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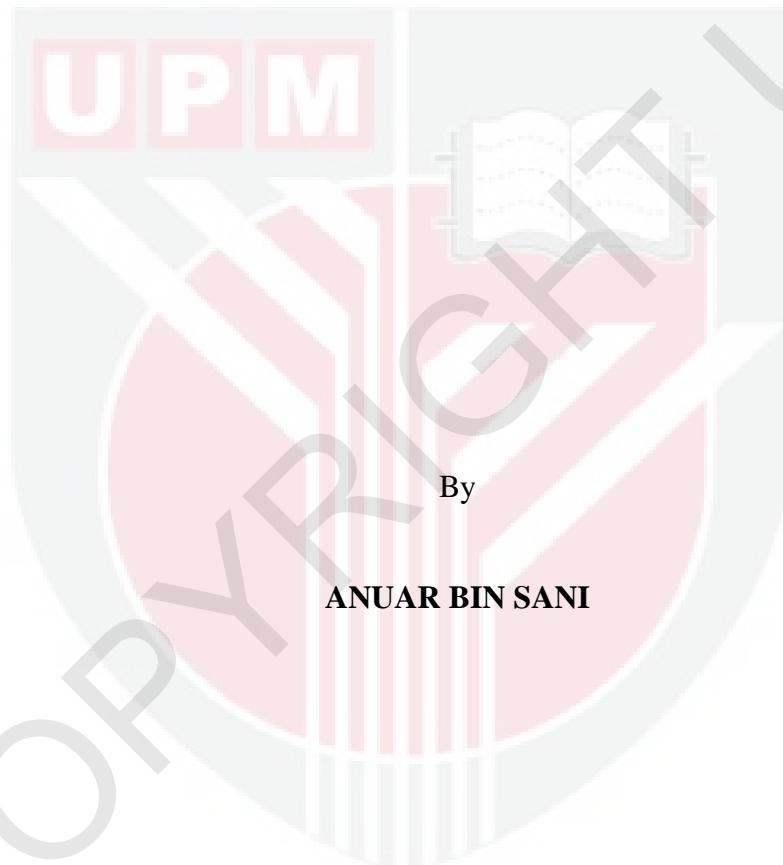
***In Vivo EVALUATION OF SWEET POTATO LEAF  
(Ipomoea batatas (L.) Lam.) EXTRACTS FOR TREATMENT OF  
PSORIASIS  
USING MOUSE MODEL***

ANUAR BIN SANI

FPSK(P) 2017 17



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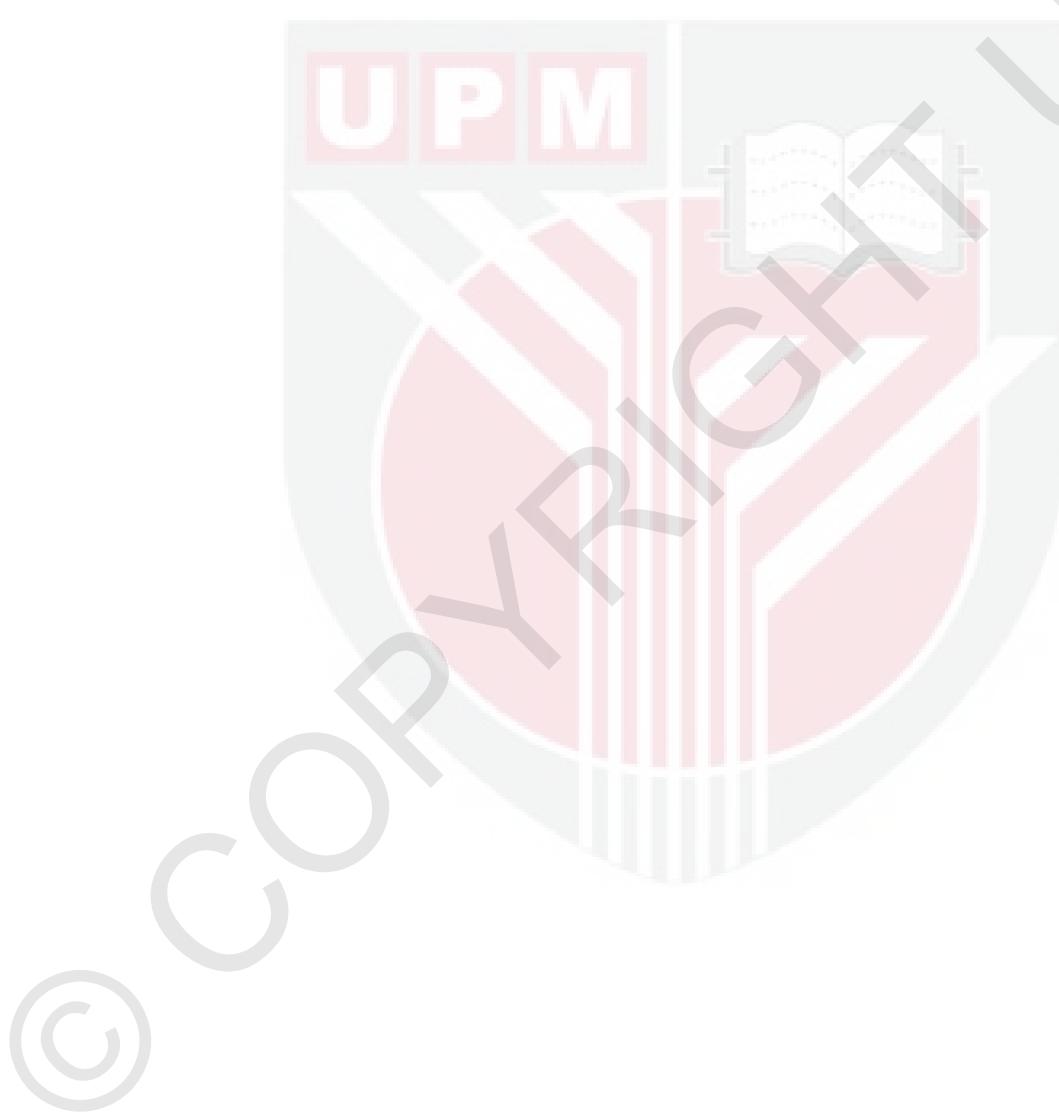
**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,  
in Fulfillments of the Requirements for the Degree of Doctor of Philosophy**

**April 2017**

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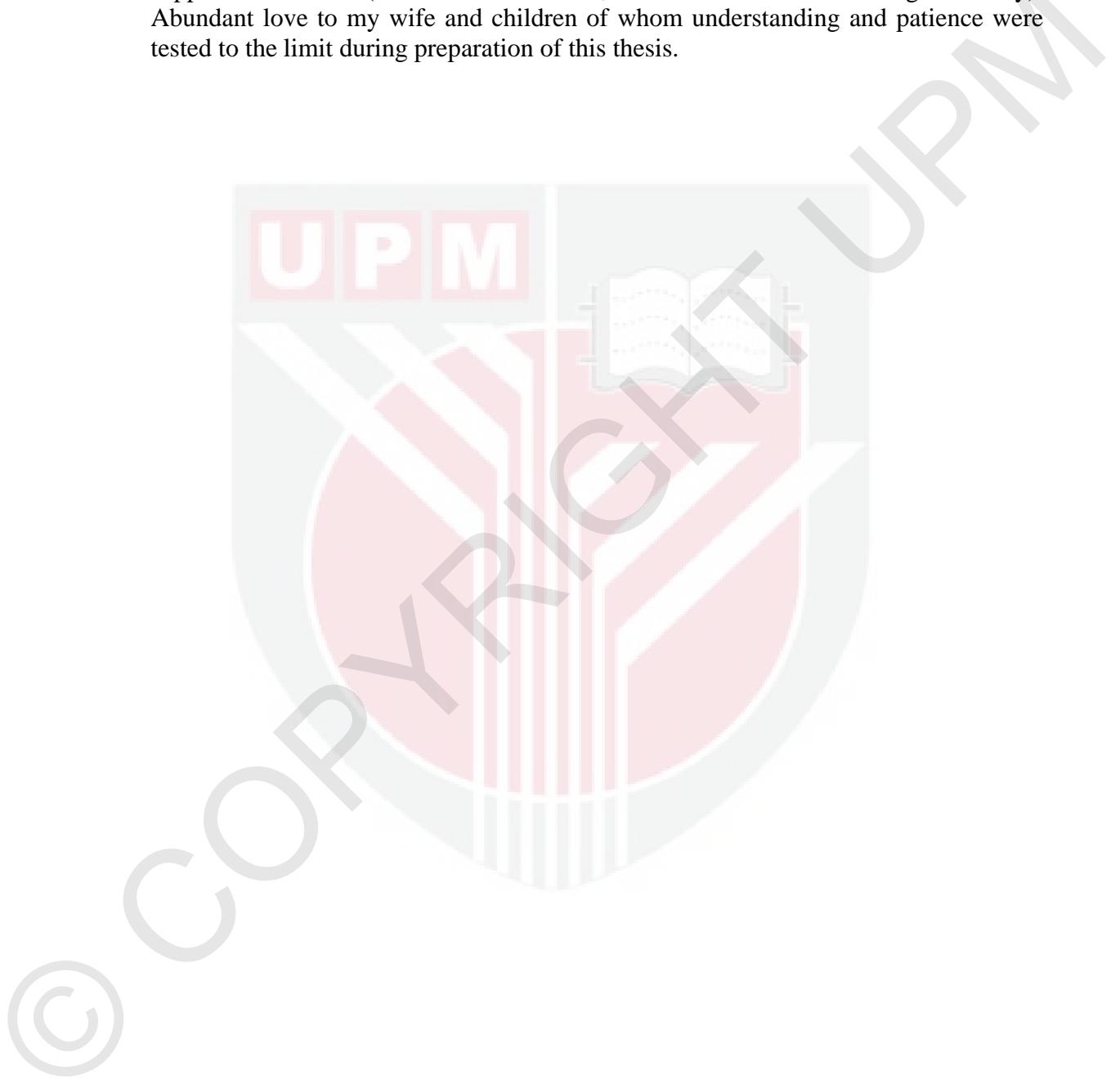
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## **DEDICATION**

It has been a roller coaster of almost seven years to finish this endeavour. Juggling study, families and work commitments was never easy. First and foremost, BIG thank you to USIM and JPA for the sponsorship and to my supervisors for never ending support and belief (Prof. Dr Wan Omar, Prof. Dr Adel and Dr. Ngah Zasmy). Abundant love to my wife and children of whom understanding and patience were tested to the limit during preparation of this thesis.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment  
of the requirement for the Degree of Doctor of Philosophy

***In Vivo EVALUATION OF SWEET POTATO LEAF  
(Ipomoea batatas (L.) Lam.) EXTRACTS FOR TREATMENT OF PSORIASIS  
USING MOUSE MODEL***

By

ANUAR SANI

April 2017

**Chairman : Professor Wan Omar Bin Abdullah, PhD**  
**Faculty : Medicine and Health Sciences**

Psoriasis is a complex genetic disease involving both autoimmune and inflammatory components though its exact aetiology remains unknown. It is prevalent worldwide and affects both men and women over a wide age range. In Malaysia, about 4% of the population are affected.

Topical medications especially corticosteroids are used for localised disease. Phototherapy and climatotherapy are effective for more extensive disease. Systemic therapies are reserved for patients with refractory disease. The use of complementary and alternative medicines is also common among psoriasis patients. In Malaysia, sweet potato leaves have been used as an alternative therapy for psoriasis. In light of these subjective observations and under continuous pressure of finding new remedies to meet the unmet needs in psoriasis treatment, the researcher examined two different extracts derived from sweet potato leaves against experimentally induced psoriasis aiming to draw a distinction between the patients' subjective observations and the scientific evidence, risks and benefits of treatments.

This study offered a novel research based on examination of lipophilic and hydrophilic extracts from sweet potato leaves (SPL) of Gendut MSP94 cultivar in experimental animal model. The hydrophilic (ethanolic) extraction of SPL was done based on the protocol described by Park *et al.* (2010). The Lipophilic (hexane) extraction was based on the protocol described by Teow *et al.* (2007). The concentrated extracts obtained were in the form of thick dark greenish pastes which were ideal for direct application onto the induced psoriasis. Hydrophilic extract showed high levels of chlorogenic and caffeic acids which were similar to earlier studies. However, it is strongly recommended that ultra-analysis of Gendut SPL extracts should be considered in the future studies to define the traces of bioactive chemicals not been identified in the

current research, as it was beyond its scope. The total phenolic content of Gendut SPL ethanolic extracts was assessed using *Amin et al.* (2004) protocol. It was found to be nearly 50% of the values obtained from other studies done on different cultivars of sweet potatoes.

The mouse model of psoriasis was induced chemically by topical Imiquimoid onto shaved skin of BALB/c mice, following the same protocol described by *van der Fits et al.* (2009). Histologically, the resulting lesions were typical to human's psoriasis vulgaris as confirmed by dermatologist and histopathologist. Flow cytometry showed increased levels of CD3<sup>+</sup>, CD4<sup>+</sup> and CD8<sup>+</sup> cells and TNF-alpha in the peripheral blood which were signs of inflammatory reaction seen in acute psoriasis. Upon treatment, results showed significant clinical improvement in the lipophilic SPL extract-treated group, as scored by modified PASI, compared to hydrophilic SPL extract-treated and negative control groups. Histopathological changes showed significant decrease of epidermal thickness and rete ridges. Immunological and molecular parameters showed significant decrease in the inflammatory biomarkers namely TNF-alpha and the percentage of double negative T cells. In conclusion, this study shows a substantial evidence of anti-psoriatic properties of SPL lipophilic extract that should encourage us to look further into its active compound(s) responsible for the effects.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

**PENILAIAN IN-VIVO EKSTRAK DAUN UBI KELEDEK  
(*Ipomoea batatas*) (L) Lam. ) TERHADAP RAWATAN PENYAKIT  
PSORIASIS MENGGUNAKAN MODEL TIKUS**

Oleh

**ANUAR SANI**

**April 2017**

**Pengerusi : Profesor Wan Omar bin Abdullah, PhD**  
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Psoriasis adalah penyakit genetik yang kompleks melibatkan kedua-dua komponen autoimun dan keradangan, namun punca bagi psoriasis masih tidak diketahui. Psoriasis mempunyai kesan penyebaran serata dunia dan melibatkan lelaki dan wanita pada julat umur yang luas. Di Malaysia, lebih kurang 4% penduduk adalah terkesan dengan psoriasis.

Rawatan topikal, terutamanya kortikosteroid, digunakan untuk psoriasis setempat. Rawatan fototerapi dan klimatoterapi berkesan kepada psoriasis yang lebih serius. Terapi sistemik dikhaskan untuk pesakit yang mempunyai psoriasis refraktori. Penggunaan rawatan pelengkap dan alternatif adalah biasa dikalangan pesakit psoriasis. Di Malaysia, ada pesakit psoriasis menggunakan daun ubi keledek sebagai rawatan alternatif bagi psoriasis. Berdasarkan pemerhatian subjektif dari pesakit psoriasis dan sememangnya ada keperluan berterusan untuk mencari rawatan yang baru untuk penyakit ini, penyelidikan ini telah memeriksa dua jenis ekstrak yang diperolehi daripada daun ubi keledek terhadap psoriasis terinduksi secara eksperimental dengan matlamat untuk membuat perbezaan antara pemerhatian subjektif pesakit dengan bukti saintifik, risiko dan faedah rawatan.

Penyelidikan ini menawarkan kajian novel berdasarkan pemerhatian ekstrak lipofilik dan hidrofilik daripada daun ubi keledek (SPL) kultivar Gendut MSP94 menggunakan model eksperimental haiwan. Pengekstrakan SPL hidrofilik (etanol) dilakukan berdasarkan protokol yang telah dinyatakan oleh Park dan rakan-rakan. (2010). Pengekstrakan SPL lipofilik (heksana) pula dilakukan berdasarkan protokol yang telah dinyatakan oleh Teow dan rakan-rakan (2007). Ekstrak pekat yang diperolehi dalam bentuk pes hijau gelap sesuai untuk aplikasi secara terus kepada psoriasis terinduksi. Ekstrak SPL hidrofilik menunjukkan tahap asid klorogenik dan caffeic yang tinggi

seperti dalam kajian-kajian yang dilakukan sebelum ini. Namun, dicadangkan supaya analisis ultra ekstrak SPL Gendut dilakukan untuk kajian yang akan datang bagi menjelaskan lagi unsur surih bahan kimia bioaktif yang tidak dikenalpasti di dalam kajian kini kerana ia berada di luar skop kajian. Jumlah kandungan fenol di dalam ekstrak Gendut SPL diuji menggunakan protokol Amin dan rakan-rakan (2004). Jumlah kandungan fenol yang adalah mendekati 50% daripada jumlah nilai yang didapati daripada kajian awal mengenai kultivar ubi keledek yang berbeza.

Tikus BALB/c telah diinduksikan dengan psoriasis secara kimia dengan mengaplikasikan topikal Imiquimoid kepada kulit tikus yang telah dicukur, mengikut kepada protokol yang dinyatakan oleh van der fits dan rakan-rakan (2009). Secara histologikal, lesi yang didapati adalah sama seperti psoriasis vulgaris manusia sebagaimana disahkan oleh pakar dermatologi dan histopatologi. Sitometri aliran menunjukkan peningkatan tahap sel CD<sup>3+</sup>, CD<sup>4+</sup>, CD<sup>8+</sup> dan TNF-alfa dalam darah periferi. Ini merupakan petanda tindak balas keradangan yang dilihat dalam psoriasis akut. Selepas rawatan, hasil menunjukkan kemajuan klinikal yang signifikan untuk kumpulan yang dirawat menggunakan ekstrak SPL lipofilik, seperti yang diskorkan menggunakan PASI terubahsuai, berbanding dengan kumpulan yang dirawat dengan ekstrak SPL hidrofilik dan kumpulan kontrol negatif. Perubahan histopatologikal menunjukkan pengurangan yang signifikan bagi ketebalan epidermal dan aluran rete. Parameter immunologi dan molekular menunjukkan pengurangan yang signifikan bagi petunjuk biologi keradangan, terutamanya TNF-alfa dan peratusan negatif berganda sel T. Kesimpulannya, kajian ini menunjukkan bukti yang kukuh bagi ciri-ciri anti-psoriasis untuk ekstrak SPL lipofilik. Ini seharusnya meyakinkan kita untuk melihat lebih dalam kepada sebatian-sebatian aktif yang bertanggungjawab kepada kesan-kesan anti-psoriasis.

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I certify that a Thesis Examination Committee has met on 21 April 2017 to conduct the final examination of Anuar bin Sani on his thesis entitled "In Vivo Evaluation of Sweet Potato Leaf (*Ipomoea batatas* (L.) Lam.) Extracts for Treatment of Psoriasis Using Mouse Model" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

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## LIST OF ABBREVIATIONS

APC	Antigen-presenting cells
CD	cluster of differentiation
CAM	Complementary and Alternative medicine
CLCL	cumulative life course impairment
DCs	Dendritic cells
dw	dry weight
FC	Folin-Ciocalteu
GAE	Gallic acid equivalent
g	gram
H&E stain	Hematoxylin and Eosin stain
HPLC	High performance liquid chromatography
HETG	Hydrophilic Extract Test Group
IFN-alpha	Interferon -alpha
IFN-γ	interferon-gamma
IL	interleukin
IB	ipomoea batatas
LETG	Lipophilic Extract Test Group
NCG	Negative Control Group
PCG	Positive Control Group
PASI	Psoriatic Assessment and Severity Index
Treg	Regulatory T cells
SPL	sweet potato leaves
Th1	T helper cell 1
Th2	T helper cell 2
TF	total flavonoids
TP	total phenolic

TPC

Total phenolic content

TNF- $\alpha$

tumour necrosis factor- alpha



# **CHAPTER 1**

## **INTRODUCTION**

### **1.1 Overview of psoriasis**

Psoriasis is a common chronic disfiguring inflammatory immunological disorder essentially of the skin. In between 1.3% and 34.7% of skin psoriasis, joints are involved as psoriatic arthritis which causes severe disability (Lebwohl, 2003).

Skin psoriasis is one of the prototypic papulosquamous diseases which involves skin and nails, and is associated with several comorbidities. The typical aetiology is unknown, however there are consistent pathological and immunological features of epidermal hyperproliferation, T cell expansion, cytokine over production and angiogenesis mostly related to dysregulation of the immune system. So, immunological, genetic and environmental factors interplay and influence the pathophysiology and the outcome of the disease (Lebwohl, 2003).

Psoriasis has a great negative impact on patients' quality of life. The emotional and social consequences should not be underestimated. Many patients have severe humiliation because of their visible lesions. They withdraw from the society, develop depression, anxiety, and have increased suicidality (Olivier, Robert, Daihung, Urbà, Catalin, Hywel & Gelfand, 2010). At the community level, psoriasis causes significant social and economic burden. Work absenteeism and other complications related to psoriasis are known as cumulative life course impairment (CLCI) (Warren, Kleyn, & Gulliver, 2011; Pearce, Singh, Balkrishnan, Kulkarni, Fleischer & Feldman, 2006).

A spectrum of anti-psoriatic treatment modalities are available. The severity of the disease usually determines the therapeutic approach and approximately 70 to 80% of all patients with psoriasis can be treated adequately with topical preparations (Mitra, Morrissey & Lim, 2013). Despite the recent advances in psoriasis therapeutics, the disease remains as lifelong incurable ailment of relapse and remission. The main goal of treatment remains to establish and prolong periods between flares (Global report on psoriasis-WHO, 2016). None of available therapies induce a permanent remission and all therapies are associated with side effects such as skin atrophy due to topical steroids or even immune suppression by anti metabolites that can place limits on their uses. As such, Complementary and Alternative Medicines (CAM) are widely used by psoriatic patients worldwide.

The common modalities of CAMs include traditional Chinese medicine (TCM), herbal therapies, dietary supplements, climatology, and mind/body interventions. Herbal and dietary therapies seem to have the most evidence for efficacy (Talbott & Duffy, 2015).

## **1.2 The rationale of studying sweet potato leaves (SPL) effects on experimentally induced psoriasis in BALB/c mice**

The idea of studying the topical effects of Sweet Potato (*Ipomoea batatas*) Leaves (SPL) on induced psoriasis lesions in animal model, came from the anecdotal observations of Malaysian patients who failed to achieve satisfactory response to conventional therapies, but observed significant remission after trying topical SPL paste. Furthermore, plenty of research were done especially in Japan, Taiwan and China on immune-modulatory potential of sweet potato leaves and considered as one of the healthiest food (Johnson & Pace, 2010). Coupled with the known side effects and high cost of conventional treatments of psoriasis, the need to look for an alternative in the form of locally available and cheap to get is even more pressing, thus sweet potato leaves fits the bill.

Scientific research based on investigating an observable event is a standard way of getting new knowledge, and correcting and integrating previous information. Successful studies based on examination of observations can help to construct a hypothesis and draw a conclusion. This study used mixed quantitative and semi-quantitative methodologies to process and analyse the obtained data.

## **1.3 Objectives**

General: To determine the potential action of topical hydrophilic and lipophilic extracts of Sweet Potato Leaves (*Ipomoea batatas*) on experimentally induced psoriasis in animal (mouse) model.

Specific:

- i) To assess response of induced psoriatic lesions when subjected to topical SPL extracts using modified PASI score.
- ii) To compare the histological response of induced psoriasis lesions following topical application SPL extracts.
- iii) To examine the liver, spleen and kidney histological sections before and following topical application of SPL extracts.
- iv) To evaluate the effects of topical application of SPL extracts on immunoparameters of cellular and molecular components of the psoriatic induced mice.

## **1.4 Hypothesis**

The hypothesis was that topical application of SPL extracts on the induced psoriatic lesions would speed up the recovery by reversing the induced lesions to near normality as compared to leaving the induced lesions to heal by itselfs. And, it would be interesting to find out, if there was such an effect, which of the extracts would work better.

It was also hypothesized that the positive healing effect would be accompanied by changes in cellular and molecular immune-parameters.



## REFERENCES

- Amin, I., Zamaliah, M. M. and Chin, W. F. 2004. Total antioxidant activity and phenolic content in selected vegetables. *Food Chemistry*, 87(4): 581-586.
- Arican, O., Aral, M., Sasmaz, S., & Ciragil, P. (2005). Serum levels of TNF- $\alpha$ , IFN- $\gamma$ , IL-6, IL-8, IL-12, IL-17, and IL-18 in patients with active psoriasis and correlation with disease severity. *Mediators of Inflammation*, 2005(5), 273-279.
- Art ICW, Hollman PCH. (2005). Polyphenols and disease risk in epidemiologic studies. *American Journal Clinical Nutrition* 81(suppl):317S–25S
- Asadullah, K., Docke, W. D., Volk, H. D., & Sterry, W. (1999). The pathophysiological role of cytokines in psoriasis. *Drugs Today (Barc)*, 35(12), 913-924.
- Balato, A., Balato, N., Megna, M., Schiattarella, M., Lembo, S., & Ayal, F. (2012). Pathogenesis of Psoriasis: The Role of Pro-Inflammatory Cytokines Produced by Keratinocytes. *Psoriasis*. doi:10.5772/26163
- Barker, C. L., McHale, M. T., Gillies, A. K., Waller, J., Pearce, D. M., Osborne, J., ... & Pringle, J. H. (2004). The development and characterization of an in vitro model of psoriasis. *Journal of Investigative Dermatology*, 123(5), 892-901.
- Bettini, M., & Vignali, D. A. (2009). Regulatory T cells and inhibitory cytokines in autoimmunity. *Current Opinion in Immunology*, 21(6), 612-618. doi:10.1016/j.coim.2009.09.011
- Bhati, D. (2016). Fruits and vegetables: Nature's gift to obtain better health through antioxidants. *Food Science Research Journal*, 7(2), 335-339. doi:10.15740/has/fsrj/7.2/335-339
- Boehncke, W., & Schön, M. P. (2015). Psoriasis. *The Lancet*, 386(9997), 983-994. doi:10.1016/s0140-6736(14)61909-7
- Bovell-Benjamin, A. C. (2007). Sweet potato: a review of its past, present, and future role in human nutrition. *Advances in Food and Nutrition Research*, 52, 1-59.
- Bowcock, A. M. (2004). The genetics of psoriasis, psoriatic arthritis and atopic dermatitis. *Human Molecular Genetics*, 13(90001). doi:10.1093/hmg/ddh094
- Boyman, O., Hefti, H. P., Conrad, C., Nickoloff, B. J., Suter, M., & Nestle, F. O. (2004). Spontaneous development of psoriasis in a new animal model shows an essential role for resident T cells and tumor necrosis factor- $\alpha$ . *Journal of Experimental Medicine*, 199(5), 731-736.
- Bravo, L. (1998). Polyphenols: chemistry, dietary sources, metabolism, and nutritional significance. *Nutrition Reviews*, 56(11), 317-333.

- Chang, W. H., Chen, C. M., Hu, S. P., Kan, N. W., Chiu, C. C., & Liu, J. F. (2007). Effect of purple sweet potato leaf consumption on the modulation of the antioxidative status in basketball players during training. *Asia Pacific Journal of Clinical Nutrition*, 16(3), 455-461.
- Chaudhari, U., Romano, P., Mulcahy, L. D., Dooley, L. T., Baker, D. G., & Gottlieb, A. B. (2001). Efficacy and safety of infliximab monotherapy for plaque-type psoriasis: a randomised trial. *The Lancet*, 357(9271), 1842-1847.
- Chen, C. M., Li, S. C., Lin, Y. L., Hsu, C. Y., Shieh, M. J., & Liu, J. F. (2005). Consumption of purple sweet potato leaves modulates human immune response: T-lymphocyte functions, lytic activity of natural killer cell and antibody production. *World Journal of Gastroenterology: WJG*, 11(37), 5777.
- Chen, C. M., Lin, Y. L., Chen, C. O., Hsu, C. Y., Shieh, M. J., & Liu, J. F. (2008). Consumption of purple sweet potato leaves decreases lipid peroxidation and DNA damage in humans. *Asia Pacific Journal of Clinical Nutrition*, 17(3), 408-414.
- Chodorowska, G. (1998). Plasma concentrations of IFN- $\gamma$  and TNF- $\alpha$ : in psoriatic patients before and after local treatment with dithranol ointment. *Journal of the European Academy of Dermatology and Venereology*, 10(2), 147-151.
- Christophers, E., Griffiths, C., Gaitanis, G., & Kerkhof, P. V. (2006). The unmet treatment need for moderate to severe psoriasis: results of a survey and chart review. *Journal of the European Academy of Dermatology and Venereology*, 0(0). doi:10.1111/j.1468-3083.2006.01667.x
- Chuang, L.T., Glew, R.H., Wang, Y.C., Yao. P.W., Lin, C.C., Presley, J.M., Schulze, J. and Hou, C.W. 2011. Comparison of the fatty acid, amino acid, mineral and antioxidant content of sweet potato leaves grown on Matsu Island and Mainland Taiwan. *Food 5 (Special Issue 1)*: 43-47
- Cimmino, M. (2007). Epidemiology of psoriasis and psoriatic arthhritis. *Reumatismo*, 20-23.
- Cimmino, M. A. (2007). Epidemiology of psoriasis and psoriatic arthritis. *Reumatismo*, 59(1s), 19-24.
- Clark, R. A., Chong, B., Mirchandani, N., Brinster, N. K., Yamanaka, K., Dowgiert, R. K., & Kupper, T. S. (2006). The Vast Majority of CLA T Cells Are Resident in Normal Skin. *The Journal of Immunology*, 176(7), 4431-4439. doi:10.4049/jimmunol.176.7.4431
- Creamer, D., Sullivan, D., Bicknell, R., & Barker, J. N. W. N. (2002). Angiogenesis in psoriasis. *Angiogenesis*, 5(4), 231-236.
- D'Acquisto, F., & Crompton, T. (2011). CD3+ CD4- CD8-(double negative) T cells: saviours or villains of the immune response?. *Biochemical Pharmacology*, 82(4), 333-340.

- Davison, S., Allen, M., Mallon, E., & Barker, J. (2001). Contrasting patterns of streptococcal superantigen-induced T-cell proliferation in guttate vs. chronic plaque psoriasis. *British Journal of Dermatology*, 145(2), 245-251. doi:10.1046/j.1365-2133.2001.04341.x
- Di Meglio, P., & Duarte, J. H. (2013). CD8 T Cells and IFN- $\gamma$  emerge as critical players for psoriasis in a novel model of mouse psoriasisform skin inflammation. *Journal of Investigative Dermatology*, 133(4), 871-874.
- Dini, I., Tenore, G. C., & Dini, A. (2006). New polyphenol derivative in Ipomoea batatas tubers and its antioxidant activity. *Journal of Agricultural and Food Chemistry*, 54(23), 8733-8737.
- Duke, J.A. & Wain, K.K., (1981). Medicinal Plants of the World, 3 vol. Computer index with more than 85,000 entries. Plants genetics and germplasm Institute. Agriculture Research Service, Beltsville, Maryland.
- Fitch, E., Harper, E., Skorcheva, I., Kurtz, S. E., & Blauvelt, A. (2007). Pathophysiology of psoriasis: Recent advances on IL-23 and Th17 cytokines. *Current Rheumatology Reports*, 9(6), 461-467. doi:10.1007/s11926-007-0075-1
- Fits, L. V., Mourits, S., Voerman, J. S., Kant, M., Boon, L., Laman, J. D., . . . Lubberts, E. (2009). Imiquimod-Induced Psoriasis-Like Skin Inflammation in Mice Is Mediated via the IL-23/IL-17 Axis. *The Journal of Immunology*, 182(9), 5836-5845. doi:10.4049/jimmunol.0802999
- Fredriksson, T., & Pettersson, U. (1978). Severe Psoriasis – Oral Therapy with a New Retinoid. *Dermatology*, 157(4), 238-244. doi:10.1159/000250839
- Fry, L., Baker, B. S., Powles, A. V., & Engstrand, L. (2014). Psoriasis is not an autoimmune disease? *Experimental Dermatology*, 24(4), 241-244. doi:10.1111/exd.12572
- Fu, Z. F., Tu, Z. C., Zhang, L., Wang, H., Wen, Q. H., & Huang, T. (2016). Antioxidant activities and polyphenols of sweet potato (Ipomoea batatas L.) leaves extracted with solvents of various polarities. *Food Bioscience*, 15, 11-18.
- Gearing, A. (1990). Cytokines in skin lesions of psoriasis. *Cytokine*, 2(1), 68-75. doi:10.1016/1043-4666(90)90045-u
- Geherin, S. A., Fintushel, S. R., Lee, M. H., Wilson, R. P., Patel, R. T., Alt, C., . . . Debes, G. F. (2012). The Skin, a Novel Niche for Recirculating B Cells. *The Journal of Immunology*, 188(12), 6027-6035. doi:10.4049/jimmunol.1102639
- Gelfand, J. M., Weinstein, R., Porter, S. B., Neumann, A. L., Berlin, J. A., & Margolis, D. J. (2005). Prevalence and Treatment of Psoriasis in the United Kingdom. *Archives of Dermatology*, 141(12). doi:10.1001/archderm.141.12.1537

- Ghasemzadeh, A. (2012). Polyphenolic content and their antioxidant activity in leaf extract of sweet potato (*Ipomoea batatas*). *Journal of Medicinal Plants Research*, 6(15). doi:10.5897/jmpr11.1353
- Ghoreschi, K., Weigert, C., & Röcken, M. (2007). Immunopathogenesis and role of T cells in psoriasis. *Clinics in Dermatology*, 25(6), 574-580.
- Global Report on Psoriasis-WHO (2016). URL <http://apps.who.int/iris/bitstream/10665/204417/1/9789241565189>
- Gudjonsson, J. E., Johnston, A., Dyson, M., Valdimarsson, H., & Elder, J. T. (2007). Mouse models of psoriasis. *Journal of Investigative Dermatology*, 127(6), 1292-1308.
- Gunderson, A. J., Mohammed, J., Horvath, F. J., Podolsky, M. A., Anderson, C. R., & Glick, A. B. (2013). CD8+ T cells mediate RAS-induced psoriasis-like skin inflammation through IFN- $\gamma$ . *Journal of Investigative Dermatology*, 133(4), 955-963.
- Henseler, T. (1998). Genetics of psoriasis. *Archives of Dermatological Research*, 290(9), 463-476. doi:10.1007/s004030050338
- Hue, S., Boyce, A. N., & Somasundram, C. (2011). Comparative Study on the Antioxidant Activity of Leaf Extract and Carotenoids Extract from *Ipomoea batatas* var. Oren (Sweetpotato) Leaves. *World Academy of Science, Engineering and Technology*, 58, 584-587.
- Hwang, S. J., Kim, Y. W., Park, Y., Lee, H. J., & Kim, K. W. (2014). Anti-inflammatory effects of chlorogenic acid in lipopolysaccharide-stimulated RAW 264.7 cells. *Inflammation Research*, 63(1), 81-90.
- Icen, M., Crowson, C. S., McEvoy, M. T., Dann, F. J., Gabriel, S. E., & Kremers, H. M. (2009). Trends in incidence of adult-onset psoriasis over three decades: A population-based study. *Journal of the American Academy of Dermatology*, 60(3), 394-401. doi:10.1016/j.jaad.2008.10.062
- Ikäheimo, I., Tiilikainen, A., Karvonen, J., & Silvennoinen-Kassinen, S. (1996). HLA risk haplotype Cw6, DR7, DQA1\*0201 and HLA-Cw6 with reference to the clinical picture of psoriasis vulgaris. *Archives of Dermatological Research*, 288(7), 363-365. doi:10.1007/bf02507104
- Ishiguro, K., Yoshimoto, M., Suzuki, M., & Yahara, S. (2008). Anti-Oxidative Activity In The Lipophilic Fraction Of Sweet Potato Tubers. *Acta Horticulturae*, (768), 571-577. doi:10.17660/actahortic.2008.768.77
- Islam S. (2006). Sweetpotato (*Ipomea batatas* L.) leaf: its potential effect on human health and nutrition. *Journal of Food Science* 71:R13–8

- Jacobson, C. C., Kumar, S., & Kimball, A. B. (2011). Latitude and psoriasis prevalence. *Journal of the American Academy of Dermatology*, 65(4), 870-873. doi:10.1016/j.jaad.2009.05.047
- Jin, Y. R., Lee, M. S., Lee, J. H., Hsu, H. K., Lu, J. Y., Chao, S. S., ... & Ger, L. P. (2007). Intake of vitamin A-rich foods and lung cancer risk in Taiwan: with special reference to garland chrysanthemum and sweet potato leaf consumption. *Asia Pacific Journal of Clinical Nutrition*, 16(3), 477-488.
- Johnson, M., & Pace, R. D. (2010). Sweet potato leaves: properties and synergistic interactions that promote health and prevent disease. *Nutrition Reviews*, 68(10), 604-615.
- Kai, H., Akamatsu, E., Torii, E., Kodama, H., Yukizaki, C., Sakakibara, Y., ... & Matsuno, K. (2011). Inhibition of proliferation by agricultural plant extracts in seven human adult T-cell leukaemia (ATL)-related cell lines. *Journal of Natural Medicines*, 65(3-4), 651-655.
- Kamili, Q. U., & Menter, A. (2009). Topical Treatment of Psoriasis. *Management of Psoriasis Current Problems in Dermatology*, 37-58. doi:10.1159/000232303
- Karczewski, J., Dobrowolska, A., Rychlewska-Hańczewska, A., & Adamski, Z. (2016). New insights into the role of T cells in pathogenesis of psoriasis and psoriatic arthritis. *Autoimmunity*, 49(7), 435-450.
- Karna, P., Gundala, S. R., Gupta, M. V., Shamsi, S. A., Pace, R. D., Yates, C., . . . Aneja, R. (2011). Polyphenol-rich sweet potato greens extract inhibits proliferation and induces apoptosis in prostate cancer cells in vitro and in vivo. *Carcinogenesis*, 32(12), 1872-1880. doi:10.1093/carcin/bgr215
- Krueger, J. G. (2005). Psoriasis pathophysiology: current concepts of pathogenesis. *Annals of the Rheumatic Diseases*, 64(Suppl\_2), II30-II36. doi:10.1136/ard.2004.031120
- Kusano, S. (2000). Antidiabetic activity of white skinned sweet potato (*Ipomoea batatas* L.) in obese Zucker fatty rats. *Biological and Pharmaceutical Bulletin*, 23(1), 23-26.
- Lako, J., Wattanapenpaiboon, N., Wahlqvist, M., & Trenerry, C. (2006). Phytochemical intakes of the Fijian population. *Asia Pacific Journal of Clinical Nutrition*, 15(2), 275.
- Langley, R. (2012). Exploring new concepts in the successful management of psoriasis. *Journal of the European Academy of Dermatology and Venereology*, 26, 1-2. doi:10.1111/j.1468-3083.2011.04409.x
- Lebot, V., Ndiaye, A., & Malapa, R. (2011). Phenotypic characterization of sweet potato [*Ipomoea batatas* (L.) Lam.] genotypes in relation to prediction of chemical quality constituents by NIRS equations. *Plant Breeding*, 130(4), 457-463. doi:10.1111/j.1439-0523.2010.01840.x

- Lebwohl, M. (2003). Psoriasis. *The Lancet*, 361(9364), 1197-1204.  
doi:10.1016/s0140-6736(03)12954-6
- Leonardi, C. L., Powers, J. L., Matheson, R. T., Goffe, B. S., Zitnik, R., Wang, A., & Gottlieb, A. B. (2003). Etanercept as monotherapy in patients with psoriasis. *New England Journal of Medicine*, 349(21), 2014-2022.
- Lew, B., Cho, Y., Kim, J., Sim, W., & Kim, N. (2006). Ceramides and Cell Signaling Molecules in Psoriatic Epidermis: Reduced Levels of Ceramides, PKC- $\alpha$ , and JNK. *Journal of Korean Medical Science*, 21(1), 95.  
doi:10.3346/jkms.2006.21.1.95
- Lew, W., Bowcock, A. M., & Krueger, J. G. (2004). Psoriasis vulgaris: cutaneous lymphoid tissue supports T-cell activation and 'Type 1' inflammatory gene expression. *Trends in Immunology*, 25(6), 295-305.  
doi:10.1016/j.it.2004.03.006
- Lin, Y., Wong, W., & Pang, J. S. (2006). Successful treatment of recalcitrant psoriasis with Indigo naturalis ointment. *Clinical and Experimental Dermatology*, 0(0).  
doi:10.1111/j.1365-2230.2006.02229.x
- Liu, M., Song, S., Li, H., Jiang, X., Yin, P., Wan, C., ... & Xu, J. (2014). The protective effect of caffeic acid against inflammation injury of primary bovine mammary epithelial cells induced by lipopolysaccharide. *Journal of Dairy Science*, 97(5), 2856-2865.
- Lowe, N. J., Breeding, J., Kean, C., & Cohn, M. (1981). Psoriasiform Dermatoses in a Rhesus Monkey. *Journal of Investigative Dermatology*, 76(2), 141-143.  
doi:10.1111/1523-1747.ep12525484
- Lowes, M. A., Suárez-Fariñas, M., & Krueger, J. G. (2014). Immunology of psoriasis. *Annual Review of Immunology*, 32, 227-255.
- Ludvik, B. H., Mahdjoobian, K., Waldhaeusl, W., Hofer, A., Prager, R., Kautzky-Willer, A., & Pacini, G. (2002). The effect of Ipomoea batatas (caipó) on glucose metabolism and serum cholesterol in patients with type 2 diabetes. *Diabetes Care*, 25(1), 239-240.
- Ludvik, B., Neuffer, B. & Pacini, G. 2004. Efficacy of Ipomoea batatas (Caiapo) on diabetes control in type 2 diabetic subjects treated with diet. *Diabetes Care* 27: 436-440.
- Lynde, C. W., Poulin, Y., Vender, R., Bourcier, M., & Khalil, S. (2014). Interleukin 17A: Toward a new understanding of psoriasis pathogenesis. *Journal of the American Academy of Dermatology*, 71(1), 141-150.  
doi:10.1016/j.jaad.2013.12.036

- Martin, D. A., Towne, J. E., Kricorian, G., Klekotka, P., Gudjonsson, J. E., Krueger, J. G., & Russell, C. B. (2013). The Emerging Role of IL-17 in the Pathogenesis of Psoriasis: Preclinical and Clinical Findings. *Journal of Investigative Dermatology*, 133(1), 17-26. doi:10.1038/jid.2012.194
- McColl, I. (2009). Retrieved on 08 Feb 2017  
[http://teachingdermatopathology.blogspot.my/2009\\_12\\_01\\_archive.html](http://teachingdermatopathology.blogspot.my/2009_12_01_archive.html)
- Mcfadden, J., Baker, B., Powles, A., & Fry, L. (2009). Psoriasis and streptococci: the natural selection of psoriasis revisited. *British Journal of Dermatology*, 160(5), 929-937. doi:10.1111/j.1365-2133.2009.09102.x
- Meira, M., da Silva, P., David, J.M. & David, J.P. 2012. Review of the genus Ipomoea: traditional uses, chemistry and biological activities. *Revista Brasileira de Farmacognosia Brazilian Journal of Pharmacognosy* 22(3): 682-713
- Michalak-Stoma, A., Bartosińska, J., Kowal, M., Juszkiwicz-Borowiec, M., Gerkowicz, A., & Chodorowska, G. (2013). Serum levels of selected Th17 and Th22 cytokines in psoriatic patients. *Disease Markers*, 35(6), 625-631.
- Milenkovic, D., Jude, B., & Morand, C. (2013). miRNA as molecular target of polyphenols underlying their biological effects. *Free Radical Biology and Medicine*, 64, 40-51.
- Mrowietz, U., Guérin, A., Mulani, P., & Gupta, S. (2010). Comorbidity prevalence in psoriasis patients: A meta-analysis. *Journal of the American Academy of Dermatology*, 62(3), AB123.
- Naik, S., Bouladoux, N., Wilhelm, C., Molloy, M. J., Salcedo, R., Kastenmuller, W., . . . Belkaid, Y. (2012). Compartmentalized Control of Skin Immunity by Resident Commensals. *Science*, 337(6098), 1115-1119. doi:10.1126/science.1225152
- Naldi, L., Svensson, A., Diepgen, T., Elsner, P., Grob, J., Coenraads, P., . . . Williams, H. (2003). Randomized Clinical Trials for Psoriasis 1977–2000: The EDEN Survey. *Journal of Investigative Dermatology*, 120(5), 738-741. doi:10.1046/j.1523-1747.2003.12145.x
- Nestle, F. O., & Nickoloff, B. J. (2006). Animal models of psoriasis: a brief update. *Journal of the European Academy of Dermatology and Venereology*, 20(s2), 24-27.
- Nestle, F. O., Kaplan, D. H., & Barker, J. (2009). Psoriasis. *New England Journal of Medicine*, 361(5), 496-509. doi:10.1056/nejmra0804595
- Oka, A., Mabuchi, T., Ozawa, A., & Inoko, H. (2012). Current understanding of human genetics and genetic analysis of psoriasis. *The Journal of Dermatology*, 39(3), 231-241. doi:10.1111/j.1346-8138.2012.01504.x
- Olthof, M. R., Hollman, P. C., & Katan, M. B. (2001). Chlorogenic acid and caffeic acid are absorbed in humans. *The Journal of Nutrition*, 131(1), 66-71.

- Padda, M. S., & Picha, D. H. (2007). Antioxidant activity and phenolic composition in 'Beauregard'sweetpotato are affected by root size and leaf age. *Journal of the American Society for Horticultural Science*, 132(4), 447-451.
- Parisi, R., Symmons, D. P., Griffiths, C. E., & Ashcroft, D. M. (2013). Global Epidemiology of Psoriasis: A Systematic Review of Incidence and Prevalence. *Journal of Investigative Dermatology*, 133(2), 377-385. doi:10.1038/jid.2012.339
- Park, K. H., Kim, J. R., Lee, J. S., Lee, H., & Cho, K. H. (2010). Ethanol and water extract of purple sweet potato exhibits anti-atherosclerotic activity and inhibits protein glycation. *Journal of Medicinal Food*, 13(1), 91-98.
- Pasparakis, M., Haase, I., & Nestle, F. O. (2014). Mechanisms regulating skin immunity and inflammation. *Nature Reviews Immunology*, 14(5), 289-301. doi:10.1038/nri3646
- Patel, T., & Gordon, K. B. (2004). Adalimumab: efficacy and safety in psoriasis and rheumatoid arthritis. *Dermatologic Therapy*, 17(5), 427-431.
- Paul, C., Gourraud, P., Bronsard, V., Prey, S., Puzenat, E., Aractingi, S., . . . Ortonne, J. (2010). Evidence-based recommendations to assess psoriasis severity: systematic literature review and expert opinion of a panel of dermatologists. *Journal of the European Academy of Dermatology and Venereology*, 24, 2-9. doi:10.1111/j.1468-3083.2009.03561.x
- Pochapski, M.T., Fosquiera, E.C., Esmerino, L.A., dos Santos, E.B., Farago, P.V., Santos, S.A. & Groppo, F.C. (2011). Phytochemical screening, antioxidant and antimicrobial activities from the crude leaves' extract from *Ipomoea batatas* (L.) Lam. *Pharmacognosy Magazine* 7(26): 165-170
- Preclinical and Clinical Findings. *Journal of Investigative Dermatology*, 133(1), 17-26. doi:10.1038/jid.2012.194
- Rabah, I. O., Hou, D. X., Komine, S. I., & Fujii, M. (2004). Potential chemopreventive properties of extract from baked sweet potato (*Ipomoea batatas* Lam. Cv. Koganesengan). *Journal of Agricultural and Food Chemistry*, 52(23), 7152-7157.
- Ragaz, A., & Ackerman, A. B. (1979). Evolution, maturation, and regression of lesions of psoriasis: new observations and correlation of clinical and histologic findings. *The American Journal of Dermatopathology*, 1(3), 199-214.
- Rather, I. A., Bajpai, V. K., Han, J., & Nam, G. J. (2016). Imiquimod-induced psoriasis-like skin inflammation in mouse model. *Bangladesh Journal of Pharmacology*, 11(4), 849-851.
- Raychaudhuri, S., & Farber, E. (2001). The prevalence of psoriasis in the world. *Journal of the European Academy of Dermatology and Venereology*, 15(1), 16-17. doi:10.1046/j.1468-3083.2001.00192.x

- Rohloff, J. (2015). Analysis of Phenolic and Cyclic Compounds in Plants Using Derivatization Techniques in Combination with GC-MS-Based Metabolite Profiling. *Molecules*, 20(2), 3431-3462. doi:10.3390/molecules20023431
- Sano, S., Chan, K. S., Carbajal, S., Clifford, J., Peavey, M., Kiguchi, K., . . . Digiovanni, J. (2004). Stat3 links activated keratinocytes and immunocytes required for development of psoriasis in a novel transgenic mouse model. *Nature Medicine*, 11(1), 43-49. doi:10.1038/nm1162
- Sato, Y., Itagaki, S., Kurokawa, T., Ogura, J., Kobayashi, M., Hirano, T., . . . & Iseki, K. (2011). In vitro and in vivo antioxidant properties of chlorogenic acid and caffeic acid. *International Journal of Pharmaceutics*, 403(1), 136-138.
- Schön, M. P. (1999). Animal models of psoriasis—what can we learn from them?. *Journal of Investigative Dermatology*, 112(4), 405-410.
- Singleton, V. L., Orthofer, R., & Lamuela-Raventós, R. M. (1999). [14] Analysis of total phenols and other oxidation substrates and antioxidants by means of folin-ciocalteu reagent. *Methods in Enzymology*, 299, 152-178.
- Sinniah B, Saraswathy Devi S, Prashant BS (2010). Epidemiology of psoriasis in Malaysia: a hospital based study. *Medical Journal of Malaysia*. 2010 Jun; 65(2):112-4)
- Soler, D. C., & McCormick, T. S. (2011). The Dark Side of Regulatory T Cells in Psoriasis. *Journal of Investigative Dermatology*, 131(9), 1785-1786. doi:10.1038/jid.2011.200
- Strohal, R., Kirby, B., & Puig, L. (2013). Psoriasis beyond the skin: an expert group consensus on the management of psoriatic arthritis and common comorbidities in patients with moderate-to-severe psoriasis. *Journal of the European Academy of Dermatology and Venereology*, 28(12), 1661-1669. doi:10.1111/jdv.12350
- Suda, I., Oki, T., Masuda, M., Kobayashi, M., Nishiba, Y., & Furuta, S. (2003). Physiological functionality of purple-fleshed sweet potatoes containing anthocyanins and their utilization in foods. *Japan Agricultural Research Quarterly: JARQ*, 37(3), 167-173.
- Sugiyama, H., Gyulai, R., Toichi, E., Garaczi, E., Shimada, S., Stevens, S. R., . . . Cooper, K. D. (2004). Dysfunctional Blood and Target Tissue CD4 CD25high Regulatory T Cells in Psoriasis: Mechanism Underlying Unrestrained Pathogenic Effector T Cell Proliferation. *The Journal of Immunology*, 174(1), 164-173.
- Sundberg, J. P., France, M., Boggess, D., Sundberg, B. A., Jenson, A., Beamer, W. G., & Shultz, L. D. (1997). Development and Progression of Psoriasiform Dermatitis and Systemic Lesions in the Flaky Skin (fsn) Mouse Mutant. *Pathobiology*, 65(5), 271-286.

- Teow, C.C., Truong, V.D., McFeeters, R.F., Thompson, R.L., Pecota, K.V. & Yencho, G.C. (2007). Antioxidant activities, phenolic and  $\beta$ -carotene contents of sweet potato genotypes with varying flesh colours. *Food Chemistry* 103: 829-838.
- Tollefson, M. M., Crowson, C. S., McEvoy, M. T., & Kremers, H. M. (2010). Incidence of psoriasis in children: A population-based study. *Journal of the American Academy of Dermatology*, 62(6), 979-987. doi:10.1016/j.jaad.2009.07.029
- Truong, V-D. & Avula, R.Y. (2010). Sweet potato purees and powders for functional food ingredients. In *Sweet potato: Post Harvest Aspects in Food*. p112-161. Nova Science Publishers, Inc. New York.
- Valdimarsson, H., Bake, B. S., Jónsdóttir, I., & Fry, L. (1986). Psoriasis: a disease of abnormal Keratinocyte proliferation induced by T lymphocytes. *Immunology Today*, 7(9), 256-259. doi:10.1016/0167-5699(86)90005-8
- Valdimarsson, H., Thorleifsdottir, R. H., Sigurdardottir, S. L., Gudjonsson, J. E., & Johnston, A. (2009). Psoriasis – as an autoimmune disease caused by molecular mimicry. *Trends in Immunology*, 30(10), 494-501. doi:10.1016/j.it.2009.07.00
- van der Fits, L., Mourits, S., Voerman, J. S., Kant, M., Boon, L., Laman, J. D., ... & Lubberts, E. (2009). Imiquimod-induced psoriasis-like skin inflammation in mice is mediated via the IL-23/IL-17 axis. *The Journal of Immunology*, 182(9), 5836-5845.
- Walter, W. M., & Catignani, G. L. (1981). Biological quality and composition of sweet potato protein fractions. *Journal of Agricultural and Food Chemistry*, 29(4), 797-799. doi:10.1021/jf00106a027
- Warren, R. B., Kleyn, C. E., & Gulliver, W. P. (2011). Cumulative life course impairment in psoriasis: patient perception of disease-related impairment throughout the life course. *British Journal of Dermatology*, 164(s1), 1-14.
- Wasmund, N., Topp, I. & Schories, D. (2006). Optimising the storage and extraction of chlorophyll samples. *Oceanologia* 48(1): 125-144.
- Watzl, B., Bub, A., Brandstetter, B. R., & Rechkemmer, G. (1999). Modulation of human T-lymphocyte functions by the consumption of carotenoid-rich vegetables. *British Journal of Nutrition*, 82(5), 383-389.
- WHO Global report on PSORIASIS, 2016-WHO. (n.d.). Retrieved October 8, 2016, from <http://apps.who.int/iris/bitstream/10665/204417/1/9789241565189eng.pdf>
- Wilson, C. D., Pace, R. D., Bromfield, E., Jones, G., & Lu, J. Y. (1998). Sweet potato in a vegetarian menu plan for NASA's Advanced Life Support Program. *Life Support & Biosphere Science*, 5(3), 347-351.

- Woodrow, J. C., & Ilchysyn, A. (1985). HLA antigens in psoriasis and psoriatic arthritis. *Journal of Medical Genetics*, 22(6), 492-495. doi:10.1136/jmg.22.6.492
- Wu, J. K., Siller, G., & Strutton, G. (2004). Psoriasis induced by topical imiquimod. *Australasian Journal of Dermatology*, 45(1), 47-50.
- Xu, W., Liu, L., Hu, B., Sun, Y., Ye, H., Ma, D., & Zeng, X. (2010). TPC in the leaves of 116 sweet potato (*Ipomoea batatas* L.) varieties and Pushu 53 leaf extracts. *Journal of Food Composition and Analysis*, 23(6), 599-604.
- Zanolli, M. D., Jayo, M. J., Jayo, J. M., Blaine, D., Hall, J., & Jorizzo, J. L. (1988). Evaluation of psoriatic plaques that spontaneously developed in a cynomolgus monkey (*Macaca fascicularis*). *Acta Dermato-venereologica Supplementum*, 146, 58-58.