



UNIVERSITI PUTRA MALAYSIA

***ANTAGONIST YEAST FOR BIOCONTROL OF POSTHARVEST
ANTHRACNOSE DISEASE ON PAPAYA (*Carica papaya* L.)***

HAMIZAH BINTI HASSAN

FP 2017 34



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By

HAMIZAH BINTI HASSAN

**Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

July 2017

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DEDICATION

To my parents, siblings, supervisors, friends and husband.
Thank you.



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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment
of the requirement for the degree of Doctor of Philosophy

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July 2017

Chairman : Mahmud Tengku Muda Mohamed, PhD
Faculty : Agriculture

Papaya (*Carica papaya* L.) contain high nutritive value and very susceptible to postharvest decay. Major postharvest disease related to papaya fruit is anthracnose, caused by *Colletotrichum gloeosporioides*. Postharvest fungicide treatments are very popular for the disease control due to its fast and broad spectrum effects. However, withdrawal of a few commercial fungicides due to its negative effects on human, environment and produce increased public concerns and demands to other safer alternatives of control measures. Among all, biological control using microorganism to fight against host pathogens provides a positive results. However, limited studies done in Malaysia regarding the use of yeast as biocontrol agent for postharvest disease. Thus, this study was carried out with aim to develop a locally isolated, yeast antagonist agent for controlling anthracnose disease on papaya. In the first experiment, isolation of natural yeast on papaya plant was carried out. A total of 110 yeasts were isolated and tested for their antagonist activity against *C. gloeosporioides*. After a series of initial screening tests for antagonism, two yeasts, namely F001 and FL013 showed higher antagonistic activity against the pathogen were selected. Both F001 and FL013 showed better antagonist activity when concentration of yeast cells were increased from 5×10^4 to 5×10^8 cells mL⁻¹ in fungal spore germination test. Both yeasts also showed their ability to work well in cold storage condition and significantly inhibited the mycelial growth up to 70% using FL013 dipping treatment. However, only F001 has the ability to produce diffusible antifungal compounds which caused total suppression (100%) on the fungal growth. Thus, by referring to previous results, only F001 was identified and used in next experiments. Yeast F001, identified as *Trichosporon asahii*, also exhibited direct attachment on pathogen to control fungal hyphae growth and able to colonize well on papaya peel. In the next experiment, suitable additives for combining with *T. asahii* to improve antagonist activity of the yeast was investigated. Results showed that combination of *T. asahii* with calcium chloride able to totally suppress the pathogen growth in *in vitro* test. For *in vivo* test, combination of *T. asahii* with calcium chloride at 2.5% (w/v) caused reduction in

disease severity until 83% as compare to control fruit. This treatment combination was chosen and used in the subsequent studies. Next, effect of utilizing this treatment combination on disease incidence, disease severity, total phenolic, total flavonoid and antioxidant activity were evaluated. Results showed that *T. asahii* with 2.5% calcium chloride coating on ‘Solo’ papaya was capable to reduce disease severity but not able to significantly increase the activity of those phytochemicals. The treatment also showed that no interaction effect with storage day on defense-related enzyme, including polyphenol oxidase, phenylalanine ammonia lyase and catalase activity. However, treatment application on the fruit did not influence the quality and storability of papaya after stored at 12 °C for 28 days.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan Ijazah Doktor Falsafah

YIS ANTAGONIS SEBAGAI KAWALAN BIOLOGI BAGI PENYAKIT ANTRAKNOS LEPAS TUAI TERHADAP BETIK (*Carica papaya* L.)

Oleh

HAMIZAH BINTI HASSAN

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Betik (*Carica papaya* L.) mengandungi nilai pemakanan yang tinggi dan sangat mudah terdedah kepada kerosakan lepas tuai. Penyakit lepas tuai utama yang berkaitan dengan buah betik adalah antraknos yang disebabkan oleh *Colletotrichum gloeosporioides*. Rawatan racun kulat lepas tuai adalah sangat popular untuk kawalan penyakit ini kerana ia memberi kesan yang lebih cepat dan meluas. Walau bagaimanapun, pengharaman beberapa racun kulat komersial disebabkan oleh kesan negatif kepada manusia, alam sekitar dan hasil tanaman telah meningkatkan kebimbangan orang ramai dan permintaan kepada alternatif lain yang lebih selamat sebagai langkah-langkah kawalan. Antara alternatifnya adalah, kawalan biologi menggunakan mikroorganisma lain untuk melawan patogen hasil tanaman turut memberikan hasil yang positif. Walau bagaimanapun, kajian yang dilakukan di Malaysia mengenai penggunaan yis sebagai agen kawalan biologi bagi penyakit lepas tuai masih terhad. Oleh itu, kajian ini dijalankan dengan tujuan untuk mengenalpasti yis yang diperolehi secara lokal, sebagai agen antagonis untuk mengawal penyakit antraknos pada buah betik. Dalam kajian pertama, pemencilan yis yang terdapat secara semula jadi pada tumbuhan betik telah dijalankan. Sebanyak 110 yis telah diperolehi dan diuji untuk aktiviti antagonis terhadap *C. gloeosporioides*. Selepas beberapa siri ujian saringan awal, dua yis, iaitu F001 dan FL013 yang menunjukkan aktiviti antagonis yang lebih berkesan ke atas patogen telah dipilih. Kedua-dua yis, F001 dan FL013 menunjukkan aktiviti antagonis yang lebih baik apabila kepekatan sel yis ditingkatkan dari 5×10^4 kepada 5×10^8 sel mL⁻¹ dalam ujian percambahan spora fungi. Kedua-dua yis turut mempamerkan keupayaan untuk bertindak dengan lebih baik dalam suhu dingin dan secara signifikannya menghalang diameter miselium sehingga 70% menggunakan rawatan celupan FL013. Namun, hanya F001 yang berupaya menghasilkan komponen antikulat yang boleh meresap seterusnya menyebabkan penindasan secara total (100%) ke atas pertumbuhan patogen. Oleh itu, merujuk keputusan sebelumnya, hanya F001 dikenal pasti dan digunakan dalam eksperimen yang seterusnya. Yis F001, dikenal pasti sebagai *Trichosporon asahii*, juga menunjukkan pelekatan secara

langsung kepada patogen untuk mengawal pertumbuhan kulat hifa dan berupaya menghuni kulit buah betik dengan baik. Dalam eksperimen seterusnya, kajian telah dijalankan bagi mencari bahan tambahan yang sesuai untuk digabung bersama *T. asahii* bagi meningkatkan aktiviti antagonis yis. Hasil kajian menunjukkan bahawa kombinasi *T. asahii* dengan kalsium klorida dapat menghalang pertumbuhan patogen sepenuhnya (100%) di dalam ujian *in vitro*. Bagi ujian *in vivo*, gabungan *T. asahii* dengan 2.5% kalsium klorida (w/v) telah menyebabkan pengurangan tahap jangkitan sehingga 83% berbanding buah yang tidak dirawat. Kombinasi rawatan ini telah dipilih dan digunakan dalam kajian seterusnya. Seterusnya, kesan menggunakan gabungan rawatan terhadap jangkitan penyakit, tahap jangkitan, jumlah kandungan fenolik dan flavonoid serta aktiviti antioksidan telah dinilai. Hasil kajian menunjukkan bahawa salutan *T. asahii* dengan 2.5% kalsium klorida pada betik 'Solo' mampu untuk mengurangkan tahap jangkitan tetapi tidak mampu untuk menyebabkan peningkatan aktiviti fitokimia secara ketara. Rawatan ini juga menunjukkan tiada kesan interaksi dengan tempoh penyimpanan ke atas enzim berkaitan pertahanan, termasuk aktiviti polifenol oksidase, fenilalanina amonia liase dan katalase. Walau bagaimanapun, aplikasi rawatan tersebut pada buah tidak menjejaskan kualiti dan tempoh simpanan betik selepas disimpan pada 12 °C selama 28 hari.

ACKNOWLEDGEMENTS

I would like to express my sincere and deepest appreciation to all my kind supervisors; Prof. Dr. Mahmud Tengku Muda Mohamed, Assoc. Prof. Dr. Siti Hajar Ahmad and Assoc. Prof. Dr. Kamaruzaman Sijam for their continuous support, enlightening and patient guidance during my study, throughout the research and preparation of this thesis. I am also grateful to my friends and laboratory staffs for their kind support, suggestion and help during my study.

I am greatly indebted to my family especially my parents and siblings for their sacrifices, unconditional support and valuable advices until the completion of this study.

My gratitude extended to my colleagues and lovely friends in Postharvest Laboratory, Department of Crop Science especially Nor Elliza Tajidin, Farah Farhanah Haron, Munirah Mohamad, Bunga Raya Ketaren, Azimah Hamidon, Indah Shukor, Aizul Azfar, Fatimahtul Hazwani Hasan, Siti Aishah Aziz, Siti Fairuz Yusof and Nor Shariah Salleh for their kind help, invaluable guidance, supports, advices, comments and suggestions.

Also, special thanks to my dearest husband, Mohd Ridzuan Mohd Nor for his continuous encouragement, trust and motivation.

I certify that a Thesis Examination Committee has met on 4 July 2017 to conduct the final examination of Hamizah binti Hassan on her thesis entitled "Antagonist Yeast for Biocontrol of Postharvest Anthracnose Disease on Papaya (*Carica papaya* L.)" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

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LIST OF ABBREVIATIONS

μL	Microliter
BEN	Benomyl 50% WP
BLAST	Basic local alignment search tool
C*	Chroma
CAT	Catalase
cells mL ⁻¹	Cells/mililiter
CFU	Colony forming units
CM	Coating material
cm day ⁻¹	Centimeter/ day
CRD	Completely randomized design
DMRT	Duncan's multiple range test
DNA	Deoxyribonucleic acid
DOA	Department of Agriculture
DPPH	2,2-diphenyl-1-picrylhydrazyl radical scavenging
DS	Disease severity
FAMA	Federal Agricultural Marketing Authority
FAOSTAT	Food and Agriculture Organization Corporate Statistical Database
FRAP	Ferric reducing antioxidant power
g L ⁻¹	Gram/liter
h°	Hue
ITS	Internal transcribed spacer
L*	Lightness
LSD	Least significant difference
M	Moles per liter
MAFC	Malaysian AgriFood Corporation
MF	<i>Metschnikowia fructicola</i>
mL	Milliliter
mm	Milimeter
mM	Milimoles per liter
MOA	Ministry of Agriculture
NCBI	National Center for Biotechnology Information
NYDA	Nutrient yeast dextrose agar
NYDB	Nutrient yeast dextrose broth
PAL	Phenylalanine ammonia-lyase
PCR	Polymerase chain reaction
pH	Potential of hydrogen
PIRG	Percent inhibition of radial growth
PPO	Polyphenol oxidase
R ²	Coefficient determination
rDNA	Ribosomal deoxyribonucleic acid
rpm	Revolutions per minute
SBC	Sodium bicarbonate
SD	Storage duration
SDW	Sterile distilled water
SEM	Scanning electron microscope
SE	Standard error

spores mL ⁻¹	Spores/mililiter
SSC	Soluble solids concentration
TA	<i>Trichosporon asahii</i> combined with 2.5% calcium chloride
TFC	Total flavonoid content
TPC	Total phenolic content
w/v	Weight/volume
WP	Wettable powders



CHAPTER 1

INTRODUCTION

Papaya (*Carica papaya* L.) is an important income generator fruit in Malaysia. According to the Ministry of Agriculture (MOA), in 2001, papaya recorded the highest total export value for fruits with more than RM 90 million followed by watermelon which was RM 60 million (MOA, 2002). However, the export value exhibited a gradual decrease since papaya production decreased from 65,000 tonnes in 2001 to 32,000 tonnes in 2013 (FAOSTAT, 2015; MOA, 2015).

Anthrachnose, caused by *Colletotrichum gloeosporioides*, is a common postharvest disease of papaya fruit observed in the market chain which limits exportation (Paull et al., 1997; Ong and Ali, 2015). The control of anthracnose has been accomplished almost exclusively by fungicide application. However, the withdrawal of a number of fungicides that previously used to manage postharvest diseases of fruit had increased public concerns on the potential harmful consequences of fungicide toxicity to the human health and environment. The shifting of global trend towards lessen use of fungicides on fresh produce, safer and eco-friendly alternatives for reducing the decay loss in the harvested commodities, such as using antagonistic microorganisms like yeasts, fungi and bacteria, seem more promising and gaining popularity (Droby, 2006; Korsten, 2006).

Considerable success has been achieved when utilizing antagonistic microorganisms to control postharvest diseases. According to Irtwange (2006) and Qing and Shiping (2000), among all antagonistic organisms, yeasts are efficient biological control agents. Yeasts own many positive natural properties that make them useful for control purposes. In addition, yeasts generally do not produce allergenic spores or mycotoxins as many mycelial fungi do or antibiotic metabolites likely to be produced by bacterial antagonists. Other than requiring only simple nutritional condition, yeasts are also able to colonize on dry surfaces for long periods of time and resistant to many pesticides used in the postharvest environment (El-Tarabily and Sivasithamparam, 2006). Furthermore, past studies suggested that modes of action of biocontrol yeasts indicated low possibility of any hazard to consumers (Chanchaichaovivat et al., 2007).

Generally, antagonistic yeasts alone are often less effective than many of the commercial fungicides currently in use (Janisiewicz and Korsten, 2002). Some substances or methods appear to be useful when in combination with biocontrol yeasts. These include chitosan (Yu et al., 2007), calcium (Janisiewicz and Korsten, 2002; Tian et al., 2002a; Cao et al., 2008), sodium bicarbonate (Droby et al., 2003b; Yao et al., 2004), commercial fruit waxes (Lahlali et al., 2014) and microwave (Zhang et al., 2006). The studies showed that an integrated disease control strategy provides more effective and stable alternatives to synthetic fungicides.

A considerable number of antagonistic microorganisms have been tested in several countries for controlling different pathogens in different fruit-pathogen interaction systems especially on temperate fruits. However, the introduction of new microorganisms from foreign sources or regions to the local host may contribute risk to the produce and ecosystem. Since very few studies have been published from Malaysia on the control of anthracnose diseases on papaya using antagonistic yeast, this study was done with the following objectives:

1. To isolate, screen and identify yeast from papaya plant for their ability to control *C. gloeosporioides*.
2. To investigate the effect of additives (sodium bicarbonate or calcium chloride) in improving the biocontrol efficacy of *T. asahii*.
3. To examine the outcome of *T. asahii* combined with calcium chloride on host defence mechanism of 'Solo' papaya.
4. To evaluate the effect of antagonistic yeast supplemented with calcium chloride on the storability and quality of postharvest papaya.

BIBLIOGRAPHY

- Abakerli, R. B., Sparrapan, R., Sawaya, A. C. H. F., Eberlin, M. N., Jara, J. L. P., Rodrigues, N. R., Fay, E. F., Luiz, A. J. B., Galvao, T. D. L., Martins, D. d. S., Yamanishi, O. K. and Toledo, H. H. B. (2015). Carbon disulfide formation in papaya under conditions of dithiocarbamate residue analysis. *Food Chemistry*, 188, 71-76.
- Abliz, P., Fukushima, K., Takizawa, K., Yang, R., Li, R. and Nishimura, K. (2002). Identification of the first isolates of *Trichosporon asahii* var. *asahii* from disseminated trichosporonosis in China. *Diagnostic Microbiology and Infectious Disease*, 44(1), 17-22.
- Abouraïcha, E., El Alaoui-Talibi, Z., El Boutachfaiti, R., Petit, E., Courtois, B., Courtois, J. and El Modafar, C. (2015). Induction of natural defense and protection against *Penicillium expansum* and *Botrytis cinerea* in apple fruit in response to bioelicitors isolated from green algae. *Scientia Horticulturae*, 181, 121-128.
- Abu Bakar, M. F., Mohamed, M., Rahmat, A. and Fry, J. (2009). Phytochemicals and antioxidant activity of different parts of bambangan (*Mangifera pajang*) and tarap (*Artocarpus odoratissimus*). *Food Chemistry*, 113(2), 479-483.
- Adikaram, N. K. B., Joyce, D. C. and Terry, L. A. (2002). Biocontrol activity and induce resistance as a possible mode of action for *Aureobasidium pullulans* against grey mould of strawberry fruit. *Australian Plant Pathology*, 31, 223-229.
- Aghdam, M. S., Dokhanieh, A. Y., Hassanpour, H. and Fard, J. R. (2013). Enhancement of antioxidant capacity of cornelian cherry (*Cornus mas*) fruit by postharvest calcium treatment. *Scientia Horticulturae*, 161, 160-164.
- Aghdam, M. S., Hassanpouraghdam, M. B., Paliyath, G. and Farmani, B. (2012). The language of calcium in postharvest life of fruits, vegetables and flowers. *Scientia Horticulturae*, 144, 102-115.
- Al Zaemey, A. B., Magan, N. and Thompson, A. K. (1993). Studies on the effect of fruit-coating polymers and organic acids on growth of *Colletotrichum musae* in vitro and on postharvest control of anthracnose of bananas. *Mycological Research*, 97(12), 1463-1468.
- Ali, A. (2006). Anthracnose incidence, biochemical changes, postharvest quality and gas exchange of chitosan-coated papaya. (Doctor of Philosophy), Universiti Putra Malaysia, Malaysia.
- Ali, A., Ong, M. K. and Forney, C. F. (2014). Effect of ozone pre-conditioning on quality and antioxidant capacity of papaya fruit during ambient storage. *Food Chemistry*, 142, 19-26.

- Ali, H. and Nadarajah, K. (2014). Evaluating the efficacy of *Trichoderma* spp and *Bacillus substilis* as biocontrol agents against *Magnaporthe grisea* in rice. *Australian Journal of Crop Science*, 8(9), 1324-1335.
- Alvindia, D. G. and Natsuaki, K. T. (2008). Evaluation of fungal epiphytes isolated from banana fruit surfaces for biocontrol of banana crown rot disease. *Crop Protection*, 27, 1200-1207.
- Antunes, A. G. V., Pasqualotto, A. C., Diaz, M. C., d'Azevedo, P. A. and Severo, L. C. (2004). Candidemia in a brazilian tertiary care hospital: Species distribution and antifungal susceptibility patterns. *Journal of the Sao Paulo Institute of Tropical Medicine*, 46, 239-241.
- Arnaldo, L. C., Padovan, A. C. B. and Chaves, G. M. (2011). Current knowledge of *Trichosporon* spp. and Trichosporonosis. *Clinical Microbiology Reviews*, 24, 682-700.
- Arras, G. and Arru, S. (1997). Mechanisms of action of some microbial antagonists against fungal pathogens. *Annal Microbiology and Enzymology*, 47, 97-120.
- Arras, G., de-Cicco, V., Arru, S. and Lima, G. (1998). Biocontrol by yeasts of blue mold of citrus fruits and the mode of action of an isolate of *Pichia guilliermondii*. *Journal of Horticultural Science and Biotechnology*, 73, 413-418.
- Arras, G., Sanna, P., Astone, V. and Arru, S. (1998). Effect of CaCl₂ on the inhibitory activity of *Rhodotorula glutinis* against *Penicillium italicum* on orange fruits. *Italus Hortus*, 5, 67-70.
- Balibrea, M. E., Martinez-Andujar, C., Cuartero, J., Bolarin, M. C. and Perez-Alfocea, F. (2006). The high fruit soluble sugar content in wild *Lycopersicon* species and their hybrids with cultivars depends on sucrose import during ripening rather than on sucrose metabolism. *Functional Plant Biology*, 33, 279-288.
- Banani, H., Spadaro, D., Zhang, D., Matic, S., Garibaldi, A. and Gullino, M. L. (2015). Postharvest application of a novel chitinase cloned from *Metschnikowia fructicola* and overexpressed in *Pichia pastoris* to control brown rot of peaches. *International Journal of Food Microbiology*, 199, 54-61.
- Bar-Shimon, M., Yehuda, H., Cohen, L., Weiss, B., Kobeshnikov, A., Daus, A., Goldway, M., Wisniewski, M. and Droby, S. (2004). Characterization of extracellular lytic enzymes produced by the yeast biocontrol agent *Candida oleophila*. *Current Genetics*, 45, 140-148.
- Barkai-Golan, R. (2001). *Postharvest Diseases of Fruit and Vegetables: Development and Control*. Elsevier Sciences, Amsterdam, Netherlands.

- Bartz, J. A. and Brecht, J. K. (2002). *Postharvest Physiology and Pathology of Vegetables*. Marcel Dekker, United States of America.
- Bastiaanse, H., de Bellaire, L., Lassois, L., Misson, C. and Jijakli, M. H. (2010). Integrated control of crown rot of banana with *Candida oleophila* strain O, calcium chloride and modified atmosphere packaging. *Biological Control*, 3, 100-107.
- Bautista-Banos, S., Hernández-Lauzardo, A. N., Velázquez-del Valle, M. G., Hernández-López, M., Ait Barka, E., Bosquez-Molina, E. and Wilson, C. L. (2006). Chitosan as a potential natural compound to control pre and postharvest diseases of horticultural commodities. *Crop Protection*, 25, 108-118.
- Bautista-Banos, S., Sivakumar, D., Bello-Pérez, A., Arce, R. V. and Hernández-López, M. (2013). A review of the management alternatives for controlling fungi on papaya fruit during the postharvest supply chain. *Crop Protection*, 49, 8-20.
- Beers, J. R. F. and Sizer, I. W. (1952). A spectrophotometric method for measuring the breakdown of hydrogen peroxide by catalase. *Journal of Biological Chemistry*, 95, 133-140.
- Benbow, J. M. and Sugar, D. (1999). Fruit surface colonization and biological control of postharvest diseases of pear by preharvest yeast applications. *Plant Disease*, 83, 839-844.
- Bourne, M. C. (1980). Texture evaluation of horticultural crops. *HortScience*, 15, 51-57.
- Breinig, F., Tipper, D. J. and Schmitt, M. J. (2002). Kre1p, the plasma membrane receptor for the yeast K1 viral toxin. *Cell*, 108, 395-405.
- Bron, I. U. and Jacomino, A. P. (2006). Ripening and quality of 'Golden' papaya fruit harvested at different maturity stages. *Brazilian Journal of Plant Physiology*, 18(3), 389-396.
- Calderone, R. A. and Clancy, C. J. (2011). *Candida and Candidiasis*. American Society for Microbiology Press, United States of America.
- Canamas, T. P., Vinas, I., Usall, J., Torres, R., Anguera, M. and Teixido, N. (2008). Control of postharvest diseases on citrus fruit by preharvest applications of biocontrol agent *Pantoea agglomerans* CPA-2: Part II. Effectiveness of different cell formulations. *Postharvest Biology and Technology*, 49(1), 96-106.
- Cao, B., Li, H., Tian, S. and Qin, G. (2012). Boron improves the biocontrol activity of *Cryptococcus laurentii* against *Penicillium expansum* in jujube fruit. *Postharvest Biology and Technology*, 68, 16-21.

- Cao, F., Guan, C., Dai, H., Li, X. and Zhang, Z. (2015). Soluble solids content is positively correlated with phosphorus content in ripening strawberry fruits. *Scientia Horticulturae*, 195, 183-187.
- Cao, S. and Zheng, Y. (2010). Effect of 1-methylcyclopropene on anthracnose rot caused by *Colletotrichum acutatum* and disease resistance in loquat fruit. *Journal of The Science of Food and Agriculture*, 90, 2289-2294.
- Cao, S., Zheng, Y., Wang, K., Tang, S. and Rui, H. (2009). Effect of yeast antagonist in combination with methyl jasmonate treatment on postharvest anthracnose rot of loquat fruit. *Biological Control*, 50, 73-77.
- Cao, S. F., Zheng, Y. H., Tang, S. S. and Wang, K. T. (2008). Improved control of postharvest anthracnose rot in loquat fruit by a combination treatment of *Pichia membranefaciens* with CaCl₂. *International Journal of Food Microbiology*, 126, 216-220.
- Capdeville, G. d., Jr., M. T. S., Santos, J. R. P., Miranda, S. d. P., Caetano, A. R. and Torres, F. A. G. (2007). Selection and testing of epiphytic yeasts to control anthracnose in post-harvest of papaya fruit. *Scientia Horticulturae*, 111, 179-185.
- Capdeville, G. d., Wilson, C. L., Beer, S. V. and Aist, J. R. (2002). Alternative disease control agents induce resistance to blue mold in harvested 'Red Delicious' apple fruit. *Phytopathology*, 92, 900-908.
- Casals, C., Elmer, P. A. G., Vinas, I., Teixido, N., Sisquella, M. and Usall, J. (2012). The combination of curing with either chitosan or *Bacillus subtilis* CPA-8 to control brown rot infections caused by *Monilinia fructicola*. *Postharvest Biology and Technology*, 64, 126-132.
- Castoria, R., Curtis, F. d., Lima, G. and Cicco, V. d. (1997). β -1,3-glucanase activity of two saprophytic yeasts and possible mode of action as biocontrol agents against postharvest diseases. *Postharvest Biology and Technology*, 12, 293-300.
- Chalutz, E., Ben-Arie, R., Droby, S., Cohen, L., Weiss, B. and Wilson, C. L. (1988). Yeasts as biocontrol agents of postharvest diseases of fruit. *Phytoparasitica*, 16, 69-75.
- Chalutz, E. and Wilson, C. L. (1990). Postharvest biocontrol of green and blue mold and sour rot of citrus by *Debaryomyces hansenii*. *Plant Disease*, 74, 134-137.
- Chan, Y. K. (2014a). Breeding the papaya-*Carica papaya*. In F. Ng (Ed.), (pp. 32-42): UTAR Agriculture Science Journal.
- Chan, Y. K. (2014b). A Practical Compendium on Advances in Plant Breeding. Genetics Society of Malaysia, Malaysia.

- Chan, Y. K. and Paull, R. E. (2008). Encyclopedia of fruit and nuts. CAB International, Wallingford, United Kingdom.
- Chan, Z. and Tian, S. P. (2005). Interaction of antagonistic yeasts against postharvest pathogens of apple fruit and possible mode of action. *Postharvest Biology and Technology*, 36, 215-223.
- Chanchaichaovivat, A., Ruenwongsa, P. and Panijpan, B. (2007). Screening and identification of yeast strains from fruits and vegetables: Potential for biological control of postharvest chilli anthracnose (*Colletotrichum capsici*). *Biological Control*, 42, 326-335.
- Chardonnet, C. O., Sams, C. E., Trigiano, R. N. and Conway, W. S. (2000). Variability of three isolates of *Botrytis cinerea* affects the inhibitory effects of calcium on the fungus. *Phytopathology*, 90, 769-774.
- Chau, K. F. and Alvarez, A. M. (1983). A histological study of anthracnose on *Carica papaya*. *Phytopathology*, 73, 1113-1116.
- Chen, N. M. and Paull, R. E. (1986). Development and prevention of chilling injury in papaya fruit. *Journal of American Society of Horticultural Science*, 111, 639-643.
- Cheng, S. H., Sheen, J., Gerrish, C. and Bolwell, G. P. (2001). Molecular identification of phenylalanine ammonia-lyase as a substrate of a specific constitutively active Arabidopsis CDPK expressed in maize protoplasts. *FEBS Letters*, 503, 185-188.
- Coelho, A. R., Celli, M. G., Ono, E. Y. S. and Wosiacki, G. (2007). *Penicillium expansum* versus antagonist yeasts with perspectives of application in biocontrol and patulin degradation. *Brazilian Archives of Biology and Technology*, 50, 725-733.
- Conway, W. S. (1982). Effect of postharvest calcium treatment on decay of delicious apples. *Plant Disease*, 66, 402-403.
- Conway, W. S., Leverentz, B., Janisiewicz, W. J., Blodgett, A. B., Saftner, R. A. and Camp, M. J. (2004). Integrating heat treatment, biocontrol and sodium bicarbonate to reduce postharvest decay of apple caused by *Colletotrichum acutatum* and *Penicillium expansum*. *Postharvest Biology and Technology*, 34, 11-20.
- Conway, W. S. and Sams, C. E. (1983). Calcium infiltration of Golden Delicious apples and its effect on decay. *Phytopathology*, 73, 1068-1071.
- Conway, W. S., Sams, C. E. and Hickey, K. D. (2002). Pre- and postharvest calcium treatment of apple fruit and its effect on quality. *Acta Horticulturae*, 594, 413-419.

- De Koch, S. (1998). *Control of post-harvest decay of fruit by means of antagonistic microorganisms*. (Doctor of Philosophy), University of Stellenbosch, Stellenbosch.
- Denner, F. D. N., Kotzé, J. M. and Putterill, J. F. (1986). The effect of temperature on spore germination, growth and appressorium formation of *Colletotrichum gloeosporioides* and *Dothiorella aromatica*. *South African Avocado Growers' Association Yearbook*, 9, 19-22.
- Diby, P. and Sarma, Y. R. (2005). *Pseudomonas fluorescens* mediated systemic resistance in black pepper (*Piper nigrum* L.) is driven through an elevated synthesis of defense enzymes. *Archives of Phytopathology and Plant Protection Journal*, 38, 139-149.
- Dickman, M. B. (1994). *Papaya Anthracnose*. American Phytopathological Society, Minneapolis, Minnesota.
- DOA. (2015). Fruit Crop Statistics Malaysia 2104. Retrieved 21 June 2016, from http://www.doa.gov.my/c/document_library/get_file?uuid=24206d88-31d7-49b0-97fe-6929d5340c7f&groupId=338810
- Dotto, G. L., Vieira, M. L. G. and Pinto, L. A. A. (2015). Use of chitosan solutions for the microbiological shelf life extension of papaya fruits during storage at room temperature. *LWT - Food Science and Technology*, 64(1), 126-130.
- Droby, S. (2006). Biological control of postharvest diseases of fruit and vegetables: difficulties and challenges. *Phytopathology*, 39, 105-117.
- Droby, S. and Chalutz, E. (1994). Mode of action of biocontrol agents for postharvest diseases. In *Biological Control of Postharvest Diseases of Fruits and Vegetables-Theory and Practice* ed. Wilson C.L. and Wisniewski M.E. CRC Press, Boca Raton, Florida.
- Droby, S., Chalutz, E. and Wilson, C. L. (1991). Antagonistic microorganisms as biocontrol agents of postharvest diseases of fruit and vegetables *Postharvest News Information* 2, 169-173.
- Droby, S., Chalutz, E., Wilson, C. L. and Wisniewski, M. E. (1989). Characterization of the biocontrol activity of *Debaryomyces hansenii* in the control of *Penicillium digitatum* on grapefruit. *Canadian Journal of Microbiology*, 35, 794-800.
- Droby, S., Cohen, L., Daus, A., Weiss, B., Horev, B., Chalutz, E., Katz, H., Kere-Tzur, M. and Shachnai, A. (1998). Commercial testing of aspire: a yeast preparation for the biological control of postharvest decay of citrus. *Biological Control*, 12, 97-101.
- Droby, S., Vinokur, V., Weiss, B., Cohen, L., Daus, A., Goldschmidt, E. E. and Porat, R. (2002). Induction of resistance to *Penicillium digitatum* in

grapefruit by the yeast biocontrol agent *Candida oleophila*. *Phytopathology*, 92, 393-399.

- Droby, S., Wilson, C., Wisniewski, M. and El Ghaouth, A. (2000). Biologically based technology for the control of postharvest diseases of fruits and vegetables. In *Microbial Food Contamination*, ed. Wilson, C. and Droby, S. CRC Press, Boca Raton, United States of America.
- Droby, S., Wisniewski, M. and Benkeblia, N. (2011). *Postharvest Biology and Technology of Tropical and Subtropical Fruits: Fundamental Issues*. Woodhead Publishing Limited.
- Droby, S., Wisniewski, M., El-Ghaouth, A. and Wilson, C. (2003a). Biological control of postharvest diseases of fruits and vegetables: current achievements and future challenges. *Acta Horticulturae*, 628, 703-713.
- Droby, S., Wisniewski, M., El-Ghaouth, A. and Wilson, C. (2003b). Influence of food additives on the control of postharvest rots of apple and peach and efficacy of the yeast based biocontrol product Aspire. *Postharvest Biology and Technology*, 27, 127-135.
- Droby, S., Wisniewski, M., Macarasin, D. and Wilson, C. (2009). Twenty years of postharvest biocontrol research: is it time for a new paradigm? *Postharvest Biology and Technology*, 52, 137-145.
- Droby, S., Wisniewski, M. E., Cohen, L., Weiss, B., Touitou, D., Eilam, Y. and Chalutz, E. (1997). Influence of CaCl₂ on *Penicillium digitatum*, grapefruit peel tissue and biocontrol activity of *Pichia guilliermondii*. *Phytopathology*, 87, 310-315.
- Druvefors, U. (2004). *Yeast biocontrol of grain spoilage mold*. (Doctor of Philosophy), Swedish University of Agricultural Sciences, Sweden. Retrieved from <http://diss-epsilon.slu.se/archive/00000552>
- Ebrahimi, L., Etebarian, H. R., Aminian, H. and Sahebani, N. (2012). Enhancement of biocontrol activity of *Torulaspora delbrueckii* with methyl jasmonate against apple blue mould disease. *Archives of Phytopathology and Plant Protection Journal*, 45, 2355-2363.
- Eckert, J. W. (1990). Recent development in chemical control of postharvest diseases. *Acta Horticulturae*, 269, 477-494.
- Eckert, J. W. and Ogawa, I. M. (1985). The chemical control of postharvest diseases: Sub tropical and tropical fruits. *Annual Review of Phytopathology*, 23, 421-454.
- Edwards, K., Johnstone, C. and Thompson, C. (1991). A simple and rapid method for the preparation of genomic plant DNA for PCR analysis. *Nucleic Acids Research*, 19, 1349.

- El-Tarabily, K. A. (2004). Suppression of *Rhizoctonia solani* diseases of sugar beet by antagonistic and plant growth-promoting yeasts. *Journal of Applied Microbiology*, 96, 69-75.
- El-Tarabily, K. A. and Sivasithamparam, K. (2006). Potential of yeasts as biocontrol agents of soil-borne fungal plant pathogens and as plant growth promoters. *Mycoscience*, 47, 25-35.
- El Ghaouth, A. (1997). Biologically based alternatives to synthetic fungicides for the control of postharvest diseases. *Journal of Industrial Microbiology and Biotechnology*, 19, 160-162.
- El Ghaouth, A., Smilanick, J. L. and Wilson, C. L. (2000). Enhancement of the performance of *Candida saitoana* by the addition of glycochitosan for control of postharvest decay of apple and citrus fruit. *Postharvest Biology and Technology*, 19, 249-253.
- El Ghaouth, A. and Wilson, C. (1995). Biologically-based technologies for the control of postharvest diseases. *Postharvest News and Information*, 6, 5-11.
- El Ghaouth, A., Wilson, C. and Wisniewski, M. (1998). Ultrastructural and cytochemical aspect of the biocontrol activity of *Botrytis cinerea* by *Candida saitoana* in apple fruit. *Phytopathology*, 88, 282-291.
- El Ghaouth, A., Wilson, C., Wisniewski, M., Droby, S., Smilanick, J. L. and Korsten, L. (2002a). Biological control of postharvest diseases of fruits and vegetables. *Applied Mycology and Biotechnology*, 2, 219-238.
- El Ghaouth, A., Wilson, C. L. and Wisniewski, M. (2003). Control of postharvest decay of apple fruit with *Candida saitoana* and induction of defense responses. *Phytopathology*, 93, 344-348.
- El Ghaouth, A., Wilson, C. L., Wisniewski, M., Droby, S., Smilanick, J. L. and Korsten, L. (2002b). *Biological control of postharvest diseases of citrus fruits*. In Gnanamanickam S S, *Biological Control of Crop Diseases*. New York, Marcel Dekker Press.
- El Ghaouth, A., Wilson, C. L. and Wisniewski, M. E. (2004). Biologically based alternatives to synthetic fungicides for the postharvest diseases of fruit and vegetables. In: Naqvi S.A.M.H. (Ed.) *Diseases of Fruit and Vegetables* (Vol. 2, pp. 511-535). Kluwer Academic Publishers, The Netherlands.
- Elad, Y. and Freeman, S. (2002). Biological control of fungal plant pathogens. In F. Kempken (Ed.), *The Mycota, A Comprehensive Treatise on Fungi as Experimental Systems for Basic and Applied Research. XI. Agricultural Applications*. Springer, Heidelberg, Germany.
- Escribano-Bailon, M. T., Santos-Buelga, C. and Rivas-Gonzalo, J. (2004). Anthocyanins in cereals. *Journal of Chromatography*, 1054, 129-141.

- Fajardo, J. E., McCollum, T. G., McDonald, R. E. and Mayer, R. T. (1998). Differential induction of proteins in orange flavedo by biologically based elicitors and challenged by *Penicillium digitatum* Sacc. *Biological Control*, 13, 143-151.
- Falguera, V., Quintero, J. P., Jiménez, A., Muñoz, J. A. and Ibarz, A. (2011). Edible films and coatings structures: active functions and trends in their use. *Trends Food Science and Technology*, 22, 292-303.
- FAMA. (2015). Statistik Utama Pemasaran FAMA 2014. Retrieved 21 June 2016, from <http://www.fama.gov.my/documents/10157/f3d27dd0-f3a3-4d94-ac01-bf5dd08022ec>
- Fan, Q., Tian, S. P., Li, Y. X., Xu, Y. and Wang, Y. (2000). Biological control of postharvest brown rot in peach and nectarine fruit by *Bacillus subtilis* (B-912). *Acta Botanica Sinica*, 42, 1137-1143.
- Fan, Q., Tian, S. P., Liu, S. and Xu, Y. (2002). Production of β -1,3-glucanase and chitinase of two biocontrol agents and their possible modes of action. *Chinese Science Bulletin*, 47, 292-296.
- Fan, Y., Xu, Y., Wang, D., Zhang, L., Sun, J., Sun, L. and Zhang, B. (2009). Effect of alginate coating combined with yeast antagonist on strawberry (*Fragaria×ananassa*) preservation quality. *Postharvest Biology and Technology*, 53(1), 84-90.
- FAOSTAT. (2015). Crop production. Retrieved 21 June 2016, from <http://faostat3.fao.org/browse/Q/QC/E>
- Farahani, L., Etebarian, H. R., Sahebani, N. and Sahebani, H. (2012). Effect of two strains of antagonistic yeasts in combination with silicon against two isolates of *Penicillium expansum* on apple fruit. *International Research Journal of Applied and Basic Sciences*, 3, 18-23.
- Ferreira, I. C., Baptista, P., Vilas-Boas, M. and Barros, L. (2007). Free-radical scavenging capacity and reducing power of wild edible mushrooms from northeast Portugal: Individual cap and stipe activity. *Food Chemistry*, 100(4), 1511-1516.
- Fishman, M. L., Levaj, B., Gillespie, D. and Scorza, R. (1993). Changes in the physico-chemical properties of peach fruit pectin during on-tree ripening and storage. *Journal of the American Society for Horticultural Science*, 118, 343-349.
- Florkowski, W. J., Shewfelt, R. L., Brueckner, B. and Prussia, S. E. (2009). *Postharvest Handling: A Systems Approach*. Academic Press, United States of America.

- Gholamnejad, J. and Etebarian, H. R. (2009). Effect of calcium chloride on the biocontrol efficacy of two antagonistic yeasts against *Penicillium expansum* on apple fruit. *Phytoparasitica*, 37, 255-261.
- Gnanamanickam, S. S. (2002). *Biological Control of Crop Diseases*. Marcel Dekker, United States of America.
- Gueho, E., Improvisi, L., de Hoog, G. S. and Dupont, B. (1994). Trichosporon on humans: a practical account. *Mycoses*, 37(1-2), 3-10. doi: 10.1111/j.1439-0507.1994.tb00277.x
- Gullino, M. L., Benzi, D., Aloï, C., Testoni, A. and Garibaldi, A. (1992). Biological control of Botrytis rot of apple. In *Recent advances in Botrytis research*, ed. Williamson, B., Verhoeff, K. and Malatrakis, N. E. Paper presented at the Proceedings of the 10th International Botrytis Symposium, Heraklion, Crete.
- Haron, F. F. (2012). Antifungal Activity of *Allamanda* spp. Extracts and Their Microemulsion Formulations against Anthracnose (*Colletotrichum gloeosporioides*) Disease of Papaya. (Doctor of Philosophy), Universiti Putra Malaysia, Malaysia.
- Hasan, F. H. (2014). Ethylene, simulated transport duration and maturity stage affect postharvest quality of vapour-heat treated *Carica papaya*. (Master of Science), Universiti Putra Malaysia, Malaysia.
- Hasan, M. F. (2012). Controlling Anthracnose and Postharvest Quality of Papaya (*Carica papaya* cv. Frangi) using *Bacillus subtilis* strain B34. (Doctor of Philosophy), Universiti Putra Malaysia, Malaysia.
- Hashem, M. and Alamri, S. (2009). The biocontrol of postharvest disease (*Botryodiplodia theobromae*) of guava (*Psidium guajava* L.) by the application of yeast strains. *Postharvest Biology and Technology*, 53, 123-130.
- Hata, E. M., Sijam, K., Ahmad, Z. A. M., Yusof, M. T. and Azman, N. A. (2015). *In vitro* Antimicrobial Assay of Actinomycetes in Rice Against *Xanthomonas oryzae* pv. *oryzicola* and as Potential Plant Growth Promoter. *Brazilian Archives of Biology and Technology*, 58, 821-832.
- Hernandez-Munoz, P., Almenar, E., Ocio, M. J. and Gavara, R. (2006). Effect of calcium dips and chitosan coatings on postharvest life of strawberries (*Fragaria x ananassa*). *Postharvest Biology and Technology*, 39, 247-253.
- HersHKovitz, V., Ben-Dayan, C., Raphael, G., Pasmanik-Chor, M., Liu, J., Belausov, E., Aly, R., Wisniewski, M. and Droby, S. (2012). Global changes in gene expression of grapefruit peel tissue in response to the yeast biocontrol agent *Metschnikowia fructicola*. *Molecular in Plant Pathology*, 13, 338-349.

- Hornby, J. M., Jensen, E. C., Lisec, A. D., Tasto, J. J., Jahnke, B., Shoemaker, R., Dussault, P. and Nickerson, K. W. (2001). Quorum sensing in the dimorphic fungus *Candida albicans* is mediated by farnesol. *Applied and Environmental Microbiology*, 67, 2982-2992.
- Hoy, J., Hsu, K. C., Rolston, K., Hopfer, R. L., Luna, M. and Bodey, G. P. (1986). *Trichosporon beigeli* infection: A Review. *Review of Infectious Disease*, 8, 959-967.
- Ilhan, K., Arslan, U. and Karabulut, O. A. (2006). The effect of sodium bicarbonate alone or in combination with a reduced dose of tebuconazole on the control of apple scab. *Crop Protection*, 25, 963-967.
- Illeperuma, C. K. and Jayasuriya, P. (2002). Prolonged storage of 'Karuthacolomban' mango by modified atmosphere packaging at low temperature. *Journal of Horticultural Science and Biotechnology*, 77(2), 153-157.
- Ippolito, A., Ghaouth, E. I. A., Wilson, L. C. and Wisniewski, M. (2000). Control of postharvest decay of apple fruit by *Aureobasidium pullulans* and induction of defense responses. *Postharvest Biology and Technology*, 19, 265-272.
- Ippolito, A. and Nigro, F. (2000). Impact of preharvest application of biological control agents on postharvest diseases of fresh fruit and vegetables. *Crop Protection*, 19, 715-723.
- Ippolito, A., Nigro, F. and Schena, L. (2004). Control of postharvest diseases of fresh fruit and vegetables by preharvest application of antagonistic microorganisms. In *Crop management and postharvest handling of horticultural products Volume: IV: Diseases and disorders of fruit and vegetables*, ed. Niskanen, D. R. and Jain, R. SM Science Publishers, Inc. Enfield, United States of America.
- Ippolito, A., Schena, L., Pentimone, I. and Nigro, F. (2005). Control of postharvest rots of sweet cherries by pre- and postharvest applications of *Aureobasidium pullulans* in combination with calcium chloride or sodium bicarbonate. *Postharvest Biology and Technology*, 36, 245-252.
- Irtwange, S. V. (2006). Application of biological control agents in pre- and postharvest operations. *International Commission of Agricultural Engineering*, 8(3), 1-11.
- Janisiewicz, W. J. (1987). Postharvest biological control of blue mold on apple. *Phytopathology*, 77, 481-485.
- Janisiewicz, W. J. (1988). Biological control of postharvest diseases of apple with antagonists' mixtures. *Phytopathology*, 78, 194-198.

- Janisiewicz, W. J. (1998). Biocontrol of postharvest diseases of temperate fruits: Challenges and opportunities. In *Plant-microbe interactions and biological control*, ed. Boland, J. and Kuykendall, L.D. Marcel Dekker, New York.
- Janisiewicz, W. J. and Conway, W. S. (2010). Combining biological control with physical and chemical treatments to control fruit decay after harvest. *Stewart Postharvest Review*. doi: 10.2212/spr.2010.1.3
- Janisiewicz, W. J., Conway, W. S., Glenn, D. M. and Sams, C. E. (1998). Integrating biological control and calcium treatment for controlling postharvest decay of apples. *HortScience*, 33, 105-109.
- Janisiewicz, W. J. and Jeffers, S. N. (1997). Efficacy of commercial formulation of two biofungicides for control of blue-mold and gray mold of apples. *Crop Protection*, 7, 629-633.
- Janisiewicz, W. J. and Korsten, L. (2002). Biological control of postharvest diseases of fruits. *Annual Review of Phytopathology*, 40, 411-441.
- Janisiewicz, W. J., Peterson, D. L. and Bors, R. (1994). Control of storage rots of apple with *Sporobolomyces roseus*. *Plant Disease*, 78, 466-470.
- Janisiewicz, W. J., Saftner, R. A., Conway, W. S. and Yoder, K. S. (2008). Control of blue mold decay of apple during commercial controlled atmosphere storage with yeast antagonists and sodium bicarbonate. *Postharvest Biology and Technology*, 49, 374-378.
- Janisiewicz, W. J., Tworowski, T. J. and Kurtzman, C. P. (2001). Biocontrol potential of *Metchnikowia pulcherrima* strains against blue mold of apple. *Phytopathology*, 91, 1098-1108.
- Janisiewicz, W. J., Tworowski, T. J. and Sharer, C. (2000). Characterizing the mechanism of biological control of postharvest diseases on fruits with a simple method to study competition for nutrients *Phytopathology*, 90, 1196-1200.
- Jiang, F., Zheng, X. and Chen, J. (2009). Microarray analysis of gene expression profile induced by the biocontrol yeast *Cryptococcus laurentii* in cherry tomato fruit. *Gene*, 430, 12-16.
- Jiang, Y. M. and Joyce, D. C. (2003). ABA effects on ethylene production, PAL activity, anthocyanin and phenolic contents of strawberry fruit. *Plant Growth Regulation*, 39, 171-174.
- Júnior, V. L., Maffia, L. A., Romeiro, R. d. S. and Mizubuti, E. S. G. (2006). Biocontrol of tomato late blight with the combination of epiphytic antagonists and rhizobacteria. *Biological Control*, 38(3), 331-340.
- Kader, A. A. (2008). Flavor quality of fruits and vegetables. *Journal Science of Food and Agriculture*, 88, 1863-1868.

- Karabulut, O. A., Smilanick, J. L., Gabler, F. M., Mansour, M. and Droby, S. (2003). Near-harvest applications of *Metschnikowia fructicola*, ethanol and sodium bicarbonate to control postharvest diseases of grape in central California. *Plant Disease*, 87, 1384-1389.
- Karabulut, O. A., Tezcan, H., Daus, A., Cohen, L., Wiess, B. and Droby, S. (2004). Control of preharvest and postharvest fruit rot in strawberry by *Metschnikowia fructicola*. *Biocontrol Science and Technology*, 14, 513-521.
- Karacaa, H., Pérez-Gago, M. B., Taberner, V. and Palou, L. (2014). Evaluating food additives as antifungal agents against *Monilinia fructicola* in vitro and in hydroxypropyl methylcellulose–lipid composite edible coatings for plums. *International Journal of Food Microbiology*, 179, 72-79.
- Kohl, J., Postma, J., Nicot, P., Ruocco, M. and Blum, B. (2011). Stepwise screening of microorganisms for commercial use in biological control of plant-pathogenic fungi and bacteria. *Biological Control*, 57, 1-12.
- Kokkinakis, D. M. and Brooks, J. L. (1979). Tomato peroxidases: Purification, characterization and catalytic properties. *Plant Physiology*, 63, 93-99.
- Korsten, L. (2006). Advances in control of postharvest diseases in tropical fresh produce. *International Journal of Postharvest Technology and Innovation*, 1, 48-61.
- Kosman, E. and Cohen, Y. (1996). Procedures for calculating and differentiating synergism and antagonism in action of fungicide mixtures. *Phytopathology*, 86, 1263-1272.
- Kurabachew, H. and Wydra, K. (2013). Characterization of plant growth promoting rhizobacteria and their potential as bioprotectant against tomato bacterial wilt caused by *Ralstonia solanacearum*. *Biological Control*, 67(1), 75-83.
- Kurtzman, C., Fell, J. W. and Boekhout, T. (2011). The Yeasts, A Taxonomic Study. In *Agriculturally Important Yeasts: Biological Control of Field and Postharvest Diseases Using Yeast Antagonists, and Yeasts as Pathogens of Plants*, ed . Schisler, D. A., Janisiewicz, W. J., Boekhout, T. and Kurtzman, C. P. Elsevier, United States of America.
- Kurtzman, C. P. and Droby, S. (2001). *Metschnikowia fructicola*, a new ascosporic yeast with potential for biocontrol of postharvest fruit rots.
- Kurtzman, C. P. and Fell, J. W. (1998). *The Yeasts: Taxonomic Study*. Elsevier Science, Amsterdam.
- Lahlali, R., Hamadi, Y., Drider, R., Misson, C., El Guilli, M. and Jijakli, M. H. (2014). Control of citrus blue mold by the antagonist yeast *Pichia guilliermondii* Z1: Compatibility with commercial fruit waxes and putative mechanisms of action. *Food Control*, 45, 8-15.

- Lazan, H., Selamat, M. K. and Ali, Z. M. (1995). β -Galactosidase, polygalacturonase and pectinesterase in differential softening and cell wall modification during papaya fruit ripening. *Physiologia Plantarum*, 95, 106-112.
- Leibinger, W., Breuker, B., Hahn, M. and Mendgen, K. (1997). Control of postharvest pathogens and colonization of the apple surface by antagonistic microorganisms in the field. *Phytopathology*, 87, 1103-1110.
- Li, B. Q., Zhou, Z. W. and Tian, S. P. (2008). Combined effects of endo- and exogenous trehalose on stress tolerance and biocontrol efficacy of two antagonistic yeasts. *Biological Control*, 46, 187-193.
- Li, L., Ma, J., Li, Y., Wang, Z., Gao, T. and Wang, Q. (2012). Screening and partial characterization of *Bacillus* with potential applications in biocontrol of cucumber *Fusarium* wilt. *Crop Protection*, 35, 29-35.
- Lima, G., Ippolito, A., Nigro, F. and Salerno, M. (1997). Effectiveness of *Aureobasidium pullulans* and *Candida oleophila* against postharvest strawberry rots. *Postharvest Biology and Technology*, 10, 169-178.
- Lima, G., Spina, A. M., Castoria, R., Curtis, F. D. and Cicco, V. D. (2005). Integration of biocontrol agents and food-grade additives for enhancing protection of stored apples from *Penicillium expansum*. *Journal of Food Protection*, 68, 2100-2106.
- Lima, J. R., Gondim, D. M. F., Oliveira, J. T. A., Oliveira, F. S. A., Goncalves, L. R. B. and Viana, F. M. P. (2013). Use of killer yeast in the management of postharvest papaya anthracnose. *Postharvest Biology and Technology*, 83, 58-64.
- Liu, H., Jiang, W., Bi, Y. and Luo, Y. (2005). Postharvest BTH treatment induces resistance of peach (*Prunus persica* L. cv. Jiubao) fruit to infection by *Penicillium expansum* and enhances activity of fruit defense mechanisms. *Postharvest Biology and Technology*, 35(3), 263-269.
- Liu, J., Sui, Y., Wisniewski, M., Droby, S. and Liu, Y. (2013). Review: Utilization of antagonistic yeasts to manage postharvest fungal diseases of fruit. *International Journal of Food Microbiology*, 167, 153-160.
- Liu, J., Wisniewski, M., Droby, S., Norelli, J., Hershkovitz, V., Tian, S. and Farrell, R. (2012). Increase in antioxidant gene transcripts, stress tolerance and biocontrol efficacy of *Candida oleophila* following sublethal oxidative stress exposure. *FEMS Microbiology and Ecology*, 80, 578-590.
- Liu, J., Wisniewski, M., Droby, S., Tian, S., Hershkovitz, V. and Tworowski, T. (2011). Effect of heat shock treatment on stress tolerance and biocontrol efficacy of *Metschnikowia fructicola*. *FEMS Microbiology and Ecology*, 76, 145-155.

- Lowry, O. H., Rosebrough, N. J., Farr, A. L. and Randall, R. J. (1951). Protein measurement with the Folin phenol reagent. *Journal of Biological Chemistry*, 193, 263-275.
- Lu, L., Lu, H., Wu, C., Fang, W., Yu, C., Ye, C., Shi, Y., Yu, T. and Zheng, X. (2013). *Rhodosporidium paludigenum* induces resistance and defense-related responses against *Penicillium digitatum* in citrus fruit. *Postharvest Biology and Technology*, 85, 196-202.
- Madani, B., Mohamed, M. T. M., Biggs, A. R., Kadir, J., Awang, Y., Tayebimeigooni, A. and Shojaei, T. R. (2014). Effect of pre-harvest calcium chloride applications on fruit calcium level and post-harvest anthracnose disease of papaya. *Crop Protection*, 55, 55-60.
- Maeda, C. and Nelson, S. (2014). Anthracnose of Papaya in Hawai'i. *Plant Disease*, 103, College of Tropical Agriculture and Human Resources, University of Hawai'i.
- Maguire, K. M., Banks, N. H. and Opara, L. U. (2001). Factors affecting weight loss of apples. *Horticultural Reviews*, 25, 197-234.
- Manso, T. and Nunes, C. (2011). *Metschnikowia andauensis* as a new biocontrol agent of fruit postharvest diseases. *Postharvest Biology and Technology*, 61, 64-71.
- Marquina, D., Santos, A. and Peinado, J. M. (2002). Biology of killer yeasts. *International Microbiology*, 5, 65-71.
- Mastrangelo, M. M., Rojas, A. M., Castro, M. A., Gerschenson, L. N. and Alzamora, S. M. (2000). Texture and structure of glucose-infused melon. *Journal of the Science of Food and Agriculture*, 80, 769-776.
- Matića, S., Spadaro, D., Garibaldia, A. and Gullinoa, M. L. (2014). Antagonistic yeasts and thermoherapy as seed treatments to control *Fusarium fujikuroi* on rice. *Biological Control*, 73, 59-67.
- McDonald, S., Prenzler, P. D., Antolovich, M. and Robards, K. (2001). Phenolic content and antioxidant activity of olive extracts. *Food Chemistry*, 73, 73-84.
- McGrath, M. T. (2004). What are Fungicides? Retrieved 22/10/2016, from <http://www.apsnet.org/edcenter/intropp/topics/pages/fungicides.aspx>
- McGuire, R. G. (1992). Reporting of objective colour measurements. *HortScience*, 27(12), 1254-1255.
- McLaughlin, R. J., Wilson, C. L., Droby, S., Ben-Arie, R. and Chalutz, E. (1992). Biological control of postharvest diseases of grape, peach and apple with the yeasts *Kloeckera apiculata* and *Candida guilliermondii*. *Plant Disease*, 76, 470-473.

- McLaughlin, R. J., Wisniewski, M. E., Wilson, C. L. and Chalutz, E. (1990). Effect of inoculum concentration and salt solutions on biocontrol of postharvest diseases of apple with *Candida* sp. *Phytopathology*, 80, 456-461.
- Meng, X. H., Qin, G. Z. and Tian, S. P. (2010). Influences of preharvest spraying *Cryptococcus laurentii* combined with postharvest chitosan coating on postharvest diseases and quality of table grapes in storage. *LWT-Food Science and Technology*, 43(596-601).
- Mercier, J. and Lindow, S. E. (2001). Field Performance of Antagonistic Bacteria Identified in a Novel Laboratory Assay for Biological Control of Fire Blight of Pear. *Biological Control*, 22(1), 66-71.
- Mercier, J. and Wilson, C. (1994). Colonization of apple wounds by naturally occurring microflora and introduced *Candida oleophila* and their effect on infection by *Botrytis cinerea* during storage. *Biological Control*, 4(138-144).
- Meshram, A. and Srivastava, N. (2013). An Insight into the Molecular Structure and Function of Polygalacturonase Inhibiting Protein (PGIP). *Journal of Proteins and Proteomics*, 4(3), 175-181.
- Mignani, I., Greve, L. C., Ben-Arie, R., Stotz, H. U., Li, C., Shackel, K. A. and Labavitch, J. M. (1995). The effects of GA3 and divalent cations on aspects of pectin metabolism and tissue softening in ripening tomato pericarp. *Physiologia Plantarum*, 93, 108-115.
- Mills, A. A. S., (Bud) Platt, H. W. and Hurta, R. A. R. (2006). Sensitivity of *Erwinia* spp. to salt compounds in vitro and their effect on the development of soft rot in the potato tubers in storage. *Postharvest Biology and Technology*, 41, 208-214.
- Mitcham, E. J. and McDonald, R. E. (1992). Effect of high temperature on cell wall modifications associated with tomato fruit ripening. *Postharvest Biology and Technology*, 1, 257-264.
- MOA. (2002). Pelan Pemasaran Komoditi Betik 2003-2010. Retrieved 16 July 2016, from http://www.moa.gov.my/c/document_library/get_file?uuid=3ae77141-1dc9-412c-b015-fd9f88785e21&groupId=41803
- MOA. (2015). *Agrofood Statistic 2014*: Information Management and Statistics Section Policy and Strategic Planning Division, Ministry of Agriculture, Malaysia.
- Morton, J. J. and Miami, F. L. (1987). *Fruits of warm climates*. CRC Incorporation, Winterville, North Carolina.
- Muccilli, S., Wemhoff, S., Restuccia, C. and Meinhardt, F. (2013). Exoglucanase-encoding genes from three *Wickerhamomyces anomalus* killer strains isolated from olive brine. *Yeast*, 30, 33-43.

- Musa, K. H., Abdullah, A., Jusoh, K. and Subramaniam, V. (2011). Antioxidant activity of pink-flesh guava (*Psidium guajava* L.): Effect of extraction techniques and solvents *Food Analysis Methods*, 4(1), 100-107.
- Nahass, G. T., Rosenberg, S. P. and Leonardi, C. L. (1993). Disseminated Infection With *Trichosporon beigelii*: Report of a Case and Review of the Cutaneous and Histologic Manifestations. *Archives Dermatology*, 129(8), 1020-1023.
- Nakasone, H. Y. and Paull, R. E. (1998). Tropical Fruits, Vol. 7 of Crop Production Science in Horticulture. CAB International, The University of Wisconsin, Madison, WI, USA.
- Nantawanit, N., Chanchaichaovivat, A., Panijpan, B. and Ruenwongsa, P. (2010). Induction of defense response against *Colletotrichum capsici* in chili fruit by the yeast *Pichia guilliermondii* strain R13. *Biological Control*, 52, 145-152.
- Narayansamy, P. (2006). Postharvest Pathogens and Disease Management. John Wiley & Sons, United States of America.
- Narayansamy, P. (2013). Biological Management of Diseases of Crops. Springer, New York.
- NAS. (1987). Report of the Research Briefing Panel on Biological Control in Managed Ecosystems. National Academy Press, Washington.
- Nghia, N. A., Kadir, J., Sunderasan, E., Abdullah, M. P., Malik, A. and Napis, S. (2008). Morphological and Inter Simple Sequence Repeat (ISSR) Markers Analyses of *Corynespora cassiicola* Isolates from Rubber Plantations in Malaysia. *Mycopathologia*, 166(4), 189-201.
- Nobile, C. J. and Mitchell, A. P. (2005). Regulation of cell-surface genes and biofilm formation by the *C. albicans* transcription factor Bcr1p. *Current Biology*, 15, 1150-1155.
- Nucci, M. and Anaissie, E. (2001). Revisiting the source of candidemia: skin or gut? *Clinical Infectious Diseases*, 33, 1959-1967.
- Nunes, C., Manso, T. and Lima-Costa, M. E. (2009). Postharvest biological control of citrus fruit. *Tree and Forestry Science and Biotechnology*, 3, 116-126.
- O'Hare, T. J. and Williams, D. J. (2014). Papaya as a medicinal plant. Springer, New York.
- Obagwu, J. and Korsten, L. (2003). Integrated control of citrus green and blue molds using *Bacillus subtilis* in combination with sodium bicarbonate or hot water. *Postharvest Biology and Technology*, 28, 187-194.
- Ong, M. K. and Ali, A. (2015). Antifungal action of ozone against *Colletotrichum gloeosporioides* and control of papaya anthracnose. *Postharvest Biology and Technology*, 100, 113-119.

- Oregel-Zamudio, E., Angoa-Pérez, M. V., Oyoque-Salcedo, G., Aguilar-González, C. N. and Mena-Violante, H. G. (2017). Effect of candelilla wax edible coatings combined with biocontrol bacteria on strawberry quality during the shelf-life. *Scientia Horticulturae*, 214, 273-279.
- Ortu, G., Demontis, M. A., Budroni, M., Goyard, S., d'Enfert, C. and Migheli, Q. (2005). Study of biofilm formation in *Candida albicans* may help understanding the biocontrol capability of a flor strain of *Saccharomyces cerevisiae* against the phytopathogenic fungus *Penicillium expansum*. *Journal of Plant Pathology*, 87, 300.
- Palou, L., Smilanick, J. L., Usall, J. and Vinas, I. (2001). Control of postharvest blue and green molds of oranges by hot water, sodium carbonate and sodium bicarbonate. *Plant Disease*, 85, 371-376.
- Parafati, L., Vitale, A., Restuccia, C. and Cirvilleri, G. (2015). Biocontrol ability and action mechanism of food-isolated yeast strains against *Botrytis cinerea* causing post-harvest bunch rot of table grape. *Food Microbiology*, 47, 85-92.
- Paull, R. E. (1999). Effect of temperature and relative humidity on fresh commodity quality. *Postharvest Biology and Technology*, 15, 263-277.
- Paull, R. E. and Chen, N. J. (1989). Waxing and plastic wraps influence water loss from papaya fruit during storage and ripening. *Journal of the American Society for Horticultural Science*, 114, 937-942.
- Paull, R. E., Nishijima, W., Reyes, M. and Cavaletto, C. (1997). Postharvest handling and losses during marketing of papaya (*Carica papaya* L.). *Postharvest Biology and Technology*, 11, 165-179.
- Pereira, T., Almeida, P. S. G. d., Azevedo, I. G. d., Cunha, M. d., Oliveira, J. G. d., Silva, M. G. d. and Vargas, H. (2009). Gas diffusion in 'Golden' papaya fruit at different maturity stages. *Postharvest Biology and Technology*, 54, 123-130.
- Pfaller, M. A. and Diekema, D. J. (2007). Epidemiology of invasive candidiasis: a persistent public health problem. *Clinical Microbiology Reviews*, 20, 133-163.
- Picchioni, G. A., Watada, A. E., Conway, W. S., Whitaker, B. D. and Sams, C. E. (1995). Phospholipid, galactolipid and steryl lipid composition of apple fruit cortical tissue following postharvest CaCl₂ infiltration. *Phytochemistry*, 39, 763-769.
- Poovaiah, B. W. (1986). Role of calcium in prolonging storage life of fruits and vegetables. *Food Technology*, 40, 86-88.
- Poovaiah, B. W. and Reddy, A. S. N. (1993). Calcium and signal transduction in plants. *Critical Reviews in Plant Sciences*, 12, 185-211.

- Pratella, G. C. and Mari, M. (1993). Effectiveness of *Trichoderma*, *Gliocladium* and *Paecilomyces* in postharvest fruit protection. *Postharvest Biology and Technology*, 3, 49-56.
- Prior, C., Elango, F. and Whitewell, A. (1992). Chemical control of *Colletotrichum* infection in mangoes. In *Colletotrichum: Biology, Pathology and Control*. CABI, Wallingford, United Kingdom.
- Qin, G. Z. and Tian, S. P. (2005). Enhancement of biocontrol activity of *Cryptococcus laurentii* by silicon and the possible mechanisms involved. *Phytopathology*, 95, 69-75.
- Qin, G. Z., Tian, S. P., Xu, Y. and Wan, Y. K. (2003). Enhancement of biocontrol efficacy of antagonistic yeasts by salicylic acid in sweet cherry fruit. *Physiological and Molecular Plant Pathology*, 62, 147-154.
- Qing, F. and Shiping, T. (2000). Postharvest biological control of *Rhizopus* rot of nectarine fruits by *Pichia membranefaciens*. *Plant Disease*, 84(11), 1212-1216.
- Rahman, M. A. (2008). Postharvest Management of Anthracnose on Quality of Papaya (*Carica Papaya* L.) Using Antagonistic Bacteria. (Doctor of Philosophy), Universiti Putra Malaysia, Malaysia.
- Rahman, M. A., Kadir, J., Mahmud, T. M. M., Abdul Rahman, R. and Begum, M. M. (2007). Screening of Antagonistic Bacteria for Biocontrol Activities on *Colletotrichum gloeosporioides* in Papaya. *Asian Journal of Plant Sciences*, 6(1), 12-20.
- Rahman, M. A., Mahmud, T. M. M., Kadir, J., Rahman, R. A. and Begum, M. M. (2009). Enhancing the Efficacy of *Burkholderia cepacia* B23 with Calcium Chloride and Chitosan to Control Anthracnose of Papaya During Storage. *The Plant Pathology Journal*, 25(4), 361-368.
- Rahman, M. S. (2007). Surface treatment and edible coatings in food preservation. In, *Handbook of Food Preservation, Second Edition*, ed. Baldwin, E. A. CRC Press, United State of America.
- Ramezani, A., Rahemi, M., Maftoun, M., Bahman, K., Eshghi, S., Safizadeh, M. R. and Tavallali, V. (2010). The ameliorative effects of spermidine and calcium chloride on chilling injury in pomegranate fruits after long-term storage. *Fruits*, 65, 169-178.
- Ramiro, D. A., Guerreiro-Filho, O. and Mazzafera, P. (2006). Phenol contents, oxidase activities and the resistance of coffee to the leaf miner *Leucoptera coffeella*. *Journal of Chemical Ecology*, 32, 1977-1988.
- Ranganna, S. (1997). *Handbook of Analysis and Quality Control for Fruit and Vegetable Products*. Tata McGraw-Hill, New Delhi, India.

- Robards, K., Prenzler, P. D., Tucker, G., Swatsitang, P. and Glover, W. (1999). Phenolic compounds and their role in oxidative processes in fruits. *Food Chemistry*, 66(4), 401-436.
- Robiglio, A., Sosa, M. C., Lutz, M. C., Lopes, C. A. and Sangorrín, M. P. (2011). Yeast biocontrol of fungal spoilage of pears stored at low temperature. *International Journal of Food Microbiology*, 147, 211-216.
- Roman, A. D. E., Salvaña, E. M. T., Guzman-Peñamora, M. A. J. D., Roxas, E. A., Leyritana, K. T. and Saniel, M. C. (2014). Invasive trichosporonosis in an AIDS patient: case report and review of the literature. *International Journal of STD and AIDS*, 25, 70-75.
- Ryals, J. A., Neuenschwander, U. H., Willits, M. G., Molina, A., Steiner, H. Y. and Hunt, M. D. (1996). Systemic acquired resistance. *The Plant Cell*, 8(10), 1809.
- Saftner, R. A., Conway, W. S. and Sams, C. E. (1998). Effects of postharvest calcium and fruit coating treatments on postharvest life, quality maintenance and fruit-surface injury in 'Golden Delicious' apples. *Journal of the American Society for Horticultural Science*, 294-298.
- Saligkarias, I. D., Gravanis, F. T. and Epton, H. A. S. (2002). Biological control of *Botrytis cinerea* on tomato plants by the use of epiphytic yeasts *Candida guilliermondii* strains 101 and US 7 and *Candida oleophila* strain I-182: II. A study on mode of action. *Biological Control*, 25, 151-161.
- Sangwanich, S., Sangchote, S. and Leelasuphakul, W. (2013). Biocontrol of Citrus green mould and postharvest quality parameters. *International Food Research Journal*, 20(6), 3381-3386.
- Santi, R., Bhowmilk, S. R. and Jung, C. P. (1992). Shelf life of mature green tomatoes stored in controlled atmosphere and high humidity. *Journal Food of Science*, 57, 948-951.
- Santos, A., San Mauro, M., Bravo, E. and Marquina, D. (2009). PMKT2, a new killer toxin from *Pichia membranifaciens* and its promising biotechnological properties for control of the spoilage yeast *Brettanomyces bruxellensis*. *Microbiology*, 155, 624-634.
- Sariah, M. (1994). Potential of *Bacillus* spp. as a biocontrol agent for anthracnose fruit rot of chilli. *Malaysian Applied Biology*, 23(1&2), 53-60.
- Schena, L., Ippolito, A., Zehavi, T., Cohen, L., Nigro, F. and Droby, S. (1999). Genetic diversity and biocontrol activity of *Aureobasidium pullulans* isolates against postharvest rots. *Postharvest Biology and Technology*, 17, 189-199.
- Schena, L., Nigro, F., Pentimone, I. A. and Ippolito, A. (2003). Control of postharvest rots of sweet cherries and table grapes with endophytic isolates

of *Aureobasidium pullulans*. *Postharvest Biology and Technology*, 30(3), 209-220.

Scherm, B., Ortu, G., Muzzu, A., Budroni, M., Arras, G. and Migheli, Q. (2003). Biocontrol activity of antagonistic yeasts against *Penicillium expansum* on apple. *Journal of Plant Pathology*, 85, 205-213.

Schmitt, M. J. and Breinig, F. (2006). Yeast viral killer toxins: lethality and self-protection. *Natural Review of Microbiology*, 4, 212-221.

Selitreffnikoff, C. P. (2001). Antifungal proteins. *Applied and Environmental Microbiology*, 67, 2883-2894.

Shahnazi, S. (2012). Biological Characterization and Genetic Diversity of *Fusarium* spp. Associated with Yellowing Disease in Black Pepper (*Piper nigrum* L.) in Malaysia. (Doctor of Philosophy), Universiti Putra Malaysia, Malaysia.

Sharma, N. (2014). Biological Controls for Preventing Food Deterioration. Wiley Blackwell, United Kingdom.

Sharma, R. R., Singh, D. and Singh, R. (2009). Biological control of postharvest diseases of fruits and vegetables by microbial antagonists: A review. *Biological Control*, 50, 205-221.

Shewfelt, R. L. (1986). Postharvest treatment for extending the shelf life of fruits and vegetables. *Food Technology*, 40(70-80).

Shi, J., Liu, A., Li, X. and Chen, W. (2011). Control of *Phytophthora nicotianae* disease, induction of defense responses and genes expression of papaya fruits treated with *Pseudomonas putida* MGP1. *Journal Science of Food and Agriculture*, 93, 568-574.

Singh, A. (1980). *Fruit Physiology and Production*. Kalyani Publisher, New Delhi

Singh, V. and Deverall, B. J. (1984). *Bacillus subtilis* as a control agent against fungal pathogens of citrus fruit. *Transactions in British Mycological Society*, 83, 487-490.

Snowdon, A. L. (1990). *Color Atlas of Post-harvest Diseases and Disorders of Fruit and Vegetables* (Vol. 1): CRC Press Inc., Boca Raton, Florida

Sobiczewski, P., Bryk, H. and Berezynski, S. (1996). Evaluation of epiphytic bacteria isolated from apple leaves in the control of postharvest diseases. *Journal of Fruit and Ornamental Plant Research*, 4, 35-45.

Spadaro, D. and Droby, S. (2016). Development of biocontrol products for postharvest diseases of fruit: The importance of elucidating the mechanisms of action of yeast antagonists. *Trends in Food Science & Technology*, 47, 39-49.

- Stintzi, A., Heitz, T., Prasad, V., Wiedemann-Merdinoglu, S., Kauffmann, S., Geoffroy, P., Legrand, M. and Fritig, B. (1993). Plant 'pathogenesis-related' proteins and their role in defense against pathogens. *Biochimie*, 75(8), 687-706.
- Sugar, D. and Spotts, R. A. (1999). Control of postharvest decay in pear by four laboratory-grown yeasts and two-registered biocontrol products. . *Plant Disease*, 83, 155-158.
- Sugita, T., Nishikawa, A., Shinoda, T. and Kume, H. (1995). Taxonomic position of deep-seated, mucosa-associated, and superficial isolates of *Trichosporon cutaneum* from trichosporonosis patients. *Journal of Clinical Microbiology*, 33, 1368–1370.
- Sulaiman, R. (2011). Aethiology of stem canker pathogen in *Jatropha curcas* L. in Peninsular Malaysia. (Master of Science), Universiti Putra Malaysia, Malaysia.
- Tamura, K., Stecher, G., Peterson, D., Filipski, A. and Kumar, S. (2013). MEGA6: Molecular Evolutionary Genetics Analysis Version 6.0. *Molecular Biology and Evolution*, 30(12), 2725-2729.
- Teixido, N., Torres, R., Viñas, I., Abadias, M. and Usall, J. (2011). Postharvest Biology and Technology of Tropical and Subtropical Fruits: Fundamental Issues. Woodhead Publishing Limited.
- Teixido, N., Usall, J. and Vinas, I. (1999). Efficacy of preharvest and postharvest *Candida sake* biocontrol treatments to prevent blue mold on apples during storage. *International Journal of Food Microbiology*, 50, 203-210.
- Teixido, N., Viñas, I., Usall, J. and Magan, N. (1998). Improving ecological fitness and environmental stress tolerance of the biocontrol yeast *Candida sake* by manipulation of intracellular sugar alcohol and sugar content. *Mycological Research*, 102, 1409-1417.
- Terry, L. A. and Joyce, D. C. (2004). Elicitors of induced disease resistance in postharvest horticultural crops: a brief review. *Postharvest Biology and Technology*, 32, 1-13.
- Tian, S. P., Fan, Q., Xu, Y. and Jiang, A. L. (2002a). Effects of calcium on biocontrol activity of yeast antagonists against the postharvest fungal pathogen *Rhizopus stolonifer*. *Plant Pathology*, 51, 352-358.
- Tian, S. P., Xu, Y., Jiang, A. L. and Gong, Q. Q. (2002b). Physiological and quality responses of longan fruit to high O₂ or high CO₂ atmospheres in storage. *Postharvest Biology and Technology*, 24, 335-340.
- Tronsmo, A. and Dennis, C. (1977). The use of *Trichoderma* species to control strawberry fruit rots. *Netherland Journal of Plant Pathology*, 83, 449-455.

- Tsantili, E., Rouskas, D., Christopoulos, M. V., Stanidis, V., Akrivos, J. and Papanikolaou, D. (2007). Effects of two pre-harvest calcium treatments on physiological and quality parameters in 'Vogue' cherries during storage. *Journal of Horticultural Science and Biotechnology*, 82, 657-663.
- Tuzun, S. and Kloepper, J. (1995). Practical application and implementation of induced resistance. In *Induced Resistance to Disease in Plants*, ed. Hammerschmidt, R. and Kluwer, J. K. Academic Publisher, Dordrecht.
- Usall, J., Smilanick, J., Palou, L., Denis-Arrue, N., Teixido, N., Torres, R. and Vinas, I. (2008). Preventive and curative activity of combined treatments of sodium carbonates and *Pantoea agglomerans* CPA-2 to control postharvest green mold of citrus fruit. *Postharvest Biology and Technology*, 50, 1-7.
- Venkatachalam, K. and Meenune, M. (2012). Changes in physiochemical quality and browning related enzyme activity of longkong fruit during four different weeks of on-tree maturation. *Food Chemistry*, 131, 1437-1442.
- Vero, S., Mondino, P., Burgueno, J., Soubes, M. and Wisniewski, M. (2002). Characterization of biocontrol activity of two yeast strains from Uruguay against blue mold of apple. *Postharvest Biology and Technology*, 26, 91-98.
- Vinas, I., Usall, J., Teixido, N. and Sanchis, V. (1998). Biological control of major postharvest pathogens on apple with *Candida sake*. *International Journal of Food Microbiology*, 40, 9-16.
- Waage, J., Gurr, G. and Wratten, S. (2012). *Biological Control: Measures of Success*: Springer Science and Business Media.
- Walling, L. L. (2001). Induced resistance: from the basic to the applied. *Trends in Plant Science*, 6, 445-447.
- Walsh, T. J., Melcher, G. P., Lee, J. W. and Pizzo, P. A. (1993). Infections due to *Trichosporon* species: new concepts in mycology, pathogenesis, diagnosis and treatment. *Current Topics in Medical Mycology*, 5, 79-113.
- Wan, Y. K., Tian, S. P. and Qin, G. Z. (2003). Enhancement of biocontrol activity of yeasts by adding sodium bicarbonate or ammonium molybdate to control postharvest disease of jujube fruits. *Letters in Applied Microbiology*, 37, 249-253.
- Wang, X., Xu, F., Wang, J., Jin, P. and Zheng, Y. (2013). *Bacillus cereus* AR156 induces resistance against *Rhizopus* rot through priming of defense responses in peach fruit. *Food Chemistry*, 136(2), 400-406.
- Wang, Y., Xie, X. and Long, L. E. (2014). The effect of postharvest calcium application in hydro-cooling water on tissue calcium content, biochemical changes, and quality attributes of sweet cherry fruit. *Food Chemistry*, 160(0), 22-30. doi: <http://dx.doi.org/10.1016/j.foodchem.2014.03.073>

- Wang, Y., Yu, T., Li, Y., Cai, D., Liu, X., Lu, H. and Zheng, X. D. (2009). Postharvest biocontrol of *Alternaria alternata* in Chinese winter jujube by *Rhodosporidium paludigenum*. *Journal of Applied Microbiology*, 107, 1492-1498.
- Wang, Y. F., Bao, Y. H., Shen, D. H., Feng, W., Yu, T., Zhang, J. and Zheng, X. D. (2008). Biocontrol of *Alternaria alternata* on cherry tomato fruit by use of marine yeast *Rhodosporidium paludigenum* Fell & Tallman. *International Journal of Food Microbiology*, 123, 234-239.
- Wang, Y. S., Tian, S. P., Xu, Y., Qin, G. Z. and Yao, H. (2004). Changes in the activities of pro- and anti-oxidant enzymes in peach fruit inoculated with *Cryptococcus laurentii* or *Penicillium expansum* at 0 or 20 °C. *Postharvest Biology and Technology*, 34, 21-28.
- War, A. R., Paulraj, M. G., Ahmad, T., Buhroo, A. A., Hussain, B., Ignacimuthu, S. and Sharma, H. C. (2012). Mechanisms of plant defense against insect herbivores. *Plant Signaling and Behaviour*, 7(10), 1306-1320.
- Wheeler, M. (2013). Pesticides and Parkinson's: UCLA researchers uncover further proof of a link. 15 August 2015, from <http://newsroom.ucla.edu/releases/pesticides-and-parkinson-s-more-242364>
- Whitaker, J. R. (1995). Polyphenol oxidase. In *Food Enzymes Structure and Mechanism*, ed. Wong D.W.S. Chapman and Hall, New York, NY
- White, T. J., Bruns, T., Lee, S. and Taylor, J. (1990). Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In *PCR protocols: a guide to methods and applications*, ed. Innis, M.A., Gelfand, D. H., Snisky, J. J. and White, T.J. Academic Press. San Diego.
- Wills, R. B. H., McGlasson, W. B., Graham, D. and D.C., J. (2007). Postharvest: An introduction to the physiology and handling of fruit, vegetables and ornamentals. CABI, Wallingford.
- Wilson, C. L. and Wisniewski, M. E. (1989). Biological control of postharvest diseases of fruit and vegetables: an emerging technology. *Annual Review of Phytopathology*, 27, 425-441.
- Wilson, C. L. and Wisniewski, M. E. (1994). Biological Control of Postharvest Diseases: Theory and Practice. CRC Press, Boca Raton, Florida.
- Wilson, C. L. and Wisniewski, M. E. (1998). Biological Control of Postharvest Diseases of Fruits and Vegetables-Theory and Practice. CRC Press, Boca Raton, Florida.
- Wilson, C. L., Wisniewski, M. E., Droby, E. and Chalutz, E. (1993). A selection strategy for microbial antagonists to control postharvest diseases of fruit and vegetables. *Scientia Horticulturae*, 53, 183-189.

- Wisniewski, M., Biles, C., Droby, S., McLaughlin, R., Wilson, C. and Chalutz, E. (1991). Mode of action of the postharvest biocontrol yeast, *Pichia guilliermondii*. I. Characterization of attachment to *Botrytis cinerea*. *Physiological and Molecular Plant Pathology*, 39, 245-258.
- Wisniewski, M., Wilson, C., Droby, S., Chalutz, E., El Ghaouth, A. and Stevens, C. (2007). Postharvest biocontrol: new concepts and applications. In *Biological Control: A Global Perspective*, ed. Vincent, C., Goettal, M. S. and Lazarovits, G. CABI, Cambridge.
- Wisniewski, M., Wilson, C. and Hershberger, W. (1989). Characterization of inhibition of *Rhizopus stolonifer* germination and growth by *Enterobacter cloacae*. *Canadian Journal of Botany*, 67, 2317-2323.
- Wisniewski, M. E., Droby, S., Chalutz, E. and Elam, Y. (1995). Effects of Ca²⁺ and Mg²⁺ on *Botrytis cinerea* and *Penicillium expansum* *in vitro* and on the biocontrol activity of *Candida oleophila*. *Plant Pathology*, 44, 1016-1024.
- Wittig, H. P. P., Johnson, K. B. and Pscheidt, J. W. (1997). Effect of epiphytic fungi on brown rot, blossom blight and latent infections in sweet cherry. *Plant Disease*, 81, 383-387.
- Xu, X. B. and Tian, S. P. (2008). Reducing oxidative stress in sweet cherry fruit by *Pichia membranaefaciens*: a possible mode of action against *Penicillium expansum*. *Journal of Applied Microbiology*, 105, 1170-1177.
- Yahia, E. M. (2011). *Postharvest Biology and Technology of Tropical and Subtropical Fruits*. Woodhead Publishing Limited, United Kingdom.
- Yamauchi, N. and Watada, A. E. (1991). Regulated chlorophyll degradation in spinach leaves during storage. *Journal of the American Society for Horticultural Science*, 116(1), 58-62.
- Yang, B., Yongcai, L., Yonghong, G. and Yi, W. (2010). Postharvest Pathology, vol. 2. In *Induced resistance in melons by elicitors for the control of postharvest diseases*, ed. Prusky, D. and Gullino, M.L. Springer, Netherlands.
- Yang, X., Pang, X., Xu, L., Fang, R., Huang, X., Guan, P., Lu, W. and Zhang, Z. (2009). Accumulation of soluble sugars in peel at high temperature leads to stay-green ripe banana fruit. *Journal of Experimental Botany*, 60, 4051-4062.
- Yao, H. and Tian, S. (2005a). Effects of pre- and post-harvest application of salicylic acid or methyl jasmonate on inducing disease resistance of sweet cherry fruit in storage. *Postharvest Biology and Technology*, 35, 253-262.
- Yao, H. J. and Tian, S. P. (2005b). Effects of a biocontrol agent and methyl jasmonate on postharvest diseases of peach fruit and the possible mechanisms involved. *Journal of Applied Microbiology*, 98, 941-950.

- Yao, H. J., Tian, S. P. and Wang, Y. S. (2004). Sodium bicarbonate enhances biocontrol efficacy of yeasts on fungal spoilage of pears. *International Journal of Food Microbiology*, 93, 297-304.
- Yon, R. M. (1994). Papaya: Fruit Development, Postharvest Physiology, Handling and Marketing in ASEAN. ASEAN Food Handling Bureau, Malaysia.
- Yu, T., Chen, J. S., Chen, R. L., Huang, B., Liu, D. H. and Zheng, X. D. (2007). Biocontrol of blue and gray mold diseases of pear fruit by integration of antagonistic yeast with salicylic acid. *International Journal of Food Microbiology*, 116, 339-345.
- Yu, T., Li, H. Y. and Zheng, X. D. (2007). Synergistic effect of chitosan and *Cryptococcus laurentii* on inhibition of *Penicillium expansum* infections. *International Journal of Food Microbiology*, 114, 261-266.
- Yu, T., Yu, C., Chen, F., Sheng, K., Zhou, T., Zunun, M., Abudu, O., Yang, S. and Zheng, X. (2012). Integrated control of blue mold in pear fruit by combined application of chitosan, a biocontrol yeast and calcium chloride. *Postharvest Biology and Technology*, 69, 49-53.
- Yu, T., Yu, C., Lu, H., Zunun, M., Chen, F., Zhou, T., Sheng, K. and Zheng, X. (2012). Effect of *Cryptococcus laurentii* and calcium chloride on control of *Penicillium expansum* and *Botrytis cinerea* infections in pear fruit. *Biological Control*, 61, 169-175.
- Yu, T. and Zheng, X. D. (2006). Salicylic acid enhances biocontrol efficacy of the antagonist *Cryptococcus laurentii* in apple fruit. *Journal of Plant Growth Regulation*, 25, 166-174.
- Zhang, D., Spadaro, D., Garibaldi, A. and Gullino, M. L. (2010). Selection and evaluation of new antagonists for their efficacy against postharvest brown rot of peaches. *Postharvest Biology and Technology*, 55, 174-181.
- Zhang, D., Spadaro, D., Garibaldi, A. and Gullino, M. L. (2011). Potential biocontrol activity of a strain of *Pichia guilliermondii* against grey mold of apples and its possible modes of action. *Biological Control*, 57, 193-201.
- Zhang, D., Spadaro, D., Valente, S., Garibaldi, A. and Gullino, M. L. (2012). Cloning, characterization, expression and antifungal activity of an alkaline serine protease of *Aureobasidium pullulans* PL5 involved in the biological control of postharvest pathogens. *International Journal of Food Microbiology*, 153, 453-464.
- Zhang, H., Ma, L., Turner, M., Xu, H., Dong, Y. and Jiang, S. (2009). Methyl jasmonate enhances biocontrol efficacy of *Rhodotorula glutinis* to postharvest blue mold decay of pears. *Food Chemistry*, 117, 621-626.
- Zhang, H., Ma, L., Turner, M., Xu, H., Zheng, X., Dong, Y. and Jiang, S. (2010). Salicylic acid enhances biocontrol efficacy of *Rhodotorula glutinis* against

postharvest *Rhizopus* rot of strawberries and the possible mechanisms involved. *Food Chemistry*, 122, 577-583.

Zhang, H., Zheng, X., Fu, C. and Xi, Y. (2005). Postharvest biological control of gray mold rot of pear with *Cryptococcus laurentii*. *Postharvest Biology and Technology*, 3, 79-86.

Zhang, H. Y., Zheng, X. D. and Su, D. M. (2006). Postharvest control of blue mold rot of pear by microwave treatment and *Cryptococcus laurentii*. *Journal of Food Engineering*, 77, 539-544.

Zhang, H. Y., Zheng, X. D., Wang, L., Li, S. S. and Liu, R. F. (2007). Effect of yeast antagonist in combination with hot water dips on postharvest *Rhizopus* rot of strawberries. *Journal of Food Engineering*, 78, 281-287.

Zhang, L. X. and Paull, R. E. (1990). Ripening behaviour of papaya genotypes. *HortScience*, 25, 454 - 455.

Zhao, Y., Tu, K., Shao, X., Jing, W. and Su, Z. (2008). Effects of the yeast *Pichia guilliermondii* against *Rhizopus nigricans* on tomato fruit. *Postharvest Biology and Technology*, 49(1), 113-120.

Zheng, M., Shi, J., Shi, J., Wang, Q. and Li, Y. (2013). Antimicrobial effects of volatiles produced by two antagonistic *Bacillus* strains on the anthracnose pathogen in postharvest mangos. *Biological Control*, 65(2), 200-206.

Zheng, X. D., Yu, T., Chen, R. L., Huang, B. and Wu, V. C. H. (2007). Inhibiting *Penicillium expansum* infection on pear fruit by *Cryptococcus laurentii* and cytokinin. *Postharvest Biology and Technology*, 45, 221-227.

Zheng, X. D., Zhang, H. Y. and Sun, P. (2005). Biological control of postharvest green mold decay of oranges by *Rhodotorula glutinis*. *European Food Research and Technology*, 220, 353-357.

Zheng, Y., Sheng, J., Zhao, R., Zhang, J., Lv, S., Liu, L. and Shen, L. (2011). Preharvest L-arginine treatment induced postharvest disease resistance to *Botrytis cinerea* in tomato fruits. *Journal of Agricultural and Food Chemistry*, 59, 6543-6549.