



UNIVERSITI PUTRA MALAYSIA

***EFFECTS OF CONJUGATED LINOLEIC ACIDS, FISH OIL AND
SOYBEAN OIL ON THE GROWTH PERFORMANCE, CARCASS TRAITS,
LIPID CHARACTERISTIC AND PPARs mRNA EXPRESSION IN
BROILER CHICKENS***

MARYAM ROYAN

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**Doctor of Philosophy
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2012

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THE GROWTH PERFORMANCE, CARCASS TRAITS, LIPID CHARACTERISTIC
AND PPARs mRNA EXPRESSION IN BROILER CHICKENS**

By

MARYAM ROYAN

**This Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
fulfillment of the Requirement for the Degree of Doctor of Philosophy**

June 2012

DEDICATION

I dedicate this thesis to

my husband Dr Bahman Navidshad, without whom I'd never have started and finished. I give my deepest expression of love and appreciation for the sacrifices you made during this PhD program;

and to our little angle Kasra.

Abstract of thesis presented to the Senate of Universiti Putra Malaysia
in fulfillment of the requirement for the degree of Doctor of Philosophy

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CHICKENS**

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June 2012

Chairman: Goh Yong Meng, PhD
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The conjugated linoleic acids (CLA) are known to decrease hepatic lipid deposition and overall lipid metabolism in selected animal models. These properties can be employed to reduce body fat accumulation in food animals, as in broiler chicken. In fact, every 1 % decrease in the chicken's body fat would result in the average Malaysian eating 300 g less fat for a year from chicken meat, based on per capita chicken meat consumption figures for 2010. However, the anti-lipogenic effects of CLA are variable and complicated because of the differences in animal species, diet composition and possibly their interactions with other dietary fatty acids. We hypothesized that CLA effects on fat metabolism in broiler chickens could be altered by the presence of dietary n-3 and n-6 fatty acids. Hence, fish oil (rich source of n-3 fatty acids) and soybean oil (rich source of n-6 fatty acids) were used in this trial.

This study aimed to investigate the effects of dietary conjugated linoleic acids, fish oil, soybean oil and their mixtures, as well as palm oil on broiler chickens. A total

of 560 day-old Ross 308 male broiler chickens allotted randomly into 7 equal treatment groups and used in 42 days experimental period. All chicks were fed a starter corn-soybean meal-based diet for 10 days. The treatment diets used were adjusted to be isocaloric and isonitrogenous comprising the corn-soybean meal plus specific dietary fat supplements consisting of conjugated linoleic acid (CLA), fish oil, soybean oil, palm oil or their combinations. The dietary fats were included in the experimental diets at 7% for single fats and 3.5% + 3.5% for dual mixes, and because of the lower metabolisable energy of palm oil as compared to PUFAs, its inclusion rate was about 12%. The conjugated linoleic acids supplement used in this study was LUTA-CLA 60, containing 60% conjugated linoleic acid. Therefore the dietary inclusions of 7 and 3.5% LUTA-CLA 60 were effectively supplying 4.2 and 2.1% CLA, respectively. The treatment groups were 7% soybean oil (SO), 7% LUTA-CLA 60 (CL), 7% fish oil (FO), 3.5% LUTA-CLA 60+ 3.5% soybean oil (CLSO), 3.5% fish oil + 3.5% soybean oil (FOSO), 3.5% LUTA-CLA 60 + 3.5% fish oil (CLFO) and 12% palm oil (PO). The experimental diets were used at grower (11-28 d) and finisher (29-42 d) phases and performance data were collected for each period. Tissue and serum sampling was carried out at the end of experiment (42 d).

The high level of palm oil inclusion in the diet did not adversely affect chicken's feed intake or growth rate. The lowest weight gains were recorded for birds from FO and CL groups during the grower (24.2 g/b/d) and finisher (50.9 g/b/d) phases, respectively. The FO diet also reduced the feed intake of chickens (34.5 g/b/d) while dietary conjugated linoleic acids had no effect on feed intake regardless of its dosage and combinations with other fats. It was also found that palm oil supplemented at about 12 % (w/w) increased the weight of abdominal fat pads significantly (%2.4 of live weight), while higher conjugated linoleic acids

content of the CL group increased post slaughter liver weights (%3.3 of live weight) ($P < 0.05$). Lipid contents of the breast tissues were higher in the PO (%2.46), SO (%2.02) and FO (%2.02) groups ($P < 0.05$) versus others (%1.11-1.55). Birds from the FO group had the highest amount of fats in their thigh muscles (%3.94) ($P < 0.05$).

Fish oil was more effective in reducing serum undesired lipoproteins (comparable effect with soybean oil), and CL diet enhanced serum favourite HDL fraction. It was also evident that conjugated linoleic acids in combination with soybean oil or fish oil resulted in less fat accumulation in both thigh and breast tissues, as compared to birds treated with soybean oil or fish oil only. Deposition and enrichment of longer chain n-3 fatty acids were also higher in the breast tissue of birds treated with conjugated linoleic acid in combination with fish oil (275 mg/100 g meat) as compared to the fish oil only treatment (254 mg/100 g meat).

The treatment oils demonstrated different effects on PPAR genes. The PPAR γ gene was up-regulated significantly in the PO group, whereas the levels of adipose PPAR γ gene expression were no different across treatments containing conjugated linoleic acids, fish oil, soybean oil or the mixture of these fats. On the other hand PPAR α gene expression in the liver tissue was up-regulated in response to dietary fish oil inclusion and the differences were significant for both FO and CLFO treatments compared to PO, SO and CL treatments.

In conclusion, the results of the present study showed that the dietary 7% fish oil or 4.2% CLA supplements reduced broiler chicken performance. This combination resulted in the enrichment of n-3 in chicken meat. The combination of CLA with soybean oil on the other hand increases the CLA levels in the

chicken meat. The PPAR α gene demonstrated anti-lipogenic effects when it is upregulated in the presence of CLA. Changes in the abdominal fat deposition in broiler chickens could be attributed to both PPAR α (in hepatocytes), and PPAR γ in adipocytes. Lower abdominal fat deposition was achieved by up-regulating the PPAR α (of hepatocytes), in tandem with the down-regulation of PPAR γ in the adipocytes.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

KESAN ASID LINOLEIK TERKONJUGAT, MINYAK IKAN DAN MINYAK SOYA PADA PRESTASI, CIRI KARKAS DAN LIPID SERTA EKSPRESI mRNA PPARs AYAM PEDAGING

Oleh

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Asid lemak terkonjugat (CLA) berupaya mengurangkan pengumpulan lipid hepar serta merendahkan metabolisme lemak haiwan. Kesan ini boleh digunapakai untuk mengurangkan pengumpulan lemak badan pada hawian ternakan, seperti ayam pedaging. Pengurangan sebanyak 1 % pada lemak badan ayam pedaging bakal mengurangkan sebanyak 300 g lemak setahun daripada daging ayam, berdasarkan data pengambilan daging ayam per kapita tahun 2010. Walaubagaimanapun, kesan anti-lipogenik CLA adalah berbeza dan kompleks mengikut spesis haiwan, kandungan diet, dan kemungkinan interaksi CLA dengan asid lemak yang lain. Hipotesis kajian ini berdasarkan andaian bahawa kesan CLA ke atas metabolisme lemak boleh diaruh oleh kehadiran asid lemak n-3 dan n-6. Sehubungan itu, minyak ikan (sumber asid lemak n-3) serta minyak kacang soya (sumber asid lemak n-6) digunakan dalam eksperimen ini.

Penyelidikan ini mengkaji kesan asid lemak terkonjugat, minyak ikan, minyak soya dan campurannya, serta minyak kelapa sawit ke atas ayam pedaging. Sejumlah 560 ayam pedaging Ross 308 jantan digunakan dalam kajian ini. Ayam tersebut dibahagikan secara rawak ke dalam 7 kumpulan dan diberi makan diet asas berteraskan jagung dan kacang soya selama 42 hari. Diet rawatan yang digunakan adalah isonitrogen dan isokalori setelah disesuaikan mengikut tahap dan jenis lemak suplemen yang digunakan. Suplemen lemak ditambah pada kadar 7% untuk lemak tunggal atau pada kadar 3.5% + 3.5% untuk campuran 2 lemak. Minyak kelapa sawit ditambah pada kadar yang lebih tinggi (12 %) memandangkan ia mempunyai tenaga metabolisme yang rendah berbanding asid lemak politaktepu yang lain.

Diet rawatan telah dirumus dan digunakan semasa peringkat tumbesaran (hari 11-28) dan peringkat penamat (hari 29-42). Data prestasi dikumpul untuk kedua-dua peringkat tersebut, sementara pensampelan tisu dan serum dilakukan di akhir eksperimen pada hari ke-42.

Suplemen asid lemak terkonjugat yang digunakan adalah LUTA-CLA 60 yang mengandungi 60% asid lemak terkonjugat. Sehubungan itu, kadar suplemen 7 % sebenarnya hanya membekalkan 4.2 % asid lemak terkonjugat, dan 3.5 % LUTA-CLA 60 hanya memberikan 2.1 % asid lemak terkonjugat. Berdasarkan maklumat ini, diet kumpulan rawatan adalah 7 % minyak soya (SO), 7 % LUTA-CLA 60 (CL), 7 % minyak ikan (FO), 3.5 % LUTA-CLA 60 + 3.5 % minyak soya (CLSO), 3.5 % minyak ikan + 3.5 % minyak soya (FOSO), 3.5 % LUTA-CLA 60 + 3.5 % minyak ikan (CLFO), dan 12 % minyak sawit (PO) ditambah kepada diet asas.

Penggunaan minyak sawit yang banyak dalam rangsum tidak menjejaskan kadar pengambilan makan dan tumbesaran ayam. Penambahan berat badan yang terendah dicatatkan pada ayam daripada kumpulan FO sewaktu tempoh tumbesaran dan kumpulan CLA untuk tempoh penyudahan. Diet FO juga mengurangkan tahap pengambilan makanan ayam, sementara asid lemak terkonjugat tidak menunjukkan kesan sedemikian apabila digunakan secara rawatan tunggal atau secara campuran dengan lemak lain. Minyak kelapa sawit yang diberikan pada tahap 12 % telah meningkatkan berat tisu lemak abdomen secara signifikan ($P < 0.05$), sementara tahap asid lemak terkonjugat dari kumpulan CLA telah menyebabkan berat hati yang lebih tinggi ($P < 0.05$). Kandungan lemak isi dada adalah tinggi di kalangan kumpulan PO, SO dan FO ($P < 0.05$) berbanding kumpulan lain. Ayam FO mencatatkan tahap lemak yang tertinggi untuk isi peha ($P < 0.05$). Secara amnya, kombinasi asid lemak terkonjugat dengan minyak soya atau minyak ikan telah menyebabkan pengumpulan lemak tisu yang lebih rendah pada isi dada dan peha, berbanding kumpulan ayam yang dirawat secara tunggal dengan minyak soya atau minyak ikan sahaja. Malah, ayam yang dirawat dengan kombinasi asid lemak terkonjugat dan minyak soya atau minyak ikan mencatatkan pengayaan tahap asid lemak n-3 yang lebih tinggi berbanding rawatan dengan minyak ikan atau minyak soya sahaja.

Keputusan kajian juga menunjukkan tiap jenis lemak/minyak mempunyai kesan yang berbeza ke atas gen PPAR. Aktiviti gen PPAR γ telah menunjukkan kenaikan dalam kumpulan PO, sementara minyak dan kombinasinya lain tidak menunjukkan sebarang kesan ke atas gen yang sama. Sementara itu, ekspresi gen PPAR α pada tisu hati telah meningkat

selepas dirawat dengan minyak ikan dan kombinasinya. Perbezaan ini adalah signifikan ($P < 0.05$) jika dibandingkan dengan PO, SO dan CLA.

Kesimpulannya, keputusan kajian ini menunjukkan bahawa penambahan 7 % minyak ikan dan 4.2 % CLA mengurangkan prestasi ayam pedaging. Walaubagaimanapun, kombinasi ini berupaya meningkatkan kandungan asid lemak n-3 dalam daging ayam. Gabungan CLA dan n-6 pula meningkatkan kandungan CLA dalam daging ayam. Kesan anti-lipogenik PPAR α ketara dengan kehadiran CLA. Perubahan kadar pengumpulan lemak abdomen bergantung kepada kesan PPAR α dan PPAR γ . Pengurangan lemak abdomen tercapai dengan peningkatan kawalaturan PPAR α (pada hepatosit), dan pengurangan kawalaturan PPAR γ pada sel adiposit.

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I certify that a Thesis Examination Committee has met on 2012 to conduct the final examination of **MARYAM ROYAN** on her Doctor of Philosophy thesis entitled “**EFFECTS OF CONJUGATED LINOLEIC ACIDS, FISH OIL AND SOYBEAN OIL ON THE GROWTH PERFORMANCE, CARCASS TRAITS, LIPID CHARACTERISTIC AND PPARs mRNA EXPRESSION IN BROILER CHICKENS**”

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DECLARATION

I hereby declare that the thesis is my original work except for quotations and citations, which have been duly acknowledged. I also declare that it has not been previously and is not correctly submitted for any other degree at Universiti Putra Malaysia or other institutions.



MARYAM ROYAN

Date: 14 June 2012



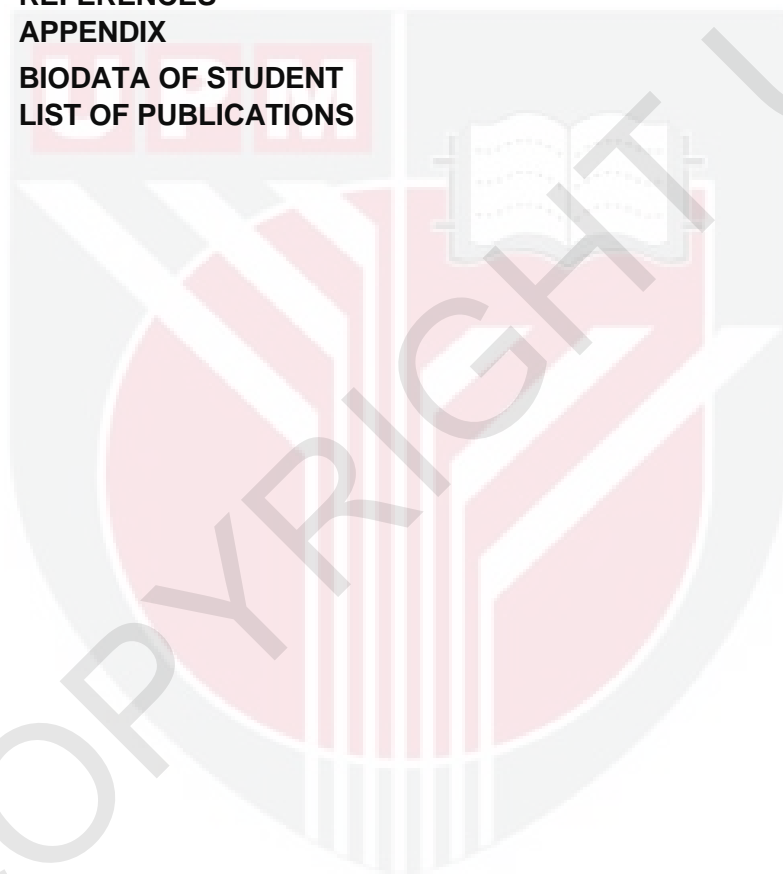
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LIST OF ABBREVIATIONS

AQP7	Aquaporin 7
°C	degrees Celsius
µg	micro gram
AA	Arachidonic Acid
ACC	Acetyl CoA carboxylase
ACO	Acyl-CoA oxidase
ACS	Acyl CoA synthetase
ANOVA	Analysis of variance
aP2	Adipocyte fatty acid binding protein
ATP	Adenosine triphosphate
Ava P	Available Phosphor
Ca	Calcium
cAMP	cyclic AMP
CD-36/FAT	Fatty acid translocase
CFIA	Canadian Food Inspection Agency
cGPDH	cytosolic glycerol phosphate dehydrogenase
CLA	Conjugated linoleic acid
CL	Diet containing 4.2% CLA
CLFO	Diet containing 2.1%CLA and 3.5% fish oil
CLSO	Diet containing 2.1%CLA and 3.5% soybean oil
CoA	Acyl coenzyme A
COX	Cyclooxygenase
CP	Crude Protein
CPT-I	Carnitine palmitoyl transferase I
Ct	Threshold cycle
Cys	Cysteine
d	Day
DAG	Diacylglycerols
DCP	Dicalcium phosphate
DHA	Docosahexaenoic acid
DM	Dry Matter
DNA	Deoxyribonucleic acid
EFA	Essential fatty acids
EFA	Essential Fatty Acid
FABP	Fatty Acid Binding Protein
FACoA	Fatty acyl CoA
FAS	Fatty acid synthase
FAT	Fatty acid transporter CD36
FATP	Fatty acid transport protein
FCR	Feed Conversion Ratio
FFAs	Free Fatty Acids
FO	Diet containing 7% fish oil
FOSO	Diet containing 3.5% fish oil and 3.5% soybean oil
FXR	Farnesoid X receptor
g	Gram
GLUT4	Glucose transporter 4
HDL	High Density Lipoprotein
HNF	Hepatic nuclear factors
IP3	Inositol triphosphate

Kd	Dissociation constant
kg	Kilo Gram
KGF	Keratinocyte Growth Factor
LA	Linoleic Acid
LCFAs	Long Chain Fatty Acids
LCPUFAs	Long Chain Poly Unsaturated Fatty Acids
LDL	Low Density Lipoprotein
LNA	Linolenic Acid
LOX	Lipoxygenase
LPL	Lipoprotein Lipase
LXR	liver X receptor
Lys	Lysine
ME	Metabolisable Energy
Met	Methionine
mg	Milligram
min	Minute
mL	Milliliter
mM	Millimole
mRNA	Messenger RNA
MUFAs	Monounsaturated fatty acids
Na	Sodium
NEFA	Non-esterified fatty acids
NFjB	Nuclear factor j B
NR	Nuclear receptor
NRC	Nutrient Requirements committee
P	Phosphor
PO	Diet containing about 12% palm oil
PPARS	Peroxisome proliferator-activated receptors
P/S	Polyunsaturated to saturated fatty acids ratio
PGE3	Prostaglandin E3
PGI3	Prostaglandin I3
PI	Peroxidizability index
PL	Phospholipids
pM/ μ L	Pico mole / micro liter
PO ₂	Oxygen partial pressure
PUFAs	Poly Unsaturated Fatty Acids
RNA	Ribonucleic acid
RQ	Respiratory quotient
RXR	Retinoid X receptor
RXR α	Retinoid X receptor
S.D	Standard Deviation
SAS	Statistical Analysis Software
SCD1	stearyl CoA desaturase-1
SFA	Saturated Fatty Acids
SO	Diet containing 7% soybean oil
SREBP-1c	Sterol regulatory element binding protein 1c
T3	Triiodothyronine
TCA cycle	Tricarboxylic acid cycle
TG	Triglycerides
TNF- α	Tumor necrosis factor- α
TXA ₃	Thromboxane A3
TZDs	Thiazolidinediones
UCPs	Uncoupling proteins
USFA	Unsaturated Fatty Acid
Vit-Min	Vitamin-Mineral Premix
VLDL	Very Low Density Lipoprotein



CHAPTER 1

GENERAL INTRODUCTION

1.1 Chicken meat as a functional food

Traditionally, poultry farmers aimed to produce birds with superior body weight and feed conversion ratio. However, in line with current developments, there are other traits that must be taken into account as well. These include producing poultry meat with lower body fat content, as well as improving the nutritiveness of the poultry meat to appeal to consumer requirements. The interrelated importance of human diet and health status is an ancient subject. In 500 B.C., Hippocrates was quoted to say that “Food is medicine, and medicine is food”. Human foods as a lifestyle factor, is involved in the incidence of many types of diseases such as cardiovascular diseases and cancer (Russo, 2009). Recent reports suggested that not only the quantity but also the composition of dietary fat is an important factor to prevent these metabolic diseases in human populations (Sealls *et al.*, 2008).

Nowadays, extra focus has been given to the production of functional foods, with components that have beneficial effects on human well-being. In definition, functional foods contain particular nutrients and (or) non-nutrients that have an effect on human health, beyond what is usually known as nutritional effects (Pisulewski, 2005). There are several compounds in foods that improve the

consumer's health status (Zanini *et al.*, 2006), among them the polyunsaturated fatty acids (PUFA, n-6 and n-3 series) and conjugated linoleic acid (CLA) (Pisulewski, 2005) have been more attractive. The fatty acid profile of chicken meat can be modified by feeding. This in turn results in chicken products that could offer a more economical source of health-effective fatty acids (Phetteplace and Watkins, 1989; Pisulewski *et al.*, 2002; Pisulewski and Kostogrys 2003; Pisulewski, 2005).

In humans, the n-3 fatty acids are important anti-inflammatory factors or precursors, with key roles in mitigating effects accompanying proinflammatory cytokines in coronary heart disease, depression, aging, cancer and autoimmune disorders such as Crohn's disease, ulcerative colitis, and lupus erythematosus (Simopoulos, 2002). CLA exerts anti-obesity, anti-carcinogenic, anti-atherogenic, and immunomodulatory effects in experimental animals and humans (Fritsche and Steinhart 1998a; Roche *et al.*, 2001). These specific fatty acids could affect chicken production efficiency too. High fat deposition in the chicken body reduces the effective energy use and can lead to lower feed efficiency (Grashorn, 2007). There are reports that PUFA and CLA could reduce fat deposition, but saturated fatty acids (SFA) from animal origin tend to result in higher fat deposition in the chicken carcass (Crespo and Esteve-Garcia, 2002; Zanini *et al.*, 2006). These reports obviously showed that dietary fats are important factors that influence both the functional properties of chicken meat and the efficiency of chicken production.

1.2 General objectives and organization of the study

The thesis consisted of two distinct sections. The literature review section briefly presents the highlights of fatty acid metabolism in birds. The later section of the literature review paid attention to the properties of PUFA and CLA, and their effects on body metabolism. In the current work, broiler chickens were fed dietary fats rich in n-3 or n-6 PUFA (fish oil and soybean oil, respectively), and CLA as the sole dietary fat source, or in dual mixes as well in order to modify the chicken's fat metabolism and meat fatty acid composition. Description of the general approach and methodology for this study is given in Chapter III.

Chapter IV presents the effects of different dietary fat types on the performance and carcass characteristics of broilers and reports on productive trait changes as a result of the dietary fat inclusion, while the experiment in chapter V investigated the effects of dietary fats on serum lipid fraction as an index of fat metabolism. The results reported in chapter VI demonstrated the effects of including different dietary fatty acid profiles on the enrichment of chicken's meat with n-3 PUFA and CLA.

In Chapter VII, Peroxisome proliferator-activated receptors (PPARs) gene expression in liver and adipose tissues as a consequence of feeding different dietary fatty acids was reported. This chapter is meant to illustrate the probable evidences that would help to clarify results obtained in Chapters IV, V and VI. At

the end of the thesis, a general discussion integrates the results of the whole research and the areas requiring further research.

It was hypothesized that the dietary n-3 fatty acids and CLA would induce changes in different aspects of lipid metabolism in broiler chickens. These fatty acids would alter both the fatty acid profile of the meat as well as the whole body fat deposition. When the mixtures of different fats were fed to broiler chickens, it was hypothesized that the fatty acids will interact to modify lipid metabolism. Based on these hypotheses, the general objectives of the present study were:

- 1- To determine the effects of soybean oil, fish oil, conjugated linoleic acid and palm oil on performance, serum lipids, chicken meat fat content and fatty acid profile and PPARs gene expression in broiler chickens.
- 2- To determine the likely synergistic effects between dietary fats on performance, serum parameters and tissue fatty acid composition of broiler chickens, focusing on anti-lipogenic effects.
- 3- To investigate the relationship between dietary n-6 : n-3 ratio and CLA concentrations on fat deposition.
- 4- To determine possible interactions between CLA and n-3 and n-6 PUFA on PPARs gene expression.

REFERENCES

- ABI (2002). Data analysis on the ABI Prism 7700 sequence detection system: setting baselines and thresholds. Applied Biosystems. USA.
- Ackman, R.G., W.M.N. Ratnayake and B. Olson. (1988). The “basic” fatty acid composition of Atlantic fish oils: potential similarities useful for enrichment of polyunsaturated fatty acids by urea complexation. *Journal of American Oil Chemistry Society*. 65:136–138.
- Adolf, R., S. Duval and E. Emeken. (2000). Biosynthesis of conjugated linoleic acid in humans. *Lipids*. 35, 131-135.
- Ahn D,U., J.L. Sell, C. Jo, M. Chamruspollert, M. Jeffrey. (1999). Effect of dietary conjugated linoleic acid on the quality characteristics of chicken eggs during refrigerated storage. *Poultry science*. 78: 922-928.
- Ajuyah, A.O., R.T. Hardin and J.S. Sim. (1993). Studies on canola seed in turkey grower diet: Effects on n-3 fatty acid composition of breast meat, breast skin and selected organs. *Canadian Journal of Animal Science*. 73:177–181.
- Akiba, Y., H. Murakami, N. Senkoylu, M. Kusanagi, K. Takahashi and K. Sato. (1995). The effects of dietary lipid on poultry performance and composition. *Proceeding of Australian Poultry Science Symposium*. 7:1-8.
- Allain, C.C., L.S. Poon, C.S.G. Chan, W. Richmond, P.C. Fu. (1974). Enzymatic Determination of Total Serum Cholesterol. *Clinical Chemistry*. 4:470-475.
- Allmann, D.W and D.W. Gibson. (1965). Fatty acid synthesis during early linoleic acid deficiency in the mouse. *Journal of Lipid Research*. 6: 51-60.
- Amri, E. Z., F. Bonino, G. Ailhaud, N. Abumrad and P.A. Grimaldi. (1995). Cloning of a protein that mediates transcriptional effects of fatty acids in preadipocytes. *The Journal of Biological Chemistry*. 270: 2367-2371.
- Annisson, E.F. (1983). Lipid metabolism. In, “Physiology and Biochemistry of the Domestic Fowl”. Ed. B.M. Freeman. Academic Press London.
- Annoni, G., B.M. Bottasso, D. Ciaci, M.F. Donato, A. Tripoli. (1982). Evaluation of a new enzymatic method for triglyceride estimation. *Journal of Research and Laboratory Medicine*. 9:115.
- AOAC. (2000). Official methods of analysis, 17th Ed, AOAC international, Gaithersburg, MD, USA, Official Method 999.11.

- Arakawa, K., and M. Sagai. (1986). Species differences in lipid peroxide levels in lung tissue and investigation of their determining factors. *Lipids*. 21:769–775.
- Arterburn, L.M., E.B. Hall and H. Oken. (2006). Distribution, interconversion, and dose response of n-3 fatty acids in humans. *The American Journal of Clinical Nutrition*. 83:1467S-1476S.
- Atkinson, R.L. (1999). Conjugated linoleic acid for altering body composition and treating obesity. In: Yurawecz, M.P., Mossoba, M.M., Kramer, J.K.G., Pariza, M.W., Nelson, G.J. (Eds.), *Advances in Conjugated Linoleic Acid Research*, vol. 1. AOCS Press, Champaign, IL, USA, pp. 348–353.
- Auboeuf, D., J. Rieusset and L. Fajas. (1997). Tissue distribution and quantification of the expression of mRNAs of peroxisome proliferator-activated receptors and liver X receptor- α in humans. No alteration in adipose tissue of obese and NIDDM patients. *Diabetes*. 46:1319–1327.
- Aviagen (2008). Ross 308 parent stock and broiler performance objectives. Ross breeders, <http://www.aviagen.com/>.
- Aydin, R., M. Pariza and M. Cook. (2001). Olive oil prevents the adverse effects of dietary conjugated linoleic acid on chick hatchability and egg quality. *Journal of Nutrition*. 131:800–806.
- Baddini, F.A., N.F. Pereira, F. da Costa and B.G. Ribeiro. (2009). Conjugated linoleic acid (CLA): effect modulation of body composition and lipid profile. *Nutrición Hospitalaria*. 24:422-428.
- Badinga, L., K.T, Selberg, A.C. Dinges, C.W. Comer and R.D. Miles. (2003). Dietary conjugated linoleic acid alters hepatic lipid content and fatty acid composition in broiler chickens. *Poultry Science*. 82: 111-116.
- Bailey, E and J.A. Horne. (1972). Formation and utilization of acetoacetate and β -hydroxybutyrate by various tissues of adult pigeon (*Columba livea*). *Comparative Biochemistry and Physiology*. 42B: 659-667.
- Balakumar, P., M. Rose and M. Singh. (2007). PPAR ligands: are they potential agents for cardiovascular disorders? *Pharmacology*. 80:1-10.
- Balao, N.C., L.J.C Lara. (2005). Oil and fat in broiler nutrition. *Brazilian Journal of Poultry Science*. 7:129-141.
- Banni, S. (2002). Conjugated linoleic acid metabolism. *Current Opinion in Lipidology*. 13:261-266.

- Banni, S., B.W. Day, R.W. Evans, F.P. Corongiu and B. Lombardi. (1995). Detection of conjugated diene isomers of linoleic acid in liver lipids of rats fed a choline-devoid diet indicates that the diet does not cause lipoperoxidation. *Journal of Nutrition. Biochemistry*. 6:281–289.
- Barceló-Coblijn, G and E.J. Murphy. (2009). Alpha-linolenic acid and its conversion to longer chain n-3 fatty acids: Benefits for human health and a role in maintaining n-3 fatty acid levels. *Progress in Lipid Research*. 48:355-374.
- Bartoň, L., M. Marounek, V. Kudrna, D. Bureš and R. Zahrádková. (2007). Growth performance and fatty acid profiles of intramuscular and subcutaneous fat from Limousin and Charolais heifers fed extruded linseed. *Meat Science*. 76:517-523.
- Bassaganya-Riera, J and R. Hontecillas. (2006). CLA and n-3 PUFA differentially modulate clinical activity and colonic PPAR-responsive gene expression in a pig model of experimental IBD. *Clinical Nutrition*. 25:454-465.
- Bee G. (2000). Dietary conjugated linoleic acid consumption during pregnancy and lactation influence on growth and tissue composition in weaned pigs. *Nutrition*. 130: 2981-2989.
- Bell, A.R., R. Savory, N.J. Horley, A.I. Choudhury, T.J. Dickins, A.M. Gray and D. Salter. (1998). Molecular basis of non-responsiveness to peroxisome proliferators: the guinea-pig PPAR α is functional and mediates peroxisome proliferator-induced hypolipidaemia. *Biochemistry Journal*. 332: 689–693.
- Benatti, P., G. Peluso, N.R. Nicolai and C. (2004). Polyunsaturated Fatty Acids: Biochemical, Nutritional and Epigenetic Properties. *American College of Nutrition*. 23:281-302.
- Berge, G.M., B. Ruyter and T. Asgard. (2004). Conjugated linoleic acid in diets for juvenile Atlantic salmon (*Salmo salar*); effects on fish performance, proximate composition, fatty acid and mineral content. *Aquaculture*. 237:365–380.
- Bessa, R.J.B., J. Santos-Silva, J.M.R. Ribeiro and A.V. Portugal. (2000). Reticulorumen biohydrogenation and the enrichment of ruminant edible products with linoleic acid conjugated isomers. *Livestock Production Science*. 63:201–211.
- Betti, M., B.L. Schneider, W.V. Wismer, V.L. Carney, M.J. Zuidhof, R.A. Renema. (2009). Omega-3 enriched broiler meat: 2. Functional properties, oxidative stability and consumer acceptance. *Poultry Science*. 88:1085–1095.
- Bezard, J., J.P. Blond, A. Bernard and P. Clouet. (1994). The metabolism and availability of essential fatty acids in animal and human tissues. *Reproduction Nutrition Development*. 34:539–568.

- Bhattacharya, A., J. Banu, M. Rahman, J. Causey and G. Fernandes. (2006). Biological effects of conjugated linoleic acids in health and disease. *The Journal of Nutritional Biochemistry*.17:789–810.
- Biely, J and B. March. (1957). Fat Studies in Poultry, Fat and Nitrogen Retention in Chicks Fed Diets Containing Different Levels of Fat and Protein. *Poultry Science*. 36:1235-1240.
- Blake, W.L and S.D. Clarke. (1990). Suppression of hepatic fatty acid synthase and S14 gene transcription by dietary polyunsaturated fat. *Journal of Nutrition*. 120:225–231.
- Bolukbasi, S.C. (2006). Effect of dietary conjugated linoleic acid (CLA) on broiler performance, serum lipoprotein content, muscle fatty acid composition and meat quality during refrigerated storage. *British Poultry Science*. 47:470–476.
- Botham, K.M., X. Zheng, M. Napolitano, M. Avella, C. Cavallari, R. Rivabene and E. Bravo. (2003). The effects of dietary n-3 polyunsaturated fatty acids delivered in chylomicron remnants on the transcription of genes regulating synthesis and secretion of very-low-density lipoprotein by the liver: modulation by cellular oxidative state. *Experimental Biology and Medicine* (Maywood). 228:143–151.
- Bretillon, L., J.M. Chardigny, O. Gregoire, O. Berdeaux and J.L. Sebedio. (1999). Effects of conjugated linoleic acid isomers on the hepatic microsomal desaturation activities in vitro. *Lipids*. 34:956-969.
- Brown, J.M., Y.D. Halvorsen, Y.R. Lea-Currie, C. Geigerman and M. McIntosh. (2001). Trans-10, cis-12, but not cis-9, trans-11, conjugated linoleic acid attenuates lipogenesis in primary cultures of stromal vascular cells from human adipose tissue. *Journal of Nutrition*. 131:2316-2321.
- Bou, R., R. Codony, A. Tres, E.A. Decker, F. Guardiola. (2009). Dietary strategies to improve nutritional value, oxidative stability, and sensory properties of poultry products. *Clinical Reviews in Food Science and Nutrition*. 49:800-822.
- Bouthegourd, J.C., P.C. Evans, D. Gripois, B. Tiffon, M.E. Blouguet, S. Roseau, C. Lutton, D. Tombe and J.C. Martin. (2002). A CLA mixture prevents body triglyceride accumulation without affecting energy expenditure in Syrian hamsters. *Journal of Nutrition*. 132:2682-2689.
- Buccioni, A., M. Antongiovanni, M. Mele, M. Gualtieri, S. Minieri and S. Rapaccini. (2009). Effect of oleic and conjugated linoleic acid in the diet of broiler chickens on the live growth performances, carcass traits and meat fatty acid profile. *Italian Journal of Animal Science*. 8:603-614.

- Bucolo, G., H. David. (1973). Quantitative determination of serum triglycerides by the use of enzymes. *Clinical Chemistry*. 19:476-482.
- Burdge, G. (2004). Alpha-linolenic acid metabolism in men and women: nutritional and biological implications. *Current Opinion in Clinical Nutrition and Metabolic Care*. 7:137-144.
- Burdge G.C., A.E. Jones and S.A. Wootton. (2002). Eicosapentaenoic and docosapentaenoic acids are the principal products of α -linoleic acid metabolism in young men. *British Journal of Nutrition*. 88:355-363.
- Burstein, M., H.R. Scholnic, R. Morfin. (1970). Rapid method for the isolation of lipoproteins from human serum by precipitation with poly anion. *Journal of Lipid Research*. 11:583-595.
- Calder, P.C. (1997). n-3 polyunsaturated fatty acids and cytokine production in health and disease. *Annals of Nutrition and Metabolism*. 41:203-234.
- Canadian Food Inspection Agency. (2003). Guide to Food Labelling and Advertising. <http://www.inspection.gc.ca/english/fssa/labeti/guide/ch7be.shtml#7.19>
- Cantor, A.H., E.A. Decker and V.P. Collins. (2000). Fatty acids in poultry and egg products. ed. Chin Kuang Chow. *Fatty Acids in Food and Their Health Implications*. Marcel Dekker, Inc. New York, NY 10016.
- Chakravarthy, M.V., Z. Pan, Y. Zhu, K. Tordjman, J.G. Schneider, T. Coleman, J. Turk and C.F. Semenkovich. (2005). "New" hepatic fat activates PPAR[α] to maintain glucose, lipid, and cholesterol homeostasis. *Cell Metabolism*. 1:309–322.
- Chambrier, C., J.P. Bastard, J. Rieusset, E. Chevillotte, D. Bonnefont-Rousselot, P. Therond, B.Hainque, J.P. Riou, M. Laville and H. Vidal. (2002). Eicosapentaenoic acid induces mRNA expression of peroxisome proliferator-activated receptor α . *Obesity Research*. 10: 518–525.
- Chawla A., J.J. Repa, R.M. Evans and D.J. Mangelsdorf. (2001). Nuclear receptors and lipid physiology: opening the X-files. *Science*. 294:1866–1870.
- Chawala, A., E.J. Schwarz, D.D. Dimaculangan and M.A. Lazar. (1994). Peroxisome proliferator activated receptor (PPAR) gamma: adipose-predominant expression and induction early in adipocyte differentiation. *Endocrinology*. 135:798-800.
- Cherian, G. and M.P. Goeger. (2004). Hepatic lipid characteristics and histopathology of laying hens fed CLA or n-3 fatty acids. *Lipids*. 39:31-36.

- Chevillotte, E., J. Rieusset, M. Roques, M. Desage and H. Vidal. (2001). The regulation of uncoupling protein-2 gene expression by w-6 polyunsaturated fatty acids in human skeletal muscle cells involves multiple pathways, including the nuclear receptor peroxisome proliferator-activated receptor. *Journal of Biological Chemistry*. 276:10853-10860.
- Chin, S.F., W. Liu, J.M. Storkson, Y.L. Ha and M.W. Pariza. (1992). Dietary sources of conjugated dienoic isomers of linoleic acid, a newly recognized class of anticarcinogens. *Journal of Food Composition and Analysis*. 5:185–197.
- Chin, S.F., J.M. Storkson, K.J. Albright, M.E. Cook and M.W. Pariza. (1994). Conjugated linoleic acid is a growth factor for rats as shown by enhanced weight gain and improved feed efficiency. *Journal of Nutrition*. 124:2344-2349.
- Choi, J.S., M.H. Jung, H.S. Park and J. Song. (2004). Effect of conjugated linoleic acid isomers on insulin resistance and mRNA levels of genes regulating energy metabolism in high-fat-fed rats. *Nutrition*. 20:1008-1017.
- Choi, Y., Y.C. Kim, Y.B. Han, Y. Park, M.W. Pariza and J.M. Ntambi. (2000). The trans-10, cis-12 isomer of conjugated linoleic acid down regulate stearyl-CoA desaturase gene expression in 3T3-L1 adipocytes. *Journal of Nutrition*. 130:1920–1924.
- Choi, Y., Y. Park, J.M. Storkson, M.W. Pariza and J.M. Ntambi. (2002). Inhibition of stearyl-CoA desaturase activity by the cis-9, trans-11 isomer and the trans-10, cis-12 isomer of conjugated linoleic acid in MDA-MB-231 and MCF-7 human breast cancer cells. *Biochemical and Biophysical Research Communications*. 294:785-790.
- Christiansen, E.N., J.S. Lund, T. Rørtveit and A.C. Rustan. (1991). Effect of dietary n-3 and n-6 fatty acids on fatty acid desaturation in rat liver. *Biochimica et Biophysica Acta*. 1082:57–62.
- Clarke, S.D. (2000). Polyunsaturated fatty acid regulation of gene transcription: a mechanism to improve energy balance and insulin resistance. *British Journal of Nutrition*. 83:S59-S66.
- Clarke, S.D., M.K. Armstrong and D.B. Jump. (1990) Nutritional control of rat liver fatty acid synthase and S14 mRNA abundance. *Journal of Nutrition*. 120:218-224.
- Clarke, B.A. and S.D. Clarke. (1982). Suppression of rat liver fatty acid synthesis by eicosa-5,8,11,14-tetraenoic acid without a reduction in lipogenic enzymes. *Journal of Nutrition*. 112:1212-1219.

- Clarke, S.D. and D.B. Jump. (1993). Fatty acid regulation of gene expression: a unique role for polyunsaturated fats. In: 'Nutrition and Gene Expression' Eds Berdanier, C. & Hargrove, J.L. CRC Press, Boca Raton, Florida, pp. 227-246.
- Clement, L., H. Poirier, I. Niot, V. Bocher, M. Guerre-Millo, S. Krief, B. Staels and P. Besnard. (2002). Dietary trans-10,cis-12 conjugated linoleic acid induces hyperinsulinemia and fatty liver in the mouse. *Journal of Lipid Research*. 43: 1400–1409.
- Cohen, C., G. Perrault, C. Voltz, R. Steinberg and P. Soubrié. (2002). SR141716, a central cannabinoid (CB(1)) receptor antagonist, blocks the motivational and dopamine-releasing effects of nicotine in rats. *Behavioural Pharmacology*. 13:451-463.
- Coleman, R.A., T.M. Lewin, C.G. Van Horn, M.R. Gonzalez-Baró. (2002). Do long-chain acyl-CoA synthases regulate fatty acid entry into synthetic versus degradative pathways? *Journal of Nutrition*. 132:2123–2126.
- Corino, C., J. Mourot, S. Magni, G. Pastorelli and F. Rosi. (2002). Influence of dietary conjugated linoleic acid on growth, meat quality, lipogenesis, plasma leptin and physiological variables of lipid metabolism in rabbits. *Journal of Animal Science*, 80:1020–1028.
- Cunningham, B.A., J.T. Moncur., J.T. Huntington., W.B. Kinlaw. (1998). "Spot 14" protein: a metabolic integrator in normal and neoplastic cells. *Thyroid*. 8: 815-825.
- Cook, H.W. (1991). Fatty acid desaturation and chain elongation in eukaryotes. In 'Biochemistry of Lipids, Lipoproteins and Membranes'. Eds Vance, D.E.
- Cook, M.E., C.C. Miller, Y. Park and M.W. Pariza. (1993). Immune modulation by altered nutrient metabolism: nutritional control of immune-induced growth depression. *Poultry Science*. 72:1301-1305.
- Corl, B.A., L.H. Baumgard, D.A. Dwyer, J.M. Griinari, B.S. Phillips, D.E. Bauman. (2001). The role of delta(9)-desaturase in the production of cis-9, trans-11 CLA. *Journal of Nutrition and Biochemistry*. 12:622–630.
- Couet, C., J. Delarue, P. Ritz, J.M. Antoine and F. Lamisse. (1997). Effect of dietary fish oil on body fat mass and basal oxidation in healthy adults. *International Journal of Obesity*. 21:637- 643.
- Crespo, N. and E. Esteve-Garcia. (2002). Nutrient and fatty acid deposition in broilers fed different dietary fatty acid profiles. *Poultry Science*. 81:1533–1542.
- Daggy, B., C. Arost and A. Bensadoun. (1987). Dietary fish oil decreases VLDL production rates. *Biochimica et Biophysica Acta*. 920:293-300.

- Darshan, S. (2001). Modulation of Human Immune and Inflammatory Responses by Dietary Fatty Acids. *Nutrition*. 17:669–673.
- Degrace, P., L. Demizieux, J. Gresti, J.M. Chardigny, J.L. Sébédio and P. Clouet. (2004). Hepatic steatosis is not due to impaired fatty acid oxidation capacities in C57BL/6J mice fed the conjugates trans-10, cis-12-isomer of linoleic acid. *Journal of Nutrition*. 134:861-867.
- Denbow, D. M. (2000). Gastrointestinal anatomy and physiology. Pages 299-325. In: *Sturkie's Avian Physiology*. G. C. Whittow, ed. Academic Press, San Diego, California.
- Denli, M., F. Okan and F. Doran. (2004). Effect of conjugated linoleic acid (CLA) on the performance and serum variables of broiler chickens intoxicated with aflatoxin B₁. *South African Journal of Animal Science*. 34:97–103.
- Denli, M., F. Okan, F. Doran and T.C. Inal. (2005). Effect of dietary conjugated linoleic acid (CLA) on carcass quality, serum lipid variables and histopathological changes of broiler chickens infected with aflatoxin B₁. *South African Journal of Animal Science*. 35:109-116.
- Desvergne, B., L. Michalik and W. Wahli. (2006). Transcriptional regulation of metabolism. *Physiological Reviews*. 86:465-514.
- Desvergne, B. and W. Wahli. (1999). Peroxisome proliferator-activated receptors: nuclear control of metabolism. *Endocrine Reviews*. 20:649-88.
- Ding, S.T., Y.C. Li, K.E. Nestor, S.G. Velleman and H.J. Mersmann. (2003). Expression of turkey transcription factors and Acyl-coenzyme oxidase in different tissues and genetic populations. *Poultry Science*. 82:17–24.
- Doreau, M and Y. Chillard. (1997). Digestion and metabolism of dietary fat in farm animals. *British Journal of Nutrition*. 78:Suppl.S15-S35.
- Drackley, J.K. (2000). Lipids Metabolism. Chap. 5. *Farm Animal Metabolism and Nutrition*. ed. J.P.F. D'Mello. New York, NY, 10003, USA.
- Dreyer, C., G. Krey, H. Keller, F. Givel, G. Helftenbein and W. Wahli. (1992). Control of the peroxisomal beta-oxidation pathway by a novel family of nuclear hormone receptors. *Cell*. 68:879–887.
- Du, M., and, D.U. Ahn. (2002). Effect of dietary conjugated linoleic acid on the growth rate of live birds and on the abdominal fat content and quality of broiler meat. *Poultry Science*. 81:428–433.

- Du, M., and, D.U. Ahn. (2003). Dietary CLA affects lipid metabolism in broiler chicks. *Lipids*. 38:505- 511.
- Du, M., D.U. Ahn and J.L. Sell. (1999). Effect of dietary conjugated linoleic acid on the composition of egg yolk lipids. *Poultry Science*. 78:1639–1645.
- Eder, K., N. Slomma and K. Becker. (2002). Trans-10, cis-12 conjugated linoleic acid suppresses the desaturation of linoleic acid and alpha-linolenic acids in HepG2 cells. *Journal of Nutrition*. 132:1115-1121.
- Eggert, J. M., M.A. Belury, A. Kempa-Steczko, S.E. Mills and A.P. Schinckel. (2001). Effects of conjugated linoleic acid on the belly firmness and fatty acid composition of genetically lean pigs. *Journal of Animal Science*. 79:2866–2872.
- Engelke, C.F., B.D. Siebert, K. Gregg, A.D.G. Wright and P.E. Vercoc. (2004). Kangaroo adipose tissue has higher concentrations of cis 9, trans 11-conjugated linoleic acid than lamb adipose tissue. *Journal of Animal and Feed Sciences*. 13:689–692.
- Enser, M., and J.L. Roberts. (1982). The regulation of hepatic stearylcoenzyme A desaturase in obese-hyperglycaemic (ob/ob) mice by food intake and the fatty acid composition of the diet. *Biochemical Journal*. 206:561–570.
- Escher, P., O. Braissant, S. Basu-Modak, L. Michalik, W. Wahli and B. Desvergne. (2001). Rat PPARs: quantitative analysis in adult rat tissues and regulation in fasting and refeeding. *Endocrinology*. 142:4195-202.
- Evans, M., J. Brown and M. McIntosh. (2002). Isomer-specific effects of conjugated linoleic acid (CLA) on adiposity and lipid metabolism. *Journal of Nutrition and Biochemistry*. 13:508-516.
- Fajas, L., D. Auboeuf and E. Raspe. (1997). The organization, promoter analysis, and expression of the human *PPAR* gene. *Journal of Biological Chemistry*. 272:18779–18789.
- Fiatmark, T., A. Nilsson, J. Krannes, T.S. Eikhom and M.H. Fukami. (1988). On the mechanism of induction of the enzyme systems for peroxisomal β -oxidation of fatty acids in rat liver by diets rich in partially hydrogenated fish oil. *Biochimica et Biophysica Acta*. 962:122-130.
- Field, C.J. (2003). Fatty acids/dietary importance.in : Encyclopaedia of food sciences and nutrition (edited by B. Caballero).Vol. 4. Pp. 2317-2324.Oxford,UK: Elsevier Sciences Ltd.
- Field, C.J., A.R. Edmond, A.B.R. Thompson and M.T. Clandinin. (1990). Diet fat composition alters membrane phospholipid composition, insulin binding, and

- glucose metabolism in adipocytes from control and diabetic animals. *Journal of Biological Chemistry*. 19:11143-11150.
- Fogerty, A.C., G.L. Ford and D. Svoronos. (1988). Octadeca-9,11-dienoic acid in foodstuffs and in the lipids of human blood and breast milk. *Nutrition Reports International*. 38:937–944.
- Folch, J., M. Lees and G.H. Sloane-Stanley. (1957). A simple method for the isolation and purification of total lipids from animal tissues. *Journal of Biological Chemistry*. 226:497-509.
- Forman, B., J. Chen and R.M. Evans. (1997). Hypolipidemic drugs, polyunsaturated fatty acids, and eicosanoids are ligands for peroxisome proliferator-activated receptors α and δ . *Proceedings of the National Academy of Sciences of the United States of America*. 94:4312–4317.
- Freeman, C.P. (1984). Digestion, absorption and transport of fats-non ruminants. In "Fats in animal nutrition". Ed. J. Wiseman, Proceedings of the 37th nottingham easter school. Butterworths, London, UK. chapter 5.
- Frenkel, B., N. Mayorek, R. Hertz and J. Bar-Tana. (1988). The hypocholesterolemic effect of beta, beta'-methyl-substituted hexadecanedioic acid (MEDICA 16) is mediated by a decrease in apolipoprotein C-III. *Journal of Biological Chemistry*. 263:8491–8497.
- Fritsche J. and H. Steinhart. (1998a). Analysis, occurrence, and physiological properties of trans fatty acids (TFA) with particular emphasis on conjugated linoleic acid isomers (CLA) – a review. *Fett/Lipid*. 100: 190-210.
- Fritsche, J. and C. Steinhart. (1998b). Amounts of conjugated linoleic acid (CLA) in German foods and evaluation of daily intake. *Zeitschrift für Lebensmitteluntersuchung und -Forschung A*. 206:77-82.
- Furuse, M., A. Murai, K. Kita, K. Asakura and J. Okumura. (1991). Lipogenesis depending on sexual maturity in female Japanese quail (*Coturnix coturnix*). *Comparative Biochemistry and Physiology*. 100B:343-345.
- Garg, M.L., E. Sebokoba, A.A. Wierzbicki, A.B.R Thomson, M.T. Clandin. (1988). Differential effects of dietary linoleic acid and α -linolenic acid in rat tissue. *Lipids*. 23:847–852.
- Gatlin, L.A., M. T. See, D.K. Larick, X. Lin and J. Odle. (2002). Conjugated linoleic acid in combination with supplemental dietary fat alters pork fat quality. *Journal of Nutrition*. 132:3105–3112.

- Gebauer, S.K., T.L. Psota, W.S. Harris and P.M. Kris-Etherton. (2006). n-3 fatty acid dietary recommendations and food sources to achieve essentiality and cardiovascular benefits. *American Journal of Clinical Nutrition*. 83(6 Suppl):1526S-1535S.
- Gonzalez-Esquerra, R. and S. Leeson. (2001). Alternatives for enrichment of eggs and chicken meat with n-3 fatty acids. *Canadian Journal of Animal Science*. 81: 295–305.
- Goodridge, A.G. (1968). Lipolysis in vitro in adipose tissue from embryonic and growing chicks. *American Journal of Physiology*. 214: 902-918.
- Goodridge, A.G., J.F. Crish, D.A. Fantozzi, M.J. Glynias, F.B. Hillgartner, S.A. Klautsky, X.J. Ma, D.A. Mitchell and L.M. Salati. (1989). Regulation of avian genes involved in fatty acid synthesis. *Journal of Animal Science*. 66:(Suppl. 3) 49.
- Gottlicher, M., A. Demoz, D. Svensson, P. Tollet, R.K. Berge and J.Å. Gustafsson. (1993). Structural and metabolic requirements for activators of the peroxisome proliferator-activated receptor. *Biochemical Pharmacology*. 46:2177-2184.
- Grashorn, M.A. (2007). Functionality of Poultry Meat. *Journal of Applied Poultry Research*. 16:99–106.
- Green S. (1992). Receptor-mediated mechanism of peroxisome proliferators. *Biochemical Pharmacology*. 43:393-401.
- Griffin, H.D. and D. Hermier. (1998). Plasma lipoprotein metabolism and fattening on poultry. In *Leanness in Domestic Birds*. B. Leclercq, and C.C. Whitehead ed. Butterworths, London. Pp:175–201.
- Griffin, H.D., K. Guo, D. Windsor and S.C. Butterwith. (1992). Adipose tissue lipogenesis and fat deposition in leaner broiler chickens. *Journal of Nutrition*. 122:363–368.
- Griffin, H.D., D. Windsor and C.C. Whitehead. (1991). Changes in lipoprotein metabolism and body composition in chickens in response to divergent selection for plasma very low density lipoprotein concentration. *British Poultry Science*. 32:195-201.
- Griinari, J.M. and D.E. Bauman. (1999). Biosynthesis of conjugated linoleic acid and its incorporation into meat and milk in ruminants. In: *Advances in Conjugated Linoleic Acid Research*, M.P. Yurawecz, M.M. Mossoba, J.K.G. Kramer, M.W. Pariza and G.J. Nelson, ed. AOCS Press, Champaign, IL. pp. 180-200.
- Griinari, J.M., B.A. Corl, S.H. Lacy, P.Y. Chouinard, K.V. Nurmela and D.E. Bauman. (2000). Conjugated linoleic acid is synthesized endogenously in lactating dairy cows by Delta(9)-desaturase. *Journal of Nutrition*. 130:2285-2291.

- Griminger, P. (1986). Lipid metabolism. In 'Avian Physiology' 4th edn. Ed. Sturkie P.D. Springer-Verlag, New York. pp:345-358.
- Grunder, A.A., J.R. Chambers and A. Fortin. (1987). Plasma very low density lipoproteins, abdominal fat lipase, and fatness during rearing in two strains of broiler chickens. *Poultry Science*. 66:471-479.
- Guan. Y., Y. Zhang, L. Davis and M.D. Breyer. (1997). Expression of peroxisome proliferator-activated receptors in urinary tract of rabbits and humans. *American Journal of Physiology*. 273:F1013-F1022.
- Gunstone, F.D., Harwood, J.L. and Padley, F.B. (Eds.) (1994) The Lipid Handbook, Second Edition. 722 p. Chapman and Hall, London.
- Ha, Y.L., N.K. Grimm and M.W. Pariza. (1987). Anticarcinogens from fried ground beef: heat-altered derivatives of linoleic acid. *Carcinogenesis*. 8:1881-1887.
- Ha, Y.L., N.K. Grimm and M.W. Pariza. (1989). Newly recognized anticarcinogenic fatty acids: identification and quantification in natural and processed cheeses. *Journal of Agriculture and Food Chemistry*. 37:75-81.
- Han, K.L., J.S. Choi, J.Y. Lee, J. Song, M.K. Joe, M.H. Jung and J.K. Hwang. (2008). Therapeutic Potential of Peroxisome Proliferators-Activated Receptor-alpha/gamma Dual Agonist With Alleviation of Endoplasmic Reticulum Stress for the Treatment of Diabetes. *Diabetes*. 57:737-745.
- Harden, R.L. and T.P. Oscar. (1993). Thyroid hormones and growth hormone regulation of broiler adipocyte lipolysis. *Poultry Science*. 72:669-676.
- Hargis, P.S., C.E. Dean and B.M. Hargis. (1991). *In ovo* endocrine regulation of growth. *British Reviews in Poultry Biology*. 3:307-323.
- Hargrave, K.M., M.J. Azain and J.L. Miner. (2005). Dietary coconut oil increases conjugated linoleic acid-induced body fat loss in mice independent of essential fatty acid deficiency. *Biochimica et Biophysica Acta*. 1737:52-60.
- Harris, W.S., W.C. Poston and C.K. Haddock. (2007). Tissue n-3 and n-6 fatty acids and risk for coronary heart disease events. *Atherosclerosis*. 193:1-10.
- Hellerstein, M.K., S.N. Meydani, M. Meydani, K. Wu and C.A. Dinarello. (1989). Interleukin-1- induced anorexia in the rat. Influence of prostaglandins. *Journal of Clinical Investigation*. 84: 228-235.
- Hertzel, A.V. and D.A. Bernlohr. (2000). The mammalian fatty acid-binding protein multigene family: molecular and genetic insights into function. *Trends in Endocrinology and Metabolism*. 11:175-180.

- Hermier, D. (1997). Lipoprotein Metabolism and Fattening in Poultry. *Journal of Nutrition*. 127: 805S–808S.
- Hojo, M., I. Takada, W. Kimura, K. Fukuda, S. Yasugi. (2006). Expression patterns of the chicken preoxisome proliferator-activated receptors (PPARs) during the development of digestive organs. *Gene Expression Patterns*. 6: 171-179.
- Holmer, G. and J.L. Beare-Rogers. (1985). Linseed oil and marine oil as sources (n-3) fatty acids in rat heart. *Nutrition Research*. 5:101 I-1014.
- Houseknecht, K.L., J.P. Vanden Heuvel, S.Y. Moya-Camarena, C.P. Portocarrero, L.W. Peck, K.P. Nickel and M.A. Belury. (1998). Dietary conjugated linoleic acid normalizes impaired glucose tolerance in the Zucker diabetic fatty fa/fa rat. *Biochemical and Biophysical Research Communications*. 244:678–682.
- Hua, X., J. Wu, J.L. Goldstein, M.S. Brown, H.H. Hobbs. (1995). Structure of the human gene encoding sterol regulatory element binding protein-1 (SREBF1) and localization of SREBF1 and SREBF2 to chromosomes 17p11.2 and 22q13. *Genomics*. 25:667–673.
- Huang, Z.B., R.G. Ackman, W.M.N. Ratnayake and F.G. Proudfoot. (1990). Effect of dietary fish oil on n-3 fatty acid levels in chicken eggs and thigh flesh. *Journal of Agriculture and Food Chemistry*. 38:743–747.
- Hulan, H.W., R.G. Ackman, W.M.N. Ratnayake, and F.G. Proudfoot. (1988). Omega-3 fatty acid levels and performance of broilers chickens fed redfish meal or redfish oil. *Canadian Journal of Animal Science*. 68:533–547.
- Hulan, H.W., F.G. Proudfoot and D.M. Nash. (1984). The effects of different dietary fat sources on general performance and carcass fatty acid composition of broiler chickens. *Poultry Science*. 63:324–332.
- Hur, S.J., H.S. Yang, G.B. Park and S.T. Joo. (2007). Effects of dietary glycine betaine on pork quality in different muscle types. *Asian Australian Journal of Animal Science*. 20:1754-1760.
- Hwang, D. (2000). Fatty acids and immune responses-a new perspective in searching for clues to mechanism. *Annual Review of Nutrition*. 20:431–456.
- Ip, C., M. Singh, H.J. Thompson and J.A. Scimeca. (1994). Conjugated linoleic acid suppresses mammary carcinogenesis and proliferative activity of the mammary gland in the rat. *Cancer Research*. 54:1212–1215.
- Issemann, I. and S. Green. (1990). Activation of a member of the steroid hormone receptor superfamily by peroxisome proliferators. *Nature*. 347:645-650.

- Javadi, M., J.H. Math, G.H. Everts, R. Hovenier, S. Javadi, H Kappert and A.C. Beynen. (2007). Effect of dietary conjugated linoleic acid on body composition and energy balance in broiler chickens. *British Journal of Nutrition*. 98:1152–1158.
- Jensen, L.S., G.W. Schumaier and J.D. Latshaw. (1970). “Extra caloric” effect of dietary fat for developing turkeys as influenced by calorie-protein ratio. *Poultry Science*. 49:1697-1704.
- Jiang, J., A. Wolk and B. Vessby. (1999). Relation between the intake of milk fat and the occurrence of conjugated linoleic acid in human adipose tissue. *American Journal of Clinical Nutrition*. 70:21–27.
- Johnson, P. T., and C. Sanders. (Eds.) (1994). Nutrition. Cincinnati: RNF Publications.
- Joo, S.T., J.I. Lee, Y.L. Ha and G.B. Park. (2002). Effects of dietary conjugated linoleic acid on fatty acid composition, lipid oxidation, color, and water-holding capacity of pork loin. *Journal of Animal Science*. 80:108–112.
- Jump, D.B., D. Botolin, Y. Wang, J. Xu, B. Christian and O. Demeure. (2005). Fatty acid regulation of hepatic gene transcription. *Journal of Nutrition*. 135:2503-2506.
- Jump, D.B., S.D. Clarke, A. Thelen and N. Liimatta. (1994). Coordinate regulation of glycolytic and lipogenic gene expression by polyunsaturated fatty acids. *Journal of Lipid Research*. 35:1076-1084.
- Kamphuis, M.M., M.P. Lejeune, W.H. Saris and M.S. Westerterp-Plantenga. (2003a). Effect of conjugated linoleic acid supplementation after weight loss on appetite and food intake in overweight subjects. *European Journal of Clinical Nutrition*. 57:1268-1274.
- Kamphuis, M.M., M.P. Lejeune, W.H. Saris and M.S. Westerterp-Plantenga. (2003b). The effect of conjugated linoleic acid supplementation after weight loss on body weight regain, body composition, and resting metabolic rate in overweight subjects. *International Journal of Obesity and Related Metabolic Disorders: Journal of the International Association for the Study of Obesity*. 27:840–847.
- Kang, K.R., G. Cherian and J.S. Sim. (2001). Dietary palm oil alters the lipid stability of polyunsaturated fatty acid-modified poultry products. *Poultry Science*. 80:228–234.
- Katsurada, A., N. Intani, H. Fukuda, Y. Matsumura and N. Nishimoto. (1990). Effects of nutrients and hormones on transcriptional and post-transcriptional regulation of acetyl-CoA carboxylase in rat liver. *European Journal of Biochemistry*. 190:435-441.

- Katsurada, A., N. Intani, H. Fukuda, T. Noguchi and T. Tanaka. (1987). Influence of diet on the transcriptional and post-transcriptional regulation of malic enzyme induction in the rat liver. *European Journal of Biochemistry*. 168:487-492.
- Kawahara, S., S. Takenoyama, K. Takuma, M. Muguruma and K. Yamauchi. (2009). Effects of dietary supplementation with conjugated linoleic acid on fatty acid composition and lipid oxidation in chicken breast meat. *Animal Science Journal*. 468–474.
- Keller, H., C. Deyer, J. Medin, A. Nahfoudi, K. Ozato, and W. Wahli. (1993). Fatty acids and retinoids control lipid metabolism through activation of peroxisome proliferator-activated receptor-retinoid X receptor heterodimers. *Proceedings of the National Academy of Sciences of the United States of America*. 90:2160-2164.
- Kennedy, S.R. (2007). Bioactive fatty acids as dietary supplements for farmed fish: effects on growth performance, lipid metabolism, gene expression and immune parameters. Ph. D. Thesis, University of Stirling, Scotland.
- Ketels E, G. DeGroot G. (1989). Effect of ratio of unsaturated to saturated fatty acids of the dietary lipid fraction on utilization and metabolizable energy of added fats in young chicks. *Poultry Science*. 68:1506-1512.
- Kepler, C.R., K.P. Hirons, J.J. McNeill and S.B. Tove. (1966). Intermediates and products of the biohydrogenation of linoleic acid by *Butyrivibrio fibrisolvens*. *Journal of Biological Chemistry*. 241:1350-1354.
- Kersten, S. (2005). Regulation of lipid metabolism via angiopoietin-like proteins. *Biochemical Society Transactions*. 33:1059-1062.
- Kersten, S. (2002). Effects of fatty acids on gene expression: role of peroxisome proliferator-activated receptor α , liver X receptor α and sterol regulatory element-binding protein-1c. *Proceedings of the Nutrition Society*. 61:371–374.
- Khanal, R.C. and T.R. Dhiman. (2004). Biosynthesis of conjugated linoleic acid (CLA): A review. *Pakistan Journal of Nutrition*. 3:72-81.
- Kim, H.Y. (2007). Novel metabolism of docosahexaenoic acid in neural cells. *Journal of Biological Chemistry*. 282:18661–18665.
- Kim, H., M. Haluzik, Z. Asghar, D. Yau, J.W. Joseph, A.M. Fernandez, M.L. Reitman, S. Yakar, B. Stannard, L. Heron-Milhavet and M.B. Wheeler. (2003). Peroxisome proliferator-activated receptor- α agonist treatment in a transgenic model of type 2 diabetes reverses the lipotoxic state and improves glucose homeostasis. *Diabetes*. 52:1770-1778.

- Klasing, K.C. (1998). Nutritional modulation of resistance to infectious diseases. *Poultry Science*. 77:1119-1125.
- Kliwer, S.A., S.S. Sundseth, S.A. Jones, P.J. Brown, G.B. Wisely, C.S. Koble, P. Devchand, W. Wahli, T.M. Wilson, J.M. Lenhard and J.M. Lehmann. (1997). Fatty acids and eicosanoids regulate gene expression through direct interactions with peroxisome proliferator-activated receptors alpha and gamma. *Proceedings of the National Academy of Sciences of the United States of America*. 94:4318–4323.
- Knight, T.W., S. Knowles, A.F. Death, J. West, M. Agnew, C.A. Morris and R.W. Purchas, (2003). Factors affecting the variation in fatty acid concentrations in lean beef from grass-fed cattle in New Zealand and the implications for human health. *New Zealand Journal of Agricultural Research*. 46:83-95.
- Komprda, T., J. Zelenka, E. Fajmonova, M. Fialova, and D. Kladroba. (2005). Arachidonic acid and long-chain n-3 polyunsaturated fatty acid contents in meat of selected poultry and fish species in relation to dietary fat sources. *Journal of Agriculture and Food Chemistry*. 53:6804- 6812.
- Konig, B., K. Kluge, K. Haase, C. Brandsh, G.I. Stangle, K. Eder. (2007a). Effects of Clofibrate Treatment in laing Hens. *Poultry Science*. 86:1187-1195.
- Konig, B, A. Koch, J. Spielmann, C. Hilgenfeld, G.I. Stangl, and, K. Eder. (2007b). Activation of PPAR α lowers synthesis and concentration of cholesterol by reduction of nuclear SREBP-2. *Biochemical Pharmacology*. 73:574–585.
- Korver, D.R. and K.C. Klasing. (1997). Dietary fish oil alters specific and inflammatory immune responses in chicks. *Journal of Nutrition*. 127: 2039-20-46.
- Kramer, J.K.G., N. Sehat, M.E.R. Dugan, M.M. Mossoba, M.P.Yurawecz, J.A.G. Roach. (1998). Distributions of conjugated linoleic acid (CLA) isomers in tissue lipid classes of pigs fed a commercial CLA mixture determined by gas chromatography and silver ion-high-performance liquid chromatography. *Lipids*. 33:549–558.
- Krogdahl, A. (1985). Digestion and absorption of lipids in poultry. *Journal of Nutrition*. 115:675–685.
- Krogdahl, A., and J.L. Sell. (1989). Influence of age on lipase, amylase, and protease activities in pancreatic tissue and intestinal contents of young turkeys. *Poultry Science*. 68:1561–1568.
- Langslow, D.R. and C.N. Hales. (1969). Lipolysis in chicken adipose tissue *in vitro*. *Journal of Endocrinology*. 43:285-294.

- Larsen, T.M., S. Toubro, O. Gudmundsen and A. Astrup. (2006). Conjugated linoleic acid supplementation for 1 y does not prevent weight or body fat regain. *The American Journal of Clinical Nutrition*. 83:606–612.
- Law, R.E., S. Goetze and X.P. Xi, S. Jackson, Y. Kawano, L. Demer, M.C. Fishbein, W.P. Meehan, W.A. Hsueh. (2000). Expression and function of PPAR γ in rat and human vascular smooth muscle cells. *Circulation*. 101:1311-1318.
- Lawson R.E., A.R. Moss, D.I. Givens. (2001). The role of dairy products in supplying conjugated linoleic acid to man's diet: a review. *Nutrition Research Reviews*. 14: 153-172.
- Leaflet, A.S. (2004). Dietary Conjugated Linoleic Acid (CLA) Effects Lipid Metabolism in Broiler Chicks R1934 Iowa State University Animal Industry Report.
- Lee, K.N., M.W. Pariza, and J.M. Ntambi. (1998). Conjugated linoleic acid decreases hepatic stearoyl-CoA desaturase mRNA expression. *Biochemical and Biophysical Research Communications*. 248:817–821.
- Lehmann, J.M., L.B. Morre, and T.A. Smith-Oliver, W.O. Wilkison, T.M. Willson, S.A. Kliewer. (1995). An antidiabetic thiazolidinedione is a high affinity ligand for peroxisome proliferator-activated receptor γ (PPAR γ). *Journal of Biological Chemistry*. 270:12953–12956.
- Lemberger. T., B. Desvergne, and W. Wahli. (1996). Peroxisome proliferator-activated receptors: a nuclear Receptor signaling sathway in lipid physiology. *Annual Review of Cell and Developmental Biology*. 12:335–63.
- Leskanich, C.O. and R.C. Noble. (1997). Manipulation of the n-3 polyunsaturated 343 fatty acid composition of avian eggs and meat. *World Poultry Science Journal*. 53:155-183.
- Lessire, M., M. Doreau, and A. Aumaitre. (1996). Digestive and metabolic utilization of fats in domestic animals. Pages 703--713 in *Oils and fats manual*, edited by A. Karleskind. Lavoisier Publishing, Paris, France.
- Lessire, M., and B. Leclercq. (1982). Metabolisable energy value of fats in chicks and adult cockerels. *Animal Feed Science and Technology*. 7:365–374.
- Leveille, G. A., D.R. Romsos, Y. Yeh and E.K. O'Hea. (1975). Lipid Biosynthesis in the Chick. A Consideration of Site of Synthesis, Influence of Diet and Possible Regulatory Mechanisms. *Poultry Science*. 54:1075-1093.
- Li, Y., B.A. and Watkins. (1998). Conjugated linoleic acids alter bone fatty acid composition and reduce ex vivo prostaglandin E2 biosynthesis in rats fed n-6 or n-3 fatty acids. *Lipids*. 33:417–425.

- Lin, Y., A. Kreeft, J.A.E. Schuurbijs and R. Draijer. (2001). Different effects of conjugated linoleic acid isomers on lipoprotein lipase activity in 3T3-L1 adipocytes. *Journal of Nutrition and Biochemistry*. 12:183–189.
- Lin, Q. S.E. Ruuska, N.S. Shaw, D. Dong and N. Noy. (1999). Ligand selectivity of the peroxisome proliferator-activated receptor alpha. *Biochemistry*. 38:185–190.
- Livak, K.J., and T.D. Schmittengen. (2001). Methods. Analysis of relative gene expression data using realtime quantitative PCR and the $2^{-\Delta\Delta C(T)}$. *Methods*. 25: 402–408.
- Lock, A.B., A.M. Mitchell and C.R. Elcombe. (1989). Biochemical mechanism of induction of hepatic peroxisome proliferation. *Annual Review of Pharmacology and Toxicology*. 29:145- 63.
- Lopez-Ferrer, S., M.D. Baucells, A.C. Barroeta and M.A. Grashorn. (2001). n-3 enrichment of chicken meat. 1. Use of very long-chain fatty acids in chicken diets and their influence on meat quality: fish oil. *Poultry Science*. 80:741-732.
- Lopes-Virella, M.F., P. Stone, S. Ellis, J.A. Colwell. (1977). Cholesterol determination in high-density lipoproteins separated by three different methods. *Clinical Chemistry*. 23:882-884.
- Luo, J., S.W. Rizkalla, J. Boillot, C. Alamowitch, H. Chaib, F. Bruzzo, N. Desplanque, A.M. Dalix, G. Durand and G. Slama. (1996). Dietary (n-3) polyunsaturated fatty acids improve adipocyte insulin action and glucose metabolism in insulinresistant rats: relation to membrane fatty acids. *Journal of Nutrition*. 126:1951-1958.
- Luquet, S., J. Lopez-Soriano, D. Holst, C. Gaudel, C. Jehl-Pietri, A. Fredenrich, and P.A. Grimaldi. (2004). Roles of peroxisome proliferator-activated receptor delta (PPARdelta) in the control of fatty acid catabolism. A new target for the treatment of metabolic syndrome. *Biochimie*. 86:833–837.
- Madsen, L., R.K. Petersen and K. Kristiansen. (2005). Regulation of adipocyte differentiation and function by polyunsaturated fatty acids. *Biochimica et Biophysica Acta*. 1740:266– 286.
- Madsen, L., A.C. Rustan, H. Vaagenes, K. Berge, H. Dyroy, R.K. Berge. (1999). Eicosapentaenoic and docosahexaenoic acid affect mitochondrial and peroxisomal fatty acid oxidation in relation to substrate preference. *Lipids*. 34:951–963.
- Mandard, S., M. Müller and S. Kersten. (2004). Peroxisome proliferator-activated receptor alpha target genes. *Cellular and Molecular Life Sciences*. 61:393–416.

- Manilla, H., A.F. Husveth, and K. Nemeth. (1999). Effects of dietary fat origin on the performance of broiler chickens and composition of selected tissues. *Acta Agrar. Kaposváriensis*. 3:47–57.
- Martin, J.C., S. Grégoire, M.H. Siess, M. Genty, J.M. Chardigny, O. Berdeaux, P. Juanéda and J.L. Sébédio. (2000). Effects of conjugated linoleic acid isomers on lipid-metabolizing enzymes in male rats. *Lipids*. 35:91-98.
- Matsubara, Y., K. Sato, H. Ishii, and Y. Akiba. (2005). Changes in mRNA expression of regulatory factors involved in adipocyte differentiation during fatty acid induced adipogenesis in chicken. *Comparative Biochemistry and Physiology - Part A*. 141:108–115.
- McCue, M.D., O. Amitai, I. Khozin-Goldberg, S.R. McWilliams and B. Pinshow. (2009). Effect of dietary fatty acid composition on fatty acid profiles of polar and neutral lipid tissue fractions in zebra finches, *Taeniopygia guttata*, *Comparative Biochemistry and Physiology - Part A*. 154: 165–172.
- Meng, H., H. Li, J.G. Zhao and Z.L. Gu. (2005). Differential expression of peroxisome proliferator-activated receptors alpha and gamma gene in various chicken tissues. *Domestic Animal Endocrinology*. 28:105-110.
- Mersmann, H.J. (2002). Mechanisms for conjugated linoleic acid-mediated reduction in fat deposition. *Journal of Animal Science*. 80:E126-E134.
- Mezentseva, N.V., J.S. Kumaratilake., S.T. Newman. (2008). The brown adipocyte differentiation pathway in birds: An evolutionary road not taken. *BMC Biology*. 6:17.
- Mikkelsen, L., H.S. Hansen, N. Grunnet and J. Dich. (1993). Inhibition of fatty acid synthesis in rat hepatocytes by exogenous polyunsaturated fatty acids is caused by lipid peroxidation. *Biochimica et Biophysica Acta*. 1166: 99-104.
- Miller, C.C., Y. Park, M.W. Pariza and M.E. Cook. (1994). Feeding conjugated linoleic acid to animals partially overcomes catabolic response due to endotoxin injection. *Biochemical and Biophysical Research Communications*. 198:1107-1112.
- Mohrhauer, H., K. Christiansen, M.V. Gan, M. Deubig, R.T. Holman. (1967). Chain elongation of linoleic acid and its inhibition by other fatty acids in vitro. *The Journal of Biological Chemistry*. 242:4507–14.
- Moore, S.A., E. Hurt, E. Yoder, H. Sprecher, and A.A. Spector. (1995). Docosahexaenoic acid synthesis in human skin fibroblasts involves peroxisomal retroconversion of tetracosahexaenoic acid. *Journal of Lipid Research*. 36:2433–2443.

- Mosley, E.E., M.K. McGuire, J.E. Williams and M.A. McGuire. (2006a). *cis-9, trans-11* conjugated linoleic acid is synthesized from vaccenic acid in lactating women. *Journal of Nutrition*. 136:2297-2301.
- Mosley, E.E., B. Shafi Dagger, P.J. Moate and M.A. McGuire. (2006b). *cis-9, trans-11* conjugated linoleic acid is synthesized directly from vaccenic acid in lactating dairy cattle. *Journal of Nutrition*. 136:570-575.
- Moya-Camarena, S.Y. and M.A. Belury. (1999a). CLA and PPAR γ activation. *Journal of Nutrition*. 129:602-606.
- Moya-Camarena, S.Y., J.P.V. Heuvel and S.G. Blanchard. (1999b). Conjugated linoleic acid is a potent naturally occurring ligand and activator of PPAR α . *Journal of Lipid Research*. 40:1426–1433.
- Mukherjee, R., L. Jow, G.E. Croston and Jr. Paterniti. (1997). Identification, characterization, and tissue distribution of human peroxisome proliferator-activated receptor (PPAR) isoforms PPAR γ 2 versus PPAR γ 1 and activation with retinoid X receptor agonists and antagonists. *Journal of Biological Chemistry*. 272:8071–8076.
- Nagao, K., N. Inoue, Y.M. Wang, B. Shirouchi and T. Yanagita. (2005). Dietary conjugated linoleic acid alleviates nonalcoholic fatty liver disease in Zucker (fa/fa) rats. *Journal of Nutrition*. 135:9-13.
- Nagao, K., N. Inoue, Y.M. Wang and T. Yanagita. (2003). Conjugated linoleic acid enhances plasma adiponectin level and alleviates hyperinsulinemia and hypertension in Zucker diabetic fatty (fa/fa) rats. *Biochemical and Biophysical Research Communications*. 310:562-566.
- Nakatani, Y., Kaneto, H., Kawamori, D., Yoshiuchi, K., Hatazaki, M., Matsuoka, T.A., Ozawa, K., Ogawa, S., Hori, M., Yamasaki, Y., et al. 2005. Involvement of endoplasmic reticulum stress in insulin resistance and diabetes. *Journal of Biological Chemistry*. 280: 847–851.
- Narciso-Gaytan, C., D. Shin, A.R. Sams, C.A. Bailey, R.K. Miller, S.B. Smith, O.R. Leyva-Ovalle and M.X. Sanchez-Plata. (2010). Soybean, palm kernel, and animal-vegetable oils and vitamin E supplementation effect on lipid oxidation stability of sous vide chicken meat. *Poultry Science*. 89 :721–728
- Nash, D.M., R.M.G. Hamilton and H.W. Hulan. (1995). The effect of dietary herring meal on the omega-3 fatty acid content of plasma and egg yolk lipids of laying hens. *Canadian Journal of Animal Science*. 75:247–253.

- Nehlig, A., M.C. Crone, and P.R. Lehr. (1980). Variations of β -hydroxybutyrate dehydrogenase activity in brain and liver mitochondria of the developing chick. *Biochimica et Biophysica Acta*. 633:22-32.
- Nestel, P.J. (1990). Effects of n-3 fatty acids on lipid metabolism. *Annual Review of Nutrition*. 10:149-67.
- Newman, R.E. (2000). Modulation of avian metabolism by dietary fatty acids. PhD. Thesis, University of Sydney, Australia.
- Newman, R.E., W.L. Bryden, E. Fleck, J.R. Ashes, W.A. Buttemer, L.H. Storlien and J.A. Downing. (2002) Dietary n-3 and n-6 fatty acids alter avian metabolism: metabolism and abdominal fat deposition. *British Journal of Nutrition*. 88:11–18.
- Nicolosi, R.J., E.J. Rogers, D. Kritchevsky, J.A. Scimeca, P.J. Huth. (1997). Dietary conjugated linoleic acid reduces plasma lipoproteins and early aortic atherosclerosis in hypercholesterolemic hamsters. *Artery* 22:266–277.
- Nir, I. and H. Lin. (1982). The skeleton, an important site of lipogenesis in the chick. *Annales de la nutrition et de l'alimentation*. 26:100-105.
- Noy, Y., and D. Sklan. (1995). Digestion and absorption in the young chick. *Poultry Science*. 74:366–373.
- NRC, 1994. Nutrient Requirements of Poultry. National Academy Press, Washington, D. C. USA.
- Ntambi, J.M. (1999). Regulation of stearoyl-CoA desaturase by polyunsaturated fatty acids and cholesterol. *Journal of Lipid Research*. 40:1549–1558.
- Ntambi, J.M. (1991). Dietary regulation of stearoyl-CoA desaturase I gene expression in mouse liver. *Journal of Biological Chemistry*. 267:10925-10930.
- O'Hagan, S., A. Menzel. (2003). A subchronic 90-day oral rat toxicity study and in vitro genotoxicity studies with a conjugated linoleic acid product. *Food and Chemical Toxicology: An International Journal Published for the British Industrial Biological Research Association*. 41:1749–1760.
- Ohnuki, K., S. Haramizu, K. Ishihara and T. Fushiki. (2001). Increased energy metabolism and suppressed body fat accumulation in mice by a low concentration of conjugated linoleic acid. *Bioscience, Biotechnology, and Biochemistry*. 65:2200-2204.
- Ojano-Dirain, C., M. Toyomizu, T. Wing, M. Cooper, W.G. Bottje. (2007). Gene Expression in Breast Muscle and Duodenum from Low and High Feed Efficient Broilers. *Poultry Science*. 86:372–381.

- Oscar T.P. (1995). Lipid mobilization from chicken fat cells. In "The Biology of Fat in Meat Animals Current Advances". Eds. S.B. Smith & D.R. Smith. American Society of Animal Science, Champaign, Illinois.
- Palmer, C.N.A., M.H. Hsu and K.J. Griffin, J.L. Raucy and E.F. Johnson. (1998). Peroxisome proliferator activated receptor- α expression in human liver. *Molecular Pharmacology*. 53:14–22.
- Pariza, M.W., Y. Park and M.E. Cook. (2000). Mechanisms of action of conjugated linoleic acid: evidence and speculation. *Proceedings of the Society for Experimental Biology and Medicine*. 223:8–13.
- Pariza, M.W., Y. Park, and M.E. Cook. (2001). The biologically active isomers of conjugated linoleic acid. *Progress in Lipid Research*. 40:283–298.
- Pariza, M.W. (2004). Perspective on the safety and effectiveness of conjugated linoleic acid. *The American Journal of Clinical Nutrition*. 79:1132S–1136S.
- Pariza, M.W. and W.A. Hargraves. (1985). A beef-derived mutagenesis modulator inhibits initiation of mouse epidermal tumors by 7, 12-dimethylbenz[a]anthracene. *Carcinogenesis*. 6:591-593.
- Pariza, M.W., S.H. Ashoor, F.S. Chu and D.B. Lund. (1979). Effects of temperature and time on mutagen formation in pan-fried hamburger. *Cancer Letters*. 7:63-69.
- Park, Y., K. J. Albright, W. Liu, J. M. Storkson, M. E. Cook, and M. W. Pariza. (1997). Effect of conjugated linoleic acid on body composition in mice. *Lipids* 32: 853-858.
- Park, Y., K.J. Albright and M.W. Pariza. (2005). Effects of conjugated linoleic acid on long term feeding in Fischer 344 rats. *Food and Chemical Toxicology: An International Journal Published for the British Industrial Biological Research Association*. 43:1273–1279.
- Park, Y. and M.W. Pariza. (2007). Mechanisms of body fat modulation by conjugated linoleic acid (CLA). *Food Research International*. 40:311–323.
- Park, Y., J.M. Storkson, W. Liu, K.J. Albright, M.E. Cook and M.W. Pariza. (2004). Structure-activity relationship of conjugated linoleic acid and its cognates in inhibiting heparin-releasable lipoprotein lipase and glycerol release from fully differentiated 3T3-L1 adipocytes. *Journal of Nutritional Biochemistry*. 15:561-568.
- Park, Y., J.M. Storkson, J.M. Ntambi, M.E. Cook, C.J. Sih, M.W. Pariza. (2000). Inhibition of hepatic stearyl-CoA desaturase activity by trans-10, cis-12 conjugated linoleic acid and its derivatives. *Biochimica et Biophysica Acta*. 1486:285–292.

- Patsouris, D. (2006). Transcriptional regulation of nutrient A2 p2. PhD Thesis. University Wageningen, Netherlands.
- Pearce J. (1980). A comparison of the response of hepatic enzyme activity in domestic fowl (*Gallus domesticus*) and the rat (*Rattus norvegicus*) to the feeding of diets containing large proportions of glucose and fructose. *Comparative Biochemistry and Physiology*. 651B: 423-426.
- Peters, J.M., Y. Park, F.J. Gonzalez and M.W. Pariza, M.W. (2001). Influence of Conjugated Linoleic Acid on Body Composition and Target Gene Expression in Peroxisome-Proliferator-Activated Receptor α -Null Mice. *Biochimica et Biophysica Acta*. 1533:233-241.
- Phetteplace, H.W. and B.A. Watkins. (1989). Effects of various n-3 sources on fatty acid composition in chicken tissues. *Journal of Food Composition and Analysis*. 2:104-117.
- Phetteplace, H. W. and B.A. Watkins. (1990). Lipid measurements in chickens fed different combinations of chicken fat and menhaden oil. *Journal of Agriculture and Food Chemistry*. 38:1848-1853.
- Picard, F. and J. Auwerx. (2002). PPAR(γ) and glucose homeostasis. *Annual Review of Nutrition*. 22:167-197.
- Pineda Torra, I., P. Gervois and B. Staels. (1999). Peroxisome proliferator activated receptor alpha in metabolic disease, inflammation, atherosclerosis and aging. *Current Opinion in Lipidology*. 10:151-159.
- Piperova, L.S., B.B. Teter, I. Bruckental, J. Sampugna, S.E. Mills, M.P. Yurawecz, J. Fritsche, K. Ku and R.A. Erdman. (2000). Mammary lipogenic enzyme activity, *trans* fatty acids and conjugated linoleic acids are altered in lactating dairy cows fed a milk fat-depressing diet. *Journal of Nutrition*. 130:2568-2574.
- Pisulewski, P.M. (2005). Nutritional potential for improving meat quality in poultry. *Animal Science Papers and Reports*. 23:303-315.
- Pisulewski, P.M. and R.B. Kostogrys. (2003). Functional properties of foods of animal origin and methods of their assessment. *Polish Journal of Food and Nutrition Sciences*. 53:65-73.
- Pisulewski, P.M., B. Szymczyk and R.B. Kostogrys. (2002). Health-promoting properties of conjugated linoleic acid isomers and perspectives of production of CLA-enriched foods of animal origin. In Polish, summary in English. *Żywnieć Człowieka XXIX*: 87-103.

- Polin, D. and T.H. Hussein. (1982). The effect of bile acid on lipid and nitrogen retention, carcass composition, and dietary metabolizable energy in very young chicks. *Poultry Science*. 61:1697–1707.
- Poureslami, R., G.M. Turchini, K. Raes, G. Huyghebaert and S. De Smet. (2010). Effect of diet, sex and age on fatty acid metabolism in broiler chickens: SFA and MUFA. *British Journal of Nutrition*. 104:204–213.
- Power, G.W. and E.A. Newsholme. (1997). Dietary fatty acids influence the activity and metabolic control of mitochondrial carnitine palmitoyltransferase I in rat heart and skeletal muscle. *Journal of Nutrition*. 127:2142–2150.
- Purushotham, A., G.E. Shrode, A.A. Wendel, L.F. Liu and M.A. Belury. (2007). Conjugated linoleic acid does not reduce body fat but decreases hepatic steatosis in adult Wistar rats. *Journal of Nutritional Biochemistry*. 18:676- 684.
- Raes K., S. De Smet and D. Demeyer. (2004). Effects of dietary fatty acids on incorporation of long chain polyunsaturated fatty acids and conjugated linoleic acid in lamb, beef and pork meat. *Animal Feed Science Technology*. 113:199-221.
- Ramenofsky, M. (1990). Fat storage and fat metabolism in relation to migration. In *Bird Migration: Physiology and Ecophysiology* (ed. E. Gwinner), pp. 214-231. New York: Springer-Verlag.
- Ramsay, T.G., C.M. Evock-Clover, N.C. Steele, M.J. Azain, (2001). Dietary conjugated linoleic acid alters fatty acid composition of pig skeletal muscle and fat. *Journal of Animal Science*. 79:2152–2161.
- Ratnayake, W.M.N., R.G. Ackman, and H.W. Hulan. (1989). Effect of redfish meal enriched diets on the taste and the n- 3 PUFA of 42-day-old broiler chickens. *Journal of the Science of Food and Agriculture*. 49:59–74.
- Rosen, E.D., P. Sarraf and A.E. Troy, G. Bradwin, K. Moore, D.S. Milstone, B.M. Spiegelman, R.M. Mortensen. (1999). PPAR gamma is required for the differentiation of adipose tissue in vivo and in vitro. *Molecular Cell*. 4:611–617.
- Roche, H.M. (2005). Fatty acids and the metabolic syndrome. *Proceedings Nutrition Society Journal*. 64:23-29.
- Roche, H.M., E. Noone, A. Nugent, M.J. Gibney, M.J. (2001). Conjugated linoleic acid: a novel therapeutic nutrient? *Nutrition Research Review*. 14:173-187.
- Russo, G.L. (2009). Dietary n - 6 and n - 3 polyunsaturated fatty acids: From biochemistry to clinical implications in cardiovascular prevention. *Biochemical Pharmacology*. 77:937-946.

- Rustan, A.C., E.N. Christiansen and C.A. Drevon. (1992). Serum lipids, hepatic glycerolipid metabolism and peroxisomal fatty acid oxidation in rats fed n-3 and n-6 fatty acids. *Biochemistry Journal* . 283:333-39.
- Rustan A.C and C.A. Drevon. (2005). Fatty Acids: Structures and Properties. Encyclopedia of life sciences. John Wiley & Sons, Ltd, New York.
- Rustan, A.C., J.Ø. Nossen, H. Osmundsen and C.A. Drevon. (1988). Eicosapentaenoic acid inhibits cholesterol esterification in cultured parenchymal cells and isolated microsomes from rat liver. *Journal of Biological Chemistry*. 263:8126-8132.
- Ruyter, B., O. Andersen, A. Dehli, A.K. Ostlund Farrants, T. Gjøen and M.S. Thomassen. (1997). Peroxisome proliferator activated receptors in Atlantic salmon (*Salmo salar*): effects on PPAR transcription and acyl-CoA oxidase activity in hepatocytes by peroxisome proliferators and fatty acids. *Biochimica et Biophysica Acta*. 1348:331-338.
- Rymer, C. and D.I. Givens. (2005). n-3 Fatty acid enrichment of edible tissues of poultry: a review. *Lipids*. 40:121-130.
- Saebø, A. (2003). Commercial synthesis of CLA. Pages 71–81 in Advances in Conjugated Linoleic Acid Research. Volume 2. J. L. Sebe'dio, W. W. Christie and R. Adlof, ed. AOCS Press, Champaign.
- Sanz, M., C.J. Lopez-Bote, D. Menoyo, J.M. Bautista. (2000). Abdominal fat deposition and fatty acid synthesis are lower and β -oxidation is higher in broiler chickens fed diets containing unsaturated rather than saturated fat. *Journal of Nutrition*. 130:3034–3037.
- Sampath, H. and J.M. Ntambi. (2005). Polyunsaturated fatty acid regulation of genes of lipid metabolism. *Annual Review of Nutrition*. 25:317–40.
- Sampath, H. and J.M. Ntambi. (2006). regulation of gene expression by polyunsaturated fatty acids. *Heart Metabolism*. 32:32–35.
- Santora, J.E., D.L. Palmquist and K.L. Roehrig. (2000). Trans-vaccenic acid is desaturated to conjugated linoleic acid in mice. *Journal of Nutrition*. 130:208-215.
- Sargent, J.R., D.R. Tocher and J.G. Bell.(2002). The lipids. pp. 181-257. In: Fish Nutrition. Edited by J. E. Halver and R. W. Hardy. Academic Press, San Diego.
- SAS software version 9.1. (2002). SAS Institute Inc., Cary, NC, USA.

- Sato, K. and Y. Akiba. (2002). Lipoprotein lipase mRNA expression in abdominal adipose tissue is little modified by age and nutritional state in broiler chickens, *Poultry Science*. 81:846–852.
- Sato, K., Y. Akiba, Y. Chida and K. Takahashi. (1999). Lipoprotein hydrolysis and fat accumulation in chicken adipose tissues are reduced by chronic administration of lipoprotein lipase monoclonal antibodies. *Poultry Science*. 78:1286–1291.
- Sato, K, Fukao, K. Seki, Y and Akiba, Y. (2004). Expression of the Chicken Peroxisome Proliferator-Activated Receptor Gene Is Influenced by Aging, Nutrition, and Agonist Administration. *Poultry Science*. 83:1342–1347.
- Sato, O., C. Kuriki, Y. Fukui, K. Motojima. (2002). Dual promoter structure of mouse and human fatty acid translocase/cd36 genes and unique transcriptional activation by peroxisome proliferator-activated receptor alpha and gamma ligands. *Journal of Biological Chemistry*. 277:15703–15711.
- Sato, K., K. Matsushita, Y. Matsubara, T. Kamada, Y. Akiba. (2008). Adipose Tissue Fat Accumulation Is Reduced by a Single Intraperitoneal Injection of Peroxisome Proliferator-Activated Receptor Gamma Agonist When Given to Newly Hatched Chicks. *Poultry Science*. 87:2281-2286.
- Sato, K., H. Abe, T. Kono, M. Yamazaki, K. Nakashima, T. Kamada, Y. Akiba.(2009). Changes in peroxisome proliferator-activated receptor gamma gene expression of chicken abdominal adipose tissue with different age, sex and genotype. *Animal Science Journal*. 80: 322-327.
- Scanes, C.G. (1987). The physiology of growth, growth hormone, and other growth factors in poultry. *Critical Reviews in Poultry Science*. 1:51-105.
- Scanes, C.G. (1995). Adipose tissue and its hormonal control in poultry. In “The Biology of Fat in Meat Animals Current Advances”. Eds. S.B. Smith & D.R. Smith. American Society of Animal Science, Champaign, Illinois.
- Schmidt, A., N. Endo, S.J. Rutledge, R. Vogel, D. Shinar, G.A. Rodan. (1992). Identification of a new member of the steroid hormone receptor superfamily that is activated by a peroxisome proliferator and fatty acids. *Molecular Endocrinology*. 6:1634-1641.
- Schmalz, K., P. Kerrigan. (2003). Lipid review and teaching tips in nutrition. *California Journal of Health Promotion*. 1:164-173.
- Schoonjans. K, J. Peinado-Onsurbe and A.M. Lefebvre. (1996a). PPARalpha and PPARgamma activators direct a distinct tissue-specific transcriptional response via a PPRE in the lipoprotein lipase gene. *EMBO Journal*. 15:5336–5348.

- Schoonjans, K., B. Staels and J. Auwerx. (1996b). The peroxisome proliferator-activated receptors (PPARs) and their effects on lipid metabolism and adipocyte differentiation. *Biochimica et Biophysica Acta*.1302:93–109.
- Schoonjans, K., B. Staels, and J. Auwerx. (1996c). Role of the peroxisome proliferator-activated receptor (PPAR) in mediating the effects of fibrates and fatty acids on gene expression. *Journal of Lipid Research*. 37:907–925.
- Schwartz, R.S. and S. Abraham. (1983). Effect of dietary fat on the activity, content, rates of synthesis and degradation, and translation of mRNA coding malic enzyme. *Archives of Biochemistry and Biophysics*. 221:206-215.
- Sealls, W., M. Gonzalez, M.J. Brosnan, P.N. Black, C.C. DiRusso. (2008). Dietary polyunsaturated fatty acids (C18:2 omega6 and C18:3 omega3) do not suppress hepatic lipogenesis. *Biochimica et Biophysica Acta*. 1781: 406-614.
- Sell, J. L., S. Jin, and M. Jeffrey. (2001). Metabolizable energy value of conjugated linoleic acid for broiler chicks and laying hens. *Poultry Science*. 80:209–214.
- Senköylü, N. (2001). Feed fats. *University of Trakya, Faculty of Agriculture. Tekirdag*. 21: 247-254.
- Shearer, B.G. and W.J. Hoekstra. (2003). Recent advances in peroxisome proliferator-activated receptor science. *Current Medicinal Chemistry*. 10:267-280.
- Sher, T., H.F. Yi, O.W. McBride AND F.J. Gonzalez. (1993). cDNA cloning, chromosomal mapping, and functional characterization of the human peroxisome proliferator activated receptor. *Biochemistry*. 32:5598-5604.
- Shimomura, Y., T. Tamura and M. Suzuki. (1990). Less body fat accumulation in rats fed a sunflower oil diet than in rats fed a beef tallow diet. *Journal of Nutrition*. 120:1291-1296.
- Simon, O., K. Manner, K. Schafer, A. Sagredos, K. Eder. (2000). Effects of conjugated linoleic acids on protein to fat proportions, fatty acids, and plasma lipids in broilers. *European Journal of Lipid Science and Technology*. 102:402–410.
- Simopoulos, A.P. (1999). Essential fatty acids in health and chronic disease. *American Journal of Nutrition*.70:560S-569S.
- Simopoulos, A,P. (2002). Omega-3 fatty acids in inflammation and autoimmune diseases. *Journal of American College of Nutrition*. 21:495-505.
- Sirri, F., G. Minelli, N. Iaffaldano, N. Tallarico, and A. Franchini. (2003). Oxidative stability and quality traits of n-3 PUFA enriched chicken meats. *Italian Journal of Animal Science*. 2:450–452.

- Sklan, D. (1976). Digestion and absorption of lipids in chicks fed triglycerides or free fatty acids: synthesis of monoglycerides in the intestine. *Poultry Science*. 58:885-897.
- Sklan, D., P. Budowski, I. Ascarelli, and S. Hurwitz. (1973). Lipid absorption and secretion in the chick: Effect of raw soybean meal. *Journal of Nutrition*. 103:1299–1305.
- Smink, W., W.J.J. Gerrits, R. Hovenier, M.J.H. Geelen, H.W.J. Lobee, M.W.A. Verstegen and A.C. Beynen. (2008). Fatty acid digestion and deposition in broiler chickens fed diets containing either native or randomized palm oil. *Poultry Science*. 87:506–513.
- Song, H.J., A.A. Sneddon, P.A. Barker, C. Bestwick, S.N. Choe, S. McClinton, I. Grant, D. Rotondo, S.D. Heys, K.W. Wahle. (2004). Conjugated linoleic Acid inhibits proliferation and modulates protein kinase C isoforms in human prostate cancer cells. *Nutrition and Cancer*. 49:100–108.
- Spiegelman, B.M., E. Hu, J.B. Kim and R. Burn. (1997). PPAR γ and the control of adipogenesis. *Biochimie*. 79: 111–112.
- Stangl G.I., H. Muller and M. Kirchgessner. (1999). Conjugated linoleic acid effects on circulating hormones, metabolites and lipoproteins, and its proportion in fasting serum and erythrocyte membranes of swine. *European Journal of Nutrition*. 38:271–277.
- Stevens, L. (1996). *Avian Biochemistry and Molecular Biology*. University Press, Cambridge, U.K.
- Storkson, J.M., Y. Park, M.E. Cook, M.W. Pariza. (2005). Effects of trans-10,cis-12 conjugated linoleic acid (CLA) and cognates on apolipoprotein B secretion in HepG2 cells. *Nutrition Research*. 25:387–399.
- Suksombat, W., T. Boonmee, and P. Lounglawan. (2007). Effects of Various Levels of Conjugated Linoleic Acid Supplementation on Fatty Acid Content and Carcass Composition of Broilers. *Poultry Science*. 86:318–324.
- Szymczyk, B., and P.M. Pisulewski. (2003). Effects of dietary conjugated linoleic acid on fatty acid composition and cholesterol content of hen egg yolks. *British Journal of Nutrition*. 90:93–99.
- Szymczyk, B., P.M. Pisulewski, W. Szczurek and P. Hanczakowski. (2001). Effects of conjugated linoleic acid on growth performance, feed conversion efficiency and subsequent carcass quality in broiler chickens. *British Journal of Nutrition*. 85:465-473.

- Takahashi, K., K. Kawamata, Y. Akiba, T. Iwata and M. Kasai. (2003). Effect of a mixture of conjugated linoleic acid isomers on growth performance and antibody production in broiler chicks. *British Journal of Nutrition*. 89:691-694.
- Takeuchi, H., T. Matsuo, K. Tokuyama, Y. Shimomura, M. Suzuki. (1995). Diet-induced thermogenesis is lower in rats fed a lard diet than in those fed a high oleic acid safflower oil diet, a safflower oil diet of a linseed oil diet. *Journal of Nutrition*. 125: 920–925.
- Tebbey, P.W. and T.M. Buttke. (1992). Stearoyl-CoA desaturase gene expression in lymphocytes. *Biochemical and Biophysical Research Communications*. 186:531-536.
- Terpstra, A.H., A.C. Beynen, H. Everts, S. Kocsis, M.B. Katan and P.L. Zock. (2002). The decrease in body fat in mice fed conjugated linoleic acid is due to increases in energy expenditure and energy loss in excreta. *Journal of Nutrition*. 132:940-945.
- Thiel-Cooper, R.L., F.C. Parrish, J.C. Sparks, B.R. Wiegand and R.C. Ewan. (2001). Conjugated linoleic acid changes swine performance and carcass composition. *Journal of Animal Science*. 79:1821-182.
- Tontonoz, P., E. Hu, R.A. Graves, A.I. Budavari and B.M. Spiegelman. (1994a). mPPAR- γ 2: tissue specific regulator of an adipocyte enhancer. *Genes & Development*. 8: 1224-1234.
- Tontonoz, P., E. Hu, and B.M. Spiegelman. (1994b). Stimulation of adipogenesis in fibroblasts by PPAR- γ 2 a lipid-activated transcription factor. *Cell*. 79:1147-1156.
- Tontonoz, P., J.B. Kim, R.A. Graves, B.M. Spiegelman. (1993). ADD1: a novel helix-loop-helix transcription factor associated with adipocyte determination and differentiation. *Molecular and Cellular Biology*. 13:4753–4759.
- Trugo, N.M.F. and A.G. Torress. (2003). Fats/requirements . In: Encyclopaedia of food sciences and nutrition (edited by B. Caballero). Second Ed. pp. 2280-2281. Oxford, UK: Elsevier Sciences Ltd.
- Tsuboyama-Kasaoka, N., M. Takahashi, K. Tanemura, H. J. Kim, T. Tange, H. Okuyama, M. Kasai, S. Ikemoto, and O. Ezaki. 2000. Conjugated linoleic acid supplementation reduces adipose tissue by apoptosis and develops lipodystrophy in mice. *Diabetes*. 49:1534–1542.
- Tuncer, S.D., R. Asti, B. Coskun, M.A. Tekes and H. Erer. (1987). The effects of different energy sources on fattening performance, abdomen fat accumulation and liver fat in broiler I. The effects of fattening performance and abdomen fat accumulation. *University of Selçuk. Journal of Veterinary Faculty*. 3:25-40.

- Turpeinen, A.M., M. Mutanen, A. Aro, I. Salminen, S. Basu, D.L. Palmquist and J.M. Griinari. (2002). Bioconversion of vaccenic acid to conjugated linoleic acid in humans. *American Journal of Clinical Nutrition*. 76:504-510.
- Uauy, R. and A. Valenzuela. (2000). Marine oils: The health benefits of n-3 fatty acids. *Nutrition*. 16: 680-684.
- United States Department of Agriculture. (1988). Food and the consumer. National Food Review. Washington, DC; United States Department of Agriculture, Economic Research series.
- Vaille, K., D. Grippo, M.F. Blouquit, M. Souidi, M. Riottot, J.C. Bouthegourd, C. Serougne, J.C. Martin. (2004). Lipid atherogenic risk markers can be more favourably influenced by the cis-9,trans-11-octadecadienoate isomer than a conjugated linoleic acid mixture or fish oil in hamsters. *The British Journal of Nutrition*. 91:191–199.
- Vermeersch, G. and F. Vanschoubroek. (1968). The quantification of the effect of increasing levels of various fats on body weight gain, efficiency of food conversion and food intake of growing chicks. *British Poultry Science*. 9:13-20.
- Villalba, P.G., A.M. Jimenez-Lara, and A. Aranda, A. (1996). Vitamin D interferes with transactivation of the growth hormone gene by thyroid hormone and retinoic acid. *Molecular and Cellular Biology*. 16:318–327.
- Von Schacky, C. and W.S. Harris, (2007). Cardiovascular benefits of omega-3 fatty acids. *Cardiovascular Research*. 73:310-315.
- Wahle, K.W J., S.D. Heys and D. Rotondo. (2004). Conjugated linoleic acids: are the beneficial or detrimental to health? *Progress in Lipid Research*. 43:553-587.
- Wahli, W., O. Braissant and B. Desvergne. (1995). Peroxisome proliferator-activated receptors: transcriptional regulators of adipogenesis, lipid metabolism and more. *Chemistry & Biology*. 2:261–266.
- Wakil, S.J., J.K. Stoops and V.C. Joshi. (1983). Fatty acid synthesis and its regulation. *Annual Review of Biochemistry*. 52:537-539.
- Wall, R., R.P. Ross., G.F. Fitzgerald and C. Stanton. (2010). Fatty acids from fish: the anti-inflammatory potential of long-chain omega-3 fatty acids. *Nutrition Reviews*. 68:280–289.
- Wan, J.B., L.L. Huang, R. Rong, R. Tan, J. Wang and J.X. Kang. (2010). Endogenously decreasing tissue n-6/n-3 fatty acid ratio reduces atherosclerotic lesions in apolipoprotein E^{-/-} deficient mice by inhibiting systemic and vascular inflammation. *Arterioscler Thromb Vasc Biol*. 30:2487-2494

- Wanders R.J.A., C. Jacobs and O.H. Skjeldal. (2001). Refsum disease, in *The Metabolic and Molecular Bases of Inherited Disease* (Scriver, C. R., Beaudet, A. L., Sly, W. S. and Valle, D., eds.), 8th edn., pp. 3303±3321. McGraw-Hill, New York.
- Wang, Y., Y. Mu, H. Li, N. Ding, Q. Wang, Y. Wang, S. Wang, and N. Wang. (2008). Peroxisome proliferator-activated receptor-gamma gene: A key regulator of adipocyte differentiation in chickens. *Poultry Science*. 87:226–232.
- Wang, Y.M., K. Nagao, Y. Ujino, K. Sakata, K. Higa, N. Inoue and T. Yanagita. (2005). Short-term feeding of conjugated linoleic acid does not induce hepatic steatosis in c57BL-6J mice. *Journal of Nutritional Science and Vitaminology*. 51:440-444.
- Watkins, B.A. (1995). Biochemical and physiological aspects of polyunsaturates. *Poultry and Avian boilogy Reviews*. 6: 1-18.
- West, D.B., F.Y. Blohm, A.A. Truett and J.P. DeLany. (2000). Conjugated linoleic acid persistently increases total energy expenditure in AKR/J mice without increasing uncoupling protein gene expression. *Journal of Nutrition*. 130:2471-2477.
- Wilson, M.D., W.L. Blake, L.M. Salati, S.D. Clarke. (1990). Potency of polyunsaturated and saturated fats as short-term inhibitors of hepatic lipogenesis in rats. *Journal of Nutrition*. 120:544–552.
- Wilson, M.D., R.D. Hays and S.D. Clarke. (1986). Inhibition of liver lipogenesis by dietary polyunsaturated fat in severely diabetic rats. *Journal of Nutrition*. 116:1511–1518.
- Wiseman, J. (1984). Assessment of the digestible and metabolisable energy of fats for non-ruminants. In "Fats In Animal Nutrition". Ed. J. Wiseman, Proceedings of the 37th Nottingham Easter School. Butterworths, London, UK. Chapter 14.
- Wu. Z., Y. Xie, R.F. Morrison, N.L. Bucher, S.R. Farmer. (1998). PPARgamma induces the insulin-dependent glucose transporter GLUT4 in the absence of C/EBPalpha during the conversion of 3T3 fibroblasts into adipocytes. *The Journal of Clinical Investigation*. 101:22–32.
- Xu, H.E., M.H. Lambert and V.G. Montana. (1999). Molecular recognition of fatty acids by peroxisome proliferator-activated receptors. *Molecular Cell*. 3:397–403.
- Yamasaki, M., A. Ikeda, A. Hirao, Y. Tanaka, Y. Miyazaki, T. Rikimaru, M. Shimada, K. Sugimachi, H. Tachibana and K. Yamada. (2001). Effect of dietary conjugated linoleic acid on the *in vivo* growth of rat hepatoma dRLh-84. *Nutrition and Cancer*. 40:140-148.

- Yanagita, T., Y.M. Wang, K. Nagao, Y. Ujino, N. Inoue. (2005). Conjugated linoleic acid-induced fatty liver can be attenuated by combination with docosahexaenoic acid in C57BL/6N mice. *Journal of Agricultural and Food Chemistry*. 53:9629–9633.
- Yu, K., W. Bayona and C.B. Kallen. (1995). Differential activation of peroxisome proliferator-activated receptors by eicosanoids. *Journal of Biological Chemistry*. 270: 23975–23983.
- Zanini, S.F., G.L. Colnago, B.M.S. Pessotti, M.R. Bastos, F.P. Casagrande and V.R. Lima. (2006). Body Fat of Broiler Chickens Fed Diets with Two Fat Sources and Conjugated Linoleic Acid. *International Journal of Poultry Science*. 5: 241-246.
- Zanini, S.F., E. Vicente, G.L. Colnago, B.M.S. Pessotti, M.A. Silva. (2008). Manipulation of the fatty acids composition of poultry meat and giblets by dietary inclusion of two oil sources and conjugated linoleic acid. *Arquivo Brasileiro de Medicina Veterinaria e Zootecnia*. 60:1388-1398.
- Zerehdaran, S., A.L.J. Vereijken, J.A.M. van Arendonk and E.H. van der Waaij. (2004). Estimation of Genetic Parameters for Fat Deposition and Carcass Traits in Broilers. *Poultry Science*. 83:521-525.
- Zhang, H., Y. Guo and J. Yuan. (2005). Conjugated linoleic acid enhanced the immune function in broiler chicks. *British Journal of Nutrition*. 94:746-752.
- Zhang, H.J., Y.M. Guo, Y. Yang, J.M. Yuan. (2006). Dietary conjugated linoleic acid enhances spleen PPAR- γ mRNA expression in broiler chicks. *British Poultry Science*. 47:726-733.
- Zhao, A., J. Yu, J.L. Lew, L. Huang, S.D. Wright and J. Cui. (2004). Polyunsaturated fatty acids are FXR ligands and differentially regulate expression of FXR targets. *DNA and Cell Biology*. 23:519-526.