



UNIVERSITI PUTRA MALAYSIA

***EQUINE CHORIONIC GONADOTROPHIN INFLUENCES ON
FOLLICULAR DEVELOPMENT, OVULATION RATE AND GESTATIONAL
DEVELOPMENT IN CROSS-BRED GOATS***

MOHAMAD SHUIB MOHAMAD MOHTAR

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MASTER OF SCIENCE

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By

MOHAMAD SHUIB MOHAMAD MOHTAR

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Master of Science**

July 2012

DEDICATION

I dedicate this thesis to my inspiring mom, Noriah Abu Hassan and dad, Mohd Mohtar Yusup, who wanted me to be a role model in the family, and now your wishes come true.

And to my wife, Suhaini Mohd Salleh and precious children, Nor Fatiha Insyirah & Muhamad Faris Amsyar

May this thesis be your inspiration for your future endeavors...

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

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July 2012

Chair: Professor Abd Wahid Haron, PhD

Faculty: Veterinary Medicine

Superovulation techniques are usually conducted to enhance ovarian follicular activity and subsequently increase reproductive efficiency in goat breeding. This study was carried out to depict the effects of a superovulation technique using equine chorionic gonadotrophin (eCG) treatment on increasing follicular growth, ovulation rate and numbers of kid born. In this study, 29 non-pregnant pluriparous Boer does weighing between 45 and 65 kg, with good body condition were randomly selected and allocated into 3 groups, namely Group 1 (G1, n=11), Group 2 (G2, n=8) and Group 3 (G3, n=10). All does were synchronized using CIDR containing 0.3 g progesterone intravaginally for 18 days. Blood samples from every doe were

collected from the jugular vein using test tubes containing heparin during CIDR treatment (Day 0, 6, 13, 18) and post CIDR treatment (Day 19, 20, 27, 34, 41 and 48). On Day 17 of the CIDR treatment, superovulation treatment using eCG was given intramuscularly using three different treatment dosages. Does in G1, G2 and G3 received 600 IU, 800 IU and 1000 IU eCG, respectively on the same days. Follicular identification on the left and right ovary was conducted using transrectal ultrasonography technique for 4 consecutive days starting on Day 17 of the CIDR treatment. After CIDR removal, oestrus responses in all does were recorded and natural mating was conducted. After 7 days post-mating, the transrectal ultrasonography technique was applied to identify the numbers of corpora lutea present in each side of the ovary. Subsequently, pregnancy diagnosis was carried out using transrectal ultrasonography technique at 30 days post-mating. Measurements of sizes of placentomes were carried out using transrectal ultrasonography on Day 42, 49, 56 and 63 of pregnancy. Data of does kidded were collected after the pregnancy period. Result of the study showed that the range of percentage of does showing oestrus within 48 h after CIDR removal in the three groups was 62.5% to 81.8%. The G1 does (54.5%) had a significantly higher ($P < 0.05$) compared to G2 (25%) and G3 (20%) oestrus response within 24 h after CIDR removal. No significant differences among treatments were observed in terms of the number of follicles developed except for small sized follicles on Day 19 and 20. Meanwhile, the G2 does had the highest ovulation rate ($P < 0.05$) of 2.38 ± 0.32 CL compared to G3 (1.6 ± 0.22 CL) and G1 (1.45 ± 0.15 CL). The average pregnancy rate showed 11 out of 21 oestrus does were pregnant (52.4%) with G3 does (71.4%) having the highest pregnancy rate

compared to the other groups at 55.5% (G1) and 20% (G2). Based on progesterone concentration analysis, all treatments does except unsuccessful mated does in G3 showed increasing level of progesterone starting on Day 0 (CIDR inserted) until next 6 days. After Day 6, the progesterone level for all treatments does have continuously decline until Day 18 (CIDR removal), 19 and lastly Day 20. Only after Day 20, the progesterone level was tremendously increased and maintains above 20 ng/ml until 30 days pregnancy in successful mated does. Whereas, for unsuccessful mated does their progesterone level was peak on Day 34 before decline lower than 5 ng/ml until Day 48. Significant differences in the progesterone level were observed between pregnant and non-pregnant does at Day 41 pregnancies for G1 and at Day 48 for G2 and G3 does post-CIDR insertion. No significant differences were observed among treatments in terms of the size of placentomes (in early stage of pregnancy), kidding rate and gestation length. In conclusion, superovulation by eCG with a long term progesterone treatment in goats has increased the ovulatory activity to produce more matured follicles, high oestrous response and twinning rate. However, this thesis also revealed that the growth of placentome and gestation length were not affected by the different dosages of eCG.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**PENGARUH GONADOTROPHIN KORIONIK EKUIN KEATAS
PERKEMBANGAN FOLIKEL, KADAR OVULASI DAN PEMBANGUNAN
KEBUNTINGAN PADA KAMBING KACUKAN**

Oleh

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Teknik superovulasi kebiasaannya dilaksana bagi meningkatkan aktiviti perkembangan folikel dan pada masa yang sama meningkatkan kecekapan pembiakan pada kambing. Kajian ini dilaksanakan bagi memperlihatkan kesan-kesan teknik superovulasi dengan menggunakan rawatan Gonadotrophin Korionik ekuin (eCG) keatas peningkatan pertumbuhan folikel, kadar ovulasi dan bilangan anak dilahirkan. Di dalam kajian ini, 29 ekor kambing betina “pluriparous” Boer dengan berat badan diantara 45 dan 65 kg, beserta kondisi badan yang baik dipilih secara rambang dan ditempatkan kepada 3 kumpulan, iaitu Kumpulan 1 (G1, n=11), Kumpulan 2 (G2, n=8) dan Kumpulan 3 (G3, n=10). Kesemua kambing telah

disinkroni menggunakan CIDR yang mengandungi 0.3 g progesteron ke dalam vagina selama 18 hari. Sampel darah daripada kambing akan diambil daripada vena jugular menggunakan tabung uji yang mengandungi heparin semasa rawatan CIDR (hari 0, 6, 13, 18) dan selepas rawatan CIDR (hari 19, 20, 27, 34, 41 dan 48). Pada hari 17 rawatan CIDR, rawatan superovulasi menggunakan eCG diberi melalui intra-otot menggunakan tiga rawatan dos yang berbeza. Kambing betina G1, G2 dan G3 masing-masing akan menerima 600 IU, 800 IU dan 1000 IU eCG pada hari yang sama. Pengenalpastian folikel di kiri dan kanan ovari telah dilaksanakan menggunakan teknik transrektum pengimbasan ultra bunyi pada 4 hari berturut-turut bermula pada hari 17 rawatan CIDR. Selepas CIDR dikeluarkan, reaksi estrus pada semua kambing betina direkod dan pembiakan semulajadi dilaksanakan. Tujuh hari selepas dikahwinkan, teknik transrektum pengimbasan ultra bunyi telah digunakan bagi mengenalpasti bilangan korpus luteum yang hadir di setiap belah ovari. Berikutnya, diagnosa kebuntingan telah dilaksanakan menggunakan teknik transrektum pengimbasan ultra bunyi pada hari ke 30 selepas dikahwinkan. Pengukuran saiz placentom telah dilaksanakan menggunakan teknik transrektum pengimbasan ultra bunyi pada hari 42, 49, 56 dan 63 kebuntingan. Data kambing betina beranak dikumpul selepas tempoh kebuntingan. Keputusan kajian menunjukkan julat peratusan kambing betina yang menunjukkan estrus dalam lingkungan 48 jam selepas CIDR dikeluarkan pada ketiga-tiga kumpulan ialah 62.5% hingga 81.8%. Kambing-kambing G1 (54.5%) mencatat peratus tertinggi yang ketara ($P < 0.05$) apabila dibandingkan dengan G2 (25%) dan G3 (20%) reaksi estrus dalam lingkungan 24 jam selepas CIDR dikeluarkan. Tiada perbezaan ketara diantara rawatan-rawatan

tersebut merujuk kepada bilangan folikel terhasil kecuali bagi saiz folikel kecil pada hari 19 dan 20. Sementara itu, kambing-kambing G2 mempunyai kadar ovulasi tertinggi ($P < 0.05$) iaitu 2.38 ± 0.32 CL berbanding G3 (1.6 ± 0.22) dan G1 (1.45 ± 0.15). Purata kadar kebuntingan menunjukkan 11 daripada 21 kambing yang estrus telah bunting (52.4%) dengan G3 (71.4%) mencatatkan kadar kebuntingan tertinggi berbanding kumpulan-kumpulan lain pada 55.5% (G1) dan 20% (G2). Berdasarkan analisa kepekatan progesteron, kesemua kambing terawat kecuali kambing yang tidak berjaya kahwin di G3 telah menunjukkan peningkatan paras progesteron bermula pada hari 0 (CIDR dimasukkan) sehingga 6 hari kemudian. Selepas hari 6, paras progesteron untuk kesemua kambing-kambing terawat telah turun berterusan sehingga hari 18 (CIDR dikeluarkan), 19, dan akhirnya hari 20. Selepas hanya hari 20, tahap progesteron telah mendadak naik dan kekal di atas 20 ng/ml sehingga 30 hari kebuntingan bagi kambing-kambing yang berjaya dikahwinkan. Disebalik itu, bagi kambing-kambing yang gagal dikahwinkan paras progesteron ia telah memuncak pada hari 34 sebelum turun serendah daripada 5 ng/ml sehingga hari 48. Tahap progesteron yang berbeza ketara telah dilihat diantara kambing bunting dan tidak bunting di hari 41 kebuntingan bagi G1 dan pada hari 48 bagi kambing-kambing G2 dan G3 selepas CIDR dimasukkan. Tiada perbezaan bererti dilihat diantara rawatan-rawatan pada saiz plasentom (pada peringkat awal kebuntingan), kadar kelahiran dan tempoh kebuntingan. Kesimpulannya, superovulasi menggunakan eCG bersama satu tempoh rawatan progesteron yang panjang pada kambing telah meningkatkan aktiviti ovari bagi menghasilkan lebih banyak folikel matang, reaksi estrus yang tinggi dan kadar kembar. Walau bagaimanapun, kajian ini

turut mendedahkan pertumbuhan plasentom dan tempoh kebuntingan adalah tidak dipengaruhi oleh perbezaan dos-dos eCG.



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I certify that a Thesis Examination Committee has met on 26 July 2012 to conduct the final examination of Mohamad Shuib Mohamad Mohtar on his thesis entitled “Equine Chorionic Gonadotrophin Influences on Follicular Development, Ovulation Rate and Gestational Development in Cross-bred Goats” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U. (A) 106] 15 March 1998. The Committee recommends that the student be awarded the degree of Master of Science.

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DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institutions.

MOHAMAD SHUIB MOHAMAD MOHTAR

Date: 26 July 2012.

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LIST OF ABBREVIATIONS

ADG	average daily gain
AI	artificial insemination
ANOVA	analysis of variance
AV	artificial vagina
BC	before century
BE	Bailey ejaculator
BSE	breeding soundness examination
CASA	computer assisted sperm analysis
CIDR	Controlled Internal Drug Release
CL	corpus luteum
CV	coefficient of variance
D	day
DVS	Department of Veterinary Services
eCG	equine Chorionic Gonadotrophin
EE	electro-ejaculation
ET	embryo transfer
FCR	feed conversion ratio
FELDA	Federal Land Development Authority
FGA	fluorogestone acetate
FSH	follicle stimulating hormone
FSH-P	follicle stimulating hormone based porcine origin
G	force of gravity
g	gram
GnRH	gonadotropin releasing hormone
h	hours

HAP	horse anterior pituitary extract
IVF	in vitro fertilization
i.m	intramuscular
LH	luteinizing hormone
LOPU	laparoscopic ovum pick-up
MHz	Megahertz
MOA	Ministry of Agriculture and Agro-based Industry of Malaysia
MOET	multiple ovulation and embryo transfer
MPA	medroxyprogesterone acetate
MT	metric tonne
NSB	non-specific binding
OPU	ovum pick-up
PASW [®]	predictive analytics software
PD	pregnancy diagnosis
PGF2 α	prostaglandin F2 α
PMSG	pregnant mare serum gonadotrophin
PS	placentome size
P ₄	progesterone
pFSH	follicle stimulating hormone based porcine origin
QC-M	quality control medium pool
RIA	radioimmunoassay
r	correlation
r ²	regression
S.E.M	standard error of the mean
μ g	microgram

CHAPTER 1

INTRODUCTION

The purpose of goat (*Capra hircus*) rearing is much the same as the purpose of other ruminant species, which is to take advantage of the available forage in order to produce products that humans can consume. Apart from meat and milk as the main products, goats also provide leather to produce shoes, bag, water containers besides wool for clothing, and dung as fertilizers and fuel. They can also be used as a means of entertainment and religious sacrifice. In recent years, various goat parts have been utilized by manufacturing and pharmaceutical companies for the production of gelatin, glue and other products. Exactly when the individual animal species were domesticated is unknown. However, the earliest domestic food species (as most Westerners currently define it) was sheep (somewhere around 8000 BC), followed closely by goats, hogs and cattle (6500 BC), ilama (4000 BC), donkey (3500 BC) and chickens (3200 BC) (Stephen, 2003). Only a few decades ago more attention has been given by scientists and animal producers to the goat species. As with all other species, the genetics and the environment of the goat determine its performance. It is the job of the person (the breeder) in charge of the selection program to combine the best set of genes available for the environment in which the animal will be produced. The challenge of the selection is to improve the genetic potential of the next generation. Improving the ability of a species to produce a needed product in an economically beneficial way requires more research. Selection and breeding

programs for goats can have rather narrow goals, such as high daily weight gain and improvement of quality of carcass.

For most commercial goat producers, crossbreeding is the method of choice for a breeding program. The reason is simply that crossbred goats perform better than purebred goats in meat production. A crossbreeding program for goats is fairly easy to design, and a good variety of productive, adaptable breeds of goats from which to select is available. Crossbreeding is practised when the benefits of heterosis are desired in the offspring. In goats, this approach generally involves crossing bucks of the meat-type sire breeds with does of the fecund-type dam breeds to produce kids with increased growth rate and carcass quality while benefiting from the productive rate and maternal influence of female parent (Shrestha and Fahmy, 2007). In Australia, Dhanda *et al.* (1999) concluded that although crossbreeds demonstrated an advantage in carcass characteristics, there was no important influence on meat quality and nutritious composition. In crossing between German Fawn and Katjang goats in Malaysia, Hirooka *et al.* (1997) reported significant direct and maternal additive genetic effects for body weights at birth, 6 and 9 months of age, suggesting a large difference in the growth rate. Most crossbreeding is done within the confines of a system designed to maximize diverse and complementary traits in the individual parent breeds.

The Boer goat breed which excels in meat production was developed in the 1920s by South African farmers from the Eastern Cape when indigenous goats kept by the

Hottentot and Bantu tribe were crossed with imported Nubian and Indian goats (Skinner, 1972; Erasmus, 2000; Malan, 2000). If meat production is the goal and good Boer stock is available at reasonable prices, then crossbreeding and breeding-up could easily be practised to produce superior meat-type goats. These programs together with selection are required to genetically characterize local goat breeds as well as the exotic breeds for genetic improvement (Barilet, 2007; Fahmy and Shrestha, 2000; Shrestha and Fahmy, 2007).

The goat could only contribute approximately to 2,390 tonnes of meat compared with the consumption demand of 22,550 tonnes in Malaysia (Malaysia Production Agro-Food Statistics Books, MOA, 2010). The Boer goats were introduced into Malaysia in 2000 because this breed can adapt well to the hot and humid conditions. Boer goats have also a high average daily gain (ADG) at 147.1 g/day (Solaiman *et al.*, 2011) and low feed conversion ratio (FCR) compared with other breeds of goats. In addition, the percentage of meat is high with almost 54.1% meat obtained from the carcass (Solaiman *et al.*, 2011).

To increase productivity in goats, superovulation is one of the biotechnological tools which can be utilized to increase the number of kids born alive. Other approaches to multiply the number of kids born is through artificial insemination (AI), embryo transfer (ET) and in vitro fertilization (IVF). Superovulation permits multiple number of ovum release from the ovary, so that more ova can be fertilized. For superovulation, three kinds of hormones have been widely used. They are, equine

chorionic gonadotropin (eCG) formerly known as “pregnant mare serum gonadotropin” (PMSG), follicle stimulating hormone (FSH) and horse anterior pituitary (HAP) extract. These hormones are grouped as gonadotropins because they stimulate ovarian follicle growth. They differ among by their schedule of treatment, preparation and dosage regimens. Naqvi and Gulyani (1999) indicated that the different gonadotrophin regimens have variable effects on superovulation, ova recovery and the fertilization rate.

The combination of eCG and progestagen treatments eliminates the variability in the ovulatory response of goats both in the breeding season and in the seasonal anoestrus period. This resulted in an increased ovulation rate and fertility improvement (Van der Wethuysen, 1979; Ritar *et al.*, 1984). However, the dose of eCG that should be used to increase litter size during the breeding season is still not well defined (Regueiro *et al.*, 1999).

The present study aimed to compare the lower dosages of eCG at 600 IU and 800 IU compared to 1000 IU which eventually improve reproductive efficiency as well as reducing cost of superovulatory treatment in the goats.

Therefore, this study was conducted with the following objectives:

- i. to compare the effect of different dosages of eCG on oestrus response time.
- ii. to analyze the follicle development, ovulation and pregnancy rates.
- iii. to compare the size of placentomes, gestation length and kidding rate following different doses of eCG administration.
- iv. to compare the effectiveness of low dosages of eCG treatment on the number of kids born per doe.

There are several limitations to this study. They are:

- i. Small numbers of non-pregnant does with good body condition that were available at the farm.
- ii. Inconsistency of feeding times to the animals that produced bad effects to the mating season and fetal development in does.
- iii. Less experience on identification of corpora lutea during ultrasound scanning.

The hypothesis of this study is that lower dosages of eCG at 600 IU and 800 IU provide similar effectiveness of superovulation treatment to improve reproductive efficiency in the goats.

CHAPTER 7

BIBLIOGRAPHY

- Acharya, R. and Battacha, N. *Status of small ruminant production FAO round table on International cooperation on small ruminant research and development*. 5th International Conference on Goats, New Delhi, March. 2-8, 1992. pp. 7-43.
- Agga, G.E., Udala, U., Regassa, F. and Wudie, A. (2010). Body measurements of bucks of three breeds in Ethiopia and their correlation to breed, age and testicular measurements. *Small Ruminant Research*. 95, 2-3, 133-138.
- Akusu, M.O. and Egbunike, G.N. (1984). Fertility of the West-African dwarf goat in its native environment following prostaglandin F₂ α induced estrus. *Veterinary Quartely*. 6: 173-176.
- Ali, A. (2007). Effect of time of eCG administration on follicular response and reproductive performance of FGA-treated Ossimi ewes. *Small Ruminant Research*. 72: 33-37.
- Alice, F., Maria-Teresa Pellicer-Rubio. and Bernard, L. (2011). Reproductive cycle of goats. *Animal Reproduction Science*. 124: 211-219.
- Aly M. Karen., EL-Sayed M. Fattouh. and Saber S. Abu-Zeid. (2009). Estimation of gestational age in Egyptian native goats by ultrasonographic fetometry. *Animal Reproduction Science*. 114: 167-174.
- Al-Nakib, F., Lodge, G. and Oweh, J. (1986). A study of sexual development of ram lambs. *Journal of Animal Production*. 43: 459-468.
- Armstrong, D.T. and Evans, G. (1983a). Factors influencing success of embryo transfer in goats. *Theriogenology*, 19: 31-42.
- Armstrong, D.T., Pfitzner, A.P., Warnes, G.M., Ralph, M.M. and Seamark, R.F. (1983b). Endocrine responses of goats after induction of superovulation with PMSG and FSH. *Reproduction Fertility Journal*. 67: 395-401.
- Armstrong, D.T., Miller, B.G., Walton, E.A., Pfitzner, A.P. and Warnes, G.M. (1982). Endocrine response and factors which limit the response of follicles to PMSG and FSH. In Shelton, J.N., Trounson, A.O., Moore, N.W. and James, J.W. (eds.). *Embryo transfer in cattle, sheep and goats*. Australia Society Reproduction Biology Publication. pp 8-15.

- Baldassarre, H. and Karatzas, C.N. (2004). Advanced assisted reproduction technologies in goat. *Animal Reproduction Science*. 83: 255-266.
- Baril, G., Remy, B., Leboeuf, B., Beckers, J.F. and Saumande, J. (1996). Synchronization of oestrous in goats: The relationship between eCG binding in plasma, time of occurrence of oestrous and fertility following artificial insemination. *Theriogenology*. 45: 1553-1559.
- Baril, G. *Possibilidades atuais da transferencia de embriões em caprinos*. In: Proceeding of the XI Congresso Brasileiro de Reproducao Animal. (*Current possibilities of embryo transfer in goats*. In: Proceeding of the XI Brazilian Congress of Animal Reproduction) Belo Horizonte, Brazil, August. 13-18, 1995. pp 110-120.
- Baril, G., Brebion, P. and Chesne, P. (1993a). Manuel de formation pratique pour la transplantation embryonnaire chez la brebis et la chevre. Etude FAO: Production et santé animales. (Practical training manual for embryo transfer in sheep and goats. FAO: Animal Production and Health). FAO Education. ISBN: 92-5-203388-2. No 115. pp 196.
- Baril, G., Chemineau, P., Cognie, Y., Guerin, Y., Leboeuf, B., Orgeur, P. and Vallet, J.C. (1993b). Manuel de formation pour l'insemination artificielle chez les ovins et les caprins. Etude FAO: Production et santé animale (Manual of artificial insemination training in sheep and goats. FAO: Production and Animal Health). FAO Education. ISBN: 92-5-202808-0. No. 83. pp 258.
- Barilet, F. (2007). Genetic improvement for dairy production in sheep and goats. *Small Ruminant Research*. 70: 60-75.
- Baronet, D. and Vailancourt, D. (1989). Diagnostic de gestation par echotomographie chez la chevre. (Pregnancy diagnosis by ultrasonography in goats). *Medical Veterinary Quebec*. 19: 67-72.
- Barrett, D.M.W., Bartlewski, P.M., Batista-Arteaga, M., Symington, A. and Rawlings, N.C. (2004). Ultrasound and endocrine evaluation of the ovarian response to a single dose of 500 IU eCG following a 12-d treatment with progestagen-releasing intravaginal sponges in the breeding and non-breeding seasons in ewes. *Theriogenology*. 61: 311-327.
- Bartlewski, P.M., Beard, A.P., Cook, S.J., Chandolia, R.K. Honoramooz, A. and Rawlings, N.C. (1999a). Ovarian antral follicular dynamics and their relationship with endocrine variables throughout the oestros cycle in breeds of sheep differing in prolificacy. *Journal of Reproduction and Fertility*. 115: 111-124.

Biology online dictionary. Estrus definition. Retrieved 11 November 2011 from <http://www.biology-online.org/dictionary/estrus>

BonDurant, R.H. (1986). Embryo transfer in sheep and goats, In: Morrow, D.A. (Ed.), *Current Therapy in Theriogenology: Part 2*, Saunders, Philadelphia, PA, USA. pp 63-66.

Bongso, T.A., Fatima, I. and Dass, S. (1982). Synchronization of oestrus of goats treated with progestagen-impregnated intravaginal sponges and PMSG and reproductive performance following natural mating or AI with frozen semen. *Animal Reproduction Science*. 5: 111-116.

Bretzlaff, K.N. and Romano, J.E. (2001). Advanced reproductive techniques in goat. *Veterinary Clinical of North America: Food Animal Practice*. 17, 421-434.

Buckrell, B.C., Bonnett, B.N. and Johnson, W.H. (1986). The use of real-time ultrasound rectally for early pregnancy diagnosis in sheep. *Theriogenology*. 25: 665-673.

Cahill, L.P. (1982). Factors influencing the follicular response of animals to PMSG. In Shelton, J.N., Trounson, A.O., Moore, N.W. and James, J.W. (Eds.) *Embryo transfer in cattle, sheep and goats*. Australia Society Reproduction Biology Publication. pp 5-7.

Cardwell, B.E., Fitch, G.Q. and Geisert, D. (1998). Ultrasonic evaluation for the time of ovulation in ewes treated with norgestomet and norgestoment followed by pregnant mares serum gonadotropin. *Journal of Animal Science*. 76: 2235-2238.

Casey, N.H. and Van Niekerk, W.A. (1988). The Boer goat. I. Origin, adaptability, performance testing, reproduction and milk production. *Small Ruminant Research*. 1: 291-302.

Chacon, J., Perez, E., Muller, E., Soderquist, L. and Rodriguez-Martinez, H. (1999). Breeding soundness evaluation of extensively managed bulls in Costa Rica. *Theriogenology*, 52: 221-231.

Cognie, Y., Baril, G., Poulin, N. and Mermilod, P. (2003). Current status of embryo technologies in sheep and goats. *Theriogenology*, 59: 171-188.

Coulter, G.H. and Foote, R.H. (1979). Bovine testicular measurements as indicators of reproductive performance and their relationship to productive traits in cattle: A review. *Theriogenology*. 11: 297-311.

- Cox, C.F., Alfaro, V., Montenegro, V. and Rodriguez-Martinez, H. (2006). Computer-assisted analysis of sperm motion in goats and its relationship with sperm migration in cervical mucus. *Theriogenology*. 66: 860-867.
- Cumming, I.A. (1975). The ovine and bovine oestrous cycle. *Journal of Reproduction and Fertility*. 43: 583-596.
- Daudu, C. (1984). Spermatozoa output, testicular sperm reserve and epididymal storage capacity of the Red Sokoto Goats indigenous to Northern Nigeria. *Theriogenology*. 21: 317-324.
- Dawson, L.J., Sahlu, T., Hart, S.P., Detweiler, G., Gipson, T.A., The, T.H., Henry, G.A. and Bahr, R.J. (1994). Determination of fetal numbers in Alpine does by real time ultrasonography. *Small Ruminant Research*. 14: 225-231.
- de Castro, T., Rubianes, E., Menchaca, A. and Rivero, A. (1998). Ultrasonic study of follicular dynamics during the estrous cycle in goats. *Theriogenology*. 49: 399 (abstract).
- de Castro, T., Rubianes, E., Menchaca, A. and Rivero, A. (1999). Ovarian dynamic, serum estradiol and progesterone concentration during the interovulatory interval in goat. *Theriogenology*. 52: 399-411.
- de Sousa, S., de Melo, C.H.S., de Albuquerque, A.C., Filho, T., Avelar, S.R.G., Moura, A.A.A., Martins, J.A.M., Freitas, V.J.F. and Teixeira, D.I.A. (2011). Ovarian follicular response to different hormonal stimulation treatments in Caninéd goats. *Animal Reproduction Science*. 125: 88-93.
- Dhanda, J.S., Taylor, D.G., McCosker, J.E. and Murray, P.J. (1999). The influence of goat genotype on the production of Capretto and Chevon carcasses. 1. Growth and carcass characteristics. *Meat Science*. 52: 355-361.
- Dixit, S.P., Verma, N.K., Aggarwal, R.A.K., Vyas, M.K., Jyoti Rana, Anurodh Sharma, Pooja Tyagi, Pooja Arya, and Ulmek, B.R. (2010). Genetic diversity and relationship among southern Indian goat breeds based on micro satellite markers. *Small Ruminant Research*. 91: 153-159.
- Dogan, I., Nur, Z., Gunay, U., Sagirkaya, H., Soylu, M.K. and Sonmez, C. (2005). Oestrous synchronization during the natural breeding season in Anatolian black does. *Veterinary Medicine*. 50:33-38.
- Doize, F., Vaillancourt, D., Carabin, H. and Belanger, D. (1997). Determination of gestational age in sheep and goats transrectal ultrasonographic measurement of placentomes. *Theriogenology*, 48: 449-460.

- Dorn, C.G., Wolfe, B.A., Bessoudo, E. and Kraemer, D.C. (1989). Follicular detection in goats by ultrasonography. *Theriogenology*. 31: 185.
- Drion, P.V., Furtoss, V., Manfredi, G.B.E., Bouvier, F., Pougard, J.L., Bernelas, D., Caugnon, P., Edmond, M., Remy, B., Sulon, J., Beckers, J.F., Bodind, L. and Leboeuf, B. (2001). Four years of induction/ synchronization of oestrous in dairy goats: Effect on the evolution of eeg binding rate in relation with the parameters of reproduction. *Reproduction and Nutrition Development*. 41: 401-412.
- Drost, M. (1986). Embryo transfer. In Roberts, S.J. (Edition) *Veterinary Obstetrics and Genital Disease (Theriogenology)*. David and Charles Ins., Woodstock, Vermont. pp 927- 941.
- El-Gayar, M. and Holtz (1996). Technical note: Vitrification of goat embryos by the open pulled-straw method. *Journal of Animal Science*. 79: 2436-2438.
- Erasmus, J.A. (2000). Adaptation to various environments and resistance to disease of the improved Boer goat. *Small Ruminant Research*. 36:179-187.
- Fahmy, M.H. and Shretha, J.N.B. *Genetics for the improvement of goat meat production*. Proceedings of the Seventh International Conference on Goats, France, May. 15-21, 2000. pp 187-190.
- Falk S., Wolfgang T. and Carsten G. (2006). Gonadotropin-releasing hormone (GnRH) and its natural analogues: A review. *Theriogenology*, 66: 691-709.
- Francisco Carlos de Sousa., Carlos Henrique de Melo, Antonio Carlos Telles Filho Albuquerque, Suelty Renata Gaya Avelar, Arlindo de Alencar Araripe Moura, Jorge Martins Andre Matias, Vicente Jose de Figueiredo Freitas and Italo Darcio Alves Teixeira. (2011). Ovarian follicular response to different hormonal stimulation treatment in Caninde goats. *Animal Reproduction Science*. 125: 88-93.
- Freitas, V.J. and Ba, J. (1997). Estrus synchronization in dairy goats: use of fluorogestone acetate vaginal sponges or norgestomet ear implants. *Animal Reproduction Science*. 46: 237-244.
- Freitas, V.J.F., Baril, G., Bosc, M. and Saumande, J. (1996). The influence of ovarian status on response to estrus synchronization treatment in dairy goats during the breeding season. *Theriogenology*. 45: 1561-1567.
- Fringgens, N.C. (2003). Body lipid reserves and the reproductive cycle: Towards a better understanding. *Livestock Production Science*. 83: 209-226.

- Garner, D.L., Thomas, C.A., Joerg, H.W., DeJarnett, J.M. and Marshall, C.E. (1997). Fluorometric assessment of mitochondrial function and viability in cryopreserved bovine spermatozoa. *Biology Reproduction*. 57: 1401-1406.
- Garcia, A., Neary, M.K., Kelly, G.R. and Pierson, R.A. (1993). Accuracy of ultrasonography in early pregnancy diagnosis in the ewe. *Theriogenology*. 39: 847-861.
- Gearthart, M.A., Winfield, W.E., Knight, A.P., Smith, J.A., Dargatz, D.A., Boon, J.A. and Stokes, C.A. (1988). Real-time ultrasonography for determining pregnancy status and viable fetal numbers in ewes. *Theriogenology*. 30: 323-337.
- Gibbons, J.R., Kot, K., Thomas, D.L., Wiltbank, M.C. and Ginther, O.J. (1999). Follicular and FSH dynamics in ewes with a history of high and low ovulation rates. *Theriogenology*. 52: 1005-1020.
- Ginther, O.J. and Kot, K. (1994). Follicular dynamics during the ovulatory season in goats. *Theriogenology*. 42: 987-1001.
- Glboa, N., Perevolotsky, A., Landau, S., Nitsan, Z. and Silanikove, N. (2000). Increasing productivity in goats grazing Mediterranean woodland and scrubland by supplementation of polyethylene glycol. *Small Ruminant Research*. 38: 183-190.
- Gnatek, G.G., Smith, L.D., Duby, R.T. and Godkin, J.D. (1989). Maternal recognition of pregnancy in the goat: effects of conceptus removal on interestrus intervals and characterization of conceptus protein production during early pregnancy. *Biology of Reproduction*. 41: 655-663.
- Gonzales-Bulnes, A., Diaz-Delfa, C., Urrutia, B., Carrizosa, J.A. and Lopez-Sebastian, A. (2004). Ultrasonographic screening of the ovulatory process in goat. *Small Ruminant Research*. 52: 165-168.
- Gonzalez-Bulnes, A., Carrizosa, J.A., Diaz-Delfa, C., Garcia-Garcia, R.M. Urrutia, B., Santiago-Moreno, J., Cocero, M.J. and Lopez-Sebastian, A. (2003). Effects of ovarian follicular status on superovulatory responses of dairy goats to FSH treatment. *Small Ruminant Research*. 48: 9-14.
- Gonzalez-Bulnes, A., Santiago-Moreno, J., Cocero, M.J. and Lopez-Sebastian, A. (2000). Effect of FSH commercial preparation and follicular status on follicular growth and superovulatory response in Spanish Merino ewes. *Theriogenology*. 54: 1055-1064.

- Gonzalez de Bulnes, A, Osoro, K. and Lopez-Sebastian, A. (1999a). Ultrasonic assessment of the ovarian response in eCG-treated goats. *Small Ruminant Research*. 34: 65-69.
- Gonzalez de Bulnes, A., Santiago Moreno, J., Gomez-Brunet, A., Inskip, E.K., Townsend, E.M. and Lopez-Sebastian, A. (1999b). Follicular dynamics during the oestrous cycle in dairy goats. *Journal of Animal Science*. 68: 547-554.
- Gonzalez, F., Cabrera, F., Batista, M. Rordriguez, N., Alamo, D., Sulon, J., Beckers, J.F. and Gracia, A. (2004). A comparison of diagnosis of pregnancy in the goat via transrectal, ultrasound scanning, progesterone and pregnancy associated glycoprotein assay. *Theriogenology*. 62: 1108-1115.
- Gordon, I. (1997). Controlled reproduction in sheep and goats. (1st edition). CAB International, Wallingford, UK. pp 86-115.
- Graham, E.F. *Fundamental of the preservation of spermatozoa*. In: *The integrity of frozen spermatozoa*. In Proceeding 5th Technology Conference National Academic Sciences. Washington, D.C, 1978. pp 4-44.
- Gregor, B.M.A. (2007). An introduction to reproduction management of fibre and meat goats. Retrieved 24 Jan 2008 from <http://www.ces.vic.gov.au/DPI/nreninf>.
- Greyling, J.P.G., Van der nest, M., M., Schwalbach, L.M.J. and Muller, T.S. (2002). Superovulation and embryo transfer in South African Boer and indigenous Feral goats. *Small Ruminant Research*. 43: 45-51.
- Greyling, J.P. (2000). Reproduction traits in the Boer goat doe. *Small Ruminant Research*. 36 (2): 171-177.
- Greyling, J.P.C. and Van Niekerk, C.J. (1990). Effect of pregnant mare serum gonadotrophin (PMSG) and route of administration after progestagen treatment on oestrus and LH secretion in the Boer goat. *Small Ruminant Research*. 3: 511-516.
- Hafez, E.S.E. (1987). Reproduction in farm animals. 5th edition. Philadelphia: Lea and Febiger, Philadelphia. pp 398.
- Hamilton, W.J. and Harrison, R.J. (1951). Cyclical changes in the uterine mucosa and vagina of the goat. *Journal of Anatomy*. 85: 316-324.
- Heidari, F., Gharagozloo, F., Vojgani, M., Farrokhi, N., Vajhi, A.R., Masoudifard, Mirtorabi, M. and Nayeri Fasaei, B. (2010). The effect of a GnRH antagonist pre-treatment, in the superovulation of goats. *Small Ruminant Research*. 93: 140-143.

- Hirooka, H., Mukherjee, T.K., Panandam, J.M. and Horst, P. (1997). Genetic parameters for growth performance of the Malaysian local goats and their crossbreds with the German (improved) Fawn goats. *Journal of Animal Breed and Genetics*. 114: 191-199.
- Hulet, C.V. and Shelton, M. (1980). Reproductive cycles of sheep and goat. In reproduction in farm animals. 4th Edition. Hafez, E.S.E. (ed), Lea and Febiger, Philadelphia. pp 346-357.
- Hutchens, T. (2005). Overview of Artificial Insemination of Kentucky Meat and Dairy Goats Extention Associate University of Kentucky, Goat producer Newsletter from UK and KSU. Retrieved 29 Okt 2010 from <http://www.uky.edu/AG/AnimalScience/goats/newsletter/goatproducersnewsletter019052.pdf>.
- Holtz.W. (2005). Recent developments in assisted reproduction in goats. *Small Ruminant Research*. 60: 95-110.
- Ishwar A.K, and Memon, M.A. (1995). Embryo transfer in sheep and goats: a review. *Small Ruminant Research* 19: 35-43.
- Jainudeen, M.R. and Hafez, E.S.E. (1987). *Sheep and Goats*. In Hafez, E.S.E. (Edition) Reproduction in farm animals 5th edition Philadelphia: Lea and Febiger. pp 315-323.
- Jarrell, V.L. and Dziuk, P.J. (1991). Effect of number of corpora lutea and fetuses on concentrations of progesterone in blood of goats. *Journal of Animal Science*. 69: 770-773.
- Kadzere, C.T., Llewelyn, C.A. and Chivandi, E. (1996). Plasma progesterone, calcium, magnesium and zinc concentration from oestrus synchronization to weaning in indigenous goats in Zimbabwe. *Small Ruminant Research*. 24:21-26.
- Kähn, W. (2004). Ultrasonography in sheep and goats. Schlütersche Verlag GmbH & Co. KG, Hans-Böckler-Allee 7, 30173, Hannover, Germany. 3: 187-210.
- Kähn, W., Achtzehn, J., Kähn, B., Richter, A., Schulz, J. and Wolf, M. (1993). Sonography of pregnancy in sheep. II. Accuracy of tranrectal and transcutaneous pregnancy diagnosis. *Dutch Veterinary Public Health*. 100: 29-31.
- Kelidari, H.R., Souri, M., Shabankareh, H.K. and Hashemi, S.B. (2010). Repeated administration of hCG on follicular and luteal characteristics and serum progesterone concentrations in eCG-superovulated does. *Small Ruminant Research*. 90: 95-100.

- Kermani, H.M., Kohram, H., Zareh, S.A., and Saberifar, T. (2012). Ovarian response and pregnancy rate following different doses of eCG treatment in Chall ewes. *Small Ruminant Research*. 102: 63-67.
- Knight, M. and Garcia, G. (1997). The status and characteristics of the goat (*Capra hircus*) and its potential role as a significant milk producer in the tropics: a review. *Small Ruminant Research*. 26: 203–215.
- Krisher, R.L., Gwazdauskas, F.C., Page, R.L., Russell, C.G., Canseco, R.S., Sparks, A.E.T., Valender, W.H., Johnson, J.L. and Pearson, R.E. (1994). Ovulation rate, zygote recovery and follicular populations in FSH-superovulated goats treated with PGF₂ α and/or GnRH. *Theriogenology*. 41: 491-498.
- Lassoued, N. and Rekik, M. (2005). Variations saisonnieres de l'oestrus et de l'ovulation chez la chevre locale Maure en Tunisie. (Seasonal variations in estrus and ovulation in the local Maure goat in Tunisia). *Journal of Animal Husbandry and Veterinary Medicine in Tropical Countries*. 58: 69-73.
- Leboeuf, B. Extensive applications of artificial insemination in goats. In: Proceeding of the 5th International Conference on Goats, vol 2, New Delhi, India, March. 2 -8, 1992. pp 299-308.
- Leboeuf, B., Forgererit, Y., Bernelas, D., Pougard, J.L., Senty, E. and Driancourt, M.A. (2003). Efficacy of two types of vaginal sponges to control onset of oestrus, time of preovulatory LH peak and kidding rate in goats inseminated with variable numbers of spermatozoa. *Theriogenology*. 50: 1371-1380.
- Lehloenya, K.C. and Greyling, J.P.C. (2010). The ovarian response and embryo recovery rate in Boer goat does following different superovulation protocols, during the breeding season. *Small Ruminant Research*. 88: 38-43.
- Lehloenya, K.C., Greyling, J.P.C. and Grobler, S. (2008). Effect of season on the superovulatory response in Boer does. *Small Ruminant Research*. 78: 74-79.
- Lehloenya, K.C., Greyling, J.P.C. and Schwalbach, L.M.J. (2005). Reproductive performance of South African indigenous goats following oestrous synchronization and AI. *Small Ruminant Research*. 57:115-120.
- Llewelyn, C.A., Ogaa, J.S. and Obwolo, M.J. (1992). Plasma progesterone concentration during pregnancy, pseudopregnancy and onset of ovarian activity post-partum of indigenous goats of Zimbabwe. *Tropical Animal Health Production*. 24 (4): 242-250.
- Llewelyn, C.A. and Kadzere, C.T. (1992). Oestrus synchronization and fertility following treatment with fluorogestone acetate (Chronolone) impregnated

intra-vaginal sponges in indigenous goats of Zimbabwe. *Zimbabwe Veterinary Journal*. 23 (4): 159-164.

Mahmood, S., Koul, G.L. and Biswas, J.C. (1991). Comparative efficacy of FSH-P and PMSG on superovulation in Pashmina goats. *Theriogenology*. 35: 1191-1196.

Malan, S.W. (2000). The improved Boer goat. *Small Ruminant Research*. 36: 165-170.

Malhi, P.S., Adams, G.P. and Mapletoft, R.J. (2008). Superovulatory response in a bovine model of reproductive aging. *Animal Reproduction Science*. pp 1257-1266.

Martinez, M.F., Bosch, P. and Bosch, R.A. (1998). Determination of early pregnancy and embryonic growth in goat by transrectal ultrasound scanning. *Theriogenology*. 49: 1555-1565.

McBride-Johnson, B., Nuti, L.C. and Wiltz, D. (1994). Ultrasonography examination of the caprine ovary. *Veterinary Medicine*. 89: 477-480.

McIntosh, W.A.C., Robinson, J.J. and Aitken, R.P. (1975). Pregnant Mare Serum Gonadotrophin: Rate of clearance from the circulation of sheep. *Reproduction Fertility Journal*. 44: 95-100.

McNatty, K.P., Hudson, N.L., Ball, K., Mason, A. and Simons, M.H. (1989). Superovulation and embryo recovery in goats treated with Ovagen and Folltropin. *New Zealand Veterinary Journal*. 37: 27-29.

Mellado, M., Olivas, R. and Ruiz, F. (2000a). Effect of buck stimulus on mature and pre-pubertal norgestomet-treated goats. *Small Ruminant Research*. 36: 269-274.

Mellado, M., Amaro, J.L. Garcia, J.E. and Lara, L.M. (2000b). Factors affecting gestation length in goats and the effect of gestation period on kid survival. *Journal of Agricultural Science*. 135 (1), 85-89.

Memon, M.A., Bretzlaff, K.N. and Ott, R.S. (1986). Comparison of semen collection technique in goats. *Theriogenology*. 26:6: 823- 827.

Memon, M.A. (1983). Male fertility. In: Sheep and Goat disease. *Veterinary Clinics of North America*. W.B. Saunders, Philadelphia. 5: 539-555.

Menchaca, A. and Rubianes, E. (2002). Relation between progesterone levels during the early luteal phase and follicular dynamic in goats. *Theriogenology*. 57: 1411- 1419.

- Menchaca, A., Pinczak, A. and Rubianes, E. (2002). Follicular recruitment and ovulatory response to FSH treatment initiated on Day 0 or Day 3 post-ovulation in goats. *Theriogenology*. 58: 1713-1721.
- Menegatos, J., Chadio, S., Karatzas, G. and Stoforos, E. (1995). Progesterone levels throughout progestagen treatment influence the establishment of pregnancy in the goat. *Theriogenology*. 43: 1365-1370.
- Mengistu, U. (2007). *Performance of the Ethiopian Somali goats during different watering regimes*. Doctoral thesis. Department of Anatomy, Physiology and Biochemistry, Swedish University of Agricultural Sciences. Acta Universitatis Agriculturae Sueciae. Volume 53. Uppsala, Sweden. ISSN 1652-6880, ISBN 978-91-576-7352-7.
- Moaeen-ud-Din, M., Yang, L.G., Chen, S.L., Zhang, Z.R., Xiao, J.Z., Wen, Q.Y. and Dai, M. (2008). Reproductive performance of Matou goat under subtropical monsoonal climate of Central China. *Tropical Animal Health and Production*. 40: 17-23.
- Moore, N.W. and Eppleston, J. (1979). Embryo transfer in the Angora goat. *Australia Journal Agriculture Research*. 30: 973-981.
- Morand-Fehr, P., Boutonnet, J., Devendra, C., Dubef, J.P., Haenlein, G., Holst, P., Mowlem, L. and Capote, J. (2004). Strategy for goat farming in the 21st century. *Small Ruminant Research*. 51: 175-183.
- Motlomelo, K.C., Greyling, J.P.C. and Schwalbach, L.M.J. (2002). Synchronization of oestrous in goats: the use of different progestagen treatments. *Small Ruminant Research*. 45: 45-49.
- Naqvi, S.M.K. and Gulyani, R. (1999). Ovarian response and embryo recovery to different superovulatory regimens in Rambouillet ewes under semi-arid conditions. *Small Ruminant Research*. 34: 127-131.
- Noakes, E.D., Parkinson, J.T. and England, G.C.W. (2001). Fertility and infertility in male animals. In: England, G.C.W., Arthur, G.H., Noakes, D.E., Parkinson, T.J. (Eds.), *Arthur's Veterinary Reproduction and Obstetrics*, 8th ed. Saunders, Philadelphia. pp 695-750.
- Nowshari, M.A., Yuswiati, E., Puls-Kleingeld, M. and Holtz, W. (1992). Superovulation in peripubertal and adult goats treated with PMSG or pFSH. In: Lokeshwar RR (edition), *Recent advance in goat production*. Nutan Printers, New Delhi, India. pp 1358-1363.
- Ott, R.S. and Memon, M.A. (1980a). *Factors effecting reproductive performance*. In: *Sheep and Goat Manual*. Society for Theriogenology. X:5-9.

- Ott, R.S., Nelson, D.R. and Hixon, J.E. (1980b). Fertility of goats following synchronization of oestrus with prostaglandin F2 alfa. *Theriogenology*. 13: 341-345.
- Padilla-Rivas, G.R., Sohnrey, B. and Holtz, W. (2005). Early pregnancy detection by real time ultrasonography in Boer goats. *Small Ruminant Research*. 58:87-92.
- Panhwar, F. (2007). Modern reproductive methods used to enhance goat production. Retrieved 29 Okt 2010 from <http://www.goatworld.com/articles/management/repduction.shtml>
- Pierson, J.T., Baldassarre, H., Keefer, C.L. and Downey, B.R. (2003). Influence of GnRH administration on timing of the LH surge and ovulation in dwarf goats. *Theriogenology*. 60: 397-406.
- Pinczak, A., Menchaca, A. and Rubianes, E. Ovarian and uterine scanning during the early pregnancy in goats. In Proceedings the IV International Symposium on Animal Reproduction. Cordoba, Argentina, June. 22nd -24th 2001. pp 298 (abstract).
- Rege, J.E., Toe, F., Mukasa-Mugerwa, E., Tembely, S.Anindo, D. Baker, R.L.andLahlou-Kassi, A. (2000). Reproductive characteristics of Ethiopian Highland sheep. II. Genetic parameters of semen characteristics and their relationships with testicular measurement in ram lambs. *Small Ruminant Research*. 37: 173-187.
- Regueiro, M., Perez Clariget, R., Ganzabal, A., Aba, M. and Forsberg, M. (1999). Effect of medroxyprogesterone acetate and eCG treatment on the reproductive performance of dairy goats. *Small Ruminant Research*. 33: 223-230.
- Riaz, H., Sattar, A., Arshad, M.A. and Ahmad, N. (2012). Effect of synchronization protocols and GnRH treatment on the reproductive performance in goats. *Small Ruminant Research*. 104: 151-155.
- Riesenber, S., Meinecke-Tillmann, S. and Meinecke B. (2001). Ultrasonic survey of follicular development following superovulation with a single application of pFSH, eCG or hMG in goats. *Small Ruminant Research*. 40: 83-93.
- Riesenber, S., Lewalski, H., Meinecke-Tillmann, S. and Meinecke, B. *Ultrasonic documentation of follicular dynamics following superovulatory regimens in small ruminants- preliminary results*. In: Proceeding of the 11th Reunion of A.ET.E., Hannover, Germany, September. 8-9, 1995. pp 234.

- Ritar, A., Roberson, J. and Evans, G. (1994). Ovulatory activity, hormonal induction of ovulation and fertility of young cashmere and Angora female goats in a temperate environment. *Reproduction Fertility Development*. 6: 737-747.
- Ritar, A.J. (1993). Control of ovulation, storage of semen and artificial insemination of fibre-producing goats in Australia: A review. *Journal Experimental Agriculture*. 33: 807-820.
- Ritar, A., Maxwell, W. and Salamon, S. (1984). Ovulation and LH secretion in the goat after intravaginal progestagen sponges-PMSG treatment. *Journal of Reproductive Fertility*. 72: 559-563.
- Robinson, J.J., Ashworth, C.J., Rooke, J.A., Mitchell, L.M. and McEvoy, T.G. (2006). Nutrition and fertility in ruminant livestock. *Animal Feed Science and Technology*. 126: 259-276.
- Romano, B.G.C. and Wheaton, J.E. (1998). Transcervical artificial insemination of ewes out of season using frozen-thawed semen effect of equine chorionic gonadotropin on pregnancy rate. *Theriogenology*. 49: 997-1005.
- Romano, J.E. (2004). Synchronization of estrus using CIDR, FGA or MAP intravaginal passaries during the breeding season in Nubian goats. *Small Ruminant Research*. 55: 15-19.
- Rosnina, Y (1989). *Superovulation and egg recovery in goats*. Master of Sciences Thesis, Universiti Putra Malaysia, Selangor.
- Roy, F., Maurel, M.C., Combes, B., Vaiman, D., Edmond, P., Isabelle, C., Pobel, T., Deleting, F., Combarous, I. and Guillou, F. (1999). The negative effect of repeated equine Chorionic Gonadotropin treatment on subsequent fertility in alpine goats is due to a hormonal immune response involving the major histocompatibility complex. *Biology of Reproduction*. 60: 805-813.
- Rubianes, E. and Ungerfeld, R. (1993). Uterine involution and ovarian changes during early postpartum in autumn lambing corriedale ewes. *Theriogenology*. 40: 365-372.
- Sadler, R. (1986). *Embryo transfer procedures in goats*. *Asian Livestock*. 21 (11): 144-150.
- Salehi, R., Kohram, H., Towhidi, A., Kermani Moakhtar, H. and Honarvar, M. (2010). Follicular development and ovulation rate following different superovulatory treatments in Chall ewes. *Small Ruminant Research*. 93: 213-217.

- Salisbury, G.W., Fuller, H.K. and Willett, E.T. (1941). Preservation of bovine spermatozoa in yolk-citrate diluents and field results from its use. *Journal of Dairy Science*, 24: 905-910.
- Sargison, N.D. (2008). *Sheep Flock Health. A Planned Approach*. Blackwell, Oxford.
- Saumande, J., Chupin, D., Mariana, J.C., Ortavant, R. and Mauleon, P. (1978). Some studies on Pregnant Mare Serum Gonadotrophin (PMSG) and on Endocrine responses after application for superovulation in cattle. In Sreena, J.M., and Nijhoff, M. (eds) *Current topics in Veterinary Medical I*. The Hague. Page 122-143.
- Saunders Comprehensive Veterinary Dictionary (3rd edition) (2007). Saunders Elsevier, St. Louis, Missouri, USA. pp 656.
- Schrick F.N., Surface R.A., Pritchard J.Y., Dailey R.A., Townsend E.C. and Inskoop E.K. (1993). Ovarian structures during the estrous cycle and early pregnancy in ewes. *Biology Reproductive*. 49: 1133-1140.
- Schwarz, T. and Wiezhos, E. (2000). Relationship between FSH and ovarian follicular dynamics in goats during the estrous cycle. *Theriogenology*. 53: 381 (abstract).
- Selgrath, J.P., Memon, M.A., Smith, T.E. and Ebert, K.M. (1990). Collection and transfer of microinjectable embryos from dairy goats. *Theriogenology*. 34, 1195-1205.
- Selvaraju, M. and Kathiresan, D. (1997). Effect of oestrus synchronization on kidding rate in Tellcherry goats. *Indian Veterinary Journal*. 74: 35-37.
- Senn, B.J. and Richardson, M.E. (1992). Seasonal effects on caprine response to synchronization of estrus and superovulatory treatment. *Theriogenology*, 37: 579-685.
- Senthil Kumar, P., Saravanan, D., Rajasundaram, R.C., Selvaraju, M. and Kathiresan, D. (2003). Serum oestradiol and progesterone profiles and their relationship with superovulatory responses in Tellcherry goats treated with eCG and FSH. *Small Ruminant Research*. 49: 69-77.
- Sheldrick, E.L., Ricketts, A.P. and Flint, A.P.F. (1981). Placental production of 5 β -pregnane-3 α , 20 α -diol in goats. *Journal of Endocrinology*. 90: 151-158.
- Shelton, M. (1978). Reproduction and breeding of goats. *Journal of Dairy Science*. 61: 994-1010.

- Shiple, C.F.B., Buckrell, B.C., Myline, M.J.A., Pollard, J. and Hunton, J.R. (2007). Artificial insemination and embryo transfer in sheep. In: Youngquist, R.S. Theriell, W.R. (Eds.) *Current Therapy in Large Animal: Theriogenology 2*. Saunders/ Elsevier, pp. 629-641.
- Shrestha, J.N.B. and Fahmy, M.H. (2007). Breeding goats for meat production: a review (2). Crossbreeding and formation of composite population. *Small Ruminant Research*. 67: 93-112.
- Silanikove, N. (2000). The physiological basis of adaptation in goats in to harsh environments. *Small Ruminant Research*. 35: 181–193.
- Singh-Knights, D. and Knights, M. (2005). Feasibility of Goat Production in West Virginia: A Handbook for Beginners. West Virginia Agricultural and Forestry Experiment Station, Davis College of Agriculture, Forestry, and Consumer Sciences, Western Virginia University (Bulletin 728).
- Skinner, J.D. (1972). Utilization of the Boer goat for intensive animal production. *Tropical Animal Health Production*. 4: 120-128.
- Smith, M.C. and Sherman, D.M. (1994). Goat medicine. John Wiley and Sons. 2005.
- Solaiman, S., Min, B.R., Gurung, N., Behrends, J. and Taha, E. (2011). Assessing feed intake, growth performance, organ growth and carcass characteristics of purebred Boer and Kiko male kids fed high concentrate diet. *Small Ruminant Research*. 98: 98-101.
- Sreenan, J.M., Beehan, D. and Gosling, J.P. (1978). Ovarian responses in relation to endocrine status following PMSG stimulation in the cow. In Screena, J.M. (ed.) *Control of reproduction in the cow. C.E.C. Luxembourg*. Page 144-158.
- Steele, M. (1996). Goat. The tropical Agriculturalist series. CTA publication, Macmillan Education Ltd, London. 40-43.
- Stephen, W.D. (2003). Introduction to animal science: Global, biological, social and industry perspectives. Pearson Education, Incorporated, Upper Saddle River, New Jersey, 07458, USA. Part 1, Chapter 1, pp 4.
- Suguna, K. S., Mehrotra, S.K. Agarwal, M. Hoque, S.K. Singh, U. Shanker, T. Sarath (2008). Early pregnancy diagnosis and embryonic and fetal development using real time B mode ultrasound in goats. *Small Ruminant Research*. 80: 80-86.
- Tamboura, H., Sawadogo, L. and Wereme, A. (1998). Caracteristiques temporelles et endocriennes de la puberte et du cycle oestral chez la chevre locale "Mossi"

du Burkina Faso. (Features and temperelles endocriennes of puberty and the estrous cycle in local goats “Mossi” Burkina Faso). *Biotechnology Agronomy Society and Environmental*. 2 (1): 85-91.

Tervit, H.R. (1986). Embryo transfer and artificial insemination in Angora goat. Proceeding, Mohair Conference, New Zealand. pp 12-17.

Toe, F., Rege, J., Mukasa-Mugerwa, E., Tembely, S., Anindo, D., Baker, R. and Lahlou-Kassi, A. (2000). Reproductive characteristics of Ethiopian highland sheep. I. Genetic parameters of testicular measurement in ram lambs and relationship with age at puberty in ewe lambs. *Small Ruminant Research*. 36: 227-240.

Van der Westhuysen, J.M. (1979). The control of ovarian function in cycling and anoestrous Angora goat does. *Agroanimalia* 11: 23-35.

Vinoles, C., Forsberg, M., Banchero, G. and Rubianes, E. (2002). Ovarian follicular dynamics and endocrine profiles in Polwarth ewes in high and low body condition. *Journal of Animal Science*. 74: 539-545.

Vinoles, C., Forsberg, M., Banchero, G. and Rubianes, E. (2001). Effect of long-term and short-term progestagen treatment on follicular development and pregnancy rate in cyclic ewes. *Theriogenology*. 55: 993-1004.

Waldron, D.F., Willingham, T.D., Thompson, P.V. and Bretzlaff, K.N. (1999). Effect of concomitant injection of prostaglandin and PMSG on pregnancy rate and prolificacy of artificially inseminated Spanish goats synchronized with controlled internal drug release devices. *Small Ruminant Research*. 31: 177-179.

Webb, E.C., Mamabolo, M.J., Du Preez, E.R. and Morris, S.D. (2003). Reproductive status of goats in communal systems in South Africa, Retrived 9 Jun 2011 from <http://www.upac.za/asservices/ais/vet/sec41.pdf>

Whitley, N.C. and Jackson D.J. (2004). An update on estrus synchronization in goats: a minor species. *Journal of Animal Science*. 82: 270-276.

Wildeus, S. (1999). Current concepts in synchronization of oestrus: Sheep and goats. Retrieved 29 Oktober 2010 from <http://jas.fass.org/cgi/reprint/77/E-Suppl/1-am.pdf>

Wildeus, S. (2007). Reproductive management of the meat goat. Retrieved 29 October 2010 from <http://www.goatworld.com/articles/pregnancy/reproductivemanagement.shtml>

Wilson, R.T. and Durkin, J.W. (1984). Age at permanent incisor eruption in indigenous goats and sheep in semi-arid Africa. *Livestock Production Science*. 11: 451-455.

Zeder, M.A. and Hesse, B. (1999). Science news, Goat busters track domestic. The initial domestication of goats (*Capra hircus*) in the Zagros Mountains 10,000 years ago. *Science*. 287:254-257.

