



UNIVERSITI PUTRA MALAYSIA

***EFFECTS OF LIGHT SOURCES AND DRYING METHODS ON PLANT
GROWTH AND STEVIOL GLYCOSIDES CONTENT OF STEVIA
(*Stevia rebaudiana* Bertoni)***

MOHD AZIZ BIN RASHID

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(*Stevia rebaudiana* Bertoni)**

By

MOHD AZIZ BIN RASHID

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of Master of Science**

June 2017

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

EFFECTS OF LIGHT SOURCES AND DRYING METHODS ON PLANT GROWTH AND STEVIOL GLYCOSIDES CONTENT OF STEVIA
(Stevia rebaudiana Bertoni)

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June 2017

Chairman : Phebe Ding, PhD
Faculty : Agriculture

Stevia (Stevia rebaudiana Bertoni) has received great attention with the rise in demand for low-sugar food and beverage additives, and natural alternative to sugarcane sugar and artificial sweeteners. The leaves produce intensively sweet steviol glycosides (mainly stevioside and rebaudioside A). *Stevia* is an obligate short-day plant with a critical daylength of 13 h. Daylength in Malaysia which is less than 13 h causes *stevia* to flower early, resulting in a low leaf yield and a low percentage of sweetener content. In the first experiment, the effects of night interruption technique using incandescent, fluorescent cool day light (C.D.L.), fluorescent warm white (W.W.), light-emitting diode (LED) C.D.L. and LED W.W. for 60 min daily to lengthen vegetative phase, on plant biomass and steviol glycoside content of *stevia* were investigated. The study was carried out using a nested design with a 6 x 4 factorial arrangements of six light sources and four growing weeks after night interruption initiated (week 2, 4, 6 and 8) with four replications. Results showed that night interruption technique extended vegetative phase from 20 days (control) to 120 days; thus allowing accumulation of plant biomass and steviol glycosides content. Leaf biomass and steviol glycosides of all light sources treated plants increased significantly as compared to control plants especially in week 6 and 8. Fluorescent W.W. can be considered as the best light source for night interruption technique of *stevia* as total steviol glycosides content per plant increased by 190-270% as compared to other light sources. The second experiment was conducted to determine the effects of several drying methods on steviol glycosides content of *stevia* leaves. Six drying method treatments were arranged in a completely randomized design (CRD) with four replications. Stevioside content was not significantly affected by the drying of sun, air, oven and freeze at different temperature to obtain pre-determined moisture content (MC). However, rebaudioside A content was significantly reduced by 3.38% under oven drying at 70 °C to obtain 10% MC. The reduction indicated that thermal degradation of rebaudioside A occurred at higher temperature. In conclusion, sun drying is the easiest and cheapest technique to dry *stevia* leaves without causing significant reduction of stevioside and rebaudioside A.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**KESAN SUMBER CAHAYA DAN KAEDAH PENGERINGAN KEPADA
PERTUMBUHAN POKOK DAN KANDUNGAN GLIKOSIDA STEVIOL
STEVIA (*Stevia rebaudiana* Bertoni)**

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Stevia (*Stevia rebaudiana* Bertoni) telah menerima banyak perhatian dengan pertambahan permintaan untuk makanan rendah gula dan bahan tambahan minuman, dan alternatif semulajadi kepada gula tebu dan pemanis tiruan. Daunnya menghasilkan glikosida steviol yang sangat manis (terutamanya steviosida dan rebaudiosida A). *Stevia* ialah tumbuhan hari pendek obligat dengan tempoh siang kritikal sebanyak 13 jam. Tempoh siang di Malaysia yang kurang dari 13 jam menyebabkan pokok *stevia* berbunga cepat, merendahkan hasil daun dan kandungan peratus pemanis yang rendah. Dalam eksperimen pertama, kesan teknik pecahan malam menggunakan lampu pijar, pendarfluor cool day light (C.D.L.), pendarfluor warm white (W.W.), diod pancaran cahaya (LED) C.D.L. and LED W.W. selama 60 minit pada setiap hari untuk memanjangkan fasa vegetatif, ke atas biomasa tumbuhan dan kandungan glikosida steviol telah dikaji. Kajian telah dijalankan menggunakan rekabentuk tersarang dengan 6 x 4 susunan faktorial iaitu enam sumber cahaya dan empat minggu pertumbuhan selepas pecahan malam bermula (minggu 2, 4, 6, dan 8) dengan empat replikasi. Keputusan menunjukkan teknik pecahan malam memanjangkan fasa vegetatif dari 20 hari (kawalan) kepada 120 hari; seterusnya membenarkan pengumpulan biomasa tumbuhan dan kandungan glikosida steviol. Biomasa daun dan glikosida steviol bagi pokok-pokok yang didedahkan dengan semua sumber cahaya menunjukkan peningkatan signifikan berbanding kawalan terutamanya pada minggu pertumbuhan ke 6 dan 8. Pendarfluor W.W. adalah sumber cahaya terbaik untuk teknik pecahan malam bagi *stevia* dengan peningkatan kandungan glikosida steviol sepokok sebanyak 190-270% berbanding sumber cahaya lain. Eksperimen kedua dijalankan untuk menentukan kesan beberapa kaedah pengeringan terhadap kandungan glikosida steviol daun *stevia*. Enam kaedah pengeringan disusun dalam rekabentuk penuh rawak (CRD) dengan empat replikasi. Kandungan steviosida tidak terjejas secara signifikan oleh pengeringan matahari, oven dan pembekuan pada suhu berbeza untuk mencapai kandungan kelembapan (MC) yang ditentukan lebih awal. Walau bagaimanapun, kandungan rebaudiosida A telah mengurang secara signifikan sebanyak 3.38% secara pengeringan oven pada 70 °C untuk mencapai 10% MC. Pengurangan ini menunjukkan degradasi

terma berlaku pada suhu tinggi. Kesimpulannya, pengeringan matahari adalah teknik paling mudah dan murah untuk mengeringkan daun stevia tanpa menyebabkan pengurangan signifikan kepada steviosida dan rebaudiosida A.



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I certify that a Thesis Examination Committee has met on 14 June 2017 to conduct the final examination of Mohd Aziz bin Rashid on his thesis entitled "Effects of Light Sources and Drying Methods on Plant Growth and Steviol Glycosides Content of Stevia (*Stevia rebaudiana* Bertoni)" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

AGR	Absolute growth rate
ANOVA	Analysis of Variance
AOP	Antioxidant properties
C.D.L.	Cool day light
CRD	Completely Randomized Design
CV	Coefficient of variation
DMRT	Duncan's Multiple Range Test
ELSD	Evaporative Light Scattering Detector
FDA	Food and Drug Administration
Fm	Maximum fluorescence
Fo	Initial fluorescence
Fv	Variable fluorescence
GA	Gibberellins
GRAS	Generally Recognized as Safe
HPLC	High Performance Liquid Chromatography
IBA	Indole-3-butyric acid
LAI	Leaf Area Index
LAR	Leaf area ratio
LED	Light-emitting diode
LD	Long-day
LSD	Least Significant Differences
MARDI	Malaysian Agricultural Research and Development Institute
MC	Moisture content
NPK	Nitrogen, phosphorus and potassium

NS	Non-significant
<i>P</i>	Probability
PAR	Photosynthetically Active Radiation
Pfr	Phytochrome far red
ppm	parts per million
Pr	Phytochrome red
rpm	revolutions per minute
RT	Retention time
<i>r</i>	Pearson's correlation coefficients
SAS	Statistical Analysis System
S.E.	Standard error
SLA	Specific leaf area
SD	Short-day
TPC	Total phenolic content
U.S.A.	United States of America
UV	Ultra-violet
WHO	World Health Organization
W.W.	Warm white
μl	Microliter

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CHAPTER 1

INTRODUCTION

1.1 Background

Stevia has received great attention with the rise in demand for low-carbohydrate, low-sugar food and beverage additives. Leaves of this plant produce a zero-calorie, non-nutritive, high potency sweetener with a sweetening value which is 150-450 times (by weight) higher than sucrose (Kinghorn, 1987). The sweeteners, containing steviol glycosides can be used as a natural alternative sweetener to other synthetic sweeteners such as aspartame, saccharine or acesulfame-K that constantly being associated with health concerns (Puri et al., 2011). Steviol glycosides are considered safe for consumption with no major contradictions, warnings and side effects reported (Ferri et al., 2006; Geuns, 2000).

Stevia was first introduced into Malaysia in the mid-1970's but failed to turn into a commercial crop mainly due to lack of suitable varieties, crop management and production technology (Ghawas et al., 2009). All variety of stevia introduced from other producing countries are photoperiod-sensitive. Daylength in Malaysia (which is less than 13 h) causes stevia to flower early, resulting in a low leaf yield per harvest and a low percentage of sweetener content. Early flowering also causes frequent harvesting and this leads to higher labour requirement and increases in production cost (Tan et al., 2008). The plants were harvested approximately every month (with lower leaf yield) as compared to 3-5 times annually in countries with longer daylength.

The use of classic incandescent lamps (rich in far-red light) as night interruption delayed the flowering and induced stevioside content of stevia (Zaidan et al., 1980) but it was considered not economical due to long hours of exposure required and less energy-efficiency. Commercial bulbs such as fluorescent lamps and light-emitting diode (LED) are more energy efficient, however its effect on plant biomass and flowering of stevia is still lacking. In this study, several types of commercial bulbs were used to impose night interruption in stevia plants and its effects to increase leaf biomass, delay flowering and induce steviol glycosides contents were determined.

Several drying methods are used to dry stevia leaves but the most preferable and practiced method is by sun drying (Samsudin and Aziz, 2013; Gates, 1997). However, sun drying will probably reduce the steviol glycosides content in the leaves. Abou-Arab et al. (2010) found that sun drying for 48 h caused 0.81% reduction of stevioside in stevia leaves. According to Steve Marsden of Herbal Advantage Inc., very little steviol glycoside will be lost if sun-drying for 8 h or less (Richard, 1996). However, proper experiment and scientific data are required to confirm the significance of steviol glycosides reduction by drying methods. The study on the effects of drying methods on steviol glycosides content is still lacking. In the second study, the effects of several drying methods on steviol glycosides content were determined.

1.2 Objectives

The objectives of the study were:

1. To determine the effects of different light sources as night interruption on vegetative and reproductive growth, and steviol glycosides content of stevia.
2. To determine the effects of several drying methods on steviol glycosides content of stevia.

1.3 Significance of study

The demand for stevia is growing due to increase cases of *Diabetes mellitus*, cardiovascular diseases, hypertension, obesity and other health problems among Malaysians, as well as concern over the safety of chemical sweeteners. Sweeteners from stevia plant are natural, zero-calorie, has a low glycemic index, can reduce blood sugar level and safe for consumption. However, the photoperiod-sensitive in stevia causes the plants to flower early that leads to low leaf yield and low sweetener content, thus preventing the commercial and large-scale production under local conditions. Previously, the study using light sources were merely to investigate the photoperiodism characteristics in stevia and not meant for agronomic purposes and technique for large scale production.

The technique of night interruption using commercial light sources in this study can be used to manipulate photoperiodism of stevia. The success of this technique will prevent early flowering, increase plant growth, leaf yield and sweetener content of stevia. Short duration of light exposure as night interruption will be cost effective and more environment friendly as compared to other technique such as day lengthening by continuous long hours of light exposure. Therefore, the findings of this study will give many benefits for local farmers and producers as well as researchers. The farmers and stevia producers can adopt this technique to overcome the agronomic problem and plant physiological challenges; thus will increase the production of stevia in Malaysia. The results from second study, which is the effects of drying methods on sweetener content, will benefit stevia producers in maintaining good quality of stevia at post harvest stage.

REFERENCES

- Abou-Arab, A.E., Abou-Arab, A.A. and Abu-Salem, M.F. 2010. Physico-chemical assessment of natural sweeteners steviosides produced from *Stevia rebaudiana* Bertoni plant. *African Journal of Food Science* 4(5): 269-281.
- Adams, S.R., Valdes, V.M. and Langton, F.A. 2008. Why does low intensity, long-day lighting promote growth in *Petunia*, *Impatiens* and tomato? *The Journal of Horticultural Science and Biotechnology* 83: 609-615.
- Anton, S.D., Martin, C.K., Han, H., Coulon, S., Cefalu, W.T., Geiselman, P. and Williamson, D.A. 2010. Effects of stevia, aspartame, and sucrose on food intake, satiety, and postprandial glucose and insulin levels. *Appetite* 55: 37-43.
- Anwar, S.H. and Kunz, B. 2011. The influence of drying methods on the stabilization of fish oil microcapsules: Comparison of spray granulation, spray drying, and freeze drying. *Journal of Food Engineering* 105(2): 367-378.
- Arena, C., Tsonev, T., Doneva, D., Micco, D.V., Michelozzi, M., Brunetti, C., Centritto, M., Fineschi, S., Velikova, V. and Loreto, F. 2016. The effect of light quality on growth, photosynthesis, leaf anatomy and volatile isoprenoids of a monoterpene-emitting herbaceous species (*Solanum lycopersicum* L.) and an isoprene-emitting tree (*Platanus orientalis* L.). *Environmental and Experimental Botany* 130: 122-132.
- Arguello, J.A., Nunezy, S.B. and Ledesma, A. 1997. Bulbing physiology in garlic (*Allium sativum* L.) cv. "Rosado Paraguay" III. Nutrient content in garlic plants: its relation to growth dynamics and bulb morphogenesis. *Acta Horticulturae* 433: 417-426.
- Armizatul, S.A.H., Azhar, K. and Nazrul, R. 2009. Influence of lengthening day on stevia vegetative growth, In *Transactions, Malaysian Society of Plant Physiology Conference*. Negeri Sembilan, Malaysia, November 23, 2009, pp. 17-18.
- Association of Official Analytical Chemists. 1990. *Official Methods of Analysis*. AOAC International: Washington, pp. 788-914.
- Aziz, M.R., Jamil, A.Z. and Armizatul, S.A.H. 2011. Night break delay flowering and increase yield of stevia. Paper presented at Malaysian Agricultural Research and Development Institute Science and Technology Exhibition. Selangor, Malaysia, December 5-7, 2011.
- Bakker, M., Bemrah, N., Hearty, A., Konig, J., Leblanc, C. and Tlustos, C. 2011. Revised exposure assessment for steviol glycosides for the proposed uses as a food additive. *European Food Safety Authority Journal* 9(1): 1972-1991.
- Barriocanal, L., Palacios, M., Benitez, G., Benitez, S., Jimenez J.T. and Jimenez, N. 2008. Apparent lack of pharmacological effect of steviol glycosides used as sweeteners in humans, a pilot study of repeated exposures in some

- normotensive and hypotensive individuals and in type 1 and type 2 diabetics. *Regulatory Toxicology and Pharmacology* 51(1): 37-41.
- Beemnet, M.K., Hassen, N., Gebremeskel, H., Mekonnen, S.A., Damtew, Z., Philipos, M., Melka, B. and Niide, T. 2013. Participatory development of quality seedlings for stevia (*Stevia rebaudiana* Bertoni) using stem cuttings. *World Journal of Agricultural Sciences* 9(2): 148-154.
- Benford, D.J., Hill, F., Schlatter, J. and DiNovi, M. 2009. *Safety Evaluation of Certain Food Additives: Steviol glycosides (addendum)*. 60th Joint FAO/WHO Expert Committee on Food Additives (JECFA): Geneva, pp. 117-144.
- Bespalkok-Filho, J.C. and Hattori, K. 1997. Embryogenic callus formation and histological studies from *Stevia rebaudiana* (Bert.) Bertoni floret explants. *The Revista Brasileira de Fisiologia Vegetal* 9: 185-188.
- Bjorkman, O. and Demmig, B. 1987. Photon yield of O₂ evolution and chlorophyll fluorescence characteristics at 77K among vascular plants of diverse origins. *Planta* 170: 489-504.
- Blanchard, M.G. and Runkle, E.S. 2009. Use of a cyclic high-pressure sodium lamp to inhibit flowering of chrysanthemum and velvet sage. *Scientia Horticulturae* 122: 448-454.
- Blanchard, M.G. and Runkle, E.S. 2010. Intermittent light from a rotating high-pressure sodium lamp promotes flowering of long-day plants. *HortScience* 45: 236-241.
- Bondarev, N.I., Sukhamova, M.A., Reshetnyak, O.V. and Nosov, A.M. 2003. Steviol glycoside content in different organs of *Stevia rebaudiana* and its dynamics through ontogeny. *Biologia Plantarum* 47(2): 261-264.
- Bracht, A.K., Alvarez, M. and Bracht, A. 1985. Effects of *Stevia rebaudiana* natural products on rat live mitochondria. *Biochemistry Pharmacology* 34: 873-882.
- Brandle, J.E. and Rosa, N. 1992. Heritability for yield, leaf:stem ratio and stevioside content estimated from landrace cultivar of *Stevia rebaudiana*. *Canadian Journal of Plant Science* 72: 1263-1266.
- Cathey, H.M. and Borthwick, H.A. 1961. Cyclic lighting for controlling flowering of chrysanthemum. *Proceedings of the American Society for Horticultural Science* 78: 545-552.
- Ceunen, S. and Geuns, J.M.C. 2013. Influence of photoperiodism on the spatio-temporal accumulation of steviol glycosides in *Stevia rebaudiana* (Bertoni). *Plant Science* 198: 72-82.
- Ceunen, S., Werbrouck, S. and Geuns, J.M.C. 2012. Stimulation of steviol glycoside accumulation in *Stevia rebaudiana* by red LED light. *Journal of Plant Physiology* 169: 749-752.

- Chalapathi, M.V., Shivaraj, B. and Ramakrishna, P.V.R. 1997. Nutrient uptake and yield of stevia (*Stevia rebaudiana* Bertoni) as influenced by methods of planting and fertilizer levels. *Crop Research* 14: 205-208.
- Chalapathi, M.V., Thimmegowda, S., Kumar, N.D., Rao, G.G.E. and Mallikarjuna, K. 2001. Influence of length of cutting and growth regulators on vegetative propagation of stevia (*Stevia rebaudiana* Bert.). *Crop Research* 21: 53-56.
- Chan, E.W.C., Lim, Y.Y., Wong, K.K., Lim, K.K., Tan, S.P., Lianto, F.S. and Yong, M.Y. 2009. Effects of different drying methods on the antioxidant properties of leaves and tea of ginger species. *Food Chemistry* 113: 166–172.
- Chan, P., Xu, D.Y., Liu, J.C., Chen, Y.J., Tomlinson, B., Huang, W.P. and Cheng, J. T. 1998. The effect of stevioside on blood pressure and plasma catecholamines in spontaneously hypertensive rats. *Life Science* 63: 1679-1684.
- Chatsudthipong, V. and Muanprasat, C. 2009. Stevioside and related compounds: Therapeutic benefits beyond sweetness. *Pharmacology and Therapeutics* 121: 41-54.
- Chen, K., Chang, T.R. and Chen, S.T. 1978. Studies on the cultivation of stevia and seasonal variation of stevioside. *China Gartenbau* 24: 34–42.
- Chen, T.H., Chen, S.C., Chan, P., Chu, Y L., Yang, H.Y. and Cheng, J.T. 2005. Mechanism of the hypoglycemic effect of stevioside, a glycoside of *Stevia rebaudiana*. *Planta Medica* 71: 108-113.
- Chranioti, C., Chanioti, S. and Tzia, C. 2016. Comparison of spray, freeze and oven drying as a means of reducing bitter aftertaste of steviol glycosides (derived from *Stevia rebaudiana* Bertoni plant) – Evaluation of the final products. *Food Chemistry* 190: 1151-1158.
- Crammer, B. and Ikan, R. 1986. Sweet glycosides from the stevia plant. *Chemistry in Britain* 22: 915-916.
- Dacome, A.S., Silva, C.C., Costa, C.E.M., Fontana, J. D., Adelman, J. and Costa, S.C. 2005. Sweet diterpenic glycosides balance of a new cultivar of *Stevia rebaudiana* (Bert) Bertoni: isolation and quantitative distribution by chromatographic, spectroscopic and electrophoretic methods. *Process Biochemistry* 44: 3587-3594.
- Darise, M., Kohda, H., Mizutani, K., Kasai, R. and Tanaka, O. 1983. Chemical constituents of flowers of *Stevia rebaudiana* Bertoni. *Agricultural and Biological Chemistry* 47: 133-135.
- DiCristina, K. and Germino, M. 2006. Correlation of neighbourhood relationships, carbon assimilation and water status of sagebrush seedlings establishing after fire. *Western North American Naturalist* 66(4): 441-449.
- Doymaz, I. and Pala, M. 2003. The thin-layer drying characteristics of corn. *Journal of Food Engineering* 60: 125–130.

- Dube, K.G. 2011. Absolute growth rate, leaf area index, leaf:stem ratio and harvest index influenced by organic manures, biofertilizers and growth regulators in *Stevia rebaudiana* Bertoni. *Asiatic Journal of Biotechnology Resources* 2(5): 508-521.
- DuBois, G.E. 2000. Sweeteners: non-nutritive. In *Encyclopedia of Food Science and Technology*, ed. F.J. Francis, pp. 2245-2265. New York: John Wiley and Sons, Inc.
- Dwivedi, R.S. 1999. Unnurtured and untapped super sweet nonsacchariferous plant species in India. *Current Science* 76: 1454-1461.
- Ermakov, E.I. and Kochetov, A.A. 1996. Specific features in growth and development of stevia plants under various light regimes in regulated conditions. *Doklady Rossiiskoi Akademii Selskokhozyaistvennykh Nauk* 1: 8-9.
- Ferri, L.A, Alves-Do-Prado, W., Yamada, S.S., Gazola, S., Batista, M.R. and Bazotte, R.B. 2006. Investigation of the antihypertensive effect of oral crude stevioside in patients with mild essential hypertension. *Phytotherapy Research* 20(9): 732-736.
- Frederico, A.P., Ruas, P.M., Marin-Morales, M.A., Fuas, C.F. and Nakajima, J.N. 1996. Chromosome studies in some stevia (Compositae) species from southern Brazil. *Brazilian Journal of Genetics* 19: 605-609.
- Gao, Q., Zhao, P., Zeng, X., Cai, X. and Shen, W. 2002. A model of stomatal conductance to quantify the relationship between leaf transpiration, microclimate and soil water stress. *Plant, Cell & Environment* 25: 1373-1381.
- Gates, D. 1997. The stevia story-how to grow stevia. Retrieved on 10 January 2011 from <http://www.stevia.net/growingstevia.htm>
- Geuns, J.M.C. 2000. Safety of stevia and stevioside. *Phytochemistry* 4: 75-88.
- Geuns, J.M.C. 2008. Steviol glycosides: technical and pharmacological aspects, In *Proceedings of the 2nd EUSTAS Stevia Symposium*. Heverlee, Belgium, June 27, 2008, pp. 59-78.
- Gentry, A.H. 1996. *A field Guide of the Families and Genera of Woody Plants of Northwest South America (Colombia, Ecuador, Peru) with Supplementary Notes on Herbaceous Taxa*, p. 895. Chicago, Illinois: The University of Chicago Press.
- Ghawas, M.M., Puteh, Azhar, M.A.B., Zaki, W.M. and Zawayi, M. 2009. Penanaman dan pengeluaran tanaman stevia. *Buletin Teknologi Tanaman* 6: 35-45 (In Malay language).
- Gisleine, E.C., Abdol, H.A. and Claudio, C.A. 2006. Investigation of the tolerability of oral stevioside in Brazilian hyperlipidemic patients. *Brazilian Archives of Biology and Technology* 49: 583-587.

- Goettemoeller, J. and Ching, A. 1999. Seed germination in *Stevia rebaudiana*. In *Perspectives on New Crops and New Uses*, ed. J. Janick, pp. 510-511. Alexandria, Virginia: ASHS Press.
- Gvasaliya, V.P., Kovalenko, N.V. and Garguliya, M.C. 1990. Studies on the possibility of growing honey grass in Abkhazia conditions. *Subtropicheskie Kultury* 5: 149-156.
- Hanson, J.R. and White, A.F. 1968. Studies in terpenoid biosynthesis-II: The biosynthesis of steviol. *Phytochemistry* 7: 595-597.
- Hatter, K. 2010. How to dry stevia leaves? Retrieved on 1 December 2010 from http://www.ehow.com/how_7253313_dry-stevialeaves.html
- Heo, J.W., Lee, C.W., Murthy, H.N. and Paek, K.Y. 2003. Influence of light quality and photoperiod on flowering of *Cyclamen persicum* Mill. cv. 'Dixie White'. *Plant Growth Regulation* 40: 7-10.
- Higuchi, Y., Sumitomo, K., Oda, A., Shimizu, H. and Hisamatsu, T. 2012. Day light quality affects the night-break response in the short-day plant chrysanthemum, suggesting differential phytochrome-mediated regulation of flowering. *Journal of Plant Physiology* 169: 1789-1796.
- Hopkins, W.G. and Huner, N.P.A. 2008. Measuring time: Controlling development by photoperiod and endogenous clocks. In *Introduction to Plant Physiology*, pp. 413-432. London and Ontario: John Wiley and Sons, Inc.
- Hunt, R. 1990. *Basic Growth Analysis*, pp. 17-81. London: Academic Division of Unwind Hyman Ltd.
- Hunt, R. 2003. Growth and development: Growth analysis, individual plants. In *Encyclopedia of Applied Plant Sciences*, ed. B. Thomas, pp. 579-588. San Diego California: Elsevier Academic Press.
- Islam, N., Patil, G.G. and Gislerod, H.R. 2005. Effect of photoperiod and light integral on flowering and growth of *Eustoma grandiflorum* (Raf.) Shinn. *Scientia Horticulturae* 103: 441-451.
- Jaitak, V., Gupta, A.P., Kaul, V.K. and Ahuja, P.S. 2008. Validated high-performance thin-layer chromatography method for steviol glycosides in *Stevia rebaudiana*. *Journal of Pharmaceutical and Biomedical Analysis* 47: 790-794.
- Jaroslav, P., Barbora, H. and Tuulia, H. 2006. Characterization of *Stevia rebaudiana* by comprehensive two-dimensional liquid chromatography time-of-flight mass spectrometry. *Journal of Chromatography* 1150(1-2): 85-92.
- Jeppesen, P.B., Gregersen, S., Rolfsen, S.E.D., Jepsen, M., Colombo, M., Agger, A., Xiao, J., Kruhoffer, M., Orntoft, T. and Hermansen, K. 2003. Antihyperglycemic and blood pressure-reducing effects of stevioside in the diabetic gotokakizaki rat. *Metabolism* 52: 372-378.

- Ji, H., Du, A., Zhang, L., Xu, C., Yang, M. and Li, F. 2012. Effect of drying methods on antioxidant properties in *Robinia pseudoacacia* L. flower. *Journal of Medicinal Plant Research* 6(16): 3233-3239.
- Joint Food and Agriculture Organization/World Health Expert Committee on Food Additives. 2005. Evaluation of Certain Food Additives (Rep. No. 63). Geneva: World Health Organization, pp. 34-39.
- Kade, I.J., Ibukun, E.O., Nogueira, C.W. and Rocha, D.A. 2008. Sun-drying diminishes the antioxidative potentials of leaves of *Eugenia uniflora* against formation of thiobarbituric acid reactive substances induced in homogenates of rat brain and liver. *Experimental and Toxicologic Pathology* 60: 365-371.
- Kang, K.H. and Lee, E.W. 1981. Physio-ecological studies on stevia (*Stevia rebaudiana* Bertoni). *Korean Journal of Crop Science* 26: 69-89.
- Kang, K.J., Oh, W., Shin, J.H. and Kim, K.S. 2008. Night interruption and cyclic lighting promote flowering of *Cyclamen persicum* under low temperature regime. *Horticulture, Environment, and Biotechnology* 49: 72-77.
- Kennely, E.J. 2002. Sweet and non-sweet constituents of *Stevia rebaudiana*. In *Stevia, the Genus of Stevia, Medicinal and Aromatic Plants*, ed. A.D. Kinghorn, pp. 68-85. London: Taylor and Francis Group.
- Kim, H.J., Jung, H.H. and Kim, K.S. 2011b. Influence of photoperiod on growth and flowering of dwarf purple loosestrife. *Horticulture, Environment, and Biotechnology* 52: 1-5.
- Kim, Y.J., Lee, H.J. and Kim, K.S. 2011a. Night interruption promotes vegetative growth and flowering of *Cymbidium*. *Scientia Horticulturae* 130: 887-893.
- King, R.M. and Robinson, H. 1987. The genera of the Eupatorieae (Asteraceae). In *Monographs in Systematic Botany from the Missouri Botanical Garden*, pp. 1-581. St. Louis: Missouri Botanical Garden.
- Kinghorn, A.D. 1987. Biologically active compounds from plants with reputed medical and sweetening properties. *Journal of Natural Products* 50: 1009-1024.
- Kinghorn, A.D. and Soejarto, D.D. 1985. Current status of stevioside as a sweetening agent for human use. In *Economic and Medical Plant Research*, ed. H. Wagner, H. Hikino, and N.R. Farnsworth, pp 1-51. London: Academic Press.
- Kubota, S., Yamato, T., Hisamatsu, T., Esaki, S., Oi, R., Roh, M.S. and Koshioka, M. 2000. Effects of red and far-red rich spectral treatments and diurnal temperature alternation on the growth and development of *Petunia*. *Journal of the Japanese Society for Horticultural Science* 69(4): 403-409.
- Larrauri, J.A., Ruperez, P. and Saura-Calixto, F. 1997. Effect of drying temperature on the stability of polyphenols and antioxidant activity of red grape pomace peels. *Journal of Agricultural and Food Chemistry* 45: 1390-1393.

- Leuning, R. 1995. A critical appraisal of combined stomatal-photosynthesis model for C3 plants. *Plant, Cell & Environment* 18: 339-355.
- Lewis, W.H. 1992. Early uses of *Stevia rebaudiana* (Asteraceae) leaves as a sweetener in Paraguay. *Economic Botany* 46: 336-337.
- Li, J., Li, G., Wang, H. and Deng, X.W. 2011. Phytochrome Signaling Mechanisms. *The Arabidopsis Book* 9: 1-26.
- Lim, Y.Y. and Murtijaya, J. 2007. Antioxidant properties of *Phyllanthus amarus* extracts as affected by different drying methods. *Food Science and Technology* 40: 1664-1669.
- Lin, S.D., Sung, J.M. and Chen, C.L. 2011. Effect of drying and storage conditions on caffeic acid derivatives and total phenolics of *Echinacea purpurea* grown in Taiwan. *Food Chemistry* 125: 226–231.
- Ma, L., Chen, H., Zhu, W. and Wang, Z. 2013. Effect of different drying methods on physicochemical properties and antioxidant activities of polysaccharides extracted from mushroom *Inonotus obliquus*. *Food Research International* 50(2): 633-640.
- Madan, S., Ahmad, S., Singh, G.N., Kohli, K., Kumar, Y., Singh, R. and Gary, M. 2010. *Stevia rebaudiana* (Bert.) Bertoni – A review. *Indian Journal of Natural Products and Resources* 1(3): 267-286.
- Maiti, R.K. and Purohit, S.S. 2008. *Stevia: A Miracle Plant for Human Health*, pp. 1-178. Jodhpur: Agrobios.
- Megeji, N.W., Kumar, J.K., Virendra, S., Kaul, V.K. and Ahuja, P.S. 2005. Introducing *Stevia rebaudiana*, a natural zero-calorie sweetener. *Current Science* 88: 801-804.
- Melis, M.S. 1992. Renal excretion of steviosides in rats. *Journal of Natural Products* 55: 688-690.
- Melis, M.S. 1999. Effects of chronic administration of *Stevia rebaudiana* on fertility in rats. *Journal of Ethnopharmacology* 67: 157-161.
- Metivier, J. and Viana, A.M. 1979. The effect of long and short day length upon the growth of whole plants and the level of soluble proteins, sugars and stevioside in leaves of *Stevia rebaudiana* Bert. *Journal of Experimental Botany* 30: 1211-1222.
- Midmore, D.J. and Rank, A.H. 2002. *A new rural industry-stevia to replace imported chemical sweeteners*. Rural Industries Research and Development Corporation, pp. 3-16. Canberra: RIRDC Web Publication.
- Nakamura, S. and Tamura, Y. 1985. Variation in the main glycosides of stevia (*Stevia rebaudiana* Bertoni). *Japanese Journal of Tropical Agriculture* 29: 109-115.

- Nassar, A.M., Besheit, S.Y. and Allam, A.I. 2001. Effect of sowing dates, harvesting dates and their interaction on yield and quality of stevia (*Stevia rebaudiana* (Bert.) Bertoni) in Egypt. *Egyptian Journal of Applied Sciences* 16: 442-459.
- Nishiyama, P., Kusumoto, I.T., Costa, S.C., Alvarez, M. and Vieira, L.G.E. 1991. Correlation between the contents of total carbohydrates and steviosides in leaves of *Stevia rebaudiana*. *Arquivos De Biologia E Tecnologia* 34: 425-434.
- Oddone, B. 1999. *How to Grow Stevia*, pp. 1-30. Connecticut: Gaurani Botanicals, Inc.
- Ohta, M., Sasa, S., Inoue, A., Tamai, T., Fujita, I., Morita, K. and Matsuura, F. 2010. Characterization of novel steviol glycosides from leaves of *Stevia rebaudiana* Morita. *The Japanese Society of Applied Glycoscience* 57: 199-209.
- Orphanides, A., Goulas, V. and Gekas, V. 2013. Effect of drying method on the phenolic content and antioxidant capacity of Spearmint. *Czech Journal of Food Sciences* 5: 509-513.
- Park, Y.J., Kim, Y.J. and Kim, K.S. 2013. Vegetative growth and flowering of *Dianthus*, *Zinnia* and *Pelargonium* as affected by night interruption at different timings. *Horticulture, Environment, and Biotechnology* 54: 236-242.
- Periche, A., Castello, M.L., Heredia, A. and Escriche, I. 2015. Influence of drying method on steviol glycosides and antioxidants in *Stevia rebaudiana* leaves. *Food Chemistry* 172: 1-6.
- Puri, M., Sharma, D. and Tiwari, A. 2011. Downstream processing of stevioside and its potential applications. *Biotechnology Advances* 29(6): 781-791.
- Raji, A.A. and Mohamad, O. 2011. Effects of stem cutting types, position and hormonal factors on rooting in *Stevia rebaudiana* Bertoni. *Journal of Agricultural Science* 4(1): 49-57.
- Ralph, P. J. and Gademan, R. 2005. Rapid light curves: A powerful tool to assess photosynthetic activity. *Aquatic Botany* 82: 222-237.
- Ren, G., Liu, X. and Shi, Y. 2011. Effects of plant growth regulator S-Y on diurnal changes in photosynthetic parameters and yield of *Stevia Rebaudina* Bertoni. *Energy Procedia* 5: 429-434.
- Richard, D. 1996. Botany and cultivation. In *Stevia rebaudiana : Natures Sweet Secret*, pp. 10-13. Illinois: Blue Heron Press.
- Rojas, C.G. and Miranda, P. 2002. HPLC isolation and structural elucidation of diastereomeric nilolyl ester tetrasachharides from Mexican scammony root. *Tetrahedron* 58: 3145-3154.
- Roper, T.R., Keller, J.D., Loscher, W.H. and Rom, C.R. 1988. Photosynthesis and carbohydrate partitioning in sweet cherry: Fruiting effects. *Plant Physiology* 72: 42-47.

- Runkle, E. and Fisher, P., 2004. Photoperiod and flowering. In *Lighting Up Profits: Understanding Greenhouse Lighting*, pp. 25-32. Willoughby Ohio: Meister Media Worldwide.
- Samsudin, A. and Aziz, I.A. 2013. Drying of stevia leaves using laboratory and pilot scale dryers. *Journal of Tropical Agriculture and Food Science* 41(1): 137–147.
- Savita, S.M., Sheela, K., Sharan, S., Shankar, A.G., Parama, R. and Srinivasakey, M. 2004a. Health implications of *Stevia rebaudiana*. *Journal of Human Ecology* 15: 191-94.
- Savita, S.M., Sheela, K., Sharan, S., Shankar, A.G. and Parama, R. 2004b. *Stevia rebaudiana* – A functional component for food industry. *Journal of Human Ecology* 15: 261-264.
- Schaper, H. and Chacko, E.K. 1993. Effects of irradiance, leaf age, chlorophyll content and branch girdling on gas exchange of cashew (*Anacardium occidentale* L. leaves. *HortScience* 68: 541-550.
- Sekaran, T., Giridhar, P. and Ravishankar, G.A. 2007. Production of steviosides in *ex vitro* and *in vitro* grown *Stevia rebaudiana* Bertoni. *Journal of the Science of Food and Agriculture* 87: 420-424.
- Serfaty, M., Ibdah, M., Fischer, R., Chaimovitsh, D., Saranga, Y. and Dudai, N. 2013. Dynamics of yield components and stevioside production in *Stevia rebaudiana* grown under different planting times, plant stands and harvest regime. *Industrial Crops and Products* 50: 731– 736.
- Sh, K., Singh, V. and Megeji, N.W. 2006. Cultivation of stevia [*Stevia rebaudiana* (Bert.) Bertoni]: A comprehensive review. *Advances in Agronomy* 89: 137-177.
- Shivashankara, K.S. and Mathai, C.K. 2000. Inhibition of photosynthesis by flowering in mango (*Mangifera indica* L.). A study by gas exchange methods. *Scientia Horticulturae* 83: 205-212.
- Shock, C.C. 1982. Rebaudi's stevia: natural non-caloric sweeteners. *California Agriculture* 36: 4-5.
- Singh, S.D. and Rao, G.P. 2005. Stevia: The herbal sugar of the 21st century. *Sugar Technology* 7: 17-24.
- Soejarto, D.D., Compadre, C.M., Medon, P.J., Kamath, S.K. and Kinghorn, A.D. 1983. Potential sweetening agents of plant origin: Field search for sweet-tasting stevia species. *Economic Botany* 37: 71-79.
- Soejarto, D.D. 2002. Botany of stevia and *Stevia rebaudiana*. In *Stevia: The Genus Stevia*, ed. A.D. Kinghorn, pp. 18-39. London and New York: Taylor and Francis.

- Starrat, A.N., Kirby, C.W., Pocs, R. and Brandle, J.E. 2002. Rebaudioside F a diterpene glycoside from *Stevia rebaudiana*. *Phytochemistry* 59: 367-370.
- Strauss, S. 1995. The perfect sweetener? *Technology Review* 98: 18-20.
- Sumida, T. 1980. Studies on *Stevia rebaudiana* Bertoni M, introduced from Brazil as a new sweetness resources in Japan. *Japan Centre Agricultural Experiment Station* 31: 1-71.
- Suvarnakuta, P., Chaweerungrat, C. and Devahastin, S. 2011. Effects of drying methods on assay and antioxidant activity of xanthonenes in mangosteen rind. *Food Chemistry* 125: 240-247.
- Taiz, L. and Zeiger, E. 2002. The control of flowering. In *Plant Physiology*, pp. 559-590. Massachusetts: Sinauer Associates, Inc.
- Takahashi, K., Matsuda, M., Ohashi, K., Tanaguchi, K., Nakagomi, O., Abe, Y., Mori, S., Okutani, N.K. and Shigeta, S. 2001. Analysis of anti-rotavirus activity of extract from *Stevia rebaudiana*. *Antiviral Research* 49: 15-24.
- Tamura, Y., Nakamura, S., Fukui, H. and Tabata, M. 1984. Comparison of stevia plants grown from seeds, cuttings and stem tip cultures for growth and sweet diterpene glycosides. *Plant Cell Reports* 3: 180-182.
- Tan, S.L., Ghawas, M.M., Najib, M.Y. and Zawawi, M. 2008. Preliminary evaluation and selection of stevia under Malaysian conditions. *Journal of Tropical Agriculture and Food Science* 36(2): 171-177.
- Tan, S.L., Wan Zaki, W.M., Ghawas, M.M., Mansor, M.P. and Zawawi, M. 2010. *Teknologi Penanaman dan Pemprosesan Primer Tumbuhan Ubatan*. Kuala Lumpur: Penerbit Institut Penyelidikan dan Kemajuan Pertanian Malaysia (MARDI). (In Malay language)
- Thakur, D.S. and Patel, S.R. 1998. Growth and sink potential of rice as influenced by the split application of potassium with FYM in inceptisols of eastern central India. *Journal of Potassium Research* 14: 73-77.
- Thomas, B. and Vince-Prue, D. 1997. Daylength perception in short-day plants. In *Photoperiodism in Plants*, ed. B. Thomas and D. Vince-Prue, pp. 85-117. London: Academic Press.
- Tirtoboma. 1988. The effect of cutting material and internode number on the growth and yield of *Stevia rebaudiana*. *Menara Perkebunan* 56: 96-101.
- Torres, A.P. and Lopez, R.G. 2011. Photoperiod and temperature influence flowering responses and morphology of *Tecoma stans*. *HortScience* 46: 416-419.
- Toyomasu, T., Kawaide, H., Mitsuhashi, W., Inoue, Y. and Kamiya, Y. 1998. Phytochrome regulates gibberellin biosynthesis during germination of photoblastic lettuce seeds. *Plant Physiology* 118: 1517-1523.

- Urban, L., Lu, P. and Thibaud, R. 2004. Inhibitory effect of flowering and early fruit growth on leaf photosynthesis in mango. *Tree Physiology* 24: 387-399.
- Valio, I.F.M. and Rocha, R.F. 1977. Effect of photoperiod and growth regulator on growth and flowering of *Stevia rebaudiana* Bertoni. *Japan Journal of Crop Science* 46(2): 243-248.
- Vince-Prue, D. 1975. *Photoperiodism in Plants*, pp. 1-414. London: McGraw-Hill.
- Vince-Prue, D. and Canham, A.E. 1983. Horticultural significance of photomorphogenesis. In *Shropshire, Encyclopedia of Plant Physiology*, ed. W.H. Mohr, pp. 518-544. Berlin: Springer-Verlag.
- Woelwer-Rieck, U., Lankes, C., Wawrzun, A. and Wu, M. 2010. Improved HPLC method for the evaluation of the major steviol glycosides in leaves of *Stevia rebaudiana*. *European Food Research and Technology* 231: 581-588.
- Yadav, A.K., Singh, S., Dhyani, D. and Ahuja, P.S. 2011. A review on the improvement of stevia [*Stevia rebaudiana* (Bertoni)]. *Canadian Journal of Plant Science* 91: 1-27.
- Yamada, A., Tanigawa, T., Suyama, T., Matsuno, T. and Kunitake, T. 2009. Red:far-red light ratio and far-red light integral promote or retard growth and flowering in *Eustoma grandiflorum* (Raf.) Shinn. *Scientia Horticulturae* 120: 101-106.
- Yamaguchi, S., Smith, M.W., Brown, R.G., Kamiya, Y. and Sun, T. 1998. Phytochrome regulation and differential expression of gibberellin 3 β -hydroxylase genes in germinating Arabidopsis seeds. *Plant Cell* 10: 2115-2126.
- Zaidan, L.B.P., Dietrich, S.M.C. and Felipe, G.M. 1980. Effect of photoperiod on flowering and stevioside content in plant of *Stevia rebaudiana* Bertoni. *Japan Journal Crop Science* 49: 569-574.
- Zeevaart, J.A.D. and Gage, D.A. 1993. *ent*-Kaurene biosynthesis is enhanced by long photoperiods in the long-day plants *Spinacia oleracea* L. and *Agrostemma githago* L. *Plant Physiology* 101: 25-29.