



UNIVERSITI PUTRA MALAYSIA

***PHYSICO-CHEMICAL PROPERTIES OF WATER, PROXIMATE
COMPOSITION, MINERALS AND ANTIOXIDANT OF SIX SPECIES OF
SEAGRASSES AT MERAMBONG SHOAL, JOHORE, MALAYSIA***

WAN HAZMA BINTI WAN NAWI

FP 2017 57



**PHYSICO-CHEMICAL PROPERTIES OF WATER, PROXIMATE
COMPOSITION, MINERALS AND ANTIOXIDANT OF SIX SPECIES OF
SEAGRASSES AT MERAMBONG SHOAL, JOHORE, MALAYSIA**

By

WAN HAZMA BINTI WAN NAWI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Master of Science**

March 2017

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

PHYSICO-CHEMICAL PROPERTIES OF WATER, PROXIMATE COMPOSITION, MINERALS AND ANTIOXIDANT OF SIX SPECIES OF SEAGRASSES AT MERAMBONG SHOAL, JOHORE, MALAYSIA

By

WAN HAZMA BINTI WAN NAWI

March 2017

Chairman : Muta Harah binti Zakaria @ Ya, PhD
Faculty : Agriculture

Seagrass has not yet been claimed as a dietary stuff and source of medicine for human consumption. Since several seagrass species are edible, investigation of nutrient, proximate composition and antioxidant properties in marine macrophytes are very important. Thus, the objectives of this study were to assess the physico-chemical and nutrient status of seawater in the seagrass habitat, determine the proximate composition, mineral contents and screening the antioxidant properties of six seagrass species; *Enhalus acoroides*, *Thalassia hemprichii*, *Cymodocea serrulata*, *Halodule uninervis*, *Halophila ovalis* (small and big-leaved) and *Halophila spinulosa* at Merambong shoal, Johore during the low tide, from March to August 2013. Nutritive proximate compositions were estimated following analysis methods described by the Association of Official Analytical Chemists. Results showed that seagrasses were high in carbohydrate (48.99-83.03%), fiber (10.02-27.48%) and ash (11.46-41.69%), and low in lipid content (0.02-1.22%). The protein content showed variation with species ranged 5.50% to 16.04%. The moisture content range from 6.68-21.49%. Macro-(Nitrogen-N, Phosphorus-P, Potassium-K, Calcium-Ca, Magnesium-Mg) and micro-(Zinc-Zn, Iron-Fe, Manganese-Mn, Copper-Cu) minerals measured using Atomic Absorption Spectrophotometer (AAS) and their concentrations varied with species and time. Minerals content; N, P, K, Ca, Mg, Zn, Fe, Mn, and Cu ranged 3308.00-36826.67 mg kg⁻¹, 429.33-3400.00 mg kg⁻¹, 2025.33-29520.00 mg kg⁻¹, 4680.00-25280.00 mg kg⁻¹, 3244.00-13040.00 mg kg⁻¹, 85.33-180.00 mg kg⁻¹, 922.67-5657.33 mg kg⁻¹, 13.33-129.33 mg kg⁻¹ and 3.20-29.20 mg kg⁻¹ respectively. Trend in micro-nutrient contents categorically followed a simple pattern, Fe>Zn>Mn>Cu. Methanol extracts of seagrasses were used to examine antioxidant activity by DPPH radical scavenging activity, total phenolic content (TPC) content and total flavonoid content (TFC). DPPH scavenging activity, TPC and TFC significantly different (p<0.05) between seagrass species with month. Rhizomes of *E. acoroides* and *H. uninervis* had relatively higher antioxidant activity (0.34 mg ml⁻¹ in June and 0.26 mg ml⁻¹ in July and August of EC₅₀ value respectively). Total phenolic content of seagrasses at Merambong shoal ranged 1.02 to 91.77 mg GAE kg⁻¹ with lower content in March and higher in August. The values of TFC were also varied among species and months (ranged from 43.25 to

361.46 mg QE kg⁻¹). Based on biplots generated by Principle component analysis (PCA), there were clear separation of proximate composition and mineral contents of seagrass species with locations from Malaysia and other regions. In conclusion, seagrasses possessed essential nutrient content and good antioxidant properties that could be utilized for pharmaceutical and nutraceutical uses.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

SIFAT FIZIKO-KIMIA AIR, KOMPOSISI PROKSIMAT, MINERAL DAN ANTIOKSIDA PADA ENAM SPESIES RUMPUT LAUT DI BETING MERAMBONG, JOHOR, MALAYSIA

Oleh

WAN HAZMA BINTI WAN NAWI

Mac 2017

Pengerusi : Muta Harah binti Zakaria @ Ya, PhD
Fakulti : Pertanian

Rumput laut belum diiktiraf sebagai barangan makanan dan sumber perubatan untuk kegunaan manusia. Memandangkan beberapa spesies rumput laut boleh dimakan, kajian nutrien, komposisi proksimat dan antioksidan dalam makrofit marin adalah sangat penting. Oleh itu, objektif kajian ini adalah untuk menilai fiziko-kimia dan status nutrien air laut dalam habitat rumput laut, mengkaji komposisi proksimat, kandungan mineral dan pemeriksaan sifat antioksidan enam spesies rumput laut; *Enhalus acoroides*, *Thalassia hemprichii*, *Cymodocea serrulata*, *Halodule uninervis*, *Halophila ovalis* (small and big-leaved) and *Halophila spinulosa* di Beting Merambong, Johor semasa air surut, dari bulan Mac hingga Ogos 2013. Nutrisi komposisi proksimat dianggarkan mengikut kaedah analisis diterangkan oleh Persatuan Official Analytical Chemists. Keputusan menunjukkan karbohidrat dalam rumput laut adalah tinggi (48.99% hingga 83.03%), serat (10.02 % hingga 27.48%) dan abu (11.46% hingga 41.69%), dan rendah kandungan lipid (0.02% hingga 1.22%). Kandungan protein berbeza mengikut spesies dari 5.50% hingga 16.04%. Kandungan kelembapan adalah di antara 6.68% hingga 21.49%. Makro-(Nitrogen-N, Fosforus-P, Kalium-K, Kalsium-Ca, Magnesium-Mg) dan mikro-(Zink-Zn, Ferum-Fe, Mangan-Mn, Tembaga-Cu) mineral diukur menggunakan Spektrofotometer Penyerapan Atom (AAS) kekekatannya adalah berbeza mengikut spesies dan masa. Kandungan mineral; N, P, K, Ca, Mg, Zn, Fe, Mn, dan Cu adalah di antara 3308.00-36826.67 mg kg⁻¹, 429.33-3400.00 mg kg⁻¹, 2025.33-29520.00 mg kg⁻¹, 4680.00-25280.00 mg kg⁻¹, 3244.00-13040.00 mg kg⁻¹, 85.33-180.00 mg kg⁻¹, 922.67-5657.33 mg kg⁻¹, 13.33-129.33 mg kg⁻¹ dan 3.20-29.20 mg kg⁻¹ masing-masing. Kecenderungan dalam kandungan mikro-nutrien dikategorikan mengikut corak yang mudah, Fe>Zn>Mn>Cu. Ekstrak metanol rumput laut telah digunakan untuk menentukan paras antioksidan dengan aktiviti memerangkap radikal DPPH, jumlah kandungan fenolik (TPC) dan jumlah kandungan flavonoid (TFC). Aktiviti menguraikan DPPH, TPC dan TFC berbeza dengan ketara (p<0.05) antara spesies rumput laut dengan bulan. Rizom *E. acoroides* dan *H. uninervis* mempunyai aktiviti antioksidan yang lebih tinggi (nilai EC₅₀ masing-masing 0.34 mg ml⁻¹ dalam bulan Jun dan 0.26 mg ml⁻¹ dalam bulan Julai dan Ogos). Jumlah kandungan fenolik rumput laut di Beting Merambong adalah 1.02-91.77 mg GAE kg⁻¹

dengan kandungan rendah dalam bulan Mac dan tinggi dalam bulan Ogos. Nilai TFC berubah antara spesies dan bulan (adalah di antara 43.25 hingga 361.46 mg QE kg⁻¹). Berdasarkan biplots dihasilkan oleh analisis komponen Prinsip (PCA), terdapat pemisahan yang jelas komposisi proksimat dan kandungan mineral spesies rumput laut dengan lokasi dari Malaysia dan negara lain. Kesimpulannya, rumput laut memiliki kandungan nutrien penting dan ciri antioksidan baik yang boleh digunakan untuk industri farmaseutikal dan nutraseutikal.



ACKNOWLEDGEMENTS

In the name of ALLAH the Most Gracious and the Most Merciful

I am most grateful to ALLAH SWT for His Blessings and Guidance. This is the most gratifying moment to thank and acknowledge the tremendous assistance and great contributions from individuals, organization and institutions throughout my work for past four years. Foremost, I would like to express my sincere gratitude to my research advisor Assoc. Prof. Dr. Muta Harah Zakaria @ Ya for the continuous support of my master science study and research, for her patience, motivation, enthusiasm and immense knowledge. I also truly appreciate her strong positive attitude, guidance and her willingness to give her time helped me in all the time of research and writing of this thesis has been very much appreciated. I could not have imagined having a better advisor and mentor for my study.

I would also like to express my deep gratitude and very great appreciation to my research co-supervisors, Prof. Dr. Japar Sidik Bujang and Dr. Natrah Fatin Mohd Ikhsan for their valuable and constructive suggestions during the planning and development of this study. I am particularly grateful for the assistance given by my committee members for their guidance, enthusiastic, encouragement and useful critiques for this study and ensuring that I can complete my study.

I also would like to thank my fellow labmates and friends, Nordiah Bidin, Dr. Shiamala Devi Ramaiya, Nur Farahin Syed, Emmelan Lau Shen Han, Nur Fatimah Abd. Halid and Saufinas Ismail for the stimulating discussions, for the sleepless nights we were working together, supportive advices, kindness and for all the fun we have had in the last four years. I also thank the Department of Aquaculture and Department of Crop Science, Faculty of Agriculture for their facilities, support and assistance since the start of my previous postgraduate work in 2013, especially the laboratory assistant officer, Mrs. Nur Shafika Maulad Abdul Jalil, Mr. Zawawi Idris and Tuan Haji Mohd. Khoiri Kandar.

I would like to acknowledge to Ministry of Education, Malaysia and Universiti Putra Malaysia, particularly in giving me the golden opportunity to get MyBrain15 Scholarship (My Master) and Graduate Research Fellowship (GRF) that provided the necessary financial and technical supports for this study. Last but not the least I would like to thank my beloved family especially my parents, Wan Nawi Wan Ismail and Mariam @ Hamidah Mustafa for financial support and supporting me spiritually throughout my life to complete this study. All their sacrifice, patience and encouragement will remain in my mind forever.

For any errors or inadequacies that may remain in this work, of course, the responsibility is entirely my own.

I certify that a Thesis Examination Committee has met on 31 March 2017 to conduct the final examination of Wan Hazma binti Wan Nawawi on her thesis entitled "Physico-Chemical Properties of Water, Proximate Composition, Minerals and Antioxidant of Six Species of Seagrasses at Merambong Shoal, Johore, Malaysia" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Annie Christianus, PhD

Senior Lecturer
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Hishamuddin bin Omar, PhD

Senior Lecturer
Faculty of Science
Universiti Putra Malaysia
(Internal Examiner)

Sitti Raehanah binti Muhamad Shaleh, PhD

Associate Professor
University Malaysia Sabah
Malaysia
(External Examiner)



NOR AINI AB. SHUKOR, PhD
Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 8 August 2017

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Muta Harah Zakaria @ Ya, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Japar Sidik Bujang, PhD

Professor
Faculty of Science
Universiti Putra Malaysia
(Member)

Natrah Fatin Mohd Ikhsan, PhD

Senior Lecturer
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by Graduate Student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly acknowledged;
- ownership of intellectual property from the thesis is as stipulated in the Memorandum of Agreement (MoA), or as according to the Universiti Putra Malaysia (Research) Rules 2012, in the event where the MoA is absent;
- permission from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) are required prior to publishing it (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No: Wan Hazma binti Wan Nawi, GS 34500

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____
Name of
Chairman of
Supervisory Committee: _____

Signature: _____
Name of
Member of
Supervisory Committee: _____

Signature: _____
Name of
Member of
Supervisory Committee: _____

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xii
LIST OF FIGURES	xiv
LIST OF ABBREVIATIONS	xviii
CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	3
2.1 Taxonomy and morphological study of tropical seagrasses in Malaysia	3
2.2 Ecological study of seagrass	4
2.3 Habitats and distribution of seagrass	6
2.4 Importance of seagrass	6
2.5 Proximate composition in seagrass	9
2.5.1 Protein	9
2.5.2 Lipid	11
2.5.3 Fiber	12
2.5.4 Ash	13
2.5.5 Moisture	13
2.5.6 Carbohydrates and nitrogen-free extractives	13
2.5.7 Caloric values	14
2.6 Mineral contents of seagrass	14
2.6.1 Macro-mineral in seagrass	15
2.6.2 Micro-mineral in seagrass	19
2.7 Antioxidant properties in seagrass	23
3 METHODOLOGY	25
3.1 Study area	25
3.2 Sample collection	25
3.3 Sample preparation	25
3.4 Determination of physical and chemical variables of water samples at Merambong shoal	27
3.5 Identification of seagrasses	30
3.6 Proximate composition analysis	31
3.6.1 Protein	31
3.6.2 Lipid	31
3.6.3 Fiber	32
3.6.4 Moisture	32
3.6.5 Ash	33
3.6.6 Carbohydrate	33
3.7 Wet digestion and dilution of macro- and micro-minerals	33

	analysis	
3.8	Determination of antioxidant activity	34
3.8.1	Total antioxidant activity (TAA)	34
3.8.2	Total phenolic content (TPC)	34
3.8.3	Total flavonoid content (TFC)	35
3.9	Statistical analysis	35
4	RESULTS AND DISCUSSIONS	37
4.1	Physico-chemical properties of water	37
4.1.1	Physical variables of seagrass bed	37
4.1.2	Chemical variables of seagrass bed	41
4.2	Nutrient content of seagrasses at Merambong shoal	45
4.2.1	Proximate composition of seagrass	45
4.2.2	Comparison of proximate composition in seagrass from different regions	61
4.2.3	Macro-minerals of seagrass	66
4.2.4	Micro-minerals of seagrass	73
4.2.5	Comparison of multi-mineral among the seagrass species	81
4.3	Antioxidant properties of seagrass	86
4.3.1	DPPH (2,2-diphenyl-1-picrylhydrazyl) radical scavenging activity	86
4.3.2	Total phenolic content (TPC)	89
4.3.3	Total flavonoid content (TFC)	89
4.3.4	Correlation between antioxidant properties of seagrass	91
5	SUMMARY, CONCLUSION AND RECOMMENDATION FOR THE FUTURE RESEARCH	93
	REFERENCES	95
	APPENDICES	110
	BIODATA OF STUDENT	119
	LIST OF PUBLICATIONS	120

LIST OF TABLES

Table		Page
1	Summary of proximate composition of different parts of seagrass species	10
2	Summary of Recommended Dietary Allowance (RDA), according to life stage and gender group for macro-mineral (Source: NAP, 2006)	16
3	Summary of macro-mineral and content in seagrass species from different locations	18
4	Summary of Recommended Dietary Allowance (RDA), according to life stage and gender group for micro-mineral (Source: NAP, 2001)	20
5	Summary of micro-mineral content in seagrass species parts from different locations	21
6	Pearson correlation of different water physical variables	40
7	Pearson correlations of water chemical variables	42
8	Correlation between ash in dry weight basis and mineral-content of seagrass	60
9	Correlation matrix (Pearson (n)) of four proximate variables	67
10	Nitrogen concentration in mg kg ⁻¹ of seagrass parts and whole plant at Merambong shoal	67
11	Phosphorus concentration in mg kg ⁻¹ of seagrass parts and whole plant at Merambong shoal	69
12	Potassium concentration in mg kg ⁻¹ of seagrass parts and whole plant at Merambong shoal	71
13	Calcium concentration in mg kg ⁻¹ of seagrass parts and whole plant at Merambong shoal	73
14	Magnesium concentration in mg kg ⁻¹ of seagrass part and whole plant at Merambong shoal	74
15	Zinc concentration in mg kg ⁻¹ of seagrass parts and whole plant at Merambong shoal	76
16	Iron concentration in mg kg ⁻¹ of seagrass parts and whole plant at Merambong shoal	77

17	Manganese concentration in mg kg ⁻¹ of seagrass parts and whole plant at Merambong shoal	79
18	Copper concentration in mg kg ⁻¹ of seagrass parts and whole plant at Merambong shoal	80
19	Principle component analysis (PCA) for macro-mineral content: Eigenvalues and percentage of variation described by the first three components	84
20	Eigenvectors for variables, the macro-mineral content used in PCA, indicating the strength of the correlation between variables and the principle components (axes)	84
21	Principle component analysis (PCA) for micro-mineral content: Eigenvalues and percentage of variation described by the first four components	85
22	Eigenvectors for variables, the micro-mineral content used in PCA, indicating the strength of the correlation between variables and the principle components (axes)	85
23	DPPH scavenging activity as EC ₅₀ in mg mL ⁻¹ of seagrass parts and whole plant from Merambong shoal for six months sampling	87
24	Total phenolic content (TPC) in mg GAE kg ⁻¹ of seagrass parts and whole plant from Merambong shoal for six months sampling	88
25	Total flavonoid content (TFC) in mg QE kg ⁻¹ of seagrass parts and whole plant from Merambong shoal for six months sampling	90
26	Principle component analysis (PCA) for antioxidant properties: Eigenvalues and percentage of variation described by the three components	92
27	Principle component analysis (PCA) for antioxidant properties: indicating the strength of the correlation between variables	92

LIST OF FIGURES

Figure		Page
1	Morphological illustration of seagrasses (Source: McKenzie, 2008)	4
2	Seagrass distribution of Peninsular and West East of Malaysia (Source: Japar Sidik <i>et al.</i> , 2006; Japar Sidik and Muta Harah, 2011)	8
3	Merambong shoal is located at Sg. Pulai estuary, Johore, the biggest seagrass bed of Peninsular Malaysia	26
4	Mixed seagrass of Merambong shoal, Sg. Pulai estuary, Johore during the spring low tide	26
5	The plant parts that were used of seagrass for proximate composition, mineral contents and antioxidant properties analyses in different sampling month	28
6	General sequence of analyses	29
7	Monthly water (a) temperature and salinity, (b) conductivity and (c) total dissolved solid of water measured <i>in-situ</i> at Merambong shoal (Refer Appendix 1)	38
8	Monthly dissolved oxygen measured <i>in-situ</i> water at Merambong shoal (Refer Appendix 1)	41
9	Monthly water pH measured <i>in-situ</i> at Merambong shoal compared with previous study by Japar Sidik <i>et al.</i> (1996) and Matias-Peralta and Yusoff (2014) (Refer Appendix 2)	42
10	Monthly (a) ammonia, (b) nitrate, (c) nitrite, (d) total nitrogen and (e) phosphate and total phosphorus in water at Merambong shoal (Refer Appendix 2)	43
11	Dendrogram of chemical variables at Merambong shoal, Johore for March to August 2013 using Ward's method	44
12	Seagrasses that were used during this study: (1) <i>Enhalus acoroides</i> , (2) <i>Thalassia hemprichii</i> , (3) <i>Halodule uninervis</i> , (4) <i>Cymodocea serrulata</i> , (5) <i>Halophila spinulosa</i> , (6) big-leaved <i>Halophila ovalis</i> and (7) small-leaved <i>Halophila ovalis</i>	46
13	Monthly proximate composition of protein (dry weight basis) in various parts and whole plant of linear leaves seagrass species (Refer Appendix 3)	47

14	Monthly proximate composition of protein (dry weight basis) in whole plant of broad leaves seagrass species (Refer Appendix 3)	48
15	Monthly proximate composition of lipid (dry weight basis) in various parts and whole plant of linear leaves seagrass species (Refer Appendix 3)	50
16	Monthly proximate composition of lipid (dry weight basis) in whole plant of broad leaves seagrass species (Refer Appendix 3)	51
17	Monthly proximate composition of fiber (dry weight basis) in various parts and whole plant of linear leaves seagrass species (Refer Appendix 3)	53
18	Monthly proximate composition of fiber (dry weight basis) in whole plant of broad leaves seagrass species (Refer Appendix 3)	54
19	Monthly proximate composition of moisture (dry weight basis) in various parts and whole plant of linear leaves seagrass species (Refer Appendix 3)	55
20	Monthly proximate composition of moisture (dry weight basis) in whole plant of broad leaves seagrass species (Refer Appendix 3)	56
21	Monthly proximate composition of ash (dry weight basis) in various parts and whole plant of linear leaves seagrass species (Refer Appendix 3)	58
22	Monthly proximate composition of ash (dry weight basis) in whole plant of broad leaves seagrass species (Refer Appendix 3)	59
23	Monthly proximate composition of carbohydrate (dry weight basis) in various parts and whole plant of linear leaves seagrass species (Refer Appendix 3)	62
24	Monthly proximate composition of carbohydrate (dry weight basis) in whole plant of broad leaves seagrass species (Refer Appendix 3)	63
25	The fraction of proximate composition for 6 month study in each seagrass species	64
26	Comparison of ash, carbohydrate, lipid and protein with other seagrass from different regions. 1=Ea-L, India, 2=Th-L, India, 3=Hd-L, Hawaiian Islands, 4=Hd-L, Florida, 5=Hd-Rhi/R, Hawaiian Islands, 6=Hd-Rhi/R, Florida, 7=Hh-L, Hawaiian Islands, 8=Hh-Rhi/R, Hawaiian Island, 9=Hj-L, Florida, 10=Hj-Rhi/R, Florida, 11=Cs-L, India, 12=Cr-L, India, 13=Si-L, India, 14=Hp-L, India, 15=Ea-Rhi, Malaysia, 16=Ea-L, Malaysia, 17=BL Ho-W, Malaysia, 18= SL Ho-W, Malaysia, 19=Hs-W,	65

Malaysia, 20=Hu-W, Malaysia, 21=Th-W, Malaysia, 22=Cs-W, Malaysia. Seagrass species: Ea=*Enhalus acoroides*, Cs=*Cymodocea serrulata*, Hp=*Halodule pinifolia*, Hu=*Halodule uninervis*, Si=*Syringodium isoetifolium*, BL Ho=big-leaved *Halophila ovalis*, SL Ho=small-leaved *Halophila ovalis*, Hs=*Halophila spinulosa*, Hd=*Halophila decipiens*, Hh=*Halophila hawaiiiana*, Hj=*Halophila johnsonii* and Th=*Thalassia hemprichii*. Plant part: W=Whole plant, L=Leaf, Rhi=Rhizome and R=Root (Refer Appendix 4)

- 27 Principle Component Analysis (PCA) of mineral contents with plot of the macro-mineral variables in seagrass and positions of 6 seagrass species according to their F scores macro-mineral variable. EA rhi=*Enhalus acoroides* rhizome, EA leaf=*Enhalus acoroides* leaves, TH=*Thalassia hemprichii*, CS=*Cymodocea serrulata*, HU=*Halodule uninervis*, BL-HO=big-leaved *Halophila ovalis*, SL-HO=small-leaved *Halophila ovalis* and HS=*Halophila spinulosa* 82
- 28 Principle Component Analysis (PCA) of mineral contents with plot of the micro-mineral variables in seagrass and positions of 6 seagrass species according to their F scores macro-mineral variable. EA rhi=*Enhalus acoroides* rhizome, EA leaf=*Enhalus acoroides* leaves, TH=*Thalassia hemprichii*, CS=*Cymodocea serrulata*, HU=*Halodule uninervis*, BL-HO=big-leaved *Halophila ovalis*, SL-HO=small-leaved *Halophila ovalis* and HS=*Halophila spinulosa* 82
- 29 Comparison of macro-mineral content with other seagrass from different region. 1=Ea-W, India, 2=Cs-W, India, 3=Cs-L, India, 4=Cs-Rhi, India, 5=Cs-R, India, 6=Hp-W, India, 7=Si-W, India, 8=Th-W, India, 9=Po-W, Greece, 10=Hst-W, Greece, 11=Ea-Rhi, Malaysia, 12=Ea-L, Malaysia, 13=BL Ho-W, Malaysia, 14=SL Ho-W, Malaysia, 15=Hs-W, Malaysia, 16=Hu-W, Malaysia, 17=Th-W, Malaysia, 18=Cs-W, Malaysia. Seagrass species: Ea=*Enhalus acoroides*, Cs=*Cymodocea serrulata*, Hp=*Halodule pinifolia*, Hu=*Halodule uninervis*, Si=*Syringodium isoetifolium*, Po=*Posidonia oceanica*, Hst=*Halophila stipulacea*, BL Ho=big-leaved *Halophila ovalis*, SL Ho=small-leaved *Halophila ovalis*, Hs=*Halophila spinulosa* and Th=*Thalassia hemprichii*. Plant part: W=Whole plant, L=Leaf, Rhi=Rhizome and R=Root. (Refer Appendix 7) 84
- 30 Comparison of micro-mineral content with other seagrass from different region. 1=Ho-W, India, 2=Ho-W, Australia, 3=Ho-Rhi, Australia, 4=Ho-R, Australia, 5=Ea-W, India, 6=Ea-W, India, 7=Cs-W, India, 8=Cs-W, India, 9=Cr-W, India, 10=Cr-W, India, 11=Cr-W, India, 12=Hu-W, India, 13=Hu-W, India, 14=Hp-W, India, 15=Hp-W, India, 16=Hp-W, India, 17=Hp-W, India, 18=Hd-W, India, 19=Hov-W, India, 20=Si-W, India, 21=Si-W, 85

India, 22=Si-W, India, 23=Rm-W, India, 24=Ea-Rhi, Malaysia, 25-Ea-L, Malaysia, 26=BL Ho-W, Malaysia, 27= SL Ho-W, Malaysia, 28=Hs-W, Malaysia, 29=Hu-W, Malaysia, 30=Th-W, Malaysia, 31=Cs-W, Malaysia. Seagrass species: Ea=*Enhalus acoroides*, Cs=*Cymodocea serrulata*, Cr=*Cymodocea rotundata*, Hp=*Halodule pinifolia*, Hu=*Halodule uninervis*, Si=*Syringodium isoetifolium*, Hd=*Halophila decipiens*, Hov=*Halophila ovata*, Ho=*Halophila ovalis*, BL Ho=big-leaved *Halophila ovalis*, SL Ho=small-leaved *Halophila ovalis*, Hs=*Halophila spinulosa*, Th=*Thalassia hemprichii* and Rm=*Rupia megacarpa*. Plant part: W=Whole plant, L=Leaf, Rhi=Rhizome and R=Root. (Refer Appendix 8)

- 31 Plot of the variables of antioxidant properties in seagrass species at Merambong shoal for six months according to PC1 and PC2. 92
EA rhi=*Enhalus acoroides* rhizome, EA leaf=*Enhalus acoroides* leaves, TH=*Thalassia hemprichii*, CS=*Cymodocea serrulata*, HU=*Halodule uninervis*, BL-HO=big-leaved *Halophila ovalis*, SL-HO=small-leaved *Halophila ovalis* and HS=*Halophila spinulosa*

LIST OF ABBREVIATIONS

AAS	Absorption atomic spectrometer
ABTS	2,2-azinobis (3-ethylbenzothiazoline-6-sulfonic acid)
ACA	Aldehyde/carboxylic acid
AI	Adequate intake
AlCl ₃	Aluminium chloride
ANOVA	Analysis of variance
AOAC	Association of Official Analytical Chemists
B	Boron
Ca	Calcium
Ca ²⁺	Calcium ion
Cl ⁻	Chloride ion
CO ₂	Carbon dioxide
Cu	Copper
DNA	Deoxyribonucleic acid
DNMRT	Duncan's New Multiple Range Test
DO	Dissolved oxygen
DPPH	2,2-diphenyl-1-picrylhydrazyl
DW	Dry weight
EC ₅₀	Effective concentration at 50%
FAO	Food and Agriculture Organization
Fe	Iron
FNB	Food and Nutrition Board
FOX	Ferrous oxidation-xylenol orange
FRAP	Ferric reducing/antioxidant power
FTC	Ferric thiocyanate
GAE	Gallic acid equivalents
GC	Gas chromatography
GE	Gross energy
H ⁺	Hydrogen ion
H ₂ O	Water
HCl	Hydrochloric acid
HCO ₃ ⁻	Bicarbonate ion
HPLC	High-performance liquid chromatography
H ₂ O	Water
H ₂ O ₂	Hydrogen peroxide
H ₂ SO ₄	Sulphuric acid
IOM	Institute of Medicine
K	Potassium
MA	Malonaldehyde
Mg	Magnesium
Mn	Manganese
Mo	Molybdenum
MSL	Mean sea level
N	Nitrogen
Na ⁺	Sodium ion
Na ₂ CO ₃	Sodium carbonate
NaNO ₂	Sodium nitrite

NaOH	Sodium hydroxide
NAP	National Academies Press
NCCFN	National Coordinating Committee on Food and Nutrition
NFE	Nitrogen-free extractives
NO ³ -N	Nitrate-nitrogen
NO ₂ ⁻ -N	Nitrite-nitrogen
O ₂	Oxygen
ORAC	Oxygen radical absorption capacity
P	Phosphorus
PCA	Principle component analysis
PO ₄ ³⁻	Phosphate ion
QE	Quercetin equivalents
RDA	Recommended Dietary Allowance
SCF	Scientific Committee on Food
TAA	Total antioxidant activity
TBA	Thiobarbituric acid assay
TDS	Total dissolved solids
TFC	Total flavonoid content
TPC	Total phenolic content
Trolox	(S)-(-)-6-hydroxy-2,5,7,8-tetramethylchroman-2-carboxylic acid
UNEP	United Nations Environment Programme
USEPA	United States Environmental Protection Agency
UV-VIS	Ultraviolet-visible
WHO	World Health Organization
Zn	Zinc

CHAPTER 1

INTRODUCTION

Seagrasses are the only flowering plants that have adapted themselves to living in marine and estuarine habitats, and are fully submerged most of the time (Japar Sidik *et al.*, 2006a). Malaysian seagrass can be found patchy in shallow intertidal and subtidal habitat, mangrove area, semi-enclosed lagoons, coral reef, rocky shores and shoals in subtidal zones (Japar Sidik and Muta Harah, 2003; Annaletchumy *et al.*, 2005; Japar Sidik *et al.*, 2006a; Japar Sidik *et al.*, 2012). There are 16 seagrass species recorded in Malaysia along its 4800 km coastal area (Japar Sidik and Muta Harah, 2011; Nguyen *et al.*, 2014; Japar Sidik *et al.*, 2016). Seagrasses are keystone species in many estuaries and shallow lagoons as excellent indicator providing unique ecosystems that act as primary production for economically and ecologically important higher consumers (UNEP, 2004; Japar Sidik *et al.*, 2006a; Short *et al.*, 2011).

Inhabit the area within the intertidal zone as Merambong shoal whose located near the estuary of Pulai river influenced by tidal much affect the growth of seagrasses. A good environment and suitable condition of water qualities are very important for optimum growth and survival of seagrasses. Therefore, it is significant to investigate the relationship between physico-chemical parameters of the environment with biochemical processes in seagrasses. Seagrass meadows are the most productive plant communities that providing important grazing habitat and food for a variety of marine organisms (Short and Wyllie-Echeverria, 1996). In seagrasses, metals sequestered may be passed through trophic links to higher level consumers such as fish (dengkis, *Siganus* sp.; sea urchin, *Echinoidea* spp.) including dugongs (*Dugong dugon*) and green turtles (*Chelonia mydas*) in tropical ecosystems (Brand-Gardner *et al.*, 1999; Japar Sidik *et al.*, 2006a).

Besides consumed by animals, there are reports by Green and Short (2003) and Kannan *et al.* (2011) where seagrasses are used as food sources for human in Southeast Asia regions such as Philippines, Indonesia, Thailand and Malaysia especially by the coastal populations. Compared to seaweed, the chemical composition of seagrass has been poorly investigated and most of the available information only deal with the seed of *Enhalus acoroides* (Montano *et al.*, 1999). The nutrients in these seagrasses have potential as new material sources for aquatic animal feed development.

There are abundance information regarding variation between the population of a given seagrass species for the chemical components of temperate, subtropical or tropical seagrasses. However, information on the nutritional value of seagrasses, especially in seagrasses of Malaysia are still lacking. Each plant species has its own nutrient composition that is important to both animals and human needs. Since a few of seagrasses was edible, investigation nutrient, mineral composition and antioxidant properties in marine aquatic macrophytes are very important. Although there are a few

studies have been conducted around the world the nutritional properties of seagrass are not completely known.

According to the previous study by Kannan *et al.* (2013b) seagrasses such as *Halodule pinifolia* and *Cymodocea rotundata* contain high level of antioxidant acting as major signaling pathways of cells and protector from oxidative damage. Seagrasses also have a good reputation in the medical field especially as remedy. Torre-Castroa and Ronnback (2004) reported that fishermen and seaweed farmers in East Coast of Zanzibar are using the roots of *E. acoroides* as a remedy against stings of different kinds of rays, scorpion fish (Scorpaenidae) as well as for diverse rabbit fish (Siganidae). It is also used for easing wounds, muscle pains and stomach problems. Another seagrass, *Cymodocea* spp. is good remedy against cough, malaria and used as a tranquillizer for babies. *Thalassia hemprichii* is being use as a treatment for malaria and relieving fever. and *Thalassodendron ciliatum* is used to relieve smallpox disease. A mixture of *Cymodocea* and *Thalassia* is good to treat skin diseases and fever. In spite of the above information very little are known on the usage and utilization of seagrasses.

Seagrasses are threatened by many factors especially by human activities. Rapid development is one of the predominant contributed of their disappearance around the world. This study focuses on a seagrass bed of Merambong shoal, Johor which is undergoing rapid development called Forest City. Thus, the objectives of this study were to:

1. assess the physico-chemical and nutrient status of seawater in the seagrass habitat at Merambong shoal.
2. determine the proximate composition and mineral contents of seagrasses in different sampling month inhabiting the Merambong shoal.
3. screening the antioxidant properties of different seagrasses in different sampling month harvested at Merambong shoal.

REFERENCES

- Agati, G., Azzarello, E., Pollastri, S. and Tattini, M. (2012). Flavonoids as antioxidants in plants: location and functional significance. *Plant Science*, 196: 67-76.
- Aggarwal, S. K. (2012). Herbaria and Data Information Systems in Plant Taxonomy. *Plant Taxonomy: past, present and future*, pp. 167.
- Akter, M., Ahmed, M. and Eun, J. B. (2010). Solvent effects on antioxidant properties of persimmon (*Diospyros kaki* L. cv. Daebong) seeds. *International Journal of Food Science and Technology*, 45(11): 2258-2264.
- Aluko, R. E. and Yada, R. Y. (1997). Some physicochemical and functional properties of Lima Bean (*Phaseolus lunatus*) isoelectric protein isolate as function of pH and salt concentration. *International Journal of Food Sciences and Nutrition*, 48: 31-35.
- Annaletchumy, L., Japar Sidik, B., Muta Harah, Z. and Arshad, A. (2005). Morphology of *Halophila ovalis* (R.Br.) Hook. f. from Peninsular and East Malaysia. *Pertanika Journal of Tropical Agricultural Science*, 28(1): 1-11.
- AOAC. (2012). Official methods of analysis of AOAC International.
- Arber, A. (1920). Water plants. A study of aquatic angiosperms. Cambridge University Press, Cambridge (digitally reprint 2010).
- Athiperumalsami, T., Kumar, V. and Jesudass, L. L. (2008). Survey and phytochemical analysis of seagrasses in the Gulf of Mannar, southeast coast of India. *Botanica Marina*, 51: 269–277.
- Athiperumalsami, T., Devi Rajeswari, V., Hastha Poorna, S., Kumar, V. and Louis Jesudass L. (2010). Antioxidant activity of seagrasses and seaweeds. *Botanica Marina*, 53(3): 251–257.
- Barbier, E. B., Hacker, S. D., Chris Kennedy, Koch, E. W., Stier, A. C. and Silliman, B. R. (2011). The value of estuarine and coastal ecosystem services. *Ecological Monographs*, 81(2): 169–193.
- Barnabas, A. D. (1991). *Thalassodendron ciliatum* (Forssk.) Den Hartog: root structure and histochemistry in relation to apoplastic transport. *Aquatic Botany*, 40(2): 129-143.
- Benzie, I. F. and Strain, J. J. (1999). Ferric reducing/antioxidant power assay: Direct measure of total antioxidant activity of biological fluids and modified version for simultaneous measurement of total antioxidant power and ascorbic acid concentration. *Methods in Enzymology*, 299: 15-27.
- Bhaskarachary, K. (2011). Potassium and human nutrition: The soil-plant-human continuum. *Karnataka Journal of Agricultural Sciences*, 24(1): 39-44.

- Bhattacharjee, S., Sultana, A., Sazzad, M. H., Islam, M. A., Ahtashom, M. and Asaduzzaman, M. (2013). Analysis of the proximate composition and energy values of two varieties of onion (*Allium cepa* L.) bulbs of different origin: A comparative study. *International Journal of Food Sciences and Nutrition*, 2(5): 246-253.
- Bhowmik, S., Datta, B. K. and Saha, A. K. (2012). Determination of mineral content and heavy metal content of some traditionally important aquatic plants of tripura, India using atomic absorption spectroscopy. *Journal of Agricultural Technology*, 8: 1467-1476.
- Brand-Gardner, S. J., Limpus, C. J. and Lanyon, J. M. (1999). Diet selection by immature green turtles, *Chelonia mydas*, in subtropical Moreton Bay, south-east Queensland. *Australian Journal of Zoology*, 47(2): 181-191.
- Brand-Williams, W., Cuvelier, M. E. and Berset, C. L. W. T. (1995). Use of a free radical method to evaluate antioxidant activity. *LWT-Food Science and Technology*, 28(1): 25-30.
- Bridson, D. and Forman, L. (1992). The Herbarium. Handbook. Second edition. Kew Royal Botanic Gardens, Surrey.
- Campbell, S. J., McKenzie, L. J. and Kerville, S. P. (2006). Photosynthetic responses of seven tropical seagrasses to elevated seawater temperature. *Journal of Experimental Marine Biology and Ecology*, 330(2): 455-468.
- Cao, G., Alessio, H. M. and Cutler, R. G. (1993). Oxygen-radical absorbance capacity assay for antioxidants. *Free Radical Biology and Medicine*, 14(3): 303-311.
- Castellano, G., Tena, J. and Torrens, F. (2012). Classification of phenolic compounds by chemical structural indicators and its relation to antioxidant properties of *Posidonia oceanica* (L.) Delile. *Communications in Mathematical and in Computer Chemistry*, 67: 231-250.
- Cebrian, J. and Duarte, C. M. (1998). Patterns in leaf herbivory on seagrasses. *Aquatic Botany*, 60(1): 67-82.
- Chan, P. T., Matanjun, P., Yasir, S. M. and Tan, T. S. (2015). Antioxidant activities and polyphenolics of various solvent extracts of red seaweed, *Gracilaria changii*. *Journal of Applied Phycology*, 27(6): 2377-2386.
- Choi, H. G., Lee, J. H., Park, H. H. and Sayegh, F. A. (2009). Antioxidant and antimicrobial activity of *Zostera marina* L. extract. *Algae*, 24(3): 179-184.
- Cob, Z. C., Arshad, A., Bujang, J. S. and Ghaffar, M. A. (2014). Spatial and temporal variations in *Strombus canarium* (Gastropoda: Strombidae) abundance at Merambong seagrass bed, Malaysia. *Sains Malaysiana*, 43(4): 503-511.

- Conti, M. E., Bocca, B., Iacobucci, M., Finoia, M. G., Mecozzi, M., Pino, A. and Alimonti, A. (2010). Baseline trace metals in seagrass, algae, and mollusks in a southern Tyrrhenian ecosystem (Linosa Island, Sicily). *Archives of environmental contamination and toxicology*, 58(1): 79-95.
- Cornelisen, C. D. and Thomas, F. I. (2006). Water flow enhances ammonium and nitrate uptake in a seagrass community. *Marine Ecology Progress Series*, 312: 1-13.
- Cox, S., Abu-Ghannam, N. and Gupta, S. (2010). An assessment of the antioxidant and antimicrobial activity of six species of edible Irish seaweeds. *International Food Research Journal*, 17: 205-220.
- Dastagir, G., Hussain, F., Khattak, F. and Khandzadi (2013). Proximate analysis of plants of family Zygophyllaceae and Euphorbiaceae during winter. *Sarhad Journal of Agriculture*, 29(3): 395-400.
- Dawes, C. J. and Lawrence, J. M. (1983). Proximate composition and caloric content of seagrasses. *Marine Technology Society Journal*, 17(2): 53–8.
- Dawes, C. J., Lobban, C. S. and Tomasko, D. A. (1989). A comparison of the physiological ecology of the seagrasses *Halophila decipiens* Ostenfeld and *H. johnsonii* Eiseman from Florida. *Aquatic Botany*, 33(1-2): 149-154.
- den Hartog, C. (1970). *The Seagrasses of the World*. North-Holland, Amsterdam.
- Denton, G. R. W., Marsh, H., Heinsohn, G. E. and Burdon-jones, C. (1980). The unusual metal status of the dugong *Dugon dugon*. *Marine Biology*, 57(3): 201-219.
- Devlin, R. M. (1967). *Plant Physiology*, Reinhold, New York, 564. Egborge, ABM (1994) *Water Pollution in Nigeria. Biodiversity and Chemistry of Warri River*.
- Duarte, C. M. (1991). Seagrass depth limits. *Aquatic Botany*, 40(4): 363-377.
- Edeogu, C. O., Ezeonu, F. C., Okaka, A. N. C., Ekuma, C. E. and Elom, S. O. (2007). Proximate composition of staple food crops in Ebonyi State (South Eastern Nigeria). *International Journal of Biotechnology and Biochemistry*, 3(1): 1-8.
- EFSA. (2006). Tolerable upper intake levels for vitamins and minerals. Retrieve from http://www.efsa.europa.eu/sites/default/files/efsa_rep/blobserver_assets/ndatol erableuil.pdf
- Fakhrulddin, I. M., Japar Sidik, B. and Muta Harah, Z. (2013). *Halophila beccarii* Aschers (Hydrocharitaceae) responses to different salinity gradient. *Journal of Fisheries and Aquatic Science*, 8(3): 462-471.
- FAO. (2002). *World agriculture: towards 2015/2040, Summary Report*. Published by Food and Agriculture Organization of the United Nations, Rome, ISBN 92-5-104761-8, pp. 97.

- FAO/WHO. (2005). Vitamin and mineral requirements in human nutrition. Dept. of Nutrition for Health and Development. Geneva: World Health Organization.
- Fernandez-Torquemada, Y. and Sanchez-Lizaso, J. L. (2005). Effects of salinity on leaf growth and survival of the Mediterranean seagrass *Posidonia oceanica* (L.) Delile. *Journal of Experimental Marine Biology and Ecology*, 320(1): 57-63.
- Fleurence, J. (1999). Seaweed proteins: biochemical, nutritional aspects and potential uses. *Trends in Food Science and Technology*, 10(1): 25-28.
- FNB. (1997). Institute of Medicine (US) Standing Committee on the Scientific Evaluation of Dietary Reference Intakes. Origin and Framework of the Development of Dietary Reference Intakes.
- Fonseca, M. S. and Cahalan, J. A. (1992). A preliminary evaluation of wave attenuation for four species of seagrass. *Estuarine, Coastal and Shelf Science*, 35: 565-576.
- Galloway, J. N., Dentener, F. J., Capone, D. G., Boyer, E. W., Howarth, R. W., Seitzinger, S. P., Asner, G. P., Cleveland, C. C., Green, P. A., Holland, E. A., Karl, D. M., Michaels, A. F., Porter, J. H., Townsend, A. R. and Voosmarty, C. J. (2004). Nitrogen cycles: past, present, and future. *Biogeochemistry*, 70(2): 153-226.
- Garrett, W. N. and Johnson, D. E. (1983). Nutritional energetics of ruminants. *Journal of Animal Science*, 57(2): 478-497.
- Gavin, N. M. and Durako, M. J. (2011). Localization and antioxidant capacity of flavonoids from intertidal and subtidal *Halophila johnsonii* and *Halophila decipiens*. *Aquatic Botany*, 95(3): 242-247.
- Gil, M. I., Tomás-Barberán, F. A., Hess-Pierce, B. and Kader, A. A. (2002). Antioxidant capacities, phenolic compounds, carotenoids, and vitamin C contents of nectarine, peach, and plum cultivars from California. *Journal of Agricultural and Food Chemistry*, 50(17): 4976-4982.
- Govindjee, R. (1995). Sixty-three years since Kautsky: Chlorophyll a fluorescence. *Australian Journal of Plant Physiology*, 22 (2): 131-160.
- Gras, A. F., Koch, M. S. and Madden, C. J. (2003). Phosphorus uptake kinetics of a dominant tropical seagrass *Thalassia testudinum*. *Aquatic Botany*, 76(4): 299-315.
- Grattan, S. R. and Grieve, C. M. (1992). Mineral element acquisition and growth response of plants grown in saline environments. *Agriculture, Ecosystems and Environment*, 38(4): 275-300.
- Green, E. P. and Short, F. T. (2003). World atlas of seagrasses. University of California Press.

- Greve, T. M. and Binzer, T. (2004). Which factors regulate seagrass growth and distribution. *European seagrasses: an introduction to monitoring and management. Monitoring and Managing of European Seagrasses Project (M&MS)*, pp. 19-23.
- Guo, C., Yang, J., Wei, J., Li, Y., Xu, J. and Jiang, Y. (2003). Antioxidant activities of peel, pulp and seed fractions of common fruits as determined by FRAP assay. *Nutrition Research*, 23(12): 1719-1726.
- Hach Handbook. (2011). Nitrogen, Ammonia. Method 8155 for water, wastewater and seawater salicylate method. Edition 5. Hach Company, USA.
- Heck, J. K. L., Hays, G. and Orth, R. J. (2003). Critical evaluation of nursery role hypothesis for seagrass meadows. *Marine Ecology Progress Series*, 253: 123-136.
- Hemminga, M. A. and Duarte, C. M. (2000). Seagrass ecology. Cambridge University Press, Cambridge, UK.
- Hoerudin, D. (2004). Phenolic and flavonoid contents of Australian honeys from different floral sources (Doctoral dissertation, Master Thesis): Queensland University, Brisbane, Australia).
- Iizumi, H. and Hattori, A. (1982). Growth and organic production of eelgrass (*Zostera marina* L.) in temperate waters of the Pacific coast of Japan. III. The kinetics of nitrogen uptake. *Aquatic Botany*, 12: 245-256.
- IOM. (2002). Dietary Reference Intakes for energy, carbohydrate, fiber, fat, fatty acids, cholesterol, protein and amino acids. Washington, DC: National Academy Press.
- ISO. (2007). Water quality. Determination of pH. Draft International Standard ISO/DIS 26149.
- Jagtap, T. G. (1983). Metal distribution in *Halophila beccarii* (Aschers) and surrounding environment along the central west coast of India. *Mahasagar*, 16(4): 429-434.
- Japar Sidik, B., Arshad, A., Hishamuddin, O., Muta Harah, Z. and Misni, S. (1996). Seagrass and macroalgal communities of Sungai Pulai estuary, south-west Johore, Peninsular Malaysia. In Kuo, J., Walker, D. I. and Kirkman, H. (eds.), *Seagrass Biology: Scientific Discussion from an International Workshop, Rottnest Island, Western Australia*, pp. 3-12.
- Japar Sidik, B., Muta Harah, Z., Mohd Pauzi, A. and Suleika, M. (1999). *Halodule* species from Malaysia-distribution and morphological variation. *Aquatic Botany*, 65: 33-46.

- Japar Sidik, B., Muta Harah, Z. and Short, F. T. (2016). Seagrass in Malaysia: Issues and Challenges Ahead. In C. Max Finlayson, G. Randy Milton, R. Crawford Prentice and Nick C. Davidson (eds.), Volume 2. The Wetland Book: II: Distribution, Description and Conservation. p.1-9. Springer Netherlands.
- Japar Sidik, B. and Muta Harah, Z. (2003). Seagrasses of Malaysia. In Green, E. P. and Short, F. T (eds.), Chapter 14. World Atlas of seagrasses. University of California Press, Berkeley, Los Angeles, London, pp. 152-160.
- Japar Sidik, B., Muta Harah, Z. and Arshad, A. (2006a). Distribution and significance of seagrass ecosystems in Malaysia. *Aquatic Ecosystem Health and Management*, 9(2): 203-214.
- Japar Sidik, B., Muta Harah, Z., Arshad, A., Lam, S. L. and Ogawa, H. (2006b). Flowers and sexes in Malaysian seagrasses. *Coastal Marine Science*, 30(1):184-188.
- Japar Sidik B. and Muta Harah, Z. (2011). Seagrasses in Malaysia. In H. Ogawa, Japar Sidik B. and Muta Harah, Z. (eds.), *Seagrasses: Resource status and trends in Indonesia, Japan, Malaysia, Thailand and Vietnam*, Seizando-Shoten Publishing Co., Ltd., Tokyo, pp. 22-37.
- Japar Sidik, B., Muta Harah, Z. and Kawaguchi, S. (2012). Historical review of seagrass research in Malaysia before 2001. *Coastal Marine Science*, 35(1): 157-168.
- Jeevitha, M., Athiperumalsami, T. and Kumar, V. (2013). Dietary fibre, mineral, vitamin, amino acid and fatty acid content of seagrasses from Tuticorin Bay, Southeast coast of India. *Phytochemistry*, 90: 135-146.
- Jimenez-Escrig, A., Rincon, M., Pulido, R., and Saura-Calixto, F. (2001). Guava fruit (*Psidium guajava* L.) as a new source of antioxidant dietary fiber. *Journal of Agricultural and Food Chemistry*, 49(11): 5489-5493.
- Johnson, M. W., Heck, K. L. and Fourqurean, J. W. (2006). Nutrient content of seagrasses and epiphytes in the northern Gulf of Mexico: evidence of phosphorus and nitrogen limitation. *Aquatic Botany*, 85(2): 103-111.
- Kannan, R., Ganesan, M., Govindasamy, C., Rajendran, K., Sampathkumar, P. and Kannan, L. (1992). Tissue concentration of heavy metals in seagrasses of the Palk Bay, Bay of Bengal. *International Journal of Ecology and Environmental Sciences*, 18: 29-34.
- Kannan, L., Thangaradjou, T. and Anantharaman, P. (1999). Status of seagrasses of India. *Seaweed Research and Utilisation*, 21: 25-33.
- Kannan, L. and Thangaradjou, T. (2006). Identification and assessment of biomass and productivity of seagrasses. National Training Workshop on Marine and Coastal Biodiversity Assessment for Conservation and Sustainable Utilization. *SDMRI Publication*, 10: 9-15.

- Kannan, R. R. R., Arumugam, R. and Anantharaman, P. (2010). Antibacterial potential of three seagrasses against human pathogens. *Asian Pacific Journal of Tropical Medicine*, 3(11): 890-893.
- Kannan, R. R. R., Arumugam, R. and Anantharaman, P. (2011). Chemometric studies of multielemental composition of few seagrasses from Gulf of Mannar, India. *Biological Trace Element Research*, 143(2): 1149-1158.
- Kannan, R. R. R., Arumugam, R. and Anantharaman, P. (2013a). Seagrasses as potential source of medicinal food ingredients: Nutritional analysis and multivariate approach. *Biomedicine and Preventive Nutrition*, 3: 375–380.
- Kannan, R. R. R., Arumugam, R., Thangaradjou, T. and Anantharaman, P. (2013b). Phytochemical constituents, antioxidant properties and p-coumaric acid analysis in some seagrasses. *Food Research International*, 54(1): 1229–1236.
- Kawashima, L. M. and Soares, L. M. V. (2003). Mineral profile of raw and cooked leafy vegetables consumed in Southern Brazil. *Journal of Food Composition and Analysis*, 16(5): 605-611.
- Kilminster, K. (2013). Trace element content of seagrasses in the Leschenault Estuary, Western Australia. *Marine Pollution Bulletin*, 73(1): 381-388.
- Kim, C. S. and Jung, J. (1993). The susceptibility of mung bean chloroplasts to photoinhibition is increased by an excess supply of iron to plants: a photobiological aspect of iron toxicity in plant leaves. *Photochemistry and Photobiology*, 58(1): 120-126.
- Kim, Y., Mosier, N. S., Hendrickson, R., Ezeji, T., Blaschek, H., Dien, B., Cotta, M. Dale, B. and Ladisch, M. R. (2008). Composition of corn dry-grind ethanol by-products: DDGS, wet cake, and thin stillage. *Bioresource Technology*, 99(12): 5165-5176.
- Koch, E. W. (1996). Hydrodynamics of a shallow *Thalassia testudinum* bed in Florida, USA. In Kuo, J., Phillips, R. C., Walker, D. I. and Kirkman, H. (eds.), *Seagrass Biology: proceedings of an international workshop*. Western Australia Museum, Perth, Australia, pp. 105–110.
- Koch, E. W., Ackerman, J., van Keulen, M. and Verduin, J. (2006). Fluid dynamics in seagrass ecology: from molecules to ecosystems. In Larkum, A. W. D., Orth, R. J. and Duarte, C. M. (eds.), *Seagrasses: biology, ecology and conservation*. Springer-Verlag, Heidelberg, Germany, pp. 193–225.
- Kuo, J. and den Hartog, C. (2007). Seagrass morphology, anatomy, and ultrastructure. In Larkum, A., Orth, R. J. and Duarte, C. M. (eds.), *Seagrasses: Biology, Ecology and Conservation*. Springer Netherlands, pp. 51-87.
- Krinsky, N. I. (1992). Mechanism of action of biological antioxidants. *Experimental Biology and Medicine*, 200(2): 248-254.

- Kwenin, W. K. J., Wolli, M. and Dzomeku, B. M. (2011). Assessing the nutritional value of some African indigenous green leafy vegetables in Ghana. *Journal of Animal and Plant Sciences*, 10(2): 1300-1305.
- Latha, S. and Daniel, M. (2001). Phenolic antioxidants of some common pulses. *Journal of Food Science and Technology*, 38(3): 272-273.
- Lauer, M. and Aswani, S. (2010). Indigenous knowledge and long term ecological change: detection, interpretation and response to changing ecological conditions in Pacific Island communities. *Environment Management*, 45: 985-997.
- Lanyon, J. (1986). Guide to the identification of seagrasses in the Great Barrier Reef Region. Great Barrier Reef Marine Park Authority, Queensland.
- Lanyon, J. M., Limpus, C. J. and Marsh, H. (1989). Dugong and turtles: grazers in the seagrass system. In Larkum A. W. D., McComb A. J. and Shepherd S. A. (eds.), *Biology of seagrasses: a treatise on the biology of seagrasses with special reference to the Australian region*. Elsevier Science Publishing Company, Amsterdam, pp. 105-112.
- Lanyon, J. M. and Marsh, H. (1995). Digesta passage times in the dugong. *Australian Journal of Zoology*, 43(2): 119-127.
- Lee, K. S. and Dunton, K. H. (1999). Inorganic nitrogen acquisition in the seagrass *Thalassia testudinum*: Development of a whole plant nitrogen budget. *Limnology and Oceanography*, 44(5): 1204-1215.
- Leong, L. P. and Shui, G. (2002). An investigation of antioxidant capacity of fruits in Singapore markets. *Food Chemistry*, 76(1): 69-75.
- Leterme, P., Buldgen, A., Estrada, F. and Londono, A. M. (2006). Mineral content of tropical fruits and unconventional foods of the Andes and the rain forest of Colombia. *Food Chemistry*, 95(4): 644-652.
- Levin, D. A. (1971). Plant phenolics: an ecological perspective. *American Naturalist*, 105(942): 157-181.
- Lyngby, J. E. and Brix, H. (1982). Seasonal and environmental variation in cadmium, copper, lead and zinc concentrations in eelgrass (*Zostera marina* L.) in the Limfjord, Denmark. *Aquatic Botany*, 14: 59-74.
- MacArtain, P., Gill, C. I., Brooks, M., Campbell, R. and Rowland, I. R. (2007). Nutritional value of edible seaweeds. *Nutrition Reviews*, 65(12): 535-543.
- Malea, P. (1994). Seasonal variation and local distribution of metals in the seagrass *Halophila stipulacea* (Forsk.) Aschers. in the Antikyra Gulf, Greece. *Environmental Pollution*, 85(1): 77-85.

- Malea, P., Haritonidis, S. and Kevrekidis, T. (1994). Seasonal and local variations of metal concentrations in the seagrass *Posidonia oceanica* (L.) Delile in the Antikyra Gulf, Greece. *Science of the Total Environment*, 153(3): 225-235.
- Malea, P., Boubonari, T. and Kevrekidis, T. (2008). Iron, zinc, copper, lead and cadmium contents in *Ruppia maritima* from a Mediterranean coastal lagoon: monthly variation and distribution in different plant fractions. *Botanica Marina*, 51(4): 320–330.
- Mann, J. and Stewart, T. (2012). Essentials of human nutrition. Oxford University Press.
- Marion, G. M., Millero, F. J., Camoes, M. F., Spitzer, P., Feistel, R. and Chen, C. T. (2011). pH of seawater. *Marine Chemistry*, 126(1): 89-96.
- Matanjun, P. (2008) Chemical Composition, Antioxidative and Cholesterol Lowering Properties of Selected Malaysia Seaweeds. PhD thesis, Universiti Putra Malaysia.
- Matias-Peralta, H. M. and Yusoff, F. M. (2014). Seasonal environmental quality variations in a tropical seagrass ecosystem at the Straits of Malacca. *The Malayan Nature Journal*, 66(1 and 2): 63-80.
- McDermid, K. J., Stuercke, B. and Balazs, G. H. (2007). Nutritional composition of marine plants in the diet of the green sea turtle (*Chelonia mydas*) in the Hawaiian Islands. *Bulletin of Marine Science*, 81(1): 55-71.
- McKenzie, L. J. (1994). Seasonal changes in biomass and shoot characteristics of a *Zostera capricorni* Aschers. dominant meadow in Cairns Harbour, northern Queensland. *Marine and Freshwater Research*, 45(7): 1337-1352.
- McKenzie, L. J. (2008). Seagrass-watch: Proceedings of a Workshop for Mapping and Monitoring Seagrass Habitats in North East Arnhem Land, Northern Territory, 18-20 October 2008. (Seagrass-Watch HQ, Cairns), pp. 49.
- McKenzie, L. J. and Campbell, S. J. (2004). Surviving The Summer Heat: Seagrass Burns As Corals Bleach. *Seagrass-Watch News*, 19: 1.
- McMillan, C. and Moseley, F. N. (1967). Salinity tolerances of five marine spermatophytes of Redfish Bay, Texas. *Ecology*, 48(3): 503-506.
- Meteorological Department Malaysia (2013). Records of Daily Rainfall Amount at Hospital Pontian for Year 2013.
- Miller, N. J. and Rice-Evans, C. A. (1997). Factors influencing the antioxidant activity determined by the ABTS•+ radical cation assay. *Free Radical Research*, 26(3): 195-199.

- Mohammad, M. J. and Mazahreh, N. (2003). Changes in soil fertility parameters in response to irrigation of forage crops with secondary treated wastewater. *Communications in Soil Science and Plant Analysis*, 34(9-10): 1281-1294.
- Molyneux, P. (2004). The use of the stable free radical diphenylpicrylhydrazyl (DPPH) for estimating antioxidant activity. *Songklanakarin Journal of Science and Technology*, 26(2): 211-219.
- Montano, N. M., Bonifacio, R. S. and Rumbaoa, R. G. O. (1999). Proximate analysis of the flour and starch from *Enhalus acoroides* (L.f.) Royle seeds. *Aquatic Botany*, 65(1-4): 321-5.
- Moon, J. K. and Shibamoto, T. (2009). Antioxidant assays for plant and food components. *Journal of Agricultural and Food Chemistry*, 57(5): 1655-1666.
- Moses, O., Olawuni, I. and Iwouno, J. O. (2012). The proximate composition and functional properties of full-fat flour, and protein isolate of lima bean (*Phaseolus lunatus*). 1: 349. doi:10.4172/scientificreports.349.
- Murphy, L. R., Kinsey, S. T. and Durako, M. J. (2003). Physiological effects of short-term salinity changes on *Ruppia maritima*. *Aquatic Botany*, 75: 293-309.
- Muta Harah, Z., Japar Sidik, B. and Arshad, A. (2002). Flowering, fruiting and seedling of annual *Halophila beccarii* Aschers. in Peninsular Malaysia. *Bulletin of Marine Science*, 71(3): 1199-1205.
- NAP. (2001). Dietary Reference Intakes for Vitamin A, Vitamin K, Arsenic, Boron, Chromium, Copper, Iodine, Iron, Manganese, Molybdenum, Nickel, Silicon, Vanadium and Zinc. Institute of Medicine of The National Academies, The National Academies Press. Washington D. C.
- NAP. (2006). Dietary Reference Intakes: The Essential Guide to Nutrient Requirements. Institute of Medicine of The National Academies, The National Academies Press. Washington D. C.
- NCCFN. (National Coordinating Committee on Food and Nutrition) (2005). Recommended Nutrient Intakes for Malaysia. A Report of the Technical Working Group on Nutritional Guidelines. Ministry of Health Malaysia, Putrajaya, Malaysia.
- NCCFN. (National Coordinating Committee on Food and Nutrition) (2010). Malaysian Dietary Guidelines. Ministry of Health Malaysia, Putrajaya, Malaysia.
- Nielsen, O. I., Koch, M. S., Jensen, H. S. and Madden, C. J. (2006). *Thalassia testudinum* phosphate uptake kinetics at low in situ concentrations using a ³³P radioisotope technique. *Limnology and Oceanography*, 51(1): 208.
- Nienhuis, P. H. (1986). Background levels of heavy metals in nine tropical seagrass species in Indonesia. *Marine Pollution Bulletin*, 17(11): 508-511.

- Nies, D. H. (1999). Microbial heavy-metal resistance. *Applied Microbiology and Biotechnology*, 51(6): 730-750.
- Norashikin, A., Harah, Z. M. and Sidik, B. J. (2013). Intertidal seaweeds and their multi-life forms. *Journal of Fisheries and Aquatic Science*, 8(3): 452.
- Norziah, M. H. and Ching, C. Y. (2000). Nutritional composition of edible seaweed *Gracilaria changii*. *Food Chemistry*, 68(1): 69-76.
- Nguyen, V. X., Detcharoen, M., Tuntiprapas, P., Soe-Htun, U., Sidik, J. B., Harah, M. Z. and Papenbrock, J. (2014). Genetic species identification and population structure of *Halophila* (Hydrocharitaceae) from the Western Pacific to the Eastern Indian Ocean. *BMC Evolutionary Biology*, 14(1): 1.
- Ou, B., Hampsch-Woodill, M. and Prior, R. L. (2001). Development and validation of an improved oxygen radical absorbance capacity assay using fluorescein as the fluorescent probe. *Journal of Agricultural and Food Chemistry*, 49(10): 4619-4626.
- Ozkok, A., D'arcy, B. and Sorkun, K. (2010). Total phenolic acid and total flavonoid content of Turkish pine honeydew honey. *Journal of ApiProduct and ApiMedical Science*, 2(2): 65-71.
- Ozyigit, I. I., Dogan, I., Demir, G., Eskin, B., Keskin, M. and Yalcin, I. E. (2013). Distribution of some elements in *Veronica scutellata* L. from Bolu, Turkey: soil-plant interactions. *Sains Malaysiana*, 42(10): 1403-1407.
- Pedersen, M. F. and Borum, J. (1992). Nitrogen dynamics of eelgrass *Zostera marina* during low nutrient availability. *Marine Ecology Progress Series*, 80: 65-73.
- Pedersen, M. F. and Borum, J. (1993). An annual nitrogen budget for a seagrass *Zostera marina* population. *Marine Ecology Progress Series*, 101: 169-169.
- Pedersen, M. F., Paling, E. I. and Walker, D. I. (1997). Nitrogen uptake and allocation in the seagrass *Amphibolis antarctica*. *Aquatic Botany*, 56: 105-117.
- Phillips, R. C. and Menez, E. G. (1988). Seagrasses. Smithsonian Contributions to the Marine Sciences. Number 34, Smithsonian Institution Press, Washington, DC, pp. 104.
- Ponnusamy, S. and Vellaichamy, T. (2012). Nutritional assessment, polyphenols evaluation and antioxidant activity of food resource plant *Decalepis hamiltonii* Wight & Arn. *Journal of Applied Pharmaceutical Science*, 2(5): 106-110.
- Pradheeba, M., Dilipan, E., Nobi, E. P. Thangaradjou T. and Sivakumar K. (2011). Evaluation of seagrasses for their nutritional value. *Indian Journal of Geo-Marine Sciences*, 40(1): 105-111.

- Prager, E. J. and Halley, R. B. (1999). The influence of seagrass on shell layers and Florida Bay mudbanks. *Journal of Coastal Research*, 15: 1151-1162.
- Prange, J. A. and Dennison, W. C. (2000). Physiological responses of five seagrass species to trace metals. *Marine Pollution Bulletin*, 41(7): 327-336.
- Prior, R. L., Hoang, H. A., Gu, L., Wu, X., Bacchiocca, M., Howard, L., Woodill, M. H., Huang, D., Ou, B. and Jacob, R. (2003). Assays for hydrophilic and lipophilic antioxidant capacity (oxygen radical absorbance capacity (ORACFL) of plasma and other biological and food samples. *Journal of Agricultural and Food Chemistry*, 51(11): 3273-3279.
- Pulich, W. M. (1980). Heavy metal accumulation by selected *Halodule wrightii* Asch. populations in the Corpus Christi Bay area (Seagrasses; Texas). *Contributions in Marine Science*, 23: 89-100.
- Pytkowicz, R. M. (1979). pH of Seawater. *Environmental International*, 2: 417-418.
- Ramage, D. L. and Schiel, D. R. (1998). Reproduction in the seagrass *Zostera novazelandica* on intertidal platforms in southern New Zealand. *Marine Biology*, 130(3): 479-489.
- Renaud, S. C., Guéguen, R., Schenker, J. and d'Houtaud, A. (1998). Alcohol and mortality in middle-aged men from eastern France. *Epidemiology*, 9(2): 184-188.
- Rollon, R. N., de Ruyter van Steveninck, E. D. and van Vierssen, W. (2003). Spatio-temporal variation in sexual reproduction of the tropical seagrass *Enhalus acoroides* (L.f.) Royle in Cape Bolinao, NW Philippines. *Aquatic Botany*, 76: 339-354.
- Santoso, J., Anwarriyah, S., Rumiantin, R. O., Putri, A. P., Ukhty, N. and Yoshie-Stark, Y. (2012). Phenol content, antioxidant activity and fibers profile of four tropical seagrasses from Indonesia. *Journal of Coastal Development*, 15(2): 189-196.
- Shams El Din, N. G. and El-Sherif, Z. M. (2013). Nutritional value of *Cymodocea nodosa* and *Posidonia oceanica* along the western Egyptian Mediterranean coast. *The Egyptian Journal of Aquatic Research*, 39(3): 153-165.
- Short, F. T., Carruthers, T., Dennison, W. and Waycott, M. (2007). Global seagrass distribution and diversity: A Bioregional model. *Journal of Experiment Marine Biology and Ecology*, 350: 3-20.
- Short, F. T., Polidoro, B., Livingstone, S. R., Carpenter, K. E., Bandeira, S., Bujang, J. S., Calumpang, H. P., Carruthers, J. B., Coles, R. G., Dennison, W. C., Erfemeijer, P. L., Fortes, M. D., Freeman, A. S., Jagtap, T. G., Abu Hena, M. K., Kendrick, G. A., Kenworthy, W. J., La Nafie, Y. A., Nasution, I. M., Orth, R. J., Prathep, A., Sanciangco, J. C., Tussenbroek, B. V., Vergara, S. G.,

- Waycott, M. and Zieman, J. C. (2011). Extinction risk assessment of the world's seagrass species. *Biological Conservation*, 144(7): 1961-1971.
- Short, F. T. and McRoy, C. P. (1984). Nitrogen uptake by leaves and roots of the seagrass *Zostera marina* L. *Botanica Marina*, 27(12): 547-556.
- Short, F. T. and Wyllie-Echeverria, S. (1996). Natural and human-induced disturbance of seagrasses. *Environmental Conservation*, 23(01): 17-27.
- Smil, V. (1999). Nitrogen in crop production: An account of global flows. *Global Biogeochemical Cycles*, 13(2): 647-662.
- Smolders, A. J. P., Vergeer, L. H. T., Van der Velde, G. and Roelofs, J. G. M. (2000). Phenolic contents of submerged, emergent and floating leaves of aquatic and semi-aquatic macrophyte species: why do they differ? *Oikos*, 91(2): 307-310.
- Soobrattee, M. A., Neerghen, V. S., Luximon-Ramma, A., Aruoma, O. I. and Bahorun, T. (2005). Phenolics as potential antioxidant therapeutic agents: mechanism and actions. *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, 579(1): 200-213.
- Sotirakou, E. Kladitis, G., Diamantis, N. and Grigoropoulou, H. (1999). Ammonia and phosphorus removal in municipal wastewater treatment plant with extended aeration. *Global Nest: the International Journal*, 1(1): 47-53.
- Stapel, J., Aarts, T. L., van Duynhoven, B. H., de Groot, J. D., van den Hoogen, P. H. and Hemminga, M. A. (1996). Nutrient uptake by leaves and roots of the seagrass *Thalassia hemprichii* in the Spermonde Archipelago, Indonesia. *Marine Ecology Progress Series*, 134: 195-206.
- Stott, L., Cannariato, K., Thunell, R., Haug, G. H., Koutavas, A. and Lund, S. (2004). Decline of surface temperature and salinity in the western tropical Pacific Ocean in the Holocene epoch. *Nature*, 431(7004): 56-59.
- Sumner, D. M. and Belaine, G. (2005). Evaporation, precipitation, and associated salinity changes at a humid, subtropical estuary. *Estuaries*, 28(6): 844-855.
- Suzuki, N., Fujimuraa, A., Nagaib, T., Mizumotoc, I., Itamid, T., Hatated, H., Nozawaa, T., Katoa, N., Nomotoe, T. and Yodaf, B. (2004). Antioxidative activity of animal and vegetable dietary fibers. *BioFactors*, 21: 329-333.
- Tee, E. S., Mohd Ismail, N., Mohd Nasir, A. and Khatijah, I. (1988). Nutrient composition of Malaysian foods. Kuala Lumpur: ASEAN Sub-Committee on Protein, Food Habits Research and Development.
- Temple, N. J. (2000). Antioxidants and disease: more questions than answers. *Nutrition Research*, 20(3): 449-459.
- Terrados, J. and Williams, S. L. (1997). Leaf versus root nitrogen uptake by the surfgrass *Phyllospadix torreyi*. *Marine Ecology Progress Series*, 149: 267-277.

- Thaipong, K., Boonprakob, U., Crosby, K., Cisneros-Zevallos, L. and Byrne, D. H. (2006). Comparison of ABTS, DPPH, FRAP, and ORAC assays for estimating antioxidant activity from guava fruit extracts. *Journal of Food Composition and Analysis*, 19(6): 669-675.
- Thangaradjou, T., Nobi, E. P., Dilipan, E., Sivakumar, K. and Susila, S. (2010). Heavy metal enrichment in seagrasses of Andaman Islands and its implication to the health of the coastal ecosystem. *Indian Journal of Marine Sciences*, 39, 85-91.
- Thangaradjou, T., Raja, S., Subhashini, P., Nobi, E. P. and Dilipan, E. (2013). Heavy metal enrichment in the seagrasses of Lakshadweep group of islands-a multivariate statistical analysis. *Environmental Monitoring and Assessment*, 185(1): 673-685.
- Torbatinejad, N. and Sabine, J. R. (2001). Laboratory evaluation of some marine plants on South Australian beaches. *Journal of Agricultural Science and Technology*, 3: 91-100.
- Torre-Castro, M. and Ronnback, P. (2004). Links between humans and seagrasses-an example from tropical East Africa. *Ocean and Coastal Management*, 47(7): 361-387.
- Touchette, B. W. and Burkholder, J. A. M. (2000). Review of nitrogen and phosphorus metabolism in seagrasses. *Journal of Experimental Marine Biology and Ecology*, 250(1): 133-167.
- UNEP, United Nations Environment Programme. (2004). Seagrass in the South China Sea. UNEP/ GEF/ SCS Technical Publication No.3, pp. 16.
- Vermaat, J. E., Agawin, N. S. R., Duarte, C. M., Fortes, M. D., Marba, N. and Uri, J. S. (1995). Meadow maintenance, growth and productivity of a mixed Philippine seagrass bed. *Marine Ecology Progress Series*, 124: 215-225.
- Vermaat, J. E., Verhagen, F. C. and Lindenburg, D. (2000). Contrasting responses in two populations of *Zostera noltii* Hornem. to experimental photoperiod manipulation at two salinities. *Aquatic Botany*, 67(3): 179-189.
- Vermani, A., Prabhat, N. and Chauhan, A. (2010). Physico-chemical analysis of ash of some medicinal plants growing in Uttarakhand, India. *Nature and Science*, 8(6): 88-91.
- Vunchi, M. A., Umar, A. N., King, M. A., Liman, A. A., Jeremiah, G. and Aigbe C. O. (2011). Proximate, vitamins and mineral composition of *Vitex doniana* (black plum) fruit pulp. *Journal of Basic Applied Sciences*, 19(1): 97-101.
- Wahbeh, M. I. (1984). Levels of zinc, manganese, magnesium, iron and cadmium in three species of seagrass from Aqaba (Jordan). *Aquatic Botany*, 20: 179-183.

- Walker, D. I. and McComb, A. J. (1990). Salinity response of the seagrass *Amphibolis antarctica* (Labill.) Sonder et Aschers.: an experimental validation of field results. *Aquatic Botany*, 36(4): 359-366.
- Walworth, J. (2013). Nitrogen in the Soil and the Environment. Department of Soil, Water and Environmental Science. The University of Arizona, pp. 1-3.
- Waycott, M., McMahon, K., Mellors, J., Calladine, A. and Kleine, D. (2004). A guide to tropical seagrasses of the Indo-West Pacific.
- Whelton, P. K., He, J., Cutler, J. A., Brancati, F. L., Appel, L. J., Follmann, D. and Klag, M. J. (1997). Effects of oral potassium on blood pressure: meta-analysis of randomized controlled clinical trials. *Jama*, 277(20): 1624-1632.
- Yamamuro, M. and Chirapart, A. (2005). Quality of the seagrass *Halophila ovalis* on a Thai intertidal flat as food for the dugong. *Journal Oceanography*, 61:18-36.
- Zar, J. H. (1999). Biostatistic analysis. 4th Edition, Prentice Hall, New Jersey, pp. 663.
- Zieman, J. C. and Zieman, R. T. (1989). Ecology of the seagrass meadows of the west coast of Florida: a community profile. Washington, DC: U.S. Dept. of the Interior, Fish and Wildlife Service, Research and Development, pp. 155.
- Zimmerman, R. C., Smith, R. D. and Alberte, R. S. (1987). Is growth of eelgrass nitrogen limited? A numerical simulation of the effects of light and nitrogen on the growth dynamics of *Zostera marina*. *Marine Ecology Progress Series*, 41: 167-176.
- Zin, Z. M., Abdul-Hamid, A. and Osman, A. (2002). Antioxidative activity of extracts from Mengkudu (*Morinda citrifolia* L.) root, fruit and leaf. *Food Chemistry*, 78(2): 227-231.