



**UNIVERSITI PUTRA MALAYSIA**

***GASTROPROTECTIVE AND ANTI-INFLAMMATORY ACTIVITIES OF A  
MIXTURE OF *Melastoma malabathricum* L. AND *Muntingia calabura* L.  
LEAVES***

**SITI ZAWANAH BINTI HALIM**

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By

**SITI ZAWANAH BINTI HALIM**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,  
in Fulfillment of the Requirements for the Degree of Master of Science**

**June 2017**

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Master of Science

**GASTROPROTECTIVE AND ANTI-INFLAMMATORY ACTIVITIES OF A MIXTURE OF *Melastoma malabathricum* L. AND *Muntingia calabura* L. LEAVES**

By

**SITI ZAWANAH BINTI HALIM**

**June 2017**

**Chairman : Associate Professor Zainul Amiruddin Zakaria, PhD**  
**Faculty : Medicine and Health Sciences**

Gastric ulcer is a part of the peptic ulcer disease which well acknowledged as one of major human illness. Basically, the gastric ulcer is occurred due to the imbalance between the protective factors and aggressive factors in the stomach. Recently, there were a few of medicines being used to treat the gastric ulcer. However, those medicines had caused certain adverse effects. At the same time, medicinal plants have proved to give positive results as the treatment for many diseases including gastric ulcer. *Melastoma malabathricum* (MM) and *Muntingia calabura* (MC) were two plants which have been studied for their pharmacological activities. In addition, the previous study had also reported the markedly gastroprotective effect of both those plants leaves individually via *in vitro* and *in vivo* experiments. Thus, in attempt to develop a pharmaceutical product with antiulcer potential, it is necessary to make sure it is effective at lower dose range, so the present study was aimed to identify the gastroprotective activity of methanol extract of a mixture of *Melastoma malabathricum* and *Muntingia calabura* (MMMC) at various ratio using rats models and further investigate whether combination of those plant at various ratio will exert synergistic effect by enhancing or reducing the observed antiulcer activity. The small particle of MM and MC grinded leaves were mixed together based on three different ratios (1:1, 1:3 and 3:1; w/w) and soaked in methanol in ratio 1:20 for 72 hours, filtered and evaporated using the rotatory evaporator to get the concentrated crude extract. By using the crude extract obtained from the evaporation process, the doses (15, 150 or 300 mg/kg) were prepared. Ranitidine (100 mg/kg) was used as a reference drug while 2% Tween 80 was used as a negative control. The Sprague-Dawley rats were given the test solutions orally for seven consecutive days and were subjected to the ethanol-induced gastric ulcer. The rats were euthanized; macroscopic and histological observations of the gaster were done. The ulcer area (UA) was determined and the percentage of the inhibition exhibited by the MMMC was calculated. The gaster was subjected to antioxidant studies including the superoxide dismutase (SOD), catalase

(CAT), glutathione (GSH), prostaglandin E<sub>2</sub> (PGE<sub>2</sub>) and malondialdehyde (MDA) assay. The MMMC was tested for antioxidant study using the total phenolic content (TPC), oxygen radical absorbance capacity assay (ORAC) and 2,2-diphenyl-1-picrylhydrazyl radical scavenging assay (DPPH). The antisecretory potential of MMMC was investigated using the pyloric ligation model while the role of nitric oxide and endogenous sulfhydryl group were measured by testing the extract against L-NG-Nitroarginine Methyl Ester (L-NAME) and N-Ethylmaleimide (NEM) using the ethanol-induced model. Ratio 1:1 of MMMC showed the most effective gastroprotective activity in which it significantly ( $P < 0.05$ ) reduced the gastric lesion when assessed using the ethanol-induced gastric ulcer models. The macroscopic findings were validated by the microscopic observations. Furthermore, except for the pH and total acidity, the extract also significantly ( $P < 0.05$ ) reduced the volume of gastric content whereas the mucus content was significantly ( $P < 0.05$ ) increased in MMMC-treated rats in the pyloric-ligation test, where by ratio 1:1 constantly exhibited the best result among the three ratios. Besides that, the gastroprotection effect of MMMC significantly ( $P < 0.05$ ) involved the participation of nitric oxide and sulfhydryl compounds. Ratio 1:1 of MMMC showed the highest value of TPC and ORAC activity compared to the other ratios, meanwhile ratio 3:1 of MMMC exhibited the strongest IC<sub>50</sub> of DPPH scavenging activity. The *in-vitro* anti-inflammatory effect of MMMC evaluation showed moderate NO inhibitory activity at IC<sub>50</sub> of ratio 1:1 and 1:3 of MMMC, compared to ratio 3:1 which showed weak NO inhibitory activity. While the antioxidants studies of the stomach revealed the improvement of CAT, SOD, GSH, PGE<sub>2</sub> and MDA activities by MMMC with ratio 1:1 showed the best result among the three ratios. Besides that, the result obtained from the GC-MS analysis showed several peaks detected at the different real time suggested that MMMC contained several compounds that have been proven by other studies to show antioxidant, anti-inflammatory and gastroprotective effect. The UHPLC analysis suggested that the presence of quercetin and gallic acid in the most effective ratio of MMMC, which is 1:1 was confirmed in MMMC. In conclusion, MMMC at the ratio of 1:1, exerted the best gastroprotective activity against the ethanol-induced gastric ulcer assay partly by activating its antisecretory and antioxidant activities, together with modulation of the gastric tissue endogenous antioxidant system.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

**AKTIVITI PERLINDUNGAN GASTRO DAN ANTI-INFLAMASI OLEH  
CAMPURAN DAUN-DAUN *Melastoma malabathricum* L. DAN *Muntingia  
calabura* L**

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Ulser gastrik adalah sebahagian daripada penyakit ulser peptik yang telah sedia maklum merupakan salah satu penyakit manusia yang major. Secara asasnya, ulser gastrik berlaku disebabkan oleh ketidakimbangan di antara faktor-faktor perlindungan dan faktor-faktor agresif di dalam perut. Baru-baru ini, terdapat beberapa ubat-ubatan yang digunakan untuk merawat ulser gastrik. Namun, ubat-ubatan tersebut telah menyebabkan kesan-kesan buruk yang tertentu. Pada masa yang sama, tumbuh-tumbuhan perubatan telah terbukti memberi keputusan-keputusan positif sebagai rawatan untuk pelbagai penyakit termasuk ulser gastrik. *Melastoma malabathricum* (MM) dan *Muntingia calabura* (MC) merupakan dua tumbuh-tumbuhan yang telah dikaji mengenai aktiviti-aktiviti farmakologi mereka. Tambahan pula, kajian sebelum ini telah melaporkan kesan perlindungan gastro yang signifikan oleh daun daripada kedua-dua tumbuhan tersebut secara berasingan melalui eksperimen *in vitro* dan *in vivo*. Oleh itu, dalam percubaan mengembangkan sesuatu produk farmasiutikal, ianya menjadi keperluan untuk memastikan produk tersebut berkesan pada kadar dos yang lebih rendah, maka kajian ini dijalankan bertujuan untuk mengenalpasti aktiviti sinergi perlindungan gastro oleh ekstrak metanol dari campuran *Melastoma malabathricum* dan *Muntingia calabura* (MMMC) pada nisbah yang berbeza menggunakan model tikus dan seterusnya dikaji samada gabungan kedua-dua tumbuhan tersebut pada nisbah yang berbeza mampu menghasilkan kesan sinergi melalui peningkatan atau penurunan aktiviti anti ulser yang diperhatikan. Zarah-zarah kecil daripada daun-daun MM dan MC yang telah dikisar, dicampur bersama-sama berdasarkan tiga nisbah yang berbeza (1:1, 1:3 dan 3:1; w/w) dan direndam di dalam metanol dalam nisbah 1:20 selama 72 jam, ditapis dan disejat menggunakan pemutar penyejat untuk mendapatkan mentah ekstrak yang pekat. Dengan menggunakan mentah ekstrak yang diperolehi daripada proses penyejatan, dos-dos (15, 150 atau 300 mg/kg) disediakan. Ranitidine (100 mg/kg) telah digunakan sebagai dadah rujukan sementara 2% Tween 80 telah digunakan sebagai kawalan negatif. Tikus-tikus jenis *Sprague-Dawley* tersebut telah

diberikan solusi –solusi yang diuji melalui oral untuk tujuh hari berturut-turut dan telah di arah kepada ulser gastrik rangsangan etanol. Tikus-tikus tersebut telah dieutanasia; pemerhatian makroskopik dan histologi ke atas gaster telah dijalankan. Kawasan ulser (UA) telah dikenalpasti dan peratusan perencatan yang telah ditunjukkan oleh MMMC turut dikira. Gaster telah di arah kepada kajian-kajian antioksidan termasuk eksperimen superoxide dismutase (SOD), catalase (CAT), glutathione (GSH), prostaglandin E<sub>2</sub> (PGE<sub>2</sub>) dan malondialdehyde (MDA). MMMC telah diuji untuk kajian antioksidan menggunakan jumlah kandungan fenolik (TPC), eksperimen kapasiti penyerapan oksigen radikal (ORAC) dan eksperimen memerangkap radikal 2,2- diphenyl-1-picrylhydrazyl (DPPH). Potensi anti-rembesan oleh MMMC telah dikaji menggunakan model ikatan pilorik sementara peranan nitrik oksida dan kumpulan sulfhydryl dalaman disukat dengan menguji ekstrak terhadap L-NG-Nitroarginine Methyl Ester (L-NAME) dan N-Ethylmaleimide (NEM) menggunakan model rangsangan etanol. Nisbah 1:1 MMMC menunjukkan aktiviti perlindungan gastro yang paling efektif di mana ia mengurangkan luka gastrik secara signifikan ( $P < 0.05$ ) apabila diuji menggunakan model gastrik ulser rangsangan etanol. Penemuan makroskopik telah disahkan menggunakan pemerhatian mikroskopik. Tambahan pula, kecuali pH dan jumlah keasidan, ekstrak tersebut turut mengkurangkan isipadu kandungan gastrik secara signifikan ( $P < 0.05$ ) sedangkan kandungan mukus telah meningkat secara signifikan ( $P < 0.05$ ) dalam kumpulan tikus yang dirawat oleh MMMC dalam kajian ikatan pilorik, di mana nisbah 1:1 secara terus mempamerkan keputusan terbaik di antara ketiga-tiga nisbah. Selain itu, kesan perlindungan gastro oleh MMMC telah melibatkan penyertaan nitrik oksida dan sebatian sulfhydryl secara signifikan ( $P < 0.05$ ). Nisbah 1:1 MMMC telah menunjukkan nilai TPC dan aktiviti ORAC yang paling tinggi berbanding nisbah yang lain, sementara nisbah 3:1 MMMC mempamerkan IC<sub>50</sub> aktiviti memerangkap DPPH yang paling kuat. Penilaian kesan anti-keradangan secara *in-vitro* oleh MMMC menunjukkan bahawa aktiviti penyekatan NO pada IC<sub>50</sub> adalah sederhana bagi nisbah 1:1 dan 1:3 MMMC, berbanding nisbah 3:1 yang menunjukkan aktiviti penyekatan NO yang lemah. Sementara kajian-kajian antioksidan terhadap perut mendedahkan penambahbaikan aktiviti-aktiviti CAT, SOD, GSH, PGE<sub>2</sub> dan MDA oleh MMMC dengan nisbah 1:1 telah menunjukkan keputusan paling baik di antara ketiga-tiga nisbah. Selain itu, keputusan yang diperoleh daripada analisa GC-MS menunjukkan beberapa puncak-puncak telah dikesan pada masa sebenar yang berbeza yang mencadangkan bahawa MMMC mengandungi beberapa sebatian yang telah terbukti oleh kajian-kajian lain menunjukkan kesan antioksidan, anti-keradangan dan perlindungan gastro. Analisa UHPLC mencadangkan bahawa kehadiran quercetin dan gallic acid di dalam nisbah MMMC yang paling efektif iaitu 1:1 adalah pasti di dalam MMMC. Kesimpulannya, MMMC pada nisbah 1:1, telah menunjukkan aktiviti perlindungan gastro yang terbaik terhadap eksperimen ulser gastrik yang dirangsang oleh etanol, sebahagiannya melalui pengaktifan aktiviti-aktiviti anti-rembesan dan antioksidanya, bersama-sama pemodulasian sistem antioksidasi di dalam tisu gastrik.

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I certify that a Thesis Examination Committee has met on 5 June 2017 to conduct the final examination of Siti Zawanah binti Halim on her thesis entitled "Gastroprotective and Anti-Inflammatory Activities of a Mixture of *Melastoma malabathricum* L. and *Muntingia calabura* L. Leaves" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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## LIST OF ABBREVIATIONS

AAPH	2,2'-Azobis 92-methylpropionamide dihydrochloride
AchE	Acetylcholinesterase
ANOVA	Analysis of variance
ASA	Acetylsalicylic acid
cAMP	Cyclic adenosine monophosphate
CAT	Catalase
CuSO <sub>4</sub>	Copper Sulphate
DMEM	Dulbecco's Modified Eagle's Medium
DMSO	Dimethyl sulfoxide
DPPH	2,2-Diphenyl-1-Picrylhydrazyl
ELISA	Enzyme-linked immunosorbent assay
FBS	Fetal bovine serum
GAE	Gallic acid equivalent
GC-MS	Gas Chromatography – Mass Spectral
GSH	Glutathione
HCl	Hydrochloric acid
HEPES	4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid
HPLC	High-performance liquid chromatography
H <sub>2</sub> O <sub>2</sub>	Hydrogen peroxide
IACUC	Institutional Animal Care and Use Committee
IFN- $\gamma$	<i>Interferon gamma</i>
IHME	Institute for Health Metrics and Evaluation
LC-MS	Liquid chromatography- mass spectrometry

L-NAME	$N^G$ - $\omega$ -nitro-L-arginine methyl ester
LPS	<i>Lipopolysaccharide</i>
MDA	Malondialdehyde
MgCl	Magnesium Chloride
MMMC	<i>Melastoma malabathricum Muntingia calabura</i>
NaCl	Sodium Chloride
NaOH	Natrium Hydroxide
Na <sub>2</sub> CO <sub>3</sub>	Sodium carbonate
NEM	N-ethylmaleimide
nm	Nanometer
NO	Nitric oxide
NOS	<i>Nitric oxide synthase</i>
NO <sub>2</sub> <sup>-</sup>	Nitrite ions
NSAIDs	Non Steroidal anti-inflammatory drug
OD	Optical Density
ORAC	Oxygen radical absorbance capacity
PAF	Platelet-activating factor
PGE <sub>2</sub>	Prostaglandins
PGs	Prostaglandins
PUD	Peptic ulcer disease
RNS	Reactive nitrogen species
ROS	Reactive oxygen species
rpm	Revolutions per minute
SEM	Standard Error of Mean
SH	Sulfhydryl

SOD	Superoxide Dismutase
TBA	Thiobarbituric acid
TE	Trolox Equivalent
TPC	Total phenolic content
UA	Ulcer area
UHPLC-ESI	Ultra High Performance Liquid Chromatography- Electrospray Ionization
UV	Ultraviolet



## CHAPTER 1

### INTRODUCTION

Peptic ulcers have become one of the major human illness affecting nearly 8-10% of the global population (Calam and Baron, 2001), and of these number, 5% suffer from gastric ulcers (Bandyopadhyay *et al.*, 2001). Diverse factors such as alcohol consumption, a stressful lifestyle, use of steroidal and non-steroidal anti-inflammatory drugs (NSAIDs) and drugs which stimulate gastric acid and pepsin secretion, *Helicobacter pylori* infections and smoking contribute to the pathogenesis of gastric ulcers (Rang *et al.*, 2014). An imbalance between the aggressive factors such as acid and pepsin secretion, *Helicobacter pylori*, refluxed bile, release of leukotrienes and reactive oxygen species (ROS) and mucosal defensive factors that include bicarbonate secretion, mucus-bicarbonate barrier, surface active phospholipids, prostaglandins (PGs), mucosal blood flow, cell renewal and migration, non-enzymatic and enzymatic antioxidants and some growth factors (De Lira Mota *et al.*, 2009) leads to gastric damages.

The prevention or cure of peptic ulcers has become an important challenge in the current medicine world. Although gastric ulcer is linked to many causative factors, secretion of gastric acid is still believed to remain as the central component of this disease (De Lira Mota *et al.*, 2009). Thus, inhibition of gastric acid secretion is the key therapeutic target for ulcer diseases (Jain *et al.*, 2007). Therefore, current medicinal treatment of gastric ulcers available is generally based on the inhibition of gastric acid secretion by histamine H<sub>2</sub>-antagonists, proton pump inhibitors, and antimuscarinics, as well as on acid-independent therapy provided by sucralfate and bismuth cholinergic (Bighetti *et al.*, 2005). However, gastric ulcer therapy faces a major drawback nowadays as most of the drugs available in the market are often associated with adverse effects such as bradycardia and pancreatitis (Bandyopadhyay *et al.*, 2002; Rang *et al.*, 2014; Herrmann *et al.*, 1990; Johnson and Miller, 1988).

In this context, the use of medicinal plants has gained the interest of many researchers. The natural product is in continuous expansion all over the world and became the most attractive source of new drug for the treatment and prevention of many diseases. A diverse range of bioactive molecules isolated from plant natural product has been shown to produce promising results for the treatment of gastric ulcer (Borrelli and Izzo, 2000).

Two of the plants that are currently under investigation for their potential pharmacological activities in the laboratory are *Melastoma malabathricum* L. (family Melastomataceae) and *Muntingia calabura* L. (family Muntingiaceae). *Melastoma malabathricum* also was known as *senduduk ungu* in Malaysia. It is one of the most common herbs or small shrubs found throughout the tropic in the moist land mostly from India, Thailand and Malaysia. *Melastoma malabathricum*'s leaves, shoots, barks, seeds and roots have been used as a folk remedy to treat diarrhea, dysentery, hemorrhoids, cuts and wounds, toothache, and stomachache (Joffry *et al.*, 2011). Scientific evaluations on

*Melastoma malabathricum* have revealed several pharmacological activities possessed by the plant. *Melastoma malabathricum* leaves have been reported to exhibit significant antinociceptive, anti-inflammatory, wound healing, cytotoxic, antidiarrheal and antioxidant activities (Joffry *et al.*, 2011). Flavonoids, tannins, saponins, triterpenes and steroids have been detected in the leaves of *Melastoma malabathricum* (Mamat *et al.*, 2013).

While *Muntingia calabura*, commonly known as Jamaican cherry or *kerukup siam* in Malaysia, is widely cultivated in warm areas of the Asian region, including Malaysia (Chin, 1989). In Asia and tropical America, various parts of this tree have been documented for several medicinal uses. The leaves, flowers, barks and roots of *Muntingia calabura* have been used as a folk remedy to treat headaches, fever and incipient cold. Besides, they are also employed as antiseptic, antispasmodic, and antidyspeptic agent (Kaneda *et al.*, 1991; Nshimo *et al.*, 1993). It also has been scientifically validated to possess several pharmacological activities. Significant antinociception (Zakaria *et al.*, 2006b, 2007a, 2007b), antitumor (Kaneda *et al.*, 1991; Su *et al.*, 2003), anti-inflammatory, antipyretic (Zakaria *et al.*, 2007b), antibacterial (Sulaiman *et al.*, 2006), antiproliferative and antioxidant (Zakaria *et al.*, 2011b) activities have been exhibited by the leaves of *Muntingia calabura*. Several types of flavonoids have been isolated and identified from the leaves, roots and stem barks of *Muntingia calabura* ( Kaneda *et al.*, 1991; Nshimo *et al.*, 1993; Su *et al.*, 2003; Chen *et al.*, 2005; Sufian *et al.*, 2013).

The previous study has reported the significant antiulcer activity of methanol extract of *Melastoma malabathricum* and *Muntingia calabura* leaves individually using several animal models as well as *in vitro* antioxidant and anti-inflammatory activities (Balan *et al.*, 2014; Zakaria *et al.*, 2015). Therefore, in an attempt to develop a pharmaceutical product with gastroprotective potential, it is necessary to make sure it is effective at lower dose range. Since no study has been done to determine whether there is gastroprotective action between combination ratio of these two plants, which might demonstrate gastroprotective activity at lower dose range, the present study was aimed to investigate whether the combination of these plants at various ratio will exert a gastroprotective effect by enhancing or reducing the observed gastroprotective activity.

#### **Justification for studying the gastroprotective and anti-inflammatory activity of methanol extract of *Melastoma malabathricum* and *Muntingia calabura* leaves**

To date, there are many approaches (Konturek *et al.*, 2011) and advanced development of modern medicine. However, these scenario comes together with several obstacles, such as the presence of drugs adverse effect that might stop the patients with a certain health condition to consume the drugs, unaffordable and descend of drug availability. Because of that, there is calling for a research that can produce a product which is safe, effective in a low dose of consumption and affordable. The methanol extract of *Melastoma malabathricum* and *Muntingia calabura* leaves individually have been reported to show significant gastroprotective activity using several animal models as well as *in vitro* experiment of antioxidant and anti-inflammatory activities (Balan *et al.*, 2015; Mamat *et al.*, 2013; Mazura *et al.*, 2007; Zakaria *et al.*, 2011a; Zakaria *et*



*al.*, 2006a). However, in an attempt develop a pharmaceutical product with gastroprotective potential, it is necessary to make sure it is effective at lower dose range since there were many commercial drugs reported to cause an adverse effect when consumed at a certain dose which is considering high. In this study, the combination of those plant at the various ratio with gastroprotective activity will be investigated.

### **Hypothesis**

There are gastroprotective and anti-inflammatory activities of methanol extract of a mixture *Melastoma malabathricum* and *Muntingia calabura* leaves through *in vivo* and *in vitro* experiments.

### **General objectives**

- To identify the gastroprotective and anti-inflammatory activities of methanol extract of a mixture *Melastoma malabathricum* and *Muntingia calabura* leaves through *in vivo* and *in vitro* experiments.

### **Specific objectives**

- To determine the best ratio and lowest dose among the three combination ratios of mixture of *Melastoma malabathricum* and *Muntingia calabura* that gives the most effective gastroprotective and anti-inflammatory activities.
- To elucidate the physical changes through pyloric ligation model, L-NAME and NEM pre-treated ethanol-induced in rats.
- To determine the level of antioxidant activities of the extract as a part of the gastroprotective pathway.
- To screen for the compounds present in MMC using Gas Chromatography-Mass Spectrometry (GC-MS) and Ultra High Performance Liquid Chromatography-Electrospray Ionization (UHPLC-ESI).

## REFERENCES

- Aase, S. (1989). Disturbances in the balance between aggressive and protective factors in the gastric and duodenal mucosa. *Scandinavian Journal of Gastroenterology*, 24(sup163), 17-23.
- Abdelwahab, S. I. (2013). Protective mechanism of gallic acid and its novel derivative against ethanol-induced gastric ulcerogenesis: Involvement of immunomodulation markers, Hsp70 and Bcl-2-associated X protein. *International immunopharmacology*, 16(2), 296-305.
- Adinortey, M. B., Ansah, C., Galyuon, I., & Nyarko, A. (2013). In vivo models used for evaluation of potential antigastroduodenal ulcer agents. *Ulcers*, 2013.
- Agrawal, A. A., & Weber, M. G. (2015). On the study of plant defence and herbivory using comparative approaches: how important are secondary plant compounds. *Ecology letters*, 18(10), 985-991.
- Agunod, M., Yamaguchi, N., Lopez, R., Luhby, A. L., & Glass, G. B. J. (1969). Correlative study of hydrochloric acid, pepsin, and intrinsic factor secretion in newborns and infants. *Digestive Diseases and Sciences*, 14(6), 400-414.
- Akah, P. A., Onyirioha, C. A., Nworu, C. C., & Ndu, O. O. (2009). Gastro-protective effects of the leaf extract and fractions of *fleurya aestuans* L (urticaceae). *International Journal of Health Research*, 2(1).
- Alen, Y., Nakajima, S., Nitoda, T., Baba, N., Kanzaki, H., & Kawazu, K. (2000). Antinematodal activity of some tropical rainforest plants against the pinewood nematode *Bursaphelenchus xylophilus*. *Zeitschrift Fur Naturforschung - Section C Journal of Biosciences*, 55(3-4), 295-299.
- Ali, D. M. H., Keng, C. W., & Boey, P. L. (2010). *Triterpenoids, glycolipids and flavonoids of Melastoma malabathricum: Isolation, spectrometric characterization and antibacterial activity*. VDM Publishing.
- Allen, A. (1981). Structure and function of gastrointestinal mucus. *Physiology of the gastrointestinal tract*, 1, 617-639.
- Allen, A., & Garner, A. (1980). Mucus and bicarbonate secretion in the stomach and their possible role in mucosal protection. *Gut*, 21(3), 249.
- Amarowicz, R., Pegg, R. B., Rahimi-Moghaddam, P., Barl, B., & Weil, J. A. (2004). Free-radical scavenging capacity and antioxidant activity of selected plant species from the Canadian prairies. *Food chemistry*, 84(4), 551-562.
- Andersen, J., Nilsson, C., de Richelieu, T., Fridriksdottir, H., Gobilick, J., Mertz, O., & Gausset, Q. (2003). Local use of forest products in Kuyongon, Sabah, Malaysia. *ASEAN Review of Biodiversity and Environmental Conservation (ARBEC)*, 1-18.
- Ardeman, S., Chanarin, I., & Doyle, J. C. (1964). Studies on secretion of gastric intrinsic factor in man. *British medical journal*, 2(5409), 600.

- Armengaud, P., Sulpice, R., Miller, A. J., Stitt, M., Amtmann, A., & Gibon, Y. (2009). Multilevel analysis of primary metabolism provides new insights into the role of potassium nutrition for glycolysis and nitrogen assimilation in Arabidopsis roots. *Plant Physiology*, 150(2), 772-785.
- Ayesha, S., Premakumari, K. B., & Roukiya, S. (2010). Antioxidant activity and estimation of total phenolic content of *Muntingia calabura* by colorimetry. *International Journal of ChemTech Research*, 2(1), 205-208.
- Babu, P. V. A., Liu, D., & Gilbert, E. R. (2013). Recent advances in understanding the anti-diabetic actions of dietary flavonoids. *The Journal of nutritional biochemistry*, 24(11), 1777-1789.
- Bafna, P. A., & Balaraman, R. (2004). Anti-ulcer and antioxidant activity of DHC-1, a herbal formulation. *Journal of ethnopharmacology*, 90(1), 123-127.
- Balan, T., Sani, M. H. M., Ahmad, S. H. M., Suppaiah, V., Mohtarrudin, N., & Zakaria, Z. A. (2015). Antioxidant and anti-inflammatory activities contribute to the prophylactic effect of semi-purified fractions obtained from the crude methanol extract of *Muntingia calabura* leaves against gastric ulceration in rats. *Journal of ethnopharmacology*, 164, 1-15.
- Balan, T., Mohd. Sani, M. H., Suppaiah, V., Mohtarrudin, N., Suhaili, Z., Ahmad, Z., & Zakaria, Z. A. (2014). Antiulcer activity of *Muntingia calabura* leaves involves the modulation of endogenous nitric oxide and nonprotein sulfhydryl compounds. *Pharmaceutical biology*, 52(4), 410-418.
- Bandyopadhyay, D., Bandyopadhyay, A., Das, P. K., & Reiter, R. J. (2002). Melatonin protects against gastric ulceration and increases the efficacy of ranitidine and omeprazole in reducing gastric damage. *Journal of pineal research*, 33(1), 1-7.
- Bandyopadhyay, D., Biswas, K., Reiter, R. J., & Banerjee, R. K. (2001). Gastric toxicity and mucosal ulceration induced by oxygen-derived reactive species: protection by melatonin. *Current molecular medicine*, 1(4), 501-513.
- Batista, L. M., Almeida, A. B. A., Morais Lima, G. R., Sousa Falcão, H., Pietro Magri, L., Luiz-Ferreira, A., ... & Monteiro, A. R. (2014). Gastroprotective effects (in rodents) of a flavonoid rich fraction obtained from *Syngonanthus macrolepis*. *Journal of Pharmacy and Pharmacology*, 66(3), 445-452.
- Beil, W., Birkholz, C., & Sewing, K. F. (1995). Effects of flavonoids on parietal cell acid secretion, gastric mucosal prostaglandin production and *Helicobacter pylori* growth. *Arzneimittel-forschung*, 45(6), 697-700.
- Berthoud, H. R. (2008). The vagus nerve, food intake and obesity. *Regulatory peptides*, 149(1), 15-25.
- Beserra, A. M. S. E. S., Calegari, P. I., Souza, M. D. C., dos Santos, R. A. N., Lima, J. C. D. S., Silva, R. M., ... & Martins, D. T. D. O. (2011). Gastroprotective and ulcer-healing mechanisms of ellagic acid in experimental rats. *Journal of Agricultural and Food Chemistry*, 59(13), 6957-6965.

- Bighetti, A. E., Antonio, M. A., Kohn, L. K., Rehder, V. L. G., Foglio, M. A., Possenti, A., ... & Carvalho, J. E. (2005). Antiulcerogenic activity of a crude hydroalcoholic extract and coumarin isolated from *Mikania laevigata* Schultz Bip. *Phytomedicine*, 12(1), 72-77.
- Blix, H. S., Viktil, K. K., Moger, T. A., & Reikvam, A. (2010). Drugs with narrow therapeutic index as indicators in the risk management of hospitalised patients. *Pharmacy practice*, 8(1), 50-55.
- Bodhankar, S. L., Jain, B. B., Ahire, B. P., Daude, R. B., & Shitole, P. P. (2006). The effect of rabeprazole and its isomers on aspirin and histamine-induced ulcers in rats. *Indian journal of pharmacology*, 38(5), 357.
- Bogdan, C., Röllinghoff, M., & Diefenbach, A. (2000). The role of nitric oxide in innate immunity. *Immunological reviews*, 173(1), 17-26.
- Boligon, A. A., de Freitas, R. B., de Brum, T. F., Waczuk, E. P., Klimaczewski, C. V., de Ávila, D. S., ... & de Freitas Bauermann, L. (2014). Antiulcerogenic activity of *Scutia buxifolia* on gastric ulcers induced by ethanol in rats. *Acta Pharmaceutica Sinica B*, 4(5), 358-367.
- Borrelli, F., & Izzo, A. A. (2000). The plant kingdom as a source of anti-ulcer remedies. *Phytotherapy Research*, 14(8), 581-591.
- Bors, W., & Michel, C. (2002). Chemistry of the antioxidant effect of polyphenols. *Annals of the New York Academy of Sciences*, 957(1), 57-69.
- Burkill, I. H. (1966). A dictionary of the economic products of the Malay Peninsula. A *Dictionary of the Economic Products of the Malay Peninsula.*, 2(2nd edition).
- Calam, J., & Baron, J. H. (2001). Pathophysiology of duodenal and gastric ulcer and gastric cancer. *British Medical Journal*, 323(7319), 980-983.
- Chen, H., Liao, H., Liu, Y., Zheng, Y., Wu, X., Su, Z., Zhang, X., Lai, Z., Lai, X., Lin, Z.X. and Su, Z. (2015). Protective effects of pogostone from *Pogostemonis Herba* against ethanol-induced gastric ulcer in rats. *Fitoterapia* 100: 110-117.
- Chen, J. J., Lee, H. H., Shih, C. D., Liao, C. H., Chen, I. S., & Chou, T. H. (2007). New dihydrochalcones and anti-platelet aggregation constituents from the leaves of *Muntingia calabura*. *Planta medica*, 73(06), 572-577.
- Chen, J. J., Lee, H. H., Duh, C. Y., & Chen, I. S. (2005). Cytotoxic chalcones and flavonoids from the leaves of *Muntingia calabura*. *Planta medica*, 71(10), 970-973.
- Chen, J. J., Lin, R. W., Duh, C. Y., Huang, H. Y., & Chen, I. S. (2004). Flavones and cytotoxic constituents from the stem bark of *Muntingia calabura*. *Journal of the Chinese Chemical Society*, 51(3), 665-670.
- Chey, W. Y. (1993). Hormonal control of pancreatic exocrine secretion. *The pancreas: biology, pathobiology and disease*, 403-424.
- Chin, W.Y. (1989). A guide to the wayside trees of Singapore. *Singapore: Singapore Science Centre 160p.-col. illus.. ISBN, 1220074454.*

- Comalada, M., Ballester, I., Bailón, E., Sierra, S., Xaus, J., Gálvez, J., ... & Zarzuelo, A. (2006). Inhibition of pro-inflammatory markers in primary bone marrow-derived mouse macrophages by naturally occurring flavonoids: analysis of the structure–activity relationship. *Biochemical pharmacology*, 72(8), 1010-1021.
- Corne, S. J. (1974). Proceedings: A method for the quantitative estimation of gastric barrier mucus. *The Journal of physiology*, 242(2), 116-179.
- Coskun, Ö., Kanter, M., Armutçu, F., Çetin, K., Kaybolmaz, B., & Yazgan, Ö. (2004). Protective effects of quercetin, a flavonoid antioxidant, in absolute ethanol-induced acute gastric ulcer. *European Journal of General Medicine*, 1(3), 37-42.
- Croteau, R., Kutchan, T. M., & Lewis, N. G. (2000). Natural products (secondary metabolites). *Biochemistry and molecular biology of plants*, 24, 1250-1319.
- Das, K. K., & Kotoky, J. (1988). A New Aliphatic Constituent of *Melastoma Malabathricum* Linn. *Journal of the Indian Chemical Society*, 65(5), 385-386.
- Debbab, A., Aly, A. H., Lin, W. H., & Proksch, P. (2010). Bioactive compounds from marine bacteria and fungi. *Microbial biotechnology*, 3(5), 544-563.
- de-Faria, F.M., Almeida, A.C.A., ALuiz-Ferreira, A., Takayama, C., Dunder, R.J., da Silva, M.A., Salvador, M.J., Abdelnur, P.V., Eberlin, M.N., Vilegas, W., Toma, W. and Souza-Brito, A.R.M. (2012). Antioxidant action of mangrove polyphenols against gastric damage induced by absolute ethanol and ischemia-reperfusion in the rat. *The Scientific World Journal* 2012: 1-9.
- De Lira Mota, K. S., Dias, G. E. N., Pinto, M. E. F., Luiz-Ferreira, Â., Souza-Brito, A. R. M., Hiruma-Lima, C. A., ... Batista, L. M. (2009). Flavonoids with gastroprotective activity. *Molecules*, 14, 979–1012.
- Demir, S., Yilmaz, M., Koseoglu, M., Akalin, N., Aslan, D., & Aydin, A. (2003). Role of free radicals in peptic ulcer and gastritis. *Turkish Journal of Gastroenterology*, 14(1), 39-43.
- Dorđević, S., Petrović, S., Dobrić, S., Milenković, M., Vučićević, D., Žižić, S., & Kukić, J. (2007). Antimicrobial, anti-inflammatory, anti-ulcer and antioxidant activities of *Carlina acanthifolia* root essential oil. *Journal of ethnopharmacology*, 109(3), 458-463.
- Eglen, R. M. (2001). Muscarinic receptors and gastrointestinal tract smooth muscle function. *Life sciences*, 68(22-23), 2573-2578.
- El-Missiry, M. A., El-Sayed, I. H., & Othman, A. I. (2001). Protection by metal complexes with SOD-mimetic activity against oxidative gastric injury induced by indometacin and ethanol in rats. *Annals of clinical biochemistry*, 38(6), 694-700.
- Fabricant, D. S., & Farnsworth, N. R. (2001). The value of plants used in traditional medicine for drug discovery. *Environmental health perspectives*, 109(Suppl 1), 69.

- Faravani, M. (2008). *The population biology of Straits Rhododendron (Melastoma malabathricum L.)* (Doctoral dissertation, University Malaya).
- Farnsworth, N. R. (1994). Ethnopharmacology and drug development. *Ethnobotany and the search for new drugs*, 185, 42-51.
- Farnsworth, N. R. (1990). The role of ethnopharmacology in drug development. In *Ciba Foundation Symposium 154-Bioactive Compounds from Plants* (pp. 2-21). John Wiley & Sons, Ltd.
- Fenech, M. (2001). The role of folic acid and Vitamin B12 in genomic stability of human cells. *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, 475(1), 57-67.
- Furness, J. B., & Costa, M. (1987). *The enteric nervous system* (pp. 65-71). Edinburgh etc.: Churchill Livingstone.
- Ganong, W. F., & Ganong, W. (1995). *Review of medical physiology* (p. 59). Norwalk, CT: Appleton & Lange.)
- Garner, A., Flemström, G., Allen, A., Heylings, J. R., & McQueen, S. (1984). Gastric mucosal protective mechanisms: roles of epithelial bicarbonate and mucus secretions. *Scandinavian journal of gastroenterology. Supplement*, 101, 79-86.
- Gershman, G. (2011). Diagnostic upper gastrointestinal endoscopy. *Practical Pediatric Gastrointestinal Endoscopy*, 41.
- Graham, A. H. (2009). Effects of Prednisone or Prednisone with Ultralow-Dose Aspirin on the Gastroduodenal Mucosa of Healthy Dogs (Doctoral dissertation, Virginia Tech).
- Gray, H., & Lewis, W. H. (1942). *Anatomy of the human body* (Vol. 1970, pp. 993-1007). Philadelphia: Lea & Febiger.
- Grossman, M. I. (1970). Gastrin and its activities. *Nature*, 228(5277), 1147-1150.
- Grosvenor, P. W., Supriono, A., & Gray, D. O. (1995). Medicinal plants from Riau Province, Sumatra, Indonesia. Part 2: antibacterial and antifungal activity. *Journal of ethnopharmacology*, 45(2), 97-111.
- Gülçin, I., Küfrevioğlu, O., Oktay, M., Büyükkökuroğlu, M.E., (2004). Antioxidant, antimicrobial, antiulcer and analgesic activities of nettle (*Urtica dioica* L.). *Journal of Ethnopharmacology* 90, 205-215.
- Gyamfi, M. A., & Aniya, Y. (2002). Antioxidant properties of Thonningianin A, isolated from the African medicinal herb, *Thonningia sanguinea*. *Biochemical pharmacology*, 63(9), 1725-1737.
- Hajrezaie, M., Salehen, N., Karimian, H., Zahedifard, M., Shams, K., Al Batran, R., ... & Abdulla, M. A. (2015). Biochanin a gastroprotective effects in ethanol-induced gastric mucosal ulceration in rats. *PloS one*, 10(3), e0121529.
- Håkanson, R., Böttcher, G., Ekblad, E., Panula, P., Simonsson, M., Dohlsten, M., ... & Sundler, F. (1986). Histamine in endocrine cells in the stomach. *Histochemistry and Cell Biology*, 86(1), 5-17.

- Hamiza, O. O., Rehman, M. U., Tahir, M., Khan, R., Khan, A. Q., Lateef, A., ... & Sultana, S. (2012). Amelioration of 1, 2 Dimethylhydrazine (DMH) induced colon oxidative stress, inflammation and tumor promotion response by tannic acid in Wistar rats. *Asian Pacific Journal of Cancer Prevention*, 13(9), 4393-4402.
- Heo, S. J., Yoon, W. J., Kim, K. N., Ahn, G. N., Kang, S. M., Kang, D. H., ... & Jeon, Y. J. (2010). Evaluation of anti-inflammatory effect of fucoxanthin isolated from brown algae in lipopolysaccharide-stimulated RAW 264.7 macrophages. *Food and chemical toxicology*, 48(8), 2045-2051.
- Happé, R. P., & Rothuizen, J. (1995). Digestive tract. In *Medical History and Physical Examination in Companion Animals* (pp. 109-126). Springer Netherlands.
- Herrmann, R., Shaw, R. G., & Fone, D. J. (1990). Ranitidine-associated recurrent acute pancreatitis. *Internal Medicine Journal*, 20(3), 243-244.
- Herriott, R. M. (1962). Pepsinogen and pepsin. *The Journal of general physiology*, 45(4), 57-76.
- Hersey, S. J., & Sachs, G. (1995). Gastric acid secretion. *Physiological Reviews*, 75(1), 155-190.
- Hu, X. T., Ding, C., Zhou, N., & Xu, C. (2015). Quercetin protects gastric epithelial cell from oxidative damage in vitro and in vivo. *European journal of pharmacology*, 754, 115-124.
- Huh, C. H., Bhutani, M. S., Farfan, E. B., & Bolch, W. E. (2003). Individual variations in mucosa and total wall thickness in the stomach and rectum assessed via endoscopic ultrasound. *Physiological measurement*, 24(4), N15.
- Huang, D., Ou, B., Hampsch-Woodill, M., Flanagan, J. A., & Prior, R. L. (2002). High-throughput assay of oxygen radical absorbance capacity (ORAC) using a multichannel liquid handling system coupled with a microplate fluorescence reader in 96-well format. *Journal of agricultural and food chemistry*, 50(16), 4437-4444.
- Hussain, F., Abdulla, M. A., Noor, S. M., Ismail, S., & Ali, H. M. (2008). Gastroprotective effects of *Melastoma malabathricum* aqueous leaf extract against ethanol-induced gastric ulcer in rats. *American Journal of Biochemistry and Biotechnology*, 4(4), 438-441.
- Hussein, S.A., El-Senosy, Y.A. and Hassan, M.F. (2014). Gastroprotective, antiapoptotic and anti-inflammatory effect of alpha-lipoic acid on ethanol induced gastric mucosal lesions in rats. *American Journal of Biochemistry and Molecular Biology* 4: 48-63.
- Ibrahim, I. A. A., Abdulla, M. A., Abdelwahab, S. I., Al-Bayat, F., & Majid, N. (2012). Leaves extract of *Muntingia calabura* protects against gastric ulcer induced by ethanol in Sprague–Dawley rats. *Clin Exp Pharmacol.[Epub ahead of print]* S, 5, 2161-1459.

- Ismail Suhaimy, N. W., Noor Azmi, A. K., Mohtarrudin, N., Omar, M. H., Tohid, S. F. M., Cheema, M. S., ... & Zakaria, Z. A. (2017). Semipurified Ethyl Acetate Partition of Methanolic Extract of *Melastoma malabathricum* Leaves Exerts Gastroprotective Activity Partly via Its Antioxidant-Antisecretory-Anti-Inflammatory Action and Synergistic Action of Several Flavonoid-Based Compounds. *Oxidative medicine and cellular longevity*, 2017.
- Indu BJ, Razal AR. (1998). Medicinal uses of Malaysian herbs. *Flavour Magazine*. [Online]. 6(2). Available: [http:// www.foodsmart.biz/herb.php](http://www.foodsmart.biz/herb.php). Accessed Nov 2007 10.
- Institute for Health Metrics and Evaluation (2013).
- Jaganath, I. B., & Ng, L. T. (2000). Herbs. *The Green Pharmacy of Malaysia*. Kuala Lumpur, Vinpress and Malaysia Agricultural Research and Development Institute, 95-99.
- Jain, K. S., Shah, A. K., Bariwal, J., Shelke, S. M., Kale, A. P., Jagtap, J. R., & Bhosale, A. V. (2007). Recent advances in proton pump inhibitors and management of acid-peptic disorders. *Bioorganic & medicinal chemistry*, 15(3), 1181-1205.
- Jain, S. K., & DeFilipps, R. A. (1991). *Medicinal plants of India*. Reference Publications.
- Jaios, E. S., Rahman, S. A., Ching, S. M., Kadir, A. A., Desa, M., Mohd, N., & Zakaria, Z. A. (2016). Possible mechanisms of antinociception of methanol extract of *Melastoma malabathricum* leaves. *Revista Brasileira de Farmacognosia*, 26(5), 586-594.
- Jantan, I., Rafi, I. A. A., & Jalil, J. (2005). Platelet-activating factor (PAF) receptor-binding antagonist activity of Malaysian medicinal plants. *Phytomedicine*, 12(1-2), 88-92.
- Jayakumari, S., Anbu, J., Ravichandiran, V., Anjana, A. S. H. W. I. N. I., Kumar, G. S., & Maharaj, S. (2012). Antiulcerogenic and free radical scavenging activity of flavonoid fraction of *Psidium Guajavalinn*. leaves. *Int J Pharm Pharm Sci*, 4(1), 170-4.
- Joffry, S. M., Yob, N. J., Rofiee, M. S., Affandi, M. M. R., Suhaili, Z., Othman, F., ... & Zakaria, Z. A. (2011). *Melastoma malabathricum* (L.) smith ethnomedicinal uses, chemical constituents, and pharmacological properties: a review. *Evidence-Based Complementary and Alternative Medicine*, 2012.
- Johnson, W. S., & Miller, D. R. (1988). Ranitidine and bradycardia. *Annals of internal medicine*, 108(3), 493-493.
- Johnny, L., Yusuf, U. K., & Nulit, R. (2011). Antifungal activity of selected plant leaves crude extracts against a pepper anthracnose fungus, *Colletotrichum capsici* (Sydow) butler and bisby (Ascomycota: Phyllachorales). *African Journal of Biotechnology*, 10(20), 4157-4165.
- Jordan, P. H., & Yip, B. S. (1972). The presence of gastrin in fasting and stimulated gastric juice of man. *Surgery*, 72(3), 352-356.



- Jung, H. W., Seo, U. K., Kim, J. H., Leem, K. H., & Park, Y. K. (2009). Flower extract of *Panax notoginseng* attenuates lipopolysaccharide-induced inflammatory response via blocking of NF- $\kappa$ B signaling pathway in murine macrophages. *Journal of Ethnopharmacology*, 122(2), 313-319.
- Kahl, R., & Kappus, H. (1993). Toxicology of the synthetic antioxidants BHA and BHT in comparison with the natural antioxidant vitamin E. *Zeitschrift für Lebensmittel-untersuchung und-forschung*, 196(4), 329-338.
- Kahraman, A., Çakar, H., & Köken, T. (2012). The protective effect of quercetin on long-term alcohol consumption-induced oxidative stress. *Molecular biology reports*, 39(3), 2789-2794.
- Kaneda, N., Pezzuto, J. M., Soejarto, D. D., Kinghorn, A. D., Farnsworth, N. R., Tuchinda, P., ... & Reutrakul, V. (1991). New cytotoxic flavonoids from *Muntingia calabura* roots. *Planta Medica*, 56(06), 672-673.
- Kapp Jr, R. W. (2007). Gastrointestinal Tract as Major Route of Pharmaceutical Administration. *Toxicology of the Gastrointestinal Tract* SC Gad, 107-133.
- Kararli, T. T. (1995). Comparison of the gastrointestinal anatomy, physiology, and biochemistry of humans and commonly used laboratory animals. *Biopharmaceutics & drug disposition*, 16(5), 351-380.
- Karhunen, L. J., Lappalainen, R. I., Tammela, L., Turpeinen, A. K., & Uusitupa, M. I. (1997). Subjective and physiological cephalic phase responses to food in obese binge-eating women. *International Journal of Eating Disorders*, 21(4), 321-328.
- Karthyaini & Suresh K. (2012). Pharmacognostic evaluation, in vitro antioxidant and in vivo anti-inflammatory studies of *Muntingia calabura* Linn. *J Global Trends Pharm Sci* 3:805–11.
- Keeton, R. W., & Koch, F. C. (1915). The distribution of gastrin in the body. *American Journal of Physiology--Legacy Content*, 37(3), 481-504.
- Kim, H. G., Shrestha, B., Lim, S. Y., Yoon, D. H., Chang, W. C., Shin, D. J., ... & Sung, J. M. (2006). Cordycepin inhibits lipopolysaccharide-induced inflammation by the suppression of NF- $\kappa$ B through Akt and p38 inhibition in RAW 264.7 macrophage cells. *European journal of pharmacology*, 545(2), 192-199.
- Koay, S. S. (2008). Establishment Of Cell Suspension Culture Of *Melastoma Malabathricum* L. For The Production Of Anthocyanin (Doctoral dissertation, Universiti Sains Malaysia).
- König, G. M., Kehraus, S., Seibert, S. F., Abdel-Lateff, A., & Müller, D. (2006). Natural products from marine organisms and their associated microbes. *ChemBioChem*, 7(2), 229-238.
- Konovalova, O., Gergel, E., & Herhel, V. (2013). GC-MS analysis of bioactive components of *Shepherdia argentea* (Pursh.) Nutt. from Ukrainian Flora. *The Pharma Innovation*, 2(6).

- Konturek, P. C., Brzozowski, T., & Konturek, S. J. (2011). Stress and the gut: pathophysiology, clinical consequences, diagnostic approach and treatment options. *J Physiol Pharmacol*, 62(6), 591-9.
- Kossel A. (1891). Archives of Analytical Physiology, *Physiol Abteilung*, 181–186.
- Kowluru, R. A., & Chan, P. S. (2007). Oxidative stress and diabetic retinopathy. *Journal of Diabetes Research*, 2007.
- Kumar, V., Ahmed, D., Gupta, P. S., Anwar, F., & Mujeeb, M. (2013). Anti-diabetic, anti-oxidant and anti-hyperlipidemic activities of *Melastoma malabathricum* Linn. leaves in streptozotocin induced diabetic rats. *BMC complementary and Alternative Medicine*, 13(1), 222.
- Kumar, A., Singh, V., & Chaudhary, A. K. (2011). Gastric antisecretory and antiulcer activities of *Cedrus deodara* (Roxb.) Loud. in Wistar rats. *Journal of ethnopharmacology*, 134(2), 294-297.
- Kunze, W. A. A., & Furness, J. B. (1999). The enteric nervous system and regulation of intestinal motility. *Annual review of physiology*, 61(1), 117-142.
- Kupiec, T. (2004). Quality-control analytical methods: High-performance liquid chromatography. *International journal of pharmaceutical compounding*, 8, 223-227.
- Lahlou, M. (2013). The success of natural products in drug discovery. *Pharmacology & Pharmacy*, 4(3A), 17-31.
- Lai, L. S., Chou, S. T., & Chao, W. W. (2001). Studies on the antioxidative activities of *Hsian-tsao* (*Mesona procumbens* Hemsl) leaf gum. *Journal of Agricultural and Food Chemistry*, 49(2), 963-968.
- Lakhanpal, P., & Rai, D. K. (2007). Quercetin: a versatile flavonoid. *Internet Journal of Medical Update*, 2(2), 22-37.
- Latiff, A., & Zakri, A. H. (2000). Protection of traditional knowledge, innovations and practices: The Malaysian experience. In *the UNCTAD Expert Meeting on Systems and National Experiences for Protecting Traditional Knowledge, Innovations and Practices*.
- Lau, J. Y., Sung, J., Hill, C., Henderson, C., Howden, C. W., & Metz, D. C. (2011). Systematic review of the epidemiology of complicated peptic ulcer disease: incidence, recurrence, risk factors and mortality. *Digestion*, 84(2), 102-113.
- Lee, J. Y., & Park, W. (2011). Anti-inflammatory effect of myristicin on RAW 264.7 macrophages stimulated with polyinosinic-polycytidylic acid. *Molecules*, 16(8), 7132-7142.
- Leyck, S., & Parnham, M. J. (1990). Acute antiinflammatory and gastric effects of the seleno-organic compound ebselen. *Inflammation Research*, 30(3), 426-431.
- Li, H. L., Chen, H. L., Li, H., Zhang, K. L., Chen, X. Y., Wang, X. W., ... & Liu, J. (2005). Regulatory effects of emodin on NF- $\kappa$ B activation and inflammatory cytokine expression in RAW 264.7 macrophages. *International journal of molecular medicine*, 16(1), 41-47.

- Lieberman, A. P., Pitha, P. M., Shin, H. S., & Shin, M. L. (1989). Production of tumor necrosis factor and other cytokines by astrocytes stimulated with lipopolysaccharide or a neurotropic virus. *Proceedings of the National Academy of Sciences*, 86(16), 6348-6352.
- Lima, Z. P., Severi, J. A., Pellizzon, C. H., Brito, A. R. M. S., Solis, P. N., Cáceres, A., ... & Hiruma-Lima, C. A. (2006). Can the aqueous decoction of mango flowers be used as an antiulcer agent?. *Journal of Ethnopharmacology*, 106(1), 29-37.
- Lin, J. T., Chang, Y. Y., Chen, Y. C., Shen, B. Y., & Yang, D. J. (2017). Molecular mechanisms of the effects of the ethanolic extract of *Muntingia calabura* Linn. fruit on lipopolysaccharide-induced pro-inflammatory mediators in macrophages. *Food & Function*, 8(3), 1245-1253.
- Lisec, J., Schauer, N., Kopka, J., Willmitzer, L., & Fernie, A. R. (2015). Corrigendum: gas chromatography mass spectrometry-based metabolite profiling in plants. *Nat. Protoc*, 10(1457), 10-1038.
- Lohézic-Le Dévéhat, F., Bakhtiar, A., Bézivin, C., Amoros, M., & Boustie, J. (2002). Antiviral and cytotoxic activities of some Indonesian plants. *Fitoterapia*, 73(5), 400-405.
- Lowry, J. B. (1976). Anthocyanins of the Melastomataceae, Myrtaceae and some allied families. *Phytochemistry*, 15(4), 513-516.
- Lowry, J. B. (1968). The distribution and potential taxonomic value of alkylated ellagic acids. *Phytochemistry*, 7(10), 1803-1813.
- Mahmood, N. D., Nasir, N. L. M., Rofiee, M. S., Tohid, S. F. M., Ching, S. M., Teh, L. K., ... Zakaria, Z. a. (2014). *Muntingia calabura*: A review of its traditional uses, chemical properties, and pharmacological observations. *Pharmaceutical Biology*, 0(0), 1-26.
- Maji, S., Dandapat, P., Ojha, D., Maity, C., Halder, S. K., Mohapatra, P. D., ... & Mondal, K. C. (2010). In vitro antimicrobial potentialities of different solvent extracts of ethnomedicinal plants against clinically isolated human pathogens. *Journal of Phytology*, 2(4).
- Malek, N. H. A., Baek, S. H., & Asari, A. (2003). Chemical components of *Melastoma malabathricum*. *ACGC Chemical Research Communications*, 16, 28-33.
- Marks, I. N., & Shay, H. (1959). Observations on the pathogenesis of gastric ulcer. *The Lancet*, 273(7083), 1107-1110.
- Martins, J.L., Rodrigues, O.R., da Silva, D.M., Galdino, P.M., de Paula, J.R., Romão, W., da Costa, H.B., Vaz, B.G., Ghedini, P.C. and Costa, E.A. (2014). Mechanisms involved in the gastroprotective activity of *Celtis iguanaea* (Jacq.) Sargent on gastric lesions in mice. *Journal of Ethnopharmacology* 155: 1616-1624.
- Martin, M. J., La-Casa, C., Alarcon-de-La-Lastra, C., Cabeza, J., Villegas, I., & Motilva, V. (1998). Anti-oxidant mechanisms involved in gastroprotective effects of quercetin. *Zeitschrift für Naturforschung C*, 53(1-2), 82-88.

- Martin, M. J., Marhuenda, E., Perez-Guerrero, C., & Franco, J. M. (1994). Antiulcer effect of naringin on gastric lesions induced by ethanol in rats. *Pharmacology*, 49(3), 144-150.
- Mamat, S. S., Kamarolzaman, M. F. F., Yahya, F., Mahmood, N. D., Shahril, M. S., Jakius, K. F., ... & Zakaria, Z. A. (2013). Methanol extract of *Melastoma malabathricum* leaves exerted antioxidant and liver protective activity in rats. *BMC complementary and alternative medicine*, 13(1), 326.
- Manicam, C., Abdullah, J. O., Tohit, E. R. M., Seman, Z., Chin, S. C., & Hamid, M. (2013). In vitro anticoagulant activities of *Melastoma malabathricum* Linn. aqueous leaf extract: A preliminary novel finding. *Journal of Medicinal Plants Research*, 4(14), 1464-1472.
- Manzoor-I-Khuda, M., Chowdhury, S. A., Reza, T., & Chowdhury, A. K. (1981). Chemical Investigation on *Melastoma Malabathricum*. Part 1: Isolation of Melastomic Acid and Betasitosterol from the Roots. *Journal of the Bangladesh Academy of Sciences*, vol. 5, pp. 55-59.
- Masih, N. G., & Singh, B. S. (2012). Phytochemical Screening of Some Plants Used in Herbal Based Cosmetic Preparations. *Chemistry of Phytopotentials: Health, Energy and Environmental Perspectives*, 111.
- Mazura, M. P., Susanti, D., & Rasadah, M. A. (2007). Anti-inflammatory Action of Components from *Melastoma malabathricum*. *Pharmaceutical Biology*, 45(5), 372-375.
- Meyer, R. F., Essenburg, A. D., Smith, R. D., & Kaplan, H. R. (1982). Angiotensin converting enzyme inhibitors: modifications of a tripeptide analogue. *Journal of medicinal chemistry*, 25(8), 996-999.
- Mohandoss, S., & Ravindran, P. (1993). Flavonoids from *Melastoma malabathricum*. *Fitoterapia*, 64, 277-278.
- Moncada, S. R. M. J., Palmer, R. M. L., & Higgs, E. (1991). Nitric oxide: physiology, pathophysiology, and pharmacology. *Pharmacological reviews*, 43(2), 109-142.
- Mordan, L. J., Burnett, T. S., Zhang, L. X., Tom, J., & Cooney, R. V. (1993). Inhibitors of endogenous nitrogen oxide formation block the promotion of neoplastic transformation in C3H 10T1/2 fibroblasts. *Carcinogenesis*, 14(8), 1555-1559.
- Morton, H. (2005). Democracy, Self-Reviews and the 1855 Leaves of Grass. *Virginia Quarterly Review*, 81, 229-243.
- Mota, K. S., Dias, G. E. N., Pinto, M. E. F., Luiz-Ferreira, Â., Monteiro Souza-Brito, A. R., Hiruma-Lima, C. A., ... & Batista, L. M. (2009). Flavonoids with gastroprotective activity. *Molecules*, 14(3), 979-1012.
- Murakami, S., Isobe, Y., Kijima, H., Nagai, H., Muramatu, M., & Otomo, S. (1991). Inhibition of gastric H<sup>+</sup>, K<sup>+</sup>-ATPase and acid secretion by ellagic acid. *Planta medica*, 57(04), 305-308.

- Naik, Y., Jayaram, S., Nayaka, M. H., & Dharmesh, S. M. (2007). Gastroprotective effect of swallow root (*Decalepis hamiltonii*) extract: Possible involvement of H<sup>+</sup>-K<sup>+</sup>-ATPase inhibition and antioxidative mechanism. *Journal of ethnopharmacology*, 112(1), 173-179.
- Navarrete, A., Trejo-Miranda, J. L., & Reyes-Trejo, L. (2002). Principles of root bark of *Hippocratea excelsa* (Hippocrataceae) with gastroprotective activity. *Journal of ethnopharmacology*, 79(3), 383-388.
- Nazlina, I., Norha, S., Noor Zarina, A. W., & Ahmad, I. B. (2008). Cytotoxicity and antiviral activity of *Melastoma malabathricum* extracts. *Malays J Appl Biol*, 37, 53-55.
- Nshimo, C. M., Pezzuto, J. M., Kinghorn, A. D., & Farnsworth, N. R. (1993). Cytotoxic constituents of *Muntingia calabura* leaves and stems collected in Thailand. *International journal of pharmacognosy*, 31(1), 77-81.
- Nijveldt, R. J., Van Nood, E. L. S., Van Hoorn, D. E., Boelens, P. G., Van Norren, K., & Van Leeuwen, P. A. (2001). Flavonoids: a review of probable mechanisms of action and potential applications. *The American journal of clinical nutrition*, 74(4), 418-425.
- Nivethetha, M. (2009). Cardioprotective effect of *Muntingia calabura* L \_ a traditional drug source.
- Nordin, N., Salama, S. M., Golbabapour, S., Hajrezaie, M., Hassandarvish, P., Kamalidehghan, B., ... & Karimian, H. (2014). Anti-ulcerogenic effect of methanolic extracts from *Enicosanthellum pulchrum* (King) Heusden against ethanol-induced acute gastric lesion in animal models. *PloS one*, 9(11), e111925.
- Oates, P. J., & Hakkinen, J. P. (1988). Studies on the mechanism of ethanol-induced gastric damage in rats. *Gastroenterology*, 94(1), 10-21.
- Ohshima, H., & Bartsch, H. (1994). Chronic infections and inflammatory processes as cancer risk factors: possible role of nitric oxide in carcinogenesis. *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, 305(2), 253-264.
- Okado-Matsumoto, A. and Fridovich, I., (2001). Subcellular distribution of superoxide dismutases (SOD) in rat liver: Cu,Zn-SOD in mitochondria. *The Journal of Biological Chemistry* 276: 38388-38393.
- Owen, D. A. (1986). Normal histology of the stomach. *The American journal of surgical pathology*, 10(1), 48-61.
- Panda, V.S. and Khambat, P.D. (2014). Antiulcer activity of *Garcinia indica* fruit rind (kokum berry) in rats. *Biomedicine & Aging Pathology* 4: 309-316.
- Patwardhan, B., Vaidya, A. D., & Chorghade, M. (2004). Ayurveda and natural products drug discovery. *Current Science Bangalore*, 86(6), 789-799.

- Perez Arbelaez, E. (1975). Plantas medicinales y venenosas de Colombia. *Medellin.: Hernando Salazar 295pp.. Local & botanical names, parts used, uses. Some small figs. No anatomy. Toxic plants Poisonous plants, Geog, Materia medica Pharmacognosy Drug plants Medicinal plants (PMBD, 185607344).*
- Pongprom, N., Khanapan, C., Tosert, P. (2003). Chemical constituents of *Melastoma malabathricum* Linn. *The 29th Congress on Science and Technology of Thailand, p.130.*
- Preethi, K., Premasudha, P., & Keerthana, K. (2012). Anti-inflammatory activity of *Muntingia calabura* fruits. *Pharmacognosy Journal*, 4(30), 51-56.
- Preethi, K., Vijayalakshmi, N., Shamna, R., & Sasikumar, J. M. (2010). In vitro antioxidant activity of extracts from fruits of *Muntingia calabura* Linn. from India. *Pharmacognosy Journal*, 2(14), 11-18.
- Rachchh, M. A., & Jain, S. M. (2008). Gastroprotective effect of *Benincasa hispida* fruit extract. *Indian journal of pharmacology*, 40(6), 271.
- Rahim, N.A., Hassandarvish, P., Golbabapour, S., Ismail, S., Tayyab, S. and Abdulla. M.A. (2014). Gastroprotective effect of ethanolic extract of *Curcuma xanthorrhiza* leaf against ethanol-induced gastric mucosal lesions in Sprague-dawley rats. *BioMed Research International* 2014: 416409.
- Rang, H. P., Ritter, J. M., Flower, R. J., & Henderson, G. (2014). *Rang & Dale's Pharmacology: With student consult online access.* Elsevier Health Sciences.
- Repetto, M. G., & Llesuy, S. F. (2002). Antioxidant properties of natural compounds used in popular medicine for gastric ulcers. *Brazilian journal of medical and biological research*, 35(5), 523-534.
- Rice-Evans, C. A., Miller, N. J., & Paganga, G. (1996). Structure-antioxidant activity relationships of flavonoids and phenolic acids. *Free radical biology and medicine*, 20(7), 933-956.
- Roberts, A. T., Martin, C. K., Liu, Z., Amen, R. J., Woltering, E. A., Rood, J. C., ... & Greenway, F. L. (2007). The safety and efficacy of a dietary herbal supplement and gallic acid for weight loss. *Journal of medicinal food*, 10(1), 184-188.
- Roessner, U., Wagner, C., Kopka, J., Trethewey, R. N., & Willmitzer, L. (2000). Simultaneous analysis of metabolites in potato tuber by gas chromatography–mass spectrometry. *The Plant Journal*, 23(1), 131-142.
- Rood, D. (2000). Gas chromatography problem solving and troubleshooting. *Journal of chromatographic science*, 38(10), 466-466.
- Sajem, A. L., & Gosai, K. (2006). Traditional use of medicinal plants by the Jaintia tribes in North Cachar Hills district of Assam, northeast India. *Journal of Ethnobiology and ethnomedicine*, 2(1), 33.
- Saladin, K. S., & Miller, L. (1998). *Anatomy & physiology.* New York (NY): WCB/McGraw-Hill.
- San Gabriel, A., & Uneyama, H. (2013). Amino acid sensing in the gastrointestinal tract. *Amino Acids*, 45(3), 451-461.

- Sani, M. H., Zakaria, Z. A., Balan, T., Teh, L. K., & Salleh, M. Z. (2012). Antinociceptive activity of methanol extract of *Muntingia calabura* leaves and the mechanisms of action involved. *Evidence-Based Complementary and Alternative Medicine*, 2012.
- Scheible, W. R., Morcuende, R., Czechowski, T., Fritz, C., Osuna, D., Palacios-Rojas, N., ... & Stitt, M. (2004). Genome-wide reprogramming of primary and secondary metabolism, protein synthesis, cellular growth processes, and the regulatory infrastructure of *Arabidopsis* in response to nitrogen. *Plant physiology*, 136(1), 2483-2499.
- Sen, S., Asokkumar, K., Umamaheswari, M., Sivashanmugam, A., & Subhadradevi, V. (2013). Antiulcerogenic effect of gallic Acid in rats and its effect on oxidant and antioxidant parameters in stomach tissue. *Indian journal of pharmaceutical sciences*, 75(2), 149.
- Sener, G., Paskaloglu, K., & Ayanoglu-Dülger, G. (2004). Protective effect of increasing doses of famotidine, omeprazole, lansoprazole, and melatonin against ethanol-induced gastric damage in rats. *Indian Journal of Pharmacology*, 36(3), 171.
- Sharma, H. K., Chhangte, L., & Dolui, A. K. (2001). Traditional medicinal plants in Mizoram, India. *Fitoterapia*, 72(2), 146-161.
- Shaw, S., Herbert, V., Colman, N., & Jayatilleke, E. (1990). Effect of ethanol-generated free radicals on gastric intrinsic factor and glutathione. *Alcohol*, 7(2), 153-157.
- Shay, H. (1945). A simple method for the uniform production of gastric ulceration in the rat. *Gastroenterology*, 5, 43-61.
- Shen, J., Xu, X., Cheng, F., Liu, H., Luo, X., Shen, J., ... & Jiang, H. (2003). Virtual screening on natural products for discovering active compounds and target information. *Current medicinal chemistry*, 10(21), 2327-2342.
- Shih, C. D. (2009). Activation of nitric oxide/cGMP/PKG signaling cascade mediates antihypertensive effects of *Muntingia calabura* in anesthetized spontaneously hypertensive rats. *The American journal of Chinese medicine*, 37(06), 1045-1058.
- Shih, C. D., Chen, J. J., & Lee, H. H. (2006). Activation of nitric oxide signaling pathway mediates hypotensive effect of *Muntingia calabura* L.(Tiliaceae) leaf extract. *The American journal of Chinese medicine*, 34(05), 857-872.
- Sibi, G., Naveen, R., Dhananjaya, K., Ravikumar, K. R., & Mallesha, H. (2012). Potential use of *Muntingia calabura* L. extracts against human and plant pathogens. *Pharmacognosy Journal*, 4(34), 44-47.
- Silva, L. P., de Angelis, C. D., Bonamin, F., Kushima, H., Mininel, F. J., dos Santos, L. C., ... & dos Santos Ramos, M. A. (2015). *Terminalia catappa* L.: A medicinal plant from the Caribbean pharmacopeia with anti-*Helicobacter pylori* and antiulcer action in experimental rodent models. *Journal of ethnopharmacology*, 159, 285-295.

- Silva, E. M., Souza, J. N. S., Rogez, H., Rees, J. F., & Larondelle, Y. (2007). Antioxidant activities and polyphenolic contents of fifteen selected plant species from the Amazonian region. *Food Chemistry*, 101(3), 1012-1018.
- Singh, R. (2008). Digestive system.
- Sirat, H. M., Susanti, D., Ahmad, F., Takayama, H., & Kitajima, M. (2010). Amides, triterpene and flavonoids from the leaves of *Melastoma malabathricum* L. *Journal of Natural Medicines*, 64, 492-495.
- Smith, J. L. (2003). The role of gastric acid in preventing foodborne disease and how bacteria overcome acid conditions. *Journal of food protection*, 66(7), 1292-1303.
- Sowndhararajan, K. and Chin, N.L. (2014). Antioxidant and anti-ulcer effects of ethyl acetate fraction of *Merremia tridentata* (L.) Hallier F. root. *Agriculture and Agricultural Science Procedia* 2: 406-414.
- Squier, C. A., & Kremer, M. J. (2001). Biology of oral mucosa and esophagus. *JNCI Monographs*, 2001(29), 7-15.
- Sridhar, M., Thirupathi, K., Chaitanya, G., Kumar, B. R., & Mohan, G. K. (2011). Antidiabetic effect of leaves of *Muntingia calabura* L., in normal and alloxan-induced diabetic rats. *Pharmacologyonline*, 2, 626-632.
- Srivastava, V., Viswanathaswamy, A. H. M., & Mohan, G. (2010). Determination of the antiulcer properties of sodium cromoglycate in pylorus-ligated albino rats. *Indian journal of pharmacology*, 42(3), 185.
- Stening, G. F., & Grossman, M. I. (1969). Gastrin-related peptides as stimulants of pancreatic and gastric secretion. *American Journal of Physiology--Legacy Content*, 217(1), 262-266.
- Stuehr, D. J., & Marletta, M. A. (1985). Mammalian nitrate biosynthesis: mouse macrophages produce nitrite and nitrate in response to *Escherichia coli* lipopolysaccharide. *Proceedings of the National Academy of Sciences*, 82(22), 7738-7742.
- Su, B. N., Park, E. J., Vigo, J. S., Graham, J. G., Cabieses, F., Fong, H. H., ... & Kinghorn, A. D. (2003). Activity-guided isolation of the chemical constituents of *Muntingia calabura* using a quinone reductase induction assay. *Phytochemistry*, 63(3), 335-341.
- Sufian, A. S., Ramasamy, K., Ahmat, N., Zakaria, Z. A., & Yusof, M. I. M. (2013). Isolation and identification of antibacterial and cytotoxic compounds from the leaves of *Muntingia calabura* L. *Journal of ethnopharmacology*, 146(1), 198-204.
- Sulaiman, Z. M. R., Somchit, Z. M. N., Thenamutha, M., & Kasthuri, D. (2006). The in vitro antibacterial activity of *Muntingia calabura* extracts. *International Journal of Pharmacology*, 2(4), 439-442.



- Sulaiman, M. R., Somchit, M. N., Israif, D. A., Ahmad, Z., & Moin, S. (2004). Antinociceptive effect of *Melastoma malabathricum* ethanolic extract in mice. *Fitoterapia*, 75(7), 667-672.
- Sung, J. J. Y., Kuipers, E. J., & El-Serag, H. B. (2009). Systematic review: the global incidence and prevalence of peptic ulcer disease. *Alimentary pharmacology & therapeutics*, 29(9), 938-946.
- Sunilson, J. A. J., An, K., Kumari, A. V. A. G., & Mohan, S. (2009). Antidiarrhoeal activity of leaves of *Melastoma malabathricum* linn. *Indian journal of pharmaceutical sciences*, 71(6), 691.
- Susanti, D., Sirat, H. M., Ahmad, F., & Ali, R. M. (2008). Bioactive constituents from the leaves of *Melastoma malabathricum* L. *Jurnal Ilmiah Farmasi* 5 (1): 1, 7.
- Susanti, D., Sirat, H. M., Ahmad, F., Ali, R. M., Aimi, N., & Kitajima, M. (2007). Antioxidant and cytotoxic flavonoids from the flowers of *Melastoma malabathricum* L. *Food Chemistry*, 103(3), 710-716.
- Taira, J., Nanbu, H., & Ueda, K. (2009). Nitric oxide-scavenging compounds in *Agrimonia pilosa* Ledeb on LPS-induced RAW264. 7 macrophages. *Food chemistry*, 115(4), 1221-1227.
- Takagi, K., Okabe, S., & Saziki, R. (1969). A new method for the production of chronic gastric ulcer in rats and the effect of several drugs on its healing. *The Japanese Journal of Pharmacology*, 19(3), 418-426.
- Takeuchi, K., Ueshima, K., Hironaka, Y., Fujioka, Y., Matsumoto, J., & Okabe, S. (1991). Oxygen free radicals and lipid peroxidation in the pathogenesis of gastric mucosal lesions induced by indomethacin in rats. *Digestion*, 49(3), 175-184.
- Thatoi, H. N., Panda, S. K., Rath, S. K., & Dutta, S. K. (2008). Antimicrobial activity and bOrissa. *Asian Journal of plant sciences*.
- Tietze, L. F., Bell, H. P., & Chandrasekhar, S. (2003). Natural product hybrids as new leads for drug discovery. *Angewandte Chemie International Edition*, 42(34), 3996-4028.
- Tripathi, K. D. (2013). *Essentials of medical pharmacology*. JP Medical Ltd.
- Trivedi, N. P., & Rawal, U. M. (2001). Hepatoprotective and antioxidant property of *Andrographis paniculata* (Nees) in BHC induced liver damage in mice. *Indian Journal of Experimental Biology*, 39(1), 41-46.
- Tortora, G. J., & Derrickson, B. H. (2008). *Principles of anatomy and physiology*. John Wiley & Sons.
- Tumer, T.B., Rojas-Silva, P., Poulev, A., Raskin, I. and Waterman, C. (2015). Direct and indirect antioxidant activity of polyphenol- and isothiocyanate-enriched fractions from *Moringa oleifera*. *Journal of Agriculture and Food Chemistry* 63: 1505-1513.

- Uawonggul, N., Chaveerach, A., Thammasirirak, S., Arkaravichien, T., Chuachan, C., & Daduang, S. (2006). Screening of plants acting against *Heterometrus laoticus* scorpion venom activity on fibroblast cell lysis. *Journal of Ethnopharmacology*, 103(2), 201–207.
- Ukwuani, A. N., Ihebunna, O., Samuel, R. M., & Peni, I. J. (2012). Acute oral toxicity and antiulcer activity of *Piliostigma thonningii* leaf fraction in albino rats. *Bull. Env; Pharmacol. Life Sci*, 2(1), 41-45.
- Umali-Stuart, G., & Stiuart-Santiago, A. (2010). Phillippine Medicinal Plants: Family Melastomaceae.
- Van Valkenburg., & Bunyapraphatsara N, editors. Plant resources of South-East Asia No. 12(2): Medicinal and poisonous plants 2. *J Nat Prod* (2001); 365-366.
- Velioglu, Y. S., Mazza, G., Gao, L., & Oomah, B. D. (1998). Antioxidant activity and total phenolics in selected fruits, vegetables, and grain products. *Journal of agricultural and food chemistry*, 46(10), 4113-4117.
- Venables, C. W. (1986). Mucus, pepsin, and peptic ulcer. *Gut*, 27(3), 233-238.
- Wallace, J. L. (2008). Prostaglandins, NSAIDs, and gastric mucosal protection: why doesn't the stomach digest itself?. *Physiological reviews*, 88(4), 1547-1565.
- Waly, M. I., Al-Rawahi, A. S., Al Riyami, M., Al-Kindi, M. A., Al-Issaei, H. K., Farooq, S. A., ... & Rahman, M. S. (2014). Amelioration of azoxymethane induced-carcinogenesis by reducing oxidative stress in rat colon by natural extracts. *BMC complementary and alternative medicine*, 14(1), 60.
- Wang, X., Perez, E., Liu, R., Yan, L. J., Mallet, R. T., & Yang, S. H. (2007). Pyruvate protects mitochondria from oxidative stress in human neuroblastoma SK-N-SH cells. *Brain research*, 1132, 1-9.
- Waugh, A., & Grant, A. (2010). *Ross & Wilson anatomy and physiology in health and illness*. Elsevier Health Sciences.
- Wiar, C., Mogana, S., Khalifah, S., Mahan, M., Ismail, S., Buckle, M., ... & Sulaiman, M. (2004). Antimicrobial screening of plants used for traditional medicine in the state of Perak, Peninsular Malaysia. *Fitoterapia*, 75(1), 68-73.
- Winkler, F. K., Arcy, A. D., & Hunziker, W. (1990). Structure of human pancreatic lipase. *Nature*, 343(6260), 771.
- Wong, W. (2008). *Melastoma malabathricum*: Too beautiful to be called a weed. *Green Cult. Singapore Featur. Artic.*, no. August, 1-7.
- Woods, S. C. (2004). Gastrointestinal satiety signals I. An overview of gastrointestinal signals that influence food intake. *American Journal of Physiology-Gastrointestinal and Liver Physiology*, 286(1), G7-G13.
- Yasunaka, K., Abe, F., Nagayama, A., Okabe, H., Lozada-Pérez, L., López-Villafranco, E., ... & Reyes-Chilpa, R. (2005). Antibacterial activity of crude extracts from Mexican medicinal plants and purified coumarins and xanthenes. *Journal of ethnopharmacology*, 97(2), 293-299.

- Yen, G. C., Duh, P. D., & Tsai, H. L. (2002). Antioxidant and pro-oxidant properties of ascorbic acid and gallic acid. *Food chemistry*, 79(3), 307-313.
- Yoon, W. J., Ham, Y. M., Kim, S. S., Yoo, B. S., Moon, J. Y., Baik, J. S., ... & Hyun, C. G. (2009). Suppression of pro-inflammatory cytokines, iNOS, and COX-2 expression by brown algae *Sargassum micracanthum* in RAW 264.7 macrophages. *EurAsian Journal of BioSciences*, 3.
- Yoshida, T., Nakata, F., Hosotani, K., Nitta, A., & Okudat, T. (1992). Dimeric hydrolysable tannins from *Melastoma malabathricum*. *Phytochemistry*, 31(8), 2829-2833.
- Yu, C., Mei, X. T., Zheng, Y. P., & Xu, D. H. (2014). Gastroprotective effect of taurine zinc solid dispersions against absolute ethanol-induced gastric lesions is mediated by enhancement of antioxidant activity and endogenous PGE 2 production and attenuation of NO production. *European journal of pharmacology*, 740, 329-336.
- Yusof, M. M., Teh, L., Zakaria, Z., & Ahmat, N. (2011). Antinociceptive activity of the fractionated extracts of *Muntingia calabura*. *Planta Medica*, 77(12), PF21.
- Zakaria, Z. A., Jaios, E. S., Omar, M. H., Rahman, S. A., Hamid, S. S. A., Ching, S. M., ... & Taher, M. (2016). Antinociception of petroleum ether fraction derived from crude methanol extract of *Melastoma malabathricum* leaves and its possible mechanisms of action in animal models. *BMC complementary and alternative medicine*, 16(1), 488.
- Zakaria, Z. A., Balan, T., Mamat, S. S., Mohtarrudin, N., Kek, T. L., & Salleh, M. Z. (2015). Mechanisms of gastroprotection of methanol extract of *Melastoma malabathricum* leaves. *BMC complementary and alternative medicine*, 15(1), 135.
- Zakaria, Z. A., Balan, T., Suppaiah, V., Ahmad, S., & Jamaludin, F. (2014). Mechanism (s) of action involved in the gastroprotective activity of *Muntingia calabura*. *Journal of ethnopharmacology*, 151(3), 1184-1193.
- Zakaria, Z. A., Rofiee, M. S., Mohamed, A. M., Teh, L. K., & Salleh, M. Z. (2011a). In vitro antiproliferative and antioxidant activities and total phenolic contents of the extracts of *Melastoma malabathricum* leaves. *Journal of acupuncture and meridian studies*, 4(4), 248-256.
- Zakaria, Z. A., Mohamed, A. M., Jamil, N. M., Rofiee, M. S., Hussain, M. K., Sulaiman, M. R., ... & Salleh, M. Z. (2011b). In vitro antiproliferative and antioxidant activities of the extracts of *Muntingia calabura* leaves. *The American journal of Chinese medicine*, 39(01), 183-200.
- Zakaria, Z. A., Sufian, A. S., Ramasamy, K., Ahmat, N., Sulaiman, M. R., Arifah, A. K., ... & Somchit, M. N. (2010). In vitro antimicrobial activity of *Muntingia calabura* extracts and fractions. *African Journal of Microbiology Research*, 4(4), 304-308.
- Zakaria, Z. A., Somchit, M. N., Sulaiman, M. R., Mat Jais, A. M., & Fatimah, C. A. (2008). Effects of various receptor antagonists, pH and enzymes on *Muntingia calabura* antinociception in mice. *Res J Pharmacol*, 2(3), 31-37.

- Zakaria, Z. A., Mustapha, S., Sulaiman, M. R., Mat Jais, A. M., Somchit, M. N., & Abdullah, F. C. (2007e). The antinociceptive action of aqueous extract from *Muntingia calabura* leaves: the role of opioid receptors. *Medical Principles and Practice*, 16(2), 130-136.
- Zakaria, Z. A., Hassan, M. H., Nurul Aqmar, M. N. H., Abd Ghani, M., Mohd Zaid, S. N. H., Sulaiman, M. R., ... & Fatimah, C. A. (2007d). Effects of various nonopioid receptor antagonists on the antinociceptive activity of *Muntingia calabura* extracts in mice. *Methods and findings in experimental and clinical pharmacology*, 29(8), 515-520.
- Zakaria, Z. A., Mat Jais, A. M., Mastura, M., Mat Jusoh, S. H., Mohamed, A. M., Mohd, N. S., ... & Sulaiman, M. R. (2007c). In vitro antistaphylococcal activity of the extracts of several neglected plants in Malaysia. *Int J Pharmacol*, 3, 428-431.
- Zakaria, Z. A., Hazalin, N. M. N., Zaid, S. M., Ghani, M. A., Hassan, M. H., Gopalan, H. K., & Sulaiman, M. R. (2007b). Antinociceptive, anti-inflammatory and antipyretic effects of *Muntingia calabura* aqueous extract in animal models. *Journal of Natural Medicines*, 61(4), 443-448.
- Zakaria, Z. A., Kumar, G. H., Zaid, S. N., Ghani, M. A., Hassan, M. H., Hazalin, N. A., ... & Sulaiman, M. R. (2007a). Analgesic and antipyretic actions of *Muntingia calabura* leaves chloroform extract in animal models. *Oriental Pharmacy and Experimental Medicine*, 7(1), 34-40.
- Zakaria, Z. A., Sulaiman, M. R., Jais, A. M. M., Somchit, M. N., Jayaraman, K. V., Balakhrisnan, G., & Abdullah, F. C. (2006b). The antinociceptive activity of *Muntingia calabura* aqueous extract and the involvement of l-arginine/nitric oxide/cyclic guanosine monophosphate pathway in its observed activity in mice. *Fundamental & clinical pharmacology*, 20(4), 365-372.
- Zakaria, Z. A., Raden Mohd. Nor, R. N. S., Hanan Kumar, G., Abdul Ghani, Z. D. F., Sulaiman, M. R., Rathna Devi, G., ... & Fatimah, C. A. (2006a). Antinociceptive, anti-inflammatory and antipyretic properties of *Melastoma malabathricum* leaves aqueous extract in experimental animals. *Canadian Journal of Physiology and Pharmacology*, 84(12), 1291-1299.
- Zeeshan, M., Rizvi, S. M. D., Khan, M. S., & Kumar, A. (2012). Isolation, partial purification and evaluation of bioactive compounds from leaves of *Ageratum houstonianum*. *EXCLI journal*, 11, 78.
- Zimmermann, M. (1983). Ethical guidelines for investigations of experimental pain in conscious animals. *Pain*, 16(2), 109-110.