., Palmer, R., and Moncada, S. (1990). An l-arginine/nitric oxide pathway present in human platelets regulates aggregation. *Proceedings of the National Academy of Sciences*, 87(13), 5193-5197.



- Rafeeq, A. K., Zeeshan, F., Maria, J., and Mansoor, A. (2014). Hypolipidemic and antithrombotic evaluation of myrtus communis l. In cholesterol-fed rabbits. *African Journal of Pharmacy and Pharmacology*, 8(8), 235-239. doi: 10.5897/AJPP2013.3488
- Rahmat, A., and Othman, F. (2013). Antioxidant analysis of different parts of carica papaya. *International Food Research Journal*, 20(3), 1043-1048.
- Ramli, N. (2005). Ethanol induced kidney morphological changes in rats after acute dosing. (Unpublished master dissertation), Universiti Putra Malaysia, Malaysia.
- Randall, M., and Wilding, R. (1983). Acute arterial thrombosis in rabbits: Reduced platelet accumulation after treatment with dazoxiben hydrochloride (uk 37,248- 01). *British Journal Of Clinical Pharmacology*, 15(S1).
- Raskin, I., and Ripoll, C. (2004). Can an apple a day keep the doctor away? *Current Pharmaceutical Design*, 10(27), 3419-3429.
- Reininger, A. J., Brandl, R., Penz, S., Goyal, P., Rabie, T., Rother, E., Goetz, C., Engelmann, B., Farndale, R., and Nieswandt, B. (2004). Human atheromatous plaques stimulate thrombus formation by activating platelet glycoprotein vi. *Blood*, 104(11), 2623-2623.
- Rodgers, R., Bagot, C. N., Lawrence, C., Hickman, G., McGurk, M., and Tait, R. C. (2013). Correlating prothrombin time with plasma rivaroxaban level. *British Journal Of Haematology*, 163(5), 685-687.
- Rodríguez, J. A., Orbe, J., and Páramo, J. A. (2007). Metalloproteases, vascular remodeling and atherothrombotic syndromes. *Revista Española de Cardiología*, 60(09), 959-967.
- Rohani, M., Jogstrand, T., Ekberg, M., van der Linden, J., Källner, G., Jussila, R., and Agewall, S. (2005). Interrelation between the extent of atherosclerosis in the thoracic aorta, carotid intima-media thickness and the extent of coronary artery disease. *Atherosclerosis*, 179(2), 311-316.
- Rosenfeld, M. E., and Ross, R. (1990). Macrophage and smooth muscle cell proliferation in atherosclerotic lesions of whhl and comparably hypercholesterolemic fat-fed rabbits. *Arteriosclerosis, Thrombosis, And Vascular Biology*, 10(5), 680-687.
- Saedi, T. (2015). Anti- leukaemic and chemotherapy supportive effects of *Berberis vulgaris* L. Fruit crude extract on in vitro and in vivo models. (Unpublished doctoral dissertation), Universiti Putra Malaysia, Malaysia.
- Sakakura, K., Nakano, M., Otsuka, F., Ladich, E., Kolodgie, F. D., and Virmani, R. (2013). Pathophysiology of atherosclerosis plaque progression. *Heart, Lung and Circulation*, 22(6), 399-411.

- Samuelsson, B., Folco, G., Granström, E., Kindahl, H., and Malmsten, C. (1978). Prostaglandins and thromboxanes: Biochemical and physiological considerations. *Advances In Prostaglandin And Thromboxane Research*, 4, 1.
- Santos, G. R., Tovar, A. M., Capillé, N. V., Pereira, M. S., Pomin, V. H., and Mourão, P. A. (2014). Structural and functional analyses of bovine and porcine intestinal heparins confirm they are different drugs. *Drug Discovery Today*, 19(11), 1801-1807.
- Saputri, F. C., and Jantan, I. (2011). Effects of selected medicinal plants on human low-density lipoprotein oxidation, 2, 2-diphenyl-1-picrylhydrazyl (dpph) radicals and human platelet aggregation. *Journal Of Medicinal Plants Research*, 5(26), 6182-6191.
- Saravanakumar, M., Raja, B., Manivannan, J., Silambarasan, T., Prahalathan, P., Kumar, S., and Mishra, S. K. (2015). Oral administration of veratric acid, a constituent of vegetables and fruits, prevents cardiovascular remodelling in hypertensive rats: A functional evaluation. *British Journal of Nutrition*, 114(09), 1385-1394.
- Schlitt, A., Hauroeder, B., Buerke, M., Peetz, D., Victor, A., Hundt, F., Bickel, C., Meyer, J., and Rupprecht, H. J. (2002). Effects of combined therapy of clopidogrel and aspirin in preventing thrombus formation on mechanical heart valves in an ex vivo rabbit model. *Thrombosis Research*, 107(1), 39-43. doi: [http://dx.doi.org/10.1016/S0049-3848\(02\)00185-8](http://dx.doi.org/10.1016/S0049-3848(02)00185-8)
- Sekhri, A., Lisinschi, A., Furqan, M., Palaniswamy, C., Mukhi, N., Gupta, R., and Nelson, J. C. (2016). The conundrum of “warfarin hypersensitivity”: Prolonged partial thromboplastin time from factor ix propeptide mutation. *American Journal Of Therapeutics*, 23(3), e911-e915.
- Shahraki, M. R., Arab, M. R., Mirimokaddam, E., and Palan, M. J. (2007). The effect of *Teucrium polium* L(calpoureh) on liver function, serum lipids and glucose in diabetic male rats. *Iranian Biomedical Journal*, 11(1), 65-68.
- Sharififar, F., Dehghn-Nudeh, G., and Mirtajaldini, M. (2009). Major flavonoids with antioxidant activity from *Teucrium polium* l. *Food Chemistry*, 112(4), 885-888.
- Smilde, T. J., van Wissen, S., Awollersheim, H., Trip, M. D., Kastelein, J., and Stalenhoef, A. F. (2001). Effect of aggressive versus conventional lipid lowering on atherosclerosis progression in familial hypercholesterolemia (asap): A prospective, randomised, double-blind trial. *The Lancet*, 357(9256), 577-581.
- Sniderman, A. D., and Furberg, C. D. (2008). Age as a modifiable risk factor for cardiovascular disease. *The Lancet*, 371(9623), 1547-1549.
- Sobenin, I. A., Andrianova, I. V., Lakunin, K. Y., Karagodin, V. P., Bobryshev, Y. V., and Orekhov, A. N. (2015). Anti-atherosclerotic effects of garlic preparation

in freeze injury model of atherosclerosis in cholesterol-fed rabbits. *Phytomedicine*. doi: <http://dx.doi.org/10.1016/j.phymed.2015.10.014>

- Soma, M. R., Natali, M., Donetti, E., Baetta, R., Farina, P., Leonardi, A., Comparato, C., Barberi, L., and Catapano, A. L. (2009). Effect of lercanidipine and its (r)- enantiomer on atherosclerotic lesions induced in hypercholesterolemic rabbits. *British Journal Of Pharmacology*, 125(7), 1471-1476.
- Sparrow, C. P., Burton, C. A., Hernandez, M., Mundt, S., Hassing, H., Patel, S., Rosa, R., Hermanowski-Vosatka, A., Wang, P.-R., and Zhang, D. (2001). Simvastatin has anti-inflammatory and antiatherosclerotic activities independent of plasma cholesterol lowering. *Arteriosclerosis, Thrombosis, And Vascular Biology*, 21(1), 115-121.
- Sriplang, K., Adisakwattana, S., Rungsipat, A., and Yibchok-Anun, S. (2007). Effects of orthosiphon stamineus aqueous extract on plasma glucose concentration and lipid profile in normal and streptozotocin-induced diabetic rats. *Journal of Ethnopharmacology*, 109(3), 510-514.
- Srivastava, K., Bordia, A., and Verma, S. (1995). Curcumin, a major component of food spice turmeric (*curcuma longa*) inhibits aggregation and alters eicosanoid metabolism in human blood platelets. *Prostaglandins, Leukotrienes And Essential Fatty Acids*, 52(4), 223-227.
- Srivastava, K. C. (1984). Effects of aqueous extracts of onion, garlic and ginger on platelet aggregation and metabolism of arachidonic acid in the blood vascular system: In vitro study. *Prostaglandins, Leukotrienes and Medicine*, 13(2), 227-235. doi: [http://dx.doi.org/10.1016/0262-1746\(84\)90014-3](http://dx.doi.org/10.1016/0262-1746(84)90014-3)
- Srivastava, K. C. (1986). Isolation and effects of some ginger components on platelet aggregation and eicosanoid biosynthesis. *Prostaglandins, Leukotrienes and Medicine*, 25(2-3), 187-198. doi: [http://dx.doi.org/10.1016/0262-1746\(86\)90065-X](http://dx.doi.org/10.1016/0262-1746(86)90065-X)
- Strydom, H. C., Chandler, A. B., Dinsmore, R. E., Fuster, V., Glagov, S., Insull, W., Rosenfeld, M. E., Schwartz, C. J., Wagner, W. D., and Wissler, R. W. (1995). A definition of advanced types of atherosclerotic lesions and a histological classification of atherosclerosis a report from the committee on vascular lesions of the council on arteriosclerosis. *Circulation*, 92(5), 1355-1374.
- Stouffer, G. A. (2017). Arterial pressure. *Cardiovascular Hemodynamics for the Clinician*, 17(1), 56-58.
- Suau, R., Rico, R., López-Romero, J. M., Nájera, F., and Cuevas, A. (1998). Isoquinoline alkaloids from *Berberis vulgaris* Lsubsp. *Australis*. *Phytochemistry*, 49(8), 2545-2549.
- Suckow, M. A., Stevens, K. A., and Wilson, R. P. (2012). *The laboratory rabbit, guinea pig, hamster, and other rodents*, Cambridge: Academic Press.

- Taj, E., Abdalmutalab, M., Izzaldeen, H., Abdalkareem, M., and Alhassan, M. (2011). Evidence for an in vitro anticoagulant activity of red onion (*allium cepa* L.). *Sudan Journal of Medical Sciences*, 6(2).
- Tarantino, E., Amadio, P., Squellerio, I., Porro, B., Sandrini, L., Turnu, L., Cavalca, V., Tremoli, E., and Barbieri, S. S. (2016). Role of thromboxane-dependent platelet activation in venous thrombosis: Aspirin effects in mouse model. *Pharmacological Research*, 107, 415-425.
- Teng, C.-M., Chen, C.-C., Ko, F.-N., Lee, L.-G., Huang, T.-F., Chen, Y.-P., and Hsu, H.-Y. (1988). Two antiplatelet agents from *magnolia officinalis*. *Thrombosis Research*, 50(6), 757-765.
- Teng, C.-M., Li, H.-L., Wu, T.-S., Huang, S.-C., and Huang, T.-F. (1992). Antiplatelet actions of some coumarin compounds isolated from plant sources. *Thrombosis Research*, 66(5), 549-557.
- Teng, C. M., Ko, F. N., Wang, J. P., Lin, C. N., Wu, T. S., Chen, C. C., and Huang, T. F. (1991). Antihaemostatic and antithrombotic effect of some antiplatelet agents isolated from chinese herbs. *Journal of Pharmacy and Pharmacology*, 43(9), 667-669.
- Tremblay, A. J., Morrissette, H., Gagné, J.-M., Bergeron, J., Gagné, C., and Couture, P. (2004). Validation of the friedewald formula for the determination of low-density lipoprotein cholesterol compared with  $\beta$ -quantification in a large population. *Clinical Biochemistry*, 37(9), 785-790. doi: <http://dx.doi.org/10.1016/j.clinbiochem.2004.03.008>
- Troussard, X., Cornet, E., Bardet, V., Couaillac, J.-P., Fossat, C., Luce, J.-C., Maldonado, E., Siguret, V., Tichet, J., and Lantieri, O. (2013). Full blood count normal reference values for adults in france. *Journal Of Clinical Pathology*, Jclinpath-2013-201687.
- Uprichard, J., Manning, R. A., and Laffan, M. A. (2010). Monitoring heparin anticoagulation in the acute phase response. *British Journal Of Haematology*, 149(4), 613-619.
- Verhoeff, F. (1908). Some new staining methods of wide applicability including a rapid differential stain for elastic tissue. *Journal of the American Medical Association*, 50(11), 876-877.
- Viles-Gonzalez, J. F., Fuster, V., and Badimon, J. J. (2004). Atherothrombosis: A widespread disease with unpredictable and life-threatening consequences. *European Heart Journal*, 25(14), 1197-1207.
- Watson, T., Shantsila, E., and Lip, G. Y. (2009). Mechanisms of thrombogenesis in atrial fibrillation: Virchow's triad revisited. *The Lancet*, 373(9658), 155-166.
- Wong, N. D. (2012). Is diabetes really a coronary heart disease risk equivalent? *Cardiovascular Endocrinology*, 1(4), 65-67.

- World Health Organization, (2000). Obesity: Preventing and managing the global epidemic. 2000. WHO Technical Report Series.
- World Health Organization, W. (2002). The world health report 2002: Reducing risks, promoting healthy life. Geneva: World Health Organization.
- World Health Organization, W. (2014a). Global status report on noncommunicable diseases 2014 (pp. 9). Switzerland: World Health Organization.
- World Health Organization, W. (2014b). Projections of mortality and causes of death, 2015 and 2030. Retrieved 4 November 2014 [http://www.who.int/healthinfo/global\\_burden\\_disease/projections/en/](http://www.who.int/healthinfo/global_burden_disease/projections/en/), [http://www.who.int/gho/ncd/mortality\\_morbidity/en/](http://www.who.int/gho/ncd/mortality_morbidity/en/)
- World Health Organization, W. (2017, June 2017). Noncommunicable diseases. Retrieved 17 August 2017, from <http://www.who.int/mediacentre/factsheets/fs355/en/>
- World Health Organization, W. (2011). Global atlas on cardiovascular disease prevention and control. In W. H. Organization (Series Ed.), World Health Organization Report (pp. 164). Geneva: World Health Organization.
- Wysowski, D. K., Nourjah, P., and Swartz, L. (2007). Bleeding complications with warfarin use: A prevalent adverse effect resulting in regulatory action. *Archives Of Internal Medicine*, 167(13), 1414-1419.
- Yee, D. L., Sun, C. W., Bergeron, A. L., Dong, J.-f., and Bray, P. F. (2005). Aggregometry detects platelet hyperreactivity in healthy individuals. *Blood*, 106(8), 2723-2729.
- Yin, M.-J., Yamamoto, Y., and Gaynor, R. B. (1998). The anti-inflammatory agents aspirin and salicylate inhibit the activity of i $\kappa$ b kinase- $\beta$ . *Nature*, 396(6706), 77-80.
- Yoon, S.-J., Pereira, M. S., Pavão, M. S., Hwang, J.-K., Pyun, Y.-R., and Mourão, P. A. (2002). The medicinal plant *porana volubilis* contains polysaccharides with anticoagulant activity mediated by heparin cofactor ii. *Thrombosis Research*, 106(1), 51-58.
- Yuliana, N. D., Khatib, A., Link-Struensee, A. M. R., Ijzerman, A. P., Rungkat-Zakaria, F., Choi, Y. H., and Verpoorte, R. (2009). Adenosine a1 receptor binding activity of methoxy flavonoids from *orthosiphon stamineus*. *Planta Medica*, 75(02), 132-136.
- Yusuf, S., Hawken, S., Ounpuu, S., Dans, T., Avezum, A., and Lanans, F. (2004). Effect of potentially modifiable risk factors associated with myocardial infarction in 52 countries (the interheart study): Case control study. *Lancet*, 364, 937-952.

- Zaragoza, C., Gomez-Guerrero, C., Martin-Ventura, J. L., Blanco-Colio, L., Lavin, B., Mallavia, B., Tarin, C., Mas, S., Ortiz, A., and Egido, J. (2011). Animal models of cardiovascular diseases. *BioMed Research International*, 2011.
- Zarei, A., Changizi-Ashtiyani, S., Taheri, S., and Ramezani, M. (2015). A quick overview on some aspects of endocrinological and therapeutic effects of *Berberis vulgaris* L. *Avicenna Journal Of Phytomedicine*, 5(6), 485.
- Zeng, X.-H., Zeng, X.-J., and Li, Y.-Y. (2003). Efficacy and safety of berberine for congestive heart failure secondary to ischemic or idiopathic dilated cardiomyopathy. *The American Journal of Cardiology*, 92(2), 173-176.
- Zhang, C., Rexrode, K. M., van Dam, R. M., Li, T. Y., and Hu, F. B. (2008). Abdominal obesity and the risk of all-cause, cardiovascular, and cancer mortality. *Circulation*, 117(13), 1658-1667.
- Zimetti, F., Adorni, M., Ronda, N., Gatti, R., Bernini, F., and Favari, E. (2015). The natural compound berberine positively affects macrophage functions involved in atherogenesis. *Nutrition, Metabolism and Cardiovascular Diseases*, 25(2), 195-201.
- Zulkhairi, A., Zaiton, Z., Jamaluddin, M., Sharida, F., Mohd, T., Hasnah, B., Nazmi, H., Khairul, O., and Zanariyah, A. (2008). Alpha lipoic acid posses dual antioxidant and lipid lowering properties in atherosclerotic-induced new zealand white rabbit. *Biomedicine & Pharmacotherapy*, 62(10), 716-722.