

UNIVERSITI PUTRA MALAYSIA

EFFECTS OF SUPPLEMENTING BAKER'S YEAST (Saccharomyces cerevisiae) AND YEAST GLUCAN ON GROWTH PERFORMANCE, NUTRIENT DIGESTIBILITY AND MEAT QUALITY OF BROILER CHICKEN

NABILA BINTI MOHAMADDIN

FP 2017 40



EFFECTS OF SUPPLEMENTING BAKER'S YEAST (Saccharomyces cerevisiae) AND YEAST GLUCAN ON GROWTH PERFORMANCE, NUTRIENT DIGESTIBILITY AND MEAT QUALITY OF BROILER CHICKEN



Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirements for the Degree of Master of Science

May 2017

COPYRIGHT

All material contained within this thesis, including without limitation text, logos, icons, and other artwork is copyright material of Universiti of Putra Malaysia unless otherwise stated. Use may be made of any material contained within this thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti of Putra Malaysia.

Copyright © Universiti of Putra Malaysia

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Master of Science

EFFECTS OF SUPPLEMENTING BAKER'S YEAST (Saccharomyces cerevisiae) AND YEAST GLUCAN ON GROWTH PERFORMANCE, NUTRIENT DIGESTIBILITY AND MEAT QUALITY OF BROILER CHICKEN

By

NABILA BINTI MOHAMADDIN

May 2017

Chairman Faculty

: Associate Professor Anjas Asmara Samsudin, PhD : Agriculture

Increasing concerns regarding overuse of antibiotics in broiler has promoted extensive investigation into alternatives to the use of sub-therapeutic antibiotics such as the used of yeast products. Saccharomyces cerevisiae, which is also known as "Baker Yeast" is rich in crude protein (40-45%) and vitamin B complex. Yeast products have been widely reported as successful growth promoter in poultry industry. There is no relevant information on the meat quality parameter, abdominal fat in broiler fed with yeast, and also blood profile parameter in broiler chicken. Therefore, these parameters need to be investigate for additional information on the effects of yeast products on broilers. The purpose of this study was to compare the levels of baker's yeast and to study the effects of non-antibiotic feed additives, that is baker's yeast and yeast glucan on broiler performance. The first experiment involved a total of 200 a day-old broilers (Cobb 500) and were randomly divided into five treatments, four replicates with 10 birds each. The treatments were T1: control; T2: 0.1% baker's yeast; T3: 0.2% baker's yeast; T4: 0.4% baker's yeast; and T5: 0.8% baker's yeast. For the second experiment, 200 a day-old broilers were randomly divided into four treatments, five replicates with 10 birds each. The treatments were T1: control; T2: probiotic (best level of baker's yeast from first experiment); T3: prebiotic (MacroGard[®] yeast glucan); and T4: antibiotic (virginiamycin). The individual body weight and feed intake were recorded weekly, and at day 42, the chickens were slaughtered to determine carcass quality, collection of ileal and ceacal digesta, blood collection, and gut morphology. For the first experiment, data on performances, nutrient digestibility, carcass characteristics, bacteria count, and gut morphology were collected, whereas for the second experiment, data on performances, nutrient digestibility, meat quality, bacteria count, gut morphology, and blood profile were collected. The results of this experiment showed that addition of baker's yeast (probiotic) and yeast glucan (prebiotic) to broiler's feed improved body weight compared to control diet. Similar result was also observed in the feed conversion ratio, where both baker's yeast and yeast glucan showed the lowest FCR followed by control treatment. However, carcass dressing rates were not affected by the non-antibiotic feed additives. The nutrient digestibility by the broiler improved when baker's yeast and yeast glucan were fed to the birds. Addition of both baker's yeast and yeast glucan to broiler's feed had improved the tenderness of the broiler's meat and gut morphology of the broiler (P<0.05). Other than that, baker's yeast had improved immunity through WBC reduction. In addition, baker's yeast and yeast glucan supplementation had reduced the abdominal fat content of the birds. Moreover, the inclusion of baker's yeast in broiler's feed has enhanced the population of beneficial bacteria such as Lactobacillus spp. and reduced the colonization of pathogenic bacteria such as E. coli. It has been concluded that baker's yeast has improved the performance of the broiler chicken. Thus, these can be used as antibiotic alternatives in broilers feed.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Sarjana Sains

KESAN SUPLEMENTASI YIS ROTI (Saccharomyces cerevisiae) DAN YIS GLUKAN TERHADAP PRESTASI TUMBESARAN, KADAR PENYERAPAN NUTRIEN, DAN KUALITI DAGING AYAM PEDAGING

Oleh

NABILA BINTI MOHAMADDIN

Mei 2017

Pengerusi Fakulti : Profesor Madya Anjas Asmara Samsudin, PhD : Pertanian

Kebimbangan yang meningkat berikutan penggunaan antibiotik yang berlebihan dalam industri ayam pedaging telah menarik minat penyelidik dalam mencari alternatif lain untuk menggantikan pengunaan antibiotik kepada penggunaan produk vis. Saccharomyces cerevisiae, vang dikenali sebagai "yis roti" kaya dengan protein mentah (40-45%) dan vitamin B kompleks. Ekstrak yis telah dilaporkan secara meluas sebagai pemangkin tumbesaran dalam industri poltri. Tidak ada maklumat yang relevan mengenai parameter kualiti daging, lemak di bahagian abdomen dalam ayam pedaging yang telah diberi makan yis, dan juga profil darah parameter dalam ayam daging. Tujuan kajian ini adalah untuk mengkaji kesan tahap yis roti dan kesan terhadap alternatif makanan tambahan bukan antibiotik, jaitu yis roti terhadap prestasi ayam pedaging. Untuk eksperimen pertama, sebanyak 200 ekor anak ayam berumur satu hari (Cobb 500) secara rawak telah dibahagikan kepada lima kumpulan diet, empat kumpulan kecil bagi setiap kumpulan diet dengan sepuluh ekor anak ayam untuk setiap kumpulan kecil. Antara kumpulan dietnya ialah T1: diet tanpa yis roti; T2: 0.1% yis roti; T3: 0.2% yis roti; T4: 0.4% yis roti; dan T5: 0.8% yis roti. Untuk eksperimen kedua, sebanyak 200 ekor anak ayam berumur satu hari (Cobb 500) secara rawak telah dibahagikan kepada empat kumpulan diet, lima kumpulan kecil bagi setiap kumpulan diet dengan sepuluh ekor anak ayam untuk setiap kumpulan kecil. Antara kumpulan dietnya ialah T1: diet tanpa makanan tambahan; T2: probiotik (tahap terbaik yis roti dari eksperimen pertama); T3: prebiotik (MacroGard[®] yis glukan); dan T4: antibiotik (virginiamycin). Berat badan dan pengambilan makanan individu telah dicatat setiap minggu, dan pada hari ke-42, ayam disembelih untuk pemeriksaan karkas, pengumpulan digesta ilea dan sekum, pengumpulan darah, dan pemeriksaan morfologi usus. Untuk experimen pertama, data terhadap prestasi, penghadaman nutrien, ciri-ciri karkas, bilangan bakteria, dan usus morfologi telah dikumpulkan, manakala untuk eksperimen kedua, data terhadap prestasi, penghadaman nutrien, kualiti daging, bilangan bakteria, usus morphologi, dan profil darah telah dikumpulkan. Hasil keputusan daripada eksperimen ini menunjukkan bahawa dengan mencampurkan yis roti (probiotik) dan yis glukan (prebiotik) ke dalam makanan ayam pedaging, ia telah menjadikan peningkatan berat badan ayam pedaging menjadi bertambah baik berbanding diet tanpa makanan tambahan. Begitu juga dengan data dari nisbah penukaran makanan, dimana nilai makanan tambahan yis roti dan yis glukan adalah terendah berbanding diet tanpa makanan tambahan. Walau bagaimanapun, kadar persalinan karkas tidak terjejas dengan menggunakan alternatif makanan tambahan bukan antibiotik. Penghadaman nutrien oleh ayam pedaging bertambah baik dengan pemberian yis roti dan yis glukan. Penambahan yis roti dan juga yis glukan ke dalam makanan ayam pedaging telah memperbaiki kelembutan daging ayam pedaging dan usus morfologi ayam pedaging (P<0.05). Selain daripada itu, yis roti meningkatkan imuniti melalui pengurangan tahap sel darah putih. Tambahan pula, yis roti dan yis glukan berpotensi untuk mengurangkan sisa lemak oleh sebab pengurangan kandungan lemak yang berlaku di kawasan perut ayam pedaging. Selain itu, penambahan yis roti dalam makanan ayam pedaging telah meningkatkan populasi bakteria berfaedah seperti Lactobacillus spp. dan mengurangkan populasi bakteria berbahaya seperti E. coli. Ini telah disimpulkan bahawa yis roti mampu meningkatkan prestasi ayam pedaging. Oleh itu, yis roti boleh digunakan sebagai alternatif kepada penggunaan antibiotik dalam makanan ayam pedaging.

ACKNOWLEDGEMENTS

I would like to express deepest gratitude to my supervisor Assoc. Prof. Dr. Anjas Asmara Samsudin for his full support, expert guidance, understanding, and encouragement throughout my study and research. Without his incredible patience and timely wisdom and counsel, my thesis work would have been a frustrating and overwhelming pursuit. In addition, I express my appreciation to Assoc. Prof. Dr. Halimatun Yaakub for having served on my committee, and also to my former supervisor, Prof. Dr. Abdul Razak Alimon for his contribution throughout this research. Their thoughtful questions and comments were valued greatly.

I would also like to thank the lab assistant Mr. Khairul Anwar Bahari, Mr. Saparin Demin, Miss Shuhada Adnan, and Mrs. Rohaida Rashid for helping and guiding me with my labwork.

Thanks also goes to all my fellow postgraduate friends in the Animal Science Department, Faculty of Agriculture, who had helped me throughout my research journey.

Finally, I would like to thank my parents and brothers for their unconditionally love and support during my research journey, and without them, I would not have been able to complete this thesis without their continuous love and encouragement. I certify that a Thesis Examination Committee has met on 8 May 2017 to conduct the final examination of Nabila binti Mohamaddin on her thesis entitled "Effects of Supplementing Baker's Yeast (*Saccharomyces cerevisiae*) and Yeast Glucan on Growth Performance, Nutrient Digestibility and Meat Quality of Broiler Chicken" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Loh Teck Chwen, PhD Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Azhar bin Kassim, PhD Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Internal Examiner)

Wan Zahari Mohamed, PhD Professor Universiti Malaysia Kelantan Malaysia (External Examiner)

NOR AINI AB. SHUKOR, PhD Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 4 September 2017

This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Anjas Asmara Samsudin, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Halimatun Yaakub, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Member)

ROBIAH BINTI YUNUS, PhD Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature:	Date:

Name and Matric No.: Nabila Binti Mohamaddin, GS42550

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature:	
Name of Chairman	
of Supervisory	
Committee:	Associate Professor Dr. Anjas Asmara Samsudin

Signature: Name of Member of Supervisory Committee:

Associate Professor Dr. Halimatun Yaakub

TABLE OF CONTENTS

APPR DECLA LIST C LIST C	rak owl oval arat of ta of fic	EDGEMI 'ION			i iii vi viii xiv xvi xvii
CHAP	TER				
1	INTF 1.1 1.2	RODUCT Resear Objectiv	ch Hypoth	esis	1 3 3
2	LITE 2.1		Law (Ani The Purp	ndustry in Malaysia mal Feed Act) poses and Problems of Using to in Animal's Feed	4 4 5
	2.2 2.3	2 <mark>.2.1</mark> 2.2.2 P <mark>robiot</mark> i	tive to an A Pathoger Improved	Antibiotic in Animal's Feed nic Bacteria Control d Health and Production Performance	6 6 7
		2.3.1 2.3.2 2.3.3	Mode of	ed Probiotics Genera	7 8 9
		2.0.0	2.3.3.1 2.3.3.2 2.3.3.3	Types and Substrate of Yeast Nutritional Components in Yeast Baker's Yeast (<i>Saccharomyces</i> <i>cerevisiae</i>) in Animal Feeding	9 10 10
			2.3.3.4 2.3.3.5	Baker's Yeast as a Probiotics Effect of Yeast on Nutrient Absorption	11 11
			2.3.3.6 2.3.3.7	Effect of on Growth Performance Effect of Yeast on The Intestinal Microbiota and Intestinal Morphology	12 12
			2.3.3.8	Effect of Yeast on The Immune Responses	13
	2.4	Prebioti 2.4.1 2.4.2	Common Characte	u Used Prebiotics eristics of Ideal Yeast Glucan and ide of Actions	13 13 14

C

	2.5	 2.4.3 Yeast Glucan 2.4.4 Role of Yeast Glucan 2.4.5 Baker's Yeast Cell Wall β-D-Glucan Use of Probiotics and Prebiotic in Poultry nutrition 2.5.1 Reported Effects of Probiotics and Prebiotics in Broiler Chickens 	15 16 16 16 17
	2.6	Conclusion	18
3	3.1 3.2 3.3		19 19 19 19 20 20 20 21 21 21 21 22 23 23 23 23 24 24 24
4		ERMINATION OF SUITABLE INCLUSION LEVEL OF	25
		Introduction	25 26 26 26 27 27 28 28 28 28 28 28 28 28

	4.3	Results		30
		4.3.1	Growth Performance	30
		4.3.2	Nutrient Digestibility	32
		4.3.3	Carcass Characteristics	32
		4.3.4	Gut Morphology	32
		4.3.5	Lactobacillus Bacteria Count	35
	4.4	Discussi	ion	37
		4.4.1	Growth Performance	37
		4.4.2	Nutrient Digestbility	38
		4.4.3	Carcass Characteristics	39
		4.4.4	Gut Morphology	40
		4.4.5	Lactobacillus Bacteria Count	40
	4.5	Conclus		41
F	eccr		BAKER'S YEAST AND YEAST GLUCAN	43
			TATION AS AN ALTERNATIVE TO THE	43
		BIOTICS		
	5.1	Introduc		43
	5.2		s and Methods	44
	0.2		Location	44
		5.2.2	Management of Animal	44
		5.2.3	Diets and Treatments	44
		5.2.4	Digestibility Trial	45
		5.2.5	Cmemical Analysis	46
		5.2.6	Carcass Charactristics Analysis	40 46
		5.2.7	Meat Quality Sampling	46
		5.2.1	5.2.7.1 Determination of Muscle pH	46
			5.2.7.2 Determination of Drip Loss	46
			5.2.7.3 Determination of Colour	47
			Coordinates	47
			5.2.7.4 Determination of Cooking Loss	47
			5.2.7.5 Determination of Shear Force	47
		5.2.8	Villus Height and Crypt Depth	47
		5.2.9	Microbiology Analysis	48
		5.2.5	5.2.9.1 Preparation of Caecal Sample	48
			5.2.9.2 Extraction of Caecal DNA	48
		×	5.2.9.3 Primers and SYBR Green Analysis	40
		5.2.10	Haematological Analysis	49
			5.2.10.1 Blood Collection	49
	5.3	Results		52
		5.3.1	Growth Performance	52
		5.3.2	Nutrient Utilization	52
		5.3.3	Carcass Characteristics	55
		5.3.4	Meat Quality	55
		5.3.5	Blood Haematology Parameters	55
		5.3.6	Gut Morphology	55
		5.3.7	E.coli and Lactobacillus spp. Bacteria Counts	60
	5.4	Discussi	ion	62
		5.4.1	Growth Performance	62

		5.4.2	Nutrient Utiliza	ation		63
		5.4.3	Carcass Char	acteristics		64
		5.4.4	Meat Quality			64
		5.4.5	Blood Haema	tology Paramet	ters	65
		5.4.6				66
		5.4.7			Bacteria Counts	67
	5.5	Conclu			-	68
6	GEN	IERAL D	DISCUSSION			69
7	GEN			AND RECOMM	IENDATION	71
REFERENCES 73						
APPE						94
		OF STUD	DENT			97
PUBL	ICATI	ON				98

(G)

LIST OF TABLES

Table		Page
2.1	Characteristics of ideal probiotics and their desirable properties	8
2.2	Characteristics of ideal prebiotics and their desirable properties	15
4.1	Composition of the experimental diets (finisher)	27
4.2	Growth performance of broiler fed with different levels of baker's yeast (mean ± standard error)	31
4.3	Nutrient digestibility of broiler fed with different levels of baker's yeast (mean ± standard error)	33
4.4	Carcass characteristics of broiler fed with different levels of baker's yeast (mean ± standard error)	34
4.5	Gut mo <mark>rphology of broiler</mark> fed with different levels of baker's yeast (mean ± standard error)	36
4.6	Bacterial population in the ileal digesta fed with different levels of baker's yeast (mean ± standard error)	36
5.1	Composition of the experimental diets (Finisher)	45
5.2	The sequence of primers used targeting LAB, and <i>E. coli</i>	49
5.3	Growth performance of broiler fed with baker's yeast, yeast glucan, and virginiamycin (mean ± standard error)	53
5.4	Nutrient digestibility of broiler fed with baker's yeast, yeast glucan, and virginiamycin (mean ± standard error)	54
5.5	Carcass characteristics of broiler fed with baker's yeast, yeast glucan, and virginiamycin (mean ± standard error)	56
5.6	Meat quality of broiler fed with baker's yeast, yeast glucan, and virginiamycin (mean ± standard error)	57
5.7	Blood profile of broiler fed with baker's yeast, yeast glucan, and virginiamycin (mean ± standard error)	58
5.8	Gut morphology of broiler fed wit baker's yeast, yeast glucan, and virginiamycin (mean ± standard error)	59

 $\overline{\mathbf{G}}$

5.9 Bacterial population in the ileal and ceacal digesta fed with different levels of baker's yeast (mean ± standard error)



6

61

LIST OF FIGURES

Figure		Page
4.1	Summarize of sample collection	29
5.1	Summarize of sample collection	51



 (\mathcal{C})

LIST OF ABBREVIATIONS

	AGP	Antibiotic growth promoter
	CF	Crude fiber
	CFU	Colony forming unit
	СР	Crude protein
	DFM	Direct feed microbial
	DM	Dry matter
	DMD	Dry matter digestibility
	DNA	Deoxyribonucleic acid
	EE	Ether extract
	FCR	Feed conversion ratio
	FOS	Fructooligosaccharides
g		Gram
	GE	Gross energy
	GIT	Gastrointestinal tract
	GOS	Galactooligosaccharides
	H ₂ SO ₄	Sulphuric acid
	Hb	Haemoglobin
	HCI	Hydrochloric acid
	hr	Hour
	IMO	Isomalto oligosaccharides
(C)	kg	Kilogram
U	LAB	Lactic acid bacteria
	MCHC	Mean corpuscular haemoglobin concentration

	MCV	Mean corpuscular volume
	ME	Metabolizable energy
	Min	Minute
	ml	Milliliter
	mJ / kg	Millijoule per kilogram
	μΙ	Microliter
	μm	Micrometre
	mm	Millimetre
	MOS	Mannaoligosaccharides
	N₄OH	Sodium hydroxide
	NDOs	Non-digestible oligosaccharides
	NE	Necrotic enteritis
	ОМ	Organic matter
	PCV	Packed cell volume
	qPCR	Quantitative real-time polymerase chain reaction
	RBC	Red blood cell
	SAS	Statistical analysis software
	SC	Saccharomyces cerevisiae
	SCFA	Short chain fatty acid
	SE	Standard error
	sec	Second
	SOS	Soy oligosaccharides
(\mathbf{U})	spp	Subspecies
	VFA	Volatile fatty acid

WBC White blood cell

XOS Xylooligosaccharides



CHAPTER 1

INTRODUCTION

Poultry has become one of the fastest growing segments of the agriculture sector in terms of production. Feed is the biggest contributor and accounts for more than half of the total production cost. The major aim of this industry is to produce the maximum, with minimum input, like other agriculture sector. The cost of poultry feed ingredients and compounded feed has constantly increase over the years and causing the rise in production cost. Thus, effective and balanced feeding is the most important requisite to boost up the poultry production economically. The inclusion of feed additives such as antibiotic, has led to residual effect of antibiotic resistant bacteria in eggs and meat, and eventually lead to various health hazards to consumers. Therefore, the best way to overcome this problem is the use of feed additives such as probiotics or prebiotics for better and safe production (Bidarkar *et al.*, 2014).

Probiotic are single or mixed culture of beneficial live microorganisms, that when introduced into the animals will efficiently improved the properties of beneficial bacterial that naturally inhibit the gastrointestinal tract (GIT) of the animals. They are also known as DFM (direct feed microbial). The term 'probiotic' was first introduced by Parker (1974) as "microorganisms or substances, which contributes to the intestinal microbial balance". Fuller (1989), explained probiotic as a live microbial feed supplement, which positively improved the host intestinal bacteria. In opposite with antibiotics, probiotics enhance life of the microbes, whereas antibiotics kill the microbes.

Species that are mostly used in probiotic preparations are Streptococcus thermophiles. Enterobacteria faecalis, Enterococcus faecium, Toulopsis sphaerica, Streptococci spp., Saccharomyces cerevisiae, Lactobacillus bulgaricus, L. casei, L. salvarius, L. acidophilus, L. helveticus, L. plantarum, L. Faecium, and Bifidobacteria species (Simon et al., 2001). Supplying the poultry with probiotics have shown to improve immune system, protein utilization, feed conversion ratio (FCR), reduce pathogenic bacteria and strengthen the nonpathogenic bacteria population in the gastrointestinal (GIT) system. Other than that, unicellular fungi and yeast products are also known for their fermentative ability. Feeding of non-pathogenic bacteria to the poultry can beneficially stimulate the immune system. Probiotic such as yeast can trigger several enzymes such as protease, cellulose, amylase, lipase, together with B complex vitamins in the medium in which they grow. Others benefits of feeding probiotic to broiler is it improved the intestinal tract health, enhanced protein utilization, reduced colonization of harmful bacteria in GIT of broiler, improved animals feed conversion ratio (FCR), and also improved nutrient synthesis.

Animals feed supplements have been used widely in livestock and poultry farming to improve growth performance, feed efficiency, and they are primarily of disease prevention nature like antibiotics, prebiotics, probiotics, and metabolites.

In recent years, consumers tend to be more aware and special attention have been paid for the safety of animal's feed products consumption. The application of antibiotic growth promoters (AGP) may develop some pathogen. resistance (Phillips et al., 2004). Since early 2006, the European Union prohibited the use of antibiotics as growth promoter (Patterson and Burkholder, 2003). Nowadays, modern broiler production in an intensive system with automation in feed and water delivery and with environmental control, huge number of birds can be reared in one house. Regarding to the increased bacterial resistance to antibiotics in human, it has triggered an increase in community and organizational attention in rejecting sub-therapeutic use of antibiotics in livestock (Patterson and Burkholder, 2003). Unfortunately, due to the elimination of this sub-therapeutic level of antibiotic in livestock, it might exposed the livestock with infectious disease and eventually will reduce the performance of the livestock, thus, according to Halloran, (2012), antibiotic have led to a dramatic reduction in illness and death from infectious diseases. Thus, this has led the industry to look for other alternative supplements that can replace antibiotics as a growth promoter and disease prevention.

In contrast with probiotics, the definition of prebiotics is they are not microorganisms. They are actually source of nourishment for existing intestinal microflora, promoting natural growth and replication. Prebiotics is an indigestible feed ingredients that benefit the host by selectively enhance the activities of one or many number of microorganisms in the host GIT, thus, beneficially improved the animals health (Gibson and Roberfroid, 1995). A prebiotic is a selectively fermented ingredient that gives changes, in both the activity in the gastrointestinal microbiota and the composition which gives benefits to the animals (FAO/WHO, 2002). Oligosaccharides, soluble fiber, and sugar molecules of three to six chains are commonly one identified as prebiotics.

To my knowledge, there is no relevant information on the meat quality parameter, abdominal fat in broiler fed with yeast, and also blood profile parameter in broiler chicken. Therefore, these parameters need to be investigate for additional information on the effects of yeast products on broilers.

1.1 Research Hypothesis

The supplementation of baker's yeast and yeast glucan to broilers will influence the growth performance, nutrient absorption, and improve carcass characteristic, and also will decrease the population of pathogenic bacteria in the intestinal tract of broiler.

1.2 Objectives

- I. To evaluate the effect of supplementing different levels of *Saccharomyces cerevisiae* on the nutrient digestibility performance, carcass characteristic, gut morphology and *Lactobacillus* spp. count in broiler's ileum.
- II. To determine the effect of supplementing *Saccharomyces cerevisiae* as a probiotic and yeast glucan as a prebiotic on the nutrient digestibility, performance, carcass characteristics, meat quality, *Lactobacillus* spp. and *E.coli* count, gut morphology, and blood hematology of broiler chicken.

REFERENCES

- Abaza, I. (2001). The use of some medicinal plants as feed additives in broiler diets. Ph.D. Thesis, Faculty of Agriculture, Alexandria University, Egypt.
- Abaza, I., Shehata, M. and Shoieb, M. (2006). Evaluation of some natural feed additive in layer diets. *Egyptian Poultry Science*. 26: 891-909.
- Abaza, I.M., Shehata, M.A., Shoieb, M.S. and Hassan, I.I. (2008). Evaluation of some natural feed additives in growing chick's diet. *International Journal of Poultry Science*. 7 (9): 872-879.
- Abdel-Azeem, F. (2002). Digeston, neomycin and yeast supplementation in broiler diets under Egyptian summer conditions. *Egyptian Poultry Science*. 22: 235-257.
- Abdelgader, O. (2006). Effect of dietary supplementation of yeast (*Saccharomyces cerevisiae*) on performance and carcass characteristics of broiler chicks. B.Sc. Thesis, Faculty of Animal Production, Khartoum University, Sudan.
- Abdelrahman, M.M. (2013). Effects of feeding dry fat and yeast culture on broiler chicken performance. *Turkish Journal of Veterinary and Animal Sciences.* 37: 31-37.
- Abudabos, A., AL-Batshan, H. and Murshed, M. (2015). Effects of prebiotics and probiotics on the performance and bacterial colonization of broiler chickens. *South African Journal of Animal Science*. 45: 419-428.
- Adebiyi, O., Makanjuola, B., Bankole, T. and Adeyori, A. (2012). Yeast culture (*Saccharomyces cerevisae*) supplementation: Effect on the performance and gut morphology of broiler birds. *Global Journal of Science Frontier Research.* 12.
- Adejumo, D.O., Onifade, T.O., Olutende, and Babatunde, G.M. (2005). The effects of concentration, age and duration of feeding supplemental yeast (Levucel SB) in high fibre diet on the performance of broiler chickens. The Proceedings of the first Nigeria International Poultry summit, held at the temperance, ota, Ogun State, Nigeria. pp: 69-76.
- AFRC, R.F. (1989) Probiotics in man and animals. *Journal of Applied Bacteriology*, 66: 365-378.
- Afsharmanesh, M., Barani, M. and Silversided, F. (2010). Evaluation of wetfeeding wheat-based diets containing *Saccharomyces cerevisiae* to broiler chickens. *British Poultry Science*. 51: 776-783.

- Aimanianda, V., Clavaud, C., Simenel, C., Fontaie, T., Delepierre, M. and Latage, J.-P. (2009). Cell Wall (1,6)-Glucan of Saccharomyces cerevisiae: structural characterization and *in situ* synthesis. Journal of Biological Chemistry. 284: 13401-13412.
- Alfaro, D., Silva, A., Borges, S., Maiorka, F., Vargas, S. and Santin, E. (2007). Use of Yucca schidigera extract in broiler diets and its effects on performance results obtained with different coccidiosis control methods. *The Journal of Applied Poultry Research*. 16: 248-254.
- Alloui, M. N., Szczurek, W. and Aswia Tkiewicz, S. (2013). The usefulness of prebiotics and probiotics in modern poultry nutrition: a review. *Annals* of *Animal Science*. 13: 17-32.
- AOAC (1997). Official methods of analysis, 16th Edition. Association of Official Analytical Chemists, Arlington, VA, USA. ISBN 0-935584-54-4.
- Bafundo, K., Cox, L. and Bywater, R. (2003). Review lends perspective to recent scientific findings on virginiamycin, antibiotic resistance debate. *Feedstuffs.* 75: 26-27.
- Banday, M. and Risam, K. (2001). Growth performance and carcass characteristics of broiler chicken fed with probiotics. *Indian Journal of Poultry Science*. 36: 252-255.
- Baurhoo, B., Letellier, A., Zhao, X. & Ruiz-Feria, C. (2007). Cecal populations of Lactobacilli and Bifidobacteria and Escherichia coli populations after in vivo Escherichia coli challenge in birds fed diets with purified lignin or mannanoligosaccharides. Poultry Science. 86: 2509-2516.
- Baurhoo, B., Ferket, P.R. and Zhao, X. (2009). Effects of diets containing different concentrations of mannanoligosaccharide or antibiotics on growth performance, intestinal development, cecal and litter microbial populations, and carcass parameters of broilers. *Poultry Science*. 88: 2262-2272.
- Beev, G., Todorova, P. and Tchobanova, S. (2007). Yeast cultures in ruminant nutrition. *Bulgarian Journal of Agricultural Science*. 13: 357-374.
- Benites, V., Gilharry, R., Gernat, A. and Murillo, J. (2008). Effect of dietary mannan oligosaccharide from Bio-Mos or SAF-Mannan on live performance of broiler chickens. *The Journal of Applied Poultry Research.* 17: 471-475.
- Bidarkar, V.K., Swain, P.S., Ray, S. and Dominic, G. (2014). Probiotics: Potential Alternative to Antibiotics in Ruminant Feeding. *Trends in Veterinary and Animal Sciences.* 1: 01-04.

- Blake, J., Hess, J., Macklin, K., Bilgili, S., Sefton, A. and Kocher, A. (2006). Mannan oligosaccharide (Bio-Mos) supplementation of wheat-based diets for broilers. *EPC 2006-12th European Poultry Conference, Verona, Italy, 10-14 September, 2006.* World's Poultry Science Association (WPSA).
- Bohn, J.A. and Bemiller, J.N. (1995). (1-3)-B-Glucans as biological response modifiers: a review of structure-functional activity relationships. *Carbohydrate Polymers.* 28: 3-14.
- Bonomi, A. & Vassia, G. (1978). Observations and remarks on use of Saccharomyces cerevisiae and Kluyeromyces, in form of living yeast, on production and quantitative characteristics of broilers. Archivio Veterinario Italiano. 29: 3-15.
- Bradley, G.L., Savage, T.F. & Timm, K.I. (1994). The effects of supplementing diets with *Saccharomyces cerevisiae var. boulardii* on male poult performance and ileal morphology. *Poultry Science*. 73: 1766-1770.
- Brown, G.D. & Gordon, S. (2005). Immune recognition of fungal β-glucans. *Cellular Microbiology*. 7: 471-479.
- Buzzini, P. & Vaughan-Martini, A. (2006) Yeast biodiversity and biotechnology. Biodiversity and Ecophysiology of Yeasts. Springer, 533-559.
- Cao, B.H., Karasawa, Y. & Guo, Y.M. (2005). Effects of green tea polyphenols and fructo-oligosaccharides in semi-purified diets on broilers' performance and caecal microflora and their metabolites. *Asian-Australasian Journal Animal Sciences*. 18: 85-89.

Casewell, M., Friis, C., Marco, E., Mcmullin, P. and Phillips, I. (2003). The European ban on growth-promoting antibiotics and emerging consequences for human and animal health. *Journal of Antimicrobial Chemotherapy*. 52: 159-161.

- Castagliuolo, I., Lamont, J.T., Nikulasson, S.T. and Pothoulakis, C. (1996). *Saccharomyces boulardii* protease inhibits *Clostridium* difficile toxin A effects in the rat ileum. *Infection and Immunity*, 64: 5225-5232.
- Cavazzoni, V., Adami, A. and Castrovilli, C. (1998). Performance of broiler chickens supplemented with *Bacillus coagulans* as probiotic. *British Poultry Science*. 39: 526-529.
- Charalampopoulus, D. and Rastall, R.A. (2009) *Prebiotics and Probiotics Science and Technology*. New York: Springer.
- Chen, Y. and Chen, T. (2004). Mineral utilization in layers as influenced by dietary oligofructose and inulin. *International Journal of Poultry Science*, 3: 442-445.

- Chiang, S. and Hsieh, W. (1995). Effect of direct-fed microorganisms on broiler growth performance and litter ammonia level. *Asian-Australasian Journal Animal Sciences*. 8: 159-162.
- Chichlowski, M., Croom, W., Eedens, F., Mcbride, B., Qiu, R., Chiang, C., Daniel, L., Havenstein, G. and Koci, M. (2007). Microarchitecture and spatial relationship between bacteria and ileal, cecal, and colonic epithelium in chicks fed a direct-fed microbial, PrimaLac, and salinomycin. *Poultry Science*. 86: 1121-1132.
- Choct, M. (2009). Managing gut health through nutrition. *British Poultry Science*. 50: 9-15.
- Choudhari, A., Shinde, S. and Ramteke, B. (2008). Prebiotics and probiotics as health promoter. *Veterinary World*. 1: 59-61.
- Chrintensen, H.R., Frokiaer, H. and Pestka, J.J. (2002). *Lactobacilli* differentially modulate expression of cytokines and maturation surface markers in murine dendritic cells. *The Journal of Immunology*. 168: 171-178.
- Cmiljanic, R., Lukic, M. and Trenkovski, S. (2001). The effect of 'Paciflor-C' probiotic on gain, feed conversion and mortality of fattening chicks. *Biotechnology in Animal Husbandary*. 2001; 17: 33-38.
- Contreras-Castillo, C. J., Brossi, C., Previero, T. C., and Dematte, L. C. (2008). Performance and carcass quality of broilers supplemented with antibiotics or probiotics. *Revista Brasileira de Ciencia Avicola*. 10: 227-232.
- Cummings, J. and Macfarlane, G. (2002). Gastrointestinal effects of prebiotics. *British Journal of Nutrition.* 87: S145-S151.
- Dake, M., Jadhv, J. and Patil, N. (2010). Variations of two pools of glycogen and carbohydrate in *Saccharomyces cerevisiae* grown with various ethanol concentrations. *Journal of Industrial Microbiology and Biotechnology.* 37: 701-706.
- Department of Standards Malaysia. (2005). *Malaysian Standard MS*. 316: 2006.
- Di Luzio, N.R. (1983). Immunopharmacology of glucan: a broad spectrum enhancer of host defense mechanisms. *Trends in Pharmacological Sciences*. 4: 344-347.
- Dierick, N. (1989). Biotechnology aids to improve feed and feed digestion: enzymes and fermentation. *Archives of Animal Nutrition.* 39: 241-261.

- Poultry feed metabolizable energy determination using total or partial excreta collection methods. *Revista Brasileira de Ciencia Avicola*, *12*(2): 129-132.
- Dunham, H., Williams, C., Edens, F., Casas, I. and Dobrogosz, W. (1993). *Lactobacillus reuteri* immunomodulation of stressor-associated diseases in newly hatched chickens and turkeys. *Poultry Science*. 72: 103.
- DVS. (1999). Department of Veterinary Services, Ministry of Agriculture, Malaysia.
- EC. (2001). Commission of the European Communities, Commission Recommendation, 2001/459/EC. *Official Journal of European Union L.* 161, 42-44
- EC. (2003). Commission of the European Communities, Commission Regulation (EC) No. 1831/2003. Official Journal of European Union L. 268, 29-43.
- Ezema, C. and Eze, D. (2009a). Performance and economic benefit of broilers fed palm kernel cake-based diet supplemented with probiotic. *International Journal of Poultry Science*. 8: 1003-1005.
- Ezema, C., Ezejimbe, C. C., Eze., D.C. and Kamalu, T.N. (2009b). Effect of probiotic (*Saccharomyces cerevisiae*) on digestibility and growth rate of pullets fed palm kernel cake based diet. A paper presented at the 34th. Annual Conference of the Nigerian Society of Animal Production at the University of Uyo, Nigeria: 344-346.
- Faysal, S. (2015). *Effect of various yeast levels on broiler performance*, Doctoral dissertation, UOFK.
- Ferket, P. (2003). Controlling gut health without the use of antibiotics. *Proceedings of the 30th Annual Carolina Poultry Nutrition Conference.* Citeseer.
- Ferket, P.R. (2004). Alternatives to antibiotics in poultry production: responses, practical experience and recommendations. *Nutritional Biotechnology in the Feed and Food Industries*. 57-67.
- Fernandez, F., Hinton, M. and Gils, B.V. (2002). Dietary mannanoligosaccharides and their effect on chicken caecal microflora in relation to *Salmonella enteritidis* colonization. *Avian Pathology*. 31: 49-58.

- Ferreira, C.L., Salminen, S., Grzeskowiak, L., Brizuela, M.A., Sanchez, L., Carneiro, H. and Bonnet, M. (2011). Terminology concepts of probiotic and prebiotic and their role in human and animal health. *Revista De Salud Animal.* 33: 137-139.
- Frahm, E. and Obst, U. (2003). Application of the fluorogenic probe technique (TaqMan PCR) to the detection of *Enterococcus* spp. and *Escherichia coli* in water samples. *Journal of Microbiological Methods*. 52: 123-131.
- Fuller, R. (1989). A review: probiotics in man and animals. *Journal of Applied Bacteriology.* 66: 365-378.
- Fuller, R. (1992). Probiotics: the scientific basis. *Springer Science and Business Media*. (pp. 398). London: Chapman and Hall.
- Gao, J., Zhang, H.J., Yu, S.H., Wu, S.G., Yoon, I., Quigley, J., Gao, Y.P. and Qi, G.H. (2008). Effects of yeast culture in broiler diets on performance and Immunomodulatory functions. *Poultry Science*. 87: 1377-1384.
- Garcia, I., Tajadura, V., Martin, V., Toda, T. and Sanchez, Y. (2006). Synthesis of alpha glucans in fission yeast spores is carried out by three alpha glucan synthase paralogues, Mok12p, Mok13p and Mok14p. *Molecular Microbiology*. 59: 836-853.
- Gardiner, T. (2000). Beta glucan biological activities: a review. *Glycoscience* and *Nutrition*. 1: 1-6.
- Giaimis, J., Lombard, Y., Fonteneau, P., Muller, C. D., Levy, R., Makaya-Kumba, M., Lazdins, J. & Poindron, P. (1993) Both mannose and betaglucan receptors are involved in phagocytosis of unopsonized, heatkilled *Saccharomyces cerevisiae* by murine macrophages. *Journal of Leukocyte Biology*, 54: 564-571.
- Gibson, G.R. and Roberfroid, M.B. (1995). Dietary modulation of the human colonic microbiota: Introducing the concept of prebiotics. *The Journal of Nutrition.* 125: 1401-1412.
- Giec, A. and Skupin, J. (1988). Single cell protein as food and feed. *Food/ Nahrung.* 32: 219-229.
- Glade, M. and Sist, M. (1988). Dietary yeast culture supplementation enhances urea recycling in the equine large intestine. *Nutrition Reports International.* 37: 11-17.
- Grba, S., Oura, E. and Suomalainen, H. (1975). On the formation of glycogen and trehalose in baker's yeast. *European Journal of Applied Microbiology and Biotechnology.* 2: 29-37.

- Grun, C.H., Hochstenbach, F., Humbel, B.M., Verkleij, A.J., Sietsma, J.H., Klis, F.M., Kamerling, J.P. and Vliegenthart, J.F. (2005). The structure of cell wall α-glucan from fission yeast. *Glycobiology*. 15: 245-257.
- Haghighi, H.R., Gong, J., Gyles, C.L., Hayes, M.A., Sanei, B., Parvizi, P., Gisavi, H., Chambers, J.R. and Sharif, S. (2005). Modulation of antibody-mediated immune response by probiotics in chickens. *Clinical and Diagnostic Laboratory Immunology*. 12: 1387-1392.
- Halloran, J. (2012). The overuse of antibiotics in food animals threatens public health. *Consumer Union Policy and Action For Consumer Reports*.
- Han, K., Kwon, I., Lohakare, J., Heo, S. and Chae, B. (2007). Chitooligosaccharides as an alternative to antimicrobials in improving performance, digestibility and microbial ecology of the gut in weanling pigs. Asian-Australian Journal of Animal Sciences. 20: 556.
- Hayat, J., Savage, T.F. and Mirosh, L.W. 1993). The reproductive performance of two genetically distinct lines of medium white turkey hens when fed breeder diets with and without a yeast culture containing *Saccharomyces cerevisiae*. *Animal Feed Science and Technology*. 43: 291-301.
- Higgins, J., Higgins, S., Wolfenden, A., Henderson, S., Torres-Rodriguez, A., Vicente, J., Hargis, B. and Tellez, G. (2010). Effect of lactic acid bacteria probiotic culture treatment timing on *Salmonella enteritidis* in neonatal broilers. *Poultry Science*. 89: 243-247.
- Hong, F., Hansen, R.D., Yan, J., Allendorf, D.J., Baran, J.T., Oostroff, G.R. and Ross, G.D. (2003). Glucan functions as an adjuvant for monoclonal antibody immunotherapy by recruiting tumoricidal granulocytes as killer cells. *Cancer Research*. 63: 9023-9031.
- Hooge, D.M., Sims, M.D., Sefton, A.E., Connolly, A. and Spring, P. (2003). Effect of dietary mannan oligosaccharide, with or without bacitracin or virginiamycin, on live performance of broiler chickens at relatively high stocking density on new litter. *The Journal of Applied Poultry Research.* 12: 461-467.
- Houshmand, M., Azhar, K., Zulkifli, I., Bejo, M.H., and Kamyab, A. (2012). Effects of non-antibiotic feed additives on performance, immunity and intestinal morphology of broilers fed different levels of protein. *South African Journal of Animal Science*, *42*: 22-32.
- Huang, M.K., Choi, Y.J, Houde, R., Lee, W.J, Lee, B. and Zho, X. (2004). Effects of *Lactobacilli* and an acidophilic fungus on the production performance and immune responses in broiler chickens. *Poultry Science.* 83: 788-795.

- Hunter, K., Gault, R. and Berner, M. (2002). Preparation of microparticulate βglucan from *Saccharomyces cerevisiae* for use in immune potentiation. *Letters in Applied Microbiology*. 35: 267-271.
- Hutchins, K. and Bussey, H. (1983). Cell wall receptor for yeast killer toxin: involvement of (1→6)-β-D-glucan. *Journal of Bacteriology*. 154: 161-169.
- Ignacio, E. (1995). Evaluation of the effect of yeast culture on the growth performance of broiler chick. *Poultry Science*. 74: 196.
- Immerseel, F.V., Buck, J.D., Pasmans, F., Huyghebaert, G., Haesebrouck, F. and Ducatelle, R. (2004). *Clostridium perfringens* in poultry: an emerging threat for animal and public health. *Avian Pathology.* 33: 537-549.
- Ismail, S. N., Sazili, A. Q., Idrus, Z., Meng, G. Y., Aghwan, Z. A., Sabow, A. B., and Abubakar, A. (2016). Effects of shackling and cone restraining on meat quality of broiler chickens slaughtered at two categories of live weight. *Journal of Biochemistry, Microbiology and Biotechnology*. 4: 15-19.
- Jadhav, J.P., Dake, M.S. and Patil, N.B. (2008), α,B-Glucan complex in Saccharomyces carlsbergenesis. Asian Journal of Chemistry. 20: 55-65.
- Jamas, S., Easson Jr, D.D. and Ostroff, G.R., Alpha-Beta Technology, Inc., 1998. Underivatized, aqueous soluable β (1-3) glucan, composition and method of making same. U.S. Patent 5,817,643.
- Jay, M. (1986). Modern food microbiology. 3rd Edition. Van Nostranel. New York .
- Jones, J. and Grey, T. (1995). Influence of processing on product quality and yield. *Processing of Poultry*. Springer. 127-181.
- Jordan, F.M., Gault, R.A. and Hunter, K.W. (2002). Method for preparing small particle size glucan in a dry material. U.S. Patent 6476003.
- Jozefiak, D., Rutkowski, A. and Martin, S. (2004). Carbohydrate fermentation in the avian ceca: a review. *Animal Feed Science and Technology*. 113: 1-15.
- Jozefiak, D., Kaczmarek, S. and Rutkowski, A. (2008). A note on the effects of selected prebiotics on the performance and ileal microbiota of broiler chickens. *Journal of Animal and Feed Sciences*. 17: 392-397.

- Kabir, S., Rahman, M., Rahman, M., Hosain, M., Akand, M. and Das, S. (2005). Viability of probiotics in balancing intestinal flora and effecting histological changes of crop and caecal tissues of broilers. *Biotechnology.* 4: 325-330.
- Kabir, S. L., Rahman, M., Rahman, M. and Ahmed, S. (2004). The dynamics of probiotics on growth performance and immune response in broilers. *International Journal Poultry Science*. 3: 361-364.
- Kanat, R. and Calialar, S. (1996). A research on the comparison effect on broiler chickens performance of active dried yeast and inactivated and stabilized probiotic yeast supplemented to the rations in different levels. *Poultry Science*. 75: 123.
- Karaoglu, M., Aksu, M., Esenbuga, N., Kaya, M., Macit, M. and Durdag, H. (2004). Effect of dietary probiotic on the pH and colour characteristics of carcasses, breast fillets and drumsticks of broilers. *Animal Science-Glasgow Then Penicuik*. 78: 253-260.
- Kath, F., and Kulicke, W. M. (1999). Mild enzymatic isolation of mannan and glucan from yeast *Saccharomyces cerevisiae*. *Die Angewandte Makromolekulare Chemie*. 268: 59-68.
- Kermanshahi, H. and Rostami, H. (2006). Influence of supplemental dried whey on broiler performance and cecal flora. *International Journal Poultry Science*. 5: 538-543.
- Klis, F.M., Mol, P., Hellingwerf, K. and Brul, S. (2002). Dynamics of cell wall structure in *Saccharomyces cerevisiae*. *FEMS Microbiology Reviews*. 26: 239-256.
- Klis, F.M., Boorsma, A. and De Groot, P.W. (2006). Cell wall construction in *Saccharomyces cerevisiae* yeast. 23: 185-202.
- Kocher, A., Canolly, A., Zawadzki, J. and Gallet, D. (2004). The challenge of finding alternatives to antibiotic growth promoters. *International Society for Animal Hygiene-Saint Malo.* 2004: 227-229.
- Kolida, S. and Gibson, G.R. (2011). Synbiotics in health and disease. *Annual Review of Food Science and Technology*. 2: 373-393.
- Kollar, R., Reinhold, B.B., Petrakova, E., Yeh, H.J., Ashwell, G., Drgonova, J., Kapteyn, J.C., Klis, F.M. and Cabib, E. (1997). Architecture of the yeast cell wall $\beta(1\rightarrow 6)$ -glucan interconnects mannoprotein, $\beta(1\rightarrow 3)$ glucan, and chitin. *Journal of Biological Chemistry*. 272: 17762-17775.

- Konca, Y., Kirkpinar, F., and Mert, S. (2009). Effects of mannanoligosaccharides and live yeast in diets on the carcass, cut yields, meat composition and colour of finishing turkeys. *Asian-Australia Journal Animal Science*. 22: 550-556.
- Kougias, P., Wei, D., Rice, P.J., Ensley, H.E., Kalbfleisch, J., Willams, D.L. and Browder, I.W. (2001). Normal human fibroblasts express pattern recognition receptors for fungal $(1\rightarrow 3)$ - β -D-glucans. *Infection and Immunity*. 69: 3933-3938.
- Kwiatkowski, S. and Kwiatkowski, S.E. (2012). Yeast (Saccharomyces cerevisiae) glucan polysaccharides-occurrence, separation and application in food, feed and health industries. INTECH Open Access Publisher.
- Lee, Y.K. and Salminen, S. (2009). *Handbook of probiotics and prebiotics*, John Wiley & Sons.
- Lesage, G. and Bussey, H. (2006). Cell wall assembly in Saccharomyces cerevisiae. Microbiology and Molecular Biology Reviews. 70: 317-343.
- Lillie, S.H. and Pringle, J.R. (1980). Reserve carbohydrate metabolism in *Saccharomyces cerevisiae*: responses to nutrient limitation. *Journal of Bacteriology*. 143: 1384-1394.
- Lindsey, T. (1985). Virginiamycin: Unique control of gut microbial metabolism enhances nutrient availability. *Proc. Pacesetter Conference Stafac* (virginiamycin) for Broilers. Smith Kline Animal Health Products, West Chester, PA.
- Line, J.E., Bailey, J.S., Cox, N.A., Stern, N.J. and Tompkins, T. (1998). Effect of yeast-supplemented feed on *Salmonella* and *Campylobacter* populations in broilers. *Poultry Science*. 77: 405-410.
- Liu, J.Y., Li, A.H., Ji, C. and Yang, W.M. (2009). First description of a novel Weissella species as an opportunistic pathogen for rainbow trout *Oncorhynchus mykiss* (Walbaum) in China. *Veterinary Microbiology*. 136: 314-320.
- Loddi, M.M., Gonzales, E., Takita, T.N.S., Mendes, A.A.N. and Roca, R.D.O. (2000). Effect of the use of probiotic and antibiotic on the performance, yield and carcass quality of broilers. *Revista Brasileira de Zootecnia*. 29: 1124-1131.
- Loh, T. C. (2004). Livestock production and the feed industry in Malaysia. In FAO Animal Production and Health Proceedings (FAO). FAO.

- Lund, B., Hansen, S. & Kurti, P. (2005) Efficacy of GalliPro-a microbial feed additive for broilers. *Proc.* 15th European Symposium on Poultry *Nutrition World's Poultry Science Association, Budapest, Hungary.*
- Kabir, L.S.M. (2009). The role of probiotics in the poultryindustry. *International Journal of Molecular Sciences*. 10: 3531-3545.
- Macfarlane, G., Steed, H. and Macfarlan, S. (2008). Bacterial metabolism and health-related effects of galactooligosaccharides and other prebiotics. *Journal of Applied Microbiology.* 104: 305-344.
- Madrigal, S., Watkins, S., Skinner, J., Adams, M., Waldroup, A. and Waldroup, P. (1993). Effect of an active yeast culture on performance of broilers. *Poultry Science.* 72: 87.
- Mahajan, P., Sahoo, J. and Panda, P. (1999). Research Articles-Effects of probiotic feeding and seasons on the growth performance and carcass quality of broilers. *Indian Journal of Poultry Science*. 34: 167-173.
- Makras, L., Triantafyllou, V., Fayol-Messaoudi, D., Adriany, T., Zoumpopoulou, G., Tsakalidou, E., Servin, A. and De Vuyst, L. (2006). Kinetic analysis of the antibacterial activity of probiotic *lactobacilli* towards *Salmonella enterica serovar Typhimurium* reveals a role for lactic acid and other inhibitory compounds. *Research in Microbiology*. 157: 241-247.
- Manners, D.J., Masson, A.J. and Patterson, J.C. (1973). The structure of a beta- $(1\rightarrow 6)$ -D-glucan from yeast cell walls. *Biochemistry Journal*. 135: 31-36.
- Marteau, P. and Rambaud, J.C. (1993). Potential of using lactic acid bacteria for therapy and immunomodulation in man. *FEMS Microbiology. Reviews.* 12: 207-220.
- Martin, S.A., Nisbet, D.J and Dean, R.G. (1989). Influence of a commerical yeast supplement on the *in vitro* ruminal fermentation. *Nutrition Reports International*. 40: 395-403.
- Martinez Amezcua, C., Parsons, C. M., and Noll, S. L. (2004). Content and relative bioavailability of phosphorus in distillers dried grains with solubles in chicks. *Poultry Science*. 83: 971-976.
- Mathur, S. and Singh, R. (2005). Antibiotic resistance in food lactic acid bacteria review. *International Journal of Food Microbiology*. 105: 281-295.
- Mccracken, V.J. and Gaskins, H.R. (1999). Probiotics and the immune system. *Probiotics: a critical review. Wymondham, UK: Horizon Scientific Press.* 85-111.

- Mc Donald, P., Edwards, R. A., and Greenhalgh, J. F. D. (1988). Animal Nutrtion 4 th edition. Logman Scientific and Technical. Essex, UK.
- McEwen, S. A., and Fedorka-Cray, P. J. (2002). Antimicrobial use and resistance in animals. *Clinical Infectious Diseases*. 34: S93-S106.
- Mikkelsen, L.L. and Jensen, B.B. (2000). Effect of fermented liquid feed on the activity and composition of the microbiota in the gut of pigs. *Pig News and Information*. 21: 59N-66N.
- MoA (Ministry of Agriculture and Agro-based Industry). (1998). Third National Agriculture Policy. *Kuala Lumpur, Malaysia.*
- Mohamed A. and El Zubier E. A. (1988). Some of modern technology toraise the efficiency of poultry production. A paper presented in 8th Arab Veterinary Conference. Khartoum, Sudan. March 1988.
- Mountzouris, K., Tsirtsikos, P., Kalamar, E., Nitsch, S., Schatzmair, G. and Fegeros, K. (2007). Evaluation of the efficacy of a probiotic containing *Lactobacillus, Bifidobacterium, Enterococcus,* and *Pediococcus* strains in promoting broiler performance and modulating cecal microflora composition and metabolic activities. *Poultry Science.* 86: 309-317.
- Mountzouris, K., Tsitrsikos, P., Palamidi, I., Arvaniti, A., Mohnl, M., Schatzmayr, G. and Fegeros, K. (2010). Effects of probiotic inclusion levels in broiler nutrition on growth performance, nutrient digestibility, plasma immunoglobulins, and cecal microflora composition. *Poultry Science.* 89: 58-67.
- Mourao, J.L., Pinheiro, V., Alves, A., Guedes, C., Pinto, L., Saavedra, M.J., Spring, P. and Kocher, A. (2006). Effect of mannan oligosaccharides on the performance, intestinal morphology and cecal fermentation of fattening rabbits. *Animal Feed Science and Technology*. 126: 107-120.
- Muihead, S. (1992). Direct- feed products. In S. Muihead (editor). Direct Feed microbial enzyme and forage additive compendium. 45-207.
- Mutus, R., Kocabagli, N., Alp, M., Acar, N., Eren, M. and Gezen, S. S. (2006). The effect of dietary probiotic supplementation on tibial bone characteristics and strength in broilers. *Poultry Science*. 85: 1621-1625.

NAP. (1998). Third National Agriculture Policy (1998-2010).

National Research Council. (1999). *The use of drugs in food animals: benefits and risks*. National Academies Press.

- Ngoc Lan, P.T., Binh, L.T. and Benno, Y. (2003). Impact of two probiotic Lactobacillus strains feeding on fecal *lactobacilli* and weight gains in chicken. *The Journal of General and Applied Microbiology*. 49: 29-36.
- Noverr, M. C. & Huffnagle, G. B. (2004) Does the microbiota regulate immune responses outside the gut? *Trends in Microbiology*, 12: 562-568.
- Oliveira, R.J., Matuo, R., Da Silva A.F., Matiazi, H.J., Mantovani, M.R.S.R. and Ribeiro, L.C.R. (2007). Protective effect of β-glucan extracted from *Saccharomyces cerevisiae*, against DNA damage and cytotoxicity in wild-type (k1) and repair-deficient (xrs5) CHO cells. *Toxicology in vitro*. 21: 41-52.
- Onifade, A., Babatunde, G., Afonja, S., Ademola, S. and Adesina, E. (1998). The effect of a yeast culture addition to a low-protein diet on the performance and carcass characteristics of broiler chickens. *Poultry Science*. 77: 44.
- Onwurah, F., Amaefule, K. and Ahamefule, F. (2014) Effect of baker's yeast (*Saccharomyces cerevisiae*) inclusion in feed and in drinking water on performance of broiler birds. *British Journal of Applied Science & Technology*, 4: 144.
- Owings, W.J., Reynolds, D.L., Hasiak, R.J. and Ferket, P.R. (1990). Influence of dietary supplementation with *Streptococcus faecium* M-74 on broiler body weight, feed conversion, carcass characteristics, and intestinal microbial colonization. *Poultry Science*. 69: 1257-1264.
- Oyedeji, J., Ajayi, H. and Egere, T. (2008). The Effects of increasing levels of yeast culture (Levucel SB) in a high fibre-diet on the performance and nutrient retention of broiler chicks. *Asian Journal of Poultry Science*. 2: 53-57.
- Panda, A., Reddy, M., Rao, S.R., Raju, M. and Praharaj, N. (2000). Growth, carcass characteristics, immunocompetence and response to *Escherichia coli* of broilers fed diets with various levels of probiotic. *Archives Fur Geflugelkunde.* 64: 152-156.
- Panda, A., Raju, M., Rama Rao, S. and Sharma, S. (2005). The influence of supplementation of *Lactobacillus sporogenes* on the performance of broilers. *Indian Journal of Animal Nutrition*. 22: 37-40.
- Parker, R.B. (1974). Probiotics, the other half of the antibiotic story. *Animal Nutrition Health.* 29: 4–8.
- Parks, C. W., Grimes, J. L., Ferket, P. R., & Fairchild, A. S. (2001). The effect of mannanoligosaccharides, bambermycins, and virginiamycin on performance of large white male market turkeys. *Poultry Science*. 80: 718-723.

- Paryad, A. and Mahmoudi, M. (2008). Effect of different levels of supplemental yeast (*Saccharomyces cerevisiae*) on performance, blood constituents and carcass characteristics of broiler chicks. *African Journal of Agricultural Research.* 3: 835-842.
- Patel, G. and Ingledew, W. (1975). The relationship of acid-soluble glycogen to yeast flocculation. *Canadian Journal of Microbiology*. 21: 1608-1613.
- Patterson, J. and Burkholder, K. (2003). Application of prebiotics and probiotics in poultry production. *Poultry Science*. 82: 627-631.
- Pelicano, E.R.L., De Souza, P., De Souza, H., Oba, A., Norus, E., Kodawara, L. and De Lima, T. (2003). Effect of different probiotics on broiler carcass and meat quality. *Revista Brasileira de Ciencia Avicola*. 5: 207-214.
- Pelicano, E.R.L., Souza, P., Souza, H., Oba, A., Boiago, M., Zeola, N., Scatolini, A., Bertanha, V. and Lima, T. (2005). Carcass and cut yields and meat qualitative traits of broilers fed diets containing probiotics and prebiotics. *Revista Brasileira de Ciencia Avicola*. 7: 169-175.
- Petracci, M., Betti, M., Bianchi, M. and Cavani, C. (2004). Color variation and characterization of broiler breast meat during processing in Italy. *Poultry Science.* 83: 2086-2092.
- Phillips, I., Casewell, M., Cox, T., De Groot, B., Friis, C., Jones, R., Nightingale, C., Preston, R. and Waddell, J. (2004). Does the use of antibiotics in food animals pose a risk to human health? A critical review of published data. *Journal of Antimicrobial Chemotherapy*. 53: 28-52.
- Pive, A. (1998). Non-conventional feed additives. *Journal of Animal and Feed Sciences. Supplement*, 7.
- Podmaniczky, B., Kocher, Z., Vegi, S., Korosi, K. and Molnar, A. (2006). The effect of mannan oligosaccharides on growth performance of challenged broilers. *12 European Poultry Conference.*
- Raju, M., Reddy, V., Rao, S.R. and Panda, A. (2006). Yeast: A multifunctional feed supplement for poultry-A review of the benefits of yeast in poultry diets. *Poultry International.* 45: 16-21.
- Reed, G. and T.W. Nagodawithana. (1999). Yeast technology (2nd Edn), van nostrand reinhold, New York. *Cited from Pakistan Journal Biotechnology Science*.

- Rice, P.J., Kelley, J.L., Kogan, G., Ensley, H.E., Kalbfleisch, J.H., Browder, I.W. and Williams, D.L. (2002). Human monocyte scavenger receptors are pattern recognition receptors for (1→3)-β-D-glucans. *Journal of Leukocyte Biology*. 72: 140-146.
- Rosen, G. (2007). Holo-analysis of the efficacy of Bio-Mos in broiler nutrition. British Poultry Science. 48: 21-26.
- Rostagno M., Wesley, I., Trampel, D. and Hurd, H. (2006). Salmonella prevalence in market-age turkeys on-farm and at slaughter. *Poultry Science*. 85: 1838-1842.
- Rothman-Denes, L.B. and Cabib, E. (1970). Two forms of yeast glycogen synthetase and their role in glycogen accumulation. *Proceedings of the National Academy of Sciences.* 66: 967-974.
- Sabiha, M., Elizabeth, V. and Jalaludeen, A. (2005). Effect of supplementation of probiotic on the growth performance of broiler chicken. *Indian Journal of Poultry Science*. 40: 73-75.
- Saied, J., Al-Jabary, Q. and Thalij, K. (2011). Effect of dietary supplement yeast culture on production performance and hematological parameters in broiler chicks. *International Journal Poultry Science*. 10: 376-380.
- Sakata, T., Kojima, T., Fujieda, M., Takahashi, M. and Michibata, T. (2003). Influences of probiotic bacteria on organic acid production by pig caecal bacteria *in vitro*. *Proceedings of the Nutrition Society*. 62: 73-80.
- Salyers, A.A., Gupta, A. and Wang, Y. (2004). Human intestinal bacteria as reservoirs for antibiotic resistance genes. *Trends in microbiology.* 12: 412-416.
- Samanya, M. and Yamauchi, K.E. (2002). Histological alterations of intestinal villi in chickens fed dried *Bacillus subtilis var. natto. Comparative Biochemistry and Physiology Part A: Molecular and Integrative Physiology.* 133: 95-104.
- Samarasinghe, K., Wenk, C., Silva, K. and Gunasekera, J. (2003). Turmeric (*Curcuma longa*) root powder and mannanoligosaccharides as alternatives to antibiotics in broiler chicken diets. *Asian-Australasian Journal of Animal Sciences*. 16: 1495-1500.
- Samli, H.E., Senkoylu, N., Koc, F., Kanter, M. and Agma, A. (2007). Effect of *Enterococcus faecium* and dried whey on broiler performance, gut histomorphology and intestinal microbiota. *Archives of Animal Nutrition*. 61: 42-49.

- Sams, A. and Mills K. (1993). The effect of feed withdrawal duration on the responsiveness of broiler pectoralis to rigor mortis acceleration. *Poultry Science*. 72: 1789-1796.
- Santin, E., Maiorka, A., Macari, M., Grecco, M., Sanchez, J., Okada, T. and Myasaka, A. (2001). Performance and intestinal mucosa development of broiler chickens fed diets containing *Saccharomyces cerevisiae* cell wall. *The Journal of Applied Poultry Research*. 10: 236-244.
- Scott. M .(1982). Nutrition of the chicken 3rd edition, Ithaca. New York.
- Smits, G. J., van den Ende, H., and Klis, F. M. (2001). Differential regulation of cell wall biogenesis during growth and development in yeast. *Microbiology*. 147: 781-794.
- Spahat, N. (2014). Environmental impact of different production systems and consumer willingness to pay for chicken meat produced with a higher regard for the environment. Land associate publishers.
- Sarkar, S., Mandal, L. and Banerjee, G. (1996). Comparative efficacy of different types of yeasts on the performance of broilers. *Indian Veterinary Journal.* 73: 224-226.
- Scross, M.L. (2002). Microbes versus microbes: immune signals generated by probiotic lactobacilli and their role in protection against microbial pathogens. *FEMS Immunology and Medical Microbiology*. 34: 245-253.
- Sedmak, J. (2006). *Production of beta-glucans nd mannans*. U.S. Patent Application 11/418,922.
- Sekhon, B. S. and Jairath, S. (2010) Prebiotics, probiotics and synbiotics: an overview. *Journal Pharmaceutical Education and Research.* 1: 13-36.
- Senani S., Rai, R., Padhi, M. and Saha, S. (1997). Effects of feeding different levels of *Lactobacilli* on the performance of broilers. *Indian Veterinary Journal.* 74: 808-810.
- Sharmila, A., Kasim, A., Noor, M., Jahromi, M. F., and Samsudin, A. A. (2015). Quantitative real-time PCR analysis of the caecal bacteria population of broiler chickens fed with corn-soy diet containing 20% of palm kernel meal with or without enzyme supplementation. *Journal Animal Poultry Science*. 4: 1-9.
- Shashidhara, R. and Devegowda, G. (2003). Effect of dietary mannan oligosaccharide on broiler breeder production traits and immunity. *Poultry Science*. 82: 1319-1325.

- Shen, Y., Piao, X., Kim, S., Wang, L., Liu, P., Yoon, I. and Zhen, Y. (2009). Effects of yeast culture supplementation on growth performance, intestinal health, and immune response of nursery pigs. *Journal of Animal Science*. 87: 2614-2624.
- Simmering, R. and Blaut, M. (2001). Pro-and prebiotics the tasty guardian angels?. Applied Microbiology and Biotechnology. 55: 19-28.
- Simon, O., Jadamus, A. and Vahjen, W. (2001). Probiotic feed additiveseffectiveness and expected modes of action. *Journal of Animal and Feed Sciences*. 10: 51-67.
- Sims, M.D., Dawson, K.A., Newman, K.E., Spring, P. and Hoogell, D.M. (2004). Effects of dietary mannan oligosaccharide, bacitracin methylene disalicylate, or both on the live performance and intestinal microbiology of turkeys. *Poultry Science*. 83: 1148-1154.
- Singh, S.,Niranjan, P., Singh, U., Koley, S. and Verma, D. (2009). Effects of dietary supplementation of probiotics on broiler chicken. *Animal Nutrition and Feed Technology*. 9: 85-90.
- Smirnov, A., Sklan, D. and Uni, Z. (2004). Mucin dynamics in the chick small intestine are altered by starvation. *The Journal of Nutrition.* 134: 736-742.
- Spring, P., Wenk, C., Dawson, K.A. and Newman, K.E. (2000). The effects of dietary mannaoligosaccharides on cecal parameters and the concentrations of enteric bacteria in the ceca of salmonella-challenged broiler chicks. *Poultry Science*. 79: 205-211.
- Stanley, V.G., Gray, C., Daley, M., Krueger, W.F. and Sefton, A.E. (2004). An alternative to antibiotic-based drugs in feed for enhancing performance of broilers grown on *Eimeria* Spp.-infected litter. *Poultry Science*. 83: 39-44.
- Swiatkiewicz, S., Koreleski, J. and Aczewska-Waosek, A. (2011). Effect of inulin and oligofructose on performance and bone characteristics of broiler chickens fed on diets with different concentrations of calcium and phosphorus. *British Poultry Science*. 52: 483-491.
- Tabidi, M.H., Mukhtar, A.M. and El-Rashied, E.L. (2013). Response of chicks for diet containing live yeast as probiotic natural feed additive. *Journal of Current Research in Science*. 1: 316-319.
- Teo, A.L. and Tan, H.M. (2006). Effect of *Bacillus subtilis* PB6 (CloSTAT) on broilers infected with a pathogenic strain of *Escherichia coli*. The Journal of Applied Poultry Research. 15: 229-235.

- Teo, A. and Tan, H.M. (2007). Evaluation of the performance and intestinal gut microflora of broilers fed on corn-soy diets supplemented with *Bacillus subtilis* PB6 (CloSTAT). *The Journal of Applied Poultry Research.* 16: 296-303.
- Thongsong, B., Kalandakanond-Thongsong, S. and Chavananikul, V. (2008). Effects of the addition of probiotic containing both bacteria and yeast or an antibiotic on performance parameters, mortality rate and antibiotic residue in broilers. *Thai Journal of Veterinary Medicine*. 38: 17.
- Toghyani, M., Toghyani, M. and Tabeidian, S. (2011). Effect of probiotic and prebiotic as antibiotic growth promoter substitutions on productive and carcass traits of broiler chicks. *International Conference on Food Engineering and Biotechnology. Singapura., Anais Singapura, v09.*
- Torres-Rodriguez, A., Donoghue, A., Donogue, D., Barton, J., Tellez, G. and Hargis, B. (2007). Performance and condemnation rate analysis of commercial turkey flocks treated with a *Lactobacillus* spp.-based probiotic. *Poultry Science*. 86: 444-446.
- Trachoo, N., Wechakama, P., Moongngarm, A. and Suttajit, M. (2008). Stability of freeze-dried *Lactobacillus acidophilus* in banana, soybean and pearl barley powders. *Journal of Biological Sciences*. 8: 119-124.
- Tuohy, K., Rouzaud, G., Bruck, W. and Gibson, G. (2005). Modulation of the human gut microflora towards improved health using prebioticsassessment of efficacy. *Current Pharmaceutical Design*. 11: 75-90.
- Uni, Z., Noy, Y. and Sklan, D. (1995). Posthatch changes in morphology and function of the small intestines in heavy-and light-strain chicks. *Poultry Science*. 74: 1622-1629.
- Valdivie, M. (1975). *Saccharomyces* yeast as a by-product from alcohol production on final molasses in diets for broilers. *Cuban Journal of Agricultural Science*. 9: 327-331.
- Vamanu, E. and Vamanu, A. (2010). The influence of prebiotics on bacteriocin synthesis using the strain *Lactobacillus paracasei* CMGB16. *African Journal of Microbiology Research.* 4: 534-537.
- Verstegen, M. W. and Williams, B. A. (2002) Alternatives to the use of antibiotics as growth promoters for monogastric animals. *Animal Biotechnology*, 13: 113-127.
- Vetvicka, V., Dvorak, B., Vetvickova, J., Richter, J., Krizan, J., Sima, P. and Yvin, J.C. (2007). Orally administered marine (1, 3)-Beta-D-glucan Phycarine stimulates both humoral and cellular immunity. *International Journal of Biological Macromolecules*. 40: 291-298.

- Vetvicka, V., Terayama, K., Mandeville, R., Brousseau, P., Kournikakis, B. and Ostroff, G. (2002). Pilot study: orally administered yeast β1, 3-glucan prophylactically protects against anthrax infection and cancer in mice. *Journal of American Nutraceutical Association.* 5: 1-5.
- Vetvicka, V. and Vetvickova, J. (2007). An evaluation of the immunological activities of commercially available β1, 3-glucans. *Journal of American Nutraceutical Association.* 10: 25-31.
- Vetvicka, V. and Yvin, J.C. (2004) Effects. of marine B-1, 3-glucan on immune reactions. *International Immunopharmacology.* 4: 721-730.
- Vicente, J., Wolfenden, A., Torres-Rodriguez, A., Higgins, S., Tellez, G. and Hargis, B. (2007). Effect of a *Lactobacillus* species-based probiotic and dietary lactose prebiotic on turkey poult performance with or without *Salmonella enteritidis* challenge. *The Journal of Applied Poultry Research.* 16: 361-364.
- Vicente, J.L., Torres-Rodriguez, A., Higgins, S.E., Pixley, C., Tellez, G., Donoghue, A.M. and Hargis, B.M. (2008). Effect of a selected *Lactobacillus* spp.-based probiotic on *Salmonella enterica serovar Enteritidis*-infected broiler chicks. *Avian Diseases*. 52: 143-146.
- Vinogradov, E., Petersen, B., and Bock, K. (1998). Structural analysis of the intact polysaccharide mannan from *Saccharomyces cerevisiae* yeast using 1H and 13C NMR spectroscopy at 750 MHz. *Carbohydrate Research*. 307: 177-184.
- Villar-Tajadura, M.A., Coll, P.M., Madrid, M., Cansado, J., Santos, B. and Pezer, P. (2008). Rga2 is a Rho2 GAP that regulates morphogenesis and cell integrity in *S. pombe. Molecular Microbiology*. 70: 867-881.
- Volman, J.J., Ramakers, J.D. and Plat, J. (2008). Dietary modulation of immune function by β-glucans. *Physiology and Behavior.* 94: 276-284.
- Vos, A., Dekker, N., Distel, B., Leunissen, J.A. and Hochstenbach, F. (2007). Role of the synthase domain of Ags1p in cell wall α-glucan biosynthesis in fission yeast. *Journal of Biological Chemistry.* 282: 18969-18979.
- Walker, K., Skelton, H. and Smith, K. (2002). Cutaneous lesions showing giant yeast forms of *Blastomyces dermatitidis*. *Journal of Cutaneous Pathology*. 29: 616-618.
- Wang, R.F., Cao, W.W. and Cerniglia, C.E. (1996). PCR detection and quantitation of predominant anaerobic bacteria in human and animal fecal samples. *Applied and Environmental Microbiology*. 62: 1242-1247.

- Watkins, B. and Kratzer, F. (1984). Drinking water treatment with a commercial preparation of a concentrated *Lactobacillus* culture for broiler chickens. *Poultry Science*. 63: 1671-1673.
- Wattanachant, S., Benjakul, S., and Ledward, D. A. (2004). Composition, color, and texture of Thai indigenous and broiler chicken muscles. *Poultry Science*. 83: 123-128.
- White,L.A., Newman, M.C., Cromwell, G.L., and Lindemann, M.D. (2002). Brewers dried yeast as a source of mannan oligosaccharides for weanling pigs. *Journal of Animal Science*. 80: 2619-2628.
- Williams, B.A., Verstegen, M.W. and Tamminga, S. (2001). Fermentation in the large intestine of single-stomached animals and its relationship to animal health. *Nutrition Research Reviews.* 14: 207-228.
- Wilson, W.A., Wang, Z. and Roach, P.J. (2002). Systematic identification of the genes affecting glycogen storage in the yeast *Saccharomyces cerevisiae* implication of the vacuole as a determinant of glycogen level. *Molecular and Cellular Proteomics.* 1: 232-242.
- Xu, Z., Hu, C., Xia, M., Zhan, X. and Wang, M. (2003). Effects of dietary fructooligosaccharide on digestive enzyme activities, intestinal microflora and morphology of male broilers. *Poultry Science*. 82: 1030-1036.
- Yaman, H., Ulukanli, Z., Elmali, M. and Unal, Y. (2006). The effect of a fermented probiotic, the kefir, on intestinal flora of poultry domesticated geese (*Anser anser*). *Review of Medicine Veterinar*. 157: 379-386.
- Yang, Y., Iji, P.A., Kocher, A., Thomson, E., Mikkelsen, L.L. and Choct, M. (2008). Effects of mannanoligosaccharide in broiler chicken diets on growth performance, energy utilisation, nutrient digestibility and intestinal microflora. *British Poultry Science*. 49: 186-194.
- Yason, C.V., Summers, B. and Schat, K. (1987). Pathogenesis of rotavirus infection in various age groups of chickens and turkeys: pathology. *American Journal of Veterinary Research.* 48: 927-938.
- Yeo, J. and Kim, K.I. (1997). Effect of feeding diets containing an antibiotic, a probiotic, or yucca extract on growth and intestinal urease activity in broiler chicks. *Poultry Science*. 76: 381-385.
- Yin, Y., Lei, F., Zhu, L., Li, S., Wu, Z., Zhang, R., Gao, G. F., Zhu, B. and Wang, X. (2009). Exposure of different bacterial inocula to newborn chicken affects gut microbiota development and ileum gene expression. *The ISME Journal (Multidisciplinary Journal of Microbial Ecology)*. 4: 367-376.

- Yusrizal and Chen, T.C (2003a). Effect of adding chicory fructans in feed on fecal and intestinal microflora and excreta volatile ammonia. *International Journal of Poultry Science*. 2: 188-194.
- Yusrizal and Chen, T.C. (2003b). Effect of adding chicory fructans in feed on broiler growth performance, serum cholesterol and intestinal length. International Journal of Poultry Science. 2: 214-219.
- Zdunczyk, Z., Jankowski, J. and Juskiewicz, J. (2005). Performance and intestinal parameters of turkeys fed a diet with inulin and oligofructose. *Journal of Animal and Feed Sciences. Supplement,* 14(Suppl.): 511-514.
- Zdunczyk, Z., Slominski, B., Jankowski, J. and Juskiewicz, J. (2011). Dietary content and gastrointestinal function of soybean oligosaccharides in monogastric animals. *Soybean-Biochemistry, Chemistry and Physiology.* InTech Open Access Publisher. pp. 523-540.
- Zhang, A.W., Lee, B.D., Lee, S.K., Lee, K.W., An, G.H., Song, K.B., and Lee, C.H. (2005). Effects of yeast (*Saccharomyces cerevisiae*) cell components on growth performance, meat quality, and ileal mucosa development of broiler chicks. *Poultry Science*. 87: 1015-1021.
- Zhang, X., Zhang, Y., Chen, H., Yang, J., and Wang, W. (2007). Study on spatial autocorrelation in China's animal feed industry. *New Zealand Journal of Agricultural Research*, 50: 831-838.
- Zhou, T., Chen, Y., Yoo, J., Huang, Y., Lee, J., Jang, H., Shin, S., Kim, H., Cho, J. and Kim, I. (2009). Effects of chitooligosaccharide supplementation on performance, blood characteristics, relative organ weight, and meat quality in broiler chickens. *Poultry Science*. 88: 593-600.
- Ziggers, D. (2000). Tos, a new prebiotic derived from whey. *Animal Feed Science and Technology*. 5: 34-36.