



UNIVERSITI PUTRA MALAYSIA

***METABOLIC EFFECTS OF *Cosmos caudatus* Kunth (ULAM RAJA)
SUPPLEMENTATION IN TYPE-2 DIABETES MELLITUS PATIENTS***

CHENG SHI HUI

FPSK(P) 2017 3



**METABOLIC EFFECTS OF *Cosmos caudatus* Kunth (ULAM RAJA)
SUPPLEMENTATION IN TYPE-2 DIABETES MELLITUS PATIENTS**

By

CHENG SHI HUI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

January 2017

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

**METABOLIC EFFECTS OF *Cosmos caudatus* Kunth (ULAM RAJA)
SUPPLEMENTATION IN TYPE-2 DIABETES MELLITUS PATIENTS**

By

CHENG SHI HUI

January 2017

Chair : Barakatun Nisak Mohd Yusof, PhD

Faculty : Medicine and Health Sciences

Cosmos caudatus, or locally known as “*Ulam Raja*” is a medicinal plant in Southeast Asia countries with reported medicinal benefits. Previously, supplementation with *C. caudatus* extract was found able to reduce plasma blood glucose in rats, but its effect in patients with type 2 diabetes mellitus (T2DM) was not established. To address this research gap, the present study aimed to determine the metabolic effects of *C. caudatus* in T2DM patients. The study was started by identifying the best way of *C. caudatus* supplementations. The first aim was to determine the antioxidant capacity of *C. caudatus* leaf extracts and juice using different extraction solvents (100% methanol, 100% ethanol, 95% ethanol, 50% ethanol). The findings found that *C. caudatus* leaves extracts had higher antioxidant capacity than *C. caudatus* juice. Following the first study, a two-arm randomized controlled clinical trial was carried out to determine the effectiveness and safety of *C. caudatus* supplementation in T2DM patients.

A total of 101 T2DM patients (age: 49.7 ± 9.1 years; mean HbA1C: 8.8 ± 1.6 %; BMI: 29.8 ± 4.7 kg/m²; 56% male) were enrolled into the study. Participants were randomly assigned to diabetic-ulam group or diabetic controls. Patients in diabetic-ulam group consumed 15g of raw *C. caudatus* daily for 8 weeks while diabetic controls were abstained from taking *C. caudatus*. Both groups received standard lifestyle interventions. Changes in glycemic control, cardiovascular risk factors (anthropometric, blood pressure, lipid profile, high sensitivity C-reactive protein), renal profile, and liver function were measured at baseline, week 4, week 8 and week 12 (post-intervention follow-up) of the study.

As compared to diabetic controls, *C. caudatus* consumption significantly reduced serum insulin (-1.16 versus $+3.91$ μ U/ml in controls), lowered homeostasis model assessment of insulin resistance (HOMA-IR) (-1.09 versus $+1.34$ unit in controls), and increased quantitative insulin sensitivity check index (QUICKI) ($+0.05$ versus -0.03 unit in

controls) in diabetic-ulam group. Subjects in diabetic-ulam group showed greater improvement in HbA1C (-0.76%) as compared to diabetic controls (-0.37%). Furthermore, supplementation of *C. caudatus* also resulted in the reduction of inflammation marker (hs-CRP) and systolic blood pressure, indicated its beneficial effect on reducing cardiovascular risk factors. Furthermore, *C. caudatus* consumption was found to be safe throughout the duration of the study as evident by no significant difference in liver and renal profile at the end of the study. Other parameters did not change significantly between the two groups.

In addition, a proton nuclear magnetic resonance spectroscopy (^1H NMR) based metabolomics approach was performed to determine the metabolic perturbation following *C. caudatus* consumption in T2DM patients. A total of 39 healthy individuals (age: 38.7 ± 8.5 years; BMI: 22.0 ± 1.7 kg/m²; 44% male) were recruited as healthy controls, and their urine and blood serum metabolic profiles were compared with those obtained from diabetic controls and diabetic-ulam groups. As compared to healthy individuals, the concentrations of urinary lactate, branched-chain amino acids (BCAA, including valine, leucine and isoleucine), alanine, lysine, glutamate, glutamine, and pyruvate were significantly increased in T2DM patients, whereas the concentration of urinary urea was significantly decreased in T2DM patients. In addition, concentrations of blood serum lactate, BCAA (valine, leucine and isoleucine), alanine, lysine, glutamate, and N-acetylglutamate were significantly elevated in T2DM patients as compared to healthy individuals. These findings are consistent with published literature.

Following supplementation with *C. caudatus*, serum concentration of alanine, lactate and N-acetylglutamate were significantly decreased in diabetic-ulam group. The findings indicated a partial reversal of diabetes-induced metabolic changes through altered glycolysis, gluconeogenesis and glutamate metabolism. In addition, supplementation with *C. caudatus* was found to increase the concentration of urinary hippurate in diabetic-ulam group as compared to diabetic controls, suggesting changes in gut microflora metabolism.

In conclusion, the current study provided evidence that supplementation with *C. caudatus* improved insulin sensitivity in T2DM patients, evidenced by improved HOMA-IR and QUICKI parameters. It also improved hs-CRP and systolic blood pressure in T2DM patients, suggesting its effect in reducing the cardiovascular risk factors. The current results also showed that *C. caudatus* did not negatively affect liver and renal functions, suggesting that it is safe for T2DM patients. Furthermore, metabolomic data showed that *C. caudatus* supplementation partially reversed some known diabetes-induced metabolic changes such as lactate and BCAA in blood and urine. In summary, the current study uncovered the beneficial potential of *C. caudatus* for T2DM patients, and a longer term randomized controlled clinical trial is warranted to fully explore its therapeutic potential.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**KESAN METABOLIK SUPPLEMEN *Cosmos caudatus* KUNTH (ULAM RAJA)
DI KALANGAN PESAKIT DIABETES MELLITUS JENIS DUA**

Oleh

CHENG SHI HUI

Januari 2017

Pengerusi : Barakatun Nisak Mohd Yusof, PhD

Fakulti : Perubatan dan Sains Kesihatan

Cosmos caudatus, atau dikenali sebagai “Ulam Raja” adalah sejenis herba yang digunakan untuk tujuan perubatan di negara Asia Tenggara. Sebelum ini, didapati ekstrak *C. caudatus* dapat mengurangkan plasma glukosa darah dalam model tikus, tetapi kesannya di kalangan pesakit diabetes melitus jenis dua (T2DM) masih tidak diketahui. Untuk menangani jurang ini, kajian ini bertujuan untuk menentukan kesan metabolik supplemen *C. caudatus* di kalangan pesakit T2DM. Kajian ini dimulakan dengan mengenal pasti cara terbaik untuk supplemen *C. caudatus*. Matlamat pertama adalah untuk menentukan kapasiti antioksidan daun ekstrak *C. caudatus* dan jus menggunakan pelarut pengekstrakan yang berbeza (100% methanol, 100% ethanol, 95% ethanol, 50% ethanol). Hasil kajian mendapati bahawa ekstrak *C. caudatus* mengandungi antioksidan yang lebih tinggi daripada jus *C. caudatus*. Lanjutan dari kajian pertama, kajian kedua iaitu klinikal terkawal secara rawak telah dijalankan untuk menentukan keberkesanan dan keselamatan supplemen *C. caudatus* di kalangan pesakit T2DM.

Seramai 101 pesakit T2DM (umur : 49.7 ± 9.1 tahun; min HbA1C: 8.8 ± 1.6 %; BMI: 29.8 ± 4.7 kg/m²; 56% lelaki) telah menyertai kajian ini. Peserta dibahagikan secara rawak kepada kumpulan diabetes-ulam atau kumpulan kawalan diabetes. Subjek dalam kumpulan diabetes-ulam mengambil 15g *C. caudatus* setiap hari selama lapan minggu manakala kumpulan kawalan diabetes telah dikecualikan daripada pengambilan *C. caudatus*. Kedua-dua kumpulan menerima nasihat gaya hidup yang sama. Perubahan dalam kawalan glisemik, faktor risiko kardiovaskular (antropometri, tekanan darah, profil lipid, kepekaan tinggi protein C-reaktif), profil buah pinggang dan fungsi hati diukur pada minggu 0, minggu 4, minggu 8 dan minggu 12 (kajian susulan).

Berbanding dengan kumpulan kawalan diabetes, pengambilan *C. caudatus* dapat mengurangkan serum insulin (-1.16 berbanding $+3.91$ μ U/ml dalam kawalan), menurunkan penilaian model homeostasis rintangan insulin (HOMA-IR) (-1.09

berbanding +1.34 unit dalam kawalan), dan meningkatkan pengambilan kuantitatif indeks insulin sensitiviti (QUICKI) (+0.05 berbanding -0.03 unit dalam kawalan) dalam kumpulan diabetes-ulam. Pesakit dalam kumpulan diabetes-ulam menunjukkan penurunan yang lebih banyak dari segi tahap HbA1c (-0.76 %) berbanding dengan kumpulan kawalan diabetes (-0.37 %). Tambahan lagi, supplemen *C. caudatus* juga menyebabkan pengurangan penanda keradangan (hs-CRP) dan tekanan darah sistolik, menunjukkan *C. caudatus* mempunyai kesan yang baik dalam mengurangkan faktor risiko kardiovaskular. Pengambilan *C. caudatus* didapati selamat sepanjang tempoh kajian kerana tiada perbezaan yang signifikan dalam profil hati dan buah pinggang pada akhir kajian. Parameter lain tidak berubah secara signifikan antara kedua-dua kumpulan.

Di samping itu, pendekatan metabolomik menggunakan spektroskopi nuklear magnet resonans proton (^1H NMR) telah dijalankan untuk menentukan perubahan metabolit selepas supplemen *C. caudatus* di kalangan pesakit T2DM. Sejumlah 39 individu yang sihat (umur: 38.7 ± 8.5 tahun; BMI: 22.0 ± 1.7 kg/m²; 44% lelaki) telah diambil sebagai kawalan, dan profil metabolik urin and darah mereka dibandingkan dengan kumpulan kawalan diabetes dan kumpulan diabetes-ulam. Berbanding dengan individu yang sihat, kepekatan laktat, asid amino rantai bercabang (BCAA, termasuk valine, leucine, dan isoleucine), alanin, lisin, glutamat, glutamin, dan piruvat dalam urin telah meningkat dengan ketara dalam pesakit T2DM, manakala kepekatan urea dalam urin telah menurun dengan ketara dalam pesakit T2DM. Di samping itu, kepekatan laktat, BCAA, alanin, lisin, glutamat dan N-acetylglutamate dalam serum darah meningkat dengan ketara dalam pesakit T2DM berbanding individu yang sihat. Penemuan ini adalah konsisten dengan kertas jurnal yang diterbitkan.

Selepas supplemen dengan *C. caudatus*, kepekatan serum darah alanin, laktat dan N-acetylglutamate telah menurun dengan ketara dalam pesakit kumpulan diabetes-ulam. Dapatan kajian ini menunjukkan terdapat perubahan secara separa dalam metabolik diabetes melalui metabolisme glikolisis, glukoneogenesis dan glutamat. Di samping itu, supplemen *C. caudatus* didapati meningkatkan kepekatan hippurate urin dalam kumpulan diabetes-ulam berbanding dengan kumpulan kawalan diabetes, menunjukkan perubahan dalam metabolisme usus mikroflora.

Kesimpulannya, kajian ini membuktikan bahawa supplemen *C. caudatus* dapat meningkatkan sensitiviti insulin dalam pesakit T2DM melalui HOMA-IR dan QUICKI parameter. Ia juga mengurangkan hs-CRP dan tekanan darah sistolik dalam pesakit T2DM, menunjukkan kesannya dalam mengurangkan faktor risiko kardiovaskular. Kajian ini juga menunjukkan bahawa supplemen *C. caudatus* tidak memberikan kesan negatif terhadap fungsi hati dan profil buah pinggang, menunjukkan bahawa ia adalah selamat untuk pesakit T2DM. Tambahan pula, data metabolomik menunjukkan bahawa supplemen *C. caudatus* dapat mengubah sebahagian metabolik seperti laktat dan BCAA dalam darah dan urin. Secara ringkasnya, kajian ini menunjukkan supplemen *C. caudatus* adalah berfaedah untuk pesakit T2DM, dan satu percubaan klinikal terkawal dan rawak bagi tempoh masa yang lebih lama adalah wajar dijalankan untuk menerokai potensi terapeutik *C. caudatus* dengan sepenuhnya.

ACKNOWLEDGEMENTS

I would like to convey my deepest gratitude to the following remarkable individuals who helped in making my thesis possible. First and foremost, sincere thanks to my supervisor, Dr. Barakatun Nisak Mohd Yusof for her guidance and support throughout the research. I am grateful to Prof. Dr. Amin Ismail for his support and generosity in sharing with me his knowledge. I also thank Dr. Alfi Khatib for his helpful suggestions. In addition, I would like to express my gratitude to Dr. Joseph Anthony and Dr Ng Ooi Chuan for kindly reviewed the subjects throughout the study.

I am truly grateful to be the recipient of the Mybrain Myphd scholarship under Ministry of Higher Education Malaysia. I would like to acknowledge the grant from Agrobiotechnology Institute Malaysia for funding this research. In addition, I would like to extend my sincere gratitude to all the subjects who participated in this clinical trial for their time, support and co-operation. Special thanks to all the staffs in medical clinic Hospital Serdang for their kind assistance and support throughout the research.

My appreciation also goes to all my friends for their moral support and encouragement throughout the research journey. Last but not least, my sincere gratitude goes to my beloved family members who have always been my tower of strength and source of inspiration throughout my research.

I certify that a Thesis Examination Committee has met on 6th January 2017 to conduct the final examination of Cheng Shi Hui on her thesis entitled “Metabolic effects of *Cosmos caudatus* Kunth (Ulam Raja) supplementation in Type-2 Diabetes Mellitus patients” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Azrina Azlan, PhD

Associate Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Chairman)

Norhaizan Mohd Esa, PhD

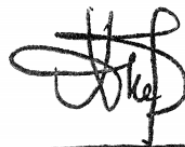
Associate Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Internal Examiner)

Loh Su Peng, PhD

Associate Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Internal Examiner)

Manohar Garg, PhD

Professor
University of Newcastle
Australia
(External Examiner)



NOR AINI AB. SHUKOR, PhD

Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 22 March 2017

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Barakatun Nisak Mohd Yusof, PhD

Associate Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Chairperson)

Amin Ismail, PhD

Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

Joseph Anthony, M.B.B.S

Senior Medical Lecturer
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

Alfi Khatib, PhD

Associate Professor
Faculty of Pharmacy
International Islamic University Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: _____

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____
Name of Chairman of
Supervisory
Committee: Barakatun Nisak Mohd Yusof, PhD

Signature: _____
Name of Member of
Supervisory
Committee: Amin Ismail, PhD

Signature: _____
Name of Member of
Supervisory
Committee: Joseph Anthony, M.B.B.S

Signature: _____
Name of Member of
Supervisory
Committee: Alfi Khatib, PhD

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xv
LIST OF FIGURES	xvi
LIST OF APPENDICES	xviii
LIST OF ABBREVIATIONS	xix
CHAPTER	
1 INTRODUCTION	1
1.1 Research Background	1
1.2 Statement of Problem	2
1.3 Significance of the study	3
1.4 General Objective	3
1.5 Specific Objectives	3
1.6 Research conceptual framework	4
2 LITERATURE REVIEW	6
2.1 Overview of diabetes mellitus	6
2.2 Epidemiology of diabetes mellitus	6
2.3 Clinical pathophysiology of T1DM	7
2.4 Clinical pathophysiology of T2DM	7
2.5 Impact of T2DM	10
2.6 Diagnostic criteria for T2DM	10
2.7 Management of T2DM	11
2.8 Lifestyle intervention	12
2.9 Current drug treatment for T2DM	12
2.10 Therapeutic effect of Malaysian herbs	14
2.11 <i>Cosmos caudatus</i> (Ulam Raja)	16
2.11.1 Mechanism of action of <i>C. caudatus</i> on glucose metabolism	18
2.11.2 Safety of <i>C. caudatus</i> consumption	19
2.12 Health benefits of polyphenols	20
2.13 Antioxidant compounds and their effect on glucose metabolism	21
2.14 Metabolomics	23
2.14.1 Metabolomics application in T2DM	26
3 ANTIOXIDANT CAPACITY OF <i>Cosmos caudatus</i> EXTRACTED USING DIFFERENT SOLVENTS	29

3.1	Introduction	29
3.2	Materials and Methods	29
3.2.1	Solvents and chemicals	29
3.2.2	Plant material	30
3.2.3	Preparation of <i>C. caudatus</i> extracts and juice	30
3.2.4	Determination of total phenolic content	30
3.2.5	Determination of total flavonoid content	30
3.2.6	DPPH radical scavenging assay	31
3.2.7	ABTS radical scavenging assay	31
3.2.8	Ferric reducing antioxidant power (FRAP) assay	32
3.2.9	HPLC determination of ascorbic acid	32
3.2.10	Statistical analysis	32
3.3	Results	33
3.3.1	Extraction yield	33
3.3.2	Total phenolic content	33
3.3.3	Total flavonoid content	34
3.3.4	DPPH radical scavenging assay	34
3.3.5	ABTS radical scavenging assay	35
3.3.6	Ferric reducing antioxidant power (FRAP) assay	35
3.3.7	Ascorbic acid equivalent antioxidant capacity (AEAC)	36
3.3.8	HPLC determination of ascorbic acid content	37
3.3.9	Correlation analysis	37
3.4	Discussion	38
3.5	Conclusion	39
4	EFFECT OF <i>Cosmos caudatus</i> (ULAM RAJA) SUPPLEMENTATION IN T2DM PATIENTS: A RANDOMIZED CONTROLLED TRIAL	40
4.1	Introduction	40
4.2	Materials and Methods	41
4.2.1	Trial design	41
4.2.2	Ethical approval and funding	41
4.2.3	Sample size determination	41
4.2.4	Screening and recruitment	42
4.2.5	Inclusion criteria	42
4.2.6	Exclusion criteria	42
4.2.7	Randomization	42
4.2.8	Intervention	43
	4.2.8.1 Diabetic-ulam group	43
	4.2.8.2 Diabetic controls	44
4.2.9	Study visits	45
4.2.10	Adherence	46
4.2.11	Management of withdrawal and drop-out process	46
4.2.12	Safety of <i>C. caudatus</i> consumption in T2DM patients	46
4.2.13	Outcomes measurements	46
4.2.14	Anthropometric measurements	47
4.2.15	Blood pressure measurements	48
4.2.16	Blood sampling	49
4.2.17	Glycemic status assessments	49

4.2.18	Lipid profile and high sensitivity C-reactive protein assessment	50
4.2.19	Renal profile assessments	50
4.2.20	Liver profile assessments	50
4.2.21	Dietary intake assessments	50
4.2.22	Physical activity levels assessment	51
4.2.23	Statistical analysis	52
4.3	Results	53
4.3.1	Subject screening	53
4.3.2	Subject enrollment	54
4.3.3	Baseline characteristics	55
4.3.4	Socio-demographic of the subjects	55
4.3.5	Health status and lifestyle habits of the subjects	56
4.3.6	Anthropometry measurement of the subjects at baseline	58
4.3.7	Body mass index category of the subjects at baseline	59
4.3.8	Blood pressure measurement of the subjects at baseline	60
4.3.9	Metabolic control of the subjects at baseline	60
4.3.10	Dietary intake assessment of the subjects at baseline	63
4.3.11	Evaluation of energy intake using EI:BMR ratio at baseline	64
4.3.12	Physical activity levels of the subjects at baseline	64
4.3.13	Changes in glycemic control over 8 weeks	65
4.3.14	Changes in anthropometry measurements over 8 weeks	70
4.3.15	Changes in blood pressure over 8 weeks	70
4.3.16	Changes in high sensitivity C-reactive protein over 8 weeks	72
4.3.17	Changes in lipid profile over 8 weeks	73
4.3.18	Changes in renal profile over 8 weeks	73
4.3.19	Changes in liver profile over 8 weeks	73
4.3.20	Adverse effect of <i>C. caudatus</i> consumption over 8 weeks	75
4.3.21	Changes in dietary intake and physical activity level over 8 weeks	75
4.3.22	Changes in glycemic control at week 12	77
4.3.23	Changes in metabolic parameters at week 12	77
4.3.24	Changes in renal profile and liver profile at week 12	79
4.3.25	Changes in dietary intake and physical activity at week 12	79
4.4	Discussion	82
4.5	Conclusion	87
5	¹H-NMR-BASED METABOLOMIC PROFILING FOLLOWING <i>Cosmos caudatus</i> CONSUMPTION	88
5.1	Introduction	88
5.2	Materials and Methods	89
5.2.1	Participant selection and study design	89
5.2.2	Blood and urine sampling	91
5.2.3	NMR metabolite profiling in serum	91

5.2.4	NMR metabolite profiling in urine	91
5.2.5	¹ H NMR spectral acquisition and preprocessing	93
5.2.6	Multivariate and univariate data analysis	93
5.3	Results	95
5.3.1	Socio-demographic characteristics of healthy individuals	95
5.3.2	Anthropometry and blood pressure measurements of the three groups	97
5.3.3	Glycemic status and hs-CRP measurements of the three groups	98
5.3.4	Lipid profile measurements of the three groups	99
5.3.5	Renal profile measurements of the three groups	100
5.3.6	Liver profile measurements of the three groups	101
5.3.7	Multivariate analysis of metabolic parameters between healthy individuals and diabetic-controls	101
5.3.8	Multivariate analysis of metabolic parameters between healthy individuals and diabetic-ulam group	103
5.3.9	Multivariate analysis of overall metabolic parameters in the three groups	105
5.3.10	Multivariate analysis of NMR urinary data between healthy individuals and diabetic-controls	107
5.3.11	Multivariate analysis of NMR urinary data between healthy individuals and diabetic-ulam group	109
5.3.12	Multivariate analysis of NMR urinary data in the three groups	111
5.3.13	Urinary NMR spectra and assignment of urinary metabolites	113
5.3.14	Relative concentration of metabolites in NMR urinary data	114
5.3.15	Multivariate analysis of blood serum NMR data between healthy individuals and diabetic-controls	115
5.3.16	Multivariate analysis of blood serum NMR data between healthy individuals and diabetic-ulam group	117
5.3.17	Multivariate analysis of overall blood serum NMR data in all three studied groups	119
5.3.18	Blood serum NMR spectra and assignment of blood serum metabolites	121
5.3.19	Relative concentration of metabolites in blood serum NMR data	122
5.4	Discussion	123
5.5	Conclusion	129
6	GENERAL DISCUSSION	130
7	CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	134
7.1	Conclusion	134
7.2	Study strengths	135
7.3	Study limitations	136
7.4	Recommendations for future work	136

REFERENCES	137
APPENDICES	162
BIODATA OF STUDENT	204
LIST OF PUBLICATIONS	205



LIST OF TABLES

Table		Page
2.1	Diagnostic criteria for T2DM based on OGTT	10
2.2	Mechanism of action and side effect of oral anti-diabetic drugs	14
2.3	Common medicinal plants used to treat T2DM	15
2.4	Summary of potential medicinal effect of <i>C. caudatus</i>	17
2.5	Biological active compounds in <i>C. caudatus</i>	18
2.6	Antioxidant compounds and their effect on glucose metabolism in humans	22
2.7	Examples of metabolomic applications on dietary intervention	27
3.1	Extraction yield of <i>C. caudatus</i> extracts	33
3.2	Total flavonoid content of <i>C. caudatus</i> extracts and juice	34
3.3	Antioxidant activities of <i>C. caudatus</i> extracts and juice	34
3.4	ABTS assay of <i>C. caudatus</i> leaf extracts and juice	35
3.5	AEAC of <i>C. caudatus</i> extracts and juice	36
3.6	Pearson correlation coefficient (r) between antioxidant compounds and antioxidant activity of <i>C. caudatus</i> leaf extracts and juice	38
4.1	Nutritional facts of <i>C. caudatus</i> (15g)	43
4.2	Standardized nutritional prescriptions for all subjects based on Malaysian Medical Nutrition Therapy for T2DM	44
4.3	Quick method factor used for energy requirement calculation	44
4.4	Timeline for outcome measurements	47
4.5	Classification of hypertension	48
4.6	Category of physical activity level	52
4.7	Demographic characteristic of the subjects by gender, age and ethnicity	55
4.8	Socio-demographic of the subjects	56
4.9	Health status of the subjects	57
4.10	Lifestyle habits of the subjects	58
4.11	Anthropometry measurement of the subjects at baseline	59
4.12	Blood pressure measurement of the subjects at baseline	60
4.13	Glycemic status, inflammatory marker and lipid profile of the subjects at baseline	61
4.14	Renal function test and liver function test of the subjects at baseline	62
4.15	Daily dietary intake of the subjects at baseline	63
4.16	Changes in glycemic status over 8 weeks	66
4.17	Changes in cardiovascular risk factors over 8 weeks	71
4.18	Changes in renal and liver profile over 8 weeks	74
4.19	Changes in dietary intake and physical activity level over 8 weeks	76
4.20	Changes in metabolic parameters in week 12	78
4.21	Changes in renal profile and liver profile at week 12	80
4.22	Changes in dietary intake and physical activity level at week 12	81
5.1	Socio-demographic characteristics of healthy individuals	96

LIST OF FIGURES

Figure		Page
1.1	Conceptual framework of the study	5
2.1	Overview of glucose homeostasis	8
2.2	Management of T2DM over time (years)	11
2.3	Picture of <i>C. caudatus</i>	16
2.4	Polyphenol classification	21
2.5	Overview of metabolomics	23
2.6	Steps involve in an NMR-based metabolomic study	25
3.1	Total phenolic content of <i>C. caudatus</i> extracts and juice	33
3.2	FRAP assay of <i>C. caudatus</i> extracts and juice	36
3.3	Ascorbic acid content of <i>C. caudatus</i> extracts and juice	37
4.1	Flow chart of study protocol	45
4.2	Screening details of the subjects	53
4.3	Subjects enrollment and follow-up based on CONSORT statement	54
4.4	BMI category of the subjects at baseline	59
4.5	Evaluation of energy intake using EI:BMR at baseline	64
4.6	Physical activity levels of the subjects at baseline	65
4.7	Mean changes in HbA1C (%) from baseline	65
4.8	Mean changes in fasting blood glucose from baseline	67
4.9	Mean changes in fructosamine from baseline	68
4.10	Mean changes in serum insulin from baseline	68
4.11	Mean changes in HOMA-IR from baseline	69
4.12	Mean changes in QUICKI from baseline	70
4.13	Mean changes in systolic blood pressure from baseline	72
4.14	Mean changes in hs-CRP from baseline	72
5.1	Study design of metabolomics study	90
5.2	Sample preparations for metabolomic analysis	92
5.3	Procedure of NMR spectra processing and analysis	94
5.4	Anthropometry and blood pressure measurements of the three groups	97
5.5	Glycemic status and inflammatory marker measurements	98
5.6	Lipid profile measurements of the three groups	99
5.7	Renal profile measurements of the three groups	100
5.8	Liver profile measurements of the three groups	101
5.9	PLS-DA of metabolic parameters comparing healthy individuals and diabetic-controls	102
5.10	Satisfactory validation plot for PLS-DA model in Figure 5.9	103
5.11	PLS-DA of metabolic parameters comparing healthy individuals and diabetic-ulam group	104
5.12	Satisfactory validation plot for PLS-DA model in Figure 5.11	105
5.13	OPLS-DA of overall metabolic parameters in the three groups	106
5.14	Validation plot for OPLS-DA model in Figure 5.13	107
5.15	OPLS-DA of ¹ H-NMR urinary data between healthy individuals and diabetic controls	108
5.16	Validation plot for OPLS-DA model in Figure 5.15	109

5.17	OPLS-DA of ¹ H-NMR urinary data between healthy individuals and diabetic-ulam group	110
5.18	Validation plot for OPLS-DA model in Figure 5.17	111
5.19	OPLS-DA of overall ¹ H-NMR urinary data in the three groups	112
5.20	Validation plot for OPLS-DA model in Figure 5.19	113
5.21	Assignment of urinary metabolites in a representative urinary NMR spectrum	114
5.22	Relative concentrations of metabolites in NMR urinary data	115
5.23	OPLS-DA of blood serum ¹ H-NMR data between healthy individuals and diabetic-controls	116
5.24	Validation plot for OPLS-DA model in Figure 5.23	117
5.25	OPLS-DA of blood serum ¹ H-NMR data between healthy individuals and diabetic-ulam group	118
5.26	Validation plot for OPLS-DA model in Figure 5.25	119
5.27	OPLS-DA of overall blood serum ¹ H-NMR data in the three groups	120
5.28	Validation plot for OPLS-DA model in Figure 5.27	121
5.29	Assignment of blood serum metabolites in a representative blood serum NMR spectrum	122
5.30	Relative concentrations of metabolites in blood serum NMR data	123
5.31	Consumption and regeneration of NAD ⁺	127
5.32	Metabolic pathway affected by <i>C. caudatus</i> consumption	129

LIST OF APPENDICES

Appendix		Page
A	Approval letter from Ethics Committee for Research involving Human Subjects Universiti Putra Malaysia (JKEUPM)	162
B	Approval letter from Herbal Medicine Research Centre, Institute for Medical Research Malaysia	164
C	Approval letter from Medical Research and Ethics Committee Ministry of Health Malaysia	165
D	Informed consent form (English)	169
	Informed consent form (Malay)	170
E	Respondent's information sheet (English)	171
	Respondent's information sheet (Malay)	179
F	Questionnaire (English/Malay)	186
G	International Physical Activity Questionnaire (English)	200
	International Physical Activity Questionnaire (Malay)	202

LIST OF ABBREVIATIONS

AA	Antioxidant activity
AACE	American Association of Clinical Endocrinologists
ABTS	2,2'-azinobis-(3-ethylbenzothiazoline-6-sulfonate)
ACD	Advance Chemistry Department
ADA	American Diabetes Association
AEAC	Ascorbic acid equivalent antioxidant capacity
ALP	Alkaline phosphatase
ALT	Alanine aminotransferase
AR	Analytical reagent
AST	Aspartate aminotransferase
ATP	Adenosine triphosphate
ANOVA	Analysis of variance
BMI	Body mass index
BCAA	Branched-chain amino acids
CHO	Carbohydrate
CI	Confidence interval
CONSORT	Consolidated standards of reporting trials
CPG	Clinical Practice Guidelines
CPMG	Carr-Putcell-Meiboom-Gill
DBP	Diastolic blood pressure
DNA	Deoxyribonucleic acid
D ₂ O	Deuterium oxide
DPP-4	Dipeptidyl peptidase-4
DPPH	2,2-Diphenyl-1-picrylhydrazyl
DR	Degradation rate
DW	Dry weight
EC ₅₀	Effective concentration (50%)
EDTA	Ethylenediaminetetraacetic acid
EI:BMR	Energy intake to basal metabolic rate
EtOH	Ethanol
FBG	Fasting blood glucose
FID	Free induction decays
FRAP	Ferric-reducing antioxidant power
FW	Fresh weight
GAE	Gallic acid equivalent
GC-MS	Gas Chromatography-Mass spectrometry
GLP-1	Glucagon-like peptide 1
GLUT4	Glucose transporter type 4
GGT	Gamma-glutamyl transpeptidase
¹ H	Proton
HbA1C	Glycated hemoglobin
HCl	Hydrochloric acid
HDL	High-density lipoprotein
HMDB	The Human Metabolome Database
HOMA-IR	Homeostasis model of assessment for insulin resistance
HPLC	High-performance liquid chromatography
Hs-CRP	High sensitivity C-reactive protein
IL	Interleukin

IPAQ	International physical activity questionnaire
ITT	Intention-to-treat
LC-MS	Liquid Chromatography-Mass spectrometry
LDL	Low-density lipoprotein
MDA	Malondialdehyde
MeOH	Methanol
MET	Metabolic equivalent of task
MS	Mass spectrometry
MTF	Metformin
NHMS	National Health and Morbidity Survey
NMR	Nuclear magnetic resonance
NOESY	Nuclear overhauser effect spectroscopy
OPLS-DA	Orthogonal partial least squares-discriminant analysis
PBG	Postprandial blood glucose
PCA	Principal component analysis
PLS-DA	Partial least squares-discriminant analysis
PPAR- γ	Peroxisome proliferator-activated receptor gamma
ppm	Parts per million
QE	Quercetin equivalent
QUICKI	Quantitative insulin sensitivity check index
RE	Retinol equivalent
ROS	Reactive oxygen species
SBP	Systolic blood pressure
SD	Standard deviation
SIMCA	Soft independent modeling of class analogy
SOD	Superoxide dismutase
SPSS	Statistical package for the social science
SU	Sulphonylureas
T1DM	Type 1 diabetes mellitus
T2DM	Type 2 diabetes mellitus
TAS	Total antioxidant status
TC	Total cholesterol
TCA	Tricarboxylic acid
TG	Triglycerides
TEAC	Trolox equivalent antioxidant capacity
TFC	Total flavonoid content
TPC	Total phenolic content
TPTZ	2,4,6-tris(2-pyridyl)-s-triazine
TSP	Sodium 3-trimethylsilyl-(2,2,3,3-d ₄)-1-propionate
WC	Waist circumference

CHAPTER 1

INTRODUCTION

1.1 Research Background

Type 2 diabetes mellitus (T2DM) is a metabolic condition characterized by hyperglycemia resulting from insulin resistance and impaired insulin secretion (American Diabetes Association, 2015). T2DM is the most common form of diabetes which accounts for about 90-95% of all diabetes cases (American Diabetes Association, 2015). The prevalence of T2DM has been rising rapidly worldwide. In 2014, about 387 millions of people suffered from T2DM worldwide, and this number is projected to rise to 592 million people by 2035 (International Diabetes Federation, 2014).

Currently, management of T2DM involves multi-dimensional approach including the prescription of oral anti-diabetic drugs (such as metformin and sulphonylureas) and lifestyle interventions (Nauck et al., 2009). While the efficacy of sulphonylureas and metformin has been established, their use is associated with side effects such as increased weight gain and elevated risk of hypoglycemia and gastrointestinal disturbance (Inzucchi et al., 2012). In addition, researchers have shown that long-term treatment with oral anti-diabetic drugs is ineffective in protecting the declining function of the pancreatic beta cell (Ball et al., 2000; Van Raalte & Diamant, 2011). The deterioration of pancreatic beta-cell function has also been associated with the elevated oxidative stress in T2DM patients (Figueroa-Romero et al., 2008; Giacco & Brownlee, 2010). Despite the multi-approaches treatments in managing T2DM, poor glycemic control is still prevalent in T2DM patients (Ramachandran et al., 2010).

Medicinal plants have been used as an alternative treatment for treating T2DM (Surya et al., 2014). *Cosmos caudatus*, or known locally as Ulam Raja, is a medicinal herb that popularly consumed in South East Asia. It has been identified as one of the ten commonly used medicinal plants in Malaysia for the treatment of T2DM (Sekar et al., 2014). In addition, *C. caudatus* has been reported to contain a variety of bioactive compounds, including ascorbic acid, quercetin, proanthocyanidins, chlorogenic acid and catechin (Abas et al., 2003; Mustafa et al., 2010; Shui et al., 2005; Sukrasno et al., 2011). Notably, treatment with *C. caudatus* was found to confer beneficial effect in the animal model, but its effect in T2DM patients has not been established.

Previous studies showed that metabolomics applications in dietary interventions enable researchers to study the therapeutic mechanism effects of the dietary interventions (Martin et al., 2009; Moazzami et al., 2012; Van Dorsten et al., 2006). Metabolomics measures metabolites within a biological system at a given time (Zhang et al., 2014), and the profiling of these metabolites can provide detailed information on how the dietary intervention affects the metabolites in the biological system (Friedrich, 2012). In view of this, metabolomics approach can provide a clearer understanding on the effects of *C. caudatus* consumption in T2DM patients.

1.2 Statement of Problem

Today T2DM is a common chronic metabolic disease worldwide. One in twelve people worldwide has T2DM (International Diabetes Federation, 2014). The prevalence of T2DM in Malaysia showed the same worrying trend. The most recent NHMS IV has revealed that one in every five Malaysians age over 30 is having diabetes (Feisul, 2012). Despite the drug treatment, a majority (78%) of T2DM patients in Malaysia still have poor glycemic control with mean HbA1C of 8.7% (Mafauzy et al., 2011).

Medicinal plants played a crucial role in T2DM research (Surya et al., 2014). Indeed, the important role of plants as T2DM treatment was evidenced by the discovery of the metformin from *Galega officinalis* (Bailey et al., 2007). *C. caudatus* (*Ulam raja*) is widely consumed among the local Malays in Malaysia. It has been used since ancient times for its curative properties such as boosting blood circulation, strengthening the bone, and treating infectious disease (Bodeker, 2009). In addition, *C. caudatus* has been reported to have the highest antioxidant capacity as compared to other 25 plants using 1,1-diphenyl-2-picrylhydrazyl (DPPH) radical *scavenging* activity and ferric ion antioxidant potential (FRAP) assays (Wong et al., 2006). This result was in agreement with a recent study which reported that *C. caudatus* have the highest antioxidant activity when compared to four common *ulam*s in Malaysia (including *pegaga*, *selom*, curry leaf and *petai*) (Reihani & Azhar, 2012). This high antioxidant content may suggest its potential in reducing oxidative stress in humans (Shui et al., 2005).

In addition to its beneficial effect on antioxidant, *C. caudatus* has been shown to exhibit anti-diabetic (Perumal et al., 2014), anti-hypertensive (Amalia et al., 2012) and anti-inflammatory effect (Ajaykumar et al., 2012) in animal studies. Recent study in rats demonstrated a significant improvement in fasting blood glucose and lipid profile after 4 weeks of *C. caudatus* supplementation (Perumal et al., 2014). However, its effect in T2DM patients remains unclear. To address this gap, therefore, the objective of this study is to investigate the effect of an eight weeks *C. caudatus* supplementation on glycemic status, cardiovascular risk factors and metabolic profile in T2DM patients.

Furthermore, there is a lack of knowledge on the potential mechanism of action of *C. caudatus* supplementation in T2DM patients. To date, no studies have been reported on the metabolite changes following the *C. caudatus* supplementation. Metabolomics is a powerful tool to study the altered metabolism, identify short-term changes in biological fluids and serve as biomarker detection (Friedrich, 2012). We hypothesized that the therapeutic effect of *C. caudatus* in T2DM patients would reflect as a change of metabolite profile in urine and blood serum. Therefore, we perform metabolomic analysis in an attempt to elucidate the altered metabolite concentration following *C. caudatus* supplementation in T2DM patients.

1.3 Significance of the Study

Herbs have received increasing interest among researchers because of its health benefits. To the best of knowledge, there is no study reported on the effect of *C. caudatus* in T2DM patients. Considering *C. caudatus* is widely consumed among the locals in South East Asian countries, the findings of this study will provide useful insight into effectiveness and safety of *C. caudatus* supplementation in T2DM patients. Results from this study will contribute to the knowledge on the potential use of *C. caudatus* as an adjuvant therapy in the management of T2DM.

Besides, metabolomics approach used in this study will fill in the gap and provide a better understanding of metabolite perturbation following the supplementation of *C. caudatus* in T2DM patients. Likewise, it can undoubtedly enhance the knowledge on the potential anti-diabetic mechanism of *C. caudatus* supplementation.

1.4 General Objective

To investigate the metabolic effect of *C. caudatus* supplementation in T2DM patients.

1.5 Specific Objectives

1. To determine the antioxidant capacity of *C. caudatus* extracted by different solvents.
2. To determine the effect of *C. caudatus* supplementation on glycemic status (fasting glucose, insulin, HbA1C, fructosamine) in T2DM patients
3. To determine the effect of *C. caudatus* supplementation on cardiovascular risk factors (including blood pressure, lipid profile and high sensitivity C-reactive protein) in T2DM patients.
4. To determine the safety of *C. caudatus* supplementation on liver and renal profile in T2DM patients.
5. To determine and compare the metabolomic profiles of urine and blood serum between diabetic-ulam group, diabetic controls and healthy individuals.

1.6 Research conceptual framework

The conceptual framework of this study is presented in Figure 1.1. T2DM patients are usually advised to make lifestyle modifications which include dietary intervention and physical activity recommendations. In addition, medications including insulin therapy, anti-diabetic, anti-hypertensive and lipid-lowering drugs are used to achieve the targeted blood glucose and reduced cardiovascular risk factors in T2DM patients. Hence, the confounding factors in this study namely dietary intake, physical activity and medications were controlled throughout the study.

Oxidative stress plays a significant role in the development of insulin resistant (Styskal et al., 2012). Hyperglycemia and hyperlipidemia increase mitochondrial reactive oxygen species production and lead to oxidative stress (Evans et al., 2002). Oxidative stress affects insulin secretion and action, subsequently leads to beta cell dysfunction and insulin resistance (Bonnard et al., 2008; Lowell & Shulman, 2005).

It was hypothesized that *C. caudatus* used as a dietary antioxidant in this trial may reduce the oxidative stress, subsequently reduce the insulin resistance and improve the outcomes measurements (including glycemic status, cardiovascular risk factors, inflammation) in T2DM patients. In order to measure the altered metabolite in the urine and blood serum following *C. caudatus* supplementation in T2DM patients, a metabolomic approach is used in this study.

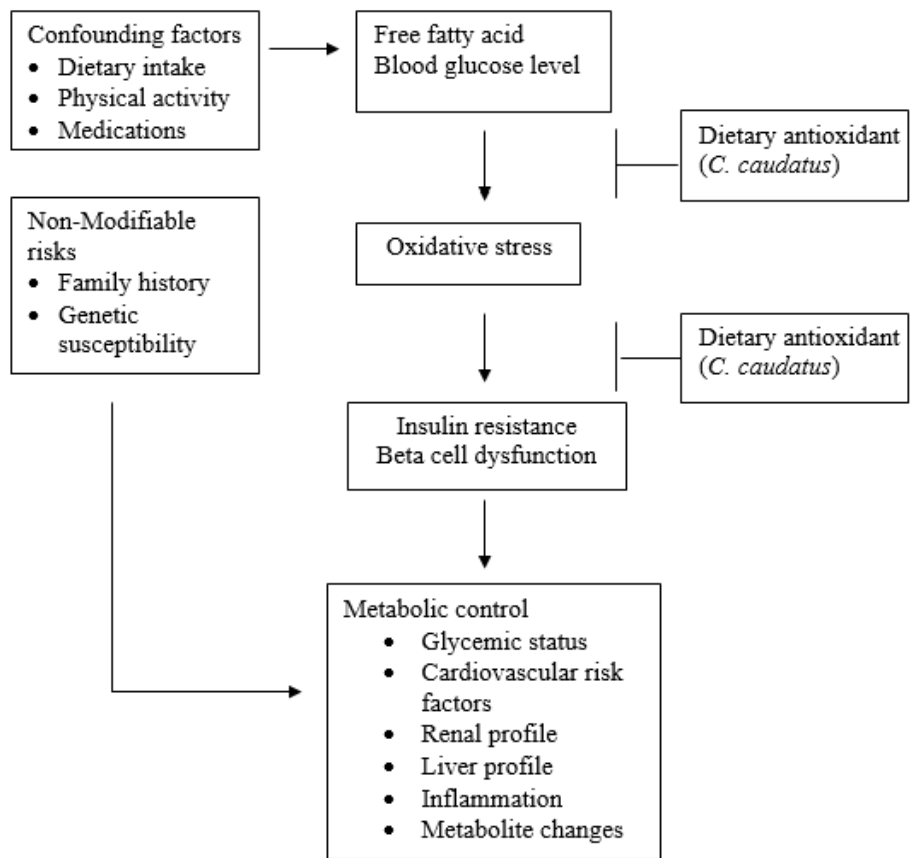


Figure 1.1: Conceptual framework of the study

REFERENCES

- Abas, F., Shaari, K., Lajis, N. H., Israf, D. A., & Kalsom, Y. U. (2003). Antioxidative and radical scavenging properties of the constituents isolated from *Cosmos caudatus* Kunth. *Natural Product Research*, 9(4), 245–248.
- Abougambou, S. S. I., Mohamed, M., Sulaiman, S. A. S., Abougambou, A. S., & Hassali, M. A. (2010). Current clinical status and complications among type 2 diabetic patients in Universiti Sains Malaysia hospital. *International Journal of Diabetes Mellitus*, 2(3), 184–188.
- Abramovitch, S. L., Reddigan, J. I., Hamadeh, M. J., Jamnik, V. K., Rowan, C. P., & Kuk, J. L. (2012). Underestimating a serving size may lead to increased food consumption when using Canada's Food Guide. *Applied Physiology, Nutrition, and Metabolism*, 37(5), 923–30.
- Abu Bakar, M. H., Sarmidi, M. R., Cheng, K.-K., Ali Khan, A., Suan, C. L., Zaman Huri, H., & Yaakob, H. (2015). Metabolomics – the complementary field in systems biology: a review on obesity and type 2 diabetes. *Molecular BioSystems*, 11(7), 1742–1774.
- Adam, Z., Muhajir, H., Ismail, a, Khamis, S., & Marsidi, N. (2010). Antihyperglycemic and glucose tolerance activity of *Ficus deltoidea* ethanolic extract in diabetic rats. *Jurnal Sains Kesihatan Malaysia*, 8(1), 25–30.
- Adeva-Andany, M., López-Ojén, M., Funcasta-Calderón, R., Ameneiros-Rodríguez, E., Donapetry-García, C., Vila-Altesor, M., & Rodríguez-Seijas, J. (2014). Comprehensive review on lactate metabolism in human health. *Mitochondrion*, 17, 76–100.
- Ajaykumar, T. V, Anandarajagopal, K., Sunilson, J. A. J., Arshad, A., & Venkateshan, N. (2012). Anti-inflammatory activity of *Cosmos Caudatus*. *International Journal of Universal Pharmacy and Bio Sciences.*, 1(2), 40–48.
- Algariri, K., Meng, K. Y., Atangwho, I. J., Asmawi, M. Z., Sadikun, A., Murugaiyah, V., & Ismail, N. (2013). Hypoglycemic and anti-hyperglycemic study of *Gynura procumbens* leaf extracts. *Asian Pacific Journal of Tropical Biomedicine*, 3(5), 358–66.
- Ali, S., & Jusoff, K. (2009). Barriers to optimal control of type 2 diabetes in Malaysian Malay patients. *Global Journal of Health Science*, 1(2), 106–118.
- Amalia, L., Anggadiredja, K., Sukrasno, Fidrianny, I., & Inggraini, R. (2012). antihypertensive potency of wild cosmos leaf extract. *Journal of Pharmacology and Toxicology*, 7(8), 359–368.
- American Diabetes Association. (2010). Standards of medical care in diabetes-2010.

Diabetes Care, 33, S11–S61.

American Diabetes Association. (2013). Diagnosis and classification of diabetes mellitus. *Diabetes Care*, 36(Suppl 1), S67–S74.

American Diabetes Association. (2015). Standards of medical care in diabetes - 2015. *Diabetes Care*, 38 (Suppl.1), S1–S94.

Amessis-Ouchemoukh, N., Abu-Reidah, I. M., Quirantes-Piné, R., Madani, K., & Segura-Carretero, A. (2014). Phytochemical profiling, in vitro evaluation of total phenolic contents and antioxidant properties of *Marrubium vulgare* (horehound) leaves of plants growing in Algeria. *Industrial Crops and Products*, 61, 120–129.

Amna, O. F., Nooraain, H., Norihama, a, Azizah, a H., & Husna, R. N. (2013). Acute and oral subacute toxicity study of ethanolic extract of *Cosmos Caudatus* leaf in sprague dawley rats. *International Journal of Bioscience, Biochemistry and Bioinformatics*, 3(4), 301-305.

Anan, F., Masaki, T., Umeno, Y., Iwao, T., Yonemochi, H., Eshima, N., ... Yoshimatsu, H. (2007). Correlations of high-sensitivity C-reactive protein and atherosclerosis in Japanese type 2 diabetic patients. *European Journal of Endocrinology / European Federation of Endocrine Societies*, 157(3), 311–7.

Andarwulan, N., Batari, R., Sandrasari, D. A., Bolling, B., & Wijaya, H. (2010). Flavonoid content and antioxidant activity of vegetables from Indonesia. *Food Chemistry*, 121(4), 1231–1235.

Andrade-Cetto, A., & Wiedenfeld, H. (2001). Hypoglycemic effect of *Cecropia obtusifolia* on streptozotocin diabetic rats. *Journal of Ethnopharmacology*, 78(2-3), 145–149.

Andrews, R., Cooper, A., Montgomery, A., Norcross, A., Peters, T., Sharp, D., ... Dayan, C. (2011). Diet or diet plus physical activity versus usual care in patients with newly diagnosed type 2 diabetes: the early ACTID randomised controlled trial. *Lancet*, 378(9786), 129–139.

Anne-Helen, S., Kaytee, R., & Ailsa, N. (2008). Plasma vitamin C level, fruit and vegetable consumption, and the risk of new-onset type 2 diabetes mellitus. *Archives of Internal Medicine*, 168(14), 1493–1499.

Antonakoudis, G., Poulimenos, I., Kifnidis, K., Zouras, C., & Antonakoudis, H. (2007). Blood pressure control and cardiovascular risk reduction. *Hippokratia*, 11(3), 114–119.

Arnlöv, J., Zethelius, B., Risérus, U., Basu, S., Berne, C., Vessby, B., ... Helmersson, J. (2009). Serum and dietary beta-carotene and alpha-tocopherol and incidence of type 2 diabetes mellitus in a community-based study of Swedish men: report from the Uppsala Longitudinal Study of Adult Men (ULSAM) study. *Diabetologia*,

52(1), 97–105.

- Arts, I. C. W., & Hollman, P. C. H. (2005). Polyphenols and disease risk in epidemiologic studies. *American Journal of Clinical Nutrition*, *81*, 317S-325S.
- Ascaso, J. F., Pardo, S., Real, J. T., Lorente, R. I., Priego, A., & Carmena, R. (2003). Diagnosing insulin resistance by simple quantitative methods in subjects with normal glucose metabolism. *Diabetes Care*, *26*(12), 3320–3325.
- Atkinson, M. a, Eisenbarth, G. S., & Michels, A. W. (2014). Type 1 diabetes. *Lancet*, *383*(9911), 69–82.
- Atoui, A. K., Mansouri, A., Boskou, G., & Kefalas, P. (2005). Tea and herbal infusions: Their antioxidant activity and phenolic profile. *Food Chemistry*, *89*, 27–36.
- Avery, L., Flynn, D., Van Wersch, A., Sniehotta, F. F., & Trenell, M. I. (2012). Changing physical activity behavior in type 2 diabetes: A systematic review and meta-analysis of behavioral interventions. *Diabetes Care*, *35*, 2681–2689.
- Bahrani, M., Ataie-Jafari, A., Hosseini, S., Foruzanfar, M. H., Rahmani, M., & Pajouhi, M. (2009). Effects of natural honey consumption in diabetic patients: an 8-week randomized clinical trial. *International Journal of Food Science and Nutrition*, *60*(7), 618–626.
- Bailey, C. J., Campbell, I. W., Chan, J. C. N., Davidson, J. A., Howlett, H. C. S., & Ritz, P. (2007). *Metformin- the Gold Standard: A Scientific handbook*. Chichester: Wiley.
- Ball, A. J., Flatt, P. R., & McClenaghan, N. H. (2000). Desensitization of sulphonylurea- and nutrient-induced insulin secretion following prolonged treatment with glibenclamide. *European Journal of Pharmacology*, *408*, 327–333.
- Ball, G. F. M. (2006). *Vitamins in foods: analysis, bioavailability and stability*. Boca Raton: Taylor & Francis.
- Barakatun Nisak, M. Y., Ruzita, a. T., Norimah, a. K., & Nor Azmi, K. (2013). Medical nutrition therapy administered by a dietitian yields favourable diabetes outcomes in individual with type 2 diabetes mellitus. *Medical Journal of Malaysia*, *68*(1), 18–23.
- Befroy, D. E., Petersen, K. F., Dufour, S., Mason, G. F., de Graaf, R. A., Rothman, D. L., & Shulman, G. I. (2007). Impaired mitochondrial substrate oxidation in muscle of insulin-resistant offspring of type 2 diabetic patients. *Diabetes*, *56*(5), 1376–1381.
- Benayad, Z., Martinez-Villaluenga, C., Frias, J., Gomez-Cordoves, C., & Es-Safi, N. E. (2014). Phenolic composition, antioxidant and anti-inflammatory activities of extracts from Moroccan *Opuntia ficus-indica* flowers obtained by different

extraction methods. *Industrial Crops and Products*, 62, 412–420.

- Bennett, W. L., Maruthur, N. M., Singh, S., Segal, J. B., Wilson, L. M., Chatterjee, R., ... Bolen, S. (2011). Comparative effectiveness and safety of medications for type 2 diabetes: an update including new drugs and 2-drug combinations. *Annals of Internal Medicine*, 154(9), 602–613.
- Benzie, I. F., & Strain, J. J. (1996). The ferric reducing ability of plasma (FRAP) as a measure of “antioxidant power”: the FRAP assay. *Analytical Biochemistry*, 239(1), 70–76.
- Berhane, F., Fite, A., Daboul, N., Al-Janabi, W., Msallaty, Z., Caruso, M., ... Seyoum, B. (2015). Plasma lactate levels increase during hyperinsulinemic euglycemic clamp and oral glucose tolerance test. *Journal of Diabetes Research*, 102054, 1–7.
- Beydoun, M. A., Shroff, M. R., Chen, X., Beydoun, H. A., Wang, Y., & Zonderman, A. B. (2011). Serum antioxidant status is associated with metabolic syndrome among U. S. adults in recent national surveys 1 – 3. *The Journal of Nutrition*, 141, 903–913.
- Bingham, C., Ellard, S., Nicholls, A. J., Pennock, C. A., Allen, J., James, A. J., ... Hattersley, A. T. (2001). The generalized aminoaciduria seen in patients with hepatocyte nuclear factor-1alpha mutations is a feature of all patients with diabetes and is associated with glucosuria. *Diabetes*, 50(9), 2047–2052.
- Black, A. E. (2000). Critical evaluation of energy intake using the Goldberg cut-off for energy intake: basal metabolic rate. A practical guide to its calculation, use and limitations. *International Journal of Obesity and Related Metabolic Disorders*, 24(9), 1119–1130.
- Boccard, J., & Rutledge, D. N. (2013). A consensus orthogonal partial least squares discriminant analysis (OPLS-DA) strategy for multiblock Omics data fusion. *Analytica Chimica Acta*, 769, 30–39.
- Bodeker, G. (2009). *Health & beauty from the rainforest: Malaysian traditions of ramuan*. Kuala Lumpur: Didier Millet.
- Boden, G. (2003). Effects of free fatty acids (FFA) on glucose metabolism: significance for insulin resistance and type 2 diabetes. *Experimental and Clinical Endocrinology & Diabetes*, 111(3), 121–4.
- Boden, G. (2011). Obesity, insulin resistance and free fatty acids. *Current Opinion in Endocrinology, Diabetes and Obesity*, 18(2), 139–143.
- Boeing, J. S., Barizão, E. O., E Silva, B. C., Montanher, P. F., de Cinque Almeida, V., & Visentainer, J. V. (2014). Evaluation of solvent effect on the extraction of phenolic compounds and antioxidant capacities from the berries: application of principal component analysis. *Chemistry Central Journal*, 8(1), 48.

- Bolen, S., Feldman, L., Vassy, J., Wilson, L., & Yeh, H. (2007). Systematic review: comparative effectiveness and safety of oral medications for type 2 diabetes mellitus. *Annals of Internal Medicine*, *147*, 386–399.
- Bonnard, C., Durand, A., Peyrol, S., Chanseaux, E., Chauvin, M. A., Morio, B., Vidal, H., Rieusset, J. (2008). Mitochondrial dysfunction results from oxidative stress in the skeletal muscle of diet-induced insulin-resistant mice. *The Journal of Clinical Investigation*, *118*(2), 789–800.
- Borai, A., Livingstone, C., Kaddam, I., & Ferns, G. (2011). Selection of the appropriate method for the assessment of insulin resistance. *BMC Medical Research Methodology*, *11*(1), 158.
- Brasnyó, P., Molnár, G. A., Mohás, M., Markó, L., Laczy, B., Cseh, J., ... Wittmann, I. (2011). Resveratrol improves insulin sensitivity, reduces oxidative stress and activates the Akt pathway in type 2 diabetic patients. *The British Journal of Nutrition*, *106*(3), 383–9.
- Brennan, L. (2014). NMR-based metabolomics: From sample preparation to applications in nutrition research. *Progress in Nuclear Magnetic Resonance Spectroscopy*, *83*, 42–49.
- Brown, A. L., Lane, J., Coverly, J., Stocks, J., Jackson, S., Stephen, A., ... Hendrickx, H. (2009). Effects of dietary supplementation with the green tea polyphenol epigallocatechin-3-gallate on insulin resistance and associated metabolic risk factors: randomized controlled trial. *The British Journal of Nutrition*, *101*(6), 886–94.
- Butler, A. E., Janson, J., Bonner-weir, S., Ritzel, R., Rizza, R. A., & Butler, P. C. (2003). Beta-cell deficit and increased beta-cell apoptosis in humans with type 2 diabetes. *Diabetes*, *52*(1), 102–110.
- Cardona, F., Andrés-Lacueva, C., Tulipani, S., Tinahones, F. J., & Queipo-Ortuño, M. I. (2013). Benefits of polyphenols on gut microbiota and implications in human health. *Journal of Nutritional Biochemistry*, *24*(8), 1415–1422.
- Caspersen, C. J., Powell, K. E., & Christenson, G. M. (1985). Physical activity, exercise, and physical fitness: definitions and distinctions for health-related research. *Public Health Reports*, *100*(2), 126–131.
- Chalmers, J., & Cooper, M. E. (2008). UKPDS and the legacy effect. *The New England Journal of Medicine*, *359*(15), 1618–1620.
- Chang, S. A., Kim, H. S., Yoon, K. H., Ko, S. H., Kwon, H. S., Kim, S. R., ... Kang, S. K. (2004). Body mass index is the most important determining factor for the degree of insulin resistance in non-obese type 2 diabetic patients in Korea. *Metabolism*, *53*(2), 142–146.

- Cheng, S. H., Barakatun-nisak, M. Y., Anthony, J., & Ismail, A. (2015). Potential medicinal benefits of *Cosmos caudatus* (Ulam Raja): A scoping review. *Journal of Research in Medical Sciences*, 20, 1000–1006.
- Cheng, S. H., Ismail, A., Anthony, J., Ng, O. C., Hamid, A. A., & Barakatun-nisak, M. Y. (2015). Eight Weeks of *Cosmos caudatus* (Ulam Raja) supplementation improves glycemic status in patients with type 2 diabetes :a randomized controlled trial. *Evidence-Based Complementary and Alternative Medicine*, 2015, 1–7.
- Cheng, S., Rhee, E. P., Larson, M. G., Lewis, G. D., McCabe, E. L., Shen, D., ... Wang, T. J. (2012). Metabolite profiling identifies pathways associated with metabolic risk in humans. *Circulation*, 125(18), 2222–31.
- Chuang, C., Martinez, K., Xie, G., Kennedy, A., Bumrungpert, A., Overman, A., ... Mcintosh, M. K. (2010). Quercetin is equally or more effective than resveratrol in attenuating tumor necrosis factor-alpha-mediated inflammation and insulin resistance in primary human adipocytes. *The American Journal of Clinical Nutrition*, 92, 1511–1521.
- Cohen, J. (1988). *Statistical power analysis for the behavioral sciences* (2nd editio.). New Jersey: Lawrence Erlbaum.
- Cole, H. S., & Camerini-Davalos, R. A. (1970). Diet therapy of diabetes mellitus. *The Medical Clinics of North America*, 54(6), 1577–1587.
- Costacou, T., Ma, B., King, I. B., & Mayer-Davis, E. J. (2008). Plasma and dietary vitamin E in relation to insulin secretion and sensitivity. *Diabetes, Obesity & Metabolism*, 10(3), 223–8.
- Craig, C. L., A. L. Marshall, M. Sjostrom, A. E. Bauman, M. L. Booth, B. E. Ainsworth, M. Pratt, U. Ekelund, A. Yngve, J. F. Sallis, and P. O. (2003). International physical activity questionnaire: 12 country reliability and validity. *Medicine & Science in Sports & Exercise*, 35(8), 3508–1381.
- Crawford, P. (2009). Effectiveness of cinnamon for lowering hemoglobin A1C in patients with type 2 diabetes: a randomized, controlled trial. *Journal of the American Board of Family Medicine*, 22(5), 507–12.
- Crawford, S. O., Hoogeveen, R. C., Brancati, F. L., Astor, B. C., Ballantyne, C. M., Schmidt, M. I., & Young, J. H. (2010). Association of blood lactate with type 2 diabetes: The atherosclerosis risk in communities carotid MRI study. *International Journal of Epidemiology*, 39(6), 1647–1655.
- Cusi, K., Kashyap, S., Gastaldelli, A., Bajaj, M., & Cersosimo, E. (2007). Effects on insulin secretion and insulin action of a 48-h reduction of plasma free fatty acids with acipimox in nondiabetic subjects genetically predisposed to type 2 diabetes. *American Journal of Physiology. Endocrinology and Metabolism*, 292(6), E1775–81.

- Daisy, P., Balasubramanian, K., Rajalakshmi, M., Eliza, J., & Selvaraj, J. (2010). Insulin mimetic impact of Catechin isolated from *Cassia fistula* on the glucose oxidation and molecular mechanisms of glucose uptake on Streptozotocin-induced diabetic Wistar rats. *Phytomedicine*, *17*(1), 28–36.
- Dandu, A. M., & M, I. N. (2009). Evaluation of beneficial effects of antioxidant properties of aqueous leaf extract of *Andrographis Paniculata* in STZ-induced diabetes. *Pakistan Journal of Pharmaceutical Sciences*, *22*(1), 49–52.
- de Oliveira, A. M., Rondó, P. H. C., Luzia, L. A., D'Abronzio, F. H., & Illison, V. K. (2011). The effects of lipoic acid and α -tocopherol supplementation on the lipid profile and insulin sensitivity of patients with type 2 diabetes mellitus: a randomized, double-blind, placebo-controlled trial. *Diabetes Research and Clinical Practice*, *92*(2), 253–60.
- DeFronzo, R. A. (2004). Pathogenesis of type 2 diabetes mellitus. *The Medical Clinics of North America*, *88*(4), 787–835.
- DeFronzo, R. A., & Tripathy, D. (2009). Skeletal muscle insulin resistance is the primary defect in type 2 diabetes. *Diabetes Care*, *32*(Suppl 2), S157–63.
- Deurenberg, P., Deurenberg-Yap, M., & Guricci, S. (2002). Asians are different from Caucasians and from each other in their body mass index/body fat per cent relationship. *Obesity Reviews*, *3*(3), 141–146.
- Dicker, D. (2011). DPP-4 Inhibitors: Impact on glycemic control and cardiovascular risk factors. *Diabetes Care*, *34*(Supplement_2), S276–S278.
- Ding, L., Jin, D., & Chen, X. (2010). Luteolin enhances insulin sensitivity via activation of PPAR γ transcriptional activity in adipocytes. *The Journal of Nutritional Biochemistry*, *21*(10), 941–7.
- Duarte, I. F., Diaz, S. O., & Gil, A. M. (2014). NMR metabolomics of human blood and urine in disease research. *Journal of Pharmaceutical and Biomedical Analysis*, *93*, 17–26.
- Dunn, W. B., Broadhurst, D. I., Atherton, H. J., Goodacre, R., & Griffin, J. L. (2011). Systems level studies of mammalian metabolomes: the roles of mass spectrometry and nuclear magnetic resonance spectroscopy. *Chemical Society Reviews*, *40*(1), 387–426.
- Edmands, W. M., Beckonert, O. P., Stella, C., Campbell, A., Lake, B. G., Lindon, J. C., ... Gooderham, N. J. (2011). Identification of human urinary biomarkers of cruciferous vegetable consumption by metabolomic profiling. *Journal of Proteome Research*, *10*(10), 4513–4521.
- Efird, J. (2011). Blocked randomization with randomly selected block sizes. *International Journal of Environmental Research and Public Health*, *8*(1), 15–20.

- Eid, H. M., Vallerand, D., Muhammad, A., Durst, T., Haddad, P. S., & Martineau, L. C. (2010). Structural constraints and the importance of lipophilicity for the mitochondrial uncoupling activity of naturally occurring caffeic acid esters with potential for the treatment of insulin resistance. *Biochemical Pharmacology*, *79*(3), 444–54.
- El-Shatanovi, G. a. T. a., Ashoush, I. S., Ahmed, E. K., & Ali, S. a. (2012). Antiatherogenic properties of vegetable juice rich in antioxidants in cholesterol-fed rats. *Annals of Agricultural Sciences*, *57*(2), 167–173.
- Esteghamati, A., Ashraf, H., Khalilzadeh, O., Zandieh, A., Nakhjavani, M., Rashidi, A., ... Asgari, F. (2010). Optimal cut-off of homeostasis model assessment of insulin resistance (HOMA-IR) for the diagnosis of metabolic syndrome: third national surveillance of risk factors of non-communicable diseases in Iran (SuRFNCD-2007). *Nutrition and Metabolism*, *7*(26), 1–8.
- Evans, J. D. (1996). *Straightforward Statistics for the Behavioral Sciences*. Pacific Grove: Brooks/Cole Publishing Company.
- Evans, J. L., Goldfine, I. R. A. D., Maddux, B. A., Grodsky, G. M., & Francisco, S. (2002). Oxidative stress and stress-activated signaling pathways: a unifying hypothesis of type 2 diabetes. *Endocrine Reviews*, *23*(5), 599–622.
- Evert, A. B., Boucher, J. L., Cypress, M., Dunbar, S. A., Franz, M. J., Mayer-Davis, E. J., ... Yancy, W. S. (2013). Nutrition Therapy Recommendations for the Management of Adults With Diabetes. *Diabetes Care*, *36*(11), 3821–3842.
- Feisul, I. M. (2012). Current burden of diabetes in Malaysia. In *Proceedings of the 1st National institutes of Health (NIH) Scientific Meeting*. Selangor.
- Figuroa-Romero, C., Sadidi, M., & Feldman, E. L. (2008). Mechanisms of disease: The oxidative stress theory of diabetic neuropathy. *Reviews in Endocrine and Metabolic Disorders*, *9*, 301–314.
- Fine, A. M., & Candidate, N. D. (2000). Oligomeric Proanthocyanidin Complexes: Applications Oligomeric Proanthocyanidins. *Alternative Medicine Review*, *5*(2), 144–151.
- Firouzi, S., Barakatun-Nisak, M. Y., & Azmi, K. N. (2015). Nutritional status, glycemic control and its associated risk factors among a sample of type 2 diabetic individuals, a pilot study. *Journal of Research in Medical Sciences*, *20*(1), 40–46.
- Frankenfield, D., Roth-Yousey, L., & Compher, C. (2005). Comparison of predictive equations for resting metabolic rate in healthy nonobese and obese adults: a systematic review. *Journal of the American Dietetic Association*, *105*(5), 775–789.
- Franz, M. J., Powers, M. A., Leontos, C., Holzmeister, L. A., Kulkarni, K., Monk, A., ... Gradwell, E. (2010). The evidence for medical nutrition therapy for type 1 and

- type 2 diabetes in adults. *Journal of the American Dietetic Association*, 110(12), 1852–1889.
- Friedrich, N. (2012). Metabolomics in diabetes research. *The Journal of Endocrinology*, 215(1), 29–42.
- Fuzzati, N. (1995). Phenylpropane derivatives from roots of *Cosmos caudatus*. *Phytochemistry*, 39(2), 409–412.
- Gan, M. J., Albanese-O'Neill, A., & Haller, M. J. (2012). Type 1 diabetes: current concepts in epidemiology, pathophysiology, clinical care, and research. *Current Problems in Pediatric and Adolescent Health Care*, 42(10), 269–91.
- Garber, A. J. (2011). Long-Acting Glucagon-Like Peptide 1 Receptor Agonists: A review of their efficacy and tolerability. *Diabetes Care*, 34(Supplement_2), S279–S284.
- Garcia-Perez, L.-E., Álvarez, M., Dilla, T., Gil-Guillén, V., & Orozco-Beltrán, D. (2013). Adherence to Therapies in Patients with Type 2 Diabetes. *Diabetes Therapy*, 4(2), 175–194.
- Gaster, M., Nehlin, J. O., & Minet, A. D. (2012). Impaired TCA cycle flux in mitochondria in skeletal muscle from type 2 diabetic subjects: marker or maker of the diabetic phenotype? *Archives of Physiology and Biochemistry*, 118(3), 156–189.
- Gauthier-Chelle, K., Mennen, L., Arnault, N., Rigalleau, V., Hercberg, S., & Gin, H. (2004). Comparison of the diet of self-declared diabetics with non-diabetic patients in the SU.VI.MAX study: Did the diabetics modify their nutritional behavior? *Diabetes and Metabolism*, 30(6), 535–542.
- Ghazali, S., Seman, Z., Cheong, K., Hock, L., Manickam, M., Kuay, L., ... Mustafa, A. (2015). Sociodemographic factors associated with multiple cardiovascular risk factors among Malaysian adults. *BMC Public Health*, 15(1), 68.
- Giacco, F., & Brownlee, M. (2010). Oxidative Stress and Diabetic Complications. *Circulation Research*, 107(9), 1058–1070.
- Goldberg, G. R., Black, A. E., Jebb, S. A., Cole, T. J., Murgatroyd, P. R., Coward, W. A., & Prentice, A. M. (1991). Critical evaluation of energy intake data using fundamental principles of energy physiology: 1. Derivation of cut-off limits to identify under-recording. *European Journal of Clinical Nutrition*, 45(12), 569–581.
- Golozar, A., Khademi, H., Kamangar, F., Poutschi, H., Islami, F., Abnet, C. C., ... Etemadi, A. (2011). Diabetes mellitus and its correlates in an Iranian adult population. *PLoS One*, 6(10), e26725.

- Griffin, J. L., Atherton, H., Shockcor, J., & Atzori, L. (2011). Metabolomics as a tool for cardiac research. *Nature Reviews Cardiology*, 8(11), 630–643.
- Gupta, S. K. (2011). Intention-to-treat concept: A review. *Perspectives in Clinical Research*, 2(3), 109–12.
- Guyton, A. G., & Hall, J. E. (2006). Urine formation by the kidneys. In E. Saunders (Ed.), *Textbook of Medical Physiology* (11th editi., pp. 307–346). Philadelphia, USA.
- Ha, C. Y., Kim, J. Y., Paik, J. K., Kim, O. Y., Paik, Y.-H., Lee, E. J., & Lee, J. H. (2012). The association of specific metabolites of lipid metabolism with markers of oxidative stress, inflammation and arterial stiffness in men with newly diagnosed type 2 diabetes. *Clinical Endocrinology*, 76(5), 674–682.
- Halliwell, B. (2007). Dietary polyphenols: good, bad, or indifferent for your health? *Cardiovascular Research*, 73(2), 341–7.
- Hamnvik, O. R., & McMahon, G. T. (2009). Balancing risk and benefit with oral hypoglycemic drugs. *The Mount Sinai Journal of Medicine*, 76(3), 234–243.
- Handelsman, Y., Mechanick, J. I., Dagogo-jack, S., & Davidson, J. a. (2011). American Association of Clinical Endocrinologists medical guidelines for clinical practice for developing a diabetes mellitus comprehensive care plan. *Endocrine Practice*, 17, 1–53.
- Harris, E. H. (2005). Elevated Liver Function Tests in Type 2 Diabetes. *Clinical Diabetes*, 23(3), 115–119.
- Hassan, W. E. (2006). *Healing Herbs of Malaysia*. Kuala Lumpur: Federal land Development Authority (FELDA).
- He, Y., Jiang, B., Wang, J., Feng, K., Chang, Q., Zhu, S., ... Hu, F. B. (2007). BMI versus the metabolic syndrome in relation to cardiovascular risk in elderly Chinese individuals. *Diabetes Care*, 30(8), 2128–2134.
- Health Promotion Board. (2003). *Food Composition Guide Singapore*. Singapore: Health Promotion Board.
- Heinzmann, S. S., Brown, I. J., Chan, Q., Bictash, M., Dumas, M. E., Kochhar, S., ... Nicholson, J. K. (2010). Metabolic profiling strategy for discovery of nutritional biomarkers: Proline betaine as a marker of citrus consumption. *American Journal of Clinical Nutrition*, 92(2), 436–443.
- Hirst, J. A., Stevens, R. J., & Farmer, A. J. (2014). Changes in HbA1c level over a 12-week follow-up in patients with type 2 diabetes following a medication change. *PLoS ONE*, 9(3), 1–7.

- Hu, F. B. (2011). Globalization of diabetes: The role of diet, lifestyle, and genes. *Diabetes Care*, *34*(6), 1249–1257.
- Huang, M., & Joseph, J. W. (2012). Metabolomic analysis of pancreatic beta-cell insulin release in response to glucose. *Islets*, *4*(3), 210–222.
- Huang, M.-C., Hsu, C.-C., Huang-Sen, W., & Shyi-Jang, S. (2010). Prospective randomized controlled trial to evaluate effectiveness of registered dietitian-led diabetes management on glycemic and diet control in a primary care setting in Taiwan. *Diabetes Care*, *33*(2), 233–239.
- Huffman, K. M., Shah, S. H., Stevens, R. D., Bain, J. R., Muehlbauer, M., Slentz, C. A., ... Kraus, W. E. (2009). Metabolic intermediates and insulin action in overweight to obese, inactive Men and woman. *Diabetes Care*, *32*(9), 1678–1683.
- Hulley, S. B., Cummings, S. R., Browner, W. S., Grady, D. G., & Newman, T. B. (2013). *Designing clinical research* (4th Editio.). Philadelphia, USA: Lippincott Williams and Wilkins.
- Hurtado, N. H., Morales, A. L., González, M. L., Escudero-Gilete, M. L., & Heredia, F. J. (2009). Colour, pH stability and antioxidant activity of anthocyanin rutinosides isolated from tamarillo fruit (*Solanum betaceum* Cav.). *Food Chemistry*, *117*(1), 88–93.
- Institute for Public Health. (2015). National Health & Morbidity Survey (NHMS) 2015. *Fact Sheet*, 5–6.
- International Diabetes Federation. (2014). *IDF Diabetes Atlas* (6th editio.). Brussels, Belgium: International Diabetes Federation.
- International Society for the Advancement of Kinanthropometry. (2001). *International standards for anthropometric assessment*. Potchefstroom, South Africa: International Society for the Advancement of Kinanthropometry.
- Inzucchi, S. E., Bergenstal, R. M., Buse, J. B., Diamant, M., Ferrannini, E., Nauck, M., ... Matthews, D. R. (2012). Management of hyperglycemia in type 2 diabetes: a patient-centered approach: position statement of the American Diabetes Association (ADA) and the European Association for the Study of Diabetes (EASD). *Diabetes Care*, *35*(6), 1364–79.
- Ismail, M., Chee, S., Roslee, R., & Zawiah, H. (1998). Predictive equations for the estimation of basal metabolic rate in Malaysian adults. *Malaysian Journal of Nutrition*, *4*(1), 73–80.
- Jayagopal, V., Kilpatrick, E. S., Jennings, P. E., Hepburn, D. A., & Atkin, S. L. (2002). Biological variation of homeostasis model assessment-derived insulin resistance in type 2 diabetes. *Diabetes Care*, *25*(11), 2022–2025.

- Jing, Y., Han, G., Hu, Y., Bi, Y., Li, L., & Zhu, D. (2009). Tea consumption and risk of type 2 diabetes: a meta-analysis of cohort studies. *Journal of General Internal Medicine*, *24*(5), 557–62.
- Jurgens, C. A., Toukatly, M. N., Fligner, C. L., Udayasankar, J., Subramanian, S. L., Zraika, S., ... Hull, R. L. (2011). B-cell loss and b-cell apoptosis in human type 2 diabetes are related to islet amyloid deposition. *The American Journal of Pathology*, *178*(6), 2632–40.
- Kahn, S. E., Cooper, M. E., & Del Prato, S. (2014). Pathophysiology and treatment of type 2 diabetes: perspectives on the past, present, and future. *Lancet*, *383*(9922), 1068–83.
- Kannappan, S., & Anuradha, C. V. (2009). Insulin sensitizing actions of fenugreek seed polyphenols, quercetin & metformin in a rat model. *Indian Journal of Medical Research*, *129*(4), 401–408.
- Katz, A., Nambi, S. S., Mather, K., Baron, A., Follmann, D., Sullivan, G., & Quon, M. J. (2000). Quantitative insulin sensitivity check index: A simple, accurate method for assessing insulin sensitivity in humans. *J Clin Endocrinol Metab*, *85*(7), 2402–2410.
- Kim, J.-A., Wei, Y., & Sowers, J. R. (2008). Role of mitochondrial dysfunction in insulin resistance. *Circulation Research*, *102*(4), 401–14.
- Kosalec, I., Bakmaz, M., Pepeljnjak, S., & Vladimir-Knezevic, S. (2004). Quantitative analysis of the flavonoids in raw propolis from northern Croatia. *Acta Pharmaceutica*, *54*(1), 65–72.
- Krebs, M., Brehm, a., Krssak, M., Anderwald, C., Bernroider, E., Nowotny, P., ... Roden, M. (2003). Direct and indirect effects of amino acids on hepatic glucose metabolism in humans. *Diabetologia*, *46*(7), 917–925.
- Krebs, M., Krssak, M., Bernroider, E., Anderwald, C., Brehm, A., Meyerspeer, M., ... Roden, M. (2002). Mechanism of amino acid-induced skeletal muscle insulin resistance in humans. *Diabetes*, *51*(3), 599–605.
- Lagouge, M., Argmann, C., Gerhart-Hines, Z., Meziane, H., Lerin, C., Daussin, F., ... Auwerx, J. (2006). Resveratrol improves mitochondrial function and protects against metabolic disease by activating SIRT1 and PGC-1 α . *Cell*, *127*(6), 1109–1122.
- Le, C., Lin, L., Jun, D., Jianhui, H., Keying, Z., Wenlong, C., ... Tao, W. (2013). The economic burden of type 2 diabetes mellitus in rural southwest China. *International Journal of Cardiology*, *165*(2), 273–277.
- Lee, S., Choi, S., Kim, H. J., Chung, Y. S., Lee, K. W., Lee, H. C., ... Kim, D. J. (2006). Cutoff values of surrogate measures of insulin resistance for metabolic syndrome

- in Korean non-diabetic adults. *Journal of Korean Medical Science*, 21(4), 695–700.
- Lee, T., & Vairappan, C. (2011). Antioxidant, antibacterial and cytotoxic activities of essential oils and ethanol extracts of selected South East Asian herbs. *Journal of Medicinal Plants Research*, 5(21), 5284–5290.
- Leong, L., & Shui, G. (2002). An investigation of antioxidant capacity of fruits in Singapore markets. *Food Chemistry*, 76(1), 69–75.
- Li, X., Xu, Z., Lu, X., Yang, X., Yin, P., Kong, H., ... Xu, G. (2009). Comprehensive two-dimensional gas chromatography/time-of-flight mass spectrometry for metabonomics: Biomarker discovery for diabetes mellitus. *Analytica Chimica Acta*, 633(2), 257–62.
- Libby, P., Ridker, P. M., & Hansson, G. K. (2009). Inflammation in Atherosclerosis: From Pathophysiology to Practice. *Journal of the American College of Cardiology*, 54(23), 2129–2138.
- Little, R. R., Rohlfing, C. L., & Sacks, D. B. (2011). Status of hemoglobin A1c measurement and goals for improvement: From Chaos to order for improving diabetes care. *Clinical Chemistry*, 57(2), 205–214.
- Lim, Y. Y., & Murtijaya, J. (2007). Antioxidant properties of *Phyllanthus amarus* extracts as affected by different drying methods. *LWT-Food Science and Technology*, 40(9), 1664–1669.
- Liu, C.-C., Wu, Y.-F., Feng, G.-M., Gao, X.-X., Zhou, Y.-Z., Hou, W.-J., ... Tian, J.-S. (2015). Plasma-metabolite-biomarkers for the therapeutic response in depressed patients by the traditional Chinese medicine formula Xiaoyaosan: A 1H NMR-based metabolomics approach. *Journal of Affective Disorders*, 185, 156–163.
- Loh, S. P., & Hadira, O. (2011). In vitro inhibitory potential of selected Malaysian plants against key enzymes involved in hyperglycemia and hypertension. *Malaysian Journal of Nutrition*, 17(1), 77–86.
- Loh, S. Y., Lee, S. Y., Quek, K. F., & Murray, L. (2012). Barriers to participation in a randomized controlled trial of Qigong exercises amongst cancer survivors: lessons learnt. *Asian Pacific Journal of Cancer Prevention*, 13(12), 6337–6342.
- Lorber, D. (2014). Importance of cardiovascular disease risk management in patients with type 2 diabetes mellitus. *Diabetes, Metabolic Syndrome and Obesity: Targets and Therapy*, 7, 169–183.
- Lowell, B., & Shulman, G. (2005). Mitochondrial dysfunction and type 2 diabetes. *Science*, 307(5708), 384–388.
- Ma, Z. A., Zhao, Z., & Turk, J. (2012). Mitochondrial dysfunction and β -cell failure in

- type 2 diabetes mellitus. *Experimental Diabetes Research*, 2012, 11–22.
- Mafauzy, M., Hussein, Z., & Chan, S. (2011). The status of diabetes control in Malaysia: results of DiabCare 2008. *Med J Malaysia*, 66(3), 175–181.
- Mahajan, A., Tabassum, R., Chavali, S., Dwivedi, O. P., Bharadwaj, M., Tandon, N., & Bharadwaj, D. (2009). High-sensitivity C-reactive protein levels and type 2 diabetes in urban North Indians. *The Journal of Clinical Endocrinology and Metabolism*, 94(6), 2123–7.
- Malaysian Dietitians' Association. (2005). *Medical nutrition therapy guidelines for type 2 diabetes*. Kuala Lumpur: Ministry of Health Malaysia.
- Manach, C., Hubert, J., Llorach, R., & Scalbert, A. (2009). The complex links between dietary phytochemicals and human health deciphered by metabolomics. *Molecular Nutrition and Food Research*, 53(10), 1303–1315.
- Martin, F. P. J., Rezzi, S., Peré-Trepat, E., Kamlage, B., Collino, S., Leibold, E., ... Kochhar, S. (2009). Metabolic effects of dark chocolate consumption on energy, gut microbiota, and stress-related metabolism in free-living subjects. *Journal of Proteome Research*, 8(12), 5568–5579.
- Mastura, I., Chew, B. H., Lee, P. Y., Cheong, A. T., Sazlina, G., Jamaiah, H., ... Zaiton, A. (2011). Control and treatment profiles of 70, 889 adult type 2 diabetes mellitus patients in Malaysia - a cross sectional survey in 2009. *International Journal of Collaborative Research on Internal Medicine & Public Health*, 3(1), 98–113.
- Matthews, D. R., Hosker, J. P., Rudenski, A. S., Naylor, B. A., Treacher, D. F., & Turner, R. C. (1985). Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia*, 28(7), 412–419.
- Mediani, A., Abas, F., Khatib, A., Maulidiani, H., Shaari, K., Choi, Y. H., & Lajis, N. H. (2012). 1H-NMR-based metabolomics approach to understanding the drying effects on the phytochemicals in *Cosmos caudatus*. *Food Research International*, 49(2), 763–770.
- Messana, I., Forni, F., & Ferrari, F. (1998). Proton nuclear magnetic resonance spectral profiles of urine in type II diabetic patients. *Clinical Chemistry*, 44(7), 1529–1534.
- Milburn, M. V., & Lawton, K. A. (2013). Application of metabolomics to diagnosis of insulin resistance. *Annual Review of Medicine*, 64, 291–305.
- Ministry of Health Malaysia. (2015). *Clinical Practice Guidelines (CPG) on Management of Type 2 diabetes Mellitus* (5th editio.). Putrajaya.
- Mirnalini, K., Zalilah, M. S., Safiah, M. Y., Tahir, a., Siti, H. M. D., Siti, R. D., ... Normah, H. (2008). Energy and nutrient intakes: Findings from the Malaysian

Adult Nutrition Survey (MANS). *Malaysian Journal of Nutrition*, 14(1), 1–24.

- Moazzami, A. a, Bondia-Pons, I., Hanhineva, K., Juntunen, K., Antl, N., Poutanen, K., & Mykkänen, H. (2012). Metabolomics reveals the metabolic shifts following an intervention with rye bread in postmenopausal women- a randomized control trial. *Nutrition Journal*, 11(1), 88.
- Mohamed, E. A. H., Yam, M. F., Siddiqol, M. J. A., Asmawi, M. Z., Sadikun, A., Ang, L. F., ... Chan, S. H. (2012). Potent alpha-glucosidase and alpha-amylase inhibitory activities of standardized 50% ethanolic extracts and sinensetin from *Orthosiphon stamineus* Benth as anti-diabetic mechanism. *BMC Complementary and Alternative Medicine*, 12(1), 176.
- Mohamed, N., Gwee Sian Khee, S., Shuid, A. N., Muhammad, N., Suhaimi, F., Othman, F., ... Soelaiman, I. N. (2012). The Effects of *Cosmos caudatus* on structural bone histomorphometry in ovariectomized rats. *Evidence-Based Complementary and Alternative Medicine*, 2012.
- Mohamed, N., Sahhugi, Z., Ramli, E. S., & Muhammad, N. (2013). The effects of *Cosmos caudatus* (ulam raja) on dynamic and cellular bone histomorphometry in ovariectomized rats. *BMC Research Notes*, 6(1), 239.
- Mozaffari-Khosravi, H., Talaei, B., Jalali, B.-A., Najarzadeh, A., & Mozayan, M. R. (2014). The effect of ginger powder supplementation on insulin resistance and glycemic indices in patients with type 2 diabetes: A randomized, double-blind, placebo-controlled trial. *Complementary Therapies in Medicine*, 22(1), 9–16.
- Mukai, K., Nagai, S., & Ohara, K. (2005). Kinetic study of the quenching reaction of singlet oxygen by tea catechins in ethanol solution. *Free Radical Biology and Medicine*, 39(6), 752–761.
- Mulder, T. P., Rietveld, A. G., van Amelsvoort, J. M., & Amelsvoort, J. M. Van. (2005). Consumption of both black tea and green tea results in an increase in the excretion of hippuric acid into urine. *The American Journal of Clinical Nutrition*, 81(1 Suppl), 256–260.
- Muniyappa, R., Lee, S., Chen, H., & Quon, M. J. (2008). Current approaches for assessing insulin sensitivity and resistance in vivo: advantages, limitations, and appropriate usage. *American Journal of Physiology - Endocrinology And Metabolism*, 294(1), E15–E26.
- Mustafa, R. A., Abdul Hamid, A., Mohamed, S., & Bakar, F. A. (2010). Total phenolic compounds, flavonoids, and radical scavenging activity of 21 selected tropical plants. *Journal of Food Science*, 75(1), C28–35.
- Nauck, M., Frid, A., Hermansen, K., Shah, N. S., Tankova, T., Mitha, I. H., ... LEAD-2 Study Group. (2009). Efficacy and safety comparison of liraglutide, glimepiride, and placebo, all in combination with metformin, in type 2 diabetes: the LEAD

- (liraglutide effect and action in diabetes)- 2 study. *Diabetes Care*, 32, 84–90.
- Nazaimoon, W. M. W., Isa, S. H., Mohamad, W. B. W., Khir, A. S., Kamaruddin, N. A., Kamarul, I. M., ... Khalid, B. A. K. (2013). Prevalence of diabetes in Malaysia and usefulness of HbA1c as a diagnostic criterion. *Diabetic Medicine*, 30(7), 825–828.
- Nesto, R. W., Bell, D., Bonow, R. O., Fonseca, V., Grundy, S. M., Horton, E. S., ... Kahn, R. (2003). Thiazolidinedione use, fluid retention, and congestive heart failure: a consensus statement from the American Heart Association and American Diabetes Association. *Circulation*, 108(23), 2941–2948.
- Newgard, C. B., An, J., Bain, J. R., Muehlbauer, M. J., Stevens, R. D., Lien, L. F., ... Laura, P. (2009). A branched-chain amino acid-related metabolic signature that differentiates obese and lean humans and contributes to insulin resistance. *Cell Metabolism*, 9(4), 311–326.
- Ng, M., Fleming, T., Robinson, M., Thomson, B., Graetz, N., Margono, C., ... Gakidou, E. (2014). Global, regional, and national prevalence of overweight and obesity in children and adults during 1980–2013: a systematic analysis for the Global Burden of Disease Study 2013. *Lancet*, 6736(14), 1–16.
- Nolan, C. J., Damm, P., & Prentki, M. (2011). Type 2 diabetes across generations: from pathophysiology to prevention and management. *Lancet*, 378(9786), 169–81.
- Norazlina, M., Ehsan, S. Z., Noor 'Adilah, K., Lee, C. P., Farhana, E., Derick, P., ... Norliza, M. (2013). Acute toxicity study of *Cosmos caudatus* on biochemical parameters in male rats. *Sains Malaysiana*, 42(9), 1247–1251.
- Ojewole, J. A. (2003). Laboratory evaluation of the hypoglycemic effect of *Anacardium occidentale* Linn (*Anacardiaceae*) stem-bark extracts in rats. *Methods and Findings in Experimental and Clinical Pharmacology*, 25(3), 199–204.
- Okoli, C. O., Ibiam, A. F., Ezike, A. C., Akah, P. A., & Okoye, T. C. (2010). Evaluation of antidiabetic potentials of *Phyllanthus niruri* in alloxan diabetic rats. *African Journal of Biotechnology*, 9(2), 248–259.
- Olendzki, B. C., Ma, Y., Hebert, J. R., Pagoto, S. L., Merriam, P. A., Rosal, M. C., & Ockene, I. S. (2013). Underreporting of energy intake and associated factors in a Latino population at risk of developing type 2 diabetes. *Journal of the American Dietetic Association*, 18(9), 1199–1216.
- Ong, K. W., Hsu, A., & Tan, B. K. H. (2013). Anti-diabetic and anti-lipidemic effects of chlorogenic acid are mediated by ampk activation. *Biochemical Pharmacology*, 85(9), 1341–51.
- Pandey, K. B., & Rizvi, S. I. (2009). Plant polyphenols as dietary antioxidants in human health and disease. *Oxidative Medicine and Cellular Longevity*, 2(5), 270–8.

- Pearson, T. A., Mensah, G. A., Alexander, R. W., Anderson, J. L., Cannon, R. O., Criqui, M., ... Vinicor, F. (2003). Markers of inflammation and cardiovascular disease: application to clinical and public health practice: a statement for healthcare professionals from the Centers for Disease Control and Prevention and the American Heart Association. *Circulation*, *107*(3), 499–511.
- Perl, S., Kushner, J. A., Buchholz, B. A., Meeker, A. K., Stein, G. M., Hsieh, M., ... Tisdale, J. F. (2010). Significant human beta-cell turnover is limited to the first three decades of life as determined by in vivo thymidine analog incorporation and radiocarbon dating. *The Journal of Clinical Endocrinology and Metabolism*, *95*(10), E234–9.
- Perumal, V., Hamid, A. A., Ismail, A., Saari, K., Abas, F., Ismail, I. S., ... Khatib, A. (2014). Effect of *Cosmos caudatus* kunth leaves on the lipid profile of a hyperlipidemia-induced animal model. *Journal of Food Chemistry and Nutrition*, *2*(1), 43–51.
- Peungvicha, P., S. Thirawarapan, S., Temsiririrkkul, R., Watanabe, H., Kumar Prasain, J., & Kadota, S. (1998). Hypoglycemic effect of the water extract of *Piper sarmentosum* in rats. *Journal of Ethnopharmacology*, *60*(1), 27–32.
- Pickering, T. G., Hall, J. E., Appel, L. J., Falkner, B. E., Graves, J., Hill, M. N., ... Roccella, E. J. (2005). Recommendations for blood pressure measurement in humans and experimental animals. Part 1: Blood pressure measurement in humans. *Hypertension*, *45*(1), 142–161.
- Piette, J. D., & Kerr, E. a. (2006). The impact of comorbid chronic conditions on diabetes care. *Diabetes Care*, *29*(3), 725–731.
- Pories, W. J., & Dohm, G. L. (2012). Diabetes: have we got it all wrong? Hyperinsulinism as the culprit: surgery provides the evidence. *Diabetes Care*, *35*(12), 2438–2442.
- Preis, S. R., Pencina, M. J., Hwang, S., Sr, B. D. A., Savage, P. J., Levy, D., & Fox, C. S. (2009). Trends in cardiovascular disease risk factors in individuals with and without diabetes in the Framingham Heart Study. *Circulation*, *120*(3), 212–220.
- Rahier, J., Guiot, Y., Goebbels, R. M., Sempoux, C., & Henquin, J. C. (2008). Pancreatic beta-cell mass in European subjects with type 2 diabetes. *Diabetes, Obesity & Metabolism*, *10*(Suppl 4), 32–42.
- Ramachandran, A., Ma, R. C. W., & Snehalatha, C. (2010). Diabetes in Asia. *Lancet*, *375*(9712), 408–18.
- Rasdi, N. H., Samah, O. A., Sule, A., & Ahmed, Q. U. (2010). Antimicrobial studies of *Cosmos caudatus* Kunth . (*Compositae*). *Journal of Medicinal Plants Research*, *4*(April), 669–673.

- Razali, N., Mat-Junit, S., Abdul-Muthalib, A. F., Subramaniam, S., & Abdul-Aziz, A. (2012). Effects of various solvents on the extraction of antioxidant phenolics from the leaves, seeds, veins and skins of *Tamarindus indica* L. *Food Chemistry*, *131*(2), 441–448.
- Rebello, S. a, Chen, C. H., Naidoo, N., Xu, W., Lee, J., Chia, K. S., ... van Dam, R. M. (2011). Coffee and tea consumption in relation to inflammation and basal glucose metabolism in a multi-ethnic Asian population: a cross-sectional study. *Nutrition Journal*, *10*(1), 61.
- Reihani, S., & Azhar, M. (2012). Antioxidant activity and total phenolic content in aqueous extracts of selected traditional Malay salads (Ulam). *International Food Research Journal*, *19*(4), 1439–1444. Retrieved from [http://ifrj.upm.edu.my/19\(04\)2012/21IFRJ19\(04\)2012Azhar\(036\).pdf](http://ifrj.upm.edu.my/19(04)2012/21IFRJ19(04)2012Azhar(036).pdf)
- Rhee, Y., & Brunt, A. (2011). Flaxseed supplementation improved insulin resistance in obese glucose intolerant people: a randomized crossover design. *Nutrition Journal*, *10*, 44.
- Rhodes, C. J. (2005). Type 2 Diabetes - a matter of beta-cell life and death? *Science*, *307*(5708), 380–384.
- Riccardi, G., Giacco, R., & Rivellese, A. . (2004). Dietary fat, insulin sensitivity and the metabolic syndrome. *Clinical Nutrition*, *23*(4), 447–456.
- Roberts, L. D., Koulman, A., & Griffin, J. L. (2014). Towards metabolic biomarkers of insulin resistance and type 2 diabetes: progress from the metabolome. *The Lancet. Diabetes & Endocrinology*, *2*(1), 65–75.
- Rytter, E., Vessby, B., Asgard, R., Ersson, C., Moussavian, S., Sjodin, A., ... Basu, S. (2010). Supplementation with a combination of antioxidants does not affect glycaemic control, oxidative stress or inflammation in type 2 diabetes subjects. *Free Radical Research*, *44*(12), 1445–1453.
- Sachdewa, A., & Khemani, L. D. (2003). Effect of *Hibiscus rosa sinensis* Linn. ethanol flower extract on blood glucose and lipid profile in streptozotocin induced diabetes in rats. *Journal of Ethnopharmacology*, *89*(1), 61–66.
- Salek, R. M., Maguire, M. L., Bentley, E., Rubtsov, D. V, Hough, T., Cheeseman, M., ... Griffin, J. L. (2007). A metabolomic comparison of urinary changes in type 2 diabetes in mouse, rat, and human. *Physiological Genomics*, *29*, 99–108.
- Samy, J., Sugumaran, M., & Lee, K. L. . (2005). *Herbs of Malaysia: an introduction to medicinal, culinary, aromatic and cosmetic use of herbs*. Selangor: Federal Publications Sdn. Bhd.
- Sanchez, M. C., Larrauri, J. A., & Saura, C. F. (1998). A procedure to measure the antiradical efficiency of polyphenols. *Journal of the Science of Food and*

Agriculture, 76(2), 270–276.

- Saudek, C. D., & Brick, J. C. (2009). The clinical use of hemoglobin A1c. *Journal of Diabetes Science and Technology*, 3(4), 629–34.
- Savorani, F., Rasmussen, M. A., Mikkelsen, M. S., & Engelsen, S. B. (2013). A primer to nutritional metabolomics by NMR spectroscopy and chemometrics. *Food Research International*, 54(1), 1131–1145.
- Scalbert, A., Johnson, I. T., & Saltmarsh, M. (2005). Polyphenols: antioxidants and beyond. *The American Journal of Clinical Nutrition*, 81(1 Suppl), 215–217.
- Scartezzini, P., Antognoni, F., Raggi, M. a., Poli, F., & Sabbioni, C. (2006). Vitamin C content and antioxidant activity of the fruit and of the Ayurvedic preparation of *Emblca officinalis* Gaertn. *Journal of Ethnopharmacology*, 104(1-2), 113–118.
- Scazzocchio, B., Vari, R., Filesi, C., D'Archivio, M., Santangelo, C., Giovannini, C., ... Masella, R. (2011). Cyanidin-3-O- β -glucoside and protocatechuic acid exert insulin-like effects by upregulating PPAR γ activity in human omental adipocytes. *Diabetes*, 60(9), 2234–44.
- Scheijen, J. L. J. M., Hanssen, N. M. J., van de Waarenburg, M. P. H., Jonkers, D. M. A. E., Stehouwer, C. D. A., & Schalkwijk, C. G. (2012). L(+) and D(-) lactate are increased in plasma and urine samples of type 2 diabetes as measured by a simultaneous quantification of L(+) and D(-) lactate by reversed-phase liquid chromatography tandem mass spectrometry. *Experimental Diabetes Research*, 2012, 234812.
- Schulz, K. F., Altman, D. G., & Moher, D. (2010). CONSORT 2010 Statement: updated guidelines for reporting parallel group randomised trials. *Trials*, 11, 32.
- Sekar, M., Zuhlilmi, M., Hamdi, A. Y., Nabila, N., Zahida, Z., & Syafiq, M. (2014). Ten commonly available medicinal plants in Malaysia used for the treatment of diabetes-a review. *Asian Journal of Pharmaceutical and Clinical Research*, 7(1), 1–5.
- Shahar, S., Earland, J., & Abdulrahman, S. (2000). Validation of a dietary history questionnaire against a 7-D weighed record for estimating nutrient intake among rural elderly Malays. *Malaysian Journal of Nutrition*, 6(1), 33–44.
- Shahidi, F., & Naczk, M. (2004). *Phenolic in Food and Nutraceutical*. Boca Raton: CRC press.
- Shazwani, M. N. N., Suzana, S., Lim, C. J., Teh, C. S., Fauzee, M. Z. M., Lim, H. C., ... Norliza, M. (2010). Assessment of physical activity level among individuals with type 2 Diabetes Mellitus at Cheras Health Clinic, Kuala Lumpur. *Malaysian Journal of Nutrition*, 16(1), 101–112.

- Shui, G., Leong, L. P., & Wong, S. P. (2005). Rapid screening and characterisation of antioxidants of *Cosmos caudatus* using liquid chromatography coupled with mass spectrometry. *Journal of Chromatography. B, Analytical Technologies in the Biomedical and Life Sciences*, 827(1), 127–38.
- Shyam, S., Arshad, F., Abdul Ghani, R., Wahab, N. A., Safii, N., Nisak, M. Y., ... Kamaruddin, N. (2013). Low glycaemic index diets improve glucose tolerance and body weight in women with previous history of gestational diabetes: a six months randomized trial. *Nutrition Journal*, 12(1), 68.
- Singh, B., & Saxena, A. (2010). Surrogate markers of insulin resistance: A review. *World Journal of Diabetes*, 1(2), 36–47.
- Smolinska, A., Blanchet, L., Buydens, L. M. C., & Wijmenga, S. S. (2012). NMR and pattern recognition methods in metabolomics: from data acquisition to biomarker discovery: a review. *Analytica Chimica Acta*, 750, 82–97.
- Solanky, K. S., Bailey, N. J., Beckwith-hall, B. M., Bingham, S., Davis, A., Holmes, E., ... Cassidy, A. (2005). Biofluid ¹H NMR-based metabonomic techniques in nutrition research — metabolic effects of dietary isoflavones in humans. *Journal of Nutritional Biochemistry*, 16, 236–244.
- Song, Y., Cook, N. R., Albert, C. M., Denburgh, M. Van, & Manson, J. E. (2009). Effects of vitamins C and E and beta-carotene on the risk of type 2 diabetes in women at high risk of cardiovascular disease: a randomized controlled trial. *American Journal of Clinical Nutrition*, 90(2), 429–437.
- Song, Y., Manson, J. E., Buring, J. E., Sesso, H. D., & Liu, S. (2005). Associations of dietary flavonoids with risk of type 2 diabetes, and markers of insulin resistance and systemic inflammation in women: a prospective study and cross-sectional analysis. *Journal of the American College of Nutrition*, 24(5), 376–384.
- Sousa, E. O., Miranda, C. M. B. a., Nobre, C. B., Boligon, A. a., Athayde, M. L., & Costa, J. G. M. (2015). Phytochemical analysis and antioxidant activities of *Lantana camara* and *Lantana montevidensis* extracts. *Industrial Crops and Products*, 70, 7–15.
- Stagos, D., Portesis, N., Spanou, C., Mossialos, D., Aligiannis, N., Chaita, E., ... Kouretas, D. (2012). Correlation of total polyphenolic content with antioxidant and antibacterial activity of 24 extracts from Greek domestic Lamiaceae species. *Food and Chemical Toxicology*, 50(11), 4115–4124.
- Stancakova, A., Civelek, M., Saleem, N. K., Soininen, P., Kangas, A. J., Cederberg, H., ... Laakso, M. (2012). Hyperglycemia and a common variant of GCKR are associated with the levels of eight amino acids in 9,369 Finnish men. *Diabetes*, 61(7), 1895–902.
- Stella, C., Beckwith-Hall, B., Cloarec, O., Holmes, E., Lindon, J. C., Powell, J., ...

- Nicholson, J. K. (2006). Susceptibility of human metabolic phenotypes to dietary modulation. *Journal of Proteome Research*, 5(10), 2780–2788.
- Stratton, I. M., Adler, A. I., Neil, H. A., Matthews, D. R., Manley, S. E., Cull, C. A., ... Holman, R. R. (2000). Association of glycaemia with macrovascular and microvascular complications of type 2 diabetes (UKPDS 35): prospective observational study. *British Medical Journal*, 321(7258), 405–412.
- Stumvoll, M., Goldstein, B. J., & van Haeften, T. W. (2005). Type 2 diabetes: principles of pathogenesis and therapy. *Lancet*, 365(9467), 1333–1346.
- Styskal, J., Van Remmen, H., Richardson, A., & Salmon, A. B. (2012). Oxidative stress and diabetes: What can we learn about insulin resistance from antioxidant mutant mouse models? *Free Radical Biology and Medicine*, 52(1), 46–58.
- Suhre, K., Meisinger, C., Doring, A., Altmayer, E., Belcredi, P., Gieger, C., ... Illig, T. (2010). Metabolic footprint of diabetes: a multiplatform metabolomics study in an epidemiological setting. *PLoS One*, 5(11), e13953.
- Sukrasno, Fidriany, I., Anggadiredja, K., Handayani, W. A., & Anam, K. (2011). Influence of drying method on flavonoid content of *Cosmos caudatus* (kunth) leaves.pdf. *Research Journal of Medicinal Plant*, 5(2), 189–195.
- Sullivan, G. M., & Feinn, R. (2012). Using effect size - or why the P Value is not enough. *Journal of Graduate Medical Education*, 4(3), 279–82.
- Surya, S., Salam, A. D., Tomy, D. V., Carla, B., Kumar, R. A., & Sunil, C. (2014). Diabetes mellitus and medicinal plants-a review. *Asian Pacific Journal of Tropical Disease*, 4(5), 337–347.
- Tan, B. K. H., Tan, C. H., & Pushparaj, P. N. (2005). Anti-diabetic activity of the semi-purified fractions of *Averrhoa bilimbi* in high fat diet fed-streptozotocin-induced diabetic rats. *Life Sciences*, 76(24), 2827–2839.
- Tan, M. C., Ng, O. C., Wong, T. W., Joseph, A., Hejar, A. R., & Rushdan, A. A. (2015). Dietary compliance, dietary supplementation and traditional remedy usage of type 2 diabetic patients with and without cardiovascular disease. *Clinical Nutrition Research*, 4(1), 18–31.
- Tee, E. S., Mohd. Ismail, N., Mohd Nasir, A., & Khatijah, I. (1997). *Nutrient composition of Malaysian foods* (4th editio.). Kuala Lumpur: Institute for Medical Research.
- Tesfaye, F., Nawi, N. G., Van Minh, H., Byass, P., Berhane, Y., Bonita, R., & Wall, S. (2007). Association between body mass index and blood pressure across three populations in Africa and Asia. *Journal of Human Hypertension*, 21(1), 28–37.
- Thaipong, K., Boonprakob, U., Crosby, K., Cisneros-Zevallos, L., & Hawkins Byrne, D. (2006). Comparison of ABTS, DPPH, FRAP, and ORAC assays for estimating

antioxidant activity from guava fruit extracts. *Journal of Food Composition and Analysis*, 19(6-7), 669–675.

- Tharkar, S., Devarajan, A., Kumpatla, S., & Viswanathan, V. (2010). The socioeconomics of diabetes from a developing country: A population based cost of illness study. *Diabetes Research and Clinical Practice*, 89(3), 334–340.
- Thilagam, E., Parimaladevi, B., Kumarappan, C., & Chandra Mandal, S. (2013). α -Glucosidase and α -Amylase Inhibitory Activity of *Senna surattensis*. *Journal of Acupuncture and Meridian Studies*, 6(1), 24–30.
- Thrower, S. L., & Bingley, P. J. (2014). What is type 1 diabetes? *Medicine*, 42(12), 682–686.
- Timmers, S., Konings, E., Bilet, L., Houtkooper, R. H., van de Weijer, T., Goossens, G. H., ... Schrauwen, P. (2011). Calorie restriction-like effects of 30 days of resveratrol supplementation on energy metabolism and metabolic profile in obese humans. *Cell Metabolism*, 14(5), 612–22.
- Tojo, A., & Kinugasa, S. (2012). Mechanisms of glomerular albumin filtration and tubular reabsorption. *International Journal of Nephrology*, 2012, 481520.
- Traber, M. G., & Stevens, J. F. (2011). Vitamins C and E: Beneficial effects from a mechanistic perspective. *Free Radical Biology and Medicine*, 51(5), 1000–1013.
- Traynor, J., Mactier, R., Geddes, C. C., & Fox, J. G. (2006). How to measure renal function in clinical practice. *British Medical Journal*, 333(7571), 733–7.
- Trifonova, O., Likhov, P., & Archakov, A. (2013). Postgenomics diagnostics: metabolomics approaches to human blood profiling. *Omics*, 17(11), 550–9.
- Trygg, J., & Wold, S. (2002). Orthogonal projections to latent structures (O-PLS). *Journal of Chemometrics*, 16(3), 119–128.
- van Dieren, S., Beulens, J. W. J., van der Schouw, Y. T., Grobbee, D. E., & Neal, B. (2010). The global burden of diabetes and its complications: an emerging pandemic. *European Journal of Cardiovascular Prevention and Rehabilitation*, 17(Suppl 1), S3–S8.
- van Doorn, M., Vogels, J., Tas, A., van Hoogdalem, E. J., Burggraaf, J., Cohen, A., & van der Greef, J. (2007). Evaluation of metabolite profiles as biomarkers for the pharmacological effects of thiazolidinediones in type 2 diabetes mellitus patients and healthy volunteers. *British Journal of Clinical Pharmacology*, 63(5), 562–74.
- Van Dorsten, F. a, Daykin, C. a, Mulder, T. P. J., & Van Duynhoven, J. P. M. (2006). Metabonomics approach to determine metabolic differences between green tea and black tea consumption. *Journal of Agricultural and Food Chemistry*, 54(18), 6929–38.

- Van Raalte, D. H., & Diamant, M. (2011). Glucolipotoxicity and beta cells in type 2 diabetes mellitus: Target for durable therapy? *Diabetes Research and Clinical Practice*, *93*, S37–S46.
- Velioglu, Y. S., Mazza, G., Gao, L., & Oomah, B. D. (1998). Antioxidant activity and total phenolics in selected fruits, vegetables, and grain products. *Journal of Agricultural and Food Chemistry*, *46*(10), 4113–4117.
- Walsh, M. C., Brennan, L., Pujos-guillot, E., Sébédio, J., Scalbert, A., Fagan, A., ... Gibney, M. J. (2007). Influence of acute phytochemical intake on human urinary metabolomic profiles. *The American Journal of Clinical Nutrition*, *86*, 1687–1693.
- Wang, T. J., Larson, M. G., Vasan, R. S., Cheng, S., Rhee, E. P., McCabe, E., ... Gerszten, R. E. (2011). Metabolite profiles and the risk of developing diabetes. *Nature Medicine*, *17*(4), 448–53.
- Wang, Y., Tang, H. R., Nicholson, J. K., Hylands, P. J., Sampson, J., & Holmes, E. (2005). A metabonomic strategy for the detection of the metabolic effects of Chamomile (*Matricaria recutita* L.) Ingestion. *Journal of Agricultural and Food Chemistry*, *53*, 191–196.
- Weyer, C., Bogardus, C., Mott, D. M., & Pratley, R. E. (1999). The natural history of insulin secretory dysfunction and insulin resistance in the pathogenesis of type 2 diabetes mellitus. *The Journal of Clinical Investigation*, *104*(6), 787–94.
- WHO Expert Consultation. (2004). Appropriate body-mass index for Asian populations and its implications for policy and intervention strategies. *Lancet*, *363*(9403), 157–63.
- Wilding, J. P. H. (2007). The importance of free fatty acids in the development of Type 2 diabetes. *Diabetic Medicine*, *24*(9), 934–45.
- Williams, H. R. T., Cox, I. J., Walker, D. G., Cobbold, J. F. L., Taylor-Robinson, S. D., Marshall, S. E., & Orchard, T. R. (2010). Differences in gut microbial metabolism are responsible for reduced hippurate synthesis in Crohn's disease. *BMC Gastroenterology*, *10*, 108.
- Woerle, H. J., Neumann, C., Zschau, S., Tenner, S., Irsigler, A., Schirra, J., ... Göke, B. (2007). Impact of fasting and postprandial glycemia on overall glycemic control in type 2 diabetes Importance of postprandial glycemia to achieve target HbA1c levels. *Diabetes Research and Clinical Practice*, *77*(2), 280–5.
- Wolf, A., Conaway, M., Crowther, J., Hazen, K., Nadler, L., Oneida, B., & Bovbjerg, V. (2004). Translating lifestyle intervention to practice in obese patients. *Diabetes Care*, *27*, 1570–1576.
- Wong, S., Leong, L., & Williamkoh, J. (2006). Antioxidant activities of aqueous extracts of selected plants. *Food Chemistry*, *99*(4), 775–783.

- World Health Organization. (2002). Diabetes: the cost of diabetes, fact sheet no. 236. Geneva.
- World Health Organization. (2004). *WHO guidelines on safety monitoring of herbal medicines in pharmacovigilance systems*. Geneva.
- Wu, S., Hopper, I., Skiba, M., & Krum, H. (2014). Dipeptidyl peptidase-4 inhibitors and cardiovascular outcomes: meta-analysis of randomized clinical trials with 55,141 participants. *Cardiovascular Therapeutics*, 32(4), 147–158.
- Xi, Y., & Roetze, D. M. (2008). Baseline Correction for NMR Spectroscopic Metabolomics Data Analysis. *BMC Bioinformatics*, 9(1), 324.
- Yang, W., Dall, T. M., Halder, P., Gallo, P., Kowal, S. L., Hogan, P. F., & Petersen, M. (2013). Economic costs of diabetes in the U.S. in 2012. *Diabetes Care*, 36, 1033–1046.
- Yeni-Komshian, H., Carantoni, M., Abbasi, F., & Reaven, G. M. (2000). Relationship between several surrogate estimates of insulin resistance and quantification of insulin-mediated glucose disposal in 490 healthy nondiabetic volunteers. *Diabetes Care*, 23(2), 171–175.
- Yki-Jarvinen, H. (2004). Thiazolidinediones. *The New England Journal of Medicine*, 351(11), 1106–1118.
- Yokoyama, H., Emoto, M., Fujiwara, S., Motoyama, K., Morioka, T., Komatsu, M., ... Nishizawa, Y. (2003). Quantitative insulin sensitivity check index and the reciprocal index of homeostasis model assessment in normal range weight and moderately obese type 2 diabetic patients. *Diabetes Care*, 26(8), 2426–2432.
- Yoon, K. H., Ko, S. H., Cho, J. H., Lee, J. M., Ahn, Y. B., Song, K. H., ... Bonner-Weir, S. (2003). Selective beta-cell loss and alpha-cell expansion in patients with type 2 diabetes mellitus in Korea. *The Journal of Clinical Endocrinology and Metabolism*, 88(5), 2300–8.
- Yousri, N. A., Mook-Kanamori, D. O., Selim, M. M. E.-D., Takiddin, A. H., Al-Homsi, H., Al-Mahmoud, K. A. S., ... Suhre, K. (2015). A systems view of type 2 diabetes-associated metabolic perturbations in saliva, blood and urine at different timescales of glycaemic control. *Diabetologia*, 58(8), 1855–1867.
- Zamora-Ros, R., Urpí-Sardà, M., Lamuela-Raventós, R. M., Estruch, R., Martínez-González, M. Á., Bulló, M., ... Andres-Lacueva, C. (2009). Resveratrol metabolites in urine as a biomarker of wine intake in free-living subjects: The PREDIMED Study. *Free Radical Biology and Medicine*, 46(12), 1562–1566.
- Zhang, A., Qiu, S., Xu, H., Sun, H., & Wang, X. (2014). Metabolomics in diabetes. *Clinica Chimica Acta*, 429, 106–110.

Zhang, Y., & Zhang, H. (2013). Microbiota associated with type 2 diabetes and its related complications. *Food Science and Human Wellness*, 2(3–4), 167–172.

Ziener, D. C., Berkowitz, K. J., Panayioto, R. M., El-Kebbi, I. M., Musey, V. C., Anderson, L. a, ... Phillips, L. S. (2003). A simple meal plan emphasizing healthy food choices is as effective as an exchange-based meal plan for urban African Americans with type 2 diabetes. *Diabetes Care*, 26(6), 1719–24.

